

British Pharmacopoeia 2016

Volume II

British Pharmacopoeia 2016

Volume II

The British Pharmacopoeia Commission has caused this British Pharmacopoeia 2016 to be prepared under regulation 317(1) of the Human Medicines Regulations 2012 and, in accordance with regulation 317(4), the Ministers have arranged for it to be published. It has been notified in draft to the European Commission in accordance with Directive 98/34/EEC.

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See General Notices

Effective date: 1 January 2016

see Notices

London: The Stationery Office

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THE DEPARTMENT OF HEALTH

In respect of Northern Ireland:

THE DEPARTMENT OF HEALTH, SOCIAL SERVICES AND
PUBLIC SAFETY

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First Published 2015

ISBN 978 011 3230 006

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Patents

In this Pharmacopoeia certain drugs and preparations have been included notwithstanding the existence of actual or potential patent rights. In so far as such substances are protected by Letters Patent their inclusion in this Pharmacopoeia neither conveys, nor implies, licence to manufacture.

Effective dates

New and revised monographs of national origin enter into force on 1 January 2016. The monographs are brought into effect under regulation 320(2) of the Human Medicines Regulations 2012.

Monographs of the European Pharmacopoeia have previously been published by the European Directorate for the Quality of Medicines & HealthCare in accordance with the Convention on the Elaboration of a European Pharmacopoeia and have been brought into effect under European Directives 2001/82/EC, 2001/83/EC and 2003/63/EC, as amended, on medicines for human and veterinary use.

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General Notices

Part I

The British Pharmacopoeia comprises the entire text within this publication. The word 'official' is used in the Pharmacopoeia to signify 'of the Pharmacopoeia'. It applies to any title, substance, preparation, method or statement included in the general notices, monographs and appendices of the Pharmacopoeia. The abbreviation for British Pharmacopoeia is BP.

Monographs of the European Pharmacopoeia are reproduced in this edition of the British Pharmacopoeia by incorporation of the text published under the direction of the Council of Europe (Partial Agreement) in accordance with the Convention on the Elaboration of a European Pharmacopoeia (Treaty Series No. 32 (1974) CMND 5763) as amended by the Protocol to the Convention (Treaty Series No. MISC16 (1990) CMND 1133). They are included for the convenience of users of the British Pharmacopoeia. In cases of doubt or dispute reference should be made to the Council of Europe text.



Monographs of the European Pharmacopoeia are distinguished by a chapter of stars against the title and by reference to the European Pharmacopoeia monograph number included immediately below the title in italics. The beginning and end of text from the European Pharmacopoeia are denoted by means of horizontal lines with the symbol 'Ph Eur' ranged left and right, respectively. The general provisions of the European Pharmacopoeia relating to different types of dosage form are included in the appropriate general monograph in that section of the British Pharmacopoeia entitled Monographs: Formulated Preparations. These general provisions apply to all dosage forms of the type defined, whether or not an individual monograph is included in the British Pharmacopoeia. In addition, the provisions of the European Pharmacopoeia General Monograph for Pharmaceutical Preparations apply to all dosage forms, whether or not an individual monograph is included in the British Pharmacopoeia. Texts of the European Pharmacopoeia are governed by the General Notices of the European Pharmacopoeia. These are reproduced as Part III of these notices.

Part II

The following general notices apply to the statements made in the monographs of the British Pharmacopoeia other than those reproduced from the European Pharmacopoeia and to the statements made in the Appendices of the British Pharmacopoeia other than a method, test or other matter described in an appendix is invoked in a monograph reproduced from the European Pharmacopoeia.

Official Standards

The requirements stated in the monographs of the Pharmacopoeia apply to articles that are intended for medicinal use but not necessarily to articles that may be sold under the same name for other purposes. An article intended for medicinal use that is described by means of an official title must comply with the requirements of the relevant monograph. A formulated preparation must comply throughout its assigned shelf-life (period of validity). The subject of any other monograph must comply throughout its period of use.

A monograph is to be construed in accordance with any general monograph or notice or any appendix, note or other explanatory material that is contained in this edition and that is applicable to that monograph. All statements contained in the monographs, except where a specific general notice indicates otherwise and with the exceptions given below, constitute standards for the official articles. An article is not of pharmacopoeial quality unless it complies with all of the requirements stated. This does not imply that a manufacturer is obliged to perform all the tests in a monograph in order to assess compliance with the Pharmacopoeia before release of a product. The manufacturer may assure himself that a product is of pharmacopoeial quality by other means, for example, from data derived from validation studies of the manufacturing process, from in-process controls or from a combination of the two. Parametric release in appropriate circumstances is thus not precluded by the need to comply with the Pharmacopoeia. The general notice on Assays and Tests indicates that analytical methods other than those described in the Pharmacopoeia may be employed for routine purposes.

Requirements in monographs have been framed to provide appropriate limitation of potential impurities rather than to provide against all possible impurities. Material found to contain an impurity not detectable by means of the prescribed tests is not of pharmacopoeial quality if the nature or amount of the impurity found is incompatible with good pharmaceutical practice.

The status of any statement given under the headings Definition, Production, Characteristics, Storage, Labelling or Action and use is defined within the general notice relating to the relevant heading. In addition to any exceptions indicated by one of the general notices referred to above, the following parts of a monograph do not constitute standards: (a) a graphic or molecular formula given at the beginning of a monograph; (b) a molecular weight; (c) a Chemical Abstracts Service Registry Number; (d) any information given at the end of a monograph concerning impurities known to be limited by that monograph; (e) information in any annex to a

monograph. Any statement containing the word 'should' constitutes non-

mandatory advice or recommendation.

The expression 'unless otherwise justified and authorised' means that the requirement in question has to be met, unless a competent authority

authorises a modification or exemption where justified in a particular case. The term 'competent authority' means the national, supranational or

international body or organisation vested with the authority for making

decisions concerning the issue in question. It may, for example, be a

licensing authority or an official control laboratory. For a formulated

preparation that is the subject of monograph in the British Pharmacopoeia

any justified and authorised modification to, or exemption from, the

requirements of the relevant general monograph of the European

Pharmacopoeia is stated in the individual monograph. For example, the

general monograph for Tablets requires that Uncoated Tablets, except for

chewable tablets, disintegrate within 15 minutes; for Calcium Lactate

Tablets a time of 30 minutes is permitted.

Many of the general monographs for formulated preparations include

statements and requirements additional to those of the European

Pharmacopoeia that are applicable to the individual monographs of the

British Pharmacopoeia. Such statements and requirements apply to all

monographs for that dosage form included in the Pharmacopoeia unless

otherwise indicated in the individual monograph.

Where a monograph on a biological substance or preparation refers to a

strain, a test, a method, a substance, etc., using the qualifications 'suitable'

or 'appropriate' without further definition in the text, the choice of such

strain, test, method, substance, etc., is made in accordance with any

international agreements or national regulations affecting the subject

concerned.

Definition of Terms

Where the term 'about' is included in a monograph or test it should be

taken to mean approximately (fairly correct or accurate; near to the actual

value).

Where the term 'corresponds' is included in a monograph or test it

should be taken to mean similar or equivalent in character or quantity.

Where the term 'similar' is included in a monograph or test it should be

taken to mean alike though not necessarily identical.

Further qualifiers (such as numerical acceptance criteria) for the above

terms are not included in the BP. The acceptance criteria for any individual

case is set based on the range of results obtained from known reference

samples, the level of precision of the equipment or apparatus used and the

level of accuracy required for the particular application. The user should

determine the variability seen in his/her own laboratory and set in-house

acceptance criteria that he/she judges to be appropriate based on the local

operating conditions.

Expression of Standards

Where the standard for the content of a substance described in a monograph is expressed in terms of the chemical formula for that substance an upper limit exceeding 100% may be stated. Such an upper limit applies to the result of the assay calculated in terms of the equivalent content of the specified chemical formula. For example, the statement 'contains not less than 99.0% and not more than 101.0% of $C_{20}H_{24}N_2O_2 \cdot HCl$ ' implies that the result of the assay is not less than 99.0% and not more than 101.0%, calculated in terms of the equivalent content of $C_{20}H_{24}N_2O_2 \cdot HCl$.

Where the result of an assay or test is required to be calculated with reference to the dried, anhydrous or ignited substance, the substance free from a specified solvent or to the peptide content, the determination of loss on drying, water content, loss on ignition, content of the specified solvent or peptide content is carried out by the method prescribed in the relevant test in the monograph.

Temperature

The Celsius thermometric scale is used in expressing temperatures.

Weights and Measures

The metric system of weights and measures is employed; SI Units have generally been adopted. Metric measures are required to have been graduated at 20° and all measurements involved in the analytical operations of the Pharmacopoeia are intended, unless otherwise stated, to be made at that temperature. Graduated glass apparatus used in analytical operations should comply with Class A requirements of the appropriate International Standard issued by the International Organization for Standardization. The abbreviation for litre is 'L' throughout the Pharmacopoeia. In line with European Directive 80/181/EEC, the abbreviation 'l' is also permitted for use.

Atomic Weights

The atomic weights adopted are the values given in the Table of Relative Atomic Weights 2001 published by the International Union of Pure and Applied Chemistry (Appendix XXV).

Constant Weight

The term 'constant weight', used in relation to the process of drying or the process of ignition, means that two consecutive weighings do not differ by more than 0.5 mg, the second weighing being made after an additional period of drying or ignition under the specified conditions appropriate to the nature and quantity of the residue (1 hour is usually suitable).

Expression of Concentrations

The term 'per cent' or more usually the symbol '%' is used with one of four different meanings in the expression of concentrations according to circumstances. In order that the meaning to be attached to the expression in each instance is clear, the following notation is used:

Per cent w/w (% w/w) (percentage weight in weight) expresses the number of grams of solute in 100 g of product.

Per cent w/v (% w/v) (percentage weight in volume) expresses the number of grams of solute in 100 mL of product.

Per cent v/v (% v/v) (percentage volume in volume) expresses the number of millilitres of solute in 100 mL of product.

Per cent v/w (% v/w) (percentage volume in weight) expresses the number of millilitres of solute in 100 g of product.

Usually the strength of solutions of solids in liquids is expressed as percentage weight in volume, of liquids in liquids as percentage volume in volume and of gases in liquids as percentage weight in weight.

When the concentration of a solution is expressed as parts per million (ppm), it means weight in weight, unless otherwise specified.

When the concentration of a solution is expressed as parts of dissolved substance in parts of the solution, it means parts by weight (g) of a solid in parts by volume (mL) of the final solution; or parts by volume (mL) of a liquid in parts by volume (mL) of the final solution; or parts by weight (g) of a gas in parts by weight (g) of the final solution.

When the concentration of a solution is expressed in molarity designated by the symbol *M* preceded by a number, it denotes the number of moles of the stated solute contained in sufficient Purified Water (unless otherwise stated) to produce 1 litre of solution.

Water Bath

The term 'water bath' means a bath of boiling water, unless water at some other temperature is indicated in the text. An alternative form of heating may be employed providing that the required temperature is approximately maintained but not exceeded.

Reagents

The reagents required for the assays and tests of the Pharmacopoeia are defined in appendices. The descriptions set out in the appendices do not imply that the materials are suitable for use in medicine.

Indicators

Indicators, the colours of which change over approximately the same range of pH, may be substituted for one another but in the event of doubt or dispute as to the equivalence of indicators for a particular purpose, the indicator specified in the text is alone authoritative. The quantity of an indicator solution appropriate for use in acid-base titrations described in assays or tests is 0.1 mL unless otherwise stated in the text.

Any solvent required in an assay or test in which an indicator is specified is previously neutralised to the indicator, unless a blank test is prescribed.

Caution Statements

A number of materials described in the monographs and some of the reagents specified for use in the assays and tests of the Pharmacopoeia may be injurious to health unless adequate precautions are taken. The principles of good laboratory practice and the provisions of any appropriate regulations such as those issued in the United Kingdom in accordance with the Health and Safety at Work *etc.* Act 1974 should be observed at all times in carrying out the assays and tests of the Pharmacopoeia. Attention is drawn to particular hazards in certain monographs by means of an italicised statement; the absence of such a statement should not however be taken to mean that no hazard exists.

Titles

Subsidiary titles, where included, have the same significance as the main titles. An abbreviated title constructed in accordance with the directions given in Appendix XXI A has the same significance as the main title. Titles that are derived by the suitable inversion of words of a main or subsidiary title, with the addition of a preposition if appropriate, are also official titles. Thus, the following are all official titles: Aspirin Tablets, Tablets of Aspirin; Atropine Injection, Injection of Atropine. A title of a formulated preparation that includes the full nonproprietary name of the active ingredient or ingredients, where this is not included in the title of the monograph, is also an official title. For example, the title Promethazine Hydrochloride Oral Solution has the same significance as Promethazine Oral Solution and the title Brompheniramine Maleate Tablets has the same significance as Brompheniramine Tablets. Where the English title at the head of a monograph in the European Pharmacopoeia is different from that at the head of the text incorporated into the British Pharmacopoeia, an Approved Synonym has been created on the recommendation of the British Pharmacopoeia Commission. Approved Synonyms have the same significance as the main title and are thus official

titles. A cumulative list of such Approved Synonyms is provided in

Appendix XXI B.

Where the names of pharmacopoeial substances, preparations and other materials occur in the text they are printed with capital initial letters and this indicates that materials of Pharmacopoeial quality must be used. Words in the text that name a reagent or other material, a physical characteristic or a process that is described or defined in an appendix are printed in italic type, for example, *methanol*, *absorbance*, *gas chromatography*, and these imply compliance with the requirements specified in the appropriate appendix.

Chemical Formulae

When the chemical composition of an official substance is known or generally accepted, the graphic and molecular formulae, the molecular weight and the Chemical Abstracts Service Registry Number are normally given at the beginning of the monograph for information. This information refers to the chemically pure substance and is not to be regarded as an indication of the purity of the official material. Elsewhere, in statements of standards of purity and strength and in descriptions of processes of assay, it is evident from the context that the formulae denote the chemically pure substances.

Where the absolute stereochemical configuration is specified, the International Union of Pure and Applied Chemistry (IUPAC) *R/S* and *E/Z* systems of designation have been used. If the substance is an enantiomer of unknown absolute stereochemistry the sign of the optical rotation, as determined in the solvent and under the conditions specified in the monograph, has been attached to the systematic name. An indication of sign of rotation has also been given where this is incorporated in a trivial name that appears on an IUPAC preferred list.

All amino acids, except glycine, have the *L*-configuration unless otherwise indicated. The three-letter and one-letter symbols used for amino acids in peptide and protein sequences are those recommended by the Joint Commission on Biochemical Nomenclature of the International Union of Pure and Applied Chemistry and the International Union of Biochemistry and Molecular Biology.

In the graphic formulae the following abbreviations are used:

Me	-CH ₃	Bu ^s	-CH(CH ₃)CH ₂ CH ₃	Bu ^s	-CH(CH ₃)CH ₂ CH ₃
Et	-CH ₂ CH ₃	Bu ⁿ	-CH ₂ CH ₂ CH ₂ CH ₃	Bu ⁿ	-CH ₂ CH ₂ CH ₂ CH ₃
Pr ⁱ	-CH(CH ₃) ₂	Bu ^t	-C(CH ₃) ₃	Bu ^t	-C(CH ₃) ₃
Pr ⁿ	-CH ₂ CH ₂ CH ₃	Ph	-C ₆ H ₅	Ph	-C ₆ H ₅
Bu ⁱ	-CH ₂ CH(CH ₃)CH ₂ CH ₃	Ac	-COCH ₃	Ac	-COCH ₃

Definition

Statements given under the heading Definition constitute an official definition of the substance, preparation or other article that is the subject of the monograph. They constitute instructions or requirements and are mandatory in nature. Certain medicinal or pharmaceutical substances and other articles are defined by reference to a particular method of manufacture. A statement that a substance or article is prepared or obtained by a certain method constitutes part of the official definition and implies that other methods are not permitted. A statement that a substance may be prepared or obtained by a certain method, however, indicates that this is one possible method and does not imply that other methods are proscribed.

Additional statements concerning the definition of formulated preparations are given in the general notice on Manufacture of Formulated Preparations.

Production

Statements given under the heading Production draw attention to particular aspects of the manufacturing process but are not necessarily comprehensive. They constitute mandatory instructions to manufacturers. They may relate, for example, to source materials, to the manufacturing process itself and its validation and control, to in-process testing or to testing that is to be carried out by the manufacturer on the final product (bulk material or dosage form) either on selected batches or on each batch prior to release. These statements cannot necessarily be verified on a sample of the final product by an independent analyst. The competent authority may establish that the instructions have been followed, for example, by examination of data received from the manufacturer, by inspection or by testing appropriate samples.

The absence of a section on Production does not imply that attention to features such as those referred to above is not required. A substance, preparation or article described in a monograph of the Pharmacopoeia is to be manufactured in accordance with the principles of good manufacturing practice and in accordance with relevant international agreements and supranational and national regulations governing medicinal products. Where in the section under the heading Production a monograph on a vaccine defines the characteristics of the vaccine strain to be used, any test methods given for confirming these characteristics are provided as examples of suitable methods. The use of these methods is not mandatory. Additional statements concerning the production of formulated preparations are given in the general notice on Manufacture of Formulated Preparations.

Manufacture of Formulated Preparations

Attention is drawn to the need to observe adequate hygienic precautions in the preparation and dispensing of pharmaceutical formulations. The principles of good pharmaceutical manufacturing practice should be observed.

The Definition in certain monographs for pharmaceutical preparations is given in terms of the principal ingredients only. Any ingredient, other than those included in the Definition, must comply with the general notice on Excipients and the product must conform with the Pharmacopoeial requirements.

The Definition in other monographs for pharmaceutical preparations is presented as a full formula. No deviation from the stated formula is permitted except those allowed by the general notices on Colouring Agents and Antimicrobial Preservatives. Where additionally directions are given under the heading Extremoporeous Preparation these are intended for the extemporaneous preparation of relatively small quantities for short-term supply and use. When so prepared, no deviation from the stated directions is permitted. If, however, such a pharmaceutical preparation is manufactured on a larger scale with the intention that it may be stored, deviations from the stated directions are permitted provided that the final product meets the following criteria:

- (1) compliance with all of the requirements stated in the monograph;
 - (2) retention of the essential characteristics of the preparation made strictly in accordance with the directions of the Pharmacopoeia.
- Monographs for yet other pharmaceutical preparations include both a Definition in terms of the principal ingredients and, under the side-heading Extemporaneous Preparation, a full formula together with, in some cases, directions for their preparation. Such full formulae and directions are intended for the extemporaneous preparation of relatively small quantities for short-term supply and use. When so prepared, no deviation from the stated formula and directions is permitted. If, however, such a pharmaceutical preparation is manufactured on a larger scale with the intention that it may be stored, deviations from the formula and directions stated under the heading Extemporaneous Preparation are permitted, provided that any ingredient, other than those included in the Definition, complies with the general notice on Excipients and that the final product meets the following criteria:

- (1) accordance with the Definition stated in the monograph;
 - (2) compliance with all of the requirements stated in the monograph;
 - (3) retention of the essential characteristics of the preparation made strictly in accordance with the formula and directions of the Pharmacopoeia.
- In the manufacture of any official preparation on a large scale with the intention that it should be stored, in addition to following any instruction under the heading Production, it is necessary to ascertain that the product is satisfactory with respect to its physical and chemical stability and its state of preservation over the claimed shelf-life. This applies irrespective of whether the formula of the Pharmacopoeia and any instructions given under the heading Extemporaneous Preparation are followed precisely or modified. Provided that the preparation has been shown to be stable in other respects, deterioration due to microbial contamination may be inhibited by the incorporation of a suitable antimicrobial preservative. In such circumstances the label states appropriate storage conditions, the date after which the product should not be used and the identity and concentration of the antimicrobial preservative.

Freshly and Recently Prepared

The direction, given under the heading Extemporaneous Preparation, that a preparation must be freshly prepared indicates that it must be made not more than 24 hours before it is issued for use. The direction that a preparation should be recently prepared indicates that deterioration is likely if the preparation is stored for longer than about 4 weeks at 15° to 25°.

Methods of Sterilisation

The methods of sterilisation used in preparing the sterile materials described in the Pharmacopoeia are given in Appendix XVIII. For aqueous preparations, steam sterilisation (heating in an autoclave) is the method of choice wherever it is known to be suitable. Any method of sterilisation must be validated with respect to both the assurance of sterility and the integrity of the product and to ensure that the final product complies with the requirements of the monograph.

Water

The term water used without qualification in formulae for formulated preparations means either potable water freshly drawn direct from the public supply and suitable for drinking or freshly boiled and cooled Purified

Water. The latter should be used if the public supply is from a local storage tank or if the potable water is unsuitable for a particular preparation.

Excipients

Where an excipient for which there is a pharmacopoeial monograph is used in preparing an official preparation it shall comply with that monograph. Any substance added in preparing an official preparation shall be innocuous, shall have no adverse influence on the therapeutic efficacy of the active ingredients and shall not interfere with the assays and tests of the Pharmacopoeia. Particular care should be taken to ensure that such substances are free from harmful organisms.

Colouring Agents

If in a monograph for a formulated preparation defined by means of a full formula a specific colouring agent or agents is prescribed, suitable alternatives approved in the country concerned may be substituted.

Antimicrobial Preservatives

When the term 'suitable antimicrobial preservative' is used it is implied that the preparation concerned will be effectively preserved according to the appropriate criteria applied and interpreted as described in the test for efficacy of antimicrobial preservation (Appendix XVI C). In certain monographs for formulated preparations defined by means of a full formula, a specific antimicrobial agent or agents may be prescribed; suitable alternatives may be substituted provided that their identity and concentration are stated on the label.

Characteristics

Statements given under the heading Characteristics are not to be interpreted in a strict sense and are not to be regarded as official requirements. Statements on taste are provided only in cases where this property is a guide to the acceptability of the material (for example, a material used primarily for flavouring). The status of statements on solubility is given in the general notice on Solubility. Solubility Statements on solubility given under the heading Characteristics are intended as information on the approximate solubility at a temperature between 15° and 25°, unless otherwise stated, and are not to be considered as official requirements. Statements given under headings such as Solubility in ethanol express exact requirements and constitute part of the standards for the substances under which they occur. The following table indicates the meanings of the terms used in statements of approximate solubilities.

Descriptive term	Approximate volume of solvent in millilitres per gram of solute
very soluble	less than 1
freely soluble	from 1 to 10
soluble	from 10 to 30
sparingly soluble	from 30 to 100
slightly soluble	from 100 to 1000
very slightly soluble	from 1000 to 10,000
practically insoluble	more than 10,000

The term 'partly soluble' is used to describe a mixture of which only some of the components dissolve.

Identification

The tests described or referred to under the heading Identification are not necessarily sufficient to establish absolute proof of identity. They provide a means of verifying that the identity of the material being examined is in accordance with the label on the container.

Unless otherwise prescribed, identification tests are carried out at a temperature between 15° and 25°.

Reference spectra Where a monograph refers to an infrared reference spectrum, this spectrum is provided in a separate section of the

Pharmacopoeia. A sample spectrum is considered to be concordant with a reference spectrum if the transmission minima (absorption maxima) of the principal bands in the sample correspond in position, relative intensities and shape to those of the reference. Instrumentation software may be used to calculate concordance with a previously recorded reference spectrum.

When tests for infrared absorption are applied to material extracted from formulated preparations, strict concordance with the specified reference spectrum may not always be possible, but nevertheless a close resemblance between the spectrum of the extracted material and the specified reference spectrum should be achieved.

Assays and Tests

The assays and tests described are the official methods upon which the standards of the Pharmacopoeia depend. The analyst is not precluded from employing alternative methods, including methods of micro-analysis, in any assay or test if it is known that the method used will give a result of equivalent accuracy. Local reference materials may be used for routine analysis, provided that these are calibrated against the official reference materials. In the event of doubt or dispute, the methods of analysis, the reference materials and the reference spectra of the Pharmacopoeia are alone authoritative.

Where the solvent used for a solution is not named, the solvent is Purified Water.

Unless otherwise prescribed, the assays and tests are carried out at a temperature between 15° and 25°.

A temperature in a test for Loss on drying, where no temperature range

is given, implies a range of $\pm 2^\circ$ about the stated value.

Visual comparative tests, unless otherwise prescribed, are carried out using identical tubes of colourless, transparent, neutral glass with a flat base. The volumes of liquid prescribed are for use with tubes 16 mm in internal diameter; tubes with a larger internal diameter may be used but the volume of liquid examined must be increased so that the depth of liquid in the tubes is not less than that obtained when the prescribed volume of liquid and tubes 16 mm in internal diameter are used. Equal volumes of the liquids to be compared are examined down the vertical axis of the tubes against a white background or, if necessary, against a black background.

The examination is carried out in diffuse light.

Where a direction is given that an analytical operation is to be carried out in subdued light, precautions should be taken to avoid exposure to direct sunlight or other strong light. Where a direction is given that an analytical operation is to be carried out 'protected from light', precautions should be taken to exclude actinic light by the use of low-actinic glassware, working in a dark room or similar procedures.

For preparations other than those of fixed strength, the quantity to be

taken for an assay or test is usually expressed in terms of the active

ingredient. This means that the quantity of the active ingredient expected to

be present and the quantity of the preparation to be taken are calculated from the strength stated on the label.

In assays the approximate quantity to be taken for examination is indicated but the quantity actually used must not deviate by more than 10% from that stated. The quantity taken is accurately weighed or measured and the result of the assay is calculated from this exact quantity. Reagents are measured and the procedures are carried out with an accuracy commensurate with the degree of precision implied by the standard stated for the assay.

In tests the stated quantity to be taken for examination must be used unless any divergence can be taken into account in conducting the test and calculating the result. The quantity taken is accurately weighed or measured with the degree of precision implied by the standard or, where the standard is not stated numerically (for example, in tests for Clarity and colour of solution), with the degree of precision implied by the number of significant figures stated. Reagents are measured and the procedures are carried out with an accuracy commensurate with this degree of precision.

The limits stated in monographs are based on data obtained in normal analytical practice; they take account of normal analytical errors, of acceptable variations in manufacture and of deterioration to an extent considered acceptable. No further tolerances are to be applied to the limits prescribed to determine whether the article being examined complies with the requirements of the monograph.

In determining compliance with a numerical limit, the calculated result of a test or assay is first rounded to the number of significant figures stated, unless otherwise prescribed. The last figure is increased by 1 when the part rejected is equal to or exceeds one half-unit, whereas it is not modified when the part rejected is less than a half-unit.

In certain tests, the concentration of impurity is given in parentheses either as a percentage or in parts per million by weight (ppm). In chromatographic tests such concentrations are stated as a percentage irrespective of the limit. In other tests they are usually stated in ppm unless the limit exceeds 500 ppm. In those chromatographic tests in which a secondary spot or peak in a chromatogram obtained with a solution of the substance being examined is described as corresponding to a named impurity and is compared with a spot or peak in a chromatogram obtained with a reference solution of the same impurity, the percentage given in parentheses indicates the limit for that impurity. In those chromatographic tests in which a spot or peak in a chromatogram obtained with a solution of the substance being examined is described in terms other than as corresponding to a named impurity (commonly, for example, as any (other) *secondary spot* or *peak*) but is compared with a spot or peak in a chromatogram obtained with a reference solution of a named impurity, the percentage given in parentheses indicates an impurity limit expressed in terms of a nominal concentration of the named impurity. In chromatographic tests in which a comparison is made between spots or peaks in chromatograms obtained with solutions of different concentrations of the substance being examined, the percentage given in parentheses indicates an impurity limit expressed in terms of a nominal concentration of the medicinal substance itself. In some monographs, in particular those for certain formulated preparations, the impurity limit is expressed in terms of a nominal concentration of the active moiety rather than of the medicinal

Biological Assays and Tests

substance itself. Where necessary for clarification the terms in which the limit is expressed are stated within the monograph.

In all cases where an impurity limit is given in parentheses, the figures given are approximations for information only; conformity with the requirements is determined on the basis of compliance or otherwise with the stated test.

The use of a proprietary designation to identify a material used in an assay or test does not imply that another equally suitable material may not be used.

Methods of assay described as Suggested methods are not obligatory, but when another method is used its precision must be not less than that required for the Suggested method.

For those antibiotics for which the monograph specifies a microbiological assay the potency requirement is expressed in the monograph in International Units (IU) per milligram. The material is not of pharmacopoeial quality if the upper fiducial limit of error is less than the stated potency. For such antibiotics the required precision of the assay is stated in the monograph in terms of the fiducial limits of error about the estimated potency.

For other substances and preparations for which the monograph specifies a biological assay, unless otherwise stated, the precision of the assay is such that the fiducial limits of error, expressed as a percentage of the estimated potency, are within a range not wider than that obtained by multiplying by a factor of 10 the square roots of the limits given in the monograph for the fiducial limits of error about the stated potency.

In all cases fiducial limits of error are based on a probability of 95% ($P = 0.95$).

Where the biological assay is being used to ascertain the purity of the material, the stated potency means the potency stated on the label in terms of International Units (IU) or other Units per gram, per milligram or per millilitre. When no such statement appears on the label, the stated potency means the fixed or minimum potency required in the monograph. This interpretation of stated potency applies in all cases except where the monograph specifically directs otherwise.

Where the biological assay is being used to determine the total activity in the container, the stated potency means the total number of International Units (IU) or other Units stated on the label or, if no such statement appears, the total activity calculated in accordance with the instructions in the monograph.

Wherever possible the primary standard used in an assay or test is the respective International Standard or Reference Preparation established by the World Health Organization for international use and the biological activity is expressed in International Units (IU).

In other cases, where Units are referred to in an assay or test, the Unit for a particular substance or preparation is, for the United Kingdom, the specific biological activity contained in such an amount of the respective primary standard as the appropriate international or national organisation indicates. The necessary information is provided with the primary standard. Unless otherwise directed, animals used in an assay or a test are healthy animals, drawn from a uniform stock, that have not previously been treated with any material that will interfere with the assay or test. Unless otherwise stated, guinea-pigs weigh not less than 250 g or, when used in systemic

toxicity tests, not less than 350 g. When used in skin tests they are white or light coloured. Unless otherwise stated, mice weigh not less than 17 g and not more than 22 g.

Certain of the biological assays and tests of the Pharmacopoeia are such that in the United Kingdom they may be carried out only in accordance with the Animals (Scientific Procedures) Act 1986. Instructions included in such assays and tests in the Pharmacopoeia, with respect to the handling of animals, are therefore confined to those concerned with the accuracy and reproducibility of the assay or test.

Certain monographs require the use of a reference substance, a reference preparation or a reference spectrum. These are chosen with regard to their intended use as prescribed in the monographs of the Pharmacopoeia and are not necessarily suitable in other circumstances.

Any information necessary for proper use of the reference substance or reference preparation is given on the label or in the accompanying leaflet or brochure. Where no drying conditions are stated in the leaflet or on the label, the substance is to be used as received. No certificate of analysis or other data not relevant to the prescribed use of the product are provided. The products are guaranteed to be suitable for use for a period of three months from dispatch when stored under the appropriate conditions. The stability of the contents of opened containers cannot be guaranteed. The current lot is listed in the BP Laboratory website catalogue. Additional information is provided in Supplementary Chapter III E.

Chemical Reference Substances

The abbreviation BPCRS indicates a Chemical Reference Substance established by the British Pharmacopoeia Commission. The abbreviation CRS or EPCRS indicates a Chemical Reference Substance established by the European Pharmacopoeia Commission. Some Chemical Reference Substances are used for the microbiological assay of antibiotics and their activity is stated, in International Units, on the label or on the accompanying leaflet and defined in the same manner as for Biological Reference Preparations.

Biological Reference Preparations

The majority of the primary biological reference preparations referred to are the appropriate International Standards and Reference Preparations established by the World Health Organisation. Because these reference materials are usually available only in limited quantities, the European Pharmacopoeia has established Biological Reference Preparations (indicated by the abbreviation BRP or EBBRP) where appropriate. Where applicable, the potency of the Biological Reference Preparations is expressed in International Units. For some Biological Reference Preparations, where an international standard or reference preparation does not exist, the potency is expressed in European Pharmacopoeia Units.

Reference Substances and Preparations

Storage

Statements under the side-heading Storage constitute non-mandatory advice. The substances and preparations described in the Pharmacopoeia are to be stored under conditions that prevent contamination and, as far as possible, deterioration. Unless otherwise stated in the monograph, the substances and preparations described in the Pharmacopoeia are kept in well-closed containers and stored at a temperature not exceeding 25°. Precautions that should be taken in relation to the effects of the atmosphere, moisture, heat and light are indicated, where appropriate, in

the monographs. Further precautions may be necessary when some materials are stored in tropical climates or under other severe conditions. The expression 'protected from moisture' means that the product is to be stored in an airtight container. Care is to be taken when the container is opened in a damp atmosphere. A low moisture content may be maintained, if necessary, by the use of a desiccant in the container provided that direct contact with the product is avoided.

The expression 'protected from light' means that the product is to be stored either in a container made of a material that absorbs actinic light sufficiently to protect the contents from change induced by such light or in a container enclosed in an outer cover that provides such protection or stored in a place from which all such light is excluded.

The expression 'tamper-evident container' means a closed container fitted with a device that reveals irreversibly whether the container has been opened, whereas, the expression 'tamper-proof container' means a closed container in which access to the contents is prevented under normal conditions of use. The two terms are considered to be synonymous by the European Pharmacopoeia Commission.

Labelling

The labelling requirements of the Pharmacopoeia are not comprehensive, and the provisions of regulations issued in accordance with the requirements of the territory in which the medicinal product is to be used should be met.

Licensed medicines intended for use within the United Kingdom must comply with the requirements of The Human Medicines Regulations 2012 and European Directive 2001/83/EC, Title V (as amended) in respect of their labelling and package leaflets, together with those regulations for the labelling of hazardous materials.

Best practice guidance on the labelling and packaging of medicines for use in the United Kingdom advises that certain items of information are deemed critical for the safe use of the medicine (see 'Best Practice Guidance on the Labelling and Packaging of Medicines' issued by the MHRA, 2012). Further information and guidance on the labelling of medicinal products can be found in Supplementary Chapter I G.

Such matters as the exact form of wording to be used and whether a particular item of information should appear on the primary label and additionally, or alternatively, on the package or exceptionally in a leaflet are, in general, outside the scope of the Pharmacopoeia. When the term 'label' is used in Labelling statements of the Pharmacopoeia, decisions as to where the particular statement should appear should therefore be made in accordance with relevant legislation.

The label of every official formulated preparation other than those of fixed strength also states the content of the active ingredient or ingredients expressed in the terms required by the monograph. Where the content of active ingredient is required to be expressed in terms other than the weight of the official medicinal substance used in making the formulation, this is specifically stated under the heading Labelling. Unless otherwise stated in the monograph, the content of the active ingredient is expressed in terms of the official medicinal substance used in making the formulation.

These requirements do not necessarily apply to unlicensed preparations supplied in accordance with a prescription. For requirements for unlicensed medicines see the general monograph on Unlicensed Medicines.

Action and Use

The statements given under this heading in monographs are intended only as information on the principal pharmacological actions or the uses of the materials in medicine or pharmacy. It should not be assumed that the substance has no other action or use. The statements are not intended to be binding on prescribers or to limit their discretion.

**Crude Drugs;
Traditional Herbal
and Complementary
Medicines**

Herbal and complementary medicines are classed as medicines under European Directive 2001/83/EC as amended. It is emphasised that, although requirements for the quality of the material are provided in the monograph to assist the registration scheme by the UK Licensing Authority, the British Pharmacopoeia Commission has not assessed the safety or efficacy of the material in traditional use.

Monograph Title For traditional herbal medicines, the monograph title is a combination of the binominal name together with a description of use. Monographs for the material that has not been processed (the herbal drug) and the processed material (the herbal drug preparation) are published where possible. To distinguish between the two, the word 'Processed' is included in the relevant monograph title.

Definition Under the heading Definition, the botanical name together with any synonym is given. Where appropriate, for material that has not been processed, information on the collection/harvesting and/or treatment/drying of the whole herbal drug may be given. For processed materials, the method of processing, where appropriate, will normally be given in a separate section.

Characteristics References to odour are included only where this is highly characteristic. References to taste are not included.

Control methods Where applicable, the control methods to be used in monographs are:

- macroscopical and microscopical descriptions and chemical/chromatographic tests for identification
- tests for absence of any related species
- microbial test to assure microbial quality
- tests for inorganic impurities and non-specific purity tests, including extractive tests, Sulfated ash and Heavy metals where appropriate
- test for Loss on drying or Water
- wherever possible, a method for assaying the active constituent(s) or suitable marker constituent(s).

The macroscopical characteristics include those features that can be seen by the unaided eye or by the use of a hand lens. When two species/subspecies of the same plant are included in the Definition, individual differences between the two are indicated where possible. The description of the microscopical characteristics of the powdered drug includes information on the dominant or the most specific characters. Where it is considered to be an aid to identification, illustrations of the powdered drug may be provided.

The following aspects are controlled by the general monograph for Herbal Drugs: they are required to be free from moulds, insects, decay, animal matter and animal excreta. Unless otherwise prescribed the amount of foreign matter is not more than 2% w/w. Microbial contamination should be minimal.

Homoeopathic Medicines

In determining the content of the active constituents or the suitable marker substances measurements are made with reference to the dried or anhydrous herbal drug. In the tests for Acid-insoluble ash, Ash, Extractive soluble in ethanol, Loss on drying, Sulfated ash, Water, Water-soluble ash and Water-soluble extractive of herbal drugs, the calculations are made with reference to the herbal drug that has not been specifically dried unless otherwise prescribed in the monograph.

Homoeopathic medicines are classed as medicines under European Directive 2001/83/EC as amended. It is emphasised that, although requirements for the quality of the material are provided in the relevant monograph in order to assist the simplified registration scheme by the UK Licensing Authority, the British Pharmacopoeia Commission has not assessed the safety or efficacy of the material in use.

All materials used for the production of homoeopathic medicines, including excipients, must comply with European Pharmacopoeia or British Pharmacopoeia monographs for those materials. Where such European Pharmacopoeia or British Pharmacopoeia monographs do not exist, each material used for the production of homoeopathic medicines must comply with an official national pharmacopoeia of a Member State.

British Pharmacopoeia monographs for homoeopathic medicines apply to homoeopathic stocks and mother tinctures only, but may be prefaced by a section which details the quality requirements applicable to the principle component where there is no European Pharmacopoeia or British Pharmacopoeia monograph for the material. These monographs also include either general statements on the methods of preparation or refer to specific methods of preparation given in the European Pharmacopoeia. Homoeopathic stocks and mother tinctures undergo the further process referred to as potentisation. Potentisation is a term specific to homoeopathic medicine and is a process of dilution of stocks and mother tinctures to produce the final product.

Identification tests are established for the components in homoeopathic stocks and usually relate to those applied to the materials used in the production of the homoeopathic stocks. An assay is included for the principal component(s) where possible. For mother tinctures, an identification test, usually chromatographic, is established and, where applicable, an assay for the principle component(s), where appropriate, other tests, related to the solvent, dry matter or known adulterants, are included. Specifications have not been set for final homoeopathic products due to the high dilution used in their preparation and the subsequent difficulty in applying analytical methodology. Statements under Crude Drugs, Traditional Herbal and Complementary Medicines also apply to homoeopathic stocks and mother tinctures, when appropriate.

Unlicensed Medicines

The General Monograph for Unlicensed Medicines applies to those formulations used in human medicine that are prepared under a Manufacturer's 'Specials' Licence or prepared extemporaneously under the supervision of a pharmacist, whether or not there is a published monograph for the specific dosage form.

An article intended for medicinal use that is described by means of an official title must comply with the requirements of the relevant monograph.

A formulated preparation must comply throughout its assigned shelf-life (period of validity). The subject of any other monograph must comply throughout its period of use.

Unlicensed medicines that are prepared under a Manufacturer's 'Specials' Licence comply with the requirements of the General Monograph for Pharmaceutical Preparations, the requirements of the General Monograph for Unlicensed Medicines and, where applicable, the requirements of the individual monograph for the specific dosage form.

Unlicensed medicines prepared extemporaneously under the supervision of a pharmacist comply with the requirements of the General Monograph for Pharmaceutical Preparations, the requirements of the General Monograph for Unlicensed Medicines and, where applicable, the requirements of the individual monograph for the specific dosage form.

While it is expected that extemporaneous preparations will demonstrate pharmaceutical compliance when tested, it is recognised that it might not be practicable to carry out the pharmaceutical tests routinely on such formulations. In the event of doubt or dispute, the methods of analysis, the reference materials and the reference spectra of the Pharmacopoeia are alone authoritative.

Part III

Monographs and other texts of the European Pharmacopoeia that are incorporated in this edition of the British Pharmacopoeia are governed by the general notices of the European Pharmacopoeia; these are reproduced below.

GENERAL NOTICES OF THE EUROPEAN

PHARMACOPOEIA

1.1. GENERAL STATEMENTS

The General Notices apply to all monographs and other texts of the

European Pharmacopoeia.

The official texts of the European Pharmacopoeia are published in

English and French. Translations in other languages may be prepared by

the signatory States of the European Pharmacopoeia Convention. In case of

doubt or dispute, the English and French versions are alone authoritative.

In the texts of the European Pharmacopoeia, the word 'Pharmacopoeia'

without qualification means the European Pharmacopoeia. The official

abbreviation Ph. Eur. may be used to indicate the European

Pharmacopoeia.

The use of the title or the subtitle of a monograph implies that the article

complies with the requirements of the relevant monograph. Such references

to monographs in the texts of the Pharmacopoeia are shown using the

monograph title and reference number in *italics*.

A preparation must comply throughout its period of validity; a distinct

period of validity and/or specifications for opened or broached containers

may be decided by the competent authority. The subject of any other

monograph must comply throughout its period of use. The period of

validity that is assigned to any given article and the time from which that

period is to be calculated are decided by the competent authority in light of

experimental results of stability studies.

Unless otherwise indicated in the General Notices or in the monographs,

statements in monographs constitute mandatory requirements. General

chapters become mandatory when referred to in a monograph, unless such

reference is made in a way that indicates that it is not the intention to make

the text referred to mandatory but rather to cite it for information.

The active substances, excipients, pharmaceutical preparations and other

articles described in the monographs are intended for human and veterinary

use (unless explicitly restricted to one of these uses).

The quality standards represented by monographs are valid only where the

articles in question are produced within the framework of a suitable quality

system. The quality system must assure that the articles consistently meet

the requirements of the Pharmacopoeia.

Alternative methods

The tests and assays described are the official methods upon which the standards of the Pharmacopoeia are based. With the agreement of the competent authority, alternative methods of analysis may be used for control purposes, provided that the methods used enable an unequivocal decision to be made as to whether compliance with the standards of the

monographs would be achieved if the official methods were used. In the event of doubt or dispute, the methods of analysis of the Pharmacopoeia are alone authoritative.

Demonstration of compliance with the Pharmacopoeia

(1) An article is not of Pharmacopoeia quality unless it complies with all the requirements stated in the monograph. This does not imply that performance of all the tests in a monograph is necessarily a prerequisite for a manufacturer in assessing compliance with the Pharmacopoeia before release of a product. The manufacturer may obtain assurance that a product is of Pharmacopoeia quality on the basis of its design, together with its control strategy and data derived, for example, from validation studies of the manufacturing process.

(2) An enhanced approach to quality control could utilise process analytical technology (PAT) and/or real-time release testing (including parametric release) strategies as alternatives to end-product testing alone. Real-time release testing in circumstances deemed appropriate by the competent authority is thus not precluded by the need to comply with the Pharmacopoeia.

(3) Reduction of animal testing: the European Pharmacopoeia is dedicated to phasing out the use of animals for test purposes, in accordance with the 3Rs (Replacement, Reduction, Refinement) set out in the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes. In demonstrating

compliance with the Pharmacopoeia as indicated above (1), manufacturers may consider establishing additional systems to monitor consistency of production. With the agreement of the competent authority, the choice of tests performed to assess compliance with the Pharmacopoeia when animal tests are prescribed is established in such a way that animal usage is minimised as much as possible.

Grade of materials

Certain materials that are the subject of a pharmacopoeial monograph may exist in different grades suitable for different purposes. Unless otherwise indicated in the monograph, the requirements apply to all grades of the material. In some monographs, particularly those on excipients, a list of functionality-related characteristics that are relevant to the use of the substance may be appended to the monograph for information. Test methods for determination of one or more of these characteristics may be given, also for information.

General monographs

Substances and preparations that are the subject of an individual monograph are also required to comply with relevant, applicable general monographs. Cross-references to applicable general monographs are not normally given in individual monographs. General monographs apply to all substances and preparations within the scope of the Definition section of the general monograph, except where a preamble limits the application, for example to substances and preparations that are the subject of a monograph of the Pharmacopoeia. General monographs on dosage forms apply to all preparations of the type defined. The requirements are not necessarily comprehensive for a given specific preparation and requirements additional to those prescribed in the general monograph may be imposed by the competent authority.

General monographs and individual monographs are complementary. If the provisions of a general monograph do not apply to a particular product, this is expressly stated in the individual monograph.

Validation of pharmacopoeial methods

The test methods given in monographs and general chapters have been validated in accordance with accepted scientific practice and current recommendations on analytical validation. Unless otherwise stated in the monograph or general chapter, validation of the test methods by the analyst is not required.

Implementation of pharmacopoeial methods

When implementing a pharmacopoeial method, the user must assess whether and to what extent the suitability of the method under the actual conditions of use needs to be demonstrated according to relevant monographs, general chapters and quality systems.

Conventional terms

The term 'competent authority' means the national, supranational or international body or organisation vested with the authority for making decisions concerning the issue in question. It may, for example, be a national pharmacopoeia authority, a licensing authority or an official control laboratory.

The expression 'unless otherwise justified and authorised' means that the requirements have to be met, unless the competent authority authorises a modification or an exemption where justified in a particular case.

Statements containing the word 'should' are informative or advisory. In certain monographs or other texts, the terms 'suitable' and 'appropriate' are used to describe a reagent, micro-organism, test method etc.; if criteria for suitability are not described in the monograph, suitability is demonstrated to the satisfaction of the competent authority.

Medicinal product (a) Any substance or combination of substances presented as having properties for treating or preventing disease in human beings and/or animals; or (b) any substance or combination of substances that may be used in or administered to human beings and/or animals with a view either to restoring, correcting or modifying physiological functions by exerting a pharmacological, immunological or metabolic action, or to making a medical diagnosis.

Herbal medicinal product Any medicinal product, exclusively containing as active ingredients one or more herbal drugs or one or more herbal drug preparations, or one or more such herbal drugs in combination with one or more such herbal drug preparations.

Active substance Any substance intended to be used in the manufacture of a medicinal product and that, when so used, becomes an active ingredient of the medicinal product. Such substances are intended to furnish a pharmacological activity or other direct effect in the diagnosis, cure, mitigation, treatment or prevention of disease, or to affect the structure and function of the body.

Excipient (auxiliary substance). Any constituent of a medicinal product that is not an active substance. Adjuvants, stabilisers, antimicrobial preservatives, diluents, antioxidants, for example, are excipients.

Interchangeable methods

Certain general chapters contain a statement that the text in question is harmonised with the corresponding text of the Japanese Pharmacopoeia and/or the United States Pharmacopoeia and that these texts are interchangeable. This implies that if a substance or preparation is found to

comply with a requirement using an interchangeable method from one of these pharmacopoeias it complies with the requirements of the European Pharmacopoeia. In the event of doubt or dispute, the text of the European Pharmacopoeia is alone authoritative.

References to

regulatory

documents

Monographs and general chapters may contain references to documents issued by regulatory authorities for medicines, for example directives and notes for guidance of the European Union. These references are provided for information for users for the Pharmacopoeia. Inclusion of such a reference does not modify the status of the documents referred to, which may be mandatory or for guidance.

1.2. OTHER PROVISIONS APPLYING TO GENERAL CHAPTERS AND MONOGRAPHS

Quantities

In tests with numerical limits and assays, the quantity stated to be taken for examination is approximate. The amount actually used, which may deviate by not more than 10 per cent from that stated, is accurately weighed or measured and the result is calculated from this exact quantity. In tests where the limit is not numerical, but usually depends upon comparison with the behaviour of a reference substance in the same conditions, the stated quantity is taken for examination. Reagents are used in the prescribed amounts.

Quantities are weighed or measured with an accuracy commensurate with the indicated degree of precision. For weighings, the precision corresponds to plus or minus 5 units after the last figure stated (for example, 0.25 g is to be interpreted as 0.245 g to 0.255 g). For the measurement of volumes, if the figure after the decimal point is a zero or ends in a zero (for example, 10.0 mL or 0.50 mL), the volume is measured using a pipette, a volumetric flask or a burette, as appropriate; otherwise, a graduated measuring cylinder or a graduated pipette may be used. Volumes stated in microlitres are measured using a micropipette or microsyringe.

It is recognised, however, that in certain cases the precision with which

quantities are stated does not correspond to the number of significant figures stated in a specified numerical limit. The weighings and

measurements are then carried out with a sufficiently improved accuracy.

Apparatus and procedures

Volumetric glassware complies with Class A requirements of the appropriate International Standard issued by the International Organisation for Standardisation.

Unless otherwise prescribed, analytical procedures are carried out at a temperature between 15 °C and 25 °C.

Unless otherwise prescribed, comparative tests are carried out using identical tubes of colourless, transparent, neutral glass with a flat base; the volumes of liquid prescribed are for use with tubes having an internal diameter of 16 mm, but tubes with a larger internal diameter may be used provided the volume of liquid used is adjusted (2.1.5). Equal volumes of the liquids to be compared are examined down the vertical axis of the tubes against a white background, or if necessary against a black background. The examination is carried out in diffuse light.

Any solvent required in a test or assay in which an indicator is to be used is previously neutralised to the indicator, unless a blank test is prescribed.

Water-bath

The term 'water-bath' means a bath of boiling water unless water at another temperature is indicated. Other methods of heating may be substituted provided the temperature is near to but not higher than 100 °C or the indicated temperature.

Drying and ignition to constant mass

The terms 'dried to constant mass' and 'ignited to constant mass' mean that 2 consecutive weighings do not differ by more than 0.5 mg, the 2nd weighing following an additional period of drying or of ignition respectively appropriate to the nature and quantity of the residue.

Where drying is prescribed using one of the expressions 'in a desiccator' or 'in vacuo', it is carried out using the conditions described in chapter 2.2.32. *Loss on drying*.

Reagents

The proper conduct of the analytical procedures described in the Pharmacopoeia and the reliability of the results depend, in part, upon the quality of the reagents used. The reagents are described in general chapter 4. It is assumed that reagents of analytical grade are used, for some reagents, tests to determine suitability are included in the specifications.

Solvents

Where the name of the solvent is not stated, the term 'solution' implies a solution in water.

Where the use of water is specified or implied in the analytical procedures described in the Pharmacopoeia or for the preparation of reagents, water complying with the requirements of the monograph *Purified water* (0008) is used, except that for many purposes the requirements for bacterial endotoxins (*Purified water in bulk*) and microbial contamination (*Purified water in containers*) are not relevant. The term 'distilled water' indicates purified water prepared by distillation.

The term 'ethanol' without qualification means anhydrous ethanol. The term 'alcohol' without qualification means ethanol (96 per cent). Other dilutions of ethanol are indicated by the term 'ethanol' or 'alcohol' followed by a statement of the percentage by volume of ethanol (C₂H₆O) required.

Expression of content

In defining content, the expression 'per cent' is used according to circumstances with one of 2 meanings:

- per cent *m/m* (percentage, mass in mass) expresses the number of grams of substance in 100 g of final product;
- per cent *V/V* (percentage, volume in volume) expresses the number of millilitres of substance in 100 mL of final product.

The expression 'parts per million' (or ppm) refers to mass in mass, unless otherwise specified.

Temperature

Where an analytical procedure describes temperature without a figure, the general terms used have the following meaning:

- in a deep-freeze: below -15 °C;
- in a refrigerator: 2 °C to 8 °C;
- cold or cool: 8 °C to 15 °C;
- room temperature: 15 °C to 25 °C.

1.3. GENERAL CHAPTERS

Containers

Materials used for containers are described in general chapter 3.1. General names used for materials, particularly plastic materials, each cover a range of products varying not only in the properties of the principal constituent but also in the additives used. The test methods and limits for materials depend on the formulation and are therefore applicable only for materials whose formulation is covered by the preamble to the specification. The use of materials with different formulations, and the test methods and limits applied to them, are subject to agreement by the competent authority. The specifications for containers in general chapter 3.2 have been developed for general application to containers of the stated category, but in view of the wide variety of containers available and possible new developments, the publication of a specification does not exclude the use, in justified circumstances, of containers that comply with other specifications, subject to agreement by the competent authority. Reference may be made within the monographs of the Pharmacopoeia to the definitions and specifications for containers provided in chapter 3.2. Containers. The general monographs for pharmaceutical dosage forms may, under the heading Definition/Production, require the use of certain types of container; certain other monographs may, under the heading Storage, indicate the type of container that is recommended for use.

1.4. MONOGRAPHS

Titles

Monograph titles are in English and French in the respective versions and there is a Latin subtitle.

Relative Atomic and Molecular Masses

The relative atomic mass (A_r) or the relative molecular mass (M_r) is shown, as and where appropriate, at the beginning of each monograph. The relative atomic and molecular masses and the molecular and graphic formulae do not constitute analytical standards for the substances described.

Chemical Abstracts Service (CAS) Registry Number

CAS registry numbers are included for information in monographs, where applicable, to provide convenient access to useful information for users. CAS Registry Number[®] is a registered trademark of the American Chemical Society.

Definition

Statements under the heading Definition constitute an official definition of the substance, preparation or other article that is the subject of the monograph.

Limits of content Where limits of content are prescribed, they are those determined by the method described under Assay. **Herbal drugs** In monographs on herbal drugs, the definition indicates whether the subject of the monograph is, for example, the whole drug or the drug in powdered form. Where a monograph applies to the drug in several states, for example both to the whole drug and the drug in powdered form, the definition states this.

Production

Statements under the heading Production draw attention to particular aspects of the manufacturing process but are not necessarily comprehensive. They constitute mandatory requirements for manufacturers, unless otherwise stated. They may relate, for example, to source materials, to the manufacturing process itself and its validation and control, to in-process

testing; or to testing that is to be carried out by the manufacturer on the final article, either on selected batches or on each batch prior to release. These statements cannot necessarily be verified on a sample of the final article by an independent analyst. The competent authority may establish that the instructions have been followed, for example, by examination of data received from the manufacturer, by inspection of manufacture or by testing appropriate samples.

The absence of a Production section does not imply that attention to features such as those referred to above is not required.

Choice of vaccine strain, Choice of vaccine composition The

Production section of a monograph may define the characteristics of a vaccine strain or vaccine composition. Unless otherwise stated, test methods given for verification of these characteristics are provided for information as examples of suitable methods. Subject to approval by the competent authority, other test methods may be used without validation against the method shown in the monograph.

Potential

Due to the increasing number of fraudulent activities and cases of

Adulteration

detect adulterated materials (i.e. active substances, excipients, intermediate products, bulk products and finished products).

To this purpose, a method for the detection of potential adulterants and relevant limits, together with a reminder that all stages of production and sourcing are subjected to a suitable quality system, may be included in this section of monographs on substances for which an incident has occurred or that present a risk of deliberate contamination. The frequency of testing by manufacturers or by users (e.g. manufacturers of intermediate products, bulk products and finished products, where relevant) depends on a risk assessment, taking into account the level of knowledge of the whole supply chain and national requirements.

This section constitutes requirements for the whole supply chain, from manufacturers to users (e.g. manufacturers of intermediate products, bulk products and finished products, where relevant). The absence of this section does not imply that attention to features such as those referred to above is not required.

Characters

The statements under the heading Characters are not to be interpreted in a strict sense and are not requirements.

Solubility In statements of solubility in the Characters section, the terms used have the following significance, referred to a temperature between 15 °C and 25 °C.

Descriptive term	Approximate volume of solvent in millilitres per gram of solute
Very soluble	less than 1
Freely soluble	from 1 to 10
Soluble	from 10 to 30
Sparingly soluble	from 30 to 100
Slightly soluble	from 100 to 1000
Very slightly soluble	from 1000 to 10 000
Practically insoluble	more than 10 000

The term 'partly soluble' is used to describe a mixture where only some of the components dissolve. The term 'miscible' is used to describe a liquid that is miscible in all proportions with the stated solvent.

Identification

Scope The tests given in the Identification section are not designed to give a full confirmation of the chemical structure or composition of the product; they are intended to give confirmation, with an acceptable degree of assurance, that the article conforms to the description on the label.

First and second identifications Certain monographs have subdivisions entitled 'First identification' and 'Second identification'. The test or tests that constitute the 'First identification' may be used in all circumstances. The test or tests that constitute the 'Second identification' may be used in pharmacies provided it can be demonstrated that the substance or preparation is fully traceable to a batch certified to comply with all the other requirements of the monograph.

Certain monographs give two or more sets of tests for the purpose of the first identification, which are equivalent and may be used independently. One or more of these sets usually contain a cross-reference to a test prescribed in the Tests section of the monograph. It may be used to simplify the work of the analyst carrying out the identification and the prescribed tests. For example, one identification set cross-refers to a test for enantiomeric purity while the other set gives a test for specific optical rotation; the intended purpose of the two is the same, that is, verification that the correct enantiomer is present.

Powdered herbal drugs Monographs on herbal drugs may contain schematic drawings of the powdered drug. These drawings complement the description given in the relevant identification test.

Tests and Assays

Scope The requirements are not framed to take account of all possible impurities. It is not to be presumed, for example, that an impurity that is not detectable by means of the prescribed tests is tolerated if common sense and good pharmaceutical practice require that it be absent. See also below under Impurities.

Calculation Where the result of a test or assay is required to be calculated with reference to the dried or anhydrous substance or on some other specified basis, the determination of loss on drying, water content or other property is carried out by the method prescribed in the relevant test in the monograph. The words 'dried substance' or 'anhydrous substance' etc. appear in parentheses after the result.

Where a quantitative determination of a residual solvent is carried out and a test for loss on drying is not carried out, the content of residual solvent is taken into account for the calculation of the assay content of the substance, the specific optical rotation and the specific absorbance. No further indication is given in the specific monograph.

Limits The limits prescribed are based on data obtained in normal analytical practice; they take account of normal analytical errors, of acceptable variations in manufacture and compounding and of deterioration to an extent considered acceptable. No further tolerances are to be applied to the limits prescribed to determine whether the article being examined complies with the requirements of the monograph.

In determining compliance with a numerical limit, the calculated result of a test or assay is first rounded to the number of significant figures stated, unless otherwise prescribed. The limits, regardless of whether the values are

expressed as percentages or as absolute values, are considered significant to the last digit shown (for example 140 indicates 3 significant figures). The last figure of the result is increased by one when the part rejected is equal to or exceeds one half-unit, whereas it is not modified when the part rejected is less than a half-unit.

Indication of permitted limit of impurities

The acceptance criteria for related substances are expressed in monographs either in terms of

comparison of peak areas (comparative tests) or as numerical values. For comparative tests, the approximate content of impurity tolerated, or the sum of impurities, may be indicated in brackets for information only.

Acceptance or rejection is determined on the basis of compliance or non-compliance with the stated test. If the use of a reference substance for the

named impurity is not prescribed, this content may be expressed as a

nominal concentration of the substance used to prepare the reference

solution specified in the monograph, unless otherwise described.

Herbal drugs For herbal drugs, the sulfated ash, total ash, water-soluble

matter, alcohol-soluble matter, water content, content of essential oil and

content of active principle are calculated with reference to the drug that has

not been specially dried, unless otherwise prescribed in the monograph.

Equivalents Where an equivalent is given, for the purposes of the

Pharmacopoeia only the figures shown are to be used in applying the

requirements of the monograph.

Culture media The culture media described in monographs and general

chapters have been found to be satisfactory for the intended purpose.

However, the components of media, particularly those of biological origin,

are of variable quality, and it may be necessary for optimal performance to

modulate the concentration of some ingredients, notably:

— peptones and meat or yeast extracts, with respect to their nutritive

properties;

— buffering substances;

— bile salts, bile extract, deoxycholate, and colouring matter, depending

on their selective properties;

— antibiotics, with respect to their activity.

Storage

The information and recommendations given under the heading Storage do

not constitute a pharmacopoeial requirement but the competent authority

may specify particular storage conditions that must be met.

The articles described in the Pharmacopoeia are stored in such a way as

to prevent contamination and, as far as possible, deterioration. Where

special conditions of storage are recommended, including the type of

container (see section 1.3. General chapters) and limits of temperature, they

are stated in the monograph.

The following expressions are used in monographs under Storage with

the meaning shown.

In an airtight container Means that the product is stored in an airtight

container (3.2). Care is to be taken when the container is opened in a damp

atmosphere. A low moisture content may be maintained, if necessary, by

the use of a desiccant in the container provided that direct contact with the

product is avoided.

Protected from light Means that the product is stored either in a

container made of a material that absorbs actinic light sufficiently to protect

the contents from change induced by such light, or in a container enclosed

in an outer cover that provides such protection, or is stored in a place from which all such light is excluded.

Labelling

In general, labelling of medicines is subject to supranational and national regulation and to international agreements. The statements under the heading Labelling are not therefore comprehensive and, moreover, for the purposes of the Pharmacopoeia only those statements that are necessary to demonstrate compliance or non-compliance with the monograph are mandatory. Any other labelling statements are included as recommendations. When the term 'label' is used in the Pharmacopoeia, the labelling statements may appear on the container, the package, a leaflet accompanying the package, or a certificate of analysis accompanying the article, as decided by the competent authority.

Warnings

Materials described in monographs and reagents specified for use in the Pharmacopoeia may be injurious to health unless adequate precautions are taken. The principles of good quality control laboratory practice and the provisions of any appropriate regulations are to be observed at all times. Attention is drawn to particular hazards in certain monographs by means of a warning statement; absence of such a statement is not to be taken to mean that no hazard exists.

Impurities

A list of all known and potential impurities that have been shown to be detected by the tests in a monograph may be given. See also chapter 5.10. *Control of impurities in substances for pharmaceutical use*. The impurities are designated by a letter or letters of the alphabet. Where a letter appears to be missing, the impurity designated by this letter has been deleted from the list during monograph development prior to publication or during monograph revision.

Functionality-Related Characteristics of Excipients

Monographs on excipients may have a section on functionality-related characteristics. The characteristics, any test methods for determination and any tolerances are not mandatory requirements; they may nevertheless be relevant for use of the excipient and are given for information (see also section 1.1. General statements).

Reference Standards

Certain monographs require the use of reference standards (chemical reference substances, herbal reference standards, biological reference preparations, reference spectra). See also chapter 5.12. *Reference standards*. The European Pharmacopoeia Commission establishes the official reference standards, which are alone authoritative in case of arbitration. These reference standards are available from the European Directorate for the Quality of Medicines & HealthCare (EDQM). Information on the available reference standards and a batch validity statement can be obtained via the EDQM website.

1.5. ABBREVIATIONS AND SYMBOLS

A	Absorbance	mp	Melting point
$A_{1\text{ cm}}^1$	Specific absorbance	n_D^{20}	Refractive index
A_r	Relative atomic mass	Ph. Eur. U.	European Pharmacopoeia Unit
$[\alpha]_D^{20}$	Specific optical rotation	ppb	Parts per billion (micrograms per kilogram)
bp	Boiling point	ppm	Parts per million (milligrams per kilogram)
BRP	Biological Reference Preparation	R	Substance or solution defined under 4. Reagents
CRS	Chemical Reference Substance	R_F	Retardation factor (see chapter 2.2.46)
d_{20}^{20}	Relative density	R_{st}	Used in chromatography to indicate the ratio of the distance travelled by a substance to the distance travelled by a reference substance
HRS	Herbal reference standard	RV	Substance used as a primary standard in volumetric analysis (chapter 4.2.1)
λ	Wavelength		
IU	International Unit		
M	Molarity		
M_r	Relative molecular mass		

LD_{50}	The statistically determined quantity of a substance that, when administered by the specified route, may be expected to cause the death of 50 per cent of the test animals within a given period	Lf dose	The quantity of toxin or toxoid that flocculates the test animals within a given period
L+/10 dose	The smallest quantity of a toxin that, in the conditions of the test, when mixed with 0.1 IU of antitoxin and administered by the specified route, causes the death of the test animals within a given period	CCID ₅₀	The statistically determined quantity of virus that may be expected to infect 50 per cent of the cell cultures to which it is added
L+ dose	The smallest quantity of a toxin that, in the conditions of the test, when mixed with 1 IU of antitoxin and administered by the specified route, causes the death of the test animals within a given period	EID ₅₀	The statistically determined quantity of virus that may be expected to infect 50 per cent of fertilised eggs into which it is inoculated
ID ₅₀	The statistically determined quantity of a virus that may be expected to infect 50 per cent of the animals into which it is inoculated	PD ₅₀	The statistically determined dose of a vaccine that, in the conditions of the test, may be expected to protect 50 per cent of the animals against a challenge dose of the micro-organisms or toxins against which it is active
Ir/100 dose	The smallest quantity of a toxin that, in the conditions of the test, when mixed with 0.01 IU of antitoxin and injected intracutaneously causes a characteristic reaction at the site of injection within a given period	BD ₅₀	The statistically determined dose of a vaccine that, in the conditions of the test, may be expected to induce specific antibodies in 50 per cent of the animals for the relevant vaccine antigens
Lp/10 dose	The smallest quantity of toxin that, in the conditions of the test, when mixed with 0.1 IU of antitoxin and administered by the specified route, causes paralysis in the test animals within a given period	PFU	Pock-forming units or plaque-forming units
		SPF	Specified-pathogen-free.

Abbreviations used in the monographs on immunoglobulins, immunosera and vaccines

Collections of micro-organisms

ATCC	American Type Culture Collection 10801 University Boulevard Manassas, Virginia 20110-2209, USA	NCTC	National Collection of Type Cultures Central Public Health Laboratory Colindale Avenue London NW9 5HT, Great Britain
C.I.P.	Collection de Bactéries de l'Institut Pasteur B.P. 52, 25 rue du Docteur Roux 75724 Paris Cedex 15, France	NCYC	National Collection of Yeast Cultures AFRC Food Research Institute Colney Lane Norwich NR4 7UA, Great Britain
IMI	International Mycological Institute Bakeham Lane Surrey TW20 9TY, Great Britain	NITE	Biological Resource Center Department of Biotechnology National Institute of Technology and Evaluation 2-5-8 Kazusakamatarai, Kisarazu-shi, Chiba, 292-0818 Japan
NCIMB	National Collection of Industrial and Marine Bacteria Ltd 23 St Machar Drive Aberdeen AB2 1RY, Great Britain	S.S.I.	Statens Serum Institut 80 Artager Boulevard, Copenhagen, Denmark
NCPF	National Collection of Pathogenic Fungi London School of Hygiene and Tropical Medicine Keppel Street London WC1E 7HT, Great Britain		

1.6. UNITS OF THE INTERNATIONAL SYSTEM (SI) USED IN THE PHARMACOPOEIA AND EQUIVALENCE WITH OTHER UNITS

International System Of Units (SI)
The International System of Units comprises 3 classes of units, namely base units, derived units and supplementary units¹. The base units and their definitions are set out in Table 1.6-1.

The derived units may be formed by combining the base units according to the algebraic relationships linking the corresponding quantities. Some of these derived units have special names and symbols. The SI units used in the Pharmacopoeia are shown in Table 1.6-2.
Some important and widely used units outside the International System are shown in Table 1.6-3.
The prefixes shown in Table 1.6-4 are used to form the names and symbols of the decimal multiples and submultiples of SI units.

¹ The definitions of the units used in the International System are given in the booklet "Le Système International d'Unités (SI)" published by the Bureau International des Poids et Mesures, Pavillon de Breteuil, F-92310 Sèvres.

Notes

1. In the Pharmacopoeia, the Celsius temperature is used (symbol t). This is defined by the following equation:

$$t = T - T_0$$

where $T_0 = 273.15$ K by definition. The Celsius or centigrade

temperature is expressed in degree Celsius (symbol $^{\circ}\text{C}$). The unit

'degree Celsius' is equal to the unit 'kelvin'.

2. The practical expressions of concentrations used in the Pharmacopoeia

are defined in the General Notices.

3. The radian is the plane angle between two radii of a circle that cut off

on the circumference an arc equal in length to the radius.

4. In the Pharmacopoeia, conditions of centrifugation are defined by

reference to the acceleration due to gravity (g):

$$g = 9.80665 \text{ m} \cdot \text{s}^{-2}$$

5. Certain quantities without dimensions are used in the Pharmacopoeia:

relative density (2.2.5), absorbance (2.2.25), specific absorbance

(2.2.25) and refractive index (2.2.6).

6. The microkatal is defined as the enzymic activity that, under defined

conditions, produces the transformation (e.g. hydrolysis) of 1 micromole

of the substrate per second.

Table 1.6.1. – SI base units

Quantity	Symbol	Name	Unit		Definition
			Symbol	Unit	
Length	l	metre	m		The metre is the length of the path travelled by light in a vacuum during a time interval of $1/299\,792\,458$ of a second.
Mass	m	kilogram	kg		The kilogram is equal to the mass of the international prototype of the kilogram.
Time	t	second	s		The second is the duration of 9 192 631 770 periods of the radiation corresponding to the transition between the two hyperfine levels of the ground state of the caesium-133 atom.
Electric current	I	ampere	A		The ampere is that constant current which, maintained in two straight parallel conductors of infinite length, of negligible circular cross-section and placed 1 metre apart in vacuum would produce between these conductors a force equal to 2×10^{-7} newton per metre of length.
Thermodynamic temperature	T	kelvin	K		The kelvin is the fraction $1/273.16$ of the thermodynamic temperature of the triple point of water.
Amount of substance	n	mole	mol		The mole is the amount of substance of a system containing as many elementary entities as there are atoms in 0.012 kilogram of carbon-12.
Luminous intensity	I_p	candela	cd		The candela is the luminous intensity in a given direction of a source emitting monochromatic radiation with a frequency of 540×10^{12} hertz and whose energy intensity in that direction is $1/683$ watt per steradian.

* When the mole is used, the elementary entities must be specified and may be atoms, molecules, ions, electrons, other particles or specified groups of such particles.

Table 1.6-2. – SI units used in the European Pharmacopoeia and equivalence with other units

Quantity		Unit				Conversion of other units into SI units	
Name	Symbol	Name	Symbol	Expression in SI base units	Expression in other SI units		
Wave number	ν	one per metre	$1/m$	m^{-1}			
Wavelength	λ	micrometre	μm	$10^{-6} m$			
Area	A, S	square metre	m^2	m^2			
Volume	V	cubic metre	m^3	m^3		$1 mL = 1 cm^3 = 10^{-6} m^3$	
Frequency	ν	hertz	Hz	s^{-1}			
Density	ρ	kilogram per cubic metre	kg/m^3	$kg \cdot m^{-3}$		$1 g/mL = 1 g/cm^3 = 10^3 kg \cdot m^{-3}$	
Velocity	v	metre per second	m/s	$m \cdot s^{-1}$		$1 dyne = 1 g \cdot cm \cdot s^{-2} = 10^{-5} N$	
Force	F	newton	N	$kg \cdot m \cdot s^{-2}$		$1 kp = 9.806 65 N$	
Pressure	p	pascal	Pa	$kg \cdot m^{-1} \cdot s^{-2}$	$N \cdot m^{-2}$	$1 dyne/cm^2 = 10^{-1} Pa = 10^{-1} N \cdot m^{-2}$	
Dynamic viscosity	η	pascal second	Pas	$kg \cdot m^{-1} \cdot s^{-1}$	$N \cdot s \cdot m^{-2}$	$1 atm = 101 325 Pa = 101.325 kPa$	
						$1 bar = 10^5 Pa = 0.1 MPa$	
Kinematic viscosity	ν	square metre per second	m^2/s	$m^2 \cdot s^{-1}$	$Pa \cdot s \cdot m^3 \cdot kg^{-1}$	$1 mm Hg = 133.322 387 Pa$	
						$1 Torr = 133.322 368 Pa$	
Energy	W	joule	J	$kg \cdot m^2 \cdot s^{-2}$	Nm	$1 cal = 4.1868 J$	
						$1 erg = 1 cm^2 \cdot g \cdot s^{-2} = 1 dyne \cdot cm = 10^{-7} J$	
Power	P	watt	W	$kg \cdot m^2 \cdot s^{-3}$	$N \cdot m \cdot s^{-1}$	$1 cal/s = 1 dyne \cdot cm \cdot s^{-1} = 10^{-7} W$	
Radiant flux						$10^{-7} W = 10^{-7} N \cdot m \cdot s^{-1} = 10^{-7} J \cdot s^{-1}$	
Absorbed dose (of radiant energy)	D	gray	Gy	$m^2 \cdot s^{-2}$	$J \cdot kg^{-1}$	$1 rad = 10^{-2} Gy$	
Electric potential, electromotive force	U	volt	V	$kg \cdot m^2 \cdot s^{-3} \cdot A^{-1}$	$W \cdot A^{-1}$		
Electric resistance	R	ohm	Ω	$m^2 \cdot kg \cdot s^{-3} \cdot A^{-2}$	$V \cdot A^{-1}$		
Quantity of electricity	Q	coulomb	C	As			
Activity of a radionuclide	A	becquerel	Bq	s^{-1}		$1 Ci = 37 \cdot 10^9 Bq = 37 \cdot 10^9 s^{-1}$	
Concentration (of amount of substance), molar concentration	c	mole per cubic metre	mol/m^3	$mol \cdot m^{-3}$		$1 mol/L = 1 M = 1 mol/dm^3 = 10^3 mol \cdot m^{-3}$	
Mass concentration	ρ	kilogram per cubic metre	kg/m^3	$kg \cdot m^{-3}$		$1 g/L = 1 g/dm^3 = 1 kg \cdot m^{-3}$	

Table 1.6-3. – Units used with the International System

Quantity	Unit		Value in SI units
	Name	Symbol	
Time	minute	min	1 min = 60 s
	hour	h	1 h = 60 min = 3600 s
	day	d	1 d = 24 h = 86 400 s
Plane angle	degree	°	1° = ($\pi/180$) rad
Volume	litre	L	1 L = 1 dm ³ = 10 ⁻³ m ³
Mass	tonne	t	1 t = 10 ³ kg
Rotational frequency	revolution per minute	r/min	1 r/min = (1/60) s ⁻¹

Table 1.6-4. – Decimal multiples and sub-multiples of units

Factor	Prefix	Symbol	Factor	Prefix	Symbol
10 ¹⁸	exa	E	10 ⁻¹	deci	d
10 ¹⁵	peta	P	10 ⁻²	centi	c
10 ¹²	tera	T	10 ⁻³	milli	m
10 ⁹	giga	G	10 ⁻⁶	micro	µ
10 ⁶	mega	M	10 ⁻⁹	nano	n
10 ³	kilo	k	10 ⁻¹²	pico	p
10 ²	hecto	h	10 ⁻¹⁵	femto	f
10 ¹	deca	da	10 ⁻¹⁸	atto	a

**Medicinal and Pharmaceutical
Substances J to Z**

Monographs

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Substances

for Pharmaceutical Use

(Ph. Eur. monograph 2034)

Ph. Eur.

DEFINITION

Substances for pharmaceutical use are any organic or inorganic substances that are used as active substances or excipients for the production of medicinal products for human or veterinary use. They may be obtained from natural sources or produced by extraction from raw materials, fermentation or synthesis.

This general monograph does not apply to herbal drugs, herbal drugs for homeopathic preparations, herbal drug preparations, extracts, or mother tinctures for homeopathic preparations, which are the subject of separate general monographs (*Herbal drugs* (1433), *Herbal drug preparations* (1434), *Extracts* (0765), *Mother tinctures for homeopathic preparations* (2029)). It does not apply to raw materials for homeopathic preparations, except where there is an individual monograph for the substance in the non-homeopathic part of the Pharmacopoeia.

Where a substance for pharmaceutical use is not described in an individual monograph of the Pharmacopoeia, it is used in a medicinal product prepared for the special needs of individual patients, the need for compliance with the present general monograph is decided in the light of a risk assessment that takes account of the available quality of the substance and its intended use.

Where medicinal products are manufactured using substances for pharmaceutical use of human or animal origin, the requirements of chapter 5.1.7. *Vital safety* apply. Substances for pharmaceutical use may be used as such or as starting materials for subsequent formulation to prepare medicinal products. Depending on the formulation, certain substances may be used either as active substances or as excipients. Solid substances may be compacted, coated, granulated, powdered to a certain fineness, or processed in other ways. A monograph is applicable to a substance processed with an excipient only where such processing is mentioned in the definition section of the monograph. *Substance for pharmaceutical use of special grade* Unless otherwise indicated or restricted in the individual monographs, a substance for pharmaceutical use is intended for human and veterinary use, and is of appropriate quality for the manufacture of all dosage forms in which it can be used.

Polyorphism Individual monographs do not usually specify crystalline or amorphous forms, unless bioavailability is affected. All forms of a substance for pharmaceutical use comply with the requirements of the monograph, unless otherwise indicated.

PRODUCTION

Substances for pharmaceutical use are manufactured by procedures that are designed to ensure a consistent quality and comply with the requirements of the individual monograph or approved specification.

The manufacture of active substances must take place under conditions of good manufacturing practice. The provisions of general chapter 5.10 apply to the control of impurities in substances for pharmaceutical use. Whether or not it is specifically stated in the individual monograph that the substance for pharmaceutical use:

— is a recombinant protein or another substance obtained as a direct gene product based on genetic modification, where applicable, the substance also complies with the requirements of the general monograph *Products of recombinant DNA technology* (0784);

— is obtained from animals susceptible to transmissible spongiform encephalopathies other than by experimental challenge, where applicable, the substance also complies with the requirements of the general monograph *Products with risk of transmitting agents of animal spongiform encephalopathies* (1483);

— is a substance derived from a fermentation process, whether or not the micro-organisms involved are modified by traditional procedures or recombinant DNA (rDNA) technology, where applicable, the substance also complies with the requirements of the general monograph *Products of fermentation* (1468).

If solvents are used during production, they are of suitable quality. In addition, their toxicity and their residual level are taken into consideration (5.4). If water is used during production, it is of suitable quality. If substances are produced or processed to yield a certain form or grade, that specific form or grade of the substance complies with the requirements of the monograph. Certain functionally-related tests may be described to control properties that may influence the suitability of the substance and subsequently the properties of dosage forms prepared from it.

Powdered substances May be processed to obtain a certain degree of fineness (2.9.35). *Compacted substances* Are processed to increase the particle size or to obtain particles of a specific form and/or to obtain a substance with a higher bulk density. *Coated active substances* Consist of particles of the active substance coated with one or more suitable excipients. *Granulated active substances* Are particles of a specified size and/or form produced from the active substance by granulation directly or with one or more suitable excipients. If substances are processed with excipients, these excipients comply with the requirements of the relevant monograph or, where no such monograph exists, the approved specification. Where active substances have been processed with excipients to produce, for example, coated or granulated substances, the processing is carried out under conditions of good manufacturing practice and the processed substances are regarded as intermediates in the manufacture of a medicinal product.

CHARACTERS

The statements under the heading *Characters* (e.g. statements about the solubility or a decomposition point) are not to be interpreted in a strict sense and are not requirements. They are given for information. Where a substance may show polymorphism, this may be stated under *Characters* in order to draw this to the attention of the user who may have to take this characteristic into consideration during formulation of a preparation.

IDENTIFICATION

Where under *Identification* an individual monograph contains subdivisions entitled 'First identification' and 'Second identification', the test or tests that constitute the 'First identification' may be used in all circumstances. The test or tests that constitute the 'Second identification' may be used in pharmacies provided it can be demonstrated that the substance or preparation is fully traceable to a batch



certified to comply with all the other requirements of the

monograph.

Certain monographs give two or more sets of tests for the

purpose of the first identification, which are equivalent and

may be used independently. One or more of these sets

usually contain a cross-reference to a test prescribed in the

Tests section of the monograph. It may be used to simplify

the work of the analyst carrying out the identification and the

prescribed tests. For example, one identification set cross-

refers to a test for enantiomeric purity while the other set

gives a test for specific optical rotation: the intended purpose

of the two is the same, that is, verification that the correct

enantiomer is present.

TESTS

Polymorphism (5.9)

If the nature of a crystalline or amorphous form imposes

restrictions on its use in preparations, the nature of the

specific crystalline or amorphous form is identified, its

morphology is adequately controlled and its identity is stated

Related substances

Unless otherwise prescribed or justified and authorised,

organic impurities in active substances are to be reported,

identified wherever possible, and qualified as indicated in

Table 2034-1 or in Table 2034-2 for peptides obtained by

chemical synthesis.

Table 2034-1. – Reporting, identification and qualification of organic impurities in active substances

Use	Maximum daily dose	Report- ing threshold	Identification threshold	Qualification threshold
Human use or human and veterinary use of > 1.0 mg daily intake of > 1.0 mg (whichever is the lower)	< 0.15 per cent or a daily intake of > 1.0 mg (whichever is the lower)	< 0.10 per cent or a daily intake of > 1.0 mg (whichever is the lower)	< 0.05 per cent	< 0.15 per cent
Human use or human and veterinary use of > 2 g/day	< 0.05 per cent	< 0.05 per cent	< 0.03 per cent	< 0.05 per cent
Veterinary use only	Not applicable	< 0.10 per cent	< 0.20 per cent	< 0.50 per cent

Table 2034-2. – Reporting, identification and qualification of organic impurities in peptides obtained by chemical synthesis

Reporting threshold	Identification threshold	Qualification threshold
> 0.1 per cent	> 0.5 per cent	> 1.0 per cent

Specific thresholds may be applied for impurities known to be unusually potent or to produce toxic or unexpected pharmacological effects.

If the individual monograph does not provide suitable control for a new impurity, a suitable test for control must be developed and included in the specification for the substance. The requirements above do not apply to biological and biotechnological products, oligonucleotides, radiopharmaceuticals, products of fermentation and semi-synthetic products derived therefrom, to crude products of animal or plant origin or herbal products.

For active substances in a new application for a medicinal product for human use, the requirements of the guideline on the limits of genotoxic impurities and the corresponding

LABELLING

In general, labelling is subject to supranational and national regulation and to international agreements. The statements

ASSAY

Unless justified and authorised, contents of substances for pharmaceutical use are determined. Suitable methods are used.

Additional properties

Control of additional properties (e.g. physical characteristics, functionality-related characteristics) may be necessary for individual manufacturing processes or formulations. Grades (such as sterile, endotoxin-free, pyrogen-free) may be produced with a view to manufacture of preparations for parenteral administration or other dosage forms and appropriate requirements may be specified in an individual monograph.

Pyrogens (2.6.8)

If the test for pyrogens is justified rather than the test for bacterial endotoxins and if a pyrogen-free grade is offered, the substance for pharmaceutical use complies with the test for pyrogens. The limit and test method are stated in the individual monograph or approved by the competent authority. Based on appropriate test validation for bacterial endotoxins and pyrogens, the test for bacterial endotoxins may replace the test for pyrogens.

Bacterial endotoxins (2.6.14)

If offered as bacterial endotoxin-free grade, the substance for pharmaceutical use complies with the test for bacterial endotoxins. The limit and test method (if not gelation method A) are stated in the individual monograph. The limit is calculated in accordance with the recommendations in general chapter 5.1.10. *Guidelines for using the test for bacterial endotoxins*, unless a lower limit is justified from results from production batches or is required by the competent authority. Where a test for bacterial endotoxins is prescribed, a test for pyrogens is not required.

Sterility (2.6.1)

If intended for use in the manufacture of sterile dosage forms without a further appropriate sterilisation procedure, or if offered as sterile grade, the substance for pharmaceutical use complies with the test for sterility.

acceptance criteria may be justified.

Microbiological quality

Individual monographs give acceptance criteria for microbiological quality wherever such control is necessary. Table 5.1.4-2. – *Acceptance criteria for microbiological quality of non-sterile substances for pharmaceutical use* in chapter 5.1.4. *Microbiological quality of non-sterile pharmaceutical preparations and substances for pharmaceutical use* gives recommendations on microbiological quality that are of general relevance for substances subject to microbial contamination. Depending on the nature of the substance and its intended use, different

Residual solvents

are limited according to the principles defined in chapter 5.4, using general method 2.4.24 or another suitable method. Where a quantitative determination of a residual solvent is carried out and a test for loss on drying is not carried out, the content of residual solvent is taken into account for calculation of the assay content of the substance, the specific optical rotation and the specific absorbance.

questions and answers documents published on the website of the European Medicines Agency (or similar evaluation principles for non-European Union member states) must be followed.

under the heading 'Labelling therefore are not comprehensive and, moreover, for the purposes of the Pharmacopoeia only those statements that are necessary to demonstrate compliance or non-compliance with the monograph are mandatory. Any other labelling statements are included as recommendations. When the term 'label' is used in the container, the package, a leaflet accompanying the package or a certificate of analysis accompanying the article, as decided by the competent authority.

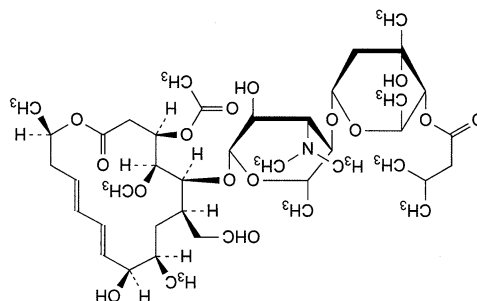
Where appropriate, the label states that the substance is: intended for a specific use; of a distinct crystalline form; of a specific degree of fineness; compacted; coated; granulated; sterile; free from bacterial endotoxins; free from pyrogens; containing gliding agents.

Where applicable, the label states: the degree of hydration; the name and concentration of any excipient.

Ph Eur

Josamycin

(Ph. Eur. monograph 1983)

C₄₂H₆₉NO₁₅ 828 16846-24-5

Action and use
Antibacterial.

Ph Eur

DEFINITION

Josamycin is a macrolide antibiotic obtained by fermentation using, for example, certain strains of *Streptomyces narbonensis* var. *josamyceticus* var. *nova*. The main component is (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-[2,6-dideoxy-3-C-methyl-4-O-(3-methylbutanoyl)-α-L-rbo-hexopyranosyl]-3-(dimethylamino)-β-D-glucopyranosyl]oxy]-10-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadecan-1,13-dien-2-one.

Content
Minimum 900 Ph. Eur. U/mg (dried substance).

CHARACTERS

Appearance

White or slightly yellowish powder, slightly hygroscopic.

Solubility
Very slightly soluble in water, freely soluble in methanol and in methylene chloride, soluble in acetone.

IDENTIFICATION

First identification A, C
Second identification A, B

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 0.10 g in methanol R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 50.0 mL with methanol R.

Spectral range 220–350 nm.

Absorption maximum At 232 nm.

Specific absorbance at the absorption maximum 330 to 370.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in 2.5 mL of methanol R.

Reference solution (a) Dissolve 10 mg of josamycin CRS in 2.5 mL of methanol R.

Reference solution (b) Dissolve 10 mg of josamycin propionate CRS in 2.5 mL of methanol R.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase methanol R, acetone R, ethyl acetate R, toluene R, hexane R (8:10:20:25:30 V/V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 100 °C for 10 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a) and different in position from the principal spot in the chromatogram obtained with reference solution (b).

C. Examine the chromatograms obtained in the test for related substances.

Results The principal peak in the chromatogram obtained with the test solution is similar in position and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₄ (2.2.2, Method II).

Dissolve 2.0 g in methanol R and dilute to 20 mL with the same solvent.

Specific optical rotation (2.2.7)

–65 to –75 (dried substance).
Dissolve 1.000 g in methanol R and dilute to 100.0 mL with the same solvent. Allow to stand for 30 min before measuring the angle of rotation.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, water R (30:70 V/V).
Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 25.0 mg of josamycin CRS in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 20.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 50.0 mL with the solvent mixture.

Reference solution (d) To 10 mL of the test solution add 0.1 mL of strong hydrogen peroxide solution R and heat in a water-bath for 10 min. Mix 1.0 mL of this solution and 1.0 mL of the test solution.

Reference solution (e) Dissolve 12.5 mg of josamycin for peak identification CRS (containing impurities A, B, C, D and E) in 5 mL of the solvent mixture.

Column:
— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 45 °C.

Mobile phase:
— mobile phase A: mix 3 volumes of a 67.9 g/L solution of tetrabutylammonium hydrogen sulfate R, 5 volumes of a 27.6 g/L solution of sodium dihydrogen phosphate monohydrate R adjusted to pH 3.0 with dilute phosphoric acid R, and dilute to 100 volumes with water R;

— mobile phase B: mix 5 volumes of a 27.6 g/L solution of sodium dihydrogen phosphate monohydrate R adjusted to pH 3.0 with dilute phosphoric acid R, and dilute to 100 volumes with water R;

— mobile phase C: mix 5 volumes of a 27.6 g/L solution of sodium dihydrogen phosphate monohydrate R adjusted to pH 3.0 with dilute phosphoric acid R, and dilute to 100 volumes with water R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 38	100	0
38 - 55	100 \rightarrow 0	0 \rightarrow 100

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 232 nm.

Injection 10 μ L of the test solution and reference solutions (b), (c), (d) and (e).

Identification of impurities Use the chromatogram supplied with josamycin for peak identification CRS and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to josamycin (retention time = about 35 min): impurity A = about 0.5; impurity B = about 0.8; impurity C = about 0.9; impurity D = about 1.2; impurity E = about 1.4.

System suitability Reference solution (d):
— resolution: minimum 1.7 between the 2 peaks due to josamycin and the peak eluted with a relative retention with reference to josamycin of about 1.1;
— retention time of josamycin: between 32 min and 38 min.

If necessary, adjust the concentration of acetonitrile in the mobile phases.

Limits:
— impurities A, B, C, D, E (any shoulder observed on the peak due to impurity A and/or the peak due to impurity B is not to be integrated separately): for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (5.0 per cent);
— any other impurity: not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
— total: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (20.0 per cent).

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Heavy metals (2.4.8)

Maximum 30 ppm.

1.0 g complies with test C. Prepare the reference solution using 3 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in vacuo at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 30.0 mg in 5 mL of methanol R and dilute to 100.0 mL with water R.

Carry out the microbiological assay of antibiotics (2.7.2).

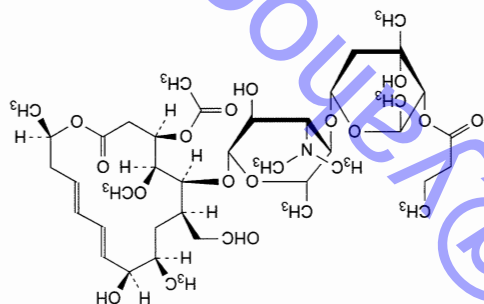
STORAGE

In an airtight container.

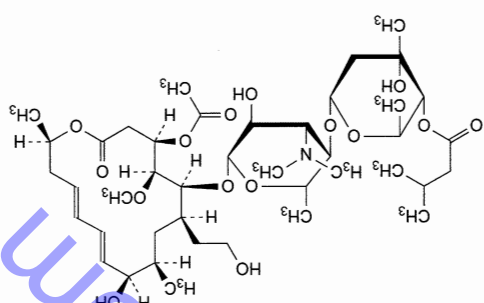
IMPURITIES

Specified impurities A, B, C, D, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): F, G, H, I, J, K.

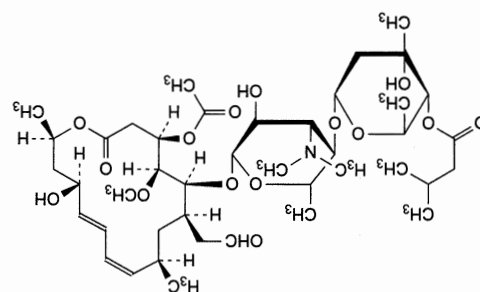


A. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-(2,6-dideoxy-3-C-methyl-α-L-rhamnopyranosyl)-3-(dimethylamino)-β-D-glucopyranosyl]oxy]-10-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,13-dien-2-one,

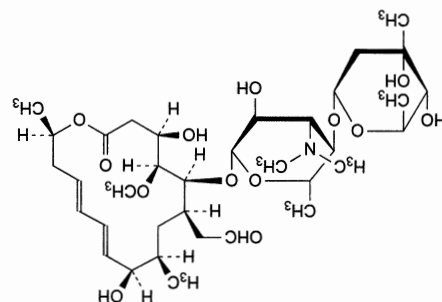


B. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-(2,6-dideoxy-3-C-methyl-α-L-rhamnopyranosyl)-3-(dimethylamino)-β-D-glucopyranosyl]oxy]-10-hydroxy-7-(2-hydroxyethyl)-5-methoxy-9,16-dimethyl-oxacyclohexadeca-1,13-dien-2-one,

C. unknown structure,

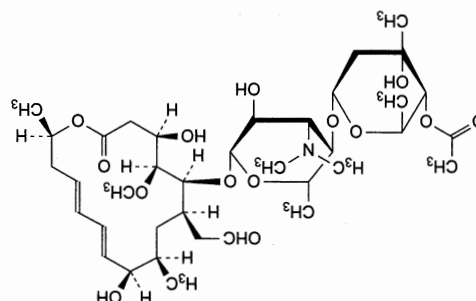


D. (4R,5S,6S,7R,9R,10Z,12E,14R,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-[(2,6-dideoxy-3-C-methyl-4-O-[(3-methylbutanoyl)- α -L-*ndo*-hexopyranosyl]-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-1,4-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-10,12-dien-2-one, (isofosamycin)

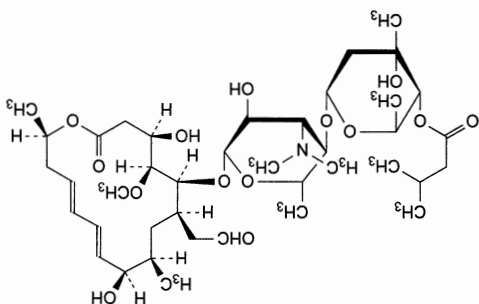


E. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-6-[[3,6-dideoxy-4-O-[(2,6-dideoxy-3-C-methyl-4-O-(3-methylbutanoyl)- α -L-*ndo*-hexopyranosyl]-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-1,0-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one,

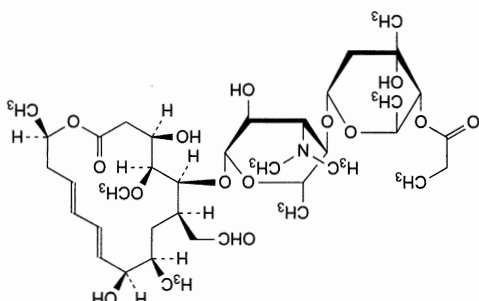
F. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-6-[[3,6-dideoxy-4-O-[(2,6-dideoxy-3-C-methyl- α -L-*ndo*-hexopyranosyl]-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-4,10-dihydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one,



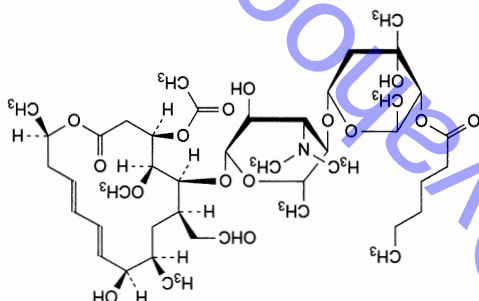
G. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-6-[[4-O-(4-O-acetyl-2,6-dideoxy-3-C-methyl- α -L-*ndo*-hexopyranosyl)-3,6-dideoxy-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-4,10-dihydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one,



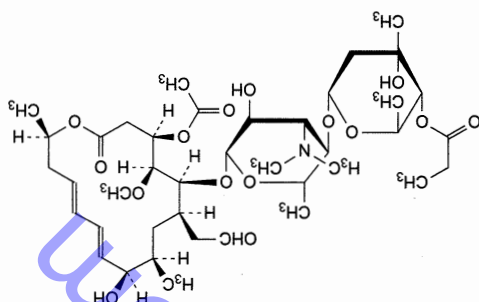
H. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-6-[[3,6-dideoxy-4-O-[(2,6-dideoxy-3-C-methyl-4-O-(3-methylbutanoyl)- α -L-*ndo*-hexopyranosyl]-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-4,10-dihydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one,



I. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-6-[[3,6-dideoxy-4-O-[(2,6-dideoxy-3-C-methyl-4-O-propanoyl)- α -L-*ndo*-hexopyranosyl]-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-4,10-dihydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one,



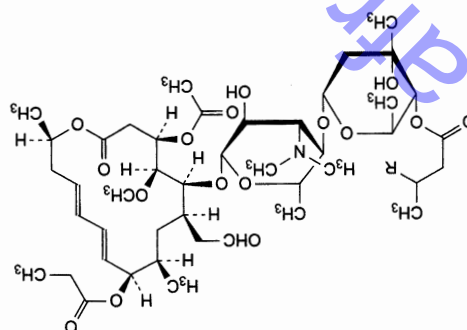
J. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-(2,6-dideoxy-4-O-hexanoyl-3-C-methyl- α -L-*ndo*-hexopyranosyl)-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-1,0-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one,



K. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-(2,6-dideoxy-3-C-methyl-4-O-propanoyl- α -L-*ndo*-hexopyranosyl)-3-(dimethylamino)- β -D-glucopyranosyl]oxy]-1,0-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadeca-1,1,1,3-dien-2-one.

Josamycin Propionate

(Ph. Eur. monograph 1982)



Leucomycin propionate	R	Mol. Formula	M _r
A3	CH ₃	C ₄₅ H ₇₃ NO ₁₆	884
A4	H	C ₄₄ H ₇₁ NO ₁₆	870

40922-77-8

(leucomycin A3 propionate)

Action and use

Antibacterial.

Ph. Eur.

DEFINITION

Propionyl ester of a macrolide antibiotic produced by certain strains of *Streptomyces narbonensis* var. *josamyceticus* var. *nova*, or obtained by any other means. The main component is (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-[2,6-dideoxy-3-C-methyl-4-O-

(3-methylbutanoyl)-α-L-rbo-hexopyranosyl]-3-(dimethylamino)-β-D-glucopyranosyl]oxy]-5-methoxy-9,16-

dimethyl-7-(2-oxoethyl)-10-(propanoyloxy)oxacyclohexadeca-

1,1,3-dien-2-one propionate (leucomycin A3 propionate).

Semi-synthetic product derived from a fermentation product.

Content

— minimum 843 Ph. Eur. U./mg (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline, slightly hygroscopic powder.

Solubility

Practically insoluble in water, freely soluble in methanol and in methylene chloride, soluble in acetone.

IDENTIFICATION

First identification A, B

Second identification B, C

Prepare solutions in methanol immediately before use.

A. Dissolve 0.10 g in methanol R and dilute to 100.0 mL

with the same solvent. Dilute 1.0 mL of the solution to

50.0 mL with methanol R. Examined between 220 nm and

350 nm (2.2.25), the solution shows an absorption maximum

at 231 nm. The specific absorbance at the absorption

maximum is 310 to 350.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in methanol R and dilute to 1 mL with the same solvent.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₄ (2.2.2, Method II).

Dissolve 1 g in methanol R and dilute to 10 mL with the

same solvent.

Specific optical rotation (2.2.7)

Dissolve 1.000 g in methanol R and dilute to 100.0 mL with

the same solvent. Allow to stand for 30 min before

measuring the angle of rotation.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be

examined in acetonitrile for chromatography R and dilute to

100.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of josamycin

propionate CRS in acetonitrile for chromatography R and dilute

to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of the substance to be

examined in 10 mL of methanol R and add 40 µL of dilute

phosphoric acid R. Mix, allow to stand for 5 min and inject.

Reference solution (c) Dilute 2.0 mL of reference solution (a)

to 100.0 mL with acetonitrile for chromatography R.

Column:

— size: l = 0.15 m, Ø = 3.9 mm,

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 µm),

— temperature: 30 °C.

Mobile phase acetonitrile R, a 15.4 g/L solution of ammonium

acetate R previously adjusted to pH 6.0 with dilute phosphoric

acid R (60:40 V/V).

Ph Eur



Kanamycin Acid Sulfate

Kanamycin Acid Sulphate
(Ph. Eur. monograph 0033)

Action and use
Aminoglycoside antibacterial.

DEFINITION

Kanamycin acid sulfate is a form of kanamycin sulfate prepared by adding sulfuric acid to a solution of kanamycin monosulfate and drying by a suitable method. The potency is not less than 670 IU/mg, calculated with reference to the dried substance.

Fermentation product.

PRODUCTION

It is produced by methods of manufacture designed to eliminate or minimise substances lowering blood pressure. The method of manufacture is validated to demonstrate that the product if tested would comply with the following test.

Abnormal toxicity (2.6.9)

Inject into each mouse 0.5 mL of a solution containing 2 mg per millilitre of the substance to be examined.

CHARACTERS

A white or almost white powder, hygroscopic, soluble in about 1 part of water, practically insoluble in acetone and in alcohol.

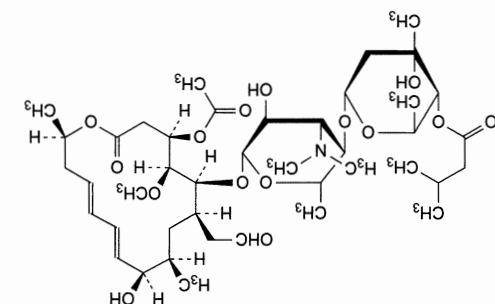
IDENTIFICATION

A. Examine by thin-layer chromatography (2.2.27), using a plate coated with a 0.75 mm layer of the following mixture: mix 0.3 g of carborer R with 240 mL of water R and allow to stand, with moderate shaking, for 1 h; adjust to pH 7 by the gradual addition, with continuous shaking, of dilute sodium hydroxide solution R and add 30 g of silica gel H R.

Heat the plate at 110 °C for 1 h, allow to cool and use immediately.

Test solution Dissolve 10 mg of the substance to be examined in water R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of kanamycin monosulfate CRS in water R and dilute to 10 mL with the same solvent.



B. (4R,5S,6S,7R,9R,10R,11E,13E,16R)-4-(acetyloxy)-6-[[3,6-dideoxy-4-O-[2,6-dideoxy-3-C-methyl-4-O-(3-methylbutanoyl)-α-L-ribo-hexopyranosyl]-3-(dimethylamino)-β-D-glucopyranosyl]oxy]-10-hydroxy-5-methoxy-9,16-dimethyl-7-(2-oxoethyl)oxacyclohexadecan-11,13-dien-2-one (fosamycin).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 232 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 3 times the retention time of leucomycin A3

proportionate.

Relative retention With reference to leucomycin A3 proportionate

(retention time = about 18 min): impurity B = about 0.2;

impurity A = about 0.3; impurity B = about 0.5;

leucomycin A4 proportionate = about 0.7;

impurity C = about 1.4; impurity D = about 2.0.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the 2 peaks eluting with a relative retention with reference to leucomycin A3

proportionate of about 0.5 and 0.7 respectively.

Limits:

— impurity D: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (c),

— impurities A, B, C, E: for each impurity, not more than

the area of the principal peak in the chromatogram

obtained with reference solution (c),

— any other impurity: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (c),

— total: not more than 7 times the area of the principal peak

in the chromatogram obtained with reference solution (c),

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (c).

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven *in vacuo* at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 40.0 mg in 20 mL of methanol R and dilute to

100.0 mL with phosphate buffer solution pH 5.6 R.

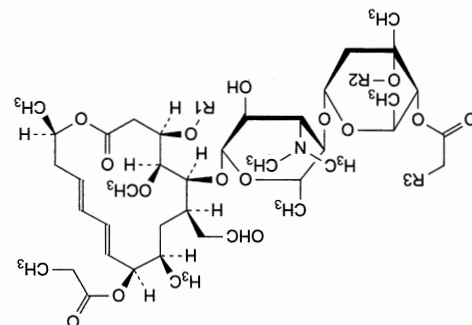
Carry out the microbiological assay of antibiotics (2.7.2).

STORAGE

In an airtight container.

IMPURITIES

Specified impurities A, B, C, D, E



A. R1 = CO-CH₃, R2 = R3 = H:

leucomycin A8 9-proportionate,

B. R1 = R2 = H, R3 = C₂H₅: leucomycin A5 9-proportionate,

C. R1 = CO-C₂H₅, R2 = H, R3 = CH(CH₃)₂:

platenomycin A1 9-proportionate,

D. R1 = CO-CH₃, R2 = CO-C₂H₅, R3 = CH(CH₃)₂:

leucomycin A3 3'',9-dipropionate,

Reference solution (b) Dissolve 10 mg of kanamycin monosulfate CRS, 10 mg of neomycin sulfate CRS and 10 mg of streptomycin sulfate CRS in water R and dilute to 10 mL with the same solvent.

Apply separately to the plate 10 µL of each solution. Develop over a path of 12 cm using a 70 g/L solution of potassium

dihydrogen phosphate R. Dry the plate in a current of warm air and spray with a mixture of equal volumes of a 2 g/L

solution of *1,3-dihydroxyphenylalanine* R in alcohol R and a 460 g/L solution of *sulfuric acid* R. Heat at 150 °C for 5 min

to 10 min. The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to

the principal spot in the chromatogram obtained with reference solution (a). The test is not valid unless the

chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

B. Dissolve 0.5 g in 10 mL of water R. Add 10 mL of *picric acid* solution R. Initiate crystallisation if necessary by

scratching the wall of the tube with a glass rod and allow to stand. Collect the crystals, wash with 20 mL of water R and

filter. Dry at 100 °C. The crystals melt (2.2.14) at about 235 °C, with decomposition.

C. Dissolve about 50 mg in 2 mL of water R. Add 1 mL of a 10 g/L solution of *minhydrin* R and heat for a few minutes on

a water-bath. A violet colour develops. **D.** It gives the reactions of sulfates (2.3.1).

TESTS

Solution S Dissolve 0.20 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

pH (2.2.3) The pH of solution S is 5.5 to 7.5.

Specific optical rotation (2.2.7) +103 to +115, determined on solution S and calculated with

reference to the dried substance. **Kanamycin B**

Examine by thin-layer chromatography (2.2.27), using a plate prepared as prescribed under identification test A.

Heat the plate at 110 °C for 1 h, allow to cool and use immediately.

Test solution Dissolve 0.11 g of the substance to be examined in water R and dilute to 20 mL with the same solvent.

Reference solution B sulfate CRS Dissolve 4 mg of kanamycin B sulfate CRS in water R and dilute to 20 mL with the same solvent.

Apply separately to the plate 4 µL of each solution. Develop over a path of 12 cm using a 70 g/L solution of potassium

dihydrogen phosphate R. Dry the plate in a current of warm air and spray with *minhydrin* and *stannous chloride reagent* R. Heat

the plate at 110 °C for 15 min. Any spot corresponding to kanamycin B in the chromatogram obtained with the test

solution is not more intense than the spot in the chromatogram obtained with the reference solution

(4.0 per cent). **Loss on drying** (2.2.32) Not more than 5.0 per cent, determined on 1.00 g by drying

at 60 °C at a pressure not exceeding 670 Pa for 3 h. **Sulfated ash** (2.4.14) Not more than 0.5 per cent, determined on 1.0 g.

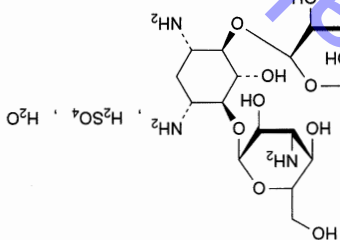
Sulfate 23.0 per cent to 26.0 per cent of sulfate (SO₄), calculated with reference to the dried substance. Dissolve 0.175 g in

100 mL of water R and adjust the solution to pH 11 using concentrated ammonia R. Add 10.0 mL of 0.1 M barium

Kanamycin Sulfate

Kanamycin Sulphate

(Kanamycin Monosulfate, Ph Eur monograph 0032)



C₁₈H₃₈N₄O₁₅S₂H₂O 601

Action and use

Aminoglycoside antibacterial.

DEFINITION

6-O-(3-Amino-3-deoxy-α-D-glucopyranosyl)-4-O-(6-amino-6-deoxy-α-D-glucopyranosyl)-2-deoxy-D-streptamine sulfate. Antimicrobial substance produced by the growth of certain strains of *Streptomyces kanamyceticus*.

Content

Minimum 750 IU/mg (dried substance).

PRODUCTION

It is produced by methods of manufacture designed to eliminate or minimise substances lowering blood pressure. The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test. **Abnormal toxicity** (2.6.9) Inject into each mouse 0.5 mL of a 2 mg/mL solution of the substance to be examined.

CHARACTERS

Appearance

White or almost white, crystalline powder.



Ph Eur

Solubility

Freely soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in water R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of kanamycin monosulfate CRS in water R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of kanamycin monosulfate CRS, 10 mg of neomycin sulfate CRS and 10 mg of streptomycin sulfate CRS in water R and dilute to 10 mL with the same solvent.

Plate Suitable plate coated with a 0.75 mm layer of a mixture prepared as follows: mix 0.3 g of carbomer R with 240 mL of water R and allow to stand, with moderate shaking, for 1 h; adjust to pH 7 by the gradual addition, with continuous shaking, of dilute sodium hydroxide solution R and add 30 g of silica gel H R.

Pre-treatment Heat the plate at 110 °C for 1 h, allow to cool and use immediately.

Mobile phase 70 g/L solution of potassium dihydrogen phosphate R.

Application 10 µL.

Development Over a path of 12 cm.

Drying In a current of warm air.

Detection Spray with a mixture of equal volumes of 2 g/L solution of 1,3-dihydroxynaphthalene R in ethanol

(96 per cent) R and a 460 g/L solution of sulfuric acid R. Heat at 150 °C for 5 min to 10 min.

System suitability: reference solution (b):

— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

B. Dissolve 0.5 g in 10 mL of water R. Add 10 mL of picric acid solution R. Initiate crystallisation if necessary by scratching the wall of the tube with a glass rod and allow to stand. Collect the crystals, wash with 20 mL of water R and filter. Dry at 100 °C. The crystals melt (2.2.14) at about 235 °C, with decomposition.

C. Dissolve about 50 mg in 2 mL of water R. Add 1 mL of a 10 g/L solution of ninhydrin R and heat for a few minutes on a water-bath. A violet colour develops.

D. It gives the reactions of sulfates (2.3.1).

TESTS**Solution S**

Dissolve 0.20 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

pH (2.2.3)

6.5 to 8.5 for solution S.

Specific optical rotation (2.2.7)

+ 112 to + 123 (dried substance), determined on solution S.

Kanamycin B

Thin-layer chromatography (2.2.27) as described under Identification A with the following modifications.

Test solution Dissolve 0.1 g of the substance to be examined in water R and dilute to 20 mL with the same solvent.

Reference solution Dissolve 4 mg of kanamycin B sulfate CRS in water R and dilute to 20 mL with the same solvent.

IDENTIFICATION

Practically insoluble in water and in organic solvents.

Solubility

Fine, white or greyish-white, unctuous powder.

Appearance**CHARACTERS**

composition.

Purified, natural, hydrated aluminium silicate of variable composition.

DEFINITION

Ph Eur

Kaolin (Natural) is required.

When kaolin is prescribed or demanded, Light Kaolin shall be dispensed or supplied, unless it is ascertained that Light Kaolin (Natural) is required.

Preparation

(Ph. Eur. monograph 0503)

Heavy Kaolin

Ph Eur

container.

If the substance is sterile, store in a sterile, tamper-proof container.

STORAGE

Carry out the microbiological assay of antibiotics (2.7.2).

ASSAY

injections R.

solution of the substance to be examined in water for

Inject per kilogram of the rabbit's mass 1 mL of a 10 mg/mL solution of the substance to be examined in water for

removal of pyrogens, it complies with the test for pyrogens. preparations without a further appropriate procedure for the

If intended for use in the manufacture of parenteral

Pyrogens (2.6.8)

SO₄.

1 mL of 0.1 M barium chloride is equivalent to 9.606 mg of

violet-blue colour disappears.

solution begins to change and continue the titration until the

50 mL of ethanol (96 per cent) R when the colour of the

phthalate purple R. Titrate with 0.1 M sodium edetate adding

solution to pH 11 with concentrated ammonia R.

Dissolve 0.250 g in 100 mL of water R and adjust the

15.0 per cent to 17.0 per cent of sulfate (dried substance).

Sulfate

Maximum 0.5 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

60 °C at a pressure not exceeding 670 Pa for 3 h.

Maximum 1.5 per cent, determined on 1.000 g by drying at

Loss on drying (2.2.32)

with the reference solution (4.0 per cent).

— **kanamycin B:** any spot corresponding to kanamycin B in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained

Limit:

Heat at 110 °C for 15 min.

Detection Spray with ninhydrin and stannous chloride reagent R.

Application 4 µL.

A. To 0.5 g in a metal crucible add 1 g of potassium nitrate R and 3 g of sodium carbonate R and heat until the mixture melts. Allow to cool. To the residue add 20 mL of boiling water R, mix and filter. Wash the residue with 50 mL of

water R. To the residue add 1 mL of *hydrochloric acid R* and 5 mL of *water R*. Filter. To the filtrate add 1 mL of *strong sodium hydroxide solution R* and filter. To the filtrate add 3 mL of *ammonium chloride solution R*. A gelatinous white precipitate is formed.

B. Add 2.0 g in 20 portions to 100 mL of a 10 g/L solution of *sodium laurylsulfate R* in a 100 mL graduated cylinder about 30 mm in diameter. Allow 2 min between additions for each portion to settle. Allow to stand for 2 h. The apparent volume of the sediment is not greater than 5 mL.

C. 0.25 g gives the reaction of silicates (2.3.1).

TESTS

Solution S
To 4 g add a mixture of 6 mL of *acetic acid R* and 34 mL of *distilled water R*, shake for 1 min and filter.

Acidity or alkalinity
To 1.0 g add 20 mL of *carbon dioxide-free water R*, shake for 2 min and filter. To 10 mL of the filtrate add 0.1 mL of *phenolphthalein solution R*. The solution is colourless. Not more than 0.25 mL of 0.01 M *sodium hydroxide* is required to change the colour of the indicator to pink.

Organic impurities
Heat 0.3 g to redness in a calcination tube. The residue is only slightly more coloured than the original substance.

Adsorption power
To 1.0 g in a ground-glass-stoppered test-tube add 10.0 mL of a 3.7 g/L solution of *methylene blue R* and shake for 2 min. Allow to settle. Centrifuge and dilute the solution 1 to 100 with *water R*. The solution is not more intensely coloured than a 0.03 g/L solution of *methylene blue R*.

Swelling power
Triturate 2 g with 2 mL of *water R*. The mixture does not flow.

Substances soluble in dilute hydrochloric acid
Maximum 1 per cent.
To 5.0 g add 7.5 mL of *dilute hydrochloric acid R* and 27.5 mL of *water R* and boil for 5 min. Filter, wash the residue on the filter with *water R* and dilute the combined filtrate and washings to 50.0 mL with *water R*. To 10.0 mL of the solution add 1.5 mL of *dilute sulfuric acid R*, evaporate to dryness on a water-bath and ignite. The residue weighs a maximum of 10 mg.

Chlorides (2.4.4)
Maximum 250 ppm.
Dilute 2 mL of solution S to 15 mL with *water R*.
Sulfates (2.4.13)
Maximum 0.1 per cent.
Dilute 1.5 mL of solution S to 15 mL with *distilled water R*.
Calcium (2.4.3)
Maximum 250 ppm.
Dilute 4 mL of solution S to 15 mL with *distilled water R*.
Extractable heavy metals (2.4.8)
Maximum 50 ppm.

To 5 mL of the solution prepared for the test for substances soluble in *dilute hydrochloric acid* add 5 mL of *water R*, 10 mL of *hydrochloric acid R* and 25 mL of *methyl isobutyl ketone R*. Shake for 2 min. Separate the layers. Evaporate the aqueous layer to dryness on a water-bath. Dissolve the residue in 1 mL of *acetic acid R* and dilute to 25 mL with *water R*. Filter. 12 mL of the solution complies with test A. Prepare the reference solution using *lead standard solution* (1 ppm Pb) R.

Light Kaolin

Action and use

Antidiarrhoeal.

Preparations

Kaolin Mixture

Kaolin and Morphine Mixture

When Kaolin or Light Kaolin is prescribed or demanded, Light Kaolin shall be dispensed or supplied unless it is ascertained that Light Kaolin (Natural) is required.

DEFINITION

Light Kaolin is a native hydrated aluminium silicate, freed from most of its impurities by elutriation and dried. It contains a suitable dispersing agent.

CHARACTERISTICS

A light, white powder free from gritty particles; odourless or almost odourless; unctuous.

Practically insoluble in *water* and in mineral acids.

IDENTIFICATION

A. To 0.5 g in a metal crucible add 1 g of *potassium nitrate* and 3 g of *sodium carbonate* and heat until the mixture melts. Allow to cool. To the residue add 20 mL of *boiling water*, mix and filter. Wash the residue with 50 mL of *water*, add to the residue 1 mL of *hydrochloric acid* and 5 mL of *water*, mix and filter. To the filtrate add 1 mL of *strong sodium hydroxide solution*, filter and add to the filtrate 3 mL of *ammonium chloride solution*. A gelatinous white precipitate is produced.

B. 0.25 g yields the reaction characteristic of *silicates*, Appendix VI.

C. Triturate 2 g with 2 mL of *water*. The resulting mixture flows.

TESTS

Coarse particles

Transfer 5 g to a stoppered cylinder (about 16 cm × 35 mm), add 60 mL of a 1% w/v solution of *sodium pyrophosphate*, shake thoroughly and allow to stand for 5 minutes. Using a pipette, withdraw 50 mL from a point about 5 cm below the surface of the liquid. To the remaining liquid add 50 mL of *water*, shake, allow to stand for 5 minutes and withdraw 50 mL in the same manner as

DEFINITION

Light Kaolin (Natural) is a native hydrated aluminium silicate, freed from most of its impurities by elutriation and dried. It does not contain a dispersing agent.

CHARACTERISTICS

A light, white powder free from gritty particles; odourless or almost odourless; unctuous.
Practically insoluble in water and in mineral acids.

IDENTIFICATION

A. To 0.5 g in a metal crucible add 1 g of potassium nitrate

and 3 g of sodium carbonate and heat until the mixture melts. Allow to cool. To the residue add 20 mL of boiling water.

the residue 1 mL of hydrochloric acid and 5 mL of water, mix and filter. Wash the residue with 50 mL of water, add to

and filter. To the filtrate add 1 mL of strong sodium hydroxide solution, filter and add to the filtrate 3 mL of ammonium

chloride solution. A gelatinous white precipitate is produced.

B. 0.25 g yields the reaction characteristic of silicates,

Appendix VI.

C. Triturate 2 g with 2 mL of water. The resulting mixture does not flow.

TESTS

Coarse particles; Arsenic; Heavy metals; Chloride;

Loss on drying; Loss on ignition; Soluble matter

Complies with the requirements stated under Light Kaolin.

Fine particles

Disperse 5 g in 250 mL of water containing 50 mg of sodium pyrophosphate by shaking vigorously for 2 minutes in a

stoppered flask, pour immediately into a glass cylinder 5 cm in diameter and transfer 20 mL to a glass dish using a

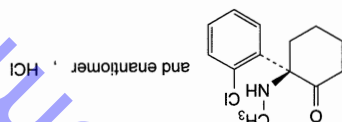
pipette. Evaporate to dryness and dry to constant weight at 105°. Allow the remainder of the suspension to stand for

4 hours at 20° and withdraw a second 20 mL portion using a pipette with its tip exactly 5 cm below the surface and

without disturbing the sediment. Transfer the second portion to a glass dish, evaporate to dryness and dry to constant

weight at 105°. The weight of the residue from the second portion is not less than 70% of the weight of the residue

(Ph Eur monograph 1020)

Ketamine Hydrochloride

$C_{13}H_{17}Cl_2NO$

274.2

1867-66-9

Ph Eur

DEFINITION

(2*RS*)-2-(2-Chlorophenyl)-2-(methylamino)cyclohexanone

hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Light Kaolin (Natural)**Action and use**

Antidiarrhoeal.

Preparations

Kaolin Mixture

Kaolin and Morphine Mixture

When Kaolin or Light Kaolin is prescribed or demanded, Light Kaolin shall be dispensed or supplied unless it is

ascertained that Light Kaolin (Natural) is required.

Boil 2 g with 100 mL of 0.2*M* hydrochloric acid under a reflux condenser for 5 minutes, cool, filter and evaporate 50 mL of the filtrate to dryness. The residue, after ignition at about 600° for 30 minutes, weighs not more than 10 mg.

Soluble matter

weight. Use 1 g.

When ignited at 600°, loses not more than 15.0% of its

Loss on ignition

1.5% of its weight. Use 1 g.

When dried to constant weight at 105°, loses not more than

Loss on drying

Appendix VII (330 ppm).

15 mL of the filtrate complies with the limit test for chlorides,

under a reflux condenser for 5 minutes, cool and filter.

Boil 1.0 g with 80 mL of water and 20 mL of 2*M* nitric acid

Chloride

(20 ppm).

standard solution (1 ppm Pb) to prepare the standard

with limit test A for heavy metals, Appendix VII. Use lead

produce 60 mL. 12 mL of the resulting solution complies

aqueous layer add 2 g of citric acid and sufficient water to

mixture of equal volumes of isomyl alcohol and ether. To the

thiocyanate and extract with two 10 mL quantities of a

of water, 2 g of ammonium chloride and 2 g of ammonium

0.5 mL of nitric acid and evaporate to low bulk. Add 20 mL

hydrochloric acid and filter. To 40 mL of the filtrate add

water bath with a mixture of 70 mL of water and 10 mL of

Heat 6.0 g for 15 minutes under a reflux condenser on a

Heavy metals

test for arsenic, Appendix VII (2 ppm).

0.50 g dispersed in 25 mL of water complies with the limit

Arsenic

residue from the first portion.

the second portion is not less than 70% of the weight of the

to constant weight at 105°. The weight of the residue from

second portion to a glass dish, evaporate to dryness and dry

surface and without disturbing the sediment. Transfer the

portion using a pipette with its tip exactly 5 cm below the

stand for 4 hours at 20° and withdraw a second 20 mL

weight at 105°. Allow the remainder of the suspension to

using a pipette. Evaporate to dryness and dry to constant

cylinder 5 cm in diameter and transfer 20 mL to a glass dish

2 minutes in a stoppered flask, pour immediately into a glass

Disperse 5 g in 250 mL of water by shaking vigorously for

Fine particles

drying at 105°, weighs not more than 25 mg.

and evaporate to dryness on a water bath. The residue, after

suspension has been withdrawn under the prescribed

before. Repeat the operation until a total of 400 mL of

Solubility

Freely soluble in water and in methanol, soluble in ethanol (96 per cent).

mp

About 260 °C, with decomposition.

IDENTIFICATION

A. Optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison ketamine hydrochloride CRS.

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

3.5 to 4.1.

Dilute 10 mL of solution S to 20 mL with carbon dioxide-free water R.

Optical rotation (2.2.7)

−0.2° to +0.2°.

Dilute 2.5 mL of solution S to 25.0 mL with water R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a)

Dissolve 5 mg of ketamine impurity A CRS in the mobile phase and dilute to 10.0 mL with the mobile phase (using ultrasound if necessary).

To 1.0 mL of the solution, add 0.5 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Column:

size: $l = 0.125$ m, $\phi = 4.0$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve 0.95 g of sodium hexanesulfonate R in 1 L of a mixture of 25 volumes of acetonitrile R1 and 75 volumes of water R and add 4 mL of acetic acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 μ L.

Run time 10 times the retention time of ketamine.

Relative retention With reference to ketamine (retention time = about 3 min): impurity A = about 1.6;

impurity B = about 3.3; impurity C = about 4.6.

System suitability: reference solution (a):

— resolution: minimum 1.5 between the peaks due to ketamine and impurity A.

Limits:

— impurities A, B, C: for each impurity, not more than area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— total: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference

solution (b) (0.5 per cent);

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dilute 10 mL of solution S to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of methanol R and add 1.0 mL of 0.1 M hydrochloric acid. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.

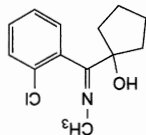
1 mL of 0.1 M sodium hydroxide is equivalent to 27.42 mg of $C_{13}H_{17}Cl_2NO$.

STORAGE

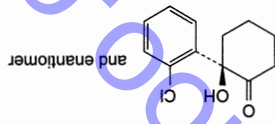
Protected from light.

IMPURITIES

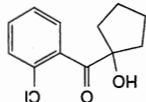
Specified impurities A, B, C



A. 1-(2-chloro-N-methylbenzimidoyl)cyclopentanol,



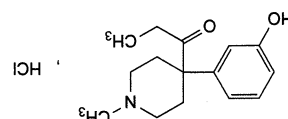
B. (2RS)-2-(2-chlorophenyl)-2-hydroxycyclohexanone,



C. (2-chlorophenyl)(1-hydroxycyclopentyl)methanone.

Ketobemidone Hydrochloride

(Ph Eur monograph 1746)


 $C_{15}H_{22}ClNO_2$ 283.8 5965-49-1

Action and use
Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION

1-[4-(3-Hydroxyphenyl)-1-methylpiperidin-4-yl]propan-1-one hydrochloride.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of ketobemidone hydrochloride.

B. Solution S (see Tests) gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.250 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₈ (2.2.2, Method II).

pH (2.2.3)

4.5 to 5.5 for solution S.

Related substances

Liquid chromatography (2.2.29).

Solution A 1.54 g/L solution of ammonium acetate R adjusted to pH 8.0 with dilute ammonia R1.

Test solution Dissolve 50.0 mg of the substance to be examined in solution A and dilute to 25.0 mL with the same solution.

Reference solution (a) Dissolve 1 mg of ketobemidone impurity B CRS and 1 mg of ketobemidone impurity C CRS in solution A and dilute to 25 mL with the same solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 20.0 mL of this solution to 100.0 mL with solution A.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

stationary phase: phenylhexylsilyl silica gel for chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase acetonitrile R, solution A (20:80 V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 278 nm.
Injection 20 μ L.
Run time 4.5 times the retention time of ketobemidone.

Relative retention With reference to ketobemidone (retention time = about 10 min): impurity A = about 0.4; impurity B = about 0.6; impurity C = about 0.7; impurity D = about 3.5.

System suitability: reference solution (a):
— resolution: minimum 4.0 between the peaks due to impurity B and impurity C.

Limits:

— impurities A, B, C, D: for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference

solution (b) (0.7 per cent);

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.50 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 5.0 mL of

0.01 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20) using

0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 28.38 mg of $C_{15}H_{22}ClNO_2$.

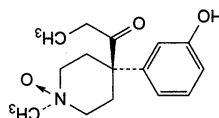
IMPURITIES

Specified impurities A, B, C, D

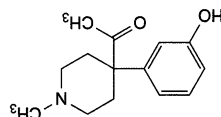
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E.

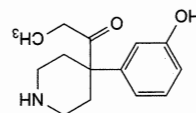
and trans isomer



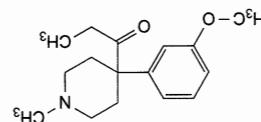
A. 1-[4-(3-hydroxyphenyl)-1-methyl-1-oxidopiperidin-4-yl]propan-1-one (cis and trans isomers),



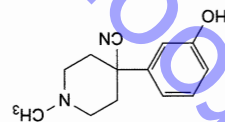
B. 1-[4-(3-hydroxyphenyl)-1-methylpiperidin-4-yl]ethanone,



C. 1-[4-(3-hydroxyphenyl)piperidin-4-yl]propan-1-one,



D. 1-[4-(3-methoxyphenyl)-1-methylpiperidin-4-yl]propan-1-one,

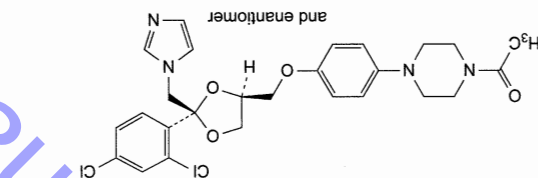


E. 4-(3-hydroxyphenyl)-1-methylpiperidin-4-carbonitrile.

Ph Eur

Ketoconazole

(Ph. Eur. monograph 0921)

C₂₆H₂₈Cl₂N₄O₄ 531.4 65277-42-1Antifungal.
Action and use

Preparation

Ketoconazole Cream

Ketoconazole Shampoo

Ph Eur

DEFINITION

1-Acetyl-4-[4-[(2R,5,4SR)-2-(2,4-dichlorophenyl)-2-(1H-imidazol-1-yl)methyl]-1,3-dioxolan-4-yl]methoxy]piperazine.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B

Second identification A, C, D

A. Melting point (2.2.14): 148 °C to 152 °C.
B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison ketoconazole CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 5 mL with the mobile phase.

Reference solution (a) Dissolve 30 mg of ketoconazole CRS in the mobile phase and dilute to 5 mL with the mobile phase.

Reference solution (b) Dissolve 30 mg of ketoconazole CRS and 30 mg of econazole nitrate CRS in the mobile phase, then dilute to 5 mL with the mobile phase.

Plate TLC octadecylsilyl silica gel plate R.

Mobile phase ammonium acetate solution R, dioxan R, methanol R (20:40:40 V/V/V).

Application 5 µL.

Development Over a path of 15 cm.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To about 30 mg in a porcelain crucible add 0.3 g of anhydrous sodium carbonate R. Heat over an open flame for 10 min. Allow to cool. Take up the residue with 5 mL of dilute nitric acid R and filter. To 1 mL of the filtrate add 1 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S Dissolve 1.0 g in methylene chloride R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₄ (2.2.2, Method II).

Optical rotation (2.2.7)

−0.10° to + 0.10°, determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2.5 mg of ketoconazole CRS and 2.5 mg of loperamide hydrochloride CRS in methanol R, then dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Column:

— size: l = 0.10 m, Ø = 4.6 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (3 µm).

Mobile phase:

— mobile phase A: acetonitrile R1, 3.4 g/L solution of tetrabutylammonium hydrogen sulfate R (50:50 V/V);

— mobile phase B: acetonitrile R1, 3.4 g/L solution of tetrabutylammonium hydrogen sulfate R (5:95 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	100 → 0	0 → 100
10 - 15	0	100

Flow rate 2 mL/min.

Detection Spectrophotometer at 220 nm.

Equilibration With acetonitrile R for at least 30 min and then

with mobile phase A for at least 5 min.

Injection 10 µL; inject methanol R as a blank.

Retention time Ketocanazole = about 6 min;

loperamide = about 8 min.

System suitability: reference solution (a):

— resolution: minimum 15 between the peaks due to

ketocanazole and loperamide; if necessary, adjust the final

concentration of acetonitrile in the mobile phase or adjust

the time programme for the linear gradient elution.

Limits:

— total: not more than the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.5 per cent);

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 70 mL of a mixture of 1 volume of

anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R.

Titrate with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

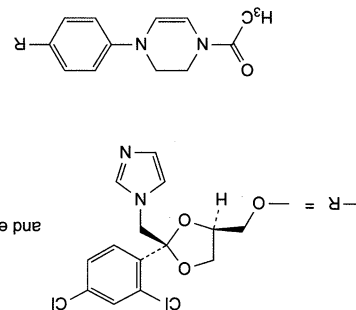
1 mL of 0.1 M perchloric acid is equivalent to 26.57 mg

of $C_{26}H_{28}Cl_2N_4O_4$.

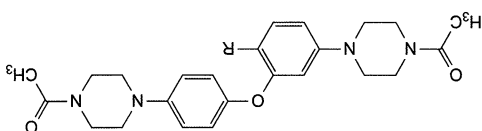
STORAGE

Protected from light.

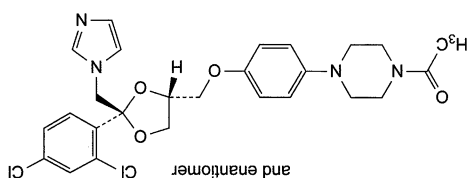
IMPURITIES



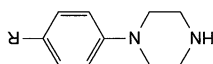
A. 1-acetyl-4-[4-[[[(2R,4S)-2-((2,4-dichlorophenyl)-2-(1H-imidazol-1-ylmethyl)-1,3-dioxolan-4-yl)methoxy]phenyl]1,2,3,4-tetrahydropyrazine,



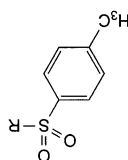
B. 1-acetyl-4-[4-[[[(2R,4S)-2-((2,4-dichlorophenyl)-2-(1H-imidazol-1-ylmethyl)-1,3-dioxolan-4-yl)methoxy]-3-[4-(4-acetyl)piperazin-1-yl]phenoxy]phenyl]piperazine,



C. 1-acetyl-4-[4-[[[(2R,4S)-2-((2,4-dichlorophenyl)-2-(1H-imidazol-1-ylmethyl)-1,3-dioxolan-4-yl)methoxy]phenyl]piperazine,



D. 1-[4-[[[(2R,4S)-2-((2,4-dichlorophenyl)-2-(1H-imidazol-1-ylmethyl)-1,3-dioxolan-4-yl)methoxy]phenyl]piperazine,



E. [[(2R,4S)-2-((2,4-dichlorophenyl)-2-(1H-imidazol-1-ylmethyl)-1,3-dioxolan-4-yl)methoxy]phenyl]piperazine,

Ketoprofen (Ph. Eur. monograph 0922)

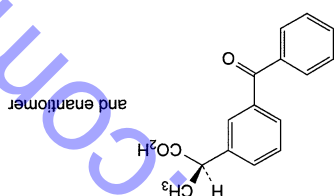


Ph Eur

$C_{16}H_{14}O_3$

254.3

22071-15-4



Ketoprofen

(Ph. Eur. monograph 0922)

Preparations
Ketoprofen Capsules

Ketoprofen Gel

Content

99.0 per cent to 100.5 per cent (dried substance).

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Melting point (2.2.14): 94 °C to 97 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 50.0 mL with ethanol (96 per cent) R.

Spectral range 230-350 nm.

Absorption maximum At 255 nm.

Specific absorbance at the absorption maximum 615 to 680.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison ketoprofen CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in acetone R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of ketoprofen CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of indometacin CRS in acetone R and dilute to 10 mL with the same solvent.

To 1 mL of this solution add 1 mL of reference solution (a).

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase glacial acetic acid R, methylene chloride R, acetone R (1:49:50 V/V/V).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results

The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference

solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 1.0 g in acetone R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of ketoprofen impurity A CRS in the mobile phase and dilute to 50.0 mL

with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of ketoprofen impurity C CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. To 1.0 mL of this solution add 1.0 mL of reference solution (b).

Column:

— size: l = 0.15 m, Ø = 4.6 mm;

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 µm) with a specific surface area of 350 m²/g and a pore size of 10 nm.

Mobile phase Mix 2 volumes of freshly prepared phosphate buffer solution pH 3.5 R, 43 volumes of acetonitrile R and 55 volumes of water R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 233 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity C.

Run time 7 times the retention time of ketoprofen.

Relative retention With reference to ketoprofen (retention time = about 7 min): impurity C = about 0.3;

impurity B = about 0.69; impurity A = about 1.5;

impurity D = about 1.35; impurity A = about 1.5;

impurity F = about 2.0.

System suitability Reference solution (d):

— resolution: minimum 7.0 between the peaks due to ketoprofen and impurity A.

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— impurity C: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— impurities B, D, E, F: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— sum of impurities other than A and C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying at 60 °C at a pressure not exceeding 0.67 kPa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

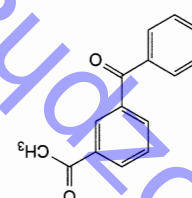
ASSAY

Dissolve 0.200 g in 25 mL of ethanol (96 per cent) R. Add 25 mL of water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M sodium hydroxide is equivalent to 25.43 mg of $C_{16}H_{14}O_3$.

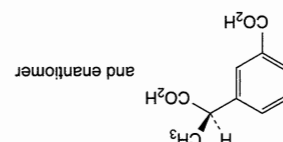
IMPURITIES

Specified impurities A, B, C, D, E, F

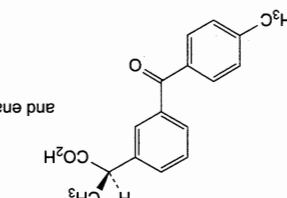
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): G, H, I, J, K, L.



A. 1-(3-benzoylphenyl)ethanone,

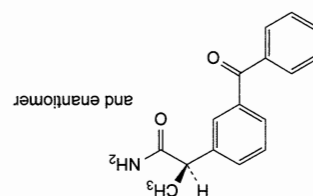


B. (3-benzoylphenyl)acetic acid,

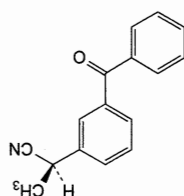


C. 3-[(1R)-1-carboxyethyl]benzoic acid,

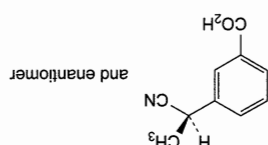
D. (2R)-2-[3-(4-methylbenzoyl)phenyl]propanoic acid,



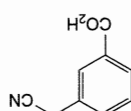
E. (2R)-2-(3-benzoylphenyl)propanamide,



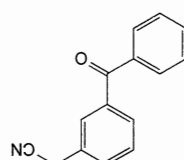
F. (2R)-2-(3-benzoylphenyl)propanenitrile,



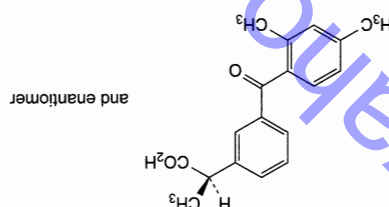
G. 3-[(1R)-1-cyanoethyl]benzoic acid,



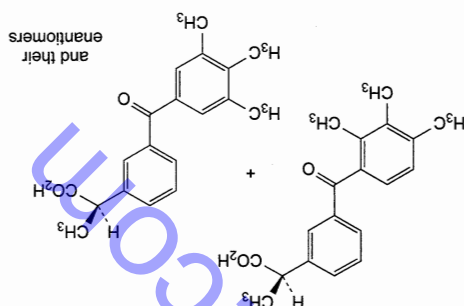
H. 3-(cyanomethyl)benzoic acid,



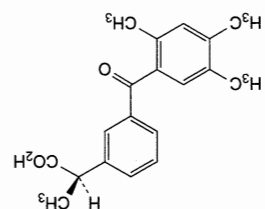
I. (3-benzoylphenyl)ethanenitrile,



J. (2R)-2-[3-(2,4-dimethylbenzoyl)phenyl]propanoic acid,



K. mixture of (2R)-2-[3-(2,3,4-trimethylbenzoyl)phenyl]propanoic acid and (2R)-2-[3-(3,4,5-trimethylbenzoyl)phenyl]propanoic acid,

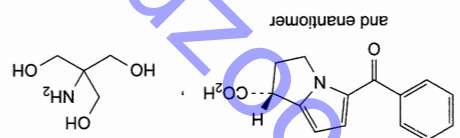


L. (2*R,S*)-2-[3-(2,4,5-trimethylbenzoyl)phenyl]propanoic acid.

Ph Eur

Ketorolac Trometamol

(Ph. Eur. monograph 1755)



74103-07-4

$C_{19}H_{24}N_2O_6$

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION

2-Amino-2-(hydroxymethyl)propane-1,3-diol (1*R,S*)-5-benzoyl-2,3-dihydro-1*H*-pyrrolizine-1-carboxylate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in methanol, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison ketorolac trometamol CRS.

TESTS

Solution S

Dissolve 0.75 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1).

pH (2.2.3)

5.7 to 6.7.

Dilute 5 mL of solution S to 15 mL with carbon dioxide-free water R.

Absorbance (2.2.25)

Maximum 0.10, determined at 430 nm for solution S.

Related substances

Liquid chromatography (2.2.29). Protect the solutions from bright light.

Solvent mixture tetrahydrofuran R, water R (30:70 V/V).

Test solution Dissolve 20 mg of the substance to be examined in the solvent mixture and dilute to 50 mL with the solvent mixture.

of $C_{19}H_{24}N_2O_6$.

1 mL of 0.1 M perchloric acid is equivalent to 37.64 mg

potentiometrically (2.2.20).

with 0.1 M perchloric acid, determining the end-point

Dissolve 0.300 g in 60 mL of anhydrous acetic acid R. Titrate

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 3 h.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

using 2 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test F. Prepare the reference solution

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

solution (a) (1.0 per cent);

peak in the chromatogram obtained with reference

— total: not more than 10 times the area of the principal

with reference solution (a) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram

— impurities A, B, C, D: for each impurity, not more than

impurity B = 0.52; impurity C = 2.2;

corresponding correction factor: impurity A = 0.67;

the peak areas of the following impurities by the

— correction factors: for the calculation of content, multiply

Limits:

impurity B and ketorolac.

— resolution: minimum 1.5 between the peaks due to

System suitability: reference solution (b):

impurity B = about 0.9.

impurity A = about 0.6; impurity D = about 0.7;

time = about 10 min; impurity C = about 0.5;

Relative retention With reference to ketorolac (retention

identify the peaks due to impurities A, B, C and D.

chromatogram obtained with reference solution (b) to

with ketorolac trometamol for peak identification CRS and the

Identification of impurities Use the chromatogram supplied

Run time 3 times the retention time of ketorolac.

Injection 10 µL.

Detection Spectrophotometer at 313 nm.

Flow rate 1.5 mL/min.

1000 mL with water R.

adjust to pH 3.0 with phosphoric acid R and dilute to

70 volumes of a solution prepared as follows: dissolve 5.75 g

Mobile phase Mix 30 volumes of tetrahydrofuran R with

— temperature: 40 °C.

(5 µm);

— stationary phase: octylsilyl silica gel for chromatography R

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

Column:

in 5 mL of the solvent mixture.

Reference solution (b) Dissolve 2 mg of ketorolac trometamol for

peak identification CRS (containing impurities A, B, C and D)

solution to 100.0 mL with the solvent mixture.

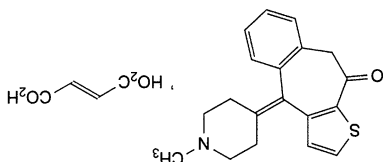
10.0 mL with the solvent mixture. Dilute 1.0 mL of this

Reference solution (a) Dilute 1.0 mL of the test solution to



Ketotifen Fumarate

(Ketotifen Hydrogen Fumarate,
Ph Eur monograph 1592)



$C_{23}H_{23}NO_5$ 425.5 34580-14-8

Action and use

Histamine H_1 receptor antagonist.

Ph Eur

DEFINITION

4-(1-Methylpiperidin-4-ylidene)-4,9-dihydro-10H-benzof[4,5-c]cyclohepta[1,2-b]thiophen-10-one hydrogen (E)-butenedioate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or brownish-yellow, fine, crystalline powder.

Solubility

Sparingly soluble in water, slightly soluble in methanol,

Practically insoluble in heptane.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison ketotifen hydrogen fumarate CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₄, BY₄ or B₄ (2.2.2, Method II). Dissolve 0.2 g in methanol R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture methanol R, water R (50:50 V/V).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

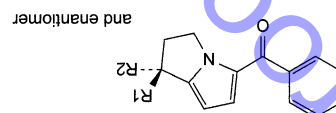
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of ketotifen impurity G CRS in 1.0 mL of a solution prepared as follows: mix 1.0 mL of the test solution with 9.0 mL of the solvent mixture. Sonicate until dissolution of impurity G is complete.

Reference solution (c) Dissolve 5 mg of ketotifen for peak identification CRS (containing impurity A) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.0$ mm;
— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (3 μ m);
— temperature: 40 °C.

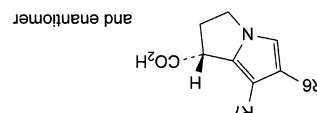


G, H, I, J.
Control of impurities in substances for pharmaceutical use): E, F, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these by the general monograph Substances for pharmaceutical use acceptance criterion for other/unspecified impurities and/or present at a sufficient level, be detected by one or other of Other detectable impurities (the following substances would, if Specified impurities A, B, C, D

IMPURITIES

Protected from light.

STORAGE



A, R1 = H, R2 = OH: (1RS)-5-benzoyl-2,3-dihydro-1H-pyrrrolizin-1-ol,
B, R1 + R2 = O: 5-benzoyl-2,3-dihydro-1H-pyrrrolizin-1-one,
D, R1 = CO₂H, R2 = OCH₃: (1RS)-5-benzoyl-1-methoxy-2,3-dihydro-1H-pyrrrolizin-1-carboxylic acid,
E, R1 = H, R2 = CO-NH-C(CH₂OH)₃: (1RS)-5-benzoyl-N-[2-hydroxy-1,1-bis(hydroxymethyl)ethyl]-2,3-dihydro-1H-pyrrrolizin-1-carboxamide,
G, R1 = CO₂CH₃, R2 = OH: methyl (1RS)-5-benzoyl-1-hydroxy-2,3-dihydro-1H-pyrrrolizin-1-carboxylate,
H, R1 = H, R2 = CO₂CH₃: methyl (1RS)-5-benzoyl-2,3-dihydro-1H-pyrrrolizin-1-carboxylate,
I, R1 = R2 = H: phenyl(2,3-dihydro-1H-pyrrrolizin-5-yl)methanone,
J, R1 = H, R2 = CO₂C₆H₅: ethyl (1RS)-5-benzoyl-2,3-dihydro-1H-pyrrrolizin-1-carboxylate,
C, R6 = CO-C₆H₅, R7 = H: (1RS)-6-benzoyl-2,3-dihydro-1H-pyrrrolizin-1-carboxylic acid,
F, R6 = H, R7 = CO-C₆H₅: (1RS)-7-benzoyl-2,3-dihydro-1H-pyrrrolizin-1-carboxylic acid.

Ph Eur

Mobile phase:
— mobile phase A: mix 175 µL of triethylamine R and 500 mL of water R;
— mobile phase B: mix 175 µL of triethylamine R and 500 mL of methanol R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 12	40	60
12 - 20	40 → 10	60 → 90
20 - 25	10	90

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 297 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied with ketotifen for peak identification *CRS* and the chromatogram obtained with reference solution (c) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity G.

Relative retention With reference to ketotifen retention

time = about 11 min; fumaric acid = about 0.1;

impurity G = about 0.8; impurity A = about 1.9.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to

impurity G and ketotifen.

Limits:

— correction factor: for the calculation of content, multiply the

peak area of impurity G by 1.4;

— impurity A: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.2 per cent);

— impurity G: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.05 per cent); disregard the peak due to fumaric acid.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in a mixture of 30 mL of acetic anhydride R

and 30 mL of anhydrous acetic acid R. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 42.55 mg of

C₂₃H₂₃NO₅.

IMPURITIES

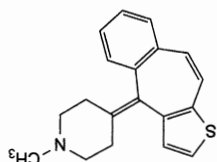
Specified impurities: A, G.

Other detectable impurities (the following substances would, if

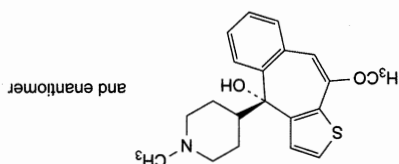
present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

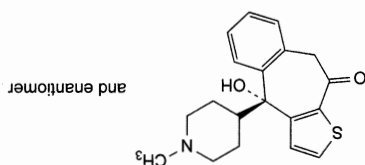
acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: B, C, D, E, F.



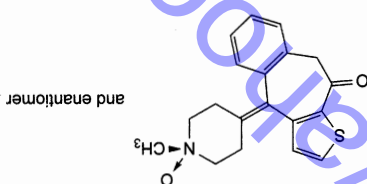
A. 4-(4H-benzo[4,5]cyclohepta[1,2-b]thiophen-4-ylidene)-1-methylpiperidine,



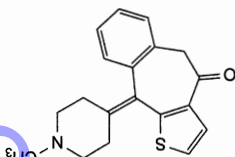
B. (4RS)-4-(10-methoxy-4-(1-methylpiperidin-4-yl)-4H-benzo[4,5]cyclohepta[1,2-b]thiophen-4-yl)-10H-benzo[4,5]cyclohepta[1,2-b]thiophen-10-one,



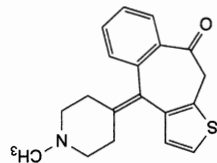
C. (4RS)-4-(10-hydroxy-4-(1-methylpiperidin-4-yl)-4H-benzo[4,5]cyclohepta[1,2-b]thiophen-4-yl)-10H-benzo[4,5]cyclohepta[1,2-b]thiophen-10-one,



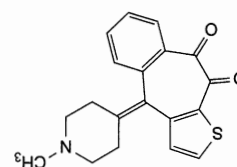
D. 4-[(R,S)-1-methylpiperidin-4-ylidene]-4,9-dihydro-10H-benzo[4,5]cyclohepta[1,2-b]thiophen-10-one N-oxide (ketotifen N-oxide),



E. 10-(1-methylpiperidin-4-ylidene)-5,10-dihydro-4H-benzo[5,6]cyclohepta[1,2-b]thiophen-4-one,



F. 4-(1-methylpiperidin-4-ylidene)-4,10-dihydro-9H-benzo[4,5]cyclohepta[1,2-b]thiophen-9-one,

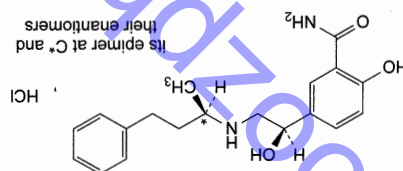


G. 4-(1-methylpiperidin-4-ylidene)-4H-benzo[4,5]cyclohepta[1,2-b]thiophen-9,10-dione.

Ph Eur

Labetalol Hydrochloride

(Ph. Eur. monograph 0923)



$C_{19}H_{25}ClN_2O_3$ 364.9 32780-64-6

Action and use

Alpha- and beta-adrenoceptor antagonist.

Preparations

Labetalol Injection

Labetalol Tablets

DEFINITION

Mixture of 4 stereoisomers of 2-hydroxy-5-[1-hydroxy-2-[(1-methyl-3-phenylpropyl)amino]ethyl]benzamide hydrochloride.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Sparsely soluble in water and in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification A, C, E

Second identification A, B, D, E

A. Optical rotation (2.2.7): -0.05° to $+0.05^\circ$, determined on solution S (see Tests).

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 25.0 mg in 0.1 M hydrochloric acid and dilute to 250.0 mL with the same acid.

Spectral range 230-350 nm.

Absorption maximum At 302 nm.

Specific absorbance at the absorption maximum 83 to 88.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison labetalol hydrochloride CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in 1 mL of ethanol (96 per cent) R.

TESTS

Solution S

Dissolve 0.50 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent. Solution S must be freshly prepared.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

pH (2.2.3)

4.0 to 5.0 for solution S.

Diastereoisomer ratio

Gas chromatography (2.2.28).

Test solution Dissolve 2.0 mg of the substance to be examined in 1.0 mL of a 12.0 g/L solution of boric acid R in anhydrous pyridine R and allow to stand for 20 min.

Column:

material: glass;

size: $l = 1.5$ m, $\varnothing = 4$ mm;

stationary phase: silanised diatomaceous earth for gas chromatography R (125-150 μ m) impregnated with 3 per cent m/m of polymethylphenylsiloxane R.

Carrier gas nitrogen for chromatography R.

Flow rate 40 mL/min.

Temperature:

column, injection port and detector: 300 $^\circ$ C.

Detection Flame ionisation.

Injection 2 μ L.

System suitability:

the height of the trough separating the 2 peaks due to the pairs of diastereoisomers is less than 5 per cent of the full scale of the recorder.

Limit:

each pair of diastereoisomers: for the area of each peak, 45 per cent to 55 per cent of the total area of the 2 peaks.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Test solution (b) Dilute 1.0 mL of test solution (a) to 50.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with mobile phase A. Dissolve 5 mg of labetalol impurity A CRS in this solution.

Reference solution (c) Dissolve 25.0 mg of labetalol hydrochloride CRS in mobile phase A and dilute to 10.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 50.0 mL with mobile phase A.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl amorphous

organosilica polymer R (3.5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: phosphoric acid R, water R (0.1:99.9 V/V);

— mobile phase B: acetonitrile R, mobile phase A (50:50 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	100	0
5 - 40	100 \rightarrow 0	0 \rightarrow 100
40 - 45	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to labetalol (retention time = about 22 min): impurity A = about 1.1.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to labetalol and impurity A.

Calculation of percentage contents:

— for each impurity, use the concentration of labetalol in reference solution (a).

Limits:

— unspecified impurities: for each impurity, maximum

0.05 per cent;

— total: maximum 0.2 per cent;

— reporting threshold: 0.03 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent ethanol (96 per cent) R.

0.25 g complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C at a pressure not exceeding 0.7 kPa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

IMPURITIES

hydrochloride CRS.

Calculate the percentage content of $C_{19}H_{25}ClN_2O_3$ taking into account the assigned content of labetalol

Retention time Labetalol = about 2 min.

Run time Twice the retention time of labetalol.

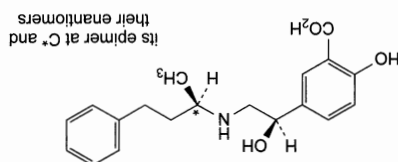
Injection Test solution (b) and reference solution (c).

Mobile phase Mobile phase A, mobile phase B (45:55 V/V).

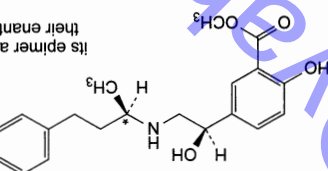
Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

ASSAY

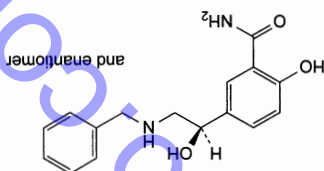
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, G.



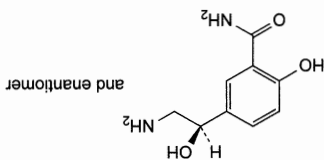
A. mixture of 4 stereoisomers of 2-hydroxy-5-[1-[(1-methyl-3-phenylpropyl)amino]ethyl]benzoic acid.



B. mixture of 4 stereoisomers of methyl 2-hydroxy-5-[1-[(1-methyl-3-phenylpropyl)amino]ethyl]benzoate.



C. 5-[(1R,2S)-2-(benzylamino)-1-hydroxyethyl]-2-hydroxybenzamide.



D. 5-[(1R,2S)-2-(benzylamino)-1-hydroxyethyl]-2-hydroxybenzamide.

Practically insoluble in water; freely soluble in acetone; sparingly soluble in absolute ethanol.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of Lisdipine (RS 407). B. In the Assay, the principal peak in the chromatogram obtained with solution (1) has the same retention time as the principal peak in the chromatogram obtained with solution (2).

TESTS

Propan-2-ol

Carry out the method for gas chromatography, Appendix III B. Prepare a 0.002% v/v solution of 1,4-dioxan (internal standard) in dimethylacetamide (solution A).
(1) 0.002% v/v solution of *propen-2-ol* in solution A.
(2) 2% w/v of the substance being examined in solution A.

CHROMATOGRAPHIC CONDITIONS

- (a) Use a glass column (60 m \times 0.32 mm) bonded with a film (5 μ m) of *polymethylsiloxane* (CP-Sil 5CB is suitable).
- (b) Use *helium* as the carrier gas at 1.7 mL per minute.
- (c) Use a temperature gradient as described below.
- (d) Use an injection temperature of 170°.
- (e) Use a detector temperature of 250°.
- (f) Inject 1 μ L of each solution.

Time (Minutes)	Temperature	Comment
0 - 1	60°	isothermal
1 - 18	60° → 110°	linear increase 3°/minute
18 - 20	110° → 200°	linear increase 50°/minute
20 - 27	200°	isothermal

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with solution (1) shows two clearly separated peaks. The retention time for propan-2-ol is about 6.2 minutes and that for dioxan is about 15 minutes.

In the chromatogram obtained with solution (2):
the percentage content of propan-2-ol is not more than 0.5% w/w.

Related substances

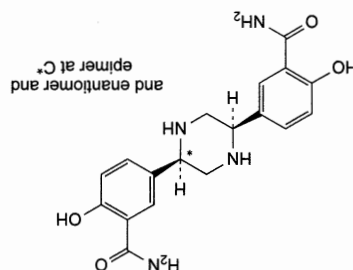
Appendix III D, using the following solutions.

(1) Dilute 1 volume of a 0.1% solution of the substance being examined in *absolute ethanol* to 5 volumes with the mobile phase.

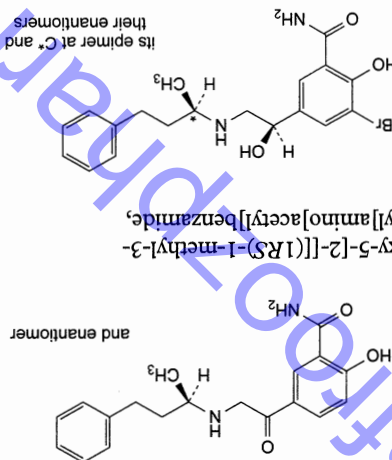
(2) Dilute 1 volume of solution (1) to 500 volumes with the mobile phase.

(3) Dilute 1 volume of a 0.1% w/v solution of lacticidipine impurity standard BPCRS in absolute ethanol to 5 volumes with the mobile phase.

E. mixture of 3 stereoisomers of 5,5'-piperazine-2,5'-diylbis(2-hydroxybenzamide),



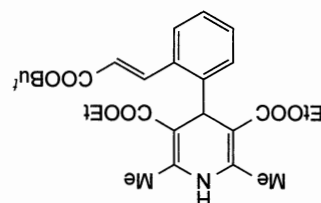
F, 2-hydroxy-5-[2-[[[(1*RS*)-1-methyl-3-phenylpropyl]amino]acetyl]benzamide,



G. mixture of 4 stereoisomers of 3-bromo-2-hydroxy-5-[1-hydroxy-2-[(1-methyl-3-phenylpropyl)amino]ethyl]benzamide.

phenylpropyl)amino]ethyl]benzamide.

Lacidipine


$$\text{C}_{26}\text{H}_{33}\text{NO}_6 \quad 455.6 \quad 103890-78-4$$

DEFINITION

Lacidipine is diethyl (B)-4-[2-(*tert*-butoxycarbonyl)vinyl]piperidine-3,5-dicarboxylate. It contains not less than 97.5% and not more than 102.0% of $C_{22}H_{27}NO_6$, calculated with reference to the anhydrous, propan-2-ol-free substance.

CHARACTERISTICS

A white to pale yellow crystalline powder. It melts at about 178°.

CHROMATOGRAPHIC CONDITIONS

- (a) Use a stainless steel column (25 cm x 4.6 mm) packed with cyanosil silica gel for chromatography (5 µm) (Spherisorb CN is suitable).
- (b) Use isocratic elution using the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 240 nm.

(f) Inject 20 µL of each solution.

- (g) If necessary adjust the composition of the mobile phase so that, in the chromatogram obtained with solution (3), the retention time of the peak due to lacidipine is about 10 minutes.
- (h) For solution (1), allow the chromatography to proceed for 2 times the retention time of the principal peak.

MOBILE PHASE

3 volumes of absolute ethanol and 97 volumes of n-hexane.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with solution (3) closely resembles the chromatogram supplied with lacidipine impurity standard BPCRS.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak due to lacidipine impurity B is not greater than 0.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.2%, taking into account the correction factor of 0.5);

the area of any other secondary peak is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.2%);

the total nominal content of impurities is not greater than 0.5%.

Water

Not more than 0.2% w/w, Appendix IX C. Use 0.5 g.

ASSAY

Carry out the method for liquid chromatography, Appendix III D, using the following solutions.

(1) Dilute 5 volumes of a 0.1% w/v solution of the substance being examined in absolute ethanol to 100 volumes with the mobile phase.

(2) Dilute 5 volumes of a 0.1% w/v solution of lacidipine BPCRS in absolute ethanol to 100 volumes with the mobile phase.

(3) Dilute 1 volume of a 0.1% w/v solution of lacidipine impurity standard BPCRS in absolute ethanol to 5 volumes with the mobile phase.

CHROMATOGRAPHIC CONDITIONS

The chromatographic procedure described under Related substances may be used.

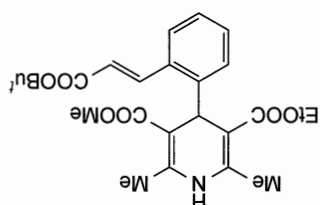
SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with solution (3) closely resembles the corresponding chromatogram supplied with lacidipine impurity standard BPCRS.

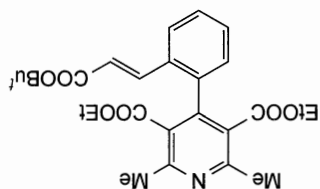
DETERMINATION OF CONTENT

Calculate the content of $C_{26}H_{33}NO_6$ from the chromatograms obtained and using the declared content of $C_{26}H_{33}NO_6$ in lacidipine BPCRS.

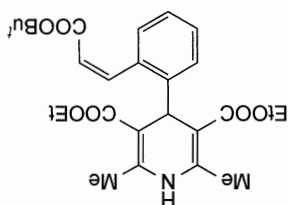
IMPURITIES



A. Ethyl methyl (E)-4-{2-[2-(tert-butoxycarbonyl)vinyl]phenyl}-1,4-dihydro-2,6-dimethylpyridine-3,5-dicarboxylate,



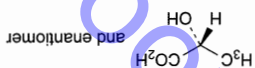
B. Diethyl (E)-4-{2-[2-(tert-butoxycarbonyl)vinyl]phenyl}-1,4-dihydro-2,6-dimethylpyridine-3,5-dicarboxylate,



C. Diethyl (Z)-4-{2-[2-(tert-butoxycarbonyl)vinyl]phenyl}-1,4-dihydro-2,6-dimethylpyridine-3,5-dicarboxylate.

Lactic Acid

(Ph. Eur. monograph 0458)



$C_3H_6O_3$ 90.1 50-21-5

Preparations

Sodium Lactate Infusion
Compound Sodium Lactate Infusion
Lactic Acid Pessaries

DEFINITION

Mixture of 2-hydroxypropionic acid, its condensation products, such as lactic acid and polylactic acids, depends on the concentration and temperature. It is usually the racemate ((RS)-lactic acid).

Content

88.0 per cent m/m to 92.0 per cent m/m of $C_3H_6O_3$.

CHARACTERS

Appearance

Colourless or slightly yellow, syrupy liquid.

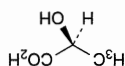
Solubility

Miscible with water and with ethanol (96 per cent).



(S)-Lactic Acid

(Ph. Eur. monograph 1771)



77-33-4

90.1

C₃H₆O₃

Ph Eur

DEFINITION

Mixture of (S)-2-hydroxypropionic acid, its condensation products, such as lactoyl-lactic acid and polylactic acids depends on the concentration and temperature.

Content

88.0 per cent *m/m* to 92.0 per cent *m/m* of C₃H₆O₃, not less than 95.0 per cent of which is the (S)-enantiomer.

CHARACTERS

Appearance

Colourless or slightly yellow, syrupy liquid.

Solubility

Miscible with water and with ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 1 g in 10 mL of water R. The solution is strongly acidic (2.2.4).

B. Relative density (2.2.5): 1.20 to 1.21.

C. It gives the reaction of lactates (2.3.1).

D. It complies with the limits of the assay.

TESTS

Solution S
Dissolve 5.0 g in 42 mL of 1 M sodium hydroxide and dilute to 50 mL with distilled water R.

Appearance

The substance to be examined is not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Ether-insoluble substances

Dissolve 1.0 g in 25 mL of ether R. The solution is not more opalescent than the solvent used for the test.

Sugars and other reducing substances

To 1 mL of solution S add 1 mL of 1 M hydrochloric acid, heat to boiling, allow to cool and add 1.5 mL of 1 M sodium hydroxide and 2 mL of cupri-tartaric solution R. Heat to boiling. No red or greenish precipitate is formed.

Methanol (2.4.24)

Maximum 50 ppm, if intended for use in the manufacture of parenteral preparations.

Citric, oxalic and phosphoric acids

To 5 mL of solution S add dilute ammonia R1 until slightly alkaline (2.2.4). Add 1 mL of calcium chloride solution R. Heat on a water-bath for 5 min. Both before and after heating, any opalescence in the solution is not more intense than that in a mixture of 1 mL of water R and 5 mL of solution S.

Sulfates (2.4.13)

Maximum 200 ppm.

Dilute 7.5 mL of solution S to 15 mL with distilled water R.

Calcium (2.4.3)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

TESTS

Dissolve 5.0 g in 42 mL of 1 M sodium hydroxide and dilute to 50 mL with distilled water R.

Appearance

The substance to be examined is not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Ether-insoluble substances

Dissolve 1.0 g in 25 mL of ether R. The solution is not more opalescent than the solvent used for the test.

Sugars and other reducing substances

To 1 mL of solution S add 1 mL of 1 M hydrochloric acid, heat to boiling, allow to cool and add 1.5 mL of 1 M sodium hydroxide and 2 mL of cupri-tartaric solution R. Heat to boiling. No red or greenish precipitate is formed.

Methanol (2.4.24)

Maximum 50 ppm, if intended for use in the manufacture of parenteral preparations.

Citric, oxalic and phosphoric acids

To 5 mL of solution S add dilute ammonia R1 until slightly alkaline (2.2.4). Add 1 mL of calcium chloride solution R. Heat on a water-bath for 5 min. Both before and after heating, any opalescence in the solution is not more intense than that in a mixture of 1 mL of water R and 5 mL of solution S.

Sulfates (2.4.13)

Maximum 200 ppm.

Dilute 7.5 mL of solution S to 15 mL with distilled water R.

Calcium (2.4.3)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with limit test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 5 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins. Before use, neutralise the test solution to pH 7.0-7.5 with strong sodium hydroxide solution R and shake vigorously.

ASSAY

Place 1.000 g in a ground-glass-stoppered flask and add 10 mL of water R and 20.0 mL of 1 M sodium hydroxide. Close the flask and allow to stand for 30 min. Using 0.5 mL of phenolphthalein solution R as indicator, titrate with 1 M hydrochloric acid until the pink colour is discharged.

1 mL of 1 M sodium hydroxide is equivalent to 90.1 mg of C₃H₆O₃.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

12 mL of solution S complies with limit test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 5 IU/g if intended for use in the manufacture of parenteral preparations without a further appropriate

procedure for the removal of bacterial endotoxins. Before

use, neutralise the test solution to pH 7.0-7.5 with strong

sodium hydroxide solution R and shake vigorously.

ASSAY

Place 1.000 g in a ground-glass-stoppered flask and add

10 mL of water R and 20.0 mL of 1 M sodium hydroxide.

Close the flask and allow to stand for 30 min. Using 0.5 mL

of phenolphthalein solution R as indicator, titrate with 1 M

hydrochloric acid until the pink colour is discharged.

1 mL of 1 M sodium hydroxide is equivalent to 90.1 mg of

$C_6H_{12}O_5$.

(S)-enantiomer

Transfer an amount of the substance to be examined

equivalent to 2.00 g of lactic acid into a round-bottomed

flask, add 25 mL of 1 M sodium hydroxide and boil gently for

15 min. Cool down and adjust to pH 7.0 using 1 M

hydrochloric acid. Add 5.0 g of ammonium molybdate R,

dissolve and dilute to 50.0 mL with water R. Filter and

measure the angle of optical rotation (2.2.7). Calculate the

percentage content of (S)-enantiomer using the expression:

$$50 + \left(24.18 \times \alpha \times \frac{m}{2.222} \times \frac{c}{90} \right)$$

α = angle of optical rotation (absolute value),

m = mass of the substance to be examined, in grams,

c = percentage content of $C_6H_{12}O_5$ in the substance to

be examined.

The complex of (S)-lactic acid formed under these test

conditions is laevorotatory.

LABELLING

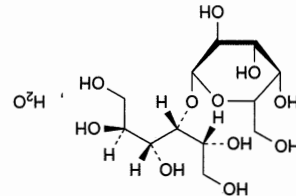
The label states, where applicable, that the substance is

suitable for use in the manufacture of parenteral

preparations.

Lactitol Monohydrate

(Ph. Eur. monograph 1337)



Action and use
Osmotic laxative.

$C_{12}H_{24}O_{11} \cdot H_2O$

362.3

81025-04-9

To 10 mL of solution S add 10 mL of carbon dioxide-free water R. To 10 mL of this solution add 0.05 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to pink. To a further 10 mL of the solution add 0.05 mL of methyl red solution R. Not more than 0.3 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to red.

Acidity or alkalinity

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Appearance of solution

Dissolve 5.000 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

TESTS

Solution S
Dissolve 5.000 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

System suitability: the chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Detection: Spray with 4-aminobenzoic acid solution R and dry in a current of cold air until the solvent is removed; heat at 100 °C for 15 min and allow to cool; spray with a 2 g/L

solution of sodium periodate R and dry in a current of cold air; heat at 100 °C for 15 min.

Drying: In air.

Development: Over 2/3 of the plate.

Application: 2 µL.

Mobile phase: water R, acetonitrile R (25:75 V/V).

Plate: TLC silica gel G plate R.

10 mL with methanol R.

(impurity E) in 1 mL of reference solution (a) and dilute to

10 mL with methanol R and dilute to 2 mL with the

same solvent.

Reference solution (a): Dissolve 5 mg of lactitol

monohydrate CRS in methanol R and dilute to 2 mL with the

same solvent.

Test solution: Dissolve 50 mg of the substance to be examined

in methanol R and dilute to 20 mL with the same solvent.

C: Thin-layer chromatography (2.2.27).

Comparison: lactitol monohydrate CRS.

B: Infrared absorption spectrophotometry (2.2.24).

A: Specific optical rotation (see Tests).

Second identification: A, C.

First identification: B.

IDENTIFICATION

Very soluble in water, slightly soluble in ethanol

(96 per cent), practically insoluble in methylene chloride

Solubility

White or almost white, crystalline powder.

Appearance

CHARACTERS

96.5 per cent to 102.0 per cent (anhydrous substance).

Content

4-O-β-D-Galactopyranosyl-D-glucitol monohydrate.

DEFINITION

Ph. Eur.

Specific optical rotation (2.2.7)

+ 13.5 to + 15.5 (anhydrous substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).
Test solution (a) Dissolve 50.0 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.
Test solution (b) Dilute 2.0 mL of test solution (a) to 50.0 mL with water R.
Reference solution (a) Dissolve 5.0 mg of lactitol monohydrate CRS and 5 mg of glycerol R in water R and dilute to 25.0 mL with the same solvent.
Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with water R. Dilute 5.0 mL of this solution to 100.0 mL with water R.
Reference solution (c) Dilute 2.5 mL of reference solution (a) to 10.0 mL with water R.

Column:
 — size: $l = 0.30$ m, $\varnothing = 7.8$ mm;
 — stationary phase: strong cation-exchange resin (calcium form) R;
 — temperature: 60 °C.
Mobile phase water R.
Flow rate 0.6 mL/min.
Detection Refractive index detector maintained at a constant temperature.

Injection 100 µL; inject test solution (a) and reference solutions (b) and (c).

Run time 2.5 times the retention time of lactitol.

Relative retention With reference to lactitol (retention time = about 13 min); impurity A = about 0.7;

impurity B = about 0.8; glycerol = about 1.3;

impurity C = about 1.5; impurity D = about 1.8;

impurity E = about 1.9.

System suitability: reference solution (c):

— resolution: minimum 5 between the peaks due to lactitol and glycerol.

Limits:

— impurity B: not more than the area of the peak due to lactitol in the chromatogram obtained with reference solution (c) (1.0 per cent);

— total of other impurities: not more than the area of the peak due to lactitol in the chromatogram obtained with reference solution (c) (1.0 per cent);

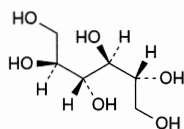
— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to the solvent.

Reducing sugars

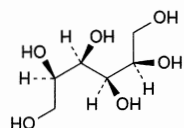
Maximum 0.2 per cent.

Dissolve 5.0 g in 3 mL of water R with gentle heating. Cool and add 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R. When the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.

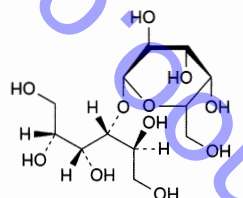
D. galactitol (dulcitol),



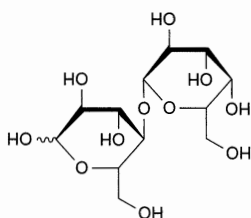
C. D-mannitol,



B. 3-O-β-D-galactopyranosyl-D-glucitol (lactulitol),



A. 4-O-β-D-galactopyranosyl-D-glucopyranose (lactose),



Specified impurities A, B, C, D, E.

IMPURITIES

monohydrate CRS.

Calculate the percentage content of $C_{12}H_{24}O_{11}$ using the chromatograms obtained with test solution (b) and reference solution (a) and the declared content of lactitol.

ASSAY
 Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

TYMC: acceptance criterion 10^2 CFU/g (2.6.12).

TAMC: acceptance criterion 10^3 CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

Absence of *Pseudomonas aeruginosa* (2.6.13).

Microbial contamination

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

4.5 per cent to 5.5 per cent, determined on 0.30 g.

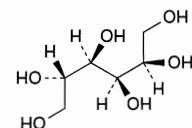
Water (2.5.12)

Maximum 1 ppm.

Nickel (2.4.15)

Maximum 0.5 ppm.

Lead (2.4.10)



E. D-glucitol (D-sorbitol).

Ph Eur

principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Dissolve 3.0 g in 25 mL of water R.

Specific optical rotation (2.2.7)

+ 23.0 to + 29.0 (anhydrous substance).

Dissolve 1.0 g in 80 mL of water R and dilute to 100.0 mL with the same solvent. Allow to stand for 24 h.

Reducing sugars

Maximum 0.2 per cent, calculated as glucose.

Dissolve 5.0 g in 25 mL of water R with the aid of gentle heat. Cool and add 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R and, when the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with limit test B. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.50 g.

Use a mixture of 1 volume of formamide R and 2 volumes of methanol R as solvent.

Total ash (2.4.16)

Maximum 0.2 per cent.

ASSAY

Dissolve 0.350 g in 50 mL of carbon dioxide-free water R, previously heated to 30 °C. Immediately titrate with 0.1 M sodium hydroxide and determine the 2 equivalence points potentiometrically (2.2.20).

The first equivalence point (V₁) corresponds to the acid form of lactobionic acid and the second equivalence point (V₂ - V₁) corresponds to the δ-lactone form.

1 mL of 0.1 M sodium hydroxide is equivalent to 35.83 mg of C₁₂H₂₂O₁₂.

1 mL of 0.1 M sodium hydroxide is equivalent to 34.03 mg of C₁₂H₂₀O₁₁.

The sum of the 2 results is expressed as a percentage content of lactobionic acid.

Ph Eur

Lactobionic Acid

(Ph. Eur. monograph 1647)

C ₁₂ H ₂₂ O ₁₂ (acid form)	358.3	96-82-2
C ₁₂ H ₂₀ O ₁₁ (δ-lactone)	340.3	5965-65-1

Ph Eur

DEFINITION

Mixture in variable proportions of 4-O-β-D-galactopyranosyl-D-glucosonic acid and 4-O-β-D-galactopyranosyl-D-glucosonic acid.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, slightly soluble in glacial acetic acid, in anhydrous ethanol and in methanol.

mp

About 125 °C with decomposition.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison lactobionic acid CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in water R, dry at 105 °C and record new spectra using the residues.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in water R and dilute to 1 mL with the same solvent.

Reference solution Dissolve 10 mg of lactobionic acid CRS in water R and dilute to 1 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R₁, ethyl acetate R, water R, methanol R (2:2:2:4 V/V/V/V).

Application 5 µL.

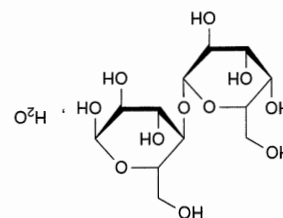
Development Over 3/4 of the plate.

Detection Spray 3 times with ammonium molybdate solution R₆ and heat in an oven at 110 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and colour to the

Lactose

(Lactose Monohydrate, Ph Eur monograph 0187)

 $C_{12}H_{22}O_{11} \cdot H_2O$

360.3

10039-26-6

Action and use

Excipient.

Ph Eur

DEFINITION

 α -D-Galactopyranosyl-(1 \rightarrow 4)-D-glucopyranose

monohydrate.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely but slowly soluble in water, practically insoluble in

ethanol (96 per cent).

IDENTIFICATION

First identification A, D.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison lactose CRS.

B. Thin-layer chromatography (2.2.27).

Solvent mixture water R, methanol R (2:3 V/V).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of lactose CRS in the

solvent mixture and dilute to 20 mL with the solvent

mixture.

Reference solution (b) Dissolve 10 mg of fructose CRS, 10 mg

of glucose CRS, 10 mg of lactose CRS and 10 mg of

sucrose CRS in the solvent mixture and dilute to 20 mL with

the solvent mixture.

Plate TLC silica gel G plate R.

Mobile phase water R, methanol R, glacial acetic acid R, ethylene

chloride R (10:15:25:50 V/V/V/V); measure the volumes

accurately, as a slight excess of water produces cloudiness.

Application 2 μ L; thoroughly dry the points of application.

Development A Over a path of 15 cm.

Drying A In a current of warm air.

Development B Immediately, over a path of 15 cm, after

renewing the mobile phase.

Drying B In a current of warm air.

Detection Spray with a solution of 0.5 g of thymol R in a

mixture of 5 mL of sulfuric acid R and 95 mL of ethanol

(96 per cent) R; heat at 130 °C for 10 min.

System suitability: reference solution (b):

— the chromatogram shows 4 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

(a).

C. Dissolve 0.25 g in 5 mL of water R. Add 5 mL of

ammonia R and heat in a water-bath at 80 °C for 10 min.

A red colour develops.

D. Water (see Tests).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Dissolve 1.0 g in boiling water R and dilute to 10 mL with

the same solvent.

Acidity or alkalinity

Dissolve 6.0 g by heating in 25 mL of carbon dioxide-free

water R, cool and add 0.3 mL of phenolphthalein solution R1.

The solution is colourless. Not more than 0.4 mL of 0.1 M sodium hydroxide is required to change the colour of the

indicator to pink or red.

Specific optical rotation (2.2.7)

+ 54.4 to + 55.9 (anhydrous substance).

Dissolve 10.0 g in 80 mL of water R, heating to 50 °C. Allow

to cool and add 0.2 mL of dilute ammonia R1. Allow to stand for 30 min and dilute to 100.0 mL with water R.

Absorbance (2.2.25)

Test solution (a) Dissolve 1.0 g in boiling water R and dilute

to 10.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to

10.0 mL with water R.

Spectral range 400 nm for test solution (a) and 210–300 nm

for test solution (b).

Results:

— at 400 nm: maximum 0.04 for test solution (a);

— from 210 nm to 220 nm: maximum 0.25 for test

solution (b);

— from 270 nm to 300 nm: maximum 0.07 for test

solution (b).

Heavy metals (2.4.8)

Maximum 5 ppm.

Dissolve 4.0 g in water R with warming, add 1 mL of 0.1 M

hydrochloric acid and dilute to 20 mL with water R. 12 mL of

the solution complies with test A. Prepare the reference

solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

4.5 per cent to 5.5 per cent, determined on 0.50 g, using a

mixture of 1 volume of formamide R and 2 volumes of

methanol R as the solvent.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10² CFU/g (2.6.12).Absence of *Escherichia coli* (2.6.13).

STORAGE

In an airtight container.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate

compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for lactose monohydrate used as a filler/diluent in solid dosage forms (compressed and powder).

Particle size distribution (2.9.31 or 2.9.38).

Bulk and tapped density (2.9.34)

Determine the bulk density and the tapped density. Calculate the Hausner Index using the following expression

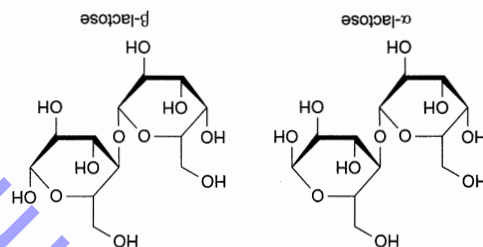
$$\frac{V_0}{V_f}$$

V_0 = volume of bulk substance;
 V_f = volume of tapped substance.

Ph Eur

Anhydrous Lactose

(Ph. Eur. monograph 1061)



$C_{12}H_{22}O_{11}$ 342.3 63-42-3

Action and use
 Excipient.

Ph Eur

DEFINITION

O-β-D-Galactopyranosyl-(1→4)-β-D-glucopyranose or mixture of O-β-D-galactopyranosyl-(1→4)-α-D-glucopyranose and O-β-D-galactopyranosyl-(1→4)-β-D-glucopyranose.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely but slowly soluble in water, practically insoluble in

ethanol (96 per cent).

IDENTIFICATION

First identification A, D.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison anhydrous lactose CRS.

B. Thin-layer chromatography (2.2.27).

Solvent mixture water R, methanol R (40:60 V/V).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of anhydrous lactose CRS in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of glucose CRS, 10 mg of fructose CRS, 10 mg of sucrose CRS in the solvent mixture and dilute to 20 mL with the solvent mixture.

Plate TLC silica gel G plate R.

Mobile phase water R, methanol R, glacial acetic acid R, ethylene chloride R (10:15:25:50 V/V/V/V); measure the volumes accurately, as a slight excess of water produces cloudiness.

Application 2 µL, thoroughly dry the starting points.

Development A Over a path of 15 cm.

Drying A In a current of warm air.

Development B Immediately, over a path of 15 cm, after renewing the mobile phase.

Drying B In a current of warm air.

Detection Spray with a solution of 0.5 g of thymol R in a mixture of 5 mL of sulfuric acid R and 95 mL of ethanol (96 per cent) R; heat at 130 °C for 10 min.

System suitability: reference solution (b):

— the chromatogram shows 4 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.25 g in 5 mL of water R. Add 5 mL of ammonia R and heat in a water-bath at 80 °C for 10 min.

A red colour develops.

D. Water (see Tests).

TESTS

Solution S

Dissolve 1.0 g in boiling water R, allow to cool and dilute to 10.0 mL with water R.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Acidity or alkalinity

Dissolve 6.0 g by heating in 25 mL of carbon dioxide-free water R, cool and add 0.3 mL of phenolphthalein solution R1.

The solution is colourless. Not more than 0.4 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to pink or red.

Specific optical rotation

+ 54.4 to + 55.9 (anhydrous substance).

Dissolve 10.0 g in 80 mL of water R, heating to 50 °C. Allow to cool and add 0.2 mL of dilute ammonia R1. Allow to stand for 30 min and dilute to 100.0 mL with water R.

Absorbance

Test solution (a) Solution S.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with water R.

Spectral range 400 nm for test solution (a) and 210-300 nm for test solution (b).

Results:

— at 400 nm: maximum 0.04 for test solution (a);

— from 210 nm to 220 nm: maximum 0.25 for test solution (b);

— from 270 nm to 300 nm: maximum 0.07 for test solution (b).

Heavy metals (2.4.8)

Maximum 5 ppm.

2.0 g complies with test C. Prepare the reference solution using 1.0 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.00 g, using a mixture of 1 volume of formamide R and 2 volumes of methanol R as the solvent.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (2.6.12).
Absence of *Escherichia coli* (2.6.13).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for anhydrous lactose used as filler/infilluent in solid dosage forms (compressed and powder).

Particle-size distribution (2.9.31 or 2.9.38).**Bulk and tapped density (2.9.34)**

Determine the bulk density and the tapped density. Calculate the Hausner index using the following expression:

$$\frac{V_0}{V_f}$$

V_0 = volume of bulk substance;
 V_f = volume of tapped substance.

 α -Lactose and β -lactose

Gas chromatography (2.2.28).

Silylation reagent dimethyl sulfoxide R, N-

trimethylsilylimidazole R, pyridine R (19.5:22:58.5 V/V/V).

Test solution Introduce 10 mg of the substance to be examined into a vial with a screw cap and add 4 mL of the

silylation reagent. Sonicate for 20 min at room temperature, allow to cool and transfer 400 μ L to an injection vial.

Add 1 mL of pyridine R, close the vial and mix well.

Reference solution Prepare a mixture of α -lactose monohydrate R and β -lactose R to obtain an anomeric ratio of about 1:1 based on the labelled anomeric contents of the α -lactose monohydrate and the β -lactose. Introduce 10 mg of the mixture into a vial with a screw cap and add 4 mL of the

silylation reagent. Sonicate for 20 min at room temperature, allow to cool and transfer 400 μ L to an injection vial.

Add 1 mL of pyridine R, close the vial and mix well.

Reference solution Prepare a mixture of α -lactose monohydrate R and β -lactose R to obtain an anomeric ratio of about 1:1 based on the labelled anomeric contents of the α -lactose monohydrate and the β -lactose. Introduce 10 mg of the mixture into a vial with a screw cap and add 4 mL of the

silylation reagent. Sonicate for 20 min at room temperature, allow to cool and transfer 400 μ L to an injection vial.

Add 1 mL of pyridine R, close the vial and mix well.

Precolumn:

— material: intermediate-polarity deactivated fused silica;
— size: $l = 2$ m, $\varnothing = 0.53$ mm.

Column:

— material: fused silica;
— size: $l = 15$ m, $\varnothing = 0.25$ mm.

Preparation

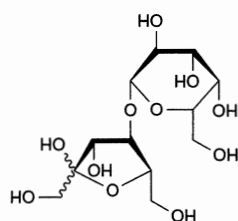
Osmotic laxative.

Action and use

$C_{12}H_{22}O_{11}$

342.3

4618-18-2



(Ph. Eur. monograph 1230)

Lactulose

Ph Eur

Determine on 1.000 g by drying in an oven at 80 °C for 2 h.

Loss on drying (2.2.32)

S_a = area of the peak due to α -lactose;
 S_b = area of the peak due to β -lactose.

$$\frac{100S_a}{S_a + S_b}$$

Calculate the percentage content of β -lactose using the following expression:

$$\frac{100S_b}{S_a + S_b}$$

following expression:

Calculate the percentage content of α -lactose using the following expression:

α -lactose and β -lactose.

— resolution: minimum 3.0 between the peaks due to

System suitability: reference solution:

time = about 12 min): α -lactose = about 0.9.

Relative retention With reference to β -lactose (retention time = about 12 min): α -lactose = about 0.9.

Detection Flame ionisation.

Temperature	Time	Column	Injection port	Detector
(°C)	(min)			
80	0 - 1			
80 \rightarrow 150	1 - 3			
150 \rightarrow 300	3 - 15.5			
300	15.5 - 17.5			
275 or use cold on-column injection				325

Temperature:

Flow rate 2.8 mL/min.

Carrier gas helium for chromatography R.

thickness 0.25 μ m).

— stationary phase: poly(dimethyl) (diphenyl) siloxane R (film

DEFINITION

4-O-β-D-Galactopyranosyl-D-arabino-hex-2-ulofuranose.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble in methanol, practically insoluble in toluene.

mp

About 168 °C.

IDENTIFICATION

First identification B, C, D, E

Second identification A, C, D, E

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 50.0 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 50.0 mg of lactulose CRS in water R and dilute to 10.0 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase Glacial acetic acid R, 50 g/L solution of boric acid R, methanol R, ethyl acetate R (10:15:20:55 V/V/V/V).

Application 2 µL.

Development Over 3/4 of the plate.

Drying At 100–105 °C for 5 min; allow to cool.

Detection Spray with a 1.0 g/L solution of 1,3-dihydroxyphenylhydrazine R in a mixture of 10 volumes of sulfuric acid R and 90 volumes of methanol R; heat at 110 °C for 5 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution (b).

C. Dissolve 50 mg in 10 mL of water R. Add 3 mL of cupric tartrate solution R and heat. A red precipitate is formed.

D. Dissolve 0.125 g in 5 mL of water R. Add 5 mL of ammonia R. Heat in a water-bath at 80 °C for 10 min. A red colour develops.

E. Specific optical rotation (see Tests).

TESTS

Solution S

Dissolve 3.0 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

pH (2.2.3)

3.0 to 7.0.

To 10 mL of solution S add 0.1 mL of a saturated solution of potassium chloride R.

Specific optical rotation (2.2.7)

–50.0 to –46.0 (anhydrous substance).

Dissolve 1.25 g in water R, add 0.2 mL of concentrated ammonia R and dilute to 25.0 mL with water R.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 1.00 g of the substance to be examined in 10 mL of water R. Add 12.5 mL of acetonitrile R with gentle heating and dilute to 25.0 mL with water R.

Reference solution (a) To 3.0 mL of the test solution add 47.5 mL of acetonitrile R with gentle heating and dilute to 100.0 mL with water R.

Reference solution (b) Dissolve 1.00 g of lactulose CRS in 10 mL of water R. Add 12.5 mL of acetonitrile R with gentle heating and dilute to 25.0 mL with water R.

Reference solution (c). Dissolve 10 mg of lactulose R, 10 mg of epilactose R (impurity A) and 10 mg of lactose R (impurity C) in water R and dilute to 5.0 mL with the same solvent.

Reference solution (d) To 5.0 mL of the test solution add 47.5 mL of acetonitrile R with gentle heating and dilute to 100.0 mL with water R. Dilute 5.0 mL of this solution to 100.0 mL with a mixture of equal volumes of acetonitrile R and water R.

Column 1:

— size: $l = 0.05$ m, $\phi = 4.6$ mm;

— stationary phase: aminopropylsilyl silica gel for chromatography R (3 µm);

— temperature: 38 ± 1 °C.

Column 2:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: aminopropylsilyl silica gel for chromatography R (3 µm);

— temperature: 38 ± 1 °C.

Columns 1 and 2 are coupled in series.

Mobile phase Dissolve 0.253 g of sodium dihydrogen phosphate R in 200 mL of water R and dilute to 1000 mL with acetonitrile R.

Flow rate 1.0 mL/min.

Detection Refractometer maintained at a constant temperature.

Injection 20 µL of the test solution and reference solutions (a), (c) and (d).

Run time Twice the retention time of lactulose.

Identification of impurities Use the chromatogram obtained with reference to identify the peaks due to impurities A and C.

Relative retention With reference to lactulose (retention time = about 18 min): impurity A = about 0.9;

impurity C = about 1.2.

System suitability: reference solution (c):

— peak-to-valley ratio: minimum 5.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to lactulose.

Limits:

— impurity C: not more than the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (3.0 per cent);

— unspecified impurities: for each impurity, not more than twice the area of the peak due to lactulose in the chromatogram obtained with reference solution (d) (0.5 per cent);

To each flask add 4.0 mL of acetate-edate buffer solution pH 5.5 R. Mix and add 4.0 mL of freshly prepared azomethine H solution R. Mix and allow to stand for 1 h. Measure the absorbance (2.2.25) of solutions A, B and C at 420 nm, using solution D as the compensation liquid. The test is not valid unless the absorbance of solution C is at least 0.25. The absorbance of solution B is not less than twice that of solution A.

Lead (2.4.10)

Maximum 0.5 ppm.

Water (2.5.12)

Maximum 2.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

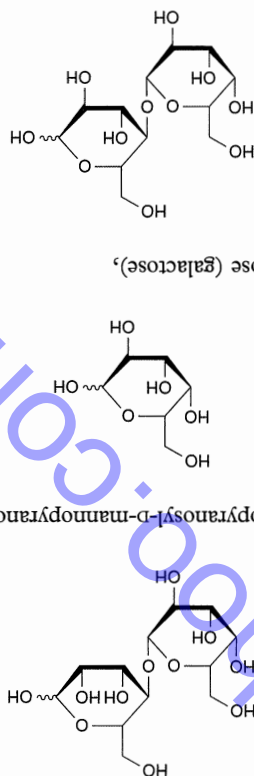
Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{12}H_{22}O_{11}$ taking into account the assigned content of lactulose CRS.

IMPURITIES

Specified impurities C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D, E.



B. D-galactopyranose (galactose),

A. 4-O-β-D-galactopyranosyl-D-mannopyranose (epilactose),

total: not more than the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (3.0 per cent); disregard limit: the area of the peak due to lactulose in the chromatogram obtained with reference solution (d) (0.25 per cent).

The thresholds indicated under Related substances

(Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Methanol

Head-space gas chromatography (2.2.28).

Internal standard solution Mix 0.5 mL of propanol R and 100.0 mL of water R. Dilute 1.0 mL of the solution to 100.0 mL with water R. Dilute 5.0 mL of this solution to 50.0 mL with water R.

Test solution To 79 mg of the substance to be examined in a 20 mL vial add 1.0 mL of the internal standard solution and 5 μL of a 0.1 per cent V/V solution of methanol R.

Reference solution To 1.0 mL of the internal standard solution in a 20 mL vial add 5 μL of a 0.1 per cent V/V solution of methanol R.

Stationary phase: ethylnvinylbenzene-divinylbenzene copolymer R (180 μm).

Carrier gas helium for chromatography R.

Flow rate 30 mL/min.

Static head-space conditions that may be used:

equilibration temperature: 60 °C;

equilibration time: 1 h;

pressurisation time: 1 min.

Temperature:

column: 140 °C;

injection port: 200 °C;

detection: 220 °C.

Detection Flame ionisation.

Injection 1 mL of the gaseous phase.

Calculate the content of methanol, taking its density (2.2.5) at 20 °C to be 0.79 g/mL.

Limit:

methanol: calculate the ratio (R) of the area of the peak due to methanol to the area of the peak due to the internal standard in the chromatogram obtained with the reference solution; calculate the ratio of the area of the peak due to methanol to the area of the peak due to the internal standard in the chromatogram obtained with the test solution: this ratio is not greater than 2R (50 ppm).

Boron

Maximum 9 ppm.

Avoid where possible the use of glassware.

Reference solution Dissolve 50.0 mg of boric acid R in water R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with water R. Keep in a well-closed polyethylene container.

In 4 polyethylene 25 mL flasks, place separately:

— 0.50 g of the substance to be examined dissolved in 2.0 mL of water R (solution A);

— 0.50 g of the substance to be examined dissolved in 1.0 mL of the reference solution and 1.0 mL of water R (solution B);

— 1.0 mL of the reference solution and 1.0 mL of water R (solution C);

— 2.0 mL of water R (solution D).

principal spot in the chromatogram obtained with the

reference solution.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference

solution (b).

C. To 0.1 g add 10 mL of water R and 3 mL of cupri-tartrate

solution R and heat. A red precipitate is formed.

D. To 0.25 g add 5 mL of water R and 5 mL of ammonia R.

Heat in a water-bath at 80 °C for 10 min. A red colour

develops.

TESTS

Solution S

Mix 10 g with carbon dioxide-free water R and dilute to

100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution BY₅ (2.2.2, Method II).

pH (2.2.3)

3.0 to 7.0.

To 10 mL of solution S, add 0.1 mL of a saturated solution

of potassium chloride R.

Related substances

Liquid chromatography (2.2.29).

Test solution Mix 4.00 g of the substance to be examined and 20 mL of water R. Add 25.0 mL of acetonitrile R with gentle

heating and dilute to 50.0 mL with water R.

Reference solution (a) To 5.0 mL of the test solution, add

47.5 mL of acetonitrile R with gentle heating and dilute to

100.0 mL with water R.

Reference solution (b) Dissolve 2.00 g of lactulose CRS in

20 mL of water R. Add 25.0 mL of acetonitrile R with gentle

heating and dilute to 50.0 mL with water R.

Reference solution (c) Dissolve 65 mg of fructose CRS

(impurity D) in a mixture of equal volumes of acetonitrile R

and water R and dilute to 100.0 mL with the same mixture

of solvents.

Reference solution (d) Dissolve 1 g of lactulose for peak

identification CRS (containing impurities A, B, C, E, F, G

and H) in reference solution (c) and dilute to 25.0 mL with

reference solution (c).

Reference solution (e) Dilute 5.0 mL of reference solution (a)

to 100.0 mL with A mixture of equal volumes of

acetonitrile R and water R.

Column 1:

— size: $l = 0.05$ m, $\phi = 4.6$ mm;

— stationary phase: aminopropylsilyl silica gel for

chromatography R (3 μ m);

— temperature: 38 ± 1 °C.

Column 2:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: aminopropylsilyl silica gel for

chromatography R (3 μ m);

— temperature: 38 ± 1 °C.

Columns 1 and 2 are coupled in series.

Mobile phase Dissolve 0.253 g of sodium dihydrogen

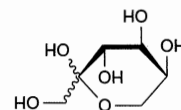
phosphate R in 200 mL of water R and dilute to 1000 mL

with acetonitrile R.

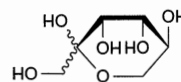
Flow rate 1.0 mL/min.

Detection Refractometer maintained at a constant

temperature.



D. D-arabino-hex-2-ulopyranose, (fructose),



E. D-lyxo-hex-2-ulopyranose (tagatose).

Ph Eur



Lactulose Solution

(Liquid Lactulose, Ph Eur monograph 0924)

Action and use
Osmotic laxative.

Ph Eur

DEFINITION

Aqueous solution of 4-O-β-D-galactopyranosyl-D-arabino-hex-2-ulofuranose normally prepared by alkaline isomerisation of lactose. It may contain other sugars including lactose, epilactose, galactose, tagatose and fructose.

Content

Minimum 620 g/L of lactulose ($C_{12}H_{22}O_{11}$; M_r 342.3) and 95.0 per cent to 105.0 per cent of the content of lactulose

stated on the label.

It may contain a suitable antimicrobial preservative.

CHARACTERS

Appearance

Clear, viscous liquid, colourless or pale brownish-yellow.

Solubility

Miscible with water. It may be a supersaturated solution or

may contain crystals that disappear on heating.

A 10 per cent V/V solution is laevorotatory.

IDENTIFICATION

First identification B, C, D

Second identification A, C, D

A. Thin-layer chromatography (2.2.27).

Test solution Dilute 0.50 g of the substance to be examined to

50 mL with water R.

Reference solution Dissolve 60 mg of lactulose CRS in water R

and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase glacial acetic acid R, 50 g/L solution of boric

acid R, methanol R, ethyl acetate R (10:15:20:55 V/V/V/V).

Application 2 μ L.

Development Over 3/4 of the plate.

Drying At 100-105 °C for 5 min and allow to cool.

Detection Spray with a 1.0 g/L solution of 1,3-

dihydroxynaphthalene R in a mixture of 10 volumes of sulfuric

acid R and 90 volumes of methanol R; heat at 110 °C for

5 min.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

Injection 20 µL of the test solution and of reference solutions (a), (d) and (e).
Run time Twice the retention time of lactulose.
Identification of impurities Use the chromatogram supplied with lactulose for peak identification CRS and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, E, F, G and H.

Relative retention With reference to lactulose (retention time = about 18 min): impurity F = about 0.2;

impurity E = about 0.38; impurity D = about 0.42; impurity B = about 0.6; impurity G = about 0.8;

impurity A = about 0.9; impurity C = about 1.2; impurity H = about 1.5.

System suitability Reference solution (d):

— **peak-to-valley ratio**: minimum 5.0, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to lactulose.

Limits:

— **impurity B**: not more than 3 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (15.0 per cent);

— **impurities A, C**: for each impurity, not more than twice the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (10.0 per cent);

— **impurities E, F**: for each impurity, not more than 0.8 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (4.0 per cent);

— **impurities G, H**: for each impurity, not more than 0.3 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (1.5 per cent);

— **impurity D**: not more than 0.2 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (1.0 per cent);

— **unspecified impurities**: for each impurity, not more than 0.1 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (0.5 per cent);

— **sum of impurities eluting after impurity H**: not more than 0.26 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (1.3 per cent);

— **total (excluding impurities B and C)**: not more than 2.4 times the area of the peak due to lactulose in the chromatogram obtained with reference solution (a) (12.0 per cent);

— **lactulose** in the chromatogram obtained with reference solution (e) (0.25 per cent).

The thresholds indicated under Related substances (Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Methanol

Head-space gas chromatography (2.2.28).

Internal standard solution Mix 0.5 mL of propanol R and 100.0 mL of water R. Dilute 1.0 mL of the solution to 50.0 mL with water R. Dilute 5.0 mL of this solution to 100.0 mL with water R.

Test solution To 0.13 g of the substance to be examined in a 20 mL vial add 1.0 mL of the internal standard solution and 5 µL of a 0.1 per cent V/V solution of methanol R.

Reference solution To 1.0 mL of the internal standard solution in a 20 mL vial add 5 µL of a 0.1 per cent V/V solution of methanol R.

Column:

— **size**: $l = 2$ m, $\varnothing = 2$ mm;

— **stationary phase**: ethylnvinylbenzene-divinylbenzene copolymer R (180 µm).

Carrier gas helium for chromatography R.

Flow rate 30 mL/min.

Static head-space conditions which may be used:

— **equilibration temperature**: 60 °C;

— **equilibration time**: 1 h;

— **pressurisation time**: 1 min.

Temperature:

— **column**: 140 °C;

— **injection port**: 200 °C;

— **detector**: 220 °C.

Detection Flame ionisation.

Injection 1 mL of the gaseous phase.

Calculate the content of methanol, taking its density (2.2.5) at 20 °C to be 0.79 g/mL.

Limit:

— **methanol**: calculate the ratio (R) of the area of the peak due to methanol to the area of the peak due to the internal standard in the chromatogram obtained with the reference solution; calculate the ratio of the area of the peak due to methanol to the area of the peak due to the internal standard in the chromatogram obtained with the test solution: this ratio is not greater than 2R (30 ppm).

Sulfites

Maximum 30 ppm.

Mix 5.0 g with 40 mL of water R, add 2.0 mL of 0.1 M sodium hydroxide and dilute to 100 mL with water R.

To 10.0 mL of this solution, add 1.0 mL of hydrochloric acid R, 2.0 mL of decolorised fuchsin solution R1 and 2.0 mL of a 0.5 per cent V/V solution of formaldehyde R. Allow to stand for 30 min and measure the absorbance (2.2.25) at 583 nm using as the compensation liquid a solution prepared at the same time and in the same manner with 10.0 mL of water R instead of the solution of the substance to be examined. The absorbance is not greater than that of a reference solution prepared at the same time and in the same manner using 10.0 mL of sulfite standard solution (1.5 ppm SO_2) R instead of the solution of the substance to be examined.

Boron

Maximum 5 ppm.

Avoid where possible the use of glassware.

Reference solution Dissolve 56.0 mg of boric acid R in water R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with water R. Keep in a well-closed polyethylene container.

In 4 polyethylene 25 mL flasks, place separately: 1.00 g of the substance to be examined and 1 mL of the reference solution (solution B);

— 1 mL of the reference solution and 1 mL of water R (solution C);

— 2 mL of water R (solution D).

To each flask, add 4.0 mL of acetate-acetate buffer solution pH 5.5 R. Mix and add 4.0 mL of freshly prepared azomethine H solution R. Mix and allow to stand for 1 h.

Measure the absorbance (2.2.25) of solutions A, B and C at 420 nm, using solution D as the compensation liquid. The test is not valid unless the absorbance of solution C is at least 0.25. The absorbance of solution B is not less than twice that of solution A.

Lead (2.4.10)

Maximum 0.5 ppm, calculated with reference to the declared content of lactulose.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.5 g and calculated with reference to the declared content of lactulose.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (2.6.12).

TYMC: acceptance criterion 10^1 CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

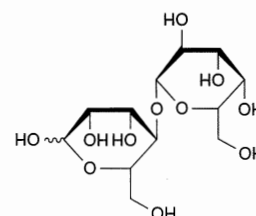
Calculate the percentage content of $C_{12}H_{22}O_{11}$ taking into account the assigned content of lactulose CRS.

LABELLING

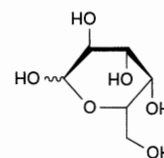
The label states the declared content of lactulose.

IMPURITIES

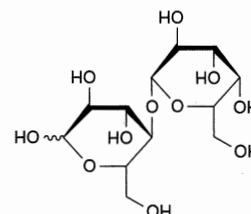
Specified impurities A, B, C, D, E, F, G, H



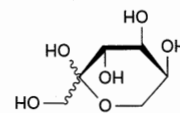
A. 4-O-β-D-galactopyranosyl-D-mannopyranose (epilactose),



B. D-galactopyranose (galactose),



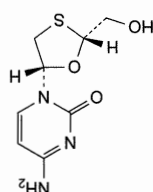
C. 4-O-β-D-galactopyranosyl-D-glucopyranose (lactose),



D. D-arabinohex-2-ulopyranose (fructose),

Lamivudine

(Ph. Eur. monograph 2217)



$C_8H_{11}N_3O_5$

229.3

134678-17-4

Action and use

Nucleoside reverse transcriptase inhibitor; antiviral (HIV).

Preparations

Lamivudine Tablets
Zidovudine and Lamivudine Tablets
Abacavir, Zidovudine and Lamivudine Tablets
Abacavir and Lamivudine Tablets

DEFINITION

4-Amino-1-[(2*R*,5*S*)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]pyrimidin-2(1*H*)-one.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Soluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

First identification B, C
Second identification A, B

A. Specific optical rotation (2.2.7): -99 to -97 (dried substance).

Dissolve 0.250 g in water R and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison lamivudine CRS.

TESTS

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *methanol* R, evaporate to dryness and record new spectra using the residues.

C. Enantiomeric purity (see Tests).

Absorbance (2.2.25)

Maximum 0.3 at 440 nm, using a path length of 4 cm. Dissolve 1.00 g in *water* R, using sonication if necessary, and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be

examined in the mobile phase and dilute to 100.0 mL with

the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *sallyclic acid* R in the

mobile phase and dilute to 100.0 mL with the mobile phase.

Dilute 1.0 mL of the solution to 100.0 mL with the mobile

phase.

Reference solution (c) Dissolve 50.0 mg of *lamivudine* CRS in

the mobile phase and dilute to 100.0 mL with the mobile

phase.

Reference solution (d) Dissolve 5 mg of *cytosine* R and 5 mg of

uracil R in the mobile phase and dilute to 100.0 mL with the

mobile phase. Dilute 2.0 mL of the solution to 10.0 mL with

the mobile phase.

Reference solution (e) Dissolve 5 mg of *lamivudine* for system

suitability 1 CRS (containing impurities A and B) in 2 mL of

the mobile phase. Add 1.0 mL of reference solution (d) and

dilute to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for

chromatography R (5 μ m);

— temperature: 35 °C.

Mobile phase Mix 5 volumes of *methanol* R and 95 volumes of

a 1.9 g/L solution of *ammonium acetate* R, previously adjusted

to pH 3.8 with *glacial acetic acid* R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 277 nm.

Injection 10 μ L.

Run time 3 times the retention time of *lamivudine*.

Identification of impurities Use the chromatograms obtained

with reference solutions (b) and (e) to identify the peaks due

to impurities A, B, E, F and C.

Relative retention With reference to *lamivudine* (retention

time = about 9 min): impurity E = about 0.28;

impurity F = about 0.32; impurity A = about 0.36;

impurity B = about 0.91; impurity J = about 1.45;

impurity C = about 2.32.

System suitability: reference solution (e):

— resolution: minimum 1.5 between the peaks due to

impurities F and A; minimum 1.5 between the peaks due

to impurity B and *lamivudine*.

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity B = 0.6;

impurity F = 2.2; impurity J = 2.2;

Enantiomeric purity

Liquid chromatography (2.2.29): use the normalisation

procedure.

Test solution Dissolve 25.0 mg of the substance to be

examined in *water* R and dilute to 100.0 mL with the same

solvent.

Reference solution Dissolve the contents of a vial of *lamivudine*

for system suitability 2 CRS (containing impurity D) in 1.0 mL

of *water* R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel BC for chiral chromatography R

(5 μ m);

— temperature: maintain at constant temperature between

15 °C and 30 °C; the temperature may be adjusted to

optimise the resolution between *lamivudine* and

impurity D; a lower temperature favours improved

resolution.

Mobile phase Mix 5 volumes of *methanol* R and 95 volumes of

a 7.7 g/L solution of *ammonium acetate* R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 10 μ L.

Run time Twice the retention time of *lamivudine*.

Relative retention With reference to *lamivudine* (retention

time = about 8 min): impurity D = about 1.2; impurity B

and enantiomer = about 1.3 and 1.5.

System suitability: reference solution:

— peak-to-valley-ratio: minimum 1.5, where H_p = height

above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

lamivudine.

Calculate the sum of the percentage contents of all

impurity peaks with a relative retention from 1.2 to 1.5.

Subtract the percentage content of impurity B as obtained in

the test for related substances.

Limit:

— impurity D: maximum 0.3 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution

using 2 mL of *lead standard solution* (10 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

Calculate the percentage content of $C_8H_{11}N_3O_5$ using the chromatograms obtained with the test solution and reference solution (c) and the declared content of $C_8H_{11}N_3O_5$ in lamivudine CRS.

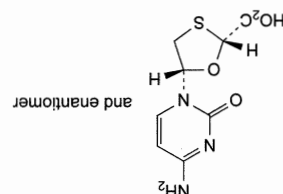
STORAGE

Protected from light.

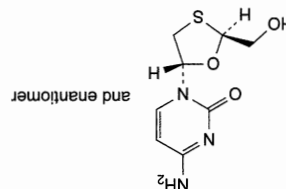
IMPURITIES

Specified impurities A, B, C, D

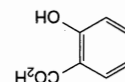
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: E, F, G, H, I, J.



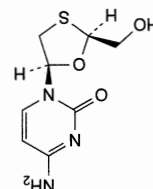
A. (2R,5SR)-5-(4-amino-2-oxopyrimidin-1(2H)-yl)-1,3-oxathiolane-2-carboxylic acid,



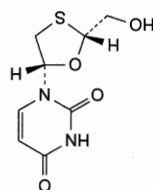
B. 4-amino-1-[(2R,5RS)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]pyrimidin-2(1H)-one ((±)-trans-lamivudine),



C. 2-hydroxybenzenecarboxylic acid (salicylic acid),

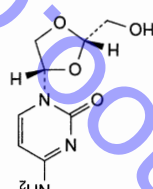


D. 4-amino-1-[(2S,5R)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]pyrimidin-2(1H)-one,

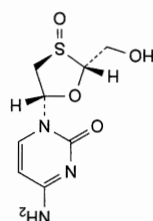


J. 1-[(2R,5S)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]pyrimidine-2,4(1H,3H)-dione.

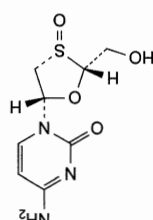
I. 4-amino-1-[(2S,4S)-2-(hydroxymethyl)-1,3-dioxolan-4-yl]pyrimidin-2(1H)-one,



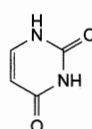
H. 4-amino-1-[(2R,3R,5S)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]pyrimidin-2(1H)-one S-oxide,



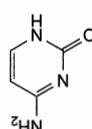
G. 4-amino-1-[(2R,3S,5S)-2-(hydroxymethyl)-1,3-oxathiolan-5-yl]pyrimidin-2(1H)-one S-oxide,



F. pyrimidine-2,4(1H,3H)-dione (uracil),

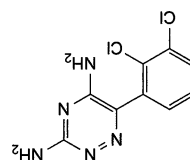


E. 4-aminopyrimidin-2(1H)-one (cytosine),



Lamotrigine

(Ph. Eur. monograph 1756)

C₉H₇Cl₂N₅ 256.1 84057-84-1

Action and use

Antiepileptic.

Preparations

Dispersible Lamotrigine Tablets

Lamotrigine Tablets

Ph. Eur.

DEFINITION

6-(2,3-Dichlorophenyl)-1,2,4-triazine-3,5-diamine.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, slightly soluble in anhydrous ethanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison lamotrigine CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in 5 mL of methanol R and dilute to 100.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Reference solution (a) Dissolve 5 mg of lamotrigine for system suitability CRS (containing impurity G) in 2.5 mL of methanol R and dilute to 25.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Dilute 2.0 mL of this solution to 10.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Reference solution (c) Dissolve 5.0 mg of lamotrigine with a 10.3 g/L solution of hydrochloric acid R.

Reference solution (d) Dissolve 10 mg of lamotrigine for peak identification CRS (containing impurities A, E and F) in 2.5 mL of methanol R and dilute to 50.0 mL with a 10.3 g/L solution of hydrochloric acid R. To 4.0 mL of this solution add 5 mL of methanol R and dilute to 100.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Blank solution Mix 5 volumes of methanol R and 95 volumes of a 10.3 g/L solution of hydrochloric acid R.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

Injection Test solution and reference solutions (d) and (c).

Run time 10 min.

Detection Spectrophotometer at 210 nm.

Mobile phase acetronitrile for chromatography R, mobile phase A (35:65 V/V).

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Impurity E

any peak due to impurity E.

— the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to the blank and

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

chromatogram obtained with reference solution (b) 0.5 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than 0.1 per cent;

chromatogram obtained with reference solution (b) 0.5 times the area of the principal peak in the

— impurities A, G: for each impurity, not more than 0.2 per cent;

in the chromatogram obtained with reference solution (b) peak area of impurity F by 1.3;

— impurity F: not more than the area of the principal peak in the chromatogram obtained with reference solution (b)

— correction factor: for the calculation of content, multiply the

Limits:

separating this peak from the peak due to lamotrigine.

above the baseline due to impurity G and H_p = height— peak-to-valley ratio: minimum 1.2, where H_p = height

System suitability: reference solution (a):

impurity F = about 1.8.

impurity A = about 1.3; impurity E = about 1.7;

time = about 7 min; impurity G = about 1.1;

Relative retention With reference to lamotrigine (retention

reference solution (a) to identify the peak due to impurity G.

suitability CRS and the chromatogram obtained with

chromatogram supplied with lamotrigine for system

identify the peaks due to impurities A, E and F; use the

chromatogram obtained with reference solution (d) to

with lamotrigine for peak identification CRS and the

Identification of impurities Use the chromatogram supplied

solutions (a), (b) and (d) and the blank solution.

Injection 10 μ L of the test solution, reference

Detection Spectrophotometer at 270 nm.

Flow rate 1.0 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 4	85	15
4 - 14	85 \rightarrow 20	15 \rightarrow 80

— mobile phase B: acetonitrile R;

phosphate R; adjust to pH 2.0 with phosphoric acid R;

150 volumes of a 2.7 g/L solution of potassium dihydrogen

— mobile phase A: mix 1 volume of triethylamine R and

Mobile phase:

— temperature: 35 °C.

silica gel for chromatography R (5 μ m);

— stationary phase: base-deactivated end-capped octadecylsilyl

Retention time Impurity E = about 5.5 min;
impurity F = about 8.5 min.

System suitability Reference solution (d):

— the chromatogram obtained is similar to the

chromatogram supplied with lamotrigine for peak

identification CRS.

Limit:

— impurity E: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

To the residue obtained in the test for sulfated ash add 2 mL of hydrochloric acid R and evaporate slowly to dryness on a water-bath. Moisten the residue with 0.05 mL of hydrochloric acid R, add 10 mL of boiling water R and heat the mixture for 10 min on a water-bath. Allow to cool to room

temperature, filter if necessary and adjust the volume of the filtrate and washings to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 2.000 g by drying in an oven at 105 °C at a pressure not exceeding 0.7 kPa for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 2.0 g.

ASSAY

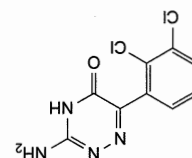
Dissolve 0.200 g in 60 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 25.61 mg of C₉H₇Cl₂N₅.

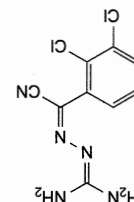
IMPURITIES

Specified impurities A, E, F, G.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use):

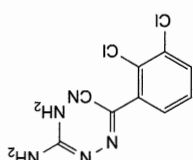


A. 3-amino-6-(2,3-dichlorophenyl)-1,2,4-triazin-5(4H)-one,

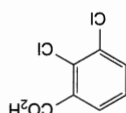


B. (2E)-[2-(diaminomethylidene)diazanylidene]diazanylidene[(2,3-

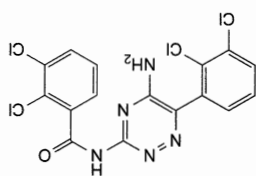
C. (2Z)-[2-(diaminomethylidene)diazanylidene]diazanylidene[(2,3-dichlorophenyl)acetonitrile],



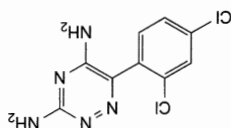
D. 6-(2,3-dichlorophenyl)-1,2,4-triazine-3,5-(2H,4H)-dione,



E. 2,3-dichlorobenzoic acid,



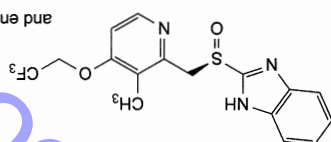
F. N-[5-amino-6-(2,3-dichlorophenyl)-1,2,4-triazin-3-yl]-2,3-dichlorobenzamide,



G. 6-(2,4-dichlorophenyl)-1,2,4-triazine-3,5-diamine.

Lansoprazole

(Ph. Eur. monograph 2219)



C₁₆H₁₄F₃N₃O₂S 369.4 103577-45-3

Action and use

Proton pump inhibitor; treatment of peptic ulcer disease.

Preparations

Gastro-resistant Lansoprazole Capsules

Gastro-resistant Lansoprazole Tablets

DEFINITION

2-[(R,S)-[[3-Methyl-4-(2,2,2-trifluoroethoxy)pyridin-2-yl]methyl]sulfinyl]-1H-benzimidazole.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or brownish powder.

Solubility

Practically insoluble in water, soluble in anhydrous ethanol, very slightly soluble in acetonitrile.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison lansoprazole CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in anhydrous ethanol R, evaporate to dryness and record new spectra using the residues.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution B₂ or BY₂ (2.2.2, Method II).

Dissolve 1.0 g in dimethylformamide R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and protect them from light.

Solvent mixture Mix 1 volume of triethylamine R and 60 volumes of water R and adjust to pH 10.5 with phosphoric acid R; mix this solution with 40 volumes of acetonitrile R₁.

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of lansoprazole for peak identification CRS (containing

impurities A and B) in 1.0 mL of the solvent mixture.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of 2-

hydroxybenzimidazole R (impurity D) and 5 mg of 2-mercaptobenzimidazole R (impurity E) in the solvent mixture and dilute to 100 mL with the solvent mixture. Dilute 1 mL of this solution to 10 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: amido-hexadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 1 volume of triethylamine R and 60 volumes of water R and adjust to pH 6.2 with phosphoric acid R; mix this solution with 40 volumes of acetonitrile R₁.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 285 nm.

Injection 10 μ L.

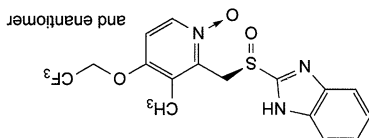
Run time 3 times the retention time of lansoprazole.

Identification of impurities Use the chromatogram supplied with lansoprazole for peak identification CRS and the

chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B; use the

chromatogram obtained with reference solution (c) to identify the peaks due to impurities D and E.

Relative retention With reference to lansoprazole (retention time = about 7 min): impurity D = about 0.4;



A, 2-[(R,S)-[[3-methyl-1-oxido-4-(2,2,2-trifluoroethoxy)pyridin-2-yl]methyl]sulfinyl]-1H-benzimidazole,

Specified impurities A, B, D, E
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C, F.

IMPURITIES

In an airtight container, protected from light.

STORAGE

1 mL of 0.1 M sodium hydroxide is equivalent to 36.94 mg of C₁₆H₁₄F₃N₃O₂S. (2.2.20).

Dissolve 0.300 g in 40 mL of ethanol (96 per cent) R and dilute to 50 mL with water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically.

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

— flow rate: 150 mL/min.

— heating time: 15 min;

— temperature: 50–70 °C;

the evaporation technique:

Maximum 0.1 per cent, determined on 0.150–0.200 g using

Water (2.5.32)

(0.05 per cent).

the chromatogram obtained with reference solution (b)

— disregard limit: 0.25 times the area of the principal peak in

(0.6 per cent);

in the chromatogram obtained with reference solution (b)

— total: not more than 3 times the area of the principal peak

(0.10 per cent);

chromatogram obtained with reference solution (b)

0.5 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than

(0.1 per cent);

chromatogram obtained with reference solution (b)

0.5 times the area of the principal peak in the

— impurities A, D, E: for each impurity, not more than

solution (b) (0.4 per cent);

peak in the chromatogram obtained with reference

— impurity B: not more than twice the area of the principal

peak area of impurity E by 0.4;

— correction factor: for the calculation of content, multiply the

Limits:

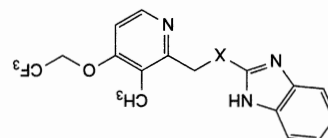
lansoprazole and impurity B.

— resolution: minimum 3.0 between the peaks due to

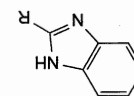
System suitability: reference solution (a):

impurity B = about 1.2.

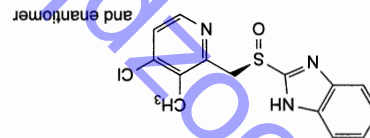
impurity A = about 0.5; impurity E = about 0.6;



B. X = SO₂; 2-[[[3-methyl-4-(2,2,2-trifluoroethoxy)pyridin-2-yl]methyl]sulfonyl]-1H-benzimidazole;
C. X = S; 2-[[[3-methyl-4-(2,2,2-trifluoroethoxy)pyridin-2-yl]methyl]sulfonyl]-1H-benzimidazole;



D. R = OH; 1H-benzimidazol-2-ol;
E. R = SH; 1H-benzimidazole-2-thiol;



F. 2-[(R,S)-[(4-chloro-3-methylpyridin-2-yl)methyl]sulfonyl]-1H-benzimidazole.

Lauromacrogol 400

(Ph. Eur. monograph 2046)

Action and use
Non-ionic surfactant.

DEFINITION

Mixture of lauryl alcohol (dodecanol) monomers of mixed macrogols. It may contain some free macrogols and it contains various amounts of free lauryl alcohol. The number of moles of ethylene oxide reacted per mole of lauryl alcohol is 9. The name of the substance is followed by a number (400) corresponding approximately to the average molecular mass of the macrogol portion.
This monograph applies to lauromacrogol 400 used as active substance.

CHARACTERS

Appearance
White or almost white, unctuous and hygroscopic mass, melting at 24 °C into a colourless or yellowish, viscous liquid.

Solubility

Freely soluble in water, very soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

A. Hydroxyl value (see Tests).
B. Saponification value (see Tests).
C. Warm the substance to be examined in an incubator at 50 °C for 1 h until fully molten and clear. Transfer 50 mL to a warmed cloud-point tube (flat-bottomed glass tube 30-33.5 mm in internal diameter and 115-125 mm high). Insert the tube into a cooling bath that allows the outer

surface of the tube to be in contact with chilled air, contained within a cylindrical metal container (internal diameter 9.5-12.5 mm greater than the external diameter of the sample tube, 115 mm high) that is surrounded by iced water. The base of the glass tube rests on a 6 mm thick cork disc, which prevents direct thermal contact with the cooled metal cylinder. Stir the substance to be examined continuously with a thermometer so that the temperature is constant throughout the substance. Periodically lift the tube out of the cooling bath to check for signs of cloudiness at the bottom of the tube. Examine the tube against a bright light source. When cloudiness is first observed, check more frequently until the substance becomes completely cloudy and the thermometer, suspended in the centre of the substance, is only just visible when viewed horizontally. Record the temperature. It is 20 °C to 25 °C.

TESTS

Appearance
The molten substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution GY₆ (2.2.2, Method I).

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of carbon dioxide-free water R and 10 mL of ethanol (96 per cent) R. Add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator to yellow.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

90 to 105, determined on 0.35 g.

Iodine value (2.5.4, Method A)

Maximum 2.0.

Peroxide value

Maximum 5.0.

Introduce 10.0 g into a 100 mL beaker, dissolve with glacial acetic acid R and dilute to 20 mL with the same solvent. Add 1 mL of saturated potassium iodide solution R, mix and allow to stand for 1 min. Add 50 mL of carbon dioxide-free water R. Titrate with 0.01 M sodium thiosulfate, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.
Determine the peroxide value using the following expression:

$$\frac{(n_1 - n_2) \times M \times 1000}{m}$$

n_1 = volume of 0.01 M sodium thiosulfate required for the substance to be examined, in millilitres;
 n_2 = volume of 0.01 M sodium thiosulfate required for the blank titration, in millilitres;
 M = molarity of the sodium thiosulfate solution, in moles per litre;
 m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)

Maximum 3.0.

Free lauryl alcohol (dodecanol)

Gas chromatography (2.2.28).
Test solution Dissolve 0.200 g of the substance to be examined in acetone R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 2.00 g of lauryl alcohol R in acetone R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 50.0 mL with acetone R.

Column: fused silica;

— size: $l = 30$ m, $\varnothing = 0.25$ mm;

— stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 0.1 μ m).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 50:1.

Temperature:

Temperature (°C)	Time (min)	Column
120	0 - 1	120
120 \rightarrow 350	1 - 23	350
350	23 - 33	300
350		Detector

Detection Flame ionisation.

Injection 1.0 μ L.

Retention time Lauryl alcohol = about 5 min.

Limit:

— free lauryl alcohol: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (2.0 per cent).

Free macroglols

Size-exclusion chromatography (2.2.30).

Test solution Dissolve 5.0 g of the substance to be examined in the mobile phase and dilute to 250.0 mL with the mobile phase.

Reference solution (a) Dissolve about 0.4 g of macroglol 1000 R in the mobile phase and dilute to 250.0 mL with the mobile phase.

Reference solution (b) Dilute 50.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Precolumns (2):

— size: $l = 0.125$ m, $\varnothing = 4$ mm;

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m) with a pore size of 10 nm.

Column:

— size: $l = 0.30$ m, $\varnothing = 7.8$ mm;

— stationary phase: hydroxylated polymethacrylate gel R (6 μ m) with a pore size of 12 nm.

Connect both precolumns to the column using a 3-way valve and switch the mobile phase flow according to the following programme:

— 0-14 s: precolumn 1 and column;

— 115 s to the end: precolumn 2 and column;

— 115 s to 8 min: flow back of precolumn 1.

Mobile phase water R, methanol R (2:8 V/V).

Flow rate 1.1 mL/min.

Detection Refractometer.

Injection 20 μ L.

Calculate the percentage content of free macroglols using the following expression:

$$\frac{A_1 \times m_2 \times 200}{A_1 \times (A_2 + 2A_3) + m_1}$$

m_1 = mass of the substance to be examined in the test solution, in grams;

m_2 = mass of macroglol 1000 R in reference solution (a), in grams;

A_1 = area of the peak due to free macroglols in the chromatogram obtained with the test solution;

A_2 = area of the peak due to macroglol 1000 in the chromatogram obtained with reference solution (a);

A_3 = area of the peak due to macroglol 1000 in the chromatogram obtained with reference solution (b).

Limit:

— free macroglols: maximum 3.0 per cent.

Average chain length of the fatty alcohol and average number of moles of ethylene oxide

Nuclear magnetic resonance spectrometry (2.2.33).

Test solution If the substance is in the solid state at room temperature, heat gently before sampling. Dissolve 0.4 mL of the substance to be examined in 0.3 mL of a mixture of 1 volume of deuterated methanol R and 2 volumes of deuterated chloroform R, containing 0.1 mol/L of chromium(III) acetylacetonate R as a relaxation aid.

Apparatus High resolution FT-NMR spectrometer operating at minimum 300 MHz.

Acquisition of ^{13}C NMR spectra The following parameters may be used:

— sweep width: 250 ppm (–15 ppm to 235 ppm);

— irradiation frequency offset: 110 ppm;

— time domain: 64 K;

— pulse delay: 3 s;

— pulse program: zgpg 30 (inverse gated, 30° excitation pulse);

— dummy scans: 4;

— number of scans: 2048.

Processing and plotting The following parameters may be used:

— size: 64 K (zero-filling);

— window multiplication: exponential;

— Lorentzian broadening factor: 1 Hz.

Use the CD₃OD signal for shift referencing. The shift of the central peak of the multiplet is set to 49.0 ppm.

Plot the spectral region 8 0.0–80.0 ppm. Compare the spectrum with the spectrum in Figure 2046-1. The shift values lie near the values given in Table 2046-1.

Table 2046-1. – Shift values

Signal	Shift (ppm)	Normalised integrals
CH ₃	14.4	0.989
CH ₂ (alkyl chain)	23.2	1.000
CH ₂ (alkyl chain)	25.5	1.001
CH ₂ (alkyl chain)	30	7.410
CH ₂ (alkyl chain)	32.5	0.963
CH ₂ (–CH ₂ –OH) (end CH ₂ -group of macroglol)	61.6	1.001
CH ₂ s (macroglol)	70.7	16.25
CH ₂ (R-CH-O-macroglol) (CH ₂ in alpha position)	72.6	0.998
CH ₂ (macroglol)	73.1	0.929

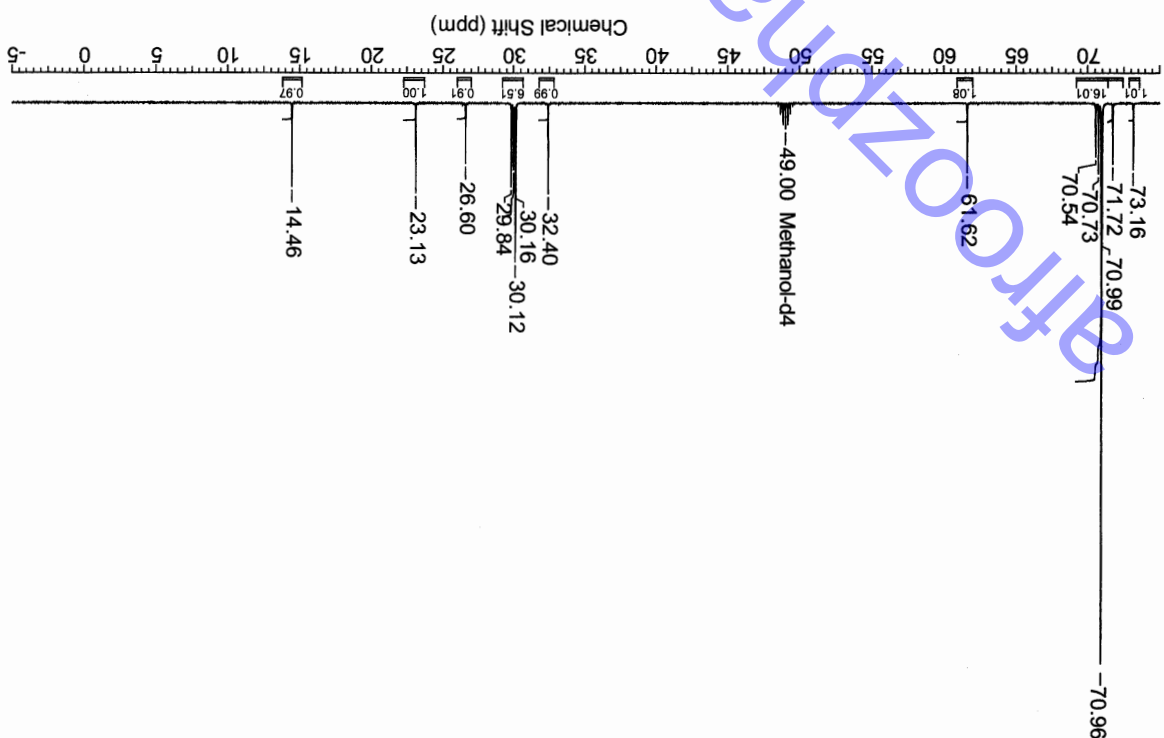


Figure 2046-1. ^{13}C NMR spectrum of lauroyl macroglyglycerol 400

Figure 2046-1. ^{13}C NMR spectrum of lauroyl macroglyglycerol 400

System suitability:

— signal-to-noise ratio: minimum 150, for the smallest

relevant peak (CH_2 at 73.1 ppm);

— peak width at half-height: maximum 0.05 ppm, for the

central CDCl_3 signal (at 8 78.6 ppm).

Calculation of the average chain length of the fatty alcohol and

the average number of moles of ethylene oxide Define the signal

at 23.2 ppm as 1.000 and normalise the integrals of the other

signals listed in Table 2046-1.

The average chain length of the fatty alcohol is calculated

using the following expression:

$$\Sigma I_{n,14-33} I_{n,i} + I_{n,72.6}$$

$\Sigma I_{n,14-33}$ = sum of the normalised integrals of the signals

$I_{n,i}$ = from 14 ppm to 33 ppm;

$I_{n,72.6}$ = normalised integral of the signal at 72.6 ppm.

The average number of moles of ethylene oxide is calculated

using the following expression:

$$0.5 \times (I_{n,62} + I_{n,71} + I_{n,73})$$

$I_{n,62}, I_{n,71}$ = normalised integral of the signals at 62 ppm,

$I_{n,73}$ = 71 ppm and 73 ppm respectively.

The sum of the normalised integrals of the signals at

62 ppm, 71 ppm and 73 ppm corresponds to the average

number of methylene groups in the macrogly part of

lauroyl macroglyglycerol 400.

Limits:

— average chain length of the fatty alcohol: 10.0 to 14.0;
— average number of moles of ethylene oxide: 7.0 to 11.0.

Appearance
Pale yellow waxy solid.

CHARACTERS

hydrogenated oils.

They are obtained by partial alcoholysis of saturated oils mainly containing triglycerides of lauric (dodecanoic) acid, using macrogly, or by esterification of glycerol and macrogly with saturated fatty acids, or by mixing glycerol esters and condensates of ethylene oxide with the fatty acids of these

Mixtures of monoesters, diesters and triesters of glycerol and monoesters and diesters of macrogly with a mean relative molecular mass between 300 and 1500.

DEFINITION

Ph Eur

Excipient.

Action and use

(Ph. Eur. monograph 1231)

Lauroyl Macroglyglycerides



Ph Eur

Maximum 0.2 per cent, determined on 2.0 g.

Total ash (2.4.16)

Maximum 2.0 per cent, determined on 0.500 g.

Water (2.5.12)

of dioxan.

Ethylene oxide and dioxan (2.4.25, Method A)
Maximum 1 ppm of ethylene oxide and maximum 10 ppm

Solubility

Dispersible in hot water, freely soluble in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 1.0 g of the substance to be examined in methylene chloride R and dilute to 20 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase hexane R, ether R (30:70 V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of rhodamine B R in ethanol (96 per cent) R and examine in ultraviolet light at 365 nm.

Results The chromatogram shows a spot due to triglycerides

with an R_f value of about 0.9 (R_{st} 1) and spots due to

1,3-diglycerides (R_{st} 0.7), to 1,2-diglycerides (R_{st} 0.6), to

monoglycerides (R_{st} 0.1) and to esters of macrogol (R_{st} 0).

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Fatty acid composition (see Tests).

TESTS

Drop point (2.2.17)

Introduce into the cup the substance to be examined, which has been melted by heating for 1 h in an oven at $100 \pm 2^\circ\text{C}$, and allow to stand for 5 h at about 5°C .

Ethylene oxide units per molecule (nominal value)	Type of macrogol	Drop point
---	------------------	------------

6 300 33 - 38

8 400 36 - 41

12 600 38 - 43

32 1500 42.5 - 47.5

Acid value (2.5.1)

Maximum 2.0, determined on 2.0 g.

Hydroxyl value (2.5.3, Method A)

Use 1.0 g.

Ethylene oxide units per molecule (nominal value)	Type of macrogol	Hydroxyl value
---	------------------	----------------

6 300 65 - 85

8 400 60 - 80

12 600 50 - 70

32 1500 36 - 56

Peroxide value (2.5.5, Method A)

Maximum 6.0, determined on 2.0 g.

Saponification value (2.5.6)

Use 2.0 g.

Ethylene oxide units per molecule (nominal value)	Type of macrogol	Saponification value
---	------------------	----------------------

6 300 190 - 204

8 400 170 - 190

12 600 150 - 170

32 1500 79 - 93

Alkaline impurities

Introduce 5.0 g into a test tube and carefully add a mixture, neutralised if necessary with 0.01 M hydrochloric acid or with 0.01 M sodium hydroxide, of 0.05 mL of a 0.4 g/L solution of bromophenol blue R in ethanol (96 per cent) R, 0.3 mL of water R and 10 mL of ethanol (96 per cent) R. Shake and allow to stand. Not more than 1.0 mL of 0.01 M hydrochloric acid is required to change the colour of the upper layer to yellow.

Free glycerol

Maximum 3.0 per cent.

Dissolve 1.20 g in 25.0 mL of methylene chloride R. Heat if necessary. After cooling, add 100 mL of water R. Shake and add 25.0 mL of periodic acid solution R. Shake and allow to stand for 30 min. Add 40 mL of a 75 g/L solution of potassium iodide R. Allow to stand for 1 min. Add 1 mL of starch solution R. Titrate the iodine with 0.1 M sodium thiosulfate. Carry out a blank titration.

1 mL of 0.1 M sodium thiosulfate is equivalent to 2.3 mg of glycerol.

Composition of fatty acids (2.4.22, Method A)

Composition of the fatty-acid fraction of the substance:

— caprylic acid: maximum 15.0 per cent;

— lauric acid: maximum 12.0 per cent;

— myristic acid: 30.0 per cent to 50.0 per cent;

— palmitic acid: 5.0 per cent to 25.0 per cent;

— stearic acid: 5.0 per cent to 35.0 per cent.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.0 g. Use a mixture

of 30 volumes of anhydrous methanol R and 70 volumes of methylene chloride R as solvent.

Total ash (2.4.16)

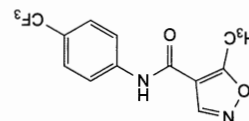
Maximum 0.1 per cent.

LABELLING

The label states the type of macrogol used (mean relative molecular mass) or the number of units of ethylene oxide per molecule (nominal value).

Leftunomide

(Ph Eur monograph 2330)


 $C_{12}H_9F_3N_2O_2$ 270.2 757-06-12-6

Action and use

Immunomodulator.

DEFINITION

5-Methyl-N'-[4-(trifluoromethyl)phenyl]phenylisoxazole-4-carboxamide.

CHARACTERS

98.0 per cent to 102.0 per cent (dried substance).

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methanol, sparingly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Heat the substance to be examined and the reference substance at 130 °C for 10 min.

Comparison leftunomide CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Store all solutions protected from light.

Test solution (a) Dissolve 25.0 mg of the substance to be

examined in 5 mL of acetonitrile for chromatography R and

dilute to 50.0 mL with the mobile phase.

Test solution (b) Dissolve 0.125 g of the substance to be

examined in 5 mL of acetonitrile for chromatography R and

dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of test solution (a) to

50.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 12.5 mg of leftunomide

impurity A CRS in 5 mL of acetonitrile for chromatography R

and dilute to 100.0 mL with the mobile phase. Dilute

10.0 mL of the solution to 100.0 mL with the mobile phase.

Dilute 2.0 mL of this solution to 100.0 mL with the mobile

phase.

Reference solution (c) Dissolve 25.0 mg of leftunomide CRS in

5 mL of acetonitrile for chromatography R and dilute to

50.0 mL with the mobile phase.

Reference solution (d) Dissolve the contents of 1 vial of

leftunomide for peak identification CRS (containing impurities B

and C) in 2.0 mL of the mobile phase and sonicate for

10 min.

Column: — size: $l = 0.125$ m, $\varnothing = 4.0$ mm;

stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 μ m).

Mobile phase Mix 5 volumes of triethylamine R with 650 volumes of water for chromatography R, adjust to pH 3.4 ± 0.1 with phosphoric acid R and add 350 volumes of acetonitrile for chromatography R.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L of test solutions (a) and (b) and reference

solutions (a), (b) and (d).

Run time Twice the retention time of leftunomide.

Identification of impurities Use the chromatogram supplied

with leftunomide for peak identification CRS and the

chromatogram obtained with reference solution (d) to

identify the peaks due to impurities B and C.

Relative retention With reference to leftunomide (retention

time = about 25 min): impurity B = about 0.2;

impurity A = about 0.4; impurity C = about 0.9.

System suitability Reference solution (d):

— peak-to-valley ratio: minimum 3, where H_p = height above

the baseline of the peak due to impurity C and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

leftunomide.

Limits: test solution (a):

— impurity B: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

— sum of impurities C and E: not more than the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent);

Limit: test solution (b):

— impurity A: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.01 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.3 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum

crucible.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution (a) and reference solution (c).

Calculate the percentage content of $C_{12}H_9F_3N_2O_2$ from the

declared content of leftunomide CRS.

STORAGE

Protected from light.

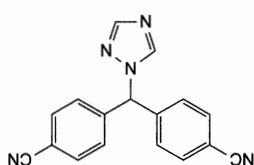
IMPURITIES

Specified impurities A, B



Letrozole

(Ph Eur monograph 2334)



C₁₇H₁₁N₅ 285.3 112809-51-5

Action and use

Aromatase inhibitor; treatment of breast carcinoma.

Ph Eur

DEFINITION

4,4'-(1*H*-1,2,4-Triazol-1-ylmethylene)dibenzonitrile.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison letrozole CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in 15 mL of acetonitrile R1 and dilute to 50.0 mL with water R.

Test solution (b) To 2.0 mL of test solution (a) add 30 mL of acetonitrile R1 and dilute to 100.0 mL with water R.

Reference solution (a) Dissolve 5.0 mg of letrozole CRS (containing impurities A and B) in 3 mL of acetonitrile R1 and dilute to 10.0 mL with water R.

Reference solution (b) To 2.0 mL of test solution (a) add 30 mL of acetonitrile R1 and dilute to 100.0 mL with water R.

Reference solution (c) Dissolve 25.0 mg of letrozole CRS in 15 mL of acetonitrile R1 and dilute to 50.0 mL with water R.

To 2.0 mL of this solution add 30 mL of acetonitrile R1 and dilute to 100.0 mL with water R.

To 1.0 mL of this solution add 6 mL of acetonitrile R1 and dilute to 20.0 mL with water R.

Column: dilute to 100.0 mL with water R.

size: $l = 0.125$ m, $\phi = 4.6$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R

Mobile phase: (5 μ m).

Mobile phase A: water R;

Mobile phase B: acetonitrile R1;

Time (min)

0 - 4 70

4 - 29 30 \rightarrow 30

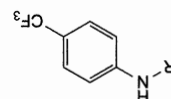
29 - 30 30 \rightarrow 70

Mobile phase B (per cent V/V)

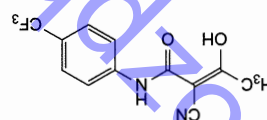
Mobile phase A (per cent V/V)

Mobile phase B (per cent V/V)

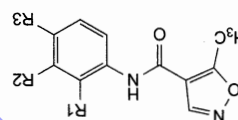
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C, D, E, F, G, H.



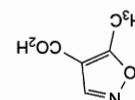
A. R = H: 4-(trifluoromethyl)aniline,
H. R = CO.CH₂.CN: 2-cyano-N-[4-(trifluoromethyl)phenyl]acetamide,



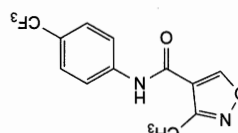
B. (2*Z*)-2-cyano-3-hydroxy-N-[4-(trifluoromethyl)phenyl]but-2-enamide (trifluoromethyl),



C. R1 = R3 = H, R2 = CF₃: 5-methyl-N-[3-(trifluoromethyl)phenyl]isoxazole-4-carboxamide,
F. R1 = CF₃, R2 = R3 = H: 5-methyl-N-[2-(trifluoromethyl)phenyl]isoxazole-4-carboxamide,
G. R1 = R2 = H, R3 = CH₃: 5-methyl-N-(4-methylphenyl)isoxazole-4-carboxamide,



D. 5-methylisoxazole-4-carboxylic acid,



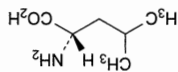
E. 3-methyl-N-[4-(trifluoromethyl)phenyl]isoxazole-4-carboxamide.

Ph Eur



Leucine

(Ph. Eur. monograph 0771)



131.2

C₆H₁₃NO₂

61-90-5

Action and use

Amino acid.

Ph Eur

DEFINITION

(2S)-2-Amino-4-methylpentanoic acid.

Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or shiny flakes.

Solubility

Sparingly soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification A, B.

Second identification A, C.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24)

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a 10.3 g/L solution of hydrochloric acid R and dilute to

50 mL with the same solution.

Reference solution Dissolve 10 mg of leucine CRS in a 10.3 g/L

solution of hydrochloric acid R and dilute to 50 mL with the

same solution.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R

(20:20:60 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C

for 15 min.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

reference solution.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured

than reference solution BY₆ (2.2.2, Method II).

Dissolve 0.5 g in a 103 g/L solution of hydrochloric acid R and

dilute to 10 mL with the same solution.

Specific optical rotation (2.2.7)

+ 14.5 to + 16.5 (dried substance).

Dissolve 1.00 g in hydrochloric acid R1 and dilute to 25.0 mL

with the same acid.

Flow rate 1.0 mL/min.
Detection Spectrophotometer at 230 nm.
Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Relative retention With reference to leucine (retention

time = about 13 min): impurity A = about 0.6;

impurity B = about 1.9.

System suitability: reference solution (a):

— resolution: minimum 5 between the peaks due to

impurity A and leucine;

— the chromatogram obtained is similar to the

chromatogram supplied with leucine CRS.

Limits:

— impurity A: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.3 per cent);

— impurity B: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Water (2.5.12)

Maximum 0.3 per cent, determined on 1.000 g. Use a

validated pyridine-free iodosulfurous reagent.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modifications.

Injection Test solution (b) and reference solution (c).

System suitability: reference solution (c):

— symmetry factor: maximum 1.7 for the peak due to

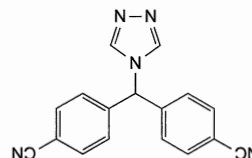
leucine.

Calculate the percentage content of C₁₇H₁₁N₅ from the

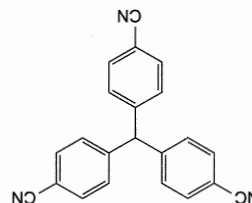
declared content of leucine CRS.

IMPURITIES

Specified impurities: A, B.



A, 4,4'-(4H-1,2,4-triazol-4-ylmethylene)dibenzonitrile,



B, 4,4',4''-methanetriyltribenzonitrile.

Ph Eur

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method 1.

The concentrations of the test solutions and the reference

solutions may be adapted according to the sensitivity of the

equipment used. The concentrations of all solutions are

adjusted so that the system suitability requirements described

in general chapter 2.2.46 are fulfilled, keeping the ratios of

concentrations between all solutions as described.

Solution A dilute hydrochloric acid R1 or a sample preparation

buffer suitable for the apparatus used.

Test solution (a) Dissolve 30.0 mg of the substance to be

examined in solution A and dilute to 50.0 mL with

solution A.

Test solution (b) Dilute 1.0 mL of test solution (a) to

25.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of test solution (a) to

100.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of isoleucine R

(impurity A) in solution A and dilute to 100.0 mL with

solution A. Dilute 1.0 mL of the solution to 250.0 mL with

solution A. Dilute 1.0 mL of this solution to 10.0 mL with

solution A.

Reference solution (c) Dissolve 30.0 mg of proline R in

solution A and dilute to 100.0 mL with solution A. Dilute

1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (d) Dilute 6.0 mL of ammonium standard

solution (100 ppm NH₄⁺) R to 50.0 mL with solution A. Dilute

1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (e) Dissolve 30 mg of isoleucine R

(impurity A) and 30 mg of leucine R in solution A and dilute

to 50.0 mL with solution A. Dilute 1.0 mL of the solution to

200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference

solutions into the amino acid analyser. Run a program

suitable for the determination of physiological amino acids.

System suitability: reference solution (e):

— resolution: minimum 1.5 between the peaks due to

impurity A and leucine.

Calculation of percentage contents:

— for impurity A in test solution (b), use the concentration

of impurity A in reference solution (b);

— for any ninhydrin-positive substance detected at 570 nm

in test solution (a), use the concentration of leucine in

reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm

in test solution (a), use the concentration of proline in

reference solution (c); if a peak is above the reporting

threshold at both wavelengths, use the result obtained at

570 nm for quantification;

— for ammonium in test solution (a), use the concentration

of ammonium in reference solution (d) taking into

account the corresponding peak in the chromatogram

obtained with the blank solution.

Limits:

— impurity A at 570 nm: maximum 0.8 per cent;

— any ninhydrin-positive substance: for each impurity,

maximum 0.2 per cent;

— ammonium at 570 nm: maximum 0.02 per cent;

— total: maximum 1.0 per cent;

— reporting threshold (excluding ammonium): 0.05 per cent.

The thresholds indicated under Related substances (Table 2034-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Chlorides (2.4.4)

Maximum 200 ppm.

Dissolve 0.25 g in water R and dilute to 15 mL with the

same solvent.

Sulfates (2.4.13)

Maximum 300 ppm.

Dissolve 0.5 g in 3 mL of dilute hydrochloric acid R and dilute

to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of dilute

hydrochloric acid R. Shake with 3 quantities, each of 10 mL,

of methyl isobutyl ketone R1, shaking for 3 min each time.

To the combined organic layers add 10 mL of water R and

shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent water R.

0.25 g complies with test H. Prepare the reference solution

using 0.25 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 3 mL of anhydrous formic acid R.

Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 13.12 mg of

C₆H₁₃NO₂.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A.

Other detectable impurities: the following substances would, if

present at a sufficient level, be detected by one or other of

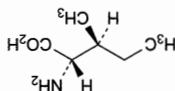
the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities. It is

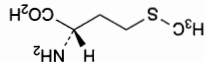
therefore not necessary to identify these impurities for

demonstration of compliance. See also 5.10. Control of

impurities in substances for pharmaceutical use: B, C, D, E.



A. (2S,3S)-2-amino-3-methylpentanoic acid (isoleucine).



B. (2S)-2-amino-4-(methylsulfonyl)butanoic acid (methionine).

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids taking one seventh of the sum of the number of moles of histidine, glutamic acid, leucine, proline, tyrosine and arginine as equal to 1. The values fall within the following limits: serine present; glutamic acid = 0.85 to 1.1; proline = 0.85 to 1.1; leucine = 1.8 to 2.2; tyrosine = 0.85 to 1.1; histidine = 0.85 to 1.1 and arginine = 0.85 to 1.1. Not more than traces of other amino acids are present, with the exception of tryptophan.

TESTS

Specific optical rotation (2.2.7)

— 38.0 to – 42.0 (anhydrous and acetic acid-free substance). Dissolve the substance to be examined in a 1 per cent *W/V* solution of *glacial acetic acid R* to obtain a concentration of 10.0 mg/mL.

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution (a) Dissolve the substance to be examined in the mobile phase to obtain a concentration of 1.0 mg/mL.

Test solution (b) Dilute 0.5 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve *leuporelin CRS* in the mobile phase to obtain a concentration of 1.0 mg/mL.

Reference solution (b) Dilute 0.5 mL of reference solution (a) to 10.0 mL with the mobile phase.

Resolution solution Dilute 5.0 mL of reference solution (a) to 50.0 mL with *water R*. To 5 mL of the solution add 100 µL of 1 M *sodium hydroxide* and shake vigorously. Heat in an oven at 100 °C for 60 min, cool immediately and add 50 µL of dilute phosphoric acid *R*. Shake vigorously.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography *R* (3 µm).

Mobile phase Dissolve about 15.2 g of *triethylamine R* in 800 mL of *water R*, adjust to pH 3.0 with phosphoric acid *R* and dilute to 1000 mL with *water R*. Add 850 mL of this solution to 150 mL of a mixture of 2 volumes of *propanol R* and 3 volumes of *acetonitrile R*.

Flow rate 1.0–1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and the resolution solution.

Run time 90 min.

Relative retention With reference to leuporelin (retention time = 41–49 min): impurity E = about 0.7;

impurity F = about 0.7; impurity H = about 0.78;

impurity A = about 0.8; impurity B = about 0.9;

impurity I = about 0.94; impurity J = about 1.09;

impurity C = about 1.2; impurity G = about 1.3;

impurity K = about 1.31; impurity D = about 1.5.

System suitability Resolution solution:

— resolution: minimum 1.5 between the peaks due to impurity B and leuporelin.

Limits:

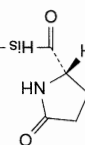
— impurity D: maximum 1.0 per cent;

— impurities A, B, C: for each impurity, maximum 0.5 per cent;

— unspecified impurities: for each impurity, maximum 0.5 per cent;

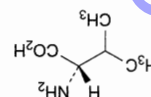
— total: maximum 2.5 per cent;

Leuporelin

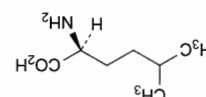


Ph Eur

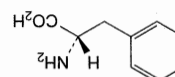
E. (2*S*)-2-amino-3-methylbutanoic acid (valine).



D. (2*S*)-2-amino-5-methylhexanoic acid (5-methylnorleucine),



C. (2*S*)-2-amino-3-phenylpropanoic acid (phenylalanine),



Ph Eur

DEFINITION

5-Oxo-L-prolyl-L-histidyl-L-tyrosyl-L-seryl-L-tyrosyl-D-leucyl-L-leucyl-L-arginyl-L-ethyl-L-prolinamide.

Synthetic nonapeptide analogue of the hypothalamic peptide, gonadorelin. It is obtained by chemical synthesis and is available as an acetate.

Content

97.0 per cent to 103.0 per cent (anhydrous and acetic acid-free substance).

CHARACTERS

Appearance

Hygrosopic, white or almost white powder.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of potassium bromide *R*.

Comparison Ph. Eur. reference spectrum of leuporelin.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (b).

C. Amino acid analysis (2.2.56). For hydrolysis use Method 1 and for analysis use Method 1.

— disregard limit: 0.1 per cent.

Acetic acid (2.5.34)

4.7 per cent to 9.0 per cent.

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Water (2.5.32)

Maximum 5.0 per cent.

Sulfated ash (2.4.14)

Maximum 0.3 per cent.

Bacterial endotoxins (2.6.14, Method D)

Less than 16.7 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Run time 60 min.

Injection 20 µL of test solution (b) and reference solution (b). Calculate the content of leuporelin ($C_{59}H_{84}N_{16}O_{12}$) using the areas of the peaks and the declared content of $C_{59}H_{84}N_{16}O_{12}$ in leuporelin CRS.

STORAGE

In an airtight container, protected from light, at a temperature not exceeding 30 °C.

If the substance is sterile, store in a sterile, tamper-proof container.

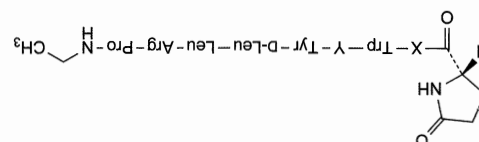
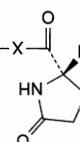
LABELLING

The label states the mass of peptide in the container.

IMPURITIES

Specified impurities A, B, C, D

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, F, G, H, I, J, K.

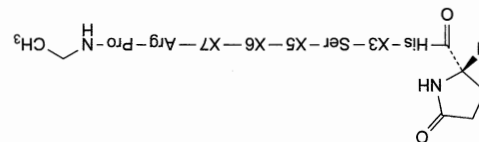
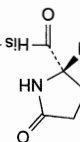


A. X = L-His, Y = D-Ser: [4-D-serine]leuporelin,

B. X = D-His, Y = L-Ser: [2-D-histidine]leuporelin,

F. X = D-His, Y = D-Ser: [2-D-histidine,4-D-

serine]leuporelin,



C. X3 = L-Trp, X5 = L-Tyr, X6 = X7 = L-Leu: [6-L-leucine]leuporelin,

E. X3 = D-Trp, X5 = L-Tyr, X6 = D-Leu, X7 = L-Leu:

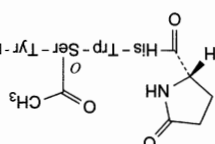
[3-D-typtophane]leuporelin,

G. X3 = L-Trp, X5 = D-Tyr, X6 = D-Leu, X7 = L-Leu:

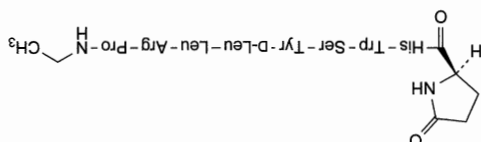
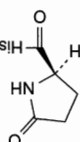
[5-D-tyrosine]leuporelin,

H. X3 = L-Trp, X5 = L-Tyr, X6 = X7 = D-Leu: [7-D-

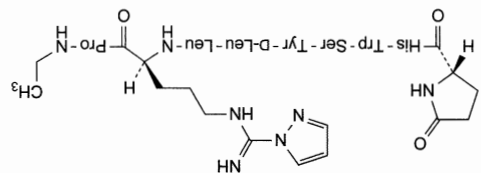
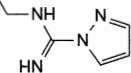
leucine]leuporelin,



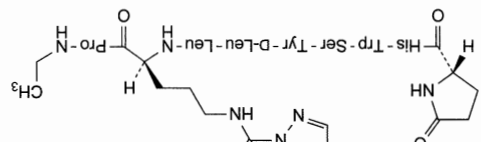
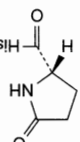
D. [4-(O-acetyl-L-serine)]leuporelin,



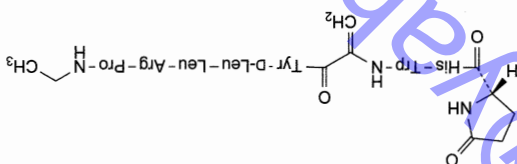
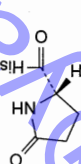
I. [1-(5-oxo-D-proline)]leuporelin,



J. [8-[5-N-[imino(1H-pyrazol-1-yl)methyl]-L-ornithine]]leuporelin,

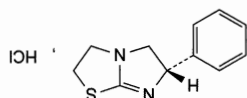


K. [4-dehydroalanine]leuporelin.



Levamisole Hydrochloride

(Ph. Eur. monograph 0726)



C₁₁H₁₃ClN₂S 240.8 16595-80-5

Action and use

Immunostimulant; antihelminthic.

DEFINITION

(6S)-6-Phenyl-2,3,5,6-tetrahydroimidazo[2,1-b]thiazole hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison levamisole hydrochloride CRS.

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y; (2.2.2, Method II).

pH (2.2.3)

3.0 to 4.5 for solution S.

Specific optical rotation (2.2.7)

–128 to –121 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use, protect from light and keep below 25 °C.

Test solution Dissolve 0.100 g of the substance to be examined in methanol R, add 1.0 mL of concentrated

ammonia R and dilute to 10.0 mL with methanol R.

Reference solution (a) Dissolve 10 mg of levamisole

hydrochloride for system suitability CRS (containing impurities

A, B, C, D and E) in methanol R, add 0.1 mL of concentrated

ammonia R and dilute to 1.0 mL with methanol R.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with methanol R. Dilute 5.0 mL of this solution to

25.0 mL with methanol R.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for

chromatography R (3 μ m).

Mobile phase:

— mobile phase A: dissolve 0.5 g of ammonium dihydrogen

phosphate R in 90 mL of water R, adjust to pH 6.5 with a

40 g/L solution of sodium hydroxide R and dilute to

100 mL with water R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	90 \rightarrow 30	10 \rightarrow 70
8 - 10	30	70

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 215 nm.

Equilibration At least 4 min with the mobile phase at the

initial composition.

Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with levamisole hydrochloride for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to levamisole (retention

time = about 3 min): impurity A = about 0.9;

impurity B = about 1.4; impurity C = about 1.5;

impurity D = about 1.6; impurity E = about 2.0.

— the chromatogram obtained with reference solution (a) is similar to the chromatogram supplied with levamisole

hydrochloride for system suitability CRS;

— **symmetry factor:** maximum 3.5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the

corresponding correction factor: impurity A = 2.0;

impurity B = 1.7; impurity C = 2.9; impurity D = 1.3;

impurity E = 2.7;

— **impurities A, B, C, D, E:** for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— **total:** not more than 1.5 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.3 per cent);

— **disregard limit:** 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 30 mL of ethanol (96 per cent) R and add

5.0 mL of 0.01 M hydrochloric acid. Carry out a

potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 24.08 mg of

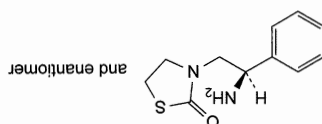
C₁₁H₁₃ClN₂S.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C, D, E



A. 3-[(2*R*S)-2-amino-2-phenylethylthio]thiazolidin-2-one,

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Dissolve 2.0 g in water R and dilute to 10.0 mL with the same solvent.

Enantiomeric purity

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 0.200 g of the substance to be examined in 2-propanol R and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution Dissolve 5 mg of the substance to be examined and 5 mg of levetiracetam impurity D CRS in the mobile phase.

mobile phase and dilute to 5.0 mL with the mobile phase. Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel OD for chiral separations R.

Mobile phase 2-propanol R, hexane R (18:82 V/V).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L.

Run time 1.4 times the retention time of levetiracetam.

Relative retention With reference to levetiracetam (retention time = about 12 min): impurity D = about 0.8.

System suitability: reference solution:

— resolution: minimum 1.5 between the peaks due to impurity D and levetiracetam;

— symmetry factor: maximum 2.0 for the peak due to levetiracetam.

Limit

— impurity D: maximum 0.8 per cent.

Impurity C

Liquid chromatography (2.2.29).

Solvent mixture water R, acetonitrile R1 (7:93 V/V).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 1 mg of the substance to be examined and 1 mg of levetiracetam impurity C CRS in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (a) Dissolve 1 mg of the substance to be examined and 1 mg of levetiracetam impurity C CRS in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of levetiracetam impurity C CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel for chromatography R (5 μ m).

Mobile phase 1.96 g/L solution of sulfuric acid R, acetonitrile R1 (7:93 V/V).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L.

Run time Twice the retention time of levetiracetam.

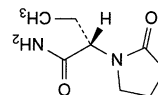
Relative retention With reference to levetiracetam (retention time = about 14 min): impurity C = about 1.2.

System suitability: reference solution (a):

— resolution: minimum 4.0 between the peaks due to levetiracetam and impurity C.

Levetiracetam

(Ph. Eur. monograph 2535)



C₈H₁₄N₂O₂ 170.2 102767-28-2

Action and use
Antiepileptic.

DEFINITION

(2S)-2-(2-Oxopropylidino-1-yl)butanamide.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Very soluble in water, soluble in acetonitrile, practically insoluble in hexane.

IDENTIFICATION

Carry out either tests A, B or tests B, C.

A. Specific optical rotation (2.2.7): -82 to -76.

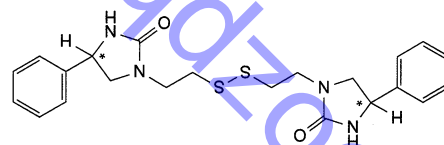
Dissolve 0.500 g in water R and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

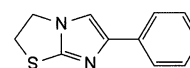
Comparison levetiracetam CRS.

C. Enantiomeric purity (see Tests).

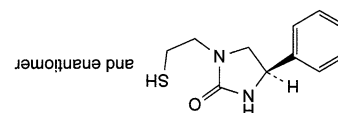
E. 1,1'-[(disulfane-1,2-diyl)bis(ethylene)]bis[(4R)-4-phenylimidazolidin-2-one].



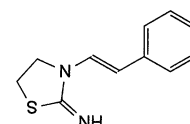
D. 6-phenyl-2,3-dihydroimidazo[2,1-b]thiazole,



C. (4R)-4-phenyl-1-(2-sulfanyylethyl)imidazolidin-2-one,



B. 3-[(E)-2-phenylethenyl]thiazolidin-2-imine,



Limit:

— *impurity C*: not more than 0.25 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (250 ppm).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture water R, acetonitrile R1 (4:96 V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of the substance to be examined and 5 mg of 2-pyrrolidone R in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (b) Dissolve 50.0 mg of levettiracetam CRS in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5 mg of levettiracetam

impurity A CRS and 5 mg of levettiracetam *impurity B* CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 100.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel for chromatography R (5 μ m).

Mobile phase 1.96 g/L solution of sulfuric acid R, acetonitrile R1 (4:96 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 10 μ L of the test solution and reference solutions (a), (c) and (d).

Run time Twice the retention time of levettiracetam.

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peaks due to

impurities A and B.

Relative retention With reference to levettiracetam (retention time = about 10 min): *impurity A* = about 0.5;

2-pyrrolidone = about 1.1; *impurity B* = about 1.2.

System suitability: reference solution (a):

— *resolution*: minimum 1.5 between the peaks due to levettiracetam and 2-pyrrolidone.

Limits:

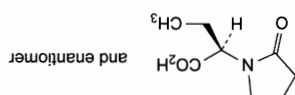
— *correction factor*: for the calculation of content, multiply the peak area of *impurity B* by 0.5;

— *impurity A*: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

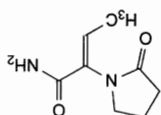
— *impurity B*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent);

— *unspecified impurities*: for each *impurity*, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent);

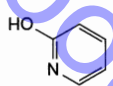
— *sum of unspecified impurities*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);



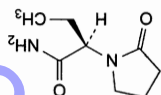
A. (2R,5S)-2-(2-oxopyrrolidin-1-yl)butanoic acid,



B. (2Z)-2-(2-oxopyrrolidin-1-yl)but-2-enamide,



C. pyridin-2-ol,



D. (2R)-2-(2-oxopyrrolidin-1-yl)butanamide ((R)-citracetam).

IMPURITIES

Specified impurities A, B, C, D.

declared content of levettiracetam CRS.

Calculate the percentage content of $C_8H_{14}N_2O_2$ from the

Injection Test solution and reference solution (b).

related substances with the following modification.

Liquid chromatography (2.2.29) as described in the test for

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 0.300 g.

Water (2.5.32)

lead standard solution (1 ppm Pb) R.

Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution

Maximum 10 ppm.

Heavy metals (2.4.8)

(0.03 per cent).

the chromatogram obtained with reference solution (c)

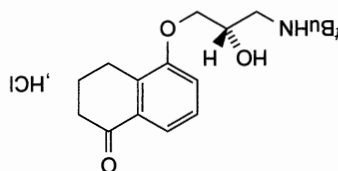
— *disregard limit*: 0.6 times the area of the principal peak in

(0.4 per cent);

in the chromatogram obtained with reference solution (c)

— *total*: not more than 8 times the area of the principal peak

Levobunolol Hydrochloride


 $C_{17}H_{25}NO_3 \cdot HCl$

327.9

27912-14-7

Action and use

Beta-adrenoceptor antagonist.

Preparation

Levobunolol Eye Drops

DEFINITION

Levobunolol Hydrochloride is (S)-5-(3-*tert*-butylamino-2-hydroxypropoxy)-1,2,3,4-tetrahydronaphthalen-1-one hydrochloride. It contains not less than 98.5% and not more than 101.0% of $C_{17}H_{25}NO_3 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or pinkish white, crystalline powder. Freely soluble in water; sparingly soluble in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of levobunolol hydrochloride (RS 200).

B. Yields the reactions characteristic of chlorides, Appendix VI.

TESTS

Acidity

pH of a 5% w/v solution, 4.5 to 6.5, Appendix V L.

Specific optical rotation

In a 3% w/v solution in methanol, -19.0 to -20.0, calculated with reference to the dried substance, Appendix V F.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using solutions in the mobile phase containing (1) 0.10% w/v of the substance being examined, (2) 0.0050% w/v of the substance being examined, (3) 0.0050% w/v of each of levobunolol hydrochloride BPGRS and atenolol.

The chromatographic procedure described under Assay may be used.

For solution (1) allow the chromatography to proceed for 3 times the retention time of the principal peak. The test is not valid unless, in the chromatogram obtained with solution (3), the resolution factor between the two principal peaks is at least 8.

In the chromatogram obtained with solution (1) the area of any secondary peak is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.5%) and the sum of the areas of any secondary peaks is not greater than twice the area of the principal peak in the chromatogram obtained with solution (2) (1%).

Loss on drying

When dried over phosphorus pentoxide at 110° at a pressure not exceeding 2 kPa for 4 hours, loses not more than 0.5% of its weight. Use 1 g.

IMPURITIES

Levobunolol Hydrochloride should be protected from light.

STORAGE

hydrochloride BPGRS.

Calculate the content of $C_{17}H_{25}NO_3 \cdot HCl$ from the declared content of $C_{17}H_{25}NO_3 \cdot HCl$ in levobunolol

The test is not valid unless in the chromatogram obtained with solution (3) the resolution factor between the two principal peaks is at least 8.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 3.9 mm) packed with end-capped octylsilyl silica gel for chromatography (10 μm) (Lichrosorb RP 8 is suitable), (b) a solution prepared by mixing 53 volumes of 0.005M sodium heptanesulfonate in methanol with 47 volumes of 0.005M sodium heptanesulfonate in water containing 1 mL of 0.5M sulfuric acid as the mobile phase with a flow rate of 1 mL per minute and (c) a detection wavelength of 223 nm.

Carry out the method for liquid chromatography, Appendix III D, using solutions in the mobile phase containing (1) 0.01% w/v of the substance being examined, (2) 0.01% w/v of levobunolol hydrochloride BPGRS and (3) 0.0050% w/v of each of levobunolol hydrochloride BPGRS and atenolol.

ASSAY

Carry out the method for liquid chromatography, Appendix III D, using solutions in the mobile phase containing (1) 0.01% w/v of the substance being examined, (2) 0.01% w/v of levobunolol hydrochloride BPGRS and (3) 0.0050% w/v of each of levobunolol hydrochloride BPGRS and atenolol.

The test is not valid unless in the chromatogram obtained with solution (3) the resolution factor between the two principal peaks is at least 8.

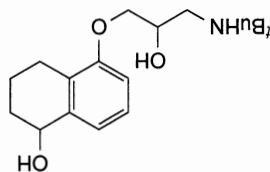
Calculate the content of $C_{17}H_{25}NO_3 \cdot HCl$ from the declared content of $C_{17}H_{25}NO_3 \cdot HCl$ in levobunolol

hydrochloride BPGRS.

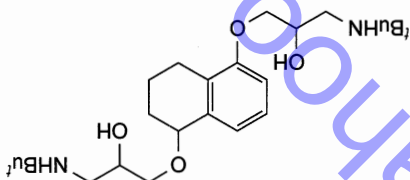
STORAGE

Levobunolol Hydrochloride should be protected from light.

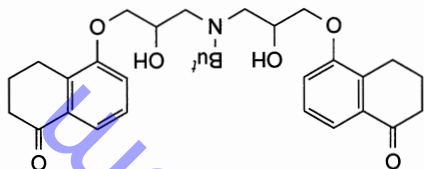
IMPURITIES



A. 5-(3-*tert*-butylamino-2-hydroxypropoxy)-1,2,3,4-tetrahydro-1-naphthol



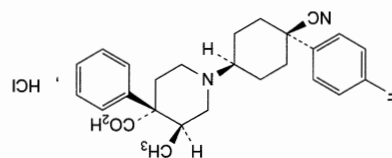
B. 1,1'-(1,2,3,4-tetrahydro-1,5-naphthalenedioxy)bis(3-*tert*-butylamino)-2-propanol



C. meso-5,5'-[(3,3'-*tert*-butylamino)bis(2-hydroxypropoxy)]bis-3,4-dihydronaphthalen-1(2H)-one

Levocabastine Hydrochloride

(Ph. Eur. monograph 1484)



$C_{26}H_{30}ClFN_2O_2$ 457.0 79547-78-7

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Ph Eur

DEFINITION

(3S,4R)-1-[*cis*-4-(4-fluorophenyl)cyclohexyl]-3-methyl-4-piperidine-4-carboxylic acid hydrochloride.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, sparingly soluble in methanol,

slightly soluble in ethanol (96 per cent) and in a 2 g/L

solution of sodium hydroxide.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison levocabastine hydrochloride CRS.

C. Dissolve 50 mg in a mixture of 0.4 mL of ammonia R and

2 mL of water R. Mix, allow to stand for 5 min and filter.

Acidify the filtrate with dilute nitric acid R. It gives

reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.250 g in methanol R and dilute to 25.0 mL with

the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution Y₇ (2.2.2. Method II).

Specific optical rotation (2.2.7)

–106 to –102 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected

from light.

Test solution Dissolve 50 mg of the substance to be examined

in methanol R1 and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of

levocabastine for system suitability 1 CRS (containing impurities

A, B, E, J and K) in 1.0 mL of methanol R1.

Reference solution (b) Dilute 1.0 mL of the test solution to

10.0 mL with methanol R1.

Column:

— size: $l = 0.10$ m, $\phi = 2.1$ mm;

— stationary phase: end-capped phenylsilyl silica gel for

chromatography R (1.7 μ m);

— temperature: 60 °C.

Mobile phase:

— mobile phase A: 17 g/L solution of tetrabutylammonium

hydrogen sulfate R₃

— mobile phase B: acetonitrile R1;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 0.5	95	5
0.5 - 3.5	95 → 90	5 → 10
3.5 - 6.0	90 → 85	10 → 15
6.0 - 11.0	85 → 70	15 → 30
11.0 - 14.5	70 → 20	30 → 80
14.5 - 15.5	20	80

Flow rate 0.45 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 2.0 μ L.

Identification of impurities Use the chromatogram supplied

with levocabastine for system suitability 1 CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities A, B, E, J and K.

Relative retention With reference to levocabastine (retention

time = about 6.5 min): impurity A = about 0.85;

impurity J = about 0.86; impurity B = about 0.90;

impurity E = about 0.94; impurity K = about 1.07.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 2.9, where H_p = height

above the baseline of the peak due to impurity K and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

levocabastine; minimum 5.0, where H_p = height above

the baseline of the peak due to impurity J and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

impurity A.

Calculation of percentage contents:

— for each impurity, use the concentration of levocabastine

in reference solution (b).

Limits:

— impurity E: maximum 0.4 per cent;

— impurity A: maximum 0.2 per cent;

— impurity B: maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.6 per cent;

— reporting threshold: 0.05 per cent.

Impurity C

Liquid chromatography (2.2.29). Carry out the test protected

from light.

Test solution Dissolve 50 mg of the substance to be examined

in methanol R1 and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of

levocabastine for system suitability 2 CRS (containing

impurity C) in 1.0 mL of methanol R1.

Reference solution (b) Dilute 1.0 mL of the test solution to

20.0 mL with methanol R1. Dilute 1.0 mL of this solution to

10.0 mL with methanol R1.

Column:

— size: $l = 0.15$ m, $\phi = 2.1$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (1.8 μ m);

— temperature: 35 °C.

Mobile phase:
— mobile phase A: 17 g/L solution of tetrabutylammonium hydrogen sulfate R₃
— mobile phase B: acetonitrile R₁;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 0.5	90	10
0.5 - 15.5	90 → 80	10 → 20
15.5 - 20.5	80 → 50	20 → 50

Flow rate 0.30 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 2.0 µL.

Identification of impurities Use the chromatogram supplied

with levocabastine for system suitability 2 CRS and the

chromatogram obtained with reference solution (a) to

identify the peak due to impurity C.

Relative retention With reference to levocabastine (retention

time = about 16 min): impurity C = about 0.98.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 10.0, where H_p = height

above the baseline of the peak due to impurity C and

H_b = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

levocabastine.

Calculation of percentage content:

— for impurity C, use the concentration of levocabastine in

reference solution (b).

Limit:

— impurity C: maximum 0.3 per cent.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum

crucible.

ASSAY

Dissolve 0.175 g in 50 mL of ethanol (96 per cent) R₃,

previously neutralised to phenol red solution R₃, and add

5.0 mL of water R₃. Carry out a potentiometric titration

the 2nd point of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 22.85 mg of

C₂₆H₃₀ClFN₂O₂.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C, E

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

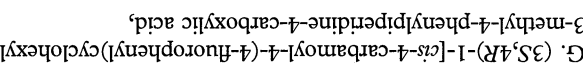
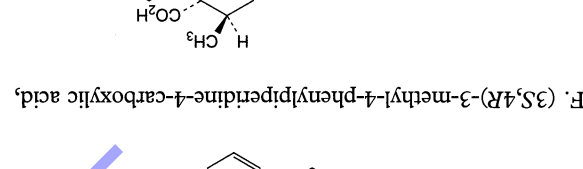
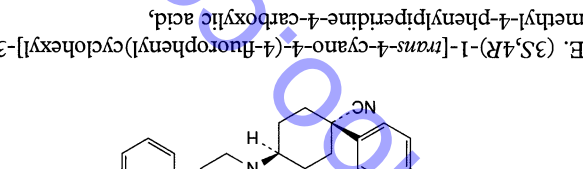
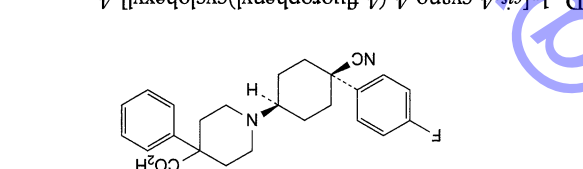
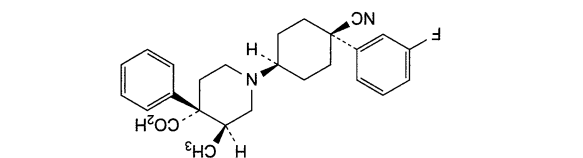
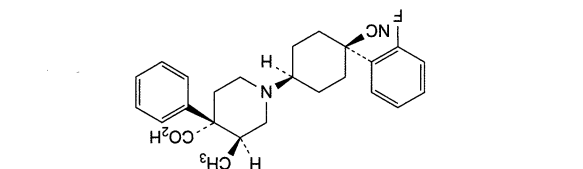
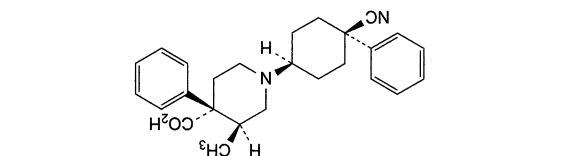
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: D, F,

G, H, I, J, K, L.



CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals, hygroscopic.

Solubility

Freely soluble in water, soluble in warm ethanol

(96 per cent), practically insoluble in acetone.

IDENTIFICATION

First identification A, B.

Second identification A, C.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs, prepared using substance previously dried

in vacuo at 50 °C for 5 h.

Comparison levocarnitine CRS.

C. To 1 mL of solution S (see Tests) add 9 mL of water R,

10 mL of dilute sulfuric acid R and 30 mL of ammonium

reineckate solution R. A pink precipitate is formed. Allow to

stand for 30 min. Filter and wash with water R, with ethanol

(96 per cent) R and then with acetone R and dry at 80 °C.

The precipitate melts (2.2.14) at 147 °C to 150 °C.

TESTS

Solution S

Dissolve 5.00 g in carbon dioxide-free water R prepared from

distilled water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

6.5 to 8.5.

Dilute 10 mL of solution S to 20 mL with carbon dioxide-free

water R.

Specific optical rotation (2.2.7)

–29.0 to –32.0 (anhydrous substance), determined on

solution S at 25 °C.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.10 g of the substance to be examined

in the mobile phase and dilute to 20.0 mL with the mobile

phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 12.5 mg of levocarnitine

impurity A CRS in water R and dilute to 50.0 mL with the

same solvent. Dilute 2.0 mL of this solution to 20.0 mL with

the mobile phase.

Reference solution (c) Dissolve 10.0 mg of levocarnitine

impurity A CRS in water R and dilute to 10.0 mL with the

same solvent. Dilute 2.0 mL of this solution to 20.0 mL with

the mobile phase.

Reference solution (d) Dissolve 0.100 g of levocarnitine CRS in

reference solution (c) and dilute to 10.0 mL with the same

solution.

Column:

size: l = 0.30 m, Ø = 3.9 mm;

stationary phase: aminopropylmethylsilyl silica gel for

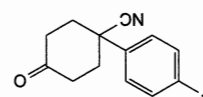
chromatography R (10 µm);

temperature: 30 °C.

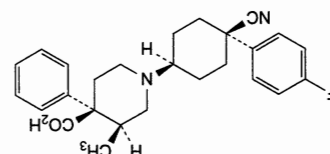
Mobile phase Mix 35 volumes of a 6.81 g/L solution of

potassium dihydrogen phosphate R adjusted to pH 4.7 with

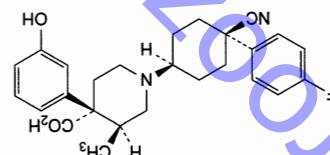
H. 1-(4-fluorophenyl)-4-oxocyclohexanecarbonitrile,



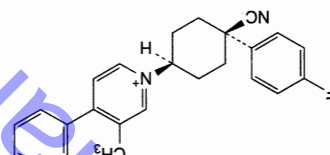
I. (3S,4S)-1-[cis-4-cyano-4-(4-fluorophenyl)cyclohexyl]-3-methyl-4-phenylpiperidine-4-carboxylic acid,



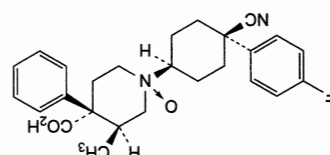
J. (3S,4R)-1-[cis-4-cyano-4-(4-fluorophenyl)cyclohexyl]-4-(3-hydroxyphenyl)-3-methylpiperidine-4-carboxylic acid,



K. 1-[cis-4-cyano-4-(4-fluorophenyl)cyclohexyl]-3-methyl-4-phenylpyridinium,



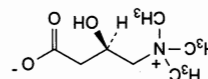
L. (3S,4R)-1-[cis-4-cyano-4-(4-fluorophenyl)cyclohexyl]-3-methyl-4-phenylpiperidine-4-carboxylic acid 1-oxide.



Ph Eur



(Ph. Eur. monograph 1339)



C₇H₁₅NO₃

161.2

541-15-1

Action and use

Carnitine substitute.

Ph Eur

DEFINITION

(3R)-3-Hydroxy-4-(trimethylammonio)butanoate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

dilute sodium hydroxide solution R, and 65 volumes of acetonitrile R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 25 µL of the test solution and reference

solutions (a), (b) and (d).

Retention time Levocarnitine = about 9.6 min;

impurity A = about 10.6 min.

System suitability Reference solution (d):

— resolution: minimum 0.9 between the peaks due to

levocarnitine and impurity A when the chromatogram is

Limits:

recorded over 15 min.

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.5 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Maximum 1.0 per cent, determined on 2.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.125 g in a mixture of 3 volumes of anhydrous formic acid R and 50 volumes of anhydrous acetic acid R.

Add 0.2 mL of crystal violet solution R. Titrate with 0.1 M perchloric acid until the colour changes from violet to green.

1 mL of 0.1 M perchloric acid is equivalent to 16.12 mg of C₇H₁₅NO₃.

STORAGE

In an airtight container.

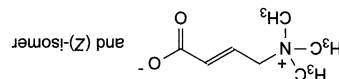
IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use):

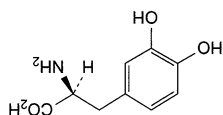
B, C, D.

A. (E)- or (Z)-4-(trimethylammonio)but-2-enoate,



Levodopa

(Ph. Eur. monograph 0038)



C₉H₁₁NO₄

197.2

59-92-7

Action and use

Dopamine precursor; treatment of Parkinson's disease.

Preparations

Co-beneldopa Capsules

Dispersible Co-beneldopa Tablets

Co-careldopa Tablets

Levodopa Capsules

Levodopa Tablets

Prolonged-release Co-beneldopa Capsules

When L-dopa is prescribed or demanded, Levodopa shall be dispensed or supplied.

Ph Eur

DEFINITION

(2S)-2-Amino-3-(3,4-dihydroxyphenyl)propanoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

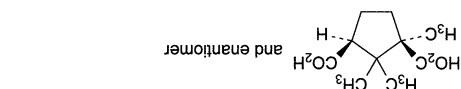
Solubility

Slightly soluble in water, practically insoluble in ethanol (96 per cent). It is freely soluble in 1 M hydrochloric acid and sparingly soluble in 0.1 M hydrochloric acid.

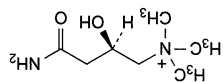
IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

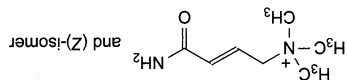
Comparison levodopa CRS.



B. (1R,3S,3SR)-1,2,2-trimethylcyclopentane-1,3-dicarboxylic acid (camphoric acid),



C. (2R)-4-amino-2-hydroxy-N,N,N-trimethyl-4-oxobutan-1-aminium (carnitinaamide),



D. (E)- or (Z)-4-amino-N,N,N-trimethyl-4-oxobut-2-en-1-aminium.

Ph Eur



TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Dissolve 1.0 g in a 103 g/L solution of hydrochloric acid R and dilute to 25 mL with the same solution.

pH (2.2.3)

4.5 to 7.0.

Shake 0.10 g with 10 mL of carbon dioxide-free water R for 15 min.

Related substances

Liquid chromatography (2.2.29). Use freshly prepared solutions.

Solution A 10.3 g/L solution of hydrochloric acid R.

Test solution Dissolve 0.100 g of the substance to be examined in solution A and dilute to 25 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with solution A. Dilute 5.0 mL of this solution to 100.0 mL with solution A.

Reference solution (b) Dissolve 8 mg of tyrosine R (impurity B) and 4 mg of 3-methoxy-L-tyrosine R (L-isomer of impurity C) in 2 mL of the test solution and dilute to 50 mL with solution A. Dilute 5 mL of this solution to 100 mL with

solution A. Dilute 5 mL of this solution to 100 mL with

in 2 mL of the test solution and dilute to 50 mL with

solution A. Dilute 5 mL of this solution to 100 mL with

Column: size: $l = 0.15$ m, $\phi = 3.9$ mm;

stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve separately 200 mg of copper acetate R and 387 mg of N,N-dimethyl-L-phenylalanine R in 250 mL of water R; mix the 2 solutions and adjust immediately to pH 4.0 with acetic acid R; add 50 mL of methanol R and dilute to 1000 mL with water R; mix and filter.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Relative retention With reference to levodopa (retention time = about 7 min): impurity D = about 0.4.

System suitability: reference solution (b):

— resolution: minimum 5 between the peaks due to impurity D and levodopa.

Limit:

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g, heating if necessary, in 5 mL of anhydrous formic acid R. Add 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 19.72 mg of $C_9H_{11}NO_4$.

STORAGE

Protected from light.

LIMITS

— correction factor: for the calculation of content, multiply the peak area of impurity B by 2.2;

— impurity B: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— resolution: minimum 10 between the peaks due to levodopa and impurity B.

System suitability: reference solution (b):

— impurity B = about 2; impurity C = about 3.5.

time = about 6 min; impurity A = about 0.7;

Relative retention With reference to levodopa (retention time = about 6 min): impurity A = about 0.7;

impurities B and C.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to

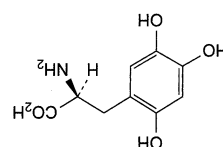
impurities B and C.

Relative retention With reference to levodopa (retention time = about 6 min): impurity A = about 0.7;

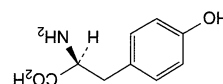
impurity B = about 2; impurity C = about 3.5.

IMPURITIES

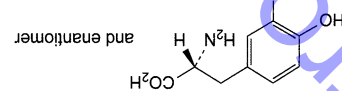
Specified impurities: A, B, C, D.



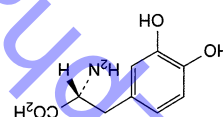
A. (2S)-2-amino-3-(2,4,5-trihydroxyphenyl)propanoic acid,



B. (2S)-2-amino-3-(4-hydroxyphenyl)propanoic acid



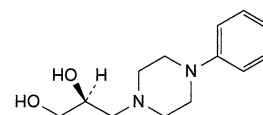
C. (2RS)-2-amino-3-(4-hydroxy-3-methoxyphenyl)propanoic acid (3-methoxy-DL-tyrosine),



D. (2R)-2-amino-3-(3,4-dihydroxyphenyl)propanoic acid (D-dopa).

Levodropropizine

(Ph. Eur. monograph 1535)



$C_{13}H_{20}N_2O_2$

236.3

99291-25-5

Action and use

Cough suppressant.

Ph Eur

DEFINITION

(2S)-3-(4-phenylpiperazin-1-yl)propane-1,2-diol.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, freely soluble in dilute acetic acid and in methanol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Carry out either tests A, B or tests B, C.

Dissolve 1.50 g in a 21 g/L solution of hydrochloric acid R and dilute to 50.0 mL with the same acid.

B. Infrared absorption spectrophotometry (2.2.24).

TESTS

pH (2.2.3)
9.2 to 10.2.

Suspend 2.5 g in carbon dioxide-free water R, heat to dissolve, cool to room temperature and dilute to 100 mL with the same solvent.

Impurity B and related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of levodropropizine impurity B CRS in methanol R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Mix 1.0 mL of the test solution with 1.0 mL of reference solution (a).

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 12 volumes of methanol R and 88 volumes of a 6.81 g/L solution of potassium dihydrogen phosphate R

adjusted to pH 3.0 with phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Run time Twice the retention time of levodropropizine.

Relative retention With reference to levodropropizine

(retention time = about 7 min): impurity B = about 1.2.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to levodropropizine and impurity B.

Limits:

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— unspecified impurities: for each impurity, not more than 0.2 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (a)

(0.10 per cent);
— total: not more than 1.2 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (a) (0.6 per cent);

— disregard limit: 0.1 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurity C

Gas chromatography (2.2.28). Prepare the solutions immediately before use.

Test solution Dissolve 0.50 g of the substance to be examined in methylene chloride R and dilute to 2.5 mL with the same solvent.

Reference solution (a) Dissolve 0.20 g of levodropropizine impurity C CRS in methylene chloride R and dilute to 100.0 mL with the same solvent. Dilute 0.5 mL of this solution to 100.0 mL with methylene chloride R.

Reference solution (b) Dissolve 0.50 g of the substance to be examined in methylene chloride R, add 250 μ L of reference solution (a) and dilute to 2.5 mL with methylene chloride R.

Column:
— material: fused silica;
— size: $l = 30$ m, $\varnothing = 0.53$ mm;
— stationary phase:
poly[(cyanopropyl)(phenyl)[dimethyl]siloxane R (film thickness 3 μ m).
Carrier gas helium for chromatography R.
Flow rate 2.5 mL/min.
Split ratio 1:8.
Temperature:
— column: 140 °C;
— injection port: 170 °C;
— detector: 250 °C.
Detection Flame ionisation.
Injection 1 μ L of the test solution and reference solution (b).
Use an appropriate split-liner, e.g. consisting of a column about 1 cm long packed with glass wool.
Run time 2.5 times the retention time of impurity C.
System suitability: reference solution (b):
— signal-to-noise ratio: minimum 10 for the peak due to impurity C.
At the end of a series of tests, heat the column at 250 °C for 4–6 h.
Limit:
— impurity C: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (5 ppm).
Enantiomeric purity
Liquid chromatography (2.2.29).
Solvent mixture anhydrous ethanol R, hexane R (40:60 V/V).
Test solution Dissolve 10.0 mg of the substance to be examined in 10.0 mL of the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.
Reference solution (a) Dissolve 10 mg of levodropropizine CRS in 10.0 mL of the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.
Reference solution (b) Dissolve 10.0 mg of levodropropizine impurity A CRS in 10.0 mL of the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.
Reference solution (c) Dilute 1.0 mL of reference solution (b) to 50.0 mL with the solvent mixture.
Reference solution (d) Dilute 0.5 mL of reference solution (b) to 25 mL with reference solution (a).
Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: silica gel OD for chiral separations R.
Mobile phase diethylamine R, anhydrous ethanol R, hexane R (0.2:5:95 V/V/V).
Flow rate 0.8 mL/min.
Detection Spectrophotometer at 254 nm.
Injection 20 μ L of the test solution and reference solutions (a), (c) and (d).
Elution order Impurity A, levodropropizine.
System suitability:
— retention times: the retention times of the principal peaks in the chromatograms obtained with the test solution and reference solution (a) are similar;
— resolution: minimum 1.3 between the peaks due to impurity A and levodropropizine in the chromatogram obtained with reference solution (d).

Limit:
— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (2 per cent).
Loss on drying (2.2.32)
Maximum 1.0 per cent, determined on 0.500 g by drying in vacuo at 60 °C over diphosphorus pentoxide R at a pressure of 0.15–0.25 kPa for 4 h.
Sulfated ash (2.4.14)
Maximum 0.2 per cent, determined on 1.0 g.

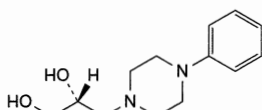
ASSAY
Dissolve 0.100 g in 50 mL of anhydrous acetic acid R. Carry out a potentiometric titration (2.2.20), using 0.1 M perchloric acid. Read the volume added at the 2nd point of inflexion. 1 mL of 0.1 M perchloric acid is equivalent to 11.82 mg of $C_{13}H_{20}N_2O_2$.

STORAGE

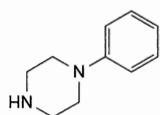
Protected from light.

IMPURITIES

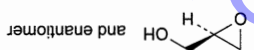
Specified impurities A, B, C



A, (2R,3S)-4-phenylpiperazin-1-ylpropane-1,2-diol (dextrodropropizine),



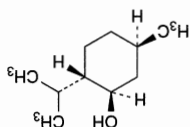
B, 1-phenylpiperazine,



C, [(2R,5S)-oxiran-2-yl]methanol (glycidol), and enantiomer

Levomenthol

(Ph. Eur. monograph 0619)



$C_{10}H_{20}O$

156.3

2216-51-5

Action and use

Decongestant.

Preparations

Levomenthol Cream
Menthol and Benzoin Inhalation

DEFINITION

(1R,2S,5R)-5-Methyl-2-(1-methylethyl)cyclohexanol.

CHARACTERS**Appearance**

Prismatic or acicular, colourless, shiny crystals.

Solubility

Practically insoluble in water, very soluble in ethanol

(96 per cent) and in light petroleum, freely soluble in fatty

oils and in liquid paraffin, very slightly soluble in glycerol.

mp
About 43 °C.

IDENTIFICATION

First identification A, C.

Second identification B, D.

A. Specific optical rotation (see Tests).

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution Dissolve 25 mg of menthol CRS in methanol R and dilute to 5 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase ethyl acetate R, toluene R (5:95 V/V).

Application 2 µL.

Development Over a path of 15 cm.

Drying In air, until the solvents have evaporated.

Detection Spray with anisaldehyde solution R and heat at 100-105 °C for 5-10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Examine the chromatograms obtained in the test for related substances.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in position and approximate dimensions to the principal peak in the chromatogram

obtained with reference solution (c).

D. Dissolve 0.20 g in 0.5 mL of anhydrous pyridine R. Add 3 mL of a 150 g/L solution of diminobenzoyl chloride R in anhydrous pyridine R. Heat on a water-bath for 10 min.

Add 7.0 mL of water R in small quantities with stirring and allow to stand in iced water for 30 min. A precipitate is formed. Allow to stand and decant the supernatant. Wash the precipitate with 2 quantities, each of 5 mL, of iced water R, recrystallise from 10 mL of acetone R, wash with iced acetone R and dry at 75 °C at a pressure not exceeding 2.7 kPa for 30 min. The crystals melt (2.2.14) at 154 °C to 157 °C.

TESTS**Solution S**

Dissolve 2.50 g in 10 mL of ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

Dissolve 1.0 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent. Add 0.1 mL of phenolphthalein solution R. The solution is colourless. Not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to pink.

Specific optical rotation (2.2.7)

—48 to —51, determined on solution S.

Related substances

Gas chromatography (2.2.28).

Test solution (a) Dissolve 0.20 g of the substance to be examined in methylene chloride R and dilute to 50.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with methylene chloride R.

Reference solution (a) Dissolve 40.0 mg of the substance to be examined and 40.0 mg of isomenthol R in methylene chloride R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 0.10 mL of test solution (a) to 100.0 mL with methylene chloride R.

Reference solution (c) Dissolve 40.0 mg of menthol CRS in methylene chloride R and dilute to 100.0 mL with the same solvent.

Column:

— material: glass;

— size: $l = 2.0$ m, $\varnothing = 2$ mm;

— stationary phase: diatomaceous earth for gas chromatography R impregnated with 15 per cent *m/m* of macrogol 1500 R.

Carrier gas nitrogen for chromatography R.

Flow rate 30 mL/min.

Temperature:

— column: 120 °C;

— injection port: 150 °C;

— detector: 200 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time Twice the retention time of menthol.

System suitability:

— resolution: minimum 1.4 between the peaks due to menthol and isomenthol in the chromatogram obtained with reference solution (a);

— signal-to-noise ratio: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits: test solution (a);

— total: not more than 1 per cent of the area of the principal peak;

— disregard limit: 0.05 per cent of the area of the principal peak.

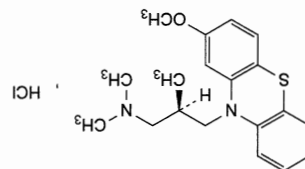
Residue on evaporation

Maximum 0.05 per cent.

Evaporate 2.00 g on a water-bath and heat in an oven at 100-105 °C for 1 h. The residue weighs not more than 1.0 mg.

Levomepromazine Hydrochloride

(Ph. Eur. monograph 0505)



$C_{19}H_{25}ClN_2OS$ 364.9 4185-80-2

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Levomepromazine Injection

DEFINITION

Levomepromazine hydrochloride contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of (2*R*)-3-(2-methoxy-10*H*-phenothiazin-10-yl)-*N,N*,2-trimethylpropan-1-amine hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or very slightly yellow, crystalline powder, slightly hygroscopic, freely soluble in water and in alcohol. It deteriorates when exposed to air and light. It exists in two forms, one melting at about 142 °C and the other at about 162 °C.

IDENTIFICATION

A. Prepare the solution protected from bright light and carry out the measurements immediately. Dissolve 50.0 mg in water *R* and dilute to 500.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with water *R*. Examined between 230 nm and 340 nm (2.2.25), the solution shows two absorption maxima, at 250 nm and 302 nm. The specific absorbance at the maximum at 250 nm is 640 to 700.

B. It complies with the identification test for phenothiazines by thin-layer chromatography (2.3.3): use levomepromazine hydrochloride *CRS* to prepare the reference solution. C. Introduce 0.2 g into a 100 mL separating funnel. Add 5 mL of water *R* and 0.5 mL of strong sodium hydroxide solution *R*. Shake vigorously with two quantities, each of 10 mL, of ether *R*. Combine the ether layers, dry over anhydrous sodium sulfate *R* and evaporate to dryness. Keep the residue at 100 °C for 15 min and allow to crystallise in iced water. Initiate crystallisation if necessary by scratching the wall of the flask with a glass rod. Dry the crystals at 60 °C for 2 h. The crystals melt (2.2.14) at 122 °C to 128 °C. D. It gives reaction (b) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.50 g in carbon dioxide-free water *R* and dilute to 25.0 mL with the same solvent.

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of bromocresol green solution *R*. Not more than 0.5 mL of 0.01 *M* sodium hydroxide or 1.0 mL of 0.01 *M* hydrochloric acid is required to change the colour of the indicator.

DEFINITION

Levomepromazine maleate contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of (2*R*)-3-(2-methoxy-10*H*-phenothiazin-10-

Ph Eur

Levomepromazine Tablets

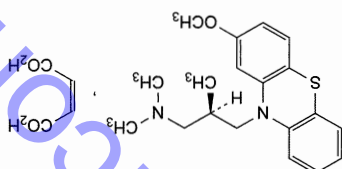
Preparation

Dopamine receptor antagonist; neuroleptic.

Action and use

$C_{23}H_{28}N_2O_5$ 444.6

7104-38-3



(Ph. Eur. monograph 0925)

Levomepromazine Maleate



Ph Eur

STORAGE

Store in an airtight container, protected from light.

$C_{19}H_{25}ClN_2OS$

1 mL of 0.1 *M* sodium hydroxide is equivalent to 36.49 mg of the end-point potentiometrically (2.2.20).

Dissolve 0.300 g in 5 mL of water *R* and add 50 mL of 2-propanol *R*. Titrate with 0.1 *M* sodium hydroxide, determining

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

drying in an oven at 105 °C for 3 h.

Not more than 1.0 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

obtained with the reference solution (0.5 per cent).

is not more intense than the spot in the chromatogram obtained with the test solution, apart from the principal spot, ultraviolet light at 254 nm. Any spot in the chromatogram cyclohexane *R*. Allow the plate to dry in air and examine in acetone *R*, 10 volumes of diethylamine *R* and 80 volumes of over a path of 15 cm using a mixture of 10 volumes of Apply separately to the plate 10 µL of each solution. Develop 95 volumes of methanol *R*.

100 mL with a mixture of 5 volumes of diethylamine *R* and Reference solution Dilute 0.5 mL of the test solution to solvents. Prepare immediately before use.

Test solution Dissolve 0.2 g of the substance to be examined in a mixture of 5 volumes of diethylamine *R* and 95 volumes of methanol *R* and dilute to 10 mL with the same mixture of

Carry out the test protected from bright light. Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ *R* as the coating substance.

Related substances

with reference to the dried substance.

Specific optical rotation (2.2.7) + 9.5 to + 11.5, determined on solution S and calculated

CHARACTERS

yl)-N,N,2-trimethylpropan-1-amine (Z)-butenedioate, calculated with reference to the dried substance.

A white or slightly yellowish, crystalline powder, slightly soluble in water, sparingly soluble in methylene chloride, slightly soluble in alcohol. It deteriorates when exposed to air and light.

It melts at about 186 °C, with decomposition.

IDENTIFICATION

First identification A, B.
Second identification A, C, D.

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with levomepromazine maleate CRS. Examine the substances prepared as discs.

C. Identification test for phenothiazines by thin-layer chromatography (2.3.3): use levomepromazine maleate CRS to prepare the reference solution.

D. Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.20 g of the substance to be examined in a mixture of 10 volumes of water R and 90 volumes of acetone R and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 50 mg of maleic acid CRS in a mixture of 10 volumes of water R and 90 volumes of acetone R and dilute to 10 mL with the same mixture of solvents.

Apply separately to the plate as bands 10 mm by 2 mm 5 µL of each solution. Develop over a path of 12 cm using a mixture of 3 volumes of water R, 7 volumes of anhydrous formic acid R and 90 volumes of di-isopropyl ether R. Dry the plate at 120 °C for 10 min and examine in ultraviolet light at 254 nm. The chromatogram obtained with the test solution shows a zone at the point of application and another zone similar in position and size to the principal zone in the chromatogram obtained with the reference solution.

TESTS

pH (2.2.3)

Carry out the test protected from bright light. Introduce 0.50 g into a conical flask and add 25.0 mL of carbon dioxide-free water R. Shake and allow the solids to settle. The pH of the supernatant solution is 3.5 to 5.5.

Specific optical rotation (2.2.7)

Dissolve 1.25 g in dimethylformamide R and dilute to 25.0 mL with the same solvent. The specific optical rotation is –7.0 to –8.5, calculated with reference to the dried substance.

Related substances

Carry out the test protected from bright light and prepare the solutions immediately before use. Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.20 g of the substance to be examined in a mixture of 10 volumes of water R and 90 volumes of acetone R and dilute to 10 mL with the same mixture of solvents.

Reference solution Dilute 0.5 mL of the test solution to 100 mL with a mixture of 10 volumes of water R and 90 volumes of acetone R.

DEFINITION

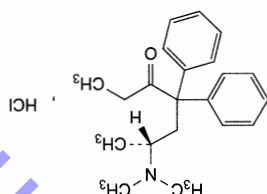
(6R)-6-(Dimethylamino)-4,4-diphenylheptan-3-one hydrochloride.

Action and use
Opioid analgesic.

C₂₁H₂₈N₂O

345.9

5967-73-7



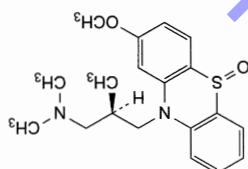
(Ph. Eur. monograph 1787)

Levomethadone Hydrochloride

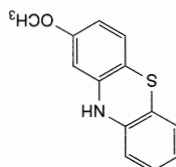
Ph Eur

10H-phenothiazine 5-oxide.

B. 10-[(2R)-3-(dimethylamino)-2-methylpropyl]-2-methoxy-



A. 2-methoxyphenothiazine,

**IMPURITIES**

Store protected from light.

STORAGE

C₂₃H₂₈N₂O₅S.

1 mL of 0.1 M perchloric acid is equivalent to 44.46 mg of

potentiometrically (2.2.20).

with 0.1 M perchloric acid, determining the end-point

Dissolve 0.350 g in 50 mL of anhydrous acetic acid R. Titrate

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

drying in an oven at 105 °C for 3 h.

Not more than 0.5 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

obtained with the reference solution (0.5 per cent).

is not more intense than the spot in the chromatogram obtained with the test solution, apart from the principal spot, ultraviolet light at 254 nm. Any spot in the chromatogram cyclohexane R. Allow the plate to dry in air and examine in acetone R, 10 volumes of diethylamine R and 80 volumes of over a path of 15 cm using a mixture of 10 volumes of Apply separately to the plate 10 µL of each solution. Develop

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, C, D

Second identification A, B, D

A. Specific optical rotation (see Tests).

B. Melting point (2.2.14): 239 °C to 242 °C.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of methadone hydrochloride.

D. Dilute 1 mL of solution S (see Tests) to 5 mL with water R and add 1 mL of dilute ammonia R1. Mix, allow to stand for 5 min and filter. The filtrate gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

Dilute 10 mL of solution S to 25 mL with carbon dioxide-free water R. To 10 mL of the solution add 0.2 mL of methyl red solution R and 0.2 mL of 0.01 M sodium hydroxide.

The solution is yellow. Add 0.4 mL of 0.01 M hydrochloric acid. The solution is red.

Specific optical rotation (2.2.7)

–125 to –135 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of the

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 12.0 mg of imipramine hydrochloride CRS in the mobile phase and dilute to 10 mL with the mobile phase. To 1 mL of the solution add 5 mL of the test solution and dilute to 10 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\phi = 4.6$ mm;

(5 μ m);

— temperature: 25 °C.

Mobile phase Mix 35 volumes of acetonitrile R and 65 volumes of an 11.5 g/L solution of phosphoric acid R adjusted to pH 3.6 with tetraethylammonium hydroxide solution R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Equilibration About 30 min.

Injection 10 μ L.

Run time 7 times the retention time of levomethadone.

Retention time Levomethadone = about 5 min.

IMPURITIES

Protected from light.

STORAGE

C₂₁H₂₈ClNO.

1 mL of 0.1 M silver nitrate is equivalent to 34.59 mg of electrode.

Dissolve 0.300 g in a mixture of 40 mL of water R and 5 mL of acetic acid R. Titrate with 0.1 M silver nitrate. Determine the end-point potentiometrically (2.2.20), using a silver

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

solution (0.5 per cent).

— dextromethadone: not more than the area of the principal peak in the chromatogram obtained with the reference

Limit:

levomethadone.

— tailing factor: maximum 3 for the peak due to

the peak due to levomethadone;

— number of theoretical plates: minimum 2000, calculated for

System suitability Test solution:

dextromethadone = about 1.4.

Relative retention With reference to levomethadone:

Injection 10 μ L.

Equilibration About 30 min.

Detection Spectrophotometer at 210 nm.

Flow rate 0.7 mL/min.

phosphate R.

and 85 volumes of a 13.6 g/L solution of potassium dihydrogen

pH 4.0 with phosphoric acid R, 15 volumes of acetonitrile R

Mobile phase Mix 1 volume of methyamine R adjusted to

— temperature: 10 °C.

chromatography R (5 μ m);

— stationary phase: 2-hydroxypropylbetadex for

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

Column:

solution to 20.0 mL with the mobile phase.

Reference solution Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this

the mobile phase.

examined in the mobile phase and dilute to 100.0 mL with

Test solution Dissolve 40.0 mg of the substance to be

Liquid chromatography (2.2.29).

Dextromethadone

(0.05 per cent).

— the chromatogram obtained with reference solution (a)

— disregard limit: 0.25 times the area of the principal peak in

solution (a) (0.5 per cent);

peak in the chromatogram obtained with reference

— total: not more than 2.5 times the area of the principal

reference solution (a) (0.1 per cent);

principal peak in the chromatogram obtained with

— any impurity: not more than 0.5 times the area of the

Limits:

imipramine and levomethadone.

— resolution: minimum 2.5 between the peaks due to

System suitability: reference solution (b):

Ph Eur

DEFINITION

13-Ethyl-17-hydroxy-18,19-dinor-17 α -pregn-4-en-20-yn-3-one.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison levonorgestrel CRS.

TESTS**Specific optical rotation (2.2.7)**

–35 to –30.

Dissolve 0.200 g in methylene chloride R and dilute to 20.0 mL with the same solvent.

Related substances

A. Impurities A, B, H, K, M, O, S, U. Liquid

chromatography (2.2.29).

Solvent mixture water for chromatography R, acetone R1 (30:70 V/V).

Test solution Dissolve 10.0 mg of the substance to be

examined in 7 mL of acetone R1 using sonication and

dilute to 10.0 mL with water for chromatography R.

Reference solution (a) Dissolve 5 mg of levonorgestrel for system suitability 1 CRS (containing impurities A, H, K, M, O and S) in 3.5 mL of acetone R1 using sonication and

dilute to 5.0 mL with water R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of levonorgestrel

impurity B CRS in 35 mL of acetone R1 and dilute to

50.0 mL with water for chromatography R. Dilute 1.0 mL of

the solution to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5.0 mg of norethisterone CRS

(impurity U) in 35 mL of acetone R1 and dilute to 50.0 mL with water for chromatography R. Dilute 1.0 mL of

the solution to 100.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octylsilyl silica gel for

chromatography with polar incorporated groups R (5 μ m);

— temperature: 30 °C.

Mobile phase:

— mobile phase A: acetone R1, water for

chromatography R (40:60 V/V);

— mobile phase B: acetone R1;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 50	100 \rightarrow 20	0 \rightarrow 80

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 215 nm and, for impurity O, at 200 nm.

C₂₁H₂₈O₂

312.5

797-63-7

Action and use

Progestogen.

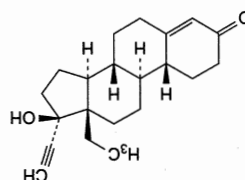
Preparations

Levonorgestrel Tablets

Levonorgestrel and Ethinylestradiol Tablets

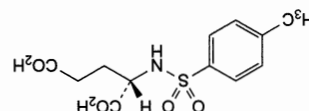
Levonorgestrel

(Ph. Eur. monograph 0926)

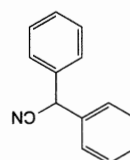


Ph Eur

F. (2S)-2-[[[(4-methylphenyl)sulfonyl]amino]pentanedioic acid (N-p-tosyl-L-glutamic acid).



E. diphenylacetone R1,

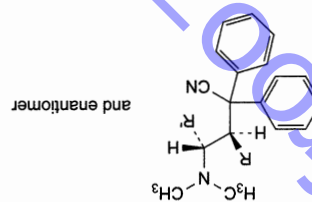


2,2-diphenylbutanenitrile,

C. R = CH₃, R' = H: (3RS)-4-(dimethylamino)-3-methyl-

diphenylpentanenitrile,

B. R = H, R' = CH₃: (4RS)-4-(dimethylamino)-2,2-



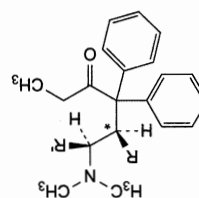
and enantiomer

4,4-diphenylhexan-3-one,

D. R = CH₃, R' = H: (5RS)-6-(dimethylamino)-5-methyl-

diphenylheptan-3-one,

A. R = H, R' = CH₃: (6S)-6-(dimethylamino)-4,4-



and epimer at C*

Injection 50 µL.

Identification of impurities Use the chromatograms supplied

chromatograms obtained with reference solution (a) at

215 nm to identify the peaks due to impurities A, H, K, M

and S, and at 200 nm to identify the peak due to

impurity O; use the chromatogram obtained with reference

solution (c) to identify the peak due to impurity B; use the

chromatogram obtained with reference solution (d) to

identify the peak due to impurity U.

Relative retention With reference to levonorgestrel (retention

time = about 20 min): impurity H = about 0.5;

impurity U = about 0.8; impurity K = about 0.85;

impurity A = about 0.91; impurity M = about 0.95;

impurity O = about 1.16; impurity B = about 1.26;

impurity S = about 1.9.

System suitability:

— signal-to-noise ratio: minimum 60 for the principal peak

in the chromatogram obtained with reference

solution (b);

— peak-to-valley ratio: minimum 3.0, where H_p = height

above the baseline of the peak due to impurity M and

H_p = height above the peak from the lowest point of

the curve separating this peak from the peak due to

impurity A in the chromatogram obtained with

reference solution (a).

Calculation of percentage contents:

— correction factors: multiply the peak areas of the

following impurities by the corresponding correction

factor: impurity A = 0.4; impurity M = 3.1;

impurity O = 2.6;

— for impurity B, use the concentration of impurity B in

reference solution (c);

— for impurity U, use the concentration of impurity U in

reference solution (d);

— for impurities other than B and U, use the

concentration of levonorgestrel in reference

solution (b).

Limits:

— impurities A, B, K: for each impurity, maximum

0.3 per cent;

— impurity O at 200 nm: maximum 0.3 per cent;

— impurities M, S, U: for each impurity, maximum

0.2 per cent;

— impurity H: maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— sum of impurities other than O: maximum 1.0 per cent;

— reporting threshold: 0.05 per cent.

B. Impurities V and W. Liquid chromatography (2.2.29).

Solvent mixture water for chromatography R, acetonitrile R1

(30:70 V/V).

Test solution Dissolve 10.0 mg of the substance to be

examined in 7 mL of acetonitrile R1 using sonication and

dilute to 10.0 mL with water for chromatography R.

Reference solution (a) Dissolve 5 mg of levonorgestrel for system

suitability 2 CRS (containing impurities V and W) in 3.5 mL

of acetonitrile R1 using sonication and dilute to 5.0 mL with

water for chromatography R.

Reference solution (b) Dissolve 5.0 mg of ethinylestradiol CRS

in 35 mL of acetonitrile R1 using sonication and dilute to

50.0 mL with water for chromatography R. Dilute 3.0 mL of

the solution to 100.0 mL with the solvent mixture.

Column: — size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (3 µm).

Mobile phase: — mobile phase A: acetonitrile R1, water for

chromatography R (40:60 V/V);

— mobile phase B: water for chromatography R,

acetonitrile R1 (10:90 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 1	92	8
1 - 3	92 → 82	8 → 18
3 - 6	82	18
6 - 16	82 → 60	18 → 40
16 - 21	60 → 0	40 → 100
21 - 32	0	100

Flow rate 1 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 50 µL.

Identification of impurities Use the chromatogram supplied

with levonorgestrel for system suitability 2 CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities V and W.

Relative retention With reference to levonorgestrel (retention

time = about 12 min): impurity W = about 0.9;

impurity V = about 1.9.

System suitability: reference solution (a):

— resolution: minimum 2.8 between the peaks due to

impurity W and levonorgestrel.

Calculation of percentage contents:

— for each impurity, use the concentration of

ethinylestradiol in reference solution (b).

Limits:

— impurity W: maximum 0.3 per cent;

— impurity V: maximum 0.15 per cent.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 45 mL of tetrahydrofuran R. Add 10 mL

of a 100 g/L solution of silver nitrate R. After 1 min, titrate

with 0.1 M sodium hydroxide, determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M sodium hydroxide is equivalent to 31.25 mg of

C₂₁H₂₈O₂.

STORAGE

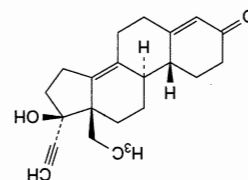
Protected from light.

IMPURITIES

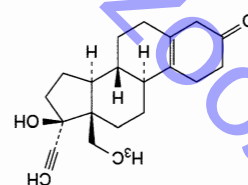
Specified impurities A, B, H, K, M, O, S, U, V, W

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

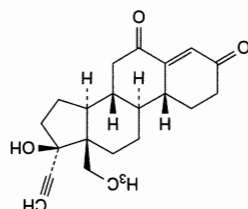
impurities for demonstration of compliance. See also 5.10.
Control of impurities in substances for pharmaceutical use): C, D,
G, I, J, L, N, P, Q, R, T.



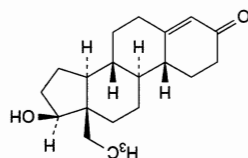
A. 13-ethyl-17-hydroxy-18,19-dinor-17 α -pregna-4,8(14)-dien-20-yn-3-one,



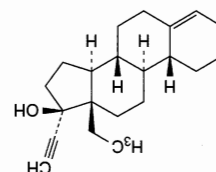
B. 13-ethyl-17-hydroxy-18,19-dinor-17 α -pregna-5(10)-en-20-yn-3-one,



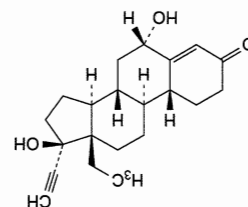
J. 13-ethyl-17-hydroxy-18,19-dinor-17 α -pregna-4-en-20-yn-3,6-dione (6-oxolevonorgestrel),



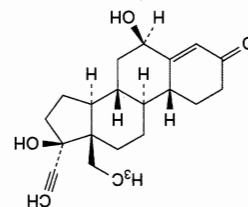
K. 13-ethyl-17 β -hydroxygon-4-en-3-one (18-methylnandrolone),



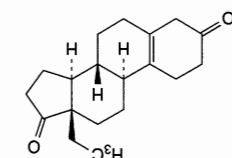
D. 13-ethyl-18,19-dinor-17 α -pregna-4-en-20-yn-17-ol (3-deoxolevonorgestrel),



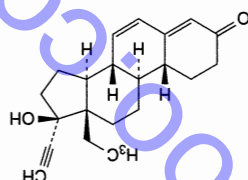
G. 13-ethyl-6 α ,17-dihydroxy-18,19-dinor-17 α -pregna-4-en-20-yn-3-one (6 α -hydroxy[levonorgestrel]),



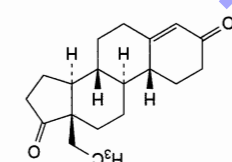
H. 13-ethyl-6 β ,17-dihydroxy-18,19-dinor-17 α -pregna-4-en-20-yn-3-one (6 β -hydroxy[levonorgestrel]),



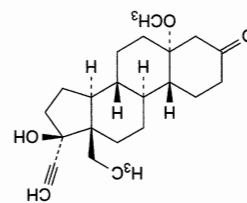
N. 13-ethylgon-5(10)-ene-3,17-dione (Δ 5(10)-levodione),



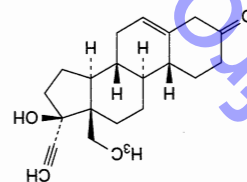
M. 13-ethyl-17-hydroxy-18,19-dinor-17 α -pregna-4,6-dien-20-yn-3-one (Δ 6-levonorgestrel),



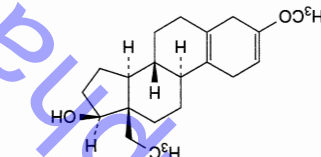
L. 13-ethylgon-4-ene-3,17-dione (levodione),



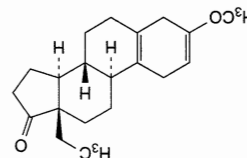
O. 13-ethyl-17-hydroxy-5α-methoxy-18,19-dimor-17α-pregn-20-yn-3-one (4,5-dihydro-5α-methoxy-levonorgestrel),



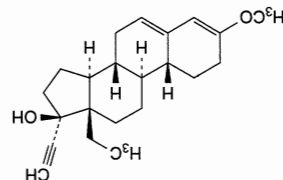
P. 13-ethyl-17-hydroxy-18,19-dimor-17α-pregn-5-en-20-yn-3-one (Δ5-levonorgestrel),



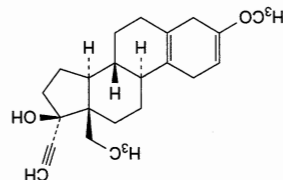
Q. 13-ethyl-3-methoxygon-2,5(10)-dien-17β-ol,



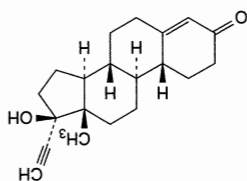
R. 13-ethyl-3-methoxygon-2,5(10)-dien-17-one,



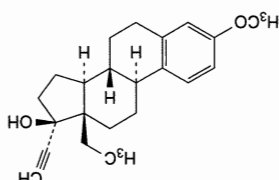
S. 13-ethyl-3-methoxy-18,19-dimor-17α-pregn-3,5-dien-20-yn-17-ol,



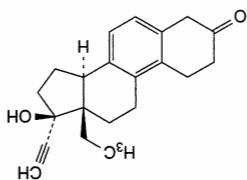
T. 13-ethyl-3-methoxy-18,19-dimor-17α-pregn-2,5(10)-dien-20-yn-17-ol,



U. 17-hydroxy-19-nor-17α-pregn-4-en-20-yn-3-one (norethisterone),



V. 13-ethyl-3-methoxy-18,19-dimor-17α-pregn-1,3,5(10)-trien-20-yn-17-ol,



W. 13-ethyl-17-hydroxy-18,19-dimor-17α-pregn-5,7,9-trien-20-yn-3-one.

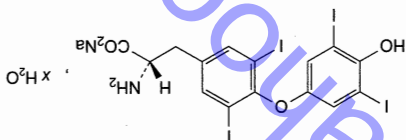
Levothyroxine Sodium

(Ph Eur Monograph 0401)



Ph Eur

$C_{15}H_{10}I_4NaO_4 \cdot xH_2O$ ($x \approx 5$) 799 25416-65-3



(anhydrous substance)

Action and use

Thyroid hormone replacement.

Preparations

Levothyroxine Oral Solution

Levothyroxine Tablets

Ph Eur

DEFINITION

Sodium (2S)-2-amino-3-[4-(4-hydroxy-3,5-diiodophenoxy)-3,5-diiodophenyl]propanoate.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

It contains a variable quantity of water.

CHARACTERS

Appearance

Almost white or slightly brownish-yellow, fine, slightly hygroscopic, crystalline powder.

Solubility

Very slightly soluble in water, slightly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison levothyroxine sodium CRS.

B. To 200 mg add 2 mL of dilute sulfuric acid R. Heat on a water-bath and then carefully over a naked flame, increasing the temperature gradually up to $600 \pm 50^\circ\text{C}$. Continue the ignition until most of the black particles have disappeared.

Dissolve the residue in 2 mL of water R. The solution gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 0.500 g in 23 mL of a gently boiling mixture of 1 volume of 1 M hydrochloric acid and 4 volumes of ethanol (96 per cent) R. Cool and dilute to 25.0 mL with the same mixture of solvents.

Appearance of solution

Freshly prepared solution S is not more intensely coloured than reference solution BY₃ (2.2.2, Method II).

Specific optical rotation (2.2.7)

+16 to +20 (anhydrous substance), determined on freshly prepared solution S.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture Mobile phase A, ethanol (96 per cent) R (1:2 V/V).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 10.0 mL of the solution to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2.5 mg of levothyroxine sodium CRS and 2.5 mg of isotyrosine sodium CRS (impurity A) in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 25.0 mg of levothyroxine sodium CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 10.0 mL of the solution to 25.0 mL with the solvent mixture.

Reference solution (d) Dissolve 2.0 mg of levothyroxine for peak identification CRS (containing impurities F and G) in 10.0 mL of the solvent mixture and sonicate for 10 min.

Column:

— size: $l = 0.15\text{ m}$, $\varnothing = 4.0\text{ mm}$;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μm).

Mobile phase:

— mobile phase A: dissolve 1.97 g of phosphoric acid R in water R and dilute to 2 L with the same solvent;

— mobile phase B: dissolve 1.97 g of phosphoric acid R in acetonitrile R1 and dilute to 2 L with the same solvent;

IMPURITIES

Specified impurities A, F, G

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of

STORAGE

In an airtight container, protected from light, at a temperature of 2°C to 8°C .

sodium CRS.

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification. Injection Test solution and reference solution (c). Calculate the percentage content of $\text{C}_{15}\text{H}_{10}\text{I}_4\text{NNaO}_4$ taking into account the assigned content of levothyroxine sodium CRS.

ASSAY

6.0 per cent to 12.0 per cent, determined on 0.100 g.

Water (2.5.32)

(Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

The thresholds indicated under Related substances

reference solution (b) (0.05 per cent).

levothyroxine in the chromatogram obtained with

— disregard limit: 0.5 times the area of the peak due to

total: maximum 2.0 per cent;

(0.2 per cent);

chromatogram obtained with reference solution (b)

twice the area of the peak due to levothyroxine in the

— unspecified impurities: for each impurity, not more than

reference solution (b) (0.3 per cent);

due to levothyroxine in the chromatogram obtained with

— impurity G: not more than 3 times the area of the peak

reference solution (b) (0.5 per cent);

due to levothyroxine in the chromatogram obtained with

— impurity F: not more than 5 times the area of the peak

solution (a) (1.0 per cent);

peak in the chromatogram obtained with reference

— impurity A: not more than the area of the corresponding

Limits:

impurity A and levothyroxine.

— resolution: minimum 5.0 between the peaks due to

System suitability: reference solution (a):

impurity F = about 2.0; impurity G = about 2.4.

time = about 11 min; impurity A = about 0.5;

Relative retention With reference to levothyroxine (retention

identify the peaks due to impurities F and G.

chromatogram obtained with reference solution (d) to

with levothyroxine for peak identification CRS and the

Identification of impurities Use the chromatogram supplied

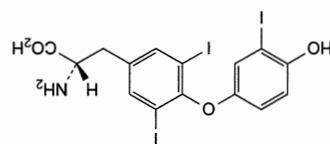
solutions (a), (b) and (d).

Injection 25 μL of the test solution and reference

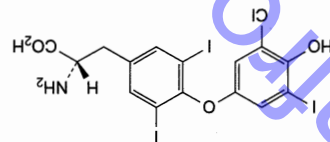
Flow rate 1 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	70	30
10 - 40	70 \rightarrow 20	30 \rightarrow 80
40 - 50	20	80

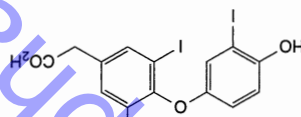
impurities in substances for pharmaceutical use): B, C, D, E, H, I, J, K



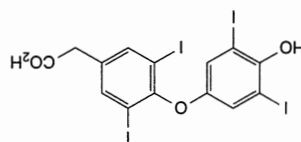
A. (2S)-2-amino-3-[4-(4-hydroxy-3-iodophenoxy)-3,5-diiodophenyl]propanoic acid (lithothionine),



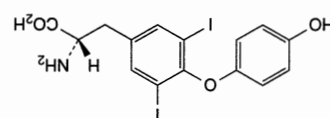
B. (2S)-2-amino-3-[4-(3-chloro-4-hydroxy-5-iodophenoxy)-3,5-diiodophenyl]propanoic acid,



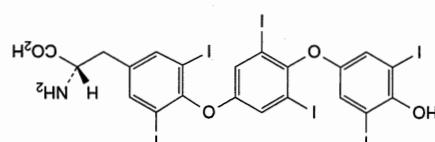
C. [4-(4-hydroxy-3-iodophenoxy)-3,5-diiodophenyl]acetic acid (triiodothyroacetic acid),



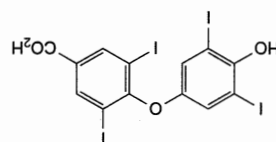
D. [4-(4-hydroxy-3,5-diiodophenoxy)-3,5-diiodophenyl]acetic acid (tetraiodothyroacetic acid),



E. (2S)-2-amino-3-[4-(4-hydroxyphenoxy)-3,5-diiodophenyl]propanoic acid (diiodothyronine),



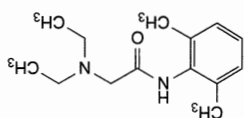
F. (2S)-2-amino-3-[4-[4-(4-hydroxy-3,5-diiodophenoxy)-3,5-diiodophenoxy]]propanoic acid,



H. 4-(4-hydroxy-3,5-diiodophenoxy)-3,5-diiodobenzoic acid,

Lidocaine

(Ph Eur Monograph 0727)



C₁₄H₂₂N₂O

234.3

137-58-6

Action and use

Local anaesthetic; Class I antiarrhythmic.

Preparation

Lidocaine Ointment

Ph Eur

DEFINITION

2-(Diethylamino)-N-(2,6-dimethylphenyl)acetamide.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, very soluble in ethanol

(96 per cent) and in methylene chloride.

IDENTIFICATION

First identification A.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison lidocaine CRS.

B. Melting point (2.2.14): 66 °C to 70 °C, determined

without previous drying.

C. To about 5 mg add 0.5 mL of fuming nitric acid R.

Evaporate to dryness on a water-bath, cool, and dissolve the

TESTS

Related substances

residue in 5 mL of acetone R. Add 0.2 mL of alcoholic potassium hydroxide solution R. A green colour develops.

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with

the mobile phase.

Reference solution (a) Dissolve 50.0 mg of 2,6-

dimethylaniline R (impurity A) in the mobile phase and dilute

to 100.0 mL with the mobile phase. Dilute 10.0 mL of this

solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of 2-chloro-N-(2,6-

dimethylphenyl)acetamide R (impurity H) in the mobile phase

and dilute to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to

10.0 mL with the mobile phase.

Reference solution (d) Mix 1.0 mL of reference solution (a),

1.0 mL of reference solution (b) and 1.0 mL of reference

solution (c), then dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;

— stationary phase: end-capped polar-embedded octadecylsilyl

amorphous organosilica polymer R (5 μ m);

— temperature: 30 °C.

Mobile phase Mix 30 volumes of acetonitrile for

chromatography R and 70 volumes of a 4.85 g/L solution of

potassium dihydrogen phosphate R previously adjusted to

pH 8.0 with strong sodium hydroxide solution R.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time 3.5 times the retention time of lidocaine.

Relative retention With reference to lidocaine (retention

time = about 17 min): impurity H = about 0.37;

impurity A = about 0.40.

System suitability Reference solution (d):

— resolution: minimum 1.5 between the peaks due to

impurities H and A.

Limits:

— impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (d) (0.01 per cent);

— unspecified impurities: for each impurity, not more than the

area of the peak due to lidocaine in the chromatogram

obtained with reference solution (d) (0.10 per cent);

— total: not more than 5 times the area of the peak due to

lidocaine in the chromatogram obtained with reference

solution (d) (0.05 per cent).

Chlorides (2.4.4)

Maximum 35 ppm.

Dissolve 1.4 g in a mixture of 3 mL of dilute nitric acid R and

12 mL of water R.

Sulfates (2.4.13)

Maximum 0.1 per cent.

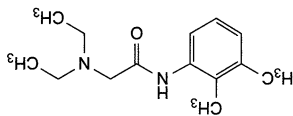
Dissolve 0.2 g in 5 mL of ethanol (96 per cent) R and dilute

to 20 mL with distilled water R.

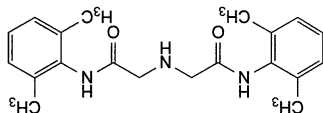
Water (2.5.12)

Maximum 1.0 per cent, determined on 1.00 g.

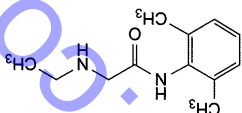
F. 2-(diethylamino)-N-(2,3-dimethylphenyl)acetamide;



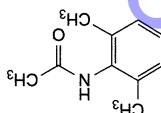
E. 2,2'-iminobis(N-(2,6-dimethylphenyl)acetamide),



D. N-(2,6-dimethylphenyl)-2-(ethylamino)acetamide,

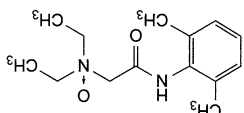


C. N-(2,6-dimethylphenyl)acetamide,

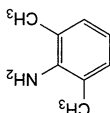


B. 2-(diethylaziridinyl)-N-(2,6-dimethylphenyl)acetamide

(lidocaine N-oxide),



A. 2,6-dimethylaniline,



D, E, F, G, H, I, J.

Control of impurities in substances for pharmaceutical use: B, C, impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph Substances for pharmaceutical use

acceptance criterion for other/unspecified impurities and/or

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities A.

IMPURITIES

of $C_{14}H_{22}N_2O$.

1 mL of 0.1 M perchloric acid is equivalent to 23.43 mg

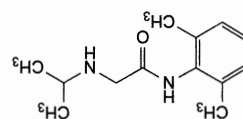
acid, determining the end-point potentiometrically (2.2.20).

To 0.200 g add 50 mL of anhydrous acetic acid R and stir

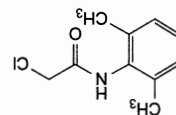
ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

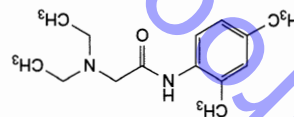
Sulfated ash (2.4.14)



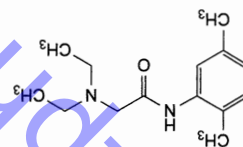
G. N-(2,6-dimethylphenyl)-2-((1-methylethyl)amino)acetamide,



H. 2-chloro-N-(2,6-dimethylphenyl)acetamide,



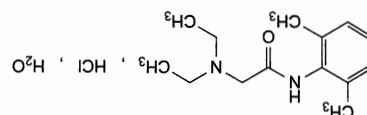
I. 2-(diethylamino)-N-(2,4-dimethylphenyl)acetamide,



J. 2-(diethylamino)-N-(2,5-dimethylphenyl)acetamide,

Lidocaine Hydrochloride

(Ph. Eur. monograph 0227)



$C_{14}H_{23}ClN_2O_2 \cdot H_2O$

288.8

6108-05-0



Solubility
Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B, D
Second identification A, C, D
A. Melting point (2.2.14): 74 °C to 79 °C, determined without previous drying.

B. Infrared absorption spectrophotometry (2.2.24).

C. To about 5 mg add 0.5 mL of fuming nitric acid R.

Evaporate to dryness on a water-bath, cool and dissolve the residue in 5 mL of acetone R. Add 0.2 mL of alcoholic potassium hydroxide solution R. A green colour is produced.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S
Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.0 to 5.5.

Dilute 1 mL of solution S to 10 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of 2,6-dimethylamine R (impurity A) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of 2-chloro-N-(2,6-dimethylphenyl)acetamide R (impurity H) in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase.

Reference solution (d) Mix 1.0 mL of reference solution (a), 1.0 mL of reference solution (b) and 1.0 mL of reference solution (c) and dilute to 100.0 mL with the mobile phase.

Column: stationary phase: end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R (5 µm);

— size: l = 0.15 m, Ø = 3.9 mm;

— temperature: 30 °C.

Mobile phase Mix 30 volumes of acetonitrile for chromatography R and 70 volumes of a 4.85 g/L solution of potassium dihydrogen phosphate R adjusted to pH 8.0 with strong sodium hydroxide solution R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Run time 3.5 times the retention time of lidocaine.

Relative retention With reference to lidocaine (retention time = about 17 min): impurity H = about 0.37;

impurity A = about 0.40.

System suitability Reference solution (d):

— resolution: minimum 1.5 between the peaks due to impurities H and A.

DEFINITION

2-(Diethylamino)-N-(2,6-dimethylphenyl)acetamide hydrochloride monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Limits:

— **impurity A**: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (d) (0.01 per cent);

— **unspecified impurities**: for each impurity, not more than area of the peak due to lidocaine in the chromatogram obtained with reference solution (d) (0.10 per cent);

— **total**: not more than 5 times the area of the peak due to lidocaine in the chromatogram obtained with reference solution (d) (0.5 per cent);

— **disregard limit**: 0.5 times the area of the peak due to lidocaine in the chromatogram obtained with reference solution (d) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 5 ppm.

Dissolve 1.0 g in water R and dilute to 25 mL with the same solvent. Carry out the prefiltration. 10 mL of the prefiltrate complies with test E. Prepare the reference solution using

2 mL of lead standard solution (1 ppm Pb) R.

Water (2.5.12)

5.5 per cent to 7.0 per cent, determined on 0.25 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.220 g in 50 mL of ethanol (96 per cent) R and add 5.0 mL of 0.01 M hydrochloric acid. Carry out a

potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 27.08 mg

STORAGE

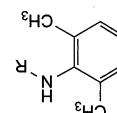
Protected from light.

IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E, F, G, H, I, J, K.



A, R = H: 2,6-dimethylaniline,

C, R = CO-CH₃; N-(2,6-dimethylphenyl)acetamide,

D, R = CO-CH₂-NH-C₂H₅; N-(2,6-dimethylphenyl)-2-

(ethylamino)acetamide,

G, R = CO-CH₂-NH-CH(CH₃)₂; N-(2,6-dimethylphenyl)-2-

2-[(1-methylethyl)amino]acetamide,

H, R = CO-CH₂-Cl; 2-chloro-N-(2,6-

dimethylphenyl)acetamide,

K, R = CO-CH₂-N(CH₃)C₂H₅; N-(2,6-dimethylphenyl)-2-

(ethylmethylanilino)acetamide,

Content
— sum of the contents of lincomycin hydrochloride and lincomycin B hydrochloride: 96.0 per cent to 102.0 per cent (anhydrous substance);

Mixture of antibiotics produced by *Streptomyces lincolnensis* var. *lincolnensis* or obtained by any other means, the main component being methyl 6,8-dideoxy-6-[[[(2S,4R)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-erythro-α-D-galacto-octopyranoside (lincomycin) hydrochloride monohydrate.

DEFINITION

Lincomycin Injection

Lincomycin Capsules

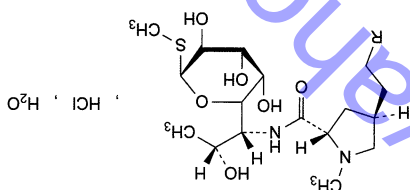
Preparations

Antibacterial.

Action and use

7179-49-9

Compound	R	Molecular formula	M _r
Lincomycin	CH ₃	C ₁₈ H ₃₅ ClN ₂ O ₈ ·H ₂ O	461.0
Lincomycin B	H	C ₁₇ H ₃₃ ClN ₂ O ₈ ·H ₂ O	447.0

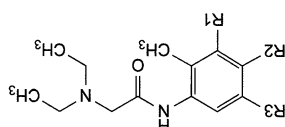


(Ph. Eur. monograph 0583)

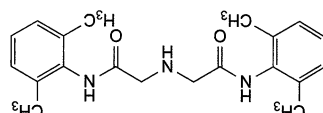
Lincomycin Hydrochloride

Ph Eur

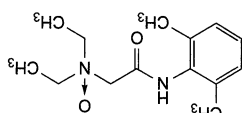
F, R₁ = CH₃, R₂ = R₃ = H: 2-(diethylamino)-N-(2,3-dimethylphenyl)acetamide,
I, R₁ = R₃ = H, R₂ = CH₃: 2-(diethylamino)-N-(2,4-dimethylphenyl)acetamide,
J, R₁ = R₂ = H, R₃ = CH₃: 2-(diethylamino)-N-(2,5-dimethylphenyl)acetamide.



E, 2,2'-(azanediy)bis[N-(2,6-dimethylphenyl)acetamide],



B, 2-(diethylazirino)-N-(2,6-dimethylphenyl)acetamide (lidocaine N²-oxide),



— lincomycin B hydrochloride: maximum 5.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, slightly soluble in ethanol (96 per cent), very slightly soluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison lincomycin hydrochloride CRS.

B. Dissolve 0.1 g in water R and dilute to 10 mL with the same solvent. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

pH (2.2.3)

3.5 to 5.5 for solution S.

Specific optical rotation (2.2.7)

+135 to +150 (anhydrous substance).

Dissolve 1.000 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of lincomycin hydrochloride CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of lincomycin hydrochloride CRS (containing impurities A, B and C) in 2 mL of the mobile phase.

Reference solution (c) Dilute 2.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped base-deactivated octylsilyl silica gel for chromatography R (5 μ m);

— temperature: 50 °C.

Buffer solution pH 6.1 Dissolve 34 g of phosphoric acid R in 900 mL of water for chromatography R, adjust to pH 6.1 with concentrated ammonia R and dilute to 1000 mL with water for chromatography R.

Mobile phase methanol R, acetonitrile R1, buffer solution pH 6.1 (8:17:75 V/V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L of the test solution and reference solutions (b), (c) and (d).

Run time 5.5 times the retention time of lincomycin.

Relative retention With reference to lincomycin (retention time = about 10 min); impurity C = about 0.4;

lincomycin B = about 0.5; impurity A = about 0.7;

impurity B = about 1.2 and 1.3.

System suitability: reference solution (b):

— resolution: minimum 1.8 between the peak due to lincomycin and the 1st peak due to impurity B.

Limits:

— impurity A: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);

— sum of the areas of the peaks due to impurity B: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (d) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.10 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (2.0 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 5 ppm.

2.0 g complies with test C. Prepare the reference solution using 1.0 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

3.1 per cent to 4.6 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 0.50 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solutions (a) and (c).

Calculate the percentage content of C₁₈H₃₅ClN₂O₆S (lincomycin) and C₁₇H₃₃ClN₂O₆S (lincomycin B) taking into account the assigned content of C₁₈H₃₅ClN₂O₆S in lincomycin hydrochloride CRS. Determine the content of lincomycin by comparing with the area of the peak due to lincomycin in the chromatogram obtained with reference solution (a). Determine the content of lincomycin B by comparing with the area of the peak due to lincomycin in the chromatogram obtained with reference solution (c).

STORAGE

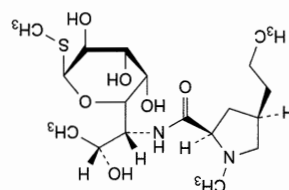
At a temperature not exceeding 30 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

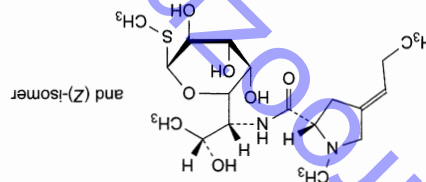
Specified impurities A, B, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

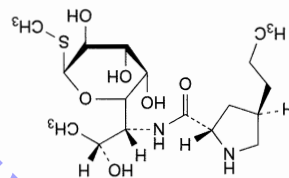
Control of impurities in substances for pharmaceutical use):
D, E, F.



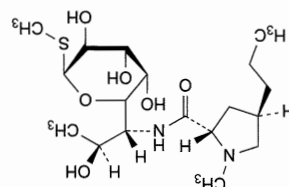
A. methyl 6,8-dideoxy-6-[[[(2R,4R)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-erythro-α-D-galactopyranoside (α-amide epimer),



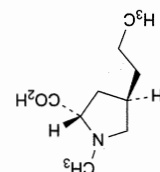
B. methyl 6,8-dideoxy-6-[[[(2S,4E)-1-methyl-4-propylidene]pyrrolidin-2-yl]carbonyl]amino]-1-thio-D-erythro-α-D-galactopyranoside (propylidene analogues),



C. methyl 6,8-dideoxy-6-[[[(2S,4R)-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-D-erythro-α-D-galactopyranoside (N-desmethyl lincomycin),



D. methyl 6,8-dideoxy-6-[[[(2S,4R)-1-methyl-4-propylpyrrolidin-2-yl]carbonyl]amino]-1-thio-L-threo-α-D-galactopyranoside (7-epi-lincomycin),



E. (2S,4R)-1-methyl-4-propylpyrrolidine-2-carboxylic acid (4-propyl hygric acid),

Linoyleyl Macrogliglycerides

(Ph. Eur. monograph 1232)

Action and use
Excipient.

Ph. Eur.

DEFINITION

Mixtures of monoesters, diesters and triesters of glycerol and monoesters and diesters of macrogols. They are obtained by partial alcoholysis of an unsaturated oil mainly containing triglycerides of linoleic (*cis*, *cis*-9,12-octadecadienoic) acid, using macrogol with a mean relative molecular mass between 300 and 400, or by esterification of glycerol and macrogol with unsaturated fatty acids, or by mixing glycerol esters and condensates of ethylene oxide with the fatty acids of this unsaturated oil.

CHARACTERS

Appearance
Amber, oily liquid which may give rise to a deposit after prolonged periods at 20 °C.

Solubility
Practically insoluble but dispersible in water, freely soluble in methylene chloride.

Viscosity
About 35 mPa.s at 40 °C.

Relative density
About 0.95 at 20 °C.

Refractive index
About 1.47 at 20 °C.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 1.0 g of the substance to be examined in methylene chloride R and dilute to 20 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase hexane R, ether R (30:70 V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of rhodamine B R in ethanol (96 per cent) R and examine in ultraviolet light at

365 nm.

Results The chromatogram shows a spot due to triglycerides with an R_F value of about 0.9 (R_F 1) and spots due to 1,3-diglycerides (R_F 0.7), to 1,2-diglycerides (R_F 0.6), to monoglycerides (R_F 0.1) and to esters of macrogol (R_F 0).

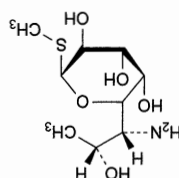
B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Fatty acid composition (see Tests).

Ph. Eur.

F. methyl 6-amino-6,8-dideoxy-1-thio-D-erythro-α-D-galactopyranoside (methyl-1-thiolinosaminide).



Virgin Linseed Oil

(Ph. Eur. monograph 1908)



Ph Eur

DEFINITION

Fatty oil obtained by cold expression from ripe seeds of *Linum usitatissimum* L. A suitable antioxidant may be added.

CHARACTERS

Appearance

Clear, yellow or brownish-yellow liquid, on exposure to air turning dark and gradually thickening. When cooled, it becomes a soft mass at about -20°C .

Solubility

Very slightly soluble in ethanol (96 per cent), miscible with light petroleum.

Relative density

About 0.931.

Refractive index

About 1.480.

IDENTIFICATION

First identification: B, C.

Second identification: A, B.

A. Identification of fatty oils by thin-layer chromatography (2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2-1.

B. Iodine value (see Tests).

C. Composition of fatty acids (see Tests).

TESTS

Acid value (2.5.1)

Maximum 4.5.

Iodine value (2.5.4)

160 to 200.

Peroxide value (2.5.5, Method A)

Maximum 15.0.

Saponification value (2.5.6)

188 to 195; carry out the saponification for 1 h.

Unsaponifiable matter (2.5.7)

Maximum 1.5 per cent, determined on 5.0 g.

Composition of fatty acids

Gas chromatography (2.4.22, Method C). Use the calibration mixture in Table 2.4.22-3.

Composition of the fatty-acid fraction of the oil:

— fatty acids with a chain length less than C_{16} : maximum

1.0 per cent,

— palmitic acid: 3.0 per cent to 8.0 per cent,

— palmitoleic acid: maximum 1.0 per cent,

— stearic acid: 2.0 per cent to 8.0 per cent,

— oleic acid: 11.0 per cent to 35.0 per cent,

— linoleic acid: 11.0 per cent to 24.0 per cent,

— linolenic acid: 35.0 per cent to 65.0 per cent,

— arachidic acid: maximum 1.0 per cent.

Cadmium

Maximum 0.5 ppm, determined as described in general method 2.4.27. Heavy metals in herbal drugs and herbal drug preparations.

Water (2.5.32)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

In an airtight container, protected from light.

Ph Eur

TESTS

Acid value (2.5.1)

Maximum 2.0, determined on 2.0 g.

Hydroxyl value (2.5.3, Method A)

45 to 65, determined on 1.0 g.

Iodine value (2.5.4, Method A)

90 to 110.

Peroxide value (2.5.5, Method A)

Maximum 12.0, determined on 2.0 g.

Saponification value (2.5.6)

150 to 170, determined on 2.0 g.

Alkaline impurities

Into a test tube introduce 5.0 g and carefully add a mixture, neutralised if necessary with 0.01 M hydrochloric acid or with 0.01 M sodium hydroxide, of 0.05 mL of a 0.4 g/L solution of bromophenol blue R in ethanol (96 per cent) R, 0.3 mL of water R and 10 mL of ethanol (96 per cent) R. Shake and allow to stand. Not more than 1.0 mL of 0.01 M hydrochloric acid is required to change the colour of the upper layer to yellow.

Free glycerol

Maximum 3.0 per cent.

Dissolve 1.20 g in 25.0 mL of methylene chloride R. Heat if necessary. After cooling, add 100 mL of water R. Shake and add 25.0 mL of peracetic acid solution R. Shake and allow to stand for 30 min. Add 40 mL of a 75 g/L solution of potassium iodide R. Allow to stand for 1 min. Add 1 mL of starch solution R. Titrate the iodine with 0.1 M sodium thiosulfate. Carry out a blank titration.

1 mL of 0.1 M sodium thiosulfate is equivalent to 2.3 mg of glycerol.

Composition of fatty acids (2.4.22, Method A).

Composition of the fatty-acid fraction of the substance:

— palmitic acid: 4.0 per cent to 20.0 per cent;

— stearic acid: maximum 6.0 per cent;

— oleic acid: 20.0 per cent to 35.0 per cent;

— linoleic acid: 50.0 per cent to 65.0 per cent;

— linolenic acid: maximum 2.0 per cent;

— arachidic acid: maximum 1.0 per cent;

— eicosenoic acid: maximum 1.0 per cent.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.0 g. Use a mixture of 30 volumes of anhydrous methanol R and 70 volumes of methylene chloride R as solvent.

Total ash (2.4.16)

Maximum 0.1 per cent.

STORAGE

Protected from light.

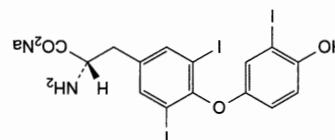
LABELLING

The label states the type of macrogol used (mean relative molecular mass) or the number of units of ethylene oxide per molecule (nominal value).

Ph Eur

Liothyronine Sodium

(Ph. Eur. monograph 0728)


 $C_{15}H_{11}I_2NNaO_4$ 673

55-06-1

Action and use

Thyroid hormone replacement.

Preparation

Liothyronine Tablets

Ph Eur

DEFINITION

Sodium (2S)-2-amino-3-[4-(4-hydroxy-3-iodophenoxy)-3,5-diodophenyl]propanoate.

Content

95.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly coloured, hygroscopic powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification A, C, E.

Second identification A, B, D, E.

A. Specific optical rotation (see Tests).

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 10.0 mg in 0.1 M sodium hydroxide and dilute to 100.0 mL with the same solvent.

Spectral range 230-350 nm.

Absorption maximum At 319 nm.

Specific absorbance at the absorption maximum 63 to 69 (dried substance).

C. Infrared absorption spectrophotometry (2.2.24).

Comparison liothyronine sodium CRS.

D. To about 50 mg in a porcelain dish add a few drops of sulfuric acid R and heat. Violet vapour is evolved.

E. To 200 mg add 2 mL of dilute sulfuric acid R. Heat on a water-bath and then carefully over a naked flame, increasing the temperature gradually up to about 600 °C. Continue the ignition until most of the particles have disappeared. Dissolve the residue in 2 mL of water R. The solution gives

TESTS

Specific optical rotation (2.2.7)

+ 18.0 to + 22.0 (dried substance).

Dissolve 0.200 g in a mixture of 1 mL of 1 M hydrochloric acid and 4 volumes of ethanol (96 per cent) R and dilute to 20.0 mL with the same mixture of solvents.

Related substances

Liquid chromatography (2.2.29). Protect the solutions from light throughout the test.

Solution A Mix 10 volumes of mobile phase A with 90 volumes of methanol R.

Solution B Mix 30 volumes of mobile phase B and 70 volumes of mobile phase A. Mix equal volumes of this solution with solution A.

Test solution Dissolve 20.0 mg of the substance to be examined in 20 mL of solution A. Dilute 4.0 mL of this solution to 20.0 mL with solution B.

Reference solution (a) Dissolve 2.5 mg of levothyronine sodium CRS (impurity A) and 2.5 mg of liothyronine sodium CRS in solution A and dilute to 25 mL with the same solution. Dilute 1.0 mL of this solution to 50.0 mL with solution B.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with solution B.

Reference solution (c) Dissolve the contents of a vial of liothyronine for peak identification CRS (containing impurities A, B, C, D and E) in solution B and dilute to 1.0 mL with the same solution.

Reference solution (d) Dissolve 20.0 mg of liothyronine sodium CRS in 20 mL of solution A. Dilute 4.0 mL of this solution to 20.0 mL with solution B.

Blank solution Solution B.

Column:

— size: $l = 0.15$ m, $\phi = 4.0$ mm;— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m).

Mobile phase:

— mobile phase A: dissolve 9.7 g of sulfuric acid R in water R and dilute to 2000 mL with the same solvent; add 1.5 g of sodium hydroxide R and adjust to pH 2.0 with 2 M sodium hydroxide;

— mobile phase B: acetonitrile R1;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	75	25
3 - 4	75 \rightarrow 70	25 \rightarrow 30
4 - 14	70	30
14 - 44	70 \rightarrow 20	30 \rightarrow 80
44 - 54	20	80

Flow rate 1 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 25 μ L of the test solution and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D and E.

— relative retention with reference to liothyronine (retention time = about 14 min): impurity B = about 0.2; impurity E = about 0.5; impurity A = about 1.4; impurity C = about 2; impurity D = about 2.4.

System suitability:

— resolution: minimum 5.0 between the peaks due to impurity A and liothyronine in the chromatogram obtained with reference solution (a).

Limits:

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

- *impurity E*: not more than 5 times the area of the peak due to liothyronine in the chromatogram obtained with reference solution (b) (0.5 per cent);
- *impurities B, C*: for each impurity, not more than 3 times the area of the peak due to liothyronine in the chromatogram obtained with reference solution (b) (0.3 per cent);
- *impurity D*: not more than twice the area of the peak due to liothyronine in the chromatogram obtained with reference solution (b) (0.2 per cent);
- *unspecified impurities*: for each impurity, not more than the area of the peak due to liothyronine in the chromatogram obtained with reference solution (b) (0.10 per cent);
- *total*: not more than twice the area of the peak due to liothyronine in the chromatogram obtained with reference solution (a) (2.0 per cent);
- *disregard limit*: 0.5 times the area of the peak due to liothyronine in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides

Maximum 2.0 per cent, expressed as NaCl (dried substance).

Dissolve 0.500 g in a 2 g/L solution of sodium hydroxide R and dilute to 100 mL with the same solvent. Add 15 mL of dilute nitric acid R and titrate with 0.05 M silver nitrate, determining the end-point potentiometrically (2.2.20).

1 mL of 0.05 M silver nitrate is equivalent to 2.93 mg of NaCl.

Loss on drying (2.2.32)

Maximum 4.0 per cent, determined on 0.500 g by drying *in vacuo* at 60 °C.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (d).

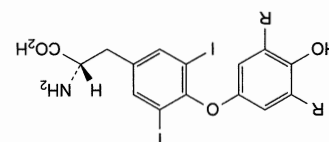
Calculate the percentage content of $C_{15}H_{11}I_3NNaO_4$ from the declared content of liothyronine sodium CRS.

STORAGE

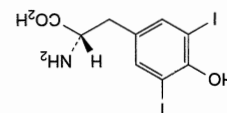
In an airtight container, protected from light, at a temperature between 2 °C and 8 °C.

IMPURITIES

Specified impurities A, B, C, D, E



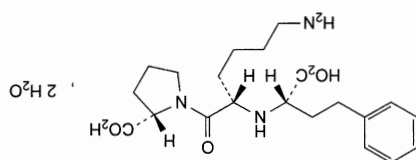
A, R = I: levothyronine,
E, R = H: (2S)-2-amino-3-[4-(4-hydroxyphenoxy)-3,5-diododiphenyl]propanoic acid (diodothyronine).



B, (2S)-2-amino-3-[4-(4-hydroxy-3,5-diododiphenyl)propanoic acid (diodothyronine),

Lisinopril Dihydrate

(Ph. Eur. monograph 1120)



$C_{21}H_{31}N_3O_5 \cdot 2H_2O$ 441.5 83915-83-7

Action and use

Angiotensin converting enzyme inhibitor.

Preparations

Lisinopril Oral Solution

Lisinopril Tablets

DEFINITION

(2S)-1-[(2S)-6-amino-2-[[[(1S)-1-carboxy-3-phenylpropyl]amino]hexanoyl]pyrrolidine-2-carboxylic acid dihydrate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility

Soluble in water, Practically insoluble in anhydrous ethanol and in heptane.

IDENTIFICATION

A. Specific optical rotation (2.2.7): -47 to -43 (anhydrous substance).

Dissolve 0.5 g in zinc acetate solution R and dilute to 50.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison lisinopril dihydrate CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 5 mg of lisinopril for system suitability A CRS (containing impurities A and E) in mobile phase A and dilute to 1.0 mL with mobile phase A.

— reporting threshold: 0.05 per cent; disregard any peak with a retention time less than 3 min.

Water (2.5.12)

8.0 per cent to 9.5 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

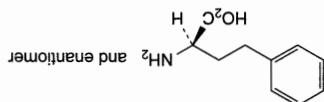
Dissolve 0.350 g in 50 mL of water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 40.55 mg of $C_{21}H_{31}N_3O_5$.

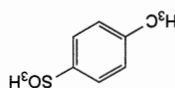
IMPURITIES

Specified impurities A, E, F, G

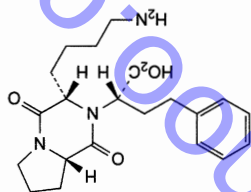
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, H, I.



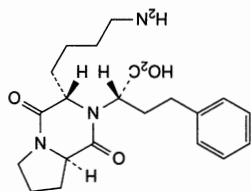
A. (2R)-2-amino-4-phenylbutanoic acid,



B. 4-methylbenzenesulfonic acid,



C. (2S)-2-[(3S,8aS)-3-(4-aminobutyl)-1,4-dioxohexahydropyrido[1,2-a]pyrazin-2(1H)-yl]-4-phenylbutanoic acid (S,S,S-diketopiperazine),



D. (2S)-2-[(3S,8aR)-3-(4-aminobutyl)-1,4-dioxohexahydropyrido[1,2-a]pyrazin-2(1H)-yl]-4-phenylbutanoic acid (R,S,S-diketopiperazine),

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve the contents of a vial of lisinopril impurity F CRS in 1.0 mL of mobile phase A.

Reference solution (d) Dissolve 5 mg of lisinopril for peak identification CRS (containing impurity G) in mobile phase A and dilute to 1.0 mL with mobile phase A.

Column:

size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

temperature: 50 °C.

Mobile phase:

mobile phase A: mix 3 volumes of acetonitrile R1 and 97 volumes of 3.12 g/L solution of sodium dihydrogen phosphate R previously adjusted to pH 3.8 with dilute phosphoric acid R;

mobile phase B: mix 20.5 volumes of acetonitrile R1 and 79.5 volumes of 3.12 g/L solution of sodium dihydrogen

phosphate R previously adjusted to pH 3.5 with dilute phosphoric acid R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	100	0
2 - 37	100 \rightarrow 0	0 \rightarrow 100
37 - 62	0	100

Flow rate 1.6 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 50 μ L.

Identification of impurities Use the chromatogram supplied

with lisinopril for system suitability A CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and E; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity F; use the chromatogram supplied with lisinopril for peak identification CRS and the chromatogram obtained with reference solution (d) to identify the peak due to impurity G.

Relative retention With reference to lisinopril (retention time = about 14 min): impurity A = about 0.7;

impurity B = about 1.2; impurity F = about 1.9;

impurity G = about 2.9.

System suitability:

resolution: minimum 1.5 between the peaks due to lisinopril and impurity E in the chromatogram obtained with reference solution (a);

signal-to-noise ratio: minimum 45 for the principal peak in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:

— correction factor: multiply the peak area of impurity F by 2.1;

— for each impurity, use the concentration of lisinopril in reference solution (b).

Limits:

impurities A, E, F: for each impurity, maximum

0.2 per cent;

impurity G: maximum 0.15 per cent;

unspecified impurities: for each impurity, maximum 0.10 per cent;

total: maximum 0.5 per cent.

Lithium Carbonate

(Ph. Eur. monograph 0228)

 Li_2CO_3

73.9

554-13-2



Action and use

Prophylaxis of affective disorders.

Preparations

Lithium Carbonate Tablets

Prolonged-release Lithium Carbonate Tablets

DEFINITION

Content

98.5 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. When moistened with hydrochloric acid R, it gives a red

colour to a non-luminous flame.

B. Dissolve 0.2 g in 1 mL of hydrochloric acid R. Evaporate to

dryness on a water-bath. The residue dissolves in 3 mL of

C. It gives the reaction of carbonates (2.3.1).

TESTS

Solution S

Suspend 10.0 g in 30 mL of distilled water R and dissolve by

the addition of 22 mL of nitric acid R. Add dilute sodium

hydroxide solution R until the solution is neutral and dilute to

100 mL with distilled water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 200 ppm.

Disperse 1.25 g in 5 mL of distilled water R and dissolve by

adding 5 mL of hydrochloric acid R. Boil for 2 min. Cool

and add dilute sodium hydroxide solution R until neutral. Dilute

to 25 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Calcium (2.4.3)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Magnesium (2.4.6)

Maximum 150 ppm.

Dilute 1 mL of solution S to 10 mL with water R. Dilute

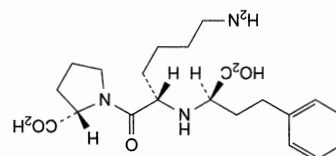
6.7 mL of this solution to 10 mL with water R.

Potassium

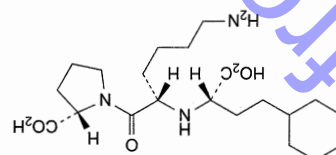
Maximum 300 ppm.

Atomic emission spectrometry (2.2.22, Method I).

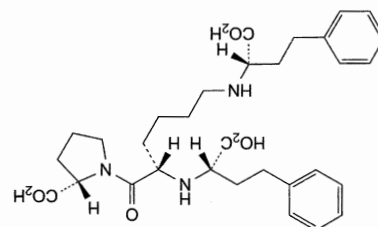
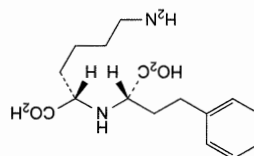
E. (2S)-1-[(2S)-6-amino-2-[[[(1R)-1-carboxy-3-phenylpropyl]amino]hexanoyl]pyrrolidine-2-carboxylic acid (lisinopril R,S,S-isomer),



F. (2S)-1-[(2S)-6-amino-2-[[[(1S)-1-carboxy-3-cyclohexylpropyl]amino]hexanoyl]pyrrolidine-2-carboxylic acid (cyclohexyl analogue),



G. (2S)-1-[(2S)-6-amino-2-[[[(2S)-1-[[[(5S)-5-[(1S)-1-carboxy-3-phenylpropyl]amino]-6-[(2S)-2-carboxy-3-phenylpyrrolidin-1-yl]-6-oxohexyl]amino]carbonyl]-3-phenylpropyl]amino]hexanoyl]pyrrolidine-2-carboxylic acid (lisinopril dimer),



I. (2S)-1-[(2S)-2,6-bis-[[[(1S)-1-carboxy-3-phenylpropyl]amino]hexanoyl]pyrrolidine-2-carboxylic acid.

Ph Eur

Test solution Dissolve 1.0 g in 10 mL of hydrochloric acid R1 and dilute to 50.0 mL with water R.

Reference solutions Prepare the reference solutions using a solution of potassium chloride R containing 500 µg of K per millilitre, diluted as necessary with water R.

Wave length 766.5 nm.

Sodium Maximum 300 ppm.

Atomic emission spectrometry (2.2.22, Method I). Test solution Dissolve 1.0 g in 10 mL of hydrochloric acid R1 and dilute to 50.0 mL with water R.

Reference solutions Prepare the reference solutions using a solution of sodium chloride R containing 500 µg of Na per millilitre, diluted as necessary with water R.

Wave length 589 nm.

Heavy metals (2.4.8) Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

ASSAY

Dissolve 0.500 g in 25.0 mL of 1 M hydrochloric acid. Titrate with 1 M sodium hydroxide, using methyl orange solution R as indicator.

1 mL of 1 M hydrochloric acid is equivalent to 36.95 mg of Li_2CO_3 .

Lithium Citrate

(Ph. Eur. monograph 0621)

$\text{C}_6\text{H}_5\text{LiO}_7 \cdot 4\text{H}_2\text{O}$

282.0

6080-58-6



Ph Eur

Ph Eur

DEFINITION

Trilithium 2-hydroxypropylpropane-1,2,3-tricarboxylate tetrahydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, fine crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. When moistened with hydrochloric acid R, it gives a red colour to a non-luminous flame.

B. Dilute 3 mL of solution S (see Tests) to 10 mL with water R. Add 3 mL of potassium ferriperoxide solution R.

A white or yellowish-white precipitate is formed.

C. To 1 mL of solution S add 4 mL of water R. The solution gives the reaction of citrates (2.3.1).

TESTS

Solution S Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of phenolphthalein solution R. Not more than 0.2 mL of 0.1 M hydrochloric acid or 0.1 M sodium hydroxide is required to change the colour of the indicator.

Readily carbonisable substances

To 0.20 g of the powdered substance to be examined add 10 mL of sulfuric acid R and heat in a water-bath at $90 \pm 1^\circ\text{C}$ for 60 min. Cool rapidly. The solution is not more intensely coloured than reference solution Y₂ or GY₂ (2.2.2, Method II).

Chlorides (2.4.4)

Maximum 100 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Oxalates

Maximum 300 ppm, calculated as anhydrous oxalate ion.

Dissolve 0.50 g in 4 mL of water R, add 3 mL of hydrochloric acid R and 1 g of zinc R in granules and heat on a water-bath for 1 min. Allow to stand for 2 min, decant the liquid into a test-tube containing 0.25 mL of a 10 g/L solution of phenylhydrazine hydrochloride R and heat to boiling. Cool rapidly, transfer to a graduated cylinder and add an equal volume of hydrochloric acid R and 0.25 mL of potassium ferricyanide solution R. Shake and allow to stand for 30 min. Any pink colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 4 mL of a 0.05 g/L solution of oxalic acid R.

Sulfates (2.4.13)

Maximum 500 ppm.

To 3 mL of solution S add 2 mL of hydrochloric acid R1 and dilute to 17 mL with distilled water R. Prepare the reference solution using 15 mL of a mixture of 2 mL of hydrochloric acid R1 and 15 mL of sulfate standard solution (10 ppm SO_4) R and compare the opalescence after 15 min.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

24.0 per cent to 27.0 per cent, determined on 0.100 g. After adding the substance to be examined, stir for 15 min before titrating. Carry out a blank titration.

ASSAY

Dissolve 80.0 mg in 50 mL of anhydrous acetic acid R, heating to about 50°C . Allow to cool. Titrate with 0.1 M perchloric acid, using 0.25 mL of naphtholbenzene solution R as indicator, until the colour changes from yellow to green.

1 mL of 0.1 M perchloric acid is equivalent to 7.00 mg of $\text{C}_6\text{H}_5\text{Li}_3\text{O}_7$.

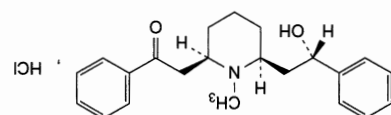
STORAGE

In an airtight container.

Ph Eur

Lobeline Hydrochloride

(Ph. Eur. monograph 1988)



$C_{22}H_{28}ClNO_2$

373.9

134-63-4

Action and use
Respiratory stimulant.

DEFINITION

2-[(2R,6S)-6-[(2S)-2-Hydroxy-2-phenylethyl]-1-methylpiperidin-2-yl]-1-phenylethanol hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, microcrystalline powder.

Solubility

Sparingly soluble in water, freely soluble in ethanol (96 per cent), soluble in methylene chloride.

IDENTIFICATION

First identification: A, B.
Second identification: B, C.

A. Infrared absorption spectrophotometry (2.2.24).
Comparison lobeline hydrochloride CRS.

B. Solution S (see Tests) gives reaction (a) of chlorides (2.3.1).

C. Examine the chromatograms obtained in the test for foreign alkaloids.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (b).

TESTS

Solution S

Dissolve 0.250 g in carbon dioxide-free water R prepared from distilled water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.6 to 6.4 for solution S.

Specific optical rotation (2.2.7)

–55 to –59 (dried substance), determined on solution S.

Foreign alkaloids

Test solution (a) Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 5.0 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with methanol R.

Reference solution (a) Dilute 0.1 mL of test solution (a) to 10 mL with methanol R.

Reference solution (b) Dissolve 10 mg of lobeline hydrochloride CRS in methanol R and dilute to 5 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of phenytoin CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

To 1 mL of the solution add 0.1 mL of the test solution and dilute to 25 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4$ mm,

— stationary phase: spherical end-capped octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve 1.0 g of sodium methanesulfonate R and 2.50 g of disodium hydrogen phosphate dihydrate R in a mixture of 3 volumes of a 6.7 per cent V/V solution of phosphoric acid R, 29 volumes of acetonitrile R and 70 volumes of water R and dilute to 1000 mL with the same mixture of solvents.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 μ L.

Run time 2 times the retention time of lobeline which is about 17 min.

System suitability: reference solution (b):
— resolution: minimum 4.0 between the peaks due to phenytoin and to lobeline.

Limits:

— any impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),

— total: maximum of 2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),

— disregard level: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Sulfates (2.4.13)

Maximum 0.1 per cent, determined on solution S.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g in vacuo.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY

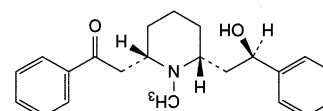
Dissolve 0.300 g in 50 mL of ethanol (96 per cent) R. Add 5 mL of 0.01 M hydrochloric acid. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of inflexion.
1 mL of 0.1 M sodium hydroxide is equivalent to 37.39 mg of $C_{22}H_{28}ClNO_2$.

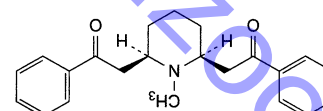
STORAGE

Protected from light.

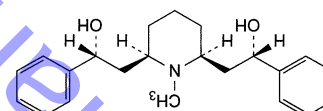
IMPURITIES



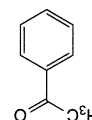
A. 2-[(2S,6R)-6-[(2R)-2-hydroxy-2-phenylethyl]-1-methylpiperidin-2-yl]-1-phenylethanol ((+)-lobeline),



B. 2,2'-[(2R,6S)-1-methylpiperidine-2,6-bis(1-phenylethanol)] (lobelamine),

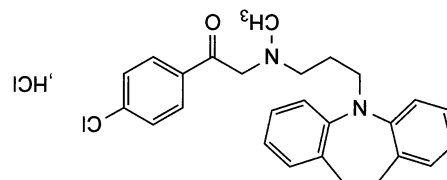


C. *meso*-(1R,1'S)-2,2'-[(2R,6S)-1-methylpiperidine-2,6-bis(1-phenylethanol)] (lobelaneline),



D. acetophenone.

Lofepramine Hydrochloride



$C_{26}H_{27}ClN_2O_2 \cdot HCl$

455.4

26786-32-3

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

Preparation

Lofepramine Tablets

DEFINITION

Lofepramine Hydrochloride is 5-(3-[N-(4-chlorophenyl)-N-methylamino]propyl)-1,1,1-dihydro-5H-dibenz[b,f]azepine hydrochloride. It contains not less than 98.5% and not more than 101.0% of $C_{26}H_{27}ClN_2O_2 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A fine, yellowish white to greenish yellow powder. Very soluble in ethanol (96%) and in methanol; slightly soluble in acetone; very slightly soluble in water.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of lofepramine hydrochloride (form A) (RS 3994).

B. 2 mL of a 1% w/v solution in ethanol (96%) complies with the test for chlorides, Appendix VI.

TESTS

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions. Solution (1) contains 0.1% w/v of the substance being examined in the mobile phase. For solution (2) dilute 1 volume of solution (1) to 200 volumes with the mobile phase. Solution (3) contains 0.002% w/v each of desipramine hydrochloride BPQRS and imipramine hydrochloride BPQRS in the mobile phase.

Inject 20 µL of solutions (1) and (2). Inject 20 µL of solution (3) and allow the chromatography to continue for 4 times the retention time of the principal peak.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with base-deactivated end-capped octylsilyl silica gel for chromatography (5 µm) (Lichrospher 60 RP-select B is suitable) maintained at 50°, (b) as the mobile phase with a flow rate of 1.5 mL per minute a 0.09% w/v solution of sodium dodecyl sulfate in a mixture of 550 volumes of acetonitrile, 325 volumes of water and 125 volumes of a buffer solution of pH 1.0 containing 0.015% w/v of glycine, 0.018% w/v of sodium chloride and 0.44% w/v of hydrochloric acid and (c) a detection wavelength of 254 nm.

The test is not valid unless, in the chromatogram obtained with solution (3), the resolution factor between the two principal peaks is at least 0.9.

In the chromatogram obtained with solution (1) the area of any secondary peak is not greater than the area of the peak in the chromatogram obtained with solution (2) (0.5%) and the sum of the areas of any secondary peaks is not greater than twice the area of the peak in the chromatogram obtained with solution (2) (1%).

Loss on drying

When dried at a temperature of 100° at a pressure not exceeding 0.2 kPa, loses not more than 0.5% of its weight.

ASSAY

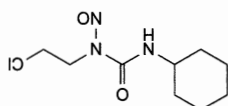
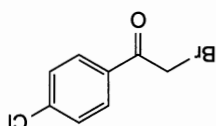
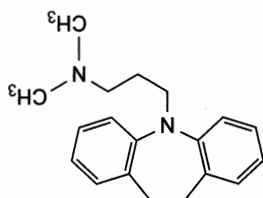
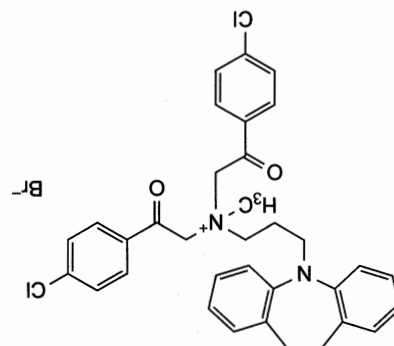
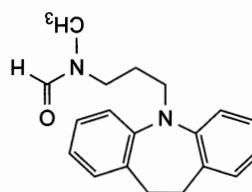
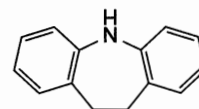
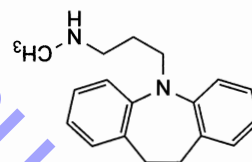
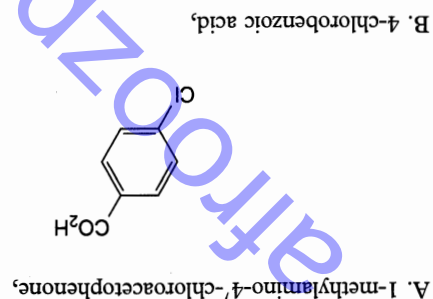
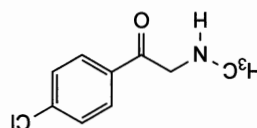
Carry out the method for liquid chromatography, Appendix III D, using the following solutions. Solution (1) contains 0.02% w/v of the substance being examined in the mobile phase. Solution (2) contains 0.02% w/v of lofepramine hydrochloride BPQRS in the mobile phase. The chromatographic procedure described under the test for Related substances may be used.

Inject 20 µL of each solution. Calculate the content of $C_{26}H_{27}ClN_2O_2 \cdot HCl$ from the chromatograms obtained and using the declared content of $C_{26}H_{27}ClN_2O_2 \cdot HCl$ in lofepramine hydrochloride BPQRS.

STORAGE

Lofepiramine Hydrochloride should be kept in an airtight container and protected from light.

IMPURITIES



G. imipramine,

H. 2-bromo-4'-chloroacetophenone.

(Ph. Eur. monograph 0928)



Lomustine

Action and use
Cytotoxic alkylating agent.

Preparation
Lomustine Capsules

Ph Eur

DEFINITION

1-(2-Chloroethyl)-3-cyclohexyl-1-nitrosourea.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone and in methylene chloride, soluble in ethanol (96 per cent).

Carry out the tests protected from light and prepare all the solutions immediately before use.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison lomustine CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Phosphate buffer solution Dissolve 1.36 g of potassium

dihydrogen phosphate R in 900 mL of water R, adjust to

pH 3.0 with dilute phosphoric acid R and dilute to 1000 mL

with water R.

Test solution Dissolve 50.0 mg of the substance to be

examined in acetonitrile R1 and dilute to 10.0 mL with the

same solvent.

Reference solution (a) Dissolve 5.0 mg of dicyclohexylurea R

(impurity C) in methanol R and dilute to 10.0 mL with the

same solvent. Dilute 1.0 mL of the solution to 50.0 mL with acetamintrile R1. Mix 1.0 mL of this solution and 1.0 mL of the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with acetamintrile R1. Dilute 1.0 mL of this solution to 10.0 mL with acetamintrile R1.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: acetamintrile R1, phosphate buffer solution (20:80 V/V);

— mobile phase B: phosphate buffer solution, acetamintrile R1 (24:76 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	75	25
2 - 17	75 \rightarrow 40	25 \rightarrow 60
17 - 34	40 \rightarrow 30	60 \rightarrow 70
34 - 42	30	70

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to

impurity C.

Relative retention With reference to lornustine (retention time = about 23 min): impurity C = about 0.7.

System suitability: reference solution (a):

— resolution: minimum 5.0 between the peaks due to

impurity C and lornustine.

Calculation of percentage contents:

— for each impurity, use the concentration of lornustine in

reference solution (b).

Limits:

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.2 per cent;

— reporting threshold: 0.05 per cent.

Chlorides (2.4.4)

Maximum 500 ppm.

Dissolve 0.24 g in 4 mL of methanol R and add 20 mL of

water R. Allow to stand for 20 min and filter. To 10 mL of

the filtrate, add 5 mL of methanol R. When preparing the

standard, replace the 5 mL of water R with 5 mL of

methanol R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

a desiccator over diposphorus pentoxide R at a pressure not

exceeding 0.7 kPa for 24 h.

ASSAY

Dissolve 0.200 g in about 3 mL of ethanol (96 per cent) R

and add 20 mL of a 200 g/L solution of potassium

hydroxide R and boil under a reflux condenser for 2 h.

Add 75 mL of water R and 4 mL of nitric acid R. Cool and

titrate with 0.1 M silver nitrate, determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

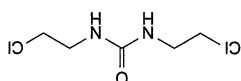
1 mL of 0.1 M silver nitrate is equivalent to 23.37 mg of $C_{29}H_{34}Cl_2N_2O_2$.

STORAGE

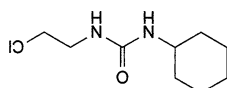
Protected from light.

IMPURITIES

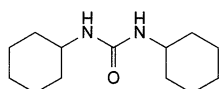
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C.



A. 1,3-bis(2-chloroethyl)urea,



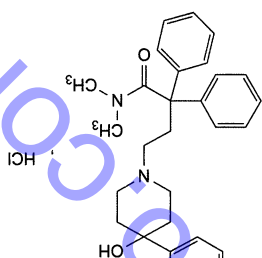
B. 1-(2-chloroethyl)-3-cyclohexylurea,



C. 1,3-dicyclohexylurea.

Loperamide Hydrochloride

(Ph. Eur. monograph 0929)



$C_{29}H_{34}Cl_2N_2O_2$ 513.5

34552-83-5

Action and use

Opioid receptor agonist; antidiarrhoeal.

Preparation

Loperamide Capsules

DEFINITION

[4-(4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl)-N,N-dimethyl-2,2-diphenylbutanamide hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, freely soluble in ethanol (96 per cent) and in methanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison loperamide hydrochloride CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of methylene chloride R, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 10.0 mg of loperamide hydrochloride for system suitability CRS in methanol R and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with methanol R. Dilute 1.0 mL of this solution to 25.0 mL with methanol R.

Column:

size: $l = 0.10$ m, $\varnothing = 4.6$ mm,

stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (3 μ m),

temperature: 35 °C.

Mobile phase:

mobile phase A: 17.0 g/L solution of tetrabutylammonium hydrogen sulfate R1,

mobile phase B: acetonitrile R,

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L.

System suitability: reference solution (a):

H_p = height above the baseline of the peak due to impurity G and

H_p = height above the baseline of the peak due to impurity H;

peak-to-valley ratio: minimum 1.5, where H_p = height above the baseline of the peak due to impurity E and

H_p = height above the baseline of the peak due to impurity F;

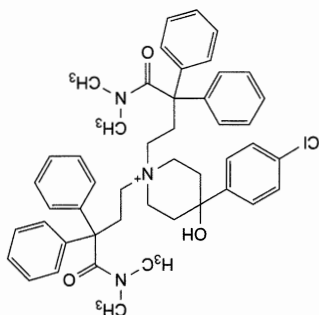
curve separating this peak from the peak due to

impurity A;

the chromatogram obtained is concordant with the chromatogram supplied with loperamide hydrochloride for system suitability CRS.

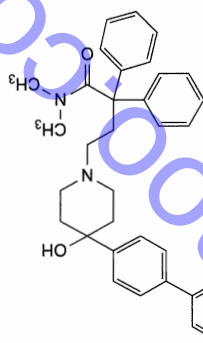
Limit:

— correction factors: for the calculation of contents, multiply the peak areas of the following impurities by the



B. 4-(4-chlorophenyl)-1,1-bis[4-(dimethylamino)-4-oxo-3,3-diphenylbutyl]-4-hydroxypiperidinium,

A. 4-[4-(4'-chlorobiphenyl-4-yl)-4-hydroxypiperidin-1-yl]-N,N-dimethyl-2,2-diphenylbutanamide,



Specified impurities: A, B, C, D, E, F, G, H.

IMPURITIES

Protected from light.

STORAGE

$C_{29}H_{34}Cl_2N_2O_2$.

1 mL of 0.1 M sodium hydroxide is equivalent to 51.35 mg of

infusion.

hydroxide. Read the volume added between the 2 points of

potentiometric titration (2.2.20), using 0.1 M sodium

5.0 mL of 0.01 M hydrochloric acid. Carry out a

Dissolve 0.400 g in 50 mL of ethanol (96 per cent) R and add

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C for 4 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

(0.05 per cent).

the chromatogram obtained with reference solution (b)

— disregard limit: 0.25 times the area of the principal peak in

solution (b) (0.3 per cent);

peak in the chromatogram obtained with reference

total: not more than 1.5 times the area of the principal

(0.10 per cent);

chromatogram obtained with reference solution (b)

0.5 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than

(0.2 per cent);

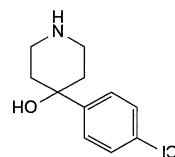
chromatogram obtained with reference solution (b)

more than the area of the principal peak in the

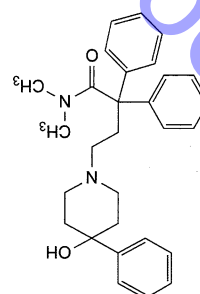
— impurities A, B, C, D, E, F, G, H: for each impurity, not

impurity D = 1.7;

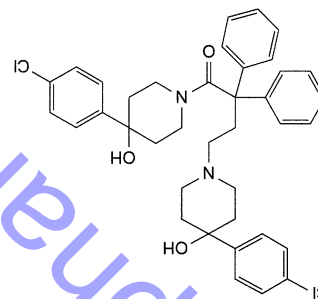
corresponding correction factor: impurity A = 1.3;



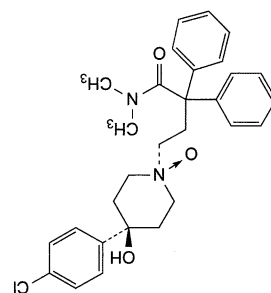
C. 4-(4-chlorophenyl)piperidin-4-ol,



D. 4-(4-hydroxy-4-phenylpiperidin-1-yl)-N,N-dimethyl-2,2-diphenylbutanamide,



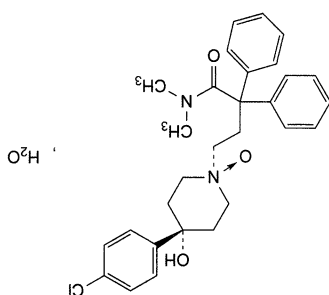
E. 4-(4-chlorophenyl)-1-[4-(4-chlorophenyl)-1-yl]-2,2-diphenylbutanoyl]piperidin-4-ol,



G. 4-[cis-4-(4-chlorophenyl)-4-hydroxy-1-oxido-1-yl]-N,N-dimethyl-2,2-diphenylbutanamide,

Loperamide Oxide Monohydrate

(Ph. Eur. monograph 1729)

 $C_{29}H_{33}ClN_2O_3 \cdot H_2O$

511.1

106900-12-3

Action and use

Opioid receptor agonist; antidiarrhoeal.

Ph Eur

DEFINITION

4-[trans-4-(4-chlorophenyl)-4-hydroxy-1-oxido-1-yl]-N,N-dimethyl-2,2-diphenylbutanamide monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder, slightly hygroscopic.

Solubility

Practically insoluble in water, freely soluble in alcohol and in methylene chloride.

mp

About 152 °C, with decomposition.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison loperamide oxide monohydrate CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of lopinavir hydrochloride CRS in methanol R, add 0.5 mL of the test solution and dilute to 100.0 mL with methanol R.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with methanol R. Dilute 1.0 mL of this solution to 25.0 mL with methanol R.

Column:
— size: $l = 0.10$ m, $\phi = 4.6$ mm,
— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (3 μ m),
— temperature: 35 °C.

Mobile phase:
— mobile phase A: 17.0 g/L solution of tetrabutylammonium hydrogen sulfate R1,
— mobile phase B: acetonitrile R,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	90 → 30	10 → 70
15 - 17	30	70

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 220 nm.

Injection 10 μ L.
Relative retention With reference to lopinavir hydrochloride (retention time = about 7 min): impurity A = about 0.9, impurity B = about 1.11; impurity C = about 1.13.
System suitability: reference solution (a):
— resolution: minimum 3.8 between the peaks due to lopinavir hydrochloride and impurity A.

Limits:
— impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent),
— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent),
— total: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent),
— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (2.5.12)
3.4 per cent to 4.2 per cent, determined on 0.500 g.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.350 g in 50 mL of a mixture of 1 volume of anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R. Titrate with 0.1 M perchloric acid using 0.2 mL of naphtholbenzenesulfonic acid R as indicator.
1 mL of 0.1 M perchloric acid is equivalent to 49.30 mg of $C_{29}H_{33}ClN_2O_3$.

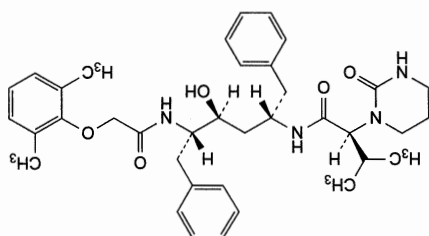
IMPURITIES

In an airtight container, protected from light.

Specified impurities: A, B, C.

Lopinavir

(Ph. Eur. monograph 2615)



Action and use
Protease inhibitor; antiviral (HIV).

$C_{27}H_{48}N_4O_5$

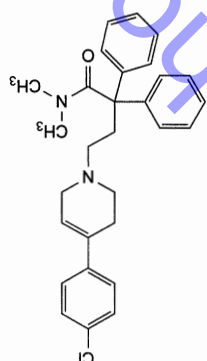
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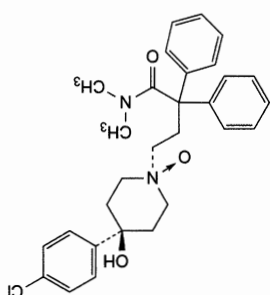


Ph Eur

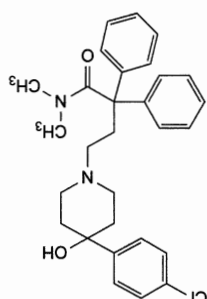
C. 4-[4-(4-chlorophenyl)-3,6-dihydropyridin-1(2H)-yl]-N,N-dimethyl-2,2-diphenylbutanamide



B. 4-[4-(4-chlorophenyl)-4-hydroxy-1-oxopiperidin-1-yl]-N,N-dimethyl-2,2-diphenylbutanamide



A. 4-[4-(4-chlorophenyl)-4-hydroxypiperidin-1-yl]-N,N-dimethyl-2,2-diphenylbutanamide (lopinavir)



Ph Eur

DEFINITION

(2*S*)-*N*-[[(1*S*,3*S*,4*S*)-1-*Benzy*l-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-3-methyl-2-[2-oxotetrahydropyrimidin-1(2*H*)-yl]butanamide.

CHARACTERS

Content 98.0 per cent to 102.0 per cent (anhydrous substance).

Appearance

White or yellowish-white, slightly hygroscopic powder.

Solubility

Practically insoluble in water, very soluble in methanol and in

methylethylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison lopinavir CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in methanol *R*, evaporate to dryness and

record new spectra using the residues.

TESTS

Specific optical rotation (2.2.7)

–27.0 to –22.0 (anhydrous substance).

Dissolve 0.200 g in methanol *R* and dilute to 25.0 mL with

the same solvent.

Related substances

A. Liquid chromatography (2.2.29).

Solvent mixture acetonitrile *R*₁, water *R* (50:50 *V/V*).

Phosphate buffer solution Dissolve 0.9 g of dipotassium hydrogen

phosphate *R* and 2.7 g of potassium dihydrogen phosphate *R* in

900 mL of water *R* and mix well. Adjust to pH 6.0 with

phosphoric acid *R*, dilute to 1000 mL with water *R* and filter.

Test solution (a) Dissolve 50.0 mg of the substance to be

examined in the solvent mixture and dilute to 100.0 mL with

the solvent mixture.

Test solution (b) Dilute 5.0 mL of test solution (a) to

100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of lopinavir CRS in

the solvent mixture and dilute to 100.0 mL with the solvent

mixture. Dilute 5.0 mL of the solution to 100.0 mL with the

solvent mixture.

Reference solution (b) Dilute 5.0 mL of test solution (b) to

250.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2.5 mg of lopinavir for system

suitability CRS (containing impurities A, B, C, F, G, I, N,

Q, R, S and T) in the solvent mixture and dilute to 5.0 mL

with the solvent mixture.

Reference solution (d) Dissolve 2.5 mg of lopinavir for peak

identification CRS (containing impurities D and O) in the

solvent mixture and dilute to 5.0 mL with the solvent

mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography *R* (4 μ m);

— temperature: 50 °C.

Mobile phase:

— mobile phase *A*: acetonitrile *R*₁, phosphate buffer

solution (45:55 *V/V*);

— mobile phase *B*: phosphate buffer solution, acetonitrile *R*₁ (25:75 *V/V*);

Time	Mobile phase A	Mobile phase B
(min)	(per cent <i>V/V</i>)	(per cent <i>V/V</i>)
0 - 60	100	0
60 - 61	100 → 0	0 → 100
61 - 81	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 μ L of test solution (a) and reference

solutions (b), (c) and (d).

Identification of impurities Use the chromatogram supplied

with lopinavir for system suitability CRS and the chromatogram

obtained with reference solution (c) to identify the peaks due

to impurities A, B, C, F, G, I and N; use the chromatogram

supplied with lopinavir for peak identification CRS and the

chromatogram obtained with reference solution (d) to

identify the peak due to impurity D.

Relative retention *r* (not *r*_G) with reference to lopinavir

(retention time = about 37 min): impurity A = about 0.03;

impurity B = about 0.07; impurity C = about 0.10;

impurity D = about 0.13; impurity F = about 0.59;

impurity G = about 0.62; impurity I = about 1.1;

impurity N = about 1.4.

System suitability: reference solution (c):

— resolution: minimum 1.5 between the peaks due to

impurities F and G.

Calculation of percentage contents:

— for impurity A, multiply the peak area by the

correction factor 1.6;

— for impurity B, multiply the peak area by the

correction factor 1.3;

— for impurity C, multiply the peak area by the

correction factor 1.5;

— for impurity D, multiply the peak area by the

correction factor 1.3;

— for each impurity, use the concentration of lopinavir in

reference solution (b).

Limits:

— impurities B, I: for each impurity, maximum

0.2 per cent;

— impurities A, C, D, F, G: for each impurity, maximum

0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— reporting threshold: 0.05 per cent; disregard any peak

eluting after impurity N.

B. Liquid chromatography (2.2.29) as described in test A for

related substances with the following modifications.

Mobile phase Mobile phase A, mobile phase B (30:70 *V/V*).

Run time 8.3 times the retention time of lopinavir.

Identification of impurities Use the chromatogram supplied

with lopinavir for system suitability CRS and the chromatogram

obtained with reference solution (c) to identify the peaks due

to impurities Q, R, S and T; use the chromatogram supplied

with lopinavir for peak identification CRS and the

chromatogram obtained with reference solution (d) to

identify the peak due to impurity O.

Relative retention *r* (not *r*_G) with reference to lopinavir

(retention time = about 6 min): impurity N = about 1.4;

impurity O = about 1.5; impurity Q = about 4.4;
impurity R = about 6.0; impurity S = about 7.1;
impurity T = about 8.5.

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to impurities S and T.

Calculation of percentage contents:

— for impurity O, multiply the peak area by the correction factor 1.3;
— for impurity Q, multiply the peak area by the correction factor 0.7;
— for each impurity, use the concentration of lopinavir in reference solution (b).

Limits:

— impurities O, Q, R, T: for each impurity, maximum 0.15 per cent;
— unspecified impurities: for each impurity, maximum 0.10 per cent;
— reporting threshold: 0.05 per cent; disregard any peak eluting before and including impurity N;
— total of all impurities eluting before and including impurity N in test A and after impurity N in test B: maximum 0.7 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture water R, ethanol (96 per cent) R (5:95 V/V).

0.25 g complies with test H. Prepare the reference solution using 0.25 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 4.4 per cent, determined on 0.250 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in test A for related substances with the following modifications.

Mobile phase Mobile phase A.

Injection Test solution (b) and reference solution (a).

Run time 1.6 times the retention time of lopinavir.

Calculate the percentage content of $C_{37}H_{48}N_4O_5$ taking into account the assigned content of lopinavir CRS.

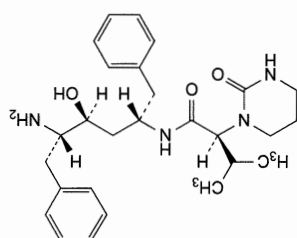
STORAGE

In an airtight container.

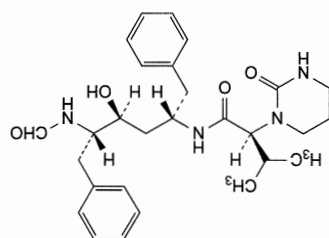
IMPURITIES

Specified impurities A, B, C, D, F, G, I, O, Q, R, T

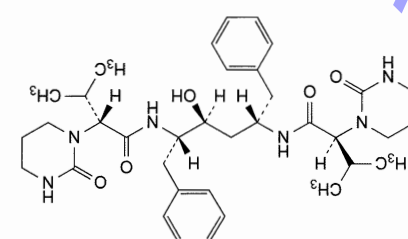
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, H, J, K, L, M, N, P, S.



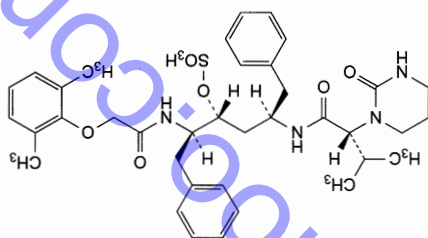
A. (2S)-N-[(1S,3S,4S)-1-benzyl-4-amino-3-hydroxy-5-phenylpentyl-2-[2-oxotetrahydropyrimidin-1(2H)-yl]butanamide,



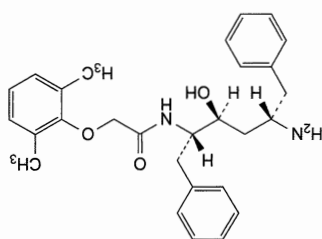
B. (2S)-N-[(1S,3S,4S)-1-benzyl-4-(formylamino)-3-hydroxy-5-phenylpentyl-2-[2-oxotetrahydropyrimidin-1(2H)-yl]butanamide,



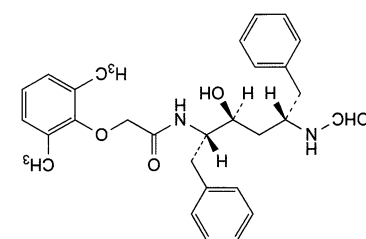
C. (2R)-N-[(1S,2S,4S)-1-benzyl-2-hydroxy-4-[(2S)-3-methyl-2-[2-oxotetrahydropyrimidin-1(2H)-yl]butanoyl]amino]-5-phenylpentyl-3-methyl-2-[2-oxotetrahydropyrimidin-1(2H)-yl]butanamide,



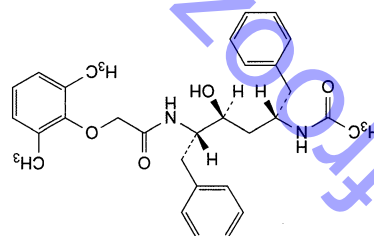
D. (1R,3R)-1-[(1R)-1-[(2-2,6-dimethylphenoxy)acetyl]amino]-2-phenylethyl-3-[(2R)-3-methyl-2-[2-oxotetrahydropyrimidin-1(2H)-yl]butanoyl]amino]-4-phenylbutyl hydrogen sulfate,



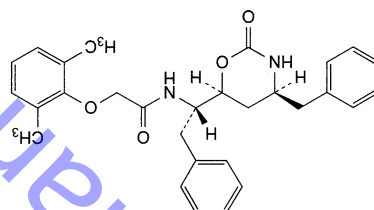
E. N-[(1S,2S,4S)-4-amino-1-benzyl-2-hydroxy-5-phenylpentyl-2-(2,6-dimethylphenoxy)acetamide,



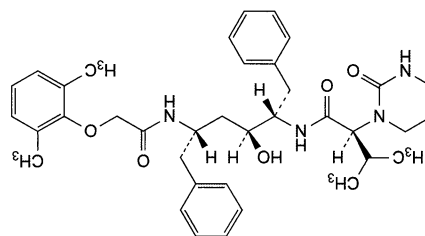
F. N-[(1S,2S,4S)-1-benzyl-4-(formylamino)-2-hydroxy-5-phenylpentyl]-2-(2,6-dimethylphenoxy)acetamide,



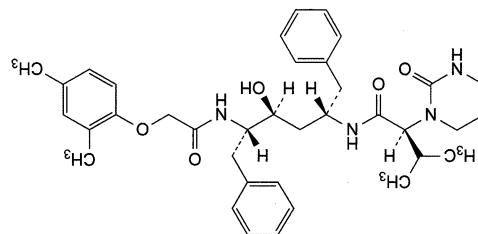
G. N-[(1S,2S,4S)-4-acetyl(1-amino)-1-benzyl-2-hydroxy-5-phenylpentyl]-2-(2,6-dimethylphenoxy)acetamide,



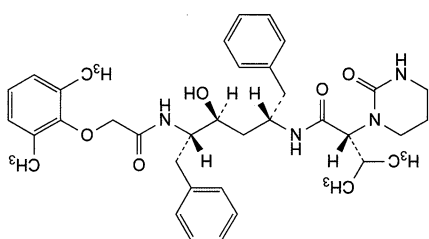
H. N-[(1S)-1-[(4S,6S)-4-benzyl-2-oxo-1,3-oxazinan-6-yl]-2-phenylethyl]-2-(2,6-dimethylphenoxy)acetamide,



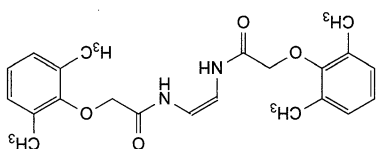
I. (2S)-N-[(1S,2S,4S)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanamide,



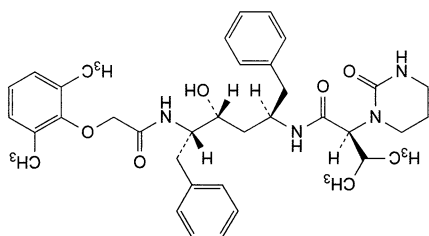
J. (2S)-N-[(1S,3S,4S)-1-benzyl-4-[[2-(2,4-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-3-methyl-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanamide,



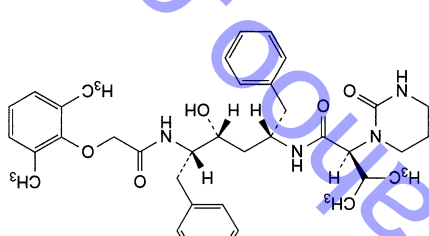
K. (2R)-N-[(1S,3S,4S)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-3-methyl-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanamide,



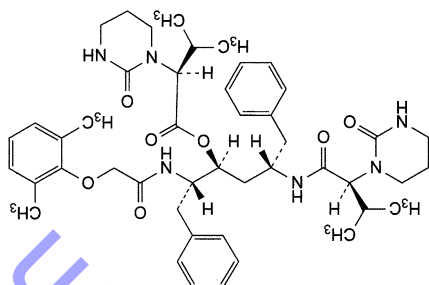
L. N,N'-(Z)-ethene-1,2-diylbis[2-(2,6-dimethylphenoxy)acetamide],



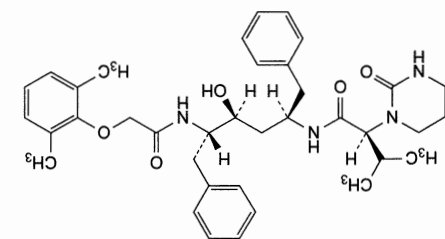
M. (2S)-N-[(1R,3R,4S)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-3-methyl-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanamide,



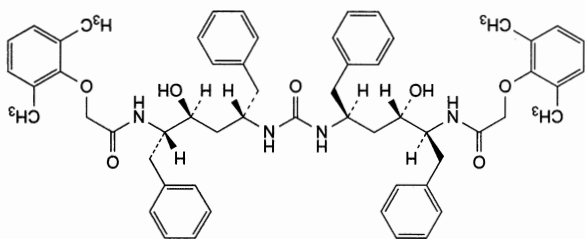
N. (2S)-N-[(1S,3R,4S)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-3-methyl-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanamide,



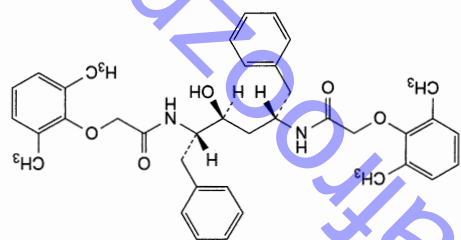
O. (1S,3S)-1-1-[[2-(2,6-dimethylphenoxy)acetyl]amino]-2-phenylethyl]-3-[[[(2S)-3-methyl-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanoyl]amino]-4-phenylbutyl] (2S)-3-methyl-2-(2-oxotetrahydropyrimidin-1(2H)-yl)butanoate,



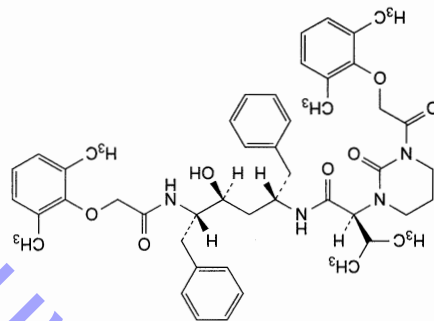
P. (2*S*)-*N*-[(1*R*,3*S*,4*S*)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-3-methyl-2-[2-oxotetrahydropyrimidin-1(2*H*)-yl]butanamide,



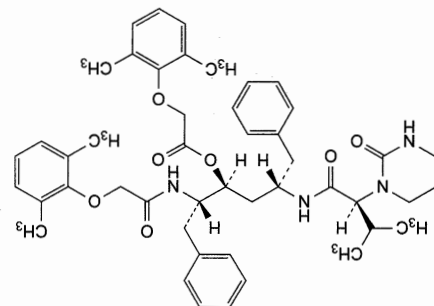
T. N, N'-bis[(1S,3S,5S)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]urea.



Q. N-[(1S,2S,4S)-1-benzyl-4-[[2-(2,6-dimethylphenoxy)acetyl]amino]-2-hydroxy-5-phenylpentyl]-2-(2,6-dimethylphenoxy)acetamide;

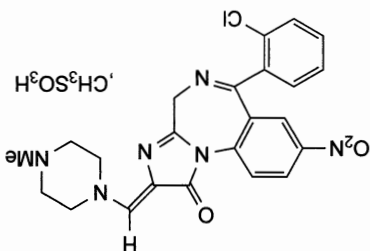


R. (2S)-N-[(1S,3S,4S)-1-benzyl-4-[(2,6-dimethylphenoxy)acetyl]amino]-3-hydroxy-5-phenylpentyl]-2-[2-(2,6-dimethylphenoxy)acetyl]-2-oxotetrahydropyrimidin-1-(2H)-yl]-3-methylbutanamide,



S. (1S,3S)-1-[(1S)-1-[2-(2,6-dimethylphenoxy)acetyl]amino]-2-phenylethyl]-3-[[[(1S)-3-methyl-2-[2-oxotetrahydropyrimidin-1(2H)-yl]butanoyl]amino]-4-phenylbutyl 2-(2,6-dimethylphenoxy)acetate,

Loprazolam Mesilate


$$\text{C}_{23}\text{H}_{21}\text{ClN}_6\text{O}_3, \text{CH}_4\text{O}_3\text{S}, \text{H}_2\text{O} \quad 579.1$$

Action and use

Preparation

Loprazolam Tablets

DEFINITION

Loprazolam Mesilate is (2)-(6-(2-chlorophenyl)-2,4-dihydro-2,4-methylenepiperazin-1-yl)methyl)-8-nitroimidazo [1,2-a] [1,4] benzodiazepin-1-one methanesulfonate monohydrate. It contains not less than 98.5% and not more than 101.0% of $C_{23}H_{21}ClN_6O_5 \cdot CH_4O_3$, calculated with reference to the dried substance.

CHARACTERISTICS

A yellow, crystalline powder. Slightly soluble in water and in ethanol (96%); very slightly soluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of loperzalam mesilate (RS 205).
B. The light absorption, Appendix II B, in the range 210 to 370 nm of a 0.001% w/v solution in ethanol (96%) exhibits a maximum at 330 nm and a shoulder at 240 nm. The absorbance at the maximum is about 0.7.

TESTS

Carry out the method for *thin-layer chromatography*, Appendix III A, using the following solutions in a mixture of equal volumes of *chloroform* and *methanol*.

(1) 2.0% w/v of the substance being examined.

(2) 0.0050% w/v of *N-methylpiperazine*.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel (Merck silica gel 60 plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, allow it to dry in air, heat at 110° for 15 minutes and spray with a mixture of 50 volumes

of a 2% w/v solution of potassium iodide and 1 volume of a 10% w/v solution of chloroplatinic(IV) acid.

MOBILE PHASE

2 volumes of 13.5M ammonia, 20 volumes of methanol and 80 volumes of chloroform.

LIMITS

Any spot corresponding to *N*-methylpiperazine in the

chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.25%).

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in a mixture of 8 volumes of water, 46 volumes of chloroform and 46 volumes of methanol.

(1) 2.0% w/v of the substance being examined.

(2) 0.0020% w/v of the substance being examined.

(3) 0.010% w/v of 6-(2-chlorophenyl)-2,4-dihydro-2-[(dimethylamino)methylene]-8-nitroimidazo[1,2-a]

[1,4]benzodiazepin-1-one BPCRS (impurity A).

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel (Merck silica gel 60 plates are suitable). Before use, stand the plate in methanol, allowing the solvent front to ascend 17 cm, allow to dry in air, heat the plate at 100° for 1 hour and use with the flow of mobile phase in the same direction as that used for the pretreatment.

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, allow it to dry in air, spray the plate with a 5% w/v solution of titanium(III) chloride in a solution of hydrochloric acid containing 10% w/v of HCl and then spray with a solution containing 0.4 g of 4-dimethyl-aminocinnamaldehyde in a mixture of 20 mL of 6M hydrochloric acid and 100 mL of ethanol (96%). Heat at 100° until spots appear (about 10 minutes).

MOBILE PHASE

20 volumes of methanol and 80 volumes of chloroform.

LIMITS

Any spot corresponding to impurity A in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (3) (0.5%) and any other secondary spot is not more intense than the spot in the chromatogram obtained with solution (2) (0.1%).

Loss on drying

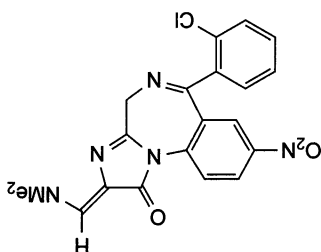
When dried at 100° to 105° for 3 hours, loses 2.5 to 4.5% of its weight. Use 1 g.

Sulfated ash

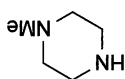
Not more than 0.1%, Appendix IX A.

IMPURITIES

ASSAY
Dissolve 0.25 g in 60 mL of a 50% v/v solution of propan-2-ol and titrate with 0.05M sodium hydroxide VS determining the end point potentiometrically. Each mL of 0.05M sodium hydroxide VS is equivalent to 28.05 mg of $C_{23}H_{21}ClN_6O_3 \cdot CH_4O_3S$.



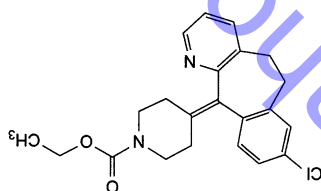
A. 6-(2-chlorophenyl)-2,4-dihydro-2-[(dimethylamino)methylene]-8-nitroimidazo[1,2-a][1,4]benzodiazepin-1-one; dimethylamino analogue.



B. *N*-methylpiperazine.

Loratadine

(Ph. Eur. monograph 2124)



$C_{22}H_{23}ClN_2O_2$ 382.9

79794-75-5

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparation

Loratadine Tablets

DEFINITION

Edyl 4-(8-chloro-5,6-dihydro-1*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridin-1-ylidene)piperidine-1-carboxylate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone and in methanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).



Comparison loratadine CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *acetone* R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II). Dissolve 1.0 g in *methanol* R and dilute to 20.0 mL with the same solvent.

Impurity H

Gas chromatography (2.2.28).

Internal standard solution Dissolve 25 mg of *isoamyl benzoate* R in *methylethyl chloride* R and dilute to 100 mL with the same solvent. Dilute 5.0 mL of this solution to 50 mL with *methylethyl chloride* R.

Test solution Dissolve 25.0 mg of the substance to be examined in *methylethyl chloride* R, add 1.0 mL of reference solution (a) and 1.0 mL of the internal standard solution and dilute to 5.0 mL with *methylethyl chloride* R.

Reference solution (a) Dissolve 25.0 mg of *loratadine* impurity H CRS in *methylethyl chloride* R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 50.0 mL with *methylethyl chloride* R.

Reference solution (b) To 1.0 mL of reference solution (a) add 1.0 mL of the internal standard solution and dilute to 5.0 mL with *methylethyl chloride* R.

Column:
— material: fused silica;
— size: $l = 25$ m, $\varnothing = 0.32$ mm;
— stationary phase: poly(dimethyl)siloxane R (film thickness 0.52 μ m).

Carrier gas helium for chromatography R.
Flow rate 1.0 mL/min.
Split ratio 1:30.
Temperature:

Temperature (°C)	Time (min)	Column
80	0 - 1	23 - 33
80 → 300	1 - 23	
300	23 - 33	260
300	33 - 300	
300	300 - 330	300

Detection Flame ionisation.

Injection 1 μ L of the test solution and reference solution (b).
Relative retention With reference to *loratadine* (retention time = about 32 min): impurity H = about 0.33; *isoamyl benzoate* = about 0.37.
System suitability: reference solution (b):
— resolution: minimum 2.0 between the peaks due to impurity H and *isoamyl benzoate*;
— signal-to-noise ratio: minimum 10 for the peak due to impurity H.

Limit:

— **impurity H:** calculate the ratio (*R*) of the area of the peak due to impurity H to the area of the peak due to *isoamyl benzoate* from the chromatogram obtained with reference solution (b); from the chromatogram obtained with the

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of *loratadine* impurity F CRS in the mobile phase and dilute to 25 mL with the mobile phase. Dilute 1 mL of this solution to 10 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of *loratadine* for system suitability CRS (containing impurities A and E) in the mobile phase, add 0.5 mL of reference solution (a) and dilute to 5 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m) with very low silanol activity; — temperature: 40 °C.

Mobile phase Mix 30 volumes of *methanol* R, 35 volumes of a 6.8 g/L solution of *potassium dihydrogen phosphate* R previously adjusted to pH 2.80 \pm 0.05 with *phosphoric acid* R and 40 volumes of *acetonitrile* R.

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 220 nm.
Injection 20 μ L of the test solution and reference solutions (b) and (c).

Run time 5 times the retention time of *loratadine*.
Identification of impurities Use the chromatogram supplied with *loratadine* for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and E.

Relative retention With reference to *loratadine* (retention time = about 12 min): impurity D = about 0.2; impurity B = about 0.4; impurity F = about 0.9; impurity E = about 1.1; impurity A = about 2.4; impurity C = about 2.7.

System suitability: reference solution (b):
— peak-to-valley ratio: minimum 2.5, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to *loratadine*.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.7; impurity F = 1.6; impurity E = 1.9;
— **impurity F:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— **impurities A, B, C, D, E:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);
— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent).

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Sulfates (2.4.13)

Maximum 150 ppm.

Ignite 1.33 g at $800 \pm 25^\circ\text{C}$ and take up the residue with

20 mL of *distilled water* R. Filter, if necessary, through paper

free from sulfates. Repeat the titration with new paper filters

until the filtrate is no longer turbid.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105°C .

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of *glacial acetic acid* R. Titrate

with 0.1 M *perchloric acid*, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M *perchloric acid* is equivalent to 38.29 mg

of $\text{C}_{22}\text{H}_{23}\text{ClN}_2\text{O}_2$.

IMPURITIES

Specified impurities A, B, C, D, E, F, H.

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

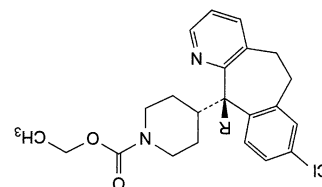
acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

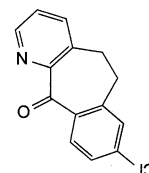
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: G.



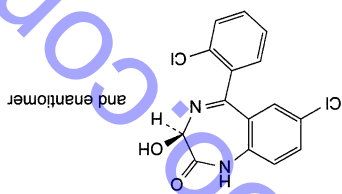
A. R = OH: ethyl 4-[(1*RS*)-8-chloro-11-hydroxy-6,11-dihydro-5*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridin-1-yl]piperidine-1-carboxylate,
F. R = F: ethyl 4-[(1*RS*)-8-chloro-11-fluoro-6,11-dihydro-5*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridin-1-yl]piperidine-1-carboxylate,
B. 8-chloro-5,6-dihydro-11*H*-benzo[5,6]cyclohepta[1,2-*b*]pyridin-1-one, carboxylate,



pyridin-1-one,

Lorazepam

(Ph. Eur. monograph 1121)



$\text{C}_{22}\text{H}_{23}\text{ClN}_2\text{O}_2$

321.2

846-49-1

Action and use

Benzydiazepine.

Preparations

Lorazepam Injection

Lorazepam Oral Solution

Lorazepam Tablets

DEFINITION

(3*RS*)-7-Chloro-5-(2-chlorophenyl)-3-hydroxy-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Content

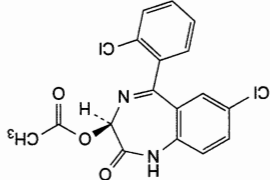
98.5 per cent to 102.0 per cent (dried substance).

Ph Eur



impurity D and lorazepam;

B. (3*RS*)-7-chloro-5-(2-chlorophenyl)-2-oxo-2,3-dihydro-1*H*-1,4-benzodiazepin-3-yl acetate,

Nc1cc(Cl)ccc1C(=O)c2cc(Cl)ccc232.12 mg of $C_{15}H_{10}Cl_2N_2O_2$.

— *peak-to-valley ratio*: minimum 5.0, where H_p = height

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0-5	80	20
5-35	80 \rightarrow 30	20 \rightarrow 70
35-50	30	70
50-60	30 \rightarrow 80	70 \rightarrow 20

IDENTIFICATION

- A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of lormetazepam (RS 207).
- B. In the test for Related substances, the chromatogram obtained with solution (2) shows a peak with the same retention time as the principal peak in the chromatogram obtained with solution (4).

TESTS**Related substances**

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in methanol (70%).

- (1) 0.25% w/v of the substance being examined.
- (2) 0.00025% w/v of the substance being examined.
- (3) 0.00025% w/v of the substance being examined.
- (4) 0.0005% w/v of lormetazepam BPCRS.
- (5) 0.00025% w/v each of lormetazepam BPCRS and lorazepam BPCRS.

CHROMATOGRAPHIC CONDITIONS

- (a) Use a stainless steel column (20 cm × 4.6 mm) packed with end-capped octadecylsilyl silica gel for chromatography (5 µm) (Hypersil ODS is suitable).
- (b) Use isocratic elution and the mobile phase described below.
- (c) Use a flow rate of 2 mL per minute.
- (d) Use an ambient column temperature.
- (e) Use a detection wavelength of 230 nm.
- (f) Inject 20 µL of each solution.

MOBILE PHASE

48 volumes of methanol and 52 volumes of a phosphate buffer prepared by dissolving 4.91 g of sodium dihydrogen orthophosphate and 0.633 g of disodium hydrogen orthophosphate in sufficient water to produce 1000 mL.

SYSTEM SUITABILITY

The test is not valid unless the resolution factor between the two principal peaks in the chromatogram obtained with solution (5) is at least 4.

LIMITS

In the chromatogram obtained with solution (1):

the area of any secondary peak is not greater than that of the principal peak in the chromatogram obtained with solution (2) (0.2%);

not more than two such peaks have an area greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.1%);

the sum of the areas of all such peaks is not greater than 2.5 times the area of the principal peak obtained with solution (2) (0.5%).

Loss on drying

When dried to constant weight at 105° for 3 hours, loses not more 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.5 g in 50 mL of nitroethane and carry out Method I for non-aqueous titration, Appendix VIII A, determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 33.52 mg of $C_{16}H_{12}Cl_2N_2O_2$.

Lormetazepam
 $C_{16}H_{12}Cl_2N_2O_2$

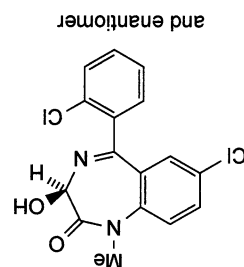
335.2

848-75-9

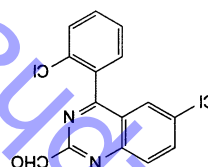
Lormetazepam Tablets

Preparation

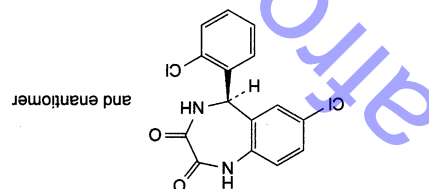
Benzodiazepine.

Action and use

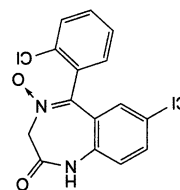
E. 6-chloro-4-(2-chlorophenyl)quinazoline-2-carbaldehyde.



D. (5RS)-7-chloro-5-(2-chlorophenyl)-4,5-dihydro-1H-1,4-benzodiazepine-2,3-dione,



C. 7-chloro-5-(2-chlorophenyl)-1,3-dihydro-2H-1,4-benzodiazepin-2-one 4-oxide,

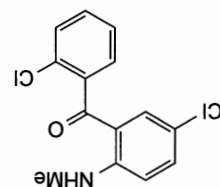
**CHARACTERISTICS**

A white, crystalline powder.

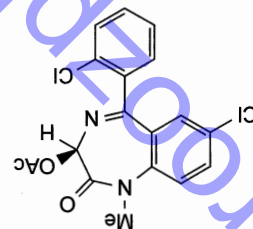
Practically insoluble in water; soluble in ethanol (96%) and in methanol.

STORAGE

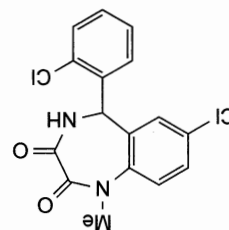
Lormetazepam should be protected from light.

IMPURITIES

A. 2-methylamino-2',5'-dichlorobenzophenone,



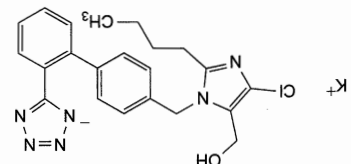
B. O'-acetyl-lormetazepam,



C. 7-chloro-1-methyl-5-(2-chlorophenyl)-4,5-dihydro-2H-1,4-benzodiazepin-2,3-(1H)-dione.

Losartan Potassium

(Ph. Eur. monograph 2232)



C₂₂H₂₂ClKNO₆

461.0

124750-99-8

**Preparation**

Losartan Potassium Tablets

Action and use

Angiotensin II (AT₁) receptor antagonist.

DEFINITION

Potassium 5-[4'-[[2-butyl-4-chloro-5-(hydroxymethyl)-1H-imidazol-1-yl]methyl]biphenyl-2-yl]tetrazol-1-ide.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder, hygroscopic.

Solubility

Freely soluble in water and in methanol, slightly soluble in

acetone/nitrite.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison losartan potassium CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in methanol R, evaporate to dryness and

record new spectra using the residues.

B. Dissolve 25 mg in 3 mL of water R. The solution gives

TESTS**Related substances**

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Test solution Dissolve 30.0 mg of the substance to be

examined in methanol R and dilute to 100.0 mL with the

same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with methanol R. Dilute 1.0 mL of this solution to

10.0 mL with methanol R.

Reference solution (b) Dissolve 6 mg of triphenylmethanol R

(impurity G) in 100.0 mL of methanol R. Dilute 1.0 mL of

the solution to 100.0 mL with methanol R. Use 1.0 mL of

this solution to dissolve the contents of a vial of losartan for

system suitability CRS (containing impurities J, K, L and M)

and sonicate for 5 min.

Reference solution (c) Dissolve 3.0 mg of losartan

impurity D CRS in methanol R and dilute to 100.0 mL with

the same solvent. Dilute 1.5 mL of this solution to 100.0 mL

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 µm);

— temperature: 35 °C.

Mobile phase:

— mobile phase A: dilute 1.0 mL of phosphoric acid R to

1000 mL with water R;

— mobile phase B: acetonitrile R₁;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 5	75	25
5 - 30	75 → 10	25 → 90
30 - 40	10	90

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 220 nm.

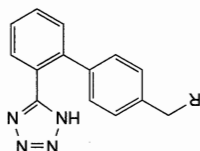
Injection 10 µL.

Identification of impurities Use the chromatogram supplied with losartan for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities G, J, K, L and M; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity D.

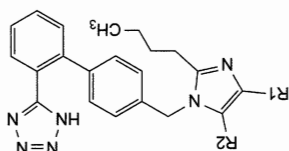
IMPURITIES

Specified impurities D, J, K, L, M

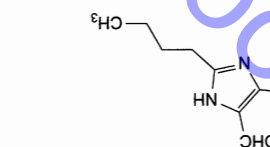
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, E, F, G, H, I



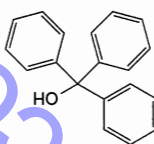
B. R = OH; [2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methanol, E. R = H; 5-(4-methylbiphenyl-2-yl)-1H-tetrazole,



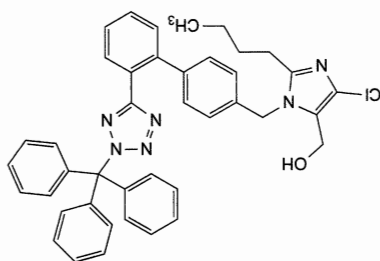
C. R1 = CH₂-OH, R2 = Cl; [2-butyl-5-chloro-1-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methoxy]-1H-imidazole-4-yl]methanol, F. R1 = Cl, R2 = CH₂-O-CH(CH₃)₂; 5-[4'-[[2-butyl-4-chloro-5-[[[(1-methylethyl)oxy]methyl]-1H-imidazol-1-yl]methyl]biphenyl-2-yl]-1H-tetrazole], I. R1 = Cl, R2 = CH₂-O-CPh₃; 5-[4'-[[2-butyl-4-chloro-5-[[[(triphenylmethyl)oxy]methyl]-1H-imidazol-1-yl]methyl]biphenyl-2-yl]-1H-tetrazole],



D. 2-butyl-4-chloro-1H-imidazole-5-carbaldehyde,



G. triphenylmethanol,



H. [2-butyl-4-chloro-1-[[2'-(2-(triphenylmethyl)-2H-tetrazol-5-yl]biphenyl-4-yl]methoxy]-1H-imidazol-5-yl]methanol,

Limits:

With reference to losartan (retention time = about 14 min): impurity D = about 0.9; impurity J = about 1.4; impurity K = about 1.5; impurity L = about 1.6; impurity M = about 1.75; impurity G = about 1.8. System suitability: reference solution (b): peak-to-valley ratio: minimum 2.0, where H_p = height above the baseline of the peak due to impurity M and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity G.

impurity D: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.15 per cent); impurities J, K, L, M: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent); unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent); total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent); disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals

Maximum 20 ppm. Prescribed solution Dissolve 1.0 g in 20 mL of a mixture of equal volumes of ethanol (96 per cent) R and water R. Test solution 12 mL of the prescribed solution. Reference solution Mix 1.0 mL of lead standard solution (10 ppm Pb) R, 2.0 mL of the prescribed solution and 9 mL of water R. Blank solution Mix 2.0 mL of the prescribed solution and 10 mL of water R.

To each solution, add 2 mL of buffer solution pH 3.5 R. Mix. The substance will precipitate. Dilute each solution to 40 mL with ethanol (96 per cent) R. The substance dissolves completely. Mix and add to 1.2 mL of thiocetamide reagent R. Mix immediately.

Filter the solutions through a membrane filter (nominal pore size 0.45 µm) (2.4.8). Compare the spots on the filters obtained with the different solutions. The test is invalid if the reference solution does not show a slight brownish-black colour compared to the blank solution. The substance to be examined complies with the test if the brownish-black colour of the spot resulting from the test solution is not more intense than that of the spot resulting from the reference solution.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

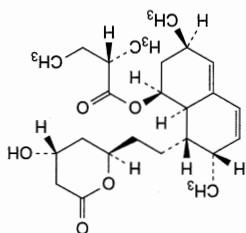
Dissolve 0.200 g in 75 mL of anhydrous acetic acid R and sonicate for 10 min. Carry out a potentiometric titration (2.2.20) using 0.1 M perchloric acid. 1 mL of 0.1 M perchloric acid is equivalent to 23.05 mg of C₂₂H₂₂ClKN₆O.

STORAGE

In an airtight container.

Lovastatin

(Ph. Eur Monograph 1538)



C₂₄H₃₆O₅ 404.5 75330-75-5

Action and use

HMG Co-A reductase inhibitor; lipid-regulating drug.

DEFINITION

(1S,3R,7S,8S,8aR)-8-[2-[(2R,4R)-4-Hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,7,8,8a-hexahydronaphthalen-1-yl (2S)-2-methylbutanoate.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone, sparingly soluble in anhydrous ethanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison lovastatin CRS.

TESTS

Specific optical rotation (2.2.7)

+ 325 to + 340 (dried substance).

Dissolve 0.125 g in acetonitrile R and dilute to 25.0 mL with the same solvent.

Impurity E

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in acetonitrile R1 and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with acetonitrile R1. Dilute 5.0 mL of this solution to 50.0 mL with acetonitrile R1.

Reference solution (b) Dissolve 4 mg of lovastatin for peak identification CRS (containing impurities A, B, C, D, E and F) in acetonitrile R1 and dilute to 10.0 mL with the same solvent.

Column:

size: l = 0.25 m, Ø = 4.6 mm;

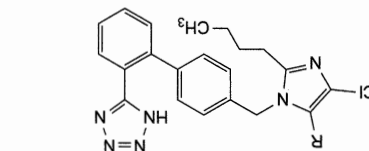
stationary phase: octylsilyl silica gel for chromatography R

temperature: 40 °C.

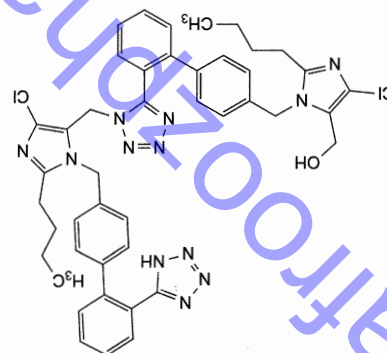
Mobile phase Mix 7 volumes of a 1.1 g/L solution of phosphoric acid R and 13 volumes of acetonitrile R1.

Flow rate 1.5 mL/min.

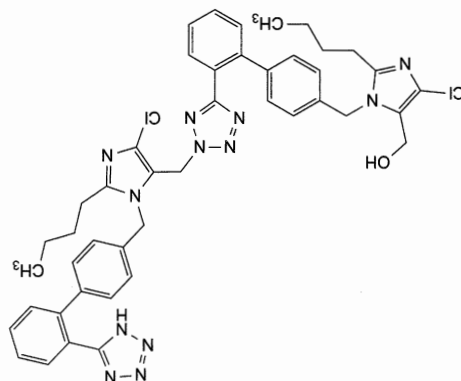
Detection Spectrophotometer at 200 nm.



J. R = CH₂O-CO-CH₃; [2-butyl-4-chloro-1-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]-1H-imidazol-5-yl]methyl acetate,
K. R = CHO; 2-butyl-4-chloro-1-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]-1H-imidazol-5-carbaldehyde,



L. [2-butyl-1-[[2'-(1-[[2-butyl-4-chloro-1-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]-1H-imidazol-5-yl]methyl]-1H-tetrazol-5-yl]biphenyl-4-yl]methyl]-4-chloro-1H-imidazol-5-yl]methanol,



M. [2-butyl-1-[[2'-(2-[[2-butyl-4-chloro-1-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]-1H-imidazol-5-yl]methyl]-2H-tetrazol-5-yl]biphenyl-4-yl]methyl]-4-chloro-1H-imidazol-5-yl]methanol.

Ph Eur

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to lovastatin.

Limits:

— **impurities A, B, C, D:** for each impurity, not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b)

— **impurity F:** not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— **reference solution (b)** (0.15 per cent);

— **unspecified impurities:** for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b)

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **disregard limit:** 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);

— **heavy metals** (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in a desiccator under high vacuum at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection: Test solution and reference solution (a).

Calculate the content of $C_{24}H_{36}O_5$ from the declared content of lovastatin CRS.

STORAGE

Under nitrogen, at a temperature of 2 °C to 8 °C.

IMPURITIES

Specified impurities A, B, C, D, E, F

A. (1S,7S,8S,8aR)-8-[2-[(2R,4R)-4-hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-7-methyl-1,2,3,7,8,8a-hexahydronaphthalen-1-yl [(2S)-2-methylbutanoate (mevastatin),

B. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

C. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

D. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

E. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

F. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

G. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

H. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

I. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

J. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

K. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

L. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

M. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

N. (3R,5R)-7-[(1S,2S,5,6R,8S,8aR)-2,6-dimethyl-8-[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (hydroxyacid lovastatin),

Injection 10 µL.

Run time 3 times the retention time of lovastatin.

Identification of impurities Use the chromatogram supplied

with lovastatin for peak identification CRS and the chromatogram obtained with reference solution (b) to

identify the peak due to impurity E.

Relative retention With reference to lovastatin (retention time = about 5 min): impurity E = about 1.3.

System suitability: reference solution (b):

— **resolution:** minimum 5.0 between the peaks due to lovastatin and impurity E.

Limit:

— **correction factor:** for the calculation of content, multiply the peak area of impurity E by 1.6;

— **impurity E:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in acetonitrile R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 20.0 mg of lovastatin CRS in acetonitrile R and dilute to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with acetonitrile R. Dilute 5.0 mL of this solution to 50.0 mL with acetonitrile R.

Reference solution (c) Dissolve 4 mg of lovastatin for peak identification CRS (containing impurities A, B, C, D, E and F) in acetonitrile R and dilute to 10.0 mL with the same solvent.

Column:

— **size:** $l = 0.25$ m, $\phi = 4.6$ mm;

— **stationary phase:** octylsilyl silica gel for chromatography R (5 µm).

Mobile phase:

— **mobile phase A:** 0.1 per cent V/V solution of phosphoric acid R;

— **mobile phase B:** acetonitrile R;

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 238 nm.

Injection 10 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with lovastatin for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D and F.

Relative retention With reference to lovastatin (retention time = about 7 min): impurity B = about 0.6;

impurity A = about 0.8; impurity F = about 0.9;

impurity C = about 1.6; impurity D = about 2.3;

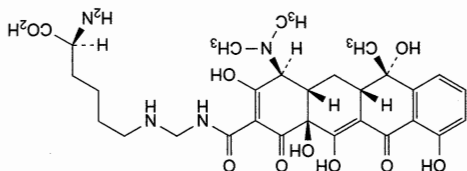
System suitability: reference solution (c):

— **peak-to-valley ratio:** minimum 3.0, where H_p = height above the baseline of the peak due to impurity F and



Lymecycline

(Ph. Eur. monograph 1654)



C₂₉H₃₈N₄O₁₀

603

992-21-2

Action and use
Tetracycline antibacterial.
Preparation
Lymecycline Capsules

DEFINITION

(2S)-2-Amino-6-[[[[(4S,4a,5a,5b,6S,12aS)-4-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracen-2-yl]carbonyl]amino]methyl]aminohexanoic acid (reaction product of formaldehyde, lysine and tetracycline).

Semi-synthetic product derived from a fermentation product.

Content

81.0 per cent to 102.0 per cent (equivalent to 60.0 per cent to 75.0 per cent of tetracycline) (anhydrous substance).

CHARACTERS

Appearance
Yellow, hygroscopic powder.

Solubility

Very soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).
Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.
Reference solution (a) Dissolve 5 mg of tetracycline hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 5 mg of tetracycline hydrochloride CRS, 5 mg of demeclocycline hydrochloride R and 5 mg of oxytetracycline hydrochloride R in methanol R and dilute to 10 mL with the same solvent.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R (2-10 µm).
Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a 63 g/L solution of oxalic acid R previously adjusted to pH 2.0 with concentrated ammonia R.

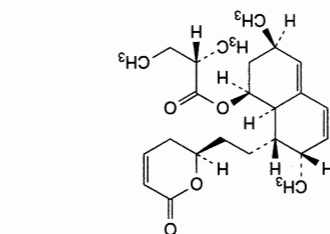
Application 2 µL.

Development Over half of the plate.

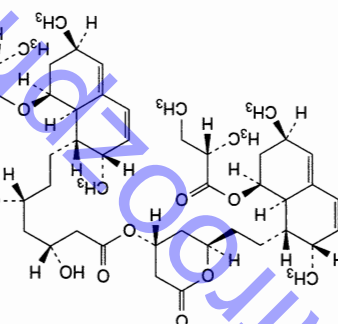
Drying In air.

Detection Examine in ultraviolet light at 254 nm.

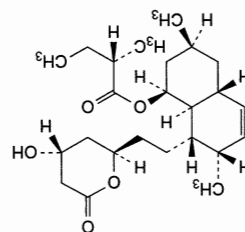
System suitability: reference solution (b):
— the chromatogram shows 3 clearly separated spots.
Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).
B. Thin-layer chromatography (2.2.27).



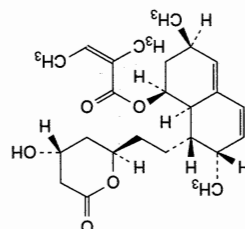
C. (1S,3R,7S,8S,8aR)-3,7-dimethyl-8-[2-[(2R)-6-oxo-3,6-dihydro-2H-pyran-2-yl]ethyl]-1,2,3,7,8,8a-hexahydronaphthalen-1-yl (2S)-2-methylbutanoate (dehydrolovastatin).



D. (2R,4R)-2-[2-[(1S,2S,6R,8S,8aR)-2,6-dimethyl-8-[[[(2S)-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]ethyl]-6-oxotetrahydro-2H-pyran-4-yl] (3R,5R)-7-oxo-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoate (lovastatin dimer),



E. (1S,3S,4aR,7S,8S,8aR)-8-[2-[(2R,4R)-4-hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,4,4a,7,8,8a-octahydronaphthalen-1-yl (2S)-2-methylbutanoate (4,4a-dihydrollovastatin),



F. (1S,3R,7S,8S,8aR)-8-[2-[(2R,4R)-4-hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,7,8,8a-hexahydronaphthalen-1-yl (2Z)-2-methylbut-2-enoate.

Ph Eur

Test solution Dissolve 50 mg of the substance to be examined in 50 mL of water R.

Reference solution (a) Dissolve 10 mg of lysine hydrochloride CRS in water R and dilute to 50 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of arginine CRS and 10 mg of lysine hydrochloride CRS in water R and dilute to 25 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, 2-propanol R (30:70 V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying At 100-105 °C until the ammonia disappears completely.

Detection Spray with ninhydrin solution R and heat at 100-105 °C for 15 min.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.2 g in 5 mL of water R, add 0.3 mL of phosphoric acid R and distil. To 1 mL of the distillate add 10 mL of chromotropic acid-sulfuric acid solution R. A violet colour is produced.

D. Specific optical rotation (see Tests).

TESTS

pH (2.2.3)

7.8 to 8.2.

Dissolve 0.1 g in 10 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7)

–180 to –210 (anhydrous substance).

Dissolve 0.250 g in water R and dilute to 50.0 mL with the same solvent.

Free tetracycline (impurity H)

Maximum 2.5 per cent (anhydrous and methanol-free substance).

To 0.5 g add 50 mL of butyl acetate R and allow to stand at 25 °C for 1 h. Filter and extract the filtrate with 2 quantities, each of 25 mL, of 0.1 M hydrochloric acid. Combine the extracts and dilute to 50.0 mL with 0.1 M hydrochloric acid.

Dilute 10.0 mL of this solution to 100.0 mL with 0.1 M hydrochloric acid. The absorbance (2.2.25) measured at 355 nm is not greater than 0.64.

Light-absorbing impurities

The absorbance (2.2.25) is not greater than 0.50 at 430 nm (anhydrous and methanol-free substance).

Dissolve 25.0 mg in 0.01 M hydrochloric acid and dilute to 10.0 mL with the same acid.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 0.125 g of the substance to be examined in 5.0 mL of water R. Add 1.0 mL of a 40 g/L solution of sodium metabisulfite R and allow to stand in the dark at 20-25 °C for 16-24 h, without stirring. Add 50 mL of 0.05 M hydrochloric acid, shake to dissolve the precipitate and dilute to 100.0 mL with water R.

Reference solution (a) Dissolve 25.0 mg of tetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (b) Dissolve 12.5 mg of 4-epitetracycline hydrochloride CRS (impurity A) in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Reference solution (c) Dissolve 10.0 mg of anhydrotetracycline hydrochloride CRS (impurity C) in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same acid.

Reference solution (d) Dissolve 10.0 mg of 4-epianhydrotetracycline hydrochloride CRS (impurity D) in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Reference solution (e) Mix 1 mL of reference solution (a), 2 mL of reference solution (b) and 5 mL of reference solution (d) and dilute to 25 mL with 0.01 M hydrochloric acid.

Reference solution (f) Mix 40.0 mL of reference solution (b), 20.0 mL of reference solution (c) and 5.0 mL of reference solution (d) and dilute to 200.0 mL with 0.01 M hydrochloric acid.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: styrene-divinylbenzene copolymer R (8 µm) with a pore size of 10 nm;
— temperature: 60 °C.

Mobile phase Weigh 80.0 g of 2-methyl-2-propanol R and transfer to a 1000 mL volumetric flask with the aid of 200 mL of water R; add 100 mL of a 35 g/L solution of dipotassium hydrogen phosphate R adjusted to pH 8.0 with dilute phosphoric acid R; 200 mL of a 10 g/L solution of tetrabutylammonium hydrogen sulfate R adjusted to pH 8.0 with dilute sodium hydroxide solution R; and 10 mL of a 40 g/L solution of sodium edetate R adjusted to pH 8.0 with dilute water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (e) and (f).

Run time 5 times the retention time of the principal peak in the chromatogram obtained with the test solution.

Relative retention With reference to tetracycline (retention time = about 8 min): impurity E = about 0.50;

impurity A = about 0.6; impurity F = about 0.68;

impurity B (eluting on the tail of the principal peak) = about 1.2; impurity D = about 1.45;

impurity G = about 1.45; impurity C = about 2.95.

System suitability: reference solution (e):

— **resolution:** minimum 3.0 between the 1st peak (impurity A) and the 2nd peak (tetracycline) and minimum 5.0 between the 2nd peak and the 3rd peak (impurity D); adjust the concentration of 2-methyl-2-propanol in the mobile phase if necessary;

— **symmetry factor:** maximum 1.25 for the peak due to tetracycline.

Limits:

— **impurity A:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (5.0 per cent),

— **impurity C:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (1.0 per cent),

— impurities B, E, F: for each impurity, not more than 0.1 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (f)

— sum of impurities D and G: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (0.5 per cent),

— any other impurity: for each impurity, not more than 0.04 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (f) (0.2 per cent),

— total: not more than 1.6 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (f) (8.0 per cent),

— disregard limit: 0.02 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (f) (0.1 per cent).

— Methanol (2.4.24, System A) Maximum 1.5 per cent.

Water (2.5.12) Maximum 5.0 per cent, determined on 0.20 g.

Sulfated ash (2.4.14) Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (a).

System suitability: maximum relative standard deviation of 1.0 per cent after 6 injections of reference solution (a).

Calculate the percentage content of tetracycline and multiply it by 1.356 to obtain the percentage content of lynestrenol.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

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Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

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Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

Chemical structures of impurities A, B, C, D, E, F, G, H

D. R1 = H, R2 = H, R3 = N(CH₃)₂: (4R,4aS,12aS)-4-

(dimethylamino)-3,10,11,12a-tetrahydroxy-6-methyl-1,12-

dioxo-1,4,4a,5,12,12a-hexahydro-2-carboxamide

(4-epianhydrotetracycline),

E. unknown structure,

F. unknown structure,

G. (4S,4aS,5aS,6S,12aS)-7-chloro-4-(dimethylamino)-

3,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-

1,4,4a,5,5a,6,11,12a-octahydro-2-carboxamide

(chlorotetracycline),

H. (4S,4aS,5aS,6S,12aS)-4-(dimethylamino)-3,6,10,12,12a-

pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-

octahydro-2-carboxamide (tetracycline).

Ph Eur

Chemical structure of Lynestrenol

Chemical structure of Lynestrenol

Chemical structure of Lynestrenol

Chemical structure of Lynestrenol

Chemical structure of Lynestrenol

Chemical structure of Lynestrenol

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Chemical structure of Lynestrenol

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.2 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

—9.5 to –11 (dried substance).

Dissolve 0.900 g in ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 0.250 g of the substance to be examined in ethyl acetate R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with ethyl acetate R. Dilute 1.0 mL of this solution to 10.0 mL with ethyl acetate R.

Reference solution (b) Dissolve 10 mg of lynestrenol for peak identification CRS (containing impurities A, B and C) in 1.0 mL of ethyl acetate R.

Column:

— material: fused silica;

— size: $l = 50$ m, $\varnothing = 0.32$ mm;

— stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 1.0 μ m).

Carrier gas helium for chromatography R.

Flow rate 3.0 mL/min.

Split ratio 1:34.

Temperature:

Time (min)	Temperature (°C)	Column
0 - 30	80 \rightarrow 230	
30 - 32	230 \rightarrow 310	
32 - 42	310	
Injection port	150	
Detector	300	

Detection Flame ionisation.

Injection 1.0 μ L.

Identification of impurities Use the chromatogram supplied with lynestrenol for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention With reference to lynestrenol (retention time = about 38 min): artefact degradation

peak = about 0.97; impurity A = about 0.99;

impurity B = about 1.005; impurity C = about 1.01.

System suitability: reference solution (b):

— peak-to-valley ratio: minimum 2.5, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to

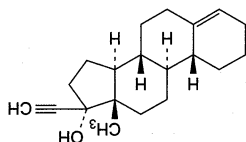
lynestrenol.

Limits:

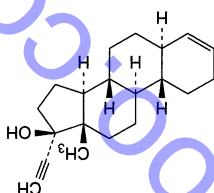
— impurity A: not more than 3 times the area of the principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

B. 19-norpregn-4-en-20-yn-17-ol,



A. 19-nor-5 α ,17 α -pregn-3-en-20-yn-17-ol,



Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*): B.

Specified impurities A, C

IMPURITIES

Protected from light.

STORAGE

$C_{20}H_{28}O$.

1 mL of 0.1 M sodium hydroxide is equivalent to 28.44 mg of

nitrate R as junction liquid. Carry out a blank titration.

and as comparison electrode a silver-silver chloride double-

potentiometrically (2.2.20), using a glass indicator electrode

0.1 M sodium hydroxide. Determine the end-point

5.0 mL of a 100 g/L solution of silver nitrate R. Titrate with

Dissolve 0.150 g in 40 mL of tetrahydrofuran R and add

ASSAY

an oven at 105 °C.

Maximum 0.5 per cent, determined on 0.500 g by drying in

Loss on drying (2.2.32)

be generated in the injection system.

(0.05 per cent). Disregard the artefact peak, which may

the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

solution (a) (1.0 per cent);

peak in the chromatogram obtained with reference

— total: not more than 10 times the area of the principal

with reference solution (a) (0.10 per cent);

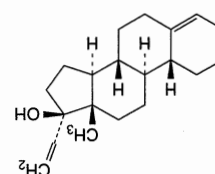
area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

solution (a) (0.2 per cent);

peak in the chromatogram obtained with reference

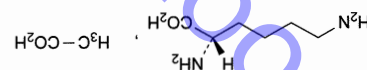
— impurity C: not more than twice the area of the principal

C. 19-nor-17 α -pregna-4,20-dien-17-ol.

Ph Eur

Lysine Acetate

(Ph. Eur. monograph 2114)

C₈H₁₈N₂O₄ 206.2 57282-49-2

Action and use

Amino acid.

Ph Eur

DEFINITION

(2S)-2,6-Diaminohexanoic acid acetate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

Solubility

Freely soluble in water, very slightly soluble in ethanol

(96 per cent).

IDENTIFICATION

First identification: A, B, E.

Second identification: A, C, D, E

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison lysine acetate CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in the minimum volume of water R,

evaporate to dryness at 60 °C and record new spectra using

the residues.

C. Examine the chromatograms obtained in the test for

ninhydrin-positive substances.

Results The principal spot in the chromatogram obtained with

test solution (b) is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

solution (a).

D. To 0.1 mL of solution S (see Tests) add 2 mL of water R

and 1 mL of a 50 g/L solution of phosphomolybdic acid R.

A yellowish-white precipitate is formed.

E. It gives reaction (a) of acetates (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in distilled water R and dilute to 50 mL with

the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Specific optical rotation (2.2.7)

+ 8.5 to + 10.0 (dried substance), determined on solution S.

Ninhydrin-positive substances

Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g of the substance to be

examined in water R and dilute to 10 mL with the same

solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 50 mL

with water R.

Reference solution (a) Dissolve 10 mg of lysine acetate CRS in

water R and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5 mL of test solution (b) to

20 mL with water R.

Reference solution (c) Dissolve 10 mg of lysine acetate CRS and

10 mg of arginine CRS in water R and dilute to 25 mL with

the same solvent.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, 2-propanol R

(30:70 V/V).

Application 5 μ L.

Development Over 2/3 of the plate.

Drying At 100-105 °C until the ammonia has evaporated.

Detection Spray with ninhydrin solution R and heat at

100-105 °C for 15 min.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated spots.

Limits: test solution (a):

— any impurity: any spot, apart from the principal spot, is

not more intense than the principal spot in the

chromatogram obtained with reference solution (b)

(0.5 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Ammonium (2.4.1, Method B)

Maximum 200 ppm, determined on 50 mg.

Prepare the standard using 0.1 mL of ammonium standard

solution (100 ppm NH₄) R.

Iron (2.4.9)

Maximum 30 ppm.

In a separating funnel, dissolve 0.33 g in 10 mL of dilute

hydrochloric acid R. Shake with 3 quantities, each of 10 mL,

of methyl isobutyl ketone R₁, shaking for 3 min each time.

To the combined organic layers add 10 mL of water R and

shake for 3 min. The aqueous layer complies with the test.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 80.0 mg in 3 mL of anhydrous formic acid R. Add 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration. 1 mL of 0.1 M perchloric acid is equivalent to 10.31 mg of $C_6H_{15}ClN_2O_2$.

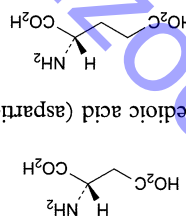
STORAGE

Protected from light.

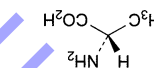
IMPURITIES

Specified impurities A, B, C, D, E, F

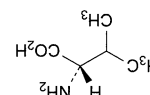
A. (2S)-2-aminobutanedioic acid (aspartic acid),



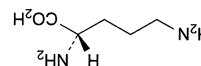
B. (2S)-2-aminopentanedioic acid (glutamic acid),



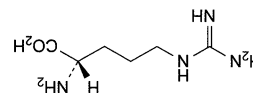
C. (S)-2-aminopropanoic acid (alanine),



D. (S)-2-amino-3-methylbutanoic acid (valine),



E. (2S)-2,5-diaminopentanoic acid (ornithine),

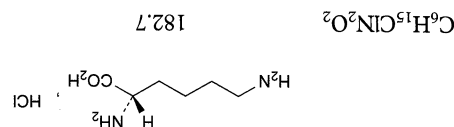


F. (S)-2-amino-5-guandipentanoic acid (arginine).

Ph Eur

Lysine Hydrochloride

(Ph. Eur. monograph 0930)



657-27-2

Action and use

Amino acid.

DEFINITION

(2S)-2,6-diaminohexanoic acid hydrochloride.

Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B, E

Second identification A, C, D, E

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison lysine hydrochloride CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of water R, evaporate to dryness at 60 °C and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in water R and dilute to 50 mL with the same solvent.

Reference solution Dissolve 10 mg of lysine hydrochloride CRS in water R and dilute to 50 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, 2-propanol R

(30:70 V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 105 °C until the ammonia disappears completely.

Detection Spray with ninhydrin solution R and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.1 mL of solution S (see Tests) add 2 mL of water R and 1 mL of a 50 g/L solution of phosphomolybdic acid R.

A yellowish-white precipitate is formed.

E. To 0.1 mL of solution S add 2 mL of water R.

The solution gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 5.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B7 or GY7 (2.2.2, Method III).

Specific optical rotation (2.2.7)

+ 21.0 to + 22.5 (dried substance).

Dissolve 2.00 g in hydrochloric acid R1 and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A water R or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of *proline R* in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of *ammonium standard solution* (100 ppm NH_4^+) *R* to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of *isoleucine R* and 30 mg of *leucine R* (impurity A) in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.
Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids. *System suitability* Reference solution (d):
— *resolution*: minimum 1.5 between the peaks due to isoleucine and impurity A.
— *Calculation of percentage contents*:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of lysine hydrochloride in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification;
— for ammonium, use the concentration of ammonium in reference solution (c) taking into account the corresponding peak in the chromatogram obtained with the blank solution.

Limits:

— any ninhydrin-positive substance: for each impurity, maximum 0.2 per cent;
— *ammonium* at 570 nm: maximum 0.02 per cent;
— *total*: maximum 1.0 per cent;
— *reporting threshold* (excluding ammonium): 0.05 per cent.
The thresholds indicated under Related substances (Table 2034-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Sulfates (2.4.13)

Maximum 300 ppm.
Dilute 5 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 30 ppm.
In a separating funnel, dissolve 0.33 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.
12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

Dissolve 0.150 g in 5 mL of anhydrous formic acid R. Add 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

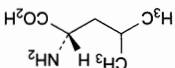
1 mL of 0.1 M perchloric acid is equivalent to 18.27 mg of $\text{C}_6\text{H}_{15}\text{ClN}_2\text{O}_2$.

STORAGE

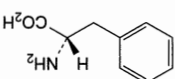
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IMPURITIES

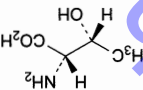
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C.



A. (2S)-2-amino-4-methylpentanoic acid (leucine).



B. (2S)-2-amino-3-phenylpropanoic acid (phenylalanine).



C. (2S,3R)-2-amino-3-hydroxybutanoic acid (threonine).

Macroglols

(Ph. Eur. monograph 1444)

Action and use

Non-ionic surfactant.

DEFINITION

Mixtures of polymers with the general formula $\text{H}[\text{OCH}_2\text{CH}_2]_n\text{OH}$ where *n* represents the average number of oxyethylene groups. The type of macrogol is defined by a number that indicates the average relative molecular mass. A suitable stabiliser may be added.



CHARACTERS

Type of macrogol	Appearance	Solubility
300	clear, viscous, colourless or almost colourless	miscible with water, very soluble in acetone, in alcohol and in
400	colourless, hygroscopic	methyle chloride, practically
600	liquid	insoluble in fatty oils and in
1000	white or almost white, hygroscopic solid with a waxy or paraffin-like appearance	very soluble in water, freely soluble in alcohol and in
1500	white or almost white solid with a waxy or paraffin-like appearance	methyle chloride, freely soluble in alcohol, practically insoluble in fatty oils and in
3000	white or almost white solid with a waxy or paraffin-like appearance	very soluble in water and in
3350	white solid with a waxy or paraffin-like appearance	methyle chloride, very slightly soluble in alcohol, practically insoluble in fatty oils and in
4000	white or almost white solid with a waxy or paraffin-like appearance	very soluble in water and in
6000	white solid with a waxy or paraffin-like appearance	methyle chloride, practically insoluble in alcohol, in fatty oils
8000	white or almost white solid with a waxy or paraffin-like appearance	very soluble in water, soluble in
20 000	white or almost white solid with a waxy or paraffin-like appearance	methyle chloride, practically insoluble in alcohol, in fatty oils
35 000	white solid with a waxy or paraffin-like appearance	methyle chloride, practically insoluble in alcohol, in fatty oils

IDENTIFICATION

A. Viscosity (see Tests).

B. To 1 g in a test-tube add 0.5 mL of *sulfuric acid* R, close the test-tube with a stopper fitted with a bent delivery tube and heat until white fumes are evolved. Collect the fumes via the delivery tube into 1 mL of *mercuric chloride solution* R. An abundant, white, crystalline precipitate is formed. C. To 0.1 g add 0.1 g of *potassium thiocyanate* R and 0.1 g of *cobalt nitrate* R and mix thoroughly with a glass rod. Add 5 mL of *methylene chloride* R and shake. The liquid phase becomes blue.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Dissolve 12.5 g in *water* R and dilute to 50 mL with the same solvent.

Acidity or alkalinity

Dissolve 5.0 g in 50 mL of *carbon dioxide-free water* R and add 0.15 mL of *bromothymol blue solution* RI. The solution is yellow or green. Not more than 0.1 mL of 0.1 M *sodium hydroxide* is required to change the colour of the indicator to blue.

Viscosity (2.2.9)

The viscosity is calculated using a density given in Table 1444-1.

For macrogols with a relative molecular mass greater than 400, determine the viscosity on a 50 per cent *m/m* solution of the substance to be examined.

Freezing point (2.2.18)

See Table 1444-2.

Hydroxyl value

Introduce *m* g (see Table 1444-3) into a dry conical flask fitted with a reflux condenser. Add 25.0 mL of *phthalic anhydride solution* R, swirl to dissolve and boil under a reflux condenser first with 25 mL of *pyridine* R and then with

Table 1444-2

Type of macrogol	Freezing point (°C)
600	15 - 25
1000	35 - 40
1500	42 - 48
3000	50 - 56
3350	53 - 57
4000	53 - 59
6000	55 - 61
8000	55 - 62
20 000	minimum 57
35 000	minimum 57

Table 1444-1

Type of macrogol	Kinematic viscosity (mm ² ·s ⁻¹)	Dynamic viscosity (mPa·s)	Density* (g/mL)
300	71 - 94	80 - 105	1.120
400	94 - 116	105 - 130	1.120
600	13.9 - 18.5	15 - 20	1.080
1000	20.4 - 27.7	22 - 30	1.080
1500	31 - 46	34 - 50	1.080
3000	69 - 93	75 - 100	1.080
3350	76 - 110	83 - 120	1.080
4000	102 - 158	110 - 170	1.080
6000	185 - 250	200 - 270	1.080
8000	240 - 472	260 - 510	1.080
20 000	2500 - 3200	2700 - 3500	1.080
35 000	10 000 - 13 000	11 000 - 14 000	1.080

* Density of the substance for macrogols 300 and 400. Density of the 50 per cent *m/m* solution for the other macrogols.

$$56.1 \times (n_2 - n_1) / m$$

Calculate the hydroxyl value using the following expression:

25 mL of *water* R, add 1.5 mL of *phenolphthalein solution* R and titrate with 1 M *sodium hydroxide* until a faint pink colour is obtained (*n*₁ mL). Carry out a blank test (*n*₂ mL).

For macrogols with a relative molecular mass greater than 1000, if the water content is more than 0.5 per cent, dry a sample of suitable mass at 100-105 °C for 2 h and carry out the determination of the hydroxyl value on the dried sample.

Table 1444-3

Type of macrogol	Hydroxyl value	m (g)
300	340 - 394	1.5
400	264 - 300	1.9
600	178 - 197	3.5
1000	107 - 118	5.0
1500	70 - 80	7.0
3000	34 - 42	12.0
3350	30 - 38	12.0
4000	25 - 32	14.0
6000	16 - 22	18.0
8000	12 - 16	24.0
20 000	-	-
35 000	-	-

Reducing substances

Dissolve 1 g in 1 mL of a 10 g/L solution of resorcinol R and warm gently if necessary. Add 2 mL of hydrochloric acid R. After 5 min the solution is not more intensely coloured than reference solution R₃ (2.2.2, Method I).

Formaldehyde

Maximum 30 ppm.
Test solution To 1.00 g add 0.25 mL of chromotropic acid, sodium salt solution R, cool in iced water and add 5.0 mL of sulfuric acid R. Allow to stand for 15 min and dilute slowly to 10 mL with water R.

Reference solution Dilute 0.860 g of formaldehyde solution R to 100 mL with water R. Dilute 1.0 mL of this solution to 100 mL with water R. In a 10 mL flask, mix 1.00 mL of this solution and 0.25 mL of chromotropic acid, sodium salt solution R, cool in iced water and add 5.0 mL of sulfuric acid R. Allow to stand for 15 min and dilute slowly to 10 mL with water R.

Blank solution In a 10 mL flask mix 1.00 mL of water R and 0.25 mL of chromotropic acid, sodium salt solution R, cool in iced water and add 5.0 mL of sulfuric acid R. Dilute slowly to 10 mL with water R.

Determine the absorbance (2.2.25) of the test solution at 567 nm, against the blank solution. It is not higher than that of the reference solution.

If the use of macrogols with a higher content of formaldehyde may have adverse effects, the competent authority may impose a limit of not more than 15 ppm.

Ethylene glycol and diethylene glycol

Carry out this test only if the macrogol has a relative molecular mass below 1000.

Gas chromatography (2.2.28).

Test solution Dissolve 5.00 g of the substance to be examined in acetone R and dilute to 100.0 mL with the same solvent. Reference solution Dissolve 0.10 g of ethylene glycol R and 0.50 g of diethylene glycol R in acetone R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with acetone R.

Column:

material: glass;

size: $l = 1.8$ m, $\varnothing = 2$ mm;

stationary phase: silanised diatomaceous earth for gas chromatography R, impregnated with 5 per cent m/m of macrogol 20 000 R.

Carrier gas nitrogen for chromatography R.

Flow rate 30 mL/min.

Temperature:

— column: if necessary, precondition the column by heating at 200 °C for about 15 h; adjust the initial temperature of the column to obtain a retention time of 14–16 min for diethylene glycol; raise the temperature of the column by about 30 °C at a rate of 2 °C/min but without exceeding 170 °C;

— injection port and detector: 250 °C.

Detection Flame ionisation.

Injection 2 μ L.

Carry out 5 replicate injections to check the repeatability of the response.

Limit Maximum 0.4 per cent, calculated as the sum of the contents of ethylene glycol and diethylene glycol.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Water (2.5.12)

Maximum 2.0 per cent for macrogols with a relative

molecular mass not greater than 1000 and maximum

1.0 per cent for macrogols with a relative molecular mass greater than 1000, determined on 2.00 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

STORAGE

In an airtight container.

LABELLING

The label states:

— the type of macrogol;

— the content of formaldehyde.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). Some of the characteristics described in the functionality-

related characteristics section may also be present in the mandatory

part of the monograph since they also represent mandatory quality

criteria. In such cases, a cross-reference to the tests described in the

mandatory part is included in the Functionality-related

characteristics section. Control of the characteristics can contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for macrogols used as

solvent.

Viscosity

(see Tests).

The following characteristics may be relevant for macrogols used as

suspension stabiliser and thickener.

Viscosity

(see Tests).

The following characteristic may be relevant for macrogols used as

lubricant in tablets.

Particle-size distribution (2.9.31)

The following characteristics may be relevant for macrogols used as

suppository base and for macrogols used in hydrophilic ointments.

Viscosity

(see Tests).

Melting point (2.2.15)

Ph Eur

**High-molecular-mass Macrolog**

(Ph. Eur. monograph 2444)

Ph Eur

DEFINITION

Mixtures of high-molecular-mass macrogols (also known as polyethylene oxides). The type of high-molecular-mass macrogol is defined by a number that indicates the nominal average molecular mass in the range of 100 000 to 7 000 000. A suitable stabiliser (e.g. butylhydroxytoluene) and a suitable flowability agent (e.g. silicon dioxide) may be added.

CHARACTERS**Appearance**

White or almost white, free-flowing powder.

Solubility

Freely soluble in water.

mp

Minimum 65 °C.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison high-molecular-mass macrogol CRS.

B. Viscosity (see Tests).

TESTS**Solution S**

Prepare a solution of the substance to be examined in a mixture of carbon dioxide-free water R and 2-propanol R according to Table 2444-1.

Table 2444-1. – Solution S and viscosity

Nominal molecular mass	Composition of solution S (g of substance / mL of carbon dioxide-free water R + mL of 2-propanol R)	Dynamic viscosity (mPa·s)	Rotation speed (r/min)
100 000	30 g / 570 + 125	12 - 50	50
200 000	30 g / 570 + 125	65 - 115	50
300 000	30 g / 570 + 125	600 - 1200	10
400 000	30 g / 570 + 125	2250 - 4500	2
600 000	30 g / 570 + 125	4500 - 8800	2
900 000	30 g / 570 + 125	8800 - 17 600	2
1 000 000	12 g / 588 + 125	400 - 800	10
2 000 000	12 g / 588 + 125	2000 - 4000	10
4 000 000	6 g / 594 + 125	1650 - 5500	2
5 000 000	6 g / 594 + 125	5500 - 7500	2
7 000 000	6 g / 594 + 125	7500 - 10 000	2

pH (2.2.3)

6.0 to 10.0 for solution S.

Viscosity (2.2.10)

Determine the dynamic viscosity of solution S at 25 °C using a rotating viscometer and a suitable spindle.

Macrolog Cetostearyl Ether

(Ph. Eur. monograph 1123)

Ph Eur

DEFINITION

Mixture of ethers of mixed macrogols with linear fatty alcohols, mainly cetostearyl alcohol. It may contain some free macrogols and it contains various amounts of free cetostearyl alcohol. The number of moles of ethylene oxide reacted per mole of cetostearyl alcohol is 2 to 33 (nominal value).

CHARACTERS**Appearance**

White or yellowish-white, waxy, unctuous mass, pellets, microbeads or flakes.

Solubility

— macrogol cetostearyl ether with low numbers of moles of ethylene oxide reacted per mole: practically insoluble in water, soluble in ethanol (96 per cent) and in methylene chloride;

LABELLING

In an airtight container.

STORAGE

examined...".

Maximum 5.0 per cent, determined on 1.00 g. Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C, until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting from "Moisten the substance to be examined...".

Sulfated ash

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 45 min.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 45 min. Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C, until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting from "Moisten the substance to be examined...".

Ethylene oxide (2.4.25, Method A) Maximum 1 ppm. Carry out the test with the following modifications. **Stock solution** Dilute 0.5 mL of ethylene oxide stock solution R2 to 10.0 mL with propanol R. Dilute 1.0 mL of this solution to 50.0 mL with propanol R. **Test solution** Weigh 0.50 g (M_T) of the substance to be examined into a 10 mL vial (other sizes may be used depending on the operating conditions). Close and allow to stand at 70 °C for 30 min. **Reference solution (a)** Weigh 0.50 g (M_R) of the substance to be examined into an identical 10 mL vial and add 10 µL of the stock solution. Close and homogenise, and allow to stand at 70 °C for 30 min. **Reference solution (b)** To 0.50 mL of ethylene oxide solution R3 in a 10 mL vial add 0.1 mL of a freshly prepared 10 mg/L solution of acetaldehyde R. Close and homogenise, and allow to stand at 70 °C for 30 min.

Loss on drying (2.2.32) Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 45 min.

Sulfated ash Maximum 5.0 per cent, determined on 1.00 g. Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C, until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting from "Moisten the substance to be examined...".

The label states the type of high-molecular-mass macrogol.



— macrologol cetostearyl ether with higher numbers of moles of ethylene oxide reacted per mole: dispersible or soluble in water, soluble in ethanol (96 per cent) and in methylene chloride.

It solidifies at 32 °C to 52 °C.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Saponification value (see Tests).

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve the prescribed amount of substance to be examined (see table below) in a mixture of 1 volume of water R and 9 volumes of methanol R and dilute to 75 mL with the same mixture of solvents.

Number of moles of ethylene oxide reacted per mole	Amount to be dissolved (g)
--	----------------------------

2 - 6	5.0
10 - 22	10.0
25 - 33	15.0

Add 60 mL of hexane R and shake for 3 min. The formation of foam can be reduced by the addition of some drops of ethanol (96 per cent) R. Filter the upper layer through anhydrous sodium sulfate R, wash the filter with 3 quantities, each of 10 mL, of hexane R and evaporate the combined filtrates to dryness. Dissolve 0.05 g of the residue in 10 mL of methanol R (the solution may be opalescent).

Reference solution Dissolve 25 mg of stearyl alcohol CRS in methanol R and dilute to 25 mL with the same solvent.

Plate TLC silica gel plate R.

Application 20 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with vanillin-sulfuric acid reagent prepared as follows: dissolve 0.5 g of vanillin R in 50 mL of ethanol (96 per cent) R and dilute to 100 mL with sulfuric acid R; allow to dry in air; heat at about 130 °C for 15 min and allow to cool in air.

Results The chromatogram obtained with the test solution shows several spots; one of these spots corresponds to the principal spot in the chromatogram obtained with the reference solution.

E. Dissolve or disperse 0.1 g in 5 mL of ethanol (96 per cent) R, add 2 mL of water R, 10 mL of dilute hydrochloric acid R, 10 mL of barium chloride solution R1 and 10 mL of a 100 g/L solution of phosphomolybdic acid R. A precipitate is formed.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Dissolve 5.0 g in ethanol (96 per cent) R and dilute to 50 mL with the same solvent.

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of ethanol (96 per cent) R and 10 mL of water R. Add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator to yellow.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

Number of moles of ethylene oxide reacted per mole (nominal value)	Hydroxyl value
--	----------------

2	150 - 180
3	135 - 155
5 - 6	100 - 134
10	75 - 90
12	67 - 77
15	58 - 67
20 - 22	40 - 55
25	36 - 46
30 - 33	32 - 40

Iodine value (2.5.4, Method A)

Maximum 2.0.

Saponification value (2.5.6)

Maximum 3.0, determined on 10.0 g.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Water (2.5.12)

Maximum 3.0 per cent, determined on 2.00 g.

Total ash (2.4.16)

Maximum 0.2 per cent, determined on 2.0 g.

STORAGE

In an airtight container.

LABELLING

The label states the number of moles of ethylene oxide reacted per mole of cetostearyl alcohol (nominal value).

Ph Eur



Macrologol 30 Dipolyhydroxystearate

(Ph Eur monograph 2584)

Ph Eur

DEFINITION

Mixture of mainly diesters of polymerised 12-hydroxystearic (12-hydroxyoctadecanoic) acid and macrogols (1444) obtained by esterification of macrologol with 12-hydroxystearic acid. The average number of moles of ethylene oxide reacted per mole of substance is 30.

CHARACTERS

Appearance

Brownish-red, waxy mass.

Solubility

Practically insoluble in water, very soluble in methylene chloride and soluble in most aliphatic and aromatic hydrocarbons.

mp: 30 °C to 40 °C.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison macrologol 30 dipolyhydroxystearate CRS.

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

TESTS**Acid value** (2.5.1) Maximum 10.0.**Hydroxyl value** (2.5.3, Method A) 12 to 30.**Iodine value** (2.5.4, Method A) Maximum 10.0.**Peroxide value** (2.5.5) Maximum 5.0.**Saponification value** (2.5.6) 125 to 145, determined on 2.0 g.

Use 30.0 mL of 0.5 M alcoholic potassium hydroxide, heat under reflux for 60 min and add 50 mL of anhydrous ethanol R before carrying out the titration.

Nickel (2.4.31) Maximum 1 ppm.**Water** (2.5.12) Maximum 1.0 per cent, determined on 0.50 g.**Sulfated ash** (2.4.14) Maximum 0.5 per cent.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.0 g in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C, until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting from "Moisten the substance to be examined..."

STORAGE

In an airtight container.

Macrolog 6 Glycerol Caprylocaprate

(Ph. Eur. monograph 1443)

Action and use

Non-ionic surfactant.

DEFINITION

Mixture of mainly mono- and diesters of polyoxyethylene glycerol ethers mainly with caprylic (octanoic) and capric (decanoic) acids. The average number of moles of ethylene oxide reacted per mole of substance is 6.

Macrolog 6 glycerol caprylocaprate may be obtained by ethoxylation of glycerol and esterification with distilled coconut or palm kernel fatty acids, or by ethoxylation of mono- and diglycerides of caprylic and capric acids.

CHARACTERS**Appearance**

Pale yellow liquid.

Solubility

Partly soluble in water, freely soluble in castor oil, in glycerol, in isopropanol and in propylene glycol.

IDENTIFICATION

A. Dissolve 1.0 g in 99 g of a mixture of 10 volumes of 2-propanol R and 90 volumes of water R. Heat the solution to about 40 °C. A turbidity is produced. Allow to cool until the turbidity disappears. The cloud point is between 15 °C and 35 °C.

Macrolog Glycerol Cocoates

(Ph. Eur. monograph 1122)

Action and use

Pharmaceutical aids.

DEFINITION

Mixtures of mono-, di- and triesters of ethoxylated glycerol with fatty acids of vegetable origin having a composition corresponding to the fatty acid composition of the oil extracted from the hard, dried fraction of the endosperm of *Cocos nucifera* L. The average number of moles of ethylene oxide reacted per mole of substance (nominal value) is either 7 (macrolog 7 glycerol cocoate) or 23 (macrolog 23 glycerol cocoate).

CHARACTERS**Appearance**

Clear, yellowish, oily liquid.

Solubility

Soluble in water and in ethanol (96 per cent) and practically insoluble in light petroleum (bp: 50-70 °C) for

Ph Eur

Ph Eur



macrologol 7 glycerol cocoate and macrologol 23 glycerol

cocoate.

Relative density

About 1.05 for macrologol 7 glycerol cocoate; about 1.09 for

macrologol 23 glycerol cocoate.

IDENTIFICATION

A. Dissolve 1.0 g of macrologol 7 glycerol cocoate in 99 g of a mixture of 10 volumes of 2-propanol R and 90 volumes of water R. Heat the solution to about 65 °C. A turbidity is

produced. Allow to cool until the turbidity disappears.

The cloud point is between 35 °C and 54 °C.

Heat a 10 g/L solution of macrologol 23 glycerol cocoate in a

100 g/L solution of sodium chloride R to about 90 °C.

A turbidity is produced. Allow to cool until the turbidity

disappears. The cloud point is between 65 °C and 85 °C.

B. Iodine value (see Tests).

C. Saponification value (see Tests).

TESTS

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution Y₂ (2.2.2,

Method D).

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of ethanol (96 per cent) R and 10 mL of water R. Add 0.1 mL of

0.1 M hydrochloric acid is required to change the colour of the

indicator to yellow.

Acid value (2.5.1)

Maximum 5.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

See Table 1122-1.

Saponification value (2.5.6)

See Table 1122-1.

Table 1122-1

Number of moles of ethylene oxide reacted per mole (nominal value)	Hydroxyl value	Saponification value (determined on 2.0 g)
7	170 - 210	85 - 105
23	80 - 100	40 - 50

Iodine value (2.5.4, Method A)

Maximum 5.0.

Composition of fatty acids

Gas chromatography (2.4.22, Method A).

Composition of the fatty-acid fraction of the substance:

— caproic acid: maximum 1.0 per cent;

— caprylic acid: 5.0 per cent to 10.0 per cent;

— lauric acid: 40.0 per cent to 55.0 per cent;

— myristic acid: 14.0 per cent to 23.0 per cent;

— palmitic acid: 8.0 per cent to 12.0 per cent;

— stearic acid: 1.0 per cent to 5.0 per cent;

— oleic acid: 5.0 per cent to 10.0 per cent;

— linoleic acid: maximum 3.0 per cent.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm

of dioxan.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.0 g.

Macrologol 20 Glycerol

Monostearate

(Ph. Eur. monograph 2044)

Action and use

Non-ionic surfactant.

Ph Eur

DEFINITION

Macrologol 20 glycerol monostearate is obtained by

ethoxylation with ethylene oxide of different types of glycerol

stearates, mainly Glycerol monostearate 40-55 (0495).

The number of moles of ethylene oxide reacted per mole of

glycerol stearate is 20 (nominal value).

CHARACTERS

Appearance

Pale yellow, oily liquid or gel.

Solubility

Soluble in water at 40 °C and above and in ethanol

(96 per cent), practically insoluble in light liquid paraffin and

in fatty oils.

Relative density

About 1.07.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Saponification value (see Tests).

C. Composition of fatty acids (see Tests).

D. Place 1 g in a test tube and add 0.1 mL of sulfuric acid R.

Heat the tube until white fumes appear. The fumes turn

filter paper impregnated with alkaline potassium

tetraiodomercurate solution R black.

TESTS

Acid value (2.5.1)

Maximum 2.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

65 to 85, determined on 0.350 g.

Iodine value (2.5.4, Method A)

Maximum 2.0.

Peroxide value (2.5.5, Method A)

Maximum 6.0.

Saponification value (2.5.6)

40 to 60.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Composition of the fatty-acid fraction of the substance:



Ph Eur

30 min. Acidify the warm solution with 20 mL of hydrochloric acid R and cool to room temperature. Shake the mixture with 50 mL of ether R and allow to stand until a separation of the layers is visible. Separate the clear upper layer, add 5 g of anhydrous sodium sulfate R, wait for 30 min, filter and evaporate to dryness on a water-bath. Dissolve 50 mg of the residue in 25 mL of ether R.

Reference solution Dissolve 50 mg of 12-hydroxystearic acid R in 25 mL of methylene chloride R.

Plate TLC octadecylsilyl silica gel plate R.

Mobile phase methylene chloride R, glacial acetic acid R, acetone R (10:40:50 V/V/V).

Application 2 μ L.

Development Over 2/3 of the plate.

Drying In a current of cold air.

Detection Spray with a 80 g/L solution of phosphomolybdic acid R in 2-propanol R and heat at 120 °C for 1-2 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and colour to the principal spot in the chromatogram obtained with the reference solution.

B. Dissolve 15.0 g in 50 mL of water R. The viscosity (2.2.9) has a maximum of 20 mPa.s.

C. Free macrogols (see Tests).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension III (2.2.1) and not more intensely coloured than reference solution B₆ or BY₆ (2.2.2, Method II).

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent.

Acid value (2.5.1)

Maximum 1.0, determined on 2.0 g.

Hydroxyl value (2.5.3, Method A)

90 to 110.

Iodine value (2.5.4, Method A)

Maximum 2.0.

Peroxide value (2.5.5, Method A)

Maximum 5.0.

Saponification value (2.5.6)

53 to 63.

Free macrogols

Size-exclusion chromatography (2.2.30).

Test solution Dissolve 1.20 g of the substance to be examined in the mobile phase and dilute to 250.0 mL with the mobile phase.

Reference solution (a) Dissolve about 0.4 g of macrologol 1000 R in the mobile phase and dilute to 250.0 mL with the mobile phase.

Reference solution (b) Dilute 50.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Precolumns (2):

— size: $l = 0.125$ m, $\varnothing = 4$ mm;

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m) with a pore size of 10 nm.

Column:

— size: $l = 0.30$ m, $\varnothing = 7.8$ mm;

— stationary phase: hydroxylated polymethacrylate gel R (6 μ m) with a pore size of 12 nm.

Type of macrologol 20 glycerol monostearate	Type of glycerol stearate used	Composition of fatty acids
Type I	Type I (obtained using stearic acid 50)	Stearic acid: 40.0 per cent to 60.0 per cent, Sum of the contents of palmitic and stearic acids: minimum 90.0 per cent
Type II	Type II (obtained using stearic acid 70)	Stearic acid: 60.0 per cent to 80.0 per cent, Sum of the contents of palmitic and stearic acids: minimum 90.0 per cent
Type III	Type III (obtained using stearic acid 95)	Stearic acid: 90.0 per cent to 99.0 per cent, Sum of the contents of palmitic and stearic acids: minimum 96.0 per cent

Ethylene oxide and dioxan (2.4.25, Method A) Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.00 g.

Total ash (2.4.16)

Maximum 0.2 per cent.

STORAGE

Protected from light.

LABELLING

The label states the type of macrologol 20 glycerol monostearate.

Macrologol 15 Hydroxystearate

(Ph. Eur. monograph 2052)

Action and use

Non-ionic surfactant.

DEFINITION

Mixture of mainly monoesters and diesters of 12-hydroxystearic (12-hydroxyoctadecanoic) acid and macrogols obtained by ethoxylation of 12-hydroxystearic acid. The number of moles of ethylene oxide reacted per mole of 12-hydroxystearic acid is 15 (nominal value). It contains free macrogols.

CHARACTERS

Appearance

Yellowish, waxy mass.

Solubility

Very soluble in water, soluble in ethanol (96 per cent), insoluble in liquid paraffin.

It solidifies at about 25 °C.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution To 1.0 g add 100 mL of a 100 g/L solution of potassium hydroxide R and boil under a reflux condenser for

Connect both precolumns to the column using a 3-way valve and switch the mobile phase flow according to the following programme:

- 0-114 s: precolumn 1 and column;
- 115 s to the end: precolumn 2 and column;
- 115 s to 7 min: flow back of precolumn 1.

Mobile phase water R, methanol R (2:8 V/V).

Flow rate 1.1 mL/min.

Detection Refractometer.

Injection 50 µL.

Calculate the percentage content of free macrogols using the following expression:

$$A_1 \times m_2 \times 200 \div (m_1 \times (A_2 + 2A_3))$$

m_1 = mass of the substance to be examined in the test

m_2 = mass of macrogol 1000 R in reference solution (a),

A_1 = area of the peak due to free macrogols in the

substance to be examined in the chromatogram

obtained with the test solution;

A_2 = area of the peak due to macrogol 1000 in the

chromatogram obtained with reference solution

(a);

A_3 = area of the peak due to macrogol 1000 in the

chromatogram obtained with reference solution

(b).

Limit:

— free macrogols: 27.0 per cent to 39.0 per cent.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 50 ppm

of dioxan.

Nickel (2.4.31)

Maximum 1 ppm.

Water (2.5.12)

Maximum 1.0 per cent, determined on 2.00 g.

Total ash (2.4.16)

Maximum 0.3 per cent, determined on 1.0 g.

STORAGE

In an airtight container.

Ph Eur

Macrogol Isotridecyl Ether

(Ph Eur monograph 2730)

Ph Eur

DEFINITION

Mixture of ethers of mixed macrogols with linear and

branched fatty alcohols, mainly $C_{13}H_{28}O$. It contains a

variable quantity of free $C_{13}H_{28}O$ and it may contain free

macrogols. The number of moles of ethylene oxide reacted

per mole of branched $C_{13}H_{28}O$ is 3 or 4 (nominal value).

CHARACTERS

Appearance

Colourless liquid.

Solubility

Practically insoluble in water, soluble or dispersible in ethanol (96 per cent), practically insoluble in light petroleum.

IDENTIFICATION

A. Gas chromatography (2.2.28).

Test solution Dissolve 0.200 g of the substance to be examined in acetone R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 0.35 g of lauryl alcohol R, 0.35 g of tridecyl alcohol R, 0.15 g of myristyl alcohol R and 0.15 g of ethylene glycol monododecyl ether R in acetone R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with acetone R.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.25$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness

0.25 µm).

Carrier gas: hydrogen for chromatography R or helium for

chromatography R.

Flow rate: 1 mL/min.

Split ratio 1:50.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 1	120			
1 - 24	120 → 350			
24 - 34	350			
				300
				350

Detection Flame ionisation.

Injection 1 µL.

Retention time Lauryl alcohol = about 6.7 min; tridecyl

alcohol = about 7.8 min; myristyl alcohol = about 9 min;

ethylene glycol monododecyl ether = about 9.3 min.

System suitability: reference solution;

— resolution: minimum 5 between the peaks due to

myristyl alcohol and ethylene glycol monododecyl

ether.

Results The sum of the areas of the peaks eluting between the

peak due to lauryl alcohol and the peak due to tridecyl

alcohol is greater than the area of the peak due to tridecyl

B. Hydroxyl value (see Tests).

C. Iodine value (see Tests).

D. Saponification value (see Tests).

E. Dissolve or disperse 0.1 g in 5 mL of ethanol (96 per cent) R. Add 10 mL of dilute hydrochloric acid R, 10 mL of barium chloride solution R1 and 10 mL of a 100 g/L solution of phosphomolybdic acid R. A precipitate is

TESTS

Appearance of solution

The solution is not more intensely coloured than reference

solution BY₅ (2.2.2, Method II).

Dissolve 5.0 g in ethanol (96 per cent) R and dilute to 50 mL

with the same solvent.

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of

ethanol (96 per cent) R and 10 mL of water R. Add 0.1 mL of

bromothymol blue solution R1. Not more than 0.5 mL of

— Macrogol lauryl ether with 9 to 23 units of ethylene oxide per molecule.

Appearance

White or almost white, waxy mass.

Solubility

Soluble or dispersible in water, soluble in alcohol, practically insoluble in light petroleum.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Saponification value (see Tests).

D. Dissolve or disperse 0.1 g in 5 mL of alcohol R, add 10 mL of dilute hydrochloric acid R, 10 mL of barium chloride solution R1 and 10 mL of a 100 g/L solution of phosphomolybdic acid R. A precipitate is formed.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Dissolve 5.0 g in alcohol R and dilute to 50 mL with the same solvent.

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of water R and 10 mL of alcohol R. Add 0.1 mL of bromothymol blue

solution R1. Not more than 0.5 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator to yellow.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

Ethylene oxide units per molecule		Hydroxyl value	
(nominal value)			
3	165 - 180	4	145 - 165
4	130 - 140	5	90 - 100
9	85 - 95	10	73 - 83
12	64 - 74	15	40 - 60
20 - 23			

The label states the number of moles of ethylene oxide reacted per mole of C₁₂H₂₆O (nominal value).

LABELLING

In an airtight container.

STORAGE

Maximum 0.2 per cent, determined on 2.0 g.

Total ash (2.4.16)

Maximum 3.0 per cent, determined on 2.00 g.

Water (2.5.12)

Maximum 1 ppm of dioxan.

Maximum 1 ppm of ethylene oxide and maximum 10 ppm

Ethylene oxide and dioxan (2.4.25)

Maximum 3.0, determined on 10.0 g.

Saponification value (2.5.6)

Maximum 2.0.

Iodine value (2.5.4)

Macrogol Lauryl Ether

(Ph. Eur. monograph 1124)



Ph Eur

The label states the number of moles of ethylene oxide reacted per mole of C₁₃H₂₈O (nominal value).

LABELLING

In an airtight container.

STORAGE

examined...".

obtained, starting from "Moisten the substance to be Carry out the test for sulfated ash (2.4.14) on the residue.

600 ± 25 °C until the substance is thoroughly charred. 100-105 °C for 1 h and ignite in a muffle furnace at Evenly distribute 1.0 g in the crucible and weigh. Dry at cool in a desiccator over a suitable desiccant and weigh.

Ignite a silica crucible at 600 ± 50 °C for 30 min, allow to Maximum 0.2 per cent.

Sulfated ash (2.4.14)

Maximum 3.0 per cent, determined on 2.00 g.

Water (2.5.12)

Maximum 1 ppm of ethylene oxide and 10 ppm of dioxan.

Ethylene oxide and dioxan (2.4.25)

Maximum 3.0, determined on 10.0 g.

Saponification value (2.5.6)

Maximum 2.0.

Iodine value (2.5.4)

Ethylene oxide units per molecule		Hydroxyl value	
(nominal value)			
3	165 - 185	4	145 - 160

Table 2730.-1

See Table 2730.-1.

Hydroxyl value (2.5.3, Method A)

Maximum 1.0, determined on 5.0 g.

Acid value (2.5.1)

indicator to yellow.

0.1 M hydrochloric acid is required to change the colour of the

Practically insoluble in water, soluble or dispersible in alcohol, practically insoluble in light petroleum.

Solubility

Colourless liquid.

Appearance

per molecule.

— Macrogol lauryl ether with 3 to 5 units of ethylene oxide

CHARACTERS

3 to 23 (nominal value).

moles of ethylene oxide reacted per mole of C₁₂H₂₆O is C₁₂H₂₆O and it may contain free macrogols. The number of mainly C₁₂H₂₆O. It contains a variable quantity of free

Mixture of ethers of mixed macrogols with fatty alcohols,

DEFINITION

Non-ionic surfactant.

Action and use

Macroglol Oleate

(Ph. Eur. monograph 1618)



Action and use
Non-ionic surfactant.

DEFINITION

A mixture of monoesters and diesters of mainly oleic (*cis*-9-octadecenoic) acid and macrogols. It may be obtained by ethoxylation of *Oleic acid* (0799) or by esterification of macrogols with oleic acid of animal or vegetable origin. It may contain free macrogols. The average polymer length is equivalent to 5-6 or 10 moles of ethylene oxide per mole (nominal value). A suitable antioxidant may be added.

CHARACTERS

Appearance
Slightly yellowish, viscous liquid.

Solubility

Dispersible in water, soluble in ethanol (96 per cent) and in 2-propanol, dispersible in oils, miscible with fatty oils and with waxes.

Refractive index

About 1.466.

IDENTIFICATION

First identification A, C.
Second identification A, B.

A. Saponification value (see Tests).

B. Thin-layer chromatography (2.2.27).

Test solution To 20 mg add 10 mL of *methylene chloride* R and mix.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase 25 per cent V/V solution of concentrated ammonia R, 2-propanol R (20:80 V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with potassium iodobismuthate solution R₄.

examine the plate about 10 min later.

Results The chromatogram obtained shows 3 principal spots, corresponding, in order of increasing R_F value, to free macrogol, macrogol mono-oleate and macrogol dioleate.

C. Composition of fatty acids (see Tests).

TESTS

Alkalinity

Dissolve 2.0 g in ethanol (96 per cent) R and dilute to 20 mL with the same solvent. To 2 mL of this solution add 0.05 mL of *phenol red solution* R. The solution is not red.

Acid value (2.5.1)

Maximum 2.0.

Hydroxyl value (2.5.3, Method A)

See Table 1618-1.

Iodine value (2.5.4, Method A)

See Table 1618-1.

Peroxide value (2.5.5, Method A)

Maximum 12.0.

Saponification value (2.5.6)

See Table 1618-1.

Appearance

Yellowish-white waxy mass.

per molecule.

— Macroglol oleyl ether with 10 to 20 units of ethylene oxide insoluble in light petroleum.

Practically insoluble in water, soluble in alcohol, practically

Solubility

Yellow liquid.

Appearance

per molecule.

— Macroglol oleyl ether with 2 to 5 units of ethylene oxide

CHARACTERS

antioxidant may be added.

The number of moles of ethylene oxide reacted per mole of free oleyl alcohol and it may contain free macrogols. alcohols, mainly oleyl alcohol. It contains a variable quantity Mixture of ethers of mixed macrogols with linear fatty

DEFINITION

Ph Eur

Non-ionic surfactant.

Action and use

(Ph. Eur. monograph 1125)

Macroglol Oleyl Ether



Ph Eur

The label states the number of moles of ethylene oxide per mole (nominal value).

LABELLING

In an airtight container.

STORAGE

Maximum 0.3 per cent, determined on 1.0 g.

Total ash (2.4.16)

anhydrous methanol R as the solvent.

Maximum 2.0 per cent, determined on 1.00 g using

Water (2.5.12)

residual dioxan.

Maximum 1 ppm of residual ethylene oxide and 10 ppm of

Residual ethylene oxide and dioxan (2.4.25)

4.0 per cent.

— fatty acids with a chain length greater than C₁₈: maximum

— *linolenic acid*: maximum 4.0 per cent;

— *linoleic acid*: maximum 18.0 per cent;

— *oleic acid*: 65.0 per cent to 88.0 per cent;

— *palmitoleic acid*: maximum 8.0 per cent;

— *palmitic acid*: maximum 16.0 per cent;

— *stearic acid*: maximum 6.0 per cent;

— *myristic acid*: maximum 5.0 per cent;

Composition of the fatty-acid fraction of the substance:

Gas chromatography (2.4.22, Method A).

Composition of fatty acids

Table 1618-1	5-6 moles of ethylene oxide	10 moles of ethylene oxide
Hydroxyl value	50 - 70	65 - 90
Iodine value	50 - 60	27 - 34
Saponification value	105 - 120	68 - 85



Macrogol Poly(vinyl alcohol) Grafted Copolymer

(Ph. Eur. monograph 2523)

DEFINITION

Grafted copolymer of macrogol and poly(vinyl alcohol), having a mean relative molecular mass of about 45 000. It consists of about 75 per cent of poly(vinyl alcohol) units and 25 per cent of macrogol units. It may contain *Amorphous colloidal silica* (0434) to improve flowability.

CHARACTERS

Appearance

White or slightly yellowish powder; opalescent solutions may be obtained during testing due to the presence of anhydrous colloidal silica.

Solubility

Very soluble in water, practically insoluble in anhydrous ethanol and in acetone. It dissolves in dilute acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison macrogol poly(vinyl alcohol) grafted copolymer CRS. Preparation Dissolve 0.2 g in 20 mL of water R, spread a few drops of the solution on a thallium bromiodide plate and evaporate the solvent at 110 °C for 30 min.
B. Dissolve 0.4 g in 2 mL of water R. Place 1 mL of the solution on a glass plate and allow to dry. A transparent film is formed.

TESTS

pH (2.2.3)

Dissolve 5.0 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Ester value

Determine the acid value (I_A) as follows. Dissolve 5.00 g in 100 mL of distilled water R while stirring with a magnetic stirrer. Titrate with 0.01 M alcoholic potassium hydroxide, determining the end-point potentiometrically (2.2.20). Carry out a blank test under the same conditions.

$$I_A = \frac{0.561(n_1 - n_2)}{m}$$

n_1 = volume of titrant used in the test, in millilitres;
 n_2 = volume of titrant used in the blank test, in millilitres;
 m = mass of the sample, in grams.

Determine the saponification value (I_S) on 5.00 g, using 50.0 mL of 0.5 M alcoholic potassium hydroxide and stirring vigorously with a magnetic stirrer.
The ester value (I_E) is calculated from the saponification value (I_S) and the acid value (I_A):

$$I_E = I_S - I_A$$

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and 10 ppm of dioxan.

Impurity A

Liquid chromatography (2.2.29).

Solubility

Dispersible or soluble in water, soluble in alcohol, practically insoluble in light petroleum.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Saponification value (see Tests).

D. Dissolve or disperse 0.1 g in 5 mL of alcohol R, add 2 mL of water R, 10 mL of dilute hydrochloric acid R, 10 mL of barium chloride solution R1 and 10 mL of a 100 g/L solution of phosphomolybdic acid R. A precipitate is formed.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Dissolve 5.0 g in alcohol R and dilute to 50 mL with the same solvent.

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of water R and 10 mL of alcohol R. Add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator to yellow.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

See Table 1125-1.

Iodine value (2.5.4)

See Table 1125-1.

Table 1125-1

Ethylene oxide units per molecule (nominal value)	Hydroxyl value	Iodine value
2	158 - 178	48 - 74*
5	110 - 125	48 - 56
10	75 - 95	24 - 38
20	40 - 65	14 - 24

* This broad range is needed since 2 different grades of oily alcohol may be used for the synthesis. The iodine value does not differ by more than 5 units from the nominal iodine value and is within the limits stated in the table.

Peroxide value (2.5.5)

Maximum 10.0.

Saponification value (2.5.6)

Maximum 3.0.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and 10 ppm of dioxan.

Water (2.5.12)

Maximum 3.0 per cent, determined on 2.00 g.

Total ash (2.4.16)

Maximum 0.2 per cent, determined on 2.0 g.

STORAGE

In an airtight container, protected from light.

LABELLING

The label states:

— the number of moles of ethylene oxide reacted per mole of oily alcohol (nominal value),
— the nominal iodine value for the type with 2 units of ethylene oxide per molecule.

Test solution Introduce 0.250 g of the substance to be examined into a 10 mL volumetric flask and add about 1 mL of methanol R2. Sonicate. Add about 8 mL of water for chromatography R and dilute to 10.0 mL with the same solvent. Filter.

Reference solution (a) Dissolve 5.0 mg of vinyl acetate CRS (impurity A) in methanol R2 and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 20.0 mL with water for chromatography R. Dilute 1.0 mL of this solution to 10.0 mL with water for chromatography R.

Reference solution (b) Dissolve 5 mg of vinyl acetate R (impurity A) and 5 mg of 1-vinylpyrrolidin-2-one R in 10 mL of methanol R2 and dilute to 50 mL with water for chromatography R. Dilute 1 mL of the solution to 20 mL with water for chromatography R.

A precolumn containing octadecylsilyl silica gel for chromatography R (5 µm) may be used if a matrix effect is observed.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R (5 µm);
— temperature: 30 °C.

Mobile phase:
— mobile phase A: acetonitrile R1, methanol R2, water for chromatography R (5:5:90 V/V/V);
— mobile phase B: methanol R2, acetonitrile R1, water for chromatography R (5:45:50 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	100	0
2 - 40	100 → 85	0 → 15
40 - 42	85 → 0	15 → 100

Flow rate 1.0 mL/min.
Detection Spectrophotometer at 205 nm.

Injection 10 µL.

Retention time Impurity A = about 19 min;

1-vinylpyrrolidin-2-one = about 25 min.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to

impurity A and 1-vinylpyrrolidin-2-one.

Limit:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (100 ppm).

Impurity B

Liquid chromatography (2.2.29).

Test solution Mix 0.200 g of the substance to be examined with water for chromatography R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 30 mg of citric acid R and 0.100 g of acetic acid R (impurity B) in the mobile phase. Shake gently to dissolve and dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R (5 µm).

Mobile phase 0.50 g/L solution of sulfuric acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 µL. After each injection, rinse the column with a mixture of equal volumes of acetonitrile for chromatography R and a 0.50 g/L solution of sulfuric acid R.

Retention time Impurity B = about 5 min; citric

acid = about 7 min.

System suitability: reference solution:

— resolution: minimum 2.0 between the peaks due to

impurity B and citric acid.

Limit:

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (1.5 per cent).

Sulfated ash (2.4.14)

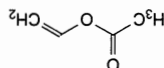
Maximum 3.0 per cent, determined on 5.0 g.

Loss on drying (2.2.32)

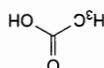
Maximum 5.0 per cent, determined on 1.000 g by drying in vacuo at 105 °C.

IMPURITIES

Specified impurities A, B



A. ethenyl acetate,



B. acetic acid.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). Some of the characteristics described in the Functionality-

related characteristics section may also be present in the mandatory

part of the monograph since they also represent mandatory quality

criteria. In such cases, a cross-reference to the tests described in the

mandatory part is included in the Functionality-related

characteristics section. Control of the characteristics can contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for macroglol

poly(vinyl alcohol) grafted copolymer used as film former in film-

coated tablets.

Viscosity (2.2.10)

Typically less than 250 mPa·s, determined on a

20 per cent *m/m* solution, using a rotating viscometer at

25 °C and rotation speed of 100 r/min.

Macrogol 40 Sorbitol Heptaoleate

(Ph. Eur. monograph 2396)

Action and use

Non-ionic surfactant.

DEFINITION

Mixture of esters of fatty acids, mainly *Oleic acid* (0799), and sorbitol ethoxylated with approximately 40 moles of ethylene oxide for each mole of sorbitol. 7 moles of oleic acid are used for each mole of sorbitol. It also contains macrogol fatty acid esters.

CHARACTERS

Appearance

Clear or slightly opalescent, yellowish, viscous, hygroscopic liquid.

Solubility

Dispersible in water, soluble in isopropyl myristate, in isopropyl palmitate, in mineral oils and in vegetable fatty oils.

Relative density

About 1.0.

Viscosity (2.2.9)

About 175 mPa·s at 25 °C.

IDENTIFICATION

First identification A, D

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).
Comparison macrogol 40 sorbitol heptaoleate CRS.

B. Hydroxyl value (see Tests).
C. Saponification value (see Tests).

D. Composition of fatty acids (see Tests).

TESTS

Acid value (2.5.1)

Maximum 12.0, determined on 3.0 g.

Hydroxyl value (2.5.3, Method A)

22 to 55.

Peroxide value

Maximum 10.0.

Introduce 10.0 g into a 100 mL beaker and dissolve with

20 mL of glacial acetic acid R. Add 1 mL of saturated

potassium iodide solution R, mix and allow to stand for 1 min.

Add 50 mL of carbon dioxide-free water R and a magnetic

stirring bar. Titrate with 0.01 M sodium thiosulfate,

determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

Determine the peroxide value using the following expression:

$$\frac{m}{(n_1 - n_2) \times M \times 1000}$$

n_1 = volume of 0.01 M sodium thiosulfate required for the titration of the substance to be examined, in millilitres;

n_2 = volume of 0.01 M sodium thiosulfate required for the blank titration, in millilitres;

M = molarity of the sodium thiosulfate solution;

m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)
90 to 110, determined on 4.0 g.

Use 30.0 mL of 0.5 M alcoholic potassium hydroxide, heat under reflux for 60 min and add 50 mL of anhydrous ethanol R before carrying out the titration.

Composition of fatty acids (2.4.22, Method C)

Use the mixture of calibrating substances in Table 2.4.22-3. *Composition of the fatty-acid fraction of the substance:*

— myristic acid: maximum 5.0 per cent;

— palmitic acid: maximum 16.0 per cent;

— palmitoleic acid: maximum 8.0 per cent;

— stearic acid: maximum 6.0 per cent;

— oleic acid: minimum 58.0 per cent;

— linoleic acid: maximum 18.0 per cent;

— linolenic acid: maximum 4.0 per cent.

Ethylene oxide and dioxan (2.4.25, Method A)
Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.50 g.

Sulfated ash

Maximum 0.25 per cent.

Heat a silica crucible to redness for 30 min, allow to cool in

a desiccator and weigh. Evenly distribute 1.0 g of the

substance to be examined in the crucible and weigh. Dry at

100-105 °C for 1 h and ignite in a muffle furnace at

600 ± 25 °C, until the substance is thoroughly charred.

Carry out the test for sulfated ash (2.4.14) on the residue

obtained, starting from 'Moisten the substance to be

STORAGE

In an airtight container, protected from light.

Macrogol Stearate

(Ph. Eur. monograph 1234)

Action and use

Non-ionic surfactant.

DEFINITION

Mixture of monoesters and diesters of mainly stearic

(octadecanoic) acid and/or palmitic (hexadecanoic) acid and

macrocols. It may be obtained by ethoxylation or by

esterification of macrocols with stearic acid 50 (type I) or

stearic acid 95 (type II) (see Stearic acid (1474)). It may

contain free macrocols. The average polymer length is

equivalent to 6 to 100 ethylene oxide units per molecule

CHARACTERS

Appearance

White or slightly yellowish waxy mass.

Solubility

Soluble in ethanol (96 per cent) and in 2-propanol. Macrogol stearate corresponding to a product with 6 to 9 units of ethylene oxide per molecule is practically insoluble, but freely dispersible in water and miscible with fatty oils and with waxes. Macrogol stearate corresponding to a product with 20 to 100 units of ethylene oxide per molecule is soluble in water and practically insoluble in fatty oils and in waxes.



IDENTIFICATION

- A. Saponification value (see Tests).
B. Composition of fatty acids (see Tests).

TESTS

Alkalinity
Dissolve 2.0 g in alcohol R and dilute to 20 mL with the same solvent. To 2 mL of this solution add 0.05 mL of phenol red solution R. The solution is not red.

Melting point (2.2.15)
See Table 1234-1.

Melt about 10 g at 80-90 °C. Introduce a sufficient amount of the substance into the tube by capillary action to form a column of the prescribed height. Allow to stand at 0 °C for 2 h.

Acid value (2.5.1)
Maximum 2.0, determined on 2.0 g.

Hydroxyl value (2.5.3, Method A)
See Table 1234-1.

Iodine value (2.5.4)
Maximum 2.0.

Saponification value (2.5.6)
See Table 1234-1.

Table 1234-1

Ethylene oxide units per molecule (nominal value)	Melting point (°C)	Hydroxyl value	Saponification value
6	90 - 110	85 - 105	85 - 105
8 - 9	26 - 35	80 - 105	88 - 100
20	33 - 40	50 - 62	46 - 56
40 - 50	38 - 52	23 - 40	20 - 35
100	48 - 60	15 - 30	5 - 20

Composition of fatty acids

Gas chromatography (2.4.22, Method C).
Composition of the fatty acid fraction of the substance:

Type of fatty acid used	Composition of fatty acids
Macroglol stearate	Stearic acid 50 Sum of the contents of palmitic and stearic acids: 40.0 per cent to 60.0 per cent.
Macroglol stearate	Stearic acid 95 Sum of the contents of palmitic and stearic acids: 90.0 per cent to 99.0 per cent.
type II	Sum of the contents of palmitic and stearic acids: not less than 96.0 per cent.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.
2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 3.0 per cent, determined on 0.50 g. Use as the solvent a mixture of equal volumes of anhydrous methanol R and methylene chloride R.

Total ash (2.4.16)

Maximum 0.3 per cent, determined on 1.0 g.

Drying In air.

Development Over a path of 15 cm.

Application 20 µL.

Mobile phase ethyl acetate R.

Plate TLC silica gel plate R.

methanol R and dilute to 25 mL with the same solvent.

Reference solution Dissolve 25 mg of stearyl alcohol CRS in

may be opalescent).

50 mg of the residue in 10 mL of methanol R (the solution

and evaporate the combined filtrates to dryness. Dissolve

wash the filter with 3 quantities, each of 10 mL, of heptane R.

Filter the upper layer through anhydrous sodium sulfate R.

by the addition of a few drops of ethanol (96 per cent) R.

and shake for 3 min. The formation of foam can be reduced

with the same mixture of solvents. Add 60 mL of heptane R

water R and 9 volumes of methanol R and dilute to 75 mL

Test solution Dissolve 10.0 g in a mixture of 1 volume of

D. Thin-layer chromatography (2.2.27).

C. Saponification value (see Tests).

B. Iodine value (see Tests).

A. Hydroxyl value (see Tests).

IDENTIFICATION

After melting, it solidifies at about 45 °C.

(96 per cent) and in methylene chloride.

reacted per mole: soluble in water, in ethanol

— macroglol stearyl ether with 20 moles of ethylene oxide

(96 per cent);

reacted per mole: soluble in water and in ethanol

— macroglol stearyl ether with 10 moles of ethylene oxide

chloride;

ethanol (96 per cent) with heating and in methylene

— macroglol stearyl ether with 2 moles of ethylene oxide

reacted per mole: practically insoluble in water, soluble in

Solubility

microbeads or flakes.

White or yellowish-white, waxy, unctuous mass, pellets,

Appearance

CHARACTERS

value).

reacted per mole of stearyl alcohol is 2 to 20 (nominal

free stearyl alcohol. The number of moles of ethylene oxide

It may contain some free macrogols and various amounts of

Mixture of ethers obtained by ethoxylation of stearyl alcohol.

DEFINITION

Ph Eur

Non-ionic surfactant.

Action and use

(Ph. Eur. monograph 1340)

Macroglol Stearyl Ether



Ph Eur

— the type of macroglol stearate.

value),

— the number of ethylene oxide units per molecule (nominal

The label states:

LABELLING

In an airtight container.

STORAGE



Magaldrate

(Ph. Eur. monograph 1539)

$\text{Al}_5\text{Mg}_{10}(\text{OH})_{31}(\text{SO}_4)_2 \cdot x\text{H}_2\text{O}$

1097 74978-16-8

Action and use

Antacid.

Preparation

Magaldrate Oral Suspension

Ph Eur

DEFINITION

Magaldrate is composed of aluminium and magnesium hydroxides and sulfates. Its composition corresponds approximately to the formula $\text{Al}_5\text{Mg}_{10}(\text{OH})_{31}(\text{SO}_4)_2 \cdot x\text{H}_2\text{O}$.

Content

90.0 per cent to 105.0 per cent (dried substance). It contains a variable quantity of water.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It is soluble in dilute mineral acids.

IDENTIFICATION

A. Dissolve 0.6 g in 20 mL of 3 M hydrochloric acid R, add about 30 mL of water R and heat to boiling. Adjust to pH 6.2 with dilute ammonia R1, continue boiling for a further 2 min, filter and retain the precipitate and the filtrate. To 2 mL of the filtrate add 2 mL of ammonium chloride solution R and neutralise with a solution prepared by dissolving 2 g of ammonium carbonate R and 2 mL of dilute ammonia R1 in 20 mL of water R; no precipitate is produced. Add disodium hydrogen phosphate solution R; a white, crystalline precipitate is produced which does not dissolve in dilute ammonia R1.

B. The precipitate retained in identification test A gives the reaction of aluminium (2.3.1).

C. The filtrate retained in identification test A gives reaction (a) of sulfates (2.3.1).

TESTS

Soluble chlorides

Maximum 3.5 per cent.

To 0.5 g add 25 mL of dilute nitric acid R and shake until completely dissolved. Add 10.0 mL of 0.1 M silver nitrate and 2 mL of ferric ammonium sulfate solution R2 as indicator. Titrate with 0.1 M ammonium thiocyanate, shaking vigorously until a persistent brownish-red colour is obtained.

1 mL of 0.1 M silver nitrate is equivalent to 3.545 mg of Cl.

Soluble sulfates

Maximum 1.9 per cent.

Disperse 0.5 g in 25 mL of water R, boil for 5 min, cool, dilute to 25.0 mL with water R, mix and filter. To 2.5 mL of the filtrate, add 30 mL of water R, neutralise to blue litmus paper R with hydrochloric acid R, add 3 mL of 1 M hydrochloric acid, 3 mL of a 120 g/L solution of barium chloride R and dilute to 50 mL with water R. Mix and allow to stand for 10 min. Any opalescence in the solution is not more intense than that in a standard prepared at the same time in the same manner using 1 mL of 0.01 M sulfuric acid instead of 2.5 mL of filtrate.

Detection Spray with vanillin-sulfuric acid reagent prepared as follows: dissolve 0.5 g of vanillin R in 50 mL of ethanol (96 per cent) R and dilute to 100 mL with sulfuric acid R; allow to dry in air; heat at about 130 °C for 15 min and allow to cool in air.

Results The chromatogram obtained with the test solution shows several spots; one of these spots corresponds to the principal spot in the chromatogram obtained with the reference solution.

E. Dissolve or disperse 0.1 g in 5 mL of ethanol (96 per cent) R, add 2 mL of water R, 10 mL of dilute hydrochloric acid R, 10 mL of barium chloride solution R1 and 10 mL of a 100 g/L solution of phosphomolybdic acid R. A precipitate is formed.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Dissolve 5.0 g in ethanol (96 per cent) R and dilute to 50 mL with the same solvent.

Alkalinity

Dissolve 2.0 g in a hot mixture of 10 mL of ethanol (96 per cent) R and 10 mL of water R. Add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator to yellow.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

Number of moles of ethylene oxide reacted per mole (nominal value)	Hydroxyl value
2	150 - 180
10	75 - 90
20	40 - 60

Iodine value (2.5.4, Method A)

Maximum 2.0.

Saponification value (2.5.6)

Maximum 3.0, determined on 10.0 g.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.00 g.

STORAGE

In an airtight container.

LABELLING

The label states the number of moles of ethylene oxide reacted per mole of stearyl alcohol (nominal value).

Ph Eur

Ph Eur



Magnesium Acetate Tetrahydrate

(Ph. Eur. monograph 2035)

 $\text{Mg}(\text{CH}_3\text{COO})_2 \cdot 4\text{H}_2\text{O}$

214,5

16674-78-5

Action and use

Used in dialysis solutions.

Ph Eur

DEFINITION

Content

98,0 per cent to 101,0 per cent of magnesium acetate (anhydrous substance).

CHARACTERS

Appearance

Colourless crystals or white or almost white, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve about 100 mg in 2 mL of water R. Add 1 mL of dilute ammonia R1 and heat. A white precipitate is formed that dissolves slowly on addition of 5 mL of ammonium chloride solution R. Add 1 mL of disodium hydrogen phosphate solution R. A white crystalline precipitate is formed.

B. It gives reaction (b) of acetates (2.3.1).

TESTS

pH (2.2.3)

7,5 to 8,5.

Dissolve 2,5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Chlorides (2.4.4)

Maximum 330 ppm.

Dissolve 1,0 g in water R and dilute to 100 mL with the same solvent.

Nitrates

Maximum 3 ppm.

Dissolve 1,0 g in distilled water R and dilute to 10 mL with the same solvent, add 5 mg of sodium chloride R, 0,05 mL of indigo carmine solution R and while stirring, 10 mL of nitrogen-free sulfuric acid R. A blue colour is produced which persists for at least 10 min.

Sulfates (2.4.13)

Maximum 600 ppm.

Dissolve 0,25 g in distilled water R and dilute to 15 mL with the same solvent.

Sulfates

16,0 per cent to 21,0 per cent (dried substance).

Dissolve 0,875 g in a mixture of 5 mL of glacial acetic acid R and 10 mL of water R and dilute to 25,0 mL with water R.

Prepare a chromatographic column of 1 cm in internal diameter containing 15 mL of cation-exchange resin R

(150-300 µm), previously washed with 30 mL of water R.

Transfer 5,0 mL of the solution to be examined to the column and elute with 15 mL of water R. To the eluate add 5 mL of a 53,6 g/L solution of magnesium acetate R, 32 mL of methanol R and 0,2 mL of alizarin S solution R. Add from a burette about 4,0 mL of 0,05 M barium chloride, add a further 0,2 mL of alizarin S solution R and slowly complete the titration until the yellow colour disappears and a violet-red tinge is visible.

1 mL of 0,05 M barium chloride is equivalent to 4,803 mg of SO_4 .

Aluminium hydroxide

32,1 per cent to 45,9 per cent (dried substance).

Dissolve 0,800 g in 10 mL of dilute hydrochloric acid R, heating on a water-bath. Cool and dilute to 50,0 mL with water R. To 10,0 mL of this solution, add dilute ammonia R1 until a precipitate begins to appear. Add the smallest quantity of dilute hydrochloric acid R needed to dissolve the precipitate and dilute to 20 mL with water R. Carry out the complexometric titration of aluminium (2.5.11).

1 mL of 0,1 M sodium edetate is equivalent to 7,80 mg of $\text{Al}(\text{OH})_3$.

Magnesium hydroxide

49,2 per cent to 66,6 per cent (dried substance).

Dissolve 0,100 g in 2 mL of dilute hydrochloric acid R and transfer to a 500 mL conical flask with the aid of water R.

Dilute to 200 mL with water R, add 20 mL of triethanolamine R with shaking, 10 mL of ammonium chloride buffer solution pH 10,0 R and about 50 mg of mordant black 11 indicator R. Titrate with 0,1 M sodium edetate until the colour changes from violet to pure blue.

1 mL of 0,1 M sodium edetate is equivalent to 5,832 mg of $\text{Mg}(\text{OH})_2$.

Sodium

Maximum 0,10 per cent.

Atomic absorption spectrometry (2.2.23, Method I).

Test solution Weigh 2,00 g into a 100 mL volumetric flask, place in an ice-bath, add 5 mL of nitric acid R and swirl to mix. Allow to warm to room temperature and dilute to 100 mL with water R. Filter, if necessary, to obtain a clear solution. Dilute 10,0 mL of the filtrate to 100,0 mL with water R.

Reference solutions Prepare the reference solutions using sodium standard solution (200 ppm Na) R, diluted as necessary with dilute nitric acid R.

Source Sodium hollow-cathode lamp.

Wavelength 589 nm.

Atomisation device Air-acetylene flame.

Heavy metals (2.4.8)

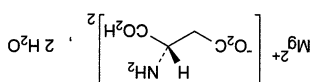
Maximum 30 ppm.

Dissolve 2,0 g in 30 mL of hydrochloric acid R1 and shake with 50 mL of methyl isobutyl ketone R for 2 min. Allow to stand, then separate and evaporate the aqueous layer to dryness. Dissolve the residue in 30 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.



Magnesium Aspartate

(Magnesium Aspartate Dihydrate,
Ph Eur monograph 1445)



$\text{C}_8\text{H}_{12}\text{MgN}_2\text{O}_8 \cdot 2\text{H}_2\text{O}$ 324.5 72231-13-1 Ph Eur

DEFINITION

Magnesium di[(5)-2-aminohydrogenobutane-1,4-dioate] dihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Examine the chromatograms obtained in the test for ninhydrin-positive substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Ignite about 15 mg until a white residue is obtained. Dissolve the residue in 1 mL of dilute hydrochloric acid R_1 , neutralise to red litmus paper R by adding dilute sodium hydroxide solution R and filter if necessary. The solution gives the reaction of magnesium (2.3.1).

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)
6.0 to 8.0 for solution S.

Specific optical rotation (2.2.7)
+ 22.0 to + 24.0 (anhydrous substance).

Dissolve 0.50 g in a 515 g/L solution of hydrochloric acid R and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances

Test solution (a) Dissolve 0.10 g of the substance to be examined in water R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with water R .

Reference solution (a) Dissolve 10 mg of magnesium aspartate dihydrate CRS in water R and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 5 mL of test solution (b) to 20 mL with water R .

Reference solution (c) Dissolve 10 mg of glutamic acid CRS and 10 mg of magnesium aspartate dihydrate CRS in 2 mL of water R and dilute to 25 mL with the same solvent.

Aluminium (2.4.17)

Maximum 1 ppm.

Prescribed solution Dissolve 4.0 g in water R and dilute to 100 mL with the same solvent. Add 10 mL of acetate buffer solution pH 6.0 R .

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R , 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R .

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R .

Calcium (2.4.3)

Maximum 100 ppm.

Dissolve 1.0 g in distilled water R and dilute to 15 mL with the same solvent.

Potassium

Maximum 0.1 per cent.

Atomic emission spectrometry (2.2.22, Method II).
Test solution Dissolve 0.5 g in water R and dilute to 100 mL with the same solvent.

Reference solutions Prepare the reference solutions using potassium standard solution (600 ppm K) R , diluted as necessary with water R .

Wavelength 766.5 nm.

Sodium

Maximum 0.5 per cent.

Atomic emission spectrometry (2.2.22, Method II).
Test solution Dissolve 1.0 g in water R and dilute to 100 mL with the same solvent.

Reference solutions Prepare the reference solutions using sodium standard solution (200 ppm Na) R , diluted as necessary with water R .

Heavy metals (2.4.8)
Wavelength 589.0 nm.

Dissolve 1.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R .

Readily oxidisable substances

Dissolve 2.0 g in 100 mL of boiling water R , add 6 mL of a 150 g/L solution of sulfuric acid R and 0.3 mL of 0.02 M potassium permanganate. Mix and boil gently for 5 min. The pink colour is not completely discharged.

Water (2.5.12)

33.0 per cent to 35.0 per cent, determined on 0.100 g.

ASSAY

Dissolve 0.150 g in 300 mL of water R . Carry out the complexometric titration of magnesium (2.5.17).
1 mL of 0.1 M sodium edetate is equivalent to 14.24 mg of $\text{C}_4\text{H}_6\text{MgO}_4$.

Ph Eur

Heavy Magnesium Carbonate

(Ph. Eur. monograph 0043)

Action and use

Antacid; osmotic laxative.

Ph Eur

DEFINITION

Hydrated basic magnesium carbonate.

Content

40.0 per cent to 45.0 per cent, calculated as MgO

(M_r 40.30).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water. It dissolves in dilute acids with effervescence.

IDENTIFICATION

A. Bulk density (2.9.34): minimum 0.25 g/mL.

B. It gives the reaction of carbonates (2.3.1).

C. Dissolve about 15 mg in 2 mL of dilute nitric acid R and neutralise with dilute sodium hydroxide solution R. The solution gives the reaction of magnesium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in 100 mL of dilute acetic acid R. When the effervescence has ceased, boil for 2 min, allow to cool and dilute to 100 mL with dilute acetic acid R. Filter, if necessary, through a previously ignited and tared porcelain or silica filter

crucible of suitable porosity to give a clear filtrate.

Appearance of solution

Solution S is not more intensely coloured than reference solution B₄ (2.2.2, Method II).

Soluble substances

Maximum 1.0 per cent.

Mix 2.00 g with 100 mL of water R and boil for 5 min. Filter whilst hot through a sintered-glass filter (40) (2.1.2), allow to cool and dilute to 100 mL with water R. Evaporate 50 mL of the filtrate to dryness and dry at 100-105 °C. The residue weighs not more than 10 mg.

Substances insoluble in acetic acid

Maximum 0.05 per cent.

Any residue obtained during the preparation of solution S, washed, dried, and ignited at 600 ± 50 °C, weighs not more than 2.5 mg.

Chlorides (2.4.4)

Maximum 700 ppm.

Dilute 1.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 0.6 per cent.

Dilute 0.5 mL of solution S to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 10 mL of solution S.

Calcium (2.4.3)

Maximum 0.75 per cent.

Dilute 2.6 mL of solution S to 150 mL with distilled water R.

Iron (2.4.9)

Maximum 400 ppm.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R

(20:20:60 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C

for 15 min.

System suitability: reference solution (c): the chromatogram

shows 2 clearly separated principal spots.

Limit:

— any impurity: any spot in the chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with reference solution (b)

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 10 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 500 ppm.

Dilute 12 mL of solution S to 15 mL with distilled water R.

Carry out the evaluation of the test after 30 min.

Ammonium (2.4.1)

Maximum 200 ppm.

50 mg complies with test B. Prepare the standard using 0.1 mL of ammonium standard solution (100 ppm NH₄) R.

Iron (2.4.9)

Maximum 50 ppm.

In a separating funnel, dissolve 0.20 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R₁, shaking for 3 min each time.

To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g with gentle heating in 20 mL of water R.

12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

10.0 per cent to 14.0 per cent, determined on 0.100 g.

Dissolve the substance in 10 mL of formamide R₁ at 50 °C

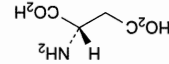
protected from moisture, add 10 mL of anhydrous methanol R and allow to cool. Carry out a blank determination.

ASSAY

Dissolve 0.260 g in 10 mL of water R and carry out the complexometric titration of magnesium (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 28.85 mg of C₈H₁₂MgN₂O₈.

IMPURITIES



A. (2S)-2-aminobutanedioic acid (aspartic acid).

Ph Eur



B. It gives the reaction of carbonates (2.3.1).
C. Dissolve about 15 mg in 2 mL of dilute nitric acid R and neutralise with dilute sodium hydroxide solution R. The solution gives the reaction of magnesium (2.3.1).

TESTS

Solution S
Dissolve 5.0 g in 100 mL of dilute acetic acid R. When the effervescence has ceased, boil for 2 min, allow to cool and dilute to 100 mL with dilute acetic acid R. Filter, if necessary, through a previously ignited and tared porcelain or silica filter crucible of suitable porosity to give a clear filtrate.

Appearance of solution

Solution S is not more intensely coloured than reference

solution B₄ (2.2.2, Method II).

Soluble substances

Maximum 1.0 per cent.
Mix 2.00 g with 100 mL of water R and boil for 5 min. Filter whilst hot through a sintered-glass filter (40) (2.1.2), allow to cool and dilute to 100 mL with water R. Evaporate 50 mL of the filtrate to dryness and dry at 100–105 °C. The residue weighs a maximum of 10 mg.

Substances insoluble in acetic acid

Maximum 0.05 per cent.
Any residue obtained during the preparation of solution S, washed, dried and ignited at 600 ± 50 °C, weighs a maximum of 2.5 mg.

Chlorides (2.4.4)

Maximum 700 ppm.
Dilute 1.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 0.3 per cent.
Dilute 1 mL of solution S to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 10 mL of solution S.

Calcium (2.4.3)

Maximum 0.15 per cent.
Dilute 2.6 mL of solution S to 150 mL with distilled water R. 15 mL of the solution complies with the test.

Iron (2.4.9)

Maximum 400 ppm.
Dissolve 0.1 g in 3 mL of dilute hydrochloric acid R and dilute to 10 mL with water R. Dilute 2.5 mL of this solution to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.
To 20 mL of solution S add 15 mL of hydrochloric acid R1 and shake with 25 mL of methyl isobutyl ketone R for 2 min. Allow to stand, separate the aqueous lower layer and evaporate to dryness. Dissolve the residue in 1 mL of acetic acid R and dilute to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

Dissolve 0.150 g in a mixture of 2 mL of dilute hydrochloric acid R and 20 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).
1 mL of 0.1 M sodium edetate is equivalent to 4.030 mg of MgO.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more

Dissolve 0.1 g in 3 mL of dilute hydrochloric acid R and dilute to 10 mL with water R. Dilute 2.5 mL of the solution to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.
To 20 mL of solution S add 15 mL of hydrochloric acid R1 and shake with 25 mL of methyl isobutyl ketone R for 2 min. Allow to stand, separate the aqueous lower layer and evaporate to dryness. Dissolve the residue in 1 mL of acetic acid R and dilute to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

Dissolve 0.150 g in a mixture of 2 mL of dilute hydrochloric acid R and 20 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).
1 mL of 0.1 M sodium edetate is equivalent to 4.030 mg of MgO.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are also to be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for heavy magnesium carbonate used as filler in tablets.

Particle-size distribution (2.9.31 or 2.9.38).

Bulk and tapped density (2.9.34)

Light Magnesium Carbonate

(Ph. Eur. monograph 0042)

Action and use

Antacid; osmotic laxative.

Preparation

Aromatic Magnesium Carbonate Mixture

DEFINITION

Hydrated basic magnesium carbonate.

Content

40.0 per cent to 45.0 per cent, calculated as MgO (M_r 40.30).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water. It dissolves in dilute acids with effervescence.

IDENTIFICATION

A. Bulk density (2.9.34): maximum 0.15 g/mL.

functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for light magnesium carbonate used as filler in oral solid dosage forms.

Particle-size distribution (2.9.31 or 2.9.38),
Bulk and tapped density (2.9.34)

Ph Eur

Magnesium Chloride Hexahydrate

(Ph. Eur. monograph 0402)
 $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$ 203.3 7791-18-6

Action and use
Used in the treatment of electrolyte deficiencies and in dialysis solutions.

Preparation
Magnesium Chloride Injection

Ph Eur

DEFINITION

Content
98.0 per cent to 101.0 per cent of $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$.

CHARACTERS

Appearance
Colourless crystals, hygroscopic.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Water (see Tests).
B. It gives reaction (a) of chlorides (2.3.1).
C. It gives the reaction of magnesium (2.3.1).

TESTS

Solution S
Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 5 mL of solution S add 0.05 mL of phenol red solution R. Not more than 0.3 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Bromides

Maximum 500 ppm.
Dilute 2.0 mL of solution S to 10.0 mL with water R. To 1.0 mL of this solution add 4.0 mL of water R, 2.0 mL of phenol red solution R3 and 1.0 mL of chloramine solution R2

and mix immediately. After exactly 2 min, add 0.30 mL of 0.1 M sodium thiosulfate, mix and dilute to 10.0 mL with water R. The absorbance (2.2.25) of the solution measured at 590 nm, using water R as the compensation liquid, is not greater than that of a standard prepared at the same time and in the same manner using 5.0 mL of a 3 mg/L solution of potassium bromide R.

Sulfates (2.4.13)

Maximum 100 ppm, determined on solution S.

Aluminium (2.4.17)

Maximum 1 ppm, if intended for use in the manufacture of peritoneal dialysis solutions, haemodialysis solutions, or haemofiltration solutions.

Prescribed solution Dissolve 4 g in 100 mL of water R and add 10 mL of acetate buffer solution pH 6.0 R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Calcium (2.4.3)

Maximum 0.1 per cent.

Dilute 1 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 10 ppm, determined on solution S.

Potassium

Maximum 500 ppm, if intended for use in the manufacture of parenteral preparations.

Atomic emission spectrometry (2.2.22, Method D).

Test solution Dissolve 1.00 g in water R and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using the following solution, diluted as necessary with water R: dissolve 1.144 g of potassium chloride R, previously dried at 100-105 °C for 3 h in water R and dilute to 1000.0 mL with the same solvent (600 µg of K per millilitre).

Wavelength 766.5 nm.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

51.0 per cent to 55.0 per cent, determined on 50.0 mg.

ASSAY

Dissolve 0.300 g in 50 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).
1 mL of 0.1 M sodium edetate is equivalent to 20.33 mg of $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$.

STORAGE

In an airtight container.

LABELLING

The label states:

— where applicable, that the substance is suitable for use in the manufacture of peritoneal dialysis solutions, haemodialysis solutions or haemofiltration solutions, — where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Partially Hydrated Magnesium Chloride

(Magnesium Chloride 4.5-Hydrate,
Ph Eur monograph 1341)

$\text{MgCl}_2 \cdot x\text{H}_2\text{O}$ with $x \approx 4.5$ 95.21 (anhydrous)

DEFINITION

Content

52.5 per cent to 55.5 per cent (calculated on an as-is basis, without allowing for the results of the test for water).

CHARACTERS

Appearance

White or almost white, hygroscopic, granular powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Water (see Tests).

B. It gives reaction (a) of chlorides (2.3.1).

C. It gives the reaction of magnesium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 5 mL of solution S add 0.05 mL of phenol red solution R.

Not more than 0.3 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Bromides

Maximum 500 ppm.

Dilute 2.0 mL of solution S to 10.0 mL with water R.

To 1.0 mL of the solution add 4.0 mL of water R, 2.0 mL of phenol red solution R3 and 1.0 mL of chloramine solution R2

and mix immediately. After exactly 2 min, add 0.30 mL of 0.1 M sodium thiosulfate, mix and dilute to 10.0 mL with water R. The absorbance (2.2.25) of the solution measured at 590 nm, using water R as the compensation liquid, is not greater than that of a standard prepared at the same time and in the same manner using 5.0 mL of a 3 mg/L solution of potassium bromide R.

Sulfates (2.4.13)

Maximum 100 ppm, determined on solution S.

Aluminium (2.4.17)

Maximum 1 ppm, if intended for use in the manufacture of peritoneal dialysis solutions, haemodialysis solutions, or haemofiltration solutions.

Prescribed solution Dissolve 4 g in 100 mL of water R and add 10 mL of acetate buffer solution pH 6.0 R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Calcium (2.4.3)

Maximum 0.1 per cent.

Dilute 1 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 10 ppm, determined on solution S.

Potassium

Maximum 500 ppm, if intended for use in the manufacture of parenteral preparations.

Atomic emission spectrometry (2.2.22, Method I).

Test solution Dissolve 1.00 g in water R and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using the following solution, diluted as necessary with water R: dissolve 1.144 g of potassium chloride R, previously dried at 100-105 °C for 3 h, in water R and dilute to 1000.0 mL with the same solvent (600 µg of K per millilitre).

Wavelength 766.5 nm.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

44.0 per cent to 48.0 per cent, determined on 50.0 mg.

ASSAY

Dissolve 0.250 g in 50 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 9.521 mg of MgCl_2 .

STORAGE

In an airtight container.

LABELLING

The label states:

— where applicable, that the substance is suitable for use in haemodialysis solutions or haemofiltration solutions;

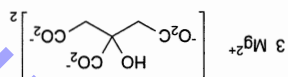
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Ph Eur



Anhydrous Magnesium Citrate

(Ph. Eur. monograph 2339)



$\text{Mg}_3(\text{C}_6\text{H}_5\text{O}_7)_2$

451.1

3344-18-1

Ph Eur

DEFINITION

Trimagnesium bis(2-hydroxypropane-1,2,3-tricarboxylate).

Content

15.0 per cent to 16.5 per cent of Mg (dried substance).

CHARACTERS

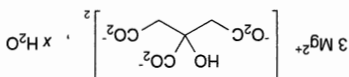
Appearance

White or almost white, fine, slightly hygroscopic powder.



Magnesium Citrate Dodecahydrate

(Ph Eur monograph 2401)



$\text{Mg}_3(\text{C}_6\text{H}_5\text{O}_7)_{2,x}\text{H}_2\text{O}$ 451.1 (anhydrous substance) with $x \approx 12$

DEFINITION

Trimagnesium bis(2-hydroxypropane-1,2,3-tricarboxylate) dodecahydrate.

Content

15.0 per cent to 16.5 per cent of Mg (dried substance).

CHARACTERS

Appearance

White or almost white, fine powder.

Solubility

Sparsely soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute hydrochloric acid.

IDENTIFICATION

- A. It gives the reaction of citrates (2.3.1).
B. It gives the reaction of magnesium (2.3.1).
C. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 2.5 g in 15 mL of dilute hydrochloric acid R with heating. Cool and dilute to 100 mL with distilled water R.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)

6.0 to 8.5.

Disperse 5.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent. Centrifuge and measure the pH of the clear supernatant.

Oxalates

Maximum 280 ppm.

Dissolve 0.50 g in a mixture of 3 mL of hydrochloric acid R and 4 mL of water R and add 1 g of activated zinc R. Allow to stand for 5 min. Transfer the liquid to a tube containing 0.25 mL of a 10 g/L solution of phenylhydrazine

hydrochloride R. Heat to boiling. Cool rapidly, transfer to a graduated cylinder and add an equal volume of hydrochloric acid R and 0.25 mL of potassium ferricyanide solution R. Shake and allow to stand for 30 min. Any pink colour in the solution is not more intense than that of a standard prepared at the same time and in the same manner using 4 mL of a 50 mg/L solution of oxalic acid R.

Sulfates (2.4.13)

Maximum 0.2 per cent.

Dilute 3.0 mL of solution S to 15 mL with distilled water R.

Calcium (2.4.3)

Maximum 0.2 per cent.

To a mixture of 2 mL of solution S and 8 mL of distilled water R, add about 0.2 mL of ammonia R and dilute to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 100 ppm.

TESTS

D. Loss on drying (see Tests).

C. pH (see Tests).

B. It gives the reaction of magnesium (2.3.1).

A. It gives the reaction of citrates (2.3.1).

IDENTIFICATION

Soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute hydrochloric acid.

Solubility

Solution S

Dissolve 5.0 g in carbon dioxide-free water R, heating at 60 °C, cool and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension III (2.2.1) and not more intensely coloured than reference solutions Y₇ or BY₆ (2.2.2, Method II).

pH (2.2.3)

6.0 to 8.5 for solution S.

Oxalates

Maximum 280 ppm.

Dissolve 0.50 g in 4 mL of water R. Add 3 mL of hydrochloric acid R and 1 g of activated zinc R. Allow to stand for 5 min. Transfer the liquid to a tube containing 0.25 mL of a 10 g/L solution of phenylhydrazine hydrochloride R. Heat to boiling.

Cool rapidly, transfer to a graduated cylinder and add an equal volume of hydrochloric acid R and 0.25 mL of potassium ferricyanide solution R. Shake and allow to stand for 30 min.

Any pink colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 4 mL of a 50 mg/L solution of oxalic acid R.

Sulfates (2.4.13)

Maximum 0.2 per cent.

Dilute 1.5 mL of solution S to 15 mL with distilled water R.

Calcium (2.4.3)

Maximum 0.2 per cent.

Dilute 1.0 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 100 ppm.

Dilute 2.0 mL of solution S to 10 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 5.0 g in 15 mL of dilute hydrochloric acid R with heating. Adjust to pH 3.5 with ammonia R and dilute to 50 mL with distilled water R. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 3.5 per cent, determined on 1.000 g by drying in an oven at 180 ± 10 °C for 5 h.

ASSAY

Dissolve 0.150 g in 50 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 2.431 mg of Mg.

STORAGE

In a non-metallic, airtight container.

Ph Eur

Dilute 4.0 mL of solution S to 10 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 5.0 g in 15 mL of dilute hydrochloric acid R with

heating. Adjust to pH 3.5 with ammonia R and dilute to

50 mL with distilled water R. 12 mL of the solution complies

solution (1 ppm Pb) R.

Loss on drying (2.2.32)

29.0 per cent to 36.0 per cent, determined on 1.000 g by

drying in an oven at $180 \pm 10^\circ\text{C}$ for 5 h.

ASSAY

Dissolve 0.200 g in 5 mL of dilute hydrochloric acid R with

heating. Cool and add 50 mL of water R. Adjust to pH 7.0

with ammonia R. Carry out the complexometric titration of

magnesium (2.5.11).

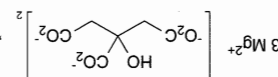
1 mL of 0.1 M sodium edetate is equivalent to 2.431 mg of

Mg.

Ph Eur

Magnesium Citrate Nonahydrate

(Ph. Eur. monograph 2402)



$\text{Mg}_3(\text{C}_6\text{H}_5\text{O}_7)_2 \cdot 9\text{H}_2\text{O}$

613

153531-96-5

Ph Eur

DEFINITION

Trimagnesium bis(2-hydroxypropane-1,2,3-tricarboxylate)

nonahydrate.

Content

15.0 per cent to 16.5 per cent of Mg (dried substance).

CHARACTERS

Appearance

White or almost white, fine powder.

Solubility

Sparingly soluble in water, practically insoluble in ethanol

(96 per cent). It dissolves in dilute hydrochloric acid.

IDENTIFICATION

A. It gives the reaction of citrates (2.3.1).

B. It gives the reaction of magnesium (2.3.1).

C. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 2.5 g in 15 mL of dilute hydrochloric acid R with

heating. Cool and dilute to 100 mL with distilled water R.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)

6.0 to 8.5.

Disperse 5.0 g in carbon dioxide-free water R and dilute to

100 mL with the same solvent. Centrifuge and measure the

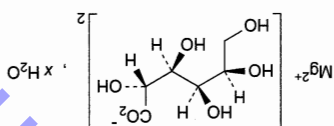
pH of the clear supernatant.

Oxalates

Maximum 280 ppm.

Magnesium Gluconate

(Ph. Eur. monograph 2161)



$\text{C}_{12}\text{H}_{22}\text{MgO}_{14} \cdot x\text{H}_2\text{O}$ 414.6 (anhydrous substance)

Ph Eur

DEFINITION

Anhydrous or hydrated magnesium bis[(2R,3S,4R,5R)-

2,3,4,5,6-pentahydroxyhexanoate] (anhydrous or hydrated

magnesium di(D-gluconate)).

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, amorphous, hygroscopic, crystalline or granular powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of water R.

Reference solution Dissolve 20 mg of calcium gluconate CRS in 1 mL of water R, heating if necessary in a water-bath at 60 °C.

Plate TLC silica gel plate R (5–40 µm) [or TLC silica gel plate R (2–10 µm)].

Mobile phase concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R (10:10:30:50 V/V/V/V).

Application 1 µL.

Development Over 3/4 of the plate.

Drying At 105 °C for 20 min, then allow to cool to room temperature.

Detection Spray with a solution containing 25 g/L of ammonium molybdate R and 10 g/L of cerium sulfate R in dilute sulfuric acid R, then heat at 105 °C for about 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. To 10 mL of solution S (see Tests) add 3 mL of ammonium chloride solution R. A slight opalescence may be observed. Add 10 mL of disodium hydrogen phosphate solution R. A white precipitate is formed that does not dissolve upon the addition of 2 mL of dilute ammonia R1.

TESTS

Solution S
Dissolve 1.0 g in water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Sucrose and reducing sugars

Dissolve 0.5 g in a mixture of 2 mL of hydrochloric acid R1 and 10 mL of water R. Boil for 5 min, allow to cool, add 10 mL of sodium carbonate solution R and allow to stand for 10 min. Dilute to 25 mL with water R and filter. To 5 mL of the filtrate add 2 mL of cupri-tartrate solution R and boil for 1 min. Allow to stand for 2 min. No red precipitate is formed.

Chlorides (2.4.4)

Maximum 500 ppm.
Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 500 ppm.
Dissolve 2.0 g in a mixture of 10 mL of acetic acid R and 90 mL of distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.
Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

TESTS

A. Mix 1 g with 1 g of potassium hydrogen sulfate R in a test tube fitted with a glass tube. Heat strongly and direct the white vapour towards a piece of filter paper impregnated with a freshly prepared 10 g/L solution of sodium nitroprusside R. The filter paper develops a blue colour in contact with piperidine R.
B. Ignite 0.1 g in a crucible. Take up the residue with 5 mL of nitric acid R and heat on a water-bath for 1 min. Filter. The filtrate gives reaction (b) of phosphates (2.3.1).
C. It gives the reaction of magnesium (2.3.1).

IDENTIFICATION

A. Mix 1 g with 1 g of potassium hydrogen sulfate R in a test tube fitted with a glass tube. Heat strongly and direct the white vapour towards a piece of filter paper impregnated with a freshly prepared 10 g/L solution of sodium nitroprusside R. The filter paper develops a blue colour in contact with piperidine R.

Solubility

Practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of acids.

Appearance

White or almost white powder, hygroscopic.
Maximum 12.0 per cent of Mg (dried substance).

CHARACTERS

Content
11.0 per cent to 12.5 per cent of Mg (dried substance).

DEFINITION

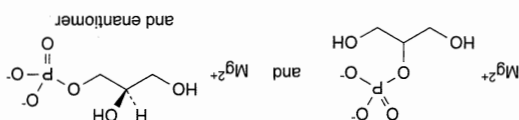
Mixture, in variable proportions, of magnesium salts of (RS)-2,3-dihydroxypropyl phosphate and 2-hydroxy-1-(hydroxymethyl)ethyl phosphate, which may be hydrated.

Preparations

Action and use

Excipient.

$C_3H_7MgO_6P$ 194.4



(Ph. Eur. monograph 1446)

Magnesium Glycerocephosphate



STORAGE

In an airtight container.
I mL of 0.1 M sodium edetate is equivalent to 41.46 mg of $C_{12}H_{22}MgO_{14}$.
Dissolve 0.350 g in 100 mL of water R and carry out the complexometric titration of magnesium (2.5.11).

ASSAY

TYMC: acceptance criterion 10^2 CFU/g (2.6.12).
TAMC: acceptance criterion 10^3 CFU/g (2.6.12).

Microbial contamination

Maximum 12.0 per cent, determined on 80 mg.

Water (2.5.32)

Dissolve 2.5 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.



1309-42-8

58.32

Mg(OH)₂

(Ph. Eur. monograph 0039)

Magnesium Hydroxide

Action and use

Antacid; osmotic laxative.

Preparations

Co-magaldrox Oral Suspension

Co-magaldrox Tablets

Ph. Eur.

DEFINITION

Content

95.0 per cent to 100.5 per cent of Mg(OH)₂.

CHARACTERS

Appearance

White or almost white, fine, amorphous powder.

Solubility

Practically insoluble in water. It dissolves in dilute acids.

IDENTIFICATION

A. Dissolve about 15 mg in 2 mL of dilute nitric acid R and neutralise with dilute sodium hydroxide solution R. The solution

gives the reaction of magnesium (2.3.1).

B. Loss on ignition (see Tests).

TESTS

Solution S

Dissolve 5.0 g in a mixture of 50 mL of acetic acid R and 50 mL of distilled water R. Not more than slight effervescence is produced. Boil for 2 min, cool and dilute to 100 mL with dilute acetic acid R. Filter, if necessary, through a previously ignited and tared porcelain or silica filter crucible of suitable porosity to give a clear filtrate.

Appearance of solution

Solution S is not more intensely coloured than reference solution B₃ (2.2.2, Method II).

Soluble substances

Maximum 2.0 per cent.

Mix 2.00 g with 100 mL of water R and boil for 5 min. Filter whilst hot through a sintered-glass filter (40) (2.1.2), allow to cool and dilute to 100 mL with water R. Evaporate 50 mL of the filtrate to dryness and dry at 100-105 °C. The residue weighs not more than 20 mg.

Substances insoluble in acetic acid

Maximum 0.1 per cent.

Any residue obtained during the preparation of solution S, washed, dried, and ignited at 600 ± 50 °C, weighs not more than 5 mg.

Chlorides (2.4.4)

Maximum 0.1 per cent.

Dilute 1 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 1.0 per cent.

Dilute 0.3 mL of solution S to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 4 ppm, determined on 5 mL of solution S.

Calcium (2.4.3)

Maximum 1.5 per cent.

Dilute 1.3 mL of solution S to 150 mL with distilled water R.

Appearance of solution

Solution S is not more opalescent than reference suspension III (2.2.1).

Acidity

Dissolve 1.0 g in 100 mL of carbon dioxide-free water R. Add 0.1 mL of phenolphthalein solution R. Not more than 1.5 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Glycerol and ethanol (96 per cent)-soluble substances

Maximum 1.5 per cent.

Shake 1.0 g with 25 mL of ethanol (96 per cent) R for 2 min. Filter and wash the residue with 5 mL of ethanol (96 per cent) R. Combine the filtrate and the washings, evaporate to dryness on a water-bath and dry the residue at 70 °C for 1 h. The residue weighs a maximum of 15 mg.

Chlorides (2.4.4)

Maximum 0.15 per cent.

Dissolve 1.0 g in water R and dilute to 100 mL with the same solvent. Dilute 3.5 mL of this solution to 15 mL with water R.

Phosphates (2.4.11)

Maximum 0.5 per cent.

Dilute 4 mL of solution S to 100 mL with water R. Dilute 1 mL of this solution to 100 mL with water R.

Sulfates (2.4.13)

Maximum 0.1 per cent.

Dilute 3 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 150 ppm.

Dissolve 67 mg in water R and dilute to 10 mL with the same solvent.

Heavy metals (2.4.8)

Maximum 20 ppm.

To 20 mL of solution S add 15 mL of hydrochloric acid R and shake with 25 mL of methyl isobutyl ketone R for 2 min. Allow to stand, then separate and evaporate the aqueous layer to dryness. Dissolve the residue in 2.5 mL of acetic acid R and dilute to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 12.0 per cent, determined on 1.000 g by drying in an oven at 150 °C for 4 h.

ASSAY

Dissolve 0.200 g in 40 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).

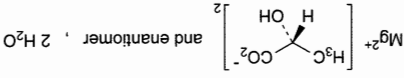
1 mL of 0.1 M sodium edetate is equivalent to 2.431 mg of Mg.

STORAGE

In an airtight container.

Ph. Eur.

Iron (2.4.9)
Maximum 0.07 per cent.
Dissolve 0.15 g in 5 mL of dilute hydrochloric acid R and dilute to 10 mL with water R.
Heavy metals (2.4.8)
Maximum 30 ppm.
Dissolve 2.0 g in 20 mL of hydrochloric acid R1 and shake with 25 mL of methyl isobutyl ketone R for 2 min. Allow to stand, separate the aqueous layer and evaporate to dryness. Dissolve the residue in 30 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.
Loss on ignition
Heat 0.5 g gradually to 900 ± 50 °C and ignite to constant mass.
ASSAY
Dissolve 0.100 g in a mixture of 2 mL of dilute hydrochloric acid R and 20 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).
1 mL of 0.1 M sodium edetate is equivalent to 5.832 mg of Mg(OH)₂.

Magnesium Lactate Dihydrate
(Ph. Eur. monograph 2160)

 $C_6H_{10}MgO_6 \cdot 2H_2O$ 238.5
Ph Eur

DEFINITION
Magnesium bis(2-hydroxypropionate) or mixture of magnesium (2R)-, (2S)- and (2RS)-2-hydroxypropionate dihydrate.
Content
98.0 per cent to 102.0 per cent (dried substance).
CHARACTERS
Appearance
White or almost white, crystalline or granular powder.
Solubility
Slightly soluble in water, soluble in boiling water, practically insoluble in ethanol (96 per cent).
IDENTIFICATION
A. It gives the reaction of lactates (2.3.1).
B. It gives the reaction of magnesium (2.3.1).
TESTS
Solution S
Dissolve 5.0 g with heating in carbon dioxide-free water R prepared from distilled water R, allow to cool and dilute to 100 mL with the same solvent.
Appearance of solution
Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)
6.5 to 8.5 for solution S.
Chlorides (2.4.4)
Maximum 200 ppm.
Dilute 5 mL of solution S to 15 mL with water R.
Sulfates (2.4.13)
Maximum 400 ppm.
Dilute 7.5 mL of solution S to 15 mL with distilled water R.
Iron (2.4.9)
Maximum 50 ppm.
Dilute 4 mL of solution S to 10 mL with water R.
Heavy metals (2.4.8)
Maximum 20 ppm.
12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.
Loss on drying (2.2.32)
14.0 per cent to 17.0 per cent, determined on 0.500 g by drying in an oven at 125 °C.
ASSAY
Dissolve 0.180 g in water R and dilute to 300 mL with the same solvent. Carry out the complexometric titration of magnesium (2.5.11).
1 mL of 0.1 M sodium edetate is equivalent to 20.25 mg of C₆H₁₀MgO₆.

Heavy Magnesium Oxide
(Ph. Eur. monograph 0041)
MgO 40.30
1309-48-4
Ph Eur

DEFINITION
Content 98.0 per cent to 100.5 per cent of MgO (ignited substance).
CHARACTERS
Appearance
Fine, white or almost white powder.
Solubility
Practically insoluble in water. It dissolves in dilute acids with at most slight effervescence.
IDENTIFICATION
A. Bulk density (2.9.34): minimum 0.25 g/mL.
B. Dissolve about 15 mg in 2 mL of dilute nitric acid R and neutralise with dilute sodium hydroxide solution R. The solution gives the reaction of magnesium (2.3.1).
C. Loss on ignition (see Tests).
TESTS
Solution S
Dissolve 5.0 g in a mixture of 30 mL of distilled water R and 70 mL of acetic acid R, boil for 2 min, cool and dilute to 100 mL with dilute acetic acid R. Filter, if necessary, through a previously ignited and tared porcelain or silica filter crucible of suitable porosity to give a clear filtrate.

Ph Eur



1309-48-4

Light Magnesium Oxide

Light Magnesia

(Ph. Eur. monograph 0040)

MgO 40.30

Action and use

Antacid; osmotic laxative.

Preparation

Magnesium Hydroxide Mixture

Ph Eur

DEFINITION

Content

98.0 per cent to 100.5 per cent of MgO (ignited substance).

CHARACTERS

Appearance

Fine, white or almost white, amorphous powder.

Solubility

Practically insoluble in water. It dissolves in dilute acids with at most slight effervescence.

IDENTIFICATION

A. Bulk density (2.9.34): maximum 0.15 g/mL.

B. Dissolve about 15 mg in 2 mL of dilute nitric acid R and neutralise with dilute sodium hydroxide solution R. The solution gives the reaction of magnesium (2.3.1).

C. Loss on ignition (see Tests).

TESTS

Solution S

Dissolve 5.0 g in a mixture of 30 mL of distilled water R and 70 mL of acetic acid R, boil for 2 min, allow to cool and dilute to 100 mL with dilute acetic acid R. Filter, if necessary, through a previously ignited and tared porcelain or silica filter crucible of a suitable porosity to give a clear filtrate.

Appearance of solution

Solution S is not more intensely coloured than reference solution B₃ (2.2.2, Method II).

Soluble substances

Maximum 2.0 per cent.

To 2.00 g add 100 mL of water R and boil for 5 min. Filter whilst hot through a sintered-glass filter (40) (2.1.2), allow to cool and dilute to 100 mL with water R. Evaporate 50 mL of the filtrate to dryness and dry at 100-105 °C. The residue weighs a maximum of 20 mg.

Substances insoluble in acetic acid

Maximum 0.1 per cent.

Any residue obtained during the preparation of solution S, washed, dried, and ignited at 600 ± 50 °C, weighs a maximum of 5 mg.

Appearance of solution

Solution S is not more intensely coloured than reference solution B₃ (2.2.2, Method II).

Soluble substances

Maximum 2.0 per cent.

To 2.00 g add 100 mL of water R and boil for 5 min. Filter whilst hot through a sintered-glass filter (40) (2.1.2), allow to cool and dilute to 100 mL with water R. Evaporate 50 mL of the filtrate to dryness and dry at 100-105 °C. The residue weighs a maximum of 20 mg.

Substances insoluble in acetic acid

Maximum 0.1 per cent.

Any residue obtained during the preparation of solution S, washed, dried and ignited at 600 ± 50 °C, weighs a maximum of 5 mg.

Chlorides (2.4.4)

Maximum 0.1 per cent.

Dilute 1 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 1.0 per cent.

Dilute 0.3 mL of solution S to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 4 ppm, determined on 5 mL of solution S.

Calcium (2.4.3)

Maximum 1.5 per cent.

Dilute 1.3 mL of solution S to 150 mL with distilled water R.

15 mL of the solution complies with the test.

Iron (2.4.9)

Maximum 0.07 per cent.

Dissolve 0.15 g in 5 mL of dilute hydrochloric acid R and dilute to 10 mL with water R. Dilute 1 mL of the solution to

10 mL with water R.

Heavy metals (2.4.8)

Maximum 30 ppm.

To 20 mL of solution S add 15 mL of hydrochloric acid R1 and shake with 25 mL of methyl isobutyl ketone R for 2 min.

Allow to stand, then separate and evaporate the aqueous layer to dryness. Dissolve the residue in 1 mL of acetic acid R

and dilute to 30 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on ignition

Maximum 8.0 per cent, determined on 1.00 g at 900 ± 25 °C.

ASSAY

Dissolve 0.320 g in 20 mL of dilute hydrochloric acid R and dilute to 100.0 mL with water R. Using 20.0 mL of the

solution, carry out the complexometric titration of magnesium (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 4.030 mg of MgO.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are

Magnesium Peroxide

(Ph. Eur. monograph 1540)



DEFINITION

Mixture of magnesium peroxide and magnesium oxide.

Content

22.0 per cent to 28.0 per cent of MgO_2 (M_r -56.30).

CHARACTERS

Appearance

White or slightly yellow, amorphous, light powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute mineral acids.

IDENTIFICATION

A. Dissolve about 15 mg in 2 mL of dilute nitric acid R and neutralise with dilute sodium hydroxide solution R. The solution gives the reaction of magnesium (2.3.1).

B. Dissolve 50 mg in 2 mL of dilute sulfuric acid R.

Add 2 mL of a 5 g/L solution of potassium permanganate R and shake. The solution becomes colourless with evolution of gas.

TESTS

Solution S1

Dissolve cautiously 5.0 g in 40 mL of hydrochloric acid R1.

Cautiously evaporate the solution to 10 mL and dilute to 100 mL with a mixture of equal volumes of acetic acid R and distilled water R. Filter, if necessary, through a previously

ignited and tared porcelain or silica filter crucible of suitable porosity to give a clear filtrate. Keep the residue for the test

Solution S2

Dilute 5 mL of solution S1 to 25 mL with distilled water R.

Appearance of solution

Solution S1 is not more intensely coloured than reference solution B₄ (2.2.2, Method II).

Acidity or alkalinity

To 2.0 g add 100 mL of carbon dioxide-free water R and heat to boiling for 5 min. Filter whilst hot through a sintered-glass filter (40) (2.1.2), allow to cool and dilute to 100 mL with carbon dioxide-free water R. To 15 mL of the filtrate, add 0.1 mL of phenolphthalein solution R. The solution is red. Not more than 0.2 mL of 0.1 M hydrochloric acid is necessary to change the colour of the indicator. Keep the filtrate for the test for soluble substances.

Acid insoluble substances

Maximum 0.1 per cent.

Any residue obtained during the preparation of solution S1, washed, dried and ignited at $600 \pm 50^\circ\text{C}$, weighs a maximum of 5 mg.

Soluble substances

Maximum 1.5 per cent.

Take 50 mL of the filtrate obtained in the test for acidity or alkalinity, evaporate to dryness and dry at 100 – 105°C .

The residue weighs a maximum of 15 mg.

Chlorides (2.4.4)

Maximum 0.1 per cent.

Dissolve 50 mg in 5 mL of dilute nitric acid R and dilute to 15 mL with water R.

Dilute 0.7 mL of solution S to 15 mL with water R.

Chlorides (2.4.4)

Maximum 0.15 per cent.

Sulfates (2.4.13)

Maximum 1.0 per cent.

Dilute 0.3 mL of solution S to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 4 ppm, determined on 5 mL of solution S.

Calcium (2.4.3)

Maximum 1.5 per cent.

Dilute 1.3 mL of solution S to 150 mL with distilled water R.

15 mL of this solution complies with the test.

Iron (2.4.9)

Maximum 0.1 per cent.

Dissolve 50 mg in 5 mL of dilute hydrochloric acid R and dilute to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 30 ppm.

To 20 mL of solution S add 15 mL of hydrochloric acid R1 and shake with 25 mL of methyl isobutyl ketone R for 2 min.

Allow to stand, then separate and evaporate the aqueous layer to dryness. Dissolve the residue in 1.5 mL of acetic acid R and dilute to 30 mL with water R. 12 mL of the

solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on ignition

Maximum 8.0 per cent, determined on 1.00 g at $900 \pm 25^\circ\text{C}$.

ASSAY

Dissolve 0.320 g in 20 mL of dilute hydrochloric acid R and dilute to 100.0 mL with water R. Using 20.0 mL of this

solution, carry out the complexometric titration of magnesium (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 4.030 mg of MgO .

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for light magnesium oxide used as filler in oral solid dosage forms.

Particle-size distribution (2.9.31 or 2.9.38).

Bulk and tapped density (2.9.34)

Ph Eur

Sulfates (2.4.13)

Maximum 0.5 per cent.

Dilute 3 mL of solution S2 to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 4 ppm, determined on 5 mL of solution S1.

Calcium (2.4.3)

Maximum 1.0 per cent.

Dilute 1 mL of solution S2 to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 500 ppm.

Dilute 2 mL of solution S2 to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 30 ppm.

To 20 mL of solution S1 add 15 mL of hydrochloric acid R1

and shake with 25 mL of methyl isobutyl ketone R for 2 min.

Allow to stand, then separate and evaporate the aqueous

layer to dryness. Dissolve the residue in 1.5 mL of acetic

acid R and dilute to 30 mL with water R. 12 mL of the

solution complies with test A. Prepare the reference solution

using lead standard solution (1 ppm Pb) R.

ASSAY

Dissolve 80.0 mg, shaking cautiously, in a mixture,

previously cooled to 20 °C, of 10 mL of sulfuric acid R and

90 mL of water R. Titrate with 0.02 M potassium

permanganate until a pink colour is obtained.

1 mL of 0.02 M potassium permanganate is equivalent to

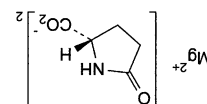
2.815 mg of MgO₂.

STORAGE

Protected from light.

Magnesium Pidolate

(Ph. Eur. monograph 1619)



C₁₀H₁₂N₂O₆Mg 280.5 62003-27-4

DEFINITION

Magnesium bis[(2S)-5-oxopyrrolidine-2-carboxylate].

Content

8.49 per cent to 8.84 per cent of Mg ($A_r = 24.31$)

(anhydrous substance).

CHARACTERS**Appearance**

Amorphous, white or almost white powder, hygroscopic.

Solubility

Very soluble in water, soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 60 mg of the substance to be examined in 2 mL of water R and dilute to 10 mL with methanol R.

Reference solution Dissolve 55 mg of pidolic acid CRS in 2 mL of water R and dilute to 10 mL with methanol R.

Plate TLC silica gel plate R.

Mobile phase methanol R, glacial acetic acid R, methylene chloride R (15:20:65 V/V/V).

Application 1 µL.

Development Over 2/3 of the plate.

Drying At 100-105 °C for 15 min.

Detection Spray with strong sodium hypochlorite solution R.

Allow to stand for 10 min and spray abundantly with glacial

acetic acid R. Allow to stand again for 10 min and dry at

100-105 °C for 2 min. Spray with potassium iodide and starch

solution R until spots appear.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

reference solution. The chromatogram obtained with the test

solution may show 2 faint secondary spots.

B. To 0.15 mL of solution S (see Tests) add 1.8 mL of

water R. The solution gives the reaction of magnesium

TESTS**Solution S**

Dissolve 5.00 g in carbon dioxide-free water R prepared from

distilled water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution B₈ (2.2.2, Method I).

pH (2.2.3)

5.5 to 7.0 for solution S.

Specific optical rotation (2.2.7)

−23.3 to −26.5 (anhydrous substance), determined on

solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.500 g of the substance to be

examined in the mobile phase and dilute to 100.0 mL with

the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase.

Reference solution (b) Dissolve 50.0 mg of pidolate

impurity B CRS in the mobile phase and dilute to 100.0 mL

with the mobile phase. Dilute 5.0 mL of the solution to

50.0 mL with the mobile phase.

Reference solution (c) Dilute 10.0 mL of reference solution (b)

to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of nitrate standard solution

(100 ppm NO₃) R to 100.0 mL with the mobile phase.

Reference solution (e) Dilute 6.0 mL of reference solution (a)

to 10.0 mL with reference solution (b).

Column:

— size: l = 0.25 m, Ø = 4.6 mm,

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 µm).

Mobile phase Dissolve 1.56 g of sodium dihydrogen phosphate R

in 1000 mL of water R and adjust to pH 2.5 with a

10 per cent V/V solution of phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of the test solution and reference

solutions (b), (c), (d) and (e).

Run time 4 times the retention time of pidolic acid.

Retention times Pictolic acid = about 4.5 min; impurity B = about 7.5 min.
System suitability: reference solution (e):
Resolution: minimum 10 between the peaks due to pictolic acid and impurity B.
Limits:

— **impurity B:** not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
 — **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
 — **total of other impurities:** not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
 — **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard any peak corresponding to the nitrate ion (NO_3^-).
Impurity A
Thin-layer chromatography (2.2.27).
Test solution Dissolve 0.250 g of the substance to be examined in 4 mL of water R and dilute to 50.0 mL with methanol R.
Reference solution (a) Dissolve 60.0 mg of glutamic acid R in 50 mL of water R and dilute to 100.0 mL with methanol R.
Reference solution (b) Dissolve 10 mg of aspartic acid R and 10 mg of glutamic acid R in water R and dilute to 25 mL with the same solvent. Dilute 1 mL of the solution to 10 mL with water R.
Plate TLC silica gel plate R.
Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).
Application 5 µL.
Development Over 2/3 of the plate.
Drying In air.
Detection Spray with ninhydrin solution R and heat at 100–105 °C for 15 min.
System suitability: reference solution (b):
 — the chromatogram shows 2 clearly separated spots.
Limits:
 — **impurity A:** any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.6 per cent).

Chlorides (2.4.4)
 Maximum 500 ppm.
 Dilute 1.0 mL of solution S to 15.0 mL with water R.
Nitrates
 Examine the chromatogram obtained with the test solution in the test for related substances.
Limits:
 — **nitrates:** not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (200 ppm).
Sulfates (2.4.13)
 Maximum 0.1 per cent.
 Dilute 1.5 mL of solution S to 15.0 mL with distilled water R.
Arsenic (2.4.2, Method A)
 Maximum 2 ppm, determined on 5.0 mL of solution S.

Magnesium Stearate (1)



(Ph. Eur. monograph 0229)

Action and use
 Excipient.

DEFINITION

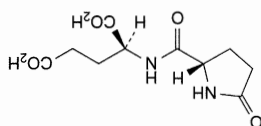
Compound of magnesium with a mixture of solid organic acids and consisting mainly of variable proportions of magnesium stearate and magnesium palmitate obtained from sources of vegetable or animal origin.

Content

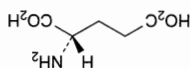
— **magnesium** (Mg ; A_r 24.305): 4.0 per cent to 5.0 per cent (dried substance);
 — **stearic acid in the fatty acid fraction:** minimum 40.0 per cent;
 — **sum of stearic acid and palmitic acid in the fatty acid fraction:** minimum 90.0 per cent.

CHARACTERS

Appearance
 White or almost white, very fine, light powder, greasy to the touch.



A. (2S)-2-aminopentanedioic acid (glutamic acid),



Specified impurities A, B

IMPURITIES

In an airtight container.

STORAGE

ASSAY
 Dissolve 0.300 g in 50 mL of water R. Carry out the complexometric titration of magnesium (2.5.11).
 1 mL of 0.1 M sodium edetate is equivalent to 2.431 mg of Mg.

Water (2.5.12)
 Maximum 8.0 per cent, determined on 0.200 g.
Heavy metals (2.4.8)
 Dilute 0.5 mL of solution S to 10 mL with water R.
 Maximum 200 ppm.
 12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Solubility

Practically insoluble in water and in anhydrous ethanol. ◆

IDENTIFICATION

First identification C, D

◆ Second identification: A, B, D.

A. Freezing point (2.2.18): minimum 53 °C, determined on

(see Tests).

B. Acid value (2.5.1): 195 to 210.

Dissolve 0.200 g of the residue obtained in the preparation of solution S in 25 mL of the prescribed mixture of solvents. ◆

C. Examine the chromatograms obtained in the assay of

stearic acid and palmitic acid.

Results The 2 principal peaks in the chromatogram obtained

with the test solution are similar in retention time to the

2 principal peaks in the chromatogram obtained with the

reference solution.

D. To 1 mL of solution S add 1 mL of dilute ammonia R₁; a white precipitate is formed that dissolves on addition of

1 mL of ammonium chloride solution R. Add 1 mL of a

120 g/L solution of disodium hydrogen phosphate R; a white

crystalline precipitate is formed.

TESTS**Solution S**

To 5.0 g add 50 mL of peroxide-free ether R, 20 mL of dilute

nitric acid R and 20 mL of water R and heat under a reflux

condenser until dissolution is complete. Allow to cool. In a

separating funnel, separate the aqueous layer and shake the

ether layer with 2 quantities, each of 4 mL, of water R.

Combine the aqueous layers, wash with 15 mL of peroxide-

free ether R and dilute to 50.0 mL with water R (solution S).

Evaporate the organic layer to dryness and dry the residue at

100-105 °C. Keep the residue for identification tests A

and B.

Acidity or alkalinity

To 1.0 g add 20 mL of carbon dioxide-free water R and boil

for 1 min with continuous shaking. Cool and filter.

To 10 mL of the filtrate add 0.05 mL of bromothymol blue

solution R₄. Not more than 0.05 mL of 0.1 M hydrochloric

acid or 0.1 M sodium hydroxide is required to change the

colour of the indicator.

Chlorides

Maximum 0.1 per cent.

Dilute 10.0 mL of solution S to 40 mL with water R.

Neutralise if necessary with nitric acid R using litmus R as

indicator. Add 1 mL of nitric acid R and 1 mL of 0.1 M silver

nitrate and dilute to 50 mL with water R. Mix and allow to

stand for 5 min protected from light. The turbidity, if any, is

not greater than that produced in a solution containing

1.4 mL of 0.02 M hydrochloric acid.

Sulfates

Maximum 1.0 per cent.

Dilute 6.0 mL of solution S to 40 mL with water R.

Neutralise if necessary with hydrochloric acid R using litmus R

as indicator. Add 1 mL of 3 M hydrochloric acid R and 3 mL

of a 120 g/L solution of barium chloride R and dilute to

50 mL with water R. Mix and allow to stand for 10 min.

The turbidity, if any, is not greater than that produced in a

solution containing 3.0 mL of 0.02 M sulfuric acid.

Cadmium

Maximum 3 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

For the preparation of all aqueous solutions and for the rinsing of glassware before use, employ water that has been passed through a

strong-acid, strong-base, mixed-bed ion-exchange resin before use.

Select all reagents to have as low a content of cadmium, lead and

nickel as practicable and store all reagent solutions in containers of

borosilicate glass. Clean glassware before use by soaking in warm

8 M nitric acid for 30 min and by rinsing with deionised water.

Blank solution Dilute 25 mL of cadmium- and lead-free nitric

acid R to 100.0 mL with water R.

Modifier solution Dissolve 20 g of ammonium dihydrogen

phosphate R and 1 g of magnesium nitrate R in water R and

dilute to 100 mL with the same solvent. Alternatively, use an

appropriate matrix modifier as recommended by the graphite

furnace atomic absorption (GFAA) spectrometer

manufacturer.

Test solution Place 0.100 g of the substance to be examined in

a polytetrafluoroethylene digestion bomb and add 2.5 mL of

cadmium- and lead-free nitric acid R. Close and seal the bomb

according to the manufacturer's operating instructions (when

using a digestion bomb, be thoroughly familiar with the safety and

operating instructions. Carefully follow the bomb manufacturer's

instructions regarding care and maintenance of these digestion

bombs. Do not use metal jacketed bombs or liners which have been

used with hydrochloric acid due to contamination from corrosion of

the metal jacket by hydrochloric acid). Heat the bomb in an

oven at 170 °C for 3 h. Cool the bomb slowly in air to room

temperature according to the bomb manufacturer's

instructions. Place the bomb in a fume cupboard and open

carefully as corrosive gases may be expelled. Dissolve the

residue in water R and dilute to 10.0 mL with the same

solvent.

Reference solution Prepare a solution of 0.0030 µg/mL of Cd

by suitable dilutions of a 0.00825 µg/mL solution of cadmium

nitrate tetrahydrate R in the blank solution.

Dilute 1.0 mL of the test solution to 10.0 mL with the blank

solution. Prepare mixtures of this solution, the reference

solution and the blank solution in the following proportions:

To each mixture add 50 µL of modifier solution and mix.

These solutions contain respectively 0 µg, 0.00075 µg and

0.0015 µg of cadmium per millilitre from the reference

solution (keep the remaining test solution for use in the test

for lead and nickel).

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Furnace.

Platform Pyrolytically coated with integrated tube.

Operating conditions Use the temperature programme

recommended for cadmium by the GFAA spectrometer

manufacturer. An example of temperature parameters for

GFAA analysis of cadmium is shown below.

Stage	Final temperature (°C)	Ramp time (s)	Hold time (s)
Drying	110	10	20
Ashing	600	10	30
Atomisation	1800	0	5

Lead

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

For the preparation of all aqueous solutions and for the rinsing of glassware before use, employ water that has been passed through a

strong-acid, strong-base, mixed-bed ion-exchange resin before use. Select all reagents to have as low a content of cadmium, lead and nickel as practicable and store all reagent solutions in containers of borosilicate glass. Clean glassware before use by soaking in warm 8 M nitric acid for 30 min and by rinsing with deionised water. Blank solution Use the solution described in the test for cadmium. Modifier solution Use the solution described in the test for cadmium. Test solution Use the solution described in the test for cadmium. Reference solution Prepare a solution of 0.100 µg/mL of Pb by suitable dilutions of lead standard solution (100 ppm Pb) R with the blank solution. Prepare mixtures of the test solution, the reference solution and the blank solution in the following proportions: (1.0:0.1:1.0 V/V/V), (1.0:0.5:0.5 V/V/V), (1.0:1.0:0.0 V/V/V). To each mixture add 50 µL of modifier solution and mix. These solutions contain respectively 0 µg, 0.025 µg and 0.05 µg of lead per millilitre from the reference solution. Source Lead hollow-cathode lamp. Wavelength 283.3 nm. Atomisation device Furnace. Platform Pyrolytically coated with integrated tube. Operating conditions Use the temperature programme recommended for lead by the GFAA spectrometer. manufacturer. An example of temperature parameters for GFAA analysis of lead is shown below.

Stage	Final temperature (°C)	Ramp time (s)	Hold time (s)
Drying	110	10	20
Ashing	450	10	30
Atomisation	2000	0	5

Nickel

Maximum 5 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

For the preparation of all aqueous solutions and for the rinsing of glassware before use, employ water that has been passed through a strong-acid, strong-base, mixed-bed ion-exchange resin before use. Select all reagents to have as low a content of cadmium, lead and nickel as practicable and store all reagent solutions in containers of borosilicate glass. Clean glassware before use by soaking in warm 8 M nitric acid for 30 min and by rinsing with deionised water. Blank solution Use the solution described in the test for cadmium. Modifier solution Dissolve 20 g of ammonium dihydrogen phosphate R in water R and dilute to 100 mL with the same solvent. Alternatively, use an appropriate matrix modifier as recommended by the GFAA spectrometer manufacturer. Test solution Use the solution described in the test for cadmium. Reference solution Prepare a solution of 0.050 µg/mL of Ni by suitable dilutions of a 0.2477 µg/mL solution of nickel nitrate hexahydrate R in the blank solution. Prepare mixtures of the test solution, the reference solution and the blank solution in the following proportions: (1.0:0.1:1.0 V/V/V), (1.0:0.5:0.5 V/V/V), (1.0:1.0:0.0 V/V/V). To each mixture add 50 µL of matrix modifier solution and mix. These reference solutions contain respectively 0 µg,

Stage	Final temperature (°C)	Ramp time (s)	Hold time (s)
Drying	110	10	20
Ashing	1000	20	30
Atomisation	2300	0	5

0.0125 µg and 0.025 µg of nickel per millilitre from the reference solution. Source Nickel hollow-cathode lamp. Wavelength 232.0 nm. Atomisation device Furnace. Platform Pyrolytically coated with integrated tube. Operating conditions Use the temperature programme recommended for nickel by the GFAA spectrometer manufacturer. An example of temperature parameters for GFAA analysis of nickel is shown below.

Loss on drying (2.2.32)

Maximum 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12). TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13). ◆

ASSAY

Magnesium

To 0.500 g in a 250 mL conical flask add 50 mL of a mixture of equal volumes of anhydrous ethanol R and butanol R, 5 mL of concentrated ammonia R, 30.0 mL of 0.1 M sodium edetate and 15 mg of mordant black 11 titrate with 0.1 M zinc sulfate until the colour changes from blue to violet. Carry out a blank titration. 1 mL of 0.1 M sodium edetate is equivalent to 2.431 mg of Mg.

Stearic acid and palmitic acid

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution In a conical flask fitted with a reflux condenser, dissolve 0.10 g of the substance to be examined in 5 mL of boron trifluoride-methanol solution R. Boil under a reflux condenser for 10 min. Add 4 mL of heptane R through the condenser and boil again under a reflux condenser for 10 min. Allow to cool. Add 20 mL of saturated sodium chloride solution R. Shake and allow the layers to separate. Dry the organic layer over 0.1 g of anhydrous sodium sulfate R (previously washed with heptane R). Dilute 1.0 mL of the solution to 10.0 mL with heptane R. Reference solution Prepare the reference solution in the same manner as the test solution using 50.0 mg of palmitic acid CRS and 50.0 mg of stearic acid CRS instead of the substance to be examined.

Column:

— material: fused silica;

— size: l = 30 m, Ø = 0.32 mm;

— stationary phase: macrogol 20 000 R (film thickness 0.5 µm).

Carrier gas helium for chromatography R.

Flow rate 2.4 mL/min.



Magnesium Sulfate Heptahydrate

Epsum Salts; Magnesium Sulphate Heptahydrate

(Ph. Eur. monograph 0044)

MgSO₄·7H₂O

246.5

10034-99-8

Action and use

Osmotic laxative; used in treatment of electrolyte deficiency.

Preparations

Magnesium Sulfate Injection

Magnesium Sulfate Mixture

DEFINITION

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or brilliant,

colourless crystals.

Solubility

Freely soluble in water, very soluble in boiling water,

practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. It gives the reactions of sulfates (2.3.1).

B. It gives the reaction of magnesium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in water R and dilute to 50 mL with the same

solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.05 mL of phenol red solution R.

Not more than 0.2 mL of 0.01 M hydrochloric acid or

0.01 M sodium hydroxide is required to change the colour of

the indicator.

Chlorides (2.4.4)

Maximum 300 ppm.

Dilute 1.7 mL of solution S to 15 mL with water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

48.0 per cent to 52.0 per cent, determined on 0.500 g by

drying in an oven at 110-120 °C for 1 h and then at 400 °C

to constant mass.

ASSAY

Dissolve 0.450 g in 100 mL of water R and carry out the

complexometric titration of magnesium (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 12.04 mg

of MgSO₄.

Ph Eur

Temperature:

Time (min)	Temperature (°C)
0 - 2	70
2 - 36	70 → 240
36 - 41	240
Injection port	220
Detector	260

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to methyl stearate: methyl

palmitate = about 0.9.

System suitability: reference solution:

— resolution: minimum 5.0 between the peaks due to methyl

palmitate and methyl stearate;

— relative standard deviation: maximum 3.0 per cent for the

areas of the peaks due to methyl palmitate and methyl

stearate, determined on 6 injections; maximum

1.0 per cent for the ratio of the areas of the peaks due to

methyl palmitate to the areas of the peaks due to methyl

stearate, determined on 6 injections.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). Some of the characteristics described in the Functionality-

related characteristics section may also be present in the mandatory

part of the monograph since they also represent mandatory quality

criteria. In such cases, a cross-reference to the tests described in the

mandatory part is included in the Functionality-related

characteristics section. Control of the characteristics can contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for magnesium

stearate used as lubricant in tablets and capsules.

Particle-size distribution (2.9.31)

Specific surface area (2.9.26, Method I)

Determine the specific surface area in the P/P₀ range of

0.05 to 0.15.

Sample outgassing 2 h at 40 °C.

Thermogravimetry (2.2.34)

¹ This monograph has undergone pharmacopoeial harmonisation.

See chapter 5.8 Pharmacopoeial Harmonisation.

Ph Eur

Dried Magnesium Sulfate

Dried Epsom Salts; Dried Magnesium Sulphate

Preparation

Magnesium Sulfate Paste

DEFINITION

Dried Magnesium Sulfate contains not less than 62.0% and not more than 70.0% of MgSO_4 . It may be prepared by drying magnesium sulfate at 100° until it has lost approximately 25% of its weight.

CHARACTERISTICS

A white powder.

Freely soluble in water; dissolves more rapidly in hot water.

IDENTIFICATION

Yields the reactions characteristic of magnesium salts and of sulfates, Appendix VI.

TESTS

To 10 mL of a 7.5% w/v solution in carbon dioxide-free water add 0.05 mL of phenol red solution. Not more than 0.2 mL of either 0.01M hydrochloric acid VS or 0.01M sodium hydroxide VS is required to change the colour of the solution.

Arsenic

0.33 g dissolved in 25 mL of water complies with the limit test for arsenic, Appendix VII (3 ppm).

Heavy metals

Dissolve 1.3 g in 20 mL of water and add 1 g of ammonium chloride. 12 mL of the resulting solution complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (1 ppm Pb) to prepare the standard (15 ppm).

Iron

0.33 g dissolved in 10 mL of water complies with the limit test for iron, Appendix VII (30 ppm).

Chloride

0.13 g dissolved in 15 mL of water complies with the limit test for chlorides, Appendix VII (400 ppm).

Insoluble matter

7.5 g dissolves in 20 mL of water, producing a solution which may be slightly turbid at first but which becomes clear in a few minutes.

ASSAY

Dissolve 0.3 g in 50 mL of water and carry out the complexometric titration of magnesium, Appendix VIII D. Each mL of 0.01M disodium edetate VS is equivalent to 12.04 mg of MgSO_4 .

Magnesium Trisilicate

(Ph. Eur. monograph 0403)

Action and use

Antacid.

Preparations

Magnesium Trisilicate Mixture

Chewable Compound Magnesium Trisilicate Tablets

Compound Magnesium Trisilicate Oral Powder

DEFINITION

It has a variable composition corresponding approximately to $\text{Mg}_2\text{Si}_3\text{O}_8 \cdot x\text{H}_2\text{O}$.



Content

— magnesium oxide (MgO ; 40.30): minimum 29.0 per cent (ignited substance),
— silicon dioxide (SiO_2 ; 60.1): minimum 65.0 per cent (ignited substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. 0.25 g gives the reaction of silicates (2.3.1).

B. 1 mL of solution S (see Tests) neutralised with dilute sodium hydroxide solution R gives the reaction of magnesium (2.3.1).

TESTS

Solution S
To 2.0 g add a mixture of 4 mL of nitric acid R and 4 mL of distilled water R. Heat to boiling with frequent shaking. Add 12 mL of distilled water R and allow to cool. Filter or centrifuge to obtain a clear solution and dilute to 20 mL with distilled water R.

Alkalinity

To 10.0 g in a 200 mL conical flask, add 100.0 g of water R and heat on a water-bath for 30 min. Allow to cool and make up to the initial mass with water R. Allow to stand and filter or centrifuge until a clear liquid is obtained. To 10 mL of this liquid add 0.1 mL of phenolphthalein solution R. Not more than 1.0 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator.

Water-soluble salts

Maximum 1.5 per cent.
In a platinum dish, evaporate to dryness on a water-bath. The residue, ignited to constant mass at $900 \pm 50^\circ\text{C}$, weighs a maximum of 30 mg.

Chlorides (2.4.4)

Maximum 500 ppm.
Dilute 0.5 mL of solution S to 15 mL with water R. Prepare the standard using a mixture of 5 mL of chloride standard solution (5 ppm Cl) R and 10 mL of water R.

Sulfates (2.4.13)

Maximum 0.5 per cent.
Dilute 0.3 mL of solution S to 15 mL with distilled water R. Arsenic (2.4.2, method A)
Maximum 4 ppm, determined on 2.5 mL of solution S.

Heavy metals (2.4.8)

Maximum 40 ppm.
Neutralise 10 mL of solution S with dilute ammonia R1, using methyl yellow solution R as an external indicator. Dilute to 20 mL with water R and filter if necessary. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on ignition

17 per cent to 34 per cent, determined on 0.5 g by ignition to constant mass at $900 \pm 50^\circ\text{C}$ in a platinum crucible.

Acid-absorbing capacity

Suspend 0.25 g in 0.1 M hydrochloric acid, dilute to 100.0 mL with the same acid and allow to stand for 2 h in a water-bath at $37 \pm 0.5^\circ\text{C}$, with frequent shaking. Allow to cool. To 20.0 mL of the supernatant solution add 0.1 mL of

brופןهنول blue solution R and titrate with 0.1 M sodium hydroxide until a blue colour is obtained. The acid-absorbing capacity is not less than 100.0 mL of 0.1 M hydrochloric acid per gram.

ASSAY

Magnesium oxide

To 1.000 g in a 200 mL conical flask, add 35 mL of

hydrochloric acid R and 60 mL of water R and heat in a water-

and the residue with water R and dilute the combined filtrate

and washings to 250.0 mL with water R. Neutralise 50.0 mL

of the solution with strong sodium hydroxide solution R (about

8 mL). Carry out the complexometric titration of magnesium

(2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 4.030 mg

of MgO.

Silicon dioxide

To 0.700 g add 10 mL of dilute sulfuric acid R and 10 mL of

water R. Heat for 90 min on a water-bath with frequent

shaking, replacing the evaporated water. Allow to cool and

decant onto an ashless filter paper (diameter 7 cm). Wash

the precipitate by decantation with 3 quantities, each of

5 mL, of hot water R, transfer it to the filter and wash it with

hot water R until 1 mL of the filtrate remains clear after the

addition of 0.05 mL of dilute hydrochloric acid R and 2 mL of

barium chloride solution R1. Incinerate the filter and its

contents in a platinum crucible, then ignite the residue

(SiO₂) at 900 ± 50 °C to constant mass.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate

compliance. Control of these characteristics can however contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for magnesium

trisilicate used as a lubricant in tablets and capsules.

Particle-size distribution (2.9.31)

Specific surface area (2.9.26, Method I)

Ph Eur



Refined Maize Oil

(Ph. Eur. monograph 1342)

Ph Eur

DEFINITION

Fatty oil obtained from the seeds of *Zea mays* L. by expression or by extraction. It is then refined.

CHARACTERS

Appearance

Clear, light yellow or yellow oil.

Solubility

Practically insoluble in water and in ethanol (96 per cent), miscible with light petroleum (bp: 40–60 °C) and with methylene chloride.

Relative density

About 0.920.

Refractive index About 1.474.

IDENTIFICATION

A. Identification of fatty oils by thin-layer chromatography

(2.3.2).

Results The chromatogram obtained with the test solution is

similar to the chromatogram obtained with the reference

solution.

B. Composition of fatty acids (see Tests).

TESTS

Acid value (2.5.1)

Manufacture of parenteral preparations, determined on

10.0 g.

Peroxide value (2.5.5, Method A)

Maximum 10.0, or maximum 5.0 if intended for use in the

manufacture of parenteral preparations.

Unsaturation matter (2.5.7)

Maximum 2.8 per cent, determined on 5.0 g.

Alkaline impurities (2.4.19)

It complies with the test.

Composition of fatty acids (2.4.22, Method A)

Use the mixture of calibrating substances in Table 2.4.22-3.

Composition of the fatty-acid fraction of the oil:

— fatty acids of chain length less than C₁₆: maximum

0.6 per cent;

— palmitic acid: 8.6 per cent to 16.5 per cent;

— stearic acid: maximum 3.3 per cent;

— oleic acid: 20.0 per cent to 42.2 per cent;

— linoleic acid: 39.4 per cent to 65.6 per cent;

— linolenic acid: 0.5 per cent to 1.5 per cent;

— arachidic acid: maximum 0.8 per cent;

— eicosenoic acid: maximum 0.5 per cent;

— behenic acid: maximum 0.5 per cent;

— other fatty acids: maximum 0.5 per cent.

Sterols (2.4.23)

Maximum 0.3 per cent of brassicasterol in the sterol fraction

of the oil.

Water (2.5.32)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

Protected from light, at a temperature not exceeding 25 °C.

LABELLING

The label states:

— where applicable, that the substance is suitable for use in

the manufacture of parenteral preparations;

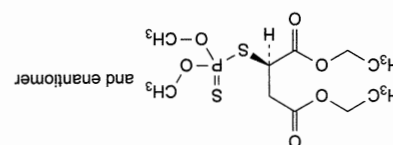
— whether the oil is obtained by mechanical expression or

by extraction.

Ph Eur

Malathion

(Ph. Eur. monograph 1343)

C₁₀H₁₉O₆PS₂ 330.4

121-75-5

Action and use

Organophosphorus insecticide.

Preparations

Malathion Lotion

Malathion Shampoo

DEFINITION

Diethyl (2*RS*)-2-(dimethoxyphosphinodithioyl)butanedioate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, colourless or slightly yellowish liquid.

Solubility

Slightly soluble in water, miscible with acetone, with cyclohexane, with ethanol (96 per cent) and with vegetable oils.

It solidifies at about 3 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison malathion CRS.

TESTS

Relative density (2.2.5)

1.220 to 1.240.

Optical rotation (2.2.7)

-0.1° to +0.1°.

Dissolve 2.50 g in ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture water R, acetonitrile R (1:3 V/V).

Test solution (a) Dissolve 0.10 g of the substance to be examined in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 0.100 g of malathion CRS in 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 0.5 mL of test solution (a) to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of malathion impurity A CRS and 5.0 mg of malathion impurity B CRS in the solvent mixture, then dilute 50.0 mL with the solvent mixture.

Reference solution (d) Dilute 2.0 mL of reference solution (c) to 10.0 mL with the solvent mixture.

Column: — size: $l = 0.15$ m, $\varnothing = 4.6$ mm; — stationary phase: octadecylsilyl silica gel for chromatography R (10 μ m); — temperature: 35 °C.

Mobile phase acetonitrile R, water R (45:55 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L of test solution (a) and reference solutions (b), (c) and (d).

Retention time Impurity B = about 3.5 min;

impurity A = about 5 min; malathion = about 16 min.

System suitability: reference solution (c):

— resolution: minimum 2.0 between the peaks due to impurities B and A.

Limits:

— impurity A: not more than 3 times the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.1 per cent);

— sum of impurities other than A and B: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (2.5.12)

Maximum 0.1 per cent, determined on 2.000 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

System suitability: reference solution (a):

— repeatability: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of C₁₀H₁₉O₆PS₂ from the declared content of malathion CRS.

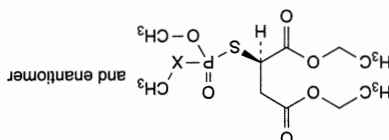
STORAGE

In an airtight container, protected from light.

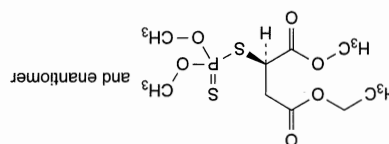
IMPURITIES

Specified impurities A, B.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



A. X = S: diethyl (2*RS*)-2-[(methoxy)(methylsulfanyl)-5-phosphinodithioyl]butanedioate (isomalathion), B. X = O: diethyl (2*RS*)-2-(dimethoxy-5-phosphinodithioyl)butanedioate (malathion).



C. ethyl and methyl (2RS)-2-(dimethoxyphosphinodithioyl) butanediolate (methyl analogue).

Ph Eur



(Ph. Eur. monograph 0365)

Maleic Acid



110-16-7

DEFINITION

Maleic acid contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of

(Z)-butenedioic acid, calculated with reference to the

CHARACTERS

A white or almost white, crystalline powder, freely soluble in water and in alcohol.

IDENTIFICATION

A. Dilute 5 mL of solution S (see Tests) to 10 mL with

water R. The pH of the dilution is less than 2.

B. Examine the chromatograms obtained in the test for fumaric acid. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size

to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.1 g in 10 mL of water R (solution a).

To 0.3 mL of solution (a) add a solution of 10 mg of

resorcinol R in 3 mL of sulfuric acid R. Heat on a water-bath

for 15 min; no colour develops. To 3 mL of solution (a) add

1 mL of bromine water R. Heat on a water-bath to remove

the bromine (15 min), heat to boiling and cool. To 0.2 mL

of this solution add a solution of 10 mg of resorcinol R in

3 mL of sulfuric acid R. Heat on a water-bath for 15 min.

A violet-pink colour develops.

TESTS

Solution S

Dissolve 5.0 g in water R and dilute to 50 mL with the same

solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution Y₇ (2.2.2, Method II).

Fumaric acid

Examine by thin-layer chromatography (2.2.27), using silica

gel GF₂₅₄ R as the coating substance.

Test solution (a) Dissolve 0.5 g of the substance to be

examined in acetone R and dilute to 5 mL with the same

solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL

with acetone R.

Iron

To 10 mL of solution S add 2 mL of dilute hydrochloric

acid R and 0.05 mL of bromine water R. After 5 min, remove

the excess of bromine by passing a current of air and add

3 mL of potassium thiocyanate solution R. Shake. Prepare a

standard at the same time and in the same manner, using a

mixture of 5 mL of iron standard solution (1 ppm Fe) R, 1 mL

of dilute hydrochloric acid R, 6 mL of water R and 0.05 mL of

bromine water R. Allow both solutions to stand for 5 min.

Any red colour in the test solution is not more intense than

that in the standard (5 ppm).

Heavy metals (2.4.8)

1.0 g complies with test D for heavy metals (10 ppm).

Prepare the reference solution using 1 mL of lead standard

solution (10 ppm Pb) R.

Water (2.5.12)

Not more than 2.0 per cent, determined on 1.00 g by the

semi-micro determination of water.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.500 g in 50 mL of water R. Titrate with

1 M sodium hydroxide using 0.5 mL of phenolphthalein

solution R as indicator.

1 mL of 1 M sodium hydroxide is equivalent to 58.04 mg of

C₄H₄O₄.

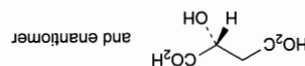
STORAGE

Store in a glass container, protected from light.

Ph Eur

Malic Acid

(Ph. Eur. monograph 2080)



$C_4H_6O_5$ 134.1 6915-15-7

Action and use

Excipient.

DEFINITION

(2R,3S)-2-Hydroxybutanedioic acid.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in alcohol, sparingly soluble in acetone.

IDENTIFICATION

A. Melting point (2.2.14): 128 °C to 132 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of malic acid.

TESTS

Solution S

Dissolve 5.00 g in water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

Water-insoluble substances

Maximum 0.1 per cent.

Dissolve 25.0 g in 100 mL of water R, filter the solution through a tared sintered-glass filter (16) (2.1.2), wash the filter with hot water R and dry at 100–105 °C to constant weight. The residue weighs a maximum of 25 mg.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 100.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of fumaric acid R and 4.0 mg of maleic acid R in 25 mL of the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 2.5 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 20.0 mg of the substance to be examined in the mobile phase, add 1.0 mL of reference solution (a) and dilute to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.30$ m, $\phi = 7.8$ mm,

— stationary phase: ion-exclusion resin for chromatography R (9 μ m),

— temperature: 37 °C.

Mobile phase 0.005 M sulfuric acid.

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L.

Run time Twice the retention time of the principal peak in the chromatogram obtained with the test solution.

Relative retention With reference to malic acid (retention time = about 10 min): impurity B = about 0.8;

impurity A = about 1.5.

System suitability: reference solution (c):

— resolution: minimum 2.5 between the peaks due to impurity B and malic acid.

Limits:

— impurity A: not more than twice the area of the corresponding peak in the chromatogram obtained with reference solution (b) (1.0 per cent),

— impurity B: not more than 0.25 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.05 per cent),

— any other impurity: for each impurity, not more than 0.5 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) (0.1 per cent),

— total of other impurities: not more than 2.5 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (b) (0.02 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 2.0 per cent, determined on 1.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

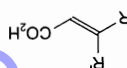
ASSAY

Dissolve 0.500 g in 50 mL of carbon dioxide-free water R. Titrate with 1 M sodium hydroxide determining the end-point potentiometrically (2.2.20).

1 mL of 1 M sodium hydroxide is equivalent to 67.05 mg of $C_4H_6O_5$.

IMPURITIES

Specified impurities: A, B.



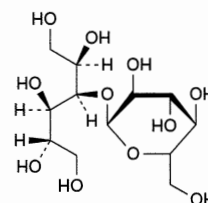
A. $R = CO_2H$, $R' = H$: (E)-butenedioic acid (fumaric acid),

B. $R = H$, $R' = CO_2H$: (Z)-butenedioic acid (maleic acid).

Ph Eur

Maltilitol

(Ph. Eur. monograph 1235)

C₁₂H₂₄O₁₁

344.3

585-88-6

Action and use
Sweetening agent.

Ph. Eur.

DEFINITION

4-O-α-D-Glucopyranosyl-D-glucitol (D-maltilitol).

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, practically insoluble in anhydrous ethanol.

IDENTIFICATION

First identification A.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison maltilitol CRS.

B. Melting point (2.2.14): 148 °C to 151 °C.

C. Specific optical rotation (2.2.7): + 105.5 to + 108.5

(anhydrous substance).

Dissolve 5.00 g in water R and dilute to 100.0 mL with the

same solvent.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined

in water R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of maltilitol CRS in

water R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of maltilitol CRS and

25 mg of sorbitol CRS in water R and dilute to 10 mL with

the same solvent.

Plate TLC silica gel G plate R.

Mobile phase water R, ethyl acetate R, propanol R

(10:20:70 V/V/V).

Application 2 µL.

Development Over a path of 17 cm.

Drying In air.

Detection Spray with 4-aminobenzoic acid solution R. Dry in a

current of cold air until the acetone is removed. Heat at

100-105 °C for 15 min. Allow to cool and spray with a 2 g/L

solution of sodium periodate R. Dry in a current of cold air.

Heat at 100 °C for 15 min.

System suitability: test solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 5.0 g in water R and dilute to 50 mL with the same

solvent.

Conductivity (2.2.38)

Maximum 20 µS·cm⁻¹.

Dissolve 20.0 g in carbon dioxide-free water R prepared from

distilled water R and dilute to 100.0 mL with the same

solvent. Measure the conductivity of the solution, while

gently stirring with a magnetic stirrer.

Reducing sugars

Maximum 0.2 per cent, expressed as glucose equivalent.

Dissolve 5.0 g in 6 mL of water R with the aid of gentle heat.

Cool and add 20 mL of cupri-citric solution R and a few glass

beads. Heat so that boiling begins after 4 min and maintain

boiling for 3 min. Cool rapidly and add 100 mL of a

2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL

of 0.025 M iodine. With continuous shaking, add 25 mL of a

mixture of 6 volumes of hydrochloric acid R and 94 volumes

of water R and, when the precipitate has dissolved, titrate the

excess of iodine with 0.05 M sodium thiosulfate using 1 mL of

starch solution R, added towards the end of the titration as

indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate

is required.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 5.0 g of the substance to be examined

in 20 mL of water R and dilute to 100.0 mL with the same

solvent.

Reference solution (a) Dissolve 0.50 g of maltilitol CRS in

2.0 mL of water R and dilute to 10.0 mL with the same

solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with water R.

Reference solution (c) Dilute 10.0 mL of reference solution (b)

to 100.0 mL with water R.

Reference solution (d) Dissolve 0.5 g of maltilitol R and 0.5 g of

sorbitol R in 5 mL of water R and dilute to 10.0 mL with the

same solvent.

Column:

— size: l = 0.3 m, Ø = 7.8 mm;

— stationary phase: strong cation-exchange resin (calcium

form) R (9 µm);

— temperature: 85 ± 1 °C.

Mobile phase Degassed water R.

Flow rate 0.5 mL/min.

Detection Refractometer maintained at a constant

temperature.

Injection 20 µL of the test solution and reference

solutions (b), (c) and (d).

Run time 3 times the retention time of maltilitol.

Relative retention With reference to maltilitol (retention

time = about 16 min): impurity B = about 0.8;

impurity A = about 1.8.

System suitability Reference solution (d):
— **resolution**: minimum 2 between the peaks due to maltitol and impurity A.
— **limits**:
— **any impurity**: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— **total**: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);
— **disregard limit**: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Lead (2.4.10)
Maximum 0.5 ppm.

Nickel (2.4.15)
Maximum 1 ppm.

Water (2.5.12)
Maximum 1.0 per cent, determined on 1.00 g.

Microbial contamination

If intended for use in the manufacture of parenteral preparations:
— TAMC: acceptance criterion: 10^2 CFU/g (2.6.12).
If not intended for use in the manufacture of parenteral preparations:
— TAMC: acceptance criterion 10^3 CFU/g (2.6.12);
— TYMC: acceptance criterion 10^2 CFU/g (2.6.12);
— absence of *Escherichia coli* (2.6.13);
— absence of *Salmonella* (2.6.13).

Bacterial endotoxins (2.6.14)

If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins:
— less than 4 IU/g for parenteral preparations having a concentration of less than 100 g/L of maltitol;
— less than 2.5 IU/g for parenteral preparations having a concentration of 100 g/L or more of maltitol.

ASSAY

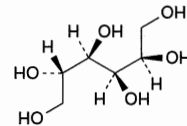
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection Test solution and reference solution (a).
Calculate the percentage content of D-maltitol from the declared content of maltitol CRS.

LABELLING

The label states:

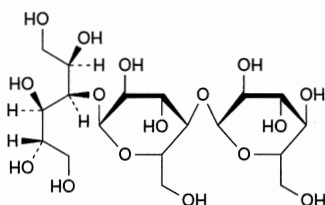
— where applicable, the maximum concentration of bacterial endotoxins;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES



A. D-glucitol (D-sorbitol),

B. O-α-D-glucopyranosyl-(1→4)-O-α-D-glucopyranosyl-(1→4)-D-glucitol (maltotrititol).



Liquid Maltitol

(Ph. Eur. monograph 1236)

Action and use

Excipient.

DEFINITION

Aqueous solution of a hydrogenated, partly hydrolysed starch, composed of a mixture of mainly 4-O-α-D-glucopyranosyl-D-glucitol (D-maltitol) with D-glucitol (D-sorbitol) and hydrogenated oligo- and polysaccharides.

Content

— D-maltitol ($C_{12}H_{24}O_{11}$): minimum 50.0 per cent m/m (anhydrous substance) and 95.0 per cent to 105.0 per cent of the content stated on the label;
— D-sorbitol ($C_6H_{14}O_6$): maximum 8.0 per cent m/m (anhydrous substance);
— anhydrous substance: 68.0 per cent m/m to 85.0 per cent m/m.

CHARACTERS

Appearance
Clear, colourless, syrupy liquid.

Solubility

Miscible with water and with glycerol.

IDENTIFICATION

First identification A.
Second identification B, C.
A. Examine the chromatograms obtained in the assay. Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).
B. Thin-layer chromatography (2.2.27).
Test solution Dilute 0.35 g of the substance to be examined to 100 mL with water R.
Reference solution (a) Dissolve 20 mg of maltitol CRS in water R and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 20 mg of maltitol CRS and 20 mg of sorbitol CRS in water R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase water R, ethyl acetate R, propanol R (10:20:70 V/V/V).

Application 2 µL.

Development Over a path of 17 cm.

Drying In air.

Detection Spray with 4-aminobenzoic acid solution R. Dry in a current of cold air until the acetone is removed. Heat at 100-105 °C for 15 min. Allow to cool and spray with a 2 g/L solution of sodium periodate R. Dry in a current of cold air. Heat at 100 °C for 15 min.

System suitability: reference solution (b):
— the chromatogram shows 2 clearly separated spots.
Results The principal spot in the chromatogram obtained with the test solution is similar in position and colour to the principal spot in the chromatogram obtained with reference solution (a).
C. To 3 mL of a freshly prepared 100 g/L solution of pyrocatechol R, add 6 mL of sulfuric acid R while cooling in iced water. To 3 mL of the cooled mixture, add 0.3 mL of solution S (see Tests). Heat gently over a naked-flame for about 30 s. A pink colour develops.

TESTS

Solution S
Dilute 7.0 g to 50 mL with water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Conductivity (2.2.38)

Maximum 10 µS·cm⁻¹, measured on undiluted liquid maltitol while gently stirring with a magnetic stirrer.

Reducing sugars

Maximum 0.2 per cent, calculated as glucose equivalent. To 5.0 g add 6 mL of water R, 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R and, when the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.

Lead (2.4.10)

Maximum 0.5 ppm.

Nickel (2.4.15)

Maximum 1 ppm.

Water (2.5.12)

15.0 per cent m/m to 32.0 per cent m/m, determined on 0.100 g. Use as solvent a mixture of equal volumes of anhydrous methanol R and formamide R. Carry out the titration at about 50 °C.

ASSAY

Liquid chromatography (2.2.29).

Test solution Mix 1.00 g of the solution to be examined with 20 mL of water R and dilute to 50.0 mL with the same solvent.
Reference solution (a) Dissolve 50.0 mg of maltitol CRS in 2 mL of water R and dilute to 5.0 mL with the same solvent.
Reference solution (b) Dissolve 8.0 mg of sorbitol CRS in 2 mL of water R and dilute to 5.0 mL with the same solvent.
Reference solution (c) Dissolve 50 mg of maltitol R and 50 mg of sorbitol R in 2 mL of water R and dilute to 5.0 mL with the same solvent.

Column:

— size: l = 0.3 m, Ø = 7.8 mm;
— stationary phase: strong cation-exchange resin (calcium form) R (9 µm);

— temperature: 85 ± 2 °C.

Mobile phase Degassed water R.

Flow rate 0.5 mL/min.

Detection Refractometer maintained at a constant temperature.

Injection 20 µL.

Run time 3 times the retention time of maltitol.

Relative retention With reference to maltitol (retention time = about 16 min): sorbitol = about 1.8.

System suitability: reference solution (c):

— resolution: minimum 2 between the peaks due to sorbitol and maltitol.

Calculate the percentage contents of D-maltitol and D-sorbitol from the declared contents of maltitol CRS and sorbitol CRS.

LABELLING

The label states the content of D-maltitol.

Maltodextrin

(Ph. Eur. monograph 1542)

Action and use

Excipient.

Ph. Eur.

DEFINITION

Mixture of glucose, disaccharides and polysaccharides, obtained by the partial hydrolysis of starch.

The degree of hydrolysis, expressed as dextrose equivalent (DE), is less than 20 (nominal value).

CHARACTERS

Appearance

White or almost white, slightly hygroscopic powder or granules.

Solubility

Freely soluble in water.

IDENTIFICATION

A. Dissolve 0.1 g in 2.5 mL of water R and heat with 2.5 mL of cupri-tartaric solution R. A red precipitate is formed.

B. Dip, for 1 s, a suitable stick with a reactive pad containing glucose-oxidase, peroxidase and a hydrogen-donating substance, such as tetramethylbenzidine, in a 100 g/L solution of the substance to be examined. Observe the colour of the reactive pad; within 60 s the colour changes from yellow to green or blue.

C. It is a powder or granules.

D. Dextrose equivalent (see Tests).

TESTS

Solution S
Dissolve 12.5 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

pH (2.2.3)

4.0 to 7.0.

Mix 1 mL of a 223.6 g/L solution of potassium chloride R and 30 mL of solution S.

Sulfur dioxide (2.5.29)

Maximum 20 ppm.



Heavy metals (2.4.8)

Maximum 10 ppm.

Dilute 4 mL of solution S to 30 mL with water R.

The solution complies with test E. Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 6.0 per cent, determined on 10.00 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.

Dextrose equivalent

(DE): within 2 DE units of the nominal value.

Weigh an amount of the substance to be examined equivalent to 2.85-3.15 g of reducing carbohydrates, calculated as dextrose equivalent, into a 500 mL volumetric flask. Dissolve in water R and dilute to 500.0 mL with the same solvent. Transfer the solution to a 50 mL burette.

Pipette 25.0 mL of cupri-tartaric solution R into a 250 mL flask and add 18.5 mL of the test solution from the burette, previously adjusted so that the solution begins to boil within 2 min ± 15 s. Allow to boil for exactly 120 s, add 1 mL of a 1 g/L solution of methylene blue R and titrate with the test solution (V₁) until the blue colour disappears. Maintain the solution at boiling throughout the titration.

Standardise the cupri-tartaric solution using a 6.00 g/L solution of glucose R (V₀).

Calculate the dextrose equivalent using the following expression:

$$\frac{300 \times V_0 \times 100}{V_1 \times M \times D}$$

V₀ = total volume of glucose standard solution, in

millilitres;

V₁ = total volume of test solution, in millilitres;

M = sample mass, in grams;

D = percentage content of dry matter in the substance.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

LABELLING

The label states the dextrose equivalent (DE) (= nominal

value).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate

compliance. Control of these characteristics can however contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for maltodextrin used as filler and binder in tablets and capsules

acid R.

B. Dissolve 50 mg in 5 mL of water R. Add 0.5 mL of ammonium sulfide solution R. A pale pink precipitate is formed that dissolves upon the addition of 1 mL of glacial acetic

reference solution.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the

Detection Spray with a solution containing 25 g/L of

ammonium molybdate R and 10 g/L of cerium sulfate R in dilute

sulfuric acid R, and heat at 105 °C for about 10 min.

Drying At 105 °C for 20 min, then allow to cool to room

temperature.

Development Over 3/4 of the plate.

Application 1 µL.

Mobile phase concentrated ammonia R, ethyl acetate R, water R,

plate R (2-10 µm).

Plate TLC silica gel plate R (5-40 µm) [or TLC silica gel

60 °C.

Reference solution Dissolve 20 mg of calcium gluconate CRS in

1 mL of water R, heating if necessary in a water-bath at

Test solution Dissolve 20 mg of the substance to be examined

in 1 mL of water R.

A. Thin-layer chromatography (2.2.27).

insoluble in water, practically insoluble in anhydrous ethanol,

Soluble in water, practically insoluble in anhydrous ethanol,

insoluble in methylene chloride.

White or pale pink, slightly hygroscopic, crystalline powder.

Appearance

CHARACTERS

98.0 per cent to 102.0 per cent (anhydrous substance).

Content

2,3,4,5,6-pentahydroxyhexanoate] (anhydrous or hydrated

managanese(II) di-(D-gluconate)).

Anydrous or hydrated managanese(II) bis[(2R,3S,4R,5R)-

DEFINITION

Ph Eur

(anhydrous substance)

445.2

C₁₂H₂₂MnO₁₄·xH₂O

Chemical structure diagram of Manganese Gluconate showing the gluconate anion and the manganese cation.

(Ph. Eur. monograph 2162)

Manganese Gluconate

Powder flow (2.9.36)

Particle-size distribution (2.9.31 or 2.9.38).

(see Tests).

Dextrose equivalent

Ph Eur

(Ph. Eur. monograph 2162)

Manganese Gluconate

Powder flow (2.9.36)

Particle-size distribution (2.9.31 or 2.9.38).

(see Tests).

Dextrose equivalent

Ph Eur

(Ph. Eur. monograph 2162)

Manganese Gluconate

Powder flow (2.9.36)

Particle-size distribution (2.9.31 or 2.9.38).

(see Tests).

Dextrose equivalent

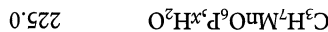
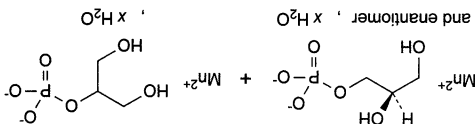
Ph Eur

(Ph. Eur. monograph 2162)

Manganese Gluconate

Hydrated Manganese Glycerocephosphate

(Ph. Eur. monograph 2163)



(anhydrous substance)



TESTS

Solution S Dissolve 1.0 g in water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Sucrose and reducing sugars

Dissolve 0.5 g in a mixture of 2 mL of hydrochloric acid R1 and 10 mL of water R. Boil for 5 min, allow to cool, add 10 mL of sodium carbonate solution R and allow to stand for 10 min. Dilute to 25 mL with water R and filter. To 5 mL of the filtrate add 2 mL of cupri-tartaric solution R and boil for 1 min. Allow to stand for 2 min. No red precipitate is formed.

Chlorides (2.4.4)

Maximum 500 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 500 ppm.

Dissolve 2.0 g in a mixture of 10 mL of acetic acid R and 90 mL of distilled water R.

Zinc

Maximum 50 ppm.

To 10 mL of solution S add 1 mL of sulfuric acid R and 0.1 mL of potassium ferrioxalate solution R. After 30 s, any opalescence in the solution is not more intense than that in a mixture of 1.0 mL of zinc standard solution (10 ppm Zn) R, 9 mL of water R, 1 mL of sulfuric acid R and 0.1 mL of potassium ferrioxalate solution R.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in 20 mL of water R, heating in a water-bath at 60 °C. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.32)

Maximum 9.0 per cent, determined on 80 mg.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (2.6.12).

TYMC: acceptance criterion 10^2 CFU/g (2.6.12).

ASSAY

Dissolve 0.400 g in 50 mL of water R. Add 10 mg of ascorbic acid R, 20 mL of ammonium chloride buffer solution pH 10.0 R and 0.2 mL of a 2 g/L solution of mordant black 11 R in triethanolamine R. Titrate with 0.1 M sodium edetate until the colour changes from violet to pure blue.

1 mL of 0.1 M sodium edetate is equivalent to 44.52 mg of $\text{C}_{12}\text{H}_{22}\text{MnO}_{14}$.

STORAGE

In a non-metallic, airtight container.

Ph Eur

Chlorides (2.4.4)

Maximum 0.15 per cent.

Dissolve 0.22 g in a mixture of 1 mL of nitric acid R and 10 mL of water R and dilute to 100 mL with water R.

Phosphates (2.4.11)

Maximum 0.3 per cent.

Dilute 1.0 mL of solution S to 100.0 mL with water R. To 10 mL of this solution add 140 mL of water R.

Sulfates (2.4.13)

Maximum 0.2 per cent.

Dilute 5 mL of solution S to 50 mL with distilled water R.

Iron (2.4.9)

Maximum 50 ppm.

Dilute 4 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 12.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

ASSAY

To 0.200 g add 1.5 mL of 1 M hydrochloric acid, 50 mL of

water R, 10 mg of ascorbic acid R and 20 mL of ammonium

chloride buffer solution pH 10.0 R. Stir until dissolution.

Immediately add 0.3 mL of a 2 g/L solution of mordant black

11 R in triethanolamine R and titrate with 0.1 M sodium

edetate until the colour changes from violet to pure blue.

1 mL of 0.1 M sodium edetate is equivalent to 22.50 mg

of $C_3H_7MnO_6P$.

STORAGE

In an airtight container.

Manganese Sulfate

Manganese Sulphate

$MnSO_4 \cdot 4H_2O$

223.1

7785-87-7

Ph Eur

Manganese Sulfate Monohydrate

Manganese Sulphate Monohydrate

$MnSO_4 \cdot H_2O$

169.0

10034-96-5

**DEFINITION****Content**

99.0 per cent to 101.0 per cent (ignited substance).

CHARACTERS**Appearance**

Pale pink crystalline powder, slightly hygroscopic.

Solubility

Freely soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. Solution S (see Tests) gives reaction (a) of sulfates (2.3.1).

B. Dissolve 50 mg in 5 mL of water R. Add 0.5 mL of

ammonium sulfide solution R. A pale pink precipitate is formed

which dissolves on the addition of 1 mL of anhydrous acetic

acid R.

C. Loss on ignition (see Tests).

TESTS**Solution S**

Dissolve 10.0 g in distilled water R and dilute to 100 mL with the same solvent.

Iron

mannan (40 ppm).

standard solution (2 ppm Pb) diluted to 50 mL in the same

more intense than that obtained by treating 10 mL of lead

20 seconds. The colour produced within 2 minutes is not

acid and pass hydrogen sulfide through the solution for

Dissolve 0.50 g in 50 mL of water, add 1 mL of 1 M acetic

Heavy metals

(4 ppm).

0.25 g complies with the limit test for arsenic, Appendix VII

Arsenic**TESTS**

C. Yields the reactions characteristic of sulfates, Appendix VI.

A purple solution is produced.

boil gently for a few minutes, add 100 mL of water and filter.

B. To 0.1 g add 2 g of lead(IV) oxide and 5 mL of nitric acid,

soluble in 6 M acetic acid.

sulfide solution. A pink precipitate is produced which is

A. Dissolve 0.5 g in 10 mL of water and add 1 mL of sodium

IDENTIFICATION

Freely soluble in water, practically insoluble in ethanol (96%).

Pale pink crystals or crystalline powder.

CHARACTERISTICS

at 450° to 500°.

of $MnSO_4$, calculated with reference to the substance ignited

It contains not less than 98.0% and not more than 100.5%

Manganese sulfate is manganese(II) sulfate tetrahydrate.

DEFINITION

to 3.020 mg of $MnSO_4$.

Each mL of 0.1 M ammonium iron(II) sulfate VS is equivalent

represents the amount of ammonium iron(II) sulfate required

being examined. The difference between the titrations

permanganate VS. Repeat the operation without the substance

sulfate VS, and titrate immediately with 0.02 M potassium

filtrate and washings in 50 mL of 0.1 M ammonium iron(II)

wash the residue with 40 mL of the mixture, collecting the

of 3 volumes of nitric acid and 97 volumes of water, filter,

boiled and cooled nitric acid, cool, add 1.5 g of sodium

Dissolve 0.15 g in 40 mL of water, add 8 mL of freshly

ASSAY

31.0 to 34.0% of its weight. Use 1 g.

When ignited to constant weight at 450° to 500°, loses

Loss on ignition

chlorides, Appendix VII (330 ppm).

15 mL of a 1.0% w/v solution complies with the limit test for

Chloride

solution (100 ppm Zn) in the same manner (500 ppm).

intense than that obtained by treating 10 mL of zinc standard

stand for 15 minutes. Any turbidity produced is not more

solution of potassium hexacyanoferrate(II), mix and allow to

1 M hydrochloric acid and 0.3 mL of a freshly prepared 3% w/v

Dissolve 2.0 g in 10 mL of water, add 3 mL of

Zinc

(Fe) diluted to 10 mL in the same manner (40 ppm).

obtained by treating 2 mL of iron standard solution (20 ppm

Any colour in the upper layer is not more intense than that

and amyl acetate, shake well and allow to separate.

and 20 mL of a mixture of equal volumes of isoamyl alcohol

Add 5 mL of a 10% w/v solution of ammonium thiocyanate

permanganate until a permanent pink colour is produced.

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Specific optical rotation (2.2.7): + 23 to + 25 (dried

substance).

Dissolve 2.00 g of the substance to be examined and 2.6 g of

disodium tetraborate R in about 20 mL of water R at 30 °C;

shake continuously for 15-30 min without further heating.

Dilute the resulting clear solution to 25.0 mL with water R.

B. Melting point (see Tests).◇

C. Infrared absorption spectrophotometry (2.2.24).

Comparison mannitol CRS.

If the spectra obtained in the solid state show differences,

dissolve separately in 2 glass vials 25 mg of the substance to

be examined and 25 mg of the reference substance in

0.25 mL of distilled water R without heating. The solutions

obtained are clear. Evaporate to dryness by heating in a

microwave oven with a power range of 600-700 W for

20 min or by heating in an oven at 100 °C for 1 h then

gradually applying vacuum until a dry residue is obtained.

Non-sticky, white or slightly yellowish powders are obtained.

Record new spectra using the residues.

◇D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined

in water R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of mannitol CRS in

water R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of mannitol R and

25 mg of sorbitol R in water R and dilute to 10 mL with the

same solvent.

Plate TLC silica gel plate R.

Application 2 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with 4-aminobenzoic acid solution R and dry in

a current of cold air until the acetone is removed; heat at

100 °C for 15 min, allow to cool then spray with a 2 g/L

solution of sodium periodate R; dry in a current of cold air and

heat at 100 °C for 15 min.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

solution (a).◇

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dissolve 5.0 g in water R and dilute to 50 mL with the same

solvent.

Conductivity (2.2.38)

Maximum 20 µS·cm⁻¹.

Dissolve 20.0 g in carbon dioxide-free water R prepared from

distilled water R by heating at 40-50 °C and dilute to

100.0 mL with the same solvent. After cooling, measure the

conductivity of the solution while gently stirring with a

magnetic stirrer.

Appearance of solution

Solution S is not more opalescent than reference

suspension II (2.2.1).

Chlorides (2.4.4)

Maximum 100 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Iron (2.4.9)

Maximum 10 ppm, determined on solution S.

Zinc

Maximum 50 ppm.

To 10 mL of solution S add 1 mL of sulfuric acid R and

0.1 mL of potassium ferrocyanide solution R. After 30 s, any

opalescence in the solution is not more intense than that in a

mixture of 5 mL of zinc standard solution (10 ppm Zn) R,

5 mL of water R, 1 mL of sulfuric acid R and 0.1 mL of

potassium ferrocyanide solution R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (2 ppm Pb) R.

Loss on ignition

10.0 per cent to 12.0 per cent, determined on 1.00 g at

500 ± 50 °C.

ASSAY

Dissolve 0.150 g in 50 mL of water R. Add 10 mg of ascorbic

acid R, 20 mL of ammonium chloride buffer solution pH 10.0 R

and 0.2 mL of 2 g/L solution of mordant black 11 R in

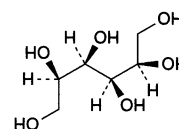
triethanolamine R. Titrate with 0.1 M sodium edetate until the

colour changes from violet to pure blue.

1 mL of 0.1 M sodium edetate is equivalent to 15.10 mg

of MnSO₄.**Mannitol**

(Ph. Eur. monograph 0559)



Ph Eur

69-65-8

Preparation

Mannitol Infusion

DEFINITION

D-Mannitol.

Content

97.0 per cent to 102.0 per cent (dried substance).

◆CHARACTERS

Appearance

White or almost white crystals or powder.

Solubility

Freely soluble in water, practically insoluble in ethanol

(96 per cent).

It shows polymorphism (5.9).◆

Melting point (2.2.14)

165 °C to 170 °C.

Reducing sugars

Maximum 0.1 per cent (calculated as glucose equivalent). To 7.0 g add 13 mL of water R. Boil gently with 40 mL of cupri-tartrate solution R for 3 min, and allow to stand for 2 min. A precipitate is formed. Filter through a sintered-glass filter (16) (2.1.2) coated with diatomaceous earth R or a sintered-glass filter (10) (2.1.2). Wash the precipitate with hot water R (about 50–60 °C) until the washing is no longer alkaline, and filter the washings through the same sintered-glass filter. Discard the filtrate. Immediately dissolve the precipitate in 20 mL of ferric sulfate solution R, filter through the same sintered-glass filter, and wash the filter with 15–20 mL of water R. Combine the washings and the filtrate, heat to 80 °C, and titrate with 0.02 M potassium permanganate. Not more than 3.2 mL is required to change the colour of the solution from green to pink so that the colour persists for at least 10 s.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.50 g of the substance to be examined in 2.5 mL of water R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.50 g of mannitol CRS in 2.5 mL of water R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with water R.

Reference solution (c) Dilute 0.5 mL of reference solution (b) to 20.0 mL with water R.

Reference solution (d) Dissolve 0.25 g of mannitol R and 0.25 g of sorbitol R (impurity A) in 5 mL of water R and dilute to 10.0 mL with the same solvent.

Reference solution (e) Dissolve 0.5 g of mannitol R (impurity B) and 0.5 g of isomalt R (impurity C) in 5 mL of water R and dilute to 100 mL with the same solvent.

Column:
— size: $l = 0.3$ m, $\phi = 7.8$ mm;
— stationary phase: strong cation-exchange resin (calcium form) R (9 μ m);
— temperature: 85 ± 2 °C.

Mobile phase Degassed water R.
Flow rate 0.5 mL/min.

Detection Refractometer maintained at a constant temperature (40 °C for example).
Injection 20 μ L of the test solution and reference solutions (b), (c), (d) and (e).

Run time 1.5 times the retention time of mannitol.
Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peak due to impurity A and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities B and C. **Relative retention** With reference to mannitol (retention time = about 20 min): impurity C (1st peak) = about 0.6; impurity B = about 0.7; impurity C (2nd peak) = about 0.73; impurity A = about 1.2. Impurity C elutes in 2 peaks. Coelution of impurity B and the 2nd peak due to impurity C may be observed.

System suitability Reference solution (d):
— resolution: minimum 2.0 between the peaks due to mannitol and impurity A.

Limits:

— **impurity A**: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— **sum of impurities B and C**: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— **unspecified impurities**: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— **total**: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— **disregard limit**: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Nickel (2.4.15)

Maximum 1 ppm.

Suspend 10.0 g in 30.0 mL of dilute acetic acid R and dilute to 100.0 mL with water R. Use water-saturated methyl isobutyl ketone R.

Heavy metals

Maximum 5 ppm.

Test solution Introduce 5.0 g into a 50 mL colour comparison tube and dissolve with 40 mL of water R. Add 2 mL of dilute acetic acid R1 and dilute to 50 mL with water R.

Reference solution Introduce 2.5 mL of lead standard solution (10 ppm Pb) R into a 50 mL colour comparison tube, add 2 mL of dilute acetic acid R1 and dilute to 50 mL with water R.

Add about 50 μ L of sodium sulfide solution R1 to each of the test solution and the reference solution, mix thoroughly, and allow to stand for 5 min. Examine the solutions by viewing the tubes vertically or horizontally against a white background. The test solution is not more intensely coloured than the reference solution.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Microbial contamination

If intended for use in the manufacture of parenteral preparations:

— TAMC: acceptance criterion 10^2 CFU/g (2.6.12).
If not intended for use in the manufacture of parenteral preparations:

— TAMC: acceptance criterion 10^3 CFU/g (2.6.12);
— TYMC: acceptance criterion 10^2 CFU/g (2.6.12);
— absence of *Escherichia coli* (2.6.13);
— absence of *Salmonella* (2.6.14).

Bacterial endotoxins (2.6.14)

If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins:

— less than 4 IU/g for parenteral preparations having a concentration of 100 g/L or less of mannitol;
— less than 2.5 IU/g for parenteral preparations having a concentration of more than 100 g/L of mannitol. ♦

ASSAY

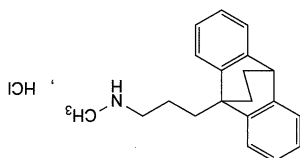
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).
Calculate the percentage content of D-mannitol taking into account the assigned content of mannitol CRS.



Maprotiline Hydrochloride

(Ph. Eur. monograph 1237)



$C_{20}H_{24}ClN$

313.9

10347-81-6

Action and use
Antidepressant.

Ph Eur

DEFINITION
3-(9,10-Ethanoanthracen-9(10H)-yl)-N-methylpropan-1-amine hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent), sparingly soluble in methylene chloride, very slightly soluble in acetone.

It shows polymorphism (5.9).

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 10 mg in 1 M hydrochloric acid and dilute to 100 mL with the same acid.

Spectral range 250-300 nm.

Absorption maxima At 265 nm and 272 nm.

Absorption minimum At 268 nm.

Absorbance ratio $A_{272}/A_{265} = 1.1$ to 1.3.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison maprotiline hydrochloride CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of maprotiline hydrochloride CRS in methanol R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of maprotiline impurity D CRS in reference solution (a) and dilute to 2 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ethyl acetate R, dilute ammonia R1, 2-butanone R (4:5:14 V/V/V).

Application 5 μ L.

Development Over half of the plate.

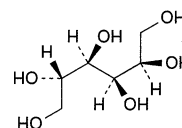
LABELLING

The label states:

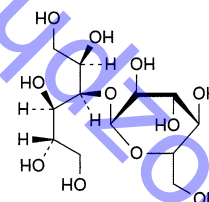
- where applicable, the maximum concentration of bacterial endotoxins;
- where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES

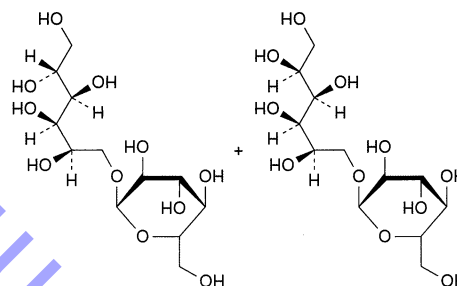
Specified impurities A, B, C.



A. D-glucitol (D-sorbitol),



B. 4-O-α-D-glucopyranosyl-D-glucitol (D-maltitol).



C. mixture of 6-O-α-D-glucopyranosyl-D-glucitol and 1-O-α-D-glucopyranosyl-D-mannitol (isomalt).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15).

Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for mannitol used as filler in tablets and capsules.

Particle-size distribution (2.9.31 or 2.9.38).

Powder flow (2.9.36)

Ph Eur

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.

Drying In a current of warm air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dilute 0.5 mL of solution S (see Tests) to 2 mL with methanol R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S Dissolve 1.0 g in methanol R and dilute to 20 mL with the same solvent.

Appearance of solution Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Related substances Liquid chromatography (2.2.29):

Test solution Dissolve 0.10 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 1.0 mg of maprotiline impurity D CRS in the test solution and dilute to 10.0 mL with the test solution.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: silica gel for chromatography R (5 μ m).

Mobile phase Dissolve about 0.580 g of ammonium acetate R in 200 mL of water R and add 2 mL of a 70 g/L solution of concentrated ammonia R; add 150 mL of 2-propanol R and 650 mL of methanol R; the resulting apparent pH value is between 8.2 and 8.4.

Flow rate 1 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of maprotiline.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.

Relative retention With reference to maprotiline (retention time = about 10 min): impurity A = about 0.3;

impurity B = about 0.5; impurity C = about 0.7;

impurity D = about 0.8; impurity E = about 1.3.

System suitability: reference solution (b):

— resolution: 1.8 to 3.2 between the peaks due to impurity D and maprotiline; if necessary, adjust the pH of the mobile phase, in steps of 0.1 pH unit, by adding a

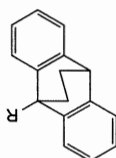
50 per cent *V/V* solution of acetic acid R if the resolution is less than 1.8, or by adding a 70 g/L solution of

concentrated ammonia R if the resolution is greater than 3.2.

Limits:

— impurities A, B, C, D, E: for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);



Specified impurities A, B, C, D, E.

IMPURITIES

1 mL of 0.1 M sodium hydroxide is equivalent to 31.39 mg of $C_{20}H_{24}ClN$.

ASSAY

Dissolve 0.250 g in a mixture of 5 mL of 0.1 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 31.39 mg of $C_{20}H_{24}ClN$.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

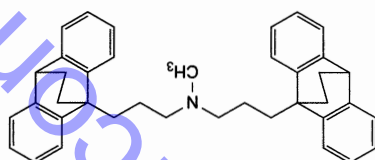
Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 80 °C at a pressure not exceeding 2.5 kPa for 6 h.

B. 3-(9,10-ethanoanthracen-9(10H)-yl)-N-[3-(9,10-ethanoanthracen-9(10H)-yl)propyl]-N-methylpropan-1-amine.



9(10H)-yl)-N,N-dimethylpropylpropan-1-amine,

E. R = $CH_2-CH_2-CH_2-N(CH_3)_2$; 3-(9,10-ethanoanthracen-

(dehydro)maprotiline),

9(10H)-yl)-N-methylprop-2-en-1-amine

D. R = $CH=CH-CH_2-NH-CH_3$; 3-(9,10-ethanoanthracen-

9(10H)-yl)propan-1-amine,

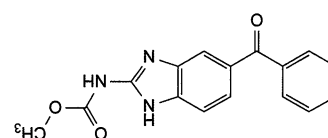
C. R = $CH_2-CH_2-CH_2-NH_2$; 3-(9,10-ethanoanthracen-

yl)prop-2-enal,

A. R = $CH=CH-CH=O$; 3-(9,10-ethanoanthracen-9(10H)-

Mebednadazole

(Ph. Eur. monograph 0845)


 $C_{16}H_{13}N_3O_3$ 295.3 31431-39-7

Action and use

Benzimidazole antihelminthic.

Ph Eur

DEFINITION

Methyl (5-benzoyl-1H-benzimidazol-2-yl)carbamate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (5.9). The acceptable crystalline form corresponds to mebednadazole CRS.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison mebednadazole CRS.

Preparation Examine the substances without prior treatment.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in dimethylformamide R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of mebednadazole for system suitability CRS (containing impurities A, B, C, D, E, F and G) in dimethylformamide R and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with dimethylformamide R. Dilute 5.0 mL of this solution to 20.0 mL with dimethylformamide R.

Column:

size: $l = 0.10$ m, $\varnothing = 4.6$ mm;stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (3 μ m);

temperature: 40 °C.

Mobile phase:

mobile phase A: 7.5 g/L solution of ammonium acetate R;

mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	80 \rightarrow 70	20 \rightarrow 30
15 - 20	70 \rightarrow 10	30 \rightarrow 90
20 - 25	10	90

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 250 nm.
Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with mebednadazole for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E, F and G.

Relative retention With reference to mebednadazole (retention time = about 12 min): impurity A = about 0.4; impurity B = about 0.5; impurity C = about 0.7; impurity D = about 1.1; impurity E = about 1.3; impurity F = about 1.4; impurity G = about 1.6.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 4, where H_p = height above the baseline of the peak due to impurity D and H_b = height above the baseline of the lowest point of the curve separating this peak from the peak due to mebednadazole.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity G by 1.4;

— impurity G: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurities A, B, C, D, E, F: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);

— unspecified impurities: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— disintegration limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 3 mL of anhydrous formic acid R and add 50 mL of a mixture of 1 volume of anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

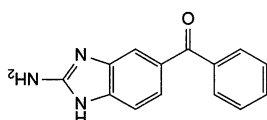
1 mL of 0.1 M perchloric acid is equivalent to 29.53 mg of $C_{16}H_{13}N_3O_3$.

STORAGE

Protected from light.

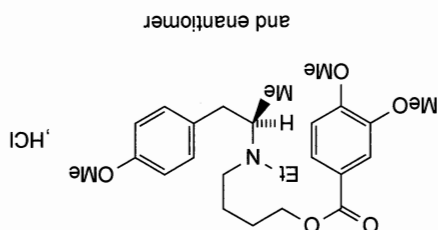
IMPURITIES

Specified impurities A, B, C, D, E, F, G



A. (2-amino-1H-benzimidazol-5-yl)phenylmethanone.

Mebeverine Hydrochloride



$C_{25}H_{35}NO_5 \cdot HCl$ 466.0 2753-45-9

Action and use

Smooth muscle relaxant; antispasmodic.

Preparation

Mebeverine Tablets

DEFINITION

Mebeverine Hydrochloride is (RS)-4-[ethyl(4-methoxy- α -methylphenethyl)amino]butyl veratrate hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{25}H_{35}NO_5 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder. Very soluble in water; freely soluble in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of mebeverine hydrochloride (RS 209).
B. Dissolve 25 mg in 2 mL of water, acidity with 2M nitric acid and centrifuge. The supernatant liquid yields reaction A characteristic of chlorides, Appendix VI, beginning at the words 'add 0.4 mL of...'.
TESTS

Acidity

pH of a 2% w/v solution, 4.5 to 6.5, Appendix V L.

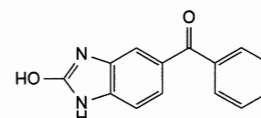
Ether-soluble extractive

Dissolve 40 mg in 25 mL of 2M hydrochloric acid and shake with 50 mL of ether for 1 minute. Wash the ether layer with three 25 mL quantities of water; evaporate the ether to dryness using a rotary evaporator and dissolve the residue in sufficient methanol to produce 20 mL. The absorbance of the resulting solution at 260 nm is not more than 0.23, Appendix II B.

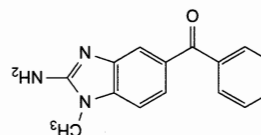
Non-tertiary amine

Dissolve 0.5 g in 5 mL of pyridine, add 5 mL of copper chloride-pyridine reagent and heat at 50° for 30 minutes. Cool, add sufficient acetone to produce 50 mL and measure the absorbance of the resulting solution at 405 nm, Appendix II B, using in the reference cell a solution obtained by treating 5 mL of pyridine in the same manner. The absorbance is not more than that obtained by repeating the test using 5 mL of a 0.0060% w/v solution of di-n-butylamine in pyridine and beginning at the words 'add 5 mL of copper chloride-pyridine reagent...'.
Related substances

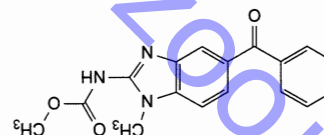
Carry out the method for thin-layer chromatography, Appendix III A, using a silica gel F₂₅₄ precoated plate (Merck silica gel 60 F₂₅₄ plates are suitable) and a mixture of 50 volumes of absolute ethanol, 50 volumes of chloroform and



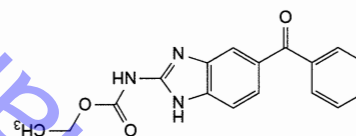
B. (2-hydroxy-1H-benzimidazol-5-yl)phenylmethanone, phenylmethanone,



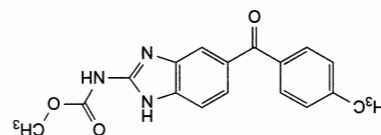
C. (2-amino-1-methyl-1H-benzimidazol-5-yl)phenylmethanone,



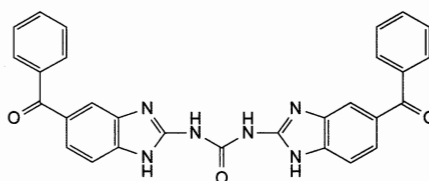
D. methyl (5-benzoyl-1-methyl-1H-benzimidazol-2-yl)carbamate,



E. ethyl (5-benzoyl-1H-benzimidazol-2-yl)carbamate,



F. methyl [5-(4-methylbenzoyl)-1H-benzimidazol-2-yl]carbamate,



G. N,N'-bis(5-benzoyl-1H-benzimidazol-2-yl)urea.

Ph Eur

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).
Test solution Dissolve 15.0 mg in 0.1 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of this solution to 100.0 mL with 0.1 M hydrochloric acid.

Spectral range 220-350 nm.
Absorption maximum At 232 nm.

Specific absorbance at the absorption maximum 345 to 380 (anhydrous substance).

The solution also shows a weak absorbance without a defined maximum between 260 nm and 300 nm.

B. Infrared absorption spectrophotometry (2.2.24).
Comparison meclozine dihydrochloride CRS.

C. Thin-layer chromatography (2.2.27).
Test solution Dissolve 50 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of meclozine dihydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase diethylamine R, toluene R, cyclohexane R (10:15:75 V/V/V).

Application 10 µL.
Development Over 2/3 of the plate.

Drying In a current of warm air for 5 min.
Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 15 mg in 2 mL of ethanol (96 per cent) R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Acidity or alkalinity
Calculate the acidity or alkalinity from the titration volumes obtained in the assay using the following equation:

$$A = V_2 - 2V_1$$

V_1 = volume of 0.1 M sodium hydroxide added at the 1st point of inflexion;
 V_2 = volume of 0.1 M sodium hydroxide added at the 2nd point of inflexion.

A is not less than -0.3 mL and not more than 0.3 mL for 0.350 g of the substance to be examined.

Related substances
Liquid chromatography (2.2.29).
Solvent mixture acetonitrile R, water R (50:50 V/V).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

Reference solution (b) Dissolve 7.5 mg of meclozine impurity B CRS and 7.5 mg of meclozine impurity H CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Column: — size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

Second identification A, C, D.

First identification B, D.

IDENTIFICATION
Slightly soluble in water, soluble in ethanol (96 per cent) and in methylene chloride.

Solubility
White or yellowish-white, slightly hygroscopic, crystalline powder.

Appearance
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Content

1-[(R,S)-(4-Chlorophenyl)phenylmethyl]-4-[(3-methylphenyl)methyl]piperazine dihydrochloride.

1 volume of 18M ammonia as the mobile phase. Apply separately to the plate 10 µL of each of three solutions in acetone containing (1) 2.0% w/v of the substance being examined, (2) 0.010% w/v of the substance being examined and (3) 0.0020% w/v of veratric acid. After removal of the plate, allow it to dry in air and examine under ultraviolet light (254 nm). Expose the plate to iodine vapour for 1 hour.

When viewed under ultraviolet light any spot corresponding to veratric acid in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (3). By each method of visualisation any other secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

Loss on drying
When dried at 105° for 1 hour, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.1%, Appendix IX A.

ASSAY
Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.4 g, adding 7 mL of mercury(II) acetate solution and determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 46.60 mg of C₂₅H₂₉Cl₃N₂·2HCl.

STORAGE
Meclozine Hydrochloride should be kept in an airtight container, protected from light and stored at a temperature not exceeding 30°.

Meclozine Hydrochloride
(Meclozine Dihydrochloride, Ph Eur monograph 0622)

Action and use
Histamine H₁ receptor antagonist; antihistamine.

Chemical structure
C₂₅H₂₉Cl₃N₂ 463.9 1104-22-9

Ph Eur

DEFINITION
1-[(R,S)-(4-Chlorophenyl)phenylmethyl]-4-[(3-methylphenyl)methyl]piperazine dihydrochloride.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or yellowish-white, slightly hygroscopic, crystalline powder.

Solubility
Slightly soluble in water, soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION
First identification B, D.
Second identification A, C, D.

— stationary phase: end-capped octadecylsilyl amorphous organosilica polymer R (3.5 µm);

— temperature: 35 °C.

Mobile phase:

— mobile phase A: 0.1 per cent V/V solution of concentrated ammonia R₃

— mobile phase B: acetonitrile R₁;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0-3	60	40
3-13	60 → 15	40 → 85
13-23	15 → 5	85 → 95
23-33	5	95

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and H.

Relative retention With reference to meclozine (retention

time = about 18 min): impurity B = about 0.45;

impurity H = about 0.49.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to

impurities B and H.

Limits:

— impurity B: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Water (2.5, 12)

Maximum 5.0 per cent, determined on 0.200 g.

Sulfated ash (2.4, 14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 50 mL of ethanol (96 per cent) R. Carry

out a potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 46.39 mg

of C₂₅H₂₉Cl₃N₂.

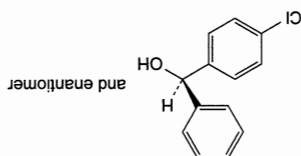
STORAGE

In an airtight container.

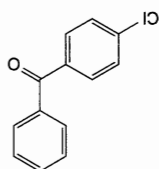
IMPURITIES

Specified impurities B

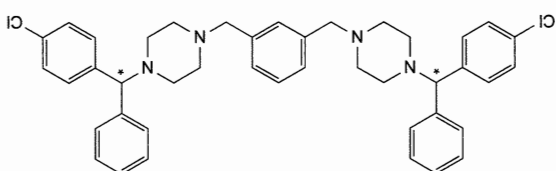
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these



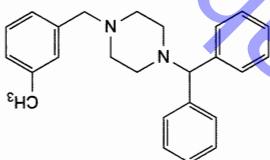
B. (R,S)-(4-chlorophenyl)phenylmethanol,



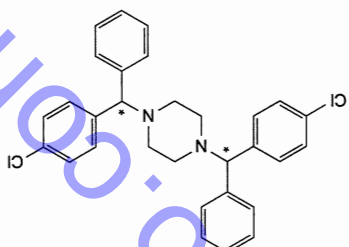
C. (4-chlorophenyl)phenylmethanone (4-chlorobenzophenone),



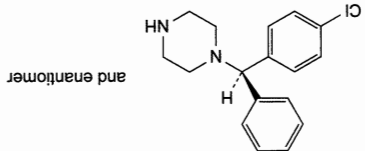
D. 1,1'-(1,3-phenylenebis(methylene))bis[4-((4-chlorophenyl)phenylmethyl)piperazine],



E. 1-(diphenylmethyl)-4-((3-methylphenyl)methyl)piperazine,



F. 1,4-bis[4-chlorophenyl]phenylmethylpiperazine,

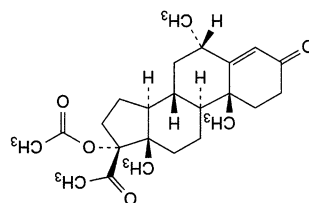


H. 1-[(R,S)-(4-chlorophenyl)phenylmethyl]piperazine.

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: C, D, E, F, H.

Medroxyprogesterone Acetate

(Ph. Eur. monograph 0673)

C₂₁H₃₄O₄ 386,5

71-58-9

Action and use

Progestogen.

Preparations

Medroxyprogesterone Injection

Medroxyprogesterone Tablets

Ph Eur

DEFINITION

6 α -Methyl-3,20-dioxopregn-4-en-17-yl acetate.

Content

97,0 per cent to 103,0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in acetone, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison medroxyprogesterone acetate CRS.

TESTS

Specific optical rotation (2.2.7)

+ 47 to + 53 (dried substance).

Dissolve 0,250 g in acetone R and dilute to 25,0 mL with the same solvent.

Impurity F

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in 5,0 mL of acetonitrile R1 and dilute to 10,0 mL with water for chromatography R.

Reference solution (a) Dilute 1,0 mL of the test solution to 20,0 mL with the mobile phase. Dilute 1,0 mL of this solution to 10,0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of medroxyprogesterone acetate for peak identification CRS (containing impurity F) in 3,0 mL of acetonitrile R1 and dilute to 5,0 mL with water for chromatography R.

Column: size: $l = 0,10$ m, $\varnothing = 4,6$ mm; stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m).

Mobile phase water for chromatography R, acetonitrile R1 (44:56 V/V).

Flow rate 1,0 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 25 μ L.

Identification of impurities Use the chromatogram supplied with medroxyprogesterone acetate for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity F.

Relative retention With reference to medroxyprogesterone acetate (retention time = about 8 min):

Limit:

— **correction factor:** for the calculation of content, multiply the peak area of impurity F by 1,8;

— **impurity F:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0,5 per cent).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, water R (50:50 V/V).

Test solution Dissolve 20 mg of the substance to be examined in the solvent mixture and dilute to 10,0 mL with the solvent mixture.

Reference solution (a) Dissolve 4 mg of medroxyprogesterone acetate for system suitability CRS (containing impurities A, B, C, D, E, G and I) in the solvent mixture and dilute to 2,0 mL with the solvent mixture.

Reference solution (b) Dilute 1,0 mL of the test solution to 100,0 mL with the solvent mixture.

Reference solution (c) Dilute 1,0 mL of reference solution (b) to 10,0 mL with the solvent mixture.

Column:

size: $l = 0,25$ m, $\varnothing = 3,0$ mm;

— **stationary phase:** end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— **temperature:** 60 °C.

Mobile phase tetrahydrofuran R, acetonitrile R, water R (12:23:65 V/V/V).

Flow rate 0,9 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 μ L.

Run time Twice the retention time of medroxyprogesterone acetate.

Identification of impurities Use the chromatogram supplied with medroxyprogesterone acetate for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E, G and I.

Relative retention With reference to medroxyprogesterone acetate (retention time = about 20 min):

impurity A = about 0,3; impurity I = about 0,5;

impurity H = about 0,65; impurity B = about 0,7;

impurity C = about 0,8; impurity G = about 0,85;

impurity D = about 0,9; impurity E = about 0,95.

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 2,5, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to medroxyprogesterone acetate.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1,5;

impurity G = 2,6;

— **impurity D:** not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1,0 per cent);

— *impurity B*: not more than 0.7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);

— *impurity A*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

— *impurities C, E, G, I*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total*: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C for 3 h.

ASSAY

Dissolve 50.0 mg in ethanol (96 per cent) *R* and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with ethanol (96 per cent) *R*. Measure the absorbance (2.2.25) at the absorption maximum at 241 nm.

Calculate the content of $C_{24}H_{34}O_4$ taking the specific absorbance to be 420.

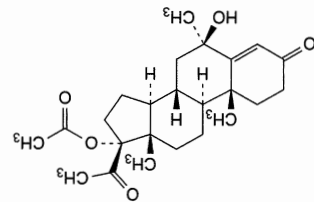
STORAGE

Protected from light.

IMPURITIES

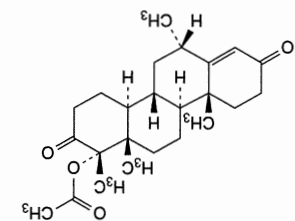
Specified impurities A, B, C, D, E, F, G, I.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): *H*.

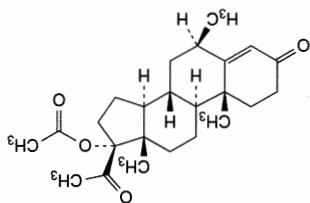


A. 6β-hydroxy-6-methyl-3,20-dioxopregn-4-en-17-yl acetate (6-hydroxymedroxyprogesterone acetate),

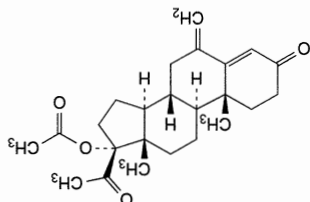
B. 17-hydroxy-6α-methylpregn-4-ene-3,20-dione (medroxyprogesterone),



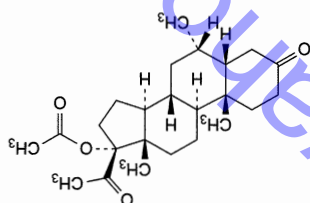
C. 6α,17α-dimethyl-3,17-dioxo-D-homoandroster-4-en-17α-yl acetate,



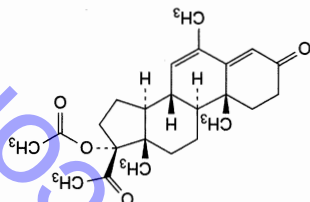
D. 6β-methyl-3,20-dioxopregn-4-en-17-yl acetate (6-epimedroxyprogesterone acetate),



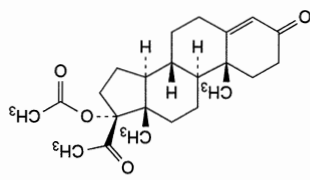
E. 6-methylidene-3,20-dioxopregn-4-en-17-yl acetate (6-methylenedroxyprogesterone acetate),



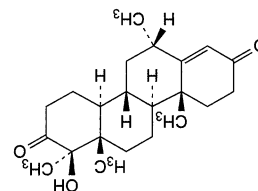
F. 6α-methyl-3,20-dioxo-5β-pregn-17-yl acetate (4,5-dihydrodromedroxyprogesterone acetate),



G. 6-methyl-3,20-dioxopregn-4,6-dien-17-yl acetate (mestrol acetate),



H. 3,20-dioxopregn-4-en-17-yl acetate (hydroxyprogesterone acetate),



I. 17α-hydroxy-6,17a-dimethyl-17a-homoandrost-4-ene-3,17-dione.

Ph Eur

Mefenamic Acid

(Ph. Eur. monograph 1240)



61-68-7

Action and Use
C₁₅H₁₅NO₂
Cyo-oxygenase Inhibitor, anergic anti-inflammatory.

Preparations

Mefenamic Acid capsules

Mefenamic Acid tablets

Ph Eur

DEFINITION

2-[(2,3-Dimethylphenyl)amino]benzoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, microcrystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison mefenamic acid CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in ethanol (96 per cent) R, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 50 mg of 2-chlorobenzoic acid R (impurity C) and 50 mg of benzoic acid R (impurity D) in the

mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 10.0 mg of mefenamic acid impurity A CRS in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve 20.0 mg of benzoic acid R in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

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Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

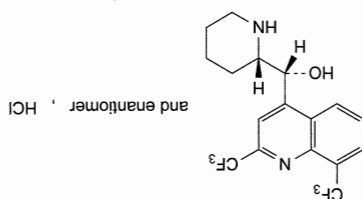
Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Mefloquine Hydrochloride

(Ph Eur monograph 1241)



C₁₇H₁₇ClF₆N₂O 414.8 51773-92-3

Action and use

Antiprotozoal (malaria).

Ph Eur

DEFINITION

(RS)-[2,8-Bis(trifluoromethyl)quinolin-4-yl][(2S)-piperidin-2-yl]methanol hydrochloride.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly yellow, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

mp: about 260 °C, with decomposition.

IDENTIFICATION

First identification A, E.

Second identification B, C, D, E.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison mefloquine hydrochloride CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in methanol, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 8 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 8 mg of mefloquine hydrochloride CRS in methanol R and dilute to 5 mL with the same solvent.

Reference solution (b) Dilute 2.5 mL of the test solution to 100 mL with methanol R.

Reference solution (c) To 1 mL of reference solution (b) add 1 mL of a 0.016 g/L solution of quinidine sulfate R in methanol R.

Plate TLC silica gel F₂₅₄ plate R.

Development Develop the plate with a mixture of 20 volumes of methanol R and 80 volumes of methylene chloride R, and dry at 100-105 °C for 15 min before use.

Mobile phase anhydrous acetic acid R, methanol R, methylene chloride R (10:10:80 V/V/V).

Application 20 µL.

Drying In a current of warm air for 15 min.

in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solutions Prepare the reference solutions using copper standard solution (0.1 per cent Cu) R, diluting with 0.1 M nitric acid.

Source Copper hollow-cathode lamp.

Wavelength 324.8 nm.

Atomisation device Air-acetylene flame.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

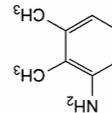
Dissolve with the aid of ultrasound 0.200 g in 100 mL of warm anhydrous ethanol R, previously neutralised to phenol red solution R. Add 0.1 mL of phenol red solution R and titrate with 0.1 M sodium hydroxide.

1 mL of 0.1 M sodium hydroxide is equivalent to 24.13 mg of C₁₇H₁₅NO₂.

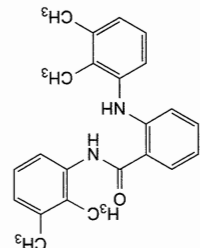
IMPURITIES

Specified impurities A, C, D.

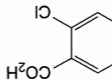
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, E.



A. 2,3-dimethylaniline,

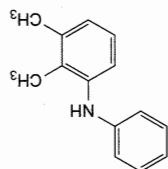


B. N-(2,3-dimethylphenyl)-2-[(2,3-dimethylphenyl)amino]benzamide,



C. 2-chlorobenzoic acid,

D. benzoic acid,



E. 2,3-dimethyl-N-phenylaniline.

Ph Eur



Detection Examine in ultraviolet light at 254 nm; lightly spray with a mixture prepared immediately before use of 1 volume of *sulfuric acid R* and 40 volumes of *iodoplatinate reagent R*; spray with strong hydrogen peroxide solution *R*.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Mix about 10 mg with 45 mg of heavy magnesium oxide *R* and ignite in a crucible until a practically white residue is obtained. Allow to cool, then add 2 mL of water *R*, 0.05 mL of phenolphthalein solution *R1* and about 1 mL of dilute hydrochloric acid *R* to make the solution colourless. Filter.

To the filtrate add a freshly prepared mixture of 0.1 mL of alizarin *S* solution *R* and 0.1 mL of zirconyl nitrate solution *R*. Mix, allow to stand for 5 min and compare the colour of the solution with a blank prepared in the same manner. The test solution is yellow and the blank is red.

D. To about 20 mg add 0.2 mL of sulfuric acid *R*. Blue fluorescence appears in ultraviolet light at 365 nm.

E. It gives reaction (b) of chlorides (2.3.1).

TESTS

Solution S Dissolve 2.50 g in methanol *R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution *S* is clear (2.2.1) and not more intensely coloured than reference solution *BY₇* (2.2.2, Method *I*).

Optical rotation (2.2.7)

–0.2° to +0.2°, determined on solution *S*.

Related substances

Test solution Dissolve 0.10 g of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 8 mg of mefloquine hydrochloride *CRS* and 8 mg of quinidine sulfate *R* in the mobile phase, then dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Precolumn:

— size: $l = 0.025$ m, $\phi = 4$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography *R* (5 μ m).

Column:

— size: $l = 0.25$ m, $\phi = 4$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography *R* (5 μ m).

Mobile phase Dissolve 1 g of tetraheptylammonium bromide *R* in a mixture of 200 volumes of methanol *R*, 400 volumes of 1.5 g/L solution of sodium hydrogen sulfate *R* and 400 volumes of acetonitrile *R*.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 280 nm.

Equilibration With the mobile phase at a flow rate of 2 mL/min for about 30 min.

Injection 20 μ L.

System suitability: reference solution (b):

— resolution: minimum 8.5 between the peaks due to quinidine and mefloquine.

Limits:

— impurity with a relative retention with reference to mefloquine of about 0.7: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— sum of impurities other than the impurity with a relative retention with reference to mefloquine of about 0.7: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test *C*. Prepare the reference solution using 2 mL of lead standard solution (10 ppm *Pb*) *R*.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.000 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

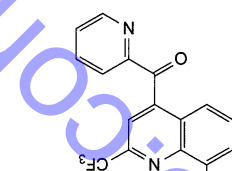
ASSAY

Dissolve 0.350 g in 15 mL of anhydrous formic acid *R* and add 40 mL of acetic anhydride *R*. Titrate with 0.1 *M* perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 *M* perchloric acid is equivalent to 41.48 mg of $C_{17}H_{17}ClF_6N_2O$.

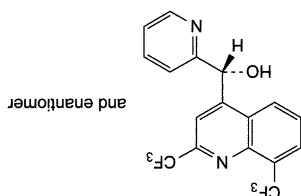
STORAGE

Protected from light.

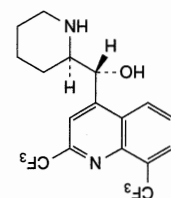
IMPURITIES



A. [2,8-bis(trifluoromethyl)quinolin-4-yl](pyridin-2-yl)methanone,



B. (R,S)-[2,8-bis(trifluoromethyl)quinolin-4-yl](pyridin-2-yl)methanol,

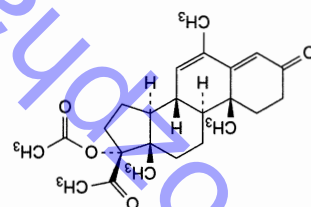


C. (R)-[2,8-bis(trifluoromethyl)quinolin-4-yl]piperidin-2-ylmethanol.

Ph Eur

Megestrol Acetate

(Ph. Eur. monograph 1593)



C₂₄H₃₂O₄ 384.5 595-33-5

Action and use

Progestogen.

Preparation

Megestrol Tablets

Ph Eur

DEFINITION

6-Methyl-3,20-dioxopregna-4,6-dien-17-yl acetate.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone, sparingly

soluble in ethanol (96 per cent).

mp

About 217 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison megestrol acetate CRS.

TESTS

Specific optical rotation (2.2.7)

+ 14.0 to + 17.0 (dried substance).

Dissolve 2.50 g in methylene chloride R and dilute to 25.0 mL

with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetic acid R, water R, acetonitrile R1

(0.1:20:80 V/V/V).

Test solution (a) Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of megestrol acetate for system suitability CRS (containing impurities A, D, G, H, I, J

and L) in 1.0 mL of the solvent mixture.

Reference solution (c) Dissolve 10 mg of megestrol acetate for

peak identification CRS (containing impurities B, C and E) in

1.0 mL of the solvent mixture.

Reference solution (d) Dissolve 50.0 mg of megestrol

acetate CRS in the solvent mixture and dilute to 50.0 mL

with the solvent mixture.

Reference solution (e) Dissolve the contents of a vial of

megestrol acetate for impurity K identification CRS in 1.0 mL of the solvent mixture.

Column:

— size: l = 0.15 m; Ø = 4.6 mm;

— stationary phase: base-deactivated end-capped octadecylsilyl

silica gel for chromatography R (3 µm);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: tetrahydrofuran R, acetonitrile R1, water R

(7.5:12.5:80 V/V/V);

— mobile phase B: water R, tetrahydrofuran R, acetonitrile R1

(20:30:50 V/V/V);

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 245 nm and, for impurity J,

at 210 nm.

Injection 20 µL of test solution (a) and reference solutions (a),

(b), (c) and (e).

Identification of impurities Use the chromatogram supplied

with megestrol acetate for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, D, G, H, I, J and L;

use the chromatogram supplied with megestrol acetate for peak

identification CRS and the chromatogram obtained with

reference solution (c) to identify the peaks due to

impurities B, C and E; use the chromatogram obtained with

reference solution (e) to identify the peak due to impurity K.

Relative retention With reference to megestrol acetate

(retention time = about 22 min): impurity B = about 0.75;

impurity E = about 0.80; impurity K = about 0.83;

impurity C = about 0.9; impurity D = about 1.11;

impurity A = about 1.14; impurity I = about 1.2;

impurity G = about 1.3; impurity J = about 1.4;

impurity H = about 1.5; impurity L = about 1.9.

System suitability: reference solution (b):

— peak-to-valley ratio: minimum 5.0, where H_p = height

above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

impurity A.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.2; impurity D = 0.4; impurity E = 0.4; impurity I = 0.5; impurity K = 0.2; impurity L = 0.6;

— **impurity A:** not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— **impurities D, H:** for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— **impurity J at 210 nm:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) at 210 nm (0.3 per cent);

— **impurity G:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **impurities B, C, E, I, K, L:** for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **sum of impurities other than J:** not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{24}H_{32}O_4$ taking into account the assigned content of megestrol acetate CRS.

STORAGE

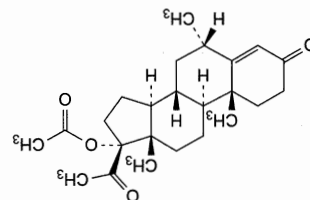
Protected from light.

IMPURITIES

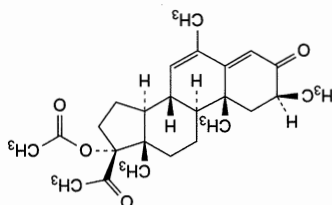
Specified impurities A, B, C, D, E, G, H, I, J, K, L

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): F.

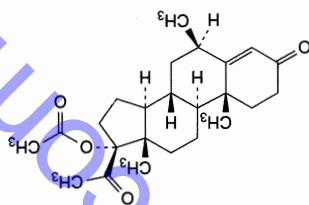
A. 6 α -methyl-3,20-dioxopregna-4-en-17-yl acetate (medroxyprogesterone acetate),



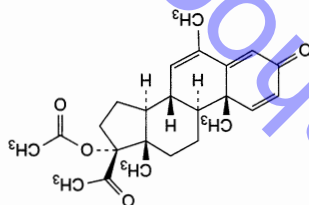
G. 2 β ,6-dimethyl-3,20-dioxopregna-4,6-dien-17-yl acetate,



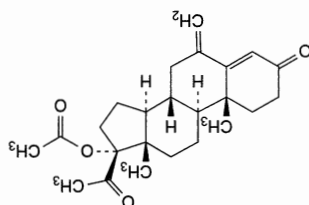
F. 6 β -methyl-3,20-dioxopregna-4-en-17-yl acetate,



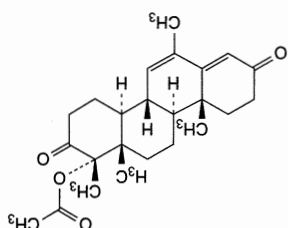
E. 6-methyl-3,20-dioxopregna-1,4,6-trien-17-yl acetate,



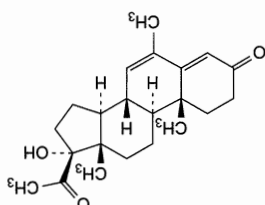
D. 6-methylene-3,20-dioxopregna-4-en-17-yl acetate (6-methylene hydroxyprogesterone acetate),



C. 6,17 α -dimethyl-3,17-dioxo-D-homoandrost-4,6-dien-17 α -yl acetate (D-homo megestrol acetate),

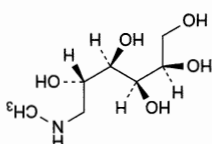


B. 6-methyl-17-hydroxypregna-4,6-diene-3,20-dione (megestrol),



Meglumine

(Ph. Eur. monograph 2055)



C₇H₁₇NO₅ 195.2 6284-40-8

Action and use

Organic base used in the preparation of organic acids.

DEFINITION

1-Deoxy-1-(methylamino)-D-glucitol.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

mp

About 128 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison meglumine CRS.

TESTS

Solution S

Dissolve 20.0 g in distilled water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and its absorbance (2.2.25) at 420 nm is not greater than 0.03.

Dissolve the residue obtained in the test for loss on drying in water R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

–16.0 to –17.0 (dried substance).

Dilute 12.5 mL of solution S to 25.0 mL with water R.

Reducing substances

Maximum 0.2 per cent, expressed as glucose.

Dilute 1.25 mL of solution S to 2.5 mL with water R, add 2 mL of cupri-tartrate solution R and heat on a water-bath for 10 min. Cool under running water for 1 min, then sonicate for 20 s. Immediately filter through a filter 25 mm in diameter and 0.5 µm in pore size. Rinse with 10 mL of water R. Prepare a standard in the same manner using 2.5 mL of a solution obtained by dissolving 20 mg of glucose R in water R and diluting to 100 mL with the same solvent. Any precipitate on the membrane filter obtained with the test solution is not more intensely coloured than the precipitate obtained with the standard.

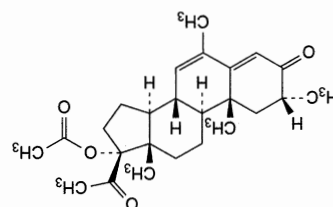
Chlorides (2.4.4)

Maximum 100 ppm.

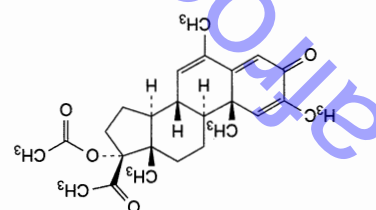
To 2.5 mL of solution S add 12.5 mL of water R.

Sulfates (2.4.13)

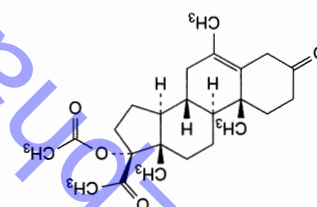
Maximum 150 ppm.



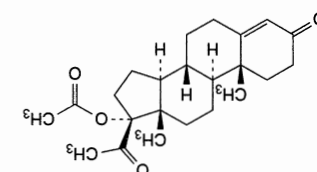
H. 2α,6-dimethyl-3,20-dioxopregna-4,6-dien-17-yl acetate,



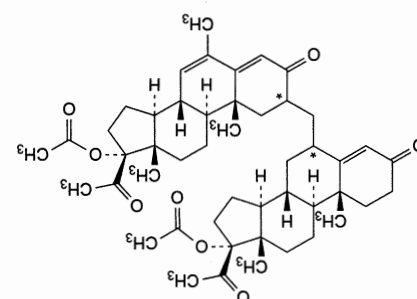
I. 2,6-dimethyl-3,20-dioxopregna-1,4,6-trien-17-yl acetate,



J. 6-methyl-3,20-dioxopregna-5-en-17-yl acetate,



K. 3,20-dioxopregna-4-en-17-yl acetate,

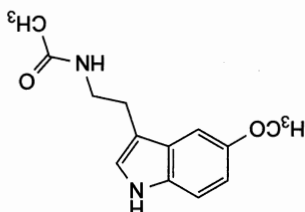


L. 2ξ-[[17-(acetyloxy)-3,20-dioxopregna-4-en-6ξ-yl]methyl]-6-methyl-3,20-dioxopregna-4,6-dien-17-yl acetate (mestrol acetate dimer).

Ph Eur



Melatonin



$C_{13}H_{16}N_2O_2$ 232.3 73-31-4

Action and use

Treatment of sleep onset insomnia.

Preparation

Melatonin Capsules

DEFINITION

Melatonin is *N*-acetyl-5-methoxytryptamine. It contains not less than 98.0% and not more than 102.0% of $C_{13}H_{16}N_2O_2$ calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white to off-white crystalline powder. It melts at about 117°.

Slightly soluble in water; soluble in acetone, ethyl acetate and methanol.

IDENTIFICATION

The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of melatonin (RS 455).

TESTS

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in a mixture of 20 volumes of acetonitrile and 80 volumes of water.

(1) Dissolve sufficient of the substance being examined to produce a solution containing 0.05% w/v of Melatonin.

(2) 0.0005% w/v of melatonin BPCRS.

(3) Dilute a mixture of 10 volumes of solution (2) and 1 volume of a 0.025% w/v solution of

5-methoxytryptamine BPCRS to 100 volumes.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (5 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (1.8 μm) (Zorbax Eclipse Plus is suitable).

(b) Use gradient elution and the mobile phase described below.

(c) Use a flow rate of 1.5 mL per minute.

(d) Use a column temperature of 25°.

(e) Use a detection wavelength of 225 nm.

(f) Inject 10 μL of each solution.

MOBILE PHASE

Mobile phase A 0.245% w/v of potassium dihydrogen orthophosphate, adjusted to pH 3.0 with 20% v/v acetonitrile.

Mobile phase B acetonitrile.

Ph Eur

1 mL of 0.05 M sulfuric acid is equivalent to 19.52 mg of $C_{13}H_{16}N_2O_2$.

potentiometrically (2.2.20).

0.05 M sulfuric acid, determining the end-point

Dissolve 0.180 g in 30 mL of water R. Titrate with

ASSAY

procedure for the removal of bacterial endotoxins.

parenteral preparations without a further appropriate

Less than 1.5 IU/g, if intended for use in the manufacture of

Bacterial endotoxins (2.6.14)

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105° C for 3 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

lead standard solution (1 ppm Pb) R.

complies with test A. Prepare the reference solution using

acid R. Dilute to 20 mL with water R. 12 mL of the solution

Adjust 10 mL of solution S to pH 3-4 with dilute acetic

Maximum 10 ppm.

Heavy metals (2.4.8)

Wavelength 231.604 nm.

water R.

standard solution (10 ppm Ni) R, diluted as necessary with

Reference solutions Prepare the reference solutions using nickel

with water R.

10.0 mL of lead-free hydrochloric acid R and dilute to 50.0 mL

Test solution Dissolve 5.00 g in 30 mL of water R, add

(ICP-AES) (2.2.57).

Inductively coupled plasma-atomic emission spectrometry

Maximum 5 ppm.

Nickel

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

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Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

Maximum 5 ppm.

When the chromatograms are recorded under the prescribed conditions the retention time of meldonium is about 7.3 minutes and the retention of 5-methoxytryptamine relative to that of meldonium is about 0.29.

Time (minutes)	Mobile phase A (% v/v)	Mobile phase B (% v/v)	Comment
0-20	90-70	10-30	linear gradient
20-36	70-30	30-70	linear gradient
36-41	30-90	70-10	re-equilibration

LIMITS

In the chromatogram obtained with solution (1): the area of any peak due to 5-methoxytryptamine is not greater than the area of the peak due to 5-methoxytryptamine in the chromatogram obtained with solution (3) (0.5%); the area of any other secondary peak is not greater than 0.1 times the area of the principal peak in the chromatogram obtained with solution (2) (0.1%); the sum of the areas of all the secondary peaks is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (1%). Disregard any peak with an area less than 0.5 times the area of the peak due to meldonium in the chromatogram obtained with solution (3) (0.05%).

Heavy metals

1 g complies with limit test D for heavy metals, Appendix VII (20 ppm). Use 2 mL of lead standard solution (10 ppm Pb) to prepare the standard solution.

Water

Not more than 0.3%, Appendix IX C. Use 0.2 g.

Sulfated ash

Not more than 0.1%, Appendix IX A. Use 1 g.

ASSAY

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in the mobile phase.

(1) Dissolve 0.1 g of the substance being examined in 100 mL and dilute 1 volume of the resulting solution to 20 volumes.

(2) 0.005% w/v of meldonium BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (5 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (1.8 µm) (Zorbax Eclipse Plus is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1.5 mL per minute.

(d) Use a column temperature of 25°.

(e) Use a detection wavelength of 225 nm.

(f) Inject 5 µL of each solution.

MOBILE PHASE

20 volumes of acetonitrile and 80 volumes of a 0.245% w/v solution of potassium dihydrogen orthophosphate, the pH of which has been adjusted to 3.0 with 20% v/v orthophosphoric acid.

When the chromatograms are recorded under the prescribed conditions, the retention time of meldonium is about 2.3 minutes.

DETERMINATION OF CONTENT

Calculate the content of $C_{13}H_{16}N_2O_2$ in the substance being examined using the declared content of $C_{13}H_{16}N_2O_2$ in meldonium BPCRS.

STORAGE

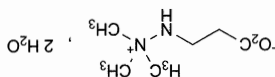
Meldonium should be protected from light and stored at a temperature not exceeding 30°.

IMPURITIES

The impurities limited by the requirements of this monograph include 5-methoxytryptamine.

Meldonium Dihydrate

(Ph. Eur. monograph 2624)



$C_6H_{14}N_2O_2 \cdot 2H_2O$ 182.2 86426-17-7

DEFINITION

3-(2,2,2-Trimethylhydrazin-2-ium-1-yl)propanoate dihydrate. Content 99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white crystals or crystalline powder,

Solubility

Very soluble in water, freely soluble in methanol, practically insoluble in acetone.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24). Comparison meldonium dihydrate CRS.

TESTS

Solution S

Dissolve 10.0 g in 50 mL of distilled water R.

Appearance of solution

Solution S is not more opalescent than reference suspension I (2.2.1) and not more intensely coloured than reference solution B₉ (2.2.2, Method II).

Related substances

Liquid chromatography (2.2.29) coupled with mass spectrometry (2.2.43). Test solution Dissolve 10.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve 10.0 mg of meldonium impurity A CRS, 10.0 mg of meldonium impurity B CRS, 10.0 mg of meldonium impurity C CRS, 10.0 mg of meldonium impurity D CRS, 10.0 mg of meldonium impurity E CRS and 10.0 mg of meldonium impurity F CRS in mobile phase A and dilute to 100.0 mL with mobile phase A. Reference solution (b) Dilute 1.5 mL of reference solution (a) to 100.0 mL with mobile phase A. Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.



— **repeatability**: maximum relative standard deviation of 10 per cent after 6 injections.

— **Calculation of percentage contents**: for impurities A, B, C, D, E and F (MRM mode), use the concentration of the corresponding impurity in reference solution (b); for impurities other than A, B, C, D, E and F (complete spectrum mode), use the concentration of meldonium in reference solution (c).

Limits: impurities A, B, C, D, E, F: for each impurity, maximum 0.15 per cent; unspecified impurities: for each impurity, maximum 0.10 per cent; total: maximum 0.3 per cent; reporting threshold: 0.05 per cent.

Chlorides (2.4.4) Maximum 100 ppm. Dilute 5 mL of solution S to 30 mL with distilled water R.

Sulfates (2.4.13) Maximum 100 ppm. Dilute 10 mL of solution S to 20 mL with distilled water R.

Water (2.5.12) 19.7 per cent to 21.0 per cent, determined on 0.120 g.

Sulfated ash (2.4.14) Maximum 0.1 per cent, determined on 1.0 g.

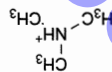
ASSAY Dissolve 0.100 g in 40 mL of glacial acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 14.62 mg of $C_6H_{11}N_2O_2$.

STORAGE

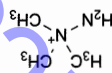
In an airtight container.

IMPURITIES

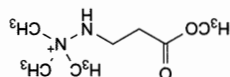
Specified impurities A, B, C, D, E, F



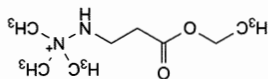
A. N,N-dimethylethanaminium,



B. 1,1,1-trimethylhydrazin-1-ium,



C. 2-(3-methoxy-3-oxopropyl)-1,1,1-trimethylhydrazin-1-ium,



D. 2-(3-ethoxy-3-oxopropyl)-1,1,1-trimethylhydrazin-1-ium,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0-7	90 → 65	10 → 35
7-12	65 → 40	35 → 60
12-18	40 → 25	60 → 75

— **stationary phase**: end-capped octadecylsilyl silica gel for chromatography R (3.5 µm).

— **mobile phase A**: 0.1 per cent V/V solution of heptafluorobutyric acid R in water for chromatography R;

— **mobile phase B**: 0.1 per cent V/V solution of heptafluorobutyric acid R in methanol R;

Substance	MRM channels	Collision energy (eV) for the interval 0-25 min
Meldonium	147.00 → 59.00	18.0
Impurity A	59.97 → 44.98	18.0
Impurity B	74.99 → 59.04	15.0
Impurity F	115.19 → 71.92	19.0
Impurity C	161.19 → 59.00	23.0
Impurity D	175.19 → 58.07	23.0
Impurity E	189.26 → 58.01	22.0

Injection 20 µL of the test solution and reference solutions (b) and (c).

Relative retention With reference to meldonium (retention time = about 9 min): impurity F = about 0.7;

impurity B = about 0.8; impurity A = about 0.9;

impurity C = about 1.5; impurity D = about 1.8;

impurity E = about 2.0.

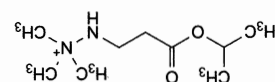
System suitability: reference solution (b):

— **signal-to-noise ratio**: impurity A = minimum 10;

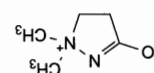
impurity B = minimum 50; impurity C = minimum 200;

impurity D = minimum 1000;

impurity E = minimum 1000; impurity F = minimum 50;



E. 1,1,1-trimethyl-2-[3-(1-methylethoxy)-3-oxopropyl]hydrazin-1-ium,

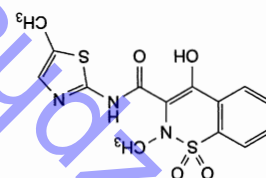


F. 1,1-dimethyl-4,5-dihydro-1H-pyrazol-1-ium-3-olate.

Ph Eur

Meloxicam

(Ph. Eur. monograph 2373)



C₁₄H₁₃N₃O₄S 351.4 7125-38-7

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations

Meloxicam Injection
Meloxicam Oral Suspension
Meloxicam Tablets

Ph Eur

DEFINITION

4-Hydroxy-2-methyl-N-(5-methylthiazol-2-yl)-2H-1,2-benzothiazine-3-carboxamide 1,1-dioxide.

Content

99.0 per cent to 101.0 per cent (dried substance).

Appearance

Pale yellow powder.

Solubility

Practically insoluble in water, soluble in dimethylformamide, very slightly soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison meloxicam CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in acetone, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 40 mg of the substance to be examined in a mixture of 5 mL of methanol R and 0.3 mL of 1 M sodium hydroxide and dilute to 20.0 mL with methanol R.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with methanol R. Dilute 5.0 mL of this solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve 2 mg of the substance to be examined, 2 mg of meloxicam impurity A CRS, 2 mg of meloxicam impurity B CRS, 2 mg of meloxicam impurity C CRS and 2 mg of meloxicam impurity D CRS in a mixture of 5 mL of methanol R and 0.3 mL of 1 M sodium hydroxide and dilute to 25 mL with methanol R. If complete dissolution cannot be achieved, filter the solution through a membrane filter (nominal pore size 0.45 µm) before injection.

Column:

size: l = 0.15 m, Ø = 4.6 mm;

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm);

temperature: 45 °C.

Mobile phase:

mobile phase A: 1 g/L solution of potassium dihydrogen phosphate R adjusted to pH 6.0 with 1 M sodium hydroxide;

mobile phase B: methanol R;

Detection Spectrophotometer at 260 nm and 350 nm. Injection 10 µL.

Relative retention With reference to meloxicam (retention time = about 7 min): impurity B = about 0.5;

impurity A = about 1.4; impurity C = about 1.7;

impurity D = about 1.9.

System suitability: reference solution (b):

— resolution: minimum 3.0 between the peaks due to meloxicam and impurity A at 350 nm; minimum 3.0 between the peaks due to impurity B and meloxicam at 260 nm.

Limits:

— correction factor for the calculation of contents, multiply the peak area of impurity A by 2.0;

— impurity A at 350 nm: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 350 nm (0.1 per cent);

— impurity B at 260 nm: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 350 nm (0.1 per cent);

— impurities C, D at 350 nm: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) at 350 nm (0.05 per cent);

— unspecified impurities: for each impurity, at the wavelength giving the higher value for the impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at the same wavelength (0.10 per cent);

— total: not more than 0.3 per cent;

— disregard limit: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) at the same wavelength (0.03 per cent).

the same wavelength (0.03 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating during the titration, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.250 g in a mixture of 5 mL of anhydrous formic acid R and 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 35.14 mg of $C_{14}H_{13}N_3O_4S_2$.

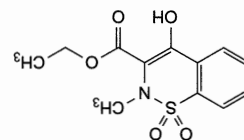
STORAGE

Protected from light.

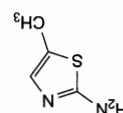
IMPURITIES

Specified impurities A, B, C, D

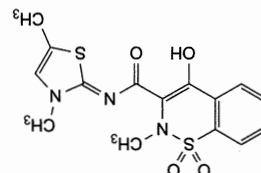
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, F.



A. ethyl 4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxylate 1,1-dioxide,



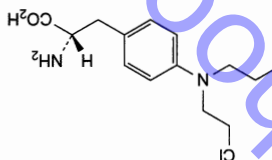
B. 5-methylthiazol-2-amine,



C. N-[(2Z)-3,5-dimethylthiazol-2(3H)-ylidene]-4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxamide 1,1-dioxide,

Melphalan

(Ph Eur monograph 1698)



$C_{13}H_{18}Cl_2N_2O_2$

305.2

148-82-3

Action and use

Cytotoxic alkylating agent.

DEFINITION

4-[Bis(2-chloroethyl)amino]-L-phenylalanine.

Content

94.0 per cent to 102.0 per cent (anhydrous and diethylamine-free substance).

CHARACTERS**Appearance**

White or almost white, hygroscopic powder.

Solubility

Practically insoluble in water, slightly soluble in methanol. It dissolves in dilute mineral acids.

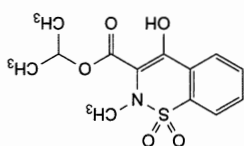
IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

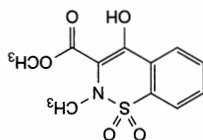
Comparison Ph. Eur. reference spectrum of melphalan.

Ph Eur

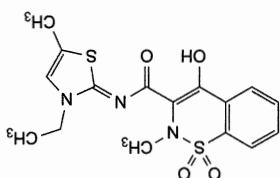
F. isopropyl 4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxylate 1,1-dioxide.



E. methyl 4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxylate 1,1-dioxide,



D. N-[(2Z)-3-ethyl-5-methylthiazol-2(3H)-ylidene]-4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxamide 1,1-dioxide,



TESTS

Appearance of solution

If intended for use in the manufacture of parenteral preparations, the solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.25 g in dilute hydrochloric acid R and dilute to 25 mL with the same acid.

Specific optical rotation (2.2.7)

–36.0 to –30.0 (anhydrous and diethylamine-free substance).

Dissolve 0.175 g at 45 °C for 10 min in methanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Use freshly prepared solutions and protect from light.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in methanol R1 and dilute to 50.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with methanol R1.

Reference solution (a) Dissolve 50.0 mg of mephalan hydrochloride CRS in methanol R1 and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with methanol R1.

Reference solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with methanol R1.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 100.0 mL with methanol R1.

Reference solution (d) Dilute 5.0 mL of reference solution (b) to 100.0 mL with methanol R1.

Reference solution (e) In order to prepare impurity I *in situ*, dissolve 5 mg of mephalan for system suitability CRS (containing impurities B, D, G, H and J) in methanol R1, dilute to 5.0 mL with the same solvent and heat at 60 °C for 15 min.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: mixture of 5 volumes of acetonitrile for chromatography R and 95 volumes of water R containing 0.01 per cent V/V of triethylamine R, 0.05 per cent m/m of ammonium acetate R and 0.05 per cent V/V of glacial acetic acid R;

— mobile phase B: mixture of 40 volumes of water R containing 0.01 per cent V/V of triethylamine R, 0.05 per cent m/m of ammonium acetate R and 0.05 per cent V/V of glacial acetic acid R;

— mobile phase C: mixture of 40 volumes of water R containing 0.01 per cent V/V of triethylamine R, 0.05 per cent m/m of ammonium acetate R and 0.05 per cent V/V of glacial acetic acid R, and 60 volumes of acetonitrile for chromatography R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 20	100 \rightarrow 0	0 \rightarrow 100
20 - 25	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 μ L of test solution (a) and reference solutions (c), (d) and (e).

Identification of impurities Use the chromatogram supplied with mephalan for system suitability CRS and the

chromatogram obtained with reference solution (e) to identify the peaks due to impurities B, D, G, H, I and J.

Relative retention With reference to mephalan (retention time = about 10 min): impurity B = about 0.3; impurity D = about 0.6; impurity I = about 0.8; impurity J = about 1.04; impurity G = about 1.4; impurity H = about 1.5.

System suitability: reference solution (e):

- peak-to-valley ratio: minimum 2.5, where H_p = height above the baseline of the peak due to impurity J and H_b = height above the baseline of the lowest point of the curve separating this peak from the peak due to mephalan.

Limits:

- impurity D: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (d) (3.0 per cent);
- impurity G: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (d) (1.0 per cent);
- impurities J, H: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
- impurity B: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
- impurity I: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
- unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
- total: not more than 11 times the area of the principal peak in the chromatogram obtained with reference solution (d) (5.5 per cent);
- disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Impurity K (diethylamine)

Gas chromatography (2.2.28).

Test solution Dissolve 0.125 g of substance to be examined in 0.15 mL of hydrochloric acid R and dilute to 5.0 mL with dimethyl sulfoxide R.

Reference solution Dilute 1 mL of methanol R and 0.125 g of diethylamine R1 (impurity K) to 10.0 mL with dimethyl sulfoxide R. Dilute 1.0 mL of the solution to 100.0 mL with dimethyl sulfoxide R.

Column:

— material: glass;

— size: $l = 1.6$ m, $\phi = 4$ mm;

— stationary phase: styrene-divinylbenzene copolymer R coated with potassium carbonate (149–177 μ m).

Carrier gas nitrogen for chromatography R.

Flow rate 42.5 mL/min.

Temperature:

— column: 170 °C;

— injection port: 190 °C;

— detector: 250 °C.

Detection Flame ionisation.

Injection 1 μ L.

Elution order Methanol, impurity K.

System suitability: reference solution:

— resolution: minimum 2.0 between the peaks due to methanol and impurity K.

Limit:
— impurity K: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.5 per cent).

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection: Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{13}H_{18}Cl_2N_2O_2$ taking into account the assigned content of mephalan

hydrochloride CRS and a conversion factor of 0.8933.

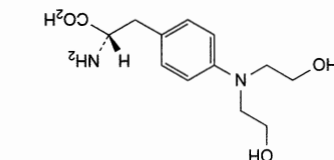
STORAGE

In an airtight container, protected from light.

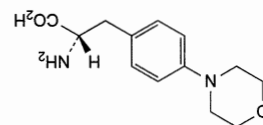
IMPURITIES

Specified impurities B, D, G, H, I, J, K

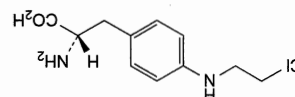
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, E, F.



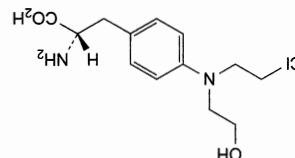
A. 4-[[bis(2-hydroxyethyl)amino]-1-phenyl]alanine,



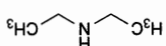
B. 4-morpholin-4-yl-1-phenylalanine,



C. 4-[(2-chloroethyl)amino]-1-phenylalanine,

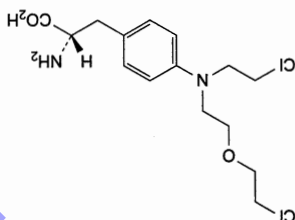


D. 4-[(2-chloroethyl)(2-hydroxyethyl)amino]-1-phenylalanine,

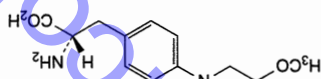


K. N-ethylethanamine (diethylamine).

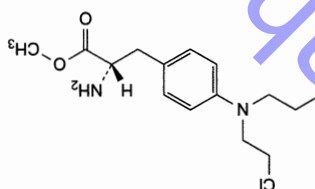
J. 4-[[2-(2-chloroethoxy)ethyl](2-chloroethyl)amino]-1-phenylalanine,



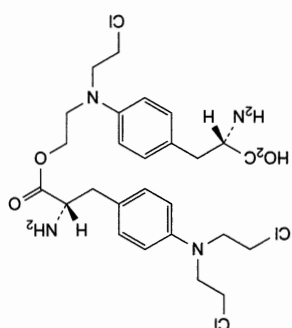
I. 4-[[2-(2-chloroethyl)(2-methoxyethyl)amino]-1-phenyl]alanine,



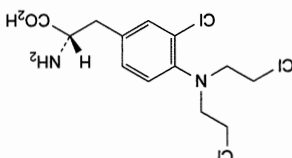
H. methyl 4-[[bis(2-chloroethyl)amino]-1-phenyl]alaninate,



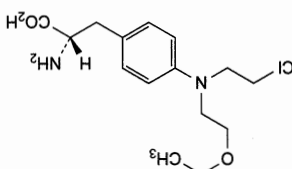
G. 4-[[2-[[4-[[bis(2-chloroethyl)amino]-1-phenyl]oxy]ethyl](2-chloroethyl)amino]-1-phenyl]alanine (mephalan dimer),



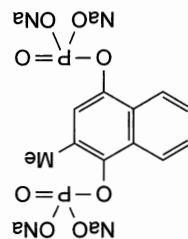
F. 4-[[bis(2-chloroethyl)amino]-3-chloro-1-phenyl]alanine (3-chloromephalan),



E. 4-[(2-chloroethyl)(2-ethoxyethyl)amino]-1-phenylalanine,



Menadiol Sodium Phosphate



$C_{11}H_8Na_4O_8P_2$ 530.2 6700-42-1

Action and use

Vitamin K analogue.

Preparation

Menadiol Phosphate Tablets

DEFINITION

Menadiol Sodium Phosphate is tetrasodium 2-methylnaphthalene-1,4-diyli di(orthophosphate) hexahydrate. It contains not less than 98.0% and not more than 100.5% of $C_{11}H_8Na_4O_8P_2$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white to pink, crystalline powder; hygroscopic. Very soluble in water; practically insoluble in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of menadiol sodium phosphate (RS 213).

B. To 10 mL of a 2% w/v solution add 10 mL of 1M sulfuric acid, 10 mL of 0.1M cerium(IV) sulfate and 1 mL of hydrogen peroxide solution (20 vol) and extract with two 10 mL quantities of chloroform. Evaporate the combined chloroform extracts to dryness on a water bath and dry the residue at 40° at a pressure not exceeding 0.7 kPa. The infrared absorption spectrum of the residue, Appendix II A, is concordant with the reference spectrum of menadiol sodium phosphate.

C. Dissolve 40 mg in 2 mL of water, heat gently with 2 mL of sulfuric acid until white fumes are evolved, add nitric acid dropwise until digestion is complete and cool. Add 2 mL of water, heat until white fumes are evolved again, cool, add a further 10 mL of water and neutralise to litmus paper with 5M ammonia. The solution yields reaction A characteristic of sodium salts and reaction B characteristic of phosphates, Appendix VI.

TESTS

Inorganic phosphate
Dissolve 25 mg in 10 mL of water, add 4 mL of 1M sulfuric acid, 1 mL of a 10% w/v solution of ammonium molybdate and 2 mL of methylaminophenyl-sulfite reagent and allow to stand for 15 minutes. The absorbance of the solution at 730 nm, Appendix II B, is not more than the absorbance of a 0.0025% w/v solution of potassium dihydrogen orthophosphate in place of the solution of the substance being examined (0.7%).

Total phosphate

Dissolve 85 mg in 50 mL of glacial acetic acid, add 5 mL of mercuric(ii) acetate solution and carry out Method I for non-aqueous titration, Appendix VIII A, determining the end point

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, in subdued light using the following solutions in methanol (50%).

- (1) 4.0% w/v of the substance being examined.
- (2) 0.020% w/v of the substance being examined.
- (3) 0.0080% w/v of 2-methyl-1,4-naphthoquinone.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel GF₂₅₄.

(b) Use the mobile phase as described below.

(c) Apply 5 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, examine under ultraviolet light (254 nm).

MOBILE PHASE

1.5 volumes of diethylamine, 5 volumes of butan-1-ol, 50 volumes of propan-1-ol and 50 volumes of a 2% w/v solution of ammonium chloride.

LIMITS

Any spot corresponding to 2-methyl-1,4-naphthoquinone (menadiol) in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (3) and any other secondary spot is not more intense than the spot in the chromatogram obtained with solution (2).

Water

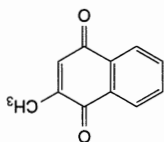
19.0 to 21.5% w/w, Appendix IX A. Use 0.25 g.

ASSAY

Dissolve 0.1 g in 25 mL of water, add 25 mL of glacial acetic acid and 25 mL of 3M hydrochloric acid and titrate with 0.02M cerium(IV) sulfate VS using platinum and calomel electrodes and determining the end point potentiometrically. Each mL of 0.02M cerium(IV) sulfate VS is equivalent to 4.221 mg of $C_{11}H_8Na_4O_8P_2$.

Menadione

(Ph. Eur. monograph 0507)



$C_{11}H_8O_2$

172.2

58-27-5

Action and use

Vitamin K analogue.

DEFINITION

Menadione contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of 2-methylnaphthalene-1,4-dione, calculated with reference to the dried substance.

Menotrophin

9002-68-0

Action and use

Gonadotrophin; treatment of infertility.

Preparation

Menotrophin Injection

DEFINITION

Menotrophin is a dry preparation containing glycoprotein gonadotrophins possessing follicle-stimulating and luteinising activities. The follicle-stimulating activity is of pituitary origin. The luteinising activity is predominantly of placental origin, but may also contain luteinising activity of placental origin. Menotrophin is obtained from the urine of post-menopausal women but, where necessary, chorionic gonadotrophin obtained from the urine of pregnant women may be added to achieve the ratio stated below.

Potency

It contains not less than 40 IU of follicle-stimulating hormone activity per mg. The ratio of IU of luteinising hormone (LH) activity to IU of follicle-stimulating hormone (FSH) activity is approximately 1. For each component the estimated potency is not less than 80% and not more than 125% of the stated potency.

PRODUCTION

Menotrophin may be prepared by a suitable fractionation procedure followed by ion-exchange chromatography. It is prepared in conditions designed to minimise microbial contamination and to be in compliance with the requirements of Appendix XXII A: Viral Safety.

CHARACTERISTICS

An almost white or slightly yellow powder.
Soluble in water.

IDENTIFICATION

Causes enlargement of the ovaries of immature female rats and increases the weight of the seminal vesicles and prostate gland of immature male rats when administered as directed in the Assay.

TESTS

Hepatitis antigens
Examined by a suitably sensitive immunochemical method, Appendix XIV B, hepatitis antigens are not detected.

HIV antigens

Examined by a suitably sensitive immunochemical method, Appendix XIV B, HIV antigens are not detected.

Bacterial endotoxins

Carry out the test for bacterial endotoxins, Appendix XIV C. The endotoxin limit is not more than 0.78 IU per IU of follicle stimulating hormone.

Water

Not more than 5.0% w/w, Appendix IX C, Method III.

ASSAY

Carry out the biological assay of menotrophin described below. The fiducial limits of error are not less than 64% and not more than 156% of the stated potency.

STORAGE

Menotrophin should be kept in an airtight, tamper-evident container and protected from light.

LABELLING

The label states (1) the number of IU (Units) of follicle-stimulating hormone activity and the number of IU (Units)

CHARACTERS

A pale-yellow, crystalline powder, practically insoluble in water, freely soluble in toluene, sparingly soluble in alcohol and in methanol. It is unstable in light.

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Melting point (2.2.14): 105 °C to 108 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with menadione CRS.

C. Dissolve about 1 mg in 5 mL of alcohol R, add 2 mL of ammonia R and 0.2 mL of ethyl cyanoacetate R. An intense bluish-violet colour develops. Add 2 mL of hydrochloric acid R. The colour disappears.

D. Dissolve about 10 mg in 1 mL of alcohol R, add 1 mL of hydrochloric acid R and heat in a water-bath. A red colour develops.

TESTS

Related substances

Carry out the test protected from bright light. Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.2 g of the substance to be examined in acetone R and dilute to 10 mL with the same solvent. Reference solution Dilute 0.5 mL of the test solution to 100 mL with acetone R.

Apply separately to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 1 volume of nitromethane R, 2 volumes of acetone R, 5 volumes of ethylene chloride R and 90 volumes of cyclohexane R. Dry the plate in a current of hot air. Repeat the development and drying a further two times. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying over diphosphorus pentoxide R at a pressure of 2 kPa to 3 kPa for 4 h.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 15 mL of glacial acetic acid R in a flask with a stopper fitted with a valve. Add 15 mL of dilute hydrochloric acid R and 1 g of zinc powder R. Close the flask. Allow the mixture to stand for 60 min, protected from light, with occasional shaking. Filter the solution over a cotton wad, wash with three quantities, each of 10 mL, of carbon dioxide-free water R. Add 0.1 mL of ferrous R and immediately titrate the combined filtrate and washings with 0.1 M ammonium and cerium nitrate.

STORAGE

Store protected from light.

Ph Eur

of luteinising hormone activity in the container; (2) the number of IU (Units) of follicle-stimulating hormone per mg and the number of IU (Units) of luteinising hormone activity per mg; (3) where applicable, the number of IU (Units) of chorionic gonadotrophin activity per mg; (4) the date after which the material is not intended to be used; (5) the conditions under which it should be stored; (6) where applicable, that it is sterile.

Sterility

Complies with the test for sterility, Appendix XVI A.

BIOLOGICAL ASSAY OF MENOTROPHIN

The potency of menotropin with respect to its follicle-

Standard Preparation of human urinary FSH and human urinary LH under the conditions of a suitable method of assaying. The potency of menotrophin with respect to its luteinising hormone activity is estimated by comparing its effect in increasing the weight of the seminal vesicles or the prostate gland of immature male rats with that of the Standard Preparation of human urinary FSH and human urinary LH under the conditions of a suitable method of

Standard Preparation

The Standard Preparation is the International Standard preparation of human urinary follicle-stimulating hormone and luteinising hormone, consisting of a freeze-dried extract of urine from post-menopausal women together with lactose, or another suitable preparation the potency of which has been determined in relation to the International Standard.

SUGGESTED METHOD

Select female rats of the same strain, 19 to 28 days old and differing in age by not more than 3 days, and having weights such that the difference between the heaviest rat and the lightest rat is not more than 10 g. Assign the rats at random to six equal groups of at least five animals. If sets of six litter

Choose three doses of the Standard Preparation and three doses of the preparation being examined such that the smallest dose produces a positive response in some of the rats and the largest dose does not produce a maximum response in all of the rats. Use doses in geometric progression. As an initial approximation, total doses of 1.5, 3.0 and 6.0 IU may be tried although the dose will depend on the sensitivity of the animals used, which may vary widely.

Dissolve separately the total quantities of the preparation being examined, and of the Standard Preparation, corresponding to the daily doses to be used in sufficient *aluminum-phosphate buffer pH 7.2* containing not less than 70 IU of *Chromococcus gonadotrophum* per mL so that the daily dose is about 0.2 mL. Add a suitable antimicrobial preservative such as 0.4% w/v of *phenol* or 0.002% w/v of *thimerosal*. Store the solutions at a temperature of 2° to 8°. Inject subcutaneously into each rat the daily dose allocated to its group. Repeat the procedure after 24 and 48 hours. About 24 hours after the last injection, kill the rats and remove the ovaries. Remove any extraneous fluid and tissue and immediately weigh the ovaries from each animal. Record the combined weight of both ovaries from each rat. Calculate the

result of the assay by standard statistical methods using the weight of the ovaries as the response. (The precision of the assay may be improved by a suitable correction of the organ weight with reference to the weight of the animal from which it was taken; an analysis of covariance may be used).

Luteinizing hormone activity

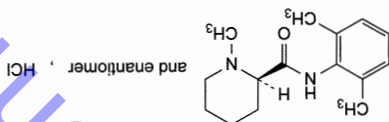
Select male rats of the same strain, approximately 19 to 28 days old and differing in age by not more than 3 days, and having weights such that the difference between the heaviest rat and the lightest rat is not more than 10 g. Assign the rats at random to six equal groups of at least five animals. If sets of six litter mates are available, allot one litter mate from each set to each group and mark according to

Choose three doses of the Standard Preparation and three doses of the preparation being examined such that the smallest dose is sufficient to produce a positive response in some of the rats and the largest dose does not produce a maximum response in all of the rats. Use doses in geometric progression. As an initial approximation, total doses of 7, 14 and 28 IU may be tried although the dose will depend on the sensitivity of the animals used, which may vary widely. Dissolve separately the total quantities of the preparation being examined, and of the Standard Preparation, corresponding to the daily doses to be used in sufficient amounts, dissolve in 10 ml of water and the daily dose is about 2.3 mg.

0.2 mL. Add a suitable antimicrobial preservative such as 0.4% w/v of phenol or 0.002% w/v of thimerosal. Store the solutions at a temperature of 2° to 8°. Inject subcutaneously into each rat the daily dose allocated to its group on 4 consecutive days at the same time each day. On the fifth day, about 24 hours after the last injection, kill the rats and remove the seminal vesicles or the prostate gland. Remove any extraneous fluid and tissue and weigh immediately the seminal vesicles or the prostate gland. Calculate the result of the assay by standard statistical methods, using the weight of the vesicles or the prostate gland as the response. (The precision of the assay may be improved by a suitable correction of the organ weight with reference to the weight of the animal from which it was taken; an analysis of covariance may be used.)

Mepivacaine Hydrochloride

(Ph. Eur. monograph 1242)


$$\text{C}_{15}\text{H}_{23}\text{ClN}_2\text{O}$$

Action and use

Local anesthetic.

DEFINITION

(RS)-N-(2,6-Dimethylphenyl)-1-methylpiperidine-2-carboxamide hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), very slightly soluble in methylene chloride.

mp

About 260 °C, with decomposition.

IDENTIFICATION

First identification A, B, D.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison mepivacaine hydrochloride CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of mepivacaine hydrochloride CRS in ethanol (96 per cent) R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of mepivacaine hydrochloride CRS and 20 mg of lidocaine hydrochloride CRS in ethanol (96 per cent) R and dilute to 5 mL with the same solvent.

Development Over a path of 12 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 5 mL of solution S (see Tests) add 1 mL of dilute sodium hydroxide solution R and shake with 2 quantities, each of 10 mL, of ether R. Dry the combined upper layers over anhydrous sodium sulfate R. Filter and evaporate the ether on a water-bath. Dry the residue at 100–105 °C for 2 h.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 1.5 g in carbon dioxide-free water R and dilute to 30 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

pH (2.2.3)

Dilute 2 mL of solution S to 5 mL with carbon dioxide-free water R.

Optical rotation (2.2.7)

–0.10° to +0.10°, determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of the substance to be examined and 30.0 mg of mepivacaine impurity B CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (e) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (f) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (g) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (h) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (i) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (j) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (k) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (l) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (m) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (n) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (o) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (p) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (q) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (r) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (s) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (t) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (u) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (v) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (w) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (x) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (y) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (z) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (aa) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ab) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ac) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ad) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ae) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (af) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ag) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ah) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ai) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (aj) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ak) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (al) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (am) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (an) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ao) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ap) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (aq) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ar) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (as) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

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Reference solution (au) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (av) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (aw) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ax) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ay) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (az) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ba) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (bb) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

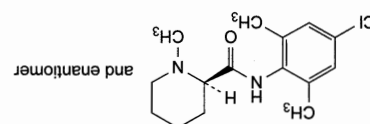
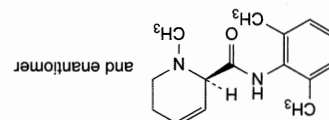
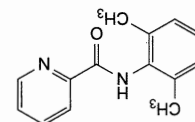
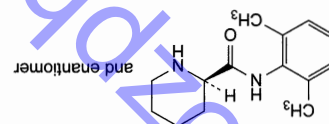
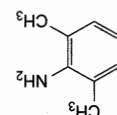
Reference solution (bc) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.
1 mL of 0.1 M sodium hydroxide is equivalent to 28.28 mg of $C_{15}H_{23}ClN_2O_4$.

IMPURITIES

Specified impurities A, B, C, D, E.



Ph Eur

Action and use

Hypnotic.

Ph Eur

DEFINITION

Meprobamate contains not less than 97.0 per cent and not more than the equivalent of 101.0 per cent of 2-methyl-2-propylpropane-1,3-diyldicarbamate, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, amorphous or crystalline powder, slightly soluble in water, freely soluble in alcohol.

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Melting point (2.2.14): 104 °C to 108 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with meprobamate CRS.

C. To 0.5 g add 1 mL of acetic anhydride R and 0.05 mL of sulfuric acid R, mix and allow to stand for 30 min, shaking frequently. Pour the solution dropwise into 50 mL of water R, mix and allow to stand. Initiate crystallisation by scratching the wall of the tube with a glass rod. Collect the precipitate by filtration, wash and dry at 60 °C.

The precipitate melts (2.2.14) at 124 °C to 128 °C.

D. Dissolve 0.2 g in 15 mL of 0.5 M alcoholic potassium hydroxide and boil under a reflux condenser for 15 min.

Add 0.5 mL of glacial acetic acid R and 1 mL of a 50 g/L solution of cobalt nitrate R in ethanol R. A deep-blue colour develops.

TESTS

Appearance of solution

Dissolve 1.0 g in 20 mL of ethanol R. The solution is clear (2.2.1) and colourless (2.2.2, Method II).

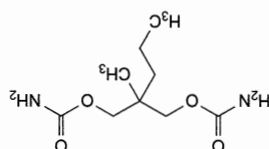
Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel G R as the coating substance.

Test solution Dissolve 0.20 g of the substance to be examined in alcohol R and dilute to 10 mL with the same solvent. Reference solution Dilute 0.1 mL of the test solution to 10 mL with alcohol R.

Apply separately to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of pyridine R, 30 volumes of acetone R and 70 volumes of hexane R. Dry the plate at 120 °C for 30 min, allow to cool and spray with a solution of 0.25 g of vanillin R in a cooled mixture of 10 mL of alcohol R and 40 mL of sulfuric acid R and heat at 100 °C to 105 °C for 30 min. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the

(Ph. Eur. monograph 0407)



$C_9H_{18}N_2O_4$

218.3

57-53-4



chromatogram obtained with the reference solution

(1.0 per cent).

Heavy metals (2.4.8)

Dissolve 2.0 g in a mixture of 15 volumes of water R and 85 volumes of acetone R and dilute to 20 mL with the same mixture of solvents. 12 mL of the solution complies with test B for heavy metals (10 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with the mixture of water R and acetone R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

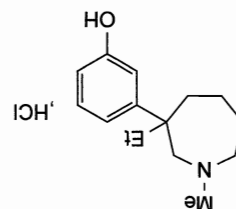
Dissolve 0.1000 g in 15 mL of a 25 per cent V/V solution of sulfuric acid R and boil under a reflux condenser for 3 h.

Cool, dissolve by cautiously adding 30 mL of water R, cool again and place in a steam-distillation apparatus. Add 40 mL of strong sodium hydroxide solution R and distil immediately by passing steam through the mixture. Collect the distillate into 40 mL of a 40 g/L solution of boric acid R until the total volume in the receiver reaches about 200 mL.

Add 0.25 mL of methyl red mixed solution R. Titrate with 0.1 M hydrochloric acid until the colour changes from green to violet. Carry out a blank titration.

1 mL of 0.1 M hydrochloric acid is equivalent to 10.91 mg of $C_{15}H_{23}NO_4$.

Meptazinol Hydrochloride



$C_{15}H_{23}NO_4 \cdot HCl$

269.8

59263-76-2

Action and use

Opioid receptor partial agonist; analgesic.

Preparations

Meptazinol Injection

Meptazinol Tablets

DEFINITION

Meptazinol Hydrochloride is 3-(3-ethyl-1-methylpiperidino-3-yl)phenol hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{15}H_{23}NO_4 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder.
Very soluble in water and in methanol; freely soluble in ethanol (96%); very slightly soluble in acetone. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.01% w/v solution in ethanol (96%) exhibits a maximum only at 275 nm. The absorbance at 275 nm is about 0.80.
B. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of meptazinol hydrochloride (RS 215).

TESTS

Acidity or alkalinity

To 10 mL of a 2% w/v solution in carbon dioxide-free water add 0.2 mL of 0.01M sodium hydroxide VS and 0.1 mL of methyl red solution; the solution is yellow. Add 0.4 mL of 0.01M hydrochloric acid VS; the solution is red.

Colour of solution

A 10.0% w/v solution is not more intensely coloured than reference solution Y, Appendix IV B, Method II.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in methanol.

(1) 1.0% w/v of the substance being examined.

(2) 0.01% w/v of the substance being examined.

(3) 0.005% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel (Merck silica gel 60 F₂₅₄ plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, examine under ultraviolet light (254 nm), expose to iodine vapour for 2 hours and examine again.

MOBILE PHASE

1 volume of 18M ammonia, 30 volumes of chloroform and 70 volumes of ethyl acetate.

LIMITS

By each method of visualisation, any secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (1%) and not more than one such spot is more intense than the spot in the chromatogram obtained with solution (3) (0.5%).

Loss on drying

When dried at 105° for 3 hours, loses not more than 0.5% of its weight. Use 1 g.

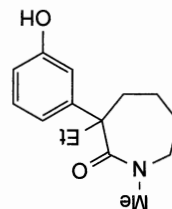
Sulfated ash

Not more than 0.1%, Appendix IX A.

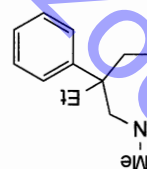
ASSAY

Dissolve 0.15 g in 50 mL of anhydrous acetic acid and add 5 mL of mercury(II) acetate solution. Carry out Method I for non-aqueous titration, Appendix VIII A, determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 26.98 mg of $C_{15}H_{23}NO_4 \cdot HCl$.

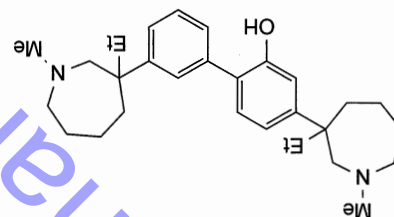
IMPURITIES



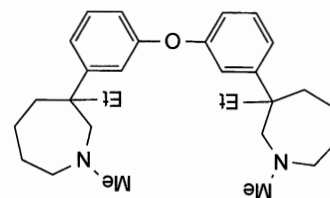
A. 3-ethyl-3-(3-hydroxyphenyl)-1-methylperhydroazepin-2-one



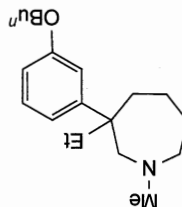
B. 3-ethyl-1-methyl-3-phenylperhydroazepine



C. 4-(3-ethyl-1-methylperhydroazepin-3-yl)-2-[3-(3-ethyl-1-methylperhydroazepin-3-yl)phenyl]phenol



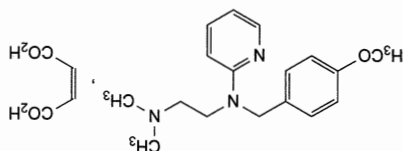
D. 3,3'-oxybis(3-ethyl-1-methylperhydroazepin-3-yl)benzene



E. 3-ethyl-3-(3-butoxyphenyl)-1-methylperhydroazepine

Mepyramine Maleate

(Ph. Eur. monograph 0278)


 $C_{21}H_{27}N_3O_5$ 401.5 59-33-6

Action and use

Anti-emetic.

Preparation

Mepyramine Tablets

Ph Eur

DEFINITION

N-(4-Methoxybenzyl)-*N,N'*-dimethyl-*N*-(pyridin-2-yl)ethane-1,2-diamine (*Z*)-butenedioate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B.

Second identification A, C, D, E.

A. Melting point (2.2.14): 99 °C to 103 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation 50 g/L solutions in methylene chloride *R* using a

0.1 mm cell.

C. Dissolve 0.100 g in 0.01 *M* hydrochloric acid and dilute to

100.0 mL with the same acid. Dilute 1.0 mL of this solution

to 100.0 mL with 0.01 *M* hydrochloric acid. Examined

between 220 nm and 350 nm (2.2.25), the solution shows

2 absorption maxima, at 239 nm and 316 nm. The specific

absorbances at the absorption maxima are 431 to 477 and

196 to 220, respectively.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 40 mg of the substance to be examined

in methylene chloride *R* and dilute to 10 mL with the same

solvent.

Reference solution Dissolve 40 mg of mepyramine maleate CRS

in methylene chloride *R* and dilute to 10 mL with the same

solvent.

Plate TLC silica gel GF₂₅₄ plate *R*.

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.



— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard the peak due to maleic acid.

Chlorides (2.4.4)

Maximum 100 ppm.

Dilute 2.5 mL of solution S to 15 mL with *water R*.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution

using 2 mL of *lead standard solution* (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.25 per cent, determined on 1.000 g by drying in

an oven at 80 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on the residue obtained

in the test for loss on drying.

ASSAY

Dissolve 0.150 g in 40 mL of *anhydrous acetic acid R*. Titrate

with 0.1 M *perchloric acid*, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M *perchloric acid* is equivalent to 20.07 mg

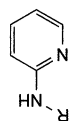
of $C_{21}H_{27}N_3O_5$.

STORAGE

Protected from light.

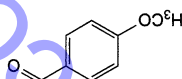
IMPURITIES

Specified impurities A, B, C



A, R = $CH_2-C_6H_4-p-OCH_3$; N-(4-methoxybenzyl)pyridin-2-

amine, C, R = H; pyridin-2-amine,



B, 4-methoxybenzaldehyde (anisaldehyde).

E. Triturate 0.1 g with 3 mL of *water R* and 1 mL of *strong sodium hydroxide solution R*. Shake with 3 quantities, each of 5 mL, of *ether R*. To 0.1 mL of the aqueous layer add a solution of 10 mg of *resorcinol R* in 3 mL of *sulfuric acid R*. Heat on a water-bath for 15 min; no colour develops. To the rest of the aqueous layer add 1 mL of *bromine water R*. Heat on a water-bath for 15 min and then heat to boiling and cool. To 0.2 mL of this solution add a solution of 10 mg of *resorcinol R* in 3 mL of *sulfuric acid R*. Heat on a water-bath for 15 min; a violet-pink colour develops.

TESTS

Solution S

Dissolve 5.0 g in *carbon dioxide-free water R* and dilute to

25 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured

than reference solution Y₆ (2.2.2, Method II).

Dilute 5 mL of solution S to 25 mL with *carbon dioxide-free*

water R.

pH (2.2.3)

4.9 to 5.2.

Dilute 1.0 mL of solution S to 10 mL with *carbon dioxide-free*

water R.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be

examined in the mobile phase and dilute to 50.0 mL with

the mobile phase.

Reference solution (a) Dissolve 5 mg of *anisaldehyde R*

(impurity B), 5.0 mg of *mepyramine impurity A CRS* and

5.0 mg of *mepyramine impurity C CRS* in the mobile phase

and dilute to 100.0 mL with the mobile phase. Dilute

2.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: *phenylsilyl silica gel for chromatography R1*

(5 μ m).

Mobile phase Mix 0.1 volume of *triethylamine R*, 40 volumes

of a 0.771 g/L solution of *ammonium acetate R* and

60 volumes of *methanol R*.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time 3 times the retention time of *mepyramine*.

Relative retention With reference to *mepyramine* (retention

time = about 13 min): maleic acid = about 0.2;

impurity C = about 0.3; impurity B = about 0.4;

impurity A = about 0.5.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to

impurities C and B.

Limits:

— impurities A, C: for each impurity, not more than the area

of the corresponding peak in the chromatogram obtained

with reference solution (a) (0.2 per cent);

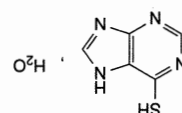
— impurity B: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

Mercaptopurine

(Ph. Eur. monograph 0096)

 $C_5H_4N_4S$, H_2O

170.2

6112-76-1

Action and use

Thiopurine cytotoxic.

Preparations

Mercaptopurine Oral Suspension

Mercaptopurine Tablets

DEFINITION

7H-Purine-6-thiol monohydrate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent). It dissolves in solutions of alkali hydroxides.

IDENTIFICATION

A. Dissolve 20 mg in 5 mL of dimethyl sulfoxide R and dilute

solution to 200 mL with 0.1 M hydrochloric acid. Examined

between 230 nm and 350 nm (2.2.25), the solution shows

only 1 absorption maximum, at 325 nm.

B. Dissolve about 20 mg in 20 mL of ethanol (96 per cent) R

heated to 60 °C and add 1 mL of a saturated solution of

mercuric acetate R in ethanol (96 per cent) R. A white

precipitate is formed.

C. Dissolve about 20 mg in 20 mL of ethanol (96 per cent) R

heated to 60 °C and add 1 mL of a 10 g/L solution of lead

acetate R in ethanol (96 per cent) R. A yellow precipitate is

TESTS

Impurity A

Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined

in 1 mL of dimethyl sulfoxide R and dilute to 10 mL with

methanol R.

Reference solution Dissolve 10 mg of hypoxanthine R in 10 mL

of dimethyl sulfoxide R and dilute to 100 mL with methanol R.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase concentrated ammonia R, water R, acetone R

Application 5 µL.

Development Over a path of 10 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Limit:

— impurity A: any spot corresponding to hypoxanthine in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with the reference solution (2.0 per cent).

Mercuric Chloride

(Ph. Eur. monograph 0120)

 $HgCl_2$

271.5

7487-94-7



DEFINITION

Content

99.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless or white or almost white crystals or heavy crystalline masses.

Solubility

Soluble in water and in glycerol, freely soluble in ethanol

(96 per cent).

IDENTIFICATION

A. It gives the reactions of chlorides (2.3.1).

B. Solution S (see Tests) gives the reactions of mercury

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to

20 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference

suspension II (2.2.1) and is colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of methyl red solution R.

The solution is red. Add 0.5 g of sodium chloride R.

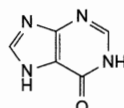
The solution becomes yellow. Not more than 0.5 mL of

0.01 M hydrochloric acid is required to change the colour to

Mercurous chloride

Dissolve 1.0 g in 30 mL of ether R. The solution shows no

opalescence.



A. 1,7-dihydro-6H-purin-6-one (hypoxanthine).

Ph Eur

IMPURITIES

Protected from light.

STORAGE

15.22 mg of $C_5H_4N_4S$.

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to

end-point potentiometrically (2.2.20).

Dissolve 0.100 g in 50 mL of dimethylformamide R. Titrate

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

10.0 per cent to 12.0 per cent, determined on 0.250 g.

Water (2.5.12)

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 2.00 g by drying *in vacuo* for 24 h.

ASSAY

Dissolve 0.500 g in 100 mL of water R. Add 20.0 mL of

0.1 M sodium edetate and 5 mL of buffer solution pH 10.9 R.

Allow to stand for 15 min. Add 0.1 g of mordan black 11

triurate R and titrate with 0.1 M zinc sulfate until the colour

changes to purple. Add 3 g of potassium iodide R, allow to

stand for 2 min, add a further 0.1 g of mordan black 11

triurate R and titrate with 0.1 M zinc sulfate. The second titration is

equivalent to 27.15 mg of HgCl₂.

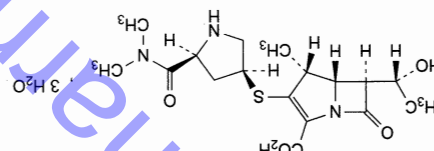
STORAGE

Protected from light.

Ph Eur

Meropenem Trihydrate

(Ph. Eur. monograph 2234)

C₁₇H₂₅N₃O₅·3H₂O

437.5

119478-56-7

Action and use
Carbapenem antibacterial.

Ph Eur

DEFINITION

(4R,5S,6S)-3-[[[(3S,5S)-5-[(D)dimethylamino]carbonyl]pyrrolidin-3-yl]sulfonyl]-6-[[[(1R)-1-hydroxyethyl]-4-methyl-oxo-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid trihydrate.

Content
Semi-synthetic product derived from a fermentation product. 97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or light yellow, crystalline powder.

Solubility

Sparsely soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison meropenem trihydrate CRS.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Dissolve 1.0 g in 20 mL of a 50 g/L solution of sodium hydrogen carbonate R.

pH (2.2.3)

4.0 to 6.0.

Dissolve 0.20 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Specific optical rotation (2.2.7)
-17 to -21 (anhydrous substance).
Dissolve 0.125 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare test solutions (a) and (b) and reference solution (c) immediately before use. Prepare and store reference solution (a) at 4 °C and use within 6 h.

Solvent mixture To 1.0 mL of triethylamine R add 900 mL of water for chromatography R. Adjust to pH 5.0 with dilute phosphoric acid R and dilute to 1000.0 mL with water for chromatography R.

Test solution (a) Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) In order to prepare impurities A and B *in-situ*, heat 10 mL of test solution (a) to 60 °C for about 20 min or, alternatively, allow 10 mL of test solution (a) to stand at ambient temperature for about 8 h.

Reference solution (c) Dissolve 50.0 mg of meropenem trihydrate CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:

— size: 1 = 0.25 m, Ø = 4.6 mm;

— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 µm);

— temperature: 40 °C.

Mobile phase acetonitrile R1, solvent mixture (7:100 V/V).

Flow rate 1.6 mL/min.

Detection Spectrophotometer at 220 nm. Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Run time 4 times the retention time of meropenem. Relative retention With reference to meropenem (retention time = about 6 min): impurity A = about 0.5; impurity B = about 2.2.

System suitability: reference solution (b):
— resolution: minimum 5.0 between the peaks due to impurity A and meropenem.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity A by 1.6;

— impurity A: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

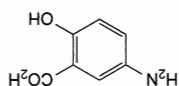
— impurity B: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— sum of impurities other than A and B: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

Mesalazine

(Ph. Eur. monograph 1699)



$C_7H_7NO_3$ 153.1

89-57-6

Action and use

Aminosalicylate; treatment of ulcerative colitis.

Preparations

Mesalazine Enema
Mesalazine Foam Enema
Prolonged-release Mesalazine Granules
Mesalazine Suppositories
Gastro-resistant Mesalazine Tablets
Prolonged-release Mesalazine Tablets

DEFINITION

5-Amino-2-hydroxybenzoic acid.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

Almost white or light grey or light pink powder or crystals.

Solubility

Very slightly soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides and in dilute hydrochloric acid.

IDENTIFICATION

First identification B.

Second identification A, C.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 50.0 mg in 10 mL of a 10.3 g/L solution of hydrochloric acid R and dilute to 100.0 mL with the same acid. Dilute 5.0 mL of this solution to 200.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Absorption maximum At about 230 nm.

Specific absorbance at the absorption maximum 430 to 450.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison mesalazine CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of a mixture of equal volumes of glacial acetic acid R and water R and dilute to 10.0 mL with methanol R.

Reference solution Dissolve 25 mg of mesalazine CRS in 5 mL of a mixture of equal volumes of glacial acetic acid R and water R and dilute to 10.0 mL with methanol R.

Plate A suitable silica gel as the coating substance.

Mobile phase glacial acetic acid R, methanol R, methyl isobutyl ketone R (10:40:50 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 365 nm.

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

0.50 g complies with test G. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

11.4 per cent to 13.4 per cent, determined on 0.100 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 0.125 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{17}H_{15}NO_5$ from the declared content of mesopren trihydrate CRS.

STORAGE

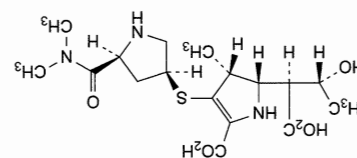
If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

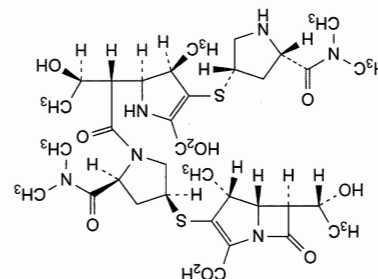
The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES

Specified impurities A, B



A. (4R,5S)-5-[(1S,2R)-1-carboxy-2-hydroxypropyl]-3-[[[(3S,5S)-5-[(dimethylamino)carbonyl]pyrrolidin-3-yl]-4-methyl-4,5-dihydro-1H-pyrrole-2-carboxylic acid, azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid.



B. (4R,5S,6S)-3-[[[(3S,5S)-1-[(2S,3R)-2-[(2S,3R)-5-carboxysulfany]-3-methyl-2,3-dihydro-1H-pyrrol-2-yl]-3-sulfany]-5-[(dimethylamino)carbonyl]pyrrolidin-3-yl]-4-methyl-4,5-dihydro-1H-pyrrole-2-carboxylic acid, azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

Maintain the solutions at 40 °C during preparation and measurements. Dissolve 0.5 g in 1 M hydrochloric acid and dilute to 20 mL with the same acid. The solution is clear (2.2.1). Immediately measure the absorbance (2.2.25) of the solution at 440 nm and 650 nm. The absorbance is not greater than 0.15 at 440 nm and 0.10 at 650 nm.

Reducing substances

Dissolve 0.10 g in dilute hydrochloric acid R and dilute to 25 mL with the same acid. Add 0.2 mL of starch solution R and 0.25 mL of 0.01 M iodine. Allow to stand for 2 min. The solution is blue or violet-brown.

Impurities A and C

Liquid chromatography (2.2.29). Prepare the solutions and mobile phases immediately before use.

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a). Dissolve 5.0 mg of mesalazine impurity C CRS in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (b). Dissolve 5.0 mg of mesalazine impurity A CRS in mobile phase A and dilute to 250.0 mL with mobile phase A. To 1.0 mL of the solution add 1.0 mL of reference solution (a) and dilute to 100.0 mL with mobile phase A.

Reference solution (c). Dilute 1.0 mL of the test solution to 200.0 mL with mobile phase A. To 5.0 mL of this solution add 5.0 mL of reference solution (a).

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (3 μ m).

Mobile phases:

— mobile phase A: dissolve 1.0 g of phosphoric acid R and 2.2 g of perchloric acid R in water R and dilute to 1000.0 mL with the same solvent;
— mobile phase B: dissolve 1.0 g of phosphoric acid R and 1.7 g of perchloric acid R in acetonitrile R1 and dilute to 1000.0 mL with the same solvent;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	100	0
8 - 25	100 \rightarrow 40	0 \rightarrow 60

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C.

Relative retention With reference to mesalazine (retention time = about 9 min): impurity A = about 0.5; impurity C = about 0.9.

0.01 M hydrochloric acid.

Reference solution (c) Dissolve 5 mg of 4-aminosalicylic acid R (impurity E), 5 mg of 2,5-dihydroxybenzoic acid R (impurity G), 15 mg of salicylic acid R (impurity H), 5 mg of 2-chlorobenzoic acid R (impurity L), 5 mg of 2-chloro-5-nitrobenzoic acid R (impurity M), 10 mg of sulfanilic acid R (impurity O) and 5 mg of 3-nitrosalicylic acid R (impurity R) in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 1.0 mL of the solution to 50.0 mL of

0.01 M hydrochloric acid and dilute to 5.0 mL with the same solvent.
Reference solution (b) Dissolve 5 mg of mesalazine for system suitability CRS (containing impurities F, J and P) in this solution to 10.0 mL with 0.01 M hydrochloric acid.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with 0.01 M hydrochloric acid. Dilute 1.0 mL of this solution to 10.0 mL with 0.01 M hydrochloric acid.
Test solution Dissolve 0.100 g of the substance to be examined in 0.01 M hydrochloric acid, with the aid of ultrasound, and dilute to 100.0 mL with the same solvent.

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Related substances

(10 ppm).
in the chromatogram obtained with the reference solution — impurity K: not more than the area of the principal peak
Limit:
— signal-to-noise ratio: minimum 10 for the principal peak.
System suitability: reference solution:

Retention time Impurity K = about 15 min.

Run time 1.5 times the retention time of impurity K.

Injection 50 μ L.

Detection Spectrophotometer at 205 nm.

Flow rate 1.0 mL/min.

Mobile phase Mix 15 volumes of methanol R2 with 85 volumes of a solution containing 1.41 g/L of potassium dihydrogen phosphate R and 0.47 g/L of disodium hydrogen phosphate R previously adjusted to pH 8.0 with a 42 g/L solution of sodium hydroxide R.

temperature: 40 °C.

chromatography R (5 μ m);

— stationary phase: spherical octadecylsilyl silica gel for

— size: $l = 0.25$ m, $\varnothing = 4$ mm;

Column:

with the mobile phase.

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.
Reference solution Dissolve 27.8 mg of aniline hydrochloride R in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 0.20 mL of the solution to 20.0 mL with the mobile phase. Dilute 0.20 mL of this solution to 20.0 mL with the mobile phase.

Impurity K

Liquid chromatography (2.2.29).

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.
Reference solution Dissolve 27.8 mg of aniline hydrochloride R in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 0.20 mL of the solution to 20.0 mL with the mobile phase. Dilute 0.20 mL of this solution to 20.0 mL with the mobile phase.

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (200 ppm);

— impurity C: not more than 4 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (200 ppm);

— impurity F: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (200 ppm).

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to

impurity C and mesalazine.

Limits:

Reference solution (d) Dissolve 3.0 mg of 2-chlorobenzoic acid R (impurity L) in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL of 0.01 M hydrochloric acid.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl amorphous

organosilica polymer for mass spectrometry R (5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: dissolve 6.9 g of sodium dihydrogen

phosphate monohydrate R in 950 mL of water R, adjust to

pH 6.2 with dilute sodium hydroxide solution R and dilute

to 1000 mL with water R;

— mobile phase B: acetonitrile R, mobile phase A (40:60 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	100	0
8 - 20	100 \rightarrow 85	0 \rightarrow 15
20 - 40	85 \rightarrow 25	15 \rightarrow 75
40 - 60	25 \rightarrow 0	75 \rightarrow 100

Detection Spectrophotometer at 240 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied

with mesalazine for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities F, J and P; use the

chromatogram obtained with reference solution (c) to identify

the peaks due to impurities E, G, H, L, M, O and R.

Relative retention With reference to mesalazine (retention

time = about 6 min): impurity O = about 0.5;

impurity J = about 0.6; impurity E = about 0.8;

impurity F = about 1.36; impurity G = about 1.44;

impurity P = about 1.5; impurity L = about 2.0;

impurity M = about 3.3; impurity H = about 3.5;

impurity R = about 5.1.

System suitability:

— peak-to-valley ratio: minimum 3.0, where H_p = height

above the baseline of the peak due to impurity F and

H_o = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

mesalazine in the chromatogram obtained with reference

solution (b);

— signal-to-noise ratio: minimum 10 for the peak due to

impurity L in the chromatogram obtained with reference

solution (d).

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity E = 1.3;

impurity G = 1.4; impurity H = 1.4; impurity J = 2.0;

impurity L = 4.5; impurity M = 1.7; impurity O = 0.6;

impurity P = 0.6; impurity R = 1.3;

— impurity H: not more than 3 times the area of the

principal peak in the chromatogram obtained with

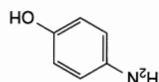
reference solution (a) (0.3 per cent);

— impurities F, J, O, P: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.1 per cent);

A. 4-aminophenol,



I, N, Q, S.

Control of impurities in substances for pharmaceutical use: B, D,

impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph *Substances for pharmaceutical use*

acceptance criterion for other/unspecified impurities and/or

the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

specified impurities A, C, E, F, G, H, J, K, L, M, O, P, R

IMPURITIES

In an airtight container, protected from light.

STORAGE

$C_{17}H_{15}NO_3$.

1 mL of 0.1 M sodium hydroxide is equivalent to 15.31 mg of

determining the end-point potentiometrically (2.2.20).

Dissolve 50.0 mg in 100 mL of boiling water R. Cool rapidly

to room temperature and titrate with 0.1 M sodium hydroxide,

ASSAY

Maximum 0.2 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Loss on drying (2.2.32)

using 1 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test F. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

filter. 15 mL of the filtrate complies with the test.

Shake 1.0 g with 20 mL of distilled water R for 1 min and

Maximum 200 ppm.

Sulfates (2.4.13)

of Cl.

1 mL of 0.005 M silver nitrate is equivalent to 0.1773 mg

potentiometrically (2.2.20).

with 0.005 M silver nitrate, determining the end-point

Add 100 mL of water R and 5 mL of 2 M nitric acid. Titrate

Dissolve 1.50 g in 50 mL of anhydrous formic acid R.

Maximum 0.1 per cent.

Chlorides

(0.03 per cent).

— the chromatogram obtained with reference solution (a)

disregard limit: 0.3 times the area of the principal peak in

(0.5 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.05 per cent);

— chromatogram obtained with reference solution (a)

0.5 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than

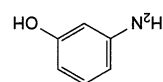
(0.05 per cent);

— chromatogram obtained with reference solution (a)

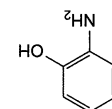
than 0.5 times the area of the principal peak in the

— impurities E, G, L, M, R: for each impurity, not more

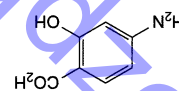
B. 3-aminophenol,



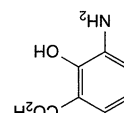
C. 2-aminophenol,



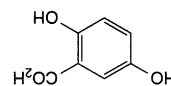
D. 3-aminobenzoic acid,



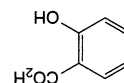
E. 4-amino-2-hydroxybenzoic acid (4-aminosalicylic acid),



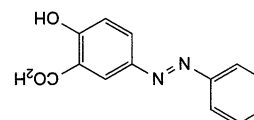
F. 3-amino-2-hydroxybenzoic acid (3-aminosalicylic acid),



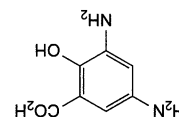
G. 2,5-dihydroxybenzoic acid,



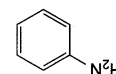
H. 2-hydroxybenzoic acid (salicylic acid),



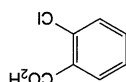
I. 2-hydroxy-5-(phenyldiazenyl)benzoic acid (phenylazosalicylic acid),



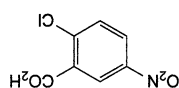
J. 3,5-diamino-2-hydroxybenzoic acid (3,5-diaminosalicylic acid),



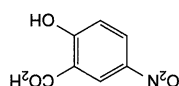
K. aniline,



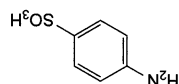
L. 2-chlorobenzoic acid,



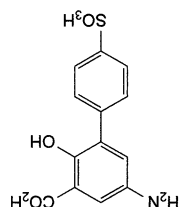
M. 2-chloro-5-nitrobenzoic acid,



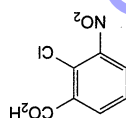
N. 2-hydroxy-5-nitrobenzoic acid (5-nitrosalicylic acid),



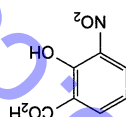
O. 4-aminobenzenesulfonic acid (sulfanilic acid),



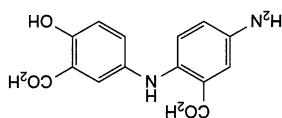
P. 5-amino-2-hydroxy-3-(4-sulphophenyl)benzoic acid (3-(4-sulphophenyl)-5-aminosalicylic acid),



Q. 2-chloro-3-nitrobenzoic acid,



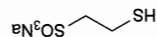
R. 2-hydroxy-3-nitrobenzoic acid (3-nitrosalicylic acid),



S. 2-hydroxy-5-[(2-carboxy-4-aminophenyl)amino]benzoic acid.

Mesna

(Ph. Eur. monograph 1674)



$C_2H_5NaO_3S_2$ 164.2 19767-45-4

Action and use

Preventing adverse effects of cyclophosphamide and ifosfamide.

Ph. Eur.

DEFINITION

Sodium 2-sulfanylethanesulfonate.

Content

96.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellow, crystalline powder, hygroscopic.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of mesna.

B. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference

suspension II (2.2.1) and not more intensely coloured than

reference solution Y₇ (2.2.2, Method II).

pH (2.2.3)

4.5 to 6.0.

Dilute 10 mL of solution S to 20 mL with carbon dioxide-free

water R.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.10 g of the substance to be examined

in the mobile phase and dilute to 25.0 mL with the mobile

phase.

Reference solution (a) Dissolve 4.0 mg of mesna

impurity C CRS in the mobile phase and dilute to 50.0 mL

with the mobile phase. Dilute 2.0 mL of the solution to

20.0 mL with the mobile phase.

Reference solution (b) Dissolve 6.0 mg of mesna

impurity D CRS in the mobile phase and dilute to 50.0 mL

with the mobile phase.

Reference solution (c) Dilute 3.0 mL of the test solution to

10.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (c)

to 100.0 mL with the mobile phase.

Reference solution (e) Dilute 6.0 mL of reference solution (c)

to 20.0 mL with the mobile phase. To 10 mL of the solution

add 10 mL of reference solution (a).

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R

(10 μ m).

Mobile phase Dissolve 2.94 g of potassium dihydrogen phosphate R, 2.94 g of dipotassium hydrogen phosphate R and 2.6 g of tetrabutylammonium hydrogen sulfate R in about 600 mL of water R. Adjust to pH 2.3 with phosphoric acid R, add 335 mL of methanol R and dilute to 1000 mL with water R.

Detection Spectrophotometer at 235 nm.

Injection 20 μ L.

Run time 4 times the retention time of mesna.

Relative retention With reference to mesna (retention

time = about 4.8 min): impurities A and B = about 0.6;

impurity E = about 0.8; impurity C = about 1.4;

impurity D = about 2.3.

System suitability: reference solution (e):

— resolution: minimum 3.0 between the peaks due to mesna

and impurity C.

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of impurities A, B and E by 0.01,

— impurity C: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (a) (0.2 per cent),

— impurity D: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (b) (3.0 per cent),

— impurities A, B, E: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (d) (0.3 per cent),

— any other impurity: for each impurity, not more than one

third of the area of the principal peak in the

chromatogram obtained with reference solution (d)

(0.1 per cent),

— sum of other impurities: not more than the area of the

principal peak in the chromatogram obtained with

reference solution (d) (0.3 per cent),

— disregard limit: 0.15 times the area of the principal peak in

the chromatogram obtained with reference solution (d)

(0.045 per cent).

Chlorides (2.4.4)

Maximum 250 ppm.

Dilute 1 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.

Dilute 5 mL of solution S to 30 mL with distilled water R.

15 mL of the solution complies with the test.

Dissodium edetate

Maximum 500 ppm.

Dissolve 4.000 g in 90 mL of water R and adjust to pH 4.5

using 0.1 M hydrochloric acid. Add 10 mL of acetate buffer

solution pH 4.5 R and 50 mL of 2-propanol R. Titrate with

0.25 g/L solution of diethylenetriamine R in 2-propanol R. Titrant with

0.01 M zinc sulfate until the colour changes from bluish-grey

to pink.

1 mL of 0.01 M zinc sulfate is equivalent to 3.72 mg

of $C_{10}H_{14}N_2Na_2O_8 \cdot 2H_2O$.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dilute 10 mL of solution S to 20 mL with water R. 12 mL of

the solution complies with limit test A. Prepare the reference

solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g under high

vacuum at 60 °C for 2 h.

ASSAY

Dissolve 0.120 g in 10 mL of water R. Add 10 mL of 1 M sulfuric acid and 10.0 mL of 0.1 M iodine. Titrate with 0.1 M sodium thiosulfate adding 1 mL of starch solution R near the endpoint. Carry out a blank titration.

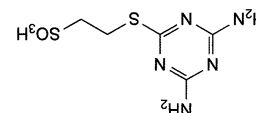
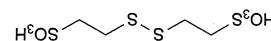
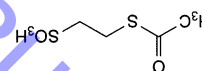
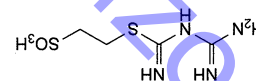
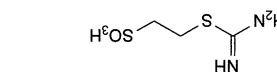
1 mL of 0.1 M sodium thiosulfate is equivalent to 16.42 mg of $C_{20}H_{32}O_2NaO_3S_2$.

STORAGE

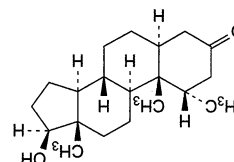
In an airtight container.

IMPURITIES

Specified impurities: A, B, C, D, E.

**Mestosterone**

(Ph. Eur. monograph 1730)



$C_{20}H_{32}O_2$

304.5

1424-00-6

Action and use

Androgen.

Ph. Eur.

DEFINITION

17β-Hydroxy-1α-methyl-5α-androstan-3-one.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS**Appearance**

White or yellowish crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in acetone, in ethyl acetate and in methanol.

IDENTIFICATION

A. Melting point (2.2.14): 206 °C to 211 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison mestosterone CRS.

TESTS

Specific optical rotation (2.2.7)

+ 20 to + 24 (dried substance).

Dissolve 0.200 g in methylene chloride R and dilute to 10.0 mL with the same solvent.

Impurity B

Maximum 0.5 per cent.

Thin-layer chromatography (2.2.27).

Test solution Dissolve 100.0 mg of the substance to be examined in a mixture of equal volumes of methanol R and methylene chloride R and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with a mixture of equal volumes of methanol R and methylene chloride R.

Reference solution (b) Dissolve 5.0 mg of mestosterone impurity A CRS in reference solution (a) and dilute to 100.0 mL with the same solution.

Plate TLC silica gel plate R.

Mobile phase methanol R, acetone R, toluene R (2:15:85 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 366 nm; spray with a 200 g/L solution of toluenesulfonic acid R in alcohol R and heat the plate for 10 min at 120 °C.

System suitability The chromatogram obtained with reference solution (b) shows 2 clearly separated spots (blue spot due to mestosterone and yellow spot due to impurity A).

Limit:

— impurity B: any blue spot, apart from the main spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in a mixture of 20 volumes of water R and 80 volumes of acetonitrile R and dilute to 25.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 50.0 mg of mestosterone CRS in a mixture of 20 volumes of water R and 80 volumes of acetonitrile R and dilute to 25.0 mL with the same mixture of solvents.

Reference solution (b). Dissolve 10.0 mg of mestosterone impurity A CRS in a mixture of 20 volumes of water R and 80 volumes of acetonitrile R and dilute to 5.0 mL with the same mixture of solvents.

Reference solution (c) Dilute 0.5 mL of reference solution (a) and 0.5 mL of reference solution (b) to 100.0 mL with a

mixture of 20 volumes of water R and 80 volumes of acetonitrile R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R

(3 μ m).

Mobile phase acetonitrile R, water R, methanol R

(20:40:60 V/V/V).

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 50 μ L; inject the test solution and reference

solution (c).

Run time 3 times the retention time of mestrolone.

Relative retention With reference to mestrolone (retention

time = about 22 min); impurity A = about 0.7.

System suitability: reference solution (c):

— resolution: minimum 0.0 between the peaks due to

impurity A and to mestrolone.

Limits:

— impurity A: not more than the area of the peak due to

impurity A in the chromatogram obtained with reference

solution (c) (0.5 per cent),

— any other impurity: not more than half the area of the peak

due to mestrolone in the chromatogram obtained with

reference solution (c) (0.25 per cent),

— total: not more than 1.5 times the area of the peak due to

mestrolone in the chromatogram obtained with reference

solution (c) (0.75 per cent),

— disregard limit: 0.1 times the area of the peak due to

mestrolone in the chromatogram obtained with reference

solution (c) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

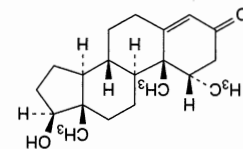
related substances.

Injection 10 μ L; inject the test solution and reference

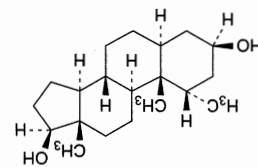
solution (a).

Calculate the percentage content of $C_{20}H_{32}O_2$.

IMPURITIES



A. 17β-hydroxy-1α-methylandrosta-4-en-3-one,

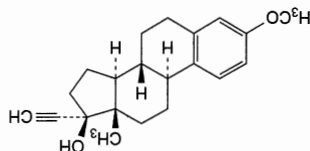


B. 1α-methyl-5α-androsta-3β,17β-diol.

Ph Eur

Mestranol

(Ph. Eur. monograph 0509)



$C_{21}H_{26}O_2$

310.4

72-33-3

Action and use

Estrogen.

Ph Eur

DEFINITION

Mestranol contains not less than 98.0 per cent and not more

than the equivalent of 102.0 per cent of 3-methoxy-19-nor-

17α-pregna-1,3,5(10)-trien-20-yn-17-ol, calculated with

reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, practically

insoluble in water, sparingly soluble in alcohol.

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Melting point (2.2.14): 150 °C to 154 °C.

B. Examine by infrared absorption spectrophotometry

(2.2.24), comparing with the spectrum obtained with

mestranol CRS.

C. Examine the chromatograms obtained in the test for

related substances in daylight and in ultraviolet light at

365 nm. The principal spot in the chromatogram obtained

with test solution (b) is similar in position, colour,

fluorescence and size to the principal spot in the

chromatogram obtained with reference solution (a).

D. Dissolve about 5 mg in 1 mL of sulfuric acid R. A red

colour develops with a greenish-yellow fluorescence in

ultraviolet light at 365 nm. Add the solution to 10 mL of

water R and mix. The solution becomes pink and a pink to

violet precipitate is formed on standing.

TESTS

Specific optical rotation (2.2.7)

Dissolve 0.100 g in anhydrous pyridine R and dilute to

10.0 mL with the same solvent. The specific optical rotation

is –20 to –24, calculated with reference to the dried

substance.

Absorbance (2.2.25)

Dissolve 25.0 mg in alcohol R and dilute to 25.0 mL with the

same solvent. Dilute 10.0 mL of this solution to 100.0 mL

with alcohol R. Examine between 260 nm and 310 nm, the

solution shows two absorption maxima, at 279 nm and

288 nm, and a minimum at 286 nm. The specific

absorptivities at the maxima are 62 to 68 and 59 to 64,

respectively.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica

gel G R as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be

examined in chloroform R and dilute to 10 mL with the same

solvent.



Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with chloroform R.

Reference solution (a) Dissolve 10 mg of *mestranol* CRS in chloroform R and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (b) to 10 mL with chloroform R.

Reference solution (c) Dilute 5 mL of reference solution (b) to 10 mL with chloroform R.

Apply separately to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of *alcohol* R and 90 volumes of *toluene* R. Allow the plate to dry in air until the solvent has evaporated. Heat at 110 °C for 10 min. Spray the hot plate with *alcoholic solution of sulfuric acid* R. Heat again at 110 °C for 10 min. Examine in daylight and in ultraviolet light at 365 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (1.0 per cent) and at most one such spot is more intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent).

Loss on drying (2.2.32)

Not more than 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C for 3 h.

ASSAY

Dissolve 0.200 g in 40 mL of *tetrahydrofuran* R and add 5 mL of a 100 g/L solution of *silver nitrate* R. Titrate with 0.1 M *sodium hydroxide*, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

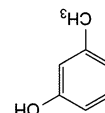
1 mL of 0.1 M *sodium hydroxide* is equivalent to 31.04 mg of $C_{21}H_{26}O_2$.

STORAGE

Store protected from light.

Metacresol

(Ph. Eur. monograph 2077)



C_7H_8O

108.1

108-39-4

Action and use

Antiseptic; antimicrobial preservative.

Ph Eur

DEFINITION

3-Methylphenol.

CHARACTERS

Appearance

Colourless or yellowish liquid.

Solubility

Sparingly soluble in water, miscible with ethanol (96 per cent) and with methylene chloride.

Relative density

About 1.03.

mp

About 11 °C.

bp

About 202 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of metacresol.

TESTS

Solution S

Dissolve 1.5 g in *carbon dioxide-free water* R and dilute to 100 mL with the same solvent.

Appearance of solution

Freshly prepared solution S is not more opalescent than reference suspension III (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Acidity

To 25 mL of solution S add 0.15 mL of *methyl red solution* R. The solution is red. Not more than 0.5 mL of 0.01 M *sodium hydroxide* is required to change the colour of the indicator to yellow.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution Dissolve 1.00 g of the substance to be examined in *methanol* R and dilute to 100.0 mL with the same solvent. **Reference solution (a)** Dissolve 0.10 g of *cresol* R, 0.10 g of *p-cresol* R and 0.10 g of the substance to be examined in *methanol* R and dilute to 20.0 mL with the same solvent. **Reference solution (b)** Dilute 1.0 mL of the test solution to 100.0 mL with *methanol* R. Dilute 1.0 mL of this solution to 20.0 mL with *methanol* R.

Column:

— material: fused silica,

— size: l = 25 m, Ø = 0.25 mm,

— stationary phase:

poly[(cyanopropyl)(methyl)][(phenyl)(methyl)]siloxane R (0.2 µm).

Carrier gas *helium* for chromatography R.

Flow rate 1.8 mL/min.

Split ratio 1:30.

Temperature:

Time (min)	Temperature (°C)
0 - 35	100
35 - 40	100 → 150
40 - 50	150
Injection port	200
Detector	200

Detection Flame ionisation.

Injection 1.0 µL.

Relative retention With reference to metacresol (retention time = about 28 min): impurity B = about 0.75;

impurity C = about 0.98.

System suitability: reference solution (a):

— resolution: minimum 1.4 between the peaks due to impurity C and metacresol.

Limits:
— **impurities B, C:** for each impurity, not more than 0.5 per cent,
— **any other impurity:** for each impurity, not more than 0.1 per cent,
— **total:** not more than 1.0 per cent.
— **disregard limit:** the area of the peak due to metacresol in the chromatogram obtained with reference solution (b) (0.05 per cent).

Residue on evaporation

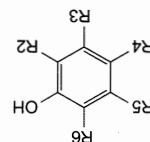
Maximum 0.1 per cent.
Evaporate 2.0 g to dryness on a water-bath in a fume cupboard and dry at 100-105 °C for 1 h. The residue weighs a maximum of 2 mg.

STORAGE

In an airtight container, protected from light.

IMPURITIES

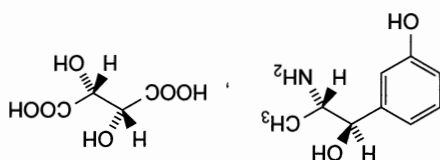
Specified impurities B, C.
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D, E, F, G, H, I, J, K, L, M.



- A. R2 = R3 = R4 = R5 = R6 = H: phenol,
B. R2 = CH₃, R3 = R4 = R5 = R6 = H: 2-methylphenol (o-cresol, cresol),
C. R2 = R3 = R4 = R5 = R6 = H, R4 = CH₃: 4-methylphenol (p-cresol),
D. R2 = R6 = CH₃, R3 = R4 = R5 = H:
2,6-dimethylphenol (2,6-xyleneol),
E. R2 = C₂H₅, R3 = R4 = R5 = R6 = H: 2-ethylphenol (o-ethylphenol),
F. R2 = R4 = CH₃, R3 = R5 = R6 = H:
2,4-dimethylphenol (2,4-xyleneol),
G. R2 = R5 = CH₃, R3 = R4 = R6 = H:
2,5-dimethylphenol (2,5-xyleneol),
H. R2 = CH(CH₃)₂, R3 = R4 = R5 = R6 = H:
2-(1-methylethyl)phenol,
I. R2 = R3 = CH₃, R4 = R5 = R6 = H:
2,3-dimethylphenol (2,3-xyleneol),
J. R2 = R4 = R6 = H, R3 = R5 = CH₃:
3,5-dimethylphenol (3,5-xyleneol),
K. R2 = R3 = R5 = R6 = H, R4 = C₂H₅: 4-ethylphenol (p-ethylphenol),
L. R2 = R5 = R6 = H, R3 = R4 = CH₃:
3,4-dimethylphenol (3,4-xyleneol),
M. R2 = R3 = R5 = CH₃, R4 = R6 = H:
2,3,5-trimethylphenol.

Ph Eur

Metaraminol Tartrate



C₉H₁₃NO₂·C₄H₆O₆

317.3

17171-57-2

Action and use

Adrenoceptor agonist.

Preparation

Metaraminol Injection

DEFINITION

Metaraminol Tartrate is (1R,2S)-2-amino-1-(3-hydroxyphenyl)propan-1-ol hydrogen (2R,3R)-tartrate. It contains not less than 99.0% and not more than 101.0% of C₉H₁₃NO₂·C₄H₆O₆, calculated with reference to the dried substance.

CHARACTERISTICS

A white, crystalline powder.
Freely soluble in water; sparingly soluble in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

A. In the test for Related substances the principal spot in the chromatogram obtained with solution (2) corresponds to that in the chromatogram obtained with solution (4).
B. To 0.5 mL of a 0.05% w/v solution add 0.5 mL of phosphomolybdic reagent and 5 mL of dilute sodium carbonate solution and allow to stand for 5 minutes. An intense blue colour is produced.
C. To 4 mL of a 0.05% w/v solution add 5 mL of borate buffer pH 9.6 and 1 mL of a freshly prepared 0.5% w/v solution of sodium 1,2-naphthoquinone-4-sulfonate and allow to stand for 1 minute. Add 0.2 mL of a 2% v/v solution of benzaldehyde in chloroform and 5 mL of toluene and shake. A mauve colour is immediately produced in the toluene layer (distinction from phenylephrine).

TESTS

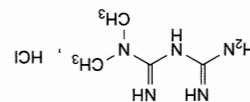
Acidity
pH of a 5% w/v solution, 3.2 to 3.5, Appendix V L.
Phenones
Absorbance of a 0.2% w/v solution at 310 nm, not more than 0.2, calculated with reference to the dried substance, Appendix II B.

Related substances

Carry out in subdued light the method for thin-layer chromatography, Appendix III A, using the following solutions in methanol.
(1) 1.0% w/v of the substance being examined.
(2) 0.050% w/v of the substance being examined.
(3) 0.0050% w/v of the substance being examined.
(4) 0.050% w/v of metaraminol tartrate BPCRS.
CHROMATOGRAPHIC CONDITIONS
(a) Use a silica gel precoated plate (Merck silica gel 60 plates are suitable).
(b) Use the mobile phase as described below.
(c) Apply 10 µL of each solution.

Metformin Hydrochloride

(Ph. Eur. monograph 0931)



$C_4H_{12}ClN_5$

165.6

1115-70-4

Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.6 g and crystal violet solution as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 31.73 mg of $C_4H_{12}NO_2 \cdot C_4H_6O_6$.

ASSAY

Not more than 0.1%, Appendix IX A.

Sulfated ash

0.5% of its weight. Use 1 g.

When dried to constant weight at 105°, loses not more than

Loss on drying

chromatogram obtained with solution (3) (0.5%).

Any secondary spot in the chromatogram obtained with

solution (1) is not more intense than the spot in the

LIMITS

10 volumes of 13.5M ammonia, 80 volumes of chloroform and 80 volumes of methanol.

MOBILE PHASE

2M sodium carbonate.

stand for 5 minutes and mix cautiously with 25 mL of with 1.5 mL of a 5% w/v solution of sodium nitrite, allow to 0.45% w/v solution of sulfanilic acid in 1M hydrochloric acid solution prepared in the following manner. Mix 25 mL of a

(d) Develop the plate to 15 cm.

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white crystals.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in acetone and in methylene chloride.

IDENTIFICATION

First identification B, E

Second identification A, C, D, E

Comparison metformin hydrochloride CRS.

B. Infrared absorption spectrophotometry (2.2.24).

A. Melting point (2.2.14): 222 °C to 226 °C.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in water R and dilute to 5 mL with the same solvent.

Reference solution Dissolve 20 mg of metformin hydrochloride CRS in water R and dilute to 5 mL with the

same solvent.

Plate TLC silica gel G plate R.

Mobile phase glacial acetic acid R, butanol R, water R (10:40:50 V/V/V); use the upper layer.

Application 5 µL.

Development Over 3/4 of the plate.

Drying At 100-105 °C for 15 min.

Detection Spray with a mixture of equal volumes of a 100 g/L solution of sodium nitroprusside R, a 100 g/L solution of potassium ferricyanide R and a 100 g/L solution of sodium hydroxide R, prepared 20 min before use.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 5 mg in water R and dilute to 100 mL with the same solvent. To 2 mL of the solution add 0.25 mL of strong sodium hydroxide solution R and 0.10 mL of α-naphthol solution R. Mix and allow to stand in iced water for 15 min. Add 0.5 mL of sodium hypobromite solution R and mix. A pink colour develops.

E. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II). Heat the solution to 50 °C and cool to room temperature.

Impurity F

Liquid chromatography (2.2.29).

Derivatisation solution. Prepare the solution immediately before use. Dissolve 1 mL of fluorodinitrobenzene R in 100.0 mL of acetonitrile for chromatography R.

Blank solution To 5.0 mL of acetonitrile for chromatography R add 100 µL of triethylamine R1 and 1.0 mL of the derivatisation solution. Shake well and heat at 60 °C for 30 min. After cooling, dilute to 10.0 mL with acetonitrile for chromatography R.

Test solution Prepare the solution immediately before use. Suspend 10.0 mg of the substance to be examined in 5.0 mL of acetonitrile for chromatography R and sonicate for 5 min. Add 100 µL of triethylamine R1 and 1.0 mL of the derivatisation solution. Shake well and heat at 60 °C for 30 min. After cooling, dilute to 10.0 mL with acetonitrile for chromatography R. Filter or centrifuge at 800 g for 5 min before use.

Reference solution Dissolve 1.0 mL of metformin impurity F CRS in 100.0 mL of acetonitrile for

chromatography R. Dilute 2.5 mL of the solution to 100.0 mL with acetonitrile for chromatography R. To 1.0 mL of this solution add successively 5.0 mL of acetonitrile for chromatography R, 100 µL of triethylamine R1 and 1.0 mL of the derivatisation solution. Shake well and heat at 60 °C for 30 min. After cooling, dilute to 10.0 mL with acetonitrile for chromatography R.

Column:
— size: $l = 0.125$ m, $\phi = 3$ mm;
— stationary phase: spherical end-capped octadecylsilyl silica gel
for chromatography R1 (5 μ m);
— temperature: 30 °C.
Mobile phase:
— mobile phase A: phosphoric acid R, water R (0.1:99.9 V/V);
— mobile phase B: acetonitrile for chromatography R₅.

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 10	60 → 45	40 → 55
10 - 11	45 → 25	55 → 75
11 - 15	25	75

Flow rate 0.7 mL/min.
Detection Spectrophotometer at 380 nm.
Injection 5 μ L.

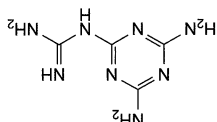
Identification of impurities Use the chromatograms obtained with the blank solution and the reference solution to identify the peak due to the impurity F derivative.
Retention time Impurity F derivative = about 4 min.
System suitability: reference solution:
— **resolution:** minimum 3.0 between the peak due to the impurity F derivative and the nearby eluting peaks due to the derivatisation reagent.
Limit:
— **impurity F:** not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.05 per cent).

Related substances

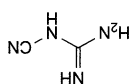
Liquid chromatography (2.2.29).
Test solution Dissolve 0.50 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 20.0 mg of metformin impurity A CRS in water R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 200.0 mL with the mobile phase.
Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.
Reference solution (c) Dissolve 10 mg of melamine R (impurity D) in about 90 mL of water R. Add 5 mL of the test solution and dilute to 100 mL with water R. Dilute 1 mL of this solution to 50 mL with the mobile phase.

Column:
— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: strong cation-exchange silica gel for chromatography R (10 μ m).
Mobile phase 17 g/L solution of ammonium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R.
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 218 nm.

Run time Twice the retention time of metformin.
Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.



A. cyanoguanidine,



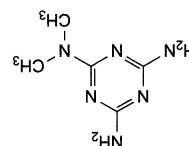
Specified impurities A, F
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: B, C, D, E.

IMPURITIES

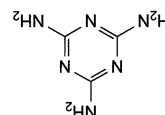
ASSAY
Dissolve 0.100 g in 4 mL of anhydrous formic acid R. Add 80 mL of acetonitrile R. Carry out the titration immediately. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 16.56 mg of C₄H₁₂N₅.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 5 h.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.
Heavy metals (2.4.8)
Maximum 10 ppm.
12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

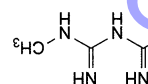
Relative retention With reference to metformin (retention time = about 10 min): impurity A = about 0.2; impurity D = about 0.3.
System suitability: reference solution (c):
— **resolution:** minimum 10 between the peaks due to impurity D and metformin.
Limit:
— **impurity A:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.02 per cent);
— **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);
— **total:** maximum 0.2 per cent;
— **disregard limit:** 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);
— **the chromatogram obtained with reference solution (b)** (0.03 per cent); do not disregard the peak due to impurity A.



C. N,N' -dimethyl-1,3,5-triazine-2,4,6-triamine (N,N' -dimethylmelamine),



D. 1,3,5-triazine-2,4,6-triamine (melamine),



E. 1-methylbiguanide,

F. N -methylethylmethanamine (dimethylamine),

Basic Butylated Methacrylate Copolymer

(Ph. Eur. monograph 1975)

Action and use

Excipient.

DEFINITION

Copolymer of 2-(dimethylamino)ethyl methacrylate, butyl methacrylate and methyl methacrylate having a mean relative molecular mass of about 150 000. The ratio of 2-(dimethylamino)ethyl methacrylate groups to butyl methacrylate and methyl methacrylate groups is about 2:1:1. Content of dimethylaminoethyl groups 20.8 per cent to 25.5 per cent (dried substance).

CHARACTERS

Appearance
Colourless or yellowish granules or white or almost white powder, slightly hygroscopic.

Solubility

Practically insoluble in water, freely soluble in methylene chloride. It dissolves slowly in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison basic butylated methacrylate copolymer CRS.

TESTS

B. It complies with the limits of the assay.

Solution S

Dissolve 12.5 g in a mixture of 35.0 g of acetone R and 52.5 g of 2-propanol R.

Viscosity (2.2.10)

3 mPa·s to 6 mPa·s, determined on solution S.

Apparatus Rotating viscometer.

Dimensions:

— spindle: diameter = 25.15 mm, height = 90.74 mm, shaft diameter = 4 mm;
— cylinder: diameter = 27.62 mm, height = 0.135 mm.

Rotating speed 30 r/min.

Volume of solution 16 mL of solution S.

Temperature 20 °C.

Absorbance (2.2.25)

Maximumum 0.30 at 420 nm, determined on solution S.

Appearance of a film

Spread 1.0 mL of solution S evenly on a glass plate. Upon drying a clear film is formed.

Monomers

Maximumum 0.1 per cent for each monomer (butyl methacrylate, methyl methacrylate and

2-(dimethylamino)ethyl methacrylate), determined by procedures A and B.

A. Butyl methacrylate and methyl methacrylate. Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R1, phosphate buffer solution pH 2.0 R (40:60 V/V).

Test solution Dissolve 1.00 g of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution Dissolve 20.0 mg of butyl methacrylate CRS (impurity A) and 10.0 mg of methyl methacrylate CRS (impurity B) in 3.0 mL of butanol R and dilute to 10.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 250.0 mL with the solvent mixture.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (7 μ m).

Mobile phase phosphate buffer solution pH 2.0 R, methanol R (45:55 V/V).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 50 μ L.

System suitability: reference solution:

— resolution: minimum 5 between the peaks due to impurities A and B.

Calculate the percentage content of each monomer using the following expression:

$$100 \times 10^{-6} \times \frac{M}{C} \times \frac{A_T}{A_R} \times 50$$

C = concentration of the monomer in the reference solution, in micrograms per millilitre;

M = mass of substance to be examined in the test solution, in grams;

A_T = area of the peak due to the monomer in the chromatogram obtained with the test solution;

A_R = area of the peak due to the monomer in the chromatogram obtained with the reference solution.

B. 2-(Dimethylamino)ethyl methacrylate. Liquid chromatography (2.2.29).

Test solution Dissolve 1.00 g of the substance to be examined in tetrahydrofuran R and dilute to 50.0 mL with the same solvent.

Reference solution Dissolve 10.0 mg of 2-(dimethylamino)ethyl methacrylate CRS (impurity C) in tetrahydrofuran R and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 50.0 mL with tetrahydrofuran R.

Column:
— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;
— stationary phase: aminopropylsilyl silica gel for chromatography R (7 μ m).
Mobile phase Mix 25 volumes of a 3.404 g/L solution of potassium dihydrogen phosphate R and 75 volumes of tetrahydrofuran R.
Flow rate 2.0 mL/min.
Detection Spectrophotometer at 215 nm.
Injection 50 μ L.

Calculate the percentage content of impurity C as described under procedure A.
Heavy metals (2.4.8)
Maximum 20 ppm.
2.0 g complies with test C. Prepare the reference solution using 4.0 mL of lead standard solution (10 ppm Pb) R.

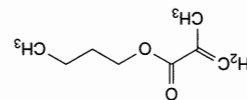
Loss on drying (2.2.32)
Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 110 °C for 3 h.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in a mixture of 4 mL of water R and 96 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 7.21 mg of C_4H_9NO .

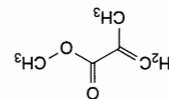
STORAGE

In an airtight container.

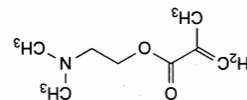
IMPURITIES



A. butyl 2-methylprop-2-enoate (butyl methacrylate),



B. methyl 2-methylprop-2-enoate (methyl methacrylate),



C. 2-(dimethylamino)ethyl 2-methylprop-2-enoate (2-(dimethylamino)ethyl methacrylate).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the mandatory related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the

Methacrylic Acid - Ethyl Acrylate Copolymer (1:1)



Action and use
Pharmaceutical aid.

(Ph. Eur. monograph 1128)

DEFINITION

Copolymer of methacrylic acid and ethyl acrylate having a mean relative molecular mass of about 250 000. The ratio of carboxylic groups to ester groups is about 1:1. The substance is in the acid form (type A) or partially neutralised using sodium hydroxide (type B). It may contain suitable surface-active agents such as sodium dodecyl sulfate and polysorbate 80.

Content

— type A: 46.0 per cent to 50.6 per cent of methacrylic acid units (dried substance);
— type B: 43.0 per cent to 48.0 per cent of methacrylic acid units (dried substance).

CHARACTERS

Appearance

White or almost white, free-flowing powder.

Solubility

Practically insoluble in water (type A) or dispersible in water (type B), freely soluble in anhydrous ethanol, practically insoluble in ethyl acetate. It is freely soluble in a 40 g/L solution of sodium hydroxide.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 0.1 g of the substance to be examined in 1 mL of ethanol (90 per cent V/V) R, and place 2 drops of the solution on a sodium chloride plate; dry to allow the formation of a film and cover with another sodium chloride plate.

Comparison methacrylic acid - ethyl acrylate copolymer (1:1) (type A or type B) CRS.

B. It complies with the limits of the assay.

C. Sulfated ash (see Tests).

TESTS

Viscosity (2.2.10)

— *Type A*: 100 mPa·s to 200 mPa·s.

Dissolve a quantity of the substance to be examined corresponding to 37.5 g of the dried substance in a mixture of 7.9 g of water R and 254.6 g of 2-propanol R. Determine the viscosity at 20 °C using a rotating viscometer at a shear rate of 10 s⁻¹.

— *Type B*: not more than 100 mPa·s.

Disperse a quantity of the substance to be examined corresponding to 80.0 g of the dried substance in water R and make up to 320 g with the same solvent. Stir for 3 h and determine the viscosity at 23 °C using a rotating viscometer and a spindle rotating at 100 r/min.

Dimensions of the spindle Diameter = 47.0 mm; height = 27.0 mm; shaft diameter = 3.18 mm.

Appearance of a film

Place 1 mL of the solution (type A) or dispersion (type B) prepared for the test for viscosity on a glass plate and allow to dry. A clear, brittle film is formed.

Ethyl acrylate and methacrylic acid

Liquid chromatography (2.2.29).

Blank solution To 50.0 mL of methanol R add 25.0 mL of the mobile phase.

Test solution Dissolve 40 mg of the substance to be examined in 50.0 mL of methanol R and add 25.0 mL of the mobile phase.

Reference solution Dissolve 10 mg of ethyl acrylate R and 10 mg of methacrylic acid R in methanol R and dilute to 50.0 mL with the same solvent. Dilute 0.1 mL of the solution to 50.0 mL with methanol R and add 25.0 mL of the mobile phase.

Column: size: $l = 0.10$ m, $\varnothing = 4$ mm; stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase methanol R, phosphate buffer solution pH 2.0 R (30:70 V/V).

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 202 nm.

Injection 50 μ L.

System suitability:

— *resolution*: minimum 2.0 between the peaks due to ethyl acrylate and methacrylic acid in the chromatogram

obtained with the reference solution; the chromatogram obtained with the same retention time as the peaks due to ethyl acrylate or methacrylic acid.

Limit:

— *sum of the contents of ethyl acrylate and methacrylic acid*: maximum 0.1 per cent.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.4 per cent (type A) or 0.5 per cent to 3.0 per cent (type B), determined on 1.0 g.

ASSAY

Dissolve 1.000 g in a mixture of 40 mL of water R and 60 mL of 2-propanol R. Titrate slowly while stirring with

0.5 M sodium hydroxide, using phenolphthalein solution R as indicator.

1 mL of 0.5 M sodium hydroxide is equivalent to 43.05 mg of C₄H₆O₂ (methacrylic acid units).

LABELLING

The label states the type (type A or type B).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for methacrylic acid - ethyl acrylate copolymer (1:1) used as gastro-resistant coating agent.

Viscosity

(see Tests).

Appearance of a film

(see Tests).

Solubility of a film

Take a piece of the film obtained in the test for appearance of a film (see Tests), place it in a flask containing 0.1 M hydrochloric acid and stir. It does not dissolve within 2 h. Take another piece of the film and place it in a flask containing phosphate buffer solution pH 6.0 R with stirring. It dissolves within 1 h.

Ph Eur

Methacrylic Acid - Ethyl Acrylate Copolymer (1:1) Dispersion 30 per cent

(Ph Eur monograph 1129)

Action and use

Excipient.

Ph Eur

DEFINITION

Dispersion in water of a copolymer of methacrylic acid and ethyl acrylate having a mean relative molecular mass of about 250 000. The ratio of carboxylic groups to ester groups is about 1:1.

Content

46.0 per cent to 50.6 per cent of methacrylic acid units (residue on evaporation).

It may contain suitable surface-active agents such as sodium dodecyl sulfate and polysorbate 80.

CHARACTERISTICS

Appearance

Opaque, white or almost white, slightly viscous liquid.

Solubility

Miscible with water. On addition of solvents such as acetone, anhydrous ethanol or 2-propanol, a precipitate is formed which dissolves on addition of excess solvent. It is miscible with a 40 g/L solution of sodium hydroxide.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of methacrylic acid - ethyl acrylate copolymer (1:1) dispersion 30 per cent.

B. It complies with the limits of the assay.

TESTS

Viscosity (2.2.10)

Maximum 15 mPa·s, determined using a rotating viscometer at 20 °C and at a shear rate of 50 s⁻¹.

Appearance of a film

Place 1 mL on a glass plate and allow to dry. A clear, brittle film is formed.

Particulate matter

Filter 100.0 g through a tared stainless steel sieve (90). Rinse with water R until a clear filtrate is obtained and dry at 100-105 °C. The residue weighs a maximum of 1.00 g.

Ethyl acrylate and methacrylic acid

Liquid chromatography (2.2.29).
Blank solution T to 50.0 mL of methanol R add 25.0 mL of the mobile phase.

Test solution Dissolve 40 mg of the dispersion to be examined in 50.0 mL of methanol R and add 25.0 mL of the mobile phase.

Reference solution Dissolve 10 mg of ethyl acrylate R and 10 mg of methacrylic acid R in methanol R, then dilute to 50.0 mL with the same solvent. Dilute 0.1 mL of the solution to 50.0 mL with methanol R and add 25.0 mL of the mobile phase.

Column:

— size: $l = 0.10$ m, $\phi = 4$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).
Mobile phase methanol R, phosphate buffer solution pH 2.0 R (30:70 V/V).

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 202 nm.

Injection 50 μ L.

System suitability:

— resolution: minimum 2.0 between the peaks due to ethyl acrylate and methacrylic acid in the chromatogram

— obtained with the reference solution;

— the chromatogram obtained with the blank solution does not show peaks with the same retention times as ethyl acrylate or methacrylic acid.

Limit:

— sum of the contents of ethyl acrylate and methacrylic acid: maximum 0.1 per cent.

Residue on evaporation

28.5 per cent to 31.5 per cent.

Dry 1.000 g at 110 °C for 5 h. The residue weighs not less than 0.285 g and not more than 0.315 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

ASSAY

Dissolve 1.500 g in a mixture of 40 mL of water R and 60 mL of 2-propanol R. Titrate slowly while stirring with 0.5 M sodium hydroxide, using phenolphthalein solution R as indicator.

1 mL of 0.5 M sodium hydroxide is equivalent to 43.05 mg of C₄H₆O₂ (methacrylic acid units).

STORAGE

At a temperature of 5 °C to 25 °C, protected from freezing.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.
The following characteristics may be relevant for methacrylic acid-ethyl acrylate copolymer (1:1) dispersion 30 per cent used as a gastro-resistant coating agent.

Viscosity

(see Tests).

Appearance of a film

(see Tests).

Solubility of a film

Take a piece of the film obtained in the test for appearance of a film and place it in a flask containing a 10.3 g/L solution of hydrochloric acid R with stirring. It does not dissolve within 2 h. Take another piece of the film and place it in a flask containing phosphate buffer solution pH 6.0 R with stirring. It dissolves within 1 h.

Action and use

Excipient.

DEFINITION

Copolymer of methacrylic acid and methyl methacrylate having a mean relative molecular mass of about 135 000. The ratio of carboxylic groups to ester groups is about 1:1. Content 46.0 per cent to 50.6 per cent of methacrylic acid units (dried substance).

CHARACTERS

Appearance

White or almost white, free-flowing powder.



Methacrylic Acid - Methyl Methacrylate Copolymer (1:1)

(Ph. Eur. monograph 1127)

1 mL of 0.5 M sodium hydroxide is equivalent to 43.05 mg of $C_4H_6O_2$ (methacrylic acid units).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for methacrylic acid-methyl methacrylate copolymer (1:1) used as a gastro-resistant coating agent.

Viscosity

(see Tests).

Appearance of a film

(see Tests).

Solubility of a film

Take a piece of the film obtained in the test for appearance of a film and place it in a flask containing a 10.3 g/L solution of hydrochloric acid R with stirring. It does not dissolve within 2 h. Take another piece of the film and place it in a flask containing phosphate buffer solution pH 6.8 R with stirring. It dissolves within 1 h.

Action and use

Excipient.

(Ph. Eur. monograph 1130)

Methacrylic Acid - Methyl Methacrylate Copolymer (1:2)

Ph Eur



DEFINITION

Copolymer of methacrylic acid and methyl methacrylate having a mean relative molecular mass of about 135 000. The ratio of carboxylic groups to ester groups is about 1:2.

Content

27.6 per cent to 30.7 per cent of methacrylic acid units (dried substance).

CHARACTERS

Appearance

White or almost white, free-flowing powder.

Solubility

Practically insoluble in water, freely soluble in anhydrous ethanol and in 2-propanol, practically insoluble in ethyl acetate. It is freely soluble in a 40 g/L solution of sodium hydroxide.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Solubility

Practically insoluble in water, freely soluble in anhydrous ethanol and in 2-propanol, practically insoluble in ethyl acetate. It is freely soluble in a 40 g/L solution of sodium hydroxide.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of methacrylic acid - methyl methacrylate copolymer (1:1).

B. It complies with the limits of the assay.

TESTS

Viscosity (2.2.10)

50 mPa·s to 200 mPa·s. Dissolve a quantity of the substance to be examined corresponding to 37.5 g of the dried substance in a mixture of 7.9 g of water R and 254.6 g of 2-propanol R. Determine the viscosity using a rotating viscometer at 20 °C and at a shear rate of 10 s⁻¹.

Appearance of a film

Place 1 mL of the solution prepared in the test for viscosity on a glass plate and allow to dry. A clear, brittle film is formed.

Methyl methacrylate and methacrylic acid

Liquid chromatography (2.2.29).

Blank solution To 50.0 mL of methanol R add 25.0 mL of the mobile phase.

Test solution Dissolve 40 mg of the substance to be examined in 50.0 mL of methanol R and add 25.0 mL of the mobile phase.

Reference solution Dissolve 10 mg of methacrylic acid R and 10 mg of methyl methacrylate R in methanol R, then dilute to 50.0 mL with the same solvent. Dilute 0.1 mL of this solution to 50.0 mL with methanol R and add 25.0 mL of the mobile phase.

Column: mobile phase.

— size: $l = 0.10$ m, $\phi = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase methanol R, phosphate buffer solution pH 2.0 R (30:70 V/V).

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 202 nm.

Injection 50 μ L.

System suitability:

— resolution: minimum 2.0 between the peaks due to methyl methacrylate and methacrylic acid in the chromatogram

— the chromatogram obtained with the reference solution; not show peaks with the same retention times as methyl methacrylate or methacrylic acid.

Limit:

— sum of the contents of methyl methacrylate and methacrylic acid: maximum 0.1 per cent.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 1.000 g in a mixture of 40 mL of water R and 60 mL of 2-propanol R. Titrate slowly while stirring with 0.5 M sodium hydroxide, using phenolphthalein solution R as indicator.

Comparison Ph. Eur. reference spectrum of methacrylic acid -

methyl methacrylate copolymer (1.2).

B. It complies with the limits of the assay.

TESTS

Viscosity (2.2.10)

50 mPa.s to 200 mPa.s.

Dissolve a quantity of the substance to be examined

corresponding to 37.5 g of the dried substance in a mixture of 7.9 g of water R and 254.6 g of 2-propanol R. Determine the viscosity using a rotating viscometer at 20 °C and at a shear rate of 10 s⁻¹.

Appearance of a film

Place 1 mL of the solution prepared in the test for viscosity on a glass plate and allow to dry. A clear, brittle film is formed.

Methyl methacrylate and methacrylic acid

Liquid chromatography (2.2.29).

Blank solution To 50.0 mL of methanol R add 25.0 mL of the mobile phase.

Test solution Dissolve 40 mg of the substance to be examined in 50.0 mL of methanol R and add 25.0 mL of the mobile phase.

Reference solution Dissolve 10 mg of methacrylic acid R and 10 mg of methyl methacrylate R in methanol R, then dilute to 50.0 mL with the same solvent. Dilute 0.1 mL of this solution to 50.0 mL with methanol R and add 25.0 mL of the mobile phase.

Column: size: l = 0.10 m, Ø = 4 mm; stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase methanol R, phosphate buffer solution pH 2.0 R (30:70 V/V).

Flow rate 2.5 mL/min.

Detection Spectrophotometer at 202 nm.

Injection 50 µL.

System suitability:

— resolution: minimum 2.0 between the peaks due to methyl methacrylate and methacrylic acid in the chromatogram

— obtained with the reference solution;

— the chromatogram obtained with the blank solution does not show peaks with the same retention times as methyl methacrylate or methacrylic acid.

Limit:

— sum of the contents of methyl methacrylate and methacrylic acid: maximum 0.1 per cent.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 1.000 g in a mixture of 40 mL of water R and 60 mL of 2-propanol R. Titrate slowly while stirring with 0.5 M sodium hydroxide, using phenolphthalein solution R as indicator.

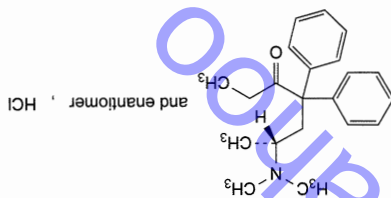
1 mL of 0.5 M sodium hydroxide is equivalent to 43.05 mg of C₁₁H₁₅O₂ (methacrylic acid units).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more

Methadone Hydrochloride

(Ph. Eur. monograph 0408)



C₂₁H₂₈ClNO

345.9

1095-90-5

Action and use

Opioid receptor agonist; analgesic.

Preparations

Methadone Injection

Methadone Linctus

Methadone Oral Solution (1 mg per ml)

Methadone Tablets

DEFINITION

(6R,S)-6-(Dimethylamino)-4,4-diphenylheptan-3-one hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Viscosity

(see Tests).

Appearance of a film

(see Tests).

Solubility of a film

Take a piece of the film obtained in the test for appearance of a film and place it in a flask containing a 10.3 g/L solution of hydrochloric acid R with stirring. It does not dissolve within 2 h. Take another piece of the film and place it in a flask containing phosphate buffer solution pH 6.8 R with stirring. It does not dissolve within 2 h. Take another piece of the film and place it in a flask containing 0.2 M phosphate buffer solution pH 7.5 R with stirring. It dissolves within 1 h.

Ph Eur



Solubility

Soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, C, D

Second identification A, B, D

A. Optical rotation (see Tests).

B. Melting point (2.2.14): 233 °C to 236 °C.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of methadone hydrochloride.

D. Dilute 1 mL of solution S (see Tests) to 5 mL with water R and add 1 mL of dilute ammonia R1. Mix, allow to stand for 5 min and filter. The filtrate gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

Dilute 10 mL of solution S to 25 mL with carbon dioxide-free water R. To 10 mL of the solution add 0.2 mL of methyl red solution R and 0.2 mL of 0.01 M sodium hydroxide.

The solution is yellow. Add 0.4 mL of 0.01 M hydrochloric acid. The solution is red.

Optical rotation (2.2.7)

–0.05° to +0.05°, determined on solution S in a 2 dm tube.

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 0.100 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with methanol R. Dilute 1.0 mL of this solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve 5 mg of impiramine hydrochloride CRS and 5 mg of cyclobenzaprine hydrochloride CRS in 100.0 mL of methanol R.

Column:

material: fused silica;

size: l = 50 m, Ø = 0.32 mm;

stationary phase: poly(dimethyl) (diphenyl)siloxane R (film thickness 1.05 µm).

Carrier gas helium for chromatography R.

Flow rate 1.2 mL/min.

Injection liner Packed with deactivated glass wool to wipe the needle.

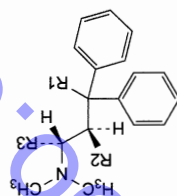
Split ratio 1:100.

Temperature:

Time (min)	Temperature (°C)
0 - 4	150 → 250
4 - 35	250
Injection port	200
Detector	250

Detection Flame ionisation.

Injection 2 µL.



Specified impurities A, B, C, D, E

IMPURITIES

Protected from light.

STORAGE

of C₂₁H₂₈ClNO.

1 mL of 0.1 M sodium hydroxide is equivalent to 34.59 mg of C₂₁H₂₈ClNO.

potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion. Carry out a blank titration.

Dissolve 0.300 g in a mixture of 5 mL of 0.01 M hydrochloric acid and 50 mL of anhydrous ethanol R. Carry out a

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

(0.05 per cent).

the chromatogram obtained with reference solution (a) — disregard limit: 0.5 times the area of the principal peak in (0.3 per cent);

in the chromatogram obtained with reference solution (a) — total: not more than 3 times the area of the principal peak with reference solution (a) (0.10 per cent);

area of the principal peak in the chromatogram obtained — unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— impurities A, B, C, D, E: for each impurity, not more than the area of the principal peak in the chromatogram

Limits:

impiramine and cyclobenzaprine.

— resolution: minimum 3.0 between the peaks due to System suitability: reference solution (b):

impiramine = about 1.19; cyclobenzaprine = about 1.24.

impurity D = about 0.98; impurity A = about 1.14;

impurity C = about 0.81; impurity B = about 0.89;

time = about 25 min): impurity E = about 0.44;

Relative retention With reference to methadone (retention time = about 1.5 times the retention time of methadone.

D. R1 = CO-C₂H₅, R2 = CH₃, R3 = H: (5RS)-6-(dimethylamino)-5-methyl-4,4-diphenylhexan-3-one (isomethadone),

(3RS)-4-(dimethylamino)-3-methyl-2,2-diphenylbutanenitrile (isodidavalol),

C. R1 = CN, R2 = CH₃, R3 = H:

(didavalol),

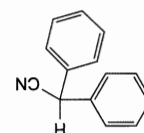
(4RS)-4-(dimethylamino)-2,2-diphenylpentanenitrile

B. R1 = CN, R2 = H, R3 = CH₃:

(isomethadone ketimine),

(2RS)-4-imino-N,N,2-trimethyl-3,3-diphenylhexan-1-amine

A. R1 = CNH-C₂H₅, R2 = CH₃, R3 = H:



E. diphenylacetoneitrile.

Ph Eur

Methane

(Ph. Eur. monograph 2413)

CH₄ 16.04 74-82-8



DEFINITION

Content
Minimum 99.5 per cent V/V of CH₄.

This monograph applies to methane obtained from natural gas and intended for medicinal use.

CHARACTERS

Appearance
Colourless gas, flammable in air.

Solubility

At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in about 27 volumes of water.

IDENTIFICATION

Examine the chromatograms obtained in the assay.
Results The principal peak in the chromatogram obtained with the gas to be examined is similar in retention time to the principal peak in the chromatogram obtained with reference gas (b).

TESTS

Nitrogen
Gas chromatography (2.2.28).
Gas to be examined The substance to be examined.
Reference gas Mixture containing 500 ppm V/V of nitrogen R1 in methane R1.

Column:
— material: stainless steel;
— size: l = 2 m, Ø = 2 mm;
— stationary phase: molecular sieve for chromatography R (0.5 nm).

Carrier gas helium for chromatography R.
Flow rate 30 mL/min.

Temperature:
— column: 80 °C;
— detector: 150 °C.
Detection Thermal conductivity.

Injection 1 mL.
Relative retention With reference to methane (retention time = about 3 min): nitrogen = about 0.7.
System suitability Reference gas:
— resolution: minimum 1.5 between the peaks due to nitrogen and methane.

Calculation of percentage content:
— use the concentration of nitrogen in the reference gas.
Limit:
— maximum 500 ppm V/V.

C₂-C₄ Hydrocarbons

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas Use the following mixture of gases in methane R: ethane R (impurity C) (100 ppm V/V), propane R (impurity D) (100 ppm V/V), i-butane R (impurity E) (100 ppm V/V) and n-butane R (impurity F) (100 ppm V/V).

Column:

— material: fused silica;
— size: l = 30 m, Ø = 0.53 mm;

— stationary phase: deactivated aluminium oxide for chromatography R.

Carrier gas helium for chromatography R.

Flow rate 113 mL/min.

Split ratio 1:10.

Temperature:

Time (min)	Temperature (°C)	Column	Detector
0 - 1	30	30 ÷ 150	300
1 - 3.4	150		
3.4 - 4.5			

Detection Flame ionisation. Use appropriate fuel gases: hydrocarbon-free air R and hydrogen for chromatography R.

Injection 0.1 mL.

Relative retention With reference to methane (retention time = about 1.2 min): impurity C = about 1.2; impurity D = about 1.8; impurity E = about 2.5; impurity F = about 2.6.

System suitability Reference gas:

— resolution: minimum 1.5 between the peaks due to methane and impurity C; minimum 1.5 between the peaks due to impurity E and impurity F.

Calculation of percentage contents:

— for each of the impurities C, D, E and F, use the corresponding concentration in the reference gas.

Limit:

— total (sum of impurities C, D, E and F): maximum 100 ppm V/V.

Water (2.5.28)

Maximum 10.0 ppm V/V.

ASSAY

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas (a) Methane R1.

Reference gas (b) Mixture containing 1 per cent V/V of nitrogen R1 in methane R1.

Column:

— material: stainless steel;

— size: l = 2 m, Ø = 2 mm;

— stationary phase: molecular sieve for chromatography R (0.5 nm).

Carrier gas helium for chromatography R.

Flow rate 30 mL/min.

Temperature:

— column: 80 °C;

— detector: 150 °C.

Detection Thermal conductivity.

Injection 10 µL.

Relative retention With reference to methane (retention time = about 3 min): nitrogen = about 0.7.

System suitability: reference gas (b):

— resolution: minimum 1.5 between the peaks due to nitrogen and methane.

Calculate the percentage content of CH₄ using the chromatogram obtained with reference gas (a).

STORAGE

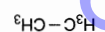
As a compressed gas, in appropriate containers complying with the legal regulations.

IMPURITIES

Specified impurities A, B, C, D, E, F

A, N₂: nitrogen,

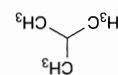
B, H₂O: water,



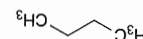
C, ethane,



D, propane,



E, 2-methylpropane (i-butane),



F, butane (n-butane).

Methanol

Methyl Alcohol

(Ph. Eur. monograph 1989)

DEFINITION

Methyl alcohol.

CHARACTERS

Appearance

Clear, colourless, volatile, hygroscopic liquid.

Solubility

Miscible with water and with methylene chloride.

bp

About 64 °C.

It is flammable.

IDENTIFICATION

A. Refractive index (2.2.6): 1.328 to 1.330.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of methanol.

$$\frac{A_2 - A_1}{2 \times A_1}$$

using the following expression:

Calculate the content of impurity A in parts per million V/V

— impurity A: maximum 2 ppm V/V.

Limit:

— impurity B (1st peak) and impurity C (2nd peak).

— resolution: minimum 4.0 between the peaks due to

System suitability: reference solution (a):

Injection 1 µL.

Detection Flame ionisation.

Temperature	Time	Column	Injection port	Detector
40	0 - 12	12 - 32	32 - 42	240
40		40 → 240		200
				280

Temperature:

Split ratio 1:20.

Linear velocity 35 cm/s.

Carrier gas helium for chromatography R.

thickness 1.8 µm).

poly[(cyanopropyl)(phenyl)]dimethylsiloxane R (film

— stationary phase:

— size: l = 30 m, Ø = 0.32 mm,

— material: fused silica,

Column:

solution (a).

Reference solution (b) Dilute 100 µL of benzene R to 100.0 mL

with test solution (a). Dilute 0.20 mL to 100.0 mL with test

test solution (a).

solution (a). Dilute 100 µL of this solution to 10.0 mL with

50 µL of acetone R and dilute to 50.0 mL with test

Reference solution (a) To 50 µL of anhydrous ethanol R add

to 100.0 mL with test solution (a).

50.0 mL with test solution (a). Dilute 5.0 mL of this solution

Test solution (b) Dilute 1.0 mL of 4-methylpentan-2-ol R to

Test solution (a) The substance to be examined.

Gas chromatography (2.2.28).

Impurity A

compensation liquid. The absorption curve is smooth.

Examine between 230 nm and 290 nm using water R as the

maximum 0.02 at 270 nm and maximum 0.01 at 290 nm.

Maximum 0.15 at 230 nm, maximum 0.05 at 250 nm,

Absorbance (2.2.25)

0.791 to 0.793.

Relative density (2.2.5)

required to change the colour of the indicator to pink.

Not more than 0.9 mL of 0.01 M sodium hydroxide is

phenolphthalein solution R1. The solution is colourless.

To 25 mL add 25 mL of water R and 0.25 mL of

Acidity or alkalinity

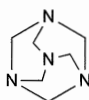
It is clear (2.2.1) and colourless (2.2.2, Method II).

Appearance

TESTS

Methenamine

(Ph. Eur. monograph 1545)



$C_6H_{12}N_4$

140.2

100-97-0

Action and use

Anti-infective.

Ph Eur

DEFINITION

1,3,5,7-Tetraazobicyclo[3.3.1.1^{3,7}.1^{3,7}]decane.

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification A

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison methenamine CRS.

B. To 1 mL of solution S (see Tests) add 1 mL of sulfuric acid R and immediately heat to boiling. Allow to cool.

To 1 mL of the solution add 4 mL of water R and 5 mL of acetylacetonate reagent R1. Heat on a water-bath for 5 min. An intense yellow colour develops.

C. To 1 mL of solution S add 1 mL of dilute sulfuric acid R and immediately heat to boiling. The solution gives the reaction of ammonium salts and salts of volatile bases (2.3.1). D. Dissolve 10 mg in 5 mL of water R and acidify with dilute hydrochloric acid R. Add 1 mL of potassium iodobismuthate solution R. An orange precipitate is formed immediately.

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 5 mL of solution S add 0.1 mL of phenolphthalein solution R. Not more than 0.2 mL of 0.1 M hydrochloric acid or 0.1 M sodium hydroxide is required to change the colour of the indicator.

Free formaldehyde

Maximum 50 ppm. Dissolve 0.8 g in water R and dilute to 8 mL with the same solvent. Add 2 mL of ammoniacal silver nitrate solution R. After 5 min, any grey colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner with a mixture of 8 mL of freshly prepared formaldehyde standard solution (5 ppm CH_2O) R and 2 mL of ammoniacal silver nitrate solution R.

Related substances
Gas chromatography (2.2.28) as described in the test for impurity A.

If necessary, the identity of impurity A can be confirmed using another suitable chromatographic system (stationary phase with a different polarity).

A_1 = area of the peak due to impurity A in the chromatogram obtained with test solution (a),
 A_2 = area of the peak due to impurity A in the chromatogram obtained with reference solution (b).

— **any impurity**: for each impurity, not more than the area of the peak due to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (0.1 per cent),

— **total**: not more than 3 times the area of the peak due to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (0.3 per cent),
— **disregard limit**: 0.05 times the area of the peak due to 4-methylpentan-2-ol in the chromatogram obtained with test solution (b) (50 ppm).

Reducing substances

To 20 mL add 0.1 mL of 0.02 M potassium permanganate. The pink colour is not completely discharged within 5 min.

Residue on evaporation

Maximum 10 ppm.

Evaporate 100 g to dryness on a water bath and dry in an oven at 100-105 °C. The residue weighs a maximum of 1 mg.

Water (2.5.12)

Maximum 0.10 per cent, determined on 10.0 g.

STORAGE

In an airtight container.

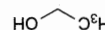
IMPURITIES

Specified impurities A

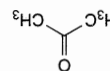
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C.



A. benzene,



B. ethanol,



C. propanone (acetone).

Ph Eur

Chlorides (2.4.4) Maximum 100 ppm. Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13) Maximum 100 ppm, determined on solution S.

Ammonium (2.4.1) Dilute 2 mL of freshly prepared solution S to 13 mL with water R. Add 2 mL of dilute sodium hydroxide solution R.

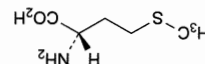
Heavy metals (2.4.8) Maximum 20 ppm. 12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32) Maximum 2.0 per cent, determined on 1.000 g by drying in a desiccator.

ASSAY Dissolve 0.100 g in 30 mL of methanol R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 14.02 mg of $C_5H_{12}N_2O_2S$.

STORAGE Protected from light.

(Ph. Eur. monograph 1027)



63-68-3

Action and use
Amino acid.

DEFINITION

Methionine contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (2S)-2-amino-4-(methylsulfonyl)butanoic acid, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, soluble in water, very slightly soluble in alcohol.

IDENTIFICATION

First identification A, B
Second identification A, C, D

A. Specific optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with methionine CRS. Examine the substances prepared as discs. C. Examine the chromatograms obtained in the test for ninhydrin-positive substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Solution S Dissolve 2.5 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

The pH of solution S is 5.5 to 6.5.

Specific optical rotation (2.2.7)

Dissolve 1.00 g in hydrochloric acid R1 and dilute to 50.0 mL with the same acid. The specific optical rotation is +22.5 to +24.0, calculated with reference to the dried substance.

Ninhydrin-positive substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel plate R.

Test solution (a) Dissolve 0.10 g of the substance to be examined in dilute hydrochloric acid R and dilute to 10 mL with the same acid.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with water R.

Reference solution (a) Dissolve 10 mg of methionine CRS in 10 g/L solution of hydrochloric acid R and dilute to 50 mL with the same acid solution.

Reference solution (b) Dilute 5 mL of test solution (b) to 20 mL with water R.

Reference solution (c) Dissolve 10 mg of methionine CRS and 10 mg of serine CRS in a 10 g/L solution of hydrochloric acid R and dilute to 25 mL with the same acid solution.

Apply separately to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 20 volumes of glacial acetic acid R, 20 volumes of water R and 60 volumes of butanol R. Allow the plate to dry in air, spray with ninhydrin solution R and heat at 100 °C to 105 °C for 15 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated spots.

Chlorides

To 10 mL of solution S add 25 mL of water R, 5 mL of dilute nitric acid R and 10 mL of silver nitrate solution R2.

Allow to stand protected from light for 5 min.

Any opalescence in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 10 mL of chloride standard solution (5 ppm Cl) R (200 ppm). Examine the tubes laterally against a black background.

Sulfates (2.4.13)

Dissolve 0.5 g in 3 mL of dilute hydrochloric acid R and dilute to 15 mL with distilled water R. The solution complies with the limit test for sulfates (300 ppm).

Ammonium (2.4.1)

0.10 g complies with limit test B for ammonium (200 ppm). Prepare the standard using 0.2 mL of ammonium standard solution (100 ppm NH_4) R.

Iron (2.4.9)

In a separating funnel, dissolve 1.0 g in 10 mL of dilute hydrochloric acid R. Shake with three quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min in each time. To the combined upper layers add 10 mL of water R and shake for 3 min. The lower layer complies with the limit test for iron (10 ppm).

Heavy metals (2.4.8)

2.0 g complies with test C for heavy metals (10 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.125 g in 5 mL of anhydrous formic acid R.

Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

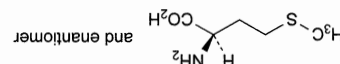
1 mL of 0.1 M perchloric acid is equivalent to 14.92 mg of $C_5H_{11}NO_2S$.

STORAGE

Store protected from light.

DL-Methionine

(Ph. Eur. monograph 0624)



59-51-8

Action and use

Used in treatment of paracetamol overdose.

DEFINITION

DL-Methionine contains not less than 99.0 per cent and more than the equivalent of 101.0 per cent of (2R,5)-2-amino-4-(methylsulfanyl)butanoic acid, calculated with reference to the dried substance.

CHARACTERS

Almost white, crystalline powder or small flakes, sparingly soluble in water, very slightly soluble in alcohol. It dissolves in dilute acids and in dilute solutions of the alkali hydroxides. It melts at about 270 °C (instantaneous method).

IDENTIFICATION

First identification A, C.
Second identification B, C, D.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with DL-methionine CRS. Dry the substances at 105 °C.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour

Chlorides

Dissolve 0.25 g in 35 mL of water R. Add 5 mL of dilute nitric acid R and 10 mL of silver nitrate solution R2. Allow to stand protected from light for 5 min. Any opalescence in the solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 10 mL of chloride standard solution (5 ppm Cl) R and 25 mL of water R (200 ppm). Examine the tubes laterally against a black background.

Sulfates (2.4.13)

Dissolve 1.0 g in 20 mL of distilled water R, heating to 60 °C. Cool to 10 °C and filter. 15 mL of the solution complies with the limit test for sulfates (200 ppm).

Heavy metals (2.4.8)

1.0 g complies with test D for heavy metals (20 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel G R as the coating substance.

Test solution (a) Dissolve 0.2 g in water R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL with water R.

Reference solution (a) Dissolve 20 mg of DL-methionine CRS in water R and dilute to 50 mL with the same solvent.

Reference solution (b) Dilute 1 mL of reference solution (a) to 10 mL with water R.

Apply separately to the plate 5 µL of each solution. Develop over a path of 10 cm using a mixture of 20 volumes of glacial acetic acid R, 20 volumes of water R and 60 volumes of butanol R. Allow the plate to dry in air and spray with ninhydrin solution R. Heat the plate at 100 °C to 105 °C for 15 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Chlorides

Dissolve 0.25 g in 35 mL of water R. Add 5 mL of dilute nitric acid R and 10 mL of silver nitrate solution R2. Allow to stand protected from light for 5 min. Any opalescence in the solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 10 mL of chloride standard solution (5 ppm Cl) R and 25 mL of water R (200 ppm). Examine the tubes laterally against a black background.

Sulfates (2.4.13)

Dissolve 1.0 g in 20 mL of distilled water R, heating to 60 °C. Cool to 10 °C and filter. 15 mL of the solution complies with the limit test for sulfates (200 ppm).

Heavy metals (2.4.8)

1.0 g complies with test D for heavy metals (20 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.140 g in 3 mL of anhydrous formic acid R. Add 30 mL of anhydrous acetic acid R. Immediately after dissolution, titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 14.92 mg of $C_5H_11NO_5$.

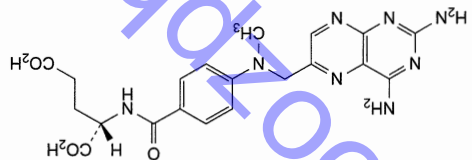
STORAGE

Store protected from light.

Ph Eur

Methotrexate

(Ph. Eur. monograph 0560)



59-05-2

Action and use

Dihydrofolate reductase inhibitor; cytostatic.

Preparations

Methotrexate Injection
Methotrexate Oral Solution
Methotrexate Tablets

DEFINITION

(2S)-2-[[4-[[[(2S)-2,4-diaminopteridin-6-yl)methyl]amino]benzoyl]amino]pentanedioic acid.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

Yellow or orange, crystalline, hygroscopic powder.

Solubility

Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides and carbonates.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison methotrexate CRS.

TESTS**Related substances**

Liquid chromatography (2.2.29). *Test solution* (b). Dissolve 25.0 mg of the substance to be examined in a mixture of 0.5 mL of dilute ammonia R1 and 5 mL of mobile phase A. Dilute 5.0 mL of this solution to 50.0 mL with mobile phase A. Dissolve 40.0 mg of the substance to be examined in a mixture of 0.5 mL of dilute ammonia R1 and 5 mL of mobile phase A and dilute to 100.0 mL with mobile phase A. *Test solution* (b). Dissolve 25.0 mg of the substance to be examined in a mixture of 0.5 mL of dilute ammonia R1 and 5 mL of mobile phase A and dilute to 50.0 mL with mobile phase A and dilute to 50.0 mL with mobile phase A.

phase A. Dilute 5.0 mL of this solution to 50.0 mL with

mobile phase A. *Reference solution* (a). Dissolve 25.0 mg of methotrexate CRS in a mixture of 0.5 mL of dilute ammonia R1 and 5 mL of mobile phase A and dilute to 50.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 50.0 mL with mobile

phase A. *Reference solution* (b). Dilute 5.0 mL of test solution (a) to 100.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 25.0 mL with mobile phase A.

Reference solution (d). Dissolve 5 mg of the substance to be examined, 5 mg of 4-aminobenzoic acid R (impurity B), 5 mg of methotrexate impurity C CRS, 5 mg of methotrexate impurity D CRS and 5 mg of methotrexate impurity E CRS in a mixture of 0.5 mL of dilute ammonia R1 and 5 mL of mobile phase A and dilute to 100 mL with mobile phase A.

Reference solution (e). Dissolve 8 mg of methotrexate for peak identification CRS (containing impurities H and I) in a mixture of 0.1 mL of dilute ammonia R1 and 1 mL of mobile phase A and dilute to 20 mL with mobile phase A.

Column: size: $l = 0.25$ m, $\phi = 4.0$ mm; stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase: mix 5 volumes of acetonitrile for chromatography R and 95 volumes of a 3.4 g/L solution of anhydrous sodium dihydrogen phosphate R previously adjusted to pH 6.0 with a 42 g/L solution of sodium hydroxide R; mobile phase B: mix 50 volumes of acetonitrile for chromatography R and 50 volumes of a 3.4 g/L solution of anhydrous sodium dihydrogen phosphate R previously adjusted to pH 6.0 with a 42 g/L solution of sodium hydroxide R;

adjusted to pH 6.0 with a 42 g/L solution of sodium hydroxide R; chromatography R and 50 volumes of a 3.4 g/L solution of anhydrous sodium dihydrogen phosphate R previously adjusted to pH 6.0 with a 42 g/L solution of sodium hydroxide R;

Time (min) Mobile phase A (per cent V/V) Mobile phase B (per cent V/V)

0 - 10 100 0
10 - 20 100 \rightarrow 95
20 - 28 95 \rightarrow 50
28 - 37 50 50

Detection Spectrophotometer at 280 nm. Injection 20 μ L of test solution (a) and reference solutions (b), (c), (d) and (e).

Identification of impurities Use the chromatogram supplied with methotrexate for peak identification CRS and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities H and I; use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities B, C and E.

Relative retention With reference to methotrexate (retention time = about 18 min): impurity B = about 0.3; impurity C = about 0.4; impurity D = about 0.9; impurity E = about 1.4; impurity I = about 1.5; impurity H = about 1.6.

System suitability: resolution: minimum 2.0 between the peaks due to impurities B and C and minimum 1.5 between the peaks

due to impurity D and methotrexate, in the chromatogram obtained with reference solution (d); minimum 1.5 between the peaks due to impurities I and H in the chromatogram obtained with reference solution (e); if the resolution between impurity D and methotrexate does not comply, increase the flow rate to meet the requirement.

Limits:

— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity E = 0.8; impurity I = 1.4;

— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *impurities B, E*: for each impurity, not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— *impurities H, I*: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent);

— *sum of impurities other than B, C and E*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.03 per cent).

Enantiomeric purity

Liquid chromatography (2.2.29).
Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.
Reference solution (b) Dissolve 4.0 mg of methotrexate for system suitability CRS (containing impurity F) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Column: size: 1 = 0.15 m, Ø = 4.0 mm; stationary phase: bovine albumin R bound to silica gel for chromatography R (7 µm) with a pore size of 30 nm. Mobile phase Add 500 mL of a 7.1 g/L solution of anhydrous disodium hydrogen phosphate R to 600 mL of a 6.9 g/L solution of sodium dihydrogen phosphate monohydrate R, mix, and adjust to pH 6.9 with dilute sodium hydroxide solution R; to 920 mL of this mixture add 80 mL of propanol R.

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 302 nm.
Injection 20 µL.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity F.
Relative retention With reference to methotrexate (retention time = about 4 min): impurity F = about 1.6.
System suitability: reference solution (b):
— resolution: minimum 2.0 between the peaks due to methotrexate and impurity F.

Limit:
— *impurity F*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3.0 per cent).

Heavy metals (2.4.8)

Maximum 50 ppm.
1.0 g complies with test C. Prepare the reference solution using 5 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 13.0 per cent, determined on 0.10 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

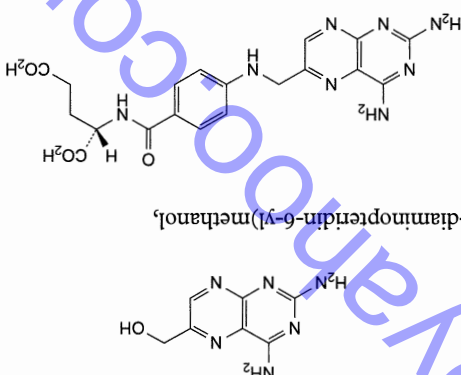
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection Test solution (b) and reference solution (a).
Calculate the percentage content of C₂₀H₂₂N₈O₅ taking into account the assigned content of methotrexate CRS.

STORAGE

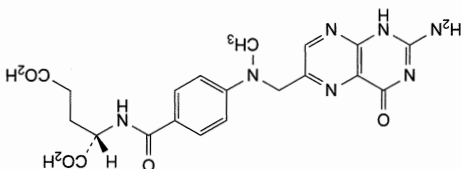
In an airtight container, protected from light.

IMPURITIES

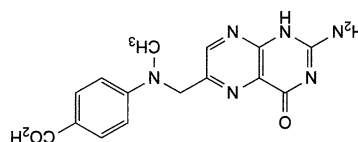
Specified impurities B, C, E, F, H, I
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D, G, J, K, L.



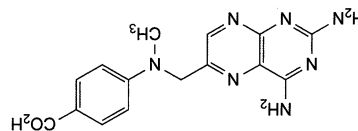
A. (2,4-diaminopteridin-6-yl)methanol,
B. (2S)-2-[[4-[[[(2,4-diaminopteridin-6-yl)methyl]amino]benzoyl]amino]pentanedioic acid (4-aminofolic acid),
aminopterin),



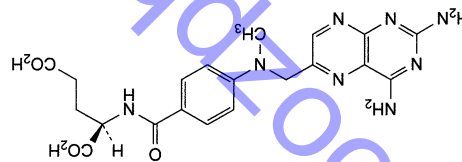
C. (2S)-2-[[4-[[[(2-oxo-1,4-dihydropyridin-6-yl)methyl]amino]benzoyl]amino]pentanedioic acid (N-methylfolic acid, methopterin),



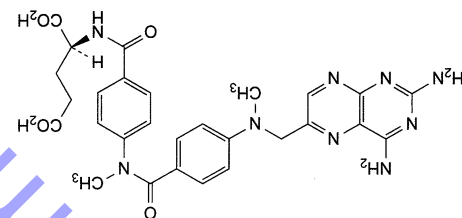
D. 4-[[[(2-amino-4-oxo-1,4-dihydropyridin-6-yl)methyl]methylamino]benzoic acid (*A*⁷¹⁰-methylpterioic acid),



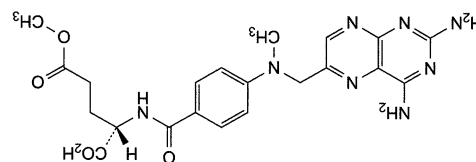
E. 4-[[[(2,4-diaminopyridin-6-yl)methyl]methylamino]benzoic acid (4-amino-*N*⁷-methylpterioic acid, APA),



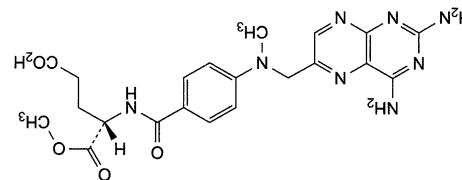
F. (2*R*)-2-[[4-[[[(2,4-diaminopyridin-6-yl)methyl]methylamino]benzoic acid ((*R*)-methotrexate),



G. (2*S*)-2-[[4-[[[(2,4-diaminopyridin-6-yl)methyl]methylamino]benzoic acid,

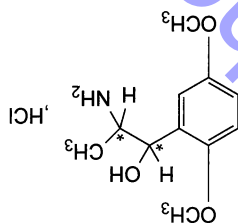


H. (2*S*)-2-[[4-[[[(2,4-diaminopyridin-6-yl)methyl]methylamino]benzoic acid (methotrexate 5-methyl ester),



I. (4*S*)-4-[[4-[[[(2,4-diaminopyridin-6-yl)methyl]methylamino]benzoic acid (methotrexate 1-methyl ester),

Methoxamine Hydrochloride



mixture of four stereoisomers

$C_{11}H_{17}NO_3 \cdot HCl$

247.7

61-16-5

Action and use

Adrenoceptor agonist.

Preparation

Methoxamine Injection

DEFINITION

Methoxamine Hydrochloride is all-*rac*-2-amino-1-(2,5-dimethoxyphenyl)propan-1-ol hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{11}H_{17}NO_3 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

Colourless crystals or white, plate-like crystals or a white, crystalline powder.
Freely soluble in *water*; soluble in *ethanol* (96%); very slightly soluble in *ether*.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of methoxamine hydrochloride (*RS* 220).

B. Yields the reactions characteristic of chlorides, Appendix VI.

Appendix VI.

TESTS

Acidity

pH of a 2% w/v solution, 4.0 to 6.0, Appendix V L.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using silica gel GF₂₅₄ as the coating substance and a mixture of 86 volumes of chloroform, 12 volumes of methanol and 2 volumes of 13.5M ammonia as the mobile phase. Apply separately to the plate 5 µL of each of three solutions in methanol containing (1) 2.0% w/v of the substance being examined, (2) 0.010% w/v of 2,5-dimethoxybenzaldehyde and (3) 0.020% w/v of the substance being examined. After removal of the plate, allow it to dry in air and examine under ultraviolet light (365 nm). Any spot corresponding to 2,5-dimethoxybenzaldehyde in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.5%). Spray the plate with a 0.5% w/v solution of ninhydrin in butan-1-ol containing 3% v/v of glacial acetic acid and heat at 105° for 5 minutes. Any other secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (3) (1%).

Loss on drying

When dried at 105° for 2 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

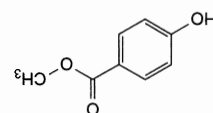
Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.5 g and 1-naphtholbenzenesulfonic acid as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 24.77 mg of C₁₁H₁₇NO₃·HCl.

Methyl Hydroxybenzoate



Methiparaben

(Methyl Parahydroxybenzoate, Ph Eur monograph 0409)



C₈H₈O₃ 152.1

99-76-3

Action and use

Antimicrobial preservative.

Ph Eur

DEFINITION

Methyl 4-hydroxybenzoate.

Content

98.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

First identification A, B

Second identification A, C

A. Melting point (2.2.14): 125 °C to 128 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g of the substance to be examined in acetone R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with acetone R.

Reference solution (a) Dissolve 10 mg of methyl parahydroxybenzoate CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of ethyl parahydroxybenzoate CRS in 1 mL of test solution (a) and dilute to 10 mL with acetone R.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R. Mobile phase glacial acetic acid R, water R, methanol R (1:30:70 V/V/V).

Application 2 µL of test solution (b) and reference solutions (a) and (b).

Development Over 2/3 of the plate. Drying In air.

Detection Examine in ultraviolet light at 254 nm. System suitability: reference solution (b): — the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 1.0 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Acidity

To 2 mL of solution S add 3 mL of ethanol (96 per cent) R, 5 mL of carbon dioxide-free water R and 0.1 mL of bromocresol green solution R. Not more than 0.1 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to blue.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of 4-hydroxybenzoic acid R (impurity A) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 50.0 mg of methyl parahydroxybenzoate CRS in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

— Mobile phase 6.8 g/L solution of potassium dihydrogen phosphate R, methanol R (35:65 V/V).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 10 μ L of the test solution and reference solutions (a) and (c).

Run time 5 times the retention time of methyl parahydroxybenzoate.

Relative retention With reference to methyl parahydroxybenzoate (retention time = about 2.3 min):

impurity A = about 0.6.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to impurity A and methyl parahydroxybenzoate.

Limits:

— peak area of impurity A by 1.4;

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_8H_8O_3$ from the declared content of methyl parahydroxybenzoate CRS.

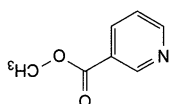
IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.

Methyl Nicotinate

(Ph. Eur. monograph 2129)



$C_7H_7NO_2$ 137.1

Action and use

Vasodilator.

Ph Eur

DEFINITION

Methyl pyridine-3-carboxylate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification B

Second identification A, C, D

A. Melting point (2.2.14): 40 °C to 42 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison methyl nicotinate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 2 mL with the same solvent.

Ph Eur

93-60-7



Reference solution Dissolve 10 mg of methyl nicotinate CRS in methanol R and dilute to 2 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase methanol R, toluene R (10:90 V/V).

Application 2 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 0.5 g add 0.1 g of citric acid R and 0.2 mL of acetic anhydride R. Heat cautiously for 1 min. A yellow colour is produced which turns first to orange, then to red and then to violet.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 25 mg of nicotinic acid R in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (e) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (f) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (g) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (h) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (i) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (j) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (k) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (l) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (m) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (n) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (o) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (p) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (q) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (r) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (s) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (t) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (u) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (v) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (w) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (x) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (y) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (z) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (aa) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ab) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ac) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ad) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ae) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (af) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ag) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ah) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ai) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (aj) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ak) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (al) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (am) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

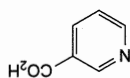
Reference solution (an) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ao) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (ap) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

IMPURITIES

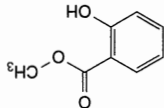
Specified impurities: A.



A. pyridine-3-carboxylic acid (nicotinic acid).

Methyl Salicylate

(Ph. Eur. monograph 0230)



152.1

C₈H₈O₃

Action and use

Counter-irritant.

Preparations

Methyl Salicylate Liniment

Methyl Salicylate Ointment

DEFINITION

Methyl 2-hydroxybenzoate.

Content

99.0 per cent m/m to 100.5 per cent m/m.

CHARACTERS

Appearance: colourless or slightly yellow liquid.

Solubility

Very slightly soluble in water, miscible with ethanol (96 per cent) and with fatty and essential oils.

IDENTIFICATION

A. Heat 0.25 mL with 2 mL of dilute sodium hydroxide solution R on a water-bath for 5 min. Add 3 mL of dilute sulfuric acid R. A crystalline precipitate is formed. Filter. The precipitate washed with water R and dried at 100-105 °C, melts (2.2.14) at 156 °C to 161 °C. B. To 10 mL of a saturated solution add 0.05 mL of ferric chloride solution R1. A violet colour develops.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

To 2 mL add 10 mL of ethanol (96 per cent) R.

Acidity

Dissolve 5.0 g in a mixture of 0.2 mL of bromocresol green

solution R and 50 mL of ethanol (96 per cent) R previously neutralised to a blue colour by addition of 0.1 M sodium

hydroxide. Not more than 0.4 mL of 0.1 M sodium hydroxide

is required to restore the blue colour.

Relative density (2.2.5)

1.180 to 1.186.

Refractive index (2.2.6)

1.535 to 1.538.

ASSAY

Dissolve 0.500 g in 25 mL of ethanol (96 per cent) R.

Add 0.05 mL of phenol red solution R and neutralise with

0.1 M sodium hydroxide. To the neutralised solution add

50.0 mL of 0.1 M sodium hydroxide and heat under a reflux

condenser on a water-bath for 30 min. Cool and titrate with

0.1 M hydrochloric acid. Calculate the volume of 0.1 M sodium

hydroxide used in the saponification. Carry out a blank

titration.

1 mL of 0.1 M sodium hydroxide is equivalent to 15.21 mg of

$C_8H_8O_3$.

STORAGE

Protected from light.

Industrial Methylated Spirit

Industrial Methylated Spirits; IMS

Industrial Denatured Alcohol

DEFINITION

Industrial Methylated Spirit is a mixture of nineteen volumes

of ethanol of an appropriate strength with one volume of

approved wood naphtha. Two strengths are available

containing 99% by volume and 95% by volume of alcohol

CHARACTERISTICS

A colourless, clear, mobile, volatile liquid, boiling at about

78°.

IDENTIFICATION

Mix 0.1 mL with 0.05 mL of an 11% w/w solution of

orthophosphoric acid and 0.25 mL of dilute potassium

permanganate solution. After 1 minute add a few mg of sodium

metabisulfite and shake until the mixture is decolorised.

Add 1.5 mL of a 50% v/v solution of sulfuric acid and a few

mg of finely powdered chromotropic acid sodium salt, shake

well and heat on a water bath for 5 minutes. A deep violet

TESTS**Acidity or alkalinity**

25 mL requires not more than 0.2 mL of 0.1 M sodium

hydroxide VS to produce a pink colour with phenolphthalein

solution R1 and not more than 1.0 mL of 0.1 M hydrochloric

acid VS to produce a red colour with methyl red solution.

Methylcellulose⁽¹⁾

(Ph. Eur. monograph 0345)

9004-67-5

Action and use

Bulk-forming laxative; excipient.

Preparation

Methylcellulose Tablets

Ph Eur

DEFINITION

Partly O-methylated cellulose. Methyl ether of cellulose.

Content

26.0 per cent to 33.0 per cent of methoxy groups ($-OCH_3$; 31.03) (dried substance).



Clarity of solution
Dilute 5.0 mL to 100 mL with water. The solution is clear.
Appendix IV A.

Apparent density
For '66 OP' grade, not greater than 811.6 kg m^{-3} , and for '74 OP' grade, not greater than 792.8 kg m^{-3} .
Appendix V G.

Aldehydes
To 5.0 mL add 5 mL of water and 1 mL of decolorised fuchsin solution and allow to stand for 30 minutes. Any colour produced is not more intense than that obtained by treating in the same manner 5 mL of a 0.005% w/v solution of redistilled acetaldehyde in aldehyde-free ethanol (96%) (50 ppm).

Non-volatile matter
When evaporated and dried at 105°, leaves not more than 0.01% w/v of residue.

Industrial Methylated Spirit (Ketone-free)

Industrial Denatured Alcohol (Ketone-free)

DEFINITION

Industrial Methylated Spirit (Ketone-free) is a mixture of nineteen volumes of ethanol of an appropriate strength with

one volume of approved wood naphtha substantially free

from ketones. Two strengths are available containing 99% by

volume and 95% by volume of alcohol (also known as 74 OP

and 66 OP respectively).

CHARACTERISTICS; IDENTIFICATION; TESTS

Acidity or alkalinity; Clarity of solution; Apparent

density; Aldehydes; Non-volatile matter

Complies with the requirements stated under Industrial

Methylated Spirit.

Ketones

Dilute 5 mL to 10 mL with water, add 1 mL of a 1.0% w/v

solution of 2-nitrobenzaldehyde in ethanol (50%) followed by

1 mL of a 15% w/v solution of sodium hydroxide in water and

allow to stand for 15 minutes. Any colour produced is not

more intense than that produced by treating in the same

manner 10 mL of a 0.025% v/v solution of acetone in

ethanol (50%) (500 ppm).

◆ CHARACTERS

Appearance

White, yellowish-white or greyish-white powder or granules, hygroscopic after drying.

Solubility

Practically insoluble in hot water, in acetone, in anhydrous ethanol and in toluene. It dissolves in cold water giving a colloidal solution. ◆

IDENTIFICATION

A. Evenly distribute 1.0 g onto the surface of 100 mL of water in a beaker, tapping the top of the beaker gently if necessary to ensure a uniform layer on the surface. Allow to stand for 1-2 min: the powdered material aggregates on the surface.

B. Evenly distribute 1.0 g into 100 mL of boiling water, stir the mixture using a magnetic stirrer with a bar 25 mm long: a slurry is formed and the particles do not dissolve. Allow the slurry to cool to 5 °C and stir using a magnetic stirrer: a clear or slightly turbid solution occurs with its thickness dependent on the viscosity grade.

C. To 0.1 mL of the solution obtained in identification test B add 9 mL of a 90 per cent *V/V* solution of *sulfuric acid*, shake, heat on a water-bath for exactly 3 min, immediately cool in an ice-bath, carefully add 0.6 mL of a 25 g/L solution of *minhydri*, shake and allow to stand at 25 °C: a red colour develops and does not change to purple within 100 min.

D. Place 2-3 mL of the solution obtained in identification test B on a glass slide as a thin film and allow the water to evaporate: a coherent, clear film forms on the glass slide. E. Add 50.0 mL of the solution obtained in identification test B to 50.0 mL of water in a beaker. Insert a thermometer into the solution. Stir the solution on a magnetic stirrer/hot plate and begin heating, increasing the temperature at a rate of 2-5 °C per minute. Determine the temperature at which a turbidity increase begins to occur and designate the temperature as the flocculation temperature: the flocculation temperature is higher than 50 °C.

TESTS

◆ Appearance of solution

The solution is not more opalescent than reference suspension III (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

While stirring, introduce a quantity of the substance to be examined equivalent to 1.0 g of the dried substance into 50 g of carbon dioxide-free water heated to 90 °C. Allow to cool, adjust the mass of the solution to 100 g with carbon dioxide-free water and stir until dissolution is complete. Allow to stand at 2-8 °C for 1 h before carrying out the test. ◆

pH (2.2.3)

5.0 to 8.0 for the solution prepared as described under Viscosity.

Read the pH after the probe has been immersed for 5 ± 0.5 min.

Viscosity

80 per cent to 120 per cent of the nominal value for samples with a viscosity of less than 600 mPa.s (Method I); 75 per cent to 140 per cent of the nominal value for samples with a viscosity of 600 mPa.s or higher (Method 2). Method 1, to be applied to samples with a viscosity of less than 600 mPa.s: Weigh a quantity of the substance to be examined equivalent to 4.000 g of the dried substance. Transfer into a wide-mouthed bottle, and adjust the total mass of the sample

Nominal viscosity* (mPa.s)	Rotor number	Revolution (r/min)	Calculation multiplier
----------------------------	--------------	--------------------	------------------------

Table 0345-1.

600 to less than 1400	3	60	20
1400 to less than 3500	3	12	100
3500 to less than 9500	4	60	100
9500 to less than 99 500	4	6	1000
99 500 or more	4	3	2000

*the nominal viscosity is based on the manufacturer's specifications.

and the water to 200.0 g with hot water. Capping the bottle, stir by mechanical means at 400 ± 50 r/min for 10-20 min until the particles are thoroughly dispersed and wetted. Scrape down the inside of the bottle with a spatula if necessary, to ensure that there is no undissolved material on the sides of the bottle, and continue the stirring in a cooling water-bath maintained at a temperature below 5 °C for another 20-40 min. Adjust the solution mass if necessary to 200.0 g using cold water. Centrifuge the solution if necessary to expel any entrapped air bubbles. Using a spatula, remove any foam. Determine the kinematic viscosity (ν) of this solution using the capillary viscometer method (2.2.9). Separately, determine the density (ρ) (2.2.5) of the solution and calculate the dynamic viscosity (η), as $\eta = \rho \nu$.

Method 2, to be applied to samples with a viscosity of 600 mPa.s or higher: Weigh a quantity of the substance to be examined equivalent to 10.00 g of the dried substance. Transfer into a wide-mouthed bottle, and adjust the total mass of the sample and the water to 500.0 g with hot water. Capping the bottle, stir by mechanical means at 400 ± 50 r/min for 10-20 min until the particles are thoroughly dispersed and wetted. Scrape down the inside of the bottle with a spatula if necessary, to ensure that there is no undissolved material on the sides of the bottle, and continue the stirring in a cooling water-bath maintained at a temperature below 5 °C for another 20-40 min. Adjust the solution mass if necessary to 500.0 g using cold water. Centrifuge the solution if necessary to expel any entrapped air bubbles. Using a spatula, remove any foam. Determine the viscosity (2.2.10) of this solution at 20 ± 0.1 °C using a rotating viscometer. Apparatus: Single-cylinder type spindle viscometer. Apply the rotor number, revolution and calculation multiplier. Apply the conditions specified in Table 0345-1.

Allow the spindle to rotate for 2 min before taking the measurement. Allow a rest period of at least 2 min between subsequent measurements. Repeat the measurement twice and determine the mean of the 3 readings.

Heavy metals (2.4.8)
Maximum 20 ppm.
1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

Sulfated ash (2.4.14)
Maximum 1.5 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (2.2.28).

Apparatus:

— **reaction vial:** a 5 mL pressure-tight vial, 50 mm in height,

20 mm in external diameter and 13 mm in internal

diameter at the mouth, equipped with a pressure-tight

butyl rubber membrane stopper coated with

polytetrafluoroethylene and secured with an aluminium

crimped cap or another sealing system providing a

sufficient air-tightness;

— **heater:** a heating module with a square aluminium block

having holes 20 mm in diameter and 32 mm in depth, so

that the reaction vials fit; mixing of the contents of the

vial is effected using a magnetic stirrer equipped in the

heating module or using a reciprocal shaker that performs

approximately 100 cycles/min.

Internal standard solution 30 g/L solution of *octane R* in

o-xylene *R*.

Test solution Weigh 65.0 mg of the substance to be examined,

place in a reaction vial, add 0.06–0.10 g of *adipic acid R*,

2.0 mL of the internal standard solution and 2.0 mL of

hydrotic acid R, immediately cap and seal the vial, and weigh

accurately. Mix the contents of the vial continuously for

60 min while heating the block so that the temperature of the

contents is maintained at $130 \pm 2^\circ\text{C}$. If a reciprocal shaker

or magnetic stirrer cannot be used, shake the vial thoroughly

by hand at 5 min intervals during the initial 30 min of the

heating time. Allow the vial to cool, and again weigh

accurately. If the loss of mass is less than 0.50 per cent of the

contents and there is no evidence of a leak, use the upper

layer of the mixture as the test solution.

Reference solution Place 0.06–0.10 g of *adipic acid R*, 2.0 mL

of the internal standard solution and 2.0 mL of *hydrotic*

acid R in another reaction vial, cap and seal the vial, and

weigh accurately. Add 45 μL of *methyl iodide R* through the

septum with a syringe, and weigh accurately. Shake the

reaction vial thoroughly and use the upper layer as the

reference solution.

Column:

— **size:** $l = 1.8\text{--}3\text{ m}$, $\varnothing = 3\text{--}4\text{ mm}$;

— **stationary phase:** *diatomaceous earth* for gas

chromatography R (125–150 μm) impregnated with

10–20 per cent of *poly(dimethyl)siloxane R*;

— **temperature:** 100 $^\circ\text{C}$.

Carrier gas *helium* for *chromatography R* or *nitrogen* for

chromatography R (flame ionisation); *helium* for

chromatography R (thermal conductivity).

Flow rate Adjusted so that the retention time of the internal

standard is about 10 min.

Detection Flame ionisation or thermal conductivity.

Injection 1–2 μL .

System suitability: reference solution.

— **resolution:** well-resolved peaks due to methyl iodide (1st

peak) and the internal standard (2nd peak).

Calculate the ratio (Q) of the area of the peak due to methyl

iodide to the area of the peak due to the internal standard in

the chromatogram obtained with the test solution, and the

ratio (Q_1) of the area of the peak due to methyl iodide to the

area of the peak due to the internal standard in the

chromatogram obtained with the reference solution.

Calculate the percentage content of methoxy groups using

the following expression:

$$\frac{Q \times m_1}{Q_1 \times m} \times 21.864$$

m_1 = mass of methyl iodide in the reference solution, in

milligrams;

m = mass of the sample (dried substance), in

milligrams.

LABELLING

The label states the viscosity in millipascal seconds.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). Some of the characteristics described in the Functionality-

related characteristics section may also be present in the mandatory

part of the monograph since they also represent mandatory quality

criteria. In such cases, a cross-reference to the tests described in the

mandatory part is included in the Functionality-related

characteristics section. Control of the characteristics can contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for methylcellulose

used as binder, viscosity-enhancing agent or film former.

Viscosity

See Tests.

Degree of substitution

See Assay.

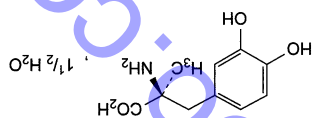
Ph Eur

¹ This monograph has undergone pharmacopoeial harmonisation.

See chapter 5.8 Pharmacopoeial Harmonisation.

Methylidopa

(*Ph. Eur. monograph 0045*)



$\text{C}_{10}\text{H}_{13}\text{NO}_4 \cdot 1\frac{1}{2}\text{H}_2\text{O}$ 238.2 41372-08-1

Action and use

α -Adrenoceptor agonist; treatment of hypertension.

Preparation

Methylidopa Tablets

Ph Eur

DEFINITION

(2*S*)-2-Amino-3-(3,4-dihydroxyphenyl)-2-methylpropanoic acid sesquihydrate (1-methylidopa sesquihydrate).

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or yellowish-white, crystalline powder or colourless or almost colourless crystals.

Solubility

Slightly soluble in water, very slightly soluble in ethanol (96 per cent). It is freely soluble in dilute mineral acids.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Carry out either tests A, B or tests A, C.

Comparison methyldopa CRS.

B. Enantiomeric purity (see Tests).

C. Specific optical rotation (2.2.7): -28.0 to -25.0 .

Dissolve a quantity equivalent to 2.20 g of the anhydrous substance in aluminium chloride solution R and dilute to

50.0 mL with the same solution.

TESTS**Appearance of solution**

Dissolve 1.0 g in 1 M hydrochloric acid and dilute to 25 mL with the same solvent. The solution is not more intensely coloured than reference solution BY₆ or B₆ (2.2.2).

Method II.**Acidity**

Dissolve 1.0 g with heating in 100 mL of carbon dioxide-free water R. Add 0.1 mL of methyl red solution R. Not more than 0.5 mL of 0.1 M sodium hydroxide is required to produce the pure yellow colour of the indicator.

Absorbance (2.2.25)

Test solution Dissolve 40.0 mg in 0.1 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of the solution to 100.0 mL with 0.1 M hydrochloric acid.

Spectral range 230 – 350 nm.

Absorption maximum At 280 nm.

(anhydrous substance).

Enantiomeric purity

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

phase.

Reference solution (a) Dilute 5.0 mL of the test solution to

20.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of racemic

methyldopa CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\phi = 3.9$ mm;

— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve separately 0.200 g of copper acetate R

and 0.387 g of *N,N*-dimethyl-1-phenylalanine R in water R;

mix the 2 solutions and adjust immediately to pH 4.3 with

acetic acid R; add 50 mL of methanol R and dilute to

1000 mL with water R; mix and filter.

Equilibrate the column with the mobile phase for about 2 h.

If necessary, decrease the concentration of methanol R so the

peak corresponding to D-methyldopa is clearly separated from

the negative system peak that appears at about 6 min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Run time Twice the retention time of L-methyldopa.

Relative retention With reference to L-methyldopa (retention

time = about 14 min): D-methyldopa = about 0.7 .

Limit:

— resolution: minimum 5.0 between the peaks due to

D-methyldopa and L-methyldopa.

— D-methyldopa (impurity D): not more than the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.5 per cent).

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Test solution Dissolve 0.100 g of the substance to be

examined in 0.1 M hydrochloric acid and dilute to 25.0 mL

with the same acid.

Reference solution (a) Dilute 1.0 mL of the test solution to

50.0 mL with 0.1 M hydrochloric acid. Dilute 5.0 mL of this

solution to 100.0 mL with 0.1 M hydrochloric acid.

Reference solution (b) Dissolve the contents of a vial of

methyldopa for system suitability CRS (containing

impurities A, B and C) in 1.0 mL of 0.1 M hydrochloric acid.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: spherical di-isobutyloctadecylsilyl silica gel

for chromatography R (5 μ m) with a pore size of 8 nm.

Mobile phase methanol R, 0.1 M phosphate buffer solution

pH 3.0 R (15:85 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Run time 6 times the retention time of methyldopa.

Identification of impurities Use the chromatogram supplied

with methyldopa for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B and C.

Relative retention With reference to methyldopa (retention

time = about 5 min): impurity A = about 1.9 ;

impurity B = about 4.3 ; impurity C = about 4.9 .

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurities B and C.

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity B = 2.6 ;

impurity C = 1.3 ;

impurities A, B, C: for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.05 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.03 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

1.0 g complies with test F. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

10.0 per cent to 13.0 per cent, determined on 0.20 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.180 g, heating if necessary, in 50 mL of glacial acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 21.12 mg of

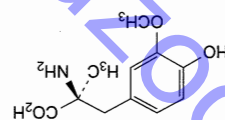
$C_{12}H_{13}NO_4$.

STORAGE

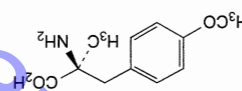
Protected from light.

IMPURITIES

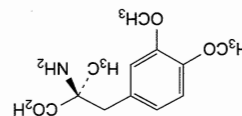
Specified impurities A, B, C, D



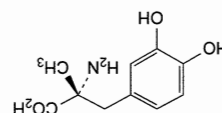
A. (2S)-2-amino-3-(4-hydroxy-3-methoxyphenyl)-2-methylpropanoic acid (3-methoxymethyldopa),



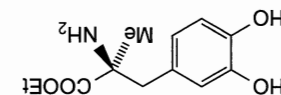
B. (2S)-2-amino-3-(4-methoxyphenyl)-2-methylpropanoic acid,



C. (2S)-2-amino-3-(3,4-dimethoxyphenyl)-2-methylpropanoic acid,



D. (2R)-2-amino-3-(3,4-dihydroxyphenyl)-2-methylpropanoic acid (D-methyldopa).



$C_{12}H_{17}NO_4 \cdot HCl$

275.7

2508-79-4

Action and use

Alpha₂-adrenoceptor agonist; treatment of hypertension.

Preparation

Methyldopate Injection

DEFINITION

Methyldopate Hydrochloride is ethyl 3-(3,4-

dihydroxyphenyl)-2-methyl-L-alaninate hydrochloride.

It contains not less than 98.5% and not more than 101.0% of $C_{12}H_{17}NO_4 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder.

Freely soluble in water, in ethanol (96%) and in methanol;

practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of methyldopate hydrochloride (RS 224).

B. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.008% w/v solution in 0.1M hydrochloric acid exhibits a maximum only at 280 nm. The absorbance at the maximum at 280 nm is about 0.80.

C. Yields reaction A characteristic of chlorides, Appendix VI.

TESTS**Acidity**

pH of a 1% w/v solution, 3.0 to 5.0, Appendix V L.

Specific optical rotation

In a 4% w/v solution in 0.1M hydrochloric acid and determined at 405 nm, -13.5 to -14.9, calculated with reference to the dried substance, Appendix V F.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in methanol.

(1) 10.0% w/v of the substance being examined.

(2) 0.25% w/v of methyldopa BPCRS.

(3) 0.040% w/v of methyldopa BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel GF₂₅₄.
(b) Use the mobile phase as described below.
(c) Apply 2 µL of each solution.
(d) Develop the plate to 12 cm.

(e) After removal of the plate, dry in air, heat at 105° for 10 minutes and examine under ultraviolet light (254 nm).
Expose the plate to iodine vapour for 10 minutes and examine again.

MOBILE PHASE

Equal volumes of acetone, butan-1-ol, glacial acetic acid, toluene and water.

LIMITS

By each method of visualisation any spot corresponding to methyldopa in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (2.5%) and any other secondary spot is not more intense than the spot in the chromatogram obtained with solution (3) (0.4%).

Loss on drying

When dried to constant weight at 105° at a pressure not exceeding 0.7 kPa, loses not more than 0.5% of its weight.

Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for non-aqueous titration,

Appendix VIII A, using 0.5 g and determining the end point

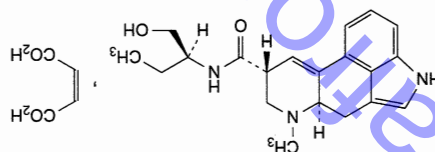
potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 27.57 mg of $C_{12}H_{17}NO_4 \cdot HCl$.

STORAGE

Methylethergometrine Hydrochloride should be protected from light.

Methylethergometrine Maleate

(Ph. Eur. monograph 1788)



$C_{24}H_{29}N_3O_6$ 455.5 57432-61-8

Action and use

Oxytocic.

Ph. Eur.

DEFINITION

(6aR,9R)-N-[(1S)-1-(Hydroxymethyl)propyl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-fg]quinoline-9-carboxamide (Z)-butenedioate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Soluble in water, slightly soluble in anhydrous ethanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison methylethergometrine maleate CRS.

TESTS

Solution S

Dissolve 0.100 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

pH (2.2.3)

4.4 to 5.2.

Dilute 2.0 mL of solution S to 50.0 mL with carbon dioxide-free water R.

Specific optical rotation (2.2.7)

+44.0 to +50.0 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Test solution Dissolve 25 mg of the substance to be examined in 15 mL of mobile phase B and dilute to 50.0 mL with water R.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with water R. Dilute 1.0 mL of this solution to 10.0 mL with water R.

Reference solution (b) Dissolve the contents of a vial of methylethergometrine for system suitability CRS (containing impurities A, B, C, D, E, F, G, H and I) in 1.0 mL of a

mixture of 30 volumes of mobile phase B and 70 volumes of water R.
Column:
— size: $l = 0.10$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3.5 μ m).
Mobile phase:
— mobile phase A: 2 g/L solution of ammonium carbonate R;
— mobile phase B: acetonitrile R, water R (50:50 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	85	15
2 - 7	85 \rightarrow 65	15 \rightarrow 35
7 - 12	65	35
12 - 17	65 \rightarrow 20	35 \rightarrow 80
17 - 19	20	80

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 310 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied with methylethergometrine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D, E, F, G, H and I.

Relative retention With reference to methylethergometrine (retention time = about 12 min): impurity A = about 0.2;

impurity B = about 0.5; impurity C = about 0.6;

impurity D = about 0.7; impurity I = about 1.10;

impurity E = about 1.14; impurity F = about 1.2;

impurity G = about 1.3; impurity H = about 1.4.

System suitability: reference solution (b):

— resolution: minimum 3.0 between the peaks due to methylethergometrine and impurity I; minimum 1.5 between the peaks due to impurities I and E.

Limits:

— impurity I: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.2 per cent);

— impurities A, B, D, E, F, G, H: for each impurity, not more than 1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

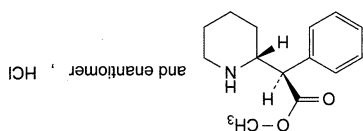
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.



Methylphenidate Hydrochloride

(Ph. Eur. monograph 2235)



$C_{14}H_{20}ClNO_2$ 269.8 298-59-9

Action and use

Narcolepsy; hyperactivity disorder in children.

Ph Eur

DEFINITION

Methyl (2*R*)-piperidin-2-yl]acetate hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, fine, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison methylphenidate hydrochloride CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in 1.0 mL of methanol R.

Reference solution Dissolve 5 mg of methylphenidate hydrochloride CRS in 1.0 mL of methanol R.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, methanol R, methylene chloride R (1:4:95 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 60 °C for 5 min.

Detection Spray with a freshly prepared 5 g/L solution of fast blue B salt R, heat to 60 °C for 1 min.

Result The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot obtained with the reference solution.

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture Mix 20 volumes of acetonitrile R1 and 80 volumes of a solution prepared as follows: dissolve 1.36 g of sodium octanesulfonate R in 950 mL of water for chromatography R, add 1.0 mL of methylamine R2, adjust to pH 2.7 with phosphoric acid R and dilute to 1000 mL with water for chromatography R.

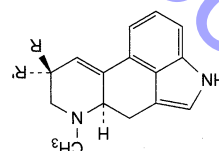
IMPURITIES

In an airtight container, protected from light.

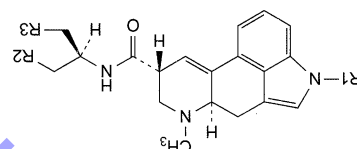
STORAGE

of $C_{14}H_{20}NO_2$.
1 mL of 0.1 M perchloric acid is equivalent to 45.55 mg
potentiometrically (2.2.20).

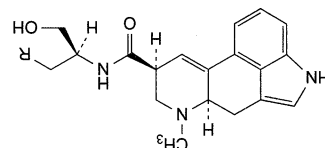
Dissolve 0.300 g in 60 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point



A. R = H, R' = CO₂H; (6*a*R,9*R*)-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxylic acid, B. R = CO₂H, R' = H; (6*a*R,9*S*)-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxylic acid, C. R = H, R' = CONH₂; (6*a*R,9*R*)-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide, E. R = CONH₂, R' = H; (6*a*R,9*S*)-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide,



D. R1 = R2 = R3 = OH; (6*a*R,9*R*)-*N*-[(1*S*)-2-hydroxy-1-methylethyl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergometrine), G. R1 = R2 = R3 = OH; (6*a*R,9*R*)-*N*-[(1*S*)-1-(hydroxymethyl)propyl]-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (methysergide), I. R1 = H, R2 = OH, R3 = CH₃; (6*a*R,9*R*)-*N*-[(1*R*)-1-(hydroxymethyl)propyl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (1'-*epi*-methylethergometrine),



F. R = H; (6*a*R,9*S*)-*N*-[(1*S*)-2-hydroxy-1-methylethyl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (ergometrine), H. R = CH₃; (6*a*R,9*S*)-*N*-[(1*S*)-1-(hydroxymethyl)propyl]-7-methyl-4,6,6a,7,8,9-hexahydroindolo[4,3-*fg*]quinoline-9-carboxamide (methylethergometrine).

Ph Eur

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of methyphenidate impurity C CRS in 100.0 mL of the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of methyphenidate impurity mixture CRS (impurities A and B) in 1.0 mL of reference solution (a).

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.075$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3.5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: dissolve 2.16 g of sodium octanesulfonate R in 950 mL of water for chromatography R, add 1.0 mL of triethylamine R₂, adjust to pH 2.7 with phosphoric acid R and dilute to 1000 mL with water for chromatography R;

— mobile phase B: acetonitrile R₁;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	80	20
15 - 35	80 → 60	20 → 40

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied with methyphenidate impurity mixture CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C.

Relative retention With reference to methyphenidate (retention time = about 20 min): impurity A = about 0.35; impurity C = about 0.40; impurity B = about 0.6.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to impurities A and C.

Limits:

— impurities A, B: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— total: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

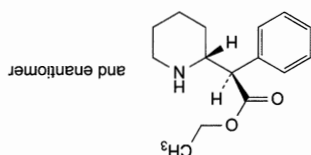
Heavy metals (2.4.8)

Maximum 20 ppm.

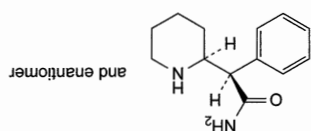
Dissolve 1.0 g in water R and dilute to 20.0 mL with the same solvent. 12 mL of the solution complies with test A.

Prepare the reference solution using lead standard solution (1 ppm Pb) R.

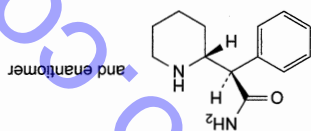
E. ethyl (2*RS*)-phenyl[(2*RS*)-piperidin-2-yl]acetate,



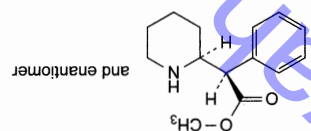
D. (2*RS*)-2-phenyl-2-[(2*SR*)-piperidin-2-yl]acetamide,



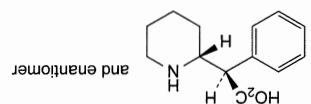
C. (2*RS*)-2-phenyl-2-[(2*RS*)-piperidin-2-yl]acetamide,



B. methyl (2*RS*)-phenyl[(2*SR*)-piperidin-2-yl]acetate,



A. (2*RS*)-phenyl[(2*RS*)-piperidin-2-yl]acetic acid,



E, F.

Control of impurities in substances for pharmaceutical use: C, D, impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these by the general monograph *Substances for pharmaceutical use* acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of other detectable impurities (the following substances would, if specified impurities A, B

IMPURITIES

Protected from light.

STORAGE

$C_{14}H_{20}ClNO_2$.

1 mL of 0.1 M sodium hydroxide is equivalent to 26.98 mg of the volume added between the 2 points of inflexion.

and an electrode for non-aqueous acid-base titrations. Read potentiometric titration (2.2.20) using 0.1 M sodium hydroxide 5.0 mL of 0.01 M hydrochloric acid. Carry out a

Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R and add

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

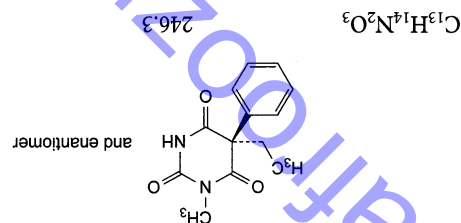
in vacuo at 60 °C for 4 h.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

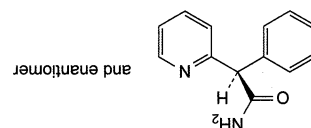
Methyphenobarbital

(Ph. Eur. monograph 0189)



Ph Eur

F. (2*RS*)-2-phenyl-2-(pyridin-2-yl)acetamide.



Action and use

Barbiturate.

Preparation

Methyphenobarbital Tablets

Ph Eur

DEFINITION

(5*RS*)-5-Ethyl-1-methyl-5-phenylpyrimidine-2,4,6-(1*H*,3*H*,5*H*)-trione.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals.

Solubility

Practically insoluble in water, very slightly soluble in ethanol

(96 per cent).

It forms water-soluble compounds with alkali hydroxides and

carbonates and with ammonia.

IDENTIFICATION

First identification A, B

Second identification A, C, D.

A. Determine the melting point (2.2.14) of the substance to

be examined. Mix equal parts of the substance to be

examined and methyphenobarbital CRS and determine the

melting point of the mixture. The difference between the

melting points (which are about 178 °C) is not greater than

2 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison methyphenobarbital CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in methylene chloride R and dilute to 10 mL with the same

solvent.

Reference solution Dissolve 10 mg of methyphenobarbital CRS

in methylene chloride R and dilute to 10 mL with the same

solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase concentrated ammonia R, ethanol (96 per cent) R,

methylene chloride R (5:15:80 V/V/V); use the lower layer.

Application 10 µL.

Development Over 2/3 of the plate.

Detection Examine immediately in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

D. To about 10 mg add 0.2 mL of sulfuric acid R and

0.1 mL of nitric acid R. Heat on a water-bath for 10 min.

Cool in iced water and add 5 mL of water R and 5 mL of

strong sodium hydroxide solution R. Add 5 mL of acetone R,

shake and allow to stand. A dark-red colour develops in the

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured

than reference solution Y₆ (2.2.2, Method II).

Dissolve 1.0 g, with gentle heating, in a mixture of 4 mL of

dilute sodium hydroxide solution R and 6 mL of water R.

Acidity

Boil 1.0 g with 50 mL of water R for 2 min, allow to cool

and filter. To 10 mL of the filtrate add 0.15 mL of methyl red

solution R. The solution is orange-yellow. Not more than

0.1 mL of 0.1 M sodium hydroxide is required to produce a

pure yellow colour.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined

in 10.0 mL of methanol R and dilute to 50.0 mL with the

mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 5.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of phenobarbital CRS

(impurity A) in 1.0 mL of methanol R and dilute to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the test solution.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 µm).

Mobile phase Dissolve 6.60 g of sodium acetate R in 900 mL of

water R, add 3 mL of glacial acetic acid R, adjust to pH 4.5

with glacial acetic acid R and dilute to 1000 mL with water R.

Mix 40 volumes of this solution with 60 volumes of

methanol R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 3.5 times the retention time of methyphenobarbital.

Identification of impurities Use the chromatogram obtained

with reference solution (b) to identify the peak due to

impurity A.

Relative retention With reference to methyphenobarbital

(retention time = about 7 min): impurity A = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to

impurity A and methyphenobarbital.

Limits:

— impurity A: not more than the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in ethanol (96 per cent), slightly soluble in acetone and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

First identification A, B

Second identification C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison methylprednisolone CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in the minimum volume of acetone R,

evaporate to dryness on a water-bath and record new spectra

using the residues.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained

with the test solution is similar in retention time and size to

the principal peak in the chromatogram obtained with

reference solution (c).

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 10 mg of the substance to be

examined in methanol R and dilute to 2 mL with the same

solvent (solution A). Dilute 1 mL of solution A to 5 mL with

methylene chloride R.

Test solution (b) Transfer 0.4 mL of solution A to a glass tube

100 mm long and 20 mm in diameter and fitted with a

ground-glass stopper or a polytetrafluoroethylene cap and

evaporate the solvent with gentle heating under a stream of

nitrogen R. Add 2 mL of a 15 per cent V/V solution of glacial

acetic acid R and 50 mg of sodium bismuthate R. Stopper the

tube and shake the suspension in a mechanical shaker

protected from light for 1 h. Add 2 mL of a 15 per cent V/V

solution of glacial acetic acid R and filter into a 50 mL

separating funnel, washing the filter with 2 quantities, each of

5 mL, of water R. Shake the clear filtrate with 10 mL of

methylene chloride R. Wash the organic layer with 5 mL of

1 M sodium hydroxide and 2 quantities, each of 5 mL, of

water R. Dry over anhydrous sodium sulfate R.

Reference solution (a) Dissolve 10 mg of

methylprednisolone CRS in methanol R and dilute to 2 mL with

the same solvent (solution B). Dilute 1 mL of solution B to

5 mL with methylene chloride R.

Reference solution (b) Transfer 0.4 mL of solution B to a glass

tube 100 mm long and 20 mm in diameter and fitted with a

ground-glass stopper or a polytetrafluoroethylene cap and

evaporate the solvent with gentle heating under a stream of

nitrogen R. Add 2 mL of a 15 per cent V/V solution of glacial

acetic acid R and 50 mg of sodium bismuthate R. Stopper the

tube and shake the suspension in a mechanical shaker

protected from light for 1 h. Add 2 mL of a 15 per cent V/V

solution of glacial acetic acid R and filter into a 50 mL

separating funnel, washing the filter with 2 quantities, each of

5 mL, of water R. Shake the clear filtrate with 10 mL of

methylene chloride R. Wash the organic layer with 5 mL of

1 M sodium hydroxide and 2 quantities, each of 5 mL, of

water R. Dry over anhydrous sodium sulfate R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase butanol R saturated with water R, toluene R,

ether R (5:10:85 V/V/V).

— unspecified impurities: for each impurity, not more than

0.2 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent);

— total: not more than 1.4 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.7 per cent);

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105°C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 70 mL of ethanol (96 per cent) R and add

20 mL of water R. Stir with a mechanical stirrer for about

30 min and sonicate to achieve complete dissolution. Titrate

with 0.1 M sodium hydroxide, determining the end-point

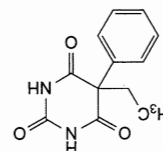
potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 24.63 mg of

C₁₃H₁₄N₂O₃.

IMPURITIES

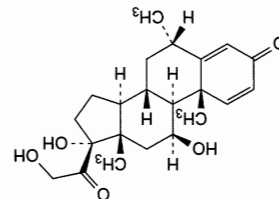
Specified impurities A



A. 5-ethyl-5-phenylpyrimidine-2,4,6-(1H,3H,5H)-trione
(phenobarbital).



Methylprednisolone
(Ph. Eur. monograph 0561)



C₂₂H₃₀O₅ 374.5 83-43-2

Action and use

Glucocorticoid.

Preparation

Methylprednisolone Tablets

DEFINITION

11β,17,21-Trihydroxy-6α-methylpregna-1,4-diene-3,20-dione.

Content

97.0 per cent to 102.0 per cent (dried substance).

Application 5 µL of test solution (a) and reference solution (a), 10 µL of test solution (b) and reference solution (b), applying the latter 2 in small quantities in order to obtain small spots.

Development Over 3/4 of the plate.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position, and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with alcoholic solution of sulfuric acid R and heat at 120 °C for 15 min. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spot in each of the chromatograms obtained with test solution (b) and reference solution (a) have an R_F value distinctly higher than that of the principal spot in each of the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of sulfuric acid R and shake to dissolve. Within 5 min, an intense red colour develops. When examined in ultraviolet light at 365 nm, brownish-red fluorescence is seen. Add this solution to 10 mL of water R and mix. The colour fades and there is a yellowish-green fluorescence in ultraviolet light at 365 nm.

TESTS

Specific optical rotation (2.2.7)

Dissolve 0.250 g in ethanol (96 per cent) R and dilute to + 97.0 to + 103.0 (dried substance).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture phosphoric acid R, acetonitrile R, water R (0.1:50:50 V/V/V).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 6 mg of methylprednisolone for system suitability CRS (containing impurities A, B, C, D, E, G and I) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 30.0 mg of methylprednisolone CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 µm);

— temperature: 45 °C.

Mobile phase:

— mobile phase A: phosphoric acid R, tetrahydrofuran R, acetonitrile R (0.1:1.5:10:90 V/V/V/V);

— mobile phase B: phosphoric acid R, tetrahydrofuran R, acetonitrile R (0.1:1.5:100 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 14	83	17
14 - 30	83 → 52	17 → 48

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 247 nm.

Injection 10 µL of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with methylprednisolone for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E, G and I.

Relative retention With reference to methylprednisolone (retention time = about 12 min): impurity B = about 0.85; impurity H = about 0.88; impurity A = about 0.92; impurities G and I = about 1.54; impurity C = about 1.7; impurity E = about 1.9; impurity D

(isomer 1) = about 2.10; impurity D (isomer 2) = about 2.2.

System suitability: reference solution (a):

— resolution: minimum 1.7 between the peaks due to impurity A and methylprednisolone.

Limits:

— impurity D: for the sum of the areas of the 2 isomer peaks, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurity A: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— sum of impurities G and I: not more than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— impurities B, H: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— impurities C, E: for each impurity, not more than 0.15 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{22}H_{30}O_5$ taking into account the assigned content of methylprednisolone CRS.

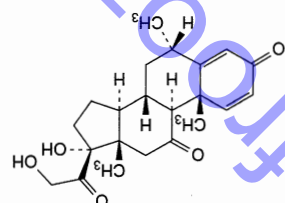
STORAGE

Protected from light, at a temperature of 2 °C to 8 °C.

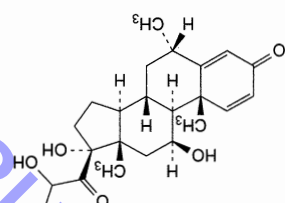
IMPURITIES

Specified impurities A, B, C, D, E, G, H, I

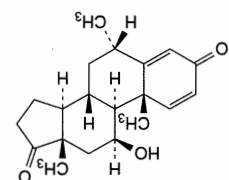
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): F, J, K, L.



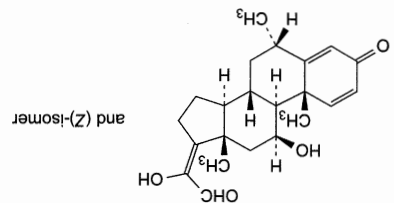
A. 17,21-dihydroxy-6α-methylpregna-1,4-diene-3,11,20-trione,



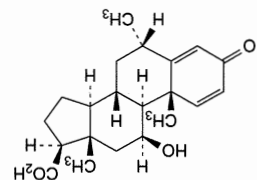
B. 11β,17,21-tetrahydroxy-6α-methylpregna-1,4-diene-3,20-dione,



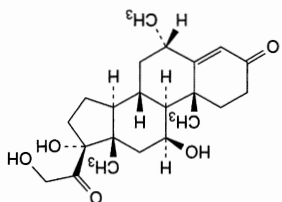
C. 11β-hydroxy-6α-methylandrosta-1,4-diene-3,17-dione,



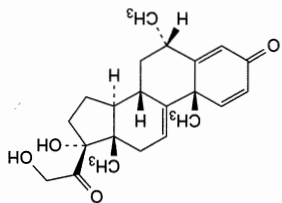
D. (E)-11β,20-dihydroxy-6α-methylpregna-1,4,17(20)-trien-3,21-dione,



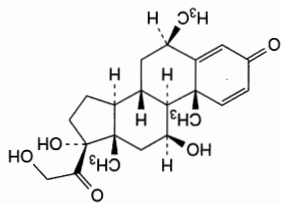
E. 11β-hydroxy-6α-methyl-3-oxoandrosta-1,4-diene-17β-carboxylic acid,



F. 11β,17,21-trihydroxy-6α-methylpregna-4-ene-3,20-dione,

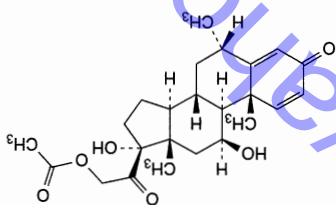


G. 17,21-dihydroxy-6α-methylpregna-1,4,9(11)-trien-3,20-dione,

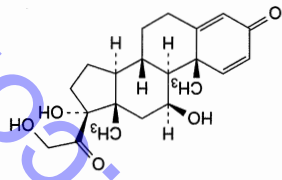


H. 11β,17,21-trihydroxy-6β-methylpregna-1,4-diene-3,20-dione,

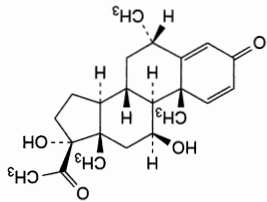
I. unknown structure,



J. 11β,17-dihydroxy-6α-methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (methylprednisolone acetate),



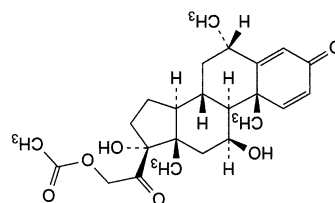
K. 11β,17,21-trihydroxypregna-1,4-diene-3,20-dione (prednisolone),



L. 11β,17-dihydroxy-6α-methylpregna-1,4-diene-3,20-dione.

Methylprednisolone Acetate

(Ph. Eur. monograph 0933)

C₂₄H₃₂O₆ 416.5 53-36-1

Action and use

Glucocorticoid.

Preparation

Methylprednisolone Acetate Injection

Ph Eur

DEFINITION

11β,17-Dihydroxy-6α-methyl-3,20-dioxopregn-1,4-dien-21-yl acetate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

First identification A, B.

Second identification C, D, E.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison methylprednisolone acetate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of acetone R, evaporate to dryness and record new spectra using the residues.

B. Thin-layer chromatography (2.2.27).

Solvent mixture methanol R, methylene chloride R (1:9 V/V).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of methylprednisolone acetate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of prednisolone acetate CRS and 10 mg of methylprednisolone acetate CRS in the solvent mixture, then dilute to 10 mL with the solvent mixture.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase butanol R, toluene R, ether R (5:10:85 V/V/V).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the test solution.

principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R. Heat

at 120 °C for 10 min or until the spots appear. Allow to

cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained

with the test solution is similar in position, colour in daylight,

fluorescence in ultraviolet light at 365 nm and size to the

principal spot in the chromatogram obtained with reference

solution (a).

System suitability: reference solution (b):

— the chromatogram shows 2 spots which, when

examined in ultraviolet light at 365 nm, may not be

completely separated.

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 25 mg of the substance to be

examined in methanol R and dilute to 5 mL with the same

solvent (solution A). Dilute 2 mL of this solution to 10 mL

with methylene chloride R.

Test solution (b) Transfer 2 mL of solution A to a 15 mL

glass tube with a ground-glass stopper or a

polytetrafluoroethylene cap. Add 10 mL of saturated

methanolic potassium hydrogen carbonate solution R and

immediately pass a current of nitrogen R through the solution

for 5 min. Stopper the tube. Heat in a water-bath at 45 °C

protected from light for 1 h. Allow to cool.

Reference solution (a) Dissolve 25 mg of methylprednisolone

acetate CRS in methanol R and dilute to 5 mL with the same

solvent (solution B). Dilute 2 mL of this solution to 10 mL

with methylene chloride R.

Reference solution (b) Transfer 2 mL of solution B to a 15 mL

glass tube with a ground-glass stopper or a

polytetrafluoroethylene cap. Add 10 mL of saturated

methanolic potassium hydrogen carbonate solution R and

immediately pass a current of nitrogen R through the solution

for 5 min. Stopper the tube. Heat in a water-bath at 45 °C

protected from light for 1 h. Allow to cool.

Mobile phase Add a mixture of 1.2 volumes of water R and

8 volumes of methanol R to a mixture of 15 volumes of

ether R and 77 volumes of methylene chloride R.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms

obtained with the test solutions is similar in position and size

to the principal spot in the chromatogram obtained with the

corresponding reference solution.

Detection B Spray with alcoholic solution of sulfuric acid R. Heat

at 120 °C for 10 min or until the spots appear. Allow to

cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms

obtained with the test solutions is similar in position, colour

in daylight, fluorescence in ultraviolet light at 365 nm and

size to the principal spot in the chromatogram obtained with

the corresponding reference solution. The principal spot in

each of the chromatograms obtained with test solution (b)

and reference solution (b) has an R_F value distinctly lower

than that of the principal spot in each of the chromatograms

obtained with test solution (a) and reference solution (a).

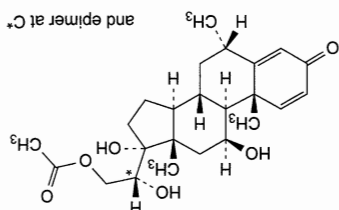
D. Add about 2 mg to 2 mL of sulfuric acid R and shake to

dissolve. Within 5 min, an intense red colour develops. When

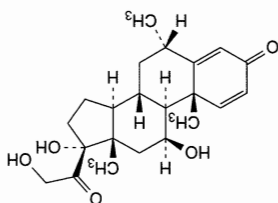
STORAGE

Protected from light.

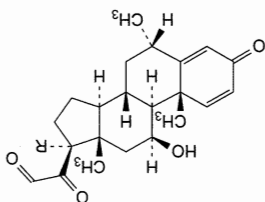
IMPURITIES



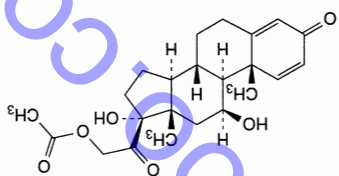
A. (20R)-11β,17,20-trihydroxy-6α-methyl-3-oxopregna-1,4-dien-21-yl acetate,



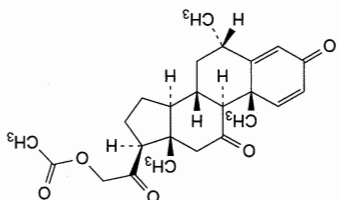
B. 11β,17,21-trihydroxy-6α-methylpregna-1,4-diene-3,20-dione (methylprednisolone),



C. R = OH. 11β,17-dihydroxy-6α-methylpregna-1,4-diene-3,20,21-trione,
D. R = H. 11β-hydroxy-6α-methylpregna-1,4-diene-3,20,21-trione,



E. 11β,17-dihydroxy-3,20-dioxopregna-1,4-dien-21-yl acetate (prednisolone acetate),



F. 6α-methyl-3,11,20-trioxopregna-1,4-dien-21-yl acetate,

TESTS

Specific optical rotation (2.2.7)

+ 97 to + 105 (dried substance).

Dissolve 0.250 g in dioxan R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in 5 mL of tetrahydrofuran R and dilute to 10.0 mL with water R.

Reference solution (a) Dissolve 4 mg of methylprednisolone acetate CRS and 4 mg of dexamethasone acetate CRS in the mobile phase, then dilute to 20.0 mL with the mobile phase.

Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase In a 1000 mL volumetric flask mix 260 mL tetrahydrofuran R and 700 mL of water R, then allow to equilibrate; dilute to 1000 mL with water R and mix again.

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for about 45 min.

Injection 20 μ L.

Run time 1.5 times the retention time of methylprednisolone acetate.

Retention time Methylprednisolone acetate = about 43 min;

dexamethasone acetate = about 57 min.

System suitability: reference solution (a):

— resolution: minimum 6.5 between the peaks due to methylprednisolone acetate and dexamethasone acetate;

if necessary, adjust the concentration of water R in the mobile phase.

Limits:

— total: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b);

solution (b) (1.0 per cent);

— disregard limit: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

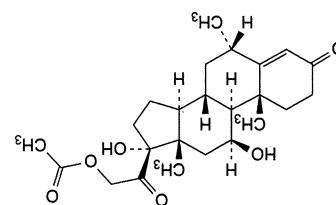
Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

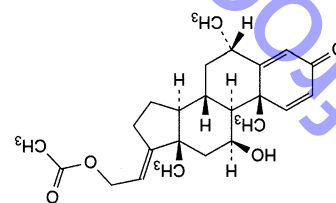
ASSAY

Dissolve 0.100 g in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with ethanol (96 per cent) R. Measure the absorbance (2.2.25) at the absorption maximum at 243 nm.

Calculate the content of $C_{24}H_{32}O_6$ taking the specific absorbance to be 355.



G. 11β,17-dihydroxy-6α-methyl-3,20-dioxopregna-4-en-21-yl acetate,

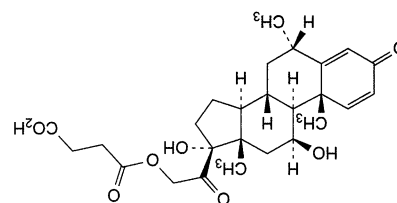


H. 11β-hydroxy-6α-methyl-3-oxopregna-1,4,17(20)-trien-21-yl acetate.

Ph Eur

Methylprednisolone Hydrogen Succinate

(Ph. Eur. monograph 1131)



2921-57-5

Action and use

Glucocorticoid.

Ph Eur

DEFINITION

4-[(11β,17-Dihydroxy-6α-methyl-3,20-dioxopregna-1,4-dien-21-yl)oxy]-4-oxobutanoic acid.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Practically insoluble in water, slightly soluble in acetone and in anhydrous ethanol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification A, B

Second identification C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison methylprednisolone hydrogen succinate CRS.

B. Thin layer chromatography (2.2.27).

Solvent mixture methanol R, methylene chloride R (1:9 V/V).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 20 mg of methylprednisolone hydrogen succinate CRS in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of hydrocortisone hydrogen succinate CRS in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase anhydrous formic acid R, anhydrous ethanol R, methylene chloride R (0.1:1:15 V/V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R; heat at 120 °C for 10 min or until the spots appear and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability: reference solution (b):

— the chromatogram shows 2 spots which may, however, not be completely separated.

C. Thin layer chromatography (2.2.27).

Test solution (a) Dissolve 25 mg of the substance to be examined in methanol R with gentle heating and dilute to 5 mL with the same solvent (solution A). Dilute 2 mL of this solution to 10 mL with methylene chloride R.

Test solution (b) Transfer 2 mL of solution A to a 15 mL glass tube with a ground-glass stopper or a

polytetrafluoroethylene cap. Add 10 mL of a 0.8 g/L solution of sodium hydroxide R in methanol R and immediately pass a stream of nitrogen R through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C, protected from light, for 30 min. Allow to cool.

Reference solution (a) Dissolve 25 mg of methylprednisolone hydrogen succinate CRS in methanol R with gentle heating and dilute to 5 mL with the same solvent (solution B). Dilute 2 mL of this solution to 10 mL with methylene chloride R.

Reference solution (b) Transfer 2 mL of solution B to a 15 mL glass tube with a ground-glass stopper or a

polytetrafluoroethylene cap. Add 10 mL of a 0.8 g/L solution of sodium hydroxide R in methanol R and immediately pass a stream of nitrogen R through the solution for 5 min. Stopper the tube. Heat in a water-bath at 45 °C, protected from light, for 30 min. Allow to cool.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and 8 volumes of methanol R to a mixture of 15 volumes of ether R and 77 volumes of methylene chloride R.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms obtained with the test solutions is similar in position and size to the principal spot in the chromatogram obtained with the corresponding reference solution.

Detection B Spray with alcoholic solution of sulfuric acid R. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spot in each of the chromatograms obtained with test solution (b) and reference solution (b) has an R_f value distinctly higher than that of the principal spot in each of the chromatograms obtained with test solution (a) and reference solution (a).

D. Add about 2 mg to 2 mL of sulfuric acid R and shake to dissolve. Within 5 min a reddish-brown colour develops. Add this solution to 10 mL of water R and mix. The colour fades and a precipitate is formed.

TESTS

Appearance of solution

The solution is clear (2.2.1).

Dissolve 0.100 g in 5 mL of sodium hydrogen carbonate solution R.

Specific optical rotation (2.2.7)

+ 87 to + 95 (dried substance).

Dissolve 0.250 g in dioxan R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 25 mg of methylprednisolone hydrogen succinate for performance test CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R, 3 per cent V/V solution of glacial acetic acid R (33:67 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for about 30 min.

Injection 20 μ L.

Run time Twice the retention time of methylprednisolone hydrogen succinate.

Retention time Methylprednisolone hydrogen succinate = about 22 min; impurity D (eluting immediately after the main peak and appearing as a shoulder) = about 24 min.

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 4, where H_p = height above the base line of the peak due to impurity D and H_v = height above the base line of the lowest point of the curve separating this peak from the peak due to methylprednisolone hydrogen succinate; if necessary,

adjust the concentration of acetonitrile in the mobile phase.

Limits:

— **impurities A, B, C, D:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— **total:** not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);

— **disregard limit:** 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 50.0 mL with ethanol (96 per cent) R. Measure the absorbance (2.2.25) at the absorption maximum at 243 nm.

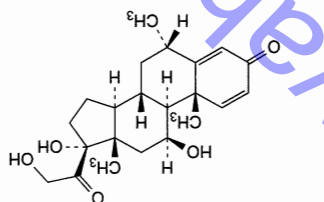
Calculate the content of $C_{26}H_{34}O_8$ taking the specific absorbance to be 316.

STORAGE

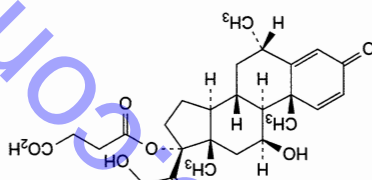
In an airtight container, protected from light.

IMPURITIES

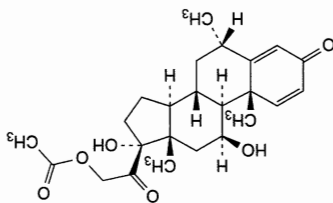
Specified impurities A, B, C, D



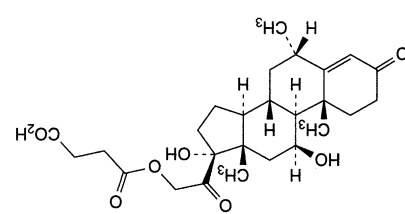
A. 11 β ,17,21-trihydroxy-6 α -methylpregna-1,4-diene-3,20-dione (methylprednisolone).



B. 4-[(11 β ,21-dihydroxy-6 α -methyl-3,20-dioxopregna-1,4-dien-17-yl)oxy]-4-oxobutanoic acid (methylprednisolone 17-(hydrogen succinate)).



C. 11 β ,17-dihydroxy-6 α -methyl-3,20-dioxopregna-1,4-dien-21-yl acetate (methylprednisolone acetate).

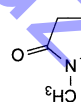


D. 4-[(11β,17-dihydroxy-6α-methyl-3,20-dioxopregn-4-en-21-yl)oxy]-4-oxobutanoic acid (methylhydrocortisone 21-(hydrogen succinate)).

Ph Eur

Methylpyrrolidone

(N-Methylpyrrolidone, Ph Eur monograph 1675)



872-50-4

C₅H₉NO

DEFINITION

1-Methylpyrrolidin-2-one.

CHARACTERS

Appearance
Clear, colourless liquid.

Solubility

Miscible with water and with alcohol.

bp

About 204 °C.

Relative density

About 1.034.

Refractive index

About 1.469.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Films.

Comparison Ph. Eur. reference spectrum of N-methylpyrrolidone.

TESTS

Appearance

The substance to be examined is clear (2.2.1) and colourless (2.2.2, Method II).

Alkalinity

Dissolve 50 mL of the substance to be examined in 50 mL of water R previously adjusted with 0.02 M potassium hydroxide or 0.02 M hydrochloric acid until a yellow colour is obtained using 0.5 mL of bromothymol blue solution R1 as indicator. Titrate with 0.02 M hydrochloric acid to the initial coloration. Not more than 8.0 mL of 0.02 M hydrochloric acid is required.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

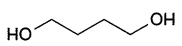
Test solution The substance to be examined.

Reference solution To 1 mL of the substance to be examined, add 1 mL of 2-pyrrolidone R and dilute to 20 mL with methylene chloride R.

Column:

— material: fused silica,

F. butane-1,4-diol,



(2-pyrrolidone),

G. R1 = R2 = R3 = R4 = H: pyrrolidin-2-one

(5R)-1,5-dimethylpyrrolidin-2-one,

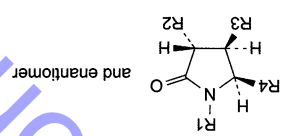
E. R1 = R4 = CH₃, R2 = R3 = H:

(4R)-1,4-dimethylpyrrolidin-2-one,

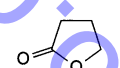
D. R1 = R3 = CH₃, R2 = R4 = H:

(3R)-1,3-dimethylpyrrolidin-2-one,

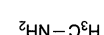
C. R1 = R2 = CH₃, R3 = R4 = H:



B. dihydrofuran-2(3H)-one (γ-butyrolactone),



A. methanamine (methylamine),



IMPURITIES

Protected from light.

STORAGE

Maximum 0.1 per cent, determined on 1.000 g.

Water (2.5.32)

(2 ppm Pb) R.

Prepare the reference solution using lead standard solution same solvent. 12 mL of the solution complies with test A.

Dissolve 4.0 g in water R and dilute to 20.0 mL with the Maximum 10 ppm.

Heavy metals (2.4.8)

— disregard limit: 0.02 per cent.

— total: maximum 0.3 per cent,

— any impurity: maximum 0.1 per cent,

Limits:

N-methylpyrrolidone and impurity G.

System suitability: reference solution:

Injection 1 µL.

Detection Flame ionisation.

Time (min)	Temperature (°C)	Column	Injection port	Detector
0	100			
23.3	170	0 - 23.3		
53	280	23.3 - 53		
	280			

Temperature:

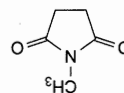
Split ratio 1:100.

Linear velocity 20 cm/s.

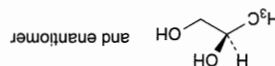
Carrier gas nitrogen for chromatography R.

— stationary phase: poly(dimethyl)siloxane R (5 µm).

— size: l = 30 m, Ø = 0.32 mm,



H. 1-methylpyrrolidine-2,5-dione (N-methylsuccinimide),

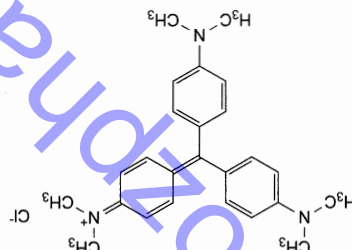


I. (R,S)-propane-1,2-diol (propylene glycol).

Ph Eur

Methylosanilinium Chloride

(Ph. Eur. monograph 1990)



$C_{25}H_{30}ClN_3$ 408.0
548-62-9

Action and use

Antiseptic dye (gentian violet).

Ph Eur

DEFINITION

N-[4-Bis[4-(dimethylamino)phenyl]methylene]cyclohexa-2,5-dienylidene]-N-methyldiamanaminium chloride (hexamethyl-*p*-rosanilinium chloride). It contains not more than 10 per cent of pentamethyl-*p*-rosanilinium chloride and is also known as crystal violet and gentian violet.

Content

95.0 per cent to 103.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Dark green, shiny powder, hygroscopic.

Solubility

Sparsely soluble in water, freely soluble in ethanol

(96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: A.

Second identification: B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison methylosanilinium chloride CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of methylosanilinium chloride CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase glacial acetic acid R, water R, butanol R (17:17:66 V/V/V).

Application 2 μ L.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution. In the chromatogram obtained with the test solution, a secondary spot may be observed.

C. Dissolve 50 mg in water R and dilute to 5 mL with the same solvent; add 3 mL of dilute sulfuric acid R, 1 g of zinc powder R and heat gently. The mixture decolourises. Filter.

To 3 mL of the filtrate add 0.5 mL of silver nitrate solution R1. A white turbidity is produced which slowly forms a dark, coagulating precipitate.

TESTS

N,N-Dimethylaniline (2.4.26, Method A)

Maximum 100 ppm.

Test solution Dissolve in a ground-glass-stoppered tube 0.50 g of the substance to be examined in 30.0 mL of water R.

Add 1.0 mL of the internal standard solution. Adjust the solution to 26-28 °C. Add 1.0 mL of strong sodium hydroxide solution R and mix for 2 min. Add 2.0 mL of trimethylpentane R. Shake for 2 min and centrifuge. Use the upper layer.

Reference solution Dissolve 50.0 mg of N,N-dimethylaniline R in 4.0 mL of 0.1 M hydrochloric acid and dilute to 50.0 mL with water R. Dilute 1.0 mL of this solution to 100.0 mL with water R. To 0.50 g of the substance to be examined add 5.0 mL of this solution and dilute to 30.0 mL with water R.

Add 1.0 mL of the internal standard solution and 1.0 mL of strong sodium hydroxide solution R. Add 2.0 mL of trimethylpentane R. Shake for 2 min and centrifuge. Use the upper layer.

Limit: calculate the ratio (R) of the area of the peak due to the N,N-dimethylaniline to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with the test solution, calculate the ratio of the area of the peak due to N,N-dimethylaniline to the area of the peak due to the internal standard; this ratio is not greater than 0.5 R.

Pentamethyl-*p*-rosanilinium

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 30.0 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 3.0 mg of methylosanilinium for system suitability CRS in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with ethanol (96 per cent) R. Dilute 1.0 mL of this solution to 100.0 mL with ethanol (96 per cent) R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: spherical octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase glacial acetic acid R, water R, methanol R (10:190:800 V/V/V).

Flow rate 1 mL/min.

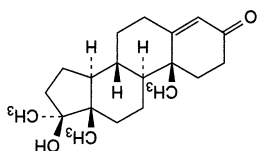
Detection Spectrophotometer at 589 nm.

Injection 20 μ L.



Methyltestosterone

(Ph. Eur. monograph 0410)



$C_{20}H_{30}O_2$

302.5

58-18-4

Action and Use

Anabolic steroid.

DEFINITION

17β-Hydroxy-17-methylandroster-4-en-3-one.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish-white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B.

Second identification A, C.

A. Melting point (2.2.14): 162 °C to 168 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Comparison methyltestosterone CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.2 g of the substance to be examined

in a mixture of 1 volume of methanol R and 9 volumes of

chloroform R and dilute to 10 mL with the same mixture of

solvents.

Reference solution Dissolve 20 mg of methyltestosterone CRS in

1 mL of a mixture of 1 volume of methanol R and 9 volumes

of chloroform R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase anhydrous acetic acid R, light petroleum R, butyl

acetate R (1:30:70 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm and spray

with a saturated solution of potassium dichromate R in a

mixture of 30 volumes of water R and 70 volumes of sulfuric

acid R. Examine immediately in daylight.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

TESTS

Specific optical rotation (2.2.7)

+ 79 to + 85 (dried substance).

Dissolve 0.250 g in ethanol (96 per cent) R and dilute to

25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Run time 2.5 times the retention time of the principal peak.

System suitability: reference solution (a):

— resolution: peak due to pentamethyl-p-rosanilinium is

baseline separated from the peak due to

methylrosanilinium.

Locate the peak due to pentamethyl-p-rosanilinium using the

chromatogram provided with methylrosanilinium for system

stability CRS.

Limits:

— pentamethyl-p-rosanilinium: maximum 10 per cent.

Liquid chromatography (2.2.29) as described in the test for

pentamethyl-p-rosanilinium.

Limits:

— impurity A: maximum 1.0 per cent;

— any other impurity: for each impurity, maximum

0.1 per cent;

— sum of impurities other than A: maximum 1.0 per cent;

— disregard limit: the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.05 per cent); disregard the peak due to pentamethyl-p-

rosanilinium.

Substances insoluble in ethanol (90 per cent V/V)

Maximum 0.5 per cent.

In a conical flask introduce 1.0 g and add 50 mL of ethanol

(90 per cent V/V) R. Boil under a reflux condenser for 1 h.

Filter the warm liquid through a weighed sintered glass

filter (16) (2.1.2) previously dried at 100-105 °C. Wash with

hot ethanol (90 per cent V/V) R until a colourless filtrate is

obtained. Dry at 100-105 °C until constant weight.

Water (2.5.12)

Maximum 10.0 per cent, determined on 0.100 g.

Sulfated ash (2.4.14)

Maximum 1.5 per cent, determined on 1.0 g.

ASSAY

Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to

100.0 mL with the same solvent. Dilute 2.0 mL of the

solution to 250.0 mL with ethanol (96 per cent) R. Measure

the absorbance (2.2.25) at the maximum at 589 nm.

Calculate the content of $C_{25}H_{30}ClN_3$ taking the specific

absorbance to be 2605.

STORAGE

In an airtight container.

IMPURITIES

Specified impurities: A.

A. impurity of unknown structure with a relative retention of

about 0.7.

Ph Eur

chromatogram obtained with reference solution (b) (0.5 per cent),
 — *sum of impurities other than A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent),
 — *disregard limit*: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Metals

Atomic emission spectrometry (2.2.22) in argon plasma, using as detector a conventional optical system or a mass spectrometer; in the case of a mass spectrometer, use indium as internal standard.

Test solution In a 10 mL volumetric flask, dissolve with stirring 100 mg of the substance to be examined in 9 mL of water R, add 100.0 µL of a 10 µg/mL solution of indium prepared from indium elementary standard solution for atomic spectrometry (1.000 g/L) R in nitric acid R which has been diluted fifty-fold with water R. Dilute to 10.0 mL with water R.

Reference solutions Into a 100 mL volumetric flask, introduce 10.0 mL of a standard solution containing 1.00 µg/mL of each of the metals to be determined and prepared by dilution, with water R, of each elementary standard solution for atomic spectrometry (1.000 g/L) R for the corresponding elements. Add 1.00 mL of a 10 µg/mL solution of indium prepared from indium elementary standard solution for atomic spectrometry (1.000 g/L) R in nitric acid R which has been diluted fifty-fold with water R. Dilute to 100.0 mL with water R.

Blank solution Dilute one hundred-fold with water R the 10 µg/mL solution of indium used for the test and reference solutions.

Element

Element	Optical detection			Mass detection
	Signal (nm)	Background 1 (nm)	Background 2 (nm)	
Aluminium	396.15	396.05	396.25	27
Cadmium	214.44	214.37	214.51	114
Chromium	283.56	283.49	283.64	*
Copper	327.40	327.31	327.48	65
Tin	190.00**	189.90	190.10	118
Iron	238.20	238.27	238.14	*
Manganese	260.57	260.50	260.64	55
Mercury	253.70***	253.60	253.80	200
Molybdenum	202.03	202.02	202.04	95
Nickel	231.60	231.54	231.66	60
Lead	217.00**	216.90	217.10	208
Zinc	213.86	213.80	213.91	66
Indium				115

*Element difficult, if not impossible, to be determined with a mass spectrometer as detector.
 **Borderline sensitivity with conventional optical spectrometry.
 ***Mercury is often impossible to determine using conventional optical spectrometry; it may be quantified using a device for the determination of hydrides.

Mobile phase anhydrous formic acid R, propanol R (20:80 V/V).
Application 2 µL.
Development Over a path of 8 cm.
Drying In air, protected from light.
Detection Examine in daylight.
Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution. A secondary spot may appear above the principal spot in both chromatograms.
C. Dissolve about 1 mg in 10 mL of water R. Add 1 mL of glacial acetic acid R and 0.1 g of zinc powder R. Heat to boiling. The solution becomes colourless. Filter and shake the filtrate. It becomes blue on contact with air.
D. Ignite 50 mg with 0.5 g of anhydrous sodium carbonate R. Cool and dissolve the residue in 10 mL of dilute nitric acid R. Filter. The filtrate, without further addition of dilute nitric acid R, gives reaction (a) of chlorides (2.3.1).

TESTS

Methanol-insoluble substances

Maximum 10.0 mg (1.0 per cent).
 To 1.0 g add 20 mL of methanol R and boil under a reflux condenser for 5 min. Filter through a tared sintered-glass filter (40) (2.1.2) and wash the filter with methanol R until a colourless filtrate is obtained. Dry the filter at 100 °C and weigh.

Related substances

Test solution Dissolve 15.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 7.5 mg of methylthionium impurity A CRS in the mobile phase and dilute to 50.0 mL 1.0 mL of the test solution and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:
 — size: $l = 0.25$ m, $\varnothing = 4$ mm,
 — stationary phase: octadecylsilyl silica gel for chromatography R (7 µm).

Mobile phase Mix 27 volumes of acetonitrile R and 73 volumes of a mixture of 3.4 mL of phosphoric acid R and 1000 mL of water R.

Detection Spectrophotometer at 246 nm.
Injection 20 µL.

Run time Twice the retention time of methylthionium. **Relative retention** With reference to methylthionium (retention time = about 11 min): impurity A = about 0.7. **System suitability:** reference solution (a):
 — resolution: minimum 1.5 between the peaks due to impurity A and methylthionium. If necessary, adjust the concentration of acetonitrile in the mobile phase.

Limits:

— **impurity A**: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (5.0 per cent),
 — **any other impurity**: for each impurity, not more than 0.5 times the area of the principal peak in the

Element	Maximum content in ppm
Aluminium	100 ppm
Cadmium	1 ppm
Chromium	100 ppm
Copper	300 ppm
Tin	10 ppm
Iron	200 ppm
Manganese	10 ppm
Mercury	1 ppm
Molybdenum	10 ppm
Nickel	10 ppm
Lead	10 ppm
Zinc	100 ppm

Loss on drying (2.2.32) 8.0 per cent to 22.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14) Maximum 0.25 per cent, determined on 1.0 g.

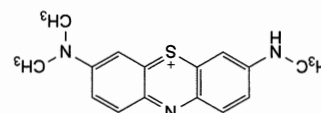
ASSAY Dissolve 0.300 g in 30 mL of water R with heating. Cool, add 50.0 mL of potassium dichromate solution R1 and dilute to 100.0 mL with water R. Allow to stand for 10 min. Filter and discard the first 20 mL of filtrate. Introduce 50.0 mL of the filtrate into a flask with a ground-glass neck, add 50 mL of dilute sulfuric acid R and 8.0 mL of potassium iodide solution R. Allow to stand protected from light for 5 min, then add 80 mL of water R. Titrate with 0.1 M sodium thiosulfate using 2 mL of starch solution R, added towards the end of the titration, as indicator. Carry out a blank titration. 1 mL of 0.1 M sodium thiosulfate is equivalent to 10.66 mg of $C_{16}H_{18}ClN_3S$.

STORAGE

In an airtight container, protected from light.

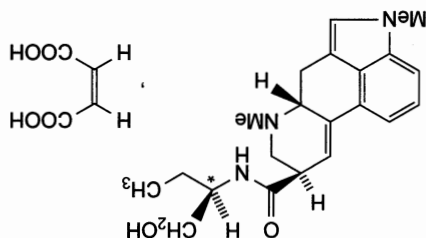
IMPURITIES

Specified impurities A.



A. 3-(dimethylamino)-7-(methyliamino)phenothiazin-5-ylum.
Ph Eur

Methysergide Maleate



and epimer at C*

$C_{21}H_{27}N_3O_2 \cdot C_4H_4O_4$

469.5

129-49-7

Action and use

Non-selective 5HT receptor antagonist.

Preparation

Methysergide Tablets

DEFINITION

Methysergide Maleate is (1R)-N-[1-(hydroxymethyl)-1-methyl-2-lysergamide hydrogen maleate. It contains not less than 98.0% and not more than 101.0% of $C_{21}H_{27}N_3O_2 \cdot C_4H_4O_4$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder which may have a yellow or pink tinge.

Slightly soluble in water and in methanol; practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of methysergide maleate (RS 227).

B. In the test for Related substances, the principal spot in the chromatogram obtained with solution (1) corresponds to that in the chromatogram obtained with solution (8).

C. Dissolve 1 mg in 1 mL of ethanol (96%) and add 1 mL of dimethylaminobenzaldehyde solution R6. A brownish red to violet colour is produced.

TESTS

Acidity

pH of a 0.2% w/v solution, 3.7 to 4.7, Appendix V L.

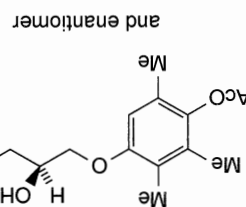
Specific optical rotation

In a 0.25% w/v solution, +35.0 to +45.0, calculated with reference to the dried substance, Appendix V F.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, protected from light using a suspension of silica gel G in 0.1M sodium hydroxide to prepare the plate and a mixture of 90 volumes of chloroform and 10 volumes of methanol as the mobile phase. Apply separately to the plate 5 µL of each of the following solutions. Solution (1) contains 2.0% w/v of the substance being examined in a mixture of 1 volume of 13.5M ammonia and 100 volumes of methanol. Solutions (2) to (8) are solutions of methysergide maleate BPCRS in a mixture of 1 volume of 13.5M ammonia and 100 volumes of methanol containing (2) 0.0050% w/v, (3) 0.010% w/v, (4) 0.015% w/v, (5) 0.020% w/v, (6) 0.030% w/v, (7) 0.040% w/v and (8) 2.0% w/v respectively. After removal of the plate, allow it to dry in air and examine under ultraviolet light (365 nm). Assess the intensities of any

Metipranolol



Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.4 g and 1-naphtholbenzenesulfonic acid VS is equivalent to 46.95 mg of $C_{21}H_{27}N_3O_2 \cdot C_4H_4O_4$.

Appendix VIII A, using 0.4 g and 1-naphtholbenzenesulfonic acid VS is equivalent to 46.95 mg of $C_{21}H_{27}N_3O_2 \cdot C_4H_4O_4$.

STORAGE

Methysergide Maleate should be protected from light and stored at a temperature of 2° to 8°.

ASSAY

Use 1 g.

When dried to constant weight at 120° at a pressure not exceeding 0.7 kPa, loses not more than 7.0% of its weight.

Loss on drying

intensities so assessed does not exceed 2%.

chromatogram obtained with solution (2). The total of the disregarding any spots less intense than the spot in the assessing the intensities of spots of different R_f values and with solutions (2) to (7), making allowance for area in (1) by reference to the spots in the chromatograms obtained secondary spots in the chromatogram obtained with solution

$C_{17}H_{27}NO_4$ 309.4 22664-55-7

Preparation

Beta-adrenoceptor antagonist.

Action and use

Metipranolol Eye Drops

DEFINITION

Metipranolol is (RS)-4-(2-hydroxy-3-isopropylaminoethoxy)-2,6-dimethylphenyl acetate. It contains not less than 99.0% and not more than 101.0% of $C_{17}H_{27}NO_4$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white, crystalline powder; melting point, about 108°. Practically insoluble in water; soluble in ethanol (96%), in acetone and in methanol. It dissolves in dilute mineral acids.

IDENTIFICATION

The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of metipranolol (RS 375).

TESTS

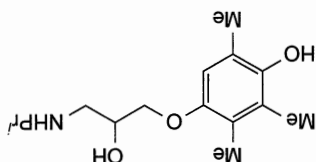
Alkalinity

Shake 0.5 g with 20 mL of water for 5 minutes and filter. The pH of the filtrate is 9.0 to 10.0, Appendix V L.

Colour of solution

A 5.0% w/v solution in 1M hydrochloric acid is not more intensely coloured than reference solution Y, Appendix IV B.

A. desacetylmethipranolol



IMPURITIES

Metipranolol should be protected from light.

STORAGE

Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.3 g and determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 30.94 mg of $C_{17}H_{27}NO_4$.

ASSAY

Not more than 0.5% w/w, Appendix IX C. Use 1 g.

Water

Not more than 0.1%, Appendix IX A. Use 2 g.

Sulfated ash

Not more than 0.1%, Appendix IX A. Use 2 g.

In the chromatogram obtained with solution (1) the area of any peak corresponding to desacetylmethipranolol is not greater than the area of the principal peak in the sum chromatogram obtained with solution (3) (1%) and the sum of the areas of any secondary peaks is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

The test is not valid unless, in the chromatogram obtained with solution (4), the resolution factor between the two principal peaks is at least 5.

In the chromatogram obtained with solution (1) the area of any peak corresponding to desacetylmethipranolol is not greater than the area of the principal peak in the sum chromatogram obtained with solution (3) (1%) and the sum of the areas of any secondary peaks is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.5%).

The chromatographic procedure may be carried out using end-capped octadecylsilyl silica gel for chromatography (5 µm) (Hypersil ODS is suitable), (b) as the mobile phase with a flow rate of 2 mL per minute a mixture of 1 volume of perchloric acid, 45 volumes of methanol and 54 volumes of water, the pH of the mixture being adjusted to 3.0 with 13.5M ammonia and (c) a detection wavelength of 275 nm. For solution (1) allow the chromatography to proceed for at least 3 times the retention time of the principal peak.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in methanol. Solution (1) contains 0.10% w/v of the substance being examined. Solution (2) contains 0.0005% w/v of the substance being examined. Solution (3) contains 0.001% w/v of desacetylmethipranolol BPCRS. Solution (4) contains 0.02% w/v of the substance being examined and 0.02% w/v of desacetylmethipranolol BPCRS.

Iron

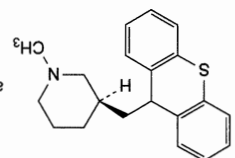
Dissolve the residue obtained in the test for Sulfated ash in 1 mL of hydrochloric acid, evaporate to dryness, dissolve the residue in a mixture of 0.5 mL of 6M acetic acid and 7 mL of water and add sufficient water to produce 20 mL. 10 mL of this solution complies with the limit test for iron, Appendix VII standard (10 ppm).

Heavy metals

2 g complies with limit test C for heavy metals, Appendix VII. Use 2 mL of lead standard solution (10 ppm Pb) to prepare the

Metixene Hydrochloride

(Ph. Eur. monograph 1347)

and enantiomer, HCl, H₂OC₂₀H₂₄ClNS, H₂O 363.9 7081-40-5

Action and use

Anticholinergic.

Ph Eur

DEFINITION

Metixene hydrochloride contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of (RS)-1-methyl-3-[(9H-thioxanthen-9-yl)methyl]piperidine hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline or fine crystalline powder, soluble in water, soluble in alcohol and in methylene chloride.

IDENTIFICATION

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with metixene hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

Dissolve 0.40 g in methanol R and dilute to 20.0 mL with the same solvent. The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2).

Method I).

pH (2.2.3)

Dissolve 0.18 g in carbon dioxide-free water R heating if necessary at about 50 °C, cool and dilute to 10.0 mL with the same solvent. The pH of the solution, measured immediately, is 4.4 to 5.8.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel plate R. Carry out the test rapidly and protected from light.

Test solution Dissolve 50 mg of the substance to be examined in methylene chloride R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of metixene hydrochloride CRS in methylene chloride R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of thioxanthene CRS in 50 mL of methylene chloride R. Dilute 1.0 mL of the solution to 20.0 mL with methylene chloride R.

Reference solution (c) Dissolve 5 mg of thioxanthone CRS in 50 mL of methylene chloride R. Dilute 1.0 mL of the solution to 20.0 mL with methylene chloride R.

Reference solution (d) Dilute 4 mL of reference solution (a) to 10.0 mL with methylene chloride R.

Apply to the plate as narrow bands 5 µL of each solution. Develop over a path of 10 cm using a mixture of 10 volumes

IMPURITIES

Store protected from light.

STORAGE

C₂₀H₂₄ClNS.

1 mL of 0.1 M sodium hydroxide is equivalent to 34.59 mg of metixene hydrochloride.

inflection. Read the volume added between the 2 points of

potentiometric titration (2.2.20), using 0.1 M sodium hydrochloric acid and 50 mL of alcohol R. Carry out a

Dissolve 0.250 g in a mixture of 5.0 mL of 0.01 M

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Not less than 4.0 per cent and not more than 6.0 per cent, determined on 0.500 g by drying in an oven at 138-142 °C.

Loss on drying (2.2.32)

are clearly visible and differentiated.

chromatograms obtained with reference solutions (b) and (c) (0.2 per cent). The test is not valid unless the bands in the

chromatogram obtained with reference solution (d)

most one such band is more intense than the band in the

obtained with reference solution (a) (0.5 per cent) and at

is not more intense than the band in the chromatogram, the bands corresponding to thioxanthene and thioxanthone,

(0.05 per cent); any band, apart from the principal band and chromatogram obtained with reference solution (c)

solution is not more intense than the band in the

to thioxanthone in the chromatogram obtained with the test reference solution (b) (0.2 per cent); any band corresponding

intense than the band in the chromatogram obtained with

chromatogram obtained with the test solution is not more

Any band corresponding to thioxanthene in the

and thioxanthone shows greenish-blue fluorescence.

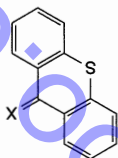
light at 365 nm. Thioxanthene shows orange fluorescence

10 min. Allow the plate to cool and examine in ultraviolet

acid R and 9 volumes of alcohol R and heat at 100 °C for

of cold air. Spray with a mixture of 1 volume of sulfuric

80 volumes of methylene chloride R. Dry the plate in a stream of glacial acetic acid R, 10 volumes of methanol R and



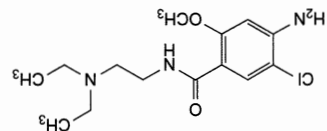
A. X = H₂: 9H-thioxanthene,

B. X = O: 9H-thioxanthen-9-one (thioxanthone).

Ph Eur

Metoclopramide

(Ph. Eur. monograph 1348)

C₁₄H₂₂ClN₃O₂ 299.8 364-62-5

Action and use

Dopamine receptor antagonists; antiemetic.

Metoclopramide Hydrochloride formulations should reflect the Hydrochloride salt.

DEFINITION

4-Amino-5-chloro-N-[2-((diethylamino)ethyl)]-2-methoxybenzamide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, fine powder.

Solubility

Practically insoluble in water, sparingly soluble or slightly soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

First identification A, B.

Second identification A, C.

A. Melting point (2.2.14): 145 °C to 149 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Examine the chromatograms obtained in test A for related

substances.

Detection Examine in ultraviolet light at 254 nm before

spraying with dimethylaminobenzaldehyde solution R1.

Results The principal spot in the chromatogram obtained with

test solution (a) is similar in position and size to the principal

spot in the chromatogram obtained with reference

solution (a).

TESTS

Appearance of solution

The freshly prepared solution is clear (2.2.1) and not more

intensely coloured than reference solution Y₆ (2.2.2,

Method II).

Dissolve 2.5 g in 25 mL of 1 M hydrochloric acid.

Related substances

A. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 40 mg of the substance to be

examined in methanol R and dilute to 10 mL with the same

solvent.

Test solution (b) Dissolve 0.160 g of the substance to be

examined in methanol R and dilute to 10 mL with the same

solvent.

Reference solution (a) Dissolve 20 mg of metoclopramide CRS

and 10 mg of sulphide CRS in methanol R and dilute to 5 mL

with the same solvent.

Reference solution (b) Dissolve 20 mg of N,N-

diethylethylenediamine R (impurity E) in methanol R and dilute

to 50 mL with the same solvent. Dilute 2 mL of this solution

to 25 mL with methanol R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, dioxan R, methanol R,

methylene chloride R (2:10:14:90 V/V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm

(identification C), then spray with dimethylaminobenzaldehyde

solution R1 and allow to dry in air.

System suitability: reference solution (a):

— the chromatogram shows 2 clearly separated spots.

Limit: test solution (b):

— impurity E: any spot due to impurity E (not visualised

in ultraviolet light at 254 nm) is not more intense than

the corresponding spot in the chromatogram obtained

with reference solution (b) (0.2 per cent).

B. Liquid chromatography (2.2.29).

Test solution Dissolve 10.0 mg of the substance to be

examined in the mobile phase and dilute to 10.0 mL with

the mobile phase.

Reference solution (a) Dilute 0.2 mL of the test solution to

100.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of metoclopramide

impurity A CRS in the mobile phase and dilute to 100 mL

with the mobile phase. Mix 1 mL of this solution with

mobile phase.

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: octylsilyl silica gel for chromatography R

(5 µm).

Mobile phase Dissolve 6.8 g of potassium dihydrogen

phosphate R in 700 mL of water R, add 0.2 mL of N,N-

dimethylethylamine R and adjust to pH 4.0 with dilute

phosphoric acid R, dilute to 1000 mL with water R, add

250 mL of acetonitrile R and mix.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time 8 times the retention time of metoclopramide.

Relative retention With reference to metoclopramide (retention

time = about 3.6 min): impurity A = about 0.82;

impurity F = about 0.89; impurity H = about 0.91;

impurity G = about 1.7; impurity C = about 2.7;

impurity D = about 2.8; impurity B = about 6.4.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurity A and metoclopramide.

Limits:

— impurities A, B, C, D, F, G, H: for each impurity, not

more than the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.2 per cent);

— total: not more than 3 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.6 per cent);

— disregard limit: 0.1 times the area of the principal peak

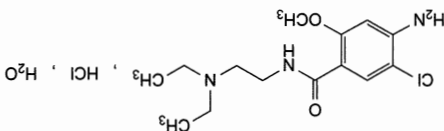
in the chromatogram obtained with reference

solution (a) (0.02 per cent).



Metoclopramide Hydrochloride

(Ph. Eur. monograph 0674)



$C_{14}H_{23}Cl_2N_3O_2 \cdot H_2O$ 354.3 54143-57-6

Action and use
Dopamine receptor antagonist; antiemetic.

Preparations
Metoclopramide Injection

Metoclopramide Oral Solution

Metoclopramide Tablets

Ph Eur

DEFINITION

Metoclopramide hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 4-amino-5-chloro-N-[2-(diethylamino)ethyl]-2-methoxybenzamide hydrochloride, calculated with reference to the anhydrous substance.

CHARACTERS

White or almost white, crystalline powder or crystals, very soluble in water, freely soluble in alcohol, sparingly soluble in methylene chloride.

It melts at about 183 °C with decomposition.

IDENTIFICATION

First identification A, B, D.
Second identification A, C, D, E.

A. The pH (2.2.3) of solution S (see Tests) is 4.5 to 6.0.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with metoclopramide hydrochloride CRS. Examine the substances as discs prepared using potassium chloride R.

C. Examine the chromatograms obtained in the test for related substances in ultraviolet light before spraying with dimethylaminobenzaldehyde solution R1. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dilute 1 mL of solution S to 2 mL with water R.

E. Dissolve about 2 mg in 2 mL of water R. The solution gives the reaction of primary aromatic amines (2.3.1).

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Related substances

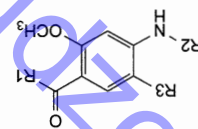
Examine by thin-layer chromatography (2.2.27), using silica gel HF₂₅₄ R as the coating substance.

Test solution (a) Dissolve 0.40 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with methanol R.

IMPURITIES

Specified impurities A, B, C, D, E, F, G, H.



A. R1 = NH-CH₂-CH₂-N(C₂H₅)₂; R2 = CO-

CH₃; R3 = Cl: 4-(acetylamino)-5-chloro-N-[2-

(diethylamino)ethyl]-2-methoxybenzamide,

B. R1 = OCH₃; R2 = CO-CH₃; R3 = Cl: methyl

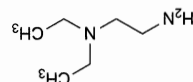
4-(acetylamino)-5-chloro-2-methoxybenzoate,

C. R1 = OH; R2 = H; R3 = Cl: 4-amino-5-chloro-2-

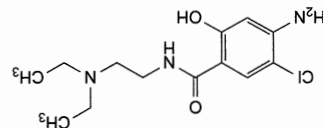
methoxybenzoic acid,

D. R1 = OCH₃; R2 = CO-CH₃; R3 = H: methyl

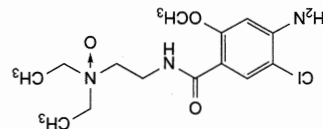
4-(acetylamino)-2-methoxybenzoate,



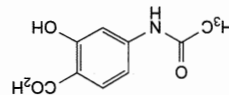
E. N,N'-diethylethane-1,2-diamine,



F. 4-amino-5-chloro-N-[2-(diethylamino)ethyl]-2-hydroxybenzamide,



G. N'-(4-amino-5-chloro-2-methoxybenzoyl)-N,N'-diethylethane-1,2-diamine N-oxide,



H. 4-(acetylamino)-2-hydroxybenzoic acid.

Ph Eur

Reference solution (a) Dissolve 20 mg of metolopranamide hydrochloride CRS in methanol R and dilute to 5 mL with the same solvent.

Reference solution (b) Dilute 5 mL of test solution (a) to 100 mL with methanol R. Dilute 1 mL of this solution to 10 mL with methanol R.

Reference solution (c) Dissolve 10 mg of N,N-diethylmethanediarnine R in methanol R and dilute to 50 mL with the same solvent.

Apply separately to the plate 5 µL of each solution. Develop over a path of 12 cm using a mixture of 2 volumes of concentrated ammonia R, 10 volumes of dioxan R, 14 volumes of methanol R and 90 volumes of methylene chloride R. Allow the plate to dry in air. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). Spray with dimethylaminobenzaldehyde solution R₁. Allow the plate to dry in air. Any spot in the chromatogram obtained with test solution (a) that has not been visualised in ultraviolet light at 254 nm is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent).

Heavy metals (2.4.8)

12 mL of solution S complies with test A for heavy metals (20 ppm). Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Water (2.5.12)

4.5 per cent to 5.5 per cent, determined on 0.500 g by the semi-micro determination of water.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.2500 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of alcohol R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume of 0.1 M sodium hydroxide added between the two points of inflexion.

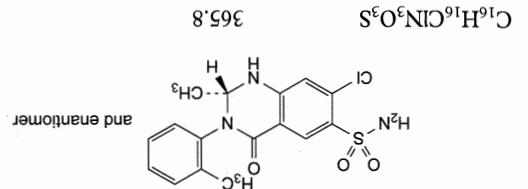
1 mL of 0.1 M sodium hydroxide is equivalent to 33.63 mg of $C_{14}H_{23}ClN_3O_2$.

STORAGE

Store protected from light.

Metolazone

(Ph. Eur. monograph 1757)



Action and use

Thiazide-like diuretic.

DEFINITION

(2*RS*)-7-Chloro-2-methyl-3-(2-methylphenyl)-4-oxo-1,2,3,4-tetrahydroquinazolin-6-sulfonamide.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder.

Solubility

Very slightly soluble in water, sparingly soluble in methanol, slightly soluble in ethyl acetate, very slightly soluble in methylene chloride.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 30.0 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Test solution (b) Dilute 2.0 mL of test solution (a) to 100.0 mL with methanol R.

Reference solution (a) Dissolve 3.0 mg of metolazone for system suitability CRS (containing impurities A, B, C, D and E) in 1 mL of methanol R.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with methanol R. Dilute 5.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (c) Dissolve 30.0 mg of metolazone CRS in methanol R and dilute to 10.0 mL with the same solvent.

Dilute 2.0 mL of this solution to 100.0 mL with methanol R. **Column:** size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 µm);

temperature: 30 °C.

Mobile phase:

— **mobile phase A:** 5.44 g/L solution of potassium dihydrogen phosphate R₃

— **mobile phase B:** methanol R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	70	30
5 - 25	70 → 50	30 → 50
25 - 35	50	50
35 - 38	50 → 70	50 → 30
38 - 48	70	30

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with metolazone for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D and E.

System suitability: reference solution (a):
— **resolution:** minimum 1.6 between the peaks due to impurities E and C and minimum 1.5 between the peaks due to impurities A and B.

Limits:

— **impurities A, B, C, D, E:** for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.5 per cent);

— **unspecified impurities:** for each impurity, not more than 0.2 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b)

(1.0 per cent);

— **disregard limit:** 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.3.2)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection: Test solution (b) and reference solution (c).

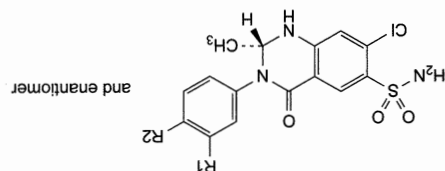
Calculate the percentage content of $C_{16}H_{16}ClN_3O_5$ from the declared content of metoprolol CRS.

STORAGE

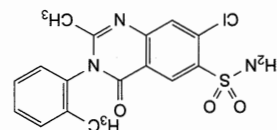
Protected from light.

IMPURITIES

Specified impurities A, B, C, D, E



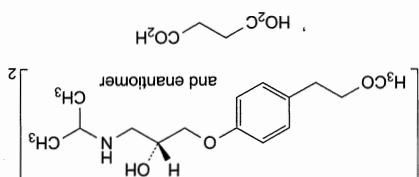
A. R₁ = CH₃, R₂ = H; (2R,5S)-7-chloro-2-methyl-3-(3-methylphenyl)-4-tetrahydroquinazolin-6-sulfonamide;
B. R₁ = H, R₂ = CH₃; (2R,5S)-7-chloro-2-methyl-3-(4-methylphenyl)-4-tetrahydroquinazolin-6-sulfonamide;
C. R₁ = R₂ = H; (2R,5S)-7-chloro-2-methyl-4-oxo-3-phenyl-1,2,3,4-tetrahydroquinazolin-6-sulfonamide;



D. 7-chloro-2-methyl-3-(2-methylphenyl)-4-tetrahydroquinazolin-6-sulfonamide;

Metoprolol Succinate

(Ph. Eur. monograph 1448)



$C_{34}H_{56}N_3O_{10}$ 653 98418-47-4

Action and use

Beta-adrenoceptor antagonist.

Ph. Eur.

DEFINITION

Bis[(2R,5S)-1-[4-(2-methoxyethyl)phenoxy]-3-[(1-methylethyl)amino]propan-2-yl] butanedioate.

Content 99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in methanol, slightly soluble in ethanol (96 per cent), very slightly soluble in ethyl acetate.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of metoprolol succinate.

TESTS

Solution S
Dissolve 0.500 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and is colourless (2.2.2, Method II).

pH (2.2.3)

7.0 to 7.6 for solution S.

Impurities M, N, O

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.50 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dilute 1 mL of the test solution to 50 mL with methanol R. Dilute 5 mL of this solution to 50 mL with methanol R.

Plate TLC silica gel plate R.

Mobile phase Place 2 beakers, each containing 30 volumes of concentrated ammonia R, at the bottom of a chromatographic

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 40 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 32.64 mg of $C_{34}H_{56}N_2O_{10}$.

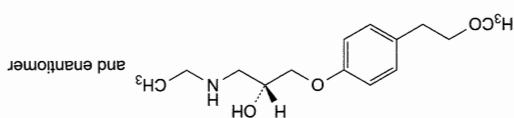
STORAGE

Protected from light.

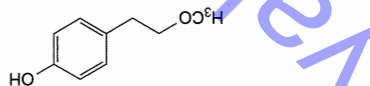
IMPURITIES

Specified impurities C, M, N, O

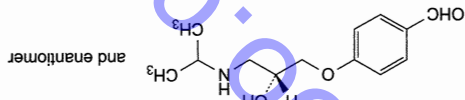
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D, E, F, G, H, J.



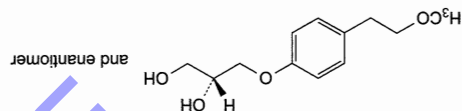
A. (2R,3S)-1-[(4-ethoxyphenyl)amino]-3-[(4-ethoxyphenyl)oxy]propan-2-ol, and enantiomer



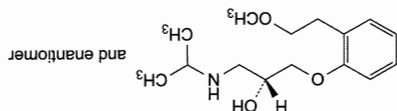
B. 4-(2-methoxyethyl)phenol, and enantiomer



C. 4-[(2R,3S)-2-hydroxy-3-[(1-methylethyl)amino]propoxy]benzaldehyde, and enantiomer



D. (2R,3S)-3-[(4-ethoxyphenyl)oxy]-1,2-diol, and enantiomer



E. (2R,3S)-1-[(4-ethoxyphenyl)oxy]-3-[(4-ethoxyphenyl)amino]propan-2-ol, and enantiomer

tank containing a mixture of 20 volumes of methanol R and 80 volumes of ethyl acetate R.

Application 10 µL.

Development Over 2/3 of the plate in a tank saturated for at least 1 h.

Drying In air for at least 3 h.

Detection Expose the plate to iodine vapour for at least 15 h.

Limits:

any impurity: any spot, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.2 per cent); disregard any spot on the line of application.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 1.5 mg of metoprolol

impurity A CRS and 2.5 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

size: $l = 0.15$ m, $\phi = 3.9$ mm;

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Dissolve 3.9 g of ammonium acetate R in 810 mL of water R, add 2.0 mL of triethylamine R, 3.0 mL of phosphoric acid R, 10.0 mL of glacial acetic acid R and 146 mL of acetonitrile R and mix.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 3 times the retention time of metoprolol.

Relative retention With reference to metoprolol (retention time = about 7 min): impurity C = about 0.4;

impurity A = about 0.8.

System suitability: reference solution (a):

resolution: minimum 6.0 between the peaks due to impurity A and metoprolol.

Limits:

correction factor: for the calculation of content, multiply the peak area of impurity C by 0.1;

impurity C: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to succinic acid.

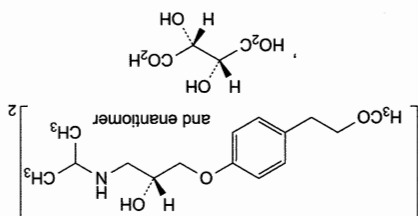
Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Metoprolol Tartrate

(Ph. Eur. monograph 1028)



C₃₄H₅₆N₂O₁₂ 685 56392-17-7

Action and use
Beta-adrenoceptor antagonist.

Preparations

Metoprolol Injection
Metoprolol Tartrate Tablets
Prolonged-release Metoprolol Tartrate Tablets

DEFINITION

Bis[(2*R*)-1-[(4-(2-methoxyethyl)phenoxy)-3-[(1-methylethyl)amino]propan-2-yl] (2*R*,3*R*)-2,3-dihydroxybutanedioate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent). It shows polymorphism (5.9).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison metoprolol tartrate CRS.

If the spectra obtained in the solid state show differences, record further spectra using discs prepared by placing 25 µL of a 100 g/L solution in methylene chloride R on a disc of potassium bromide R and evaporating the solvent. Examine immediately.

TESTS

Solution S
Dissolve 0.500 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

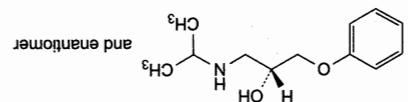
Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₈ (2.2.2, Method II).

pH (2.2.3)

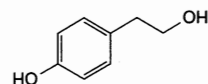
6.0 to 7.0 for solution S.

Specific optical rotation (2.2.7)

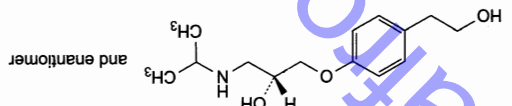
+ 7.0 to + 10.0 (dried substance), determined on solution S. Thin-layer chromatography (2.2.27).
Test solution Dissolve 0.50 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.



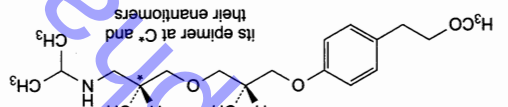
F. (2*R*)-1-[(1-methylethyl)amino]-3-phenoxypropan-2-ol,



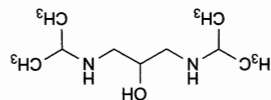
G. 2-(4-hydroxyphenyl)ethanol,



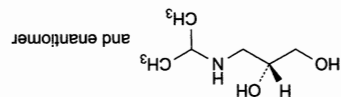
H. (2*R*)-1-[(4-(2-hydroxyethyl)phenoxy)-3-[(1-methylethyl)amino]propan-2-ol,



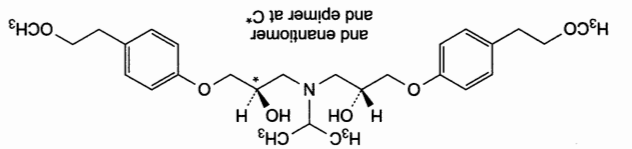
J. mixture of the 4 stereoisomers of 1-[2-hydroxy-3-[(1-methylethyl)amino]propoxy]-3-[(2-methoxyethyl)phenoxy]propan-2-ol,



M. 1,3-bis[(1-methylethyl)amino]propan-2-ol,



N. (2*R*)-3-[(1-methylethyl)amino]propan-1,2-diol,



O. mixture of the 3 stereoisomers of 1,1'-bis[(1-methylethyl)amino]bis[3-[(4-(2-methoxyethyl)phenoxy]propan-2-ol],

Ph Eur

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to tartaric acid.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* over anhydrous calcium chloride R for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

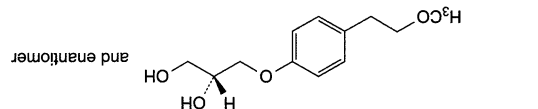
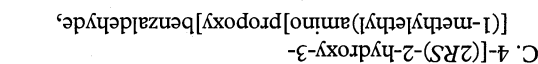
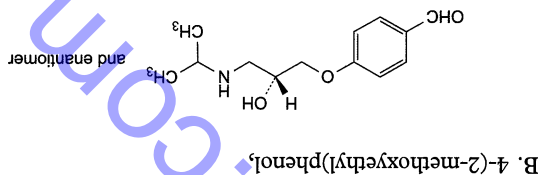
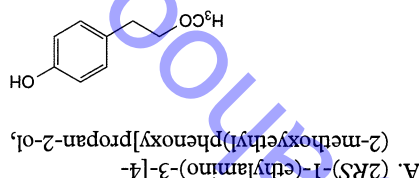
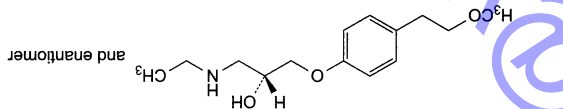
1 mL of 0.1 M perchloric acid is equivalent to 34.24 mg of $C_{34}H_{56}N_2O_{12}$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C, D, E, F, G, H, J, M, N, O



Reference solution (a) Dilute 1 mL of the test solution to 20 mL with methanol R. Dilute 5 mL of this solution to 50 mL with methanol R.

Reference solution (b) Dilute 4 mL of reference solution (a) to 10 mL with methanol R.

Plate TLC silica gel plate R.

Mobile phase Place 2 beakers, each containing 30 volumes of concentrated ammonium R, at the bottom of a chromatographic tank containing a mixture of 20 volumes of methanol R and 80 volumes of ethyl acetate R.

Application 5 µL.

Development Over 2/3 of the plate in a tank saturated for at least 1 h.

Drying In air for at least 3 h.

Detection Expose the plate to iodine vapour for at least 15 h.

Limits:

— any impurity: any spot, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent) and at most 1 such spot is more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent);

— disregard any spot on the line of application.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 1.5 mg of metoprolol impurity A CRS and 2.5 mg of metoprolol tartrate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Dissolve 3.9 g of ammonium acetate R in 810 mL of water R, add 2.0 mL of triethylamine R, 3.0 mL of phosphoric acid R, 10.0 mL of glacial acetic acid R and 146 mL of acetonitrile R and mix.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time 3 times the retention time of metoprolol.

Relative retention With reference to metoprolol (retention time = about 7 min): impurity H = about 0.3;

impurity C = about 0.4; impurity G = about 0.45;

impurity F = about 0.7; impurity A = about 0.8;

impurity J = about 1.4; impurity D = about 1.6;

impurity E = about 1.8; impurity B = about 2.

System suitability: reference solution (a):

— resolution: minimum 6.0 between the peaks due to impurity A and metoprolol.

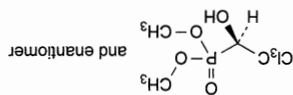
Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity C by 0.1;

— impurities A, B, C, D, E, F, G, H, J: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

Metrifonate

(Ph. Eur. monograph 1133)



52-68-6

C₄H₈Cl₃O₄P

257.4

Action and use

Anthelmintic.

Ph. Eur.

DEFINITION

Dimethyl (R₅)-(2,2,2-trichloro-1-hydroxyethyl)phosphonate.

Content

98.0 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, very soluble in methylene chloride, freely soluble in acetone and in ethanol (96 per cent).

mp

76 °C to 81 °C.

IDENTIFICATION

First identification A, B.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison metrifonate CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of metrifonate CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, dioxan R, toluene R (5:25:70 V/V/V).

Application 10 µL.

Development In an unsaturated tank over a path of 15 cm.

Drying In air.

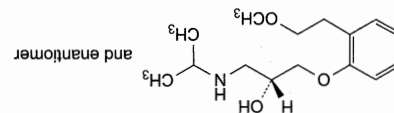
Detection Spray with a 50 g/L solution of 4-

(4-nitrobenzyl)pyridine R in acetone R and heat at 120 °C for 15 min; spray the still-warm plate with a 100 g/L solution of tetraethylene pentamine R in acetone R and examine immediately.

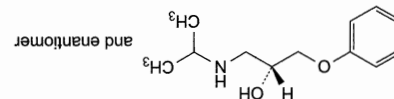
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve about 20 mg in 1 mL of dilute sodium hydroxide solution R. Add 1 mL of pyridine R. Shake and heat on a water-bath for 2 min. A red colour develops in the upper layer.

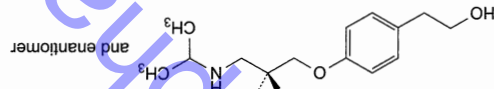
D. To 0.1 g add 0.5 mL of nitric acid R, 0.5 mL of a 500 g/L solution of ammonium nitrate R1 and 0.1 mL of strong hydrogen peroxide solution R. Heat on a water-bath for 10 min. Heat to boiling and add 1 mL of ammonium



E. (2R,5S)-1-[2-(2-methoxyethyl)phenoxy]-3-[(1-methylethyl)amino]propan-2-ol,

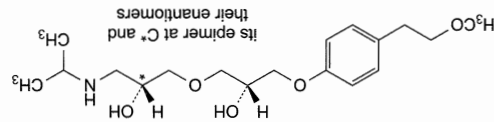


F. (2R,5S)-1-[(1-methylethyl)amino]-3-phenoxypropan-2-ol,

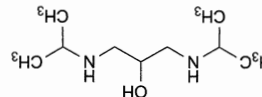


G. 2-(4-hydroxyphenyl)ethanol,

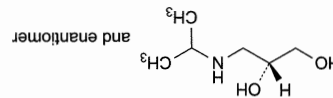
H. (2R,5S)-1-[4-(2-hydroxyethyl)phenoxy]-3-[(1-methylethyl)amino]propan-2-ol,



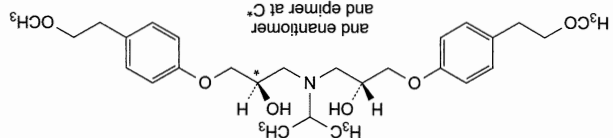
J. mixture of the 4 stereoisomers of 1-[2-hydroxy-3-[(1-methylethyl)amino]propoxy]-3-[(2-methoxyethyl)phenoxy]propan-2-ol,



M. 1,3-bis[(1-methylethyl)amino]propan-2-ol,



N. (2R,5S)-3-[(1-methylethyl)amino]propan-1,2-diol,



O. mixture of the 3 stereoisomers of 1,1'-[(1-methylethyl)imino]bis[3-[(4-(2-methoxyethyl)phenoxy)propan-2-ol],

Ph. Eur.



mobdiate solution R. A yellow colour is produced or a yellow precipitate is formed.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution *Y*₇ (2.2.2, Method II).

Dissolve 5.0 g in 20 mL of methanol *R*.

Acidity

Dissolve 2.5 g in carbon dioxide-free water *R* and dilute to

50 mL with the same solvent. Add 0.1 mL of methyl red

solution *R*. Not more than 1.0 mL of 0.1 M sodium hydroxide

is required to change the colour of the indicator to yellow.

Optical rotation (2.2.7)

–0.10° to +0.10°.

Dissolve 0.1 g in ethanol (96 per cent) *R* and dilute to

10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase B, mobile phase A (10:90 V/V).

Test solution Dissolve 0.20 g of the substance to be examined

in the solvent mixture and dilute to 10.0 mL with the solvent

mixture.

Reference solution (a) Use a freshly prepared solution. Dissolve

10.0 mg of desmethylmetrifonate CRS (impurity A) in the

solvent mixture and dilute to 20.0 mL with the solvent

mixture. Dilute 1.0 mL of this solution to 5.0 mL with the

solvent mixture.

Reference solution (b) Dissolve 0.10 g of dichlorvos *R*

(impurity B) in the solvent mixture and dilute to 50.0 mL

with the solvent mixture. Dilute 1.0 mL of this solution to

10.0 mL with the solvent mixture. Dilute 5.0 mL of this

solution to 100.0 mL with the solvent mixture.

Reference solution (d) Use a freshly prepared solution. Mix

1.0 mL of reference solution (a), 1.0 mL of reference

solution (b) and 0.025 mL of the test solution.

Reference solution (e) Dilute 4.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography *R*

(10 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: 1.36 g/L solution of potassium dihydrogen

phosphate *R*, previously adjusted to pH 2.9 with phosphoric

acid *R*;

— mobile phase B: acetonitrile *R*;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	90	10
5 - 25	90 \rightarrow 85	10 \rightarrow 15
25 - end	85 \rightarrow 45	15 \rightarrow 55

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 50 μ L.

Run time 3 times the retention time of metrifonate.

IMPURITIES

Protected from light.

Specified impurities A, B.

STORAGE

V_P = volume of silver nitrate used in the assay, in mL
 M_P = mass of substance used in the assay, in grams;
 V_{Cl} = volume of silver nitrate used in the test for chlorides, in mL
 M_{Cl} = mass of substance used in the test for chlorides, in grams.

$$\left[\frac{M_P}{V_P} - \frac{M_{Cl}}{V_{Cl} \times 0.1} \right] \times 25.74 \times 0.1$$

expression:

Calculate the percentage content of $C_4H_8Cl_3O_4P$, taking into account the content of chloride and using the following potentiometrically (2.2.20), using a silver electrode. with 0.1 M silver nitrate, determining the end-point mixture at 20–22 °C. Maintain at that temperature and titrate and 100 mL of water *R* maintaining the temperature of the 20–22 °C. Add a chilled mixture of 15 mL of nitric acid *R* Add 10 mL of ethanolic *R* and allow to stand for 1 h at Dissolve 0.300 g in 30 mL of ethanol (96 per cent) *R*.

ASSAY

Maximum 0.3 per cent, determined on 3.000 g.

Water (2.5.12)

lead standard solution (1 ppm Pb) *R*.

complies with test A. Prepare the reference solution using

Dissolve 2.0 g in 20 mL of water *R*. 12 mL of the solution

Maximum 10 ppm.

Heavy metals (2.4.8)

Cl.

1 mL of 0.01 M silver nitrate is equivalent to 0.3546 mg of

potentiometrically (2.2.20), using a silver electrode.

Titrate with 0.01 M silver nitrate determining the end-point

mixture of 15 mL of nitric acid *R* and 100 mL of water *R*.

Dissolve 5.00 g in 30 mL of ethanol (96 per cent) *R* and add a

Maximum 500 ppm.

Chlorides

(0.04 per cent).

the chromatogram obtained with reference solution (e)

— disregard limit: 0.1 times the area of the principal peak in

obtained with reference solution (c) (1 per cent);

the area of the principal peak in the chromatogram

— sum of impurities other than A and B: not more than twice

with reference solution (c) (0.5 per cent);

area of the principal peak in the chromatogram obtained

— any other impurity: for each impurity, not more than the

(0.2 per cent);

in the chromatogram obtained with reference solution (b)

— impurity B: not more than the area of the principal peak

(0.5 per cent);

in the chromatogram obtained with reference solution (a)

— impurity A: not more than the area of the principal peak

Limits:

the peaks due to metrifonate and impurity B.

— resolution: minimum 3.0 between the peaks due to

impurity A and metrifonate and minimum 4.5 between

System suitability: Reference solution (d):

Elution order Impurity A, metrifonate, impurity B.

a water-bath for 5 min. Cool. The solution gives the reaction of primary aromatic amines (2.3.1).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution GY₆ (2.2.2, Method II).

Dissolve 1.0 g in 1 M hydrochloric acid and dilute to 20 mL with the same acid.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions protected from light.

Test solution Dissolve 0.05 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase and dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of metronidazole

impurity A CRS in the mobile phase, add 10.0 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Dilute 1.0 mL to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Mix 30 volumes of methanol R and 70 volumes of a 1.36 g/L solution of potassium dihydrogen phosphate R,

Flow rate 1 mL/min.

Detection Spectrophotometer at 315 nm.

Injection 10 μ L.

Run time 3 times the retention time of metronidazole.

Relative retention With reference to metronidazole (retention time = about 7 min): impurity A = about 0.7.

System suitability: reference solution (b):

— resolution: minimum of 2.0 between the peaks due to

metronidazole and to impurity A.

Limits:

— any impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.1 per cent),

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.2 per cent),

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.01 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

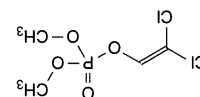
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

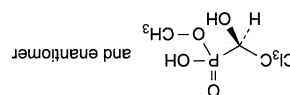
Dissolve 0.150 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

B. 2,2-dichloroethenyl dimethyl phosphite (dichlorvos).



A. methyl (RS)-(2,2,2-trichloro-1-hydroxyethyl)phosphonate acid (desmethylnetrifonate),



(Ph. Eur. monograph 0675)



Metronidazole

$C_6H_9N_3O_3$

171.2

443-48-1

Action and use

Imidazole antibacterial.

Preparations

Metronidazole Gel

Metronidazole Infusion

Metronidazole Suppositories

Metronidazole Tablets

Ph Eur

DEFINITION

2-(2-Methyl-5-nitro-1H-imidazol-1-yl)ethanol.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish, crystalline powder.

Solubility

Slightly soluble in water, in acetone, in alcohol and in

methylene chloride.

IDENTIFICATION

First identification: C.

Second identification: A, B, D.

A. Melting point (2.2.14): 159 °C to 163 °C.

B. Dissolve 40.0 mg in 0.1 M hydrochloric acid and dilute to

100.0 mL with the same acid. Dilute 5.0 mL of the solution

to 100.0 mL with 0.1 M hydrochloric acid. Examined between

230 nm and 350 nm (2.2.25), the solution shows an

absorption maximum at 277 nm and a minimum at 240 nm.

The specific absorbance at the maximum is 365 to 395.

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison metronidazole CRS.

D. To about 10 mg add about 10 mg of zinc powder R, 1 mL

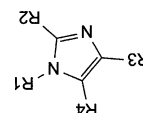
of water R and 0.25 mL of dilute hydrochloric acid R. Heat on

1 mL of 0.1 M perchloric acid is equivalent to 17.12 mg of $C_6H_9N_3O_3$.

STORAGE

Protected from light.

IMPURITIES



A. $R_1 = R_4 = H$, $R_2 = CH_3$, $R_3 = NO_2$; 2-methyl-4-nitroimidazole,

B. $R_1 = R_2 = R_4 = H$, $R_3 = NO_2$; 4-nitroimidazole,

C. $R_1 = CH_2-CH_2-OH$, $R_2 = R_4 = H$, $R_3 = NO_2$;

2-(4-nitro-1H-imidazol-1-yl)ethanol,

D. $R_1 = CH_2-CH_2-OH$, $R_2 = R_3 = H$, $R_4 = NO_2$;

2-(5-nitro-1H-imidazol-1-yl)ethanol,

E. $R_1 = CH_2-CH_2-OH$, $R_2 = CH_3$, $R_3 = NO_2$, $R_4 = H$;

2-(2-methyl-4-nitro-1H-imidazol-1-yl)ethanol,

F. $R_1 = CH_2-CH_2-O-CH_2-CH_2-$

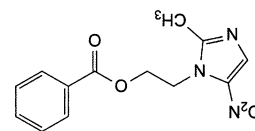
OH, $R_2 = CH_3$, $R_3 = H$, $R_4 = NO_2$; 2-[2-(2-methyl-5-nitro-1H-imidazol-1-yl)ethoxy]ethanol,

G. $R_1 = CH_2-CO_2H$, $R_2 = CH_3$, $R_3 = H$, $R_4 = NO_2$;

2-(2-methyl-5-nitro-1H-imidazol-1-yl)acetic acid.

Metronidazole Benzoate

(Ph. Eur. monograph 0934)



$C_{13}H_{13}N_3O_4$

275.3

13182-89-3

Metronidazole Oral Suspension

Preparation

Imidazole antibacterial.

Action and use

DEFINITION

2-(2-Methyl-5-nitro-1H-imidazol-1-yl)ethyl benzoate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder or flakes.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in acetone, slightly soluble in alcohol.

IDENTIFICATION

First identification C

Second identification A, B, D

A. Melting point (2.2.14): 99 °C to 102 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 0.100 g in a 103 g/L solution of hydrochloric acid R and dilute to 100.0 mL with the same acid. Dilute 1.0 mL of the solution to 100.0 mL with a 103 g/L solution of hydrochloric acid R.

Absorption maxima At 232 nm and 275 nm.

Specific absorbance at the absorption maximum at 232 nm

525 to 575.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of metronidazole benzoate.

D. To about 10 mg add about 10 mg of zinc powder R, 1 mL of water R and 0.3 mL of hydrochloric acid R. Heat on a

water-bath for 5 min and cool. The solution gives the

reaction of primary aromatic amines (2.3.1).

TESTS

Appearance of solution

The solution is not more opalescent than reference

suspension II (2.2.1) and not more intensely coloured than

reference solution GY₃ (2.2.2, Method II).

Dissolve 1.0 g in dimethylformamide R and dilute to 10 mL

with the same solvent.

Acidity

Dissolve 2.0 g in a mixture of 20 mL of dimethylformamide R

and 20 mL of water R, previously neutralised with 0.02 M

hydrochloric acid or 0.02 M sodium hydroxide using 0.2 mL of

methyl red solution R. Not more than 0.25 mL of 0.02 M

sodium hydroxide is required to change the colour of the

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase B, mobile phase A (45:55 V/V).

Test solution Dissolve 0.100 g of the substance to be

examined in the solvent mixture and dilute to 10.0 mL with

the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of metronidazole CRS

(impurity A), 5.0 mg of metronidazole impurity A CRS

(impurity B) and 5.0 mg of benzoic acid CRS (impurity C) in

the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the

solvent mixture.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: spherical diisobutyloctadecylsilyl silica gel

for chromatography R (5 μ m) with a specific surface area of

180 m²/g, a pore size of 8 nm and a carbon loading of

10 per cent.

Mobile phase:

— mobile phase A: 1.5 g/L solution of potassium dihydrogen

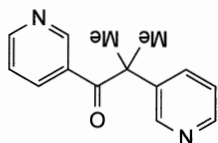
phosphate R adjusted to pH 3.2 with phosphoric acid R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	80	20
5 - 15	80 → 55	20 → 45
15 - 40	55	45

Flow rate 1.0 mL/min.

Metyrapone



$C_{14}H_{14}N_2O$ 226.3 54-36-4

Action and use

1 β -Beta-hydroxylase inhibitor; inhibition of the formation of corticosteroids.

Preparation

Metyrapone Capsules

DEFINITION

Metyrapone is 2-methyl-1,2-di(3-pyridyl)-propan-1-one. It contains not less than 97.0% and not more than 103.0% of $C_{14}H_{14}N_2O$, calculated with reference to the dried substance.

CHARACTERISTICS

A white to light amber, crystalline powder. Sparingly soluble in water; freely soluble in ethanol (96%). It dissolves in dilute mineral acids.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with reference spectrum 1 of metyrapone (RS 230). B. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.002% w/v solution in 0.1M hydrochloric acid exhibits a maximum only at 260 nm. The absorbance at 260 nm is about 1.0. C. To 5 mL of a 1% w/v solution in 1M sulfuric acid add 0.2 mL of potassium tetraiodomercurate solution. A cream precipitate is produced.

TESTS

Melting point

50° to 53°, Appendix V A.

Related substances

Carry out in subdued light the method for thin-layer chromatography, Appendix III A, using silica gel GF_{254} as the coating substance and a mixture of 90 volumes of propan-2-ol, 5 volumes of 13.5M ammonia and 5 volumes of water as the mobile phase. Apply separately to the plate 10 μ L of each of two solutions of the substance being examined in dichloromethane containing (1) 5.0% w/v and (2) 0.010% w/v. After removal of the plate, allow it to dry in air and examine under ultraviolet light (254 nm). Any secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

Loss on drying

When dried over phosphorus pentoxide at a pressure not exceeding 0.7 kPa for 6 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out the following procedure protected from light. Dissolve 0.1 g in sufficient 0.1M hydrochloric acid to produce 100 mL. Dilute 5 mL to 50 mL with 0.1M hydrochloric acid and dilute 5 mL of this solution to 50 mL with

Detection Spectrophotometer at 235 nm. Injection 10 μ L.

Relative retention With reference to metronidazole benzoate (retention time = about 20 min): impurity B = about 0.17; impurity A = about 0.20; impurity C = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurities A and B.

Limits:

— impurities A, B, C: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.01 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 80° C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

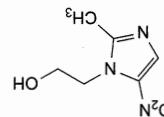
1 mL of 0.1 M perchloric acid is equivalent to 27.53 mg of $C_{14}H_{14}N_2O_4$.

STORAGE

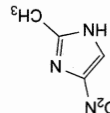
Protected from light.

IMPURITIES

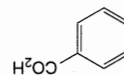
Specified impurities: A, B, C.



A. 2-(2-methyl-5-nitro-1H-imidazol-1-yl)ethanol (metronidazole),



B. 2-methyl-4-nitroimidazole,



C. benzenecarboxylic acid (benzoic acid).

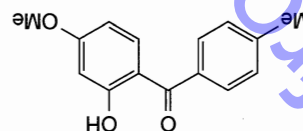
Ph Eur

0.1M hydrochloric acid. Measure the absorbance of the resulting solution at the maximum at 260 nm, Appendix II B. Calculate the content of $C_{14}H_{14}N_2O$ taking 500 as the value of A(1%, 1 cm) at the maximum at 260 nm.

STORAGE

Mexetrapone should be protected from light.

Mexenone



$C_{15}H_{14}O_3$ 242.3 1641-17-4

Action and use
Sunscreen.
Preparation
Mexenone Cream

DEFINITION

Mexenone is 2-hydroxy-4-methoxy-4'-methylbenzophenone. It contains not less than 97.0% and not more than 103.0% of $C_{15}H_{14}O_3$, calculated with reference to the dried substance.

CHARACTERISTICS

A pale yellow, crystalline powder.
Practically insoluble in water; freely soluble in acetone; sparingly soluble in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of mexenone (RS 232).

B. The high absorption, Appendix II B, in the range 230 to 350 nm of a 0.0015% w/v solution in methanol exhibits three maxima, at 243, 287 and 325 nm. The absorbances at the maxima are about 0.53, about 0.90 and about 0.66, respectively.

C. In the test for Related substances, the principal spot in the chromatogram obtained with solution (2) corresponds to the spot in the chromatogram obtained with solution (4).

TESTS

Melting point

99° to 102°, Appendix V A.

Iron

Ignite 1.0 g with 1 g of anhydrous sodium carbonate, cool, dissolve the residue in 5 mL of hydrochloric acid and dilute to 40 mL with water. 10 mL of the solution complies with the limit test for iron, Appendix VII (40 ppm).

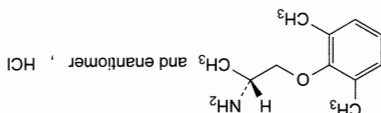
Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using silica gel GF₂₅₄ as the coating substance and a mixture of 100 volumes of toluene and 10 volumes of butan-2-one as the mobile phase. Apply separately to the plate 5 µL of each of four solutions in butan-2-one containing (1) 10% w/v of the substance being examined, (2) 0.20% w/v of the substance being examined, (3) 0.10% w/v of the substance being examined and (4) 0.20% w/v of mexenone BPCRS. After removal of the plate, allow it to dry in air and examine under ultraviolet light

Mexiletine Hydrochloride



(Ph. Eur. monograph 1029)



$C_{11}H_{18}ClNO$ 215.7 5370-01-4

Action and use
Class I antiarrhythmic.

Preparations

Mexiletine Capsules

Mexiletine Injection.

Ph Eur

DEFINITION

(2RS)-1-(2,6-Dimethylphenoxy)propan-2-amine hydrochloride.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in methanol, sparingly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison mexiletine hydrochloride CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and

record new spectra using the residues.

B. Dilute 1.5 mL of solution S (see Tests) to 15 mL with water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dilute 5 mL of solution S to 10 mL with water R.

pH (2.2.3)

4.0 to 5.5 for solution S.

Impurity D

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.500 g of the substance to be examined in methanol R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of mexiletine impurity D CRS in 4.0 mL of methanol R.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with methanol R.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 5.0 mL with methanol R.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 5.0 mL with reference solution (b).

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, ethanol (96 per cent) R, acetone R, toluene R (3:7:45:45 V/V/V/V).

Application 5 µL of the test solution and reference solutions (c) and (d).

Development Over a path of 10 cm.

Drying In air.

Detection Spray with ninhydrin solution R3 and heat at 100-105 °C for 15 min or until the spots appear.

System suitability The chromatogram obtained with reference solution (d) shows 2 clearly separated spots.

Limit:

— impurity D: any spot corresponding to impurity D in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.200 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase.

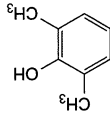
Reference solution (b) Dissolve the contents of a vial of mexiletine impurity C CRS in the mobile phase and transfer the solution quantitatively to a volumetric flask containing 16.0 mg of 2,6-dimethylphenol R (impurity A). Dilute to 20.0 mL with the mobile phase. Mix 1.0 mL of this solution with 2.0 mL of reference solution (a) and dilute the mixture to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 65 volumes of methanol R2 and 35 volumes of a solution prepared as follows: dissolve 11.5 g of anhydrous sodium acetate R in 500 mL of water R, add 3.2 mL of glacial



A. 2,6-dimethylphenol.

acetic acid R, mix and allow to cool; adjust to pH 4.8 with glacial acetic acid R and dilute to 1000 mL with water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 262 nm.

Injection 20 µL.

Run time 5.5 times the retention time of mexiletine.

Relative retention With reference to mexiletine (retention time = about 4 min): impurity C = about 0.7;

impurity A = about 1.8.

System suitability: reference solution (b): — resolution: minimum 5.0 between the peaks due to impurity C and mexiletine.

Limits:

— impurity A: not more than 2.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity C: not more than 20 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— any other impurity: for each impurity, not more than 0.5 times the area of the peak due to mexiletine in the chromatogram obtained with reference solution (b) (0.1 per cent);

— total: not more than 2.5 times the area of the peak due to mexiletine in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.25 times the area of the peak due to mexiletine in the chromatogram and reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

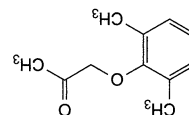
Dissolve 0.150 g in 50 mL of a mixture of equal volumes of acetic anhydride R and anhydrous acetic acid R. Titrate immediately with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20) and completing the titration within 2 min.

1 mL of 0.1 M perchloric acid is equivalent to 21.57 mg of $C_{11}H_{18}ClNO$.

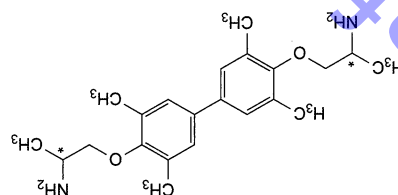
IMPURITIES

Specified impurities A, C, D

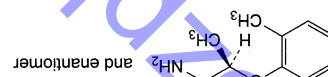
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B.



B. 1-(2,6-dimethylphenoxy)propan-2-one,



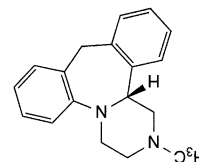
C. 1,1'-[(3,3',5,5'-tetramethylbiphenyl-4,4'-diyl)bis(oxy)]dipropylamine,



D. (2R,S)-2-(2,6-dimethylphenoxy)propan-1-amine.

Mianserin Hydrochloride

(Ph. Eur. monograph 0846)



and enantiomer, HCl

 $C_{18}H_{21}ClN_2$

300.8

21535-47-7

Action and use

Monoamine reuptake inhibitor; tetracyclic antidepressant.

Preparation

Mianserin Tablets

Ph Eur

DEFINITION

(14bR,S)-2-Methyl-1,2,3,4,10,14b-hexahydrodibenzo[*c,f*]pyrazino[1,2-*a*]azepine hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or crystals.

Solubility

Sparingly soluble in water and in methylene chloride, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B, D.
Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 50.0 mg in water R and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with water R.

Spectral range 230–350 nm.

Absorption maximum At 279 nm.

Specific absorbance at the absorption maximum 64 to 72.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison mianserin hydrochloride CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of mianserin hydrochloride CRS in methylene chloride R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of mianserin hydrochloride CRS and 10 mg of cyproheptadine hydrochloride CRS in methylene chloride R and dilute to 5 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase diethylamine R, ether R, cyclohexane R (5:20:75 V/V/V).

Application 2 µL.

Development Over 2/3 of the plate.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

pH (2.2.3)

4.0 to 5.5.

Dissolve 0.10 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Buffer solution pH 3.0 Dissolve 5.0 g of sodium octanesulfonate R in water R and dilute to 350 mL with the same solvent. Stir until complete dissolution. Adjust to pH 3.0 with a mixture of 1 volume of phosphoric acid R and 3 volumes of water R. Dilute to 400 mL with water R.

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of mianserin for system suitability CRS (containing impurities A, D and E) in 1.0 mL of the mobile phase.

Reference solution (c) Dissolve 5.0 mg of mianserin impurity B CRS in the mobile phase and dilute to 50.0 mL

with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;

— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Buffer solution pH 3.0, methanol R (37:63 V/V).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 μ L.

Run time Twice the retention time of mianserin.

Identification of impurities Use the chromatogram supplied

with mianserin for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, D and E.

Relative retention With reference to mianserin (retention

time = about 18 min): impurity B = about 0.2;

impurity A = about 0.5; impurity D = about 0.7;

impurity E = about 1.1.

System suitability: reference solution (b):

— peak-to-valley ratio: minimum 4.0, where H_p = height

above the baseline of the peak due to impurity B and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

mianserin.

Limits:

— **correction factor:** for the calculation of content, multiply the

peak areas of the following impurities by the

corresponding correction factor: impurity A = 2.4;

impurity D = 2.1;

— **impurity B:** not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (c) (0.3 per cent);

— **impurities A, D, E:** for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— **unspecified impurities:** for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— **total:** not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

over diphosphorus pentoxide R at 65 °C at a pressure not

exceeding 0.7 kPa for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 5.0 mL of 0.01 M

hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry

out a potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 30.08 mg of

$C_{18}H_{21}ClN_2$.

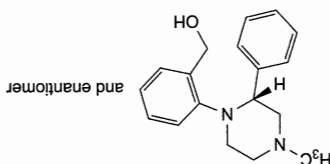
STORAGE

Protected from light.

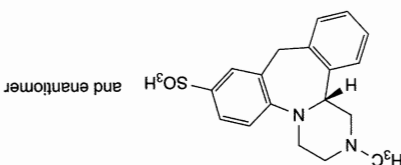
IMPURITIES

Specified impurities A, B, D, E

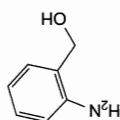
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C, F.



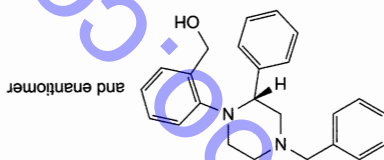
A. [2-[(2R)-4-methyl-2-phenylpiperazin-1-yl]phenyl]methanol, and enantiomer



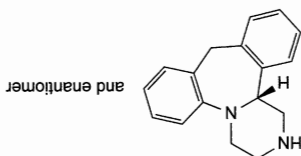
B. (14bRS)-2-methyl-1,2,3,4,10,14b-hexahydrodibenzoc[5,7]pyrazino[1,2-a]azepine-8-sulfonic acid, and enantiomer



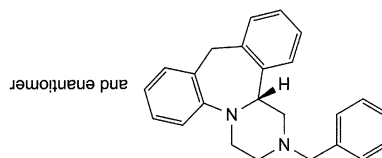
C. (2-aminophenyl)methanol, and enantiomer



D. [2-[(2R)-4-benzyl-2-phenylpiperazin-1-yl]phenyl]methanol, and enantiomer



E. (14bRS)-1,2,3,4,10,14b-hexahydrodibenzoc[5,7]pyrazino[1,2-a]azepine, and enantiomer

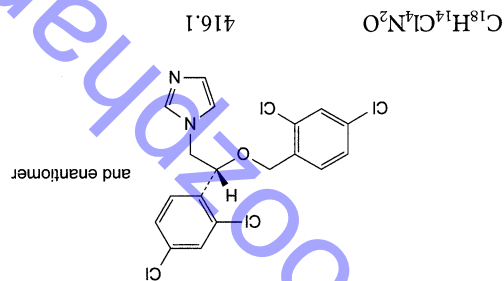


F. (14bRS)-2-benzyl-1,2,3,4,10,14b-hexahydrotrodoibenzo[c,j]pyrazino[1,2-a]azepine.

Ph Eur



(Ph. Eur. monograph 0935)



22916-47-8

Miconazole

Action and use
Antifungal azole.

Preparations
Miconazole Oromucosal Gel
Miconazole Eye Drops

DEFINITION

1-[(2RS)-2-[(2,4-Dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Melting point (2.2.14): 83 °C to 87 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of potassium bromide R.

Comparison miconazole CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 5 mL with the mobile phase.

Reference solution (a) Dissolve 30 mg of miconazole CRS in the mobile phase and dilute to 5 mL with the mobile phase.

Reference solution (b) Dissolve 30 mg of miconazole CRS and 30 mg of econazole nitrate CRS in the mobile phase and dilute to 5 mL with the mobile phase.

to 5 mL with the mobile phase.

Plate TLC octadecylsilyl silica gel plate R.
Mobile phase ammonium acetate solution R, dioxan R, methanol R (20:40:40 V/V/V).
Application 5 µL.
Development Over a path of 15 cm.
Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 30 mg in a porcelain crucible add 0.3 g of anhydrous sodium carbonate R. Heat over an open flame for 10 min.

Allow to cool. Take up the residue with 5 mL of dilute nitric acid R and filter. To 1 mL of the filtrate add 1 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.1 g in methanol R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Optical rotation (2.2.7)

—0.10° to +0.10°, determined on solution S.

Related substances

Test solution To 0.100 g of the substance to be examined add 3.0 mL of acetonitrile R and 3.2 mL of methanol R and shake until complete dissolution. Further dilute to 10.0 mL with a 15.8 g/L solution of ammonium acetate R.

Reference solution (a) Dissolve 2.5 mg of miconazole CRS and 2.5 mg of econazole nitrate CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Column:

— size: l = 0.10 m, Ø = 4.6 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (3 µm).

Mobile phase Dissolve 6.0 g of ammonium acetate R in a mixture of 300 mL of acetonitrile R, 320 mL of methanol R and 380 mL of water R.

Flow rate 2 mL/min.

Detection Spectrophotometer at 235 nm.

Equilibration With the mobile phase for about 30 min.

Injection 10 µL.

Run time 1.2 times the retention time of miconazole.

Retention time Econazole = about 10 min;

miconazole = about 20 min.

System suitability: reference solution (a):

— resolution: minimum 10 between the peaks due to econazole and miconazole; if necessary, adjust the composition of the mobile phase.

Limits:

— impurities A, B, C, D, E, F, G: for each impurity, not more than the area of the principal peak in the

chromatogram obtained with reference solution (b)
(0.25 per cent);
— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 4 h.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 50 mL of a mixture of 1 volume of anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R. Using 0.2 mL of naphtholbenzenesulfonic acid R as indicator, titrate with 0.1 M perchloric acid until the colour changes from orange-yellow to green.
1 mL of 0.1 M perchloric acid is equivalent to 41.61 mg of $C_{18}H_{14}Cl_4N_2O$.

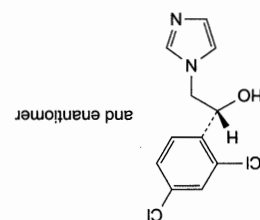
STORAGE

Protected from light.

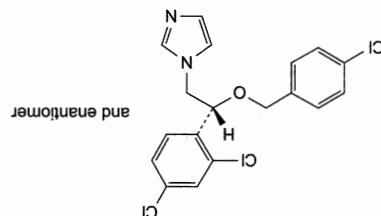
IMPURITIES

Specified impurities: A, B, C, D, E, F, G.

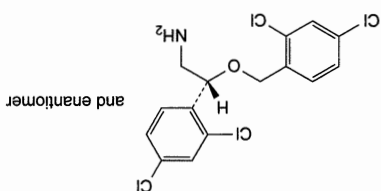
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): H, I.



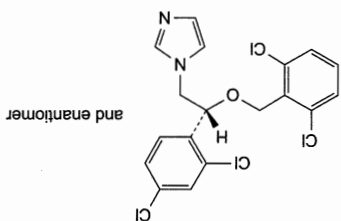
A. (1R)-1-(2,4-dichlorophenyl)-2-(1H-imidazol-1-yl)ethanol,



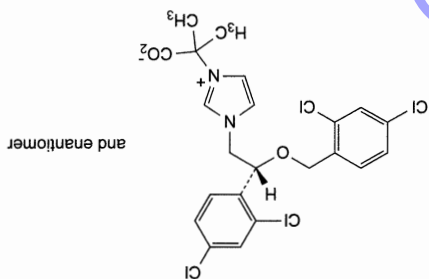
B. 1-[(2R)-2-[(4-chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole,



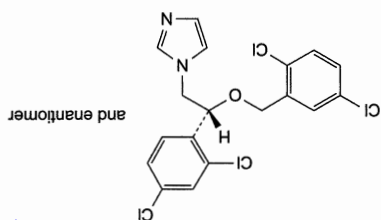
C. (2R)-2-[(2,4-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethanamine,



D. 1-[(2R)-2-[(2,6-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole,



E. 2-[1-[(2R)-2-[(2,4-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazol-3-yl]-2-methylpropanoate,



F. 1-[(2R)-2-[(3,4-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole,

G. 1-[(2R)-2-[(2,5-dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole,

Second identification A, C, D.
A. Melting point (2.2.14): 178 °C to 184 °C.
B. Infrared absorption spectrophotometry (2.2.24).
C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 5 mL with the mobile phase.
Reference solution (a) Dissolve 30 mg of miconazole nitrate CRS in the mobile phase and dilute to 5 mL with the mobile phase.

Reference solution (b) Dissolve 30 mg of miconazole nitrate CRS and 30 mg of econazole nitrate CRS in the mobile phase, then dilute to 5 mL with the mobile phase.
Plate TLC octadecylsilyl silica gel plate R.
Mobile phase ammonium acetate solution R, dioxan R, methanol R (20:40:40 V/V/V).

Application 5 µL.
Development Over 3/4 of the plate.
Drying In a current of warm air for 15 min.
Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability: reference solution (b): the chromatogram shows 2 clearly separated spots.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).
D. It gives the reaction of nitrates (2.3.1).

TESTS

Solution S
 Dissolve 0.1 g in methanol R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

Related substances

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.
Reference solution (a) Dissolve 2.5 mg of miconazole nitrate CRS and 2.5 mg of econazole nitrate CRS in the mobile phase, then dilute to 100.0 mL with the mobile phase.
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 20.0 mL with the mobile phase.

Column:

— size: l = 0.10 m, Ø = 4.6 mm;
 — stationary phase: octadecylsilyl silica gel for chromatography R (3 µm).

Mobile phase Dissolve 6.0 g of ammonium acetate R in a mixture of 300 mL of acetonitrile R, 320 mL of methanol R and 380 mL of water R.

Flow rate 2 mL/min.

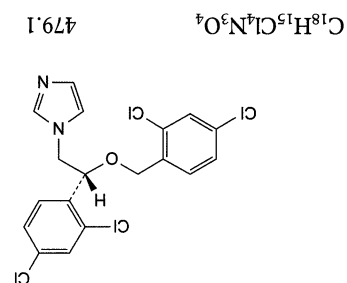
Detection Spectrophotometer at 235 nm.

Injection 10 µL.

Run time 1.2 times the retention time of miconazole.

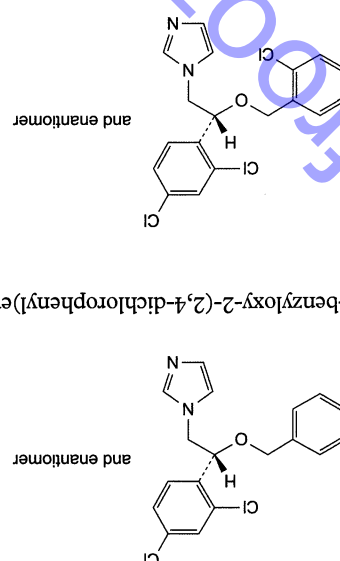
Miconazole Nitrate

(Ph. Eur. monograph 0513)

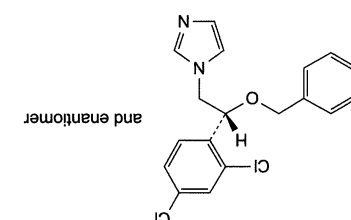


22832-87-7

I. 1-[(2RS)-2-[(2-chlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole.



H. 1-[(2RS)-2-benzyl-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole.



and enantiomer

Preparation

Miconazole and Hydrocortisone Cream
 Miconazole and Hydrocortisone Acetate Cream
 Miconazole and Hydrocortisone Ointment

DEFINITION

1-[(2RS)-2-[(2,4-Dichlorobenzyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole nitrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B.

Relative retention With reference to miconazole (retention time = about 20 min): impurity A = about 0.1; impurity E = about 0.3; impurity C = about 0.4; econazole = about 0.5; impurity B = about 0.6; impurity D = about 0.75; impurity F = about 0.85; impurity G = about 0.9.

System suitability: reference solution (a):

— **resolution:** minimum 10 between the peaks due to econazole and miconazole.

Limits:

— **impurities A, B, C, D, E, F, G:** for each impurity, not more than the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.25 per cent);

— **unspecified impurities:** for each impurity, not more than 0.4 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.5 per cent);

— **disregard limit:** 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent); disregard any peak due to the nitrate ion.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 75 mL of anhydrous acetic acid R, with slight heating if necessary. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 47.91 mg of $C_{18}H_{15}Cl_4N_3O_4$.

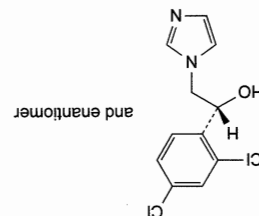
STORAGE

Protected from light.

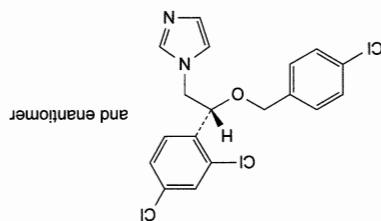
IMPURITIES

Specified impurities A, B, C, D, E, F, G

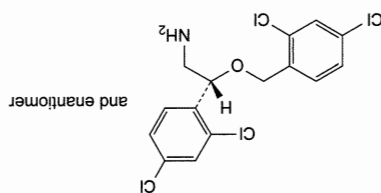
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): H, I.



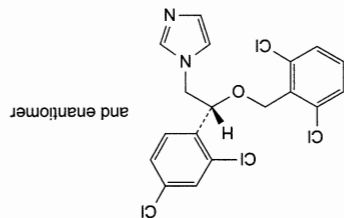
A. (1R,5S)-1-(2,4-dichlorophenyl)-2-(1H-imidazol-1-yl)ethanol, and enantiomer



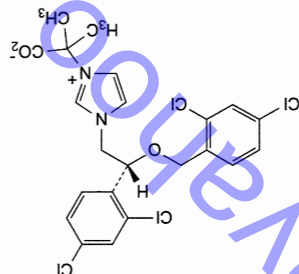
B. 1-[(2R,5S)-2-[(4-chlorobenzoyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole, and enantiomer



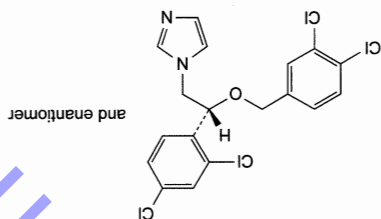
C. (2R,5S)-2-[(2,4-dichlorobenzoyl)oxy]-2-(2,4-dichlorophenyl)ethanamine, and enantiomer



D. 1-[(2R,5S)-2-[(2,6-dichlorobenzoyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole, and enantiomer



E. 2-[1-[(2R,5S)-2-[(2,4-dichlorobenzoyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazol-3-yl]-2-methylpropanoate, and enantiomer



F. 1-[(2R,5S)-2-[(3,4-dichlorobenzoyl)oxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole, and enantiomer

CHARACTERS**Appearance**

White or yellowish, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone and in ethanol (96 per cent), soluble in methanol.

IDENTIFICATION**First identification B.**

Second identification A, C, D, E.

A. Melting point (2.2.14): 161 °C to 164 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison midazolam CRS.

C. Examine the chromatograms obtained in the test for impurity C.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference

solution (b).

D. Mix 90 mg with 0.30 g of anhydrous sodium carbonate R

and ignite in a crucible until an almost white residue is

obtained (normally in less than 5 min). Allow to cool and

dissolve the residue in 5 mL of dilute nitric acid R. Filter (the

filtrate is also used in identification test E). Add 1.0 mL of

the filtrate to a freshly prepared mixture of 0.1 mL of

alizarin S solution R and 0.1 mL of zirconyl nitrate solution R.

Mix, allow to stand for 5 min and compare the colour of the

solution with that of a blank prepared in the same manner.

The test solution is yellow and the blank solution is red.

E. To 1 mL of the filtrate obtained in identification test D

add 1 mL of water R. The solution gives reaction (a) of

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 0.1 g in 0.1 M hydrochloric acid and dilute to 10 mL

with the same acid.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be

examined in methanol R and dilute to 50.0 mL with the same

solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with methanol R. Dilute 1.0 mL of this solution to

10.0 mL with methanol R.

Reference solution (b) Dissolve the contents of a vial of

midazolam for system suitability CRS (containing

impurities A, B, E, G and H) in 1.0 mL of methanol R.

Column:

— size: l = 0.25 m, Ø = 4.0 mm;

— stationary phase: end-capped octylsilyl silica gel for

chromatography R (5 µm).

Mobile phase Prepare a solution containing 7.7 g/L of

ammonium acetate R and 10 mL/L of tetrabutylammonium

hydroxide solution (400 g/L) R and adjust to pH 5.3 with

glacial acetic acid R. Mix 44 volumes of this solution with

56 volumes of methanol R.

Flow rate 1.0 mL/min.

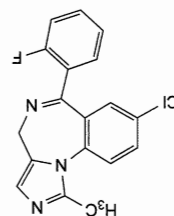
Detection Spectrophotometer at 254 nm.

Injection 10 µL.

Run time 2.5 times the retention time of midazolam.

Midazolam

(Ph. Eur. monograph 0936)



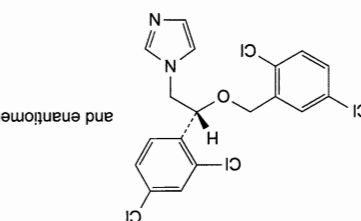
C₁₈H₁₃ClFN₃

325.8

59467-70-8



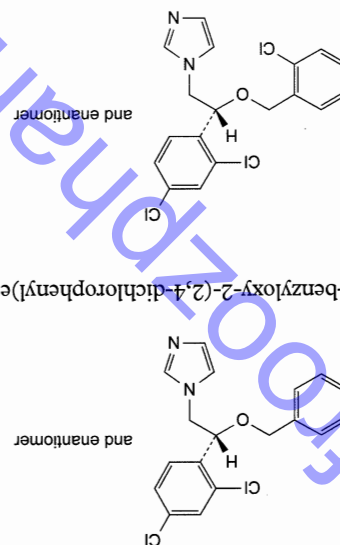
Ph. Eur.



G. 1-[(2R,5-dichlorobenzyl)oxy]-2-[(2,4-dichlorophenyl)ethyl]-1H-imidazole,

and enantiomer

H. 1-[(2R,5-dichlorobenzyl)oxy]-2-[(2,4-dichlorophenyl)ethyl]-1H-imidazole,



I. 1-[(2R,5-dichlorobenzyl)oxy]-2-[(2,4-dichlorophenyl)ethyl]-1H-imidazole.

Midazolam Injection

Midazolam Oral Solution

Midazolam Oromucosal Solution

Action and use

Benzodiazepine.

Preparations

DEFINITION

8-Chloro-6-(2-fluorophenyl)-1-methyl-4H-imidazo[1,5-a][1,4]benzodiazepine.

Content

98.5 per cent to 101.5 per cent (dried substance).

Identification of impurities Use the chromatogram supplied

with midazolam for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B, E, G and H.

Relative retention With reference to midazolam (retention

time = about 17 min): impurity B = about 0.5;

impurity A = about 0.9; impurity G = about 1.2;

impurity H = about 1.9; impurity B = about 2.2.

System suitability:

— **signal-to-noise ratio:** minimum 40 for the principal peak in

the chromatogram obtained with reference solution (a);

— **peak-to-valley ratio:** minimum 3.0, where H_p = height

above the baseline of the peak due to impurity A and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

midazolam in the chromatogram obtained with reference

solution (b).

Limits:

— **correction factors:** for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity A = 2.0;

impurity B = 2.0; impurity H = 1.7;

— **impurity B:** not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.2 per cent);

— **impurities A, E, G, H:** for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— **unspecified impurities:** for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— **total:** not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.3 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Impurity C

Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.20 g of the substance to be

examined in ethanol (96 per cent) R and dilute to 5 mL with

the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 50 mL

with ethanol (96 per cent) R.

Reference solution (a) Dissolve the contents of a vial of

midazolam impurity C CRS in 2.0 mL of methanol R.

Reference solution (b) Dissolve 8 mg of midazolam CRS in

ethanol (96 per cent) R and dilute to 10 mL with the same

solvent.

Reference solution (c) Dissolve 40 mg of the substance to be

examined in 1 mL of reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase glacial acetic acid R, water R, methanol R, ethyl

acetate R (2:15:20:80 V/V/V/V).

Application 5 µL.

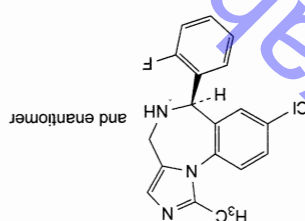
Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated spots.



Specified impurities A, B, C, E, G, H
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, F, I, J.

IMPURITIES

of C₁₈H₁₃ClFN₃.

1 mL of 0.1 M perchloric acid is equivalent to 16.29 mg

potentiometrically (2.2.20).

Dissolve 0.120 g in 30 mL of anhydrous acetic acid R and add 20 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid to the 2nd point of inflexion, determining the end-point

ASSAY

crucible.

Maximum 0.1 per cent, determined on 1.0 g in a platinum

Sulfated ash (2.4.14)

an oven at 105 °C for 2 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

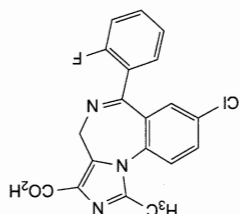
reference solution (a) (0.1 per cent).

chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with

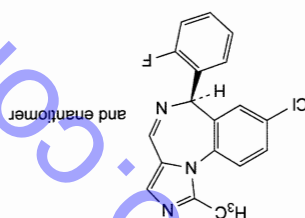
— **impurity C:** any spot due to impurity C in the

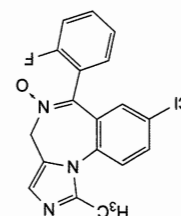
Limit:

C. 8-chloro-6-(2-fluorophenyl)-1-methyl-4H-imidazo[1,5-a][1,4]benzodiazepine-3-carboxylic acid,

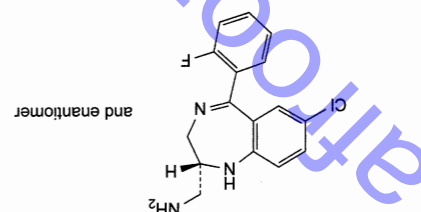


B. (6R,8S)-8-chloro-6-(2-fluorophenyl)-1-methyl-6H-imidazo[1,5-a][1,4]benzodiazepine,

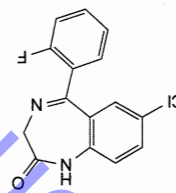




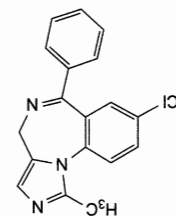
D. 8-chloro-6-(2-fluorophenyl)-1-methyl-4H-imidazo[1,5-a][1,4]benzodiazepine 5-oxide,



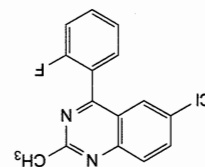
E. [(2*RS*)-7-chloro-5-(2-fluorophenyl)-2,3-dihydro-1*H*-1,4-benzodiazepin-2-yl][methanamine],



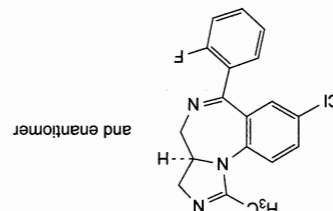
F. 7-chloro-5-(2-fluorophenyl)-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one (1-des[(diethylamino)ethyl]flurazepam),



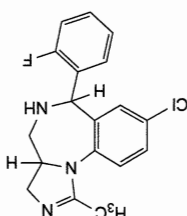
G. 8-chloro-1-methyl-6-phenyl-4*H*-imidazo[1,5-a][1,4]benzodiazepine (desfluoromidazolam),



H. 6-chloro-4-(2-fluorophenyl)-2-methylquinazoline,



I. (3*AR*)-8-chloro-6-(2-fluorophenyl)-1-methyl-3*a*,4-dihydro-3*H*-imidazo[1,5-a][1,4]benzodiazepine,



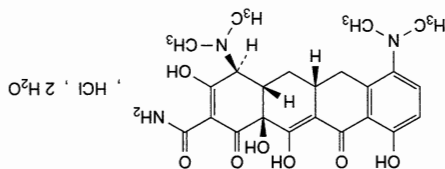
J. 8-chloro-6-(2-fluorophenyl)-1-methyl-3*a*,4,5,6-tetrahydro-3*H*-imidazo[1,5-a][1,4]benzodiazepine.

Ph Eur



Minocycline Hydrochloride Dihydrate

Minocycline Hydrochloride
(Ph. Eur. monograph 1030)



$C_{23}H_{28}ClN_3O_7 \cdot 2H_2O$ 530.0 13614-98-7

Action and use

Tetracycline antibacterial.

Preparations

Prolonged-release Minocycline Capsules

Minocycline Tablets

Ph Eur

DEFINITION

(4*S*,4*aS*,5*aR*,12*aS*)-4,7-Bis(dimethylamino)-3,10,12,12*a*-tetrahydroxy-1,11-dioxo-1,4,4*a*,5,5*a*,6,11,12*a*-octahydrotetracene-2-carboxamide hydrochloride dihydrate. Semi-synthetic product derived from a fermentation product. 96.0 per cent to 102.5 per cent (anhydrous substance).

CHARACTERS

Appearance

Yellow, hygroscopic, crystalline powder.

Solubility

Sparingly soluble in water, slightly soluble in ethanol (96 per cent). It dissolves in solutions of alkali hydroxides and carbonates.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.
Reference solution (a) Dissolve 5 mg of minocycline hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 5 mg of minocycline hydrochloride CRS and 5 mg of oxytetracycline hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a 63 g/L solution of oxalic acid R previously adjusted to pH 2 with concentrated ammonia R.

Application 1 µL

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of sulfuric acid R. A bright yellow colour develops. Add 2.5 mL of water R to the solution. The solution becomes pale yellow.

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.200 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and its absorbance (2.2.25) at 450 nm using a 1 cm cell is not greater than 0.23.

Dilute 1.0 mL of solution S to 10.0 mL with water R.

pH (2.2.3)

3.5 to 4.5 for solution S.

Light-absorbing impurities

Carry out the measurement within 1 h of preparing solution S. The absorbance (2.2.25) of solution S measured at 560 nm is not greater than 0.06.

Related substances

Liquid chromatography (2.2.29).

Carry out the test protected from bright light. Store the solutions at a temperature of 2–8 °C and use them within 3 h of preparation.

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Test solution (b) Dilute 10.0 mL of test solution (a) to 20.0 mL with the mobile phase.

Reference solution (a) Dissolve 12.5 mg of minocycline hydrochloride CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 2.0 mL of test solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.2 mL of test solution (a) to 100.0 mL with the mobile phase.

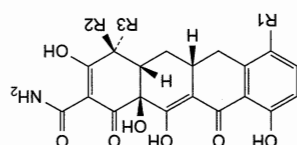
Reference solution (d) Dissolve 10 mg of minocycline hydrochloride CRS in 1 mL of water R. Boil the solution on a water-bath for 20 min. Dilute to 25 mL with the mobile phase.

Column:

— size: $l = 0.20$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 25 volumes of a 4 g/L solution of sodium edetate R, 27 volumes of dimethylformamide R and 50 volumes of a 28 g/L solution of ammonium oxalate R, and adjust to pH 7.0 with tetrabutylammonium hydroxide solution (104 g/L) R.



Specified impurities: A, B, C, D.

IMPURITIES

In an airtight container, protected from light. If the substance is sterile, store in a sterile, tamper-proof container.

STORAGE

Calculate the percentage content of $C_{23}H_{28}ClN_3O_7$ from the declared content of minocycline hydrochloride CRS.

6 injections of reference solution (a).

peak area for minocycline of 1.5 per cent after

— repeatability: maximum relative standard deviation of the

System suitability:

Injection Test solution (b) and reference solution (a).

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

ASSAY

procedure for the removal of bacterial endotoxins.

of parenteral preparations without a further appropriate

Bacterial endotoxins (2.6.14)

Less than 1.25 IU/mg, if intended for use in the manufacture

Sulfated ash (2.4.14)

5.0 per cent to 8.0 per cent, determined on 0.500 g.

Water (2.5.12)

using 2.5 mL of lead standard solution (10 ppm Pb) R.

Maximum 50 ppm.

Heavy metals (2.4.8)

reference solution (b) (2.0 per cent).

the principal peak in the chromatogram obtained with

— total of impurities other than A: not more than the area of

with reference solution (c) (1.2 per cent);

area of the principal peak in the chromatogram obtained

— any other impurity: for each impurity, not more than the

(1.2 per cent);

in the chromatogram obtained with reference solution (c)

— impurity A: not more than the area of the principal peak

Limits:

obtained with reference solution (a).

the peak due to minocycline in the chromatogram

— number of theoretical plates: minimum 3000, calculated for

obtained with reference solution (d);

impurity A and minocycline in the chromatogram

— resolution: minimum 2.0 between the peaks due to

System suitability:

Run time 1.5 times the retention time of minocycline.

(b), (c) and (d).

Injection 20 µL of test solution (a) and reference solutions (a),

Detection Spectrophotometer at 280 nm.

Flow rate 1 mL/min.

A. R1 = R3 = N(CH₃)₂, R2 = H: (4R,4aS,5aR,12aS)-4,7-bis(dimethylamino)-3,10,12,12a-tetrahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide (4-epiminocycline),

B. R1 = R3 = H, R2 = N(CH₃)₂: (4S,4aS,5aR,12aS)-4-

(dimethylamino)-3,10,12,12a-tetrahydroxy-1,11-

dioxo-1,4,4a,5,5a,6,11,12a-octahydrodrotetracene-2-carboxamide

(sancycline),

C, R1 = NH-CH₃, R2 = N(CH₃)₂, R3 = H;

(4S,4aS,5aR,12aS)-4-(dimethylamino)-3,10,12,12a-

tetrahydroxy-7-(methylamino)-1,11-dioxo-

1,4,4a,5,5a,6,11,12a-octahydrodrotetracene-2-carboxamide

(7-monodemethylminocycline),

D, R1 = NH₂, R2 = N(CH₃)₂, R3 = H;

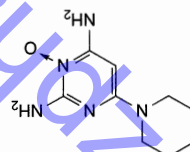
(4S,4aR,12aS)-7-amino-4-(dimethylamino)-3,10,12,12a-

tetrahydroxy-1,11-dioxo-1,4,4a,5,5a,6,11,12a-

octahydrodrotetracene-2-carboxamide (7-aminosancycline).

Ph Eur

Minoxidil
(Ph. Eur. monograph 0937)



C₉H₁₅N₅O 209.3 38304-91-5

Action and use
Vasodilator; treatment of hypertension; male pattern baldness.

Preparation
Minoxidil Scalp Application

Ph Eur

DEFINITION

6-(Piperidin-1-yl)pyrimidine-2,4-diamine 3-oxide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, soluble in methanol and in propylene glycol.

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution (a) Dissolve 20.0 mg in 0.1 M hydrochloric acid

and dilute to 100.0 mL with the same acid (solution A).

Dilute 2.0 mL of solution A to 100.0 mL with 0.1 M

hydrochloric acid.

Test solution (b) Dilute 2.0 mL of solution A to 100.0 mL

with 0.1 M sodium hydroxide.

Spectral range 200-350 nm.

Absorption maxima At 230 nm and 281 nm for test

solution (a); at 230 nm, 262 nm and 288 nm for test

solution (b).

Specific absorbances at the absorption maxima:

— at 230 nm: 1015 to 1120 for test solution (a); 1525 to

1685 for test solution (b);

— at 262 nm: 485 to 535 for test solution (b);

— at 281 nm: 1060 to 1170 for test solution (a);

— at 288 nm: 555 to 605 for test solution (b).

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of minoxidil CRS in

methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, methanol R

(1.5:100 V/V).

Application 2 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

D. Dissolve about 10 mg in 1 mL of methanol R.

Add 0.1 mL of copper sulfate solution R. A green colour

develops. The solution becomes greenish-yellow on the

addition of 0.1 mL of dilute hydrochloric acid R.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 12.5 mg of the substance to be

examined in the mobile phase and dilute to 50.0 mL with

the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of minoxidil for system

stability CRS (containing impurities A, B and E) in the

mobile phase and dilute to 20.0 mL with the mobile phase.

Column:

— size: l = 0.10 m, Ø = 4.6 mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (2.6 µm);

— temperature: 40 °C.

Mobile phase Solution containing 0.1 per cent V/V of

trifluoroacetic acid R and 2 g/L of sodium heptanesulfonate R in

a mixture of 45 volumes of methanol R and 55 volumes of

water for chromatography R.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Run time Twice the retention time of minoxidil.

Identification of impurities Use the chromatogram supplied

with minoxidil for system stability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B and E.

Relative retention With reference to minoxidil (retention

time = about 5 min): impurity A = about 0.3;

impurity B = about 0.4; impurity E = about 1.2.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to

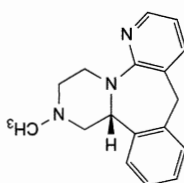
impurities A and B; minimum 2.0 between the peaks due

to minoxidil and impurity E.



Mirtazapine

(Ph. Eur. monograph 2338)



and enantiomer

C₁₇H₁₉N₃ 265.4 61337-67-5

Action and use

Inhibitor of 5HT and noradrenaline reuptake; antidepressant.

Preparations

Mirtazapine Tablets

Mirtazapine Oral Solution

Orodispersible Mirtazapine Tablets

Ph. Eur.

DEFINITION

(14bR)-2-Methyl-1,2,3,4,10,14b-hexahydropyrazino[2,1-a]pyrido[2,3-c][2]benzazepine.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder, slightly hygroscopic to

hygroscopic.

Solubility

Practically insoluble in water, freely soluble in anhydrous

ethanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison mirtazapine CRS.

If the spectra obtained in the solid state show differences,

substance separately in anhydrous ethanol R, evaporate to

dryness and record new spectra using the residues.

TESTS

Optical rotation (2.2.7)

–0.10° to +0.10° (anhydrous substance).

Dissolve 0.250 g in anhydrous ethanol R and dilute to

25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, water R (50:50 V/V).

Buffer solution Dissolve 18.0 g of tetramethylammonium

hydroxide R in 950 mL of water R. While stirring, adjust to

pH 7.4 with phosphoric acid R, then dilute to 1000 mL with

water R and mix.

Test solution Dissolve 30 mg of the substance to be examined

in the solvent mixture and dilute to 20 mL with the solvent

mixture.

Reference solution (a) Dissolve 3 mg of mirtazapine for system

suitability CRS (containing impurities A, B, C, D, E and F)

in 2 mL of the solvent mixture.

Calculation of percentage contents:

— for impurity B, multiply the peak area by the correction

factor 1.6;

— for each impurity, use the concentration of minoxidil in

reference solution (a).

Limits:

— impurity E: maximum 0.2 per cent;

— impurity B: maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.3 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent methanol R.

Dissolve 1.0 g in 25 mL of the solvent and sonicate.

The solution complies with test H. Prepare the reference

solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 50 mL of anhydrous acetic acid R. Titrate

with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 20.93 mg

of C₁₇H₁₉N₃O.

STORAGE

Protected from light.

IMPURITIES

Specified impurities B, E

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

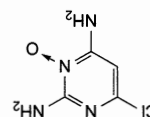
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

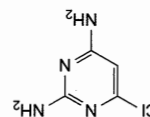
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

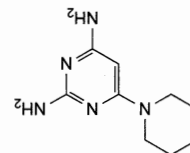
Control of impurities in substances for pharmaceutical use): A.



A. 6-chlororopyrimidine-2,4-diamine 3-oxide,



B. 6-chlororopyrimidine-2,4-diamine,



E. 6-(piperidin-1-yl)pyrimidine-2,4-diamine (deoxyminoxidil).

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase tetrahydrofuran for chromatography R , methanol R , acetonitrile R , buffer solution (7.5:12.5:15:65 V/V/V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 μ L.

Run time Twice the retention time of mirtazapine.

Identification of impurities Use the chromatogram supplied with mirtazapine for system suitability CRS and the chromatogram obtained with reference solution (a) to

identify the peaks due to impurities A, B, C, D, E and F.

Relative retention With reference to mirtazapine (retention time = about 25 min): impurity A = about 0.2;

impurity B = about 0.3; impurity C = about 0.35;

impurity D = about 0.4; impurity E = about 1.3;

impurity F = about 1.35.

System suitability:

— resolution: minimum 1.5 between the peaks due to

impurities E and F in the chromatogram obtained with

reference solution (a);

— symmetry factor: 0.8 to 2.0 for the principal peak in the

chromatogram obtained with reference solution (b).

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity A = 1.3;

impurity B = 1.3; impurity F = 0.2;

— impurities A, B, C, D, E, F: for each impurity, not more

than the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (b)

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Water (2.5.12)

Maximum 3.5 per cent, determined on 1.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 35 mL of glacial acetic acid R . Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 13.27 mg

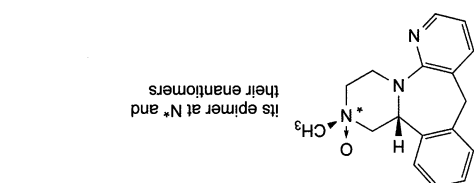
of $C_{17}H_{19}N_3$.

STORAGE

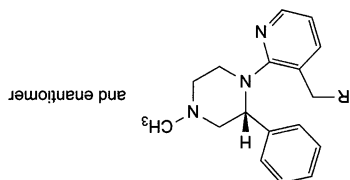
In an airtight container.

IMPURITIES

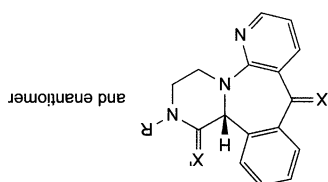
Specified impurities A, B, C, D, E, F



A. (14bRS)-2-methyl-1,2,3,4,10,14b-hexahydrotetrahydropyridino[2,1-c]pyrido[2,3-c][2]benzazepine 2-oxide,



B. $R = OH$: [2-[(2RS)-4-methyl-2-phenylpiperazin-1-yl]pyridin-3-yl]methanol,
E. $R = H$: (2RS)-4-methyl-1-(3-methylpyridin-2-yl)-2-phenylpiperazine,

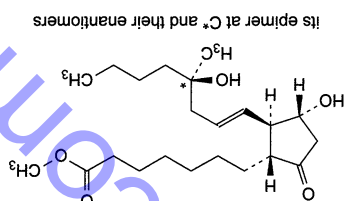


C. $R = CH_3$, $X = H_2$, $X' = O$: (14bRS)-2-methyl-3,4,10,14b-tetrahydropyridino[2,1-c]pyrido[2,3-c][2]benzazepine-1-(2H)-one,
D. $R = H$, $X = X' = H_2$: (14bRS)-1,2,3,4,10,14b-hexahydrotetrahydropyridino[2,1-c]pyrido[2,3-c][2]benzazepine,
F. $R = CH_3$, $X = O$, $X' = H_2$: (14bRS)-2-methyl-1,3,4,14b-tetrahydropyridino[2,1-c]pyrido[2,3-c][2]benzazepine-10(2H)-one.



Misoprostol

(Ph. Eur. monograph 1731)



$C_{22}H_{38}O_5$

382.5

59122-46-2

Action and use

Prostaglandin (PGE1) analogue.

DEFINITION

Mixture of methyl 7-[(1RS,2RS,3RS)-3-hydroxy-2-[(1E,4RS)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate and methyl 7-[(1RS,2RS,3RS)-3-hydroxy-2-[(1E,4SR)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate.

The 4 stereoisomers are present in approximately equal proportions.

Content

96.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, colourless or yellowish, oily liquid, hygroscopic.

Solubility

Practically insoluble in water, soluble in ethanol (96 per cent), sparingly soluble in acetonitrile.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison misoprostol CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Use freshly prepared solutions.

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 5.0 mL with the

mobile phase.

Reference solution (a) Dissolve 25.0 mg of misoprostol CRS in the mobile phase and dilute to 5.0 mL with the mobile

phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of misoprostol for system suitability CRS (containing impurities A, B and C) in 1 mL

of the mobile phase.

Column: size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: silica gel for chromatography R (5 μ m).

Mobile phase Mix 5 volumes of acetonitrile R1, 215 volumes of

dioxan R, 780 volumes of heptane R and sonicate for 10 min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L of the test solution and reference solutions (b)

and (c).

Run time 1.5 times the retention time of misoprostol.

Identification of impurities Use the chromatogram supplied

with misoprostol for system suitability CRS and the

chromatogram obtained with reference solution (c) to identify

the peaks due to impurities A, B and C.

Relative retention With reference to misoprostol (retention

time = about 18 min): impurity C = about 0.2;

impurity A = about 0.7; impurity B (1st peak) = about 0.85;

impurity B (2nd peak) = about 0.91.

System suitability: reference solution (c):

resolution: minimum 1.2 between the peaks due to

impurity B (2nd peak) and misoprostol.

Limits:

correction factor for the calculation of content, multiply the

peak area of impurity C by 0.13;

impurity B (sum of 1st and 2nd peaks): not more than

10 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(1.0 per cent);

impurity A: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.3 per cent);

impurity C: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

total: not more than 15 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (1.5 per cent);

disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Diastereoisomers

Liquid chromatography (2.2.29). Use freshly prepared solutions.

Test solution Dissolve 10.0 mg of the substance to be

examined in the mobile phase and dilute to 10.0 mL with

the mobile phase.

Column: size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: silica gel for chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase Mix 20 volumes of 2-propanol R, 40 volumes of

anhydrous ethanol R, 940 volumes of heptane R and sonicate

for 10 min.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of the 1st peak of

misoprostol.

Retention time Misoprostol 1st peak = about 19 min;

misoprostol 2nd peak = about 21 min.

System suitability Test solution:

resolution: minimum 2.0 between the 1st and 2nd peaks of

misoprostol.

Limit:

1st peak of misoprostol: 45 per cent to 55 per cent of the

sum of the areas of the 2 peaks due to misoprostol.

Water (2.5.32)

Maximum 1.0 per cent.

Use 1.0 mL of a 10 mg/mL solution of the substance to be

examined in methanol R.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modifications.

Injection 20 μ L of the test solution and reference solution (a).

System suitability: reference solution (a):

symmetry factor: maximum 3.7 for the peak due to

misoprostol.

Calculate the percentage content of C₂₂H₃₈O₅ using the

declared content of misoprostol CRS.

STORAGE

In an airtight container, at -20 °C.

IMPURITIES

Specified impurities A, B, C

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

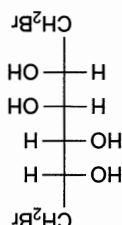
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: D, E,

F.

Mitobronitol



$\text{C}_6\text{H}_{12}\text{Br}_2\text{O}_4$ 308.0 488-41-5

Action and use

Cytotoxic.

Preparation

Mitobronitol Tablets

DEFINITION

Mitobronitol is 1,6-dibromo-1,6-dideoxy-D-mannitol. It contains not less than 98.5% and not more than 101.0% of $\text{C}_6\text{H}_{12}\text{Br}_2\text{O}_4$ calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline solid.

Slightly soluble in water, in acetone and in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of mitobronitol

(RS 236).

B. Dissolve 0.1 g in 10 mL of 1M sodium hydroxide, boil, cool, acidify with 2M nitric acid and add 1 mL of silver nitrate solution. A pale yellow, curdy precipitate is produced.
C. Dissolve 20 mg in 2 mL of a mixture of 1 volume of periodic acid solution and 24 volumes of water. Add 1 mL of 0.25M barium chloride and shake well. A white, flocculent precipitate is produced.

TESTS

Acidity

Shake 2 g with 50 mL of carbon dioxide-free water for

15 minutes and filter. 40 mL of the filtrate requires not more than 0.3 mL of 0.02M sodium hydroxide VS for neutralisation using phenolphthalein solution R1 as indicator.

Clarity and colour of solution

A 4.0% w/v solution in dimethylformamide is clear, Appendix IV A, and colourless, Appendix IV B, Method I.

Heavy metals

1.0 g complies with limit test C for heavy metals, Appendix VII (20 ppm). Use 2 mL of lead standard solution (10 ppm Pb) to prepare the standard.

Ionic halide

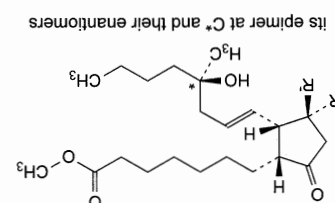
Shake 0.20 g with 30 mL of water for 5 minutes and filter. 15 mL of the filtrate complies with the limit test for chlorides, Appendix VII (500 ppm, calculated as Cl).

Loss on drying

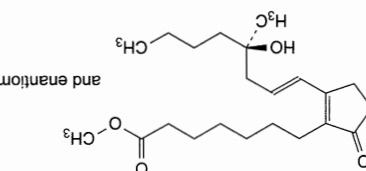
When dried at 105° for 2 hours, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

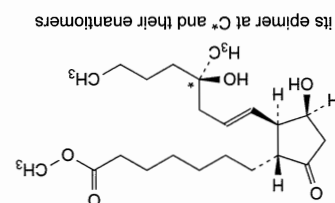
Not more than 0.2%, Appendix IX A.



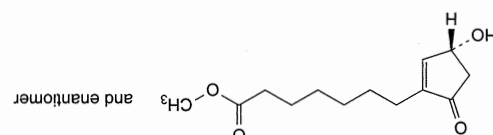
A. R = H, R' = OH: mixture of methyl 7-[(1R,2SR,3SR)-3-hydroxy-2-[(1E,4R)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate and methyl 7-[(1R,2SR,3SR)-3-hydroxy-2-[(1E,4SR)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate (8-epimisoprostol), B. R = OH, R' = H: mixture of methyl 7-[(1R,2SR,3RS)-3-hydroxy-2-[(1E,4R)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate and methyl 7-[(1R,2SR,3RS)-3-hydroxy-2-[(1E,4SR)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate (12-epimisoprostol),



D. methyl 7-2-[(1E,4R)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopent-1-enyl]heptanoate (misoprostol B),



E. mixture of methyl 7-[(1R,2RS,3SR)-3-hydroxy-2-[(1E,4R)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate and methyl 7-[(1R,2RS,3SR)-3-hydroxy-2-[(1E,4SR)-4-hydroxy-4-methyloct-1-enyl]-5-oxocyclopentyl]heptanoate (11-epi misoprostol),



F. methyl 7-[(3RS)-3-hydroxy-5-oxocyclopent-1-enyl]heptanoate.

ASSAY

Dissolve 0.2 g in 20 mL of 1M sodium hydroxide by heating gently, cool, add 25 mL of 0.1M silver nitrate VS and acidify with 5 mL of 5M nitric acid. Add 2 mL of ammonium iron(III) sulfate solution R2 and titrate the excess of silver nitrate with 0.1M ammonium thiocyanate VS. Each mL of 0.1M silver nitrate VS is equivalent to 15.40 mg of $C_{15}H_{18}N_4O_5$.

STORAGE

Mitomycinol should be protected from light.

Mitomycin

(Ph. Eur. monograph 1655)



$C_{15}H_{18}N_4O_5$ 334.3

50-07-7

Action and use

Antibacterial; cytotoxic.

Ph Eur

DEFINITION

[(1a,5,8,5,8a,8b,8c)-6-Amino-8a-methoxy-5-methyl-4,7-dioxo-1,1a,2,4,7,8a,8b-octahydrodiazirino[2',3',3',4]pyrrolo[1,2-a]indol-8-yl]methyl carbamate (mitomycin C).

Content

Substance produced by a strain of *Streptomyces caespitosus*.

CHARACTERS

Appearance

Blue-violet crystals or crystalline powder.

Solubility

Slightly soluble in water, freely soluble in dimethylacetamide, sparingly soluble in methanol, slightly soluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison mitomycin CRS.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

pH (2.2.3)

5.5 to 7.5.

Related substances

Dissolve 10 mg in 10 mL of carbon dioxide-free water R.

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 50.0 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

the same solvent.

Test solution Dissolve 50.0 mg of the substance to be examined in dimethylacetamide R and dilute to 100.0 mL with the same solvent.

Liquid chromatography (2.2.29).

ASSAY

procedure for the removal of bacterial endotoxins.

of parenteral preparations without a further appropriate

Less than 10 IU/mg, if intended for use in the manufacture

Bacterial endotoxins (2.6.14, Method B)

Maximum 2.5 per cent, determined on 0.30 g.

Water (2.5.12)

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.1 times the area of the principal peak in

(2.0 per cent),

in the chromatogram obtained with reference solution (a)

— total: not more than 4 times the area of the principal peak

with reference solution (a) (0.5 per cent),

area of the principal peak in the chromatogram obtained

— any other impurity: for each impurity, not more than the

obtained with reference solution (a) (0.5 per cent),

the area of the principal peak in the chromatogram

— impurities A, B, C, D: for each impurity, not more than

peak area of impurity A by 0.35,

— correction factor: for the calculation of content, multiply the

Limits:

mitomycin and impurity A.

— resolution: minimum 1.5 between the peaks due to

System suitability: reference solution (b):

impurity B = about 1.6.

impurity C = about 1.2; impurity A = about 1.3;

time = about 21 min; impurity D = about 0.6;

Relative retention With reference to mitomycin (retention

Injection 10 µL.

Detection Spectrophotometer at 254 nm.

Flow rate 1.0 mL/min.

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 10	100	0
10 - 30	100 → 0	0 → 100
30 - 45	0	100
45 - 50	0 → 100	100 → 0

ammonium acetate R (50:50 V/V);

— mobile phase B: methanol R, 0.77 g/L solution of

ammonium acetate R (20:80 V/V);

— mobile phase A: methanol R, 0.77 g/L solution of

Mobile phase:

— temperature: 30 °C.

— octadecylsilyl silica gel for chromatography R (5 µm),

— stationary phase: spherical base-deactivated end-capped

size

— size: $l = 0.25$ m, $\phi = 4.6$ mm,

Column:

dilute to 10 mL with methanol R.

Mix 2 mL of this solution and 1 mL of the test solution and

methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of cinnamamide R in

10.0 mL with methanol R.

100.0 mL with methanol R. Dilute 5.0 mL of this solution to

Reference solution (a) Dilute 1.0 mL of the test solution to

Reference solution (a) Dissolve 50.0 mg of mitomycin CRS in dimethylacetamide R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of cinnamamide R in methanol R and dilute to 20 mL with the same solvent.

Mix 2 mL of this solution with 2 mL of reference solution (a).

Column:

— **size:** $l = 0.30$ m, $\varnothing = 3.9$ mm,
— **stationary phase:** end-capped phenylsilyl silica gel for chromatography R (10 μ m) with a specific surface area of 330 m²/g, a carbon loading of 8 per cent and a pore size of 12.5 nm.

Mobile phase Mix 23 volumes of methanol R, 77 volumes of a solution containing 2.05 g/L of ammonium acetate R and 2.8 mL/L of dilute acetic acid R.

Flow rate 2.0 mL/min.

Detection Variable wavelength spectrophotometer capable of operating at 365 nm and 254 nm.

Injection 20 μ L.

Run time Twice the retention time of mitomycin.

Relative retention With reference to mitomycin (retention time = about 8 min): impurity A = about 1.2.

System suitability:

— **resolution:** minimum 1.8 between the peaks due to mitomycin and impurity A in the chromatogram obtained with reference solution (b) at 254 nm,
— **symmetry factor:** maximum 1.3 for the principal peak in the chromatogram obtained with reference solution (a) at 365 nm.

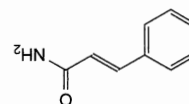
Calculate the percentage content of C₁₅H₁₈N₄O₅ from the chromatograms obtained at 365 nm and the declared content of mitomycin CRS.

STORAGE

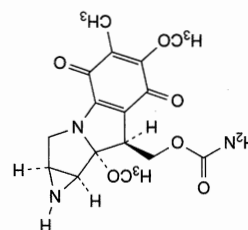
Protected from light.

IMPURITIES

Specified impurities: A, B, C, D.



A. (E)-3-phenylprop-2-enamide (cinnamamide),



B. [(1a,5,8,8a,8b,8c,8d,8e,8f,8g,8h,8i,8j,8k,8l,8m,8n,8o,8p,8q,8r,8s,8t,8u,8v,8w,8x,8y,8z,8aa,8ab,8ac,8ad,8ae,8af,8ag,8ah,8ai,8aj,8ak,8al,8am,8an,8ao,8ap,8aq,8ar,8as,8at,8au,8av,8aw,8ax,8ay,8az,8ba,8bb,8bc,8bd,8be,8bf,8bg,8bh,8bi,8bj,8bk,8bl,8bm,8bn,8bo,8bp,8bq,8br,8bs,8bt,8bu,8bv,8bw,8bx,8by,8bz,8ca,8cb,8cc,8cd,8ce,8cf,8cg,8ch,8ci,8cj,8ck,8cl,8cm,8cn,8co,8cp,8cq,8cr,8cs,8ct,8cu,8cv,8cw,8cx,8cy,8cz,8da,8db,8dc,8dd,8de,8df,8dg,8dh,8di,8dj,8dk,8dl,8dm,8dn,8do,8dp,8dq,8dr,8ds,8dt,8du,8dv,8dw,8dx,8dy,8dz,8ea,8eb,8ec,8ed,8ee,8ef,8eg,8eh,8ei,8ej,8ek,8el,8em,8en,8eo,8ep,8eq,8er,8es,8et,8eu,8ev,8ew,8ex,8ey,8ez,8fa,8fb,8fc,8fd,8fe,8ff,8fg,8fh,8fi,8fj,8fk,8fl,8fm,8fn,8fo,8fp,8fq,8fr,8fs,8ft,8fu,8fv,8fw,8fx,8fy,8fz,8ga,8gb,8gc,8gd,8ge,8gf,8gg,8gh,8gi,8gj,8gk,8gl,8gm,8gn,8go,8gp,8gq,8gr,8gs,8gt,8gu,8gv,8gw,8gx,8gy,8gz,8ha,8hb,8hc,8hd,8he,8hf,8hg,8hh,8hi,8hj,8hk,8hl,8hm,8hn,8ho,8hp,8hq,8hr,8hs,8ht,8hu,8hv,8hw,8hx,8hy,8hz,8ia,8ib,8ic,8id,8ie,8if,8ig,8ih,8ii,8ij,8ik,8il,8im,8in,8io,8ip,8iq,8ir,8is,8it,8iu,8iv,8iw,8ix,8iy,8iz,8ja,8jb,8jc,8jd,8je,8jf,8jg,8jh,8ji,8jj,8jk,8jl,8jm,8jn,8jo,8jp,8jq,8jr,8js,8jt,8ju,8jv,8jw,8jx,8jy,8jz,8ka,8kb,8kc,8kd,8ke,8kf,8kg,8kh,8ki,8kj,8kk,8kl,8km,8kn,8ko,8kp,8kq,8kr,8ks,8kt,8ku,8kv,8kw,8kx,8ky,8kz,8la,8lb,8lc,8ld,8le,8lf,8lg,8lh,8li,8lj,8lk,8ll,8lm,8ln,8lo,8lp,8lq,8lr,8ls,8lt,8lu,8lv,8lw,8lx,8ly,8lz,8ma,8mb,8mc,8md,8me,8mf,8mg,8mh,8mi,8mj,8mk,8ml,8mm,8mn,8mo,8mp,8mq,8mr,8ms,8mt,8mu,8mv,8mw,8mx,8my,8mz,8na,8nb,8nc,8nd,8ne,8nf,8ng,8nh,8ni,8nj,8nk,8nl,8nm,8nn,8no,8np,8nq,8nr,8ns,8nt,8nu,8nv,8nw,8nx,8ny,8nz,8oa,8ob,8oc,8od,8oe,8of,8og,8oh,8oi,8oj,8ok,8ol,8om,8on,8oo,8op,8oq,8or,8os,8ot,8ou,8ov,8ow,8ox,8oy,8oz,8pa,8pb,8pc,8pd,8pe,8pf,8pg,8ph,8pi,8pj,8pk,8pl,8pm,8pn,8po,8pp,8pq,8pr,8ps,8pt,8pu,8pv,8pw,8px,8py,8pz,8qa,8qb,8qc,8qd,8qe,8qf,8qg,8qh,8qi,8qj,8qk,8ql,8qm,8qn,8qo,8qp,8qq,8qr,8qs,8qt,8qu,8qv,8qw,8qx,8qy,8qz,8ra,8rb,8rc,8rd,8re,8rf,8rg,8rh,8ri,8rj,8rk,8rl,8rm,8rn,8ro,8rp,8rq,8rr,8rs,8rt,8ru,8rv,8rw,8rx,8ry,8rz,8sa,8sb,8sc,8sd,8se,8sf,8sg,8sh,8si,8sj,8sk,8sl,8sm,8sn,8so,8sp,8sq,8sr,8ss,8st,8su,8sv,8sw,8sx,8sy,8sz,8ta,8tb,8tc,8td,8te,8tf,8tg,8th,8ti,8tj,8tk,8tl,8tm,8tn,8to,8tp,8tq,8tr,8ts,8tt,8tu,8tv,8tw,8tx,8ty,8tz,8ua,8ub,8uc,8ud,8ue,8uf,8ug,8uh,8ui,8uj,8uk,8ul,8um,8un,8uo,8up,8uq,8ur,8us,8ut,8uu,8uv,8uw,8ux,8uy,8uz,8va,8vb,8vc,8vd,8ve,8vf,8vg,8vh,8vi,8vj,8vk,8vl,8vm,8vn,8vo,8vp,8vq,8vr,8vs,8vt,8vu,8vv,8vw,8vx,8vy,8vz,8wa,8wb,8wc,8wd,8we,8wf,8wg,8wh,8wi,8wj,8wk,8wl,8wm,8wn,8wo,8wp,8wq,8wr,8ws,8wt,8wu,8wv,8ww,8wx,8wy,8wz,8xa,8xb,8xc,8xd,8xe,8xf,8xg,8xh,8xi,8xj,8xk,8xl,8xm,8xn,8xo,8xp,8xq,8xr,8xs,8xt,8xu,8xv,8xw,8xx,8xy,8xz,8ya,8yb,8yc,8yd,8ye,8yf,8yg,8yh,8yi,8yj,8yk,8yl,8ym,8yn,8yo,8yp,8yq,8yr,8ys,8yt,8yu,8yv,8yw,8yx,8yy,8yz,8za,8zb,8zc,8zd,8ze,8zf,8zg,8zh,8zi,8zj,8zk,8zl,8zm,8zn,8zo,8zp,8zq,8zr,8zs,8zt,8zu,8zv,8zw,8zx,8zy,8zz,9aa,9ab,9ac,9ad,9ae,9af,9ag,9ah,9ai,9aj,9ak,9al,9am,9an,9ao,9ap,9aq,9ar,9as,9at,9au,9av,9aw,9ax,9ay,9az,9ba,9bb,9bc,9bd,9be,9bf,9bg,9bh,9bi,9bj,9bk,9bl,9bm,9bn,9bo,9bp,9bq,9br,9bs,9bt,9bu,9bv,9bw,9bx,9by,9bz,9ca,9cb,9cc,9cd,9ce,9cf,9cg,9ch,9ci,9cj,9ck,9cl,9cm,9cn,9co,9cp,9cq,9cr,9cs,9ct,9cu,9cv,9cw,9cx,9cy,9cz,9da,9db,9dc,9dd,9de,9df,9dg,9dh,9di,9dj,9dk,9dl,9dm,9dn,9do,9dp,9dq,9dr,9ds,9dt,9du,9dv,9dw,9dx,9dy,9dz,9ea,9eb,9ec,9ed,9ee,9ef,9eg,9eh,9ei,9ej,9ek,9el,9em,9en,9eo,9ep,9eq,9er,9es,9et,9eu,9ev,9ew,9ex,9ey,9ez,9fa,9fb,9fc,9fd,9fe,9ff,9fg,9fh,9fi,9fj,9fk,9fl,9fm,9fn,9fo,9fp,9fq,9fr,9fs,9ft,9fu,9fv,9fw,9fx,9fy,9fz,9ga,9gb,9gc,9gd,9ge,9gf,9gg,9gh,9gi,9gj,9gk,9gl,9gm,9gn,9go,9gp,9gq,9gr,9gs,9gt,9gu,9gv,9gw,9gx,9gy,9gz,9ha,9hb,9hc,9hd,9he,9hf,9hg,9hh,9hi,9hj,9hk,9hl,9hm,9hn,9ho,9hp,9hq,9hr,9hs,9ht,9hu,9hv,9hw,9hx,9hy,9hz,9ia,9ib,9ic,9id,9ie,9if,9ig,9ih,9ii,9ij,9ik,9il,9im,9in,9io,9ip,9iq,9ir,9is,9it,9iu,9iv,9iw,9ix,9iy,9iz,9ja,9jb,9jc,9jd,9je,9jf,9jg,9jh,9ji,9jj,9jk,9jl,9jm,9jn,9jo,9jp,9jq,9jr,9js,9jt,9ju,9jv,9jw,9jx,9jy,9jz,9ka,9kb,9kc,9kd,9ke,9kf,9kg,9kh,9ki,9kj,9kk,9kl,9km,9kn,9ko,9kp,9kq,9kr,9ks,9kt,9ku,9kv,9kw,9kx,9ky,9kz,9la,9lb,9lc,9ld,9le,9lf,9lg,9lh,9li,9lj,9lk,9ll,9lm,9ln,9lo,9lp,9lq,9lr,9ls,9lt,9lu,9lv,9lw,9lx,9ly,9lz,9ma,9mb,9mc,9md,9me,9mf,9mg,9mh,9mi,9mj,9mk,9ml,9mm,9mn,9mo,9mp,9mq,9mr,9ms,9mt,9mu,9mv,9mw,9mx,9my,9mz,9na,9nb,9nc,9nd,9ne,9nf,9ng,9nh,9ni,9nj,9nk,9nl,9nm,9nn,9no,9np,9nq,9nr,9ns,9nt,9nu,9nv,9nw,9nx,9ny,9nz,9oa,9ob,9oc,9od,9oe,9of,9og,9oh,9oi,9oj,9ok,9ol,9om,9on,9oo,9op,9oq,9or,9os,9ot,9ou,9ov,9ow,9ox,9oy,9oz,9pa,9pb,9pc,9pd,9pe,9pf,9pg,9ph,9pi,9pj,9pk,9pl,9pm,9pn,9po,9pp,9pq,9pr,9ps,9pt,9pu,9pv,9pw,9px,9py,9pz,9qa,9qb,9qc,9qd,9qe,9qf,9qg,9qh,9qi,9qj,9qk,9ql,9qm,9qn,9qo,9qp,9qq,9qr,9qs,9qt,9qu,9qv,9qw,9qx,9qy,9qz,9ra,9rb,9rc,9rd,9re,9rf,9rg,9rh,9ri,9rj,9rk,9rl,9rm,9rn,9ro,9rp,9rq,9rr,9rs,9rt,9ru,9rv,9rw,9rx,9ry,9rz,9sa,9sb,9sc,9sd,9se,9sf,9sg,9sh,9si,9sj,9sk,9sl,9sm,9sn,9so,9sp,9sq,9sr,9ss,9st,9su,9sv,9sw,9sx,9sy,9sz,9ta,9tb,9tc,9td,9te,9tf,9tg,9th,9ti,9tj,9tk,9tl,9tm,9tn,9to,9tp,9tq,9tr,9ts,9tt,9tu,9tv,9tw,9tx,9ty,9tz,9ua,9ub,9uc,9ud,9ue,9uf,9ug,9uh,9ui,9uj,9uk,9ul,9um,9un,9uo,9up,9uq,9ur,9us,9ut,9uu,9uv,9uw,9ux,9uy,9uz,9va,9vb,9vc,9vd,9ve,9vf,9vg,9vh,9vi,9vj,9vk,9vl,9vm,9vn,9vo,9vp,9vq,9vr,9vs,9vt,9vu,9vv,9vw,9vx,9vy,9vz,9wa,9wb,9wc,9wd,9we,9wf,9wg,9wh,9wi,9wj,9wk,9wl,9wm,9wn,9wo,9wp,9wq,9wr,9ws,9wt,9wu,9wv,9ww,9wx,9wy,9wz,9xa,9xb,9xc,9xd,9xe,9xf,9xg,9xh,9xi,9xj,9xk,9xl,9xm,9xn,9xo,9xp,9xq,9xr,9xs,9xt,9xu,9xv,9xw,9xx,9xy,9xz,9ya,9yb,9yc,9yd,9ye,9yf,9yg,9yh,9yi,9yj,9yk,9yl,9ym,9yn,9yo,9yp,9yq,9yr,9ys,9yt,9yu,9yv,9yw,9yx,9yy,9yz,9za,9zb,9zc,9zd,9ze,9zf,9zg,9zh,9zi,9zj,9zk,9zl,9zm,9zn,9zo,9zp,9zq,9zr,9zs,9zt,9zu,9zv,9zw,9zx,9zy,9zz,10aa,10ab,10ac,10ad,10ae,10af,10ag,10ah,10ai,10aj,10ak,10al,10am,10an,10ao,10ap,10aq,10ar,10as,10at,10au,10av,10aw,10ax,10ay,10az,10ba,10bb,10bc,10bd,10be,10bf,10bg,10bh,10bi,10bj,10bk,10bl,10bm,10bn,10bo,10bp,10bq,10br,10bs,10bt,10bu,10bv,10bw,10bx,10by,10bz,10ca,10cb,10cc,10cd,10ce,10cf,10cg,10ch,10ci,10cj,10ck,10cl,10cm,10cn,10co,10cp,10cq,10cr,10cs,10ct,10cu,10cv,10cw,10cx,10cy,10cz,10da,10db,10dc,10dd,10de,10df,10dg,10dh,10di,10dj,10dk,10dl,10dm,10dn,10do,10dp,10dq,10dr,10ds,10dt,10du,10dv,10dw,10dx,10dy,10dz,10ea,10eb,10ec,10ed,10ee,10ef,10eg,10eh,10ei,10ej,10ek,10el,10em,10en,10eo,10ep,10eq,10er,10es,10et,10eu,10ev,10ew,10ex,10ey,10ez,10fa,10fb,10fc,10fd,10fe,10ff,10fg,10fh,10fi,10fj,10fk,10fl,10fm,10fn,10fo,10fp,10fq,10fr,10fs,10ft,10fu,10fv,10fw,10fx,10fy,10fz,10ga,10gb,10gc,10gd,10ge,10gf,10gg,10gh,10gi,10gj,10gk,10gl,10gm,10gn,10go,10gp,10gq,10gr,10gs,10gt,10gu,10gv,10gw,10gx,10gy,10gz,10ha,10hb,10hc,10hd,10he,10hf,10hg,10hh,10hi,10hj,10hk,10hl,10hm,10hn,10ho,10hp,10hq,10hr,10hs,10ht,10hu,10hv,10hw,10hx,10hy,10hz,10ia,10ib,10ic,10id,10ie,10if,10ig,10ih,10ii,10ij,10ik,10il,10im,10in,10io,10ip,10iq,10ir,10is,10it,10iu,10iv,10iw,10ix,10iy,10iz,10ja,10jb,10jc,10jd,10je,10jf,10jg,10jh,10ji,10jj,10jk,10jl,10jm,10jn,10jo,10jp,10jq,10jr,10js,10jt,10ju,10jv,10jw,10jx,10jy,10jz,10ka,10kb,10kc,10kd,10ke,10kf,10kg,10kh,10ki,10kj,10kk,10kl,10km,10kn,10ko,10kp,10kq,10kr,10ks,10kt,10ku,10kv,10kw,10kx,10ky,10kz,10la,10lb,10lc,10ld,10le,10lf,10lg,10lh,10li,10lj,10lk,10ll,10lm,10ln,10lo,10lp,10lq,10lr,10ls,10lt,10lu,10lv,10lw,10lx,10ly,10lz,10ma,10mb,10mc,10md,10me,10mf,10mg,10mh,10mi,10mj,10mk,10ml,10mm,10mn,10mo,10mp,10mq,10mr,10ms,10mt,10mu,10mv,10mw,10mx,10my,10mz,10na,10nb,10nc,10nd,10ne,10nf,10ng,10nh,10ni,10nj,10nk,10nl,10nm,10nn,10no,10np,10nq,10nr,10ns,10nt,10nu,10nv,10nw,10nx,10ny,10nz,10oa,10ob,10oc,10od,10oe,10of,10og,10oh,10oi,10oj,10ok,10ol,10om,10on,10oo,10op,10oq,10or,10os,10ot,10ou,10ov,10ow,10ox,10oy,10oz,10pa,10pb,10pc,10pd,10pe,10pf,10pg,10ph,10pi,10pj,10pk,10pl,10pm,10pn,10po,10pp,10pq,10pr,10ps,10pt,10pu,10pv,10pw,10px,10py,10pz,10qa,10qb,10qc,10qd,10qe,10qf,10qg,10qh,10qi,10qj,10qk,10ql,10qm,10qn,10qo,10qp,10qq,10qr,10qs,10qt,10qu,10qv,10qw,10qx,10qy,10qz,10ra,10rb,10rc,10rd,10re,10rf,10rg,10rh,10ri,10rj,10rk,10rl,10rm,10rn,10ro,10rp,10rq,10rr,10rs,10rt,10ru,10rv,10rw,10rx,10ry,10rz,10sa,10sb,10sc,10sd,10se,10sf,10sg,10sh,10si,10sj,10sk,10sl,10sm,10sn,10so,10sp,10sq,10sr,10ss,10st,10su,10sv,10sw,10sx,10sy,10sz,10ta,10tb,10tc,10td,10te,10tf,10tg,10th,10ti,10tj,10tk,10tl,10tm,10tn,10to,10tp,10tq,10tr,10ts,10tt,10tu,10tv,10tw,10tx,10ty,10tz,10ua,10ub,10uc,10ud,10ue,10uf,10ug,10uh,10ui,10uj,10uk,10ul,10um,10un,10uo,10up,10uq,10ur,10us,10ut,10uu,10uv,10uw,10ux,10uy,10uz,10va,10vb,10vc,10vd,10ve,10vf,10vg,10vh,10vi,10vj,10vk,10vl,10vm,10vn,10vo,10vp,10vq,10vr,10vs,10vt,10vu,10vv,10vw,10vx,10vy,10vz,10wa,10wb,10wc,10wd,10we,10wf,10wg,10wh,10wi,10wj,10wk,10wl,10wm,10wn,10wo,10wp,10wq,10wr,10ws,10wt,10wu,10wv,10ww,10wx,10wy,10wz,10xa,10xb,10xc,10xd,10xe,10xf,10xg,10xh,10xi,10xj,10xk,10xl,10xm,10xn,10xo,10xp,10xq,10xr,10xs,10xt,10xu,10xv,10xw,10xx,10xy,10xz,10ya,10yb,10yc,10yd,10ye,10yf,10yg,10yh,10yi,10yj,10yk,10yl,10ym,10yn,10yo,10yp,10yq,10yr,10ys,10yt,10yu,10yv,10yw,10yx,10yy,10yz,10za,10zb,10zc,10zd,10ze,10zf,10zg,10zh,10zi,10zj,10zk,10zl,10zm,10zn,10zo,10zp,10zq,10zr,10zs,10zt,10zu,10zv,10zw,10zx,10zy,10zz,11aa,11ab,11ac,11ad,11ae,11af,11ag,11ah,11ai,11aj,11ak,11al,11am,11an,11ao,11ap,11aq,11ar,11as,11at,11au,11av,11aw,11ax,11ay,11az,11ba,11bb,11bc,11bd,11be,11bf,11bg,11bh,11bi,11bj,11bk,11bl,11bm,11bn,11bo,11bp,11bq,11br,11bs,11bt,11bu,11bv,11bw,11bx,11by,11bz,11ca,11cb,11cc,11cd,11ce,11cf,11cg,11ch,11ci,11cj,11ck,11cl,11cm,11cn,11co,11cp,11cq,11cr,11cs,11ct,11cu,11cv,11cw,11cx,11cy,11cz,11da,11db,11dc,11dd,11de,11df,11dg,11dh,11di,11dj,11dk,11dl,11dm,11dn,11do,11dp,11dq,11dr,11ds,11dt,11du,11dv,11dw,11dx,11dy,11dz,11ea,11eb,11ec,11ed,11ee,11ef,11eg,11eh,11ei,11ej,11ek,11el,11em,11en,11eo,11ep,11eq,11er,11es,11et,11eu,11ev,11ew,11ex,11ey,11ez,11fa,11fb,11fc,11fd,11fe,11ff,11fg,11fh,11fi,11fj,11fk,11fl,11fm,11fn,11fo,11fp,11fq,11fr,11fs,11ft,11fu,11fv,11fw,11fx,11fy,11fz,11ga,11gb,11gc,11gd,11ge,11gf,11gg,11gh,11gi,11gj,11gk,11gl,11gm,11gn,11go,11gp,11gq,11gr,11gs,11gt,11gu,11gv,11gw,11gx,11gy,11gz,11ha,11hb,11hc,11hd,11he,11hf,11hg,11hh,11hi,11hj,11hk,11hl,11hm,11hn,11ho,11hp,11hq,11hr,11hs,11ht,11hu,11hv,11hw,11hx,11hy,11hz,11ia,11ib,11ic,11id,11ie,11if,11ig,11ih,11ii,11ij,11ik,11il,11im,11in,11io,11ip,11iq,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IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Preparation Dissolve 2-3 mg in 1 mL of methanol R by warming in a water-bath at 40-50 °C. Evaporate to dryness under a stream of dry nitrogen, warming gently if necessary. Examine the residue.

Comparison mitoxantrone hydrochloride CRS.
B. It gives reaction (b) of chlorides (2.3.1).

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in about 40 mL of the mobile phase, sonicating if necessary, and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of mitoxantrone hydrochloride CRS in about 40 mL of the mobile phase, sonicating if necessary, and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1 mL of the test solution to 100 mL with the mobile phase.

Reference solution (c) Dissolve 2.0 mg of mitoxantrone impurity A CRS in 1.0 mL of reference solution (a).

Reference solution (d) Dilute 1 mL of reference solution (b) to 10 mL with the mobile phase.

Column: size: $l = 0.30$ m, $\phi = 3.0$ mm;

stationary phase: phenylsilyl silica gel for chromatography R (10 μ m).

Mobile phase Mix 750 volumes of water R, 250 volumes of acetonitrile R and 25 volumes of a solution prepared as follows: dissolve 22.0 g of sodium heptanesulfonate R in about 150 mL of water R and filter through a 0.45 μ m filter; wash the filter with water R and combine the filtrate and washings; add 32.0 mL of glacial acetic acid R and dilute to 250 mL with water R.

Flow rate 3 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 50 μ L of the test solution and reference solutions (b), (c) and (d).

Run time 3 times the retention time of mitoxantrone.

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to mitoxantrone and impurity A.

Limits:

— impurities A, B, C, D: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent);

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (d) (0.1 per cent).

Ethanol

Gas chromatography (2.2.28).

Internal standard solution Dilute 2.0 mL of propanol R to 100 mL with water R. Dilute 5.0 mL of this solution to 100 mL with water R.

Test solution Mix 0.100 g of the substance to be examined with 2.0 mL of the internal standard solution and dilute to 5.0 mL with water R. Place the flask in an ultrasonic bath for

2 min, then shake the flask for 2 min. If necessary, repeat the sonication and shaking until dissolution is complete. Reference solution Dilute 2.0 mL of anhydrous ethanol R to 100.0 mL with water R. Dilute 5.0 mL of the solution to 100.0 mL with water R. Dilute 10.0 mL of this solution and 10.0 mL of the internal standard solution to 25.0 mL with water R.

Column: size: $l = 2$ m, $\phi = 3$ mm;

stationary phase: ethylnonylbenzene-divinylbenzene copolymer R.

Carrier gas helium for chromatography R.

Flow rate 19 mL/min.

Temperature: column: 120 °C;

injection port: 175 °C;

detection: 210 °C.

Detection Flame ionisation.

Injection 1 μ L.

Retention time Ethanol = about 1 min; propanol = about 2 min.

System suitability: reference solution:

— resolution: minimum 6 between the peaks due to ethanol and propanol.

Calculate the content of ethanol, taking its density (2.2.5) to be 0.790 g/mL at 20 °C.

Limit:

— ethanol: maximum 1.6 per cent m/m.

Water (2.5.12)

Maximum 6.0 per cent, determined on 0.300 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

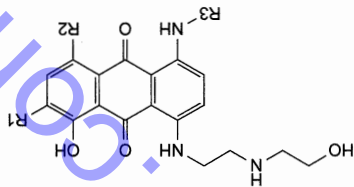
Calculate the percentage content of $C_{22}H_{30}Cl_2N_4O_6$ from the declared content of mitoxantrone hydrochloride CRS.

STORAGE

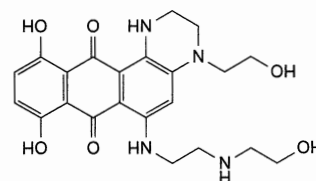
In an airtight container.

IMPURITIES

Specified impurities A, B, C, D.



A. R1 = R3 = H, R2 = OH; 1-amino-5,8-dihydroxy-4-[[2-[(2-hydroxyethyl)amino]ethyl]amino]anthracene-9,10-dione, B. R1 = R2 = H, R3 = CH₂-CH₂-NH-CH₂-CH₂-OH; 5-hydroxy-1,4-bis[[2-[(2-hydroxyethyl)amino]ethyl]amino]anthracene-9,10-dione, C. R1 = Cl, R2 = OH, R3 = CH₂-CH₂-NH-CH₂-CH₂-OH; 2-chloro-1,4-dihydroxy-5,8-bis[[2-[(2-hydroxyethyl)amino]ethyl]amino]anthracene-9,10-dione,

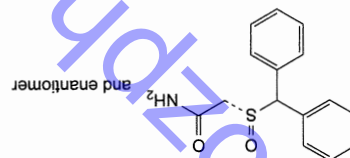


D, 8,11-dihydroxy-4-(2-hydroxyethyl)-6-[[2-[(2-hydroxyethyl)amino]ethyl]amino]-1,2,3,4-tetrahydrophthalothio[2,3-f]quininoxaline-7,12-dione.

Ph Eur

Modafinil

(Ph. Eur. monograph 2307)

C₁₅H₁₅NO₂ 273.4 68693-11-8

Action and use

Narcolepsy and sleep disorders.

Ph Eur

DEFINITION

2-[(RS)-(Diphenylmethyl)sulfinyl]acetamide.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble or practically insoluble in water,

sparingly soluble in methanol, slightly soluble in ethanol

(96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison modafinil CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in methanol R, evaporate to dryness and

record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R1, water R (35:65 V/V).

Test solution Dissolve 50.0 mg of the substance to be

examined in 35 mL of acetonitrile R1 and dilute to 50.0 mL

with water R. Dilute 1.0 mL of this solution to 10.0 mL with

the solvent mixture.

Reference solution (a) Dissolve 50.0 mg of modafinil CRS in

35 mL of acetonitrile R1 and dilute to 50.0 mL with water R.

mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this

solution to 20.0 mL with the solvent mixture.

Reference solution (c) Add 2.0 mL of the solvent mixture to a

vial of modafinil for system suitability CRS (containing

impurities A, B and C) and sonicate for 10 min.

Column:

size: $l = 0.15$ m, $\phi = 4.6$ mm;

stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase Mix 35 volumes of acetonitrile R1 and

65 volumes of a 6.8 g/L solution of potassium dihydrogen

phosphate R previously adjusted to pH 2.3 with phosphoric

acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L of the test solution and reference solutions (b)

and (c).

Run time 4 times the retention time of modafinil.

Identification of impurities Use the chromatogram supplied

with modafinil for system suitability CRS and the

chromatogram obtained with reference solution (c) to identify

the peaks due to impurities A, B and C.

Relative retention With reference to modafinil (retention

time = about 4 min): impurity A = about 1.3;

impurity B = about 1.8; impurity C = about 3.0.

System suitability: reference solution (c):

— peak-to-valley ratio: minimum 2.5, where H_p = height

above the baseline of the peak due to impurity A and

H_p = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

modafinil.

— impurity A: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— impurities B, C: for each impurity, not more than 5 times

the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.5 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 10 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (1.0 per cent);

— disregard limit: 0.5 times the area of the principal peak in

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in 40 mL of methanol R, warming slightly.

Add 7.5 mL of water R. Allow to cool, then dilute to

50.0 mL with methanol R. 12 mL of the solution complies

with test B. Prepare the reference solution using 2 mL of lead

standard solution (2 ppm Pb) R, 8 mL of methanol R and

2 mL of the test solution.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

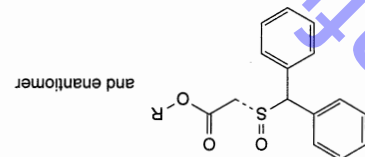
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of modafinil from the declared content of $C_{15}H_{15}NO_2S$ in modafinil CRS.

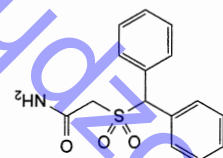
IMPURITIES

Specified impurities: A, B, C.



A, R = H; [(R,S)-(diphenylmethyl)sulfinyl]acetic acid,

C, R = CH₃; methyl [(R,S)-(diphenylmethyl)sulfinyl]acetate,



B, 2-[(diphenylmethyl)sulfonyl]acetamide.

Molgramostim Concentrated Solution

(Ph. Eur. monograph 1641)

APARSPSPST	QPEWHVNAIQ	EARRLNLNSR
DTAAEMNETV	EVISSEMFLO	EPTCLOTRLE
LYKQGLRGSL	TKLKGPLTMM	ASHYKQHCPP
TPETSCATQI	ITFESEKENT	KDFLLVIFPD
CWEPVOE		

99283-10-0

Action and use
Recombinant granulocyte macrophage colony-stimulating factor.

Ph Eur

DEFINITION

Solution of a protein having the structure of the granulocyte macrophage colony stimulating factor which is produced and secreted by various human blood cell types. The protein stimulates the differentiation and proliferation of leucocyte stem cells into mature granulocytes and macrophages.

Content

Minimum 2.0 mg of protein per millilitre.

Potency

Minimum 0.7×10^7 IU per milligram of protein.

PRODUCTION

Molgramostim concentrated solution is produced by a method based on recombinant DNA (rDNA) technology, using bacteria as host cells. It is produced under conditions

designed to minimise microbial contamination of the product.

Prior to release, the following tests are carried out on each batch of the final bulk product, unless exemption has been granted by the competent authority.

Host-cell derived proteins

The limit is approved by the competent authority.

Host-cell or vector derived DNA

The limit is approved by the competent authority.

CHARACTERS**Appearance**

Clear, colourless liquid.

IDENTIFICATION

A. It shows the expected biological activity (see Assay).

B. Isoelectric focusing (2.2.54).

Test solution Dilute the preparation to be examined with water R to obtain a concentration of 0.25 mg/mL.

Reference solution (a) Dilute molgramostim CRS with water R to obtain a concentration of 0.25 mg/mL.

Reference solution (b) Use an isoelectric point (pI) calibration solution, in the pI range of 2.5–6.5, prepared according to the manufacturer's instructions.

Focusing:

pH gradient: 4.0–6.5,

— **anolyte:** 14.7 g/L (0.1 M) solution of glutamic acid R in a 50 per cent V/V solution of dilute phosphoric acid R (0.5 M),

— **appication:** 20 µL.

Detection Immerse the gel in a suitable volume of a solution containing 115 g/L of trichloroacetic acid R and 34.5 g/L of sulfadiazine R and shake the container gently for 30 min.

acid R, 100 volumes of ethanol R and 268 volumes of water R in a staining solution prewarmed to 60 °C and prepared by adding acid blue 83 R at a concentration of 1.2 g/L to mixture A. Wash the gel in several containers with mixture A and keep the gel in this mixture until the background is clear (12–24 h). After adequate destaining, soak the gel for 1 h in a 10 per cent V/V solution of glycine R in mixture A.

System suitability:

— in the electropherogram obtained with reference solution (b), the relevant isoelectric point markers are distributed along the entire length of the gel,

— in the electropherogram obtained with reference solution (a), the pI of the principal band is 4.9 to 5.4.

Results The principal band in the electropherogram obtained with the test solution corresponds in position to the principal band in the electropherogram obtained with reference solution (a). Plot the migration distances of the relevant pI markers versus their pI and determine the isoelectric points of the principal component of each of the test solution and reference solution (a). They do not differ by more than 0.2 pI units.

C. Examine the electropherograms obtained under reducing conditions in the test for impurities with molecular masses differing from that of molgramostim. The principal band in the electropherogram obtained with test solution (a) is similar in position to the principal band in the electropherogram obtained with reference solution (a).

D. Peptide mapping (2.2.55).

conditions adjusted as indicated to achieve optimum resolution of all amino acids, — a sample obtained from a blank sequencing cycle obtained as recommended by the equipment manufacturer.

Results The first 16 amino acids are: Ala-Pro-Ala-Arg-Ser-Pro-Ser-Pro-Ser-Thr-Gln-Pro-Tyr-Glu-His-Val.

TESTS

Impurities with molecular masses differing from that of molgramostim

Polyacrylamide gel electrophoresis (2.2.31) under both reducing and non-reducing conditions. Gel dimensions 0.75 mm thick. Resolving gel 14 per cent acrylamide. Sample buffer A Mix equal volumes of water R and concentrated SDS-PAGE sample buffer R. Sample buffer B (reducing conditions) Mix equal volumes of water R and concentrated SDS-PAGE sample buffer for reducing conditions R. Dilute the preparation to be examined in water R to obtain a concentration of 1.0 mg/mL. To 1 volume of this solution add 1 volume of concentrated SDS-PAGE sample buffer R. Test solution (a) Dilute 0.020 mL of test solution (a) to 1.0 mL with sample buffer A. Test solution (b) (2 per cent control). Dilute 0.020 mL of test solution (a) to 0.20 mL of test solution (b) add 0.20 mL of sample buffer A. Test solution (c) (1 per cent control). To 0.20 mL of test solution (b) add 0.20 mL of sample buffer A. Test solution (d) (0.5 per cent control). To 0.20 mL of test solution (c) add 0.20 mL of sample buffer A. Test solution (e) (0.25 per cent control). To 0.20 mL of test solution (d) add 0.20 mL of sample buffer A. Test solution (f) (0.1 per cent control). To 0.20 mL of test solution (e) add 0.30 mL of sample buffer A. Test solution (g) (0.05 per cent control). To 0.20 mL of test solution (f) add 0.20 mL of sample buffer A. Test solution (h) (0.025 per cent control). To 0.20 mL of test solution (g) add 0.20 mL of sample buffer A. Test solution (i) Prepare as for test solution (a), but using concentrated SDS-PAGE sample buffer for reducing conditions R. Test solutions (j)-(p) Prepare as for test solutions (b)-(h), but using sample buffer B. Reference solution (a) Dilute molgramostim CRS in water R to obtain a concentration of 0.02 mg/mL. Mix 1 volume of this solution with 1 volume of concentrated SDS-PAGE sample buffer R. Reference solution (b) Prepare as for reference solution (a), but using concentrated SDS-PAGE sample buffer for reducing conditions R. Reference solution (c) Use a solution of molecular mass markers suitable for calibrating SDS-PAGE gels in the range of 14 400-94 000. Dissolve in sample buffer or sample buffer (reducing conditions), as appropriate. Sample treatment Boil for 3 min. Application 50 µL; apply reduced and non-reduced solutions to separate gels.

Detection Silver staining as described below. Immerse the gel overnight in a mixture of 10 volumes of acetic acid R₁, 40 volumes of water R and 50 volumes of methanol R. Transfer the gel to a 100 g/L solution of glutaraldehyde solution with water R, and keep the gel in

Test solution Introduce 50 µL of tris-hydrochloride buffer solution pH 8.0 R and 50 µL of the preparation to be examined at a concentration of 2 mg/mL into a polypropylene tube of 0.5 mL capacity. Add 4 µL of a 1 mg/mL solution of trypsin for peptide mapping R in a 0.01 per cent V/V solution of trifluoroacetic acid R, cap tightly and mix well. Incubate at about 37 °C for 18 h. Add 125 µL of a 764 g/L (8 M) solution of guanidine hydrochloride R and mix well. Add 10 µL of a 154.2 g/L (1 M) solution of dithiothreitol R and mix well. Place the capped tube in boiling water for 1 min. Cool to room temperature. Reference solution Prepare at the same time and in the same manner as for the test solution but use molgramostim CRS instead of the preparation to be examined. Examine the 2 tryptic digests by liquid chromatography (2.2.29).

Column:

— size: $l = 0.10$ m, $\phi = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm) with a pore size of 30 nm.

Mobile phase:

— mobile phase A: dilute 1 mL of trifluoroacetic acid R in 1000 mL of water R;

— mobile phase B: dilute 1 mL of trifluoroacetic acid R in 100 mL of water R; add 900 mL of acetonitrile for chromatography R and mix;

Flow rate 1.0 mL/min. Detection Spectrophotometer at 214 nm. Equilibration At initial conditions for at least 12 min. Injection 200 µL.

System suitability The chromatograms obtained with the reference solution and the test solution are qualitatively similar to the Ph. Eur. reference chromatogram of molgramostim digest.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

E. N-Terminal sequence analysis. Perform the Edman degradation using an automated solid-phase sequencer, operated in accordance with the manufacturer's instructions.

Load about 1 nmol of the test preparation to a sequencing cartridge using the protocol provided by the manufacturer. Run 16 sequencing cycles, noting, if appropriate, the presence of proline at positions 2, 6, 8 and 12. Identify the phenylthiohydantoin (PTH)-amino acids released at each sequencing cycle by reverse-phase liquid chromatography. The procedure may be carried out using the column and reagents recommended by the manufacturer of the sequencing equipment for the separation of PTH-amino acids.

The separation procedure is calibrated using: — the mixture of PTH-amino acids provided by the manufacturer of the sequencer, with the gradient

Time (min)	Mobile Phase A (per cent V/V)	Mobile Phase B (per cent V/V)
0 - 35.0	100 → 65	0 → 35
35.0 - 105.0	65 → 35	35 → 65
105.0 - 107.5	35 → 100	65 → 0
107.5 - 120.0	100	0

water R for 20 min. Repeat this washing-step twice. Transfer the gel to a mixture containing 0.75 g/L of sodium hydroxide R, 14 g/L of concentrated ammonia R and 8 g/L of silver nitrate R. This solution is prepared immediately before use. Place the gel on a shaker in the dark for 5 min. Wash the gel for 30 s in each of 3 containers with water R and shake the gel in a mixture consisting of 0.05 g/L of citric acid R, 0.05 per cent V/V of formaldehyde R and 0.005 per cent V/V of methanol R in water R. Protein bands become visible during this step. Keep the gel in the solution until sufficiently stained and then rinse the gel repeatedly with water R in a shaking water bath. Soak gels in a solution consisting of 10 per cent V/V of acetic acid R and 1 per cent V/V of glycerol R.

System suitability: the validation criteria are met (2.2.31), a band is seen in the electropherogram obtained with test solution (h),

— a gradation of intensity of staining is seen in the electropherograms obtained with test solutions (a)-(h) and (i)-(p),

— the molecular mass of the principal band in the electropherogram obtained with reference solution (a) or (b) is within the range of 15 100 to 17 100.

Limits Compare the staining intensity of each non-molgramostim band observed in the electropherogram obtained with test solution (a) to the staining intensity of the principal band in the electropherograms obtained with test solutions (b)-(h). Proceed similarly with the

electropherograms obtained with test solutions (i)-(p). The impurity level is estimated as the dilution, in percentage, of the solution giving the electropherogram with the closest intensity of staining.

Reducing conditions:

- impurity with an apparent molecular mass of 20 000: maximum 1 per cent,
- impurity with an apparent molecular mass of 25 000: maximum 0.1 per cent,
- impurity with an apparent molecular mass of 30 000: maximum 0.3 per cent,
- total: maximum 2 per cent.

Non-reducing conditions:

- total of all impurities of molecular masses higher than 30 000: maximum 1 per cent.

Related proteins

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution (a) Dilute the preparation to be examined with 0.05 M phosphate buffer solution pH 7.0 R to obtain a concentration of 0.5 mg/mL.

Test solution (b) Mix 1 volume of test solution (a) with 4 volumes of a 0.125 mg/mL solution of human albumin R or bovine albumin R in 0.05 M phosphate buffer solution pH 7.0 R.

Reference solution (a) Dilute molgramostim CRS with 0.05 M phosphate buffer solution pH 7.0 R to obtain a concentration of 0.5 mg/mL.

Reference solution (b) Mix 1 volume of reference solution (a) with 4 volumes of a 0.125 mg/mL solution of human albumin R or bovine albumin R in 0.05 M phosphate buffer solution pH 7.0 R.

Column:

- size: $l = 0.15$ m, $\phi = 4.6$ mm,
- stationary phase: butylsilyl silica gel for chromatography R (5 μ m) with a pore size of 30 nm.

Mobile phase:

- mobile phase A: to about 800 mL of water R add 1.0 mL of trifluoroacetic acid R and dilute to 1000 mL with water R;
- mobile phase B: to 100 mL of water R add 1.0 mL of trifluoroacetic acid R and 900 mL of acetonitrile for chromatography R;

Time (min)	Mobile Phase A (per cent V/V)	Mobile Phase B (per cent V/V)
0 - 30	64 \rightarrow 44	36 \rightarrow 56
30 - 35	44 \rightarrow 0	56 \rightarrow 100
35 - 45	0	100
45 - 50	0 \rightarrow 64	100 \rightarrow 36
50 - 60	64	36

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 100 μ L of test solution (a), reference solutions (a) and (b).

System suitability: reference solution (b):

- retention time: molgramostim = about 22 min,
- repeatability: maximum relative standard deviation of 5.0 per cent after 4 injections,
- resolution: minimum 2 between the peaks due to albumin and molgramostim.

Limits:

- any impurity: for each impurity, maximum 1.5 per cent, total of impurities eluting between 5 min and 30 min; maximum 4 per cent.

Bacterial endotoxins (2.6.14)

- Less than 5 IU in the volume that contains 1.0 mg of protein.

ASSAY

Protein

Liquid chromatography (2.2.29) as described in the test for related proteins.

Injection 150 μ L of test solution (b) and reference solution (b).

Calculate the content of molgramostim using the declared content of molgramostim in molgramostim CRS.

Potency

Determination of the biological activity of molgramostim concentrated solution based on the stimulation of proliferation of TF-1 cells by molgramostim.

The following method uses the conversion of tetrazolium bromide (MTT) as a staining method. Validated alternative stains such as Alamar blue have also been found suitable.

TF-1 cells are incubated with varying dilutions of test and reference preparations of molgramostim. They are then incubated with a solution of MTT. This cytochemical stain is converted by cellular dehydrogenases to a purple formazan product. The formazan is then measured

spectrophotometrically. The potency of the preparation to be examined is determined by comparison of the dilutions of the test preparation with the dilutions of the appropriate International Standard of molgramostim or with a reference preparation calibrated in International Units, which yield the same response (50 per cent maximal stimulation).

The International Unit is the activity contained in a stated amount of the appropriate International Standard.

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Sparsingly soluble in water, soluble in anhydrous ethanol and in methylene chloride.

mp: about 142 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison molsidomine CRS.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

Dissolve 1.0 g in anhydrous ethanol R by heating at about 50 °C for about 5 min and dilute to 20.0 mL with the same solvent.

pH (2.2.3)

5.5 to 7.5.

Dissolve 0.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Impurity B

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Detection Spectrophotometer at 240 nm.

Injection 20 µL of test solution (a) and reference solution (b).

Relative retention With reference to molsidomine (retention time = about 9 min): impurity B = about 0.43.

System suitability: reference solution (b):

— signal-to-noise ratio: minimum 20 for the principal peak.

Limit:

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (3 ppm).

Impurity E

Liquid chromatography (2.2.29).

Test solution Dissolve 0.200 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of morpholine for chromatography R in 500.0 mL of water for water for

Dilute 20.0 mL of the solution to 500.0 mL with water for chromatography R. Dilute 5.0 mL of this solution to 100.0 mL with water for chromatography R.

Reference solution (b) Mix 10.0 mL of the test solution with 10.0 mL of reference solution (a).

Column:

— size: l = 0.25 m, Ø = 4.0 mm;

— stationary phase: resin for reversed-phase ion chromatography R;

— temperature: 25 °C.

Mobile phase Mix 3.0 mL of methanesulfonic acid R and 75 mL of acetonitrile R in water for chromatography R and dilute to 500.0 mL with water for chromatography R.

Suppressor regenerant water for chromatography R.

Flow rate 1.0 mL/min.

Expected background conductivity Less than 0.5 µS.

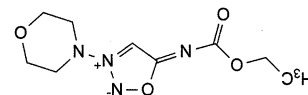
Detection Conductivity detector at 10 µS.

Injection 50 µL.

Run time 20 min.

Molsidomine

(Ph. Eur. monograph 1701)



C₉H₁₄N₄O₄

242.2

25717-80-0

Ph Eur

The label states:
— the content, in milligrams of protein per millilitre,
— the potency, in International Units per milligram of protein.

LABELLING

In an airtight container, protected from light, at a temperature below –65 °C.

STORAGE

of the stated potency.
are not less than 74 per cent and not more than 136 per cent
The confidence limits ($P = 0.95$) of the estimated potency
more than 125 per cent of the stated potency.
The estimated potency is not less than 80 per cent and not
4-parameter model (see 5.3).

using a suitable statistical method, for example the
sigmoidal dose-response curve to the data obtained and by
from the reading at 570 nm. Analyse the data by fitting a
570 nm and at 690 nm. Subtract the reading at 690 nm
using a 96-well microtitre plate reader. Read each plate at
Determine the relative quantity of purple formazan product
formed in each well by measuring the absorbance (2.2.25)

overnight.
adjusted to pH 2.7 with hydrochloric acid. Reincubate
240 g/L solution of sodium dodecyl sulfate R previously
plates from the incubator and add to each well 100 µL of a
bromide R to each well. Reincubate for 5 h. Remove the
Add 25 µL of a 5.0 g/L sterile solution of tetrazolium
a humidified incubator using 6 ± 1 per cent CO₂.
Incubate the plate at 36.0–38.0 °C for a minimum of 24 h in

in a uniform suspension during addition.
containing 3×10^5 cells per millilitre, maintaining the cells
Then add to each well 50 µL of a TF-1 cell suspension
series of 10 twofold dilutions to obtain a standard curve).
preparation at a concentration of about 65 IU/mL, plus a
to be tested in triplicate (test preparation and reference
the wells designed for the blanks. Add 50 µL of each solution
microtitre plate. Add an additional 50 µL of this solution to
Add 50 µL of dilution medium to all wells of a 96-well
Standard is stated by the World Health Organization.
The equivalence in International Units of the International

Action and use

Nitric oxide donor; treatment of angina pectoris.

DEFINITION

N-(Ethoxycarbonyl)-3-(morpholin-4-yl)sydnonimine.

Content

99.0 per cent to 101.0 per cent (dried substance).

Relative retention With reference to molsidomine (retention time = about 3 min); impurity E = about 2.4.

System suitability: reference solution (b):

- **signal-to-noise ratio:** minimum 6 for the peak due to impurity E.
- Limit:**
- **impurity E:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.01 per cent).

Related substances

Liquid chromatography (2.2.29). Protect the solutions from light.

Solvent mixture methanol R, mobile phase A (10:90 V/V).

Test solution (a) Dissolve 0.200 g of the substance to be examined in 2.5 mL of methanol R and dilute to 5.0 mL with mobile phase A.

Test solution (b) Dilute 1.0 mL of test solution (a) to 20.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (b) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2.4 mg of molsidomine impurity B CRS in 80 mL of methanol R and dilute to 100.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 10 mg of molsidomine hydrochloride R (impurity A) and 5 mg of molsidomine impurity D CRS in 10 mL of methanol R and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:

- size: $l = 0.15$ m, $\phi = 4.6$ mm;
- stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
- temperature: 30 °C.

Mobile phase:

- **mobile phase A:** dissolve 4.0 g of potassium dihydrogen phosphate R in water for chromatography R and dilute to 1000 mL with the same solvent;
- **mobile phase B:** methanol R1;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	90	10
3 - 10	90 \rightarrow 20	10 \rightarrow 80
10 - 13	20	80

Flow rate 1.3 mL/min.

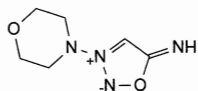
Detection Spectrophotometer at 210 nm.

Injection 20 μ L of test solution (b) and reference solutions (a) and (c).

Relative retention With reference to molsidomine (retention time = about 9 min); impurity A = about 0.2; impurity D = about 0.3.

System suitability: reference solution (c):

- **resolution:** minimum 3.5 between the peaks due to impurities A and D.
- Limits:**
- **unspecified impurities:** for each impurity, not more than the area of the peak due to molsidomine in the



A. 3-(morpholin-4-yl)sydnominine (Insidomine),

chromatogram obtained with reference solution (a) (0.10 per cent);

- **total:** not more than 3 times the area of the peak due to molsidomine in the chromatogram obtained with reference solution (a) (0.3 per cent);
- **disregard limit:** 0.5 times the area of the peak due to molsidomine in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals

Maximum 20 ppm.

Prescribed solution Dissolve 0.5 g in 20 mL of ethanol (96 per cent) R.

Test solution 12 mL of the prescribed solution.

Reference solution Mix 6 mL of lead standard solution (1 ppm Pb) (obtained by diluting lead standard solution (100 ppm Pb) R with ethanol (96 per cent) R) with 2 mL of the prescribed solution and 4 mL of water R.

Blank solution Mix 10 mL of ethanol (96 per cent) R and 2 mL of the prescribed solution.

To each solution, add 2 mL of buffer solution pH 3.5 R. Mix and add to 1.2 mL of thiocetamide reagent R.

Mix immediately. Filter the solutions through a membrane filter (nominal pore size 0.45 μ m) (2.4.8). Carry out the filtration slowly and uniformly, applying moderate and constant pressure to the piston. Compare the spots on the filters obtained with the different solutions. The test is invalid if the reference solution does not show a slight brown colour compared to the blank solution. The substance to be examined complies with the test if the brown colour of the spot resulting from the test solution is not more intense than that of the spot resulting from the reference solution.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 5 mL of acetic anhydride R and 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 24.22 mg of $C_{10}H_{14}N_4O_4$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities B, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, D.

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of mometasone furoate CRS in methylene chloride R and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of anhydrous beclomethasone dipropionate CRS in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and 8 volumes of methanol R to a mixture of 15 volumes of

ether R and 77 volumes of methylene chloride R.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R. Heat

at 120 °C for 10 min or until the spots appear. Allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight,

fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference

solution (a).

System suitability: reference solution (b):

— the chromatogram shows 2 spots which, when

examined in ultraviolet light at 365 nm, may not be completely separated.

C. Add about 2 mg to 2 mL of sulfuric acid R and shake to

dissolve. Within 15 min a light yellow colour develops. When

examined in ultraviolet light at 365 nm, no fluorescence is

seen. Add this solution to 10 mL of water R and mix.

The colour fades and there is no fluorescence.

D. Mix 80 mg with 0.30 g of anhydrous sodium carbonate R

and ignite in a crucible until an almost white residue is

obtained. Allow to cool and dissolve the residue in 5 mL of

dilute nitric acid R; filter. To 1 mL of the filtrate add 1 mL of

water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Specific optical rotation (2.2.7)

+ 50 to + 55 (dried substance).

Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to

10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture Mix 50 mL of acetonitrile R and 50 mL of

water R, then add 0.1 mL of acetic acid R.

Test solution Dissolve 20.0 mg of the substance to be

examined in 4.0 mL of acetonitrile R and dilute to 20.0 mL

with the solvent mixture.

Reference solution (a) Dissolve 2 mg of mometasone

furoate CRS and 6 mg of anhydrous beclomethasone

dipropionate CRS in the solvent mixture, then dilute to

10.0 mL with the solvent mixture. Dilute 0.25 mL of this

solution to 10.0 mL with the solvent mixture.

Comparison mometasone furoate CRS.

Preparation Discs.

A. Infrared absorption spectrophotometry (2.2.24).

Second identification B, C, D

First identification A, B

IDENTIFICATION

mp: about 220 °C, with decomposition.

Practically insoluble in water, soluble in acetone and in

methylene chloride, slightly soluble in ethanol (96 per cent).

Solubility

White or almost white powder.

Appearance

CHARACTERS

97.0 per cent to 103.0 per cent (dried substance).

Content

1,4-dien-17-yl furan-2-carboxylate.

9,21-Dichloro-11β-hydroxy-16α-methyl-3,20-dioxopregna-

DEFINITION

Ph Eur

Mometasone Scalp Application

Mometasone Ointment

Mometasone Aqueous Nasal Spray

Mometasone Cream

Preparations

Glucocorticoid.

Action and use

C₂₇H₃₀Cl₂O₆

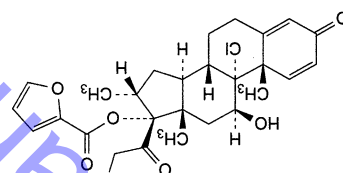
521.4

83919-23-7



Mometasone Furoate

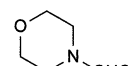
(Ph. Eur. monograph 1449)



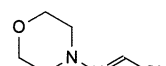
B. morpholine.



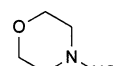
D. morpholine-4-carbaldehyde,



C. (2E)-(morpholin-4-ylimino)acetoneitrile,



B. 4-nitrosomorpholine,



Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column: — size: $l = 0.25$ m, $\phi = 4.6$ mm; — stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R , water R (50:50 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Run time T twice the retention time of mometasone furoate.

Retention time Mometasone furoate = about 17 min;

declometasone dipropionate = about 22 min.

System suitability: reference solution (a):

— resolution: minimum 6 between the peaks due to

mometasone furoate and declometasone dipropionate;

if necessary, adjust the concentration of acetonitrile in the mobile phase.

Limits:

— impurities A, B, C, D, E, F, G, H, I for each impurity,

not more than 0.6 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.3 per cent);

— total: not more than 1.2 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.6 per cent);

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to

100.0 mL with the same solvent. Dilute 2.0 mL of this

solution to 100.0 mL with ethanol (96 per cent) R . Measure

the absorbance (2.2.25) at the absorption maximum at

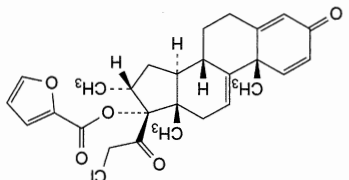
249 nm.

Calculate the content of $C_{27}H_{30}Cl_2O_6$ taking the specific

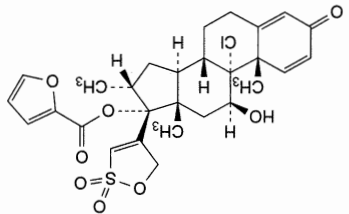
absorbance to be 481.

IMPURITIES

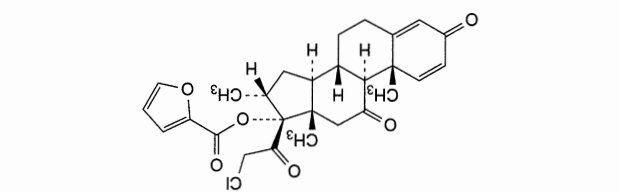
Specified impurities A, B, C, D, E, F, G, H, I



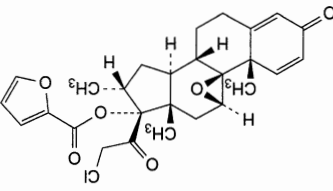
A. 21-chloro-16 α -methyl-3,20-dioxopregna-1,4,9(11)-trien-17-yl furan-2-carboxylate,



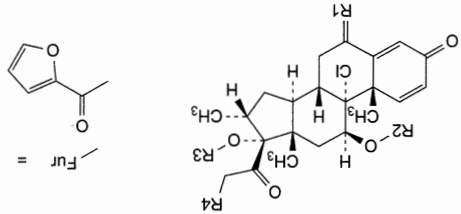
B. 4-[9-chloro-17-[(furan-2-ylcarbonyl)oxy]-1 β -hydroxy-16 α -methyl-3-oxoandrosta-1,4-dien-17 β -yl]-5H-1,2-oxathiole 2,2-dioxide,



C. 21-chloro-16 α -methyl-3,11,20-trioxopregna-1,4-dien-17-yl furan-2-carboxylate,

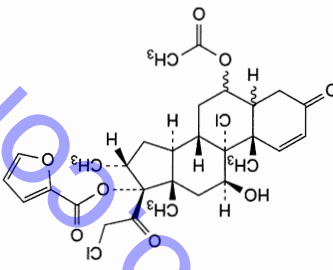


D. 21-chloro-9,11 β -epoxy-16 α -methyl-3,20-dioxo-9 β -pregna-1,4-dien-17-yl furan-2-carboxylate,



E. R1 = H, R2 = R3 = F, R4 = Cl; 9,21-dichloro-16 α -methyl-3,20-dioxopregna-1,4-diene-11 β ,17-dihydroxy-16 α -methylpregna-1,4-diene-3,20-dione (mometasone),

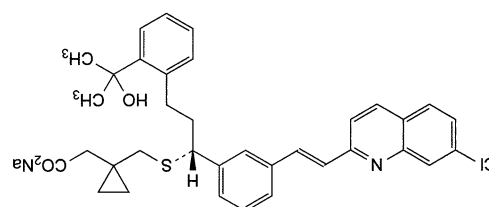
H. R1 = H, R2 = H, R3 = F, R4 = OH; 9-chloro-11 β ,21-dihydroxy-16 α -methyl-3,20-dioxopregna-1,4-dien-17-yl furan-2-carboxylate,



I. 9,21-dichloro-1 β -hydroxy-16 α -methyl-3,20-dioxo-5 ϵ -pregna-1-ene-6 ζ ,17-diyl 6-acetate 17-(furan-2-carboxylate).

Montelukast Sodium

(Ph. Eur. monograph 2583)



C₃₅H₃₅ClN₂O₃Na 608 151767-02-1

Action and use
Leukotriene CystLT₁ receptor antagonists; treatment of asthma.

Preparations
Montelukast Granules
Montelukast Chewable Tablets
Montelukast Tablets

Ph. Eur.

DEFINITION

Sodium [1-[[[1(R)-1-[3-(E)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]-3-(2-(1-hydroxy-1-methylethyl)phenyl)propyl]sulfonyl]methyl]cyclopropyl]acetate.

Content
98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water and in methylene chloride, freely soluble to very soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison *montelukast sodium* CRS.

B. Enantiomeric purity (see Tests).

C. Ignite 0.1 g in a crucible until an almost white residue is obtained. Take up the residue in 2 mL of water R and filter. The filtrate gives reaction (a) of sodium (2.3.1).

TESTS

Enantiomeric purity

Liquid chromatography (2.2.29). Carry out the test protected from light. Prepare the solutions in amber flasks.

Solvent mixture acetonitrile R, water R (50:50 V/V).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of *montelukast racemate* CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.0$ mm;

— stationary phase: silica gel AGP for chiral chromatography R (5 μ m);

— temperature: 30 °C.

(1.8 μ m);

— stationary phase: phenylsilyl silica gel for chromatography R

Column:

— size: $l = 0.05$ m, $\varnothing = 4.6$ mm;

solution to 100.0 mL with the solvent mixture.

50.0 mL with the solvent mixture. Dilute 10.0 mL of the *dicyclohexylamine* CRS in the solvent mixture and dilute to

Reference solution (d) Dissolve 65.0 mg of *montelukast*

vial and expose to ambient light for about 20 min.

transfer 1 mL of reference solution (b) to a colourless glass

Reference solution (c) In order to prepare impurity G *in situ*,

mixture.

in the solvent mixture and dilute to 10.0 mL with the solvent

Reference solution (b) Dissolve 10 mg of *montelukast* for peak

identification CRS (containing impurities B, C, D, E and F)

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

Reference solution (a) Dilute 1.0 mL of test solution (a) to

100.0 mL with the solvent mixture.

Test solution (b) Dilute 10.0 mL of test solution (a) to

the solvent mixture.

Test solution (a) Dissolve 50.0 mg of the substance to be

Solvent mixture water R, methanol R (10:90 V/V).

solutions in amber flasks.

procedure. Carry out the test protected from light. Prepare the

Liquid chromatography (2.2.29): use the normalisation

Related substances

— impurity A: maximum 0.2 per cent.

Limit:

the test solution.

and impurity A in the chromatogram obtained with

r_2 = sum of the areas of the peaks due to *montelukast*

chromatogram obtained with the test solution;

r_1 = area of the peak due to impurity A in the

$$100 \left(\frac{r_2}{r_1} \right)$$

following expression:

Calculate the percentage content of impurity A using the

the chromatogram obtained with reference solution (a).

— signal-to-noise ratio: minimum 10 for the principal peak in

obtained with reference solution (b);

impurity A and *montelukast* in the chromatogram

— resolution: minimum 2.9 between the peaks due to

System suitability:

time = about 25 min); impurity A = about 0.7.

Relative retention With reference to *montelukast* (retention

Injection 10 μ L.

Detection Spectrophotometer at 280 nm.

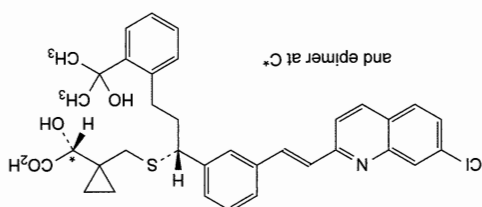
Flow rate 0.9 mL/min.

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 30	70 \rightarrow 60	30 \rightarrow 40
30 - 35	60	40

— mobile phase A: 2.3 g/L solution of ammonium acetate R adjusted to pH 5.7 with glacial acetic acid R;

— mobile phase B: acetonitrile R, methanol R (40:60 V/V);

Mobile phase:



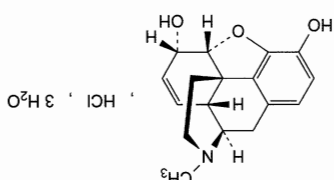
I. (2R,5S)-1-[[[1-[[3-[(E)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]-3-[2-(1-hydroxy-1-methylethyl)phenyl]propyl]sulfanyl]methyl]cyclopropyl](hydroxy)acetic acid.

Ph Eur



Morphine Hydrochloride

(Ph. Eur. monograph 0097)

C₁₇H₂₀ClNO₃·3H₂O 375.8

6055-06-7

Action and use

Opioid receptor agonist; analgesic.

Preparation

Morphine Suppositories

Ph Eur

DEFINITION

7,8-Didehydro-4,5-α-epoxy-17-methylmorphinan-3,6α-diol hydrochloride trihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless, silky needles or cubical masses, efflorescent in a dry atmosphere.

Solubility

Soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in toluene.

IDENTIFICATION

First identification A, E

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison morphine hydrochloride trihydrate CRS.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Solution A Dissolve 25.0 mg in water R and dilute to

25.0 mL with the same solvent.

Test solution (a) Dilute 10.0 mL of solution A to 100.0 mL

with water R.

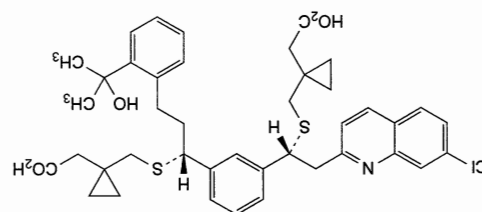
Test solution (b) Dilute 10.0 mL of solution A to 100.0 mL

with 0.1 M sodium hydroxide.

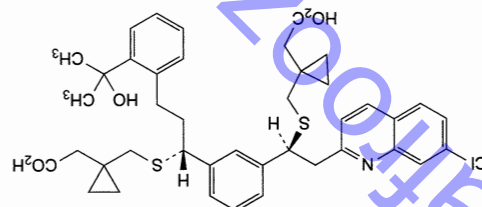
Spectral range 250–350 nm for test solutions (a) and (b).

Absorption maximum At 285 nm for test solution (a);

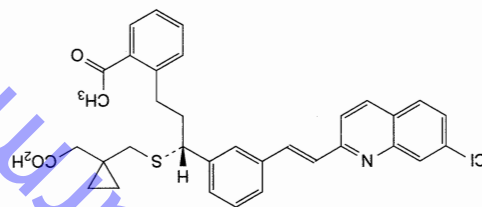
at 298 nm for test solution (b).



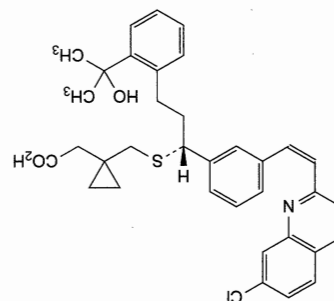
D. 1-[[[1-[[3-[(1R)-1-[[3-[(1R)-1-[[3-[(E)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]-3-[2-(1-hydroxy-1-methylethyl)phenyl]propyl]sulfanyl]methyl]cyclopropyl](hydroxy)acetic acid,



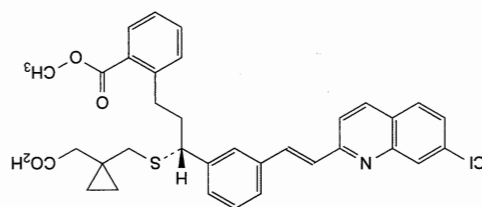
E. 1-[[[1-[[3-[(1S)-1-[[3-[(1S)-1-[[3-[(E)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]-3-[2-(1-hydroxy-1-methylethyl)phenyl]propyl]sulfanyl]methyl]cyclopropyl](hydroxy)acetic acid,



F. 1-[[[1-[[3-[(2R)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]propyl]sulfanyl]methyl]cyclopropyl](hydroxy)acetic acid,



G. 1-[[[1-[[3-[(2Z)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]propyl]sulfanyl]methyl]cyclopropyl](hydroxy)acetic acid,



H. 1-[[[1-[[3-[(E)-2-(7-chloroquinolin-2-yl)ethenyl]phenyl]-3-[2-(methoxycarbonyl)phenyl]propyl]sulfanyl]methyl]cyclopropyl](hydroxy)acetic acid,

Specific absorbance at the absorption maximum 37 to 43 for test solution (a); 64 to 72 for test solution (b).

C. To about 1 mg of powdered substance in a porcelain dish add 0.5 mL of *sulfuric acid-formaldehyde reagent R*. A purple colour develops and becomes violet.

D. It gives the reaction of alkaloids (2.3.1).

E. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S
Dissolve 0.500 g in carbon dioxide-free water *R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution *S* is clear (2.2.1) and not more intensely coloured than reference solution *Y*₆ or *BY*₆ (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution *S* add 0.05 mL of methyl red solution *R*. Not more than 0.2 mL of 0.02 *M* sodium hydroxide or 0.02 *M* hydrochloric acid is required to change the colour of the indicator.

Specific optical rotation (2.2.7)

–110 to –115 (anhydrous substance), determined on solution *S*.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.125 g of the substance to be examined in a 1 per cent *V/V* solution of acetic acid *R* and dilute to 50 mL with the same solution.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with a 1 per cent *V/V* solution of acetic acid *R*. Dilute 2.0 mL of this solution to 10.0 mL with a 1 per cent *V/V* solution of acetic acid *R*.

Reference solution (b) Dissolve 5 mg of morphine for system suitability *CRS* (containing impurities B, C, E and F) in a 1 per cent *V/V* solution of acetic acid *R* and dilute to 2 mL with the same solution.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography *R* (5 μ m);
— temperature: 35 °C.

Mobile phase:

— mobile phase A: 1.01 g/L solution of sodium heptanesulfonate *R* adjusted to pH 2.6 with a 50 per cent *V/V* solution of phosphoric acid *R*;
— mobile phase B: methanol *R*;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	85	15
2 - 35	85 \rightarrow 50	15 \rightarrow 50
35 - 40	50	50

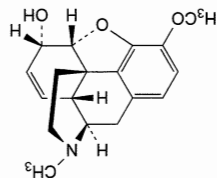
Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with morphine for system suitability *CRS* and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, E and F.

Relative retention With reference to morphine (retention time = about 12.5 min): impurity F = about 0.95;



A. 7,8-didehydro-4,5 α -epoxy-3-methoxy-17-methylmorphinan-6 α -ol (codeine)

Limits:

impurity E = about 1.1; impurity C = about 1.6; impurity B = about 1.9.

System suitability: reference solution (b):
— peak-to-valley ratio: minimum 2, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to morphine.

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.25; impurity C = 0.4; impurity E = 0.5;

— impurity B: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);

— impurities C, E: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Water (2.5.12)

12.5 per cent to 15.5 per cent, determined on 0.10 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 5 mL of 0.01 *M* hydrochloric acid and 30 mL of ethanol (96 per cent) *R*. Carry out a potentiometric titration (2.2.20), using 0.1 *M* sodium hydroxide. Read the volume added between the 2 points of inflexion.

1 mL of 0.1 *M* sodium hydroxide is equivalent to 32.18 mg of $C_{17}H_{20}ClNO_3$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities B, C, E

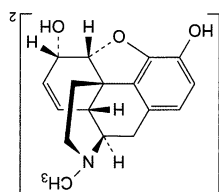
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D, F.



Morphine Sulfate

Morphine Sulphate

(Ph. Eur. monograph 1244)



$C_{17}H_{19}NO_5 \cdot H_2SO_4 \cdot 5H_2O$

759

6211-15-0

Action and use

Opioid receptor agonist; analgesic.

Preparations

Morphine Sulfate Injection
Morphine Suppositories
Morphine Tablets
Prolonged-release Morphine Tablets

Ph. Eur.

DEFINITION

Di(7,8-didehydro-4,5 α -epoxy-17-methylmorphinan-3,6 α -diol) sulfate pentahydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

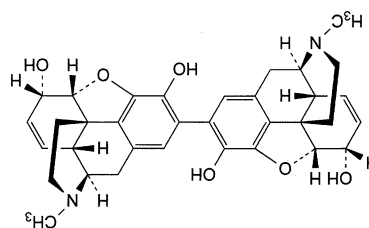
Soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in toluene.

IDENTIFICATION

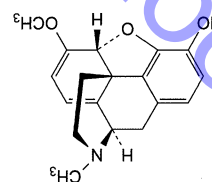
First identification A, E

Second identification B, C, D, E

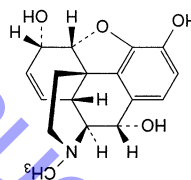
A. Infrared absorption spectrophotometry (2.2.24).
Preparation Dissolve 20 mg in 1 mL of water R, add 0.05 mL of 1 M sodium hydroxide and shake. A precipitate is formed. Filter, wash with 2 quantities, each of 0.5 mL, of water R and dry the precipitate at 145 °C for 1 h. Prepare discs using the dried precipitate.
Comparison Repeat the operations using 20 mg of morphine sulfate CRS.
B. Ultraviolet and visible absorption spectrophotometry (2.2.25).
Solution A Dissolve 25.0 mg in water R and dilute to 25.0 mL with the same solvent.
Test solution (a) Dilute 10.0 mL of solution A to 100.0 mL with water R.
Test solution (b) Dilute 10.0 mL of solution A to 100.0 mL with 0.1 M sodium hydroxide.
Spectral range 250–350 nm for test solutions (a) and (b).
Absorption maximum At 285 nm for test solution (a); at 298 nm for test solution (b).
Specific absorbance at the absorption maximum 37 to 43 for test solution (a); 64 to 72 for test solution (b).



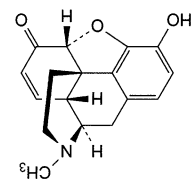
B. 7,7',8,8'-tetradidehydro-4,5 α :4',5'-diepoxo-17,17'-dimethyl-2,2'-bimorphinan-3,3',6 α ,6' α -tetrol (2,2'-bimorphine),



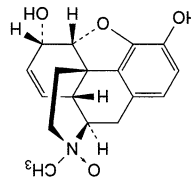
C. 6,7,8,14-tetradidehydro-4,5 α -epoxy-6-methoxy-17-methylmorphinan-3-ol (oripavine),



D. 7,8-didehydro-4,5 α -epoxy-17-methylmorphinan-3,6 α ,10 α -triol (10S-hydroxymorphine),



E. 7,8-didehydro-4,5 α -epoxy-3-hydroxy-17-methylmorphinan-6-one (morphinone),



F. (17S)-7,8-didehydro-4,5 α -epoxy-17-methylmorphinan-3,6 α -diol 17-oxide (morphine N-oxide).

Ph. Eur.

C. To about 1 mg of powdered substance in a porcelain dish add 0.5 mL of *sulfuric acid-formaldehyde reagent R*. A purple colour develops and becomes violet.

D. It gives the reaction of alkaloids (2.3.1).

E. It gives the reactions of sulfates (2.3.1).

TESTS

Solution S
Dissolve 0.500 g in carbon dioxide-free water *R* and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution *S* is clear (2.2.1) and not more intensely coloured than reference solution *Y*₆ or *BY*₆ (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution *S* add 0.05 mL of methyl red solution *R*. Not more than 0.2 mL of 0.02 *M* sodium hydroxide or 0.02 *M* hydrochloric acid is required to change the colour of the indicator.

Specific optical rotation (2.2.7)

—107 to —110 (anhydrous substance), determined on solution *S*.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 0.125 g of the substance to be examined in a 1 per cent *V/V* solution of acetic acid *R* and dilute to 50 mL with the same solution.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with a 1 per cent *V/V* solution of acetic acid *R*.

Dilute 2.0 mL of this solution to 10.0 mL with a 1 per cent *V/V* solution of acetic acid *R*.

Reference solution (b) Dissolve 5 mg of morphine for system suitability CRS (containing impurities B, C, E and F) in a 1 per cent *V/V* solution of acetic acid *R* and dilute to 2 mL with the same solution.

Column:
— size: $l = 0.15$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography *R* (5 μ m);
— temperature: 35 °C.

Mobile phase:
— mobile phase A: 1.01 g/L solution of sodium heptanesulfonate *R* adjusted to pH 2.6 with a 50 per cent *V/V* solution of phosphoric acid *R*;
— mobile phase B: methanol *R*;

Time (min)	Mobile phase A (per cent <i>V/V</i>)	Mobile phase B (per cent <i>V/V</i>)
0 - 2	85	15
2 - 35	85 \rightarrow 50	15 \rightarrow 50
35 - 40	50	50

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 230 nm.
Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with morphine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, E and F.

Relative retention With reference to morphine (retention time = about 12.5 min): impurity F = about 0.95; impurity E = about 1.1; impurity C = about 1.6; impurity B = about 1.9.

System suitability: reference solution (b):
— *peak-to-valley ratio*: minimum 2, where H_p = height above the baseline of the peak due to impurity F and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to morphine.

Limits:
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.25; impurity C = 0.4; impurity E = 0.5;

— *impurity B*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);

— *impurities C, E*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

The thresholds indicated under Related substances (Table 2034-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Iron (2.4.9)

Maximum 5 ppm.

Dissolve the residue from the test for sulfated ash in water *R* and dilute to 10.0 mL with the same solvent.

Water (2.5.12)

10.4 per cent to 13.4 per cent, determined on 0.10 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 2.0 g.

ASSAY

Dissolve 0.500 g in 120 mL of anhydrous acetic acid *R*. Titrate with 0.1 *M* perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 *M* perchloric acid is equivalent to 66.88 mg of $C_{17}H_{19}NO_{10}S$.

STORAGE

Protected from light.

IMPURITIES

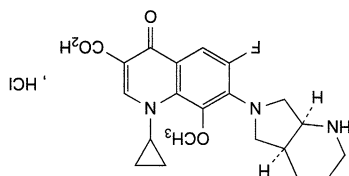
Specified impurities B, C, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D, F.



Moxifloxacin Hydrochloride

(Ph. Eur. monograph 2254)



$C_{21}H_{25}ClFN_3O_4$ 437.9 186826-86-8

Action and use

Fluoroquinolone antibacterial.

Preparations

Moxifloxacin Intracameral Injection

Ph Eur

DEFINITION

1-Cyclopropyl-6-fluoro-8-methoxy-7-[(4*a*S,7*a*S)-octahydro-6*H*-pyrrolo[3,4-*b*]pyridin-6-yl]-4-oxo-1,4-dihydroquinoline-3-carboxylic acid hydrochloride.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

The production method is validated to demonstrate the satisfactory enantiomeric purity of the final product.

CHARACTERS

Appearance

Light yellow or yellow powder or crystals, slightly hygroscopic.

Solubility

Sparsely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

- Specific optical rotation (see Tests).
- Infrared absorption spectrophotometry (2.2.24). Comparison moxifloxacin hydrochloride CRS.
- Dissolve 50 mg in 5 mL of water R, add 1 mL of dilute nitric acid R, mix, allow to stand for 5 min and filter. The filtrate gives reaction (a) of chlorides (2.3.1).

TESTS

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution GY₂ (2.2.2, Method II). If intended for use in the manufacture of parenteral preparations, the solution is clear (2.2.1) and not more intensely coloured than reference solution GY₂ (2.2.2, Method II). Dissolve 1.0 g in 20 mL of dilute sodium hydroxide solution R. pH (2.2.3) 3.9 to 4.6.

Dissolve 0.10 g in 50 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7)

–125 to –138 (anhydrous substance). Dissolve 0.200 g in 20.0 mL of a mixture of equal volumes of acetonitrile R and water R.

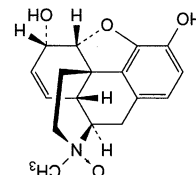
Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

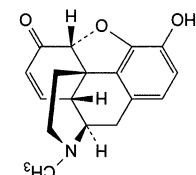
Solution A Dissolve 0.50 g of tetrabutylammonium hydrogen sulfate R and 1.0 g of potassium dihydrogen phosphate R in

Ph Eur

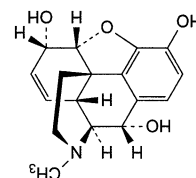
F, (1*S*)-7,8-didehydro-4,5*α*-epoxy-17-methylmorphinan-3,6*α*-diol 17-oxide (morphine *N*-oxide).



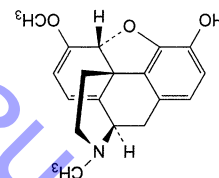
E, 7,8-didehydro-4,5*α*-epoxy-3-hydroxy-17-methylmorphinan-6-one (morphinone),



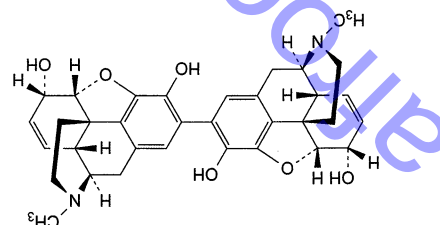
D, 7,8-didehydro-4,5*α*-epoxy-17-methylmorphinan-3,6*α*,10*α*-triol (10*S*-hydroxymorphine),



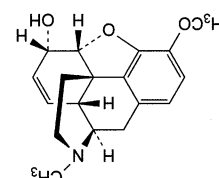
C, 6,7,8,14-tetradehydro-4,5*α*-epoxy-6-methoxy-17-methylmorphinan-3-ol (oripavine),



B, 7,7',8,8'-tetradehydro-4,5*α*,4',5'*α*-diepoxy-17,17'-dimethyl-2,2'-bimorphinan-3,3',6*α*,6'*α*-tetrol (2,2'-bimorphine),



A, 7,8-didehydro-4,5*α*-epoxy-3-methoxy-17-methylmorphinan-6*α*-ol (codeine),



about 500 mL of water R. Add 2 mL of phosphoric acid R and 0.050 g of anhydrous sodium sulfate R, then dilute to 1000.0 mL with water R.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with the same solution.

Test solution (b) Dilute 2.0 mL of test solution (a) to 20.0 mL with solution A.

Reference solution (a) Dissolve 50.0 mg of moxifloxacin hydrochloride CRS in solution A and dilute to 50.0 mL with the same solution. Dilute 2.0 mL of this solution to 20.0 mL with solution A.

Reference solution (b) Dissolve 5 mg of moxifloxacin for peak identification CRS (containing impurities A, B, C, D and E) in solution A and dilute to 5.0 mL with the same solution.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped phenylsilyl silica gel for chromatography R (5 μ m);
— temperature: 45 °C.

Mobile phase Mix 28 volumes of methanol R and 72 volumes of a solution containing 0.5 g/L of tetrabutylammonium hydrogen sulfate R, 1.0 g/L of potassium dihydrogen phosphate R, and 3.4 g/L of phosphoric acid R.

Detection Spectrophotometer at 293 nm.

Injection 10 μ L of test solution (a) and reference solutions (b) and (c).

Run time 2.5 times the retention time of moxifloxacin.

Identification of impurities Use the chromatogram supplied with moxifloxacin for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and E.

Relative retention With reference to moxifloxacin (retention time = about 14 min): impurity A = about 1.1; impurity B = about 1.3; impurity C = about 1.4; impurity D = about 1.6; impurity E = about 1.7.

System suitability: reference solution (b):
— resolution: minimum 1.5 between the peaks due to moxifloxacin and impurity A;

— the chromatogram obtained is similar to the chromatogram supplied with moxifloxacin for peak identification CRS.

Limits:
— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.4; impurity E = 3.5;

— impurities A, B, C, D, E: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);

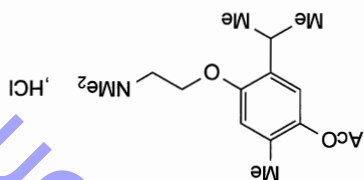
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (2.5.12)
Maximum 4.5 per cent, determined on 0.200 g.

Moxisylyte Hydrochloride



$C_{16}H_{25}NO_3 \cdot HCl$

315.8

964-52-3

Action and use

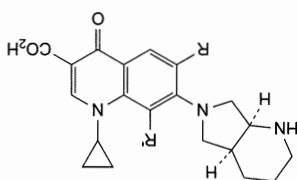
Alpha-adrenoceptor antagonist.

Preparation

Moxisylyte Tablets

DEFINITION

Moxisylyte Hydrochloride is 4-(2-dimethylaminoethoxy)-5-isopropyl-2-methylphenyl acetate hydrochloride. It contains not less than 99.0% and not more than 101.0% of



Specified impurities: A, B, C, D, E.

IMPURITIES

preparations.

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

LABELLING

In an airtight container, protected from light.

STORAGE

the declared content of moxifloxacin hydrochloride CRS.

Calculate the percentage content of $C_{21}H_{25}ClFN_3O_4$ from the declared content of moxifloxacin hydrochloride CRS.

ASSAY
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with the same solution.

Test solution (b) Dilute 2.0 mL of test solution (a) to 20.0 mL with solution A.

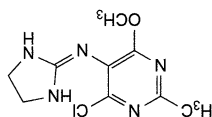
Reference solution (a) Dissolve 50.0 mg of moxifloxacin hydrochloride CRS in solution A and dilute to 50.0 mL with the same solution. Dilute 2.0 mL of this solution to 20.0 mL with solution A.

Reference solution (b) Dissolve 5 mg of moxifloxacin for peak identification CRS (containing impurities A, B, C, D and E) in solution A and dilute to 5.0 mL with the same solution.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Moxonidine

(Ph. Eur. monograph 1758)



$C_9H_{12}ClN_5O$

241.7

75438-57-2

Action and use

Imidazoline I_1 receptor antagonist; treatment of hypertension.

Ph Eur

DEFINITION

4-Chloro-N-(imidazolidin-2-ylidene)-6-methoxy-2-methylpyrimidin-5-amine.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, sparingly soluble in methanol, slightly soluble in methylene chloride, very slightly soluble in acetonitrile.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison moxonidine CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in a mixture of equal volumes of methanol R and water R and dilute to 100.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 10.0 mg of moxonidine CRS in a mixture of equal volumes of methanol R and water R and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with a mixture of equal volumes of methanol R and water R. Dilute 2.0 mL of this solution to 20.0 mL with a mixture of equal volumes of methanol R and water R.

Reference solution (c) Dissolve 5.0 mg of moxonidine impurity A CRS in a mixture of equal volumes of methanol R and water R and dilute to 100.0 mL with the same mixture of solvents.

Reference solution (d) Dilute 6.0 mL of reference solution (c) to 100.0 mL with a mixture of equal volumes of methanol R and water R.

Reference solution (e) Dilute 2.5 mL of reference solution (a) to 50.0 mL with reference solution (c).

Column:

— size: $l = 0.25$ m, $\varnothing = 4$ mm;

— stationary phase: base-deactivated octylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase Mix 136 volumes of acetonitrile R with

1000 volumes of a 3.48 g/L solution of sodium

Acidity
pH of a 5% w/v solution, 4.5 to 5.5, Appendix V L.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using solutions in the mobile phase containing (1) 0.0010% w/v of

2-(6-hydroxyethylamino)ethanolamine hydrochloride BPCRS and 0.00050% w/v of 2-thymoxethylamino

hydrochloride BPCRS and (2) 0.2% w/v of the substance being examined. For solution (3) dilute 1 volume of solution (2) to 100 volumes. For solution (4) dilute 1 volume of

solution (3) to 10 volumes.

The chromatographic procedure may be carried out using (a) a stainless steel column (30 cm \times 3.9 mm) packed with octadecylsilyl silica gel for chromatography (10 μ m) (µBondapak C18 is suitable), (b) 0.005M sodium hexanesulfonate in a mixture of 2 volumes of glacial acetic acid, 185 volumes of water and 315 volumes of the mobile phase with a flow rate of 1.0 mL per minute and (c) a detection wavelength of 276 nm.

The peaks in the chromatogram obtained with solution (1)

are due to (a) 2-(6-hydroxyethylamino)ethanolamine hydrochloride and (b) 2-thymoxethylamino hydrochloride in order of their elution. In the chromatogram obtained with solution (2) the areas of any peaks corresponding to (a) and (b) are not greater than the areas of the corresponding peaks in the chromatogram obtained with

solution (1) (0.5% of 2-(6-hydroxy-

2-thymoxethylamino hydrochloride and 0.25% of the area of any peak with a retention time relative to

moxislyte of 2.3 is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (1% of 2-(6-chloroethylamino)ethanolamine hydrochloride) and the area of any peak with a retention time relative to

moxislyte of 0.9 is not greater than the area of the principal peak in the chromatogram obtained with solution (4) (0.1% of 2-(6-acetylthymoxethylamino)ethanolamine).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for non-aqueous titration,

Appendix VIII A, using 0.5 g and 1-naphtholbenzenesulfonic acid VS is equivalent as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 31.58 mg of $C_{16}H_{25}NO_3HCl$.

STORAGE

Moxislyte Hydrochloride should be protected from light.



pentanesulfonate R previously adjusted to pH 3.5 with dilute sulfuric acid R.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 µL; inject a blank, the test solution and reference solutions (b), (d) and (e).

Run time Twice the retention time of moxonidine.

Relative retentions With reference to moxonidine (retention time = about 11.6 min): impurity A = about 0.9;

impurity B = about 1.7.

System suitability: reference solution (e):

— resolution: minimum of 2 between the peaks due to

impurity A and moxonidine.

Limits:

— impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (d) (0.3 per cent);

— impurity B: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.3 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent); disregard any peak observed with the

blank run.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of C₂₆H₄₄O from the

areas of the peaks and the declared content of

moxonidine CRS.

IMPURITIES

Specified impurities A, B

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

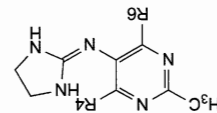
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

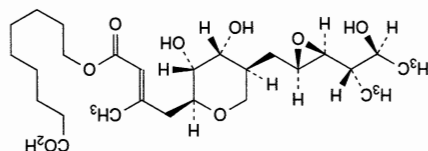
Control of impurities in substances for pharmaceutical use): C, D.



A, R4 = R6 = Cl; 4,6-dichloro-N-(imidazolidin-2-ylidene)-2-methylpyrimidin-5-amine (6-chloromoxonidine),
B, R4 = R6 = OCH₃; N-(imidazolidin-2-ylidene)-4,6-dimethoxy-2-methylpyrimidin-5-amine (4-methoxymoxonidine),

Mupirocin

(Ph. Eur. monograph 1450)



C₂₆H₄₄O₉ 500.6 12650-69-0

Action and use

Antibacterial.

Preparation

Mupirocin Ointment

Ph. Eur.

DEFINITION

9-[[[(2E)-4-[(2S,3R,4R,5S)-3,4-dihydroxy-5-[[[(2S,3S)-3-methylbut-2-en-2-yl]oxy]nonanoic acid. Substance produced by the growth of certain strains of *Pseudomonas fluorescens* or obtained by any other means.

Content

93.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, freely soluble in acetone, in

anhydrous ethanol and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of mupirocin.

TESTS

pH (2.2.3)

3.5 to 4.0 for a freshly prepared saturated solution (about

10 g/L) in carbon dioxide-free water R.

Specific optical rotation (2.2.7)

–17 to –21 (anhydrous substance).

Dissolve 0.50 g in methanol R and dilute to 10.0 mL with the

same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mix 50 volumes of methanol R and

50 volumes of a 13.6 g/L solution of sodium acetate R

adjusted to pH 4.0 with acetic acid R.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the solvent mixture.

Reference solution (b) Adjust 10 mL of reference solution (a) to pH 2.0 with hydrochloric acid R and allow to stand for 20 h.

Reference solution (c) Dissolve 25 mg of mupirocin lithium CRS in the solvent mixture and dilute to 200.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 20 volumes of water R, 30 volumes of tetrahydrofuran R and 50 volumes of a 10.5 g/L solution of ammonium acetate R adjusted to pH 5.7 with acetic acid R.

Detection Spectrophotometer at 240 nm.

Injection 20 μ L.

Run time 3.5 times the retention time of mupirocin.

Relative retention With reference to mupirocin:

impurity C = about 0.75.

System suitability: reference solution (b):

— resolution: minimum 7.0 between the 2nd of the 2 peaks due to hydrolysis products and the peak due to mupirocin.

Limits:

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (4 per cent);

— any other impurity: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (6 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent).

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in 5 mL of methanol R and dilute to 200.0 mL with a 7.5 g/L solution of ammonium acetate R adjusted to pH 5.7 with acetic acid R.

Reference solution (a) Dissolve 25.0 mg of mupirocin lithium CRS in 5 mL of methanol R and dilute to 200.0 mL with a 7.5 g/L solution of ammonium acetate R adjusted to pH 5.7 with acetic acid R.

Reference solution (b) Adjust 10 mL of the test solution to pH 2.0 with hydrochloric acid R and allow to stand for 20 h.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

IMPURITIES

Protected from light.

Specified impurities C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D, E, F.

STORAGE

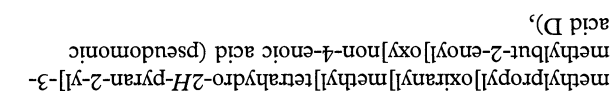
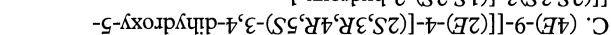
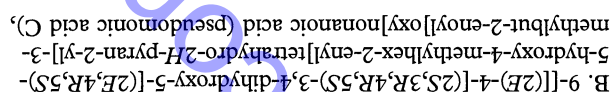
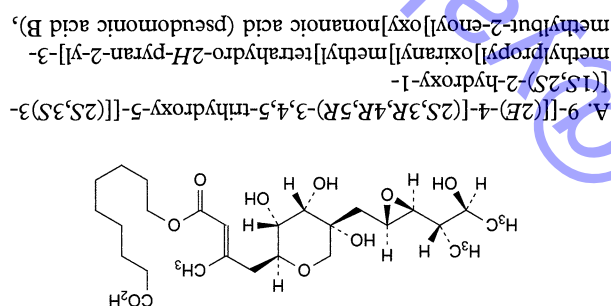
Protected from light.

IMPURITIES

Specified impurities C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D, E, F.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D, E, F.



Content
93.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility

Very slightly soluble in water, sparingly soluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison Ph. Eur. reference spectrum of mupirocin calcium.
B. It gives reaction (a) of calcium (2.3.1).

TESTS

Specific optical rotation (2.2.7)

–16 to –20 (anhydrous substance).

Dissolve 0.50 g in methanol R and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mix 50 volumes of methanol R and 50 volumes of a 13.6 g/L solution of sodium acetate R adjusted to pH 4.0 with acetic acid R.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the solvent mixture.

Reference solution (b) Adjust 10 mL of reference solution (a) to pH 2.0 with hydrochloric acid R and allow to stand for 20 h.

Reference solution (c) Dissolve 25 mg of mupirocin lithium CRS in the solvent mixture and dilute to 200.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 20 volumes of water R, 30 volumes of tetrahydrofuran R and 50 volumes of a 10.5 g/L solution of ammonium acetate R adjusted to pH 5.7 with acetic acid R. Flow rate 1 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 μ L.

Run time 3.5 times the retention time of mupirocin.

Relative retention With reference to mupirocin.

impurity C = about 0.75.

System suitability: reference solution (b):
— resolution: minimum 7.0 between the 2nd of the 2 peaks due to hydrolysis products and the peak due to mupirocin.

Limits:

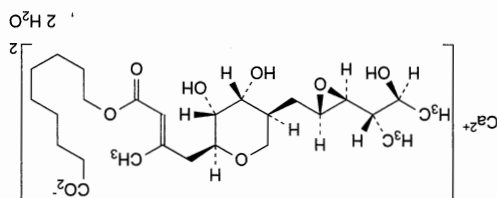
— **impurity C:** not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (2.5 per cent);
— **any other impurity:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);
— **total:** not more than 2.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (4.5 per cent);

Ph Eur



Mupirocin Calcium

(Ph. Eur. monograph 1451)



C₂H₈₆CaO₁₈·2H₂O 1075 115074-43-6

Preparations
Mupirocin Cream
Mupirocin Nasal Ointment

Action and use
Antibacterial.

Ph Eur

DEFINITION

Calcium bis[9-[[[(2E)-4-[(2S,3R,4R,5S)-3,4-dihydroxy-5-[(2S,3S)-3-[(1S,2S)-2-hydroxy-1-methylpropyl]oxiran-2-yl]tetrahydro-2H-pyran-2-yl]-3-methylbut-2-en-1-yl]oxy]nonanoate] dihydrate. Substance produced by the growth of certain strains of *Pseudomonas fluorescens* or obtained by any other means.

— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.1 per cent).

Chlorides (2.4.4)

Maximum 0.5 per cent.

Dissolve 10.0 mg in a mixture of 1 mL of dilute nitric acid R

and 15 mL of methanol R.

Water (2.5.12)

3.0 per cent to 4.5 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be

examined in 5 mL of methanol R and dilute to 200.0 mL

with a 7.5 g/L solution of ammonium acetate R adjusted to

pH 5.7 with acetic acid R.

Reference solution (a) Dissolve 25.0 mg of mupirocin

lithium CRS in 5 mL of methanol R and dilute to 200.0 mL

with a 7.5 g/L solution of ammonium acetate R adjusted to

pH 5.7 with acetic acid R.

Reference solution (b) Adjust 10 mL of the test solution to

pH 2.0 with hydrochloric acid R and allow to stand for 20 h.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Mix 19 volumes of water R, 32 volumes of

tetrahydrofuran R and 49 volumes of a 10.5 g/L solution of

ammonium acetate R adjusted to pH 5.7 with acetic acid R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

System suitability:

— resolution: minimum 7.0 between the 2nd of the 2 peaks

due to hydrolysis products and the peak due to mupirocin

in the chromatogram obtained with reference solution (b);

— repeatability: maximum relative standard deviation of

1.0 per cent after 6 injections of reference solution (a).

Calculate the percentage content of mupirocin calcium by

multiplying the percentage content of mupirocin in

mupirocin lithium by 1.038.

IMPURITIES

Specified impurities C

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

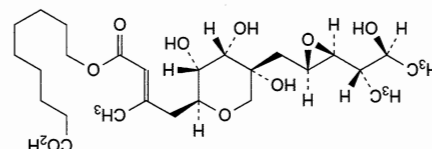
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

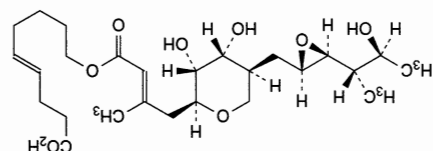
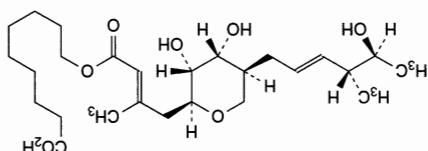
Control of impurities in substances for pharmaceutical use: A, B,

D, E, F, G, H, I.

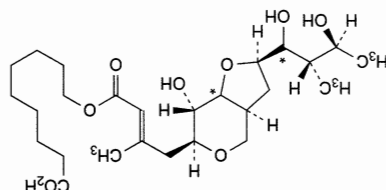


A. 9-[[[(2E)-4-[(2S,3R,4R,5R)-3,4,5-trihydroxy-5-[[[(2S,3S)-3-methylbut-2-en-1-yl]oxy]oxiranyl]tetrahydro-2H-pyran-2-yl]-3-methylbut-2-en-1-yl]oxy]nonanoic acid (pseudomonic acid B),

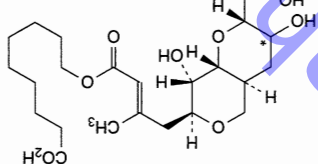
B. 9-[[[(2E)-4-[(2S,3R,4R,5S)-3,4-dihydroxy-5-[[[(2E,4R,5S)-5-hydroxy-4-methylhex-2-enyl]tetrahydro-2H-pyran-2-yl]-3-methylbut-2-en-1-yl]oxy]nonanoic acid (pseudomonic acid C),



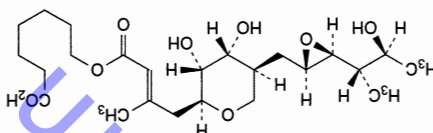
C. (4E)-9-[[[(2E)-4-[(2S,3R,4R,5S)-3,4-dihydroxy-5-[[[(2S,3S)-3-[[[(1S,2S)-2-hydroxy-1-methylpropyl]oxiranyl]tetrahydro-2H-pyran-2-yl]-3-methylbut-2-en-1-yl]oxy]non-4-enoic acid (pseudomonic acid D),



D. 9-[[[(2E)-4-[(2R,3aS,6S,7S)-2-[[[(2S,3S)-1,3-dihydroxy-2-methylbutyl]-7-hydroxyhexahydro-4H-furo[3,2-c]pyran-6-yl]-3-methylbut-2-en-1-yl]oxy]nonanoic acid,



E. 9-[[[(2E)-4-[(2R,3RS,4aS,7S,8S,8aR)-3,8-dihydroxy-2-[[[(1S,2S)-2-hydroxy-1-methylpropyl]hexahydro-2H,5H-pyrano[4,3-b]pyran-7-yl]-3-methylbut-2-en-1-yl]oxy]nonanoic acid,



F. 7-[[[(2E)-4-[(2S,3R,4R,5S)-3,4-dihydroxy-5-[[[(2S,3S)-3-methylbut-2-en-1-yl]oxy]oxiranyl]tetrahydro-2H-pyran-2-yl]-3-methylbut-2-en-1-yl]oxy]heptanoic acid,

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dissolve 0.10 g in ethanol (96 per cent) R and dilute to

10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Protect the solutions from light. Prepare the solutions immediately before use, or store them at 4–8 °C. Keep the temperature of the solutions to equilibrate in the vials for 15 min before injection.

Test solution Dissolve 20 mg of the substance to be examined in acetonitrile R and dilute to 10 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with acetonitrile R. Dilute 1.0 mL of this solution to 10.0 mL with acetonitrile R.

Reference solution (b) Dissolve 5 mg of mycophenolate mofetil for peak identification CRS (mycophenolate mofetil with

impurities A, B, D, E, F, G and H) in acetonitrile R and dilute to 2.5 mL with the same solvent.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m),

— temperature: 45 °C.

Mobile phase Mix 350 mL of acetonitrile R with a mixture of 650 mL of water R and 2.0 mL of triethylamine R previously adjusted to pH 5.3 with dilute phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 μ L.

Run time 3 times the retention time of mycophenolate

mofetil.

Relative retention With reference to mycophenolate mofetil (retention time = about 22 min): impurity F = about 0.3; impurity A = about 0.4; impurity H = about 0.5; impurity G = about 0.6; impurity B = about 0.8; impurity D = about 1.2; impurity E = about 1.6.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurity A and impurity H,

— the chromatogram obtained is similar to the

chromatogram supplied with mycophenolate mofetil for peak identification CRS.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity B by 2.1,

— **impurity F:** not more than 5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.5 per cent),

— **impurity B:** not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.2 per cent),

— **impurities A, D, E, G, H:** for each impurity, not more

than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent),

— **any other impurity:** for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.1 per cent),

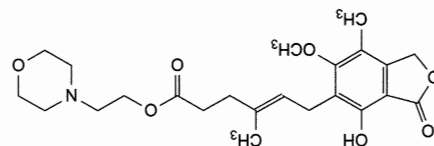
— **total:** not more than 7 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.7 per cent).

Mycophenolate Mofetil

(Ph. Eur. monograph 1700)



$C_{23}H_{31}NO_7$ 433.5

128794-94-5

Action and use

Inhibitor of nucleic acid synthesis; immunomodulator.

DEFINITION

2-(Morpholin-4-yl)ethyl (4E)-6-(4-hydroxy-6-methoxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoate.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone,

sparingly soluble in anhydrous ethanol.

mp

About 96 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24). Comparison mycophenolate mofetil CRS.

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with limit test F. Prepare the reference

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 43.35 mg

of $C_{23}H_{31}NO_7$.

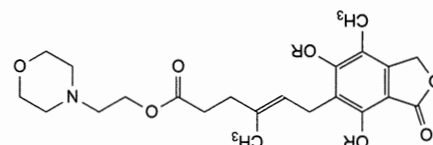
STORAGE

Protected from light.

IMPURITIES

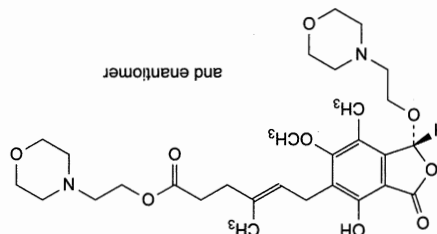
Specified impurities A, B, D, E, F, G, H

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.

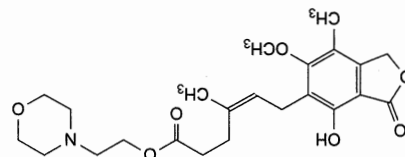


A. R = H: 2-(morpholin-4-yl)ethyl (4E)-6-(4,6-dihydroxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoate,

D. R = CH_3 : 2-(morpholin-4-yl)ethyl (4E)-6-(4,6-dimethoxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoate,



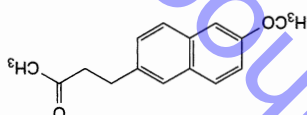
B. 2-(morpholin-4-yl)ethyl (4E)-6-[(1R)-4-hydroxy-6-methoxy-7-methyl-1-[2-(morpholin-4-yl)ethoxy]-3-oxo-1,3-dihydroisobenzofuran-5-yl]-4-methylhex-4-enoate,



C. 2-(morpholin-4-yl)ethyl (4Z)-6-(4-hydroxy-6-methoxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoate,

Nabumetone

(Ph. Eur. monograph 1350)



$C_{15}H_{16}O_2$

228.3

42924-53-8

Action and use
Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparations
Nabumetone Oral Suspension
Nabumetone Tablets

Ph. Eur.

DEFINITION

4-(6-Methoxynaphthalen-2-yl)butan-2-one.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

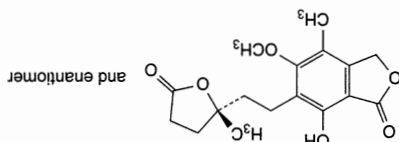
Practically insoluble in water, freely soluble in acetone, slightly soluble in methanol.

IDENTIFICATION

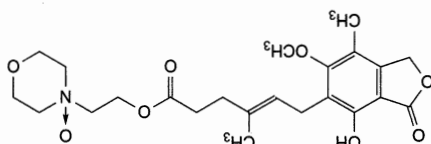
Infrared absorption spectrophotometry (2.2.24).

Ph. Eur.

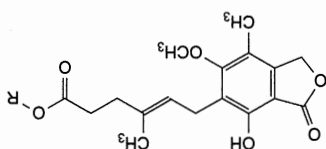
H. 7-hydroxy-5-methoxy-4-methyl-6-[2-[(2R)-2-methyl-5-oxotetrahydrofuran-2-yl]ethyl]isobenzofuran-1(3H)-one.



G. 2-(morpholin-4-yl)ethyl (4E)-6-(4-hydroxy-6-methoxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoate N-oxide,



E. R = CH_3 : methyl (4E)-6-(4-hydroxy-6-methoxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoate, F. R = H: (4E)-6-(4-hydroxy-6-methoxy-7-methyl-3-oxo-1,3-dihydroisobenzofuran-5-yl)-4-methylhex-4-enoic acid (mycophenolic acid),



Comparison nabumetone CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in acetonitrile R and dilute to 10.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 25.0 mL with acetonitrile R. Dilute 1.0 mL of this solution to 5.0 mL with acetonitrile R.

Reference solution (a) Dissolve 20.0 mg of nabumetone CRS in acetonitrile R and dilute to 10.0 mL with the same solvent.

Dilute 1.0 mL of this solution to 50.0 mL with acetonitrile R.

Reference solution (b) Dilute 0.5 mL of test solution (a) to 100.0 mL with acetonitrile R.

Reference solution (c) Dissolve 1.5 mg of nabumetone impurity F CRS in acetonitrile R and dilute to 100.0 mL with the same solvent.

Reference solution (d) Dissolve 4 mg of nabumetone impurity D CRS in acetonitrile R and dilute to 100 mL with the same solvent.

To 5 mL of this solution, add 5 mL of test solution (b).

Column: size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (4 μ m);

temperature: 40 °C.

Mobile phase: mobile phase A: mix 12 volumes of tetrahydrofuran R, 28 volumes of acetonitrile for chromatography R and 60 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

mobile phase B: mix 24 volumes of tetrahydrofuran R, 56 volumes of acetonitrile for chromatography R and 20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

20 volumes of 0.1 per cent V/V solution of glacial acetic acid R in carbon dioxide-free water R prepared from distilled water R;

— sum of impurities other than F: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8). Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12). Maximum 0.2 per cent, determined on 1.000 g.

Sulfated ash (2.4.14). Maximum 0.1 per cent, determined on 1.0 g.

ASSAY Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution (b) and reference solution (a).

System suitability: reference solution (a):

— repeatability: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of $C_{15}H_{16}O_2$ from the declared content of nabumetone CRS.

STORAGE Protected from light.

IMPURITIES Specified impurities F

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use): A, B, C, D, E.

A. 3-(6-methoxynaphthalen-2-yl)-5-methylcyclohexanone,

B. (5*RS*)-5-(6-methoxynaphthalen-2-yl)-3-methylcyclohex-2-enone,

C. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

D. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

E. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

F. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

G. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

H. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

I. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

J. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

K. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

L. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

M. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

N. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

O. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

P. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

Q. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

R. (2*RS*)-4-(6-methoxynaphthalen-2-yl)butan-2-ol,

using *liquid paraffin* R as reference. Measure the absorbance A_{90} corresponding to racemate A, at the maximum at 1266 cm^{-1} and the absorbance A_{90} corresponding to racemate B, at the maximum at 1250 cm^{-1} . The ratio A_{1266}/A_{1250} is 0.72 to 1.08 (corresponding to racemate A content of between 40 per cent and 60 per cent).

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture acetonitrile R1, water R (20:80 V/V).

Test solution Dissolve 0.100 g of the substance to be examined in 4.0 mL of the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this

solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of nadolol impurity mixture CRS (impurities A and D) in 1.0 mL of reference solution (a).

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μm);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: 5.6 g/L solution of sodium octanesulfonate adjusted to pH 3.5 with a 300 g/L solution of phosphoric acid R;

— mobile phase B: acetonitrile R1;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 7	77	23
7 - 30	77 → 65	23 → 35
30 - 35	65 → 55	35 → 45
35 - 55	55	45

Flow rate 1 mL/min.

Detection Spectrophotometer at 206 nm.

Injection 20 μL .

Identification of impurities Use the chromatogram supplied with nadolol impurity mixture CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and D.

Relative retention With reference to nadolol (retention time = about 15 min): impurity A = about 0.2; impurity C (doublet) = about 0.47 and 0.53; impurity D = about 1.5.

System suitability: reference solution (b):

— resolution: minimum 8.0 between the peaks due to nadolol and impurity D.

Limits:

— correction factor: for the calculation of content, multiply the sum of the 2 peak areas of impurity C by 0.7;

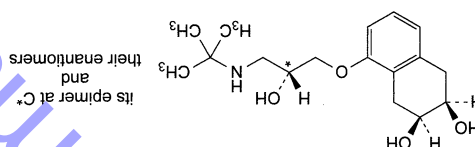
— impurities A, C, D: for each impurity, not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

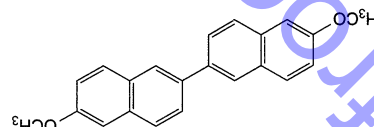
Nadolol

(Ph. Eur. monograph 1789)

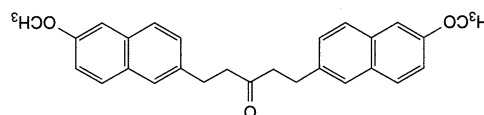


Ph Eur

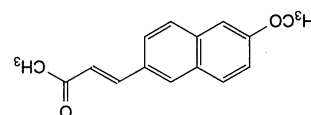
F. 6,6'-dimethoxy-2,2'-binaphthalenyl.



E. 1,5-bis-(6-methoxynaphthalen-2-yl)pentan-3-one,



D. (E)-4-(6-methoxynaphthalen-2-yl)but-3-en-2-one,



DEFINITION

It consists of 2 pairs of enantiomers that are present as *cis*-5-[(2*R*)-3-[(1,1-Dimethylethyl)amino]-2-hydroxypropoxy]-1,2,3,4-tetrahydronaphthalene-2,3-diol.

Content 98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nadolol CRS.

TESTS

Racemate content

Infrared absorption spectrophotometry (2.2.24).

Prepare a mull in *liquid paraffin* R of the substance to be examined (dried substance), adjusting the thickness of the mull to give an absorbance reading of 0.6 ± 0.1 at 1587 cm^{-1} . Record the spectrum from 1667 to 1111 cm^{-1} .

- total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- disregarded limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 30 ppm.

1.0 g complies with test D. Prepare the reference solution using 3 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 2.0 per cent, determined on 1.000 g by drying in *vacuo* at 60 °C for 3 h.

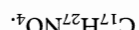
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 100 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

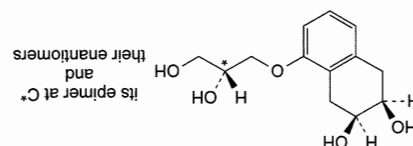
1 mL of 0.1 M perchloric acid is equivalent to 30.94 mg of



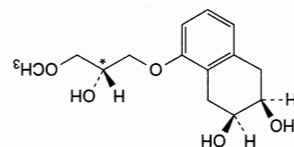
IMPURITIES

Specified impurities A, C, D

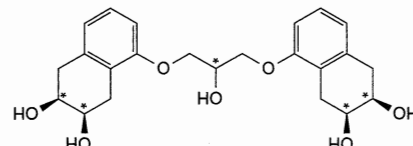
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: B, E, F, G.



A. *cis*-5-[(2*R*)-2,3-dihydroxypropoxy]-1,2,3,4-tetrahydronaphthalene-2,3-diol (tetraol),



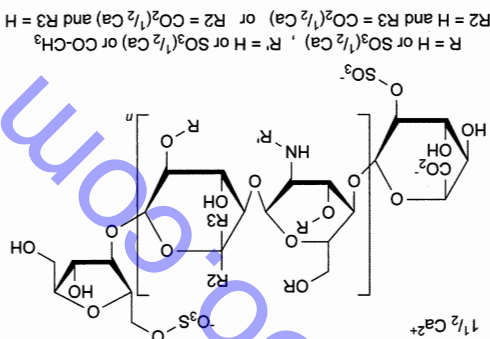
B. *cis*-5-[(2*R*)-2-hydroxy-3-methoxypropoxy]-1,2,3,4-tetrahydronaphthalene-2,3-diol,



C. 5,5'-[[(2*R*)-2-hydroxypropyl]-1,3-bis(oxymethyl)]bis(*cis*-1,2,3,4-tetrahydronaphthalene-2,3-diol) (3 diastereoisomers),

Nadroparin Calcium

(Ph. Eur. monograph 1134)

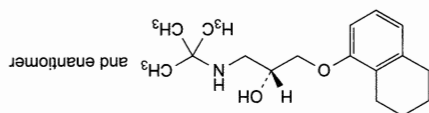


Action and use

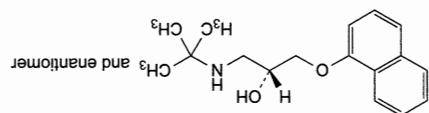
Low molecular weight heparin.

DEFINITION

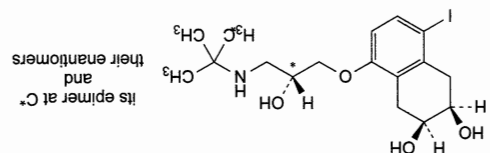
Calcium salt of low-molecular-mass heparin obtained by nitrous acid depolymerisation of heparin from pork intestinal mucosa, followed by fractionation to eliminate selectively most of the chains with a molecular mass lower than 2000. The majority of the components have a 2-O-sulfo-α-L-



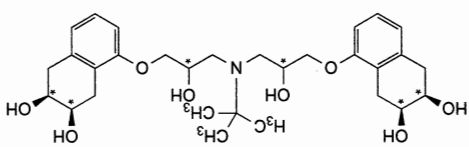
F. (2*R*)-1-[(1,1-dimethylethyl)amino]-3-(naphthalen-1-yl)oxy]propan-2-ol,



G. (2*R*)-1-[(1,1-dimethylethyl)amino]-3-(5,6,7,8-tetrahydronaphthalen-1-yl)oxy]propan-2-ol.



E. *cis*-5-[(2*R*)-3-[(1,1-dimethylethyl)amino]-2-hydroxypropoxy]-1,2,3,4-tetrahydronaphthalene-2,3-diol,



D. 5,5'-[[(1,1-dimethylethyl)imino]bis(*cis*-1,2,3,4-tetrahydronaphthalene-2,3-diol)]bis(*cis*-1,2,3,4-tetrahydronaphthalene-2,3-diol) (10 stereoisomers),

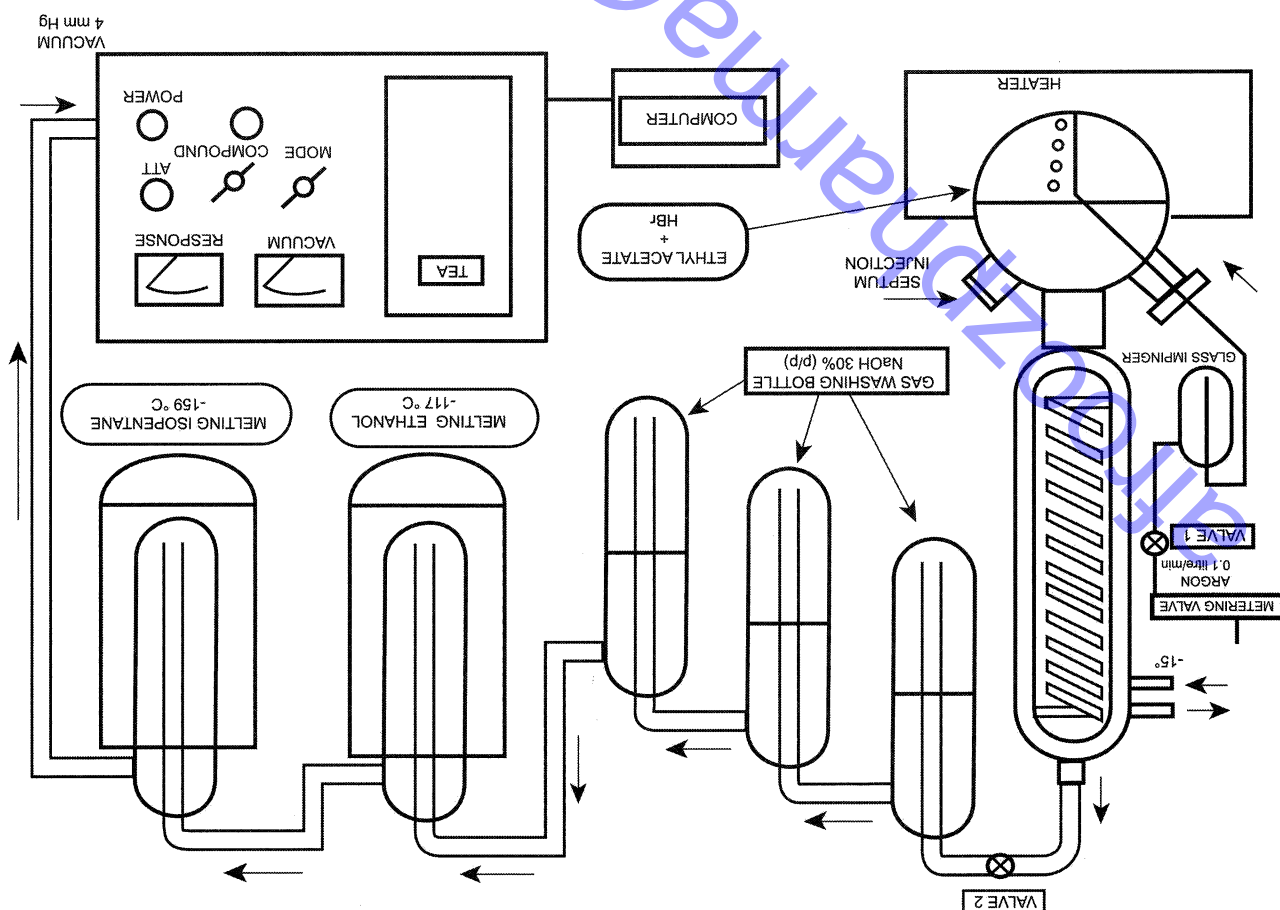


Figure 1134.-1. - Apparatus used for the assay of N-NO groups

Bubble traps. Height: 24 cm, internal diameter: 2.5 cm, internal tubing 23 cm in length by 0.5 cm internal diameter. Centrally positioned Rotulex mounting. Equipped with torion joints on the inlet and outlet.

Chemiluminescence detector.

Cold trap. Height: 16.5 cm, internal diameter: 4 cm, internal tubing 14 cm in length and internal diameter 1.3 cm. Equipped with torion joints on the inlet and outlet.

Condenser. Height: 21 cm, internal diameter: 3 cm. Lower rodavis joint and upper torion joint.

Flask. Round-bottomed borosilicate glass flask equipped with a central rodavis joint, a torion joint on the left neck and a 15 cm screw joint on the right neck.

Isothermic flask. Internal depth: 22 cm, internal diameter: 8 cm.

Septum. Silicone material, diameter: 14 mm, thickness: 3.5 mm.

Torion joint.

Tubing. Polytetrafluoroethylene FEP material, internal diameter: 3.2 mm, thickness: 0.8 mm.

idopyranosuronic acid structure at the non-reducing end and a 6-O-sulfo-2,5-anhydro-D-mannitol structure at the reducing end of their chain.

Nadroparin calcium complies with the modifications and molecular-mass heparins (0828) with the modifications and additional requirements below.

The mass-average relative molecular mass ranges between 3600 and 5000 with a characteristic value of about 4300. The degree of sulfatation is about 2 per disaccharide unit. The potency is not less than 95 IU and not more than 130 IU of anti-factor Xa activity per milligram, calculated with reference to the dried substance. The ratio of anti-factor Xa activity to anti-factor IIa activity is between 2.5 and 4.0.

IDENTIFICATION

Carry out identification test A as described in the monograph *Low-molecular-mass heparins (0828)* using *nadroparin calcium CRS*.

solvent.

Dissolve 0.5 g in *water R* and dilute to 10 mL with the same

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, *Method II*).

TESTS

The mass-average relative molecular mass ranges between 3600 and 5000. The mass percentage of chains lower than 2000 is not more than 15 per cent. The mass percentage of chains between 2000 and 8000 ranges between 75 per cent and 95 per cent. The mass percentage of chains between 2000 and 4000 ranges between 35 per cent and 55 per cent.

Carry out identification test C as described in the monograph *Low-molecular-mass heparins (0828)*. The following requirements apply.

Ethanol

Head-space gas chromatography (2.2.28).

Internal standard solution Dilute 1.0 mL of 2-propanol R to 100.0 mL with water R. Dilute 1.0 mL of the solution to 50.0 mL with water R.

Blank solution 1.0 mL of water R.

Test solution (a) To 10.0 mg of the substance to be examined, add 1.0 mL of water R.

Test solution (b) To 10.0 mg of the substance to be examined, add 0.50 mL of water R and 0.50 mL of the

internal standard solution.

Test solution (c) Dilute 1.0 mL of anhydrous ethanol R to 100.0 mL with water R. Dilute 0.5 mL of the solution to 20.0 mL with water R.

Reference solution (b) To 0.50 mL of reference solution (a), add 0.50 mL of the internal standard solution.

Column:

— material: nickel;

— size: $l = 1.5$ m; $\varnothing = 2$ mm;

— stationary phase: ethylvinylbenzene-divinylbenzene

copolymer R (150-180 μ m).

Carrier gas helium for chromatography R or nitrogen for chromatography R.

Flow rate 30 mL/min.

Static head-space conditions that may be used:

— equilibration temperature: 90 °C;

— equilibration time: 15 min;

— pressurisation time: 1 min.

Temperature:

— column: 150 °C;

— injection port and detector: 250 °C.

Detection Flame ionisation.

Identification of peaks Use the chromatogram obtained with reference solution (b) to identify the peaks due to ethanol

and 2-propanol.

Retention time Ethanol = about 2.5 min; 2-propanol = about 4 min.

Calculate the percentage content *m/m* of ethanol taking its

density at 20 °C to be 0.792 g/mL.

Limit:

— ethanol: maximum 1.0 per cent *m/m*.

N-NO groups

Maximum 0.25 ppm.

The content of N-NO-groups is determined by cleavage of the N-NO bond with hydrotromic acid in ethyl acetate under a reflux condenser and detection of the released NO by chemiluminescence.

Description of the apparatus (Figure 1134-1). Use a 500 mL borosilicate glass round-bottomed flask, above which is

attached a condenser which is equipped with:

— on one side, a torsion joint through which a stream of argon R can be introduced via a cannula;

— on the other side, a screw joint with a piston equipped with a septum through which the reference solution and

test solution will be injected.

The round-bottomed flask is connected in series to 3 bubble traps which are themselves connected to 2 cold traps, which are in turn connected to a chemiluminescence detector. Suitable tubing ensures the junctions are leak-free.

Preparation of the chemiluminescence detector Switch on the vacuum pump. The vacuum must be less than 0.5 mm Hg. 1 h before use, open the oxygen valve at a pressure of 0.2 MPa and a flow rate of 9.4 mL/min.

Preparation of the bubble trap In each bubble trap, place 30 mL of a 300 g/L solution of sodium hydroxide R in water R.

Preparation of the cold traps.

— Trap at -120 °C: Slowly add liquid nitrogen to an isothermic flask containing 250 mL of anhydrous ethanol R whilst stirring with a wooden spatula until a paste is obtained. Place the cold trap in the isothermic flask

prepared as described.

— Trap at -160 °C: Slowly add liquid nitrogen to an isothermic flask containing 250 mL of 2-methylbutane R whilst stirring with a wooden spatula until a paste is obtained. Place the cold trap in the isothermic flask

prepared as described.

Drying of the 500 mL borosilicate-glass round-bottomed flask and condenser Boil 50 mL of ethyl acetate R under reflux for 1 h under argon R without connecting the system to the

chemiluminescence detector.

Test solution Dry the substance to be examined for 12 h over diphosphorus pentoxide R at 60 °C under vacuum. Dissolve 0.10 g of the treated substance to be examined in 1.0 mL of treated formamide R. Shake the solution obtained for 30 min.

Reference solution Dilute 0.1 mL of nitrosodipropylamine solution R in 6.0 mL of anhydrous ethanol R. Dilute 0.1 mL of the solution obtained in 1.0 mL of treated formamide R. (This solution is equivalent to 0.05 ppm of N-NO groups).

Place 50 mL of treated ethyl acetate R in the dry 500 mL borosilicate glass round-bottomed flask equipped with a septum. Connect the round-bottomed flask to the condenser which has been previously cooled to -15 °C for 2 h.

Connect the argon R cannula and adjust the flow rate to 0.1 L/min. Check that the system is leak-free. Only the connector to the chemiluminescence detector remains open in order to avoid excess pressure.

Heat the treated ethyl acetate R to boiling.

Evacuate the system by slowly turning the valve of the chemiluminescence detector. At the same time tighten the inlet on the chemiluminescence detector.

When the system is equilibrated, the vacuum reaches

4 mm Hg.

The signal of the zero adjuster on the chemiluminescence detector is set to 10 per cent of the full scale of the recorder. Through the septum of the 500 mL borosilicate glass round-bottomed flask, sequentially inject 0.5 mL of water R,

2.0 mL of dilute hydrotromic acid R and then another 2.0 mL of dilute hydrotromic acid R, making sure that the recorder pen has returned to the baseline between each injection.

Inject 50.0 μ L of the reference solution, then 50.0 μ L of the test solution after the recorder pen has returned to the

baseline.

Calculate the content of N-NO groups of the substance to be examined.

Free sulfates

Liquid chromatography (2.2.29).

Test solution Dissolve 30.0 mg of the substance to be examined in water R and dilute to 10.0 mL with the same

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Freely soluble in water, freely soluble or soluble in ethanol (96 per cent), slightly or sparingly soluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 1.0 g in water R and dilute to 50 mL with the same solvent. Add 2 mL of concentrated ammonia R and shake with 3 quantities, each of 10 mL, of methylene chloride R. To the combined lower layers, add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate at a temperature not exceeding 30 °C, using a rotary evaporator. Use the residue obtained.

Comparison Ph. Eur. reference spectrum of nafidrofuryl.

B. Dissolve 0.5 g in water R and dilute to 10 mL with the same solvent. Add 2.0 mL of calcium chloride solution R. A white precipitate is formed. The precipitate dissolves after the addition of 3.0 mL of hydrochloric acid R.

TESTS**Absorbance (2.2.25)**

Maximum 0.1 at 430 nm. Dissolve 1.5 g in water R and dilute to 10 mL with the same solvent. If necessary use an ultrasonic bath.

Related substances

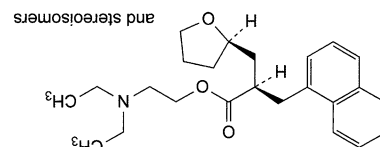
A. Liquid chromatography (2.2.29).

Test solution Dissolve 80.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase. Treat in an ultrasonic bath for 10 s. A precipitate is formed. Filter through a membrane filter (nominal pore size 0.45 µm), discarding the first 5 mL. Use a freshly prepared solution.

Reference solution (a) Dissolve 5.0 mg of nafidrofuryl impurity A CRS in acetonitrile R and dilute to 25.0 mL with the same solvent. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase. Reference solution (b) Dissolve 5 mg of the substance to be examined in acetonitrile R and dilute to 50 mL with the same solvent. Dilute 1 mL of this solution to 50 mL with the mobile phase. Column: phase. size: $l = 0.25$ m, $\varnothing = 4.6$ mm; stationary phase: spherical and capped octadecylsilyl silica gel for chromatography R (5 µm) with a specific surface area of 350 m²/g, a pore size of 10 nm and a carbon loading of 14 per cent. Mobile phase Mix 60 mL of methanol R with 150 mL of tetrabutylammonium buffer solution pH 7.0 R and dilute to 1000 mL with acetonitrile R. Flow rate 1 mL/min. Detection Spectrophotometer at 283 nm. Injection 20 µL. Run time 2.3 times the retention time of nafidrofuryl. Relative retention With reference to nafidrofuryl (retention time = about 7 min): impurity A = about 0.5; impurity B = about 0.8; impurity C = about 1.8. System suitability: reference solution (b): — resolution: minimum 3.0 between the peaks due to impurity B and nafidrofuryl.

Nafidrofuryl Oxalate

(Nafidrofuryl Hydrogen Oxalate, Ph Eur monograph 1594)



C₂₆H₃₅NO₇ 473.6 3200-06-4

Action and use

Vasodilator.

Preparation

Nafidrofuryl Capsules

Ph Eur

DEFINITION

Mixture of 4 stereoisomers of 2-(diethylamino)ethyl 2-[(naphthalen-1-yl)methyl]-3-(tetrahydrofuran-2-yl)propanoate hydrogen oxalate.

Content

99.0 per cent to 101.0 per cent (dried substance).

Reference solution Dissolve 1.4787 g of anhydrous sodium sulfate R in water R and dilute to 1000.0 mL with the same solvent. Dilute 1.0 mL of the solution to 200.0 mL with distilled water R (5 ppm of sulfate ions). Column: size: $l = 50$ mm, $\varnothing = 4.6$ mm; stationary phase: anion-exchange resin. Chemical neutralisation system Neutralisation micromembrane in line with the mobile phase for anion detection; continuously pump in counter-flow with a 2.45 g/L solution of sulfuric acid R, at a flow rate of 4 mL/min. Mobile phase: — mobile phase A: 1.91 g/L solution of disodium tetraborate R; — mobile phase B: 0.1 M sodium hydroxide.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	100	0
15 - 15.5	100 → 0	0 → 100
15.5 - 25.5	0	100

Flow rate 1.0 mL/min.

Detection Conductivity detector with a sensitivity of 30 µS.

Injection 50 µL.

Identification of peaks Use the chromatogram obtained with the reference solution to identify the principal peak due to the sulfate ion.

Retention time Sulfate ion = about 7.5 min. Change the composition of the mobile phase, if necessary, to obtain the prescribed retention time.

Limit:

— free sulfates: maximum 0.5 per cent.

Ph Eur



Limits: — impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent); — any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent); — total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent); — disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

B. Gas chromatography (2.2.28). Dissolve 1.0 g of the substance to be examined in *water* R and dilute to 50 mL with the same solvent. Add 2 mL of concentrated ammonia R and shake with 3 quantities, each of 10 mL, of methylene chloride R. To the combined lower layers, add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate at a temperature not exceeding 30 °C, using a rotary evaporator. Take up the residue with methylene chloride R and dilute to 20.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with methylene chloride R.

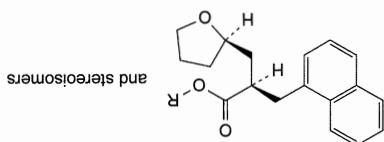
Reference solution Dissolve 5 mg of nafidrofuryl impurity F CRS in methylene chloride R and dilute to 50 mL with the same solvent.

Column: material: fused silica; size: $l = 25$ m, $\phi = 0.32$ mm; stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 0.45 μ m).

Carrier gas helium for chromatography R. Splitter flow rate 25 mL/min. Flow rate 2.9 mL/min. Temperature: 290

Temperature (°C)	Time (min)	Column
210	0 - 4	4 - 8
210 → 230	8 - 18	230 → 260
260	18 - 30	290

Detection Flame ionisation. Injection 1 μ L. **Relative retention** With reference to the second eluting peak of nafidrofuryl: impurity D = about 0.14; impurity B = about 0.55 (for the second eluting peak); impurity E = about 0.86; impurity F = about 1.04 (for the second eluting peak). **System suitability:** test solution (b): — resolution: minimum 1.0 between the 2 peaks due to the diastereoisomers of nafidrofuryl. **Limits:** test solution (a): — impurity F: for the sum of the areas of the 2 peaks, maximum 0.20 per cent of the sum of the areas of the 2 peaks due to nafidrofuryl (0.20 per cent);



A, R = H: 2-[(naphthalen-1-yl)methyl]-3-(tetrahydrofuran-2-yl)propanoic acid, B, R = C₂H₅: ethyl 2-[(naphthalen-1-yl)methyl]-3-(tetrahydrofuran-2-yl)propanoate,

IMPURITIES of C₂₆H₃₅NO₇. 1 mL of 0.1 M perchloric acid is equivalent to 47.36 mg. Dissolve 0.350 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

ASSAY Maximum 0.1 per cent, determined on 1.0 g. **Sulfated ash (2.4.14)** Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C. **Loss on drying (2.2.32)** Maximum 0.5 per cent, determined on 1.000 g by drying in (1 ppm Pb) R. reference solution using 10 mL of lead standard solution *water* R. The solution complies with test E. Prepare the filter if necessary and wash the filter. Dilute to 20 mL with until the solution is decolorised and add 0.5 mL in excess. until a pink colour is obtained. Cool, add glacial acetic acid R phenolphthalein solution R and then concentrated ammonia R hydrochloric acid R1 and *water* R. Add 0.1 mL of 2 quantities, each of 5 mL, of a mixture of equal volumes of 800 ± 50 °C for about 1 h. Take up the residue with the ignition. If necessary repeat the operation. Heat at coloured, allow to cool, mix using a fine glass rod and repeat obtained. If after 30 min of ignition the mixture remains redness until a homogeneous white or greyish-white mass is be examined with 0.5 g of magnesium oxide R1. Ignite to dull In a silica crucible, mix thoroughly 1.0 g of the substance to Maximum 10 ppm. **Heavy metals (2.4.8)** nafidrofuryl. 30 per cent of the sum of the areas of the 2 peaks due to — first eluting nafidrofuryl diastereoisomer: minimum **Limits:** test solution (b): related substances. Gas chromatography (2.2.28) as described in test B for **Diastereoisomer ratio** disregard any peaks due to impurity B. — impurity E: maximum 0.20 per cent of the sum of the areas of the 2 peaks due to nafidrofuryl (0.20 per cent); — impurity D: maximum 0.10 per cent of the sum of the areas of the 2 peaks due to nafidrofuryl (0.10 per cent); — any other impurity: for each impurity, maximum 0.10 per cent of the sum of the areas of the 2 peaks due to nafidrofuryl (0.10 per cent); — total: maximum 0.50 per cent of the sum of the areas of the 2 peaks due to nafidrofuryl (0.50 per cent); — disregard limit: 0.02 per cent of the sum of the areas of the 2 peaks due to nafidrofuryl (0.02 per cent);

methyl-4-oxo-1,4-dihydro-1,8-naphthyridine-3-carboxylic acid, calculated with reference to the dried substance.

CHARACTERS

An almost white or pale yellow, crystalline powder, practically insoluble in water, soluble in methylene chloride, slightly soluble in acetone and in alcohol. It dissolves in dilute solutions of alkali hydroxides.

It melts at about 230 °C.

IDENTIFICATION

First identification B

Second identification A, C, D

A. Dissolve 12.5 mg in 0.1 M sodium hydroxide and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with 0.1 M sodium hydroxide. Examined between 230 nm and 350 nm (2.2.25), the solution shows two absorption maxima, at 258 nm and 334 nm. The ratio of the absorbance measured at 258 nm to that measured at 334 nm is 2.2 to 2.4.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with *nalidixic acid* CRS. Examine the substances prepared as discs.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with the test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 0.1 g in 2 mL of hydrochloric acid R. Add 0.5 mL of a 100 g/L solution of β -naphthol R in alcohol R. An orange-red colour develops.

TESTS

Absorbance

Dissolve 1.50 g in methylene chloride R and dilute to 50.0 mL with the same solvent. The absorbance (2.2.25) measured at 420 nm is not greater than 0.10.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel F₂₅₄ plate R.

Test solution (a) Dissolve 0.20 g of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 20 mL with methylene chloride R.

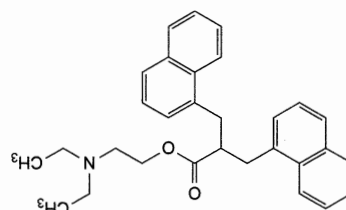
Reference solution (a) Dissolve 20 mg of *nalidixic acid* CRS in methylene chloride R and dilute to 20 mL with the same solvent.

Reference solution (b) Dilute 2 mL of test solution (b) to 10 mL with methylene chloride R.

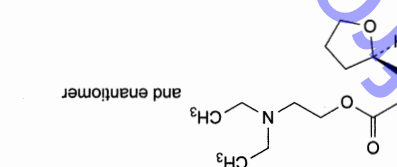
Reference solution (c) Dilute 1 mL of reference solution (b) to 10 mL with methylene chloride R.

Reference solution (d) Dilute 1 mL of reference solution (b) to 25 mL with methylene chloride R.

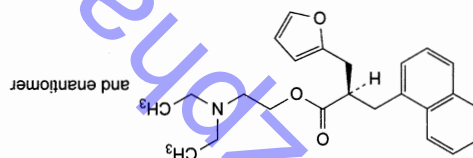
Apply to the plate 10 μ L of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of dilute ammonia R1, 20 volumes of methylene chloride R and 70 volumes of alcohol R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (c) (0.1 per cent) and not more than one such spot is more intense than the spot in the chromatogram obtained with reference solution (d).



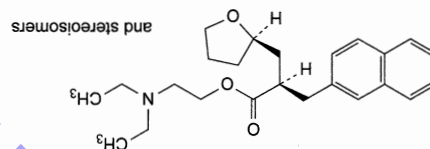
C. 2-(diethylamino)ethyl 3-(naphthalen-1-yl)-2-[(naphthalen-1-yl)methyl]propanoate,



D. 2-(diethylamino)ethyl 3-[(2R)-tetrahydrofuran-2-yl]propanoate,



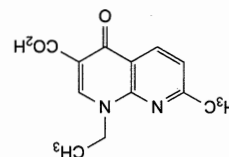
E. 2-(diethylamino)ethyl 3-[(furan-2-yl)methyl]propanoate,



F. 2-(diethylamino)ethyl 2-[(naphthalen-2-yl)methyl]-3-(tetrahydrofuran-2-yl)propanoate.

Nalidixic Acid

(Ph. Eur. monograph 0701)



C₁₂H₁₂N₂O₃

232.2

389-08-2

Action and use
Quinolone antibacterial.

Preparations

Nalidixic Acid Oral Suspension
Nalidixic Acid Tablets

DEFINITION

Nalidixic acid contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 1-ethyl-7-

Heavy metals (2.4.8)

1.0 g complies with test D for heavy metals (20 ppm).
Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 10 mL of methylene chloride R and add 30 mL of 2-propanol R and 10 mL of carbon dioxide-free water R. Keep the titration vessel covered and pass nitrogen R through the solution throughout the titration. Keep the temperature of the solution between 15 °C and 20 °C. Titrate with 0.1 M ethanolic sodium hydroxide, determining the end-point potentiometrically (2.2.20) using a silver-silver chloride comparison electrode with a sleeve diaphragm or a capillary tip, filled with a saturated solution of lithium chloride R in ethanol R, and a glass electrode as indicator electrode.

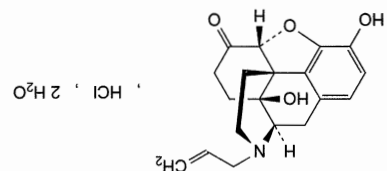
1 mL of 0.1 M ethanolic sodium hydroxide is equivalent to 23.22 mg of $C_{19}H_{22}ClNO_4 \cdot 2H_2O$.

STORAGE

Store in an airtight container, protected from light.

Naloxone Hydrochloride

(Naloxone Hydrochloride Dihydrate,
Ph Eur monograph 0729)



$C_{19}H_{22}ClNO_4 \cdot 2H_2O$ 399.9

51481-60-8

Preparation

Opioid receptor antagonist.

Naloxone Injection

DEFINITION

4,5α-Epoxy-3,14-dihydroxy-17-(prop-2-enyl)morphinan-6-one hydrochloride dihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent), practically insoluble in toluene.

IDENTIFICATION

First identification A, C
A. Infrared absorption spectrophotometry (2.2.24).
Comparison naloxone hydrochloride dihydrate CRS.
B. Thin-layer chromatography (2.2.27).
Test solution Dissolve 8 mg of the substance to be examined in 0.5 mL of water R and dilute to 1 mL with methanol R.
Reference solution Dissolve 8 mg of naloxone hydrochloride dihydrate CRS in 0.5 mL of water R and dilute to 1 mL with methanol R.
Plate TLC silica gel G plate R.
Mobile phase Mix 5 volumes of methanol R and 95 volumes of the upper layer from a mixture of 60 mL of dilute ammonia R2 and 100 mL of butanol R.
Application 5 µL.
Development Over 2/3 of the plate.
Drying In air.
Detection Spray with a freshly prepared 5 g/L solution of potassium ferricyanide R in ferric chloride solution R1; examine in daylight.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.
C. It gives reaction (a) of chlorides (2.3.1).
TESTS
Solution S
Dissolve 0.50 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.
Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).
Acidity or alkalinity
To 10.0 mL of solution S add 0.05 mL of methyl red solution R. Not more than 0.2 mL of 0.02 M sodium hydroxide or 0.02 M hydrochloric acid is required to change the colour of the indicator.
Specific optical rotation (2.2.7)
−181 to −170 (anhydrous substance), determined on solution S.
Impurity D
Liquid chromatography (2.2.29).
Solution A Dissolve 1.58 g of ammonium hydrogen carbonate R in 950 mL of water R1, adjust to pH 9.0 with concentrated ammonia R and dilute to 1000 mL with water R1.
Test solution Dissolve 0.500 g of the substance to be examined in a 10.3 g/L solution of hydrochloric acid R and dilute to 20.0 mL with the same solution.
Reference solution (a) Dissolve 10.0 mg of naloxone impurity D CRS in a 10.3 g/L solution of hydrochloric acid R and dilute to 20.0 mL with the same solution. Dilute 5.0 mL of this solution to 100.0 mL with a 10.3 g/L solution of hydrochloric acid R.
Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with a 10.3 g/L solution of hydrochloric acid R.
Reference solution (c) To 4.0 mL of the test solution add 2.0 mL of reference solution (a) and dilute to 20.0 mL with a 10.3 g/L solution of hydrochloric acid R.
Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm);
— temperature: 40 °C.

impurity D = about 1.1; impurity E = about 3.0;

impurity B = about 3.2.

Identification of impurities Use the chromatogram supplied with

naloxone for peak identification CRS and the chromatogram

obtained with reference solution (a) to identify the peaks due

to impurities A, B, C, D, E and F.

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 2.0, where H_p = height

above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to naloxone.

Limits:

— **correction factor:** for the calculation of content, multiply the

peak area of impurity E by 0.5;

— **impurities A, B, C, E, F:** for each impurity, not more than

the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— **total:** not more than 4 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.8 per cent);

— **disregard limit:** 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

Water (2.5.12)

7.5 per cent to 11.0 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 0.50 g.

ASSAY

Dissolve 0.300 g in 50 mL of ethanol (96 per cent) R and add

5.0 mL of 0.01 M hydrochloric acid. Carry out a

potentiometric titration (2.2.20), using 0.1 M ethanoic sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M ethanoic sodium hydroxide is equivalent to

36.38 mg of $C_{19}H_{22}ClNO_4$.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A, B, C, D, E, F

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

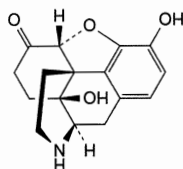
acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: G.



A. 4,5 α -epoxy-3,14-dihydroxy-3,14-dihydroxy-6-one

(noroxymorphone);

time = about 11 min; impurity C = about 0.6;

impurity A = about 0.8; impurity F = about 0.9;

Mobile phase:
— mobile phase A: acetonitrile R1, solution A (20:80 V/V);
— mobile phase B: acetonitrile R1, solution A (40:60 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 50	100	0
50 - 51	100 \rightarrow 0	0 \rightarrow 100
51 - 60	0	100

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 μ L of the test solution and reference solutions (b)

and (c).

Relative retention With reference to naloxone (retention

time = about 50 min): impurity D = about 0.8.

System suitability: reference solution (c):

— **symmetry factor:** maximum 1.8 for the peak due to

impurity D.

Limit:

— **impurity D:** not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (75 ppm).

Related substances

Liquid chromatography (2.2.29).

Solution A Dissolve 1.10 g of sodium octanesulfonate R in

950 mL of water R, adjust to pH 2.0 with a 50 per cent V/V

solution of phosphoric acid R, filter and dilute to 1000 mL

with water R.

Test solution Dissolve 0.125 g of the substance to be

examined in a 10.3 g/L solution of hydrochloric acid R and

dilute to 25.0 mL with the same solution.

Reference solution (a) Dissolve 5 mg of naloxone for peak

identification CRS (containing impurities A, B, C, D, E

and F) in 1 mL of a 10.3 g/L solution of hydrochloric acid R.

Reference solution (b) Dilute 1.0 mL of the test solution to

20.0 mL with a 10.3 g/L solution of hydrochloric acid R.

Dilute 1.0 mL of this solution to 25.0 mL with a 10.3 g/L

solution of hydrochloric acid R.

Column:

— **size:** $l = 0.125$ m, $\phi = 4.0$ mm;

— **stationary phase:** end-capped octylsilyl silica gel for

chromatography R (5 μ m);

— **temperature:** 40 °C.

Mobile phase:

— **mobile phase A:** acetonitrile R, tetrahydrofuran R, solution A

(2:4:94 V/V/V);

— **mobile phase B:** tetrahydrofuran R, acetonitrile R, solution A

(4:17:79 V/V/V);

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

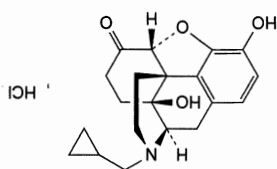
Relative retention With reference to naloxone (retention

time = about 11 min): impurity C = about 0.6;

impurity A = about 0.8; impurity F = about 0.9;

Naltrexone Hydrochloride

(Ph. Eur. monograph 1790)



$C_{20}H_{24}ClNO_4$ 377.9

Action and use

Opioid receptor antagonist.

Ph Eur

DEFINITION

17-(Cyclopropylmethyl)-4,5-epoxy-3,14-dihydromorphinan-6-one hydrochloride. It may be anhydrous, a monohydrate or a mixture or a solvate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder, very hygroscopic.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Dissolve 20 mg in water R and dilute to 5 mL with the same solvent. Make alkaline with dilute ammonia R1. Shake with 10 mL of methylene chloride R, separate the organic layer and evaporate the solvent. Dry the residue obtained *in vacuo*.

Comparison naltrexone hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.40 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ or B₆ (2.2.2, Method II).

Acidity and alkalinity

To 10 mL of solution S, add 0.05 mL of methyl red solution R. Not more than 0.2 mL of 0.02 M sodium hydroxide or 0.02 M hydrochloric acid is required to change the colour of the indicator.

Specific optical rotation (2.2.7)

–187 to –195 (anhydrous substance).

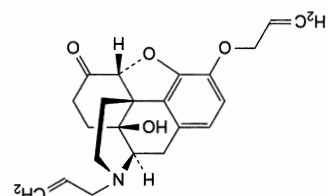
Dissolve 0.40 g in water R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

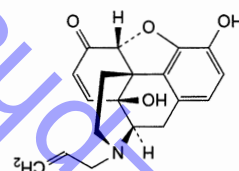
Test solution Dissolve 20.0 mg of the substance to be examined in 0.1 M hydrochloric acid and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of naltrexone impurity C CRS in 0.1 M hydrochloric acid and dilute to 2.5 mL with the same solvent.

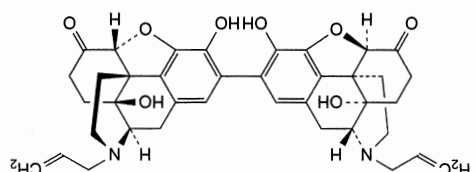


B. 4,5-epoxy-14-hydroxy-17-(prop-2-enyl)-3-(prop-2-enyloxy)morphinan-6-one (3-O-allylnaloxone),

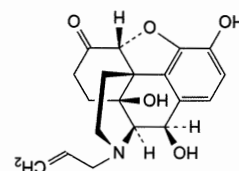
C. 4,5-epoxy-3,10-14-trihydroxy-17-(prop-2-enyl)morphinan-6-one (10- α -hydroxynaloxone),



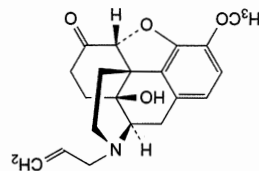
D. 7,8-didehydro-4,5-epoxy-3,14-dihydroxy-17-(prop-2-enyl)morphinan-6-one (7,8-didehydronaloxone),



E. 4,5- α ,4',5'- α -diepoxy-3,3',14,14'-tetrahydroxy-17,17'-bis(prop-2-enyl)-2,2'-bimorphinan-6,6'-dione (2,2'-binaloxone),



F. 4,5-epoxy-3,10-14-trihydroxy-17-(prop-2-enyl)morphinan-6-one (10- β -hydroxynaloxone),



G. 4,5-epoxy-14-hydroxy-3-methoxy-17-(prop-2-enyl)morphinan-6-one (3-O-methylnaloxone).

Ph Eur



Reference solution (b) Dilute 1.0 mL of the test solution and 1.0 mL of reference solution (a) to 100.0 mL with 0.1 M hydrochloric acid. Dilute 1.0 mL of this solution to 10.0 mL with 0.1 M hydrochloric acid.

Column:
— size: $l = 0.15$ m, $\phi = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R₁ (5 μ m);
— temperature: 40 °C.

Mobile phase:
— mobile phase A: 1.1 g/L solution of sodium octanesulfonate R, adjusted to pH 2.3 with phosphoric acid R;
— mobile phase B: acetonitrile R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B
0 - 45	90 \rightarrow 55	10 \rightarrow 45
45 - 47	55 \rightarrow 90	45 \rightarrow 10
47 - 55	90	10

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 230 nm.

Equilibration 8 min.

Injection 10 μ L.

Relative retention With reference to naltrexone (retention time = about 16 min): impurity A = about 0.4;

impurity B = about 0.7; impurity F = about 0.8;

impurity G = about 0.9; impurity C = about 1.05;

impurity H = about 1.1; impurity I = about 1.2;

impurity J = about 1.3; impurity D = about 1.4;

impurity E = 1.7.

System suitability: reference solution (b):

— **resolution:** minimum 2.0 between the peaks due to naltrexone and impurity C.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity D by 0.4;

— **impurities C, D, E, F, G:** for each impurity, not more than twice the area of the peak due to naltrexone in the chromatogram obtained with reference solution (b)

(0.2 per cent);

— **impurities A, B, H, I, J:** for each impurity, not more than the area of the peak due to naltrexone in the chromatogram obtained with reference solution (b)

(0.1 per cent);

— **any other impurity:** for each impurity, not more than the area of the peak due to naltrexone in the chromatogram obtained with reference solution (b)

obtained with reference solution (b) (0.1 per cent);

— **total:** not more than 10 times the area of the peak due to naltrexone in the chromatogram obtained with reference solution (b) (1.0 per cent);

— **disregard limit:** 0.5 times the area of the peak due to naltrexone in the chromatogram obtained with reference solution (b) (0.05 per cent).

Ethanol (2.4.24, System A)

Maximum 3.0 per cent.

Test solution Dissolve 0.25 g of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.

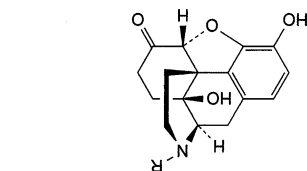
Reference solution Dilute 0.750 g of anhydrous ethanol R to 1000.0 mL with water R.

Water (2.5.12)

Maximum 10.0 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.



A. R = CHO: 17-formyl-4,5 α -epoxy-3,14-dihydroxymorphinan-6-one,

B. R = H: 4,5 α -epoxy-3,14-dihydroxymorphinan-6-one (noroxy-morphone),

C. R = CH₂-CH=CH-CH₂: 17-but-3-enyl-4,5 α -epoxy-3,14-dihydroxymorphinan-6-one,

H. R = CH₂-CH₂-CH₂-CH₃: 17-butyl-4,5 α -epoxy-3,14-dihydroxymorphinan-6-one,

D. 17,17'-bis(cyclopropylmethoxy)-4,5 α :4',5' α -dipoxy-3,3',14,14'-tetrahydroxy-2,2'-bimorphinan-6,6'-dione (pseudonaltrexone),

E. 3-(cyclopropylmethoxy)-17-(cyclopropylmethyl)-4,5 α -epoxy-14-hydroxymorphinan-6-one,

F. R = H, R' = OH: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,10 α ,14-trihydroxymorphinan-6-one,

G. R = OH, R' = H: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,10 β ,14-trihydroxymorphinan-6-one,

I. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

J. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

K. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

L. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

M. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

N. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

O. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

P. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

Q. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

R. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

S. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

T. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

U. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

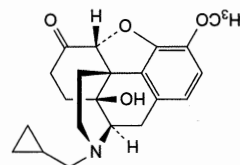
V. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

W. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

X. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

Y. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

Z. R + R' = O: 17-(cyclopropylmethyl)-4,5 α -epoxy-3,14-dihydroxymorphinan-6,10-dione,

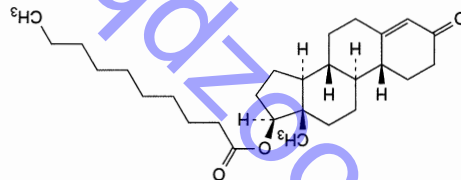


J. 17-(cyclopropylmethyl)-4,5α-epoxy-14-hydroxy-3-methoxymorphinan-6-one.

Ph Eur

Nandrolone Decanoate

(Ph. Eur. monograph 1992)



Action and use
Anabolic steroid; androgen.

Ph Eur

DEFINITION

3-Oxoestr-4-en-17β-yl decanoate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, very soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Melting point (2.2.14): 34 °C to 38 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison nandrolone decanoate CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 0.20 g in 10 mL of methanol R.

Specific optical rotation (2.2.7)

Dissolve 0.200 g in anhydrous ethanol R and dilute to + 35.0 to + 40.0 (dried substance).

20.0 mL with the same solvent.

Impurities A, B, C

Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined in methylene chloride R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with methylene chloride R.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with methylene chloride R.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 20.0 mL with methylene chloride R.

Reference solution (d) Dissolve 5 mg of nandrolone decanoate for system suitability CRS (containing impurities A, B, C) in 0.5 mL of methylene chloride R.

Plate TLC silica gel plate R.

Application 10 µL of the test solution and reference solutions (b), (c) and (d).

Development Over 2/3 of the plate.

Drying In air.

Detection Treat with alcoholic solution of sulfuric acid R and heat at 130 °C until the spots appear. Examine in ultraviolet light at 366 nm.

Retention factors Nandrolone decanoate = about 0.37; impurity A = about 0.45; impurity B = about 0.55; impurity C = about 0.58.

System suitability Reference solution (d):
the chromatogram shows 4 clearly separated spots.

Limits:

impurity A: any spot due to impurity A is not more intense than the principal spot in the chromatogram

obtained with reference solution (b) (1.0 per cent);

impurities B, C: any spot due to impurity B or C is not more intense than the principal spot in the chromatogram

obtained with reference solution (c) (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (b) Dissolve 5 mg of nandrolone decanoate for peak identification CRS (containing impurities D, F, G, H, I, K, L) in methanol R and dilute to 2.0 mL with the same solvent.

Column:

size: l = 0.15 m, Ø = 3.9 mm,

stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase:

mobile phase A: water R,

mobile phase B: acetonitrile R,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	35	65
5 - 40	35 → 0	65 → 100
40 - 75	0	100
75 - 80	0 → 35	100 → 65

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Relative retention With reference to nandrolone decanoate (retention time = about 30 min): impurity D = about 0.05;

impurity F = about 0.6; impurity K = about 0.7;

impurity L = about 0.9; impurity G = about 0.97;

impurity H = about 1.1; impurity I = about 1.2.

— **System suitability:** reference solution (b):
— **peak-to-valley ratio:** minimum 1.5, where H_p = height above the baseline of the peak due to impurity G and H_v = height above the baseline of the peak from the peak due to nandrolone decanoate.
Limits:

— **correction factors:** for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity D = 0.5; impurity F = 0.6; impurity H = 1.1; impurity I = 1.3; impurity K = 0.8;
— **impurities D, F, G, H, I, K, L:** for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— **total:** not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);
— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying over *diphosphorus pentoxide* R_5 at a pressure not exceeding 0.7 kPa for 4 h at room temperature.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 10.0 mg in *anhydrous ethanol R* and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 50.0 mL with *anhydrous ethanol R*. Measure the absorbance (2.2.25) at the absorption maximum at 240 nm. Calculate the content of $C_{28}H_{44}O_3$ taking the specific absorbance to be 407.

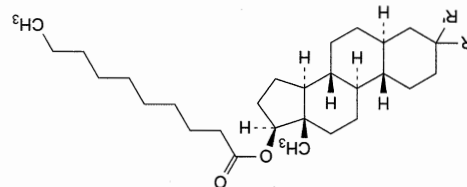
STORAGE

Under nitrogen, protected from light and at a temperature of 2 °C to 8 °C.

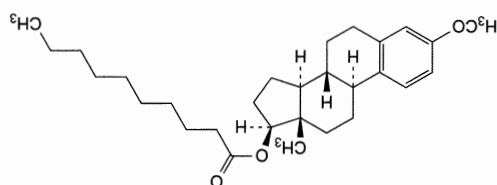
IMPURITIES

Specified impurities A, B, C, D, F, G, H, I, K, L

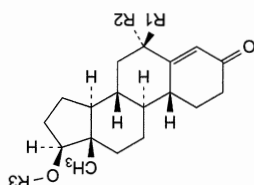
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. **Control of impurities in substances for pharmaceutical use:** E, J, J.



A. R + R' = O: 3-oxo-5 α -estrane-17 β -yl decanoate,
C. R = R' = OCH₃: 3,3-dimethoxy-5 α -estrane-17 β -yl decanoate,



B. 3-methoxyestra-1,3,5(10)-trien-17 β -yl decanoate,



D. R1 = R2 = R3 = H: 17 β -hydroxyestra-4-en-3-one,
E. R1 = H, R2 = OH, R3 = CO-[CH₂]₈-CH₃: 6 α -hydroxy-3-oxoestra-4-en-17 β -yl decanoate,

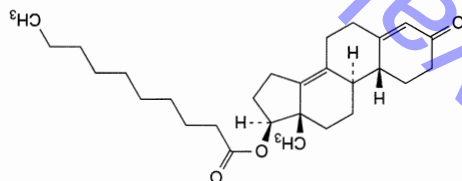
F. R1 + R2 = O, R3 = CO-[CH₂]₈-CH₃: 3,6-dioxoestra-4-en-17 β -yl decanoate,

H. R1 = R2 = H, R3 = CO-[CH₂]₉-CH₃: 3-oxoestra-4-en-17 β -yl undecanoate,

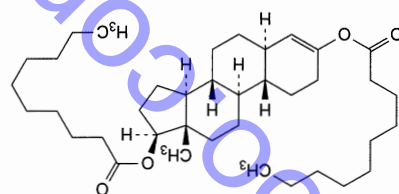
I. R1 = R2 = H, R3 = CO-[CH₂]₁₀-CH₃: 3-oxoestra-4-en-17 β -yl dodecanoate,

K. R1 = R2 = H, R3 = CO-[CH₂]₆-CH₃: 3-oxoestra-4-en-17 β -yl octanoate,

L. R1 = R2 = H, R3 = CO-[CH₂]₇-CH₃: 3-oxoestra-4-en-17 β -yl nonanoate,



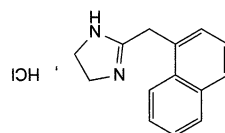
G. 3-oxoestra-4,8(14)-dien-17 β -yl decanoate,



J. 5 α -estr-3-ene-3,17 β -diyl didecanoate.

Naphazoline Hydrochloride

(Ph. Eur. monograph 0730)



$C_{14}H_{15}ClN_2$

246.7

550-99-2

Action and use
Alpha-adrenoceptor agonist.

DEFINITION
2-(Naphthalen-1-ylmethyl)-4,5-dihydro-1H-imidazole hydrochloride.

IDENTIFICATION

Appearance
White or almost white, crystalline powder.
Solubility
Freely soluble in water, soluble in ethanol (96 per cent). mp: about 259 °C, with decomposition.

CHARACTERS

Content
99.0 per cent to 101.0 per cent (dried substance).

with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (c) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.
Column:
— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;
— stationary phase: end-capped base-deactivated octylsilyl silica gel for chromatography R (4 μ m) with a pore size of 6 nm.
Mobile phase Dissolve 1.1 g of sodium octanesulfonate R in a mixture of 5 mL of glacial acetic acid R, 300 mL of acetamintrile R and 700 mL of water R.
Flow rate 1 mL/min.
Detection Spectrophotometer at 280 nm.
Injection 20 μ L.
Run time 3 times the retention time of naphazoline.

Retention time Naphazoline = about 14 min.
System suitability: reference solution (a):
— resolution: minimum 5.0 between the peaks due to naphazoline and impurity B.
Limits:
— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— unspecified impurities: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);
— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.
1 mL of 0.1 M sodium hydroxide is equivalent to 24.67 mg of $C_{14}H_{15}ClN_2$.

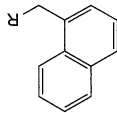
STORAGE

Protected from light.

IMPURITIES

Specified impurities: A.

Other detectable impurities: B, C, D.



A. R = CO-NH-[CH₂]₂-NH₂; N-(2-aminoethyl)-2-(naphthalen-1-yl)acetamide (naphthylacetylenediamine),
B. R = CO₂H; (naphthalen-1-yl)acetic acid (1-naphthylacetic acid),
C. R = CN; (naphthalen-1-yl)acetonitrile,
(1-naphthylacetoneitrile),

Related substances
Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 5 mg of 1-naphthylacetic acid R in the mobile phase, add 5 mL of the test solution and dilute to 100 mL with the mobile phase.
Reference solution (b) Dissolve 5.0 mg of naphazoline impurity A CRS in the mobile phase and dilute to 100.0 mL.

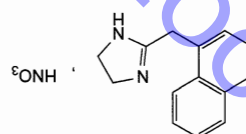
Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).
Acidity or alkalinity
To 20 mL of solution S add 0.2 mL of 0.01 M sodium hydroxide. Not more than 0.6 mL of 0.01 M hydrochloric acid is required to change the colour of the solution to red.

TESTS
C. It gives reaction (a) of chlorides (2.3.1).
Dissolve 0.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.
Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity
To 20 mL of solution S add 0.2 mL of 0.01 M sodium hydroxide. Not more than 0.6 mL of 0.01 M hydrochloric acid is required to change the colour of the solution to red.
Related substances
Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 5 mg of 1-naphthylacetic acid R in the mobile phase, add 5 mL of the test solution and dilute to 100 mL with the mobile phase.
Reference solution (b) Dissolve 5.0 mg of naphazoline impurity A CRS in the mobile phase and dilute to 100.0 mL.

Naphazoline Nitrate

(Ph Eur monograph 0147)



5144-52-5

$C_{14}H_{15}N_3O_3$

Action and use
Alpha-adrenoceptor agonist.

DEFINITION

2-(Naphthalen-1-ylmethyl)-4,5-dihydro-1H-imidazole nitrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparsingly soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification C

Second identification A, B, D

A. Melting point (2.2.14): 167 °C to 170 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in 0.01 M hydrochloric acid and

dilute to 250.0 mL with the same acid. Dilute 25.0 mL of

the solution to 100.0 mL with 0.01 M hydrochloric acid.

Spectral range 230-350 nm.

Absorption maximum At 270 nm, 280 nm, 287 nm and

291 nm.

Absorbance ratio:

— $A_{270}/A_{280} = 0.82$ to 0.86,

— $A_{291}/A_{280} = 0.65$ to 0.69.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison naphazoline nitrate CRS.

D. Dissolve 45 mg of the substance to be examined in 2 mL

of water R. Add 1 mL of sulfuric acid R. Shake carefully and

allow to cool. Add 1 mL of ferrous sulfate solution R2 dropwise

along the walls of the container. At the junction of the

2 liquids, a brown colour develops.

TESTS

Solution S

Dissolve 0.5 g in carbon dioxide-free water R, warming gently,

and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

5.0 to 6.5 for solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be

examined in the mobile phase and dilute to 100.0 mL with

the mobile phase.

Reference solution (a) Dissolve 5 mg of 1-naphthylacetic acid R

in the mobile phase, add 5 mL of the test solution and dilute

to 100 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of naphazoline

impurity A CRS in the mobile phase and dilute to 100.0 mL

with the same solvent. Dilute 5.0 mL of this solution to

100.0 mL with the mobile phase.

Reference solution (c) Dilute 2.0 mL of the test solution to

10.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm,

— stationary phase: end-capped base-deactivated octylsilyl silica

gel for chromatography R (4 μ m) with a pore size of 6 nm.

Mobile phase Dissolve 1.1 g of sodium octanesulfonate R in a

mixture of 5 mL of glacial acetic acid R, 300 mL of

acetonitrile R and 700 mL of water R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Run time 3 times the retention time of naphazoline.

Relative retention With reference to naphazoline (retention

time = about 14 min): impurity A = about 0.76;

impurity D = about 1.24; impurity B = about 1.27;

impurity C = about 2.8.

System suitability: reference solution (a):

— resolution: minimum 5.0 between the peaks due to

limits:

— impurity A: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.5 per cent),

— unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (c)

(0.10 per cent),

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (c)

(1.0 per cent),

— disregard limit: 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (c)

Chlorides (2.4.4)

Maximum 330 ppm, determined on solution S.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 30 mL of anhydrous acetic acid R. Titrate

with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 27.33 mg

of $C_{14}H_{15}N_3O_3$.

IDENTIFICATION

First identification A, D
Second identification A, B, C

A. Specific optical rotation (2.2.7): + 59 to + 62 (dried

substance).
Dissolve 0.50 g in ethanol (96 per cent) R and dilute to

25.0 mL with the same solvent.

B. Melting point (2.2.14): 154 °C to 158 °C.

C. Ultraviolet and visible absorption spectrophotometry

(2.2.25).
Test solution Dissolve 40.0 mg in methanol R and dilute to

100.0 mL with the same solvent. Dilute 10.0 mL of the

solution to 100.0 mL with methanol R.

Spectral range 230–350 nm.

Absorption maxima At 262 nm, 271 nm, 316 nm and

331 nm.

Specific absorbances at the absorption maxima:

— at 262 nm: 216 to 238;

— at 271 nm: 219 to 241;

— at 316 nm: 61 to 69;

— at 331 nm: 79 to 87.

D. Infrared absorption spectrophotometry (2.2.24).

Comparison naproxen CRS.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured

than reference solution BY₇ (2.2.2, Method II).

Dissolve 1.25 g in methanol R and dilute to 25 mL with the

same solvent.

Enantiomeric purity

Liquid chromatography (2.2.29). Protect the solutions from

light.

Test solution Dissolve 25.0 mg of the substance to be

examined in tetrahydrofuran R and dilute to 50.0 mL with the

same solvent. Dilute 2.0 mL of the solution to 20.0 mL with

the mobile phase.

Reference solution (a) Dilute 2.5 mL of the test solution to

100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of racemic naproxen CRS

in 10.0 mL of tetrahydrofuran R and dilute to 100.0 mL with

the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel π -acceptor/ π -donor for chiral

separations R (5 μ m) (S,S);

— temperature: 25 °C.

Mobile phase glacial acetic acid R, acetonitrile R, 2-propanol R,

hexane R (0.5:5:10:84.5 V/V/V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 263 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of naproxen (retention

time = about 5 min).

System suitability: reference solution (b):

— resolution: minimum 3 between the peaks due to

impurity G and naproxen.

Limit:

— impurity G: not more than the area of the principal peak

in the chromatogram obtained with reference solution (a)

(2.5 per cent).

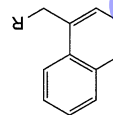
STORAGE

Protected from light.

IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.



A, R = CO-NH-[CH₂]₂-NH₂; N-(2-aminoethyl)-2-

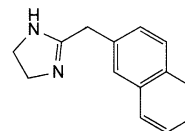
(naphthalen-1-yl)acetamide (naphthylacetylenediamine),

B, R = CO₂H; (naphthalen-1-yl)acetic acid (1-naphthylacetic

acid),

C, R = CN; (naphthalen-1-yl)acetonitrile

(1-naphthylacetonitrile),

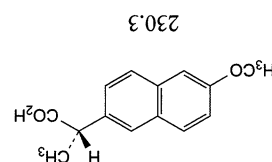


D, 2-(naphthalen-2-ylmethyl)-4,5-dihydro-1H-imidazole

(β -naphazoline).

Naproxen

(Ph. Eur. Monograph 0731)



22204-53-1

Gastro-resistant Naproxen Tablets

Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

Gastro-resistant Naproxen Tablets

Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

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Action and use

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Action and use

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Action and use

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Action and use

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Naproxen Tablets

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Naproxen Oral Suspension

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Action and use

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Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

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Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

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Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

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Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

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Naproxen Suppositories

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Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

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Naproxen Tablets

Naproxen Suppositories

Naproxen Oral Suspension

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

C₁₄H₁₄O₃

Gastro-resistant Naproxen Tablets

Naproxen Tablets

Naproxen Suppositories

Related substances

Liquid chromatography (2.2.29). Protect the solutions from light.

Test solution Dissolve 12 mg of the substance to be examined in the mobile phase and dilute to 20 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 6 mg of bromomethoxynaphthalene R (impurity N), 6.0 mg of naproxen impurity L CRS, 6 mg of 6-methoxy-2-naphthoic acid R (impurity O) and 6 mg of (1R,3S)-1-(6-methoxynaphthalen-2-yl)ethanol R (impurity K) in acetonitrile R and dilute to 10.0 mL with the same solvent. To 1.0 mL of the solution add 1.0 mL of the test solution and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.10$ m, $\phi = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m);

— temperature: 50 °C.

Mobile phase Mix 42 volumes of acetonitrile R and 58 volumes of a 1.36 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 2.0 with phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of impurity N.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to

impurities K, L, N and O.

Relative retention With reference to naproxen (retention time = about 2.5 min): impurity O = about 0.8;

impurity K = about 0.9; impurity L = about 1.4;

impurity N = about 5.3.

System suitability: reference solution (b):

— resolution: minimum 2.2 between the peaks due to impurity K and naproxen.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity O by 2.0;

— impurity O: not more than 1.5 times the area of the principal peak in the chromatogram obtained with

reference solution (a) (0.15 per cent);

— impurity L: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32) Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14) Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 25 mL of water R and 75 mL of methanol R. Titrate with 0.1 M sodium hydroxide, using 1 mL of phenolphthalein solution R as indicator.

1 mL of 0.1 M sodium hydroxide is equivalent to 23.03 mg of $C_{14}H_{14}O_3$.

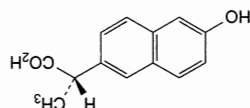
STORAGE

Protected from light.

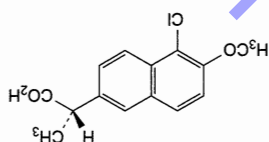
IMPURITIES

Specified impurities G, L, O

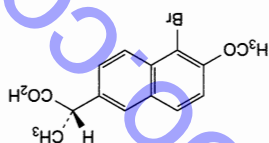
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, H, I, J, K, M, N.



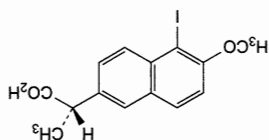
A. (2S)-2-(6-hydroxynaphthalen-2-yl)propanoic acid,



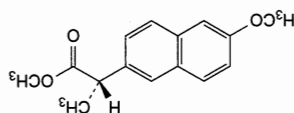
B. (2S)-2-(5-chloro-6-methoxynaphthalen-2-yl)propanoic acid,



C. (2S)-2-(5-bromo-6-methoxynaphthalen-2-yl)propanoic acid,



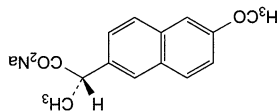
D. (2S)-2-(5-iodo-6-methoxynaphthalen-2-yl)propanoic acid,



E. methyl (2S)-2-(6-methoxynaphthalen-2-yl)propanoate,

Naproxen Sodium

(Ph. Eur. monograph 1702)

 $C_{14}H_{13}O_3Na$

252.2

26159-34-2

Ph Eur

DEFINITION

Sodium (2S)-2-(6-methoxynaphthalen-2-yl)propanoate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Freely soluble in water, freely soluble or soluble in methanol, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, C, D

Second identification A, B, D

A. Specific optical rotation (2.2.7): -17.0 to -14.7 (dried substance).Dissolve 0.50 g in a 4.2 g/L solution of sodium hydroxide R and dilute to 25.0 mL with the same solution.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 40.0 mg in methanol R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with methanol R.Spectral range 230 – 350 nm.Absorption maxima At 262 nm, 271 nm, 316 nm and 331 nm.

Specific absorbance at the absorption maxima:

— at 262 nm: 207 to 227 ;— at 271 nm: 200 to 220 ;— at 316 nm: 56 to 68 ;— at 331 nm: 72 to 84 .

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 50 mg in 5 mL of water R. Add 1 mL of dilute sulfuric acid R and 5 mL of ethyl acetate R. Shake vigorously. Allow the 2 layers to separate. Evaporate the upper layer to dryness and subsequently dry at 60°C for 15 min. Record the spectrum using the residue.

Comparison naproxen CRS.

D. It gives reaction (a) of sodium (2.3.1).

TESTS

Appearance of solution

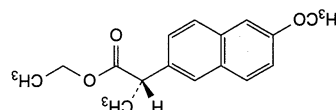
The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).Dissolve 1.25 g in water R and dilute to 25 mL with the same solvent.

pH (2.2.3)

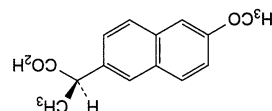
7.0 to 9.8.

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

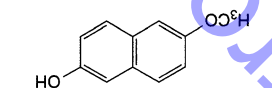
F. ethyl (2S)-2-(6-methoxynaphthalen-2-yl)propanoate,



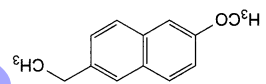
G. (2R)-2-(6-methoxynaphthalen-2-yl)propanoic acid ((R)-enantiomer),



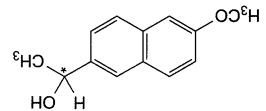
H. 6-methoxynaphthalen-2-ol,



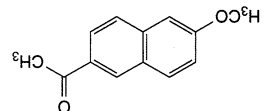
I. (6-methoxynaphthalen-2-yl)acetic acid,



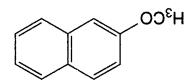
J. 2-ethyl-6-methoxynaphthalene,



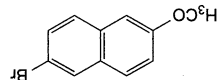
K. (1RS)-1-(6-methoxynaphthalen-2-yl)ethanol,



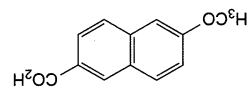
L. 1-(6-methoxynaphthalen-2-yl)ethanone,



M. 2-methoxynaphthalene (nerolin),



N. 2-bromo-6-methoxynaphthalene,



O. 6-methoxynaphthalene-2-carboxylic acid (6-methoxy-2-naphthoic acid).

Ph Eur

Enantiomeric purity

Liquid chromatography (2.2.29). Protect the solutions from

light.

Test solution Dissolve 25.0 mg of the substance to be

examined in 15 mL of water R and add 1 mL of hydrochloric acid R. Shake with 2 quantities, each of 10 mL, of ethyl

acetate R, combine the upper layers and evaporate to dryness

under reduced pressure. Dissolve the residue in 50.0 mL of

terahydrofuran R. Dilute 2.0 mL of this solution to 20.0 mL

with the mobile phase.

Reference solution (a) Dilute 2.5 mL of the test solution to

100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of racemic naproxen CRS

in 10 mL of terahydrofuran R and dilute to 100 mL with the

mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel π -acceptor/ π -donor for chiral

separations R (5 μ m) (S,S);

— temperature: 25 °C.

Mobile phase glacial acetic acid R, acetonitrile R, 2-propanol R,

hexane R (5:50:100:845 V/V/V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 263 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of naproxen (retention

time = about 5 min).

System suitability: reference solution (b):

— resolution: minimum 3 between the peaks due to

impurity G and naproxen.

Limit:

— impurity G: not more than the area of the principal peak

in the chromatogram obtained with reference solution (a)

(2.5 per cent).

Related substances

Liquid chromatography (2.2.29). Protect the solutions from

light.

Test solution Dissolve 12 mg of the substance to be examined

in the mobile phase and dilute to 20 mL with the mobile

phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

50.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 6 mg of

bromomethoxynaphthalene R (impurity N), 6.0 mg of naproxen

impurity L CRS and 6 mg of (1RS)-1-(6-methoxynaphthalen-

2-yl) ethanol R (impurity K) in acetonitrile R and dilute to

10 mL with the same solvent. To 1 mL of the solution add

1 mL of the test solution and dilute to 50 mL with the

mobile phase. Dilute 1 mL of this solution to 20 mL with

the mobile phase.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(3 μ m);

— temperature: 50 °C.

Mobile phase Mix 42 volumes of acetonitrile R and 58 volumes

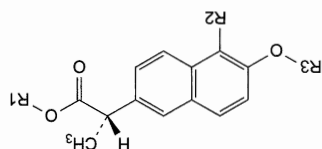
of a 1.36 g/L solution of potassium dihydrogen phosphate R

previously adjusted to pH 2.0 with phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.



A. R1 = R2 = R3 = H: (2S)-2-(6-hydroxynaphthalen-2-yl)propanoic acid,
B. R1 = H, R2 = C1, R3 = CH3: (2S)-2-(5-chloro-6-methoxynaphthalen-2-yl)propanoic acid,

C, D, E, F, H, I, J, K, M, N.
Control of impurities in substances for pharmaceutical use: A, B, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these by the general monograph Substances for pharmaceutical use acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities G, L

IMPURITIES

In an airtight container, protected from light.

STORAGE

of C₁₄H₁₃O₃Na.

1 mL of 0.1 M perchloric acid is equivalent to 25.22 mg

potentiometrically (2.2.20).

with 0.1 M perchloric acid, determining the end-point

Dissolve 0.200 g in 50 mL of anhydrous acetic acid R. Titrate

ASSAY

an oven at 105 °C for 3 h.

Maximum 1.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

result.

described in the test, filtering the solutions to evaluate the

precipitates. Dilute each solution to 40 mL with anhydrous

ethanol R; the substance dissolves completely. Proceed as

After the addition of buffer solution pH 3.5 R, the substance

2 mL of lead standard solution (10 ppm Pb) R.

complies with test A. Prepare the reference solution using

Dissolve 2.0 g in 20.0 mL of water R. 12 mL of the solution

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

— the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

(0.3 per cent);

in the chromatogram obtained with reference solution (a)

— total: not more than 3 times the area of the principal peak

with reference solution (a) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

solution (b) (0.1 per cent);

peak in the chromatogram obtained with reference

— impurity L: not more than the area of the corresponding

Limits:

impurity K and naproxen.

— resolution: minimum 2.2 between the peaks due to

System suitability: reference solution (b):

impurity L = about 1.4; impurity N = about 5.3.

time = about 2.5 min; impurity K = about 0.9;

Relative retention With reference to naproxen (retention

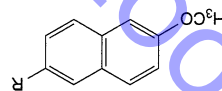
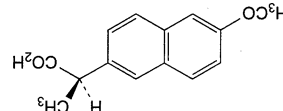
Run time 1.5 times the retention time of impurity N.

A. Specific optical rotation (2.2.7): -40.0 to -36.5 (dried substance).
 Dissolve 0.200 g in a 4 g/L solution of sodium hydroxide R and dilute to 20.0 mL with the same solution.
 B. Infrared absorption spectrophotometry (2.2.24).
 Comparison nateglinide CRS.
 If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.
 C. Test B for related substances (see Tests).

TESTS

Related substances
 A. Impurity A and unspecified impurities. Liquid chromatography (2.2.29).
 Test solution Dissolve 60.0 mg of the substance to be examined in 1 mL of acetonitrile R1 and dilute to 10.0 mL with the mobile phase.
 Reference solution (a) Dissolve 3.0 mg of nateglinide impurity A CRS in 1 mL of acetonitrile R1 and dilute to 25.0 mL with the mobile phase.
 Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.
 Reference solution (c) Dissolve 3 mg of the substance to be examined in 1 mL of acetonitrile R1, add 4.0 mL of reference solution (a) and dilute to 10 mL with the mobile phase.
 Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
 Column:
 — size: $l = 0.05$ m, $\varnothing = 3.9$ mm;
 — stationary phase: spherical end-capped octylsilyl silica gel for chromatography R (5 μ m);
 — temperature: 40°C .
 Mobile phase Mix 35 volumes of acetonitrile R1 and 65 volumes of a 7.8 g/L solution of sodium dihydrogen phosphate monohydrate R previously adjusted to pH 2.5 with phosphoric acid R.
 Flow rate 2.0 mL/min.
 Detection Spectrophotometer at 210 nm.
 Injection 100 μ L of the test solution and reference solutions (b), (c) and (d).
 Run time 5 times the retention time of nateglinide.
 Relative retention With reference to nateglinide (retention time = about 7 min): impurity A = about 0.5.
 System suitability: reference solution (c):
 — resolution: minimum 5.0 between the peaks due to impurity A and nateglinide.
 Limits:
 — impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
 — unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (0.10 per cent);
 — sum of unspecified impurities: not more than twice the area of the principal peak in the chromatogram
 — disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.05 per cent).
 B. Impurity B. Liquid chromatography (2.2.29).

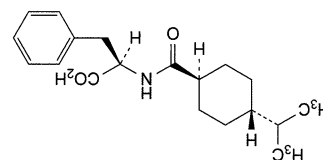
C. R1 = H, R2 = Br, R3 = CH₃: (2S)-2-(5-bromo-6-methoxynaphthalen-2-yl)propanoic acid,
 D. R1 = H, R2 = I, R3 = CH₃: (2S)-2-(5-iodo-6-methoxynaphthalen-2-yl)propanoic acid,
 E. R1 = R3 = CH₃, R2 = H: methyl (2S)-2-(6-methoxynaphthalen-2-yl)propanoate,
 F. R1 = C₂H₅, R2 = H, R3 = CH₃: ethyl (2S)-2-(6-methoxynaphthalen-2-yl)propanoate,
 G. (2R)-2-(6-methoxynaphthalen-2-yl)propanoic acid,



H. R = OH: 6-methoxynaphthalen-2-ol,
 I. R = CH₂-CO₂H: (6-methoxynaphthalen-2-yl)acetic acid,
 J. R = C₂H₅: 2-ethyl-6-methoxynaphthalene,
 K. R = CHOH-CH₃: (1R,S)-1-(6-methoxynaphthalen-2-yl)ethanol,
 L. R = CO-CH₃: 1-(6-methoxynaphthalen-2-yl)ethanone,
 M. R = H: 2-methoxynaphthalene (nerolin),
 N. R = Br: 2-bromo-6-methoxynaphthalene.

Nateglinide

(Ph. Eur. monograph 2575)



C₁₉H₂₇NO₃ 317.4 105816-04-4

Action and use
 Stimulates insulin release; treatment of diabetes mellitus.

Ph Eur

DEFINITION

N-[(trans-4-(1-Methylethyl)cyclohexyl)carbonyl]-D-phenylalanine.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methanol and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

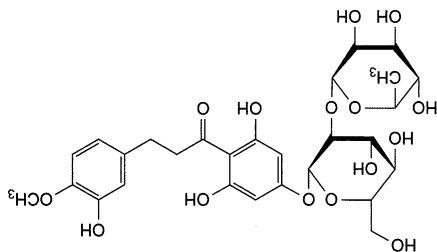
Carry out either tests A, B or tests B, C.

- Test solution** Dissolve 0.200 g of the substance to be examined in methanol R2 and dilute to 20.0 mL with the same solvent.
- Reference solution (a)** Dissolve 5 mg of nateglinide impurity B CRS in methanol R2 and dilute to 10.0 mL with the same solvent.
- Reference solution (b)** Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
- Reference solution (c)** Dissolve 0.10 g of the substance to be examined in methanol R2. Add 1.0 mL of reference solution (a) and dilute to 10.0 mL with methanol R2.
- Column:**
- size: $l = 0.25$ m, $\phi = 4.0$ mm;
 - stationary phase: urea type silica gel for chiral chromatography R (5 μ m);
 - temperature: 40 °C.
- Mobile phase** Dissolve 0.17 g of ammonium acetate R in methanol R2 and dilute to 1000 mL with the same solvent.
- Flow rate** 0.8 mL/min.
- Detection** Spectrophotometer at 220 nm.
- Injection** 10 μ L of the test solution and reference solutions (b) and (c).
- Run time** 1.5 times the retention time of nateglinide.
- Relative retention** With reference to nateglinide (retention time = about 21 min): impurity B = about 0.9.
- System suitability:** reference solution (c):
- peak-to-valley ratio: minimum 3, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to nateglinide.
- Limit:**
- impurity B: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent).
 - impurity C: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
 - impurity D: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.2 per cent).
- Limits:**
- total for impurities A, B, C, D and sum of unspecified impurities: maximum 0.5 per cent;
 - disregard limit for impurities A, B, C and D: 0.05 per cent for each impurity.
- Heavy metals** (2.4.8)
- Maximum 10 ppm.
- Solvent** methanol R.
- 0.25 g complies with test H. Prepare the reference solution using 0.25 mL of lead standard solution (10 ppm Pb) R.
- Loss on drying** (2.2.32)
- Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.
- Sulfated ash** (2.4.14)
- Maximum 0.1 per cent, determined on 1.0 g.
- ASSAY**
- Liquid chromatography** (2.2.29) as described in test C for related substances with the following modification.
- Injection** Test solution and reference solution (e).
- Calculate the percentage content of $C_{16}H_{27}NO_3$, taking into account the assigned content of nateglinide CRS.
- IMPURITIES**
- Specified impurities** A, B, C, D.
- Other detectable impurities** (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, F, G.



Neohesperidin-Dihydrochalcone

(Ph. Eur. monograph 1547)



20702-77-6 613

C₂₈H₃₆O₁₅

Ph Eur

DEFINITION

1-[4-[[2-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-2,6-dihydroxyphenyl]-3-(3-hydroxy-4-methoxyphenyl)propan-1-one.

Content

96.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish-white powder.

Solubility

Practically insoluble in water, freely soluble in dimethyl sulfoxide, soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison neohesperidin-dihydrochalcone CRS.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₄ (2.2.2, Method II).

Dissolve 0.25 g in methanol R and dilute to 25 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 0.10 g of the substance to be examined in dimethyl sulfoxide R and dilute to 50.0 mL with the same solvent.

Test solution (b) Dilute 10.0 mL of test solution (a) to 20.0 mL with dimethyl sulfoxide R.

Reference solution (a) Dissolve 50.0 mg of neohesperidin-dihydrochalcone CRS in dimethyl sulfoxide R and dilute to 50.0 mL with the same solvent.

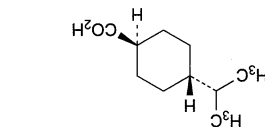
Reference solution (b) Dissolve 4.0 mg of neohesperidin-dihydrochalcone impurity B CRS in dimethyl sulfoxide R and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with dimethyl sulfoxide R.

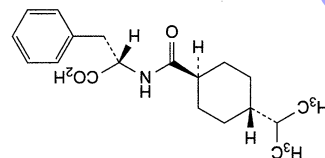
Reference solution (d) In order to prepare *in situ* impurity F and impurity G, suspend 0.10 g of the substance to be examined in 10.0 mL of a 100 g/L solution of sulfuric acid R.

II-354 Neohesperidin-Dihydrochalcone

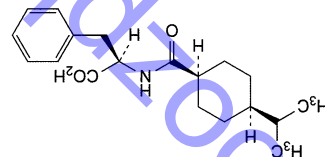
A. *trans*-4-(1-methylethyl)cyclohexanecarboxylic acid,



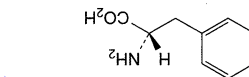
B. *N*-[[*trans*-4-(1-methylethyl)cyclohexyl]carbonyl]-L-phenylalanine (1-phenylalanine isomer),



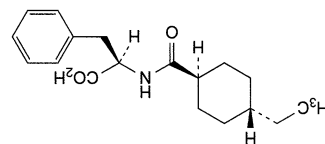
C. *N*-[[*cis*-4-(1-methylethyl)cyclohexyl]carbonyl]-D-phenylalanine (*cis*-isomer),



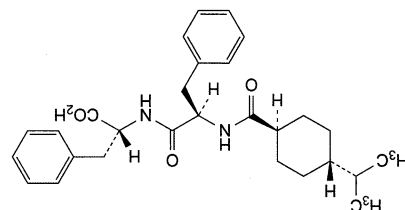
D. (2*S*)-2-amino-3-phenylpropanoic acid (phenylalanine),



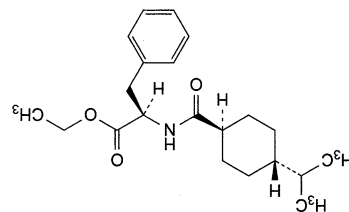
E. *N*-[[*trans*-4-ethylcyclohexyl]carbonyl]-D-phenylalanine,



F. *N*-[[*trans*-4-(1-methylethyl)cyclohexyl]carbonyl]-D-phenylalanine,



G. ethyl *N*-[[*trans*-4-(1-methylethyl)cyclohexyl]carbonyl]-D-phenylalaninate.



Ph Eur

Heat the sample for 5 min on a water-bath. Dilute immediately 1.0 mL of the resulting solution to 50.0 mL with dimethyl sulfoxide R.

Column:

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (4 µm) with a carbon loading of

— size: $l = 0.15$ m, $\phi = 3.9$ mm,

7 per cent,

— temperature: 30 °C.

Mobile phase Mix 20 volumes of acetonitrile R and 80 volumes of a solution prepared by adding 5.0 mL of glacial acetic acid R to 1000.0 mL of water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 282 nm.

Injection 10 µL; inject test solution (a) and reference solutions (a), (b), (c) and (d).

Run time 5 times the retention time of neohesperidin-dihydrochalcone which is about 10 min.

Relative retention With reference to neohesperidin-dihydrochalcone

With reference to neohesperidin-dihydrochalcone: impurity B = about 0.4;

impurity D = about 0.7; impurity F = about 1.2;

impurity G = about 3.7.

System suitability:

— resolution: minimum of 2.5 between the first peak (neohesperidin-dihydrochalcone) and the second peak (impurity F) in the chromatogram obtained with reference solution (d),

chromatogram obtained with reference solution (a) is similar to the chromatogram provided with neohesperidin-dihydrochalcone CRS.

Limits:

— impurity B: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent),

— impurity D: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (2 per cent),

— any other impurity: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent),

total of all impurities apart from impurity B: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (2.5 per cent),

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 12.0 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

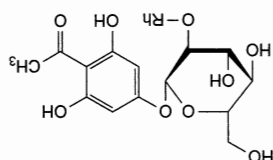
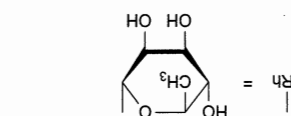
Liquid chromatography (2.2.29) as described in the test for related substances.

Injection 10 µL; inject test solution (b) and reference solutions (a) and (d).

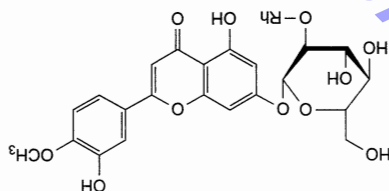
System suitability:

— resolution: minimum of 2.5 between the first peak (neohesperidin-dihydrochalcone) and the second peak (impurity F) in the chromatogram obtained with reference solution (d),

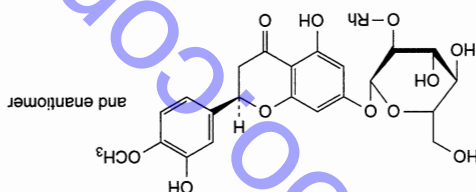
chromatogram obtained with reference solution (a) is similar to the chromatogram provided with neohesperidin-dihydrochalcone CRS.



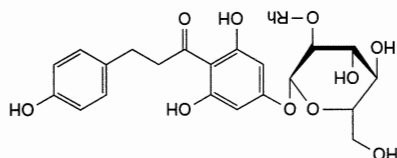
A. 1-[4-[[2-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-2,6-dihydroxyphenyl]ethanone (phloracetophenone neohesperidoside),



B. 7-[[2-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-4H-1-benzopyran-4-one (neodiosmin),



C. (2R,5R)-7-[[2-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-2,3-dihydro-4H-1-benzopyran-4-one (neohesperidin),



D. 1-[4-[[2-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-2,6-dihydroxyphenyl]propan-1-one (naringin-dihydrochalcone),

IMPURITIES

Protected from light.

STORAGE

substance to be examined.

Calculate the percentage content of $C_{28}H_{36}O_{15}$ using the chromatogram obtained with reference solution (a) and the stated content of $C_{28}H_{36}O_{15}$ in neohesperidin-dihydrochalcone CRS, correcting for the water content of the

— repeatability: reference solution (a).

reference solution (d),

(impurity F) in the chromatogram obtained with

Solubility

Very soluble in water, very slightly soluble in alcohol, practically insoluble in acetone.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for related substances.

Results:

- the retention time of the principal peak in the chromatogram obtained with the test solution is approximately the same as that of the principal peak in the chromatogram obtained with reference solution (e),
 - it complies with the limits given for impurity C.
- B. It gives reaction (a) of sulfates (2.3.1).

TESTS

pH (2.2.3)

Dissolve 0.1 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

Dissolve 1.00 g in water R and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of framycetin sulfate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (d) Dissolve the contents of a vial of neomine CRS (corresponding to 0.5 mg) in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (e) Dissolve 10 mg of neomycin sulfate CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5 μ m),

temperature: 25 °C.

Mobile phase Mix 20.0 mL of trifluoroacetic acid R, 6.0 mL of carbonate-free sodium hydroxide solution R and 500 mL of water R, allow to equilibrate, dilute to 1000 mL with water R and degas.

Flow rate 0.7 mL/min.

Post-column solution carbonate-free sodium hydroxide solution R diluted 1 in 25 previously degassed, which is added pulse-less to the column effluent using a 375 μ L polymeric mixing coil.

Flow rate 0.5 mL/min.

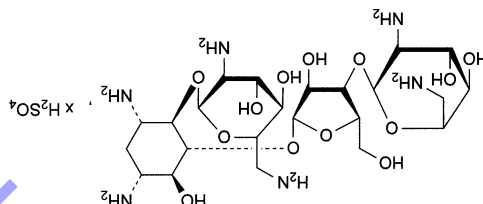
Detection Pulsed amperometric detector with a gold indicator electrode, a silver-silver chloride reference electrode and a stainless steel auxiliary electrode which is the cell body, held at respectively 0.00 V detection, + 0.80 V oxidation and -0.60 V reduction potentials, with pulse durations according to the instrument used.

Injection 10 μ L; inject the test solution and the reference solutions (b), (c), (d) and (e).

Neomycin Sulfate

Neomycin Sulfate

(Ph. Eur. monograph 0197)



$C_{23}H_{46}N_6O_{13} \cdot H_2SO_4$ 615 (base)

Action and use

Aminoglycoside antibacterial.

Preparations

Dexamethasone and Neomycin Ear Spray

Hydrocortisone and Neomycin Cream

Hydrocortisone Acetate and Neomycin Ear Drops

Hydrocortisone Acetate and Neomycin Eye Drops

Neomycin Eye Drops

Hydrocortisone Acetate and Neomycin Eye Ointment

Neomycin Eye Ointment

Neomycin Tablets

Ph. Eur.

DEFINITION

Mixture of sulfates of substances produced by the growth of certain selected strains of *Streptomyces fradiae*, the main component being the sulfate of 2-deoxy-4-O-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-5-O-[3-O-(2,6-diamino-2,6-dideoxy-β-D-ribofuranosyl)-β-D-ribofuranosyl]-D-streptamine (neomycin B).

CHARACTERS

Minimum of 680 IU/mg (dried substance).

Content

White or yellowish-white powder, hygroscopic.

Appearance

Run time 1.5 times the retention time of neomycin B.

Relative retention With reference to neomycin B (retention time = about 10 min): impurity A = about 0.65; impurity C = about 0.9; impurity G = about 1.1.

System suitability:

— **resolution:** minimum of 2.0 between the peaks due to impurity C and to neomycin B in the chromatogram obtained with reference solution (e); if necessary, adjust the volume of the carbonate-free sodium hydroxide solution in the mobile phase,

— **signal-to-noise ratio:** minimum 10 for the principal peak in the chromatogram obtained with reference solution (c).

Limits:

— **impurity A:** not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (2.0 per cent),

— **impurity C:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (15.0 per cent) and not less than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent),

— **any other impurity:** not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (5.0 per cent),

— **total of other impurities:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (15.0 per cent),

— **disregard limit:** area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent).

Sulfate

27.0 per cent to 31.0 per cent (dried substance).

Dissolve 0.250 g in 100 mL of water R and adjust the solution to pH 11 using concentrated ammonia R. Add 10.0 mL of 0.1 M barium chloride and about 0.5 mg of phthalate purple R. Titrate with 0.1 M sodium edetate until the colour of the solution begins to change, continuing the titration until the violet-blue colour disappears.

1 mL of 0.1 M barium chloride is equivalent to 9.606 mg of SO_4 .

Loss on drying (2.2.32)

Maximum 8.0 per cent, determined on 1.000 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 0.7 kPa for 3 h.

Sulfated ash (2.4.14)

Maximum 1.0 per cent, determined on 1.0 g.

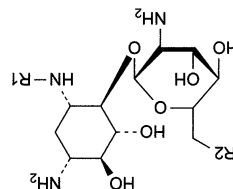
ASSAY

Carry out the microbiological assay of antibiotics (2.7.2).

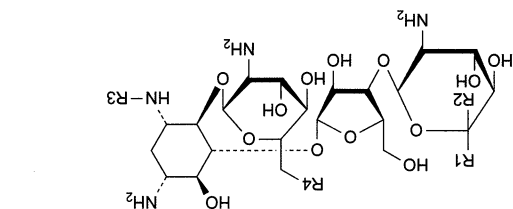
STORAGE

In an airtight container, protected from light.

IMPURITIES



A. R1 = H, R2 = NH_2 : 2-deoxy-4-O-(2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl)-D-streptamine (neamine or neomycin A-LP),



C. R1 = $\text{CH}_2\text{-NH}_2$, R2 = R3 = H, R4 = NH_2 : 2-deoxy-4-O-(2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl)-5-O-[3-O-(2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl)]- β -D-(2,6-diamino-2,6-dideoxy- α -D-ribofuranosyl)-D-streptamine (neomycin C),

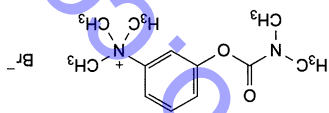
E. R1 = R3 = H, R2 = $\text{CH}_2\text{-NH}_2$, R4 = OH: 4-O-(2-amino-2-deoxy- α -D-glucopyranosyl)-2-deoxy-5-O-[3-O-(2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl)]- β -D-(2,6-diamino-2,6-dideoxy- α -D-ribofuranosyl)-D-streptamine (paromomycin I or neomycin E),

F. R1 = $\text{CH}_2\text{-NH}_2$, R2 = R3 = H, R4 = OH: 4-O-(2-amino-2-deoxy- α -D-glucopyranosyl)-2-deoxy-5-O-[3-O-(2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl)]- β -D-(2,6-diamino-2,6-dideoxy- α -D-ribofuranosyl)-D-streptamine (paromomycin II or neomycin F),

G. R1 = H, R2 = $\text{CH}_2\text{-NH}_2$, R3 = CO-CH_3 , R4 = NH_2 : 3-N-acetyl-2-deoxy-4-O-(2,6-diamino-2,6-dideoxy- α -D-glucopyranosyl)-5-O-[3-O-(2,6-diamino-2,6-dideoxy- α -D-ribofuranosyl)]-D-streptamine (neomycin B-LP),

Neostigmine Bromide

(Ph. Eur. monograph 0046)



$\text{C}_{12}\text{H}_{19}\text{BrN}_2\text{O}_2$ 303.2 114-80-7

Action and use
Cholinesterase inhibitor.

Preparation
Neostigmine Tablets

DEFINITION

3-[(Dimethylcarbamoyloxy)-N,N,N-trimethylanilinium bromide.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals, hygroscopic.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 20 mg in 0.5 M sulfuric acid and dilute

to 100 mL with the same acid.

Spectral range 230–350 nm.

Absorption maxima At 260 nm and 266 nm.

Specific absorbance at the absorption maxima:

— at 260 nm: about 16;

— at 266 nm: about 14.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison neostigmine bromide CRS.

C. To 50 mg add 0.4 g of potassium hydroxide R and 2 mL of

ethanol (96 per cent) R and heat on a water-bath for 3 min,

replacing the evaporated ethanol (96 per cent). Cool and add

2 mL of water R and 2 mL of diazobenzenesulfonic acid

solution R1. An orange-red colour develops.

D. It gives the reactions of bromides (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in distilled water R and dilute to 50 mL with

the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined

in the mobile phase and dilute to 50.0 mL with the mobile

phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 4 mg of 3-

dimethylaminophenol R (impurity B) in 50.0 mL of the mobile

phase. Dilute 1.0 mL of the solution to 200.0 mL with the

mobile phase.

Reference solution (c) Dissolve the contents of a vial of

solution (b).

Reference solution (d) Mix 1.0 mL of the mobile phase and

1.0 mL of reference solution (a).

Column:

— size: l = 0.25 m, Ø = 4.0 mm;

— stationary phase: base-deactivated end-capped octylsilyl silica

gel for chromatography R (5 µm);

— temperature: 30 °C.

Mobile phase To 710 mL of a 3.6 g/L solution of sodium

dihydrogen phosphate R previously adjusted to pH 3.2 with

290 mL of acetonitrile R1.

Flow rate 1.6 mL/min.

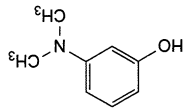
Detection Spectrophotometer at 220 nm.

Injection 50 µL Of the test solution and reference

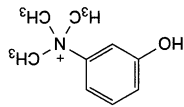
solutions (a), (c) and (d).

Run time Twice the retention time of neostigmine.

B. 3-(dimethylamino)phenol,



A. 3-hydroxy-N,N,N-trimethylanilinium,



Specified impurities B
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C.

IMPURITIES

In an airtight container, protected from light.

STORAGE

of C₁₂H₁₉BrN₂O₂.

1 mL of 0.1 M perchloric acid is equivalent to 30.32 mg

(2.2.20).

perchloric acid, determining the end-point potentiometrically

Add 50 mL of acetic anhydride R. Titrate with 0.1 M

Dissolve 0.225 g in 2 mL of anhydrous formic acid R.

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 1.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

Maximum 200 ppm, determined on solution S.

Sulfates (2.4.13)

to impurity B.

— reporting threshold: 0.05 per cent; disregard the peak due

— total: maximum 0.3 per cent;

0.10 per cent;

— unspecified impurities: for each impurity, maximum

— impurity B: maximum 0.01 per cent;

Limits:

by 0.5.

— correction factor: multiply the peak area of impurity B

reference solution (a);

— for each impurity, use the concentration of neostigmine in

Calculation of percentage contents:

the chromatogram obtained with reference solution (d).

— signal-to-noise ratio: minimum 25 for the principal peak in

reference solution (c);

impurities B and A in the chromatogram obtained with

— resolution: minimum 1.5 between the peaks due to

System suitability:

impurity A = about 0.61.

time = about 20 min): impurity B = about 0.56;

Relative retention With reference to neostigmine (retention

impurities A and B.

Identification of impurities Use the chromatogram obtained

with reference solution (c) to identify the peaks due to

TESTS

Solution S
Dissolve 2.5 g in distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 4.0 mL of solution S add 6.0 mL of water R and 0.1 mL of phenolphthalein solution R1. The solution is colourless.

Add 0.3 mL of 0.01 M sodium hydroxide; the solution becomes red. Add 0.4 mL of 0.01 M hydrochloric acid; the solution becomes colourless. Add 0.1 mL of methyl red solution R; the solution becomes red or yellowish-red.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 4 mg of 3-dimethylaminophenol R (impurity B) in 50.0 mL of the mobile phase. Dilute 1.0 mL of the solution to 200.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of neostigmine impurity A CRS in 1.0 mL of reference solution (b).

Reference solution (d) Mix 1.0 mL of the mobile phase and 1.0 mL of reference solution (a).

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: base-deactivated end-capped octylsilyl silica gel for chromatography R (5 μ m);

— temperature: 30 °C.

Mobile phase To 710 mL of a 3.6 g/L solution of sodium dihydrogen phosphate R previously adjusted to pH 3.2 with phosphoric acid R, add 4.3 g of sodium dodecyl sulfate R and 290 mL of acetonitrile R1.

Flow rate 1.6 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 μ L Of the test solution and reference solutions (a), (c) and (d).

Run time Twice the retention time of neostigmine.

Identification of impurities Use The chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to neostigmine (retention time = about 20 min): impurity B = about 0.56;

impurity A = about 0.61.

System suitability:

— resolution: minimum 1.5 between the peaks due to impurities B and A in the chromatogram obtained with reference solution (c);

— signal-to-noise ratio: minimum 25 for the principal peak in the chromatogram obtained with reference solution (d).

Calculation of percentage contents:

— for each impurity, use the concentration of neostigmine in reference solution (a);

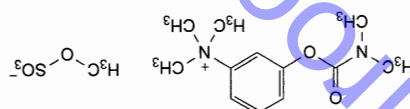
— correction factor: multiply the peak area of impurity B by 0.5.

Limits:

— impurity B: maximum 0.01 per cent;

Neostigmine Metilsulfate

(Ph. Eur. monograph 0626)



$C_{13}H_{22}N_2O_6S$ 334.4

51-60-5

Action and use

Cholinesterase inhibitor.

Preparation

Neostigmine Injection

Ph. Eur.

DEFINITION

3-[(dimethylcarbamoyloxy)-N,N,N-trimethylammonium

methyl sulfate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless

crystals, hygroscopic.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, C.

Second identification A, B, D, E.

A. Melting point (2.2.14): 144 °C to 149 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50 mg in 0.5 M sulfuric acid and dilute to 100 mL with the same acid.

Spectral range 230-350 nm.

Absorption maxima At 261 nm and 267 nm.

Resolution (2.2.25): minimum 1.9 for the absorbance ratio.

Absorbance ratio $A_{267} / A_{261} = 0.84$ to 0.87.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison neostigmine metilsulfate CRS.

D. To 50 mg add 0.4 g of potassium hydroxide R and 2 mL of ethanol (96 per cent) R and heat on a water-bath for 3 min, replacing the evaporated ethanol (96 per cent). Cool and add 2 mL of water R and 2 mL of diazobenzenesulfonic acid solution R1. An orange-red colour develops.

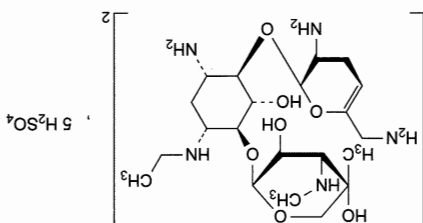
E. Dissolve 0.1 g in 5 mL of distilled water R and add 1 mL of barium chloride solution R1. No precipitate is formed.

Add 2 mL of hydrochloric acid R and heat in a water-bath for 10 min. A fine, white precipitate is formed.



Netilmicin Sulfate

Netilmicin Sulphate
(Ph. Eur. monograph 1351)



C₄₂H₉₂N₁₀O₃₄S₅ 1442 56391-57-2

Action and use

Aminoglycoside antibacterial.

DEFINITION

2-Deoxy-6-O-[3-deoxy-4-C-methyl-3-(methylamino)-β-L-arabinopyranosyl]-4-O-(2,6-diamino-2,3,4,6-tetra-deoxy-α-D-glycero-hex-4-enopyranosyl)-1-N-ethyl-D-streptamine sulfate. Substance obtained by synthesis from sisomicin. Semi-synthetic product derived from a fermentation product.

Content

Minimum 650 IU/mg (dried substance).

CHARACTERS

Appearance

White or yellowish-white powder, very hygroscopic.

Solubility

Very soluble in water, practically insoluble in acetone and in alcohol.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for related substances.
Results The retention time and size of the principal peak in the chromatogram obtained with test solution (a) are approximately the same as those of the principal peak in the chromatogram obtained with reference solution (a).
B. It gives reaction (a) of sulfates (2.3.1).

TESTS

Solution S

Dissolve 0.80 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and its absorbance at 400 nm (2.2.25) has a maximum of 0.08.

pH (2.2.3)

3.5 to 5.5 for solution S.

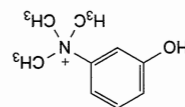
Specific optical rotation (2.2.7)

+ 88.0 to + 96.0 (dried substance).

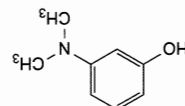
Dissolve 0.50 g in water R and dilute to 10.0 mL with the same solvent.

Related substances

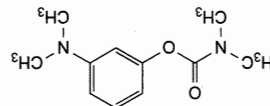
Liquid chromatography (2.2.29).
Test solution (a) Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.



A. 3-hydroxy-N,N,N-trimethylanilinium,



B. 3-(dimethylamino)phenol,



C. 3-(dimethylamino)phenyl dimethylcarbamate.

Control of impurities in substances for pharmaceutical use): A, C, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these by the general monograph Substances for pharmaceutical use acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of Other detectable impurities (the following substances would, if Specified impurities B

IMPURITIES

In an airtight container, protected from light.

STORAGE

C₁₃H₂₂N₂O₆S. 1 mL of 0.1 M hydrochloric acid is equivalent to 33.44 mg of as indicator. Carry out a blank test. hydrochloric acid, using 0.25 mL of methyl red mixed solution R the total volume in the collecting vessel is about 250 mL. Titrate the solution in the collecting vessel with 0.1 M distillate in 40 mL of a 40 g/L solution of boric acid R until dilute sodium hydroxide solution R. Distil, collecting the Dissolve 0.300 g in 150 mL of water R and add 100 mL of

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C. Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

Maximum 200 ppm, determined on solution S.

Sulfates (2.4.13)

to impurity B. — reporting threshold: 0.05 per cent; disregard the peak due — total: maximum 0.2 per cent; 0.10 per cent; — unspecified impurities: for each impurity, maximum

— *total of other impurities*: not more than twice the area of the third peak in the chromatogram obtained with reference solution (d) (2 per cent),
 — *disregard limit*: any peak with an area less than that of the principal peak in the chromatogram obtained with test solution (b) (0.1 per cent).

Sulfate
 31.5 per cent to 35.0 per cent (dried substance).
 Dissolve 0.12 g in 100 mL of water R and adjust the solution to pH 11 using concentrated ammonia R. Add 30.0 mL of 0.1 M barium chloride and about 0.5 mg of phthalate purple R. Titrate with 0.1 M sodium edetate adding 50 mL of alcohol R when the colour of the solution begins to change and continue the titration until the violet-blue colour disappears. 1 mL of 0.1 M barium chloride is equivalent to 9.606 mg of SO_4 .
Loss on drying (2.2.32)
 Maximum 15.0 per cent, determined on 0.500 g by drying at 110 °C under high vacuum for 3 h.

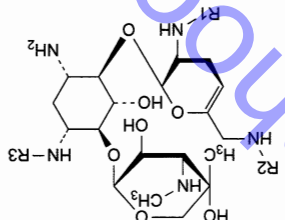
Sulfated ash (2.4.14)
 Maximum 1.0 per cent, determined on 0.5 g.

Bacterial endotoxins (2.6.14)
 Less than 1.25 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY
 Carry out the microbiological assay of antibiotics (2.7.2), using the diffusion method.

STORAGE
 In an airtight container, protected from light. If the substance is sterile, store in a sterile, tamper-proof container.

IMPURITIES



A. R1 = R2 = R3 = H: 2-deoxy-4-O-[3-deoxy-4-C-methyl-3-(methoxymethyl)-6-O-(2,6-diamino-3-(methoxymethyl)-β-L-arabinopyranosyl]-6-O-(2,6-diamino-2,3,4,6-tetrahydroxy-α-D-glycero-hex-4-enopyranosyl)-L-streptamine (sisomicin),
 C. R1 = R3 = C₂H₅, R2 = H: 4-O-[6-amino-2,3,4,6-tetrahydroxy-2-(ethylamino)-α-D-glycero-hex-4-enopyranosyl]-β-L-deoxy-6-O-[3-deoxy-4-C-methyl-3-(methoxymethyl)-β-L-arabinopyranosyl]-1-N-ethyl-D-streptamine (2'-N-arabinopyranosyl]-1-N-ethyl-D-streptamine (6'-N-ethylnetilmicin),
 D. R1 = H, R2 = R3 = C₂H₅: 4-O-[2-amino-2,3,4,6-tetrahydroxy-6-(ethylamino)-α-D-glycero-hex-4-enopyranosyl]-2-deoxy-6-O-[3-deoxy-4-C-methyl-3-(methoxymethyl)-β-L-arabinopyranosyl]-1-N-ethyl-D-streptamine (6'-N-ethylnetilmicin),

Test solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (a) Dissolve 25.0 mg of netilmicin sulfate CRS in the mobile phase and dilute to 25.0 mL with the mobile phase.
Reference solution (b) Dissolve 25.0 mg of sisomicin sulfate CRS in the mobile phase and dilute to 25.0 mL with the mobile phase.
Reference solution (c) Dissolve 20.5 mg of 1-N-ethylgarosamine sulfate CRS in the mobile phase and dilute to 25.0 mL with the mobile phase.
Reference solution (d) Dilute 1.0 mL of reference solution (a), 1.0 mL of reference solution (b) and 1.0 mL of reference solution (c) to 100.0 mL with the mobile phase.

Column:
 — size: $l = 0.25$ m, $\phi = 4.6$ mm,
 — stationary phase: styrene-divinylbenzene copolymer R (8 μm) with a pore size of 100 nm,
 — temperature: 50 °C.

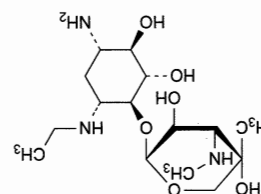
Mobile phase Prepare a solution in carbon dioxide-free water R containing 35 g/L of anhydrous sodium sulfate R, 0.5 g/L of sodium octanesulfonate R, 10 mL/L of tetrahydrofuran R, 50 mL/L of 0.2 M potassium dihydrogen phosphate R previously adjusted to pH 3.0 with a 22.5 g/L solution of phosphoric acid R and degassed.

Flow rate 1.0 mL/min.
Post-column solution 20 g/L carbonate-free solution of sodium hydroxide R previously degassed, which is added pulse-less to the column effluent using a 375 μL polymeric mixing coil.

Detection Pulsed amperometric detector with a gold indicator electrode, a silver-silver chloride reference electrode and a stainless steel auxiliary electrode which is the cell body, held at respectively + 0.05 V detection, + 0.75 V oxidation and -0.15 V reduction potentials, with pulse durations according to the instrument used.
Injection 20 μL; inject test solutions (a) and (b) and reference solution (d).
Run time 3 times the retention time of netilmicin.

Retention time Netilmicin = about 12 min.
System suitability:
 — *resolution*: minimum of 2.0 between the peaks due to impurity B (first peak) and to impurity A (second peak); minimum of 3.0 between the peaks due to impurity A (second peak) and to netilmicin (third peak) in the chromatogram obtained with reference solution (d).
 If necessary, adjust the concentration of sodium octanesulfonate in the mobile phase.
 — *signal-to-noise ratio*: minimum of 10 for the principal peak in the chromatogram obtained with test solution (b).

Limits:
 — *impurity A*: not more than the area of the second peak in the chromatogram obtained with reference solution (d) and taking into account the declared content of sisomicin sulfate CRS (1 per cent),
 — *impurity B*: not more than the area of the first peak in the chromatogram obtained with reference solution (d) and taking into account the declared content of 1-N-ethylgarosamine sulfate CRS (1 per cent),
 — *any other impurity*: not more than the area of the third peak in the chromatogram obtained with reference solution (d) (1 per cent).



B. 2-deoxy-6-O-[3-deoxy-4-C-methyl-3-(methylamino)-β-L-arabinopyranosyl]-1-N-ethyl-D-streptamine (1-N-ethyligitaramine).

Ph Eur

Anhydrous Nevirapine

(Ph. Eur. monograph 2255)



Action and use
Non-nucleoside reverse transcriptase inhibitor; antiviral (HIV).

Ph Eur

DEFINITION

1-*Cyclopropyl-4-methyl-5,11-dihydro-6H-dipyrdo* [3,2-*b*:2',3'-*e*] [1,4]diazepin-6-one.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, sparingly soluble or slightly soluble in methylene chloride, slightly soluble in methanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison anhydrous nevirapine CRS.

B. Loss on drying (see Tests).

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 24.0 mg of the substance to be examined in a mixture of 4 mL of acetonitrile R and 80 mL of the mobile phase and sonicate until dissolution is complete.

Dilute to 100.0 mL with the mobile phase.

Test solution (b) Dilute 3.0 mL of test solution (a) to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Add 2.0 mL of the mobile phase to a vial of nevirapine for peak identification CRS (containing impurities A, B and C), mix and sonicate for 1 min.

Specified impurities: A, B, C.

IMPURITIES

declared content of anhydrous nevirapine CRS.

Calculate the percentage content of $C_{15}H_{14}N_4O$ from the declared content of anhydrous nevirapine CRS.

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

using 1 mL of lead standard solution (10 ppm Pb) R.

0.50 g complies with test G. Prepare the reference solution

Maximum 20 ppm.

Heavy metals (2.4.8)

Maximum 20 ppm.

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

(0.6 per cent),

in the chromatogram obtained with reference solution (a)

— total: not more than 6 times the area of the principal peak

with reference solution (a) (0.1 per cent),

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

obtained with reference solution (a) (0.2 per cent),

the area of the principal peak in the chromatogram

— impurities A, B, C: for each impurity, not more than twice

Limits:

impurity B and nevirapine.

— resolution: minimum 5 between the peaks due to

System suitability: reference solution (b):

impurity C = 2.8.

time = about 8 min; impurity B = 0.7; impurity A = 1.5;

Relative retention With reference to nevirapine (retention

identify the peaks due to impurities A, B and C.

chromatogram obtained with reference solution (b) to

with nevirapine for peak identification CRS and the

Identification of impurities Use the chromatogram supplied

Run time 10 times the retention time of nevirapine.

and (b).

Injection 50 µL of test solution (a) and reference solutions (a)

Detection Spectrophotometer at 220 nm.

Flow rate 1.0 mL/min.

solution R.

Mobile phase Mix 20 volumes of acetonitrile R and 80 volumes of 2.88 g/L solution of ammonium dihydrogen phosphate R, previously adjusted to pH 5.0 using dilute sodium hydroxide

— temperature: 35 °C.

— chromatography R (5 µm),

— stationary phase: hexadecylammonysilyl silica gel for

— size: $l = 0.15$ m, $\phi = 4.6$ mm,

Column:

phase.

Dilute 3.0 mL of this solution to 25.0 mL with the mobile

dissolution. Dilute to 100.0 mL with the mobile phase.

80 mL of the mobile phase and sonicate until complete

nevirapine CRS in a mixture of 4 mL of acetonitrile R and

Reference solution (c) Dissolve 24.0 mg of anhydrous

Reference solution (c) Dissolve 20.0 mg of anhydrous nevirapine CRS in methanol R and sonicate until dissolution is complete. Dilute to 50.0 mL with methanol R.

Column:

— size: $l = 50$ mm, $\varnothing = 2.1$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (1.8 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: dissolve 0.77 g of ammonium acetate R in 900 mL of water R, adjust to pH 5.6 with acetic acid R

and dilute to 1000 mL with water R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 1.35	90	10
1.35 - 3.85	90 \rightarrow 67	10 \rightarrow 33
3.85 - 6.70	67 \rightarrow 60	33 \rightarrow 40
6.70 - 7.65	60	40

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 282 nm.

Injection 2.0 μ L of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied

with nevirapine for peak identification CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B and C.

Relative retention With reference to nevirapine (retention

time = about 3 min): impurity B = about 0.9;

impurity A = about 1.2; impurity C = about 1.3.

System suitability:

— resolution: minimum 5.0 between the peaks due to

impurity B and nevirapine and minimum 5.0 between the

peaks due to nevirapine and impurity A in the

chromatogram obtained with reference solution (b);

— symmetry factor: maximum 1.7 for the peak due to

nevirapine in the chromatogram obtained with reference

solution (a).

Calculation of percentage contents:

— for each impurity, use the concentration of nevirapine in

Limits:

— impurities A, B, C: for each impurity, maximum

0.2 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.6 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

0.50 g complies with test G. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

3.1 per cent to 3.9 per cent, determined on 0.300 g.

Sulfated ash (2.4.14)

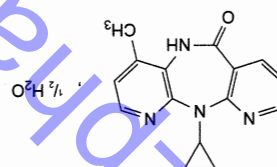
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Nevirapine Hemihydrate

(Ph Eur monograph 2479)



$C_{15}H_{14}N_4O_{1/2} \cdot H_2O$

275.3

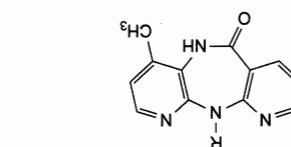
(HIV).

Action and use

Non-nucleoside reverse transcriptase inhibitor; antiviral



Ph Eur



A. R = C_2H_5 ; 11-ethyl-4-methyl-5,1,1-dihydro-6H-

dipyrrodo[3,2-b:2',3'-e][1,4]diazepin-6-one,

B. R = H; 4-methyl-5,1,1-dihydro-6H-dipyrrodo[3,2-b:2',3'-

e][1,4]diazepin-6-one,

C. R = $CH_2CH_2CH_3$; 4-methyl-11-propyl-5,1,1-dihydro-

6H-dipyrrodo[3,2-b:2',3'-e][1,4]diazepin-6-one.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be

complete. Dilute to 50.0 mL with methanol R.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with methanol R. Dilute 1.0 mL of this solution to

10.0 mL with methanol R.

Reference solution (b) Add 1.0 mL of methanol R to a vial of

nevirapine for peak identification CRS (containing

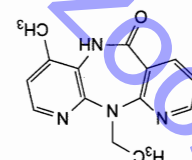
impurities A, B and C), mix and sonicate for 1 min.

Injection 2.0 mL of the test solution and reference solution (c). Calculate the percentage content of $C_{15}H_{14}N_4O$ taking into account the assigned content of anhydrous nicergoline CRS.

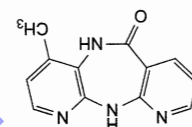
IMPURITIES

Specified impurities A, B, C

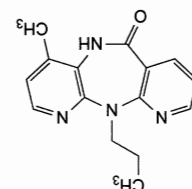
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D.



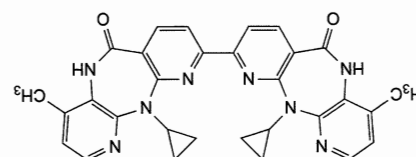
A. 11-ethyl-4-methyl-5,11-dihydro-6H-dipyrido[3,2-b:2',3'-e][1,4]diazepin-6-one,



B. 4-methyl-5,11-dihydro-6H-dipyrido[3,2-b:2',3'-e][1,4]diazepin-6-one,



C. 4-methyl-1-propyl-5,11-dihydro-6H-dipyrido[3,2-b:2',3'-e][1,4]diazepin-6-one,

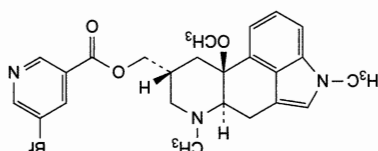


D. 11,11'-dicyclopropyl-4,4'-dimethyl-5,5',11,11'-tetrahydro-6H,6'H-9,9'-bipyrido[3,2-b:2',3'-e][1,4]diazepine-6,6'-dione.

Ph Eur

Nicergoline

(Ph Eur monograph 1998)



$C_{24}H_{26}BrN_3O_3$ 484.4 27848-84-6

Action and use

Ergot derivative.

Ph Eur

DEFINITION

[(6aR,9R,10aS)-10a-Methoxy-4,7-dimethyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinolin-9-yl]methyl 5-bromopyridine-3-carboxylate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Fine to granular, white or yellowish powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

First identification: A, B, C.

Second identification: A, B, D

A. Specific optical rotation (2.2.7): + 4.8 to + 5.8

(anhydrous substance).

Dissolve 0.50 g in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with ethanol (96 per cent) R.

Spectral range 220-350 nm.

Absorption maximum At 288 nm.

Absorption minimum At 251 nm.

Specific absorbance at the absorption maximum 175 to 185

(anhydrous substance).

C. Infrared absorption spectrophotometry (2.2.24).

Comparison nicergoline CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in ethanol (96 per cent) R, evaporate to dryness and record new spectra using the residues.

D. Dissolve 2 mg in 2 mL of sulfuric acid R. A blue colour develops.

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Dissolve 0.5 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.



Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in acetonitrile R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with acetonitrile R. Dilute 2.0 mL of this solution to 10.0 mL with acetonitrile R.

Reference solution (b) Dissolve 2.0 mg of nicergoline for system suitability CRS (containing impurities A, B, C, D, F and H) in acetonitrile R and dilute to 2.0 mL with the same solvent.

Reference solution (c) Dissolve 5.0 mg of nicergoline impurity D CRS in acetonitrile R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 50.0 mL with acetonitrile R.

Reference solution (d) Dissolve the contents of a vial of nicergoline for peak identification CRS (containing impurity I) in 1.0 mL of acetonitrile R.

Column:
 — size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
 — stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3.5 μ m);
 — temperature: 40 °C.

Mobile phase:
 — solution A: dissolve 34.02 g of potassium dihydrogen phosphate R in 930 mL of water R and dilute to 1000 mL with water R (buffer solution); dissolve 21.21 g of tetrabutylammonium hydrogen sulfate R in 225 mL of the buffer solution and dilute to 250.0 mL with the same solution; adjust to pH 7.5 with a 300 g/L solution of potassium hydroxide R;

— **mobile phase A:** mix 2.0 mL of solution A with 300 mL of acetonitrile R and 700 mL of water R;
 — **mobile phase B:** mix 2.0 mL of solution A with 300 mL of water R and 700 mL of acetonitrile R;

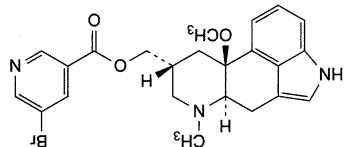
Flow rate 1.2 mL/min.
Detection Spectrophotometer at 288 nm.
Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with nicergoline for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, F and H;

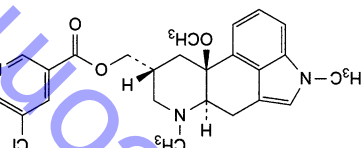
use the chromatogram obtained with reference solution (c) to identify the peak due to impurity D and the chromatogram obtained with reference solution (d) to identify the peak due to impurity I.

Relative retention With reference to nicergoline (retention time = about 3.4 min): impurity D = about 0.06; impurity C = about 0.1; impurity B = about 0.6; impurity H = about 0.8; impurity A = about 0.96; impurity F = about 1.1; impurity I = about 1.2.

System suitability: reference solution (b):
 — **resolution:** minimum 2 between the peaks due to impurity A and nicergoline.



A. [(6aR,9R,10aS)-10a-methoxy-4,7-dimethyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-g]quinolin-9-yl]methyl 5-chloropyridine-3-carboxylate (chloronicergeroline),



B. [(6aR,9R,10aS)-10a-methoxy-7-methyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-g]quinolin-9-yl]methyl 5-bromopyridine-3-carboxylate (1-desmethylnicergeroline),

IMPURITIES
 C₂₄H₂₆BrN₃O₃.
 1 mL of 0.1 M perchloric acid is equivalent to 48.44 mg of (2.2.20). Titrate to the 1st point of inflexion.
 Dissolve 0.400 g in 50 mL of acetone R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically.

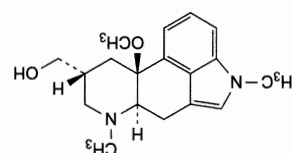
ASSAY
 Maximum 0.1 per cent, determined on 1.0 g.
Sulfated ash (2.4.14)
 Maximum 0.5 per cent, determined on 0.100 g.

Water (2.5.32)
 Maximum 0.5 per cent, determined on 0.100 g.
Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.

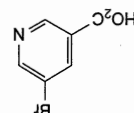
Limits:
 — **impurity B:** not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.8 per cent);
 — **impurity A:** not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
 — **impurity H:** not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
 — **impurity D:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
 — **impurities C, F, I:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
 — **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
 — **total:** maximum 1.2 per cent;
 — **disregard limit:** 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Control of impurities in substances for pharmaceutical use: E, G, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these by the general monograph *Substances for pharmaceutical use* acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of the following substances would, if

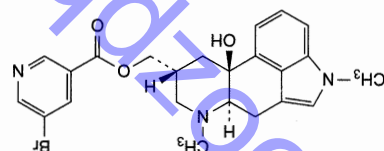
C. [(6aR,9R,10aS)-10a-methoxy-4,7-dimethyl-4,6,6a,7,8,9,10,10a-octahydridoindolo[4,3-fg]quinolin-9-yl]methanol,



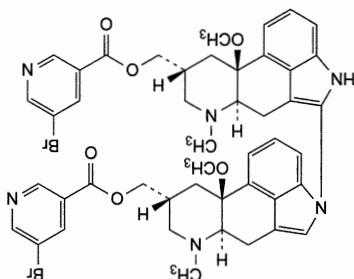
D. 5-bromopyridine-3-carboxylic acid,



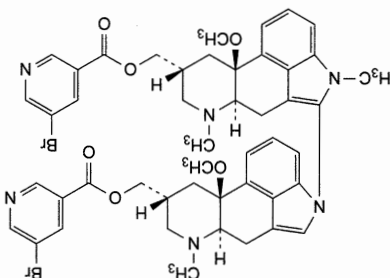
E. [(6aR,9R,10aS)-10a-hydroxy-4,7-dimethyl-4,6,6a,7,8,9,10,10a-octahydridoindolo[4,3-fg]quinolin-9-yl]methyl 5-bromopyridine-3-carboxylate,



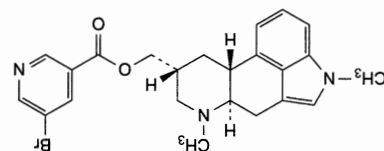
I. [(6aR,6a',9R,9',10aS,10a'S)-9'-[[[(5-bromopyridin-3-yl)carbonyl]oxy]methyl]-10a,10a'-dimethoxy-7,7'-dimethyl-4,6',6a,6a',7,7',8,8',9,9',10,10',10a,10a'-tetradecahydro-6H-4,5',-bimindolo[4,3-fg]quinoline-9-yl]methyl 5-bromopyridine-3-carboxylate,



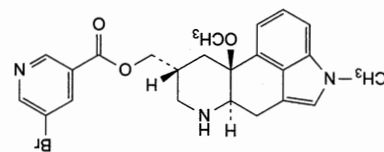
J. [(6aR,6a',9R,9',10aS,10a'S)-9'-[[[(5-bromopyridin-3-yl)carbonyl]oxy]methyl]-10a,10a'-dimethoxy-4,7,7'-trimethyl-4,6',6a,6a',7,7',8,8',9,9',10,10',10a,10a'-tetradecahydro-6H-4,5',-bimindolo[4,3-fg]quinoline-9-yl]methyl 5-bromopyridine-3-carboxylate.



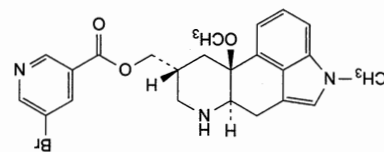
F. [(6aR,9S,10aS)-10a-methoxy-4,7-dimethyl-4,6,6a,7,8,9,10,10a-octahydridoindolo[4,3-fg]quinolin-9-yl]methyl 5-bromopyridine-3-carboxylate (isonicergoline),



G. [(6aR,9R,10aR)-4,7-dimethyl-4,6,6a,7,8,9,10,10a-octahydridoindolo[4,3-fg]quinolin-9-yl]methyl 5-bromopyridine-3-carboxylate,



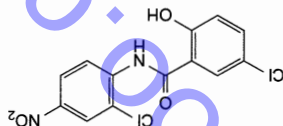
H. [(6aR,9R,10aS)-10a-methoxy-4-methyl-4,6,6a,7,8,9,10,10a-octahydridoindolo[4,3-fg]quinolin-9-yl]methyl 5-bromopyridine-3-carboxylate (6-desmethylnicergoline),



Anhydrous Niclosamide



Niclosamide
(Ph. Eur. monograph 0679)



$C_{13}H_{18}Cl_2N_2O_4$

327.1

50-65-7

Action and use

Anthelmintic.

Preparation

Niclosamide Tablets

Ph. Eur.

DEFINITION

5-Chloro-N-(2-chloro-4-nitrophenyl)-2-hydroxybenzamide.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellowish-white or yellowish, fine crystals.

Solubility

Practically insoluble in water, sparingly soluble in acetone, slightly soluble in anhydrous ethanol.

IDENTIFICATION

First identification B, E.

Second identification A, C, D, E.

A. Melting point (2.2.14): 227 °C to 232 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs prepared using about 0.5 mg of substance

and 0.3 g of potassium bromide R.

Comparison anhydrous niclosamide CRS.

C. To 50 mg add 5 mL of 1 M hydrochloric acid and 0.1 g of

zinc powder R, heat in a water-bath for 10 min, cool and

filter. To the filtrate add 1 mL of a 5 g/L solution of sodium

nitrite R and allow to stand for 3 min; add 2 mL of a 20 g/L

solution of ammonium sulfamate R, shake, allow to stand for

3 min and add 2 mL of a 5 g/L solution of

naphthylethylenediamine dihydrochloride R. A violet colour is

produced.

D. Heat the substance on a copper wire in a non-luminous

flame. The flame becomes green.

E. Loss on drying (see Tests).

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined

in methanol R, heating gently, cool and dilute to 50.0 mL

with the same solvent.

Reference solution Dilute 1.0 mL of the test solution to

100.0 mL with acetonitrile R. Dilute 1.0 mL of this solution

to 20.0 mL with acetonitrile R.

Column:

— size: $l = 0.125$ m, $\varnothing = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Mixture of equal volumes of acetonitrile R and a

solution containing 2 g/L of potassium dihydrogen phosphate R,

1 g/L of disodium hydrogen phosphate R and 2 g/L of

tetraabutylammonium hydrogen sulfate R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Limits:

— total: not more than 4 times the area of the principal peak

in the chromatogram obtained with the reference solution

(0.2 per cent);

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with the reference solution

(0.005 per cent).

5-Chlorosalicylic acid

Maximum 60 ppm.

Test solution To 1.0 g add 15 mL of water R, boil for 2 min,

cool, filter through a membrane filter (nominal pore size

0.45 μ m), wash the filter and dilute the combined filtrate and

washings to 20.0 mL with water R.

Reference solution Dissolve 30 mg of 5-chlorosalicylic acid R in

20 mL of methanol R and dilute to 100.0 mL with water R.

Dilute 1.0 mL of this solution to 100.0 mL with water R.

To 10.0 mL of the test solution and to 10.0 mL of the

reference solution add separately 0.1 mL of ferric chloride

solution R2. Any violet colour in the test solution is not more

intense than that in the reference solution.

2-Chloro-4-nitroaniline

Maximum 100 ppm.

Test solution To 0.250 g add 5 mL of methanol R, heat to

boiling, cool, add 45 mL of 1 M hydrochloric acid, heat again

to boiling, cool, filter and dilute the filtrate to 50.0 mL with

1 M hydrochloric acid.

Reference solution Dissolve 50 mg of 2-chloro-4-nitroaniline R in

methanol R and dilute to 100.0 mL with the same solvent.

Dilute 1.0 mL of the solution to 100.0 mL with methanol R.

Dilute 2.0 mL of this solution to 20.0 mL with 1 M

hydrochloric acid.

To 10.0 mL of the test solution and to 10.0 mL of the

reference solution add separately 0.5 mL of a 5 g/L solution

of sodium nitrite R and allow to stand for 3 min. Add 1 mL of

a 20 g/L solution of ammonium sulfamate R, shake, allow to

stand for 3 min and add 1 mL of a 5 g/L solution of

naphthylethylenediamine dihydrochloride R. Any pinkish-violet

colour in the test solution is not more intense than that in

the reference solution.

Chlorides (2.4.4)

Maximum 500 ppm.

To 2 g add a mixture of 1.2 mL of acetic acid R and 40 mL

of water R, boil for 2 min, cool and filter. Dilute 2 mL of the

filtrate to 15 mL with water R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.3000 g in 80 mL of a mixture of equal volumes of

acetic acid R and methanol R. Titrate with 0.1 M

tetraabutylammonium hydroxide, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M tetraabutylammonium hydroxide is equivalent to

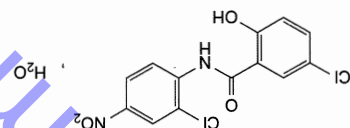
32.71 mg of $C_{13}H_8Cl_2N_2O_4$.

STORAGE

In an airtight container, protected from light.

Niclosamide Monohydrate

(Ph. Eur. monograph 0680)

 $C_{13}H_8Cl_2N_2O_4 \cdot H_2O$

345.1

Action and use

Anthelmintic.

Preparation

Niclosamide Tablets

Ph. Eur.

DEFINITION

5-Chloro-N-(2-chloro-4-nitrophenyl)-2-hydroxybenzamide monohydrate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellowish, fine crystals.

Solubility

Practically insoluble in water, sparingly soluble in acetone, slightly soluble in anhydrous ethanol.

IDENTIFICATION

First identification B, E.

Second identification A, C, D, E.

A. Melting point (2.2.14): 227 °C to 232 °C, determined after drying at 100–105 °C for 4 h.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Dry the substance to be examined at 100–105 °C for 4 h and examine as discs prepared using about 0.5 mg of substance and 0.3 g of potassium bromide R.

Comparison anhydrous niclosamide CRS.

C. To 50 mg add 5 mL of 1 M hydrochloric acid and 0.1 g of zinc powder R, heat in a water-bath for 10 min, cool and filter. To the filtrate add 1 mL of a 5 g/L solution of sodium nitrite R and allow to stand for 3 min, add 2 mL of a 20 g/L solution of ammonium sulfamate R, shake, allow to stand for 3 min and add 2 mL of a 5 g/L solution of naphthylethylenediamine dihydrochloride R. A violet colour is produced.

D. Heat the substance on a copper wire in a non-luminous flame. The flame becomes green.

E. Loss on drying (see Tests).

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in methanol R, heating gently, cool and dilute to 50.0 mL with the same solvent.

Reference solution Dilute 1.0 mL of the test solution to 100.0 mL with acetonitrile R. Dilute 1.0 mL of this solution to 20.0 mL with acetonitrile R.

Column:

— size: $l = 0.125$ m, $\phi = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mixture of equal volumes of acetonitrile R and a solution containing 2 g/L of potassium dihydrogen phosphate R, 1 g/L of disodium hydrogen phosphate R and 2 g/L of tetrabutylammonium hydrogen sulfate R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time Twice the retention time of niclosamide.

Limits:

— total: not more than 4 times the area of the principal peak in the chromatogram obtained with the reference solution

(0.2 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with the reference solution

(0.005 per cent).

5-Chlorosalicylic acid

Maximum 60 ppm.

Test solution To 1.0 g add 15 mL of water R, boil for 2 min, cool, filter through a membrane filter (nominal pore size

ASSAY

Dissolve 0.3000 g in 80 mL of a mixture of equal volumes of acetone R and methanol R. Titrate with 0.1 M

tetrabutylammonium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 32.71 mg of $C_{13}H_8Cl_2N_2O_4$.

STORAGE

Protected from light.

Chlorides (2.4.4)

Maximum 500 ppm.

To 2 g add a mixture of 1.2 mL of acetic acid R and 40 mL of water R, boil for 2 min, cool and filter. Dilute 2 mL of the filtrate to 15 mL with water R.

Loss on drying (2.2.32)

4.5 per cent to 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

2-Chloro-4-nitroaniline

Maximum 100 ppm.

Test solution To 0.250 g add 5 mL of methanol R, heat to boiling, cool, add 45 mL of 1 M hydrochloric acid, heat again to boiling, cool, filter and dilute the filtrate to 50.0 mL with 1 M hydrochloric acid.

Reference solution Dissolve 50 mg of 2-chloro-4-nitroaniline R in methanol R and dilute to 100.0 mL with the same solvent.

Dilute 2.0 mL of this solution to 20.0 mL with 1 M hydrochloric acid.

To 10.0 mL of the test solution and to 10.0 mL of the reference solution add separately 0.5 mL of a 5 g/L solution of sodium nitrite R and allow to stand for 3 min. Add 1 mL of a 20 g/L solution of ammonium sulfamate R, shake, allow to stand for 3 min and add 1 mL of a 5 g/L solution of naphthylethylenediamine dihydrochloride R. Any pinkish-violet colour produced in the test solution is not more intense than that in the reference solution.

Chlorides (2.4.4)

Maximum 500 ppm.

To 2 g add a mixture of 1.2 mL of acetic acid R and 40 mL of water R, boil for 2 min, cool and filter. Dilute 2 mL of the filtrate to 15 mL with water R.

Loss on drying (2.2.32)

4.5 per cent to 6.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Dissolve 0.3000 g in 80 mL of a mixture of equal volumes of acetone R and methanol R. Titrate with 0.1 M

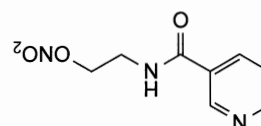
tetrabutylammonium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 32.71 mg of $C_{13}H_8Cl_2N_2O_4$.

STORAGE

Protected from light.

Nicorandil


 $C_8H_9N_3O_4$

211.2

65141-46-0

Action and use

Potassium channel opener; vasodilator.

Preparation

Nicorandil Tablets

DEFINITION

Nicorandil is N-[2-(Nitro-oxy)ethyl]nicotinamide. It contains not less than 98.5% and not more than 101.0% of $C_8H_9N_3O_4$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white to off-white crystalline powder. Freely soluble in methanol and ethanol, sparingly soluble in water.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of Nicorandil (RS 461).
B. Melting point, about 92° with decomposition, Appendix V A.

TESTS

Heavy Metals

2.0 g complies with limit test C for heavy metals, Appendix VII. Use 2 mL of lead standard solution (10 ppm Pb) to prepare the standard (10 ppm).

Sulfates

Dissolve 4.165 g in 30 mL of ethanol (50%), add 1 mL of dilute hydrochloric acid and add sufficient water to produce 50 mL. The solution complies with the limit test for sulfates, Appendix VII (100 ppm).

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in the mobile phase, prepared immediately before use.

(1) 0.2 % w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 200 volumes.

(3) Dilute 1 volume of solution (2) to 5 volumes.

(4) Equal volumes of solution (1) and 0.01% w/v of N-(2-hydroxyethyl)isonicotinamide nitric ester BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Luna C18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1.3 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 254 nm.

(f) Inject 10 µL of each solution.

(g) Allow the chromatography to proceed for 3 times the retention time of nicorandil.

MOBILE PHASE

3 volumes of trifluoroacetic acid, 5 volumes of triethylamine, 10 volumes of tetrahydrofuran and 982 volumes of water.

When the chromatograms are recorded under the prescribed conditions the retention time relative to nicorandil (retention time, about 18 minutes) for N-(2-hydroxyethyl)isonicotinamide nitric ester is about 0.86.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (4), the resolution factor between the peaks due to N-(2-hydroxyethyl)isonicotinamide nitric ester and nicorandil is at least 3.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to N-(2-hydroxyethyl)isonicotinamide nitric ester is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.5%);
the area of any other secondary peak is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.1%);
the sum of the areas of any secondary peaks other than N-(2-hydroxyethyl)isonicotinamide nitric ester is not greater than 0.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.25%).

Disregard any peak with an area less than 0.5 times the area of the principal peak in the chromatogram obtained with solution (3) (0.05%).

Sulfated ash

Not more than 0.1%, Appendix IX A.

Water

Not more than 0.5% w/w, Appendix IX C. Use 5 g.

ASSAY

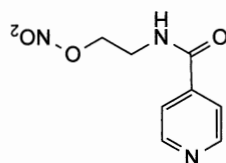
Dissolve 0.15 g in 30 mL of a mixture of 3 volumes of glacial acetic acid and 7 volumes of acetic anhydride. Carry out Method I for non-aqueous titration, Appendix VIII A, determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 21.12 mg of $C_8H_9N_3O_4$.

STORAGE

Nicorandil should be kept in an airtight container and stored at a temperature of 2° to 8°.

IMPURITIES

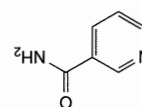
The impurities limited by the requirements of this monograph include:



1. N-(2-hydroxyethyl)isonicotinamide nitric ester

Nicotinamide

(Ph. Eur. monograph 0047)

 $C_6H_6N_2O$

122.1

98-92-0



intense than the spot in the chromatogram obtained with the

reference solution (0.25 per cent).

Heavy metals (2.4.8)

Dilute 12 mL of solution S to 18 mL with water R. 12 mL of the solution complies with test A for heavy metals (30 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.00 g by drying *in vacuo* for 18 h.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 20 mL of anhydrous acetic acid R, heating slightly if necessary, and add 5 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, using crystal violet solution R as indicator until the colour changes to greenish-blue.

1 mL of 0.1 M perchloric acid is equivalent to 12.21 mg of $C_6H_6N_2O$.

Ph Eur

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, freely soluble in water and in ethanol.

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Melting point (2.2.14): 128 °C to 131 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with nicotinamide CRS.

C. Boil 0.1 g with 1 mL of dilute sodium hydroxide solution R. Ammonia is evolved which is recognisable by its odour.

D. Dilute 2 mL of solution S (see Tests) to 100 mL with water R. To 2 mL of the solution, add 2 mL of cyanogen bromide solution R and 3 mL of a 25 g/L solution of aniline R and shake. A yellow colour develops.

TESTS**Solution S**

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

pH (2.2.3)

The pH of solution S is 6.0 to 7.5.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel GF₂₅₄ plate R.

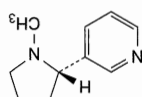
Test solution Dissolve 0.4 g of the substance to be examined in a mixture of equal volumes of alcohol R and water R and dilute to 5.0 mL with the same mixture of solvents.

Reference solution Dilute 0.5 mL of the test solution to 200 mL with a mixture of equal volumes of alcohol R and water R.

Apply to the plate 5 µL of each solution. Develop over a path of 10 cm using a mixture of 4 volumes of water R, 45 volumes of ethanol R and 48 volumes of chloroform R. Allow the plate to dry and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more

Nicotine

(Ph. Eur. monograph 1452)

 $C_{10}H_{14}N_2$

162.2

54-11-5

Action and use

Aid to smoking cessation.

Preparations

Nicotine Inhalation Cartridges

Nicotine Nasal Spray

Nicotine Sublingual Tablets

Nicotine Transdermal Patches

Ph Eur

DEFINITION

3-[(2S)-1-Methylpyrrolidin-2-yl]pyridine.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

Colourless or brownish viscous liquid, volatile, hygroscopic.

Solubility

Soluble in water, miscible with anhydrous ethanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of nicotine.

TESTS**Appearance of solution**

Dissolve 1.0 g in water R and dilute to 10 mL with the same solvent. The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅, BY₅ or R₅ (2.2.2, Method II).

Specific optical rotation (2.2.7)

— 140 to –152.
Dissolve 1.00 g in anhydrous ethanol R and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 20.0 mg of the substance to be examined in water R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of nicotine for system suitability CRS (containing impurities A, B, C, D, E, F and G) in 1.0 mL of water R.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with water R. Dilute 1.0 mL of this solution to 100.0 mL with water R.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R (5 μ m).

Mobile phase:

— mobile phase A: to 900 mL of water R, add 25 mL of a 60 g/L solution of acetic acid R, then add 6 mL of concentrated ammonia R1. Adjust to pH 10.0 with dilute ammonia R2 or dilute acetic acid R and dilute to 1000 mL with water R;
— mobile phase B: acetonitrile R₃.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	100	0
3 - 3.01	100 \rightarrow 95	0 \rightarrow 5
3.01 - 28	95 \rightarrow 74	5 \rightarrow 26
28 - 32	74 \rightarrow 60	26 \rightarrow 40

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied with nicotine for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E, F and G.

Relative retention With reference to nicotine (retention time = about 17.8 min): impurity E = about 0.3; impurity C = about 0.55; impurity F = about 0.7; impurity A = about 0.8; impurity D = about 0.86; impurity G = about 0.9; impurity B = about 1.6.

System suitability: reference solution (a):

— resolution: minimum 2.5 between the peaks due to impurity G and nicotine.

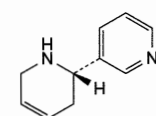
Limits:

— impurities A, B, C, D, E, F, G: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b)

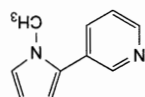
(0.3 per cent);

— area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

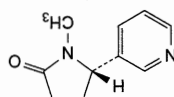
— total: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);



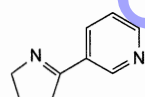
A. (2S)-1,2,3,6-tetrahydro-2,3'-bipyridyl (anatabine).



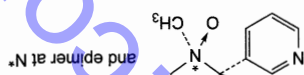
B. 3-(1-methyl-1H-pyrrol-2-yl)pyridine (β -nicotyrine).



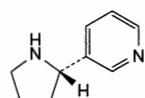
C. (5S)-1-methyl-5-(pyridin-3-yl)pyrrolidin-2-one (cotinine).



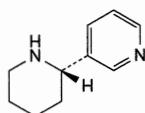
D. 3-(4,5-dihydro-3H-pyrrol-2-yl)pyridine (myosmine).



E. (1R,2S)-1-methyl-2-(pyridin-3-yl)pyrrolidine 1-oxide (nicotine N'-oxide), and epimer at N⁺



F. 3-[(2S)-pyrrolidin-2-yl]pyridine (nicotinic).



G. 3-[(2S)-piperidin-2-yl]pyridine (anabasine).

IMPURITIES

Under nitrogen, in an airtight container, protected from light.

STORAGE

$C_{10}H_{14}N_2$.

1 mL of 0.1 M perchloric acid is equivalent to 8.11 mg of

potentiometrically (2.2.20).

Dissolve 60.0 mg in 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid determining the end-point

ASSAY

Maximum 0.5 per cent, determined on 1.00 g.

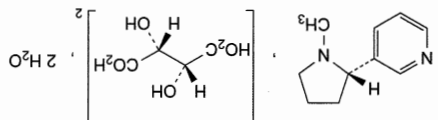
Water (2.5.12)

(0.05 per cent).

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

Nicotine Ditartrate Dihydrate

(Ph Eur monograph 2599)



$C_{18}H_{26}N_2O_{12} \cdot 2H_2O$ 498.4 6019-06-3

Action and use
Aid to smoking cessation.

Ph Eur

DEFINITION
3-[(2S)-1-Methylpyrrolidin-2-yl]pyridine bis[(2R,3R)-2,3-dihydroxybutanedioate] dihydrate.

Content
98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white powder.

Solubility

Soluble in water and in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nicotine ditartrate dihydrate CRS.

TESTS

pH (2.2.3)

3.0 to 3.4.

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

+ 21.0 to + 23.0.

Dissolve 0.25 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 60 mg of the substance to be examined in water R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve the contents of a vial of nicotine for system suitability CRS (containing impurities A, B, C, D, E, F and G) in 1.0 mL of water R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with water R. Dilute 1.0 mL of this solution to 10.0 mL with water R.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

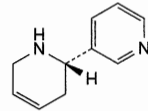
— stationary phase: end-capped polar-embedded octadecylsilyl

amorphous organosilica polymer R (5 μ m).

Mobile phase:

— mobile phase A: to 900 mL of water R add 25 mL of a 60 g/L solution of acetic acid R and 6 mL of concentrated ammonia R1; adjust to pH 10.0 with dilute ammonia R2 or dilute acetic acid R and dilute to 1000.0 mL with water R;

— mobile phase B: acetonitrile R₃.



A. (2S)-1,2,3,6-tetrahydro-2,3'-bipyridyl (anatabine);

Specified impurities A, B, C, D, E, F, G

IMPURITIES

Protected from light.

STORAGE

$C_{18}H_{26}N_2O_{12}$.

1 mL of 0.1 M perchloric acid is equivalent to 23.12 mg of

potentiometrically (2.2.20).

Dissolve 0.180 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

6.5 per cent to 8.0 per cent, determined on 0.100 g.

Water (2.5.12)

(0.05 per cent).

the chromatogram obtained with reference solution (b)

— disregard limit: 0.5 times the area of the principal peak in

(0.8 per cent);

in the chromatogram obtained with reference solution (b)

— total: not more than 8 times the area of the principal peak

with reference solution (b) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

(0.3 per cent);

chromatogram obtained with reference solution (b)

more than 3 times the area of the principal peak in the

— impurities A, B, C, D, E, F, G: for each impurity, not

Limits:

impurity G and nicotine.

— resolution: minimum 2.5 between the peaks due to

System suitability: reference solution (a):

impurity G = about 0.9; impurity B = about 1.6;

impurity A = about 0.8; impurity D = about 0.86;

impurity C = about 0.55; impurity F = about 0.7;

time = about 17.8 min; impurity E = about 0.3;

Relative retention With reference to nicotine (retention

to impurities A, B, C, D, E, F and G.

Identification of impurities Use the chromatogram supplied

with nicotine for system suitability CRS and the chromatogram

obtained with reference solution (a) to identify the peaks due

Injection 20 μ L.

Detection Spectrophotometer at 254 nm.

Flow rate 1.0 mL/min.

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 3	100	0
3 - 3.01	100 \rightarrow 95	0 \rightarrow 5
3.01 - 28	95 \rightarrow 74	5 \rightarrow 26
28 - 32	74 \rightarrow 60	26 \rightarrow 40

Solubility

Practically insoluble in water.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Shake a quantity of the substance to be examined equivalent to 100 mg of nicotine with a mixture of 10 mL of *dilute ammonia* R₂, 10 mL of *water* R, 5 mL of *strong sodium hydroxide solution* R and 20 mL of *hexane* R for 5 min. Transfer the upper layer to a beaker and evaporate to produce an oily residue. Record the spectrum of the oily residue as a thin film between *sodium chloride* R plates.

Comparison Ph. Eur. reference spectrum of nicotine.

B. Nicotine release (see Tests).

TESTS**Nicotine release**

Minimum 70 per cent of the content determined under

Assay in 10 min.

Transfer an accurately weighed quantity of the substance to be examined, equivalent to about 4 mg of nicotine, to a glass-stoppered test-tube, add 10.0 mL of a 9 g/L solution of *sodium chloride* R previously heated to 37 °C and shake vigorously for 10 min. Immediately filter the liquid through a dry filter paper discarding the 1st millilitre of filtrate. Transfer 1.0 mL of the filtrate to a 20 mL volumetric flask, dilute to 20 mL with 0.1 M *hydrochloric acid* and mix. Determine the absorbance (2.2.25) at the minima at about 236 nm and 282 nm and at the maximum at 259 nm using 1.0 mL of a 9 g/L solution of *sodium chloride* R diluted to 20 mL with 0.1 M *hydrochloric acid* as compensation liquid.

Calculate the percentage of nicotine release using the following expression:

$$\frac{20 \times 10^6 \times (A_{259} - 0.5A_{236} - 0.5A_{282})}{323 \times C \times m}$$

323 = specific absorbance of nicotine at 259 nm;

C = percentage of nicotine in the substance

to be examined determined in the assay;

m = mass of the substance to be examined, in milligrams;

$A_{236}, A_{259}, A_{282}$ = absorbances of the solution at the wavelengths indicated by the subscript.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Test solution Weigh a quantity of the substance to be examined, equivalent to 30.0 mg of nicotine, into a glass-stoppered test-tube, add 10.0 mL of *dilute ammonia* R₂ and shake vigorously for 10 min. Centrifuge for 20 min at about 3000 r/min. To 5.0 mL of the clear solution, add 5 mL of a 60 g/L solution of *acetic acid* R and dilute to 25.0 mL with *water* R.

Reference solution (a) Dissolve the contents of a vial of nicotine for system suitability CRS (containing impurities A, B, C, D, E, F and G) in 1.0 mL of *water* R.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with *water* R. Dilute 1.0 mL of this solution to 100.0 mL with *water* R.

Reference solution (c) Dissolve 46.0 mg of nicotine *ditartrate* CRS in *water* R and dilute to 25.0 mL with the same solvent.

Nicotine Resinate

(Ph Eur monograph 1792)

96055-45-7

Action and use

Aid to smoking cessation.

Preparation

Nicotine Resinate Medicated Chewing Gum

Ph Eur

DEFINITION

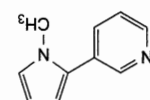
Complex of nicotine (3-[(2S)-1-methylpyrrolidin-2-yl]pyridine) with a weak cationic exchange resin.

Content

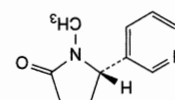
95.0 per cent to 115.0 per cent of the declared content of nicotine ($C_{10}H_{14}N_2$) stated on the label (dried substance). It may contain glycerol.

CHARACTERS**Appearance**

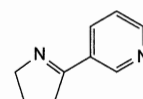
White or slightly yellowish powder, hygroscopic.



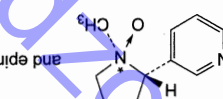
B. 3-(1-methyl-1H-pyrrol-2-yl)pyridine (β-nicotryne),



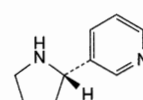
C. (5S)-1-methyl-5-(pyridin-3-yl)pyrrolidin-2-one (cotinine),



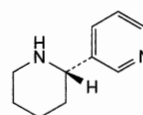
D. 3-(4,5-dihydro-3H-pyrrol-2-yl)pyridine (myosmine),



E. (1R,2S)-1-methyl-2-(pyridin-3-yl)pyrrolidine 1-oxide (nicotine N'-oxide),



F. 3-[(2S)-pyrrolidin-2-yl]pyridine (normicotine),



G. 3-[(2S)-piperidin-2-yl]pyridine (anabasine).



Ph Eur

Column:
— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R (5 μ m).

Mobile phase:

— mobile phase A: to 900 mL of water R, add 25 mL of a 60 g/L solution of acetic acid R, then add 6 mL of concentrated ammonia R1; adjust to pH 10.0 with dilute ammonia R2 or dilute acetic acid R and dilute to 1 L with water R;

— mobile phase B: acetomitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	100	0
3 - 3.01	100 \rightarrow 95	0 \rightarrow 5
3.01 - 28	95 \rightarrow 74	5 \rightarrow 26
28 - 32	74 \rightarrow 60	26 \rightarrow 40

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied

with nicotine for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E, F and G.

Relative retention With reference to nicotine (retention time = about 18 min): impurity E = about 0.3;

impurity C = about 0.55; impurity F = about 0.7;

impurity A = about 0.8; impurity D = about 0.86;

impurity G = about 0.9; impurity B = about 1.6.

System suitability: reference solution (a):

— resolution: minimum 2.5 between the peaks due to impurity G and nicotine.

Limits:

— impurities A, B, C, D, E, F, G: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b)

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 7.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

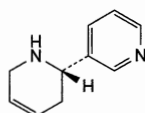
Injection Test solution and reference solution (c).

Calculate the percentage content of nicotine ($C_{10}H_{14}N_2$) taking into account the assigned content of $C_{10}H_{14}N_2$ in nicotine dihydrate CRS.

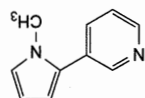
STORAGE

In an airtight container, protected from light.

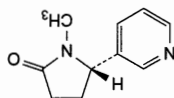
LABELLING
The label states the content of nicotine.
IMPURITIES
Specified impurities A, B, C, D, E, F, G



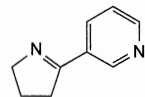
A. (2S)-1,2,3,6-tetrahydro-2,3'-bipyridine (anatabine),



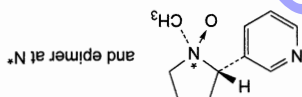
B. 3-(1-methyl-1H-pyrrol-2-yl)pyridine (β -nicotyrine),



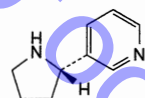
C. (5S)-1-methyl-5-(pyridin-3-yl)pyrrolidin-2-one (cotinine),



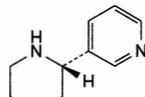
D. 3-(4,5-dihydro-3H-pyrrol-2-yl)pyridine (myosmine),



E. (1R,2S)-1-methyl-2-(pyridin-3-yl)pyrrolidine 1-oxide (nicotine N-oxide),



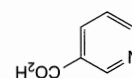
F. 3-[(2S)-pyrrolidin-2-yl]pyridine (nicotidine),



G. 3-[(2S)-piperidin-2-yl]pyridine (anabasine).

Nicotinic Acid

(Ph. Eur. monograph 0459)



123.1

59-67-6

— temperature: 15 °C.

— mobile phase: dilute 2 mL of acetic acid R in 950 mL of water R, adjust to pH 5.6 with dilute ammonia R1 and dilute to 1000 mL with water R;

— mobile phase B: acetonitrile R, methanol R (50:50 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 10	100	0
10 - 30	100→20	0→80
30 - 35	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied

with nicotinic acid impurity mixture CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A and B.

Relative retention With reference to nicotinic acid (retention

time = about 6 min): impurity A = about 2.7;

impurity B = about 2.8.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to

impurities A and B.

Limits:

— unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.05 per cent);

— total: not more than 0.5 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.05 per cent);

— disregard limit: 0.3 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.03 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

Dissolve 0.25 g in water R, heating on a water-bath, and

dilute to 15 mL with the same solvent.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 1 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of water R. Add 0.25 mL of

phenolphthalein solution R. Titrate with 0.1 M sodium hydroxide

until a pink colour is obtained. Carry out a blank titration.

1 mL of 0.1 M sodium hydroxide is equivalent to 12.31 mg of

C₆H₅NO₂.

STORAGE

Protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

Ph Eur

Nicotinic Acid Tablets

Preparation

Component of vitamin B.

Action and use

Content

Pyridine-3-carboxylic acid.

99.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water, soluble in boiling water and in

boiling ethanol (96 per cent). It dissolves in dilute solutions

of alkali hydroxides and carbonates.

IDENTIFICATION

First identification A, B.

Second identification A, C.

A. Melting point (2.2.14): 234 °C to 240 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Solvent mixture Dissolve 6.8 g of potassium dihydrogen

phosphate R in 900 mL of water R, adjust to pH 7.0 with

dilute sodium hydroxide solution R and dilute to 1000 mL with

water R.

Test solution Dissolve 50 mg in the solvent mixture and dilute

to 100.0 mL with the solvent mixture. Dilute 1.0 mL of the

solution to 25.0 mL with the solvent mixture.

Spectral range 237-262 nm.

Absorption maximum At 262 nm.

Absorption minimum At 237 nm.

Absorbance ratio A₂₃₇/A₂₆₂ = 0.46 to 0.50.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.120 g of the substance to be

examined in 200 µL of dilute ammonia R1 and dilute to

10.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with mobile phase A. Dilute 1.0 mL of this

solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of

nicotinic acid impurity mixture CRS (impurities A and B) in

1.0 mL of mobile phase A.

Column:

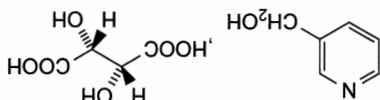
— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: end-capped silica gel for chromatography,

alkyl-bonded for use with highly aqueous mobile phase R

(4 µm);

Nicotiny Alcohol Tartrate



$C_6H_7NO_5 \cdot C_4H_6O_6$ 259.2 6164-87-0

Action and use

Vasodilator.

Preparation

Nicotiny Alcohol Tablets

DEFINITION

Nicotiny Alcohol Tartrate is 3-pyridylmethanol hydrogen (2*R*,3*R*)-tartrate. It contains not less than 98.5% and not more than 101.5% of $C_6H_7NO_5 \cdot C_4H_6O_6$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white crystalline powder.

Freely soluble in water; slightly soluble in ethanol (96%);

practically insoluble in ether.

IDENTIFICATION

A. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.004% w/v solution in 0.1M hydrochloric acid exhibits a maximum only at 261 nm. The absorbance at 261 nm is about 0.84.

B. In the test for Related substances, the principal spot in the chromatogram obtained with solution (2) corresponds to that in the chromatogram obtained with solution (4).

C. Yields reaction B characteristic of tartrates, Appendix VI.

TESTS

Acidity

pH of a 5% w/v solution, 2.8 to 3.7, Appendix V L.

Clarity and colour of solution

A 5.0% w/v solution is clear, Appendix IV A, and not more intensely coloured than reference solution V, Appendix IV B, Method II.

Melting point

146° to 150°, Appendix V A.

Heavy metals

Dissolve the residue obtained in the test for Sulfated ash in 1 mL of 2M hydrochloric acid and dilute to 20 mL with water. 12 mL of the solution complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (2 ppm) to prepare the standard (20 ppm).

Nicotinaldehyde

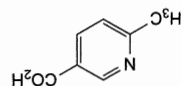
Mix 10 mL of a 10% w/v solution of the substance being examined with 10 mL of a 1.0% w/v solution of phenylhydrazine hydrochloride in 3.6M orthophosphoric acid, dilute to 50 mL with water and allow to stand for 30 minutes. Measure the absorbance of the solution at the maximum at 370 nm, Appendix II B, using in the reference cell a 0.20% w/v solution of phenylhydrazine hydrochloride in 0.72M orthophosphoric acid. The absorbance is not more than that of a solution prepared by treating 10 mL of a 0.0010% w/v solution of pyridine-3-carboxaldehyde in the same manner.

Related substances

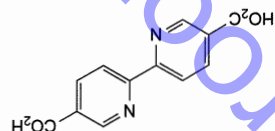
Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in 0.1M ammonia.

the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, G, H, I.

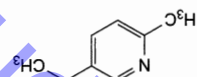
A. 6-methylpyridine-3-carboxylic acid (6-methylnicotinic acid),



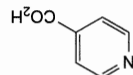
B. 2,2'-bipyridine-5,5'-dicarboxylic acid (6,6'-dinicotinic acid),



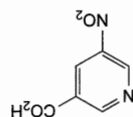
C. 5-ethyl-2-methylpyridine,



D. pyridine-2,5-dicarboxylic acid,



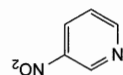
E. pyridine-4-carboxylic acid (isonicotinic acid),



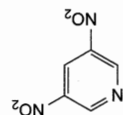
F. 5-nitropyridine-3-carboxylic acid (5-nitronicotinic acid),



G. pyridine,



H. 3-nitropyridine,



I. 3,5-dinitropyridine.

(1) 25% w/v of the substance being examined.

(2) 0.050% w/v of the substance being examined.

(3) 0.050% w/v of 3-(aminomethyl)pyridine.

(4) 0.050% w/v of nicotinic alcohol tartrate BP CRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel GF₂₅₄.

(b) Use the mobile phase as described below.

(c) Apply 5 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry it in air and examine under ultraviolet light (254 nm). Spray with a 2% w/v solution of chloro-2,4,6-trinitrobenzene in absolute ethanol, dry in a

current of air and spray with a 5% w/v solution of sodium carbonate.

MOBILE PHASE

4 volumes of 13.5M ammonia, 16 volumes of methanol,

30 volumes of 1,4-dioxan and 50 volumes of dichloromethane.

LIMITS

By each method of visualisation, in the chromatogram

obtained with solution (1):

any spot corresponding to 3-(aminomethyl)pyridine is not

more intense than the spot in the chromatogram obtained

with solution (3) (0.2%);

any other secondary spot is not more intense than the spot in

the chromatogram obtained with solution (2) (0.2%);

Disregard any spot due to tartaric acid on the line of

Loss on drying

When dried at 105° for 3 hours, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A. Use 2.0 g.

ASSAY

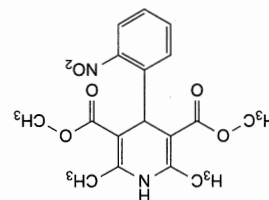
Carry out Method I for non-aqueous titration,

Appendix VIII A, using 0.25 g and determining the end-point potentiometrically. Each mL of 0.1M perchloric acid

VS is equivalent to 25.92 mg of C₁₇H₁₈N₂O₆.

Nifedipine

(Ph. Eur. monograph 0627)



346.3

21829-25-4

C₁₇H₁₈N₂O₆

Action and use

Calcium channel blocker.

Preparations

Nifedipine Capsules

Prolonged-release Nifedipine Capsules

Prolonged-release Nifedipine Tablets

Impurity D and other basic impurities

Transfer 4 g to a 250 mL conical flask and dissolve in 160 mL of glacial acetic acid R using an ultrasonic bath.

Titrate with 0.1 M perchloric acid using 0.25 mL of naphtholbenzene solution R as indicator until the colour changes from brownish-yellow to green. Not more than 0.48 mL of 0.1 M perchloric acid is required (0.14 per cent).

Related substances

Liquid chromatography (2.2.29).

TESTS

D. To 25 mg in a test tube, add 10 mL of a mixture of 1.5 volumes of hydrochloric acid R, 3.5 volumes of water R and 5 volumes of alcohol R and dissolve with gentle heating. Add 0.5 g of zinc R in granules and allow to stand for 5 min with occasional swirling. Filter into a second test tube, add 5 mL of a 10 g/L solution of sodium nitrite R to the filtrate and allow to stand for 2 min. Add 2 mL of a 50 g/L solution of ammonium sulfamate R, shake vigorously with care and add 2 mL of a 5 g/L solution of naphthylethylenediamine dihydrochloride R. An intense red colour develops which persists for not less than 5 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, appearance at 254 nm

and size to the principal spot in the chromatogram obtained with the reference solution.

Detection Examine in ultraviolet light at 254 nm.

Drying In air.

Development Over 3/4 of the plate.

Application 5 µL.

Mobile phase ethyl acetate R, cyclohexane R (40:60 V/V).

Plate TLC silica gel F₂₅₄ plate R.

Reference solution Dissolve 10 mg of nifedipine CRS in methanol R and dilute to 10 mL with the same solvent.

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

C. Thin-layer chromatography (2.2.27).

Comparison nifedipine CRS.

B. Infrared absorption spectrophotometry (2.2.24).

A. Melting point (2.2.14): 171 °C to 175 °C.

Second identification A, C, D.

First identification B.

IDENTIFICATION

Prepare solutions immediately before use in the dark or under long-wave length light (> 420 nm) and protect them from light.

Practically insoluble in water, freely soluble in acetone,

sparingly soluble in ethanol.

When exposed to daylight and to artificial light of certain wavelengths, it readily converts to a nitrosophenylpyridine

derivative. Exposure to ultraviolet light leads to the formation

of a nitrophenylpyridine derivative.

4 volumes of 13.5M ammonia, 16 volumes of methanol,

30 volumes of 1,4-dioxan and 50 volumes of dichloromethane.

By each method of visualisation, in the chromatogram

obtained with solution (1):

any spot corresponding to 3-(aminomethyl)pyridine is not

more intense than the spot in the chromatogram obtained

with solution (3) (0.2%);

any other secondary spot is not more intense than the spot in

the chromatogram obtained with solution (2) (0.2%);

Disregard any spot due to tartaric acid on the line of

the chromatogram obtained with solution (2) (0.2%);

any spot corresponding to 3-(aminomethyl)pyridine is not

more intense than the spot in the chromatogram obtained

with solution (3) (0.2%);

any other secondary spot is not more intense than the spot in

the chromatogram obtained with solution (2) (0.2%);

any spot corresponding to 3-(aminomethyl)pyridine is not

more intense than the spot in the chromatogram obtained

with solution (3) (0.2%);

any other secondary spot is not more intense than the spot in

the chromatogram obtained with solution (2) (0.2%);

any spot corresponding to 3-(aminomethyl)pyridine is not

more intense than the spot in the chromatogram obtained

with solution (3) (0.2%);

Test solution Dissolve 0.200 g of the substance to be examined in 20 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 10 mg of nifedipine impurity A CRS in methanol R and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of nifedipine impurity B CRS in methanol R and dilute to 25.0 mL with the same solvent.

Reference solution (c) Mix 1.0 mL of reference solution (a), 1.0 mL of reference solution (b) and 0.1 mL of the test solution and dilute to 20.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— size: $l = 0.15$ m, $\varnothing = 4.6$ mm,
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m),
— mobile phase: acetonitrile R, methanol R, water R (9:36:55 V/V/V).

Flow rate 1.0 mL/min.
Detection Spectrophotometer at 235 nm.
Injection 20 μ L; inject the test solution and reference solution (c).

Run time Twice the retention time of nifedipine.
Elution order Impurity A, impurity B, nifedipine.
Retention time Nifedipine = about 15.5 min.
System suitability: reference solution (c):
— resolution: minimum 1.5 between the peaks due to impurity A and impurity B and minimum 1.5 between the peaks due to impurity B and nifedipine.

Limits:
— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent),
— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent),
— any other impurity: not more than the area of the peak due to nifedipine in the chromatogram obtained with reference solution (c) (0.1 per cent),
— total: not more than 0.3 per cent,
— disregard limit: 0.1 times the area of the peak due to nifedipine in the chromatogram obtained with reference solution (c) (0.01 per cent).

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

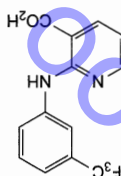
ASSAY
Dissolve 0.1300 g in a mixture of 25 mL of 2-methyl-2-propanol R and 25 mL of perchloric acid solution R. Titrate with 0.1 M cerium sulfate using 0.1 mL of ferroin R as indicator, until the pink colour disappears. Titrate slowly towards the end of the titration. Carry out a blank titration. 1 mL of 0.1 M cerium sulfate is equivalent to 17.32 mg of $C_{17}H_{18}N_2O_6$.

STORAGE
Protected from light.

IMPURITIES
Specified impurities A, B, C, D.

Niflumic Acid

(Ph. Eur. monograph 2115)



$C_{13}H_9F_3N_2O_2$ 282.2 4394-00-7 Ph. Eur.

DEFINITION

2-[[3-(Trifluoromethyl)phenyl]amino]pyridine-3-carboxylic acid.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

Pale yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone, soluble in ethanol (96 per cent) and in methanol.

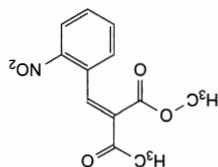
mp: about 204 °C.

IDENTIFICATION

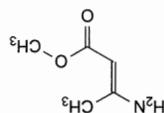
Infrared absorption spectrophotometry (2.2.24).

Comparison niflumic acid CRS.

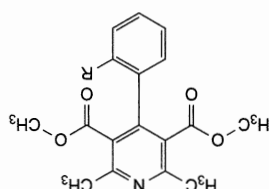
C. methyl 2-(2-nitrobenzylidene)-3-oxobutanoate,



D. methyl 3-aminobut-2-enoate.



A. R = NO₂: dimethyl 2,6-dimethyl-4-(2-nitrophenyl)pyridine-3,5-dicarboxylate (nitrosophenylpyridine analogue),
B. R = NO: dimethyl 2,6-dimethyl-4-(2-nitrophenyl)pyridine-3,5-dicarboxylate (nitrosophenylpyridine analogue),
C. methyl 2-(2-nitrobenzylidene)-3-oxobutanoate,



TESTS

Impurity C

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.50 g of the substance to be examined in 5 mL of methanol R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 25 mg of 3-trifluoromethylaniline R

(impurity C) in 20 mL of methanol R and dilute to 100 mL 100 mL with methanol R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase acetic acid R, ethyl acetate R, toluene R (5:25:90 V/V/V).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In air, until the solvents have evaporated.

Detection Spray with 4-dimethylaminocinnamaldehyde solution R and heat at 60 °C for 10 min.

Limit:

— **impurity C**: any spot due to impurity C is not more intense than the principal spot in the chromatogram obtained with the reference solution (50 ppm).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in 10 mL of acetonitrile R and dilute to 20.0 mL with water R.

Reference solution Dissolve 5.0 mg of niflumic acid impurity A CRS, 5.0 mg of niflumic acid impurity B CRS and 6.0 mg of niflumic acid impurity E CRS in 20 mL of acetonitrile R, add 5.0 mL of the test solution and dilute to 50.0 mL with water R. Dilute 1.0 mL of this solution to 100.0 mL with a mixture of equal volumes of acetonitrile R and water R.

Column:

— **size**: $l = 0.125$ m, $\varnothing = 4.0$ mm;

— **stationary phase**: octylsilyl silica gel for chromatography R (5 µm);

— **temperature**: 25 °C.

Mobile phase phosphoric acid R, acetonitrile R, water R (2.5:500:500 V/V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 267 nm.

Injection 10 µL.

Run time 4 times the retention time of niflumic acid.

Relative retention With reference to niflumic acid (retention time = about 5.5 min): impurity A = about 0.25;

impurity B = about 0.57; impurity E = about 0.64.

System suitability: reference solution:

— **resolution**: minimum 1.5 between the peaks due to impurities B and E.

Limits:

— **impurity B**: not more than 4 times the area of the corresponding peak in the chromatogram obtained with the reference solution (0.4 per cent);

— **impurity A**: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent);

— **unspecified impurities**: for each impurity, not more than the area of the peak due to niflumic acid in the chromatogram obtained with the reference solution (0.10 per cent);

IMPURITIES

$C_{13}H_8F_3N_2O_2$

1 mL of 0.1 M sodium hydroxide is equivalent to 28.22 mg of

ASSAY

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

an oven at 105 °C.

Maximum 0.3 per cent, determined on 2.000 g by drying in

Loss on drying (2.2.32)

using 2 mL of lead standard solution (10 ppm Pb) R.

2.0 g complies with test C. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

metals to 100 mL with water R.

Dilute 1.0 mL of the solution prepared in the test for heavy

Maximum 100 ppm.

Phosphates (2.4.11)

To 10 mL of this solution add 5 mL of water R.

10 mL of methanol R, and dilute to 20 mL with water R.

Dissolve 0.5 g in a mixture of 1 mL of nitric acid R and

Maximum 200 ppm.

Chlorides (2.4.4)

reference solution (0.05 per cent).

niflumic acid in the chromatogram obtained with the

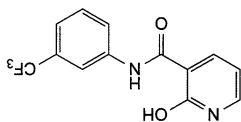
— **disregard limit**: 0.5 times the area of the peak due to

(0.2 per cent);

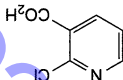
chromatogram obtained with the reference solution

area of the peak due to niflumic acid in the

— **sum of impurities other than B**: not more than twice the



A. 2-chloropyridine-3-carboxylic acid,



Control of impurities in substances for pharmaceutical use): E, F, impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these by the general monograph Substances for pharmaceutical use

acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of Other detectable impurities (the following substances would, if

Specified impurities A, B, C

B. 2-hydroxy-N-[3-(trifluoromethyl)phenyl]pyridine-3-carboxamide,

Protected from light, dissolve 10.0 mg in 10 mL of ethylene glycol monomethyl ether R and dilute to 100.0 mL with methanol R. Dilute 5.0 mL of this solution to 100.0 mL with methanol R.

Impurity A

Maximum 0.05 per cent.

Test solution (a) Dissolve 1.0 g of the substance to be examined in dimethyl sulfoxide R and dilute to 10.0 mL with the same solvent.

Test solution (b) To 5.5 mL of test solution (a) add 50.0 mL of water R while stirring. Allow to stand for 15 min and filter. **Reference solution** To 0.5 mL of test solution (a) add 5.0 mL of a 50 mg/L solution of 4-hydroxybenzoxazole R (impurity A) in dimethyl sulfoxide R. Add 50.0 mL of water R while stirring. Allow to stand for 15 min and filter. Add 0.5 mL of phosphomolybdic reagent R and 10.0 mL of sodium carbonate solution R separately to 10.0 mL of test solution (b) and to 10.0 mL of the reference solution. Allow to stand for 1 h. Examine the 2 solutions at 750 nm. The absorbance (2.2.25) of the solution obtained with test solution (b) is not greater than that obtained with the reference solution.

Related substances

Liquid chromatography (2.2.29). Use amber volumetric flasks, unless otherwise specified. **Solvent mixture** acetonitrile R, water R (40:60 V/V). **Test solution** Dissolve 10.0 mg of the substance to be examined in the solvent mixture, using sonication for not more than 5 min, and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture. **Reference solution (b)** In order to prepare impurity E *in situ*, dissolve 5 mg of the substance to be examined in the solvent mixture in a colorless volumetric flask, using sonication for 5 min, and dilute to 50 mL with the solvent mixture. Allow to stand in ambient light for 1 h. **Reference solution (c)** Dissolve 5.0 mg of methyl parahydroxybenzoate CRS (impurity B) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Column: mixture, size: $l = 0.25$ m, $\phi = 4.6$ mm; stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m); temperature: 10 °C. **Mobile phase:** — mobile phase A: tetrahydrofuran R, water R (5:95 V/V); — mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	67	33
10 - 30	67 \rightarrow 43	33 \rightarrow 57

Flow rate 1.0 mL/min.

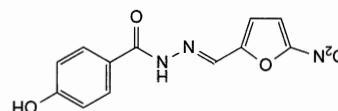
Detection Spectrophotometer at 280 nm.

Injection 50 μ L.

Relative retention With reference to nifuroxazide (retention time = about 8 min): impurity A (keto-enol

Nifuroxazide

(Ph. Eur. monograph 1999)



$C_{12}H_9N_3O_5$ 275.2 965-52-6

Action and use
Antibacterial.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

Bright yellow, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nifuroxazide CRS.

TESTS

Specific absorbance (2.2.25)

940 to 1000 at the absorption maximum at 367 nm.

tautomers) = about 0.36 and 0.39; impurity E = about 0.9; impurity B = about 1.2; impurity C = about 2.6; impurity D = about 3.4.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurity B and nitrofurazide.

Limits:

— impurity E: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

— impurities B, C, D: for each impurity, not more than

0.6 times the area of the principal peak in the

chromatogram obtained with reference solution (c)

(0.3 per cent), and not more than 1 such peak has an

area greater than 0.2 times the area of the principal peak

in the chromatogram obtained with reference solution (c)

(0.1 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— sum of impurities other than E: not more than the area of

the principal peak in the chromatogram obtained with

reference solution (c) (0.5 per cent);

— the chromatogram obtained with reference solution (c)

(0.05 per cent); disregard the peaks due to impurity A.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g, with heating if necessary, in 30 mL of

dimethylformamide R and add 20 mL of water R. Titrate with

0.1 M sodium hydroxide, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 27.52 mg of

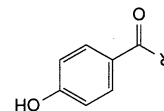
C₁₂H₉N₃O₅.

STORAGE

Protected from light.

IMPURITIES

Specified impurities: A, B, C, D, E.



A. R = NH-NH₂: 4-hydroxybenzohydrazide

(p-hydroxybenzohydrazide),

B. R = OCH₃: methyl 4-hydroxybenzoate (methyl

parahydroxybenzoate),

A. Dissolve 0.15 g in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 1.0 mL of this solution to 100.0 mL with 0.01 M hydrochloric acid. Examined between 230 nm and 350 nm (2.2.25) in a 2 cm cell, the solution shows a single absorption maximum, at 263 nm. The specific absorbance at the maximum is about 285.

Second identification A, C, D.

First identification A, B

IDENTIFICATION

An oily liquid or a crystalline mass, colourless or slightly yellowish, miscible with water and with alcohol.

CHARACTERS

the anhydrous substance.
Nikethamide contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of N₂N-dimethylpyridine-3-carboxamide, calculated with reference to

DEFINITION

Ph Eur

Nikethamide Injection

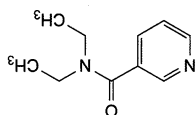
Preparation

Central nervous system stimulant.

Action and use

C₁₀H₁₄N₂O 178.2

59-26-7

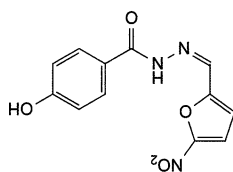


(Ph. Eur. monograph 0233)

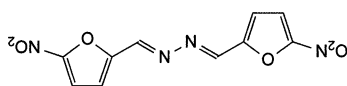
Nikethamide

Ph Eur

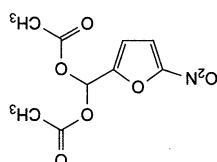
E. (Z)-4-hydroxy-N'-[(5-nitrofuran-2-yl)methylidene]benzohydrazide.



D. (E,E)-N,N'-bis[(5-nitrofuran-2-yl)methylidene]hydrazine



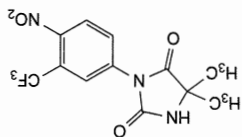
C. (5-nitrofuran-2-yl)methylidene diacetate,





Nilutamide

(Ph. Eur. monograph 2256)



$C_{12}H_{10}F_3N_3O_4$ 317.2 63612-50-0

Action and use

Cytotoxic.

Ph Eur

DEFINITION

5,5-Dimethyl-3-[4-nitro-3-(trifluoromethyl)phenyl]imidazolidine-2,4-dione.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, freely soluble in acetone,

soluble in anhydrous ethanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nilutamide CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture acetonitrile for chromatography R, water R

(35:65 V/V).

Test solution Dissolve 0.10 g of the substance to be examined in the solvent mixture and dilute to 100 mL with the solvent mixture.

Reference solution (a) Dilute 20.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of the substance to be

examined and 2 mg of nilutamide impurity B CRS in the

solvent mixture and dilute to 50 mL with the solvent

mixture.

Column: size: $l = 0.15$ m, $\phi = 4.6$ mm;

stationary phase: spherical octadecylsilyl silica gel for

chromatography R (5 μ m).

Mobile phase:

mobile phase A: 2.0 g/L solution of potassium dihydrogen

phosphate R adjusted to pH 7.5 with 1 M sodium

hydroxide;

mobile phase B: acetonitrile for chromatography R;

Flow rate 1.5 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	55	45
8 - 30	55 \rightarrow 30	45 \rightarrow 70

Ph Eur

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with nilutamide CRS.

C. Heat 0.1 g with 1 mL of dilute sodium hydroxide solution R. Diethylamine is evolved progressively and is recognisable by its characteristic odour and by its turning red litmus paper R blue.

D. Dilute 1 mL of solution S (see Tests) to 250 mL with water R. To 2 mL of this solution add 2 mL of cyanogen bromide solution R. Add 3 mL of a 25 g/L solution of aniline R and shake. A yellow colour develops.

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Appearance

The substance to be examined, in liquid form or liquefied by slight heating, is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

pH (2.2.3)
The pH of solution S is 6.0 to 7.8.

Refractive index

 (2.2.6)
1.524 to 1.526.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.4 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 40 mg of ethylnicotinamide CRS in methanol R and dilute to 100 mL with the same solvent.

Reference solution (b) Dilute 1 mL of reference solution (a) to 10 mL with methanol R.

Apply separately to the plate 10 μ L of each solution. Develop over a path of 15 cm using a mixture of 25 volumes of propanol R and 75 volumes of chloroform R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. In the chromatogram obtained with the test solution, any spot corresponding to ethylnicotinamide is not more intense than the spot in the chromatogram obtained with reference solution (b) ethylnicotinamide, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.1 per cent).

Heavy metals

 (2.4.8)
Dilute 10 mL of solution S to 25 mL with water R. 12 mL of this solution complies with test A for heavy metals (10 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water

 (2.5.12)
Not more than 0.3 per cent, determined on 2.00 g by the semi-micro determination of water.

Sulfated ash

 (2.4.14)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in a mixture of 5 mL of acetic anhydride R and 20 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 17.82 mg of $C_{10}H_{14}N_2O$.

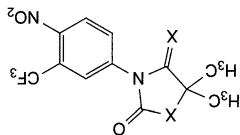
Calculate the percentage content of $C_{12}H_{10}F_3N_3O_4$ from the declared content of niflutamide CRS.

STORAGE

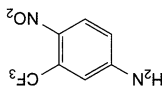
Protected from light.

IMPURITIES

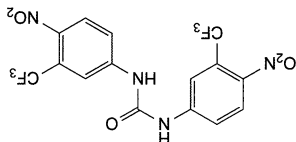
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D.



A. X = NH; 5-imino-4,4-dimethyl-1-[4-nitro-3-(trifluoromethyl)phenyl]imidazolidin-2-one,
 C. X = O; 5,5-dimethyl-3-[4-nitro-3-(trifluoromethyl)phenyl]oxazolidine-2,4-dione,



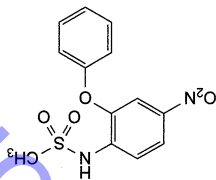
B. 4-nitro-3-(trifluoromethyl)aniline (niflutine),



D. 1,3-bis[4-nitro-3-(trifluoromethyl)phenyl]urea.

Nimesulide

(Ph. Eur. monograph 1548)



$C_{13}H_{12}N_2O_5S$ 308.3 51803-78-2

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

DEFINITION

N-(4-Nitro-2-phenoxyphenyl)methanesulfonamide.

Content

98.5 per cent to 101.5 per cent (dried substance).

Detection Spectrophotometer at 230 nm.

Injection 20 µL.

Relative retention With reference to niflutamide (retention time = about 5.3 min): impurity B = about 0.9.

System suitability: reference solution (b):

— resolution: minimum 3.0 between the peaks due to

Limits:

— unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent);

— total: not more than 1.5 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.3 per cent);

— disregard limit: 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

It complies with test B with the following modifications.

Prescribed solution Dissolve 0.5 g in a mixture of 10 volumes

of water R and 90 volumes of acetone R and dilute to 20 mL

with the same mixture of solvents.

Test solution 12 mL of the prescribed solution.

Reference solution Dilute 0.5 mL of lead standard solution

(10 ppm Pb) R to 10 mL with a mixture of 10 volumes of

water R and 90 volumes of acetone R and add 2 mL of the

prescribed solution.

Filter the solutions through a membrane filter (nominal pore

size 0.45 µm). Compare the spots on the filters obtained with

the different solutions. The substance to be examined

complies with the test if the brown colour of the spot

obtained with the test solution is not more intense than that

of the spot obtained with the reference solution.

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum

crucible.

ASSAY

Liquid chromatography (2.2.29). The solutions are stable for

24 h at room temperature and in daylight.

Solvent mixture acetonitrile for chromatography R, water R

(35:65 V/V).

Test solution Dissolve 50.0 mg of the substance to be

examined in the solvent mixture and dilute to 100.0 mL with

the solvent mixture.

Reference solution Dissolve 50.0 mg of niflutamide CRS in the

solvent mixture and dilute to 100.0 mL with the solvent

mixture.

Column: size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: spherical octadecylsilyl silica gel for

chromatography R (5 µm).

Mobile phase Mix 40 volumes of acetonitrile R and 60 volumes

of a 2.0 g/L solution of potassium dihydrogen phosphate R

adjusted to pH 7.5 with 1 M sodium hydroxide.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 267 nm.

Injection 20 µL.

Retention time About 9 min.

CHARACTERS

Appearance Yellowish, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone, slightly soluble in anhydrous ethanol.

mp

About 149 °C.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nimesulide CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in acetone R, evaporate to dryness and record new spectra using the residues.

TESTS

Absorbance (2.2.25)

Maximum 0.50 at 450 nm.

Dissolve 1.0 g in acetone R and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in 8 mL of acetonitrile R and dilute to 20.0 mL with water R.

Reference solution (a) Dissolve 5 mg of 2-phenoxyaniline R in 10 mL of acetonitrile R and dilute to 25.0 mL with water R.

Reference solution (c) Dissolve 4 mg of nimesulide for peak identification CRS (containing impurities A, B, E and F) in 4.0 mL of acetonitrile R and dilute to 10.0 mL with the mobile phase. Mix 1.0 mL of this solution with the contents of a vial of nimesulide impurity D CRS previously dissolved in 1.0 mL of acetonitrile R.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 4 mg of nimesulide for peak identification CRS (containing impurities A, B, E and F) in 4.0 mL of acetonitrile R and dilute to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 35 volumes of acetonitrile R and 65 volumes of a 1.15 g/L solution of ammonium dihydrogen phosphate R previously adjusted to pH 7.0 with ammonia R.

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time 7 times the retention time of nimesulide.

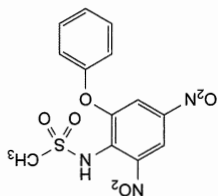
Identification of impurities Use the chromatogram supplied with nimesulide for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, E and F; use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities C and D.

Relative retention With reference to nimesulide (retention time = about 5 min): impurity A = about 0.3;

impurity B = about 2.4; impurity C = about 3.2;

impurity D = about 3.7; impurity E = about 4.2;

impurity F = about 6.1.



A. N-(2,4-dinitro-6-phenoxyphenyl)methanesulfonamide,

IMPURITIES

$C_{13}H_{12}N_2O_5S$.

1 mL of 0.1 M sodium hydroxide is equivalent to 30.83 mg of (2.2.20).

Assay Dissolve 0.240 g in 30 mL of previously neutralised acetone R and add 20 mL of water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C for 4 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Loss on drying (2.2.32)

using 2 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

the chromatogram obtained with reference solution (b)

— *disregard limit*: 0.5 times the area of the principal peak in (0.5 per cent);

in the chromatogram obtained with reference solution (b)

— *total*: not more than 5 times the area of the principal peak with reference solution (b) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— *unspecified impurities*: for each impurity, not more than (0.15 per cent);

chromatogram obtained with reference solution (b)

1.5 times the area of the principal peak in the

— *impurities A, B, C, D, E, F*: for each impurity, not more than solution (b) (0.2 per cent);

peak in the chromatogram obtained with reference

— *impurity E*: not more than twice the area of the principal

corresponding correction factor: impurity C = 0.7;

the peak areas of the following impurities by the

— *correction factors*: for the calculation of content, multiply

— *limits*:

— *resolution*: minimum 2.0 between the peaks due to

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

— *impurities C and D*.

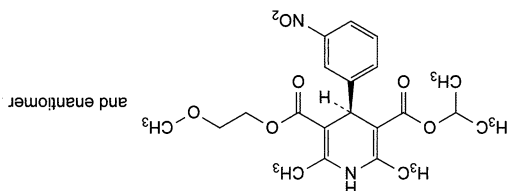
— *impurities C and D*.

— *impurities C and D*.



Nimodipine

(Ph. Eur. monograph 1245)



66085-59-4

418.4

 $C_{21}H_{26}N_2O_7$

Action and use
Calcium channel blocker.

Preparations

Nimodipine Infusion

Nimodipine Tablets

Ph Eur

DEFINITION

2-Methoxyethyl 1-methylethyl (4*R*,5)-2,6-dimethyl-4-(3-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

Light yellow or yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in ethyl acetate, sparingly soluble in anhydrous ethanol.

It shows polymorphism (5.9).

Exposure to ultraviolet light leads to the formation of a nitrophenylpyridine derivative.

Prepare solutions immediately before use either protected from light or under long-wavelength light (> 420 nm).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nimodipine CRS.

If the spectra obtained in the solid state show differences, record new spectra using 20 g/L solutions in methylene chloride *R* and a 0.2 mm cell.

TESTS

Solution S
Dissolve 1.0 g in acetone *R* and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1).

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

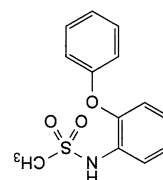
Related substances

Liquid chromatography (2.2.29).

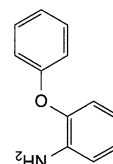
Test solution Dissolve 40.0 mg of the substance to be examined in 2.5 mL of tetrahydrofuran *R* and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

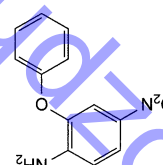
Reference solution (b) Nimodipine impurity A CRS



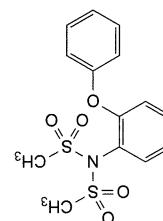
B. N-(2-phenoxyphenyl)methanesulfonamide,



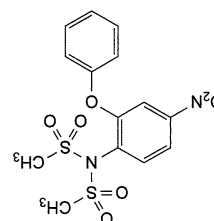
C. 2-phenoxyaniline,



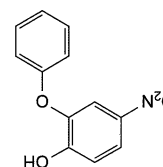
D. 4-nitro-2-phenoxyaniline,



E. N,N-bis(methylsulfonyl)-2-phenoxyaniline,



F. N,N-bis(methylsulfonyl)-4-nitro-2-phenoxyaniline,



G. 4-nitro-2-phenoxyphenol.

Ph Eur

Reference solution (c) Dilute the test solution as described in the leaflet accompanying nitrazepam impurity A CRS.
Reference solution (d) Mix reference solution (b) and reference solution (c) as described in the leaflet accompanying nitrazepam impurity A CRS.
Column:
— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 40 °C.
Mobile phase: methanol R, tetrahydrofuran R, water R (20:20:60 V/V/V).
Flow rate 2.0 mL/min.
Detection Spectrophotometer at 235 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (d).
Run time 4 times the retention time of nitrazepam.
Retention time Impurity A = about 7 min;
nitrazepam = about 8 min.

System suitability Reference solution (d):
— resolution: minimum 1.5 between the peaks due to impurity A and nitrazepam.
Limits:
— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.1 per cent);
— impurities B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— total: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— disregard limit: 0.5 times the area of the peak due to nitrazepam in the chromatogram obtained with reference solution (d) (0.05 per cent).

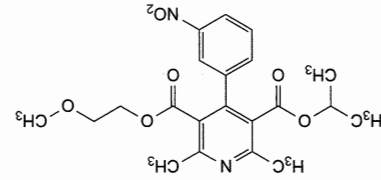
Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve with gentle heating 0.180 g in a mixture of 25 mL of 2-methyl-2-propanol R and 25 mL of perchloric acid solution R. Add 0.1 mL of ferric R. Titrate with 0.1 M cerium sulfate. Titrate slowly towards the end of the titration. Carry out a blank titration.
1 mL of 0.1 M cerium sulfate is equivalent to 20.92 mg of $C_{21}H_{26}N_2O_7$.

STORAGE
Protected from light.

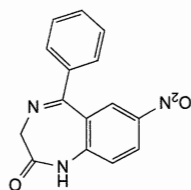
IMPURITIES
Specified impurities: A, B, C.



A. 2-methoxyethyl 1-methylethyl 2,6-dimethyl-4-(3-nitrophenyl)pyridine-3,5-dicarboxylate,

Nitrazepam

(Ph. Eur. monograph 0415)



$C_{21}H_{26}N_2O_7$ 281.3
146-22-5

Action and use

Benzodiazepine.

Preparations

Nitrazepam Oral Suspension

Nitrazepam Tablets

Ph. Eur.

DEFINITION
7-Nitro-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellow, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nitrazepam CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Test solution Dissolve 50 mg of the substance to be examined in acetone R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with acetone R. Dilute 1.0 mL of this solution to 10.0 mL with acetone R.

Reference solution (b) Dissolve 2 mg of clonazepam CRS in acetone R and dilute to 100.0 mL with the same solvent.

Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Column:
 — size: $l = 0.25$ m, $\varnothing = 4.0$ mm;
 — stationary phase: octylsilyl silica gel for chromatography R
 (5 μ m);
 — temperature: 40 °C.
Mobile phase:
 — mobile phase A: 7.8 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R;
 — mobile phase B: acetonitrile R;
 —

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0-3	65	35
3-10	65 → 50	35 → 50
10-20	50	50

Flow rate 1 mL/min.
Detection Spectrophotometer at 270 nm.
Injection 10 μ L.

Relative retention With reference to nitrzapam (retention time = about 9 min): clonazepam = about 1.1.

System suitability: reference solution (b):
 — peak-to-valley ratio: minimum 4.0, where H_p = height above the baseline of the peak due to clonazepam and H_v = height above the baseline of the peak due to the

curve separating this peak from the peak due to nitrzapam.
Limits:
 — unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
 — total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
 — disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)
 Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
 Dissolve 0.250 g in 25 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

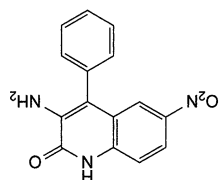
1 mL of 0.1 M perchloric acid is equivalent to 28.13 mg of $C_{15}H_{11}N_3O_3$.

STORAGE

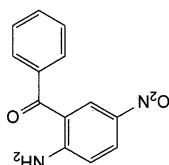
Protected from light.

IMPURITIES

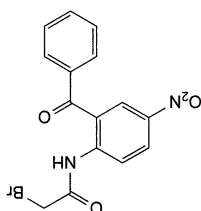
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D.



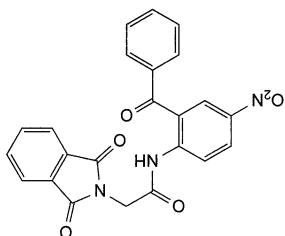
A. 3-amino-6-nitro-4-phenylquinolin-2(1H)-one,



B. (2-amino-5-nitrophenyl)phenylmethanone,



C. 2-bromo-N-[4-nitro-2-(phenylcarboxyl)phenyl]acetamide,

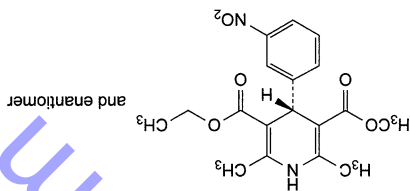


D. 2-(1,3-dioxo-1,3-dihydro-2H-isoindol-2-yl)-N-[4-nitro-2-(phenylcarboxyl)phenyl]acetamide.

Ph Eur

Nitrendipine

(Ph. Eur. monograph 1246)



$C_{18}H_{20}N_2O_6$ 360.4 39562-70-4

Action and use

Calcium channel blocker.

Ph Eur

DEFINITION

Ethyl methyl (4*R*)-2,6-dimethyl-4-(3-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate.

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in ethyl acetate, sparingly soluble in anhydrous ethanol and in methanol.

It shows polymorphism (5.9).

Exposure to ultraviolet light leads to the formation of a nitrophenylpyridine derivative.

Prepare solutions immediately before use either protected from light or under long-wavelength light (> 420 nm).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nitrendipine CRS.

If the spectra obtained in the solid state show differences, record new spectra using 20 g/L solutions in methylene chloride R and a 0.2 mm cell.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in 2.5 mL of tetrahydrofuran R and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 15.0 mg of nitrendipine impurity A CRS in 2.5 mL of tetrahydrofuran R and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 0.5 mL of the test solution to 20.0 mL with the mobile phase.

Reference solution (d) Mix 1.0 mL of reference solution (b) and 1.0 mL of reference solution (c), then dilute to 25.0 mL with the mobile phase.

Reference solution (e) Dissolve 2 mg of nitrendipine for peak identification CRS (containing impurities B and C) in 0.5 mL of tetrahydrofuran R and dilute to 1.0 mL with the mobile phase.

IMPURITIES

Protected from light.

STORAGE

of $C_{18}H_{20}N_2O_6$.

1 mL of 0.1 M cerium sulfate is equivalent to 18.02 mg

end of the titration. Carry out a blank titration.

0.1 mL of ferrous R as indicator. Titrate slowly towards the

acid solution R. Titrate with 0.1 M cerium sulfate, using

Dissolve 0.160 g with gentle heating if necessary in a mixture

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— **disregard limit**: 0.5 times the area of the principal peak in

— **total**: maximum 0.7 per cent;

with reference solution (a) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— **unspecified impurities**: for each impurity, not more than the

solution (d) (0.15 per cent);

peak in the chromatogram obtained with reference

— **impurity A**: not more than the area of the corresponding

obtained with reference solution (a) (0.4 per cent);

the area of the principal peak in the chromatogram

— **impurities B, C**: for each impurity, not more than 4 times

Limits:

impurity A and nitrendipine.

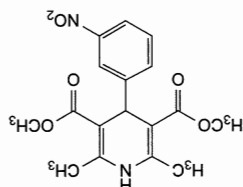
— **resolution**: minimum 2.0 between the peaks due to

System suitability Reference solution (d):

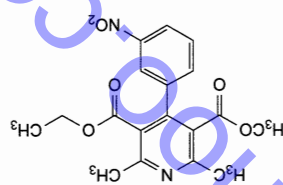
impurity A = about 0.8; impurity C = about 1.4.

time = about 9 min; impurity B = about 0.7;

Relative retention With reference to nitrendipine (retention



A. ethyl methyl 2,6-dimethyl-4-(3-nitrophenyl)pyridine-3,5-dicarboxylate,



Specified impurities A, B, C.

B. dimethyl 2,6-dimethyl-4-(3-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate,

the peaks due to impurities B and C; use the chromatogram obtained with reference solution (d) to identify the peak due

chromatogram obtained with reference solution (e) to identify

with nitrendipine for peak identification CRS and the

Identification of impurities Use the chromatogram supplied

Run time 5 times the retention time of nitrendipine.

solutions (a), (d) and (e).

Injection 10 µL of the test solution and reference

Detection Spectrophotometer at 235 nm.

Flow rate 1 mL/min.

(14:22:64 V/V/V).

Mobile phase acetonitrile R, tetrahydrofuran R, water R

— temperature: 40 °C.

chromatography R (5 µm);

— stationary phase: irregular octadecylsilyl silica gel for

— size: l = 0.125 m, Ø = 4 mm;

Column:

phase.

of tetrahydrofuran R and dilute to 1.0 mL with the mobile

identification CRS (containing impurities B and C) in 0.5 mL

Reference solution (e) Dissolve 2 mg of nitrendipine for peak

with the mobile phase.

Reference solution (d) Mix 1.0 mL of reference solution (b)

and 1.0 mL of reference solution (c), then dilute to 25.0 mL

with the mobile phase.

Reference solution (c) Dilute 0.5 mL of the test solution to

solution to 20.0 mL with the mobile phase.

10.0 mL with the mobile phase. Dilute 1.0 mL of this

impurity A CRS in 2.5 mL of tetrahydrofuran R and dilute to

Reference solution (b) Dissolve 15.0 mg of nitrendipine

solution to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

the mobile phase.

in 2.5 mL of tetrahydrofuran R and dilute to 10.0 mL with

Test solution Dissolve 20 mg of the substance to be examined

Liquid chromatography (2.2.29).

Related substances

chloride R and a 0.2 mm cell.

If the spectra obtained in the solid state show differences,

record new spectra using 20 g/L solutions in methylene

Comparison nitrendipine CRS.

Infrared absorption spectrophotometry (2.2.24).

Prepare solutions immediately before use either protected from light

or under long-wavelength light (> 420 nm).

nitrophenylpyridine derivative.

Exposure to ultraviolet light leads to the formation of a

It shows polymorphism (5.9).

Practically insoluble in water, freely soluble in ethyl acetate,

sparingly soluble in anhydrous ethanol and in methanol.

Solubility

Yellow, crystalline powder.

CHARACTERS

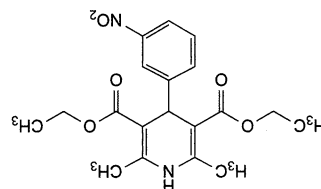
Appearance

Yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in ethyl acetate,

sparingly soluble in anhydrous ethanol and in methanol.



C, diethyl 2,6-dimethyl-4-(3-nitrophenyl)-1,4-dihydropyridine-3,5-dicarboxylate.

Nitric Acid

(Ph. Eur. monograph 1549)

HNO₃ 63.0 7697-37-2



Ph Eur

DEFINITION

Content 68.0 per cent *m/m* to 70.0 per cent *m/m*.

CHARACTERS

Appearance Clear, colourless or almost colourless liquid.

Solubility

Miscible with water.

Relative density

About 1.41.

IDENTIFICATION

A. Dilute 1 mL to 100 mL with *water R*. The solution is

strongly acid (2.2.4).

B. 0.2 mL of the solution obtained in identification test A

gives the reaction of nitrates (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dilute 2 mL to 10 mL with *water R*.

Chlorides (2.4.4)

Maximum 0.5 ppm.

To 5 g add 10 mL of *water R* and 0.3 mL of silver nitrate

solution R2 and allow to stand for 2 min protected from light.

Any opalescence is not more intense than that of a standard

prepared at the same time in the same manner using 13 mL

of *water R*, 0.5 mL of nitric acid *R*, 0.5 mL of chloride

standard solution (5 ppm Cl) *R* and 0.3 mL of silver nitrate

Sulfates (2.4.13)

Maximum 10 ppm.

To 15 g add 0.2 g of sodium carbonate *R*. After carbon

dioxide has evolved, evaporate to dryness. Dissolve the

residue in 15 mL of distilled *water R*.

Iron (2.4.9)

Maximum 10 ppm.

Dissolve the residue obtained in the test for sulfated ash in

1 mL of dilute hydrochloric acid *R* and dilute to 20 mL with

water R. Dilute 1 mL of this solution to 10 mL with *water R*.

Heavy metals (2.4.8)

Maximum 2 ppm.

Nitric Oxide

(Ph. Eur. monograph 1550)

NO 30.01 10102-43-9



Ph Eur

DEFINITION

Content

Minimum 99.0 per cent *V/V* of NO.

CHARACTERS

This monograph applies to nitric oxide for medicinal use.

Appearance

Colourless gas which turns brown when exposed to air.

Solubility

At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in

about 21 volumes of water.

PRODUCTION

Carbon dioxide

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas Mixture containing 3000 ppm *V/V* of carbon

Column:

material: stainless steel;

size: *l* = 3.5 m, \varnothing = 2 mm;

stationary phase: ethylnitrobenzene-divinylbenzene

copolymer *R*;

temperature: 50 °C.

Carrier gas helium for chromatography *R*.

Flow rate 15 mL/min.

Detection Thermal conductivity.

Injection Loop injector.

System suitability:

the chromatograms obtained show a clear separation of

carbon dioxide from nitric oxide.

Limit:

carbon dioxide: not more than the area of the

corresponding peak in the chromatogram obtained with

the reference gas (3000 ppm *V/V*).

Nitrogen

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas Mixture containing 3000 ppm V/V of nitrogen R

in helium for chromatography R.

Column:

material: stainless steel;

size: $l = 3.5$ m, $\varnothing = 2$ mm;

stationary phase: molecular sieve for chromatography R

temperature: 50°C .

Carrier gas helium for chromatography R.

Flow rate 15 mL/min.

Detection Thermal conductivity.

Injection Loop injector.

System suitability:

the chromatograms obtained show a clear separation of

nitrogen from nitric oxide.

Limit:

nitrogen: not more than the area of the corresponding

peak in the chromatogram obtained with the reference gas

(3000 ppm V/V).

Nitrogen dioxide

Maximum 400 ppm V/V.

Ultraviolet absorption spectrophotometry analyser.

Gas to be examined The substance to be examined.

Reference gas (a) nitrogen R1.

Reference gas (b) Mixture containing 400 ppm V/V of nitrogen

dioxide R in nitrogen R.

Apparatus:

an ultraviolet-visible light source (analytical wavelength

about 400 nm);

a sample gas cell through which the feed gas flows;

a closed reference gas cell containing nitrogen R1 in

parallel with the sample gas cell;

a rotating chopper which feeds light alternately through

the reference gas cell and the sample gas cell;

a semiconductor detector which generates a frequency

modulated output whose amplitude is a measure of the

difference of absorption of the sample gas and the

reference gas.

Analysis:

set the zero of the instrument using reference gas (a)

through the sample gas cell at a flow rate of 1 L/min;

adjust the span while feeding reference gas (b) through

the sample gas cell at a flow rate of 1 L/min;

at a flow rate of 1 L/min, read the value from the

instrument output and calculate, if necessary, the

concentration of nitrogen dioxide.

Nitrous oxide

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas Mixture containing 3000 ppm V/V of nitrous

oxide R in nitrogen R.

Column:

material: stainless steel;

size: $l = 3.5$ m, $\varnothing = 2$ mm;

stationary phase: ethylvinylbenzene-divinylbenzene

copolymer R;

temperature: 50°C .

Carrier gas helium for chromatography R.

Flow rate 15 mL/min.

Detection Thermal conductivity.

Injection Loop injector.

System suitability:

the chromatograms obtained show a clear separation of

nitrous oxide from nitric oxide.

Limit:

nitrous oxide: not more than the area of the corresponding

peak in the chromatogram obtained with the reference gas

(3000 ppm V/V).

Water (2.5.28)

Maximum 100 ppm V/V.

Assay

Determine the content of nitric oxide by difference using the

mass balance equation after determining the sum of the

impurities described under Production.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of nitric oxide.

STORAGE

Compressed at a pressure not exceeding 2.5 MPa (25 bars)

measured at 15°C , in suitable containers complying with the

legal regulations.

IMPURITIES

Specified impurities A, B, C, D, E

A. CO_2 : carbon dioxide;

B. N_2 : nitrogen;

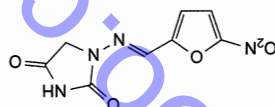
C. NO_2 : nitrogen dioxide;

D. N_2O : nitrous oxide;

E. H_2O : water.

Nitrofurantoin

(Ph. Eur. monograph 0101)



$\text{C}_8\text{H}_6\text{N}_4\text{O}_5$

238.2

67-20-9

Action and use

Antibacterial.

Preparations

Nitrofurantoin Oral Suspension

Nitrofurantoin Tablets

Ph Eur

DEFINITION

Nitrofurantoin contains not less than 98.0 per cent and not

more than the equivalent of 102.0 per cent of

1-[[[(5-nitrofurant-2-yl)methylene]amino]imidazolidine-2,4-

dione, calculated with reference to the dried substance.

CHARACTERS

A yellow, crystalline powder or yellow crystals, very slightly

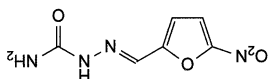
soluble in water and in ethanol (96 per cent), soluble in

dimethylformamide.



Nitrofurazone

(Nitrofurazone, Ph Eur monograph 1135)



198.1

$C_6H_6N_4O_4$

59-87-0

Action and use

Antibacterial; topical antiprotzoal.

DEFINITION

2-[(5-Nitrofuran-2-yl)methylene]diazanecarboxamide.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow or brownish-yellow, crystalline powder.

Solubility

Very slightly soluble in water, slightly soluble in ethanol

(96 per cent).

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25). Carry out the test protected from bright light.

Test solution Use the solution prepared for the assay.

Spectral range 220–400 nm.

Absorption maxima At 260 nm and 375 nm.

Absorbance ratio $A_{375}/A_{260} = 1.15$ to 1.30.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison nitrofurazone CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of nitrofurazone CRS in

methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase methanol R, nitromethane R (10:90 V/V).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with phenylhydrazine hydrochloride solution R.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

reference solution.

D. Dissolve about 1 mg in 1 mL of dimethylformamide R and

add 0.1 mL of alcoholic potassium hydroxide solution R.

A violet-red colour is produced.

TESTS

pH (2.2.3)

5.0 to 7.0.

To 1.0 g add 100 mL of carbon dioxide-free water R. Shake

and filter.

Related substances

Liquid chromatography (2.2.29).

IDENTIFICATION

A. Carry out the test protected from bright light. Use the solution

prepared for the assay. Examined between 220 nm and

400 nm (2.2.25), the solution shows two absorption maxima,

at 266 nm and 367 nm. The ratio of the absorbance at the

maximum at 367 nm to that at the maximum at 266 nm is

1.36 to 1.42.

B. Dissolve about 10 mg in 10 mL of dimethylformamide R.

To 1 mL of the solution add 0.1 mL of 0.5 M alcoholic

potassium hydroxide. A brown colour develops.

TESTS

Related substances

Examine by thin-layer chromatography (2.2.27), using silica

gel HF₂₅₄ R as the coating substance.

Test solution Dissolve 0.25 g of the substance to be examined

in a minimum of dimethylformamide R and dilute to 10 mL

with acetone R.

Reference solution Dilute 1 mL of the test solution to 100 mL

with acetone R.

Apply separately to the plate 10 µL of each solution. Develop

over a path of 15 cm using a mixture of 10 volumes of

methanol R and 90 volumes of nitromethane R. Allow the plate

to dry in air and heat at 100 °C to 105 °C for 5 min.

Examine in ultraviolet light at 254 nm. Spray with

phenylhydrazine hydrochloride solution R. Heat the plate at

100 °C to 105 °C for a further 10 min. When examined in

ultraviolet light and after spraying, any spot in the

chromatogram obtained with the test solution, apart from the

principal spot, is not more intense than the spot in the

chromatogram obtained with the reference solution

(1.0 per cent).

Loss on drying (2.2.32)

Not more than 1.0 per cent, determined on 1.00 g by drying

in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Carry out the assay protected from bright light. Dissolve 0.120 g

in 50 mL of dimethylformamide R and dilute to 1000.0 mL

with water R. Dilute 5.0 mL of the solution to 100.0 mL

with a solution containing 18 g/L of sodium acetate R and

0.14 per cent V/V of glacial acetic acid R. Measure the

absorbance (2.2.25) at the absorption maximum at 367 nm,

using the sodium acetate solution described above as

compensation liquid.

Calculate the content of $C_6H_6N_4O_4$, taking the specific

absorbance to be 765.

STORAGE

Store protected from light, at a temperature below 25 °C.

Ph Eur

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of nitrofurantoin impurity B CRS in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 10 mg of nitrofurantoin R in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute

5.0 mL of the solution to 100.0 mL with the mobile phase. **Reference solution (c)** Dissolve with the aid of ultrasound contents of a vial of nitrofurantoin for peak identification CRS (containing impurities A and B) in 1.0 mL of the mobile phase.

Column:
— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R, water R (40:60 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 310 nm.

Injection 20 μ L.

Run time 10 times the retention time of nitrofurantoin.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peaks due to

impurities A and B.

Relative retention With reference to nitrofurantoin (retention time = about 4 min): nitrofurantoin = about 1.2;

impurity B = about 4.0; impurity A = about 7.6.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

nitrofurantoin and nitrofurantoin.

Limits:

— impurities A, B: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— unspecified impurities: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a)

(1.0 per cent);

— the chromatogram obtained with reference solution (a)

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Carry out the assay protected from bright light

Dissolve 60.0 mg in 20 mL of dimethylformamide R and dilute to 500.0 mL with water R. Dilute 5.0 mL of the solution to 100.0 mL with water R. Prepare a reference solution in the same manner using 60.0 mg of nitrofurantoin CRS. Measure the absorbances (2.2.25) of the 2 solutions at the absorption maximum at 375 nm.

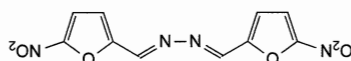
Calculate the content of $C_6H_6N_4O_4$ from the absorbances measured and the concentrations of the solutions.

STORAGE

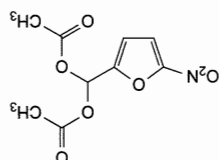
Protected from light.

IMPURITIES

Specified impurities A, B



A. 1,2-bis[(5-nitrofuran-2-yl)methylidene]diazane,



B. (5-nitrofuran-2-yl)methylene diacetate.

Nitrogen

(Ph. Eur. monograph 1247)

N₂ 28.01

7727-37-9

Nitrogen should be kept in approved metal cylinders, the shoulders of which are painted black and the remainder grey. The cylinder should carry a label stating 'Nitrogen'.

Ph. Eur.

DEFINITION

Content

Minimum 99.5 per cent V/V of N₂.

This monograph applies to nitrogen for medicinal use.

CHARACTERS

Appearance

Colourless, odourless gas.

Solubility

At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in about 62 volumes of water and about 10 volumes of ethanol

PRODUCTION

(96 per cent).

Carbon dioxide

Maximum 300 ppm V/V, determined using an infrared analyser (2.5.24).

Gas to be examined The substance to be examined. It must be filtered to avoid stray light phenomena.

Reference gas (a) Nitrogen R1.

Reference gas (b) Mixture containing 300 ppm V/V of carbon dioxide R1 in nitrogen R1.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon dioxide in the gas to be examined.

Carbon monoxide

Maximum 5 ppm V/V, determined using an infrared analyser (2.5.25).

Gas to be examined The substance to be examined. It must be filtered to avoid stray light phenomena.

Reference gas (a) Nitrogen R1.

Reference gas (b) Mixture containing 5 ppm V/V of carbon monoxide R in nitrogen R1.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon monoxide in the gas to be examined.

Oxygen

Maximum 50 ppm V/V , determined using an oxygen analyser with a detector scale ranging from 0-100 ppm V/V and equipped with an electrochemical cell.

The gas to be examined passes through a detection cell containing an aqueous solution of an electrolyte, generally potassium hydroxide. The presence of oxygen in the gas to be examined produces variation in the electric signal recorded at the outlet of the cell that is proportional to the oxygen content.

Calibrate the analyser according to the instructions of the manufacturer. Pass the gas to be examined through the analyser using a suitable pressure regulator and airtight metal tubes and operating at the prescribed flow-rates until constant readings are obtained.

Water (2.5.28)

Maximum 67 ppm V/V .

Assay

Gas chromatography (2.2.28).
Gas to be examined The substance to be examined.

Reference gas (a) Ambient air.

Reference gas (b) Nitrogen R1.

Column:

— material: stainless steel;

— size: $l = 2$ m, $\varnothing = 2$ mm;

— stationary phase: molecular sieve for chromatography R

(0.5 nm).

Carrier gas helium for chromatography R.

Flow rate 40 mL/min.

Temperature:

— column: 50 °C;

— detection: 130 °C.

Detection Thermal conductivity.

Injection Loop injector.

Inject reference gas (a). Adjust the injected volumes and operating conditions so that the height of the peak due to nitrogen in the chromatogram obtained with the reference gas is at least 35 per cent of the full scale of the recorder.

System suitability:

— the chromatograms obtained show a clear separation of oxygen and nitrogen.

Calculate the content of N_2 in the gas to be examined.

IDENTIFICATION

First identification A

Second identification B, C

A. Examine the chromatograms obtained in the assay (see Production).

Results The principal peak in the chromatogram obtained with the substance to be examined is similar in retention time to the principal peak in the chromatogram obtained with reference gas (b).

B. In a 250 mL conical flask replace the air by the substance in the flask. The splinter is extinguished.

C. In a suitable test tube, place 0.1 g of magnesium R in turnings. Close the tube with a two-hole stopper fitted with a glass tube reaching about 1 cm above the turnings. Pass the substance to be examined through the glass tube for 1 min

DEFINITION

This monograph applies to nitrogen which is used for inerting finished medicinal products which are particularly sensitive to degradation by oxygen. It does not necessarily apply to nitrogen used in earlier production steps.

Content

Minimum 99.5 per cent V/V of N_2 , calculated by deduction of the sum of impurities found when performing the test for impurities.

CHARACTERS

Colourless and odourless gas.

Solubility

At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in about 62 volumes of water and about 10 volumes of alcohol.

PRODUCTION

Oxygen

Maximum 5 ppm V/V , determined using an oxygen analyser with a detector scale ranging from 0 ppm V/V to 100 ppm V/V and equipped with an electrochemical cell.

The gas to be examined passes through a detection cell containing an aqueous solution of an electrolyte, generally potassium hydroxide. The presence of oxygen in the gas to be examined produces variation in the electric signal recorded at the outlet of the cell that is proportional to the oxygen content.

Calibrate the analyser according to the manufacturer's instructions. Pass the gas to be examined through the analyser using a suitable pressure regulator and airtight metal

Low-Oxygen Nitrogen

(Nitrogen, Low-Oxygen, Ph Eur monograph 1685)

N_2

28.01

Ph Eur

Ph Eur



D. H_2O : water.

C. O_2 : oxygen,

B. CO : carbon monoxide,

A. CO_2 : carbon dioxide,

Specified impurities A, B, C, D

IMPURITIES

complying with the legal regulations.

As a compressed gas or a liquid in appropriate containers

STORAGE

detector tube.

Maximum 67 ppm V/V , determined using a water vapour

Water vapour (2.1.6)

detector tube.

Maximum 5 ppm V/V , determined using a carbon monoxide

Carbon monoxide (2.1.6)

detector tube.

Maximum 300 ppm V/V , determined using a carbon dioxide

Carbon dioxide (2.1.6)

TESTS

of moistened red litmus paper R blue.

without heating, then for 15 min while heating the test tube to a red glow. After cooling, add 5 mL of dilute sodium hydroxide solution R. The evolving vapours change the colour

Nitrous Oxide

(Ph. Eur. monograph 0416)

N₂O 44.01

10024-97-2



Action and use

General anaesthetic; analgesic.

Nitrous oxide should be kept in approved metal cylinders which are painted blue and carry a label stating 'Nitrous Oxide'; in addition, 'Nitrous Oxide' or the symbol 'N₂O' should be stencilled in paint on the shoulder of the cylinder.

DEFINITION

Content

Minimum 98.0 per cent V/V of N₂O in the gaseous phase, when sampled at 15 °C.

CHARACTERS

Appearance

Colourless gas.

Solubility

At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in about 1.5 volumes of water.

PRODUCTION

Nitrous oxide is produced from ammonium nitrate by

thermic decomposition.

Examine the gaseous phase. If the test is performed on a cylinder, keep the cylinder at room temperature for at least 6 h before carrying out the tests. Keep the cylinder in the vertical position with the outlet valve uppermost.

Carbon dioxide

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas A mixture containing 300 ppm V/V of carbon

Column:

material: stainless steel;

size: $l = 3.5$ m, $\varnothing = 2$ mm;

stationary phase: ethylvinylbenzene-divinylbenzene copolymer R.

Carrier gas helium for chromatography R.

Flow rate 15 mL/min.

Temperature:

column: 40 °C;

detector: 90 °C.

Detection Thermal conductivity.

Injection Loop injector.

Adjust the injected volumes and operating conditions so that the height of the peak due to carbon dioxide in the chromatogram obtained with the reference gas is at least 35 per cent of the recorder. The test is not valid unless the chromatograms obtained show a clear separation of carbon dioxide from nitrous oxide.

Limit:

carbon dioxide: not more than the area of the corresponding peak in the chromatogram obtained with the reference gas (300 ppm V/V).

Carbon monoxide

Gas chromatography (2.2.28). When the test is carried out on a cylinder, use the first portion of gas to be withdrawn.

Gas to be examined The substance to be examined.

tubes and operating at the prescribed flow rates until constant readings are obtained.

Impurities

Gas chromatography (2.2.28).

Gas to be examined The substance to be examined.

Reference gas (a) Use ambient air.

Reference gas (b) Use nitrogen R1.

Column:

material: stainless steel;

size: $l = 2$ m, $\varnothing = 2$ mm,

stationary phase: appropriate molecular sieve for chromatography (0.5 nm).

Carrier gas helium for chromatography R.

Flow rate 40 mL/min.

Temperature:

column: 50 °C,

detector: 130 °C.

Detection Thermal conductivity.

System suitability: reference gas (a): adjust the injected

volumes and operating conditions so that the height of the peak due to nitrogen in the chromatogram obtained is at least 35 per cent of the full scale of the recorder.

the chromatogram obtained shows a clear separation of oxygen and nitrogen.

Limit:

total: not more than 0.5 per cent of the sum of the areas of all the peaks (0.5 per cent V/V).

IDENTIFICATION

First identification A

Second identification B, C

A. Examine the chromatograms obtained in the test for

impurities (see Production).

Results The principal peak in the chromatogram obtained

with the gas to be examined is similar in retention time to

the principal peak in the chromatogram obtained with

reference gas (b).

B. In a 250 mL conical flask replace the air by the gas to be

examined. Place a burning or glowing splinter of wood in the

flask. The splinter is extinguished.

C. In a suitable test tube, place 0.1 g of magnesium R in

turnings. Close the tube with a two-hole stopper fitted with a

glass tube reaching about 1 cm above the turnings. Pass the

gas to be examined through the glass tube for 1 min without

heating, then for 15 min while heating the test tube to a red

glow. After cooling, add 5 mL of dilute sodium hydroxide

solution R. The evolving vapours turn the colour of moistened

red litmus paper R blue.

STORAGE

Where the gas has to be stored, store as a compressed gas or a liquid in appropriate containers complying with the legal

regulations.

IMPURITIES

A. O₂: oxygen,

B. Ar: argon.

Ph Eur

Reference gas A mixture containing 5 ppm V/V of carbon

monoxide R in nitrous oxide R.

Column:

— material: stainless steel;

— size: $l = 2$ m, $\varnothing = 4$ mm;

— stationary phase: suitable molecular sieve for

chromatography (0.5 nm).

Carrier gas helium for chromatography R.

Flow rate 60 mL/min.

Temperature:

— column: 50 °C;

— injection port and detector: 130 °C.

Detection Flame ionisation with methaniser.

Injection Loop injector.

Adjust the injected volumes and the operating conditions so

that the height of the peak due to carbon monoxide in the

chromatogram obtained with the reference gas is at least

35 per cent of the full scale of the recorder.

Limit:

— carbon monoxide: not more than the area of the

corresponding peak in the chromatogram obtained with

the reference gas (5 ppm V/V).

Nitrogen monoxide and nitrogen dioxide

Maximum 2 ppm V/V in total in the gaseous and liquid

phases, determined using a chemiluminescence analyser

(2.5.26).

Gas to be examined The substance to be examined.

Reference gas (a) Nitrous oxide R.

Reference gas (b) A mixture containing 2 ppm V/V of nitrogen

monoxide R in nitrogen R1.

Calibrate the apparatus and set the sensitivity using reference

gases (a) and (b). Measure the content of nitrogen monoxide

and nitrogen dioxide, separately examining the samples

collected from the gaseous phase and the liquid phase of the

gas to be examined.

Multiply the result obtained by the quenching correction

factor in order to correct the quenching effect on the analyser

response caused by the nitrous oxide matrix effect.

The quenching correction factor is determined by applying a

known reference mixture of nitrogen monoxide in nitrous

oxide and comparing the actual content with the content

indicated by the analyser which has been calibrated with an

NO/N₂ reference mixture.

Quenching correction factor = $\frac{\text{actual nitrogen monoxide content}}{\text{indicated nitrogen monoxide content}}$

Water

Maximum 67 ppm V/V, determined using an electrolytic

hydrometer (2.5.28).

Assay

Infrared analyser (2.5.35).

Gas to be examined The substance to be examined. It must be

filtered to avoid stray light phenomena.

Reference gas (a) Nitrous oxide R.

Reference gas (b) A mixture containing 5.0 per cent V/V of

nitrogen R1 and 95.0 per cent V/V of nitrous oxide R.

Calibrate the apparatus and set the sensitivity using reference

gases (a) and (b). Measure the content of nitrous oxide in

the gas to be examined.

IDENTIFICATION

First identification: A.

(E₂)-N-[2-[[[2-(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]-N'-methyl-2-nitroethene-1,1-diamine.

DEFINITION

Ph Eur

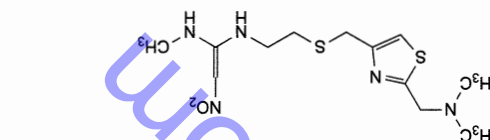
Nizatidine Infusion

Preparation

Histamine H₂ receptor antagonists; treatment of peptic ulcer.

Action and use

C₁₂H₂₁N₅O₂S₂ 331.5 76963-41-2



(Ph. Eur. monograph 1453)

Nizatidine

Ph Eur

Specified impurities A, B, C, D, E
A. CO₂: carbon dioxide,
B. CO: carbon monoxide,
C. NO: nitrogen monoxide,
D. NO₂: nitrogen dioxide,
E. H₂O: water.

IMPURITIES

Store liquefied under pressure in suitable containers complying with the legal regulations. The taps and valves are not greased or oiled.

STORAGE

Maximum 67 ppm V/V, determined using a water vapour detector tube (2.1.6).

Water vapour

Maximum 2 ppm V/V, determined using a nitrogen monoxide and nitrogen dioxide detector tube (2.1.6).

Nitrogen monoxide and nitrogen dioxide

use the first portion of the gas to be withdrawn.
detector tube (2.1.6). When the test is carried out on a cylinder, Maximum 5 ppm V/V, determined using a carbon monoxide

Carbon monoxide

Maximum 300 ppm V/V, determined using a carbon dioxide detector tube (2.1.6).

Carbon dioxide

If the test is performed on a cylinder, keep the cylinder of the substance to be examined at room temperature for at least 6 h before carrying out the tests. Keep the cylinder in the vertical position with the outlet valve uppermost.

Examine the gaseous phase.

TESTS

pyrogallol solution R. A brown colour does not develop.

C. Introduce the substance to be examined into alkaline

examined. The splinter bursts into flame.

B. Place a glowing splinter of wood in the substance to be

A. It complies with the limits of the assay.

Second identification B, C

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Almost white or slightly brownish, crystalline powder.

Solubility

Sparsingly soluble in water, soluble in methanol.

IDENTIFICATION

First identification C

Second identification A, B, D

A. Melting point (2.2.14): 131 °C to 134 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25):

Test solution Dissolve 0.10 g in methanol R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with methanol R.

Spectral range 220–350 nm.

Absorption maxima At 242 nm and 325 nm.

Absorbance ratio $A_{325}/A_{242} = 2.2$ to 2.5.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison nizatidine CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of nizatidine CRS in methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 50 mg of nizatidine CRS and 50 mg of ranitidine hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.

Mobile phase water R, concentrated ammonia R₁, 2-propanol R, ethyl acetate R (4:8:30:50 V/V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots are clearly visible. Examine in daylight.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₃ (2.2.2, Method II).

Dissolve 0.2 g in a 10 g/L solution of hydrochloric acid R and dilute to 20 mL with the same solution.

pH (2.2.3)

8.5 to 10.0.

Dissolve 0.2 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase B, mobile phase A (15:85 V/V).

Test solution (a) Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dissolve 15.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 15.0 mg of nizatidine CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of the substance to be examined and 0.5 mg of nizatidine impurity F CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5 mg of 2-(dimethylamino)thioacetamide hydrochloride R (impurity H hydrochloride) in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 20.0 mL with the solvent mixture. Use 1.0 mL of this solution to dissolve 5 mg of nizatidine for system suitability CRS (containing impurities A, B, C, D, G, J and K).

Column:

size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase:

mobile phase A: dissolve 5.9 g of ammonium acetate R in 760 mL of water R, add 1 mL of diethylamine R, and adjust to pH 7.5 with acetic acid R;

mobile phase B: methanol R;

Time (min)

Mobile phase A (per cent V/V)

Mobile phase B (per cent V/V)

0 - 3

3 - 20

20 - 45

50

85 → 50

15 → 50

50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (c) and (d).

Identification of impurities Use the chromatogram supplied with nizatidine for system suitability CRS and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, G, H, J and K; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity F.

Relative retention With reference to nizatidine (retention time = about 18 min): impurity A = about 0.19;

impurity K = about 0.21; impurity H = about 0.5;

impurity B = about 0.6; impurity C = about 0.66;

impurity J = about 0.7; impurity D = about 0.75;

impurity F = about 1.03; impurity G = about 1.5.

System suitability:

— resolution: minimum 2.0 between the peaks due to nizatidine and impurity F in the chromatogram obtained with reference solution (c); minimum 1.5 between the peaks due to impurities A and K in the chromatogram obtained with reference solution (d).

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the

corresponding correction factor: impurity B = 1.7; impurity D = 2.3; impurity H = 0.5; — *impurities A, B, C, D, F, G, H, J, K*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent); — *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent); — *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent); — *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)
Maximum 20 ppm.
Solvent methanol R.

0.5 g complies with test H. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase B, mobile phase A (35:65 V/V).
Injection Test solution (b) and reference solution (b).

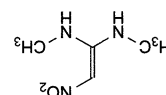
Retention time Nizatidine = 9 min.

Calculate the percentage content of $C_{12}H_{21}N_5O_2S_2$ taking into account the assigned content of nizatidine CRS.

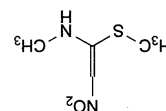
IMPURITIES

Specified impurities A, B, C, D, F, G, H, J, K

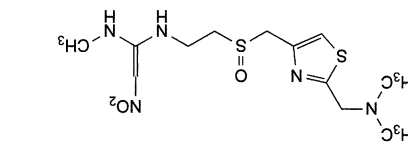
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, I.



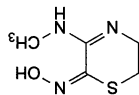
A. N,N'-dimethyl-2-nitroethene-1,1-diamine,



B. (E)-N-methyl-1-(methylsulfonyl)-2-nitroethen-1-amine,

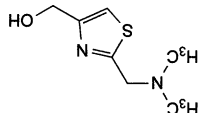


C. (E)-N-[2-[[[2-[(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]-N'-methyl-2-nitroethene-1,1-diamine,

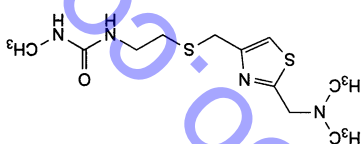


K. 3-(methylamino)-5,6-dihydro-2H-1,4-thiazin-2-one oxime.

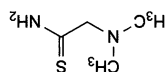
J. 2-[[[2-[(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]-N'-methylurea,



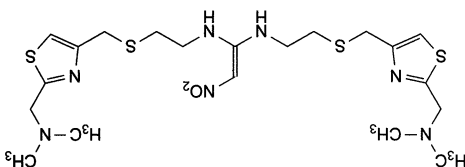
I. N-[2-[[[2-[(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]-N'-methylurea,



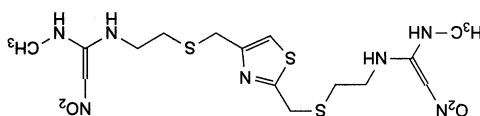
H. 2-(dimethylamino)thioacetamide,



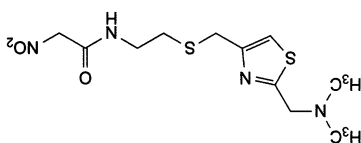
G. N,N'-bis[2-[[[2-[(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]-2-nitroethene-1,1-diamine,



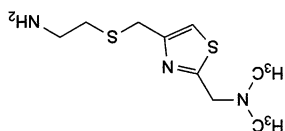
F. (E)-N-methyl-N'-[2-[[[2-[(E)-1-(methylamino)-2-nitroethenyl]amino]ethyl]thiazol-2-yl]methyl]sulfonyl]ethyl]-2-nitroethene-1,1-diamine,



E. N-[2-[[[2-[(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]-2-nitroethene-1,1-diamine,

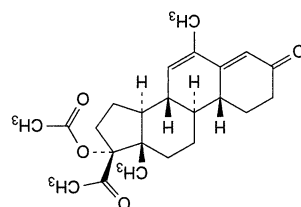


D. 2-[[[2-[(dimethylamino)methyl]thiazol-4-yl]methyl]sulfonyl]ethyl]ethanamine,



Nomegestrol Acetate

(Ph. Eur. monograph 1551)

C₂₃H₃₀O₄ 370.5 58652-20-3

Action and use

Progestogen.

Ph Eur

DEFINITION

6-Methyl-3,20-dioxo-19-norpregna-4,6-dien-17-yl acetate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone.

soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison nomegestrol acetate CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Dissolve 1.0 g in methylene chloride R and dilute to 10 mL

with the same solvent.

Specific optical rotation (2.2.7)

–60.0 to –64.0 (dried substance).

Dissolve 0.500 g in anhydrous ethanol R and dilute to

25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be

examined in methanol R and dilute to 50.0 mL with the same

solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to

200.0 mL with the mobile phase.

Reference solution (b) Dissolve 25.0 mg of nomegestrol acetate

impurity A CRS in methanol R and dilute to 50.0 mL with the

same solvent.

Reference solution (c) Dissolve 25.0 mg of nomegestrol

acetate CRS in 20 mL of methanol R, add 0.25 mL of

reference solution (b) and dilute to 50.0 mL with the mobile

phase.

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 µm).

Mobile phase acetonitrile R, methanol R, water R
(24:38:38 V/V/V).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 245 nm and at 290 nm.

Injection 10 µL.

Run time 1.5 times the retention time of nomegestrol acetate.

Retention time At 245 nm: nomegestrol acetate = about

17 min; impurity A = about 18.5 min.

System suitability Reference solution (c) at 245 nm:

— peak-to-valley ratio: minimum 5, where H_p = height above

the baseline of the peak due to impurity A and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

nomegestrol acetate.

Limits:

— impurity A at 245 nm: not more than 0.4 times the area of

the corresponding peak in the chromatogram obtained

with reference solution (c) (0.2 per cent);

— unspecified impurities at 245 nm: for each impurity, not

more than 0.2 times the area of the peak due to

impurity A in the chromatogram obtained with reference

solution (c) (0.10 per cent);

— unspecified impurities at 290 nm: for each impurity, not

more than 0.2 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent);

— sum of impurities other than A at 290 nm and 245 nm:

maximum 0.3 per cent;

— disregard limit at 245 nm: 0.1 times the area of the peak

due to impurity A in the chromatogram obtained with

reference solution (c) (0.05 per cent);

— disregard limit at 290 nm: 0.04 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.02 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

ASSAY

Dissolve 50.0 mg in anhydrous ethanol R and dilute to

100.0 mL with the same solvent. Dilute 2.0 mL of this

solution to 100.0 mL with anhydrous ethanol R. Measure the

absorbance (2.2.25) at the absorption maximum at 287 nm.

Calculate the content of C₂₃H₃₀O₄ taking the specific

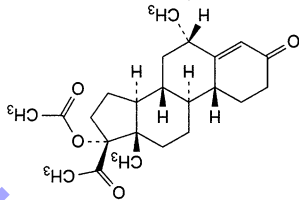
absorbance to be 685.

STORAGE

Protected from light.

IMPURITIES

Specified impurities: A.



A. 6α-methyl-3,20-dioxo-19-norpregn-4-en-17-yl acetate.

Ph Eur

Nonoxinol 9

(Ph. Eur. monograph 1454)



Action and use
Spermatoicide.

Ph Eur

DEFINITION

Mixture consisting mainly of monononylphenyl ethers of macrogols corresponding to the formula: $C_9H_{19}C_6H_4-[OCH_2-CH_2]_n-OH$ where the average value of n is 9. It may contain free macrogols.

CHARACTERS

Appearance
Clear, colourless or light yellow, viscous liquid.

Solubility

Miscible with water, with ethanol (96 per cent) and with vegetable oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison nonoxinol 9 CRS.

Preparation Film between sodium chloride R plates.

B. Cloud point (see Tests).

TESTS

Acidity or alkalinity

Boil 1.0 g with 20 mL of carbon dioxide-free water R for 1 min, with constant stirring. Cool and filter. To 10 mL of the filtrate, add 0.05 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Hydroxyl value (2.5.3, Method A)

84 to 94.

Cloud point

52 °C to 58 °C.

Dissolve 1.0 g in 99 g of water R. Transfer about 30 mL of this solution into a test-tube, heat on a water-bath and stir continuously until the solution becomes cloudy. Remove the test-tube from the water-bath (ensuring that the temperature does not increase to more than 2 °C) and continue to stir. The cloud point is the temperature at which the solution becomes sufficiently clear that the entire thermometer bulb is plainly seen.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in distilled water R and dilute to 20.0 mL with the same solvent. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Total ash (2.4.16)

Maximum 0.4 per cent, determined on 1.0 g.

STORAGE

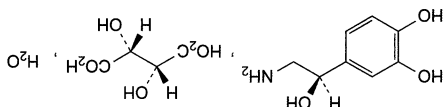
In an airtight container.

Ph Eur

Noradrenaline Acid Tartrate /



(Noradrenaline Tartrate, Ph Eur monograph 0285)



$C_{12}H_{17}NO_9 \cdot H_2O$ 337.3

108341-18-0

Action and use

Alpha-adrenoceptor agonist.

Preparation

Noradrenaline Injection/Norepinephrine Injection

Ph Eur

DEFINITION

(1R)-2-Amino-1-(3,4-dihydroxyphenyl)ethanol hydrogen (2R,3R)-2,3-dihydroxybutanedioate monohydrate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 2 g in 20 mL of a 5 g/L solution of sodium metabisulfite R and make alkaline by addition of ammonia R. Keep in iced water for 1 h and filter. Reserve the filtrate for identification test C. Wash the precipitate with 3 quantities, each of 2 mL, of water R, then with 5 mL of ethanol (96 per cent) R and finally with 5 mL of methylene chloride R and dry *in vacuo* for 3 h. The specific optical rotation (2.2.7) of the precipitate (noradrenaline base) is -48 to -44, determined using a 20.0 g/L solution in 0.5 M hydrochloric acid.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of noradrenaline base prepared as described in identification test A.

Comparison Use noradrenaline base prepared as described in identification test A from a suitable amount of noradrenaline tartrate CRS.

C. 0.2 mL of the filtrate obtained in identification test A gives reaction (b) of tartrates (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II). Dissolve 0.2 g in water R and dilute to 10 mL with the same solvent. Examine the solution immediately.

Related substances

Liquid chromatography (2.2.29). Protect the solutions from air. Remove oxygen from the mobile phases with nitrogen R immediately before use. Fill up the flasks.

Test solution Dissolve 0.20 g of the substance to be examined in mobile phase A and dilute to 50 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 10 mg of the substance to be examined in 5 mL of 0.1 M hydrochloric acid. To 1 mL of this solution add 0.1 mL of strong hydrogen peroxide solution R and expose to UV light at 254 nm for 90 min. Dilute to 10 mL with mobile phase A. The degradation of noradrenaline produces 2 peaks, one with a relative retention of about 1.2 (unidentified compound) and the other with a relative retention of about 1.5 (impurity B). Use this solution to identify the peak due to impurity B.

Reference solution (c) Dissolve 7.5 mg of noradrenaline impurity D CRS and 5 mg of noradrenaline impurity E CRS in mobile phase A and dilute to 100 mL with mobile phase A.

Reference solution (d) Dissolve 5 mg of noradrenaline impurity F CRS in mobile phase A and dilute to 10 mL with mobile phase A. To 1 mL of this solution, add 1 mL of reference solution (c) and dilute to 20 mL with mobile phase A.

Column:

— **stationary phase:** monolithic octadecylsilyl silica gel for chromatography R_s

— **temperature:** 25 °C.

Mobile phase:

— **mobile phase A:** dissolve 0.50 g of sodium heptanesulfonate R in water for chromatography R and dilute to 1000 mL with the same solvent; adjust to pH 2.2 with phosphoric acid R_s

— **mobile phase B:** dissolve 0.25 g of sodium heptanesulfonate R and dilute to 500 mL with the same solvent; add 500 mL of acetonitrile for chromatography R and adjust the apparent pH to 2.4 with phosphoric acid R_s

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Flow rate (mL/min)
0 - 2.0	98	2	1.5
2.0 - 17.0	98 → 70	2 → 30	1.5
17.0 - 24.0	70 → 50	30 → 50	1.5
24.0 - 24.1	50 → 0	50 → 100	1.5 → 4.0
24.1 - 28.0	0	100	4.0
28.0 - 28.1	0 → 98	100 → 2	4.0
28.1 - 30.0	98	2	4.0 → 1.5

Detection Spectrophotometer at 280 nm, except for impurity F: spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (a), (b) and (d).

Relative retention With reference to noradrenaline (retention time = about 3 min): impurity B = about 1.5; impurity D = about 2.8; impurity E = about 3.0; impurity F = about 6.9.

System suitability Reference solution (d):

— **resolution:** minimum 1.5 between the peaks due to impurities D and E.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.3; impurity E = 0.3; impurity F = 1.5;

— **impurity F at 254 nm:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **impurities B, D, E at 280 nm:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **unspecified impurities at 280 nm:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **sum of the impurities at 280 nm and impurity F at 254 nm:** maximum 0.3 per cent;

— **disregard limit at 280 nm:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (2.5.12)

4.5 per cent to 5.8 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of anhydrous acetic acid R, heating gently if necessary. Titrate with 0.1 M perchloric acid using 0.1 mL of crystal violet solution R as indicator, until a bluish-green colour is obtained.

1 mL of 0.1 M perchloric acid is equivalent to 31.93 mg of C₁₂H₁₇NO₉.

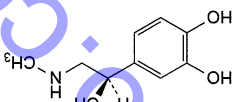
STORAGE

In an airtight container or preferably in a sealed tube under vacuum or under an inert gas, protected from light.

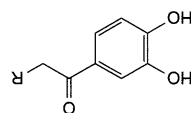
IMPURITIES

Specified impurities B, D, E, F

Other detectable impurities (the following substances would, if the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, G.

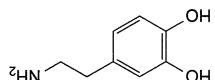


A. 4-[(1R)-1-hydroxy-2-(methylamino)ethyl]benzene-1,2-diol (adrenaline),



B. R = NH₂: 2-amino-1-(3,4-dihydroxyphenyl)ethanone (noradrenalone),

E. R = Cl: 2-chloro-1-(3,4-dihydroxyphenyl)ethanone,



C. 4-(2-aminoethyl)benzene-1,2-diol (dopamine),

Comparison Use noradrenaline base prepared as above from a suitable amount of noradrenaline tartrate CRS. C. 0.2 mL of solution S (see Tests) gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S Dissolve 0.500 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than a mixture of 0.2 mL of blue primary solution, 0.4 mL of yellow primary solution, 0.4 mL of red primary solution and 9 mL of a 13.7 per cent V/V solution of dilute hydrochloric acid R (2.2.2, Method II).

Dissolve 0.2 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent. Examine the solution immediately.

pH (2.2.3)

3.5 to 4.5 for solution S.

Specific optical rotation (2.2.7)

−37 to −41 (anhydrous substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29). Protect the solutions from air. Remove oxygen from the mobile phases with nitrogen R immediately before use. Fill up the flasks.

Test solution Dissolve 0.125 g of the substance to be examined in mobile phase A and dilute to 50 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 10 mg of the substance to be examined in 5 mL of 0.1 M hydrochloric acid. To 1 mL of this solution add 0.1 mL of strong hydrogen peroxide solution R and expose to UV light at 254 nm for 90 min. Dilute to 10 mL with mobile phase A. The degradation of noradrenaline produces 2 peaks, one with a relative retention of about 1.2 (unidentified compound) and the other with a relative retention of about 1.5 (impurity B). Use this solution to identify the peak due to impurity B.

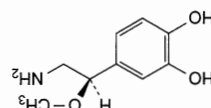
Reference solution (c) Dissolve 7.5 mg of noradrenaline impurity D CRS and 5 mg of noradrenaline impurity E CRS in mobile phase A and dilute to 100 mL with mobile phase A. **Reference solution (d)** Dissolve 5 mg of noradrenaline impurity F CRS in mobile phase A and dilute to 10 mL with mobile phase A. To 1 mL of this solution, add 1 mL of reference solution (c) and dilute to 20 mL with mobile phase A.

Column:

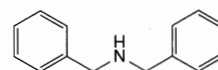
— **size:** $l = 0.10$ m, $\varnothing = 4.6$ mm;
— **stationary phase:** monolithic octadecylsilyl silica gel for chromatography R_s
— **temperature:** 25 °C.

Mobile phase:

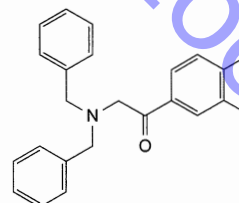
— **mobile phase A:** dissolve 0.50 g of sodium heptanesulfonate R in water for chromatography R and dilute to 1000 mL with the same solvent; adjust to pH 2.2 with phosphoric acid R_s
— **mobile phase B:** dissolve 0.25 g of sodium heptanesulfonate R in water for chromatography R and dilute to 500 mL with the same solvent; add 500 mL of acetonitrile for



D. 4[(1R)-2-amino-1-methoxyethyl]benzene-1,2-diol (noradrenaline methyl ether),

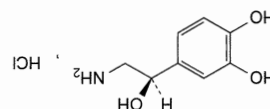


F. N-benzyl-1-phenylmethanamine,



G. 2-(di(2-aminophenyl)ethyl)ethanone.

Noradrenaline Hydrochloride / Norepinephrine Hydrochloride
(Noradrenaline Hydrochloride,
Ph Eur monograph 0732)



C₈H₁₂ClNO₃ 205.6 329-56-6

Action and use
Alpha-adrenoceptor agonist.

DEFINITION

(1R)-2-Amino-1-(3,4-dihydroxyphenyl)ethanol hydrochloride.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or brownish-white, crystalline powder.

Solubility

Very soluble in water, slightly soluble in ethanol

(96 per cent).

It becomes coloured on exposure to air and light.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of noradrenaline base prepared as follows. Dissolve 2 g in 20 mL of a 5 g/L solution of sodium metabisulfite R and make alkaline by addition of ammonia R. Keep in iced water for 1 h and filter. Wash the precipitate with 3 quantities, each of 2 mL, of water R, then with 5 mL of ethanol (96 per cent) R and finally with 5 mL of methylene chloride R and dry in vacuo for 3 h.

chromatography *R* and adjust the apparent pH to 2.4 with phosphoric acid *R*;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Flow rate (mL/min)
0 - 2.0	98	2	1.5
2.0 - 17.0	98	2	1.5
17.0 - 24.0	70	30	1.5
24.0 - 24.1	50	50	1.5
24.1 - 28.0	50	50	1.5
28.0 - 28.1	0	100	4.0
28.1 - 30.0	98	2	4.0
30.0 - 40.0	98	2	4.0

Detection Spectrophotometer at 280 nm, except for impurity F: spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (a), (b) and (d).

Relative retention With reference to noradrenaline (retention time = about 3 min): impurity B = about 1.5;

impurity D = about 2.8; impurity E = about 3.0;

impurity F = about 6.9.

System suitability Reference solution (d):

— resolution: minimum 1.5 between the peaks due to

impurities D and E.

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the

corresponding correction factor: impurity B = 0.3;

impurity E = 0.3; impurity F = 1.5;

— impurity D at 280 nm: not more than 5 times the area of

reference solution (a) (0.5 per cent);

principal peak in the chromatogram obtained with

impurity F at 254 nm: not more than the area of the

reference solution (a) (0.1 per cent);

impurities B, E at 280 nm: for each impurity, not more

than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— unspecified impurities at 280 nm: for each impurity, not

more than the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent);

— sum of impurities other than D at 280 nm: not more than

twice the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

— sum of the impurities at 280 nm and impurity F at 254 nm:

maximum 0.7 per cent;

— disregard limit at 280 nm: 0.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.05 per cent).

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.180 g in 50 mL of acetic anhydride *R* and add

10 mL of anhydrous formic acid *R*. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2.20).

G. Control of impurities in substances for pharmaceutical use): A, C, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these

by the general monograph Substances for pharmaceutical use acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of Other detectable impurities (the following substances would, if

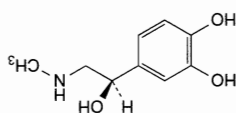
Specified impurities B, D, E, F

IMPURITIES

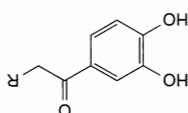
In an airtight container, or preferably in a sealed tube under vacuum or under an inert gas, protected from light.

STORAGE

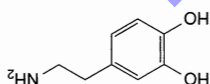
1 mL of 0.1 M perchloric acid is equivalent to 20.56 mg of C₈H₁₂ClNO₃.



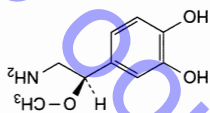
A. 4-[(1*R*)-1-hydroxy-2-(methylamino)ethyl]benzene-1,2-diol (adrenaline),



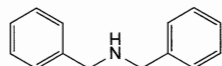
B. R = NH₂; 2-amino-1-(3,4-dihydroxyphenyl)ethanone (noradrenalone),
E. R = Cl; 2-chloro-1-(3,4-dihydroxyphenyl)ethanone,



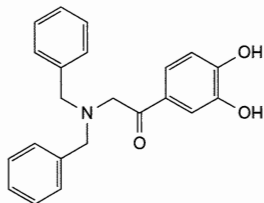
C. 4-(2-aminoethyl)benzene-1,2-diol (dopamine),



D. 4-[(1*R*)-2-amino-1-methoxyethyl]benzene-1,2-diol (noradrenaline methyl ether),



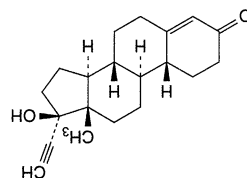
F. *N*-benzyl-1-phenylimethanamine,



G. 2-(dibenzylamino)-1-(3,4-dihydroxyphenyl)ethanone.

Norethisterone

(Ph. Eur. monograph 0234)

 $C_{20}H_{26}O_2$ 298.4

68-22-4

Action and use

Progestogen.

Preparations

Norethisterone Tablets

Estradiol and Norethisterone Tablets

DEFINITION

17-Hydroxy-19-nor-17 α -pregn-4-en-20-yn-3-one.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Practically insoluble in water, soluble in methylene chloride, sparingly soluble in acetone and in anhydrous ethanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison norethisterone CRS.

TESTS

Specific optical rotation (2.2.7)

–32.0 to –37.0 (dried substance).

Dissolve 0.250 g in acetone R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in a mixture of 40 volumes of water R and 60 volumes of acetonitrile R1 and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 5 mg of norethisterone for system suitability CRS (containing impurities A, B, C, D, E, F, G and H) in a mixture of 40 volumes of water R and 60 volumes of acetonitrile R1 and dilute to 2.0 mL with the same mixture of solvents.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of 40 volumes of water R and 60 volumes of acetonitrile R1. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of 40 volumes of water R and 60 volumes of acetonitrile R1.

Column:

size: $l = 0.15$ m, $\varnothing = 4.6$ mm,stationary phase: spherical end-capped octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

mobile phase A: water R;

mobile phase B: acetonitrile R1;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 20	63	37
20 - 25	63 \rightarrow 20	37 \rightarrow 80
25 - 35	20	80

Flow rate 1.0 mL/min.

Detection Variable wavelength spectrophotometer capable of operating at 254 nm and at 210 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram obtained with reference solution (a) and the chromatogram supplied with norethisterone CRS to identify the peaks due to the impurities A, B, C, D, E, F, G and H.

Relative retention at 254 nm With reference to norethisterone (retention time = about 10 min): impurity H = about 0.3; impurity A = about 0.8; impurity B = about 0.9; impurity G = about 1.5; impurity C (at 210 nm) = about 1.6; impurity D (at 210 nm) = about 1.7; impurity E = about 2.3; impurity F = about 2.4.

System suitability Reference solution (a) at 254 nm: resolution: baseline separation between the peaks due to impurity B and norethisterone;

peak-to-valley ratio: minimum 1.2, where H_p = height above the baseline of the peak due to impurity A and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity B.

Limits Spectrophotometer at 254 nm: correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 2.5;

impurity B = 0.7; impurity F = 1.4; impurity H = 1.7; impurities E, G, H: for each impurity, not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent); impurities A, B, F: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Limits Spectrophotometer at 210 nm:

the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

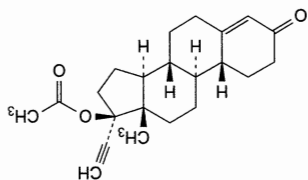
Dissolve 0.200 g in 40 mL of tetrahydrofuran R. Add 10 mL of a 100 g/L solution of silver nitrate R and titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). Rinse the electrode with acetone R after each titration.

1 mL of 0.1 M sodium hydroxide is equivalent to 29.84 mg of $C_{20}H_{26}O_2$.



Norethisterone Acetate

(Ph. Eur. monograph 0850)



$C_{22}H_{28}O_3$ 340.5 51-98-9

Action and use

Progestogen.

Preparation

Estradiol and Norethisterone Acetate Tablets

Ph Eur

DEFINITION

3-Oxo-19-nor-17 α -pregn-4-en-20-yn-17-yl acetate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison norethisterone acetate CRS.

If the spectra show differences, dissolve the substance to be examined and the reference substance separately in methylene chloride R, evaporate to dryness on a water-bath and record new spectra using the residues.

TESTS

Specific optical rotation (2.2.7)

Dissolve 0.500 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dissolve the contents of a vial of norethisterone acetate for system suitability CRS (containing impurities B, C, D, E, F, G and H) in 1.0 mL of mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

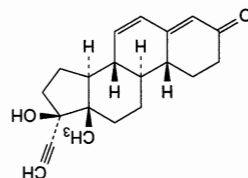
stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

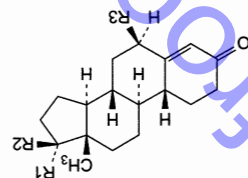
mobile phase A: water for chromatography R, acetonitrile R1 (40:60 V/V);

IMPURITIES

Specified impurities A, B, C, D, E, F, G, H



A. 17-hydroxy-19-nor-17 α -pregn-4,6-dien-20-yn-3-one,



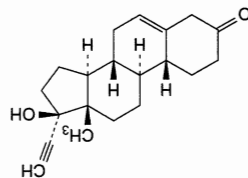
B. R1 + R2 = O, R3 = H; est-4-ene-3,17-dione

(norandrostenedione),

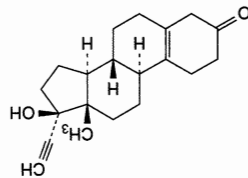
G. R1 = OH, R2 = C \equiv CH, R3 = H: 17-hydroxy-19-

norpregn-4-en-20-yn-3-one (17-*epi*-norethisterone),

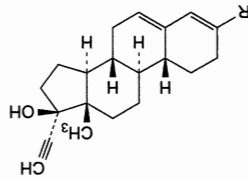
H. R1 = C \equiv CH, R2 = OH: 6 β ,17-dihydroxy-19-nor-17 α -pregn-4-en-20-yn-3-one (6 β -hydroxynorethisterone),



C. 17-hydroxy-19-nor-17 α -pregn-5-en-20-yn-3-one,



D. 17-hydroxy-19-nor-17 α -pregn-5(10)-en-20-yn-3-one,



E. R = C \equiv CH:

3-ethynyl-19-nor-17 α -pregn-3,5-dien-20-yn-17-ol,

F. R = O-C $_2$ H $_5$:

3-ethoxy-19-nor-17 α -pregn-3,5-dien-20-yn-17-ol.

Ph Eur

— mobile phase B: water for chromatography R, acetonitrile R1 (10:90 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 17	100	0
17 - 20	100 → 0	0 → 100
20 - 39	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm and, for impurities B and C, at 210 nm.

Injection 20 µL.

Identification of impurities Use the chromatograms supplied

with norethisterone acetate for system suitability CRS and the

chromatograms obtained with reference solution (a) to

identify the peaks due to impurities B, C, D+G, E, F and H.

Relative retention With reference to norethisterone acetate

(retention time = about 12 min): impurity F = about 0.4;

impurities D and G = about 0.6; impurity E = about 0.8;

impurity C = about 1.5; impurity B = about 1.6;

impurity H = about 2.8.

System suitability Reference solution (a) at 210 nm:

— peak-to-valley ratio: minimum 2.5, where H_p = height

above the baseline of the peak due to impurity C and

H_p = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

impurity B.

Calculation of percentage contents:

— for impurity C, multiply the peak area by the correction

factor 1.3;

— for impurity F, multiply the peak area by the correction

factor 1.7;

— for each impurity, use the concentration of norethisterone

acetate in reference solution (b).

Limits:

— impurities B, C at 210 nm: for each impurity, maximum

0.3 per cent;

— impurities F, H: for each impurity, maximum 0.3 per cent;

— impurity E: maximum 0.2 per cent;

— sum of impurities D and G: maximum 0.2 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 1.0 per cent;

— reporting threshold: 0.05 per cent.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

ASSAY

Dissolve 0.200 g in 40 mL of tetrahydrofuran R. Add 10 mL

of a 100 g/L solution of silver nitrate R and titrate with 0.1 M

sodium hydroxide, determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M sodium hydroxide is equivalent to 34.05 mg of

$C_{22}H_{28}O_3$.

IMPURITIES

Specified impurities B, C, D, E, F, G, H

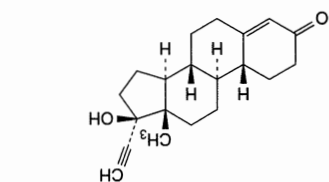
Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

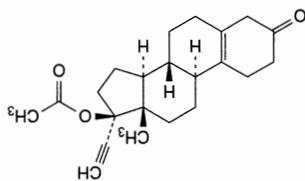
the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

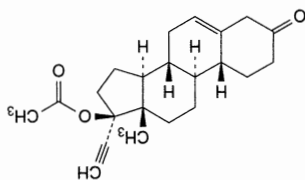
by the general monograph Substances for pharmaceutical use



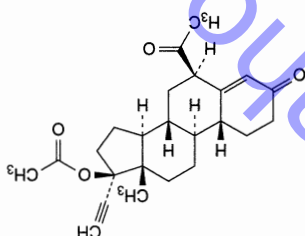
A. 17-hydroxy-19-nor-17 α -pregn-4-en-20-yn-3-one (norethisterone),



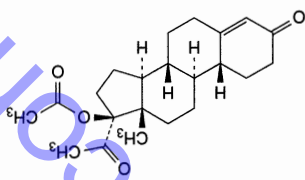
B. 3-oxo-19-nor-17 α -pregn-5(10)-en-20-yn-17-yl acetate,



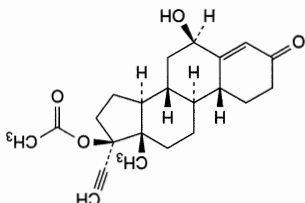
C. 3-oxo-19-nor-17 α -pregn-5-en-20-yn-17-yl acetate,



D. 6 β -acetyl-3-oxo-19-nor-17 α -pregn-4-en-20-yn-17-yl acetate,



E. 3,20-dioxo-19-nor-17 α -pregn-4-en-17-yl acetate,



F. 6 β -hydroxy-3-oxo-19-nor-17 α -pregn-4-en-20-yn-17-yl acetate,

(2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, I, J.

CHARACTERS

Appearance

White or pale yellow, hygroscopic, photosensitive, crystalline powder.

Solubility

Very slightly soluble in water, slightly soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison norfloxacin CRS.

TESTS

Appearance of solution

Dissolve 0.5 g in a previously filtered 4 g/L solution of sodium hydroxide R in methanol R and dilute to 50 mL with the same solution. The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

Related substances

Liquid chromatography (2.2.29).
Solution A Mix 5 volumes of acetonitrile R and 95 volumes of water R previously adjusted to pH 2.0 with phosphoric acid R. Test solution Dissolve 20 mg of the substance to be examined in 25 mL of solution A. Sonicate for 5 min and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (a) Dissolve 4 mg of norfloxacin for system suitability CRS (containing impurities A, E and H) in 5 mL of solution A. Sonicate for 5 min and dilute to 10 mL with solution A.
Reference solution (b) Dissolve 4 mg of norfloxacin for system suitability CRS (containing impurities A, E and H) in 5 mL of solution A. Sonicate for 5 min and dilute to 10 mL with solution A.
Reference solution (c) Dissolve 4 mg of norfloxacin for peak identification CRS (containing impurity K) in 5 mL of solution A. Sonicate for 5 min and dilute to 10 mL with solution A.

Column:

size: $l = 0.25$ m, $\phi = 4.6$ mm;
stationary phase: end-capped hexadecylammonium silica gel for chromatography R (5 μ m);
temperature: 60 °C.
Mobile phase:
mobile phase A: water R adjusted to pH 2.0 with phosphoric acid R;
mobile phase B: acetonitrile R;
Mobile phase A (per cent V/V) 95
Mobile phase B (per cent V/V) 5

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	95	5
5 - 7	95 \rightarrow 93	5 \rightarrow 7
7 - 10	93 \rightarrow 87	7 \rightarrow 13
10 - 15	87 \rightarrow 47	13 \rightarrow 53
15 - 20	47 \rightarrow 10	53 \rightarrow 90

Flow rate 1.4 mL/min.

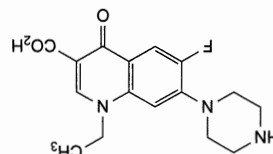
Detection Spectrophotometer at 265 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied with norfloxacin for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, E and H. Use the chromatogram supplied with norfloxacin for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peak due to impurity K.

Norfloxacin

(Ph. Eur. monograph 1248)



$C_{16}H_{18}FN_3O_3$

319.3

70458-96-7

Norfloxacin Tablets

Norfloxacin Eye Drops

Preparations

Fluoroquinolone antibacterial.

Action and use

Content

99.0 per cent to 101.0 per cent (dried substance).

DEFINITION

1-Ethyl-6-fluoro-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.

Relative retention With reference to norfloxacin (retention time = about 11 min): impurity K = about 0.6; impurity E = about 0.97; impurity A = about 1.5; impurity H = about 1.6.

System suitability: reference solution (b):
— resolution: minimum 3.0 between the peaks due to impurities A and H;

— H_p = height above the baseline of the lowest point of the curve separating this peak from the peak due to norfloxacin.

Limits:

— impurities E, K: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

— unspecified impurities: for each impurity, not more than area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 15 ppm.

2.0 g complies with test D. Prepare the reference solution using 3 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying under high vacuum at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY

Dissolve 0.240 g in 80 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 31.93 mg of $C_{16}H_{18}FN_3O_3$.

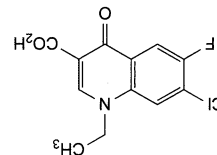
STORAGE

In an airtight container, protected from light.

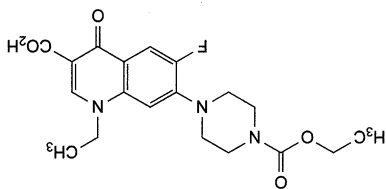
IMPURITIES

Specified impurities E, K

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, F, G, H, I, J.

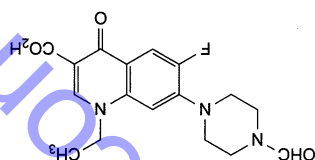


A. 7-chloro-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid.

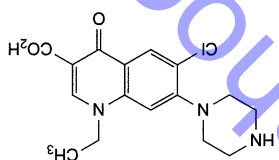


H. 7-[4-(ethoxycarbonyl)piperazin-1-yl]-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid.

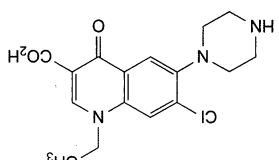
G. 1-ethyl-6-fluoro-7-(4-formylpiperazin-1-yl)-4-oxo-1,4-dihydroquinoline-3-carboxylic acid.



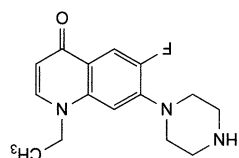
F. 6-chloro-1-ethyl-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.



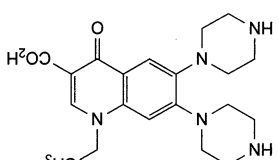
E. 7-chloro-1-ethyl-4-oxo-6-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.



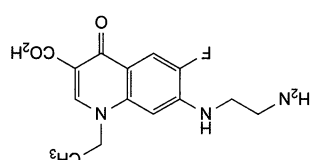
D. 1-ethyl-6-fluoro-7-(piperazin-1-yl)quinolin-4(1H)-one.



C. 1-ethyl-4-oxo-6,7-bis(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid.



B. 7-[(2-aminoethyl)amino]-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid.



Preparation Dilute the gas to be examined in nitrogen R (approximately 20:80 V/V depending on the sensitivity of the spectrophotometer).

Comparison Ph. Eur. reference spectrum of norturane.

B. Mass spectrometry (2.2.43).

Results The mass spectrum obtained with the gas to be examined is similar to the mass spectrum of norturane shown in Figure 2257.-1.

TESTS

Acidity

Maximum 0.1 ppm, expressed as HCl.

Transfer 200 mL of deionised water R previously neutralised to bromocresol purple solution R to a glass washing bottle fitted with a distribution tube with a sintered-glass disc. Pass 750 g of the gas to be examined through the water, at a rate of about 60 L/h. Titrate with 0.02 M sodium hydroxide using bromocresol purple solution R as indicator until the colour changes from yellow to bluish-violet. Carry out a blank titration using deionised water R.

1 mL of 0.02 M sodium hydroxide is equivalent to 0.729 mg of HCl.

Non-volatile matter

Carry out the test using a glass double-wall vessel (see Figure 2257.-2).

Dry the removable part in an oven at $105 \pm 2^\circ\text{C}$ for 30 min. Allow to cool in a desiccator and weigh to the nearest 0.1 mg. Connect it to the vessel.

Weigh the gas cylinder to the nearest 1 g. Fill the vessel with about 500 mL of liquefied gas and weigh the gas cylinder again. Determine the mass of the sample by weight difference. Using a suitable heating device such as a water-bath, heat the removable part such that the sample evaporates in about 2 h. Dry the removable part in an oven at $105 \pm 2^\circ\text{C}$ for 30 min. Allow to cool in a desiccator and weigh to the nearest 0.1 mg. Determine the mass of the residue by weight difference.

Calculate the content of non-volatile matter in the gas using the following expression:

$$\frac{10^3 \times m}{M}$$

m = mass of residue, in milligrams;
M = mass of sample, in grams.

Non-condensable gases

Maximum 1.5 per cent V/V.

Gas chromatography (2.2.28).

Gas to be examined The sample is taken from the vapour phase maintaining the cylinder in an upright position. Evacuate the gas loop using a multiway tap and fill cautiously with the gas to be examined.

Reference gases Mixtures of ambient air in helium for chromatography R covering a concentration range of 0.5 per cent to 2.0 per cent.

Column:

— material: stainless steel;
 — size: $l = 5\text{ m}$, $\phi = 2\text{ mm}$;
 — stationary phase: oxypropionitril-silica gel for chromatography R (150-180 μm).
 Carrier gas helium for chromatography R.
 Flow rate 21 mL/min.

Norturane

(Ph. Eur. monograph 2257)



811-97-2

C₂H₂F₄



Action and use

General anaesthetic.

DEFINITION

1,1,1,2-Tetrafluoroethane (HFC 134a).

CHARACTERS

Appearance

Clear, colourless gas, liquid under pressure.

Solubility

At 20°C and at a pressure of 101 kPa, slightly soluble in water, freely soluble in ethanol (96 per cent).

Relative density

About 1.23 at 20°C , for the gas in the liquid phase.

bp

About -26°C .

It is a hygroscopic, non-flammable gas.

IDENTIFICATION

Carry out either test A or test B.

A. Infrared absorption spectrophotometry (2.2.24). Carry out the test at atmospheric pressure.

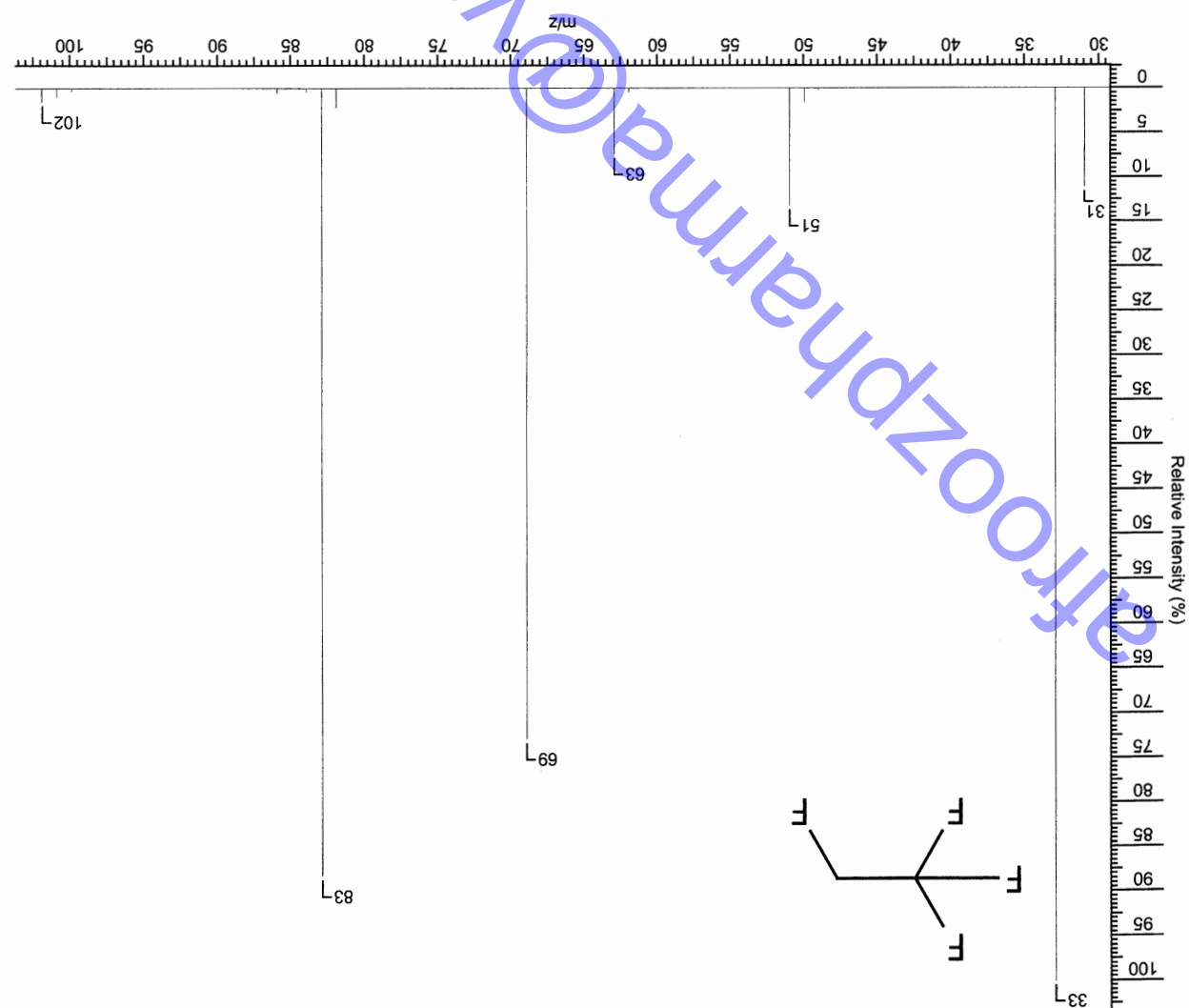


Figure 2257-1. - Mass spectrum of norflurane

m/z	RI (%)	ion	m/z	RI (%)	ion
31	11.1	[CF] ⁺	82	2.1	[CF ₂ CHF] ⁺
33	100.0	[CH ₂ F] ⁺	83	88.2	[CF ₂ CH ₂ F] ⁺
50	1.5	[CF ₂] ⁺	100	0.3	[CF ₃ CF] ⁺
51	13.2	[CHF ₂] ⁺	101	0.9	[CF ₃ CHF] ⁺
63	7.4	[CF=CHF] ⁺	102	1.5	[CF ₃ CH ₂ F] ⁺
69	72.9	[CF ₃] ⁺			

Temperature:

— column: 80 °C;

— injection port: 150 °C;

— detector: 180 °C.

Detection Thermal conductivity.

Injection 150 µL loop injector.

Run time 10 min.

Relative retention With reference to norflurane (retention

time = about 4 min): non-condensable gases = about 0.4.

Determine the concentration (V/V) of non-condensable gases

in the gas to be examined using the calibration curve

obtained with the reference gases.

Related substances

Gas chromatography (2.2.28), equipped with a gas valve sampling system and a cryogenic unit, coupled with mass

spectrometry (2.2.43).

Gas to be examined Connect the cylinder to the gas valve

sampling system and sample from the liquid phase. Then

evacuate the loop including the transfer line using a multway

tap and a vacuum pump. Open the valves of the cylinder and

fill the loop cautiously with the gas to be examined.

Reference gas (a) Prepare a mixture in helium for

chromatography R of the impurities expected in the gas to be

examined (see Table 2257-1) at a concentration of 2-6 ppm

each, always including impurity G.

— mass range: 30–300 Da;
— scan rate: ≤ 0.2 s/scan;
— temperature: ion source: 160 °C; transfer line: 200 °C.
Injection 500 μ L.

Identification of impurities Use the reconstructed ion chromatogram obtained with reference gas (a) and the information supplied in Table 2257.-1.

System suitability:

— resolution: minimum 1.4 between the peaks due to FC 1318my/c (impurity S) and FC 1318my/t (impurity T) in the chromatogram obtained with reference gas (b);

— signal-to-noise ratio: minimum 10 each for CFC 114 (impurity L) and HCC 40 (impurity W) in the chromatogram obtained with reference gas (c).

Selectivity – co-elution Several impurities elute with a resolution of less than 1; close inspection of the individual ion traces is necessary to detect and quantify potential co-eluting compounds; the relative retentions of these compounds are indicated in italics in Table 2257.-1. Before assessing the presence of individual impurities in the reconstructed ion chromatogram, a suitable background subtraction may be necessary. Two potential sources of background have to be taken into account: use of cryo-cooling (liquid carbon dioxide, m/z 44) and possible bleeding of the column (siloxanes, several ions at m/z greater than 200).

If a peak is observed at a retention time where co-elution is possible, the identification and quantification are applied to each of the co-eluting compounds.

Calculation of parts per million contents:

— for impurities A, B, C, D, E, F, G, H and I, use the concentration of the corresponding impurity in reference gas (a);
— for the other impurities, use the concentration of impurity G in reference gas (a);
— for the reporting threshold, use the concentration of impurity G in reference gas (d).

Saturated impurities

— HFC 134 (impurity C): maximum 1000 ppm;
— HFC 152a (impurity E): maximum 500 ppm;
— CFC 12 (impurity A), HCFC 124 (impurity B): for each impurity, maximum 100 ppm;
— HFC 143a (impurity D): maximum 50 ppm.

Unsaturated impurities

— CFC 1112a (impurity F), HCFC 1122 (impurity G), HFC 1225y/c (impurity H), HFC 1243z/f (impurity I): for each impurity, maximum 5 ppm.

Other detectable impurities

— other saturated or unsaturated impurities or unknown impurities: for each impurity, maximum 5 ppm;
— sum of other detectable impurities: maximum 10 ppm.

Total

Maximum 1000 ppm.

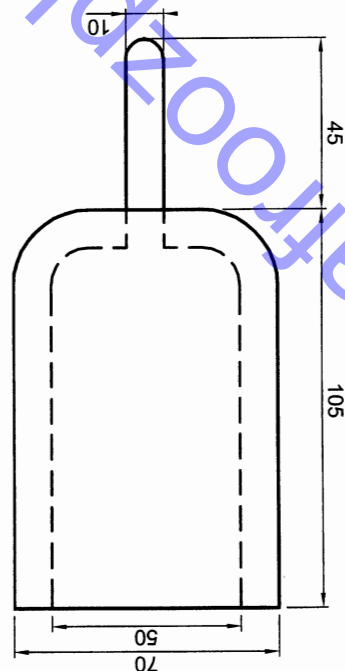
Reporting threshold

1 ppm.

Water (2.5.32)

Maximum 10 ppm, determined on 30.0 g. Take care to avoid uptake of water by the gas to be examined during the test.
Transfer the gas to be examined in liquid phase from the inverted steel cylinder to an evacuated sample can.
To transfer the sample, connect a metal tube with fittings at

Figure 2257.-2. – Glass double-wall vessel



Reference gas (b) Prepare a mixture of FC 1318my/c (impurity S) and FC 1318my/t (impurity T) in helium for chromatography R at a combined concentration of approximately 20 ppm.
Reference gas (c) Prepare a mixture of CFC 114 (impurity L) and HCC 40 (impurity W) in helium for chromatography R at a concentration of approximately 1 ppm each.
Reference gas (d) Prepare a 5-fold dilution of reference gas (a) in helium for chromatography R.
Column:
— material: fused silica;
— size: $l = 60$ m, $\varnothing = 0.18$ mm;
— stationary phase: poly[(cyanopropyl)(phenyl)][dimethylsiloxane R (film thickness 1 μ m).
Carrier gas helium for chromatography R.
Flow rate 1.1 mL/min.
Split ratio 1:75.
Temperature:

Time (min)	Temperature (°C)
0 - 2	-25
2 - 7.2	-25 \rightarrow -12
7.2 - 14	-12 \rightarrow 15
14 - 18.7	15 \rightarrow 250
18.7 - 21.2	250
Injection port	150

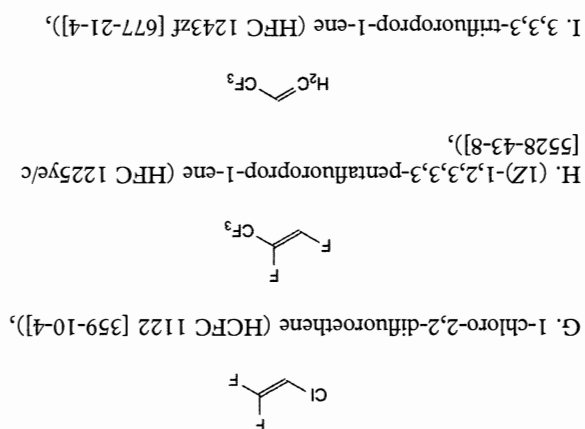
Detection Mass spectrometer; the following settings were found to be suitable and are given as examples; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criteria:
— ionisation mode: electron impact (70 eV);
— trap current: 0.2 mA;

one end to the cylinder valve, and at the other end to the needle valve on the sample can.

IMPURITIES

Specified impurities A, B, C, D, E, F, G, H, I

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: J, K, L, M, N, O, P, Q, R, S, T, U, V, W, X, Y, Z, AA, BB, CC, DD, EE, FF, GG, HH, II, JJ, KK, LL, MM, NN, OO, PP, QQ, RR, SS.



B. (2RS)-2-chloro-1,1,1,2-tetrafluoroethane (HCFC 124 [2837-89-0]),



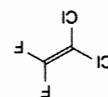
A. dichlorodifluoromethane (CFC 12 [75-71-8]),



D. 1,1,1-trifluoroethane (HFC 143a [420-46-2]),



E. 1,1-difluoroethane (HFC 152a [75-37-6]),



F. 1,1-dichloro-2,2-difluoroethene (CFC 1112a [79-35-6]),

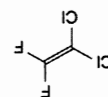


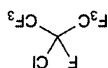
Table 2257.1. – Norflurane impurities: quantification ions and relative retentions

Impurity	Code	Structure	M _r	Quantification ion (m/z)	Relative retention
P	CFC 13	CClF ₃	104	69	0.71
Q _Q	HFC 23	CHF ₃	70	51	0.73
J	HFC 1132a	CH ₂ =CF ₂	64	64	0.73
N	CFC 115	CClF ₂ -CF ₃	154	85	0.77
D	HFC 143a	CH ₃ -CF ₃	84	65/69*	0.81
SS	HFC 32	CH ₂ F ₂	52	51/33*	0.81
II	HFC 1123	CHF=CF ₂	82	63	0.82
MM	HFC 125	CHF ₂ -CF ₃	120	101	0.84
T	FC 1318my/t	CF ₃ -CF=CF-CF ₃	200	131	0.84

Impurity	Code	Structure	M _r	Quantification ion (m/z)	Relative retention
S	FC 1318my/c	CF ₃ -CF=CF-CF ₃	200	131	0.87
Q	CFC 217ba	CF ₃ -CClF-CF ₃	204	85	0.93
KK	HFC 1234yf	CH ₂ =CF-CF ₃	114	114	0.96
RR	HFC 245cb	CH ₃ -CF ₂ -CF ₃	134	65	0.98
Norflurane	HFC 134a	CH₂F-CF₃	102	83	1
R	FC 115B1	CBF ₂ -CF ₃	198	119	1.03
H	HFC 1225ye/c	CHF=CF-CF ₃	132	113	1.10
E	HFC 152a	CH ₃ -CHF ₂	66	65	1.11
PP	HFC 161	CH ₃ -CH ₂ F	48	33/47*	1.11
I	HFC 1243zf	CH ₂ =CH-CF ₃	96	96	1.11
LL	HFC 1234ze	CHF=CH-CF ₃	114	114	1.14
A	CFC 12	CCl ₂ F ₂	120	85	1.17
C	HFC 134	CHF ₂ -CHF ₂	102	51/83*	1.21
NN	HFC 1336mzz/c	CF ₃ -CH=CH-CF ₃	164	95	1.30
GG	HFC 22	CHClF₂	86	51	1.32
L	CFC 114	CClF ₂ -CClF ₂	170	85	1.63
M	CFC 114a	CCl ₂ F-CF ₃	170	101/103*	1.64
W	HCC 40	CH ₃ Cl	50	52	1.67
G	HFC 1122	CHCl=CF ₂	98	98	1.72
DD	HFC 124a	CHF ₂ -CClF ₂	136	101	1.77
B	HFC 124	CHClF-CF ₃	136	67	1.87
HH	HFC 31	CH ₂ ClF	68	68	1.97
Y	HFC 1122a/c	CHF=CClF	98	98	2.03
O	CFC 12B1	CB ₂ ClF ₂	164	85	2.08
AA	HFC 1131/t	CHCl=CHF	80	80	2.19
OO	HFC 152	CH ₂ F-CH ₂ F	66	33	2.41
FF	HFC 133a	CH ₂ Cl-CF ₃	118	118	2.47
F	CFC 1112a	CCl ₂ =CF ₂	132	132	2.74
Z	HFC 1131/c	CHCl=CHF	80	80	2.84
J	CFC 11	CCl ₃ F	136	101	2.97
CC	HFC 123a	CHClF-CClF ₂	152	67	3.15
BB	HFC 123	CHCl-CF ₃	152	83	3.18
K	CFC 113	CCl ₂ F-CClF ₂	186	151	3.18
X	HFC 1121/t	CHCl=CClF	114	114	3.25
V	HCC 30	CH ₂ Cl ₂	84	49	3.29
EE	HFC 132b	CClF ₂ -CH ₂ Cl	134	99	3.32
U	HCC 1120	CHCl=CCl ₂	130	95	3.59

* Depending on the actual chromatographic resolution and potentially overlapping compounds, it may be necessary to select a different quantification ion.

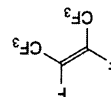
P. chlorotrifluoromethane (CFC 13 [75-72-9]),



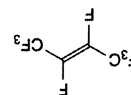
Q. 2-chloro-1,1,1,2,3,3,3-heptafluoropropane (CFC 217ba [76-18-6]),



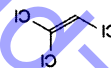
R. 1-bromo-1,1,2,2-pentafluoroethane (FC 115B1 [354-55-2]),



S. (2Z)-1,1,1,2,3,4,4,4-octafluorobut-2-ene (FC 1318my/c [1516-65-0]),



T. (2E)-1,1,1,2,3,4,4,4-octafluorobut-2-ene (FC 1318my/t [1516-64-9]),



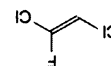
U. 1,1,2-trichloroethene (trichloroethylene, HCC 1120 [79-01-6]),



V. dichloromethane (methylene chloride, HCC 30 [75-09-2]),



W. chloromethane (methyl chloride, HCC 40 [74-87-3]),



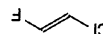
X. (E)-1,2-dichloro-1-fluoroethene (HCFC 1121/e),



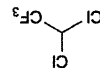
Y. (Z)-1-chloro-1,2-difluoroethene (HCFC 1122a/c [359-04-6]),



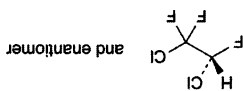
Z. (Z)-1-chloro-2-fluoroethene (HCFC 1131/c [2268-31-7]),



AA. (E)-1-chloro-2-fluoroethene (HCFC 1131/t [2268-32-8]),



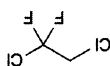
BB. 1,1-dichloro-2,2,2-trifluoroethane (HCFC 123 [306-83-2]),



CC. (2R,S)-1,2-dichloro-1,1,2-trifluoroethane (HCFC 123a [354-23-4]),



DD. 1-chloro-1,1,2,2-tetrafluoroethane (HCFC 124a [354-25-6]),



EE. 1,2-dichloro-1,1-difluoroethane (HCFC 132b [1649-08-7]),



FF. 2-chloro-1,1,1-trifluoroethane (HCFC 133a [75-88-7]),



GG. chlorodifluoromethane (HCFC 22 [75-45-6]),



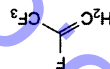
HH. chlorofluoromethane (HCFC 31 [593-70-4]),



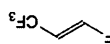
II. 1,1,2-trifluoroethene (HCFC 1123 [359-11-5]),



JJ. 1,1-difluoroethene (HCFC 1132a [75-38-7]),



KK. 2,3,3,3-tetrafluoroprop-1-ene (HFC 1234yf [754-12-1]),



LL. (1E)-1,3,3,3-tetrafluoroprop-1-ene (HFC 1234ze [1645-83-6]),



MM. 1,1,1,2,2-pentafluoroethane (HFC 125 [354-33-6]),



NN. (2Z)-1,1,1,4,4,4-hexafluorobut-2-ene (HFC 1336mzz/c [692-49-9]),



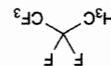
OO, 1,2-difluoroethane (HFC 152 [624-72-6]),



PP, fluoroethane (HFC 161 [353-36-6]),



QQ, trifluoromethane (HFC 23 [75-46-7]),



RR, 1,1,1,2,2-pentafluoropropane (HFC 245cb [1814-88-6]),

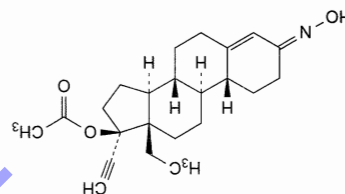


SS, difluoromethane (HFC 32 [75-10-5]).

Ph Eur

Norgestimate

(Ph. Eur. monograph 1732)



C₂₃H₃₁NO₃ 369.5 35189-28-7

Action and use
Progestogen.

Ph Eur

DEFINITION

(3*E*)-13β-Ethyl-3-(hydroxyimino)-18,19-dinor-17α-pregn-4-en-20-yn-17-yl acetate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in acetone.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison norgestimate CRS.

TESTS

Specific optical rotation (2.2.7)

Dissolve 0.200 g in methylene chloride R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture water R, methanol R (1:4 V/V).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of norgestimate for system suitability CRS (containing impurity A) in 4 mL of the solvent mixture.

Column: size: $l = 0.10$ m, $\phi = 4.6$ mm; stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m); temperature: 40 °C.

Mobile phase acetonitrile R, tetrahydrofuran for chromatography R, water R (18:22:60 V/V/V).

Flow rate 1.0 mL/min. Detection Spectrophotometer at 244 nm. Injection 25 μ L.

Run time Twice the retention time of the (E)-isomer of norgestimate.

Identification of impurities Use the chromatogram supplied with norgestimate for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

Relative retention With reference to the (E)-isomer of norgestimate (retention time = about 14 min): impurity A = about 0.7; (Z)-isomer of norgestimate = about 0.9.

System suitability: reference solution (b): resolution: minimum 1.5 between the peaks due to the (E)- and (Z)-isomers of norgestimate.

Limits: peak area of the (Z)-isomer of norgestimate by 1.33; impurity A: not more than twice the sum of the areas of the peaks due to the (E)- and (Z)-isomers of norgestimate in the chromatogram obtained with reference solution (a) (0.2 per cent); unspecified impurities: for each impurity, not more than the sum of the areas of the peaks due to the (E)- and (Z)-isomers of norgestimate in the chromatogram obtained with reference solution (a) (0.10 per cent); total: not more than 3 times the sum of the areas of the peaks due to the (E)- and (Z)-isomers of norgestimate in the chromatogram obtained with reference solution (a) (0.3 per cent); disregard limit: 0.5 times the sum of the areas of the peaks due to the (E)- and (Z)-isomers of norgestimate in the chromatogram obtained with reference solution (a) (0.05 per cent).

Ratio of (E)- to (Z)-isomers Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

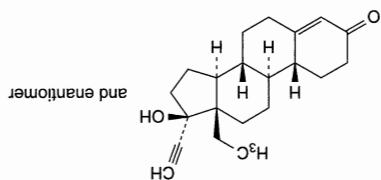
Injection Test solution. Calculate the (E)- to (Z)-isomer ratio by dividing the area of the peak due to the (E)-isomer by 1.33 times the area of the peak due to the (Z)-isomer. The ratio is 1.27 to 1.78.

Loss on drying (2.2.32) Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C for 3 h.



Norgestrel

(Ph. Eur. monograph 0940)



$C_{21}H_{28}O_2$ 312.5

6533-00-2

Action and use

Progestogen.

Preparation

Norgestrel Tablets

DEFINITION

Norgestrel contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of *rac*-13-ethyl-17-hydroxy-18,19-dinor-4-en-20-yn-3-one, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, practically insoluble in water, sparingly soluble in methylene chloride, slightly soluble in alcohol.

IDENTIFICATION

A. Dissolve 0.5 g in methylene chloride R and dilute to 10.0 mL with the same solvent. The angle of optical rotation (2.2.7) is + 0.05° to - 0.05°.
B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with norgestrel CRS.

TESTS

Related substances

Examine by thin-layer chromatography (2.2.27), using *silica gel G* R as the coating substance.
Test solution Dissolve 0.2 g of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (a) Dilute 1 mL of the test solution to 10 mL with methylene chloride R. Dilute 1 mL of this solution to 20 mL with methylene chloride R.
Reference solution (b) Dilute 4 mL of reference solution (a) to 10 mL with methylene chloride R.
Reference solution (c) Dissolve 5 mg of norgestrel CRS and 5 mg of ethinylestradiol CRS in methylene chloride R and dilute to 50 mL with the same solvent.

Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 20 volumes of ethyl acetate R and 80 volumes of methylene chloride R. Allow the plate to dry in air, spray with a 100 g/L solution of phosphomolybdic acid R in alcohol R, heat at 100-105 °C for 15 min and examine immediately. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the principal spot in the chromatogram obtained with reference solution (a) (0.5 per cent) and at most two such spots are more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent). The test is not valid unless the

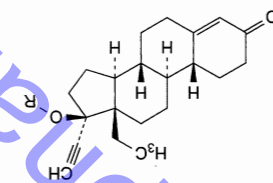
ASSAY

Dissolve 0.300 g in 40 mL of tetrahydrofuran R. Add 10 mL of a 100 g/L solution of silver nitrate R and titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). Rinse the electrode with acetone R after each titration.
If necessary, after several titrations re-equilibrate the electrode in water R for 15 min to obtain sharper titration curves.
1 mL of 0.1 M sodium hydroxide is equivalent to 36.95 mg of $C_{23}H_{31}NO_3$.

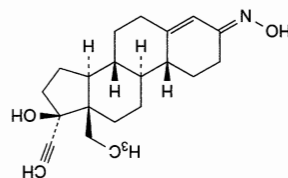
IMPURITIES

Specified impurities A

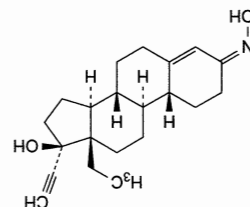
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph). They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: B, C, D.



A. R = CO-CH₃; 13β-ethyl-3-oxo-18,19-dinor-17α-pregn-4-en-20-yn-17-yl acetate (levonorgestrel acetate),
B. R = H; 13β-ethyl-17β-hydroxy-18,19-dinor-17α-pregn-4-en-20-yn-3-one (levonorgestrel),



C. (3E)-13β-ethyl-3-(hydroxyimino)-18,19-dinor-17α-pregn-4-en-20-yn-17-ol ((E)-norgestromin),



D. (3Z)-13β-ethyl-3-(hydroxyimino)-18,19-dinor-17α-pregn-4-en-20-yn-17-ol ((Z)-norgestromin).

Ph Eur

chromatogram obtained with reference solution (c) shows

two clearly separated spots.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by

drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 45 mL of *tetrahydrofuran* R. Add 10 mL

of a 100 g/L solution of *silver nitrate* R. After 1 min, titrate

with 0.1 M *sodium hydroxide* determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M *sodium hydroxide* is equivalent to 31.25 mg of

$C_{21}H_{28}O_2$.

STORAGE

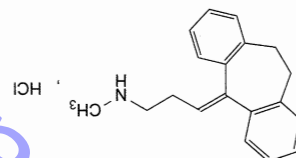
Store protected from light.

Ph Eur



Nortriptyline Hydrochloride

(Ph. Eur. monograph 0941)



$C_{21}H_{22}ClN$

299.8

894-71-3

Action and use

Preparations

Nortriptyline Capsules

Nortriptyline Tablets

Ph Eur

DEFINITION

3-(10,11-Dihydro-5H-dibenzo[a,d][7]annulen-5-ylidene)-N-

methylpropan-1-amine hydrochloride.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance White or almost white powder.

Solubility

Sparingly soluble in water, soluble in ethanol (96 per cent)

and in methylene chloride.

IDENTIFICATION

First identification C, E

Second identification A, B, D, E

A. Melting point (2.2.14): 216 °C to 220 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 20.0 mg in *methanol* R and dilute to

100.0 mL with the same solvent. Dilute 5.0 mL of this

solution to 100.0 mL with *methanol* R.

Spectral range 230–350 nm.

Absorption maximum At 239 nm.

Specific absorbance at the absorption maximum 465 to 495.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison nortriptyline hydrochloride CRS.

D. Dissolve 50 mg in 3 mL of warm *water* R, cool and add

0.05 mL of a 25 g/L solution of *quinhydrone* R in *methanol* R.

A red colour develops slowly.

E. 50 mg gives reaction (b) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured

than reference solution B₇ (2.2.2, Method II).

Dissolve 0.5 g in *water* R with gentle heating and dilute to

25 mL with the same solvent.

Acidity or alkalinity

Dissolve 0.2 g with gentle heating in *carbon dioxide-free*

water R and dilute to 10 mL with the same solvent.

Add 0.1 mL of *methyl red* solution R and 0.2 mL of 0.01 M

sodium hydroxide. The solution is yellow. Add 0.4 mL of

0.01 M *hydrochloric acid*. The solution is red.

Related substances

Liquid chromatography (2.2.29). Protect the solutions from

light.

Test solution Dissolve 50.0 mg of the substance to be

examined in the mobile phase and dilute to 25.0 mL with

the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10.0 mg of *dibenzosuberone* CRS

(impurity A) and 20 mg of *noryclodibenzazepine* CRS

(impurity B) in the mobile phase and dilute to 100.0 mL

with the mobile phase. Dilute 1.0 mL of this solution to

10.0 mL with the mobile phase.

Reference solution (c) Dissolve 10 mg of *nortriptyline* for system

suitability CRS (containing impurity D) in the mobile phase,

add 1.0 mL of reference solution (b) and dilute to 10.0 mL

with the mobile phase.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: spherical end-capped octylsilyl silica gel for

chromatography R (5 μ m);

— temperature: 45 °C.

Mobile phase Mix 70 volumes of *methanol* R2 and 30 volumes

of a solution prepared as follows: dissolve 3.25 mL of

tetrabutylammonium hydroxide solution (400 g/L) R and 0.68 g

of *potassium dihydrogen phosphate* R in 900 mL of *water* R,

adjust to pH 7.5 with *dilute phosphoric acid* R and dilute to

1000 mL with *water* R.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L of the test solution and reference solutions (a)

and (c).

Run time 3 times the retention time of *nortriptyline*.

Identification of impurities Use the chromatogram obtained

with reference solution (c) to identify the peaks due to

impurities A, B and D.

Relative retention With reference to *nortriptyline* (retention

time = about 13 min): impurity A = about 0.5;

impurity D = about 0.8; impurity B = about 0.9.

System suitability: reference solution (c):
— resolution: minimum 1.4 between the peaks due to impurities D and B, and minimum 2.0 between the peaks due to impurity B and nortriptyline.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity D by 1.7;

— impurity D: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.05 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 30 mL of ethanol (96 per cent) R.

Add 1.0 mL of 0.1 M hydrochloric acid. Carry out a

potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 29.98 mg of

C₁₉H₂₂ClN.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, D

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by the general

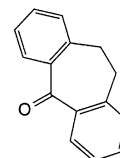
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

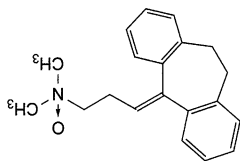
Control of impurities in substances for pharmaceutical use): B, E,

F, G, H, I, J.

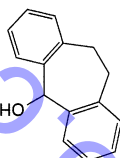


A. 10,11-dihydro-5H-dibenzo[a,d][7]annulen-5-one (dibenzosuberone),

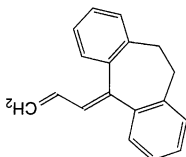
J. [3-(10,11-dihydro-5H-dibenzo[a,d][7]annulen-5-ylidene)propyl]dimethylamine oxide (amitriptyline-N-oxide).



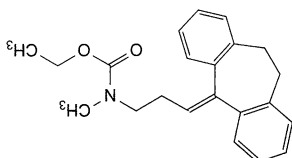
I. 10,11-dihydro-5H-dibenzo[a,d][7]annulen-5-ol (dibenzosuberol),



H. 5-prop-2-en-1-ylidene-10,11-dihydro-5H-dibenzo[a,d][7]annulene,

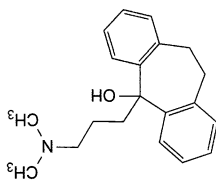


G. ethyl [3-(10,11-dihydro-5H-dibenzo[a,d][7]annulen-5-ylidene)propyl]methylcarbamate,



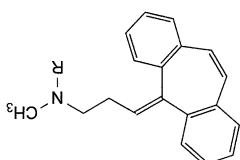
F. amitriptyline,

D. 5-[3-(dimethylamino)propyl]-10,11-dihydro-5H-dibenzo[a,d][7]annulen-5-ol,



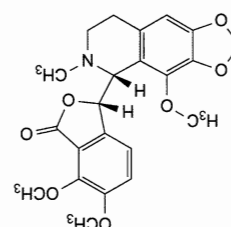
E. R = CH₃: 3-(5H-dibenzo[a,d][7]annulen-5-ylidene)-N,N-dimethylpropyl-1-amine (cyclobenzaprine),

B. R = H: 3-(5H-dibenzo[a,d][7]annulen-5-ylidene)-N-methylpropyl-1-amine (nortriptyline),



Noscaphine

(Ph. Eur. monograph 0516)



C₂₂H₂₃NO₇ 413.4

128-62-1

Action and use

Opioid receptor agonist; cough suppressant.

DEFINITION

(3S)-6,7-Dimethoxy-3-[(5R)-4-methoxy-6-methyl-5,6,7,8-tetrahydro-1,3-dioxolo[4,5-g]isquinolin-5-yl]isobenzofuran-1(3H)-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Practically insoluble in water, soluble in acetone, slightly soluble in ethanol (96 per cent). It dissolves in strong acids; on dilution of the solution with water, the base may be precipitated.

IDENTIFICATION

First identification C, E

Second identification A, B, D, E

A. Specific optical rotation (see Tests).

B. Melting point (2.2.14): 174 °C to 177 °C.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison noscaphine CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in acetone R and dilute to 100 mL with the same solvent.

Reference solution Dissolve 25 mg of noscaphine CRS in acetone R and dilute to 100 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, ethanol (96 per cent) R, acetone R, toluene R (1:3:20:20 V/V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with dilute potassium iodobismuthate solution R.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

E. To 20 mg add 10 mL of water R and shake. It does not dissolve.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 0.2 g in acetone R and dilute to 10 mL with the same solvent. Examine immediately after dissolution.

Specific optical rotation (2.2.7)

+ 42 to + 48 (dried substance).
Dissolve 0.500 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 20.0 mg of the substance to be examined in 8 mL of methanol R, with the aid of ultrasound, and dilute to 10.0 mL with the same solvent.
Reference solution (a) Dilute 1.0 mL of the test solution to 20.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.
Reference solution (b) Dissolve 5 mg of papaverine hydrochloride R in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.
Reference solution (c) Dissolve 1.5 mg of papaverine hydrochloride R in 10 mL of the test solution and dilute to 25 mL with the mobile phase.

Column:

— size: l = 0.125 m, Ø = 4.6 mm;
— stationary phase: nitrile silica gel for chromatography R (5 µm).

Mobile phase methanol R, phosphate buffer solution pH 6.0 R1 (350:650 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time 2.5 times the retention time of noscaphine.

Relative retention With reference to noscaphine (retention time = about 10 min): impurity A = about 1.3.

System suitability: reference solution (c):

— resolution: minimum 2 between the peaks due to noscaphine and impurity A.

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— any other impurity: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— sum of impurities other than A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

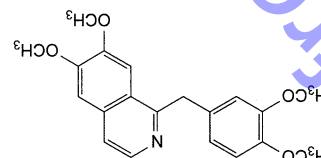
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
 Dissolve 0.350 g in 40 mL of anhydrous acetic acid R, warming gently. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
 $C_{22}H_{23}NO_7$
 1 mL of 0.1 M perchloric acid is equivalent to 41.34 mg of

STORAGE
 Protected from light.

IMPURITIES

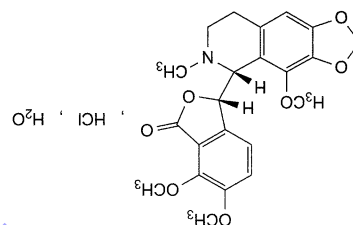


A. 1-(3,4-dimethoxybenzyl)-6,7-dimethoxyisoquinoline (papaverine).

Noscaspine Hydrochloride Hydrate



Noscaspine Hydrochloride
 (Ph. Eur. monograph 0515)



Action and use
 Opioid receptor agonist; cough suppressant

DEFINITION

(3S)-6,7-Dimethoxy-3-[(5R)-4-methoxy-6-methyl-5,6,7,8-tetrahydro-1,3-dioxolo[4,5-g]isoquinolin-5-yl]-2-benzofuran-1 (3H)-one hydrochloride hydrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals, hygroscopic.

Solubility

Freely soluble in water and in ethanol (96 per cent).

Aqueous solutions are slightly acid; the base may be precipitated when the solutions are allowed to stand.

mp

About 200 °C, with decomposition.

IDENTIFICATION

First identification C, E

Second identification A, B, D, E

A. Specific optical rotation (see Tests).

B. Melting point (2.2.14) of the precipitate obtained in identification test E: 174 °C to 177 °C.
 C. Infrared absorption spectrophotometry (2.2.24).
 Preparation Examine the precipitate obtained in identification test E.
 Comparison noscaspine CRS.
 D. Thin-layer chromatography (2.2.27).
 Test solution Dissolve 25 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 100 mL with the same solvent.
 Reference solution Dissolve 22 mg of noscaspine CRS in acetone R and dilute to 100 mL with the same solvent.
 Plate TLC silica gel plate R.
 Mobile phase concentrated ammonia R, ethanol (96 per cent) R, acetone R, toluene R (1:3:20:20 V/V/V/V).
 Application 10 µL.
 Development Over 2/3 of the plate.
 Drying In air.
 Detection Spray with dilute potassium iodobismuthate solution R. Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.
 E. Dissolve about 40 mg in a mixture of 2 mL of water R and 3 mL of ethanol (96 per cent) R and add 1 mL of dilute ammonia R2. Heat until dissolution is complete. Allow to cool, scratching the wall of the tube with a glass rod. Filter. The filtrate gives reaction (a) of chlorides (2.3.1). Wash the precipitate with water R, dry at 100-105 °C and reserve for identification tests B and C.

TESTS

The solution is not more intensely coloured than reference solution Y₆ or BY₆ (2.2.2, Method II).
 Dissolve 0.5 g in water R, add 0.3 mL of 0.1 M hydrochloric acid and dilute to 25 mL with water R.
 pH (2.2.3)
 Minimum 3.0.
 Dissolve 0.2 g in 10 mL of carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29).
 Test solution Dissolve 40.0 mg of the substance to be examined in methanol R, with the aid of ultrasound, and dilute to 20.0 mL with the same solvent.
 Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.
 Reference solution (b) Dissolve 5.0 mg of papaverine hydrochloride CRS (impurity A) in the mobile phase and dilute to 50.0 mL with the mobile phase (solution A). Dilute 1.0 mL of solution A to 10.0 mL with the mobile phase.
 Reference solution (c) Dilute 6.0 mL of solution A to 10.0 mL with the test solution.

Column:

— size: $l = 0.125$ m, $\phi = 4.6$ mm;
 — stationary phase: cyanosilyl silica gel for chromatography R (5 µm).

Mobile phase methanol R, phosphate buffer solution pH 6.0 R1

(35:65 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time 2.5 times the retention time of nystatine.

Relative retention With reference to nystatine (retention time = about 10 min): impurity A = about 1.3.

System suitability: reference solution (c):

— resolution: minimum 2 between the peaks due to nystatine and impurity A.

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.5 per cent);

— any other impurity: for each impurity, not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

— sum of impurities other than A: not more than 5 times the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

2.5 per cent to 6.5 per cent, determined on 0.200 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid over heating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.400 g in a mixture of 3 mL of anhydrous formic acid R and 30 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

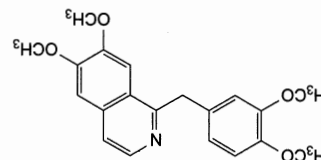
1 mL of 0.1 M perchloric acid is equivalent to 44.99 mg of C₂₂H₂₄ClNO₇.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A

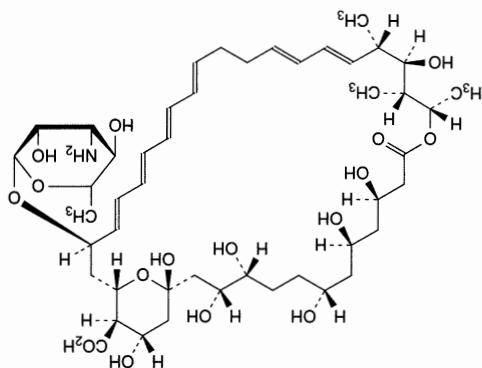


A. 1-[(3,4-dimethoxyphenyl)methyl]-6,7-dimethoxyisquinoline (papaverine).

Ph Eur

Nystatin

(Ph. Eur. monograph 0517)



C₄₇H₇₅NO₁₇ 926

Action and use

Antifungal.

Preparations

Nystatin Ointment

Nystatin Oral Suspension

Nystatin Pastilles

Nystatin Pessaries

Nystatin Tablets

Ph Eur

DEFINITION

Antifungal substance obtained by fermentation using certain strains of *Streptomyces noursei* as the production micro-organism. It contains mainly tetraenes, the principal component being (1S,3R,4R,7R,11R,15S,16R,17R,18S,19E,21E,22E,27E,29E,31E,33R,35S,36R,37S)-(3-amino-3,6-didecyl-β-D-mannopyranosyl)oxy]-1,3,4,7,9,11,17,37-octahydroxy-15,16,18-trimethyl-13-oxo-14,39-dioxabicyclo[3.3.1]nonatriacenta-19,21,25,27,29,31-hexaene-36-carboxylic acid (nystatin A1).

Content

Minimum 4400 IU/mg (dried substance) and minimum 5000 IU/mg (dried substance) if intended for oral administration.

PRODUCTION

If nystatin is not intended for cutaneous administration, the method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.

Abnormal toxicity (2.6.9)

Inject intraperitoneally into each mouse a quantity equivalent to not less than 600 IU suspended in 0.5 mL of a 5 g/L solution of *acacia* R.

CHARACTERS

Appearance

Yellow or slightly brownish powder, hygroscopic.

Solubility

Practically insoluble in water, freely soluble in dimethylformamide and in dimethyl sulfoxide, slightly soluble in methanol, practically insoluble in alcohol.

IDENTIFICATION

First identification B, E

Second identification A, C, D

A. Examine the solution prepared in the test for absorbance between 220 nm and 350 nm (2.2.25). The solution shows 4 absorption maxima at 230 nm, 291 nm, 305 nm and 319 nm, and a shoulder at 280 nm. The ratios of the absorbances at the absorption maxima at 291 nm and 319 nm to the absorbance at the absorption maximum at 305 nm are 0.61 to 0.73 and 0.83 to 0.96, respectively. The ratio of the absorbance measured at the absorption maximum at 230 nm to that measured at the shoulder at 280 nm is 0.83 to 1.25.

B. Infrared absorption spectrophotometry (2.2.24).
Comparison nystatin CRS.
C. To about 2 mg add 0.1 mL of hydrochloric acid R. A brown colour develops.
D. To about 2 mg add 0.1 mL of sulfuric acid R. A brown colour develops that becomes violet on standing.
E. Examine the chromatograms obtained in the test for composition.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

TESTS

Absorbance (2.2.25)
Dissolve 0.10 g in a mixture of 5.0 mL of glacial acetic acid R and 50 mL of methanol R and dilute to 100.0 mL with methanol R. Dilute 1.0 mL of the solution to 100.0 mL with methanol R. Determine at the maximum at 305 nm within 30 min of preparation of the solution, the absorbance is not less than 0.60.

Composition
Liquid chromatography (2.2.29): use the normalisation procedure. Carry out the test protected from light.
Test solution Dissolve 20 mg of the substance to be examined in dimethyl sulfoxide R and dilute to 50 mL with the same solvent.
Reference solution (a) Dissolve 20 mg of nystatin CRS in dimethyl sulfoxide R and dilute to 50 mL with the same solvent.
Reference solution (b) Dissolve 20 mg of the substance to be examined in 25 mL of methanol R and dilute to 50 mL with water R. To 10.0 mL of the solution add 2.0 mL of dilute hydrochloric acid R. Allow to stand at room temperature for 1 h.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with dimethyl sulfoxide R. Dilute 1.0 mL of this solution to 10.0 mL with dimethyl sulfoxide R.

Column:
— size: $l = 0.15$ m, $\varnothing = 4.6$ mm,
— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m),
— temperature: 30 °C.

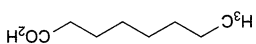
Mobile phase:
— mobile phase A: acetonitrile R, 3.85 g/L solution of ammonium acetate R (29:71 V/V),
— mobile phase B: 3.85 g/L solution of ammonium acetate R, acetonitrile R (40:60 V/V).

DEFINITION

Octanoic acid.

Action and use

Excipient.

C₈H₁₆O₂ 144.2 124-07-2

(Caprylic Acid, Ph Eur monograph 1401)

Octanoic Acid

Ph Eur

LABELLING

The label states where applicable, that the substance is only for cutaneous use.

STORAGE

In an airtight container, protected from light.

ASSAY

Carry out the microbiological assay of antibiotics (2.7.2). Protect the solutions from light throughout the assay.

Dissolve the substance to be examined and nystatin CRS separately in dimethylformamide R and dilute with a mixture of 5 volumes of dimethylformamide R and 95 volumes of buffer solution pH 6.0.

Sulfated ash (2.4.14)

Maximum 3.5 per cent, determined on 1.0 g.

exceeding 0.1 kPa for 3 h.

Maximum 5.0 per cent, determined on 1.000 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 0.1 kPa for 3 h.

Loss on drying (2.2.32)

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Heavy metals (2.4.8)

Maximum 20 ppm.

2 min.

disregard any peak with a retention time of less than

chromatogram obtained with reference solution (c);

— disregard limit: the area of the principal peak in the

— any other compound: maximum 4.0 per cent,

— nystatin A1: minimum 85.0 per cent,

Composition:

(retention time = about 13 min and 19 min).

— resolution: minimum 3.5 between the 2 principal peaks

System suitability: reference solution (b):

Retention time Nystatin A1 = about 14 min.

Injection 20 μ L

Detection Spectrophotometer at 305 nm.

Flow rate 1.0 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 25	100	0
25 - 35	100 \rightarrow 0	0 \rightarrow 100
35 - 45	0	100
45 - 50	0 \rightarrow 100	100 \rightarrow 0
50 - 55	100	0

Content

99.0 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS**Appearance**

Clear, colourless or slightly yellowish, oily liquid.

Solubility

Very slightly soluble in water, very soluble in acetone and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

A. Relative density (see Tests).

B. Examine the chromatograms obtained in the test for related substances.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

TESTS**Appearance**

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2).

Method II.**Relative density (2.2.5)**

0.909 to 0.912.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.
Test solution Dissolve 0.10 g of the substance to be examined in ethyl acetate R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 0.10 g of caprylic acid CRS in ethyl acetate R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with ethyl acetate R. Dilute 5.0 mL of this solution to 50.0 mL with ethyl acetate R.

Column:

— **material:** fused silica;

— **size:** $l = 30$ m, $\varnothing = 0.25$ mm;

— **stationary phase:** macrogol 20 000 2-nitroterephthalate R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 1.5 mL/min.

Split ratio 1:100.

Temperature:

Time (min)	Temperature (°C)
0 - 1	100
1 - 25	100 → 220
25 - 35	220
Injection port	250
Detector	250

Detection Flame ionisation.

Injection 1 μ L.

System suitability: reference solution (b):

— **signal-to-noise ratio:** minimum 5 for the principal peak.

Limits:

— **any impurity:** for each impurity, maximum 0.3 per cent;

— **total:** maximum 0.5 per cent;

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in ethanol (96 per cent) R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test B. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R and 9 mL of ethanol (96 per cent) R.

Water (2.5.12)

Maximum 0.7 per cent, determined on 1.000 g.

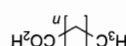
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.125 g in 25 mL of ethanol (96 per cent) R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 14.42 mg of C₈H₁₆O₂.

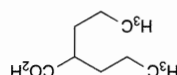
IMPURITIES

A. $n = 4$: hexanoic acid,

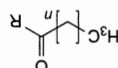
B. $n = 5$: heptanoic acid,

C. $n = 7$: nonanoic acid,

D. $n = 8$: decanoic acid,



B. 2-propylpentanoic acid (valproic acid),

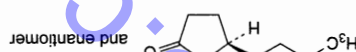


F. R = OCH₃, $n = 6$: methyl octanoate,

G. R = OC₂H₅, $n = 6$: ethyl octanoate,

H. R = OCH₃, $n = 8$: methyl decanoate,

I. R = CH₃, $n = 8$: undecan-2-one,



J. 5-butyldihydrofuran-2-one (γ -hydroxyoctanoic acid lactone).

Octoxinol 10

(Ph. Eur. monograph 1553)

Action and use

Excipient.

DEFINITION

α -[4-(1,1,3,3,3-Tetramethylbutyl)phenyl]- ω -hydroxydeca(oxyethylene).

Mixture consisting mainly of mono-octylphenyl ethers of macrogols corresponding to the formula C₈H₁₇-C₆H₄-

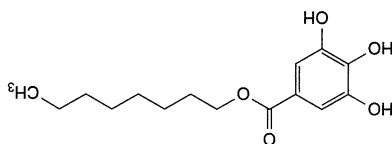


Ph Eur



Octyl Gallate

(Ph. Eur. monograph 2057)



$C_{15}H_{22}O_5$

282.3

1034-01-1

Action and use

Used in treatment of alcohol dependence.

Ph Eur

DEFINITION

Octyl 3,4,5-trihydroxybenzoate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Melting point (2.2.14).

Determine the melting point of the substance to be examined. Mix equal parts of the substance to be examined and octyl gallate CRS and determine the melting point of the mixture. The difference between the melting points (which are about 101 °C) is not greater than 2 °C.

B. Examine the chromatograms obtained in the test for impurity A.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Impurity A

Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.20 g of the substance to be examined in acetone R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 20 mL with acetone R.

Reference solution (a) Dissolve 10 mg of octyl gallate CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of gallic acid R in acetone R and dilute to 20 mL with the same solvent.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10 mL with acetone R.

Reference solution (d) Dilute 1.0 mL of reference solution (b) to 5 mL with test solution (a).

Plate TLC silica gel plate R.

Mobile phase anhydrous formic acid R, ethyl formate R, toluene R (10:40:50 V/V/V).

Application 5 µL of test solutions (a) and (b) and reference solutions (a), (c) and (d).

Development Over 2/3 of the plate.

TESTS

B. Cloud point (see Tests).

Preparation Film between sodium chloride R plates.

Comparison octoxinol 10 CRS.

A. Infrared absorption spectrophotometry (2.2.24).

IDENTIFICATION

vegetable oils.

Miscible with water, with ethanol (96 per cent) and with

Solubility

Clear, colourless or light yellow, viscous liquid.

Appearance

CHARACTERS

It may contain free macrogols.

$[OCH_2-CH_2]_n-OH$ where the average value of n is 10.

STORAGE

In an airtight container.

Maximum 0.4 per cent, determined on 1.0 g.

Total ash (2.4.16)

Maximum 0.5 per cent, determined on 2.00 g.

Water (2.5.12)

solution (1 ppm Pb) R.

test A. Prepare the reference solution using lead standard the same solvent. 12 mL of this solution complies with Dissolve 2.0 g in distilled water R and dilute to 20.0 mL with

Maximum 10 ppm.

Heavy metals (2.4.8)

of dioxan.

Maximum 1 ppm of ethylene oxide and maximum 10 ppm

Ethylene oxide and dioxan (2.4.25)

plainly seen.

becomes sufficiently clear that the entire thermometer bulb is The cloud point is the temperature at which the solution does not increase more than 2 °C, and continue to stir. test-tube from the water-bath (ensuring that the temperature continuously until the solution becomes cloudy. Remove the this solution to a test-tube, heat on a water-bath and stir Dissolve 1.0 g in 99 g of water R. Transfer about 30 mL of

63 °C to 70 °C.

Cloud point

85 to 101.

Hydroxyl value (2.5.3, Method A)

indicator.

sodium hydroxide is required to change the colour of the Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M the filtrate, add 0.05 mL of bromothymol blue solution R1. 1 min, with constant stirring. Cool and filter. To 10 mL of Boil 1.0 g with 20 mL of carbon dioxide-free water R for

Acidity or alkalinity

Drying In air for 10 min.

Detection Spray with a mixture of 1 volume of ferric chloride solution R1 and 9 volumes of ethanol (96 per cent) R.

System suitability Reference solution (d):

the chromatogram shows 2 clearly separated principal spots.

Limit: test solution (a):

intense than the spot in the chromatogram obtained with reference solution (c) (0.5 per cent).

Chlorides (2.4.4)

Maximum 100 ppm.

To 1.65 g add 50 mL of water R. Shake for 5 min. Filter.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 70 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in methanol R and dilute to 250.0 mL with the same solvent. Dilute 5.0 mL of the solution to 200.0 mL with methanol R. Measure the absorbance (2.2.25) at the absorption maximum at 275 nm.

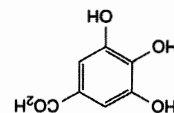
Calculate the content of C₁₅H₃₂O₂ taking the specific absorbance to be 387.

STORAGE

In a non-metallic container, protected from light.

IMPURITIES

Specified impurities: A.



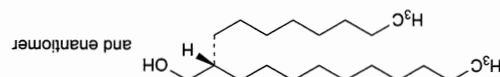
A. 3,4,5-trihydroxybenzoic acid (gallic acid).

Ph Eur



Octyldodecanol

(Ph. Eur. monograph 1136)



5333-42-6

Action and use

Excipient.

Ph Eur

DEFINITION

Condensation product of saturated liquid fatty alcohols.

Content

Minimum 90 per cent of (2R5)-2-octyldodecan-1-ol (C₂₀H₄₂O₂; 298.6), the remainder consisting mainly of related alcohols.

CHARACTERS

Appearance

Clear, colourless or yellowish, oily liquid.

Solubility

Practically insoluble in water, miscible with ethanol (96 per cent).

Relative density

About 0.840.

Refractive index About 1.455.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.20 g of the substance to be examined in toluene R and dilute to 20 mL with the same solvent.

Reference solution Dissolve 0.20 g of octyldodecanol CRS in toluene R and dilute to 20 mL with the same solvent.

Plate Suitable silica gel plate.

Mobile phase ethyl acetate R, toluene R (5:95 V/V).

Application 2 µL.

Development Over a path of 12 cm.

Drying In air.

Detection Spray with about 7 mL of a mixture of 1 volume of a 25 g/L solution of vanillin R in ethanol (96 per cent) R and 4 volumes of sulfuric acid R and heat at 130 °C for 5-10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Acidity or alkalinity

Mix 5.0 g thoroughly for 1 min with a mixture of 0.1 mL of bromothymol blue solution R1, 2 mL of heptane R and 10 mL of water R. If the aqueous layer is blue, not more than 0.15 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to yellow. If the aqueous layer is yellow, add 0.45 mL of 0.01 M sodium hydroxide and shake vigorously. After standing to ensure complete separation, the aqueous layer is blue.

Optical rotation (2.2.7)

-0.10° to +0.10°.

Dissolve 2.50 g in ethanol (96 per cent) R and dilute to 25 mL with the same solvent.

Hydroxyl value (2.5.3, Method A)

175 to 190.

Iodine value (2.5.4, Method A)

Maximum 8.0.

Peroxide value (2.5.5, Method A)

Maximum 5.0.

Saponification value (2.5.6)

Maximum 5.0.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (2.2.28).

Internal standard solution Dissolve 0.4 g of tetradecane R in hexane R and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 0.100 g of the substance to be examined in the internal standard solution and dilute to 10.0 mL with the same solution.

Reference solution Dissolve 0.100 g of octyldodecanol CRS in the internal standard solution and dilute to 10.0 mL with the same solution.

Column:

— material: stainless steel,

— size: $l = 60$ m, $\varnothing = 0.25$ mm,

— stationary phase: poly(dimethyl) (diphenyl) (divinyl)siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 0.68 mL/min.

Split ratio 1:50.

Temperature:

Time (min)	Temperature (°C)
0 - 2	180
2 - 22	180 \rightarrow 280
22 - 52	280
Injection port	290
Detector	300

Detection Flame ionisation.

Injection 1 μ L.

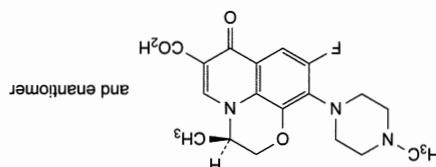
Calculate the content of $C_{20}H_{42}O$ in the substance to be examined.

STORAGE

Protected from light.

Ofloxacin

(Ph. Eur. monograph 1455)



$C_{18}H_{20}FN_3O_4$

361.4

82419-36-1

Action and use

Fluoroquinolone antibacterial.

DEFINITION

(3*R*S)-9-Fluoro-3-methyl-10-(4-methylpiperazin-1-yl)-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-*cd*]-1,4-benzoxazine-6-carboxylic acid.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture. Mix 10 mL of the impurity E CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Mix 10 mL of the solution and 5 mL of the test solution and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dissolve 10 mg of ofloxacin in the solvent mixture acetone/water R (10:60 V/V). Solvent mixture acetone/water R (10:60 V/V).

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Related substances
obtained with the reference solution (0.2 per cent).
— impurity A: any spot due to impurity A is not more intense than the corresponding spot in the chromatogram

Limit:

Detection Examine in ultraviolet light at 254 nm.

Drying In air.

Development Over 2/3 of the plate.

Application 10 μ L.

(10:10:20 V/V/V).

Mobile phase glacial acetic acid R, water R, ethyl acetate R

Plate TLC silica gel GF₂₅₄ plate R (2-10 μ m).

100.0 mL with the solvent mixture.

Impurity A CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution Dissolve 10.0 mg of ofloxacin

the solvent mixture.

Test solution Dissolve 0.250 g of the substance to be examined in the solvent mixture and dilute to 5.0 mL with

Solvent mixture methanol R, methylene chloride R (10:40 V/V).

Thin-layer chromatography (2.2.27).

Impurity A

100.0 mL with the same acid.

Dissolve 0.5 g in 0.1 M hydrochloric acid and dilute to 100.0 mL with the same acid.

Maximum 0.25 at 440 nm.

Absorbance (2.2.25)

with the same mixture of solvents.

and 40 volumes of methylene chloride R and dilute to 10.0 mL

Dissolve 0.300 g in a mixture of 10 volumes of methanol R

–0.10° to +0.10°.

Optical rotation (2.2.7)

TESTS

Comparison ofloxacin CRS.

Infrared absorption spectrophotometry (2.2.24).

IDENTIFICATION

methanol.

Slightly soluble in water, soluble in glacial acetic acid, slightly

soluble or soluble in methylene chloride, slightly soluble in

Solubility

Pale yellow or bright yellow, crystalline powder.

Appearance**CHARACTERS**

99.0 per cent to 101.0 per cent (dried substance).

Content

— stationary phase: octadecylsilyl silica gel for chromatography *R*
 — (5 µm);
 — temperature: 45 °C.
 Mobile phase Dissolve 4.0 g of ammonium acetate *R* and 7.0 g of sodium perchlorate *R* in 1300 mL of water *R*; adjust to pH 2.2 with phosphoric acid *R* and add 240 mL of acetonitrile *R*.

Flow rate Adjust so that a retention time of about 20 min is obtained for ofloxacin.

Detection Spectrophotometer at 294 nm.

Injection 10 µL.

Run time 2.5 times the retention time of ofloxacin.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to

impurity E.

Relative retention With reference to ofloxacin (retention

time = about 20 min): impurity B = about 0.3;

impurity C = about 0.5; impurity D = about 0.7;

impurity E = about 0.9; impurity F = about 1.6.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurity E and ofloxacin.

Limits:

— impurities B, C, D, E, F: for each impurity, not more than

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent);

— total: not more than 2.5 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.5 per cent);

— disregard limit: 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) *R*.

Loss on drying (2.2.32)

Maximum 0.2 per cent, determined on 1.000 g by drying at

105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 100 mL of anhydrous acetic acid *R*.

Titrate with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 36.14 mg

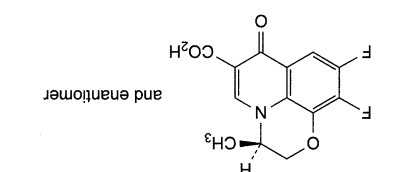
of C₁₈H₂₀FN₃O₄.

STORAGE

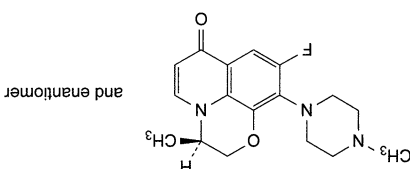
In an airtight container, protected from light.

IMPURITIES

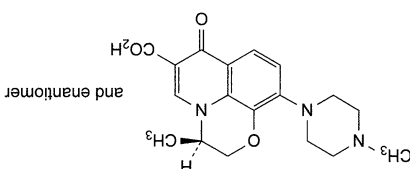
Specified impurities A, B, C, D, E, F



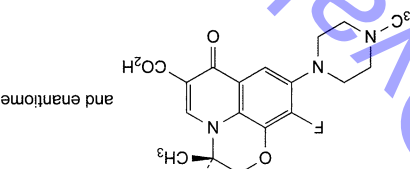
A. (3*R*)-9,10-difluoro-3-methyl-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazine-6-carboxylic acid (FPA), and enantiomer



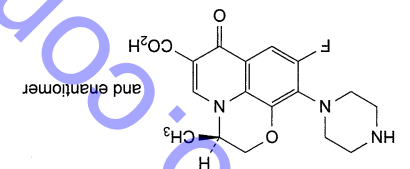
B. (3*R*)-9-fluoro-3-methyl-10-(4-methylpiperazin-1-yl)-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazine-7-one, and enantiomer



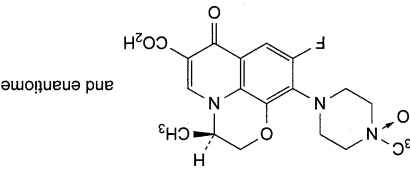
C. (3*R*)-3-methyl-10-(4-methylpiperazin-1-yl)-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazine-6-carboxylic acid, and enantiomer



D. (3*R*)-10-fluoro-3-methyl-9-(4-methylpiperazin-1-yl)-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazine-6-carboxylic acid, and enantiomer



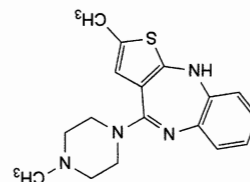
E. (3*R*)-9-fluoro-3-methyl-10-(piperazin-1-yl)-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazine-6-carboxylic acid, and enantiomer



F. 4-[(3*R*)-6-carboxy-9-fluoro-3-methyl-7-oxo-2,3-dihydro-7*H*-pyrido[1,2,3-*de*]-1,4-benzoxazin-10-yl]-1-methylpiperazine 1-oxide.

Olanzapine

(Ph. Eur. monograph 2258)



C₁₇H₂₀N₄S 312.4 132539-06-1

Action and use
Dopamine D₂ receptor antagonist; serotonin 5HT₂ receptor antagonist; neuroleptic.

Ph. Eur.

DEFINITION

2-Methyl-4-(4-methylpiperazin-1-yl)-10H-thieno[2,3-b][1,5]benzodiazepine.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, slightly soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison olanzapine CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in ethyl acetate, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the test and reference solutions immediately before use or keep them refrigerated and inject within 20 h of preparation.

Solution A Dissolve 13 g of sodium dodecyl sulfate R in about 1450 mL of water R, add 5 mL of phosphoric acid R and adjust to pH 2.5 by slowly adding strong sodium hydroxide solution R. If a precipitate is formed, this precipitate has to be re-dissolved prior to final pH adjustment. Dilute to 1500 mL with water R.

Solvent mixture Mix 4 volumes of acetonitrile R1 with 6 volumes of a 37 mg/L solution of sodium edetate R in solution A.

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 4 mg of olanzapine for system suitability CRS (containing impurities B, C and D) in 10.0 mL of the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm.

— stationary phase: octylsilyl silica gel for chromatography R

(5 μ m);

— temperature: 35 °C.

Mobile phase:

— mobile phase A: acetonitrile R1, solution A (48:52 V/V);

— mobile phase B: solution A, acetonitrile R1 (30:70 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 10	100	0
10 - 20	100 \rightarrow 0	0 \rightarrow 100
20 - 25	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied with olanzapine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C and D.

Relative retention With reference to olanzapine (retention time = about 13 min): impurity B = about 0.3;

impurity D = about 0.9; impurity C = about 1.2.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to impurity D and olanzapine.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity B by 0.4;

— impurities B, C, D: for each impurity, not more than 1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture water R, acetone R (10:90 V/V).

It complies with test H with the following modifications.

Test solution Dissolve 1.0 g of the substance to be examined in 60 mL of the solvent mixture.

Reference solution Dilute 1 mL of lead standard solution (10 ppm Pb) R to 60 mL with the solvent mixture.

Blank solution 60 mL of the solvent mixture.

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.250 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 2.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of olanzapine CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 2.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 1 mg of olanzapine impurity A CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Mix 1 volume of acetonitrile R with 1 volume of

a 6.9 g/L solution of sodium dihydrogen phosphate

monohydrate R adjusted to pH 2.5 with phosphoric acid R and

containing 12 g/L of sodium dodecyl sulfate R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 μ L.

Run time 1.2 times the retention time of olanzapine.

Relative retention With reference to olanzapine (retention

time = about 7 min): impurity A = about 0.8.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurity A and olanzapine.

Calculate the percentage content of $C_{17}H_{20}N_4S$ using the

chromatogram obtained with reference solution (a) and the

declared content of olanzapine CRS.

IMPURITIES

Specified impurities B, C, D

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

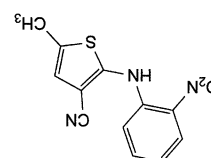
acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

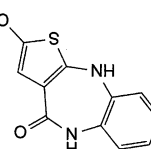
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

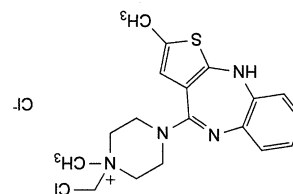
Control of impurities in substances for pharmaceutical use: A.



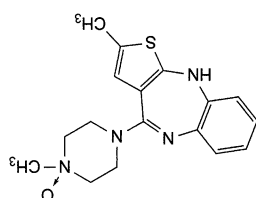
A. 5-methyl-2-[(2-nitrophenyl)amino]thiophene-3-carbonitrile,



B. 2-methyl-5,10-dihydro-4H-thieno[2,3-b][1,5]benzodiazepin-4-one,



C. 1-(chloromethyl)-1-methyl-4-(2-methyl-10H-thieno[2,3-b][1,5]benzodiazepin-4-yl)piperazin-1-ium chloride,



D. 1-methyl-4-(2-methyl-10H-thieno[2,3-b][1,5]benzodiazepin-4-yl)piperazin-1-oxide.

Ph Eur



112-80-1

Oleic Acid

(Ph. Eur. monograph 0799)

$C_{18}H_{34}O_2$

272.5

Action and use

Excipient.

DEFINITION

(Z)-Octadec-9-enoic acid ($C_{18}H_{34}O_2$; 282.5), together with varying amounts of saturated and other unsaturated fatty acids. A suitable antioxidant may be added.

Content

65.0 per cent to 88.0 per cent of $C_{18}H_{34}O_2$.

CHARACTERS

Appearance

Clear, yellowish or brownish, oily liquid.

Solubility

Practically insoluble in water, miscible with alcohol and with methylene chloride.

Relative density

About 0.892.

IDENTIFICATION

A. Acid value (see Tests).

B. Iodine value (see Tests).

C. Composition of fatty acids (see Tests).

Margarine acid Maximum 0.2 per cent for oleic acid of

vegetable origin and maximum 4.0 per cent for oleic acid of

animal origin.

TESTS

Appearance

The substance to be examined is not more intensely coloured than reference solution Y₁ or BY₁ (2.2.2, Method I).

Acid value (2.5.1)

195 to 204, determined on 0.5 g.

Iodine value (2.5.4)

89 to 105.

Peroxide value (2.5.5)

Maximum 10.0.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Test solution Prepare as described in the method but omitting the initial hydrolysis.

Composition of the fatty acid fraction of the substance:

— *myristic acid*: maximum 5.0 per cent,

— *palmitic acid*: maximum 16.0 per cent,

— *palmitoleic acid*: maximum 8.0 per cent.

1,3-diglycerides (R_f 0.7), to 1,2-diglycerides (R_f 0.6), to monoglycerides (R_f 0.1) and to esters of macrogol (R_f 0).

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Fatty acid composition (see Tests).

TESTS

Acid value (2.5.1)

Maximum 2.0, determined on 2.0 g.

Hydroxyl value (2.5.3, Method A)

45 to 65, determined on 1.0 g.

Iodine value (2.5.4, Method A)

75 to 95.

Peroxide value (2.5.5, Method A)

Maximum 12.0, determined on 2.0 g.

Saponification value (2.5.6)

150 to 170, determined on 2.0 g.

Alkaline impurities

Introduce 5.0 g into a test tube and carefully add a mixture,

neutralised if necessary with 0.01 M hydrochloric acid or with

0.01 M sodium hydroxide, of 0.05 mL of a 0.4 g/L solution of

bromophenol blue R in ethanol (96 per cent) R, 0.3 mL of

water R and 10 mL of ethanol (96 per cent) R. Shake and

allow to stand. Not more than 1.0 mL of 0.01 M hydrochloric

acid is required to change the colour of the upper layer to

Free glycerol

Maximum 3.0 per cent.

Dissolve 1.20 g in 25.0 mL of methylene chloride R. Heat if

necessary. After cooling, add 100 mL of water R. Shake and

add 25.0 mL of periodic acetic acid solution R. Shake and allow

to stand for 30 min. Add 40 mL of a 75 g/L solution of

potassium iodide R. Allow to stand for 1 min. Add 1 mL of

starch solution R. Titrate the iodine with 0.1 M sodium

thiosulfate. Carry out a blank titration.

1 mL of 0.1 M sodium thiosulfate is equivalent to 2.3 mg of

glycerol.

Composition of fatty acids (2.4.22, Method A)

Composition of the fatty-acid fraction of the substance:

— palmitic acid: 4.0 per cent to 9.0 per cent;

— stearic acid: maximum 6.0 per cent;

— oleic acid: 58.0 per cent to 80.0 per cent;

— linoleic acid: 15.0 per cent to 35.0 per cent;

— linolenic acid: maximum 2.0 per cent;

— arachidic acid: maximum 2.0 per cent;

— eicosenoic acid: maximum 2.0 per cent.

Ethylene oxide and dioxan (2.4.25)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm

of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.0 g. Use a mixture

of 30 volumes of anhydrous methanol R and 70 volumes of

methylene chloride R as solvent.

Total ash (2.4.16)

Maximum 0.1 per cent.

STORAGE

Protected from light.

Oleoyl Macrogolglycerides

(Ph. Eur. monograph 1249)

Action and use

Excipient.

DEFINITION

Mixtures of monoesters, diesters and triesters of glycerol and

monoesters and diesters of macrogols.

They are obtained by partial alcoholysis of an unsaturated oil

mainly containing triglycerides of oleic (*cis*-9-octadecenoic)

acid, using macrogol with a mean relative molecular mass

between 300 and 400, or by esterification of glycerol and

macrogl with unsaturated fatty acids, or by mixing glycerol

esters and condensates of ethylene oxide with the fatty acids

CHARACTERS

Appearance

Amber oily liquid, which may give rise to a deposit after

prolonged periods at 20 °C.

Solubility

Practically insoluble but dispersible in water, freely soluble in

methyle chloride.

Viscosity About 35 mPa.s at 40 °C.

Relative density

About 0.95 at 20 °C.

Refractive index About 1.47 at 20 °C.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 1.0 g of the substance to be examined

in methylene chloride R and dilute to 20 mL with the same

solvent.

Plate TLC silica gel plate R.

Mobile phase hexane R, ether R (30:70 V/V).

Application 10 μ L.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of rhodamine B in

ethanol (96 per cent) R and examine in ultraviolet light at

365 nm.

Results The chromatogram shows a spot due to triglycerides

with an R_f value of about 0.9 (R_f 1) and spots due to



Ph Eur

LABELLING

The label states the type of macrogol used (mean relative molecular mass) or the number of units of ethylene oxide per molecule (nominal value).

Ph Eur



Oleyl Alcohol

(Ph. Eur. monograph 2073)

Action and use

Nonionic surfactant.

Ph Eur

DEFINITION

Mixture of unsaturated and saturated long-chain fatty alcohols consisting mainly of octadec-9-enol (oleyl alcohol and elaidyl alcohol; $C_{18}H_{36}O$; M_r 268.5). It may be of vegetable or animal origin.

CHARACTERS

Appearance

Colourless or light yellow liquid.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Composition of fatty alcohols (see Tests).

TESTS

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Refractive index (2.2.6)

1.458 to 1.461, determined at 25 °C.

Cloud point

Maximum 10 °C.

Introduce about 60 g into a cylindrical flat-bottomed container, 30-33.5 mm in internal diameter and 115-125 mm high. Heat to 30 °C, cool, and immerse the container in iced water with the surfaces of the water and the sample at the same level. Insert a thermometer and, using it as a stirring rod, begin stirring rapidly and steadily when the temperature falls below 20 °C. Keep the thermometer immersed throughout the test, and remove and examine the container at regular intervals. The cloud point is the temperature at which the immersed portion of the thermometer, positioned vertically in the centre of the container, is no longer visible when viewed horizontally through the container and sample.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

205 to 215.

Saponification value (2.5.6)

Maximum 2.0.

Composition of fatty alcohols

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution Mix 25 mg of the substance to be examined with 1.0 mL of methylene chloride R.

Reference solution (a) Dissolve 25 mg of each of arachidyl alcohol R, linoleyl alcohol R, linoleyl alcohol R, oleyl alcohol R, palmityl alcohol R and stearyl alcohol R in methylene chloride R

and dilute to 5 mL with the same solvent. Dilute 1 mL of this solution to 5 mL with methylene chloride R.

Reference solution (b) Dissolve 10 mg of linoleyl alcohol R and 1 g of oleyl alcohol R in methylene chloride R and dilute to 40 mL with the same solvent.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 1 μ m).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 1:11.

Temperature:

Time (min)	Temperature (°C)
0 - 1	170
1 - 9	170 → 210
9 - 65	210
Injection port	270
Detector	280

Detection Flame ionisation.

Injection 1 μ L.

Identify the peaks using the chromatogram obtained with reference solution (a).

Relative retention With reference to oleyl alcohol (retention time = about 30 min): palmityl alcohol = about 0.6;

linoleyl alcohol = about 0.8; linoleyl alcohol = about 0.9;

stearyl alcohol = about 1.1; arachidyl alcohol = about 1.9

(elaidyl alcohol co-elutes with oleyl alcohol).

System suitability: reference solution (b):

— H_p = height above the peak due to linoleyl alcohol and above the baseline of the peak from the lowest point of the curve separating this peak from the peak due to oleyl alcohol.

— H_v = height above the peak due to linoleyl alcohol and above the baseline of the peak due to oleyl alcohol

— H_p = height above the peak due to linoleyl alcohol and above the baseline of the peak due to oleyl alcohol

— H_v = height above the peak due to linoleyl alcohol and above the baseline of the peak due to oleyl alcohol

— H_p = height above the peak due to linoleyl alcohol and above the baseline of the peak due to oleyl alcohol

— H_v = height above the peak due to linoleyl alcohol and above the baseline of the peak due to oleyl alcohol

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— H_v = height above the peak due to linoleyl alcohol and above the baseline of the peak due to oleyl alcohol



Virgin Olive Oil

Olive Oil

(Ph. Eur. monograph 0518)

Preparation

Olive Oil Ear Drops

Ph Eur

DEFINITION

Fatty oil obtained by cold expression or other suitable mechanical means from the ripe drupes of *Olea europaea* L.

CHARACTERS**Appearance**

Clear, transparent, yellow or greenish-yellow liquid.

Solubility

Practically insoluble in ethanol (96 per cent), miscible with light petroleum (bp: 50-70 °C).

When cooled, it begins to become cloudy at 10 °C and becomes a butter-like mass at about 0 °C.

Relative density

About 0.913.

IDENTIFICATION

Identification of fatty oils by thin-layer chromatography

(2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1. For certain types of olive oil, the difference in the size of spots E and F is less pronounced than in the corresponding chromatogram shown in Figure 2.3.2.-1.

TESTS**Absorbance (2.2.25)**

Maximum 0.20 at 270 nm. The ratio of the absorbance at 232 nm to that at 270 nm is greater than 8.

To 1.00 g add cyclohexane R and dilute to 100.0 mL with the same solvent.

Acid value (2.5.1)

Maximum 2.0, determined on 5.0 g.

Peroxide value (2.5.5, Method A)

Maximum 20.0.

Unsaponifiable matter

Maximum 1.5 per cent.

Place 5.0 g (m) in a 150 mL flask fitted with a reflux

condenser. Add 50 mL of 2 M alcoholic potassium hydroxide R and heat on a water-bath for 1 h, shaking frequently.

Add 50 mL of water R through the top of the condenser, shake, allow to cool and transfer the contents of the flask to a separating funnel. Rinse the flask with several portions

totalling 50 mL of light petroleum R1 and add the rinsings to the separating funnel. Shake vigorously for 1 min. Allow to

separate and transfer the aqueous layer to a 2nd separating funnel. If an emulsion forms, add small quantities of ethanol

(96 per cent) R or a concentrated solution of potassium hydroxide R. Shake the aqueous layer with 2 quantities, each

of 50 mL, of light petroleum R1. Combine the light petroleum layers in a 3rd separating funnel and wash with 3 quantities,

each of 50 mL, of ethanol (50 per cent V/V) R. Transfer the light petroleum layer to a tared 250 mL flask. Rinse the

separating funnel with small quantities of light petroleum R1 and add to the tared flask. Evaporate the light petroleum on a

water-bath and dry the residue at 100-105 °C for 15 min, keeping the flask horizontal. Allow to cool in a desiccator and

weigh (a g). Repeat the drying for successive periods of 15 min until the loss of mass between 2 successive weighings

does not exceed 0.1 per cent. Dissolve the residue in 20 mL of ethanol (96 per cent) R, previously neutralised to 0.1 mL of

bromophenol blue solution R. If necessary, titrate with 0.1 M hydrochloric acid (b mL).

Calculate the percentage content of unsaponifiable matter using the following expression:

$$100(a - 0.032b) / m$$

Relative density

About 0.913.

When cooled, it begins to become cloudy at 10 °C and becomes a butter-like mass at about 0 °C.

light petroleum (bp: 50-70 °C).

Practically insoluble in ethanol (96 per cent), miscible with

Solubility

Clear, colourless or greenish-yellow transparent liquid.

Appearance**CHARACTERS**

added.

ripe drupes of *Olea europaea* L. A suitable antioxidant may be

added.

cold expression or other suitable mechanical means from the

Partly oil obtained by refining of crude olive oil, obtained by

DEFINITION

(Ph. Eur. monograph 1456)

Refined Olive Oil

Ph Eur

In a well-filled container, protected from light, at a temperature not exceeding 25 °C.

STORAGE

Maximum 0.1 per cent, determined on 1.00 g.

Water (2.5.32)

acid R. No bluish-green colour develops.

with acetic anhydride R. To the filtrate add 0.2 mL of sulfuic

acid R. No bluish-green colour develops.

acid R. No bluish-green colour develops.

acid R. No bluish-green colour develops.

acid R. No bluish-green colour develops.

acid R. No bluish-green colour develops.

acid R. No bluish-green colour develops.

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acid R. No bluish-green colour develops.

acid R. No bluish-green colour develops.

II-432 Olmesartan Medoxomil

IDENTIFICATION

A. Acid value (see Tests).

B. Identification of fatty oils by thin-layer chromatography (2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2-1.

For certain types of olive oil, the difference in the size of spots E and F is less pronounced than in the corresponding chromatogram shown in Figure 2.3.2-1.

TESTS

Specific absorbance (2.2.25)

Maximum 1.20, determined at the absorption maximum at 270 nm.

To 1.00 g add cyclohexane R and dilute to 100.0 mL with the same solvent.

Acid value (2.5.1)

Maximum 0.3, determined on 10.0 g.

Peroxide value (2.5.5, Method A)

Maximum 10.0, or maximum 5.0 if intended for use in the manufacture of parenteral preparations.

Unasaponifiable matter

Maximum 1.5 per cent.

Place 5.0 g (*m*) in a 150 mL flask fitted with a reflux condenser. Add 50 mL of 2 M alcoholic potassium hydroxide R and heat on a water-bath for 1 h, shaking frequently.

Add 50 mL of water R through the top of the condenser, shake, allow to cool and transfer the contents of the flask to a separating funnel. Rinse the flask with several portions totalling 50 mL of light petroleum R1 and add the rinsings to the separating funnel. Shake vigorously for 1 min. Allow to separate and transfer the aqueous layer to a 2nd separating funnel. If an emulsion forms, add small quantities of ethanol (96 per cent) R or a concentrated solution of potassium hydroxide R. Shake the aqueous layer with 2 quantities, each of 50 mL, of light petroleum R1. Combine the light petroleum layers in a 3rd separating funnel and wash with 3 quantities, each of 50 mL, of ethanol (50 per cent V/V) R. Transfer the light petroleum layer to a tared 250 mL flask. Rinse the separating funnel with small quantities of light petroleum R1 and add to the flask. Evaporate the light petroleum on a water-bath and dry the residue at 100–105 °C for 15 min, keeping the flask horizontal. Allow to cool in a desiccator and weigh (*a* g). Repeat the drying for successive periods of 15 min until the loss of mass between 2 successive weighings does not exceed 0.1 per cent. Dissolve the residue in 20 mL of ethanol (96 per cent) R, previously neutralised to 0.1 mL of bromophenol blue solution R. If necessary, titrate with 0.1 M hydrochloric acid (*b* mL).

Calculate the percentage content of unasaponifiable matter using the following expression:

$$\frac{100(a - 0.032b)}{m}$$

If 0.032*b* is greater than 5 per cent of *a*, the test is not valid and must be repeated.

Alkaline impurities (2.4.19)

It complies with the test.

Composition of fatty acids (2.4.22, Method A)

Use the mixture of calibrating substances in Table 2.4.22-3. Composition of the fatty-acid fraction of the oil:
— saturated fatty acids of chain length less than C₁₆: maximum 0.1 per cent;

IDENTIFICATION

A. Acid value (see Tests).

B. Identification of fatty oils by thin-layer chromatography (2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2-1.

For certain types of olive oil, the difference in the size of spots E and F is less pronounced than in the corresponding chromatogram shown in Figure 2.3.2-1.

TESTS

Specific absorbance (2.2.25)

Maximum 1.20, determined at the absorption maximum at 270 nm.

To 1.00 g add cyclohexane R and dilute to 100.0 mL with the same solvent.

Acid value (2.5.1)

Maximum 0.3, determined on 10.0 g.

Peroxide value (2.5.5, Method A)

Maximum 10.0, or maximum 5.0 if intended for use in the manufacture of parenteral preparations.

Unasaponifiable matter

Maximum 1.5 per cent.

Place 5.0 g (*m*) in a 150 mL flask fitted with a reflux condenser. Add 50 mL of 2 M alcoholic potassium hydroxide R and heat on a water-bath for 1 h, shaking frequently.

Add 50 mL of water R through the top of the condenser, shake, allow to cool and transfer the contents of the flask to a separating funnel. Rinse the flask with several portions totalling 50 mL of light petroleum R1 and add the rinsings to the separating funnel. Shake vigorously for 1 min. Allow to separate and transfer the aqueous layer to a 2nd separating funnel. If an emulsion forms, add small quantities of ethanol (96 per cent) R or a concentrated solution of potassium hydroxide R. Shake the aqueous layer with 2 quantities, each of 50 mL, of light petroleum R1. Combine the light petroleum layers in a 3rd separating funnel and wash with 3 quantities, each of 50 mL, of ethanol (50 per cent V/V) R. Transfer the light petroleum layer to a tared 250 mL flask. Rinse the separating funnel with small quantities of light petroleum R1 and add to the flask. Evaporate the light petroleum on a water-bath and dry the residue at 100–105 °C for 15 min, keeping the flask horizontal. Allow to cool in a desiccator and weigh (*a* g). Repeat the drying for successive periods of 15 min until the loss of mass between 2 successive weighings does not exceed 0.1 per cent. Dissolve the residue in 20 mL of ethanol (96 per cent) R, previously neutralised to 0.1 mL of bromophenol blue solution R. If necessary, titrate with 0.1 M hydrochloric acid (*b* mL).

Calculate the percentage content of unasaponifiable matter using the following expression:

$$\frac{100(a - 0.032b)}{m}$$

If 0.032*b* is greater than 5 per cent of *a*, the test is not valid and must be repeated.

Alkaline impurities (2.4.19)

It complies with the test.

Composition of fatty acids (2.4.22, Method A)

Use the mixture of calibrating substances in Table 2.4.22-3. Composition of the fatty-acid fraction of the oil:
— saturated fatty acids of chain length less than C₁₆: maximum 0.1 per cent;

PREPARATION

Preparation

Olmesartan Tablets

Action and use

Angiotensin II (AT₁) receptor antagonist.

Chemical name

C₂₉H₃₀N₆O₆

Molecular weight

558.6

Ph Eur

144689-63-4

Chemical structure

Labeling

The label states:
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations;
— the name of the inert gas.

STORAGE

In a well-filled container, protected from light, at a temperature not exceeding 25 °C. If intended for use in the manufacture of parenteral preparations, store under an inert gas.

LABELLING

The label states:
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations;
— the name of the inert gas.

Sesame oil

The content of stigmasterol is not greater than that of campesterol.

sum of contents of Δ⁵,23-stigmastadienol, clerosterol, β-sitosterol, sitostanol, Δ⁵-avenasterol and Δ⁵,24-stigmastadienol: minimum 93.0 per cent.

Δ⁷-stigmastenol: maximum 0.5 per cent;
campesterol: maximum 4.0 per cent;
cholesterol: maximum 0.5 per cent;

Composition of the sterol fraction of the oil:

Sterols (2.4.23, Method B)

— lignoceric acid: maximum 0.2 per cent.
— behenic acid: maximum 0.2 per cent;
— eicosenoic acid: maximum 0.4 per cent;
— arachidic acid: maximum 0.7 per cent;
— linolenic acid: maximum 1.2 per cent;
— linoleic acid: 3.5 per cent to 20.0 per cent;
— oleic acid: 56.0 per cent to 85.0 per cent;
— stearic acid: 0.5 per cent to 5.0 per cent;
— palmitoleic acid: maximum 3.5 per cent;
— palmitic acid: 7.5 per cent to 20.0 per cent;

Water (2.5.32)

Maximum 0.1 per cent, determined on 1.00 g.

acid R. No bluish-green colour develops.

with acetic anhydride R. To the filtrate add 0.2 mL of sulfonic acid R. Filter through a filter paper impregnated with acetic anhydride R and 4.5 mL of solution of fufural R in acetic anhydride R and 4.5 mL of 1 min with a mixture of 0.5 mL of a 0.35 per cent V/V In a ground-glass-stoppered cylinder shake 10 mL for about

USP

Identification

A. Acid value (see Tests).

B. Identification of fatty oils by thin-layer chromatography (2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2-1.

For certain types of olive oil, the difference in the size of spots E and F is less pronounced than in the corresponding chromatogram shown in Figure 2.3.2-1.

TESTS

Specific absorbance (2.2.25)

Maximum 1.20, determined at the absorption maximum at 270 nm.

To 1.00 g add cyclohexane R and dilute to 100.0 mL with the same solvent.

Acid value (2.5.1)

Maximum 0.3, determined on 10.0 g.

Peroxide value (2.5.5, Method A)

Maximum 10.0, or maximum 5.0 if intended for use in the manufacture of parenteral preparations.

Unasaponifiable matter

Maximum 1.5 per cent.

Place 5.0 g (*m*) in a 150 mL flask fitted with a reflux condenser. Add 50 mL of 2 M alcoholic potassium hydroxide R and heat on a water-bath for 1 h, shaking frequently.

Add 50 mL of water R through the top of the condenser, shake, allow to cool and transfer the contents of the flask to a separating funnel. Rinse the flask with several portions totalling 50 mL of light petroleum R1 and add the rinsings to the separating funnel. Shake vigorously for 1 min. Allow to separate and transfer the aqueous layer to a 2nd separating funnel. If an emulsion forms, add small quantities of ethanol (96 per cent) R or a concentrated solution of potassium hydroxide R. Shake the aqueous layer with 2 quantities, each of 50 mL, of light petroleum R1. Combine the light petroleum layers in a 3rd separating funnel and wash with 3 quantities, each of 50 mL, of ethanol (50 per cent V/V) R. Transfer the light petroleum layer to a tared 250 mL flask. Rinse the separating funnel with small quantities of light petroleum R1 and add to the flask. Evaporate the light petroleum on a water-bath and dry the residue at 100–105 °C for 15 min, keeping the flask horizontal. Allow to cool in a desiccator and weigh (*a* g). Repeat the drying for successive periods of 15 min until the loss of mass between 2 successive weighings does not exceed 0.1 per cent. Dissolve the residue in 20 mL of ethanol (96 per cent) R, previously neutralised to 0.1 mL of bromophenol blue solution R. If necessary, titrate with 0.1 M hydrochloric acid (*b* mL).

Calculate the percentage content of unasaponifiable matter using the following expression:

$$\frac{100(a - 0.032b)}{m}$$

If 0.032*b* is greater than 5 per cent of *a*, the test is not valid and must be repeated.

Alkaline impurities (2.4.19)

It complies with the test.

Composition of fatty acids (2.4.22, Method A)

Use the mixture of calibrating substances in Table 2.4.22-3. Composition of the fatty-acid fraction of the oil:
— saturated fatty acids of chain length less than C₁₆: maximum 0.1 per cent;

DEFINITION

(5-Methyl-2-oxo-1,3-dioxol-4-yl)methyl 4-(1-hydroxy-1-methylethyl)-2-propyl-1-[[2'-(1*H*-tetrazol-5-yl)biphenyl-4-yl]methyl]-1*H*-imidazole-5-carboxylate.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in heptane.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison olmesartan medoxomil CRS.

TESTS**Related substances**

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 25 mg of the substance to be examined in acetonitrile R and dilute to 25.0 mL with the same solvent.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in acetonitrile R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of olmesartan medoxomil CRS (containing impurities A, B and C) in acetonitrile R and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 50.0 mL with acetonitrile R. Dilute 1.0 mL of this solution to 10.0 mL with acetonitrile R.

Reference solution (c) Dissolve 25.0 mg of olmesartan medoxomil CRS in acetonitrile R and dilute to 50.0 mL with the same solvent.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.6$ mm;

— stationary phase: spherical end-capped octylsilyl silica gel for chromatography R (3.5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of 2.04 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

— mobile phase A: mix 20 volumes of acetonitrile R and 80 volumes of acetonitrile R;

— mobile phase B: mix 20 volumes of a 2.04 g/L solution of potassium dihydrogen phosphate R, previously adjusted to pH 3.4 with a 1.73 g/L solution of phosphoric acid R;

Identification of impurities Use the chromatogram supplied with olmesartan medoxomil for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C.

Relative retention With reference to olmesartan medoxomil (retention time = about 10 min): impurity A = about 0.2; impurity B = about 0.7; impurity C = about 1.5.

System suitability: reference solution (a):

— resolution: minimum 3.5 between the peaks due to impurity B and olmesartan medoxomil.

Limits:

— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— impurity C: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);

— impurity B: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Acetone

Head-space gas chromatography (2.2.28) Use the direct calibration method.

Internal standard solution Dilute 1.0 mL of butanol R to 100.0 mL with dimethyl sulfoxide R.

Test solution Dissolve 0.250 g of the substance to be examined in dimethyl sulfoxide R, add 2.0 mL of the internal standard solution and dilute to 10.0 mL with dimethyl sulfoxide R.

Reference solution Dilute 0.50 mL of acetone R to 200.0 mL with dimethyl sulfoxide R. Dilute 15.0 mL of the solution to 100.0 mL with dimethyl sulfoxide R. To 25.0 mL of this solution add 10.0 mL of the internal standard solution and dilute to 50.0 mL with dimethyl sulfoxide R.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.53$ mm;

— stationary phase: macrogol 20 000 R (film thickness 1 μ m).

Carrier gas nitrogen for chromatography R or helium for chromatography R.

Flow rate 4.0 mL/min.

Split ratio 1:5.

Static head-space conditions that may be used:

— equilibration temperature: 80 °C;

— equilibration time: 30 min.

Temperature:

Column

Time (min)

Temperature (°C)

50

50 → 180

180

18 - 23

Injection port

200

200

Detection

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

200

Detection Flame ionisation.
Injection 1 mL.
 Calculate the content of acetone, taking its relative density to be 0.79 at 20 °C.
Limit:
 — acetone: maximum 0.6 per cent.
Heavy metals (2.4.8)
 Maximum 20 ppm.
Solvent mixture water R, dimethyl sulfoxide R (10:90 V/V).
 1.0 g complies with test H. Prepare the test solution with the aid of ultrasound. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.32)
 Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

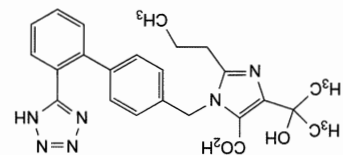
Mobile phase Mobile phase B, mobile phase A (25:75 V/V).
Injection Test solution (b) and reference solution (c).
Retention time Olmesartan medoxomil = about 10 min.
Run time 1.5 times the retention time of olmesartan medoxomil.

Calculate the percentage content of $C_{29}H_{30}N_6O_6$ taking into account the assigned content of olmesartan medoxomil CRS.

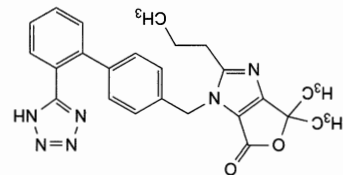
IMPURITIES

Specified impurities A, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, D.



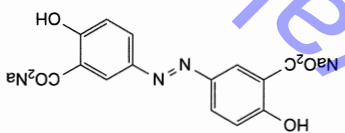
A. 4-(1-hydroxy-1-methylethyl)-2-propyl-1-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]-1H-imidazole-5-carboxylic acid (olmesartan),



B. 6,6-dimethyl-2-propyl-3-[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]-3,6-dihydro-4H-furo[3,4-d]imidazole-4-one,

Olsalazine Sodium

(Ph. Eur. monograph 1457)



Action and use

Aminosalicylate; treatment of ulcerative colitis.

DEFINITION

Disodium 3,3'-diazenediylbis(6-hydroxybenzoate).

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, fine, crystalline powder.

Solubility

Sparingly soluble in water, soluble in dimethyl sulfoxide, very slightly soluble in methanol.

It shows polymorphism (5.9).

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 40.0 mg in 5 mL of 0.1 M sodium hydroxide and dilute to 100.0 mL with a 7.8 g/L solution of

Ph Eur

6054-98-4



— stationary phase: ion-exclusion resin for chromatography R with a capacity of about 27 meq/column.

Suppressor column.

Mobile phase 0.0001 M hydrochloric acid.

Flow rate 0.9 mL/min.

Detection Conductivity detector at $10 \mu\text{S}\cdot\text{cm}^{-1}$.

Injection 0.1 mL.

System suitability: reference solution (a):

— the chromatogram shows 3 separated peaks.

Determine the concentration of acetate in the test solution using the calibration curve generated by the average of the readings obtained with the reference solutions. Measure the peak area for acetate. Calculate the percentage content of acetate using the following expression:

$$\frac{2.6}{m} \cdot c$$

c = concentration of acetate in the test solution, in

micrograms per millilitre, determined by linear

interpolation of the standard curve for reference

solution (b);

m = mass of sample, in milligrams.

Limit:

— acetate: maximum 1.0 per cent.

Methanesulfonic acid

Liquid chromatography (2.2.29).

Test solution Dissolve 0.25 g of the substance to be examined in 20 mL of water R, add 1.0 mL of dilute hydrochloric acid R

and dilute to 25.0 mL with water R. Centrifuge and then

filter the solution through a 0.45 μm filter and also through

an appropriate filter for removal of chlorides.

Reference solution (a) Dissolve 0.25 g of methanesulfonic acid R

in 50 mL of water R. Add 0.58 g of sodium acetate R and

0.08 g of sodium chloride R and dilute to 100.0 mL with

water R. Dilute 1.0 mL of this solution to 100.0 mL with

water R.

Reference solution (b) Dissolve 0.10 g of methanesulfonic acid R

in water R and dilute to 100.0 mL with water R. Dilute

3.0 mL of this solution to 100.0 mL with water R.

Precolumn:

— size: $l = 0.035 \text{ m}$, $\emptyset = 4 \text{ mm}$;

— stationary phase: resin for reversed-phase ion

chromatography R (10 μm).

Column:

— size: $l = 0.25 \text{ m}$, $\emptyset = 4 \text{ mm}$;

— stationary phase: resin for reversed-phase ion

chromatography R (10 μm).

Mobile phase Mix 10 volumes of acetonitrile for

chromatography R and 990 volumes of a solution containing

1.6 g/L of tetrabutylammonium hydroxide R and 0.053 g/L of

anhydrous sodium carbonate R.

Flow rate 1.0 mL/min.

Detection Conductivity detector at $50 \mu\text{S}\cdot\text{cm}^{-1}$.

Injection 100 μL .

System suitability: reference solution (a):

— the chromatogram shows 3 separated peaks.

Limit:

— methanesulfonic acid: not more than the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (0.3 per cent).

sodium dihydrogen phosphate R adjusted to pH 7.2 with strong sodium hydroxide solution R (buffer solution). Dilute 2.0 mL

of the solution to 100.0 mL with the buffer solution.

Spectral range 240 nm to 400 nm.

Absorption maxima At 255 nm and 362 nm.

Absorbance ratio $A_{255}/A_{362} = 0.53$ to 0.56.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison olsalazine sodium CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of 1 volume of dilute ammonia R2 and 4 volumes of ethanol (96 per cent) R and dilute to 10 mL with the same

mixture of solvents.

Reference solution (a) Dissolve 10 mg of olsalazine sodium CRS

in a mixture of 1 volume of dilute ammonia R2 and 4 volumes

of ethanol (96 per cent) R and dilute to 10 mL with the same

mixture of solvents.

Reference solution (b) Dissolve 5 mg of sulfasalazine CRS in

reference solution (a) and dilute to 5 mL with reference

solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase anhydrous formic acid R, acetone R, methylene

chloride R (5:50:60 V/V/V).

Application 10 μL .

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 separated spots.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with reference

solution (a).

D. To 0.5 g add 2 mL of sulfuric acid R. Progressively heat to

ignition and continue heating until an almost white or at

most greyish residue is obtained. Carry out the ignition at a

temperature up to $800 \pm 50^\circ\text{C}$. Dissolve the residue in

10 mL of boiling water R and filter. 2 mL of the filtrate gives

reaction (a) of sodium (2.3.1).

TESTS

Acetate

Liquid chromatography (2.2.29).

Test solution Dissolve 0.125 g of the substance to be

examined in 25.0 mL of water R and add 1.0 mL of dilute

hydrochloric acid R. Centrifuge and then filter the solution

through a 0.45 μm filter and also through an appropriate

filter for removal of chlorides.

Reference solution (a) Dissolve 0.140 g of sodium acetate R,

0.150 g of sodium formate R and 0.180 g of potassium sulfate R

in 100.0 mL of water R. Dilute 1.0 mL of this solution to

100.0 mL with water R.

Reference solution (b) Use suitable amounts of sodium acetate R

to prepare not fewer than 5 reference solutions containing

10–50 $\mu\text{g/mL}$ of acetate.

Column:

— size: $l = 0.25 \text{ m}$, $\emptyset = 9 \text{ mm}$.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in mobile phase A and dilute to 25.0 mL with

mobile phase A.

Reference solution (a) Dilute 0.5 mL of the test solution to

100.0 mL with mobile phase A.

Reference solution (b) Dissolve 20.0 mg of olsalazine sodium for

performance test CRS in mobile phase A and dilute to

25.0 mL with mobile phase A.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m);

— temperature: 30 °C.

Mobile phase:

— mobile phase A: dissolve 2.38 g of tetrabutylammonium

hydrogen sulfate R and 3.6 g of disodium hydrogen phosphate

dihydrate R in 900 mL of water R, adjust to pH 7.6 with

dilute sodium hydroxide solution R and dilute to 1000.0 mL

with water R; mix 700 mL of this buffer solution with

300 mL of methanol R;

— mobile phase B: dissolve 4.75 g of tetrabutylammonium

hydrogen sulfate R and 3.6 g of disodium hydrogen phosphate

dihydrate R in 900 mL of water R, adjust to pH 7.6 with

dilute sodium hydroxide solution R and dilute to 1000.0 mL

with water R; mix 350 mL of this buffer solution with

650 mL of methanol R;

Time

(min)

Mobile phase A (per cent V/V)

Mobile phase B (per cent V/V)

0 - 15 55 45

15 - 45 55 \rightarrow 0 45 \rightarrow 100

45 - 50 0 \rightarrow 55 100 \rightarrow 45

50 - 65 55 45

Flow rate 1 mL/min.

Detection Spectrophotometer at 360 nm.

Injection 20 μ L.

System suitability: reference solution (b):

— the chromatogram is similar to the chromatogram

obtained with olsalazine sodium for performance test CRS.

Limits:

— impurities A, B, C, D, E, F, G, H, I: for each impurity,

not more than twice the area of the principal peak in the

chromatogram obtained with reference solution (a)

(1.0 per cent), and not more than one of the peaks has an

area greater than the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.5 per cent);

— total: not more than 4 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(2.0 per cent);

— disregard limit: 0.05 times the area of the principal peak in

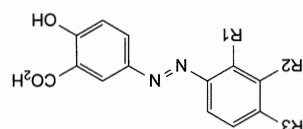
the chromatogram obtained with reference solution (a)

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.



Specified impurities: A, B, C, D, E, F, G, H, I.

IMPURITIES

C₁₄H₈N₂NaO₆.

1 mL of 0.1 M hydrochloric acid is equivalent to 17.31 mg of

taking the molecular mass of acetate to be 59.0.

Correct the volume consumed for the content of acetate,

titration.

the end-point potentiometrically (2.2.20). Carry out a blank

chloride R. Titrate with 0.1 M hydrochloric acid, determining

dioxan R and 0.2 mL of a 224 g/L solution of potassium

Dissolve 0.100 g in 15 mL of ethylene glycol R. Add 40 mL of

ASSAY

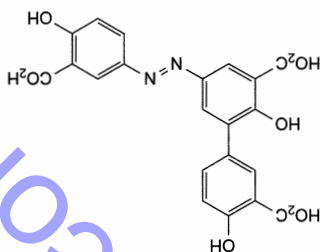
an oven at 150 °C.

Maximum 2.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

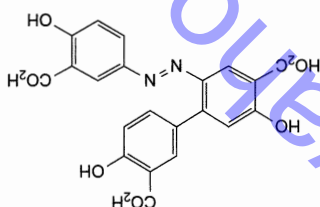
G. 5-[(3-carboxy-4-hydroxyphenyl)diazenyl]-2,4'-

dihydroxybiphenyl-3,3'-dicarboxylic acid,



F. 2'-[(3-carboxy-4-hydroxyphenyl)diazenyl]-4,5'-

dihydroxybiphenyl-3,4'-dicarboxylic acid,



E. R₁ = H, R₂ = CO-CH₂-SO₃H, R₃ = OH: 2-hydroxy-5-

6-chloro-6'-hydroxy-3,3'-diazenediylidibenzoic acid,

D. R₁ = H, R₂ = CO₂H, R₃ = Cl:

2-hydroxy-5-[(4-hydroxyphenyl)diazenyl]benzoic acid,

C. R₁ = R₂ = H, R₃ = OH:

2,6'-dihydroxy-3,3'-diazenediylidibenzoic acid,

B. R₁ = OH, R₂ = CO₂H, R₃ = H:

6-hydroxy-6'-methoxy-3,3'-diazenediylidibenzoic acid,

A. R₁ = H, R₂ = CO₂H, R₃ = OCH₃:

Solubility
Practically insoluble in water, very soluble in acetone, in ethanol (96 per cent), in heptane and in methanol.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay for EPA and DHA ethyl esters.

Results The peaks due to eicosapentaenoic acid ethyl ester and docosahexaenoic acid ethyl ester in the chromatogram obtained with test solution (b) are similar in retention time to the corresponding peaks in the chromatograms obtained with reference solutions (a₁) and (a₂).

B. It complies with the limits of the assay for total omega-3-acid ethyl esters.

TESTS

Absorbance (2.2.25)

Maximum 0.60 at 233 nm.

Dilute 0.300 g to 50.0 mL with trimethylpentane R. Dilute

2.0 mL of the solution to 50.0 mL with trimethylpentane R.

Acid value (2.5.1)

Maximum 2.0, determined on 10 g in 50 mL of the

prescribed mixture of solvents.

Anisidine value (2.5.36)

Maximum 20.0.

Peroxide value (2.5.5, Method A)

Maximum 10.0.

Oligomers and partial glycerides

Size-exclusion chromatography (2.2.30).

Test solution Dilute 50.0 mg of the substance to be examined to 10.0 mL with tetrahydrofuran R.

Reference solution Dissolve 50 mg of monodocosahexaenoic R,

30 mg of didocosahexaenoic R and 20 mg of

tridocosahexaenoic R in tetrahydrofuran R and dilute to

100.0 mL with the same solvent.

Column 3 columns to be connected in series:

— size: $l = 0.3$ m, $\varnothing = 7.8$ mm;

— stationary phase: styrene-divinylbenzene copolymer R (5 μ m)

with the following pore sizes:

— column 1: 50 nm;

— column 2: 10 nm;

— column 3: 5 nm;

— connection sequence: injector – column 1 – column 2 –

column 3 – detector.

Mobile phase tetrahydrofuran R.

Flow rate 0.8 mL/min.

Detection Differential refractometer.

Injection 40 μ L.

System suitability: reference solution:

— elution order: tridocosahexaenoic, didocosahexaenoic,

monodocosahexaenoic;

— resolution: minimum 2.0 between the peaks due to

didocosahexaenoic and monodocosahexaenoic;

minimum 1.0 between the peaks due to

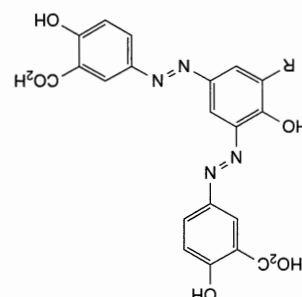
tridocosahexaenoic and didocosahexaenoic.

Calculate the percentage content of oligomers plus partial

glycerides using the following expression:

$$\frac{A}{B} \times 100$$

A = sum of the areas of all the peaks in the chromatogram;



H. R = CO₂H; 3,3',5'-carboxy-4-hydroxy-1,3-phenylenebis(diazenediyl)]bis(6-hydroxybenzoic) acid, I. R = H; 3,3',5'-[4-hydroxy-1,3-phenylenebis(diazenediyl)]bis(6-hydroxybenzoic) acid.

Ph Eur



Omega-3-Acid Ethyl Esters 60

(Ph. Eur. monograph 2063)

Action and use
Lipid-regulating drug.

Ph Eur

DEFINITION

Ethyl esters of α -linolenic acid (C18:3 n-3), morocic acid (C18:4 n-3), eicosatetraenoic acid (C20:4 n-3), timnodonic (eicosapentaenoic) acid (C20:5 n-3; EPA), henicosapentaenoic acid (C21:5 n-3), cupanodonic acid (C22:5 n-3) and ceronic (docosahexaenoic) acid (C22:6 n-3; DHA). Omega-3-acid ethyl esters 60 are obtained by transesterification of the body oil obtained from fish of families such as *Engraulidae*, *Carangidae*, *Clupeidae*, *Osmenidae*, *Salmonidae* and *Scombridae* or from animals of the class *Cephalopoda* and subsequent physico-chemical purification processes, including molecular distillation. The minimum content of total omega-3-acid ethyl esters and the minimum content of the omega-3-acids EPA and DHA ethyl esters are indicated in Table 2063.-1.

Table 2063.-1

Total omega-3-acid ethyl esters	EPA and DHA ethyl esters	EPA ethyl esters	DHA ethyl esters	Minimum content (per cent)
65	50	25	20	40
60	50	-	40	-
55	50	40	-	-

A suitable antioxidant may be added.

PRODUCTION

The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS

Appearance
Light yellow liquid.

Slight fish-like odour.

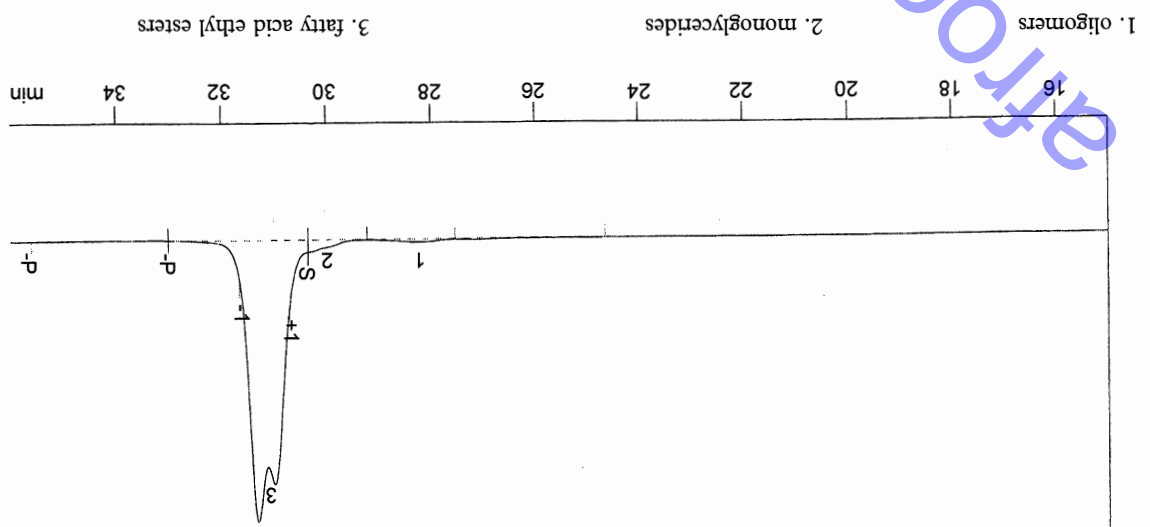


Figure 2063.-1. - Chromatogram for the test for oligomers and partial glycerides in omega-3-acid ethyl esters 60

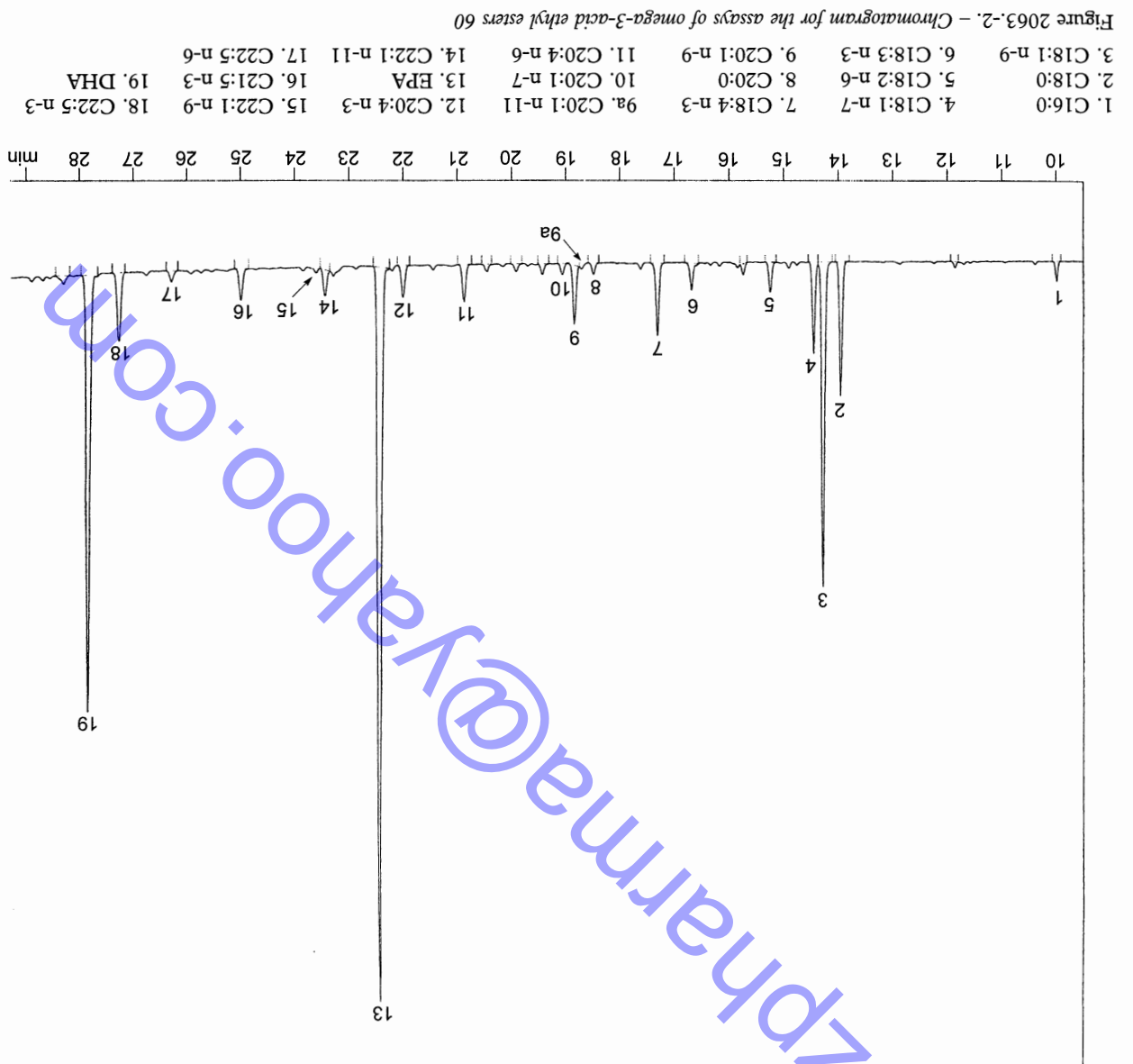


Figure 2063.-2. - Chromatogram for the assays of omega-3-acid ethyl esters 60

IDENTIFICATION

A. Examine the chromatograms obtained in the assay for EPA and DHA ethyl esters.

Results The peaks due to eicosapentaenoic acid ethyl ester and docosahexaenoic acid ethyl ester in the chromatogram obtained with test solution (b) are similar in retention time to the corresponding peaks in the chromatograms obtained with reference solutions (a₁) and (a₂).

B. It complies with the limits of the assay for total omega-3-acid ethyl esters.

TESTS

Absorbance (2.2.25)
Maximum 0.55 at 233 nm.

Dilute 0.300 g to 50.0 mL with trimethylpentane R. Dilute 2.0 mL of the solution to 50.0 mL with trimethylpentane R.

Acid value (2.5.1)
Maximum 2.0, determined on 10 g in 50 mL of the prescribed mixture of solvents.

Anisidine value (2.5.36)
Maximum 20.0.

Peroxide value (2.5.5, Method A)
Maximum 10.0.

Oligomers
Size-exclusion chromatography (2.2.30).

ASSAY

EPA and DHA ethyl esters (2.4.29)
For identification of the peaks, see Figure 2063.-2.

Total omega-3-acid ethyl esters (2.4.29)
See Figure 2063.-2.

STORAGE
Under an inert gas, in an airtight container, protected from light.

LABELLING
The label states:
— the content of total omega-3-acid ethyl esters;
— the content of EPA ethyl ester and DHA ethyl ester.

(Ph. Eur. monograph 1250)

Action and use
Lipid-regulating drug.

Ph. Eur.

Omega-3-Acid Ethyl Esters 90

DEFINITION
Ethyl esters of *alpha*-linolenic acid (C18:3 n-3), *morotic* acid (C18:4 n-3), *eicosatetraenoic acid* (C20:4 n-3), *timnodonic* (eicosapentaenoic acid) (C20:5 n-3; EPA), *heneicosapentaenoic acid* (C21:5 n-3), *clupanodonic acid* (C22:5 n-3) and *cervonic* (docosahexaenoic acid) (C22:6 n-3; DHA). Omega-3-acid ethyl esters are obtained by transesterification of the body oil obtained from fish of families such as *Engraulidae*, *Carangidae*, *Clupeidae*, *Osmenidae*, *Salmonidae* and *Scombridae* or from animals of the class *Cephalopoda* and subsequent physico-chemical purification processes, including urea fractionation followed by molecular distillation.

Content
— EPA and DHA ethyl esters: minimum 80 per cent, with minimum 40 per cent of EPA ethyl esters and minimum 34 per cent of DHA ethyl esters;
— total omega-3-acid ethyl esters: minimum 90 per cent.

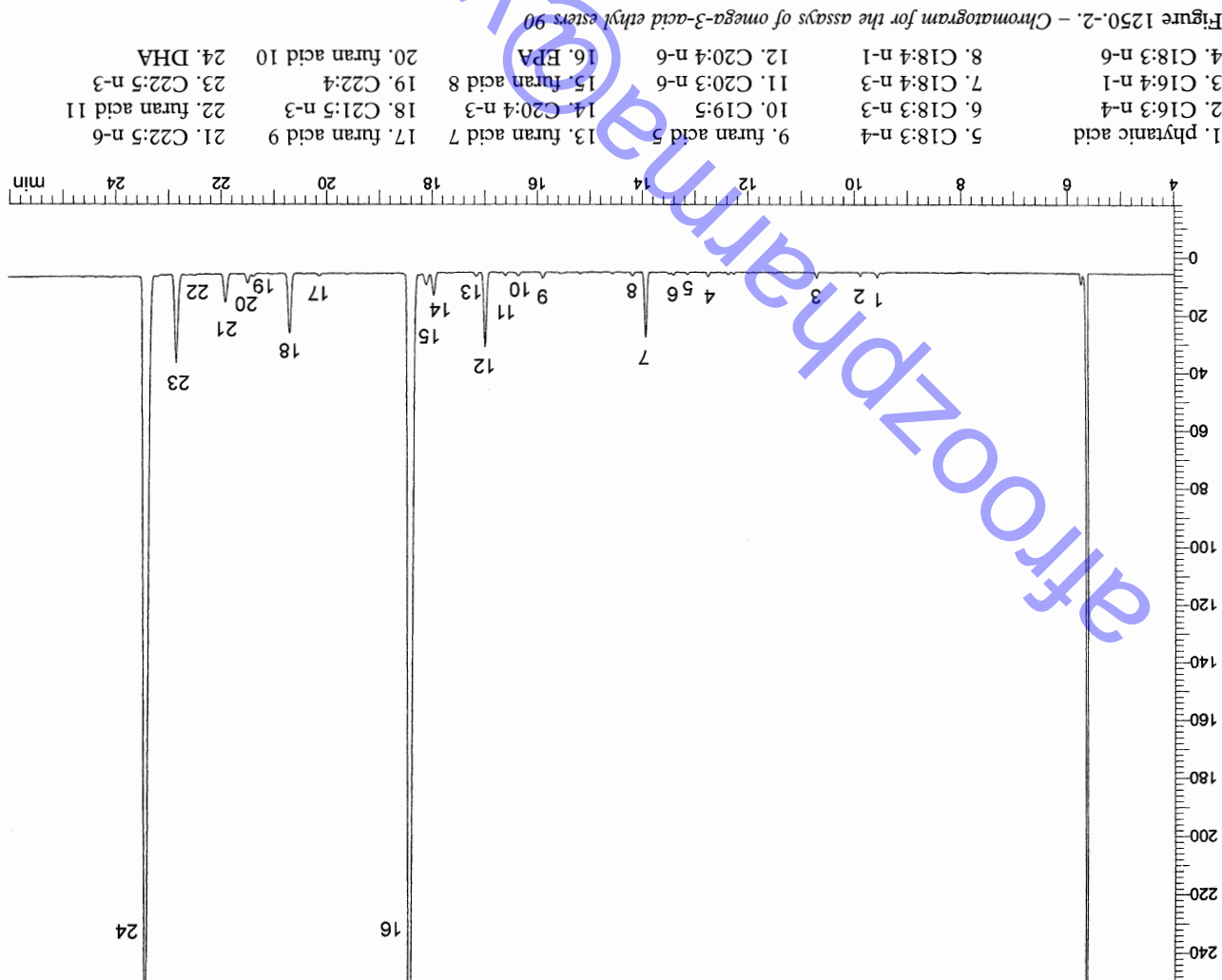
A suitable antioxidant may be added.

PRODUCTION
The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS
Appearance
Light yellow liquid.

Solubility
Practically insoluble in water, very soluble in acetone, in ethanol (96 per cent), in heptane and in methanol.





Test solution. Weigh 50.0 mg of the substance to be examined into a quartz tube. Add 1.5 mL of a 20 g/L solution of sodium hydroxide R in methanol R, cover with nitrogen R, cap tightly with a polytetrafluoroethylene-lined cap, mix and heat on a water-bath for 7 min. Allow to cool. Add 2 mL of boron trichloride-methanol solution R, cover with nitrogen R, cap tightly, mix and heat on a water-bath for 30 min. Cool to 40-50 °C, add 1 mL of trimethylpentane R, cap and shake vigorously for at least 30 s. Immediately add 5 mL of saturated sodium chloride solution R, cover with nitrogen R, cap and shake thoroughly for at least 15 s. Transfer the upper layer to a separate tube. Shake the methanol layer once more with 1 mL of trimethylpentane R. Wash the combined trimethylpentane extracts with 2 quantities, each of 1 mL, of water R. Carefully evaporate the solvent under a current of nitrogen R then add 10.0 mL of tetrahydrofuran R to the residue. Add a small amount of anhydrous sodium sulfate R and filter.

Test solution. Dilute 50.0 mg of the substance to be examined to 10.0 mL with tetrahydrofuran R.

Reference solution. Dissolve 50 mg of monodocosahexaenoic R, 30 mg of didocosahexaenoic R and 20 mg of tridocosahexaenoic R in tetrahydrofuran R and dilute to 100.0 mL with the same solvent.

Column 3 columns to be connected in series: — size: $l = 0.3$ m, $\phi = 7.8$ mm;

Column 3 columns to be connected in series:

size: $l = 0.3$ m, $\phi = 7.8$ mm;

A = sum of the areas of all the peaks in the chromatogram;

$$\frac{A}{B} \times 100$$

following expression:

Calculate the percentage content of oligomers using the minimum 1.0 between the peaks due to

tridocosahexaenoic and didocosahexaenoic.

minimum 2.0 between the peaks due to

didocosahexaenoic and monodocosahexaenoic;

— resolution: minimum 2.0 between the peaks due to

monodocosahexaenoic;

— elution order: tridocosahexaenoic, didocosahexaenoic,

System suitability: reference solution:

Injection 40 μ L.

Detection Differential refractometer.

Flow rate 0.8 mL/min.

Mobile phase tetrahydrofuran R.

column 3 – detector.

— connection sequence: injector – column 1 – column 2 –

column 3: 5 nm;

— column 2: 10 nm;

— column 1: 50 nm;

with the following pore sizes:

— stationary phase: styrene-divinylbenzene copolymer R (5 μ m)



Omega-3-Marine Triglycerides

(Omega-3-Acid Triglycerides,
Ph Eur monograph 1352)

Action and use

Lipid-regulating drug.

Ph Eur

DEFINITION

Mixture of mono-, di- and triesters of omega-3 acids with glycerol, containing mainly triesters and obtained either by esterification of concentrated and purified omega-3 acids with glycerol or by transesterification of the omega-3 acid ethyl esters with glycerol. The origin of the omega-3 acids is the body oil obtained from fish of families such as *Engraulidae*, *Carangidae*, *Chupeidae*, *Osmenidae*, *Salmonidae* and *Scombridae* or from animals of the class *Cephalopoda*. The omega-3 acids are identified as the following acids: *alpha*-linolenic acid (C18:3 n-3), moroctic acid (C18:4 n-3), eicosatetraenoic acid (C20:4 n-3), timnodonic (eicosapentaenoic) acid (C20:5 n-3; EPA), heneicosapentaenoic acid (C21:5 n-3), clupanodonic acid (C22:5 n-3) and ceronic (docosahexaenoic) acid (C22:6 n-3; DHA).

Content

— sum of the contents of the omega-3 acids EPA and DHA, expressed as triglycerides: minimum 45 per cent;
— total omega-3 acids, expressed as triglycerides: minimum 60 per cent.
A suitable antioxidant may be added.

PRODUCTION

The content of dioxins and dioxin-like PCBs (polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

CHARACTERS

Appearance

Pale yellow liquid.

Solubility

Practically insoluble in water, very soluble in acetone and in heptane, slightly soluble in anhydrous ethanol.

IDENTIFICATION

Examine the chromatograms obtained in the assay for EPA and DHA.

Results The peaks due to eicosapentaenoic acid methyl ester and docosahexaenoic acid methyl ester in the chromatogram obtained with test solution (b) are similar in retention time to the corresponding peaks in the chromatograms obtained with reference solutions (a₁) and (a₂).

TESTS

Absorbance (2.2.25)

Maximum 0.73 at 233 nm.

Dilute 0.300 g to 50.0 mL with trimethylpentane R. Dilute 2.0 mL of the solution to 50.0 mL with trimethylpentane R. Acid value (2.5.1)

Maximum 3.0, determined on 10.0 g in 50 mL of the

prescribed mixture of solvents.

Anisidine value (2.5.36)

Maximum 30.0.

Peroxide value (2.5.5, Method A)

Maximum 10.0.

B = sum of the areas of the peaks with a retention time less than the retention time of the peaks due to ethyl esters.

The ethyl ester peaks, which may be present in the form of an unresolved double peak, are identified as the major peaks in the chromatogram (see Figure 1250.-1).

Where the result obtained exceeds the limit due to the presence of monoglycerides, the following procedure is carried out.

Test solution. Weigh 50.0 mg of the substance to be examined into a quartz tube. Add 1.5 mL of a 20 g/L solution of sodium hydroxide R in methanol R, cover with nitrogen R, cap tightly with a polytetrafluoroethylene-lined cap, mix and heat on a water-bath for 7 min. Allow to cool. Add 2 mL of boron trichloride-methanol solution R, cover with nitrogen R, cap tightly, mix and heat on a water-bath for 30 min. Cool to 40–50 °C, add 1 mL of trimethylpentane R, cap and shake vigorously for at least 30 s. Immediately add 5 mL of saturated sodium chloride solution R, cover with nitrogen R, cap and shake thoroughly for at least 15 s. Transfer the upper layer to a separate tube. Shake the methanol layer once more with 1 mL of trimethylpentane R. Wash the combined trimethylpentane extracts with 2 quantities, each of 1 mL, of nitrogen R then add 10.0 mL of tetrahydrofuran R to the residue. Add a small amount of anhydrous sodium sulfate R and filter.

Calculate the percentage content of oligomers using the following expression:

$$\frac{B}{A} \times 100$$

A = sum of the areas of all the peaks in the

chromatograms;

B' = sum of the areas of the peaks with a retention time less than the retention time of the peaks due to

methyl esters.

Limit:

— oligomers: maximum 1.0 per cent.

ASSAY

EPA and DHA ethyl esters (2.4.29)

For identification of the peaks, see Figure 1250.-2.

Total omega-3-acid ethyl esters (2.4.29)

See Figure 1250.-2.

STORAGE

Under an inert gas, in an airtight container, protected from

light.

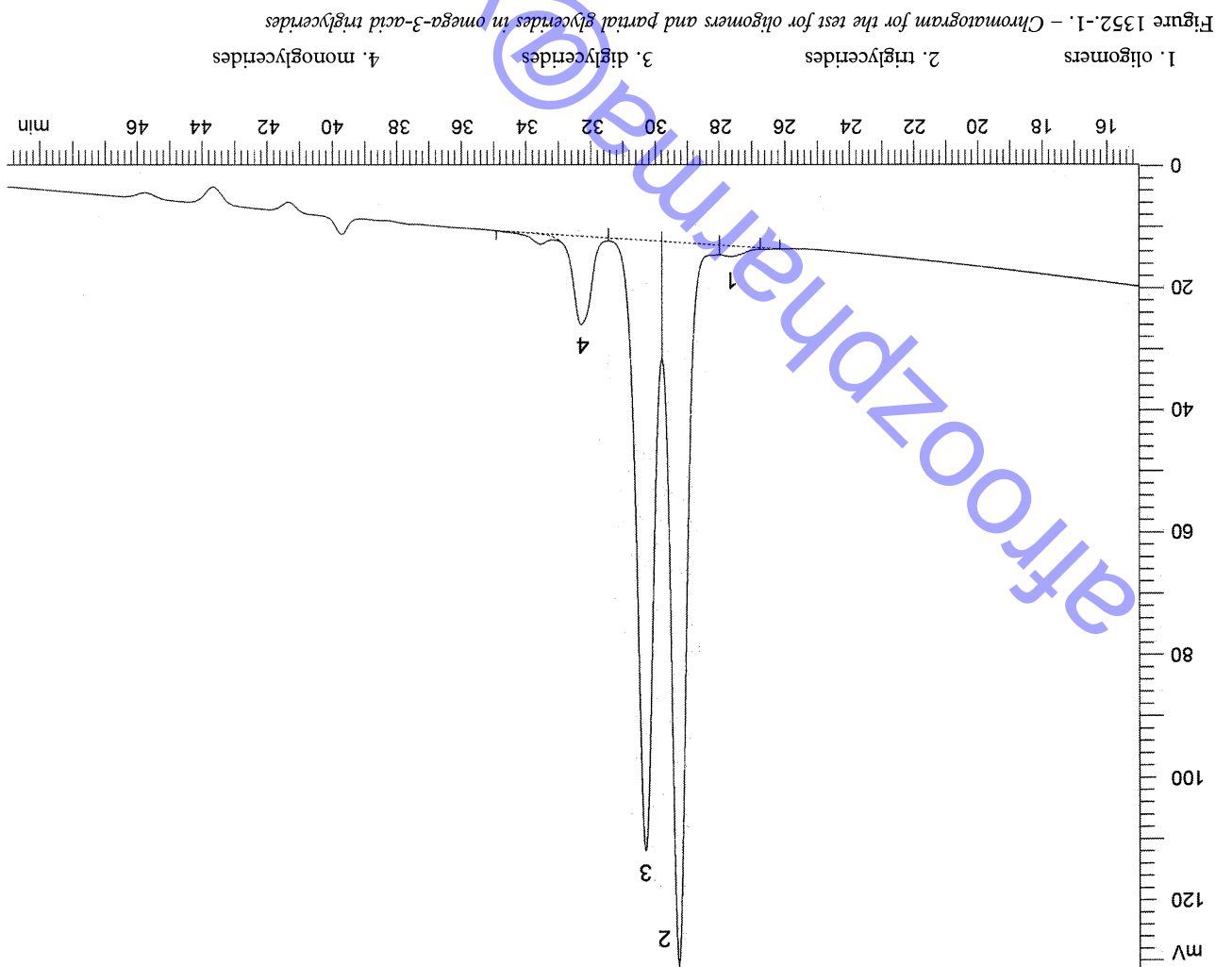
LABELLING

The label states:

— The content of total omega-3-acid ethyl esters;

— The content of EPA ethyl ester and DHA ethyl ester.

Ph Eur



Oligomers and partial glycerides

Size-exclusion chromatography (2.2.30).

Test solution Dilute 50.0 mg of the substance to be examined

to 10.0 mL with tetrahydrofuran R.

Reference solution Dissolve 50 mg of monodocosahexaenoic R₃

30 mg of didocosahexaenoic R and 20 mg of

tridocosahexaenoic R in tetrahydrofuran R and dilute to

100.0 mL with the same solvent.

Column:

3 columns to be connected in series:

— size: $l = 0.3$ m, $\varnothing = 7.8$ mm;

— stationary phase: styrene-divinylbenzene copolymer R (5 μ m)

with the following pore sizes:

— column 1: 50 nm;

— column 2: 10 nm;

— column 3: 5 nm;

— connection sequence: injector – column 1 – column 2 –

column 3 – detector.

Mobile phase tetrahydrofuran R.

Flow rate 0.8 mL/min.

Detection Differential refractometer.

Injection 40 μ L.

System suitability: reference solution:

— elution order: tridocosahexaenoic, didocosahexaenoic,

monodocosahexaenoic;

— resolution: minimum 2.0 between the peaks due to

didocosahexaenoic and monodocosahexaenoic; minimum

1.0 between the peaks due to tridocosahexaenoic and

didocosahexaenoic.

Identify the peaks using the chromatogram shown in

Figure 1352.-1. Calculate the percentage content of

oligomers using the following expression:

$$\frac{B}{A} \times 100$$

A = sum of the areas of all the peaks in the

chromatogram;

B = area of the peak with a retention time less than the

retention time of the peak due to the triglycerides.

Calculate the percentage content of partial glycerides using

the following expression:

$$\frac{C}{A} \times 100$$

A = sum of the areas of all the peaks in the

chromatogram;

C = (sum of the area(s) of the peak(s) due to the mono-

and diglycerides.

Limits:

— oligomers: maximum 3.0 per cent;

— partial glycerides: maximum 50.0 per cent.

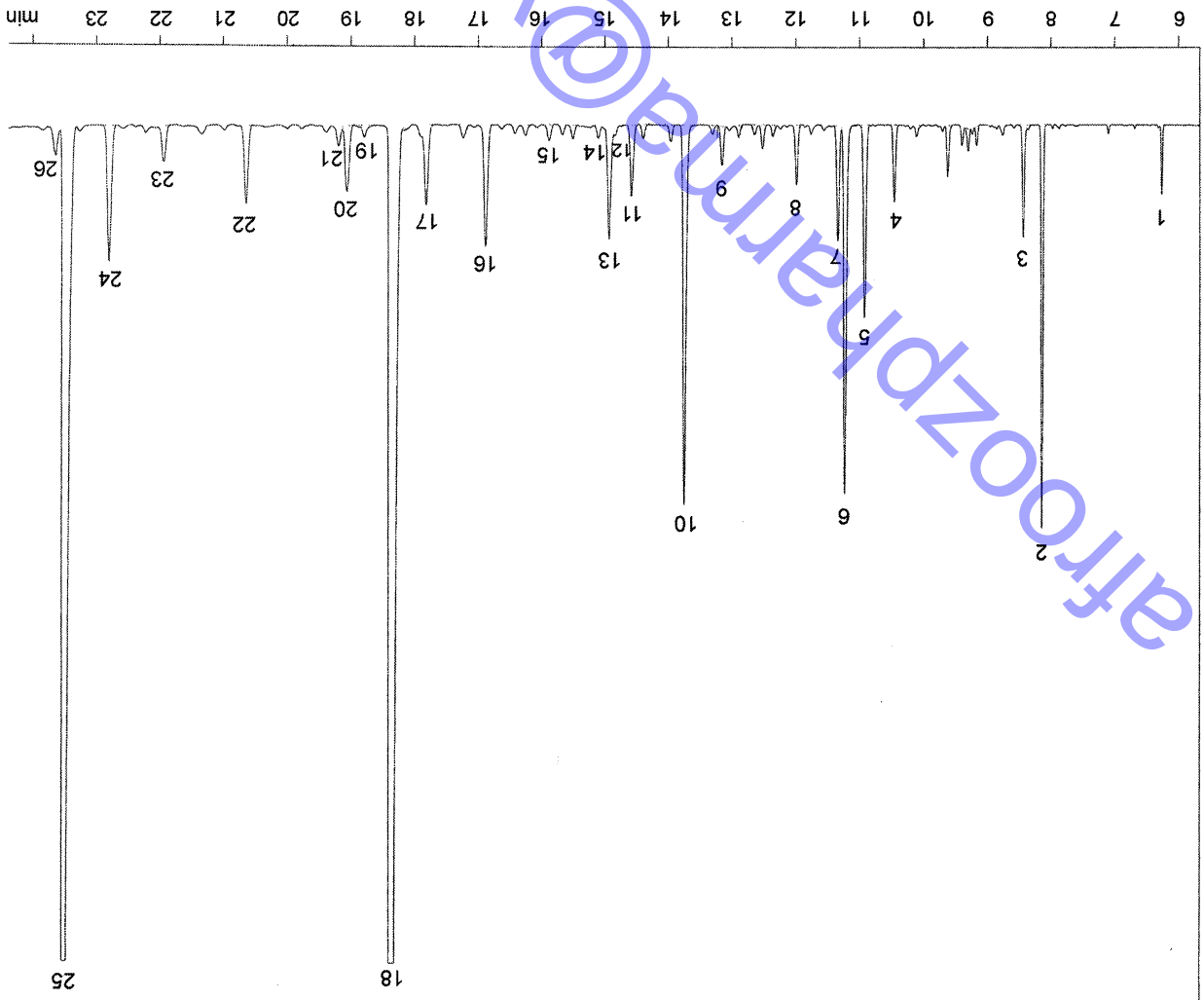
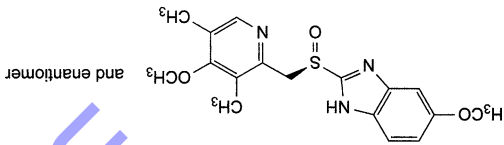


Figure 1352.-2. - Chromatogram for the assay of omega-3-acids in omeprazole

1. C14:0	4. C16:4 n-1	7. C18:1 n-7	10. C18:4 n-3	13. C20:1	16. C20:4	19. C22:0	22. C21:5	25. DHA
2. C16:0	5. C18:0	8. C18:2 n-6	11. C20:0	14. C20:1	17. C20:4	20. C22:1	23. C22:5	26. C24:1 n-9
3. C16:1	6. C18:1 n-9	9. C18:3 n-3	12. C20:1	15. C20:2	18. EPA	21. C22:1	24. C22:5	
n-7	n-11	n-6	n-11	n-7	n-3	n-11	n-6	

Omeprazole

(Ph. Eur. monograph 0942)



Under an inert gas, in a well-filled, airtight container, protected from light.

STORAGE

See Figure 1352.-2.

Total omega-3-acids (2.4.29)

For identification of the peaks, see Figure 1352.-2.

EPA and DHA (2.4.29)

ASSAY

Action and use

Proton pump inhibitor; treatment of peptic ulcer disease.

Preparations

Gastro-resistant Omeprazole Capsules

Omeprazole Oral Suspension

Gastro-resistant Omeprazole Tablets

C₁₇H₁₉N₃O₃S

345.4

73590-58-6

DEFINITION

5-Methoxy-2-[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-1H-benzimidazole.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, soluble in methylene chloride, sparingly soluble in ethanol (96 per cent) and in methanol.

It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison omepرازوله CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol, evaporate to dryness and record new spectra using the residues.

TESTS

Solution S

Dissolve 0.50 g in methylene chloride R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1).

Impurities F and G

Maximum 350 ppm for the sum of the contents.

The absorbance (2.2.25) of solution S determined at 440 nm is not greater than 0.10.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 3 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 1 mg of omepرازوله CRS and 1 mg of omepرازوله impurity D CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 3 mg of omepرازوله for peak identification CRS (containing impurity E) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 27 volumes of acetonitrile R and 73 volumes of a 1.4 g/L solution of disodium hydrogen phosphate R previously adjusted to pH 7.6 with phosphoric acid R.

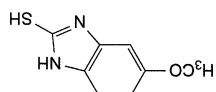
Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 40 μ L.

Run time 5 times the retention time of omepرازوله.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D; use the chromatogram supplied with omepرازوله



A. 5-methoxy-1H-benzimidazole-2-thiol,

Specified impurities D, E, F, G
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, H, I.

IMPURITIES

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

STORAGE

1 mL of 0.1 M sodium hydroxide is equivalent to 34.54 mg of $C_{17}H_{19}N_3O_3S$.
(2.2.20).

Dissolve 0.250 g in a mixture of 10 mL of water R and 40 mL of ethanol (96 per cent) R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.000 g by drying under high vacuum at 60 °C for 4 h.

Loss on drying (2.2.32)

1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) — impurities D, E: for each impurity, not more than 0.15 per cent; unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent); total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent); disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— resolution.
of acetonitrile R; an increase in the pH will improve the resolution.
impurity D and omepرازوله; if necessary, adjust the pH of the aqueous part of the mobile phase or the concentration of acetonitrile R; an increase in the pH will improve the resolution.

System suitability: reference solution (a):

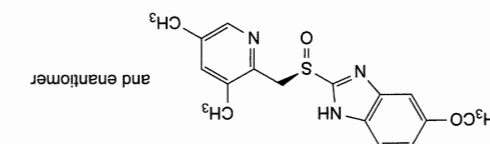
— resolution: minimum 3.0 between the peaks due to impurity D = about 0.8.

time = about 9 min): impurity E = about 0.6;

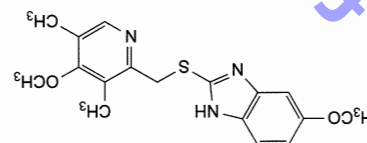
Relative retention With reference to omepرازوله (retention

impurity E.

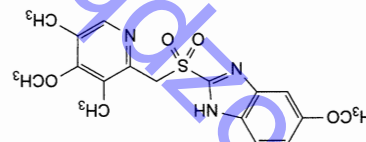
for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peak due to



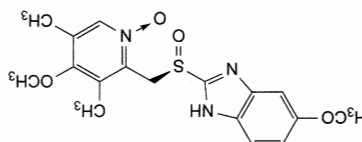
B. 2-[(R,S)-(3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-5-methoxy-1H-benzimidazole,



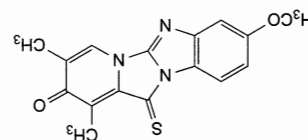
C. 5-methoxy-2-[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-1H-benzimidazole (omeprazole),



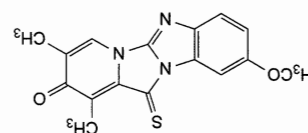
D. 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfonyl]-1H-benzimidazole (omeprazole sulfone),



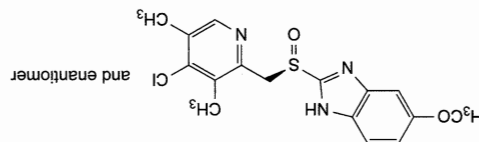
E. 4-methoxy-2-[[[(R,S)-(5-methoxy-1H-benzimidazol-2-yl)sulfinyl]methyl]-3,5-dimethylpyridine 1-oxide,



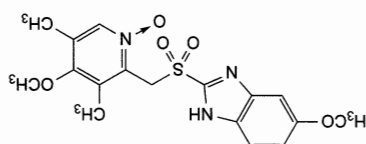
F. 8-methoxy-1,3-dimethyl-12-thioxopyrido[1',2':3,4]imidazo[1,2-a]benzimidazol-2(12H)-one,



G. 9-methoxy-1,3-dimethyl-12-thioxopyrido[1',2':3,4]imidazo[1,2-a]benzimidazol-2(12H)-one,



H. 2-[(R,S)-(4-chloro-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-5-methoxy-1H-benzimidazole,

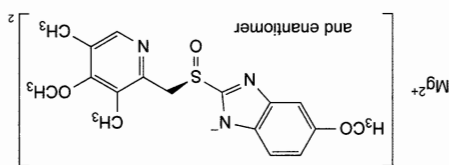


I. 4-methoxy-2-[[[(5-methoxy-1H-benzimidazol-2-yl)sulfonyl]methyl]-3,5-dimethylpyridine 1-oxide,

Ph Eur

Omeprazole Magnesium

(Ph. Eur. monograph 2374)



$C_{34}H_{36}MgN_6O_{10}S_2$ 713 95382-33-5

Action and use

Proton pump inhibitor; treatment of peptic ulcer disease.

Ph Eur

DEFINITION

Magnesium bis[5-methoxy-2-[(R,S)-(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-1H-benzimidazol-1-ide]. It contains a variable quantity of water.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Very slightly soluble in water, sparingly soluble in methanol, practically insoluble in heptane.

IDENTIFICATION

Carry out either tests A, B, C or tests A, B, D.

A. Optical rotation (2.2.7): -0.10° to $+0.10^\circ$.

Dissolve 0.250 g in methanol R and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison omeprazole magnesium CRS.

C. Atomic absorption spectrometry (2.2.23) as described in the test for magnesium.

The test solution shows the absorption maximum at 285.2 nm.

D. Ignite about 0.5 g of the substance to be examined according to the procedure for the sulfated ash test (2.4.14). Dissolve the residue in 10 mL of water R. 2 mL of this solution gives the reaction of magnesium (2.3.1).

TESTS

Absorbance (2.2.25)

Maximum 0.10 at 440 nm.

Dissolve 0.500 g in methanol R and dilute to 25.0 mL with the same solvent. Filter the solution through a membrane filter (nominal pore size 0.45 µm).

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure. Prepare the solutions immediately before use.

Test solution Dissolve 3.5 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 1 mg of omeprazole CRS and 1 mg of omeprazole impurity D CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 3 mg of omeprazole for peak identification CRS (containing impurity E) in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 27 volumes of acetonitrile R and 73 volumes of a 1.4 g/L solution of disodium hydrogen phosphate R

previously adjusted to pH 7.6 with phosphoric acid R. Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 40 μ L.

Run time 5 times the retention time of omeprazole.

Identification of impurities:

— use the chromatogram supplied with omeprazole for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity E;

— use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D = about 0.8.

— use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to omeprazole (retention time = about 9 min): impurity E = about 0.6,

impurity D = about 0.8.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to impurity D and omeprazole; if necessary, adjust the pH of the aqueous part of the mobile phase or its proportion of acetonitrile; an increase in the pH will improve the resolution.

Limits:

— impurities D, E: for each impurity, maximum 0.1 per cent; unspecified impurities: for each impurity, maximum 0.10 per cent;

— total: maximum 0.5 per cent;

— chromatogram obtained with reference solution (c) (0.05 per cent).

Magnesium

3.30 per cent to 3.55 per cent (anhydrous substance).

Atomic absorption spectrometry (2.2.23, Method D).

Test solution Dissolve 0.250 g in 20.0 mL of a 103 g/L solution of hydrochloric acid R by slow addition of the acid and dilute to 100.0 mL with water R. Dilute 10.0 mL of the solution to 200.0 mL with water R. To 10.0 mL of this solution add 4 mL of lanthanum chloride solution R and dilute to 100.0 mL with water R.

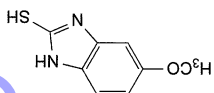
Reference solutions Prepare the reference solutions using a mixture of 1 mL of a 103 g/L solution of hydrochloric acid R and 1000.0 mL of water R.

Preparation of reference solutions Prepare the reference solutions using a mixture of 1 mL of a 103 g/L solution of hydrochloric acid R and 1000.0 mL of water R.

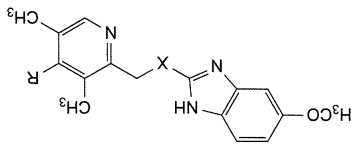
IMPURITIES

Specified impurities D, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C.



A. 5-methoxy-1H-benzimidazole-2-thiol.



B. R = H, X = SO; 2-[(3,5-dimethylpyridin-2-yl)methyl]-5-methoxy-1H-benzimidazole, (yl)methyl]sulfanyl]-5-methoxy-1H-benzimidazole, C. R = OCH₃, X = S; 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfanyl]-1H-benzimidazole, D. R = OCH₃, X = SO₂; 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfanyl]-1H-benzimidazole,

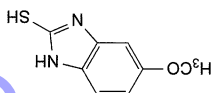
STORAGE

In an airtight container, protected from light.

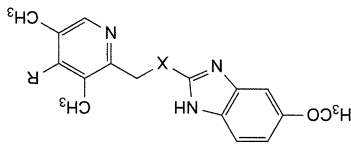
IMPURITIES

Specified impurities D, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C.



A. 5-methoxy-1H-benzimidazole-2-thiol.



B. R = H, X = SO; 2-[(3,5-dimethylpyridin-2-yl)methyl]-5-methoxy-1H-benzimidazole, (yl)methyl]sulfanyl]-5-methoxy-1H-benzimidazole, C. R = OCH₃, X = S; 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfanyl]-1H-benzimidazole, D. R = OCH₃, X = SO₂; 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfanyl]-1H-benzimidazole,

ASSAY

Liquid chromatography (2.2.29).

Buffer pH 11.0 Mix 11 mL of a 95.0 g/L solution of trisodium phosphate dodecahydrate R and 22 mL of a 179.1 g/L solution of disodium hydrogen phosphate R. Dilute to 100.0 mL with water R.

Test solution Dissolve 10.0 mg of the substance to be examined in about 10 mL of methanol R. Add 10 mL of buffer pH 11.0 and dilute to 200.0 mL with water R.

Reference solution Dissolve 10.0 mg of omeprazole CRS in about 10 mL of methanol R. Add 10 mL of buffer pH 11.0 and dilute to 200.0 mL with water R.

Column:

— size: $l = 0.125$ m, $\varnothing = 4$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 35 volumes of acetonitrile R and 65 volumes of a 1.4 g/L solution of disodium hydrogen phosphate R previously adjusted to pH 7.6 with phosphoric acid R. Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of omeprazole.

Retention time Omeprazole = about 4 min.

Calculate the percentage content of C₁₄H₁₆N₂O₆S₂ from the declared content of omeprazole CRS.

1 g of omeprazole is equivalent to 1.032 g of omeprazole magnesium.

TESTS

Solution S
Dissolve 0.50 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

pH (2.2.3)

10.3 to 11.3 for solution S.

Related substances

Liquid chromatography (2.2.29). Prepare solutions immediately before use.

Test solution Dissolve 3 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 1 mg of omeprazole CRS and 1 mg of omeprazole impurity D CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 3 mg of omeprazole for peak identification CRS (containing impurity E) in the mobile phase and dilute to 25.0 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 27 volumes of acetonitrile R and 73 volumes of a 1.4 g/L solution of disodium hydrogen phosphate R, previously adjusted to pH 7.6 with phosphoric acid R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 40 μ L.

Run time 5 times the retention time of omeprazole.

Identification of impurities Use the chromatogram supplied

chromatogram obtained with reference solution (c) to identify the peak due to impurity E; use the chromatogram obtained

with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to omeprazole (retention time = about 9 min): impurity E = about 0.6;

impurity D = about 0.8.

System suitability: reference solution (a):

— **resolution:** minimum 3.0 between the peaks due to impurity D and omeprazole; if necessary adjust the pH of the aqueous part of the mobile phase or the concentration of acetonitrile R; an increase in the pH will improve the

resolution.

Limits:

— **impurities D, E:** for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

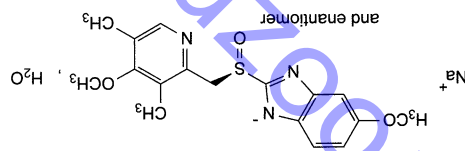
(0.15 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **total:** not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

Omeprazole Sodium

(Ph. Eur. monograph 1032)



$C_{17}H_{18}N_3NaO_3S \cdot H_2O$ 385.4 95510-70-6

Action and use
Proton pump inhibitor; treatment of peptic ulcer disease.

DEFINITION

Sodium 5-methoxy-2-[(R,S)-[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfinyl]-1H-benzimidazole monohydrate.

Content

98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), soluble in propylene glycol, very slightly soluble in methylene chloride.

IDENTIFICATION

A. Optical rotation (2.2.7): -0.10° to $+0.10^\circ$, determined on solution S.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 0.50 g of the substance to be examined in 1.50 mL of water R, add 3.0 mL of methanol R and stir

while stirring, adjust to pH 8-9 by adding, dropwise, dilute acetic acid R (about 0.4 mL); continue stirring until

crystallisation and isolate the crystalline precipitate by filtration; wash with 5 mL of water R, then 2 mL of

methanol R, and dry in vacuo at 40°C for 30 min.

Comparison omeprazole CRS.

If the spectra obtained in the solid state show differences, dissolve the crystalline precipitate and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

C. Ignite 1 g and cool. Add 1 mL of water R to the residue and neutralise with hydrochloric acid R. Filter and dilute the filtrate to 4 mL with water R. 0.1 mL of the solution gives

reaction (b) of sodium (2.3.1).

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

4.5 per cent to 10.0 per cent, determined on 0.300 g.

ASSAY

Dissolve 0.300 g in 50 mL of water R. Titrate with 0.1 M hydrochloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M hydrochloric acid corresponds to 36.74 mg of $C_{17}H_{18}N_3NaO_3S$.

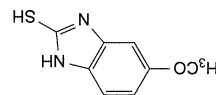
STORAGE

In an airtight container, protected from light.

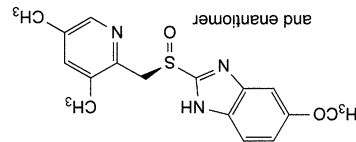
IMPURITIES

Specified impurities D, E

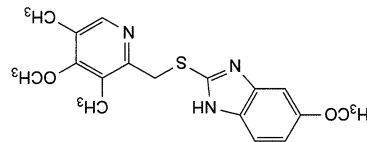
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C.



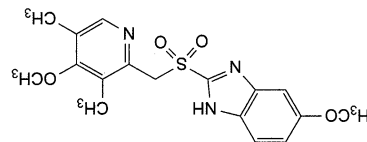
A. 5-methoxy-1H-benzimidazole-2-thiol,



B. 2-[(R,S)-(3,5-dimethylpyridin-2-yl)methyl]sulfany]-5-methoxy-1H-benzimidazole,



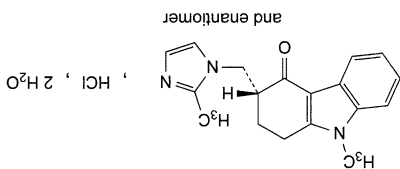
C. 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfany]-1H-benzimidazole (ufprazole),



D. 5-methoxy-2-[[[(4-methoxy-3,5-dimethylpyridin-2-yl)methyl]sulfany]-1H-benzimidazole (omeprazole-sulfone),

Ondansetron Hydrochloride Dihydrate

(Ph Eur monograph 2016)



$C_{18}H_{20}ClN_3O_2 \cdot 2H_2O$ 365.9 103639-04-9

Action and use

Serotonin 5HT₃ antagonist; treatment of nausea and vomiting.

Preparations

Ondansetron Injection

Ondansetron Tablets

Ph Eur

DEFINITION

(3R,S)-9-Methyl-3-[(2-methyl-1H-imidazol-1-yl)methyl]-1,2,3,9-tetrahydro-4H-carbazol-4-one hydrochloride dihydrate.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Sparingly soluble in water, soluble in methanol, sparingly soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Comparison ondansetron hydrochloride dihydrate CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Impurity B

Thin-layer chromatography (2.2.27).

Solvent mixture concentrated ammonia R, ethanol (96 per cent) R, methanol R (0.5:100:100 V/V/V).

Test solution Dissolve 0.125 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with

the solvent mixture.

Reference solution (a) Dissolve 12.5 mg of ondansetron for TLC system suitability CRS (containing impurities A and B) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Reference solution (b) Dilute 1 mL of the test solution to 100 mL with the solvent mixture. Dilute 4.0 mL of this solution to 10.0 mL with the solvent mixture.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, methanol R, ethyl acetate R, methylene chloride R (2:40:50:90 V/V/V/V).

Application 20 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Retardation factors Impurity A = about 0.3; impurity B = about 0.4; ondansetron = about 0.6.

System suitability The chromatogram obtained with reference solution (a) shows 3 clearly separated spots.

Limit:

— **impurity B:** any spot corresponding to impurity B in the chromatogram obtained with the test solution is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.4 per cent).

Related substances

Liquid chromatography (2.2.29):

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Test solution (b) Dissolve 90.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of ondansetron impurity A CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of ondansetron for LC system suitability CRS (containing impurities C and D) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 5.0 mg of ondansetron impurity D CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (e) Dissolve 90.0 mg of ondansetron hydrochloride CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (f) Dissolve 5.0 mg of ondansetron impurity F CRS and 5 mg of ondansetron impurity G CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (g) To 1.0 mL of reference solution (b) add 1.0 mL of reference solution (f) and dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: spherical nitrite silica gel for chromatography R (5 µm) with a specific surface area of 220 m²/g and a pore size of 8 nm.

Mobile phase Mix 20 volumes of acetonitrile R1 and 80 volumes of a 2.8 g/L solution of sodium dihydrogen phosphate monohydrate R previously adjusted to pH 5.4 with a 40 g/L solution of sodium hydroxide R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 216 nm.

Injection 20 µL of test solution (a) and reference solutions (a), (b), (c), (d), (f) and (g).

Run time 1.5 times the retention time of ondansetron.

Identification of impurities:

— use the chromatogram supplied with ondansetron for LC system suitability CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C and D;

— use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and E;

— use the chromatogram obtained with reference solution (f) to identify the peaks due to impurities F and G.

Relative retention With reference to ondansetron (retention time = about 18 min): impurity B = about 0.17; impurity F = about 0.20 (E and F may coelute); impurity C = about 0.35; impurity D = about 0.45; impurity A = about 0.80; impurity G = about 0.89 (A and G may coelute or be inverted).

System suitability: reference solution (c):

— **resolution:** minimum 2.5 between the peaks due to impurities C and D.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity C by 0.6;

— **impurity C:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **impurity D:** not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (d) (0.15 per cent);

— **sum of impurities A and G:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **sum of impurities B and F:** not more than the sum of the areas of the corresponding peaks in the chromatogram obtained with reference solution (g) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **total:** maximum 0.4 per cent;

— **disregard limit:** 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (2.5.12)

9.0 per cent to 10.5 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (e).

Calculate the percentage content of C₁₈H₂₀ClN₃O from the declared content of ondansetron hydrochloride dihydrate CRS.

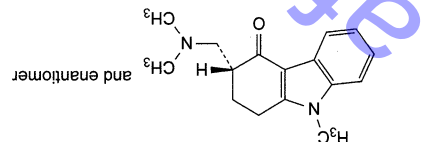
STORAGE

Protected from light.

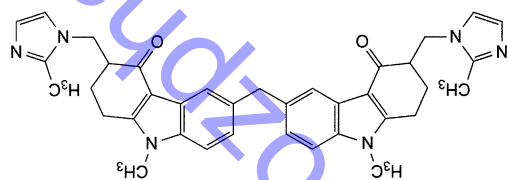
IMPURITIES

Specified impurities A, B, C, D, E, F, G

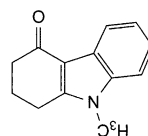
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): H.



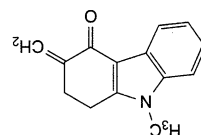
A. (3R,5S)-3-[(2-methyl-1H-imidazol-1-yl)methyl]-9-methyl-1,2,3,9-tetrahydro-4H-carbazol-4-one,



B. 6,6'-methylenbis[(3R,5S)-9-methyl-3-[(2-methyl-1H-imidazol-1-yl)methyl]-1,2,3,9-tetrahydro-4H-carbazol-4-one],



C. 9-methyl-1,2,3,9-tetrahydro-4H-carbazol-4-one,



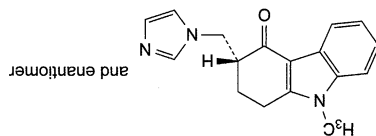
D. 9-methyl-3-methylene-1,2,3,9-tetrahydro-4H-carbazol-4-one,



E. 1H-imidazole,



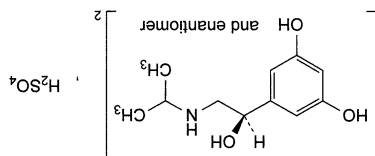
F. 2-methyl-1H-imidazole,



G. (3R,5S)-3-[(1H-imidazol-1-yl)methyl]-9-methyl-1,2,3,9-tetrahydro-4H-carbazol-4-one (C-desmethyldansetron),

Orciprenaline Sulfate

(Ph. Eur. monograph 1033)



$C_{22}H_{36}N_2O_{10}S$ 520.6 5874-97-5

Action and use

Beta₂-adrenoceptor agonist; bronchodilator.

Ph. Eur.

DEFINITION

Bis[5-[(1R,5S)-1-hydroxy-2-[(1-methylethylamino)ethyl]benzene-1,3-diol] sulfate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification B, E.

Second identification A, C, D, E.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 50.0 mg in a 0.04 per cent V/V solution of hydrochloric acid R and dilute to 50.0 mL with the same solution. Dilute 5.0 mL of this solution to 50.0 mL with a 0.04 per cent V/V solution of hydrochloric acid R.

Spectral range 240-350 nm.

Absorption maximum At 278 nm.

Specific absorbance at the absorption maximum 68.5 to 76.0 (anhydrous substance).

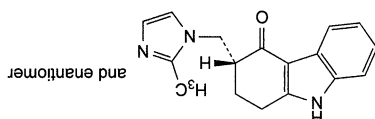
B. Infrared absorption spectrophotometry (2.2.24).

Comparison orciprenaline sulfate CRS.

If the spectra obtained show differences, dissolve separately, 50 mg of the reference substance, in the minimum volume of water R. Add 10 mL of acetone R and centrifuge. Dry the precipitates at 40 °C under reduced pressure for 3 h and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

H. (3R,5S)-3-[(2-methyl-1H-imidazol-1-yl)methyl]-1,2,3,9-tetrahydro-4H-carbazol-4-one (N-desmethyldansetron).



Test solution Dissolve 10 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of orciprenaline sulfate CRS in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of orciprenaline sulfate CRS and 10 mg of salbutamol CRS in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase ammonia R, water R, aldehyde-free methanol R (1.5:10:90 V/V/V).

Application 2 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 10 g/L solution of potassium permanganate R.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 20 mg in 2 mL of ethanol (96 per cent) R. Add 2 mL of a 1 g/L solution of dichloroquinonechlorimide R in ethanol (96 per cent) R and 1 mL of sodium carbonate solution R. A violet colour is produced, turning to brown. It gives reaction (a) of sulfates (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.0 to 5.5 for solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in the mobile phase and dilute to 20 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of orciprenaline for system suitability CRS (containing impurities A and B) in 2.0 mL of the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.0$ mm;

— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 µm);

— temperature: 45 °C.

Mobile phase Dissolve 9.1 g of potassium dihydrogen phosphate R and 4.6 g of sodium octanesulfonate R in water R, adjust to pH 4.0 with dilute phosphoric acid R and dilute to 1000 mL with water R. Add 140 mL of acetonitrile R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 µL.

Run time Twice the retention time of orciprenaline.

Identification of impurities Use the chromatogram supplied with orciprenaline for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to orciprenaline (retention time = about 7 min): impurity A = about 0.9;

impurity B = about 1.3.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurity A and orciprenaline.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity B by 0.3;

— **impurities A, B:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Phenone

Maximum 0.1 per cent.

Dissolve 0.50 g in a 0.04 per cent V/V solution of hydrochloric acid R and dilute to 25.0 mL with the same solution.

The absorbance (2.2.25) of the solution measured at 328 nm is not greater than 0.16.

Iron (2.4.9)

Maximum 20 ppm.

The residue obtained in the test for sulfated ash complies with the test. Prepare the reference solution using iron standard solution (2 ppm Fe) R.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 2.0 per cent, determined on 1.000 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 5 mL of anhydrous formic acid R and add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid using 0.1 mL of crystal violet solution R as indicator.

1 mL of 0.1 M perchloric acid is equivalent to 52.06 mg of $C_{22}H_{36}N_2O_{10}S$.

STORAGE

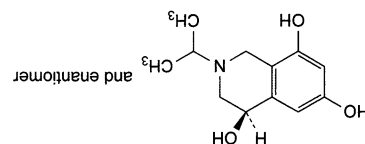
In an airtight container, protected from light.

IMPURITIES

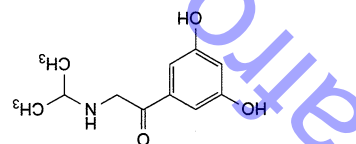
Specified impurities A, B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

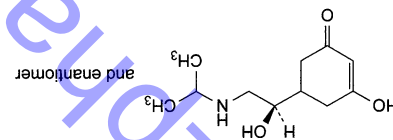
impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



A. (4R)-2-(1-methylethyl)-1,2,3,4-tetrahydroisquinoline-4,6,8-triol,



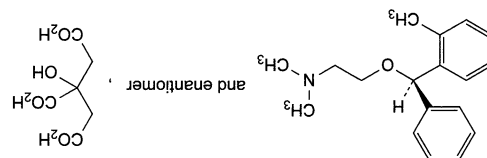
B. 1-(3,5-dihydroxyphenyl)-2-



C. 3-hydroxy-5-[(1R)-1-hydroxy-2-[(1-methylethyl)amino]ethyl]cyclohex-2-enone.

Orphenadrine Citrate

(Ph. Eur. monograph 1759)



C₂₄H₃₁NO₈ 461.5 4682-36-4

Action and use
Anticholinergic.

Ph. Eur.

DEFINITION

(RS)-N,N-Dimethyl-2-[(2-methylphenyl)phenylmethoxy]ethanamine dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparsely soluble in water, slightly soluble in ethanol

(96 per cent).

mp

About 137 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison orphenadrine citrate CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and its absorbance (2.2.25) at 436 nm has a maximum of 0.050.

Dissolve 1.0 g in 3.6 per cent V/V solution of hydrochloric acid R in ethanol (96 per cent) R and dilute to 10.0 mL with the same acid solution.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure. Identification of impurities: use the chromatogram supplied with orphenadrine for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and F. Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity E.

Test solution Dissolve 0.500 g of the substance to be examined in water R and dilute to 50 mL with the same solvent. Add 2 mL of concentrated ammonia R and shake with 3 quantities, each of 10 mL, of toluene R. To the combined upper layers add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate, at a temperature not exceeding 50 °C, using a rotary evaporator. Take up the residue with toluene R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 30 mg of orphenadrine citrate CRS and 30 mg of orphenadrine impurity E CRS in 20 mL of water R. Add 1 mL of concentrated ammonia R and shake with 3 quantities, each of 5 mL, of toluene R. To the combined upper layers add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate, at a temperature not exceeding 50 °C, using a rotary evaporator. Take up the residue with toluene R and dilute to 20.0 mL with the same solvent. Reference solution (b) Dissolve the contents of a vial of orphenadrine for peak identification CRS (containing impurities A, B, C, D and F) in 1.0 mL of toluene R.

Column: — size: l = 60 m, Ø = 0.32 mm; — stationary phase: poly (dimethyl) (diphenyl) siloxane R (film thickness 1.0 µm).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 1:25.

Temperature: — column: 240 °C;

— injection port and detector: 290 °C.

Detection Flame ionisation.

Injection 2 µL.

Run time 1.3 times the retention time of orphenadrine.

Identification of impurities: use the chromatogram supplied with orphenadrine for peak identification CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B, C, D and F.

Use the chromatogram obtained with reference solution (a)

to identify the peak due to impurity E.

Relative retention With reference to orphenadrine (retention

time = about 13 min): impurity B = about 0.5;

impurity A = about 0.6; impurity D = about 0.8;

impurity C = about 0.9; impurity E = about 0.98;

impurity F = about 1.1.

System suitability: reference solution (a):

— **resolution:** minimum of 1.5 between the peaks due to impurity E and orphenadrine.

Limits: impurities A, B, C, D, E, F: for each impurity, not more than 0.3 per cent;

— **unspecified impurities:** for each impurity, not more than 0.10 per cent;

— **total:** maximum 1.0 per cent;

— **disregard limit:** 0.05 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

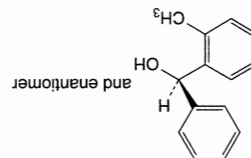
1 mL of 0.1 M perchloric acid is equivalent to 46.15 mg of $C_{24}H_{31}NO_8$.

STORAGE

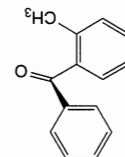
Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container, protected from light.

IMPURITIES

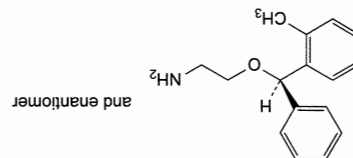
Specified impurities A, B, C, D, E, F



A. (R,S)-(2-methylphenyl)phenylmethanol (2-methylbenzhydrol),



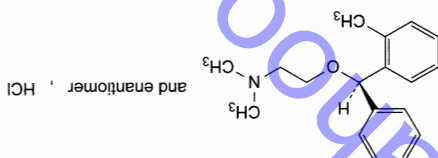
B. (2-methylphenyl)phenylmethanone (2-methylbenzophenone),



C. (R,S)-2-[(2-methylphenyl)phenylmethoxy]ethanamine,

Orphenadrine Hydrochloride

(Ph. Eur. monograph 1760)



$C_{18}H_{24}ClNO$

305.9

341-69-5

Action and use

Anticholinergic.

Preparation

Orphenadrine Hydrochloride Tablets

Ph. Eur.

DEFINITION

(R,S)-N,N-Dimethyl-2-[(2-methylphenyl)phenylmethoxy]ethanamine hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent).

mp

About 160 °C.



IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison orphenadrine hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and its absorbance (2.2.25) at 436 nm has a maximum of 0.050.

Dissolve 0.70 g in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure. Identification of impurities: use the chromatogram supplied with orphenadrine for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and F. Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity E.

Test solution Dissolve 0.300 g of the substance to be examined in water R and dilute to 50 mL with the same solvent. Add 2 mL of concentrated ammonia R and shake with 3 quantities, each of 10 mL, of toluene R. To the combined upper layers add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate, at a temperature not exceeding 50 °C, using a rotary evaporator. Take up the residue with toluene R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of orphenadrine hydrochloride CRS and 20 mg of orphenadrine impurity E CRS in 20 mL of water R. Add 1 mL of concentrated ammonia R and shake with 3 quantities, each of 5 mL, of toluene R. To the combined upper layers add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate, at a temperature not exceeding 50 °C, using a rotary evaporator. Take up the residue with toluene R and dilute to 20.0 mL with the same solvent.

Reference solution (b) Dissolve the contents of a vial of orphenadrine for peak identification CRS (containing impurities A, B, C, D and F) in 1.0 mL of toluene R. Column: — size: $l = 60$ m, $\varnothing = 0.32$ mm; — stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 1.0 μ m). Carrier gas helium for chromatography R. Flow rate 1 mL/min. Split ratio 1:25. Temperature: — column: 240 °C; — injection port and detector: 290 °C. Detection Flame ionisation. Injection 2 μ L. Run time 1.3 times the retention time of orphenadrine.

Identification of impurities: use the chromatogram supplied with orphenadrine for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D and F. Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity E. With reference to orphenadrine (retention time = about 13 min): impurity B = about 0.5; impurity A = about 0.6; impurity D = about 0.8; impurity C = about 0.9; impurity E = about 0.98; impurity F = about 1.1.

System suitability: reference solution (a): — resolution: minimum 1.5 between the peaks due to impurity E and orphenadrine.

Limits: — impurities A, B, C, D, E, F: for each impurity, not more than 0.3 per cent;

— unspecified impurities: for each impurity, not more than 0.10 per cent;

— total: not more than 1.0 per cent;

— disregard limit: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

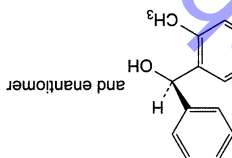
1 mL of 0.1 M perchloric acid is equivalent to 30.59 mg of $C_{18}H_{24}ClNO$.

STORAGE

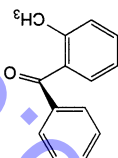
Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container, protected from light.

IMPURITIES

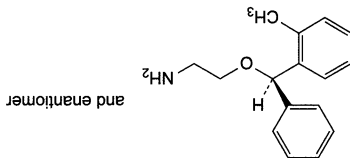
Specified impurities A, B, C, D, E, F



A. (RS)-(2-methylphenyl)phenylmethanol (2-methylbenzhydrol), and enantiomer



B. (2-methylphenyl)phenylmethanone (2-methylbenzophenone), and enantiomer



C. (RS)-2-[(2-methylphenyl)phenylmethoxy]ethanamine, and enantiomer

It shows polymorphism (5.9).

IDENTIFICATION

- A. Specific optical rotation (see Tests).
 B. Infrared absorption spectrophotometry (2.2.24).
 Comparison osetlamivir phosphate (impurity B-free) CRS.
 If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.
 C. Dissolve 200 mg in 10 mL of water R. It gives reaction (b) of phosphates (2.3.1).

TESTS

Specific optical rotation (2.2.7)
 —30.7 to –32.6 (anhydrous substance), measured at 25 °C.
 Dissolve 0.50 g in water R and dilute to 50.0 mL with the same solvent.

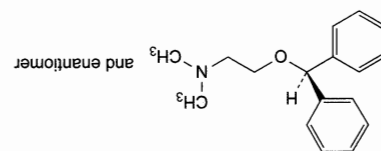
Impurity B

Liquid chromatography (2.2.29) coupled with mass spectrometry (2.2.43).
 Test solution Dissolve 0.100 g of the substance to be examined in water for chromatography R and dilute to 10.0 mL with the same solvent.
 Reference solution (a) Dissolve 2.5 mg of osetlamivir impurity B CRS in 5.0 mL of anhydrous ethanol R and dilute to 50.0 mL with water for chromatography R. Dilute 2.0 mL of the solution to 100.0 mL with water for chromatography R.
 Reference solution (b) Dissolve 50.0 mg of osetlamivir phosphate (impurity B-free) CRS in reference solution (a) and dilute to 5.0 mL with the same solution.

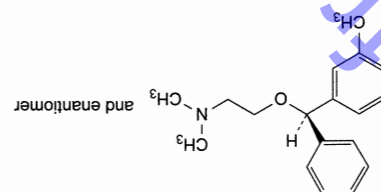
Column:
 size: $l = 0.05$ m, $\phi = 3.0$ mm;
 stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
 temperature: 40 °C.
Mobile phase Mix 10 volumes of a 1.54 g/L solution of ammonium acetate R in water for chromatography R, 30 volumes of acetonitrile R1 and 60 volumes of water for chromatography R.
Flow rate 1.5 mL/min.
Post-column split ratio Use a split ratio suitable for the mass detector (e.g. 1:3).

Detection:
 — mass detector: the following settings have been found to be suitable and are given as examples; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criterion:
 — ionisation: ESI-positive;
 — detection m/z : 356.2;
 — dwell: 580 ms;
 — gain EMV: 1;
 — fragmentator voltage: 120 V;
 — gas temperature: 350 °C;
 — drying gas flow: 13 L/min;
 — nebuliser pressure: 345 kPa;
 — capillary voltage (Vcap): 3 kV.
Injection 1 μ L of the test solution and reference solution (b).
Run time 3 min.

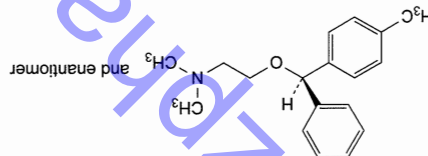
System suitability: reference solution (b):
 — repeatability: maximum relative standard deviation of 15 per cent determined on 6 injections.



D. 2-((diphenylmethoxy)-N,N-dimethylethanamine (diphenhydramine),



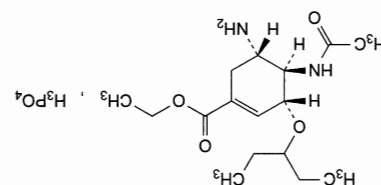
E. (R,S)-N,N-dimethyl-2-[(3-methylphenyl)phenylmethoxy]ethanamine (*meta*-methylbenzyl isomer),



F. (R,S)-N,N-dimethyl-2-[(4-methylphenyl)phenylmethoxy]ethanamine (*para*-methylbenzyl isomer).

Osetlamivir Phosphate

(Ph. Eur. monograph 2422)



$C_{16}H_{31}N_2O_8P$ 410.4 204255-11-8

Action and use
 Treatment of influenza.
Preparation
 Paediatric Osetlamivir Oral Solution

DEFINITION
 Ethyl (3R,4R,5S)-4-acetamido-5-amino-3-(1-ethylpropoxy)cyclohex-1-ene-1-carboxylate phosphate.

Content
 98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
 White or almost white powder.

Solubility
 Freely soluble in water and in methanol, practically insoluble in methylene chloride.

Limit:
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (100 ppm).

Impurity H

Gas chromatography (2.2.28).

Silylation reagent Mix 1.0 mL of chlorotrimethylsilane R, 2.0 mL of hexamethyldisilazane R and 10.0 mL of anhydrous pyridine R.

Test solution Introduce 15.0 mg of the substance to be examined into a 2 mL vial and add 1.0 mL of the silylation reagent. Close the vial, shake and heat at 60 °C for 20 min.

Reference solution Introduce 15.0 mg of oseltamivir *impurity H* CRS into a 2 mL vial and add 1.0 mL of anhydrous pyridine R. Close the vial and shake (solution A).

(Note: *impurity H* is hygroscopic.) Introduce 15.0 mg of the substance to be examined into another 2 mL vial and add 1.0 mL of the silylation reagent. Close the vial, shake and heat at 60 °C for 20 min. Centrifuge and discard the

precipitate (solution B). Introduce 10.0 µL of solution A and 10.0 µL of solution B into a volumetric flask and dilute to 10.0 mL with anhydrous pyridine R.

Column:

— *material*: fused silica;

— *size*: $l = 30$ m, $\varnothing = 0.32$ mm;

— *stationary phase*: poly(dimethyl)siloxane R (film thickness 0.25 µm).

Carrier gas helium for chromatography R.

Flow rate 1.2 mL/min.

Split ratio 1:50.

Temperature:

Time (min)	Temperature (°C)
0 - 2	180
2 - 11	180 → 250
11 - 21	250
Injection port	260
Detector	260

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to oseltamivir phosphate (retention time = about 10 min): *impurity H* = about 0.5.

System suitability: reference solution:

— **repeatability:** maximum relative standard deviation of 5 per cent for the peak due to *impurity H* after

6 injections.

Limit:

— *impurity H*: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (0.15 per cent).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R1, methanol R2, water for chromatography R (135:245:620 V/V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of oseltamivir

impurity A CRS and 5.0 mg of oseltamivir *impurity C* CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of oseltamivir phosphate

(*impurity B-free*) CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Column:

— *size*: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— *stationary phase*: end-capped octylsilyl silica gel for chromatography R (5 µm);

— *temperature*: 50 °C.

Mobile phase Mix 135 volumes of acetonitrile R1, 245 volumes of methanol R2 and 620 volumes of a 6.8 g/L solution of potassium dihydrogen phosphate R in water for chromatography R, adjusted to pH 6.0 with 1 M potassium hydroxide.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 207 nm.

Injection 15 µL of the test solution and reference solutions (a) and (b).

Run time Twice the retention time of oseltamivir phosphate.

Relative retention With reference to oseltamivir phosphate (retention time = about 17 min): *impurity A* = about 0.16;

impurity C = about 0.17.

System suitability: reference solution (b):

— **resolution:** minimum 1.5 between the peaks due to

impurities A and C.

— **impurity C**: not more than 0.3 times the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— **unspecified impurities:** for each *impurity*, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **total:** not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{16}H_{31}N_2O_8P$ from the declared content of oseltamivir phosphate (*impurity B-free*) CRS.

STORAGE

Protected from light.

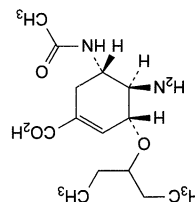
IMPURITIES

Specified impurities B, C, H

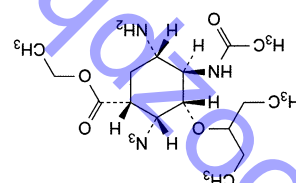
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: A, D, E, F, G.

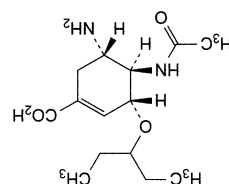
A. (3*R*,4*R*,5*S*)-5-acetamido-4-amino-3-(1-ethylpropoxy)cyclohex-1-ene-1-carboxylic acid,



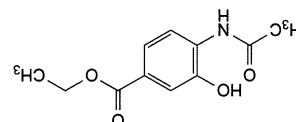
B. ethyl (1*R*,2*R*,3*S*,4*R*,5*S*)-4-acetamido-5-amino-2-azido-3-(1-ethylpropoxy)cyclohexanecarboxylate,



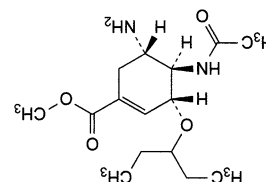
C. (3*R*,4*R*,5*S*)-4-acetamido-5-amino-3-(1-ethylpropoxy)cyclohex-1-ene-1-carboxylic acid,



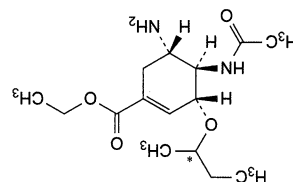
D. ethyl 4-acetamido-3-hydroxybenzoate,



E. methyl (3*R*,4*R*,5*S*)-4-acetamido-5-amino-3-(1-ethylpropoxy)cyclohex-1-ene-1-carboxylate,

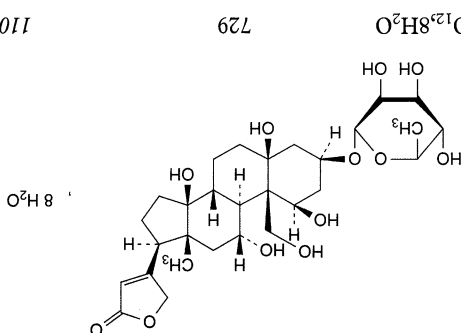


F. ethyl (3*R*,4*R*,5*S*)-4-acetamido-5-amino-3-(1-methylpropoxy)cyclohex-1-ene-1-carboxylate,



Ouabain

(Ph. Eur. monograph 0048)

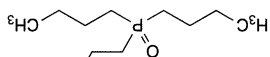


729

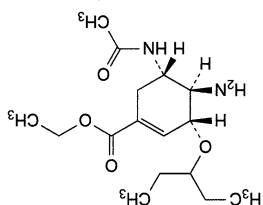
11018-89-6



H. tributylphosphane oxide.



G. ethyl (3*R*,4*R*,5*S*)-5-acetamido-4-amino-3-(1-ethylpropoxy)cyclohex-1-ene-1-carboxylate,



IDENTIFICATION

A. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the spot in the chromatogram obtained with reference solution (a).

B. Dissolve 2 mg to 3 mg in 2 mL of *sulfuric acid R*; a pink colour develops which quickly changes to red. The solution shows green fluorescence in ultraviolet light.

C. Dissolve about 1 mg in 1 mL of *dinitrobenzene solution R* and add 0.2 mL of *dilute sodium hydroxide solution R*.

D. Dissolve 0.1 g in 5 mL of a 150 g/L solution of *sulfuric acid R* and boil for a few minutes. The solution becomes yellow and turbid. Filter and add to the filtrate 5 mL of a

120 g/L solution of *sodium hydroxide R* and 3 mL of *cupri-tartaric solution R*. Heat. A red precipitate is formed.

CHARACTERS

96.0 per cent to 104.0 per cent (anhydrous substance).

Content

3β-[(6-Deoxy-α-L-mannopyranosyl)oxy]-1β,5,11α,14,19-pentahydroxy-5β,14β-card-20(22)-enolide octahydrate.

DEFINITION

Ph Eur

TESTS**Solution S**

Dissolve 0.20 g in 15 mL of water R, heating on a water-bath. Allow to cool and dilute to 20.0 mL with water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Specific optical rotation (2.2.7)

–33 to –30 (anhydrous substance), determined on solution S.

Related substances

Thin-layer chromatography (2.2.27).

Solvent mixture water R, chloroform R, methanol R

(16:50:50 V/V/V).

Test solution Dissolve a quantity of the substance to be examined corresponding to 20 mg of the anhydrous substance in 1.0 mL of the solvent mixture.

Reference solution (a) Dissolve a quantity of ouabain CRS corresponding to 20 mg of the anhydrous substance in 1.0 mL of the solvent mixture.

Reference solution (b) Dissolve a quantity of ouabain CRS 1.0 mL of the solvent mixture.

Reference solution (c) Dilute 2.5 mL of reference solution (b) to 10 mL with the solvent mixture.

Plate TLC silica gel G plate R.

Mobile phase water R, dimethyl sulfoxide R, methanol R, chloroform R (4:15:15:70 V/V/V/V); homogenise the mixture before use.

Application 5 µL.

Development Over a path of 13 cm.

Drying Immediately at 140 °C for 30 min in a ventilated oven.

Detection Allow to cool, spray with alcoholic solution of sulfuric acid R and heat at 140 °C for 15 min.

System suitability:

— the principal spot in the chromatogram obtained with the test solution and the principal spot in the chromatogram obtained with reference solution (a) migrate over a distance sufficient to give unequivocal separation of the secondary spots;

— the chromatogram obtained with reference solution (c) shows a clearly visible spot.

Limit:

— any impurity: any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (2.0 per cent).

Alkaloids and strophanthin-K

To 5.0 mL of solution S add 0.5 mL of a 100 g/L solution of *lanic acid* R. No precipitate is formed.

Water (2.5.12)

18.0 per cent to 22.0 per cent, determined on 0.100 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

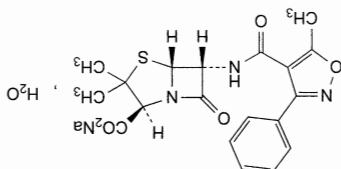
ASSAY

Test solution Dissolve 40.0 mg in ethanol (96 per cent) R and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with ethanol (96 per cent) R.

Reference solution Dissolve 40.0 mg of ouabain CRS in ethanol (96 per cent) R and dilute to 50.0 mL with the same solvent.

Oxacillin Sodium Monohydrate

(Ph Eur Monograph 2260)



$C_{19}H_{18}N_3NaO_5 \cdot H_2O$ 441.4

7240-38-2

Action and use

Penicillin antibacterial.

DEFINITION

Sodium (2S,5R,6R)-3,3-dimethyl-6-[[[(5-methyl-3-phenylisoxazol-4-yl)carbonyl]amino]-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate monohydrate.

Semi-synthetic product derived from a fermentation product.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Freely soluble in water, soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison oxacillin sodium monohydrate CRS.

B. It gives reaction (a) of sodium (2.3.1).

TESTS**Appearance of solution**

The solution is clear (2.2.1) and its absorbance (2.2.25) at 430 nm is not greater than 0.10.

Dissolve 2.50 g in water R and dilute to 25.0 mL with the same solvent.

pH (2.2.3)

4.5 to 7.5.

Dissolve 0.30 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

Dissolve 0.250 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of oxacillin sodium monohydrate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of test solution (b) to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5 mg of cloxacillin sodium CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (d) In order to prepare impurities B and D in situ, dissolve 25 mg of the substance to be examined in 1 mL of 0.05 M sodium hydroxide, allow to stand for 3 min, then dilute to 100 mL with the mobile phase. Inject immediately.

Reference solution (e) Dissolve 5 mg of oxacillin for peak identification CRS (containing impurities E, F, G, I and J) in 5 mL of the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 25 volumes of acetonitrile R and 75 volumes of 2.7 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 5.0 with dilute sodium hydroxide solution R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 μ L of test solution (a) and reference solutions (b), (c), (d) and (e).

Run time 7 times the retention time of oxacillin.

Identification of impurities:

— in the chromatogram obtained with reference solution (d), the 2 principal peaks eluting before the main peak are due to impurities B and D respectively;

— use the chromatogram supplied with oxacillin for peak identification CRS and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities E, F, G, I and J.

Relative retention With reference to oxacillin (retention time = about 5 min): impurity A = about 0.3; impurity B (isomer 1) = about 0.4; impurity B (isomer 2) = about 0.5; impurity C = about 0.65; impurity D (2 epimers) = about 0.9; impurity E = about 1.5; impurity F = about 1.9; impurity G = about 2.1; impurity I = about 3.8; impurity J = about 5.8.

System suitability:

— resolution: minimum 2.5 between the peaks due to oxacillin and impurity B in the chromatogram obtained with reference solution (c);

— the chromatogram obtained with reference solution (e) is similar to the chromatogram supplied with oxacillin for peak identification CRS.

Limits:

— impurity B: for the sum of the areas of the 2 isomer peaks, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);

— impurity E: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— impurities D (sum of the 2 epimers), F, G, I, J: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— any other impurity: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Ethyl acetate and butyl acetate

Head-space gas chromatography (2.2.28).

Test solution Dissolve 0.200 g of the substance to be examined in 6.0 mL of water R.

Reference solution Dissolve 83 mg of butyl acetate R and 83 mg of ethyl acetate R in water R and dilute to 250.0 mL with the same solvent. Use 6.0 mL of this solution.

Close the vials immediately with a rubber membrane stopper coated with polytetrafluoroethylene and secured with an aluminium crimped cap. Mix to obtain a homogeneous solution.

Column:

— material: fused silica;

— size: $l = 50$ m, $\varnothing = 0.32$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 5 μ m).

Carrier gas helium for chromatography R.

Flow rate 2 mL/min.

Static head-space conditions that may be used:

— equilibration temperature: 80 °C;

— equilibration time: 60 min;

— transfer-line temperature: 140 °C;

— pressurisation time: 30 s.

Temperature:

Column

Time

Temperature

Injection port

Detector

Limit:

— butyl acetate: maximum 1.0 per cent;

— ethyl acetate: maximum 1.0 per cent.

N,N-Dimethylamine (2.4.26, Method B)

Maximum 20 ppm.

2-Ethylhexanoic acid (2.4.28)

Maximum 0.8 per cent

Water (2.5.12)

3.5 per cent to 5.0 per cent, determined on 0.300 g.

Bacterial endotoxins (2.6.14)

Less than 0.20 IU/mg, if intended for use in the manufacture

of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{19}H_{18}N_3NaO_5$ taking into account the assigned content of oxacillin sodium monohydrate CRS.

IMPURITIES

Specified impurities B, D, E, F, G, I, J

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

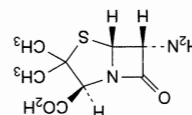
acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

(2034). It is therefore not necessary to identify these

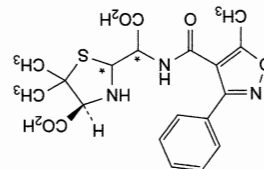
impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, C.

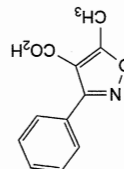


A. (2S,5R,6R)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid

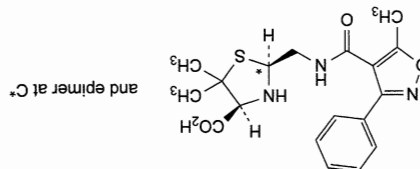
(6-aminopenicillanic acid),



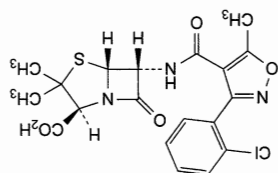
B. (4S)-2-[carboxy[[[(5-methyl-3-phenylisoxazol-4-yl) carbonyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of oxacillin),



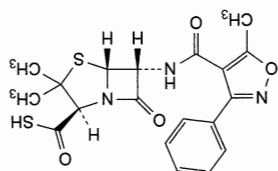
C. 5-methyl-3-phenylisoxazole-4-carboxylic acid,



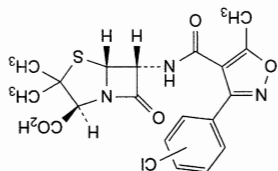
D. (2R,4S)-5,5-dimethyl-2-[[[(5-methyl-3-phenylisoxazol-4-yl) carbonyl]amino]methyl]thiazolidine-4-carboxylic acid (penicilloic acids of oxacillin),



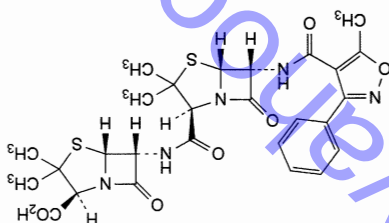
E. (2S,5R,6R)-6-[[[(3-(2-chlorophenyl)-5-methylisoxazol-4-yl) carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (cloxacillin),



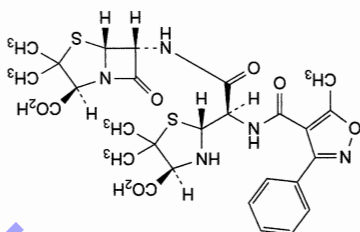
F. (2R,5R,6R)-3,3-dimethyl-6-[[[(5-methyl-3-phenylisoxazol-4-yl) carbonyl]amino]-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (thiooxacillin),



G. (2S,5R,6R)-6-[[[(3-(chlorophenyl)-5-methylisoxazol-4-yl) carbonyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (cloxacillin isomer),



I. (2S,5R,6R)-6-[[[(2S,5R,6R)-3,3-dimethyl-6-[[[(5-methyl-3-phenylisoxazol-4-yl) carbonyl]amino]-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-APA oxacillin amide),



J. (2S,5R,6R)-6-[[[(2R,4S)-4-carboxy-5,5-dimethylthiazolidine-2-yl] [[[(5-methyl-3-phenylisoxazol-4-yl) carbonyl]amino]acetyl]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (oxazolamide of 6-APA dimer).

System suitability: resolution: minimum 9 between the peaks due to nitrate and impurity A in the chromatogram obtained with reference solution (c);
— **signal-to-noise ratio:** minimum 10 for the peak due to impurity A in the chromatogram obtained with reference solution (b).
Limit:
— **impurity A:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b).
— **impurity B:** not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent).
C. Impurity C and other related substances. Liquid chromatography (2.2.29). Use vigorous shaking and very brief sonication to dissolve the substance to be examined. Inject the test solution within 20 min of preparation. Use suitable polypropylene containers for the preparation and injection of all solutions. Glass pipettes may be used for diluting solutions.
Test solution Dissolve 0.100 g of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.
Reference solution (a) Add 5.0 mg of oxaliplatin impurity B CRS to 25 mL of methanol R and dilute to 100.0 mL with water R. Sonicate for about 1.5 h until dissolved (solution A). Dilute 3.0 mL of solution A to 200.0 mL with water R.
Reference solution (b) In order to prepare impurity E *in situ*, adjust 50.0 mL of solution A to pH 6.0 with a 0.2 g/L solution of sodium hydroxide R, heat at 70 °C for 4 h and allow to cool.
Column:
— **size:** $l = 0.25$ m, $\phi = 4.6$ mm;
— **stationary phase:** octadecylsilyl silica gel for chromatography R (5 μ m);
— **temperature:** 40 °C.
Mobile phase Mix 1 volume of acetonitrile R and 99 volumes of a solution prepared as follows: dilute 0.6 mL of dilute phosphoric acid R in 1000 mL of water R and adjust to pH 3.0 with either sodium hydroxide solution R or phosphoric acid R.
Flow rate 1.2 mL/min.
Detection Spectrophotometer at 210 nm.
Injection 10 μ L of test solution (a) and reference solutions (b), (c) and (f).
Run time 3 times the retention time of oxaliplatin.
Retention time Impurity C = about 4.4 min; dichlorodiaminocyclohexaneplatinum = about 6.9 min; oxaliplatin = about 8.0 min.
System suitability:
— **resolution:** minimum 2.0 between the peaks due to dichlorodiaminocyclohexaneplatinum and oxaliplatin in the chromatogram obtained with reference solution (e);
— **signal-to-noise ratio:** minimum 50 for the peak due to impurity C and minimum 10 for the peak due to oxaliplatin in the chromatogram obtained with reference solution (a).
Limit:
— **impurity C:** not more than 0.5 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (f) (0.15 per cent);
— **unspecified impurities:** for each impurity, not more than twice the area of the peak due to oxaliplatin in the chromatogram obtained with reference solution (b) area of the peak due to oxaliplatin in the chromatogram obtained with reference solution (b) (0.15 per cent);
— **disregard limit:** the area of the peak due to oxaliplatin in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak with a retention time less than 2 min.
D. Sum of impurities other than D: maximum 0.30 per cent.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in water R and dilute to 500.0 mL with the same solvent.
Reference solution (a) Dissolve 5.0 mg of oxaliplatin CRS and 5.0 mg of oxaliplatin impurity C CRS in water R and dilute to 50.0 mL with the same solvent.
Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with water R.
Reference solution (c) Dissolve 25.0 mg of oxaliplatin CRS in water R and dilute to 250.0 mL with the same solvent.
Reference solution (d) Dissolve 5.0 mg of dichlorodiaminocyclohexaneplatinum CRS in reference solution (c) and dilute to 50.0 mL with reference solution (c).
Reference solution (e) Dilute 5 mL of reference solution (d) to 50.0 mL with water R.
Reference solution (f) To 0.100 g of the substance to be examined add 1.5 mL of reference solution (a) and dilute to 50.0 mL with water R.
Limit:
— **impurity A:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b).
— **impurity B:** not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent).
C. Impurity C and other related substances. Liquid chromatography (2.2.29). Use vigorous shaking and very brief sonication to dissolve the substance to be examined. Inject the test solution within 20 min of preparation. Use suitable polypropylene containers for the preparation and injection of all solutions. Glass pipettes may be used for diluting solutions.
Test solution Dissolve 0.100 g of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.
Reference solution (a) Add 5.0 mg of oxaliplatin impurity B CRS to 25 mL of methanol R and dilute to 100.0 mL with water R. Sonicate for about 1.5 h until dissolved (solution A). Dilute 3.0 mL of solution A to 200.0 mL with water R.
Reference solution (b) In order to prepare impurity E *in situ*, adjust 50.0 mL of solution A to pH 6.0 with a 0.2 g/L solution of sodium hydroxide R, heat at 70 °C for 4 h and allow to cool.
Column:
— **size:** $l = 0.25$ m, $\phi = 4.6$ mm;
— **stationary phase:** octadecylsilyl silica gel for chromatography R (5 μ m);
— **temperature:** 40 °C.
Mobile phase Mix 1 volume of acetonitrile R and 99 volumes of a solution prepared as follows: dilute 0.6 mL of dilute phosphoric acid R in 1000 mL of water R and adjust to pH 3.0 with either sodium hydroxide solution R or phosphoric acid R.
Flow rate 1.2 mL/min.
Detection Spectrophotometer at 210 nm.
Injection 10 μ L of test solution (a) and reference solutions (b), (c) and (f).
Run time 3 times the retention time of oxaliplatin.
Retention time Impurity C = about 4.4 min; dichlorodiaminocyclohexaneplatinum = about 6.9 min; oxaliplatin = about 8.0 min.
System suitability:
— **resolution:** minimum 2.0 between the peaks due to dichlorodiaminocyclohexaneplatinum and oxaliplatin in the chromatogram obtained with reference solution (e);
— **signal-to-noise ratio:** minimum 50 for the peak due to impurity C and minimum 10 for the peak due to oxaliplatin in the chromatogram obtained with reference solution (a).
Limit:
— **impurity C:** not more than 0.5 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (f) (0.15 per cent);
— **unspecified impurities:** for each impurity, not more than twice the area of the peak due to oxaliplatin in the chromatogram obtained with reference solution (b) area of the peak due to oxaliplatin in the chromatogram obtained with reference solution (b) (0.15 per cent);
— **disregard limit:** the area of the peak due to oxaliplatin in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak with a retention time less than 2 min.
D. Sum of impurities other than D: maximum 0.30 per cent.

Silver

Maximum 5 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 0.1000 g of the substance to be

examined in water R and dilute to 50.0 mL with the same

solvent. Dilute 20 µL of the solution to 40 µL with 0.5 M

nitric acid.

Reference solution (a) Dilute a solution of silver nitrate R

containing 1000 ppm of silver in 0.5 M nitric acid with 0.5 M

nitric acid to obtain a solution that contains 10 ppb of silver.

Reference solution (b) Mix 20 µL of the test solution and 8 µL

of reference solution (a) and dilute to 40 µL with 0.5 M nitric

acid.

Reference solution (c) Mix 20 µL of the test solution and

16 µL of reference solution (a) and dilute to 40 µL with

0.5 M nitric acid.

Source Silver hollow-cathode lamp.

Wavelength 328.1 nm.

Atomisation device Furnace.

Measure the absorbance of the test solution and reference

solutions (b) and (c).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 2 h.

Bacterial endotoxins (2.6.14)

Less than 1.0 IU/mg, if intended for use in the manufacture

of parenteral preparations without a further appropriate

procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

impurity C and other related substances with the following

modifications.

and (d).

System suitability:

— resolution: minimum 2.0 between the peaks due to

dichlorodiamminocyclohexaneplatinum and oxaliplatin in

the chromatogram obtained with reference solution (d);

— repeatability: reference solution (c).

Calculate the percentage content of oxaliplatin using the

chromatogram obtained with reference solution (c).

IMPURITIES

Specified impurities A, B, C, D

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by the general

tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: E.



A. ethanedioic acid (oxalic acid),

DEFINITION

(3R,5)-7-Chloro-3-hydroxy-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

Oxazepam Tablets

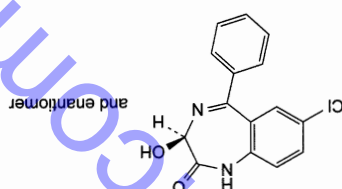
Preparation

Benzodiazepine.

Action and useC₁₅H₁₁ClN₂O₂

286.7

604-75-1



(Ph. Eur. monograph 0778)

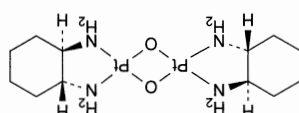
Oxazepam

Ph. Eur.

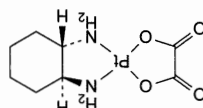
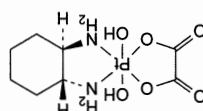
(diaquodiamminocyclohexaneplatinum dimer).

diamminekN₂kN₂[diplatinum

B. (SP-4-2)-di-μ-oxobis[(1R,2R)-cyclohexane-1,2-

D. (SP-4-2)-[(1S,2S)-cyclohexane-1,2-diamine-kN₂kN₂][ethanedioato(2-)-kO₂]₂platinum

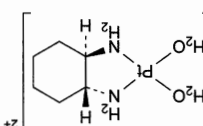
(S,S'-enantiomer of oxaliplatin),

C. (OC-6-33)-[(1R,2R)-cyclohexane-1,2-diamminekN₂kN₂][ethanedioato(2-)-kO₂]₂dihydroxyplatinum,

(diaquodiamminocyclohexaneplatinum),

diamminekN₂kN₂[platinum

B. (SP-4-2)-diqua[(1R,2R)-cyclohexane-1,2-



CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison oxazepam CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 40.0 mg of the substance to be examined in 25 mL of a mixture of equal volumes of

acetonitrile R and water R and dilute to 50.0 mL with the same mixture of solvents.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of equal volumes of acetonitrile R

and water R. Dilute 2.0 mL of this solution to 10.0 mL with a mixture of equal volumes of acetonitrile R and water R.

Reference solution (b) Dissolve the contents of a vial of oxazepam for peak identification CRS (containing

impurities A, B, C, D and E) in 1.0 mL of the test solution.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m) resistant to bases up to pH 11.

Mobile phase:

— mobile phase A: dissolve 3.48 g of dipotassium hydrogen phosphate R in 900 mL of water R, adjust to pH 10.5 with

a 40 g/L solution of sodium hydroxide R and dilute to 1000 mL with water R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 4	75	25
4 - 34	75 \rightarrow 25	25 \rightarrow 75
34 - 45	25	75
45 - 50	25 \rightarrow 75	75 \rightarrow 25
50 - 60	75	25

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 10 μ L.

Identification of impurities Use the chromatogram obtained with reference solution (b) and the chromatogram supplied

due to impurities A, B, C, D and E.

Relative retention With reference to oxazepam (retention time = about 15 min): impurity E = about 0.7;

impurity A = about 0.8; impurity B = about 1.2; impurity C = about 1.4; impurity D = about 2.0.

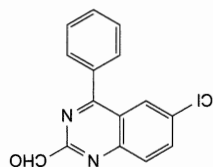
System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to impurities B and A.

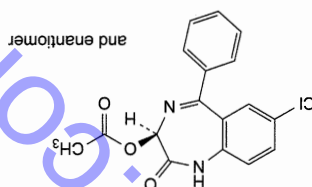
Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the

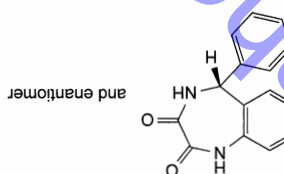
C. 6-chloro-4-phenylquinazoline-2-carbaldehyde,



B. (3*RS*)-7-chloro-2-oxo-5-phenyl-2,3-dihydro-1*H*-1,4-benzodiazepin-3-yl acetate,



A. (5*RS*)-7-chloro-5-phenyl-4,5-dihydro-1*H*-1,4-benzodiazepine-2,3-dione,



Specified impurities A, B, C, D, E

IMPURITIES

Protected from light.

STORAGE

of $C_{15}H_{11}ClN_2O_2$.

1 mL of 0.1 M perchloric acid is equivalent to 28.67 mg

(2.2.20). Dissolve 0.250 g in a mixture of 10 mL of anhydrous acetic acid R and 90 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C at a pressure not exceeding 0.7 kPa.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.25 times the area of the principal peak in

(1.0 per cent);

in the chromatogram obtained with reference solution (a)

— total: not more than 5 times the area of the principal peak

(0.10 per cent);

chromatogram obtained with reference solution (a)

0.5 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than

obtained with reference solution (a) (0.2 per cent);

the area of the principal peak in the chromatogram

— impurities A, B, C, D, E: for each impurity, not more than

impurity B = 1.1;

corresponding correction factor: impurity A = 4.0;

Solution B 1.8 g/L solution of sodium edetate R in a mixture of equal volumes of the phosphate buffer solution and water R.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in 25 mL of acetonitrile R, sonicate for 10 min, cool to room temperature and dilute to 50.0 mL with solution A.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of oxcarbazepine impurity mixture CRS (impurities A, B, I and K) in 0.5 mL of acetonitrile R and dilute to 1.0 mL with solution A.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of oxcarbazepine CRS in 25 mL of acetonitrile R, sonicate for 10 min, cool to room temperature and dilute to 50.0 mL with solution A. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: phenylhexylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: acetonitrile R, solution B, tetrahydrofuran R, water R (5:10:10:75 V/V/V/V);

— mobile phase B: solution B, tetrahydrofuran R, water R, acetonitrile R (10:10:20:60 V/V/V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 10	60	40
10 - 20	60 → 5	40 → 95
20 - 27	5	95

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 μ L of test solution (a) and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied

with oxcarbazepine impurity mixture CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities A, B, I and K.

Relative retention With reference to oxcarbazepine (retention

time = about 6 min): impurity I = about 0.8;

impurity A = about 1.3; impurities K and L = about 1.4;

impurity B = about 1.6.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 4.0, where H_p = height

above the baseline of the peak due to impurities K and L

and H_v = height above the baseline of the lowest point of

the curve separating this peak from the peak due to

impurity A.

Calculation of percentage contents:

— for each impurity, use the concentration of oxcarbazepine

Limits:

— impurities B, I: for each impurity, maximum 0.1 per cent;

— sum of impurities K and L: maximum 0.1 per cent;

— unspecified impurities: for each impurity, maximum

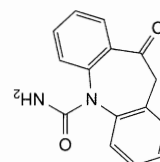
0.05 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.03 per cent.

Oxcarbazepine

(Ph Eur monograph 2577)



$C_{15}H_{12}N_2O_2$

252.3

28721-07-5

Action and use

Antiepileptic.

Ph Eur

DEFINITION

10-Oxo-10,11-dihydro-5H-dibenzo[b,f]azepine-5-

carboxamide.

CHARACTERS

Appearance

White or faintly orange, crystalline powder.

Solubility

Practically insoluble in water and in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison oxcarbazepine CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture acetonitrile R, solution A (50:50 V/V).

Phosphate buffer solution Dissolve 0.54 g of potassium

dihydrogen phosphate R and 8.9 g of disodium hydrogen

phosphate dihydrate R in 1.0 L of water R.

Solution A 1.8 g/L solution of ascorbic acid R.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase:

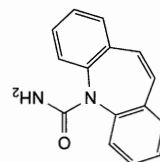
Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 7	60	40
7 - 8	60 → 5	40 → 95
8 - 13	5	95

Injection Test solution (b) and reference solution (c).
Calculate the percentage content of $C_{15}H_{12}N_2O_2$ taking into account the assigned content of oxcarbazepine CRS.

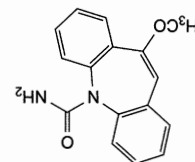
IMPURITIES

Specified impurities B, I, K, L

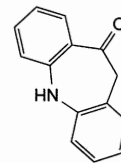
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, D, E, F, G, H, M.



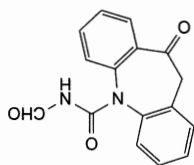
A. 5*H*-dibenzo[*b,j*]azepine-5-carboxamide (carbamazepine),



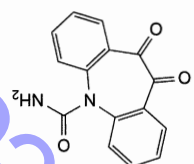
B. 10-methoxy-5*H*-dibenzo[*b,j*]azepine-5-carboxamide (10-methoxycarbamazepine),



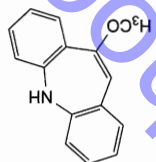
C. 5,11-dihydro-10*H*-dibenzo[*b,j*]azepin-10-one,



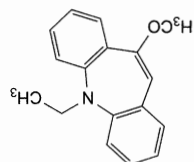
K. *N*-formyl-10-oxo-10,11-dihydro-5*H*-dibenzo[*b,j*]azepine-5-carboxamide,



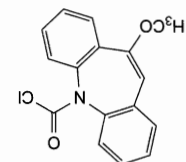
H. 10-methoxy-5*H*-dibenzo[*b,j*]azepine,



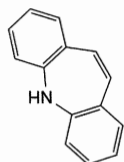
G. 5-ethyl-10-methoxy-5*H*-dibenzo[*b,j*]azepine,



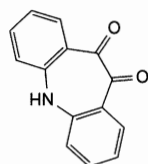
F. 10-methoxy-5*H*-dibenzo[*b,j*]azepine-5-carbonyl chloride,



E. 5*H*-dibenzo[*b,j*]azepine,



D. 5*H*-dibenzo[*b,j*]azepine-10,11-dione,



Related substances

Gas chromatography (2.2.28): use the normalisation procedure. Prepare the solutions immediately before use.

Test solution Dissolve 0.500 g of the substance to be examined in water R and dilute to 50 mL with the same solvent. Add 1 mL of a 10.3 g/L solution of hydrochloric acid R and shake with 3 quantities, each of 10 mL, of methylene chloride R. Combine

the lower layers obtained to the lower layers obtained previously, add anhydrous sodium sulfate R, shake, filter and evaporate the filtrate, at a temperature not exceeding 30 °C, using a rotary evaporator. Take up the residue with methylene chloride R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of oxeladin impurity D CRS in 10 mL of water R, add 0.5 mL of concentrated ammonia R and shake with 3 quantities, each of 2 mL, of methylene chloride R. To the combined lower layers, add 0.2 mL of the test solution and dilute to 10.0 mL with methylene chloride R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with methylene chloride R. Dilute 1.0 mL of this solution to 20.0 mL with methylene chloride R.

Reference solution (c) Dissolve 5 mg of oxeladin impurity C CRS in 10 mL of water R, add 0.5 mL of concentrated ammonia R and shake with 3 quantities, each of 2 mL, of methylene chloride R. Combine the lower layers and dilute to 10 mL with methylene chloride R.

Column:

— material: fused silica,
size: $l = 25$ m, $\varnothing = 0.32$ mm,

— stationary phase: poly(dimethyl) (diphenyl)siloxane R (film thickness 0.4 μ m).

Carrier gas helium for chromatography R.

Flow rate 1.0 mL/min. Adjust the flow rate if necessary to obtain a retention time of about 13 min for oxeladin.

Split ratio 1:15.

Temperature:

Temperature (°C)	Time (min)	Column	Injection port	Detector
160	0.4			
160 → 240	4.12			
240	12.21			
240 → 160	21.30			
280				
280				

Detection Flame ionisation.

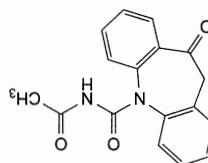
Injection 1 μ L.

Relative retention With reference to oxeladin (retention time = about 13 min): impurity A = about 0.2; impurity B = about 0.4; impurity C = about 0.8; impurity D = about 0.9.

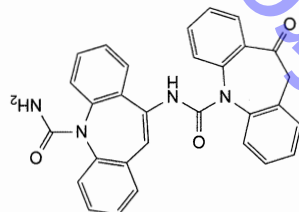
System suitability: reference solution (a):
— resolution: minimum 10 between the peaks due to impurity D and oxeladin.

Limits:

— impurity C: maximum 0.2 per cent,
— impurity D: maximum 0.3 per cent,
— any other impurity: for each impurity, maximum 0.1 per cent.



L. N-acetyl-10-oxo-10,11-dihydro-5H-dibenzo[b,j]azepine-5-carboxamide,

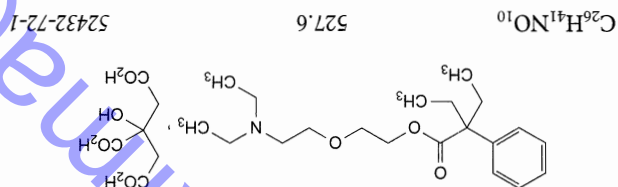


M. 10-[[[(10-oxo-10,11-dihydro-5H-dibenzo[b,j]azepin-5-yl)carbonyl]amino]-5H-dibenzo[b,j]azepine-5-carboxamide.

Ph Eur

Oxeladin Hydrogen Citrate

(Ph. Eur. monograph 1761)



Action and use
Cough suppressant.

Ph Eur

DEFINITION

2-[2-(Diethylamino)ethoxy]ethyl 2-ethyl-2-phenylbutanoate dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly to very slightly soluble in ethyl acetate.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison oxeladin hydrogen citrate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in anhydrous ethanol R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).
Dissolve 2.0 g in water R and dilute to 10.0 mL with the same solvent.

- *total*: maximum 1.0 per cent,
- *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 3 h.

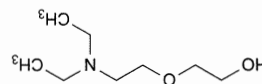
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.400 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 52.76 mg of $C_{26}H_{41}NO_{10}$.

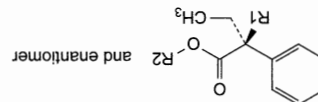
IMPURITIES

Specified impurities C, D.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B.



A. 2-[2-(diethylamino)ethoxy]ethanol,



B. R1 = C_2H_5 , R2 = H: 2-ethyl-2-phenylbutanoic acid,

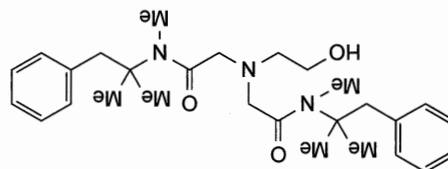
C. R1 = C_2H_5 , R2 = $[CH_2]_2-N(C_2H_5)_2$:

2-(diethylamino)ethyl 2-ethyl-2-phenylbutanoate,

D. R1 = H, R2 = $[CH_2]_2-O-[CH_2]_2-N(C_2H_5)_2$:
2-[2-(diethylamino)ethoxy]ethyl (2RS)-2-phenylbutanoate.

Ph Eur

Oxetacaine



$C_{28}H_{41}NO_3$

467.6

126-27-6

Action and use
Local anaesthetic.

DEFINITION

Oxetacaine is 2,2'-(2-hydroxyethylimino)bis[N-(α,α -dimethylphenylethyl)-N-methylacetamide]. It contains not less than 99.0% and not more than 100.5% of $C_{28}H_{41}NO_3$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder.

Practically insoluble in water; freely soluble in methanol; soluble in ethyl acetate.

IDENTIFICATION

The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of oxetacaine (RS 254).

TESTS

Melting point
100 °C to 104 °C, Appendix V A.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in ethyl acetate.

(1) 10.0% w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 200 volumes.

(3) Dilute 1 volume of solution (2) to 5 volumes.

CHROMATOGRAPHIC CONDITIONS

(a) Use a silica gel 60 precoated plate (Merck plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 5 μ L of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry it in a current of warm air and spray liberally with a solution containing 6% w/v of ammonium thiocyanate and 2% w/v of cobalt(II) chloride.

Carefully remove excess solution by applying filter paper to the plate and allow the plate to dry in air for 10 minutes or until spots appear.

MOBILE PHASE

1 volume of 18M ammonia, 20 volumes of absolute ethanol and 79 volumes of toluene.

LIMITS

In the chromatogram obtained with solution (1):

any secondary spot is not more intense than the spot in the chromatogram obtained with solution (2) (0.5%);

not more than one secondary spot is more intense than the spot in the chromatogram obtained with solution (3) (0.1%).

Loss on drying

When dried at 60 °C at a pressure not exceeding 0.7 kPa for 4 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

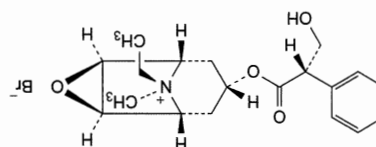
Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 1 g in 50 mL of anhydrous acetic acid and carry out Method I for non-aqueous titration, Appendix VIII A, determining the end-point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 46.76 mg of $C_{28}H_{41}NO_3$.

Oxitropium Bromide

(Ph. Eur. monograph 2170)

C₁₉H₂₆BrNO₄ 412.3 30286-75-0

Action and use

Anticholinergic; treatment of reversible airways obstruction.

Ph. Eur.

DEFINITION

(1R,2R,4S,5S,7S,9S)-9-Ethyl-7-[[[(2S)-3-hydroxy-2-phenylpropanoyl]oxy]-9-methyl-3-oxa-9-azoniatricyclo[3.3.1.0^{2,4}.0^{3,7}]nonane bromide (ethylihyosine).

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, freely soluble in methanol, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison oxitropium bromide CRS.

If the spectra obtained in the solid state show differences at about 1700 cm⁻¹ and about 3300 cm⁻¹, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

B. It gives reaction (a) of bromides (2.3.1).

TESTS

Specific optical rotation (2.2.7)

-26 to -24 (dried substance).

Dissolve 1.0 g in water R and dilute to 20.0 mL with the same solvent.

Related substances

with the mobile phase.

Reference solution (a) Dissolve 7.5 mg of oxitropium bromide impurity B CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Mix 5.0 mL of the test solution and 5.0 mL of reference solution (a).

Reference solution (d) Dilute 15.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (e) Dilute 5.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.0$ mm;— stationary phase: base-deactivated octylsilyl silica gel for chromatography R (5 μ m) with a specific surface area of 350 m²/g and a pore size of 6 nm.

Mobile phase acetonitrile for chromatography R, 7.8 g/L solution of sodium dihydrogen phosphate R (10:100 V/V).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 50 μ L of the test solution and reference solutions (b), (c), (d) and (e).

Relative retention With reference to oxitropium (retention time is about 0.8 min); impurity A is about 0.8;

impurity B is about 0.9; impurity C is about 1.3.

System suitability: reference solution (c):

— resolution: minimum 1.6 between the peaks due to impurity B and oxitropium.

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.1 per cent);

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity C: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (1.5 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.10 per cent);

— sum of unspecified impurities: not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (e) (0.05 per cent).

Impurity D

Liquid chromatography (2.2.29).

Test solution Dissolve 75.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 6.0 mg of oxitropium bromide impurity D CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 200.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) To 5.0 mL of the test solution add 5.0 mL of reference solution (a).

Column:

— size: $l = 0.125$ m, $\varnothing = 4.0$ mm;— stationary phase: base-deactivated octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile for chromatography R, 7.8 g/L solution of sodium dihydrogen phosphate R (18.5:100 V/V).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 50 µL of the test solution and reference solutions (b)

and (c).

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to

impurity D and oxitropium.

Limit:

— impurity D: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

ASSAY

Dissolve 0.350 g in 100 mL of water R and add 5.0 mL of

dilute nitric acid R. Titrate with 0.1 M silver nitrate. Determine

the end-point potentiometrically (2.2.20) using a silver

indicator electrode and a silver-silver chloride reference

electrode.

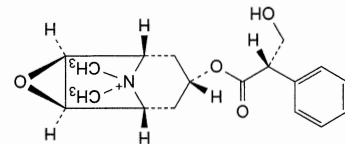
1 mL of 0.1 M silver nitrate is equivalent to 41.23 mg of

C₁₉H₂₆BrNO₄.

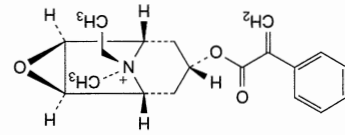
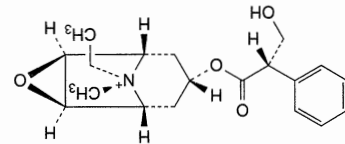
IMPURITIES

Specified impurities A, B, C, D

A. (1R,2R,4S,5S,7S)-9-ethyl-3-oxa-9-azatricyclo[3.3.1.0^{2,4}]non-7-yl [(2S)-3-hydroxy-2-phenylpropionate (N'-ethylmorphosine),



B. (1R,2R,4S,5S,7S)-9-ethyl-7-[[[(2S)-3-hydroxy-2-phenylpropionoyl]oxy]-9-methyl-3-oxa-9-azatricyclo[3.3.1.0^{2,4}]nonane (pseudo-isomer),

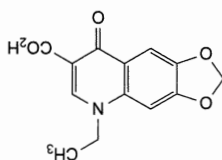


D. (1R,2R,4S,5S,7S)-9-ethyl-9-methyl-7-[(2-phenylacryloyl)oxy]-3-oxa-9-azatricyclo[3.3.1.0^{2,4}]nonane (apo-N'-ethylhyosine).

Ph Eur

Oxolinic Acid

(Ph. Eur. monograph 1353)



C₁₃H₁₁NO₅

261.2

14698-29-4

Action and use

Antibacterial.

Ph Eur

DEFINITION

5-Ethyl-8-oxo-5,8-dihydro-1,3-dioxolo[4,5-g]quinoline-7-carboxylic acid.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Almost white or pale yellow, crystalline powder.

Solubility

Practically insoluble in water, very slightly soluble in methylene chloride, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification B.

Second identification A, C.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 25.0 mg in 5 mL of 0.1 M sodium hydroxide, heating on a water-bath. Allow to cool and dilute to 100.0 mL with methanol R. Dilute 2.0 mL of this solution to 100.0 mL with 0.1 M hydrochloric acid.

Spectral range 220-350 nm.

Absorption maxima At 260 nm, 322 nm and 336 nm.

Absorbance ratio A₂₆₀/A₃₃₆ = 4.9 to 5.2.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in 3 mL of dilute sodium hydroxide solution R and dilute to 20 mL with ethanol (96 per cent) R.

Reference solution (a) Dissolve 10 mg of oxolinic acid CRS in 3 mL of dilute sodium hydroxide solution R and dilute to 20 mL with ethanol (96 per cent) R.

Reference solution (b) Dissolve 5 mg of ciprofloxacin hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 2 mL with reference solution (a).

Plate TLC silica gel plate R.

Mobile phase acetonitrile R, concentrated ammonia R, methanol R, methylene chloride R (10:20:40 V/V/V/V).

Application 10 µL.

Development At the bottom of a chromatographic tank, place an evaporating disk containing 50 mL of concentrated



ammonia R and expose the plate to the ammonia vapour for 15 min in the closed tank; withdraw the plate, transfer to a second chromatographic tank and proceed with development over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

solution (a).

TESTS

Solution S

Dissolve 0.6 g in 20 mL of a 40 g/L solution of sodium hydroxide R.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

Related substances

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in 3 mL of dilute sodium hydroxide solution R and dilute to 10 mL with ethanol (96 per cent) R.

Reference solution (a) Dilute 1 mL of the test solution to 50.0 mL with ethanol (96 per cent) R. Dilute 1.0 mL of this solution to 5.0 mL with ethanol (96 per cent) R.

Reference solution (b) Dissolve 2 mg of oxolinic acid impurity B CRS in ethanol (96 per cent) R and dilute to 10 mL with the same solvent. Dilute 1.0 mL of this solution to 10 mL with ethanol (96 per cent) R.

Reference solution (c) Dissolve 5 mg of the substance to be examined and 5 mg of oxolinic acid impurity A CRS in 2 mL of dilute sodium hydroxide solution R and dilute to 40 mL with ethanol (96 per cent) R.

Plate cellulose for chromatography R as the coating substance.

Mobile phase ammonia R, water R, propanol R (15:30:55 V/V/V).

Application 5 µL, in sufficiently small portions to obtain small spots.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated principal spots.

Limit: impurity B: any spot due to impurity B is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (0.2 per cent); impurities A, C: any spot due to impurities A or C is not more intense than the principal spot in the chromatogram obtained with reference solution (a) (0.4 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by heating in an oven at 105 °C.

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

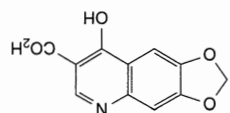
Dissolve 0.200 g in 150 mL of dimethylformamide R. Titrate with 0.1 M tetrabutylammonium hydroxide, determining the end-point potentiometrically (2.2.20). Use a glass indicator electrode and a calomel reference electrode containing, as the electrolyte, a saturated solution of potassium chloride R in methanol R. Carry out a blank titration.

1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to 26.12 mg of C₁₃H₁₁NO₅.

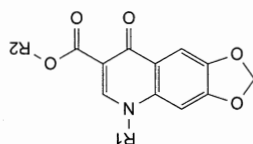
STORAGE

Protected from light.

IMPURITIES



A. 8-hydroxy-1,3-dioxolo[4,5-g]quinoline-7-carboxylic acid,

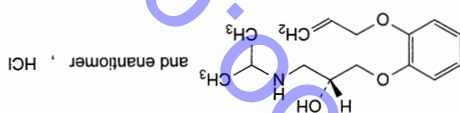


B. R₁ = R₂ = C₂H₅; ethyl 5-ethyl-8-oxo-5,8-dihydro-1,3-dioxolo[4,5-g]quinoline-7-carboxylate,

C. R₁ = CH₃, R₂ = H; 5-methyl-8-oxo-5,8-dihydro-1,3-dioxolo[4,5-g]quinoline-7-carboxylic acid.

Oxprenolol Hydrochloride

(Ph. Eur. monograph 0628)



C₁₅H₂₄ClNO₃ 301.8 6452-73-9

Action and use

Beta-adrenoceptor antagonist.

Preparation

Oxprenolol Tablets

Ph Eur

DEFINITION

(2*RS*)-1-[(1-methylethyl)amino]-3-[2-(prop-2-enyloxy)phenoxy]propan-2-ol hydrochloride.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, freely soluble in alcohol.

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Melting point (2.2.14): 107 °C to 110 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison oxprenolol hydrochloride CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference substance separately in *ethyl acetate* R, evaporate to dryness and record new spectra using the residues.

C. Examine the chromatograms obtained in the test for related substances.

Results The principal spot in the chromatogram obtained with

test solution (b) is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

solution (a).

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in carbon dioxide-free water R and dilute to

20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution GY₆ (2.2.2, Method II).

pH (2.2.3)

4.5 to 6.0 for freshly prepared solution S.

Related substances

Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g of the substance to be

examined in 2 mL of a mixture of 1 volume of methanol R

and 9 volumes of methylene chloride R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL

with a mixture of 1 volume of methanol R and 9 volumes of

methylene chloride R.

Reference solution (a) Dissolve 10 mg of oxprenolol

hydrochloride CRS in 2 mL of a mixture of 1 volume of

methanol R and 9 volumes of methylene chloride R.

Reference solution (b) Dilute 0.4 mL of test solution (a) to

100 mL with a mixture of 1 volume of methanol R and

9 volumes of methylene chloride R.

Reference solution (c) Dilute 5 mL of reference solution (b) to

10 mL with a mixture of 1 volume of methanol R and

9 volumes of methylene chloride R.

Reference solution (d) Dissolve 5 mg of alprenolol

hydrochloride CRS in 1 mL of reference solution (a).

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R, methanol R, methylene

chloride R (2:12:88 V/V/V).

Application 2 µL; allow the spots to dry in air for 15 min.

Development Over a path of 13 cm.

Drying In a current of warm air for 10 min.

Detection Allow to cool and spray with *anisaldehyde solution* R.

Heat at 100–105 °C for 5–10 min. Examine in daylight.

System suitability The test is not valid unless the

chromatogram obtained with reference solution (d) shows

2 clearly separated spots.

Limits: in the chromatogram obtained with test solution (a):

— any impurity: any spot, apart from the principal spot, is

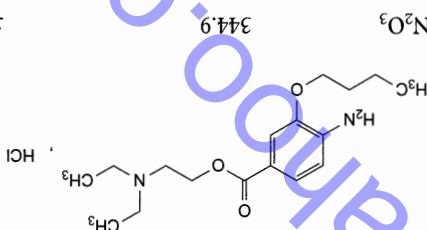
not more intense than the spot in the chromatogram

obtained with reference solution (b) (0.4 per cent);

not more than 1 such spot is more intense than the spot

Oxybuprocaine Hydrochloride

(Ph. Eur. monograph 1251)

C₁₇H₂₉ClN₂O₃

344.9

5987-82-6

Ph Eur

DEFINITION

2-(Diethylamino)ethyl 4-amino-3-butoxybenzoate

hydrochloride.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent). It shows polymorphism (5.9).

STORAGE

Protected from light.

Ph Eur

C₁₅H₂₄ClNO₃

Ph Eur

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of 0.01 M

hydrochloric acid and 50 mL of alcohol R. Carry out a

potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 30.18 mg of

C₁₅H₂₄ClNO₃.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Lead

Maximum 5 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 1.00 g of the substance to be examined

in water R and dilute to 25.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using

0.5 mL and 1.0 mL respectively of lead standard solution

(10 ppm Pb) R diluted to 25.0 mL with water R.

Source Lead hollow-cathode lamp.

Wavelength 217.0 nm.

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

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Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

IDENTIFICATION**First identification B, D.****Second identification A, C, D.****A. Melting point (2.2.14):** 158 °C to 162 °C.**B. Infrared absorption spectrophotometry (2.2.24).****Preparation Discs.****Comparison oxybuprocaine hydrochloride CRS.**

If the spectra obtained show differences, dissolve the

substance to be examined and the reference substance

separately in *methanol R*, evaporate to dryness and record

new spectra using the residues.

C. Thin-layer chromatography (2.2.27).**Test solution** Dissolve 40 mg of the substance to be examined in *methanol R* and dilute to 10 mL with the same solvent.**Reference solution (a)** Dissolve 40 mg of oxybuprocaine*hydrochloride CRS* in *methanol R* and dilute to 10 mL with the same solvent.**Reference solution (b)** Dissolve 20 mg of procaine*hydrochloride R* in reference solution (a) and dilute to 5 mL

with reference solution (a).

Plate TLC silica gel *F₂₅₄* plate *R*.**Mobile phase** anhydrous formic acid *R*, *methanol R*, *water R*, ethyl acetate *R* (10:15:15:60 *V/V/V/V*).**Application** 5 µL.**Development** Over a path of 10 cm.**Drying** In a current of warm air for 10 min.**Detection** Spray with dimethylaminobenzaldehyde solution *R₇*

and examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

solution (a).

TESTS
D. Dilute 0.2 mL of solution S (see Tests) to 2 mL with *water R*. The solution gives reaction (a) of chlorides (2.3.1).**Solution S**Dissolve 5.0 g in carbon dioxide-free *water R* and dilute to

50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution Y₅ (2.2.2, Method II).**pH** (2.2.3)

4.5 to 6.0 for solution S.

Related substances**Liquid chromatography (2.2.29).****Buffer solution** pH 2.5 Add 6 mL of perchloric acid solution *R*and 12 mL of dilute phosphoric acid *R* to 950 mL of *water R*.**Adjust** to pH 2.5 with a 40 g/L solution of sodium*hydroxide R* and dilute to 1000.0 mL with *water R*.**Test solution** Dissolve 10.0 mg of the substance to be

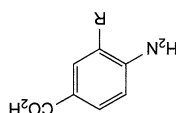
examined in the mobile phase and dilute to 25.0 mL with

the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

20.0 mL with the mobile phase. Dilute 5.0 mL of this

solution to 100.0 mL with the mobile phase.

Reference solution (b) Mix 1.0 mL of the test solution with1 mL of a 40 g/L solution of sodium hydroxide *R* and allow tostand for 20 min. Add 1 mL of dilute phosphoric acid *R* and**A. R = H:** 4-aminobenzoic acid,**B. R = O-CH₂-CH₂-CH₂-CH₃:**

4-amino-3-butoxybenzoic acid,

C. R = OH: 4-amino-3-hydroxybenzoic acid.**IMPURITIES**

Protected from light.

STORAGEof C₁₇H₂₉ClN₂O₃.

1 mL of 0.1 M perchloric acid is equivalent to 34.49 mg

(2.2.20).

ASSAY
Dissolve 0.300 g in a mixture of 20 mL of anhydrous acetic acid *R* and 20 mL of acetic anhydride *R*. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically**ASSAY**

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent determined on 1.000 g by drying in

Loss on drying (2.2.32)reference solution using lead standard solution (1 ppm Pb) *R*.

12 mL of solution S complies with test A. Prepare the

Maximum 10 ppm.

Heavy metals (2.4.8)

(0.0125 per cent).

— the chromatogram obtained with reference solution (a)

(0.25 per cent);

— *total*: not more than the area of the principal peak in the

chromatogram obtained with reference solution (a)

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

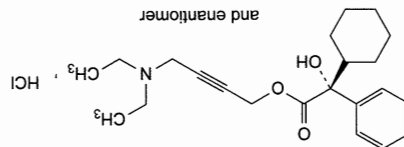
— *any impurity*: for each impurity, not more than 0.4 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

Oxybutynin Hydrochloride

(Ph. Eur. monograph 1354)



$C_{22}H_{32}ClNO_3$ 394.0 1508-65-2

Action and use

Anticholinergic.

Preparation

Oxybutynin Oral Solution

Oxybutynin Tablets

Prolonged-release Oxybutynin Tablets

DEFINITION

4-(Diethylamino)but-2-ynyl (R)-2-cyclohexyl-2-hydroxy-2-phenylacetate hydrochloride.

Content

99.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), soluble in acetone, practically insoluble in cyclohexane.

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Melting point (2.2.14): 124 °C to 129 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Comparison oxybutynin hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of oxybutynin hydrochloride CRS in ethanol (96 per cent) R and dilute to 2 mL with the same solvent.

2 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase methanol R.

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour for 30 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.00 g in water R and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of oxybutynin hydrochloride CRS and 5.0 mg of oxybutynin impurity A CRS in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Column:

— size: l = 0.15 m, Ø = 3.9 mm;

— stationary phase: octylsilyl silica gel for chromatography R₂ (5 µm).

Mobile phase Mix 49 volumes of a solution containing 3.4 g/L of potassium dihydrogen phosphate R and 4.36 g/L of dipotassium hydrogen phosphate R and 51 volumes of acetonitrile R₁.

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Run time Twice the retention time of oxybutynin.

Retention time Oxybutynin = about 15 min;

impurity A = about 24 min.

System suitability: reference solution (a):

— resolution: minimum 1.0 between the peaks due to oxybutynin and impurity A.

Limits:

— impurity A: not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (a) (1.5 per cent);

— unspecified impurities: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— sum of impurities other than A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 3.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

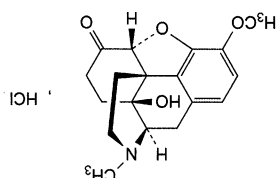
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium

Oxycodone Hydrochloride

(Ph. Eur. monograph 1793)

 $C_{18}H_{22}ClNO_4$

351.9

124-90-3

Action and use

Opioid receptor agonist; analgesic.

Preparations

Oxycodone Capsules
Oxycodone Injection
Oxycodone Oral Solution
Prolonged-release Oxycodone Tablets

Ph Eur

DEFINITION

4,5 α -Epoxy-14-hydroxy-3-methoxy-17-methylmorphinan-6-one hydrochloride.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder, hygroscopic.

Solubility

Freely soluble in water, sparingly soluble in anhydrous ethanol, practically insoluble in toluene.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Dissolve 50 mg in water R and dilute to 5 mL with the same solvent. Render the solution alkaline with dilute ammonia R1. Allow the mixture to stand until a precipitate is formed. Filter, wash the precipitate with 10 mL of cold water R, and dry for 1 h at 105 °C. Examine the precipitate.

Comparison Repeat the operations using 50 mg of oxycodone hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 1.00 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Acidity or alkalinity

To 10 mL of solution S add 0.05 mL of methyl red solution R. Not more than 0.2 mL of 0.02 M sodium hydroxide or 0.02 M hydrochloric acid is required to change the colour of the indicator.

Specific optical rotation (2.2.7)

–140 to –148 (anhydrous substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions protected from light.

hydroxide. Read the volume added between the 2 points of

1 mL of 0.1 M sodium hydroxide is equivalent to 39.4 mg of

$C_{22}H_{32}ClNO_3$.

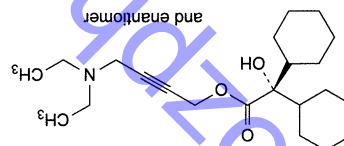
STORAGE

Protected from light.

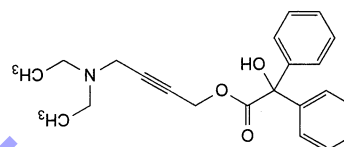
IMPURITIES

Specified impurities A.

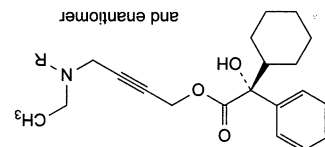
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E.



A. 4-(diethylamino)but-2-ynyl (R,S)-2-(cyclohex-3-enyl)-2-cyclohexyl-2-hydroxyacetate,

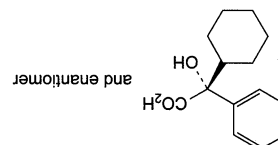


B. 4-(diethylamino)but-2-ynyl 2-hydroxy-2-phenylacetate (diphenyl analogue of oxycodone),



C. R = CH₃: 4-(ethylmethylamino)but-2-ynyl (R,S)-2-cyclohexyl-2-hydroxy-2-phenylacetate (methylmethyl analogue of oxycodone),

E. R = CH₂-CH₂-CH₃: 4-(ethylpropylamino)but-2-ynyl (R,S)-2-cyclohexyl-2-hydroxy-2-phenylacetate (ethylpropyl analogue of oxycodone),



D. (R,S)-2-cyclohexyl-2-hydroxy-2-phenylacetic acid (phenylcyclohexylglycolic acid).

Ph Eur



Test solution Dissolve 0.100 g of the substance to be examined in a 1 per cent *V/V* solution of dilute acetic acid *R* and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 20.0 mg of oxycodone impurity *D* CRS in a 1 per cent *V/V* solution of dilute acetic acid *R* and dilute to 10.0 mL with the same solution.

Reference solution (b) To 1.0 mL of the test solution, add 1.0 mL of reference solution (a) and dilute to 100.0 mL with a 1 per cent *V/V* solution of dilute acetic acid *R*. Dilute 1.0 mL of the solution to 10.0 mL with a 1 per cent *V/V* solution of dilute acetic acid *R*.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography *R* (5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase *A*: mix 830 mL of a 1.1 g/L solution of sodium heptanesulfonate monohydrate *R* previously adjusted to pH 2.0 with a mixture of equal volumes of phosphoric acid *R* and water *R*, with 70 mL of acetonitrile *R* and 100 mL of methanol *R*;

— mobile phase *B*: mix 600 mL of a 1.1 g/L solution of sodium heptanesulfonate monohydrate *R* previously adjusted to pH 2.0 with a mixture of equal volumes of phosphoric acid *R* and water *R*, with 150 mL of acetonitrile *R* and 250 mL of methanol *R*;

Time (min)	Mobile phase A (per cent <i>V/V</i>)	Mobile phase B (per cent <i>V/V</i>)
0 - 60	100 → 50	0 → 50

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Relative retention With reference to oxycodone (retention time = about 24 min): impurity *A* = about 0.4;

impurity *B* = about 0.7; impurity *C* = about 1.14;

impurity *D* = about 1.8; impurity *E* = about 1.18;

impurity *F* = about 2.4.

System suitability: reference solution (b):

— resolution: minimum 3 between the peaks due to oxycodone and impurity *D*.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity *F* by 0.5;

— sum of impurities *D* and *E*: not more than 10 times the area of the peak due to oxycodone in the chromatogram;

obtained with reference solution (b) (1.0 per cent);

— impurities *A*, *B*, *C*, *F*: for each impurity, not more than the area of the peak due to oxycodone in the

chromatogram obtained with reference solution (b) (0.1 per cent);

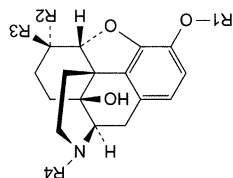
— any other impurity: for each impurity, not more than the area of the peak due to oxycodone in the chromatogram;

obtained with reference solution (b) (0.1 per cent);

— total: not more than 15 times the area of the peak due to oxycodone in the chromatogram obtained with reference

— **disregard limit:** 0.5 times the area of the peak due to oxycodone in the chromatogram obtained with reference

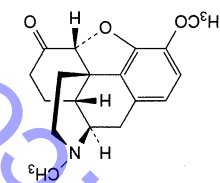
— solution (b) (0.05 per cent).



A. $R_1 = H$, $R_2 = R_3 = O$, $R_4 = CH_3$: 4,5 α -epoxy-3,14-dihydroxy-17-methylmorphinan-6-one (oxymorphone),

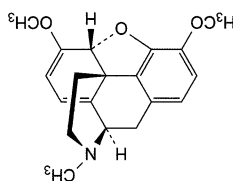
B. $R_1 = R_4 = CH_3$, $R_2 = OH$, $R_3 = H$: 4,5 α -epoxy-3-methoxy-17-methylmorphinan-6 α ,14-diol (7,8-dihydro-14-hydroxycodone),

C. $R_1 = CH_3$, $R_2 + R_3 = O$, $R_4 = H$: 4,5 α -epoxy-14-hydroxy-3-methoxymorphinan-6-one (noroxycodone),



D. 7,8-didehydro-4,5 α -epoxy-14-hydroxy-3-methoxy-17-methylmorphinan-6-one (14-hydroxycodone),

E. 4,5 α -epoxy-3-methoxy-17-methylmorphinan-6-one (hydrocodone),



F. 6,7,8,14-tetrahydro-4,5 α -epoxy-3-6-dimethoxy-17-methylmorphinan (thebain).

IMPURITIES

In an airtight container, protected from light.

STORAGE

35.19 mg of $C_{18}H_{22}ClNO_4$.

1 mL of 0.1 *M* ethanolic sodium hydroxide is equivalent to used between the 2 inflexion points.

end-point potentiometrically (2.2.20). Measure the volume with 0.1 *M* ethanolic sodium hydroxide, determining the hydrochloric acid and 60 mL of ethanol (96 per cent) *R*. Titrate Dissolve 0.250 g in a mixture of 5.0 mL of 0.01 *M*

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 7.0 per cent, determined on 0.250 g.

Water (2.5.12)

Maximum 1.0 per cent.

Ethanol (2.4.24, System *A*)

Oxygen

(Ph. Eur. monograph 0417)

32.00 7782-44-7 O₂

Oxygen should be kept in approved metal cylinders, the shoulders of which are painted white and the remainder black. The cylinder should carry a label stating 'Oxygen'. In addition, 'Oxygen' or the symbol 'O₂' should be stencilled in paint on the shoulder of the

When Oxygen is intended for use in a room in which magnetic resonance imaging (MRI) is being performed, the cylinder and fittings should be made from suitable non-ferromagnetic materials and labelled accordingly.

DEFINITION

Minimum 99.5 per cent W/V of O₂.

This monograph applies to oxygen for medicinal use.

CHARACTERS

Appearance

At 20 °C and at a pressure of 101 kPa, 1 volume dissolves in about 32 volumes of water.

PRODUCTION

Oxygen is produced by a purification process followed by

Carbon dioxide Maximum 300 ppm V/V , determined using an infrared

Reference gas (a) Oxygen R.
Reference gas (b) Mixture containing 300 ppm V/V of carbon dioxide R1 in nitrogen R1.

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon dioxide in the gas to be examined.

Carbon monoxide

Maximum 5 ppm ^{13}C , determined using an infrared analyser (2.5.25).

(Gas to be examined filter the substance to be examined to avoid stray light phenomena.

Reference gas (a) Oxygen R_1 .
Reference gas (b) Mixture containing 5 ppm V/V of carbon monoxide R in nitrogen R_1 .

Water

Maximum 67 ppm W/W , determined using an electrolytic hygrometer (2.5.28).

Assay

Determine the concentration of oxygen using a paramagnetic analyser (2.5.27).

IDENTIFICATION

It complies with the limits of the assay.

TESTS

Carbon dioxide Maximum 300 ppm *W/W*, determined using a carbon dioxide detector tube (2.1.6).

Carbon monoxide

Maximum 5 ppm W/W , determined using a carbon monoxide detector tube (2.1.6).

Water vapour

Maximum 6 ppm, determined using a water vapour detector tube (2.1.6).

STORAGE

As a compressed gas or liquid in appropriate containers, complying with the legal regulations. Oils and grease are not to be used unless they are oxygen-compatible.

IMPAIRMENTS

A. CO₂: carbon dioxide,
B. CO: carbon monoxide,
C. H₂O: water.

Ph Eur

DEFINITION

This monograph applies to oxygen (93 per cent) for medical use. It does not apply to gas produced using individual concentrators for domiciliary use.

PRODUCTION

Oxygen (93 per cent) is produced in single-stage concentrators by adsorption purification of ambient air using zeolites. During production, the oxygen content is continuously monitored by means of a paramagnetic analyser (2.5.27). Following the design and installation of the concentrator, and after any modification or significant intervention, the gas produced complies with the following requirements.

Carbon dioxide

Gas to be examined The substance to be examined. It must be filtered to avoid stray light phenomena.

Reference gas (a) Oxygen R_1 .

Reference gas (b) A mixture of 7 per cent V/V of nitrogen R_1 and 93 per cent V/V of oxygen R_1 , containing 300 ppm V/V of carbon dioxide R_1 .

Carbon monoxide

the gas to be examined.
Carbon monoxide
Maximum 5 ppm V/V , determined using an infrared analyser (2.5.25).
Gas to be examined The substance to be examined. It must be filtered to avoid stray light phenomena.
Reference gas (a) Oxygen R .
Reference gas (b) A mixture containing 5 ppm V/V of carbon monoxide R in nitrogen $R1$.



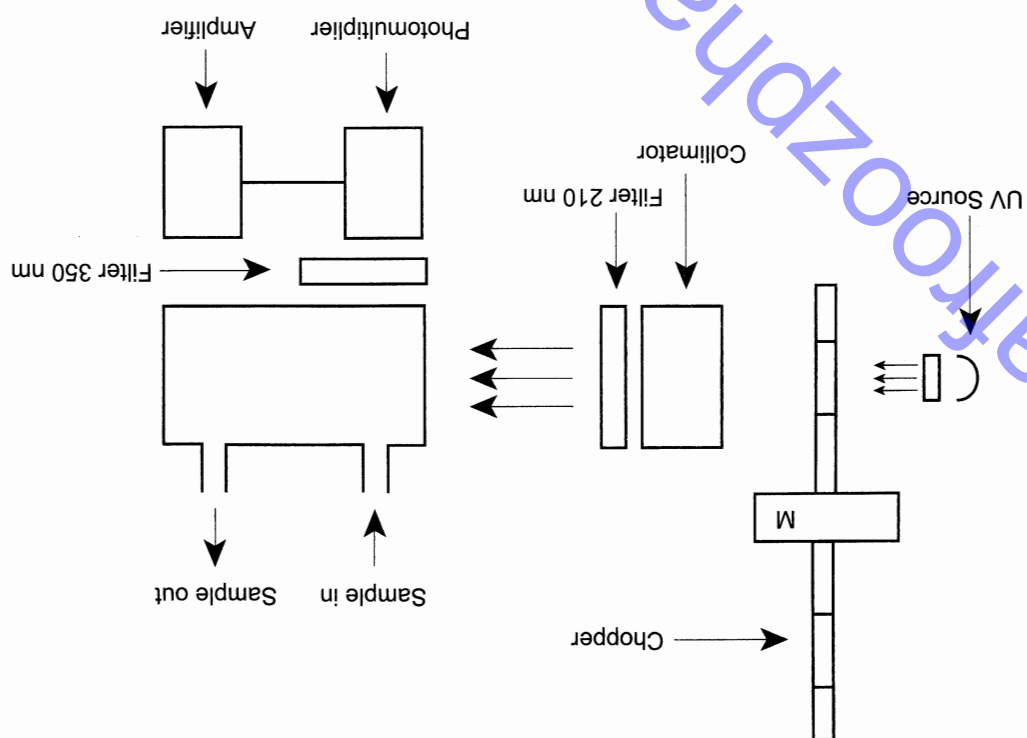


Figure 2455-1. - UV fluorescence analyser

Figure 2455-1. - UV fluorescence analyser

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of carbon monoxide in the gas to be examined.

Nitrogen monoxide and nitrogen dioxide
Maximum 2 ppm V/V in total, determined using a chemiluminescence analyser (2.5.26).

Gas to be examined The substance to be examined.
Reference gas (a) A mixture of 21 per cent V/V of oxygen R and 79 per cent V/V of nitrogen R_1 , containing less than 0.05 ppm V/V of nitrogen monoxide and nitrogen dioxide.

Reference gas (b) A mixture containing 2 ppm V/V of nitrogen dioxide R in nitrogen R_1 .

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of nitrogen monoxide and nitrogen dioxide in the gas to be examined.

Sulfur dioxide

Maximum 1 ppm V/V , determined using an ultraviolet fluorescence analyser (Figure 2455-1).

The apparatus consists of the following:
— a system generating ultraviolet radiation with a wavelength of 210 nm, made up of an ultraviolet lamp, a collimator, and a selective filter; the beam is blocked periodically by a chopper rotating at high speeds;
— a reaction chamber, through which flows the gas to be examined;

— a system that detects radiation emitted at a wavelength of 350 nm, made up of a selective filter, a photomultiplier tube and an amplifier.
Gas to be examined The substance to be examined. It must be filtered.

Reference gas (a) A mixture of 7 per cent V/V of nitrogen R_1 and 93 per cent V/V of oxygen R .

Reference gas (b) A mixture of 7 per cent V/V of nitrogen R_1 and 93 per cent V/V of oxygen R , containing 0.5 ppm V/V to 2 ppm V/V of sulfur dioxide R_1 .

Calibrate the apparatus and set the sensitivity using reference gases (a) and (b). Measure the content of sulfur dioxide in the gas to be examined.

Oil
Maximum 0.1 mg/m³, determined using an oil detector tube (2.1.6).

Water
Maximum 67 ppm V/V , determined using an electrolytic hygrometer (2.5.28).

Assay
Determine the concentration of oxygen using a paramagnetic analyser (2.5.27).

CHARACTERS

Appearance
Colourless gas.

IDENTIFICATION

It complies with the limits of the assay.

TESTS

Carbon dioxide
Maximum 300 ppm V/V , determined using a carbon dioxide detector tube (2.1.6).

Carbon monoxide
Maximum 5 ppm V/V , determined using a carbon monoxide detector tube (2.1.6).

Nitrogen monoxide and nitrogen dioxide
Maximum 2 ppm V/V in total, determined using a nitrogen monoxide and nitrogen dioxide detector tube (2.1.6).

Sulfur dioxide
Maximum 1 ppm *V/V*, determined using a sulfur dioxide detector tube (2.1.6).

Oil
Maximum 0.1 mg/m³, determined using an oil detector tube (2.1.6).

Water vapour
Maximum 67 ppm *V/V*, determined using a water vapour detector tube (2.1.6).

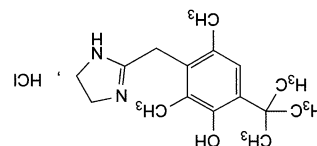
ASSAY
Determine the content of oxymetazoline using a paramagnetic analyser (2.5.27).

STORAGE
Oxygen 93 per cent obtained from an oxygen concentrator is normally used on the site where it is produced. It is fed directly into a medicinal gas pipeline or administration system. Where authorised by the competent authority, it may be stored in suitable containers complying with the legal regulations. Oils and grease are not to be used unless they are oxygen-compatible.

IMPURITIES
A. CO₂; carbon dioxide,
B. CO; carbon monoxide,
C. SO₂; sulfur dioxide,
D. NO and NO₂; nitrogen monoxide and nitrogen dioxide,
E. oil,
F. H₂O; water.

Oxymetazoline Hydrochloride

(Ph. Eur. monograph 0943)



C₁₆H₂₅ClN₂O

296.8

2315-02-8

Action and use
Alpha-adrenoceptor agonist; decongestant.

DEFINITION

3-[(4,5-Dihydro-1*H*-imidazol-2-yl)methyl]-6-(1,1-dimethylethyl)-2,4-dimethylphenol hydrochloride.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification A, D.
Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison oxymetazoline hydrochloride CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in a mixture of equal volumes of ethyl acetate R and methanol R and dilute to 5 mL with the same mixture of solvents.

Reference solution Dissolve 20 mg of oxymetazoline hydrochloride CRS in a mixture of equal volumes of ethyl acetate R and methanol R and dilute to 5 mL with the same mixture of solvents.

Plate TLC silica gel G plate R.

Mobile phase diethylamine R, cyclohexane R, anhydrous ethanol R (6:15:79 *V/V/V*).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air for 5 min, then allow to cool. Detection Spray with a freshly prepared 5.0 g/L solution of potassium ferricyanide R in ferric chloride solution R2; examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve about 2 mg in 1 mL of water R, then add 0.2 mL of a 50 g/L solution of sodium nitroprusside R and 0.2 mL of dilute sodium hydroxide solution R. Allow to stand for 10 min. Add 2 mL of sodium hydrogen carbonate solution R. A violet colour develops.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Dissolve 2.5 g in water R and dilute to 50 mL with the same solvent.

Acidity or alkalinity

Dissolve 0.25 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent. Add 0.1 mL of methyl red solution R and 0.2 mL of 0.01 M hydrochloric acid. The solution is red. Not more than 0.4 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to yellow.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 50.0 mg of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with water R. Dilute 2.0 mL of this solution to 100.0 mL with water R.

Reference solution (b) Dissolve 5.0 mg of oxymetazoline impurity A CRS and 5 mg of the substance to be examined in water R and dilute to 50.0 mL with the same solvent. Dilute 10.0 mL of the solution to 50.0 mL with water R.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 20.0 mL with water R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography with polar incorporated groups R (5 µm).

Mobile phase:
— mobile phase A: 1.36 g/L solution of potassium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R;
— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	70	30
5 - 20	70 → 15	30 → 85
20 - 35	15	85

Flow rate 1.0 mL/min.
Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Relative retention With reference to oxymetazoline (retention time = about 5.0 min): impurity A = about 0.9.

System suitability: reference solution (b):

— resolution: minimum 4.0 between the peaks due to

impurity A and oxymetazoline.

Limits:

— impurity A: not more than 1.5 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (c) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

Water (2.5, 32)

Maximum 0.3 per cent, determined on 1.00 g.

Sulfated ash (2.4, 14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 20 mL of acetic anhydride R

and 20 mL of anhydrous acetic acid R. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2, 20).

1 mL of 0.1 M perchloric acid is equivalent to 29.68 mg of

C₁₆H₂₅ClN₂O.

IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

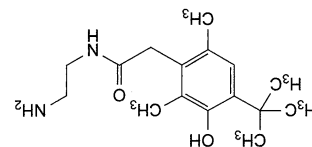
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

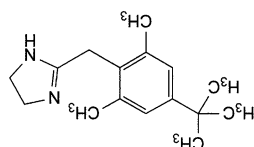
impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: B, C,

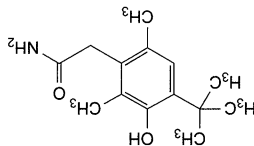
D, E.



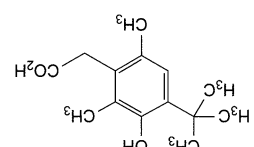
A. N-(2-aminoethyl)-2-[4-(1,1-dimethylethyl)-3-hydroxy-2,6-



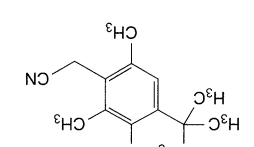
B. 2-[[4-(1,1-dimethylethyl)-2,6-dimethylphenyl]methyl]-4,5-



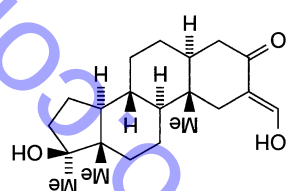
C. 2-[4-(1,1-dimethylethyl)-3-hydroxy-2,6-



D. 2-[4-(1,1-dimethylethyl)-3-hydroxy-2,6-



E. 2-[4-(1,1-dimethylethyl)-3-hydroxy-2,6-



Oxymetholone

C₂₁H₃₂O₃

332.5

434-07-1

Action and use

Anabolic steroid; androgen.

Preparation

Oxymetholone Tablets

DEFINITION

Oxymetholone is 17β-hydroxy-2-hydroxymethylene-17α-methyl-5α-androstan-3-one. It contains not less than 97.0% and not more than 103.0% of C₂₁H₃₂O₃, calculated with reference to the dried substance.

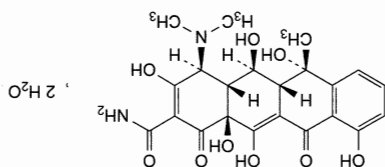
CHARACTERISTICS

A white to creamy white crystalline powder. It exhibits polymorphism.



Oxytetracycline Dihydrate

(Ph. Eur. monograph 0199)



$C_{22}H_{24}N_2O_9 \cdot 2H_2O$ 496.4

Action and use
Tetracycline antibacterial.
Preparation
Oxytetracycline Tablets

Ph. Eur.

DEFINITION

(4S,4aR,5S,5aR,6S,12aS)-4-(Dimethylamino)-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,1-dioxo-1,4,4a,5,5a,6,6a,11,12a-octahydro-2-tetracycline-2-carboxamide dihydrate.

Substance produced by the growth of certain strains of *Streptomyces rimosus* or obtained by any other means.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
Yellow, crystalline powder.

Solubility

Very slightly soluble in water. It dissolves in dilute acid and alkaline solutions.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of oxytetracycline CRS in methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of oxytetracycline CRS, 5 mg of tetracycline hydrochloride R and 5 mg of minocycline hydrochloride R in methanol R and dilute to 10 mL with the same solvent.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a 63 g/L solution of oxalic acid R previously adjusted to pH 2 with concentrated ammonia R.

Application 1 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of *sulfuric acid* R. A deep red colour develops. Add the solution to 2.5 mL of *water* R. The colour becomes yellow.

Practically insoluble in water; soluble in ether.

IDENTIFICATION

A. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.002% w/v solution in 0.01M *ethanolic sodium hydroxide* exhibits a maximum only at 315 nm. The *absorbance* at 315 nm is about 1.1.

B. The *light absorption*, Appendix II B, in the range 230 to 350 nm of a 0.003% w/v solution in 0.01M *ethanolic hydrochloric acid* exhibits a maximum only at 277 nm. The *absorbance* at 277 nm is about 1.0.

C. The *infrared absorption spectrum*, Appendix II A, is concordant with the *reference spectrum* of oxytetracycline (RS 256). If the spectra are not concordant, dissolve the substance in the minimum of *chloroform*, evaporate to dryness, dry the residue over *phosphorus pentoxide* at a pressure not exceeding 0.7 kPa and prepare a new spectrum. D. Completes with the test for identification of steroids, Appendix III A, using impregnating solvent II and mobile phase D.

TESTS

Melting point

175° to 180°, Appendix V A.

Specific optical rotation

In a 2% w/v solution in 1,4-dioxan, +34 to +38, calculated with reference to the dried substance, Appendix V R.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using silica gel G as the coating substance and a mixture of 98 volumes of *toluene* and 2 volumes of *absolute ethanol* as the mobile phase. Apply separately to the plate 10 µL of each of two solutions of the substance being examined in a mixture of equal volumes of *chloroform* and *ethanol* (96%) containing (1) 1.0% w/v and (2) 0.0050% w/v. After removal of the plate, allow it to dry in air and spray with *vanillin-ethanolic sulfuric acid reagent*. Any secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.5%).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.1 g in sufficient 0.01M *ethanolic sodium hydroxide* to produce 200 mL, dilute 5 mL to 250 mL with 0.01M *ethanolic sodium hydroxide* and measure the *absorbance* of the resulting solution at the maximum at 315 nm, Appendix II B. Calculate the content of $C_{21}H_{22}O_9$ taking 547 as the value of A(1%, 1 cm) at the maximum at 315 nm.

STORAGE

Oxytetracycline should be kept free from contact with ferrous metals and protected from light.

C. Dissolve about 10 mg in a mixture of 1 mL of dilute nitric acid R and 5 mL of water R. Shake and add 1 mL of silver nitrate solution R2. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of dilute nitric acid R, 5 mL of a 0.021 g/L solution of potassium chloride R and 1 mL of silver nitrate solution R2.

TESTS

pH (2.2.3)

4.5 to 7.5.

Suspend 0.1 g in 10 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7)

–203 to –216 (anhydrous substance).

Dissolve 0.250 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Specific absorbance (2.2.25)

290 to 310 determined at 353 nm (anhydrous substance).

Dissolve 20.0 mg in buffer solution pH 2.0 R and dilute to 100.0 mL with the same buffer solution. Dilute 10.0 mL of this solution to 100.0 mL with buffer solution pH 2.0 R.

Light-absorbing impurities

Carry out the measurements within 1 h of preparing the solutions.

Dissolve 20.0 mg in a mixture of 1 volume of 1 M

hydrochloric acid and 99 volumes of methanol R and dilute to 10.0 mL with the same mixture of solvents. The absorbance

(2.2.25), determined at 430 nm has a maximum of 0.25

(anhydrous substance).

Dissolve 0.100 g in a mixture of 1 volume of 1 M hydrochloric

acid and 99 volumes of methanol R and dilute to 10.0 mL

with the same mixture of solvents. The absorbance (2.2.25)

determined at 490 nm has a maximum of 0.20 (anhydrous

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Test solution Dissolve 20.0 mg of the substance to be

examined in 0.01 M hydrochloric acid and dilute to 25.0 mL

with the same acid.

Reference solution (a) Dissolve 20.0 mg of oxytetracycline CRS

in 0.01 M hydrochloric acid and dilute to 25.0 mL with the

same acid.

Reference solution (b) Dissolve 20.0 mg of 4-

epioxytetracycline CRS in 0.01 M hydrochloric acid and dilute

to 25.0 mL with the same acid.

Reference solution (c) Dissolve 20.0 mg of tetracycline

hydrochloride CRS in 0.01 M hydrochloric acid and dilute to

25.0 mL with the same acid.

Reference solution (d) Mix 1.5 mL of reference solution (a),

1.0 mL of reference solution (b) and 3.0 mL of reference

solution (c) and dilute to 25.0 mL with 0.01 M hydrochloric

acid.

Reference solution (e) Mix 1.0 mL of reference solution (b)

and 4.0 mL of reference solution (c) and dilute to 200.0 mL

with 0.01 M hydrochloric acid.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: styrene-divinylbenzene copolymer R (8 μ m);

— temperature: 60 °C.

Mobile phase Weigh 60.0 g of 2-methyl-2-propanol R and

transfer to a 1000 mL volumetric flask with the aid of

200 mL of water R; add 60 mL of 0.33 M phosphate buffer

solution pH 7.5 R, 50 mL of a 10 g/L solution of

tetrabutylammonium hydrogen sulfate R adjusted to pH 7.5

with dilute sodium hydroxide solution R and 10 mL of a 0.4 g/L solution of sodium edetate R adjusted to pH 7.5 with dilute sodium hydroxide solution R; dilute to 1000 mL with water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of the test solution and reference solutions (d)

and (e).

System suitability: reference solution (d):

— resolution: minimum 4.0 between the peaks due to

impurity A (1st peak) and oxytetracycline (2nd peak) and

minimum 5.0 between the peaks and due to

oxytetracycline and impurity B (3rd peak); adjust the

2-methyl-2-propanol content in the mobile phase if

necessary;

— symmetry factor: maximum 1.25 for the peak due to

oxytetracycline.

Limits:

— impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (e) (0.5 per cent);

— impurity B: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (e) (2.0 per cent);

— impurity C (eluting on the tail of the principal peak): not

more than 4 times the area of the peak due to impurity A

in the chromatogram obtained with reference solution (e)

(2.0 per cent);

— disregard limit: 0.02 times the area of the peak due to

oxytetracycline in the chromatogram obtained with

reference solution (d) (0.1 per cent).

Heavy metals (2.4.8)

Maximum 50 ppm.

0.5 g complies with test F. Prepare the reference solution

using 2.5 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

6.0 per cent to 9.0 per cent, determined on 0.250 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

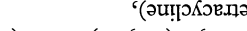
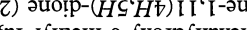
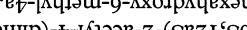
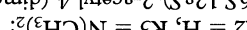
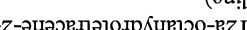
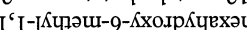
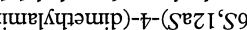
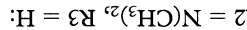
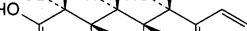
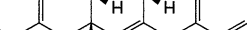
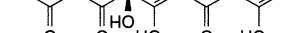
Injection Test solution and reference solution (a).

Calculate the percentage content of C₂₂H₄₄N₂O₉.

STORAGE

In an airtight container, protected from light.

IMPURITIES



(e) After removal of the plate, dry it in a current of air and examine under ultraviolet light (365 nm).

MOBILE PHASE
6 volumes of water, 35 volumes of methanol and 59 volumes of dichloromethane.

SYSTEM SUITABILITY
The test is not valid unless the chromatogram obtained with solution (3) shows two clearly separated spots.

CONFIRMATION

The principal spot in the chromatogram obtained with solution (1) corresponds in position, colour and size to that in the chromatogram obtained with solution (2).
B. To 2 mg add 5 mL of sulfuric acid; a deep red colour is produced. Add the solution to 2.5 mL of water; the colour changes to yellow.

C. Yields reaction B characteristic of calcium salts, Appendix VI.

TESTS

Acidity or alkalinity

pH of a 2.5% w/v suspension, 6.0 to 7.5, Appendix V L.

Light absorption

Absorbance of a 0.002% w/v solution in 0.1M chloride buffer pH 2.0 at the maximum at 353 nm, 0.56 to 0.61, calculated with reference to the anhydrous substance, Appendix II B.

Specific optical rotation

In a 1% w/v solution in 0.1M hydrochloric acid, -194 to -210, calculated with reference to the anhydrous substance, Appendix V F. Allow the solution to stand protected from light for 30 minutes before measurement.

Light-absorbing impurities

A. Dissolve 0.2 g in 6 mL of 1M hydrochloric acid and add sufficient methanol to produce 100 mL. The absorbance at 430 nm, when measured within 1 hour of preparing the solution, is not more than 0.30, calculated with reference to the anhydrous substance, Appendix II B.

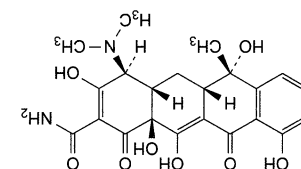
B. Dissolve 1 g in 6 mL of 1M hydrochloric acid and add sufficient methanol to produce 100 mL. The absorbance at 490 nm, when measured within 1 hour of preparing the solution, is not more than 0.20, calculated with reference to the anhydrous substance, Appendix II B.

Calcium

3.90 to 4.20%, calculated with reference to the anhydrous substance, when determined by the following method.

Transfer about 1 g, accurately weighed, to a Kjeldahl flask, cautiously add 10 mL of nitric acid and mix. Allow to stand for 5 minutes, add a glass bead and heat on a water bath for 5 minutes. Remove from the water bath, cautiously add 5 mL of 9M perchloric acid and heat, adding further 5 mL quantities of the perchloric acid at intervals until the liquid is almost colourless. Add 0.1 mL of nitric acid and allow any further reaction to subside. Do not allow the volume of the liquid in the flask to be reduced below 3 mL at any stage in the oxidation. Wash the walls of the flask with 40 mL of water, collecting the washings in the flask, and boil for 3 to 4 minutes to expel chlorine. Cool, transfer the contents of the flask to a conical flask with the aid of water and dilute to about 200 mL with water. Adjust to pH 9 with 5M sodium hydroxide and then add 100 mL of water followed by 12 mL of 10M sodium hydroxide and mix. Add about 15 mg of calcium oxalate VS until the colour changes from violet to full blue. Each mL of 0.05M disodium edetate VS is equivalent to 2.004 mg of Ca.

Oxytetracycline Calcium



B. (4S,4aS,5aS,6S,12aS)-4-(dimethylamino)-3,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracycene-2-carboxamide (tetracycline).

Ph Eur

Action and use
Tetracycline antibacterial.

DEFINITION

Oxytetracycline Calcium is the calcium salt of

(4S,4aR,5S,5aR,6S,12aS)-4-dimethylamino-1,4,4a,5,5a,6,11,12a-octahydro-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,11-dioxonaphthacene-2-carboxamide, a substance produced by the growth of certain strains of *Streptomyces rimosus* or obtained by any other means. It contains not less than 90.0% and not more than 100.5% of $(C_{22}H_{23}N_2O_9)_2Ca$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A pale yellow to greenish fawn, crystalline powder. Practically insoluble in water; soluble in dilute acids. It dissolves slowly in 5M ammonia.

IDENTIFICATION

A. Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in

0.01M methanolic hydrochloric acid.

(1) 0.05% w/v of the substance being examined.

(2) 0.05% w/v of oxytetracycline BPCRS.

(3) 0.05% w/v of each of oxytetracycline BPCRS and demeclocycline hydrochloride BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use a silica gel precoated plate (Merk silica gel 60 plates are suitable). Adjust the pH of a 10% w/v solution of disodium edetate to 7.0 with 10M sodium hydroxide and spray the solution evenly onto the plate (about 10 mL for a plate 100 mm × 200 mm). Allow the plate to dry in a horizontal position for at least 1 hour. Before use, dry the plate at 110° for 1 hour.

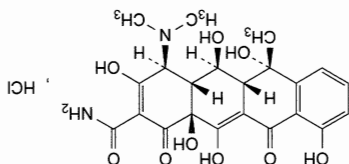
(b) Use the mobile phase as described below.

(c) Apply 1 µL of each solution.

(d) Develop the plate to 15 cm.

Oxytetracycline Hydrochloride

(Ph. Eur. monograph 0198)



$C_{22}H_{25}ClN_2O_9$ 496.9 2058-46-0

Action and use

Tetracycline antibacterial.

Preparation

Oxytetracycline Capsules

Ph Eur

DEFINITION

(4S,4aR,5S,5aR,6S,12aS)-4-(Dimethylamino)-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,1-dioxo-1,4,4a,5,5a,6,11,12a-octahydrodrotetracene-2-carboxamide hydrochloride.

Substance produced by the growth of certain strains of *Streptomyces rimosus* or obtained by any other means.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Yellow, crystalline powder, hygroscopic.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent). Solutions in water become turbid on standing, owing to the precipitation of oxytetracycline.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of oxytetracycline hydrochloride CRS in methanol R and dilute to 10 mL with the

same solvent.

Reference solution (b) Dissolve 5 mg of oxytetracycline hydrochloride CRS, 5 mg of tetracycline hydrochloride R and 5 mg of minocycline hydrochloride R in methanol R and dilute

to 10 mL with the same solvent.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a 63 g/L solution of oxalic acid R previously adjusted to pH 2 with concentrated

ammonia R.

Application 1 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference

solution (a).

ASSAY

Not more than 15.0% w/w, Appendix IX C. Use 0.25 g.

Carry out the method for liquid chromatography, Appendix III D, using the following solutions.

(1) Dilute 1 volume of a 0.1% w/v solution of the substance being examined in 0.1M hydrochloric acid to 20 volumes with the mobile phase.

(2) Dilute 1 volume of a 0.1% w/v solution of oxytetracycline BPCRS in 0.1M hydrochloric acid to 20 volumes with the mobile phase.

(3) 0.1% w/v of 4-epioxytetracycline EPCRS in

0.1M hydrochloric acid.

(4) 0.1% w/v of tetracycline hydrochloride BPCRS in

0.1M hydrochloric acid.

(5) Dilute a mixture containing 1.5 mL of a 0.1% w/v

solution of oxytetracycline BPCRS in 0.1M hydrochloric acid,

1 mL of solution (3) and 3 mL of solution (4) to 25 mL

with the mobile phase.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with styrene-divinylbenzene copolymer (8 to 10 µm) (Polymer Laboratories, PLRP-S 100A, is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use a column temperature of 60°.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

To 50.0 g of 2-methyl-propan-2-ol add 200 mL of water, 60 mL of 0.33M phosphoric buffer pH 7.5, 50 mL of a 1.0% w/v solution of tetrabutylammonium hydrogen sulfate previously adjusted to pH 7.5 with 2M sodium hydroxide and 10 mL of a 0.04% w/v solution of disodium edetate previously adjusted to pH 7.5 with 2M sodium hydroxide and dilute to 1 litre with water.

SYSTEM SUITABILITY

The Assay is not valid unless, in the chromatogram obtained with solution (5):

the resolution factor between the first peak

(4-epioxytetracycline) and the second peak (oxytetracycline)

is at least 4.0;

the resolution factor between the second peak and the third

peak (tetracycline) is at least 5.0 (if necessary, reduce the

content of 2-methylpropan-2-ol in the mobile phase to

increase the resolution);

the symmetry factor of the peak due to oxytetracycline is not

more than 1.25.

DETERMINATION OF CONTENT

Calculate the content of $(C_{22}H_{23}N_2O_9)_2Ca$ using the declared content of $C_{22}H_{24}N_2O_9$ in oxytetracycline BPCRS. Each mg of $C_{22}H_{24}N_2O_9$ is equivalent to 1.041 mg of $(C_{22}H_{23}N_2O_9)_2Ca$.

STORAGE

Oxytetracycline Calcium should be protected from light and stored at a temperature of 2° to 8°.

LABELLING

The label states (1) the date after which the material is not intended to be used; (2) the conditions under which it should be stored.

B. To about 2 mg add 5 mL of sulfuric acid R. A deep red colour develops. Add the solution to 2.5 mL of water R. The colour becomes yellow.

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

pH (2.2.3) 2.3 to 2.9.

Dissolve 0.1 g in 10 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7) –188 to –200 (anhydrous substance).

Dissolve 0.250 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Specific absorbance (2.2.25) 270 to 290 determined at 353 nm (anhydrous substance).

Dissolve 20.0 mg in buffer solution pH 2.0 R and dilute to 100.0 mL with the same buffer solution. Dilute 10.0 mL of the solution to 100.0 mL with buffer solution pH 2.0 R.

Light-absorbing impurities

Carry out the measurements within 1 h of preparing the solutions. Dissolve 20.0 mg in a mixture of 1 volume of 1 M hydrochloric acid and 99 volumes of methanol R and dilute to 10.0 mL with the same mixture of solvents. The absorbance (2.2.25) determined at 430 nm has a maximum of 0.50 (anhydrous substance).

Dissolve 0.100 g in a mixture of 1 volume of 1 M hydrochloric acid and 99 volumes of methanol R and dilute to 10.0 mL with the same mixture of solvents. The absorbance (2.2.25) determined at 490 nm has a maximum of 0.20 (anhydrous substance).

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use. Test solution Dissolve 20.0 mg of the substance to be examined in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (a) Dissolve 20.0 mg of oxytetracycline CRS in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (b) Dissolve 20.0 mg of 4-epioxytetracycline CRS in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (c) Dissolve 20.0 mg of tetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (d) Dissolve 8.0 mg of α-apo-oxytetracycline CRS in 5 mL of 0.01 M sodium hydroxide, using sonication if necessary, and dilute to 100.0 mL with 0.01 M hydrochloric acid.

Reference solution (e) Dissolve 8.0 mg of β-apo-oxytetracycline CRS in 5 mL of 0.01 M sodium hydroxide, using sonication if necessary, and dilute to 100.0 mL with 0.01 M hydrochloric acid.

Reference solution (f) Mix 1.5 mL of reference solution (a), 1.0 mL of reference solution (b), 3.0 mL of reference solution (c), 3.0 mL of reference solution (d) and 3.0 mL of reference solution (e) and dilute to 25.0 mL with 0.01 M hydrochloric acid.

Reference solution (g) Mix 1.0 mL of reference solution (b), 4.0 mL of reference solution (c) and 40.0 mL of reference solution (e) and dilute to 200.0 mL with 0.01 M hydrochloric acid.

acid.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: styrene-divinylbenzene copolymer R (8 μ m); — temperature: 60 °C.

Mobile phase Weigh 30.0 g (for mobile phase A) and 100.0 g (for mobile phase B) of 2-methyl-2-propanol R and transfer separately to 1000 mL volumetric flasks with the aid of 200 mL of water R; to each flask add 60 mL of 0.33 M phosphate buffer solution pH 7.5 R, 50 mL of a 10 g/L solution of tetrabutylammonium hydrogen sulfate R adjusted to pH 7.5 with dilute sodium hydroxide solution R and 10 mL of a 0.4 g/L solution of sodium edate R adjusted to pH 7.5 with dilute sodium hydroxide solution R; dilute each solution to 1000 mL with water R;

Time	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	70	30
15 - 30	30	70
30 - 45	70	30

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of the test solution and reference solutions (f) and (g).

System suitability: reference solution (f): — resolution: minimum 4.0 between the peaks due to impurity A (1st peak) and oxytetracycline (2nd peak), minimum 5.0 between the peaks due to oxytetracycline and impurity B (3rd peak) and minimum 3.5 between the peaks due to impurity D (4th peak) and impurity E (5th peak); if necessary, adapt the ratio mobile phase A: mobile phase B and/or adjust the time programme used to produce the 1-step gradient elution; — symmetry factor: maximum 1.25 for the peak due to oxytetracycline.

Limits: — impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (g) (0.5 per cent); — impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (g) (2.0 per cent); — impurity C (eluting on the tail of the main peak): not more than 4 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (g) (2.0 per cent); — total of impurities D, E and F (eluting between the latter two): not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (g) (2.0 per cent); — disregard limit: 0.02 times the area of the peak due to oxytetracycline in the chromatogram obtained with reference solution (f) (0.1 per cent).

Heavy metals (2.4.8) Maximum 50 ppm.

0.5 g complies with test F. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12) Maximum 2.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14) Maximum 0.5 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 0.4 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

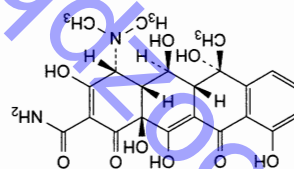
Calculate the percentage content of $C_{22}H_{25}ClN_2O_9$ taking 1 mg of oxytocin as equivalent to 1.079 mg of

oxytocin hydrochloride.

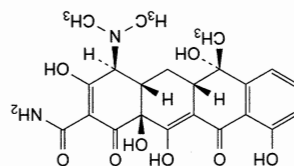
STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

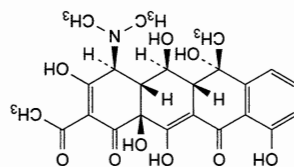
IMPURITIES



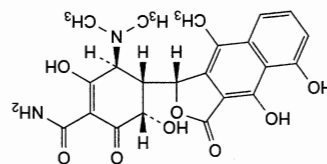
A. (4R,4aR,5S,5aR,6S,12aS)-4-(dimethylamino)-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydroretetracene-2-carboxamide (4-epioxytocycline),



B. (4S,4aR,5S,5aR,6S,12aS)-4-(dimethylamino)-3,5,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydroretetracene-2-carboxamide (tetraacycline),



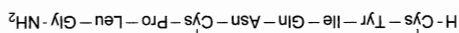
C. (4S,4aR,5S,5aR,6S,12aS)-2-acetyl-4-(dimethylamino)-3,5,6,10,12,12a-hexahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-tetrahydroretetracene-1,11(4H,5H)-dione (2-acetyl-2-decarbarmoyloxytetraacycline),



D. (3S,4S,5S)-4-[(1R)-4,5-dihydroxy-9-methyl-3-oxo-1,3-dihydronaphtho[2,3-c]furan-1-yl]-3-(dimethylamino)-2,5-dihydroxy-6-oxocyclohex-1-enecarboxamide (α-apo-oxytetraacycline),

Oxytocin

(Ph. Eur. monograph 0780)



Action and use

Oxytocic.

Preparations

Ergometrine and Oxytocin Injection
Oxytocin Injection

DEFINITION

Synthetic cyclic nonapeptide having the structure of the hormone produced by the posterior lobe of the pituitary gland that stimulates contraction of the uterus and milk ejection in receptive mammals. It is available in the freeze-dried form as an acetate.
Content
93.0 per cent to 102.0 per cent (anhydrous and acetic acid-free substance).
By convention, for the purpose of labelling oxytocin preparations, 1 mg of oxytocin peptide ($C_{43}H_{66}N_{12}O_{12}S_2$) is equivalent to 600 IU of biological activity.

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Very soluble in water. It dissolves in dilute solutions of acetic acid and of ethanol (96 per cent).

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

B. Amino acid analysis (2.2.56). For hydrolysis use Method 1 and for analysis use Method 1.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids, taking 1/6 of the sum of the number of moles of aspartic acid, glutamic acid, proline, glycine, isoleucine and leucine as equal to 1.

The values fall within the following limits: aspartic acid: 0.90 to 1.10; glutamic acid: 0.90 to 1.10; proline: 0.90 to 1.10; glycine: 0.90 to 1.10; leucine: 0.90 to 1.10; isoleucine: 0.90 to 1.10; tyrosine: 0.7 to 1.05; half-cystine: 1.4 to 2.1. Not more than traces of other amino acids are present.

TESTS

pH (2.2.3)

3.0 to 6.0.

Dissolve 0.200 g in carbon dioxide-free water R and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Prepare a 0.25 mg/mL solution of the substance to be examined in a 15.6 g/L solution of sodium dihydrogen phosphate R.

Resolution solution Dissolve the contents of a vial of oxytocin/desmopressin validation mixture CRS in 1 mL of a 15.6 g/L solution of sodium dihydrogen phosphate R.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase:

— mobile phase A: 15.6 g/L solution of sodium dihydrogen phosphate R;

— mobile phase B: acetonitrile for chromatography R, water R

(50:50 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	70 \rightarrow 40	30 \rightarrow 60
30 - 30.1	40 \rightarrow 70	60 \rightarrow 30
30.1 - 45	70	30

Flow rate 1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 μ L.

Retention time Oxytocin = about 7.5 min;

desmopressin = about 10 min.

System suitability Resolution solution:

— resolution: minimum 5.0 between the peaks due to desmopressin and oxytocin.

Limits:

— any impurity: maximum 1.5 per cent;

— total: maximum 5 per cent;

— disregard limit: 0.1 per cent.

Acetic acid (2.5.34)

6.0 per cent to 10.0 per cent.

Test solution Dissolve 15.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and

Oxytocin Concentrated Solution



Oxytocin Bulk Solution

(Ph. Eur. monograph 0779)

H-Cys-Tyr-Ile-Gln-Asn-Cys-Pro-Leu-Gly-NH₂

C₄₃H₆₆N₁₂O₁₂S₂ 1007

Action and use

Oxytocic.

Preparations

Ergometrine and Oxytocin Injection

Oxytocin Injection

DEFINITION

L-Cysteinyl-L-tyrosyl-L-isoleucyl-L-glutamyl-L-asparaginyl-L-cysteinyl-L-prolyl-L-leucylglycinamide cyclic (1 \rightarrow 6)-disulfide. Solution of oxytocin, a synthetic cyclic nonapeptide having the structure of the hormone produced by the posterior lobe of the pituitary gland and that stimulates contraction of the uterus and milk ejection in receptive mammals. It is available as a solution with a stated concentration of not less than 0.25 mg

of oxytocin per millilitre, in a solvent that may contain an appropriate antimicrobial preservative.

Content

95.0 per cent to 105.0 per cent of the amount of the peptide stated per millilitre.

By convention, for the purpose of labelling oxytocin preparations, 1 mg of oxytocin peptide (C₄₃H₆₆N₁₂O₁₂S₂) is equivalent to 600 IU of biological activity.

CHARACTERS

Appearance

Clear, colourless liquid.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

B. Amino acid analysis (2.2.56). For hydrolysis use Method 1 and for analysis use Method 1.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids taking 1/6 of the sum of the number of moles of aspartic acid, glutamic acid, proline, glycine, isoleucine and leucine as equal to 1. The values fall within the following limits: aspartic acid: 0.90 to 1.10; glutamic acid: 0.90 to 1.10; proline: 0.90 to 1.10; isoleucine: 0.90 to 1.10; leucine: 0.90 to 1.10; tyrosine: 0.7 to 1.05; half cystine: 1.4 to 2.1. Not more than traces of other amino acids are present.

TESTS

pH (2.2.3)

3.0 to 5.0.

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution The preparation to be examined.

Resolution solution Dissolve the contents of a vial of oxytocin/desmopressin validation mixture CRS in 1 mL of 15.6 g/L solution of sodium dihydrogen phosphate R.

Column:

— size: $l = 0.125$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: 15.6 g/L solution of sodium dihydrogen phosphate R;

— mobile phase B: acetonitrile for chromatography R, water R (50:50 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	70 \rightarrow 40	30 \rightarrow 60
30 - 30.1	40 \rightarrow 70	60 \rightarrow 30
30.1 - 45	70	30

Flow rate 1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 μ L.

Retention time Oxytocin = about 7.5 min;

desmopressin = about 10 min.

System suitability Resolution solution:

— resolution: minimum 5.0 between the peaks due to desmopressin and oxytocin.

Limits:

— any impurity: maximum 1.5 per cent;

— total: maximum 5 per cent;

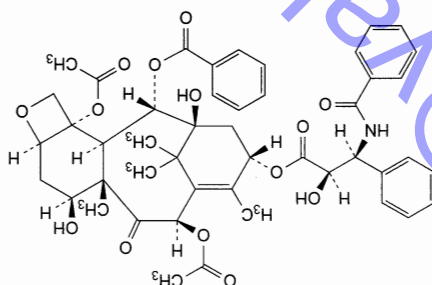
— disregard limit: 0.1 per cent.

Bacterial endotoxins (2.6.14)

Less than 300 IU in the volume that contains 1 mg of oxytocin, if intended for use in the manufacture of parenteral

Pacitaxel

(Ph. Eur. monograph 1794)



Action and use

Taxane cytotoxic.

DEFINITION

5 β ,20-Epoxy-1,7 β -dihydroxy-9-oxotax-11-ene-2 α ,4,10 β ,13 α -tetrayl 4,10-diacetate 2-benzoate 13-[(2R,3S)-3-(benzoylamino)-2-hydroxy-3-phenylpropionate]. It is isolated from natural sources or produced by fermentation or by a semi-synthetic process.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in methanol and freely soluble in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison *pacitaxel* CRS.

LABELLING

The label states the oxytocin peptide content in milligrams of $C_{43}H_{66}N_{12}O_{12}S_2$ per millilitre.

STORAGE

Calculate the content of oxytocin ($C_{43}H_{66}N_{12}O_{12}S_2$) from the declared content of $C_{43}H_{66}N_{12}O_{12}S_2$ in oxytocin CRS.

Injection 25 μ L.

Reference solution Dissolve the contents of a vial of oxytocin CRS in a 15.6 g/L solution of sodium dihydrogen phosphate R to obtain a concentration of 0.25 mg/mL.

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

ASSAY

removal of bacterial endotoxins.
preparations without a further appropriate procedure for the

Ph Eur

33069-62-4

854

$C_{47}H_{51}NO_{14}$



If the spectra obtained in the solid state show differences, dissolve 10 mg of the substance to be examined and the reference substance separately in 0.4 mL of methylene chloride R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.1 g in 10 mL of methanol R.

Specific optical rotation (2.2.7)

–49.0 to –55.0 (anhydrous substance).

Dissolve 0.250 g in methanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

A. Pachitaxel isolated from natural sources or produced by fermentation.

Test solution (a) Dissolve 20.0 mg of the substance to be examined in acetonitrile R1 and dilute to 10.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 20.0 mL with acetonitrile R1.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 10.0 mL with acetonitrile R1.

Reference solution (b) Dissolve 5.0 mg of pachitaxel CRS in acetonitrile R1 and dilute to 5.0 mL with the same solvent.

Dilute 2.0 mL of this solution to 20.0 mL with acetonitrile R1.

Reference solution (c) Dissolve 2.0 mg of pachitaxel impurity C CRS in acetonitrile R1 and dilute to 20.0 mL with the same solvent.

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 50.0 mL with acetonitrile R1.

Reference solution (e) To 1 mL of reference solution (b) add 1 mL of reference solution (c).

Reference solution (f) Dissolve 5 mg of pachitaxel natural for peak identification CRS (containing impurities A, B, C, D, E, F, H, O, P, Q and R) in acetonitrile R1 and dilute to 5 mL with the same solvent.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: diisopropylamopropylsilyl silica gel for chromatography R (5 μ m) with a specific surface area of 180 m²/g and a pore size of 8 nm;

— temperature: 20 ± 1 °C.

Mobile phase:

— mobile phase A: methanol R, water R (200:800 V/V);

— mobile phase B: methanol R, acetonitrile R (200:800 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 60	85 \rightarrow 56	15 \rightarrow 44
60 - 61	56 \rightarrow 85	44 \rightarrow 15
61 - 75	85	15

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 227 nm.

Injection 10 μ L of test solution (a) and reference solutions (a), (d), (e) and (f).

Identification of impurities Use the chromatogram supplied with pachitaxel natural for peak identification CRS and the chromatogram obtained with reference solution (f) to identify the peaks due to impurities A, B, C, D, E, F, H, O, P, Q and R.

Relative retention With reference to pachitaxel (retention time = about 50 min): impurities A and B = about 0.90;

impurity R = about 0.93; impurity H = about 0.96;

impurities Q and P = about 1.02; impurity C = about 1.05;

impurity D = about 1.07; impurities O and E = about 1.15;

impurity F = about 1.20.

System suitability: reference solution (e):

— resolution: minimum 3.5 between the peaks due to pachitaxel and impurity C.

Limits:

— sum of impurities E and O: not more than 5 times the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.5 per cent);

impurity R: not more than 5 times the area of the principal peak in the chromatogram obtained with

reference solution (a) (0.5 per cent);

sum of impurities A and B: not more than 4 times the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.4 per cent);

impurity C: not more than 3 times the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

impurity D: not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

sum of impurities F and Q: not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

impurity F: not more than the area of the principal peak in the chromatogram obtained with reference

solution (d) (0.1 per cent);

unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.10 per cent);

total: not more than 15 times the area of the principal peak in the chromatogram obtained with reference

solution (a) (1.5 per cent);

disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.05 per cent).

B. Pachitaxel produced by a semi-synthetic process.

Test solution Dissolve 10.0 mg of the substance to be examined in acetonitrile R1 and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with acetonitrile R1. Dilute 1.0 mL of this solution to 100.0 mL with acetonitrile R1.

Reference solution (b) Dissolve 5.0 mg of pachitaxel CRS in acetonitrile R1 and dilute to 5.0 mL with the same solvent.

Reference solution (c) Dissolve 5 mg of pachitaxel semi-synthetic for peak identification CRS (containing impurities A, G, I and L) in acetonitrile R1 and dilute to 5 mL with the same solvent.

Reference solution (d) Dissolve the contents of a vial of pachitaxel semi-synthetic for system suitability CRS (containing impurities E, H and N) in 1 mL of acetonitrile R1.

Column:
— size: $l = 0.15$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m) with a specific surface area of 300 m²/g and a pore size of 12 nm;
temperature: 35 °C.

Mobile phase:

— mobile phase A: acetonitrile for chromatography R,
water R (400:600 V/V);

— mobile phase B: acetonitrile for chromatography R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 20	100	0
20 - 60	100 \rightarrow 10	0 \rightarrow 90
60 - 62	10 \rightarrow 100	90 \rightarrow 0
62 - 70	100	0

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 227 nm.

Injection 15 μ L of the test solution and reference solutions (a), (c) and (d).

Identification of impurities Use the chromatogram supplied with *paclitaxel semi-synthetic for peak identification* CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, G, I and L; use the chromatogram supplied with *paclitaxel semi-synthetic for system suitability* CRS and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities E, H and N.

Relative retention With reference to paclitaxel (retention time = about 23 min): impurity N = about 0.2; impurity G = about 0.5; impurity A = about 0.8; impurities M, J and H = about 0.9; impurity E = about 1.3; impurity I = about 1.4; impurity L = about 1.5; impurity K = about 2.2.
System suitability Reference solution (d):
— resolution: minimum 1.5 between the peaks due to impurity H and paclitaxel.

Limits:

— correction factor for the calculation of content, multiply the peak area of impurity N by 1.29;
— impurity A: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.7 per cent);
— impurity L: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— impurities E, I: for each impurity, not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— sum of impurities H, J and M: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 12 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.2 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in methanol R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test B. Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb), obtained by diluting lead standard solution (100 ppm Pb) R with methanol R and 2 mL of the test solution. To 12 mL of each solution, add 2 mL of buffer solution pH 3.5 R. Mix. Add 1.2 mL of thioacetamide reagent R. The substance will precipitate. Dilute to 40 mL with methanol R; the substance re-dissolves completely. Filter the solution through a membrane filter (nominal pore size 0.45 μ m). Compare the spots on the filters obtained with the different solutions. The substance to be examined complies with the test if any brownish-black colour in the spot obtained with the test solution is not more intense than that of the spot obtained with the reference solution.

Water (2.5.32)

Maximum 3.0 per cent, determined on 0.050 g.

Microbial contamination

TAMC: acceptance criterion 10² CFU/g (2.6.12).

Bacterial endotoxins (2.6.14)

Less than 0.4 IU/mg.

ASSAY

A. Paclitaxel isolated from natural sources or produced by fermentation.

Liquid chromatography (2.2.29) as described in test A for related substances with the following modification.
Injection Test solution (b) and reference solution (b).
Calculate the percentage content of C₄₇H₅₁NO₁₄ from the declared content of *paclitaxel* CRS.

B. Paclitaxel produced by a semi-synthetic process.
Liquid chromatography (2.2.29) as described in test B for related substances with the following modification.
Injection 10 μ L of the test solution and reference solution (b).
Calculate the percentage content of C₄₇H₅₁NO₁₄ from the declared content of *paclitaxel* CRS.

STORAGE

In an airtight container, protected from light.

LABELLING

The label states the origin of the substance:

— isolated from natural sources;
— produced by fermentation;

— produced by a semi-synthetic process.

IMPURITIES

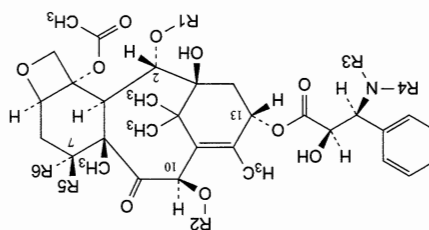
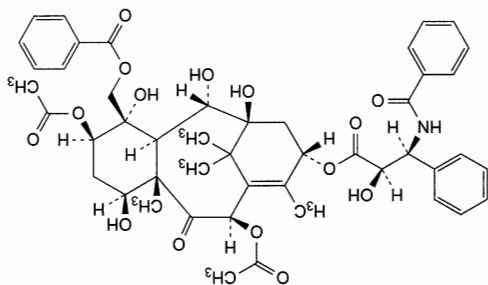
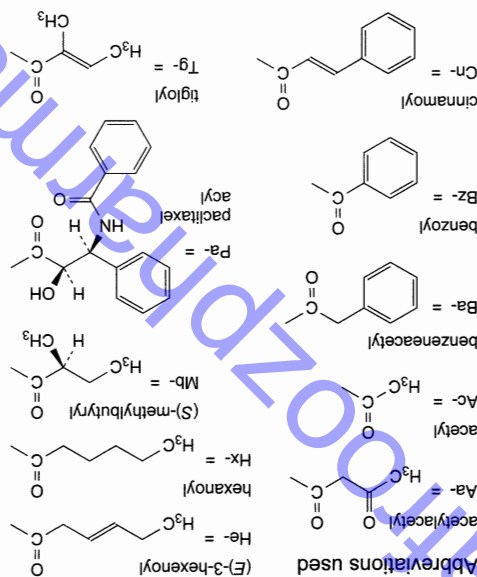
Test A

Specified impurities A, B, C, D, E, F, O, P, Q, R

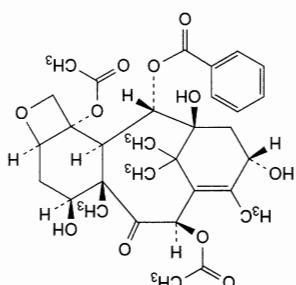
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): H.

Test B

Specified impurities A, E, G, H, I, J, K, L, M, N.

**Abbreviations used**

M. 1,2 α ,4,7 β -dihydroxy-9-oxotax-11-ene-5 β ,10 β ,13 α ,20-tetrayl 5,10-diacetate 20-benzoate 13-[(2R,3S)-3-(benzoylamino)-2-hydroxy-3-phenylpropionate]



N. 13-O-de[(2R,3S)-3-(benzoylamino)-2-hydroxy-3-phenylpropionyl]pacitaxel (bacactin III).

Palmitic Acid

(Ph. Eur. monograph 1904)

57-10-3

**Action and use**

Excipient.

DEFINITION

Hexadecanoic acid ($\text{C}_{16}\text{H}_{32}\text{O}_2$; M_r 256.4), obtained from fats or oils of vegetable or animal origin.

Content

Minimum 92.0 per cent.

CHARACTERS**Appearance**

White or almost white, waxy solid.

Solubility

Practically insoluble in water, soluble in ethanol

IDENTIFICATION

A. Freezing point (see Tests).

B. Acid value (2.5.1): 216 to 220, determined on 0.1 g.

C. Examine the chromatograms obtained in the assay.

A. R1 = Tg, R2 = Ac, R3 = Bz, R4 = R6 = H, R5 = OH;
 2-O-debenzoyl-2-O-tigloylpacitaxel,
 B. R1 = Bz, R2 = Ac, R3 = Tg, R4 = R6 = H, R5 = OH;
 N-debenzoyl-N-tigloylpacitaxel (cephalomannine),
 C. R1 = Bz, R2 = Ac, R3 = Hx, R4 = R6 = H, R5 = OH;
 N-debenzoyl-N-hexanoylpacitaxel (pacitaxel C),
 D. R1 = Bz, R2 = Ac, R3 = Tg, R4 = R5 = H, R6 = OH;
 N-debenzoyl-N-tigloyl-7-epi-pacitaxel,
 (7-epi-cephalomannine),
 E. R1 = R3 = Bz, R2 = Ac, R4 = R5 = H, R6 = OH;
 7-epi-pacitaxel,
 F. R1 = Bz, R2 = Ac, R3 = Hx, R4 = CH₃, R5 = OH,
 R6 = H: N-debenzoyl-N-hexanoyl-N-methylpacitaxel
 (N-methylpacitaxel C),
 G. R1 = R3 = Bz, R2 = R4 = R6 = H, R5 = OH;
 10-O-deacetyl-10-O-tigloylpacitaxel,
 H. R1 = R3 = Bz, R2 = R4 = R5 = H, R6 = OH;
 10-O-deacetyl-7-epi-pacitaxel,
 I. R1 = R3 = Bz, R2 = Pa, R4 = R6 = H, R5 = OH;
 10-O-[(2R,3S)-3-(benzoylamino)-2-hydroxy-3-phenylpropionyl]-10-O-deacetyl-10-O-tigloylpacitaxel,
 J. R1 = R3 = Bz, R2 = Aa, R4 = R6 = H, R5 = OH;
 10-O-deacetyl-10-O-(3-oxobutanoyl)pacitaxel,
 K. R1 = R3 = Bz, R2 = Ac, R4 = R6 = H, R5 = OH;
 O-Si(C₂H₅)₃: 7-O-(triethylsilyl)pacitaxel,
 L. R1 = R3 = Bz, R2 = Ac, R4 = R6 = H, R5 = OH;
 O-CO-CH₃: 7-O-acetyl-10-O-tigloylpacitaxel,
 O. R1 = Bz, R2 = Ac, R3 = Cn, R4 = R6 = H, R5 = OH;
 N-cinnamoyl-N-debenzoylpacitaxel,

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with the reference solution.

TESTS

Appearance

Heat the substance to be examined to about 75 °C. The resulting liquid is not more intensely coloured than reference solution Y₇ or BY₇ (2.2.2, Method I).

Acidity

Melt 5.0 g, stir for 2 min in 10 mL of hot carbon dioxide-free water R, cool slowly and filter. To the filtrate add 0.05 mL of methyl orange solution R. No red colour develops.

Freezing point (2.2.18)

60 °C to 66 °C.

Iodine value (2.5.4)

Maximum 1.

Stearic acid

Maximum 6.0 per cent, determined as prescribed in the assay.

Nickel (2.4.31)

Maximum 1 ppm.

ASSAY

Gas chromatography (2.4.22, Method C). Prepare the solutions as described in the method but omitting the initial hydrolysis.

Reference solution Prepare the reference solution in the same manner as the test solution using a mixture of 50 mg of palm stearic acid R and 50 mg of stearic acid R instead of the substance to be examined.

Relative retention With reference to methyl stearate: methyl palmitate = about 0.9.

System suitability:

— **Resolution:** minimum 5.0 between the peaks due to methyl stearate and methyl palmitate.

Fractionated Palm Kernel Oil

The standards of this monograph encompass several different suppository bases. The selection of a particular suppository formulation should be appropriate to the product concerned and it may be necessary to apply more restrictive standards for a particular application.

Action and use

Suppository basis.

DEFINITION

Fractionated Palm Kernel Oil is obtained by expression of the natural oil from the kernels of *Elaeis guineensis* Jacq. followed by selective solvent fractionation and hydrogenation.

CHARACTERISTICS

A white, solid, brittle fat; odourless or almost odourless. Practically insoluble in water; miscible with ether and with petroleum spirit (boiling range, 40° to 60°); practically insoluble in ethanol (96%).

TESTS

Acid value

Not more than 0.2, Appendix X B.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
It is sparingly soluble in dilute mineral acids and dissolves in dilute alkaline solutions.

Solubility

White or almost white, crystalline powder.

Appearance

CHARACTERS

98.0 per cent to 101.0 per cent (anhydrous substance).

Content

Disodium dihydrogen (3-amino-1-hydroxypropylidene)bisphosphonate pentahydrate.

DEFINITION

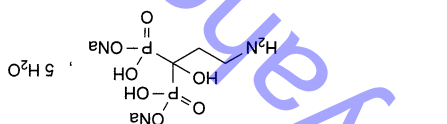
Ph Eur

Preparation

Pamidronate Disodium Infusion

Action and use

Bisphosphonate; treatment of osteolytic lesions; Paget's disease; hypercalcaemia of malignancy.



(Ph. Eur. monograph 1779)
Disodium Pamidronate

Pentahydrate

Pamidronate Disodium



Dissolve 5 g in 15 mL of chloroform, add 20 mL of glacial acetic acid and 0.5 mL of a saturated solution of potassium iodide, mix well, allow to stand in the dark for exactly 1 minute, add 30 mL of water and titrate with 0.01M sodium thiosulfate VS using starch mucilage as indicator. Not more than 0.5 mL of 0.01M sodium thiosulfate VS is required.

Peroxides

246 to 250, Appendix X G.

Saponification value

At 50°, 1.445 to 1.447, Appendix V E.

Refractive index

before carrying out the test.
22° and allow to stand at this temperature for 24 hours transfer to a vessel previously kept at a temperature of 15° to 22° and allow to stand at this temperature for 24 hours of cloudiness appear and continue to stir by hand until the substance has the consistency of a paste. Immediately stir continuously with a mechanical stirrer until the first signs stirring until the temperature falls to between 32° and 34°, temperature between 53° and 60°. Cool with occasional filter through a suitable dry filter paper, maintaining the about 30 g in an oven at a temperature of 55° to 60° and substance being examined in the following manner. Melt 31° to 36°, Appendix V A, Method IV. Prepare the

Melting point

Not more than 6.0 (iodine bromide method), Appendix X E.

Iodine value

Comparison pantothenate disodium pentahydrate CRS.
 B. Dissolve 0.5 g in 10 mL of water R. The solution gives reaction (a) of sodium (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).
 Dissolve 0.20 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

pH (2.2.3)

7.8 to 8.8.

Dissolve 0.100 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Impurity A

Thin-layer chromatography (2.2.27).

Test solution Dissolve 30 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 15 mg of 3-aminopropionic acid R in water R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with water R.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, di-isopropyl ether R, methanol R (4:8:9 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Spray with a ninhydrin solution R. Heat at 100-105 °C for 15 min.

Limit:

— impurity A: any spot due to impurity A is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

Impurities B and C

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.

Reference solution To 2.0 mL of a 0.3 g/L solution of phosphoric acid R add 2.0 mL of a 0.25 g/L solution of phosphorous acid R and dilute to 50.0 mL with water R.

Column:

— size: l = 0.10 m, Ø = 4.6 mm,

— stationary phase: anion-exchange resin R (5 µm),

— temperature: 35 °C.

Mobile phase To 0.5 mL of anhydrous formic acid R add 2500 mL of water R; adjust to pH 3.5 with an 80 g/L solution of sodium hydroxide R.

Flow rate 1.0 mL/min.

Detection Refractometer.

Injection 100 µL.

Relative retention With reference to pantothenate (retention time = about 13 min): impurity B = about 1.3; impurity C = about 1.6.

System suitability: reference solution:

— resolution: minimum 2.5 between the peaks due to impurities B and C.

Limit:

— impurities B, C: for each impurity, not more than the area of the corresponding peaks in the chromatogram obtained with the reference solution (0.5 per cent).

Pancreatic Extract

(Pancreas Powder, Ph Eur monograph 0350)



DEFINITION

Pancreas powder is prepared from the fresh or frozen pancreases of mammals. It contains various enzymes having proteolytic, lipolytic and amylolytic activities.

1 mg of pancreas powder contains not less than 1.0 Ph. Eur. U. of total proteolytic activity, 15 Ph. Eur. U. of lipolytic activity and 12 Ph. Eur. U. of amylolytic activity.

PRODUCTION

The animals from which pancreas powder is derived must fulfil the requirements for the health of animals suitable for human consumption.

CHARACTERS

Appearance

Slightly brown, amorphous powder.

Solubility

Partly soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Titrate 0.5 g with 10 mL of water R and adjust to pH 8 with 0.1 M sodium hydroxide, using 0.1 mL of cresol red solution R as indicator. Divide the suspension into 2 equal parts (suspension (a) and suspension (b)). Boil

suspension (a), heat to 38-40 °C and maintain at this temperature for 1 h. Suspension (a) is colourless or slightly pink and suspension (b) is distinctly more red.

B. Titrate 0.25 g with 10 mL of water R and adjust to pH 8 with 0.1 M sodium hydroxide, using 0.1 mL of cresol red solution R as indicator. Divide the suspension into 2 equal parts (suspension (a) and suspension (b)). Boil

suspension (a), heat to 38-40 °C and maintain at this temperature for 1 h. Suspension (a) is colourless or slightly pink and suspension (b) is distinctly more red.

Heavy metals (2.4.8)

Maximum 20 ppm.

2.0 g complies with test C. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Water (2.5.12)

23.0 per cent to 27.0 per cent, determined on 0.100 g.

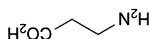
ASSAY

Dissolve 0.250 g in 70 mL of water R. Titrate with 0.1 M hydrochloric acid determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M hydrochloric acid is equivalent to 27.91 mg of C₃H₉NNaO₇P₂.

IMPURITIES

Specified impurities A, B, C



A. 3-aminopropionic acid (β-alanine),

B. H₃PO₄: phosphoric acid,

C. H₃PO₃: phosphorous acid.

Designate tubes in duplicate T₁, T₂, S_{1b}, S_{2b}, S₃, S_{3b}, designate a tube B.

Add borate buffer solution pH 7.5 R to the tubes as follows:

B: 3.0 mL,

S₁ and S_{1b}: 2.0 mL,

S₂, S_{2b}, T and T_p: 1.0 mL.

Add the reference suspension to the tubes as follows:

S₁ and S_{1b}: 1.0 mL,

S₂ and S_{2b}: 2.0 mL,

S₃ and S_{3b}: 3.0 mL.

Add 2.0 mL of the test suspension to tubes T and T_p.

Add 5.0 mL of a 50 g/L solution of trichloroacetic acid R to

tubes B, S_{1b}, S_{2b}, S_{3b} and T_p. Mix by shaking.

Place the tubes and the casein solution in a water-bath at

35 ± 0.5 °C. Place a glass rod in each tube. When

temperature equilibrium is reached, add 2.0 mL of the casein

solution to tubes B, S_{1b}, S_{2b}, S_{3b} and T_p. Mix. At time zero,

add 2.0 mL of casein solution successively and at intervals of

30 s to tubes S₁, S₂, S₃ and T. Mix immediately after each

addition. Exactly 30 min after addition of the casein solution,

taking into account the regular interval adopted, add 5.0 mL

of a 50 g/L solution of trichloroacetic acid R to tubes S₁, S₂

S₃ and T. Mix. Withdraw the tubes from the water-bath and

allow to stand at room temperature for 20 min.

Filter the contents of each tube twice through the same

suitable filter paper previously washed with a 50 g/L solution

of trichloroacetic acid R, then with water R and dried.

A suitable filter paper complies with the following test: filter

5 mL of a 50 g/L solution of trichloroacetic acid R on a 7 cm

disc of white filter paper; the absorbance (2.2.25) of the

filtrate, measured at 275 nm using unfiltered trichloroacetic

acid solution as the compensation liquid, is less than 0.04.

A schematic presentation of the above operations is shown in

Table 0350.-1.

Table 0350.-1

Tubes		Buffer solution		Reference suspension		Test suspension		Trichloroacetic acid solution		Water-bath 35 °C		Casein solution		Mix		Water-bath 35 °C 30 min		Trichloroacetic acid solution		Mix		Room temperature		Filter	
S ₁	S _{1b}	S ₂	S _{2b}	S ₃	S _{3b}	T	T _p	B																	
2	2	1	1	2	2	3	3	2	5	5	5	2	2	2	2	2	2	5	5	5	5	5	5	5	5
+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

Measure the absorbance (2.2.25) of the filtrates at 275 nm using the filtrate obtained from tube B as the compensation liquid.

suspension (a) and to the remaining 75 mL add suspension (b). Heat each mixture to 38-40 °C and maintain at this temperature for 5 min.

To 1 mL of each mixture add 10 mL of iodine solution R₂. The mixture obtained with suspension (a) has an intense blue-violet colour; the mixture obtained with suspension (b) has the colour of the iodine solution.

TESTS

Fat content

Maximum 5.0 per cent.

In an extraction apparatus, treat 1.0 g with light petroleum R₁

for 3 h. Evaporate the solvent and dry the residue at

100-105 °C for 2 h. The residue weighs a maximum of

50 mg.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 0.50 g by drying at

60 °C at a pressure not exceeding 670 Pa for 4 h.

Microbial contamination

TAMC: acceptance criterion 10⁴ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

ASSAY

Total proteolytic activity

The total proteolytic activity of pancreas powder is

determined by comparing the quantity of peptides non-

precipitable by a 50 g/L solution of trichloroacetic acid R

released per minute from a substrate of casein solution with

the quantity of such peptides released by pancreas powder

(protease) BRP from the same substrate in the same

conditions.

Casein solution Suspend a quantity of casein BRP equivalent

to 1.25 g of dried substance in 5 mL of water R, add 10 mL

of 0.1 M sodium hydroxide and stir for 1 min. (Determine the

water content of casein BRP prior to the test by heating at

60 °C in vacuo for 4 h.) Add 60 mL of water R and stir with

a magnetic stirrer until the solution is practically clear. Adjust

to pH 8.0 with 0.1 M sodium hydroxide or 0.1 M hydrochloric

acid. Dilute to 100.0 mL with water R. Use the solution on

the day of preparation.

Enterokinase solution Dissolve 50 mg of enterokinase BRP in

0.02 M calcium chloride solution R and dilute to 50.0 mL with

the same solvent. Use the solution on the day of preparation.

To avoid absorption of water formed by condensation, allow the

preparation to be examined and the reference preparation to reach

room temperature before opening the containers.

For the test suspension and the reference suspension, prepare the

suspension and carry out the dilution at 0-4 °C.

Test suspension Triturate 0.100 g of the substance to be

examined for 5 min adding gradually 25 mL of 0.02 M

calcium chloride solution R. Transfer completely to a

volumetric flask and dilute to 100.0 mL with 0.02 M calcium

chloride solution R. To 10.0 mL of this suspension add

10.0 mL of the enterokinase solution and heat on a water-

bath at 35 ± 0.5 °C for 15 min. Cool and dilute with borate

buffer solution pH 7.5 R at 5 ± 3 °C to a final concentration

of about 0.065 Ph. Eur. U. of total proteolytic activity per

millilitre calculated on the basis of the stated activity.

Reference suspension Prepare a suspension of pancreas powder

(protease) BRP as described for the test suspension but

without the addition of enterokinase so as to obtain a known

final concentration of about 0.065 Ph. Eur. U. per millilitre

calculated on the basis of the stated activity.

a very fine suspension is obtained. Dilute the suspension with *maleate buffer solution pH 7.0 R*, transfer quantitatively to a volumetric flask and dilute to 100.0 mL with the buffer solution. Keep the flask containing the test suspension in iced water during the titration.

Reference suspension Prepare a suspension of *pancreas powder (lipase) BRP* as described for the test suspension using a quantity equivalent to about 2500 Ph. Eur. U.

Carry out the titrations immediately after preparation of the test suspension and the reference suspension. Place 29.5 mL of olive oil emulsion in the reaction vessel equilibrated at $37 \pm 0.5^\circ\text{C}$. Fit the vessel with the electrodes, a stirrer and the burette (the tip being immersed in the olive oil emulsion).

Put the lid in place and switch on the apparatus. Carefully add 0.1 M sodium hydroxide with stirring to adjust to pH 9.2. Using a rapid-flow graduated pipette transfer about 0.5 mL of the previously homogenised reference suspension, start the chronometer and add continuously 0.1 M sodium hydroxide to maintain the pH at 9.0. After exactly 1 min, note the volume of 0.1 M sodium hydroxide used. Carry out the measurement a further 4 times. Discard the first reading and determine the average of the 4 others (S_1). Make 2 further determinations (S_2 and S_3). Calculate the average of the values S_1 , S_2 and S_3 . The average volume of 0.1 M sodium hydroxide used should be about 0.12 mL per minute with limits of 0.08 mL to 0.16 mL.

Carry out 3 determinations in the same manner for the test suspension (T_1 , T_2 and T_3). If the quantity of 0.1 M sodium hydroxide used is outside the limits of 0.08 mL to 0.16 mL per minute, the assay is repeated with a quantity of test suspension that is more suitable but situated between 0.4 mL and 0.6 mL. Otherwise the quantity of the substance to be examined is adjusted to comply with the conditions of the test. Calculate the average of the values T_1 , T_2 and T_3 . Calculate the activity in European Pharmacopoeia Units per milligram using the following expression:

$$\frac{n \times m_1}{m_1 \times m} \times A$$

n = average volume of 0.1 M sodium hydroxide used per minute during the titration of the test suspension, in millilitres;
 n_1 = average volume of 0.1 M sodium hydroxide used per minute during the titration of the reference suspension, in millilitres;
 m = mass of the substance to be examined, in milligrams;
 m_1 = mass of the reference preparation, in milligrams;
 A = activity of *pancreas powder (lipase) BRP*, in European Pharmacopoeia Units per milligram.

Amylolytic activity

The amylolytic activity is determined by comparing the rate at which a suspension of pancreas powder hydrolyses a substrate of starch solution with the rate at which a suspension of *pancreas powder (amylase) BRP* hydrolyses the same substrate under the same conditions.

Starch solution To a quantity of starch *BRP* equivalent to 2.0 g of the dried substance add 10 mL of water *R* and mix. (Determine the water content of starch *BRP* prior to the test by heating at 120°C for 4 h). Add this suspension, whilst stirring continuously, to 160 mL of boiling water *R*. Wash the container several times with successive quantities, each of 10 mL, of water *R* and add the washings to the hot starch

The test is not valid unless the corrected absorbance values are between 0.15 and 0.60.

Lipolytic activity

The lipolytic activity is determined by comparing the rate at which a suspension of pancreas powder hydrolyses a substrate of olive oil emulsion with the rate at which a suspension of *pancreas powder (lipase) BRP* hydrolyses the same substrate under the same conditions. The test is carried out under nitrogen.

Olive oil stock emulsion In an 800 mL beaker 9 cm in diameter, place 40 mL of olive oil *R*, 330 mL of *acacia solution R* and 30 mL of water *R*. Place an electric mixer at the bottom of the beaker. Place the beaker in a vessel containing ethanol (96 per cent) *R* and a sufficient quantity of ice as a cooling mixture. Emulsify using the mixer at an average speed of 1000-2000 r/min. Cool to $5-10^\circ\text{C}$. Increase the mixing speed to 8000 r/min. Mix for 30 min keeping the temperature below 25°C by the continuous addition of crushed ice into the cooling mixture. (A mixture of calcium chloride and crushed ice is also suitable). Store the stock emulsion in a refrigerator and use within 14 days.

The emulsion must not separate into 2 distinct layers. Check the diameter of the globules of the emulsion under a microscope. At least 90 per cent have a diameter below 3 µm and none has a diameter greater than 10 µm. Shake the emulsion thoroughly before preparing the emulsion substrate. For 10 determinations, mix the following solutions in the order indicated: 100 mL of the stock emulsion, 80 mL of *tris(hydroxymethyl)aminomethane solution R1*, 20 mL of a freshly prepared 80 g/L of sodium taurocholate *BRP* and 95 mL of water *R*. Use on the day of preparation.

Apparatus Use a reaction vessel of about 50 mL capacity provided with:

- a device that will maintain a temperature of $37 \pm 0.5^\circ\text{C}$;
- a magnetic stirrer;
- a lid with holes for the insertion of electrodes, the tip of a burette, a tube for the admission of nitrogen and the introduction of reagents.

An automatic or manual titration apparatus may be used. In the latter case, the burette is graduated in 0.005 mL and the pH-meter is provided with a wide reading scale and glass-calomel or glass-silver-silver chloride electrodes. After each test the reaction vessel is evacuated by suction and washed several times with water *R*, the washings being removed each time by suction.

To avoid absorption of water formed by condensation, allow the preparation to be examined and the reference preparation to reach room temperature before opening the containers.

For the test suspension and the reference suspension, prepare the *Test suspension* In a small mortar cooled to $0-4^\circ\text{C}$, triturate carefully a quantity of the substance to be examined equivalent to about 2500 Ph. Eur. U. of lipolytic activity with 1 mL of *maleate buffer solution pH 7.0 R* (lipase solvent) until

solution. Heat to boiling, stirring continuously. Cool to room temperature and dilute to 200 mL with water R. Use the solution on the day of preparation.

To avoid absorption of water formed by condensation, allow the preparation to be examined and the reference preparation to reach room temperature before opening the containers.

For the test suspension and the reference suspension, prepare the suspension and carry out the dilution at 0–4 °C.

Test suspension Triturate a quantity of the substance to be examined equivalent to about 1500 Ph. Eur. U. of amylolytic activity with 60 mL of phosphate buffer solution pH 6.8 RI for 15 min. Transfer quantitatively to a volumetric flask and dilute to 100.0 mL with phosphate buffer solution pH 6.8 RI. Reference suspension Prepare a suspension of pancreas powder (amylase) BRP as described for the test suspension, using a quantity equivalent to about 1500 Ph. Eur. U.

In a test-tube 200 mm long and 22 mm in diameter, fitted with a ground-glass stopper, place 25.0 mL of starch solution, 10.0 mL of phosphate buffer solution pH 6.8 RI and 1.0 mL of an 11.7 g/L solution of sodium chloride R. Close the tube, shake and place in a water-bath at 25.0 ± 0.1 °C. When the temperature equilibrium has been reached, add 1.0 mL of the test suspension and start the chronometer.

Mix and place the tube in the water-bath. After exactly 10 min, add 2 mL of 1 M hydrochloric acid. Transfer the mixture quantitatively to a 300 mL conical flask fitted with a ground-glass stopper. Whilst shaking continuously, add 10.0 mL of 0.05 M iodine immediately followed by 45 mL of 0.1 M sodium hydroxide. Allow to stand in the dark at a temperature between 15 °C and 25 °C for 15 min.

4 volumes of water R. Titrate the excess of iodine with 0.1 M sodium thiosulfate using a microburette. Carry out a blank titration adding the 2 mL of 1 M hydrochloric acid before introducing the test suspension. Carry out the titration of the reference suspension in the same manner.

The test is not valid unless both n' -n and n' - n_1 are between 1.9 mL and 3.6 mL.

Calculate the amylolytic activity in European Pharmacopoeia Units per milligram using the following expression:

$$\frac{(n' - n) m_1}{(n'_1 - n_1) m} \times A$$

n = volume of 0.1 M sodium thiosulfate used in the titration of the test suspension, in millilitres;
n₁ = volume of 0.1 M sodium thiosulfate used in the titration of the reference suspension, in millilitres;
n' = volume of 0.1 M sodium thiosulfate used in the blank titration of the test suspension, in millilitres;
n'₁ = volume of 0.1 M sodium thiosulfate used in the blank titration of the reference suspension, in millilitres;
m = mass of the substance to be examined, in milligrams;
m₁ = mass of the reference preparation, in milligrams;
A = activity of pancreas powder (amylase) BRP, in European Pharmacopoeia Units per milligram.

STORAGE

In an airtight container.

Ph Eur

Pancreatin

Action and use

Enzyme; treatment of pancreatic exocrine deficiency.

Preparations

Pancreatin Granules

Gastro-resistant Pancreatin Tablets

DEFINITION

Pancreatin is a preparation of mammalian pancreas containing enzymes having protease, lipase and amylase activity. It may contain Sodium Chloride.

Potency

Pancreatin contains in 1 mg not less than 1.4 Units of free protease activity, not less than 20 Units of lipase activity and not less than 24 Units of amylase activity.

PRODUCTION

Pancreatin is prepared in conditions designed to minimise the degree of microbial contamination.

CHARACTERISTICS

A white or buff amorphous powder.

Soluble or partly soluble in water forming a slightly turbid solution; practically insoluble in ethanol (96%) and in ether.

IDENTIFICATION

A. Triturate 0.5 g with 10 mL of water and adjust to pH 8.0 by the addition of 1 M sodium hydroxide using cresol red solution as indicator. Divide the resulting solution into two equal portions. Boil one portion [solution (1)] and leave the other untreated [solution (2)]. Dissolve 0.1 g of soluble starch in 100 mL of boiling water, boil for 2 minutes, cool and dilute to 150 mL with water. Add solution (1) to half the starch mucilage and solution (2) to the remainder and maintain the mixtures at 38° to 40° for 5 minutes. To 1 mL of each mixture add 10 mL of iodinated potassium iodide solution. The liquid containing solution (2) retains the colour of the solution of iodine and the liquid containing solution (1) acquires an intense blue colour.

B. Triturate 0.25 g with 10 mL of water and adjust to pH 8.0 by the addition of 1 M sodium hydroxide using cresol red solution as indicator. Divide the resulting solution into two equal portions. Boil one portion [solution (1)] and leave the other untreated [solution (2)]. Dissolve 0.1 g of soluble starch in 100 mL of boiling water, boil for 2 minutes, cool and dilute to 150 mL with water. Add solution (1) to half the starch mucilage and solution (2) to the remainder and maintain the mixtures at 38° to 40° for 5 minutes. To 1 mL of each mixture add 10 mL of iodinated potassium iodide solution. The liquid containing solution (2) retains the colour of the solution of iodine and the liquid containing solution (1) acquires an intense blue colour.

TESTS

Fat

Extract 1 g with petroleum spirit (boiling range, 40° to 60°) for 3 hours in an apparatus for the continuous extraction of drugs, Appendix XI F, evaporate the extract and dry the residue at 105° for 2 hours. The residue weighs not more than 30 mg.

Loss on drying

When dried at 60° at a pressure not exceeding 0.7 kPa for 4 hours, loses not more than 5.0% of its weight. Use 0.5 g.

Microbial contamination

1 g is free from *Escherichia coli*; 10 g is free from *Salmonella*. Appendix XVI B1.

ASSAY

Carry out the Assay of pancreatin, Appendix XIV I.

STORAGE

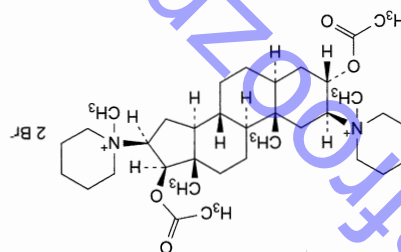
Pancreatin should be stored at a temperature not exceeding 15°.

LABELLING

The label states (1) the minimum number of Units of activity of free protease, lipase and amylase per mg; (2) the name of any added substances; (3) the date after which the material is not intended to be used; (4) the conditions under which it should be stored.

Pancuronium Bromide

(Ph. Eur. monograph 0681)



15500-66-0

Action and use

Non-depolarizing neuromuscular blocker.

Preparation

Pancuronium Injection

Ph. Eur.

DEFINITION

1,1'-[3,7-bis(1-methylpiperidinium)-5,5'-diacetyloxy]-2,2'-bis(1-methylpiperidinium) dibromide.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White, yellowish-white or slightly pink, crystalline powder, hygroscopic.

Solubility

Very soluble or freely soluble in water, very soluble in methylene chloride, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison *pancuronium bromide* CRS.

B. It gives reaction (a) of bromides (2.3.1).

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 50 mg in water R and dilute to 25 mL with the same solvent.

Specific optical rotation (2.2.7)

+38.0 to +42.0 (anhydrous substance).

Dissolve 0.75 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Thin-layer chromatography (2.2.27). Prepare the solutions immediately before use.

Test solution Dissolve 50.0 mg of the substance to be examined in methylene chloride R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with methylene chloride R. Dilute 1.0 mL of this solution to 20.0 mL with methylene chloride R.

Reference solution (b) Dissolve 10.0 mg of *pancuronium bromide* for system suitability CRS (containing 1.0 per cent of impurity D) in 1.0 mL of methylene chloride R.

Plate TLC silica gel plate R (2-10 µm).

Mobile phase 400 g/L solution of sodium iodide R, acetonitrile R, 2-propanol R (5:10:85 V/V/V).

Application 5 µL.

Development In an unlined and unsaturated tank over a path of 8 cm.

Drying In a current of air at room temperature.

Detection Spray with a 20 g/L solution of sodium nitrite R and allow to dry for 5 min. Then spray with potassium iodobismuthate solution R5. Cover the plate with a transparent glass cover.

System suitability:

— the chromatogram obtained with reference solution (b) shows 2 clearly separated spots due to *pancuronium bromide* (R_F = about 0.5) and impurity D (R_F = about 0.6);

— the chromatogram obtained with reference solution (a) shows a clearly visible spot.

Note Impurity A if present will co-migrate with impurity D. Limits:

— impurities A, D: any spot due to impurities A and/or D is not more intense than the spot due to impurity D in the chromatogram obtained with reference solution (b)

(1.0 per cent),

unspecified impurities: any other spot is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.10 per cent).

Water (2.5.12)

Maximum 8.0 per cent, determined on 0.300 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of acetic anhydride R, heating if necessary. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 36.63 mg of $C_{35}H_{60}Br_2N_2O_4$.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A, D

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use): B, C, E.

PRODUCTION

It is produced by methods of manufacture designed to guarantee the proper hydrate form and it complies, if tested, with a suitable test that demonstrates its sesquihydrate nature (for example near-infrared spectroscopy (2.2.40) or X-ray powder diffraction (2.9.33)).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), practically insoluble in hexane.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison pantoprazole sodium sesquihydrate CRS.

B. It gives reaction (a) of sodium (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Dissolve 0.20 g in water R and dilute to 20.0 mL with the same solvent.

Optical rotation (2.2.7)

−0.4° to +0.4°.

Dissolve 0.2 g in 10 mL of water R. Adjust to pH 11.5–12.0 with an 8 g/L solution of sodium hydroxide R. Dilute to 20.0 mL with water R.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile for chromatography R, 40 mg/L

solution of sodium hydroxide R (50:50 V/V).

Test solution Dissolve 23 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of pantoprazole for system suitability CRS (containing impurities A, B, C, D and E) in 1.0 mL of the solvent mixture.

Column:

size: $l = 0.125$ m, $\phi = 4$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m);

temperature: 40 °C.

Mobile phase:

mobile phase A: 1.74 g/L solution of dipotassium hydrogen phosphate R adjusted to pH 7.00 \pm 0.05 with a 330 g/L solution of phosphoric acid R;

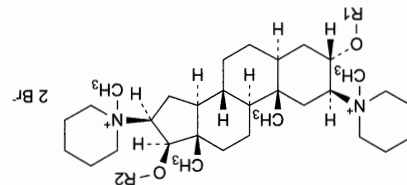
mobile phase B: acetonitrile for chromatography R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 40	80 \rightarrow 20	20 \rightarrow 80
40 - 45	20 \rightarrow 80	80 \rightarrow 20

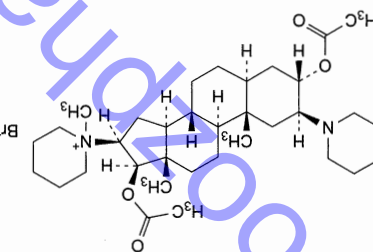
Flow rate 1.0 mL/min.

Detection Spectrophotometer at 290 nm and, for impurity C,

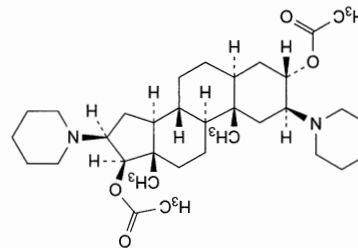
Injection 20 μ L.



A. R1 = CO-CH₃, R2 = H; 1,1'-[3 α -(acetyloxy)-17 β -hydroxy-5 α -androstan-2 β ,16 β -diyl]bis(1-methylpiperidinium) dibromide (dacuronium bromide),
B. R1 = CO-CH₃, R2 = H, R1'-[17 β -(acetyloxy)-3 α -hydroxy-5 α -androstan-2 β ,16 β -diyl]bis(1-methylpiperidinium) dibromide,
C. R1 = R2 = H; 1,1'-[3 α ,17 β -dihydroxy-5 α -androstan-2 β ,16 β -diyl]bis(1-methylpiperidinium) dibromide,



D. 1-[3 α ,17 β -bis(acetyloxy)-2 β -(piperidin-1-yl)-5 α -androstan-16 β -yl]-1-methylpiperidinium bromide (vecuronium bromide),



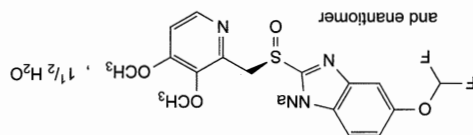
E. 2 β ,16 β -bis(piperidin-1-yl)-5 α -androstan-3 α ,17 β -diyl diacetate.

Ph Eur



Pantoprazole Sodium Sesquihydrate

(Ph. Eur. monograph 2296)



C₁₆H₁₄F₂N₃NaO₄S, 1/2 H₂O 432.4 164579-32-2

Action and use
Proton pump inhibitor; treatment of peptic ulcer disease.

Ph Eur

DEFINITION

Sodium 5-(difluoromethoxy)-2-[(R)-[(3,4-dimethoxyphenyl)methyl]sulfinyl]benzimidazol-1-ide sesquihydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

Identification of impurities Use the chromatogram supplied with pantoprazole for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, D + F and E.

Relative retention With reference to pantoprazole (retention time = about 1.1 min): impurity C = about 0.6; impurity A = about 0.9; impurities D and F = about 1.2; impurity E = about 1.3; impurity B = about 1.5.

System suitability: reference solution (b):

- **resolution:** minimum 1.5 between the peaks due to impurities E and D + F;
- the chromatogram obtained is similar to the chromatogram supplied with pantoprazole for system suitability CRS.

Limits:

- **correction factor:** for the calculation of content, multiply the peak area of impurity C by 0.3;
- **impurity A:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- **sum of impurities D and F:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
- **impurities B, C, E:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);
- **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- **total:** not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

5.9 per cent to 6.9 per cent, determined on 0.150 g.

ASSAY

Dissolve 0.200 g in 80 mL of anhydrous acetic acid R, add

5 mL of acetic anhydride R and mix for at least 10 min.

Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

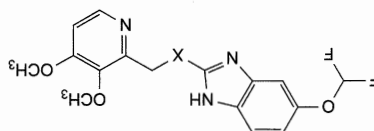
1 mL of 0.1 M perchloric acid is equivalent to 20.27 mg of $C_{16}H_{14}F_2N_3NaO_4S$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities: A, B, C, D, E, F.



A. X = SO₂: 5-(difluoromethoxy)-2-[[[(3,4-dimethoxyphenyl)methyl]sulfonyl]-1H-benzimidazole],
B. X = S: 5-(difluoromethoxy)-2-[[[(3,4-dimethoxyphenyl)methyl]sulfonyl]-1H-benzimidazole],
Yl)methyl]sulfonyl]-1H-benzimidazole,

DEFINITION

Papaveretum is a mixture of 253 parts of Morphine Hydrochloride ($C_{17}H_{19}NO_3 \cdot HCl$, 375.9), 23 parts of Papaverine Hydrochloride ($C_{20}H_{21}NO_4 \cdot HCl$, 375.9) and parts of Codeine Hydrochloride ($C_{18}H_{21}NO_3 \cdot HCl$, 371.9). It contains not less than 80.0% and not more than 88.4% of $C_{17}H_{19}NO_3 \cdot HCl$, not less than 8.3% and not more than 9.2% of $C_{20}H_{21}NO_4 \cdot HCl$ and not less than 6.6% and not more than 7.4% of $C_{18}H_{21}NO_3 \cdot HCl$, calculated with reference to the dried material.

CHARACTERISTICS

A white or almost white, crystalline powder. Soluble in water; sparingly soluble in ethanol (96%).

IDENTIFICATION

A. Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in water: (1) 1.5% w/v of the substance being examined.

Papaveretum

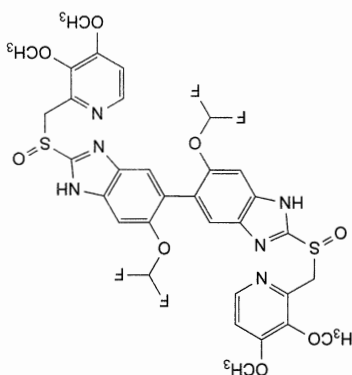
Action and use

Opioid receptor agonist; analgesic.

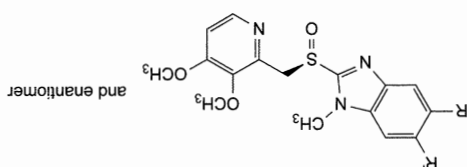
Preparation

Papaveretum Injection

E. mixture of the stereoisomers of 6,6'-bis(difluoromethoxy)-2,2'-bis[[[(3,4-dimethoxyphenyl)methyl]sulfonyl]-1H,1'-H-5,5'-bibenzimidazole].



D. R = OCHF₂, R' = H: 5-(difluoromethoxy)-2-[(RS)-[3,4-dimethoxyphenyl)methyl]sulfonyl]-1-methyl-1H-benzimidazole,
F. R = H, R' = OCHF₂: 6-(difluoromethoxy)-2-[(RS)-[3,4-dimethoxyphenyl)methyl]sulfonyl]-1-methyl-1H-benzimidazole,



C. 5-(difluoromethoxy)-1H-benzimidazole-2-thiol,

the pH of the mobile phase with either *glacial acetic acid* or *2M sodium hydroxide* in order to obtain optimum separation of the three principal components in solution (3). The retention times of codeine and papaverine relative to that of morphine are about 1.3 and 1.7 respectively.

DETERMINATION OF CONTENT

For anhydrous morphine hydrochloride
Calculate the content of $C_{17}H_{19}NO_3 \cdot HCl$ using the declared content of $C_{17}H_{19}NO_3$ in morphine sulfate BPCRS. Each mg of $C_{17}H_{19}NO_3$ is equivalent to 1.13 mg of $C_{17}H_{19}NO_3 \cdot HCl$.

For papaverine hydrochloride
Calculate the content of $C_{20}H_{21}NO_4 \cdot HCl$ using the declared content of $C_{20}H_{21}NO_4 \cdot HCl$ in papaverine hydrochloride BPCRS.

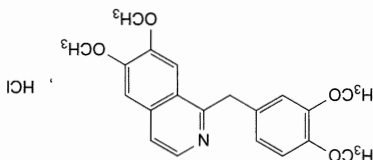
For anhydrous codeine hydrochloride
Calculate the content of $C_{18}H_{21}NO_3 \cdot HCl$ using the declared content of $C_{18}H_{21}NO_3 \cdot HCl$ in codeine hydrochloride BPCRS.

STORAGE

Papaveretum should be protected from light.

Papaverine Hydrochloride

(Ph. Eur. monograph 0102)



$C_{20}H_{22}ClNO_4$ 375.9

61-25-6

Action and use

Phosphodiesterase inhibitor; smooth muscle relaxant.

Preparation

Papaverine Injection

DEFINITION

1-(3,4-Dimethoxybenzyl)-6,7-dimethoxyisoquinoline hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder, or white or almost white crystals.

Solubility

Sparingly soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, D

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison papaverine hydrochloride CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

(2) 1.28% w/v of morphine sulfate BPCRS.

(3) 0.115% w/v of papaverine hydrochloride BPCRS.

(4) 0.10% w/v of codeine hydrochloride BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use a silica gel precoated plate (Merck silica gel 60 plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry at 100° to 105° for 15 minutes, allow it to cool and spray with potassium

iodobismuthate solution and then with a 0.4% v/v solution of sulfonic acid.

MOBILE PHASE

2 volumes of 13.5M ammonia, 6 volumes of ethanol (96%), 40 volumes of acetone and 40 volumes of toluene.

CONFIRMATION

The chromatogram obtained with solution (1) shows three principal spots corresponding in position and colour to the principal spots in the chromatograms obtained with solutions

(2), (3) and (4).

B. Yields the reactions characteristics of chlorides.

TESTS

Acidity

pH of a 1.5% w/v solution, 3.7 to 4.7, Appendix V L.

Clarity and colour of solution

A 1.5% w/v solution in water is clear, Appendix IV A, and not more intensely coloured than reference solution B_{Y5}.

Appendix IV B, Method II.

Loss on drying

When dried to constant weight at 130°, loses not less than 10.0% and not more than 14.0% of its weight. Use 0.5 g.

ASSAY

Carry out the method for liquid chromatography,

Appendix III D, using the following solutions in methanol

(60%).

(1) 0.15% w/v of the substance being examined.

(2) 0.128% w/v of morphine sulfate BPCRS.

(3) 0.128% w/v of morphine sulfate BPCRS, 0.0115% w/v of papaverine hydrochloride BPCRS and 0.010% w/v of codeine hydrochloride BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (10 cm × 4.6 mm) packed with end-capped octadecylsilyl silica gel for chromatography (5 µm) (Nucleosil C18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 285 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

0.01M sodium acetate and 0.005M diethyl sodium sulfosuccinate in methanol (60%) adjusted to pH 5.5 with glacial acetic acid.

SYSTEM SUITABILITY

If necessary, adjust the proportion of methanol in the mobile phase in the range 55% v/v to 65% v/v so that the retention time of morphine in solution (2) is 4 to 5 minutes. Adjust

Reference solution Dissolve 5 mg of papaverine hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase diethylamine R, ethyl acetate R, toluene R (10:20:70 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying At 100-105 °C for 2 h.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 10 mL of solution S (see Tests) add 5 mL of ammonia R dropwise and allow to stand for 10 min.

The precipitate, washed and dried, melts (2.2.14) at 146 °C to 149 °C.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S
Dissolve 0.4 g in carbon dioxide-free water R, heating gently if necessary, and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)
3.0 to 4.0 for solution S.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, mobile phase A (20:80 V/V).

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 12 mg of noscapine CRS in 1.0 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: base-deactivated octylsilyl silica gel for chromatography R (5 µm).

Mobile phase:

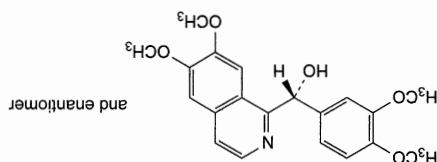
— mobile phase A: 3.4 g/L solution of potassium dihydrogen phosphate R adjusted to pH 3.0 with dilute phosphoric acid R;

— mobile phase B: acetonitrile R;

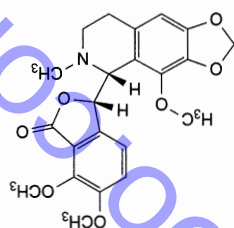
— mobile phase C: methanol R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Mobile phase C (per cent V/V)
0 - 5	85	5	10
5 - 12	85 → 60	5	10 → 35
12 - 20	60	5	35
20 - 24	60 → 40	5 → 20	35 → 40
24 - 27	40	20	40
27 - 32	40 → 85	20 → 5	40 → 10

B. (RS)-(3,4-dimethoxyphenyl)(6,7-dimethoxyisoquinolin-1-yl)methanol (papaverinol),



A. (3S)-6,7-dimethoxy-3-[(5R)-4-methoxy-6-methyl-5,6,7,8-tetrahydro-1,3-dioxolo[4,5-g]isoquinolin-5-yl]isobenzofuran-1(3H)-one (noscapine),



IMPURITIES

C₂₀H₂₂ClNO₄
1 mL of 0.1 M sodium hydroxide is equivalent to 37.59 mg of inflixion.

potentiometric titration (2.2.20), using 0.1 M sodium hydrochloric acid and 50 mL of alcohol R. Carry out a Dissolve 0.300 g in a mixture of 5.0 mL of 0.01 M

ASSAY

test for loss on drying.

Maximum 0.1 per cent, determined on the residue from the Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in Loss on drying (2.2.32)

(0.05 per cent).

the chromatogram obtained with reference solution (a) — disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— any impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) impurity C = 2.7; impurity D = 0.5;

corresponding correction factor: impurity A = 6.2;

the peak areas of the following impurities by the — correction factors: for the calculation of contents, multiply

Limits:

impurity A and papaverine.

— resolution: minimum 1.5 between the peaks due to System suitability: reference solution (b):

impurity D = about 1.2.

impurity A = about 0.9; impurity F = about 1.1;

impurity C = about 0.75; impurity B = about 0.8;

time = about 24 min; impurity E = about 0.7;

Relative retention With reference to papaverine (retention

Injection 10 µL.

Detection Spectrophotometer at 238 nm.

Flow rate 1 mL/min.

Paracetamol Suppositories

Paracetamol Tablets

Paracetamol and Caffeine Tablets

Soluble Paracetamol and Caffeine Tablets

Dispersible Paracetamol Tablets

Soluble Paracetamol Tablets

Paracetamol, Codeine Phosphate and Caffeine Capsules

Paracetamol, Codeine Phosphate and Caffeine Tablets

DEFINITION

N-(4-Hydroxyphenyl)acetamide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water, freely soluble in alcohol, very slightly soluble in methylene chloride.

IDENTIFICATION

First identification A, C

Second identification A, B, D, E

A. Melting point (2.2.14): 168 °C to 172 °C.

B. Dissolve 0.1 g in methanol R and dilute to 100.0 mL with the same solvent. To 1.0 mL of the solution add 0.5 mL of a

10.3 g/L solution of hydrochloric acid R and dilute to 100.0 mL with methanol R. Protect the solution from bright

light and immediately measure the absorbance (2.2.25) at the absorption maximum at 249 nm. The specific absorbance at

the maximum is 860 to 980.

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison paracetamol CRS.

D. To 0.1 g add 1 mL of hydrochloric acid R, heat to boiling for 3 min, add 1 mL of water R and cool in an ice bath.

No precipitate is formed. Add 0.05 mL of a 4.9 g/L solution of potassium dichromate R. A violet colour develops which does not change to red.

E. It gives the reaction of acetyl (2.3.1). Heat over a naked flame.

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 0.200 g of the substance to be examined in 2.5 mL of methanol R containing 4.0 g/L of a

400 g/L solution of tetrabutylammonium hydroxide R and dilute to 10.0 mL with a mixture of equal volumes of a

17.9 g/L solution of disodium hydrogen phosphate R and of a 7.8 g/L solution of sodium dihydrogen phosphate R.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 5.0 mL of this

solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

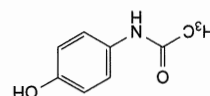
Reference solution (c) Dissolve 5.0 mg of 4-aminophenol R, 5 mg of paracetamol CRS and 5.0 mg of chloroacetanilide R in

methanol R and dilute to 20.0 mL with the same solvent.

Dilute 1.0 mL to 250.0 mL with the mobile phase.

Paracetamol

(Ph. Eur. monograph 0049)



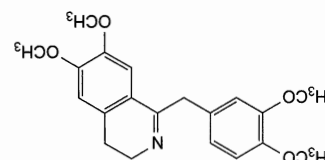
C₈H₉NO₂

151.2

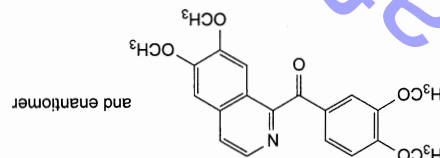
103-90-2



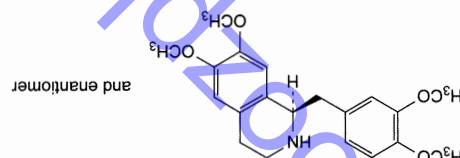
Ph. Eur.



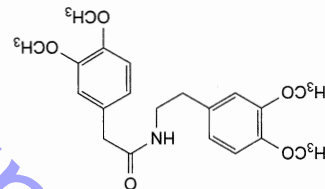
C. 1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-3,4-dihydroisoquinoline (dihydropapaverine),



D. (3,4-dimethoxyphenyl)(6,7-dimethoxyisoquinolin-1-yl)methanone (papaveraldehyde),



E. (1R,5S)-1-(3,4-dimethoxybenzyl)-6,7-dimethoxy-1,2,3,4-tetrahydroisoquinoline (tetrahydropapaverine),



F. 2-(3,4-dimethoxyphenyl)-N-[2-(3,4-dimethoxyphenyl)ethyl]acetamide.

Reference solution (d) Dissolve 20.0 mg of 4-nitrophenol R in methanol R and dilute to 50.0 mL with the same solvent.

Dilute 1.0 mL to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octylsilyl silica gel for chromatography R

(5 μ m),

— temperature: 35 °C.

Mobile phase Mix 375 volumes of a 17.9 g/L solution of

disodium hydrogen phosphate R, 375 volumes of a 7.8 g/L

solution of sodium dihydrogen phosphate R and 250 volumes of

methanol R containing 4.6 g/L of a 400 g/L solution of

tetrabutylammonium hydroxide R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 20 μ L.

Run time 12 times the retention time of paracetamol.

Relative retentions With reference to paracetamol (retention

time = about 4 min): impurity K = about 0.8;

impurity F = about 3; impurity J = about 7.

System suitability: reference solution (c):

— resolution: minimum 4.0 between the peaks due to

impurity K and to paracetamol,

— signal-to-noise ratio: minimum 50 for the peak due to

impurity J.

Limits:

— impurity J: not more than 0.2 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (c) (10 ppm),

— impurity K: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (c) (50 ppm),

— impurity F: not more than half the area of the

corresponding peak in the chromatogram obtained with

reference solution (d) (0.05 per cent),

— any other impurity: not more than half the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.1 per cent),

— disregard limit for the calculation of the total of other

impurities: the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.01 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in a mixture of 15 volumes of water R and 85 volumes of acetone R and dilute to 20 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (1 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with a mixture of 15 volumes of water R and 85 volumes of acetone R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 10 mL of water R and 30 mL of dilute sulfuric acid R. Boil under a reflux condenser for 1 h, cool and dilute to 100.0 mL with water R.

IMPURITIES

Protected from light.

STORAGE

C₈H₉NO₂.

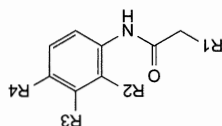
1 mL of 0.1 M cerium sulfate is equivalent to 7.56 mg of

yellow colour is obtained. Carry out a blank titration.

ferroin R. Titrate with 0.1 M cerium sulfate until a greenish-

ice, 15 mL of dilute hydrochloric acid R and 0.1 mL of

To 20.0 mL of the solution add 40 mL of water R, 40 g of



A. R₁ = R₃ = R₄ = H, R₂ = OH;
N-(2-hydroxyphenyl)acetamide,

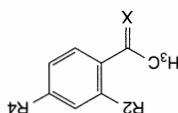
B. R₁ = CH₃, R₂ = R₃ = H, R₄ = OH;
N-(4-hydroxyphenyl)propanamide,

C. R₁ = R₂ = H, R₃ = Cl, R₄ = OH;
N-(3-chloro-4-hydroxyphenyl)acetamide,

D. R₁ = R₂ = R₃ = R₄ = H; N-phenylacetamide,

H. R₁ = R₂ = R₃ = H, R₄ = O-CO-CH₃;
4-(acetylamino)phenyl acetate,

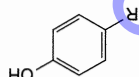
J. R₁ = R₂ = R₃ = H, R₄ = Cl;
N-(4-chlorophenyl)acetamide (chloroacetanilide),



E. X = O, R₂ = H, R₄ = OH;
1-(4-hydroxyphenyl)ethanone,

G. X = N-OH, R₂ = H, R₄ = OH;
1-(4-hydroxyphenyl)ethanone oxime,

I. X = O, R₂ = OH, R₄ = H; 1-(2-hydroxyphenyl)ethanone,



F. R = NO₂: 4-nitrophenol,

K. R = NH₂: 4-aminophenol.

Hard Paraffin

(Ph. Eur. monograph 1034)



CHARACTERS

Appearance

Colourless or white or almost white mass; the melted

substance is free from fluorescence in daylight.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, practically insoluble in ethanol (96 per cent).



Light Liquid Paraffin

(Ph. Eur. monograph 0240)

Preparation

Light Liquid Paraffin Eye Drops

Ph. Eur.

DEFINITION

Purified mixture of liquid saturated hydrocarbons obtained from petroleum.

CHARACTERS

Appearance

Colourless, transparent, oily liquid, free from fluorescence in daylight.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent), miscible with hydrocarbons.

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of liquid paraffin.

B. In a test tube cautiously boil 1 mL with 1 mL of 0.1 M

sodium hydroxide, with continuous shaking, for about 30 s.

On cooling to room temperature, 2 phases separate. To the

aqueous phase add 0.1 mL of phenolphthalein solution R.

The solution becomes red.

C. Viscosity (see Tests).

TESTS

Acidity or alkalinity

To 10 mL add 20 mL of boiling water R and shake

vigorously for 1 min. Separate the aqueous layer and filter.

To 10 mL of the filtrate, add 0.1 mL of phenolphthalein

solution R. The solution is colourless. Not more than 0.1 mL

of 0.1 M sodium hydroxide is required to change the colour of

the indicator to pink.

Relative density (2.2.5)

0.810 to 0.875.

Viscosity (2.2.9)

25 mPa·s to 80 mPa·s.

Polycyclic aromatic hydrocarbons

Use reagents for ultraviolet spectrophotometry.

Introduce 25.0 mL into a 125 mL separating funnel with

unlubricated ground-glass parts (stopper, stopcock).

Add 25 mL of hexane R which has been previously shaken

twice with one-fifth its volume of dimethyl sulfoxide R.

Mix and add 5.0 mL of dimethyl sulfoxide R. Shake vigorously

for 1 min and allow to stand until 2 clear layers are formed.

Transfer the lower layer to a 2nd separating funnel, add 2 mL

of hexane R and shake the mixture vigorously. Allow to stand

until 2 clear layers are formed. Separate the lower layer and

measure its absorbance (2.2.25) between 260 nm and

420 nm, using as the compensation liquid the clear lower

layer obtained by vigorously shaking 5.0 mL of dimethyl

sulfoxide R with 25 mL of hexane R for 1 min. Prepare a

7.0 mg/L reference solution of naphthalene R in

trimethylpentane R and measure the absorbance of the

solution at the absorption maximum at 275 nm, using

trimethylpentane R as the compensation liquid. At no

wavelength between 260 nm and 420 nm does the

absorbance of the test solution exceed one-third that of the

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison hard paraffin CRS.

Preparation Place about 2 mg on a sodium chloride plate, heat in an oven at 100 °C for 10 min, spread the melted

substance with another sodium chloride plate and remove

one of the plates.

B. Acidity or alkalinity (see Tests).

C. Melting point (2.2.16): 50 °C to 61 °C.

TESTS

Acidity or alkalinity

To 15 g add 30 mL of boiling water R and shake vigorously

for 1 min. Allow to cool and to separate. To 10 mL of the

aqueous layer add 0.1 mL of phenolphthalein solution R.

The solution is colourless. Not more than 1.0 mL of 0.01 M

sodium hydroxide is required to change the colour of the

indicator to red. To a further 10 mL of the aqueous layer

add 0.1 mL of methyl red solution R. The solution is yellow.

Not more than 0.5 mL of 0.01 M hydrochloric acid is required

to change the colour of the indicator to red.

Polycyclic aromatic hydrocarbons

Use reagents for ultraviolet absorption spectrophotometry. Dissolve

0.50 g in 25 mL of heptane R and place in a 125 mL

separating funnel with unlubricated ground-glass parts

(stopper, stopcock). Add 5.0 mL of dimethyl sulfoxide R.

Shake vigorously for 1 min and allow to stand until 2 clear

layers are formed. Transfer the lower layer to a 2nd

separating funnel, add 2 mL of heptane R and shake the

mixture vigorously. Allow to stand until 2 clear layers are

formed. Separate the lower layer and measure its absorbance

(2.2.25) between 265 nm and 420 nm using as the

compensation liquid the clear lower layer obtained by

vigorously shaking 5.0 mL of dimethyl sulfoxide R with 25 mL

of heptane R for 1 min. Prepare a 7.0 mg/L reference solution

of naphthalene R in dimethyl sulfoxide R and measure the

absorbance of this solution at the absorption maximum at

278 nm using dimethyl sulfoxide R as the compensation liquid.

At wavelengths from 265 nm to 420 nm, the absorbance of

the test solution is not greater than one-third that of the

reference solution at 278 nm.

Sulfates (2.4.13)

Maximum 150 ppm.

Introduce 2.0 g of the melted substance to be examined into

a 50 mL ground-glass-stoppered separating funnel.

Add 30 mL of boiling distilled water R, shake vigorously for

1 min and filter.

STORAGE

Protected from light.

Ph. Eur.

On cooling to room temperature, 2 phases separate. To the aqueous phase add 0.1 mL of *phenolphthalein* solution R. The solution becomes red.

C. Viscosity (see Tests).

TESTS

Acidity or alkalinity

To 10 mL add 20 mL of boiling *water* R and shake vigorously for 1 min. Separate the aqueous layer and filter. To 10 mL of the filtrate, add 0.1 mL of *phenolphthalein* solution R. The solution is colourless. Not more than 0.1 mL of 0.1 M *sodium hydroxide* is required to change the colour of the indicator to pink.

Relative density (2.2.5)

0.827 to 0.890.

Viscosity (2.2.9)

110 mPa.s to 230 mPa.s.

Polycyclic aromatic hydrocarbons

Use reagents for ultraviolet spectrophotometry.

Introduce 25.0 mL into a 125 mL separating funnel with

unlubricated ground-glass parts (stopper, stopcock).

Add 25 mL of *hexane* R which has been previously shaken

twice with one-fifth its volume of *dimethyl sulfoxide* R.

Mix and add 5.0 mL of *dimethyl sulfoxide* R. Shake vigorously

for 1 min and allow to stand until 2 clear layers are formed.

Transfer the lower layer to a 2° separating funnel, add 2 mL

of *hexane* R and shake the mixture vigorously. Allow to stand

until 2 clear layers are formed. Separate the lower layer and

measure its absorbance (2.2.25) between 260 nm and

420 nm, using as the compensation liquid the clear lower

layer obtained by vigorously shaking 5.0 mL of *dimethyl*

sulfoxide R with 25 mL of *hexane* R for 1 min. Prepare a

7.0 mg/L reference solution of *naphthalene* R in

trimethylpentane R and measure the absorbance of the

solution at the absorption maximum at 275 nm, using

trimethylpentane R as the compensation liquid. At no

wavelength between 260 nm and 420 nm does the

absorbance of the test solution exceed one-third that of the

reference solution at 275 nm.

Readily carbonisable substances

Use a ground-glass-stoppered tube about 125 mm long and

18 mm in internal diameter, graduated at 5 mL and 10 mL;

wash with hot *water* R (temperature at least 60 °C),

acetone R, *heptane* R and finally with *acetone* R, dry at

100-110 °C. Cool in a desiccator. Introduce 5 mL of the

substance to be examined and add 5 mL of *nitrogen-free*

sulfuric acid R1. Insert the stopper and shake as vigorously as

possible, in the longitudinal direction of the tube, for 5 s.

Loosen the stopper, immediately place the tube in a water-

bath, avoiding contact of the tube with the bottom or side of

the bath, and heat for 10 min. After 2 min, 4 min, 6 min

and 8 min, remove the tube from the bath and shake as

vigorously as possible, in the longitudinal direction of the

tube for 5 s. At the end of 10 min of heating, remove the

tube from the water-bath and allow to stand for 10 min.

Centrifuge at 2000 g for 5 min. The lower layer is not more

intensely coloured (2.2.2, *Method I*) than a mixture of

0.5 mL of blue primary solution, 1.5 mL of red primary

solution, 3.0 mL of yellow primary solution and 2 mL of a

10 g/L solution of *hydrochloric acid* R.

Solid paraffins

Dry a suitable quantity of the substance to be examined by

heating at 100 °C for 2 h and cool in a desiccator over

sulfuric acid R. Place in a glass tube with an internal diameter

of about 25 mm, close the tube and immerse in a bath of

Liquid Paraffin

(Ph. Eur. monograph 0239)

Action and use

Faecal softener.

Preparations

Liquid Paraffin Oral Emulsion

Liquid Paraffin and Magnesium Hydroxide Oral Emulsion

DEFINITION

Purified mixture of liquid saturated hydrocarbons obtained

from petroleum.

CHARACTERS

Appearance

Colourless, transparent, oily liquid, free from fluorescence in

daylight.

Solubility

Practically insoluble in water, slightly soluble in ethanol

(96 per cent), miscible with hydrocarbons.

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of liquid paraffin.

B. In a test tube cautiously boil 1 mL with 1 mL of 0.1 M

sodium hydroxide, with continuous shaking, for about 30 s.



White Soft Paraffin

White Petroleum Jelly

(Ph. Eur. monograph 1799)

Ph Eur

DEFINITION

Purified and wholly or nearly decolorised mixture of semi-solid hydrocarbons, obtained from petroleum. It may contain a suitable antioxidant. White soft paraffin described in this monograph is not suitable for oral use.

CHARACTERS

Appearance

White or almost white, translucent, soft unctuous mass, slightly fluorescent in daylight when melted.

Solubility

Practically insoluble in water, slightly soluble in methylene chloride, practically insoluble in ethanol (96 per cent) and in glycerol.

IDENTIFICATION

First identification A, B, D

Second identification A, C, D

A. The drop point is between 35 °C and 70 °C and does not differ by more than 5 °C from the value stated on the label, according to method (2.2.17) with the following modification

to fill the cup: heat the substance to be examined at a temperature not exceeding 80 °C, with stirring to ensure uniformity. Warm the metal cup at a temperature not exceeding 80 °C in an oven, remove it from the oven, place on a clean plate or ceramic tile and pour a sufficient quantity of the melted sample into the cup to fill it completely. Allow the filled cup to cool for 30 min on the plate or the ceramic tile and place it in a water bath at 24–26 °C for 30–40 min. Level the surface of the sample with a single stroke of a knife or razor blade, avoiding compression of the sample.

B. Infrared absorption spectrophotometry (2.2.24).
Preparation Place about 2 mg on a sodium chloride R plate, spread the substance with another sodium chloride R plate and remove 1 of the plates.
Comparison Repeat the operations using white soft paraffin CRS.

C. Melt 2 g and when a homogeneous phase is obtained, add 2 mL of water R and 0.2 mL of 0.05 M iodine. Shake. Allow to cool. The solid upper layer is violet-pink or brown.
D. Appearance (see Tests).

TESTS

Appearance

The substance is white. Melt 12 g on a water-bath. The melted mass is not more intensely coloured than a mixture of 1 volume of yellow primary solution and 9 volumes of a 10 g/L solution of hydrochloric acid R (2.2.2. Method II).

Acidity or alkalinity

To 10 g add 20 mL of boiling water R and shake vigorously for 1 min. Allow to cool and decant. To 10 mL of the aqueous layer add 0.1 mL of phenolphthalein solution R. The solution is colourless. Not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to red.

Consistency (2.9.9)

60 to 300.

Polycyclic aromatic hydrocarbons

Use reagents for ultraviolet spectrophotometry. Dissolve 1.0 g in 50 mL of hexane R which has been previously shaken twice with 10 mL of dimethyl sulfoxide R. Transfer the solution to a 125 mL separating funnel with unlubricated ground-glass parts (stopper, stopcock). Add 20 mL of dimethyl sulfoxide R. Shake vigorously for 1 min and allow to stand until 2 clear layers are formed. Transfer the lower layer to a second separating funnel. Repeat the extraction with a further 20 mL of dimethyl sulfoxide R. Shake vigorously the combined lower layers with 20 mL of hexane R for 1 min. Allow to stand until 2 clear layers are formed. Separate the lower layer and dilute to 50.0 mL with dimethyl sulfoxide R. Measure the absorbance (2.2.25) over the range 260 nm to 420 nm using a path length of 4 cm and as compensation liquid the clear lower layer obtained by vigorously shaking 10 mL of dimethyl sulfoxide R with 25 mL of hexane R for 1 min. Prepare a reference solution in dimethyl sulfoxide R containing 6.0 mg of naphthalene R per litre and measure the absorbance of the solution at the maximum at 278 nm using a path length of 4 cm and dimethyl sulfoxide R as compensation liquid. At no wavelength in the range 260 nm to 420 nm does the absorbance of the test solution exceed that of the reference solution at 278 nm.

Sulfated ash (2.4.14)

Maximum 0.05 per cent, determined on 2.0 g.

STORAGE

Protected from light.

LABELLING

The label states the nominal drop point.

Yellow Soft Paraffin

Yellow Petroleum Jelly

(Ph. Eur. monograph 1554)

Action and use

Excipient.

DEFINITION

Purified mixture of semi-solid hydrocarbons, obtained from petroleum. It may contain a suitable antioxidant.

CHARACTERS

Appearance

Yellow, translucent, unctuous mass, slightly fluorescent in daylight when melted.

Solubility

Practically insoluble in water, slightly soluble in methylene chloride, practically insoluble in ethanol (96 per cent) and in glycerol.



Ph Eur

Ph Eur

IDENTIFICATION

First identification A, B, D
Second identification A, C, D

A. The drop point (2.2.17) is 40 °C to 60 °C and does not differ by more than 5 °C from the value stated on the label, with the following modification to fill the cup: heat the substance to be examined at 118-122 °C, with stirring to ensure uniformity, then cool to 100-107 °C. Warm the metal cup at 103-107 °C in an oven, remove it from the oven, place on a clean plate or ceramic tile and pour a sufficient quantity of the melted sample into the cup to fill it completely. Allow the filled cup to cool for 30 min on the ceramic tile and place it in a water-bath at 24-26 °C for a further 30-40 min. Level the surface of the sample with a single stroke of a knife or razor blade, avoiding compression of the sample.

B. Examine by infrared absorption spectrophotometry (2.2.24).

Preparation Place about 2 mg on a sodium chloride R plate, spread the substance with another sodium chloride R plate and remove 1 of the plates.

Comparison Repeat the operations using yellow soft paraffin CRS.

C. Melt 2 g and when a homogeneous phase is obtained, add 2 mL of water R and 0.2 mL of 0.05 M iodine. Shake. Allow to cool. The solid upper layer is violet-pink or brown.

D. Appearance (see Tests).

TESTS

Appearance

The substance is yellow. Melt 12 g on a water-bath.

The melted mass is not more intensely coloured than a mixture of 7.6 volumes of yellow primary solution and 2.4 volumes of red primary solution (2.2.2, Method II).

Acidity or alkalinity

To 10 g add 20 mL of boiling water R and shake vigorously for 1 min. Allow to cool and decant. To 10 mL of the aqueous layer add 0.1 mL of phenolphthalein solution R. The solution is colourless. Not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to red.

Consistency (2.9.9)

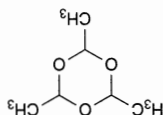
100 to 300.

Polycyclic aromatic hydrocarbons

Use reagents for ultraviolet absorption spectrophotometry. Dissolve 1.0 g in 50 mL of hexane R which has been previously shaken twice with one-fifth its volume of dimethyl sulfoxide R. Transfer the solution to a 125 mL separating funnel with unlubricated ground-glass parts (stopper, stopcock). Add 20 mL of dimethyl sulfoxide R. Shake vigorously for 1 min and allow to stand until two clear layers are formed. Transfer the lower layer to a 2nd separating funnel. Repeat the extraction with a further 20 mL of dimethyl sulfoxide R. Shake vigorously the combined lower layers with 20 mL of hexane R for 1 min. Allow to stand until 2 clear layers are formed. Separate the lower layer and dilute to 50.0 mL with between 260 nm and 420 nm using a path length of 4 cm and using as the compensation liquid the clear lower layer obtained by vigorously shaking 10 mL of dimethyl sulfoxide R with 25 mL of hexane R for 1 min. Prepare a 9.0 mg/L reference solution of naphthalene R in dimethyl sulfoxide R and measure the absorbance of this solution at the maximum at 278 nm using a path length of 4 cm and using dimethyl sulfoxide R as the compensation liquid. At no wavelength in

Paraldehyde

(Ph. Eur. monograph 0351)



$C_6H_{12}O_3$

132.2

123-63-7

Preparation
Paraldehyde Injection

Action and use
Anticonvulsant.

Ph. Eur.

DEFINITION

2,4,6-Trimethyl-1,3,5-trioxane (cyclic trimer of acetaldehyde).

It may contain a suitable quantity of an antioxidant.

CHARACTERS

Appearance

Colourless or slightly yellow, transparent liquid. It solidifies on cooling to form a crystalline mass.

Solubility

Soluble in water, but less soluble in boiling water, miscible with ethanol (96 per cent) and with essential oils.

IDENTIFICATION

A. Solution S (see Tests) is clear (2.2.1) but becomes turbid on warming.

B. To 5 mL add 0.1 mL of dilute sulfuric acid R and heat. Acetaldehyde, recognisable by its odour, is evolved.

C. To 5 mL of solution S in a test-tube add 5 mL of ammoniacal silver nitrate solution R and heat in a water-bath. Silver is deposited as a mirror on the wall of the tube.

TESTS

Solution S

Dissolve 20.0 mL in carbon dioxide-free water R and dilute to 200.0 mL with the same solvent.

Acidity

To 50.0 mL of solution S add 0.05 mL of phenolphthalein solution R. Not more than 1.5 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Refractive index (2.2.6)

1.403 to 1.406.

Relative density (2.2.5)

0.991 to 0.996.

Distillation range (2.2.11)
A maximum of 10 per cent distils below 123 °C and a minimum of 95 per cent distils below 126 °C.

Freezing point (2.2.18)
10 °C to 13 °C.

Acetaldehyde
To 5.0 mL add a mixture of 0.2 mL of methyl orange

solution R₃, 5 mL of ethanol (60 per cent V/V) R and 5 mL of alcoholic hydroxylamine solution R and shake. Not more than 0.8 mL of 0.5 M sodium hydroxide is required to change the colour of the indicator to pure yellow.

Peroxides
Place 50.0 mL of solution S in a ground-glass-stoppered

flask, add 5 mL of dilute sulfuric acid R and 10 mL of potassium iodide solution R, close the flask and allow to stand protected from light for 15 min. Titrate with 0.1 M sodium thiosulfate using 1 mL of starch solution R as indicator. Allow to stand for 5 min and, if necessary complete the titration.

Not more than 2.0 mL of 0.1 M sodium thiosulfate is required.

Non-volatile residue
Maximum 0.6 g/L.
Heat 5.0 mL in a tared evaporating dish on a water-bath and dry at 105 °C for 1 h. The residue weighs a maximum of 3 mg.

STORAGE
In a small, well-filled, airtight container, protected from light. If the substance has solidified the whole contents of the container must be liquefied before use.

Ph. Eur. monograph 1252)

Parnaparin Sodium

Chemical structure

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

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Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Distillation range (2.2.11)
A maximum of 10 per cent distils below 123 °C and a minimum of 95 per cent distils below 126 °C.

Freezing point (2.2.18)
10 °C to 13 °C.

Acetaldehyde
To 5.0 mL add a mixture of 0.2 mL of methyl orange

solution R₃, 5 mL of ethanol (60 per cent V/V) R and 5 mL of alcoholic hydroxylamine solution R and shake. Not more than 0.8 mL of 0.5 M sodium hydroxide is required to change the colour of the indicator to pure yellow.

Peroxides
Place 50.0 mL of solution S in a ground-glass-stoppered

flask, add 5 mL of dilute sulfuric acid R and 10 mL of potassium iodide solution R, close the flask and allow to stand protected from light for 15 min. Titrate with 0.1 M sodium thiosulfate using 1 mL of starch solution R as indicator. Allow to stand for 5 min and, if necessary complete the titration.

Not more than 2.0 mL of 0.1 M sodium thiosulfate is required.

Non-volatile residue
Maximum 0.6 g/L.
Heat 5.0 mL in a tared evaporating dish on a water-bath and dry at 105 °C for 1 h. The residue weighs a maximum of 3 mg.

STORAGE
In a small, well-filled, airtight container, protected from light. If the substance has solidified the whole contents of the container must be liquefied before use.

Ph. Eur. monograph 1252)

Parnaparin Sodium

Chemical structure

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

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Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

Ph. Eur. monograph 1252)

PRODUCTION

Impurity G Maximum 1 ppm, determined by a suitable, validated method.

CHARACTERS**Appearance**

White or almost white, hygroscopic, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in methanol, sparingly soluble in anhydrous ethanol and in methylene chloride. It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Comparison anhydrous paroxetine hydrochloride CRS.

If the spectra obtained in the solid state show differences, mix 1 part of the substance to be examined and 1 part of the reference substance separately with 30 parts of anhydrous

acetone R and heat to boiling to dissolve. Recrystallise and record new spectra using the residues.

B. Water (see Tests).

C. It gives reaction (b) of chlorides (2.3.1).

TESTS**Impurity D**

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 5 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of paroxetine impurity D CRS in 2 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the test solution.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.10$ m, $\phi = 4.0$ mm;
— stationary phase: silica gel AGP for chiral chromatography R (5 μ m);
— temperature: 30 °C.

Mobile phase Dissolve 8.7 g of dipotassium hydrogen phosphate R in 1000 mL of water for chromatography R and adjust to pH 6.5 with phosphoric acid R; mix 930 mL of this solution and 70 mL of acetonitrile R.

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 295 nm.

Injection 20 μ L of the test solution and reference solutions (b) and (c).

Run time 2.5 times the retention time of paroxetine which is about 12 min.

System suitability:

— **peak-to-valley ratio:** minimum 2.0, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to paroxetine in the chromatogram obtained with reference solution (b);

— **signal-to-noise ratio:** minimum 3 for the principal peak in the chromatogram obtained with reference solution (c);

— **symmetry factor:** the requirements stated in chapter 2.2.46 are not applicable.

Limit: — **impurity D:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture tetrahydrofuran R, water R (10:90 V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 5.0 mL of the test solution to 50.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of anhydrous paroxetine hydrochloride impurity H CRS in 25 mL of tetrahydrofuran R and dilute to 50.0 mL with water R.

Reference solution (c) Dissolve 5 mg of anhydrous paroxetine hydrochloride impurity C CRS in 25 mL of tetrahydrofuran R and dilute to 50.0 mL with water R.

Reference solution (d) To 5.0 mL of reference solution (a) add 1.0 mL of reference solution (b) and dilute to 100.0 mL with the solvent mixture.

Reference solution (e) To 5.0 mL of reference solution (a) add 5.0 mL of reference solution (b) and 5.0 mL of reference solution (c). Dilute to 100.0 mL with the solvent mixture.

Reference solution (f) Dissolve 2.5 mg of paroxetine impurity E CRS in the solvent mixture, add 2.5 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Reference solution (g) Dissolve 5 mg of paroxetine impurity A CRS in the solvent mixture and dilute to 50 mL with the solvent mixture. Use this solution to identify the peak due to impurity A.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m);
— temperature: 40 °C.

Mobile phase: — **mobile phase A:** trifluoroacetic acid R, tetrahydrofuran R, water R (5:100:900 V/V/V);
— **mobile phase B:** trifluoroacetic acid R, tetrahydrofuran R, acetonitrile R (5:100:900 V/V/V).

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	80	20
30 - 50	80 \rightarrow 20	20 \rightarrow 80
50 - 55	20	80
55 - 60	20 \rightarrow 80	80 \rightarrow 20
60 - 65	80	20

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 295 nm.

Injection 20 μ L of the test solution and reference solutions (d), (e), (f) and (g).

Relative retention With reference to paroxetine (retention time = about 28 min): impurity A = about 0.8; impurity B = about 0.9; impurity C = about 1.5.

— *resolution*: minimum S/N between the peaks due to impurity E and paroxetine in the chromatogram obtained with reference solution (f);

— *impurity A*: not more than 0.6 times the area of the peak due to paroxetine in the chromatogram obtained with reference solution (d) (0.3 per cent);

— *unspecified impurities*: for each impurity, not more than 0.2 times the area of the peak due to paroxetine in the chromatogram obtained with reference solution (d) (0.10 per cent);

— *total*: not more than the area of the peak due to

Impurities H and I
(0.05 per cent).
Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.
Mobile phase composition at 363 nm

impurity H = about 0.4.

System suitability: reference solution (e):

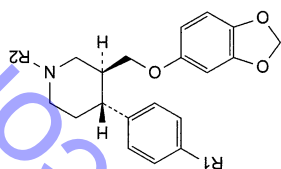
— signal-to-noise ratio: minimum 3 for the peak due to impurity H.

Maximum 3.5 per cent.
2-Propanol (2.4.24, System B)
Maximum 4.3 per cent.
Heavy metals (2.4.8)

(10 ppm 1-6) K.
Maximum 2.5.12
Water (2.5.12)
Maximum 1.5 per cent, determined on 0.500 g.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g in a platinum

Specified impurity

5-yl)oxy)methyl]-4-phenylpiperidine (desflunorparoxetine), C. R1 = F, R2 = CH₂-C₆H₅; (3S,4R)-3-[(1,3-benzodioxol-5-yl)oxy)methyl]-1-benzyl-4-(4-fluorophenyl)piperidine (N-benzylparoxetine),



IMPURITIES
In an airtight container, weigh 25.0 g. of the specified impurity.

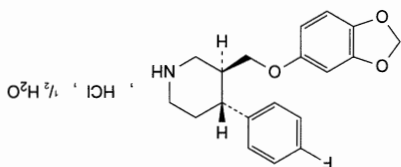
(2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, E.

A. R1 = H; R2 = (3S,4R)-3-[(1,3-benzodioxol-5-yl)oxy]methyl-4-phenylpiperidine (desfluraneoxetane).

F: K1 = H, K2 = C₆H₅; (3S,4R)-5-[(1,3-benzodioxol-5-yloxy)methyl]-1-benzyl-4-phenylpiperidine
(*N*-benzyl-4-phenylpiperidine),

Paroxetine Hydrochloride Hemihydrate

(Ph. Eur. monograph 2018)



$C_{19}H_{21}ClFN_3 \cdot \frac{1}{2}H_2O$ 374.8 110429-35-1

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Ph Eur

DEFINITION

(3*S*,4*R*)-3-[(1,3-benzodioxol-5-yl)oxy)methyl]-4-(4-fluorophenyl)piperidine hydrochloride hemihydrate.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

Impurity G

Maximum 1 ppm, determined by a suitable, validated method.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in methanol, sparingly soluble in ethanol (96 per cent) and in methylene chloride.

It shows pseudopolymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison paroxetine hydrochloride hemihydrate CRS.

If the spectra obtained show differences, dissolve 1 part of the substance to be examined and 1 part of the reference substance separately in 10 parts of a mixture of 1 volume of water *R* and 9 volumes of 2-propanol *R* and heat to 70 °C to dissolve. Recrystallise and record new spectra using the residues.

B. Examine the chromatograms obtained in the test for impurity D.

Injection Test solution and reference solution (c).

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution (c).

C. Water (see Tests).

D. It gives reaction (b) of chlorides (2.3.1).

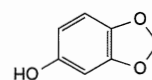
TESTS

Impurity D

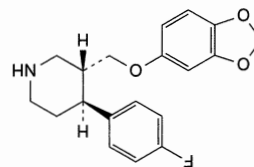
Liquid chromatography (2.2.29).

Test solution Dissolve 0.1000 g of the substance to be examined in 20 mL of methanol *R* and dilute to 100.0 mL with the mobile phase.

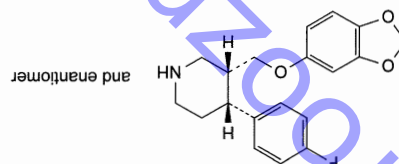
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.



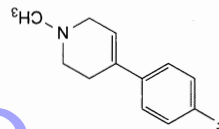
B. 1,3-benzodioxol-5-ol (sesamol),



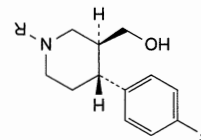
D. (3*R*,4*S*)-3-[(1,3-benzodioxol-5-yl)oxy)methyl]-4-(4-fluorophenyl)piperidine ((+)-*trans*-paroxetine),



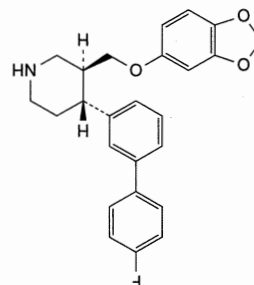
E. (3*R*,4*R*)-3-[(1,3-benzodioxol-5-yl)oxy)methyl]-4-(4-fluorophenyl)piperidine (*cis*-paroxetine),



G. 4-(4-fluorophenyl)-1-methyl-1,2,3,6-tetrahydropyridine,



H. $R = CH_2-C_6H_5$; [(3*S*,4*R*)-1-benzyl-4-(4-fluorophenyl)piperidin-3-yl] methanol, I. $R = H$; [(3*S*,4*R*)-4-(4-fluorophenyl)piperidin-3-yl] methanol,



J. (3*S*,4*R*)-3-[(1,3-benzodioxol-5-yl)oxy)methyl]-4-(4'-fluorobiphenyl-3-yl)piperidine.

Ph Eur



Reference solution (b) Dissolve 5 mg of paroxetine impurity D CRS and 5 mg of paroxetine hemihydrate CRS in 2 mL of methanol R and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dissolve 10 mg of paroxetine hemihydrate CRS in 2 mL of methanol R and dilute to 10.0 mL with the mobile phase.

Column:
— size: $l = 0.10$ m, $\varnothing = 4.0$ mm;
— stationary phase: silica gel AGP for chiral chromatography R (5 μ m).

Mobile phase Mix 2 volumes of methanol R and 8 volumes of a 5.8 g/L solution of sodium chloride R.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 295 nm.

Injection 10 μ L of the test solution and reference solutions (a) and (b).

Run time 2.5 times the retention time of paroxetine.

Retention time Paroxetine = about 30 min.

System suitability: reference solution (b):
— resolution: minimum 2.2 between the peaks due to impurity D and paroxetine.

Limit:
— impurity D: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent).

Related substances
Liquid chromatography (2.2.29).
Solvent mixture tetrahydrofuran R, water R (1:9 V/V).
Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the same solvent mixture.

Reference solution (a) Dilute 5.0 mL of the test solution to 50.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 200.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2 mg of paroxetine for system suitability CRS (containing impurity C) in the solvent mixture and dilute to 10 mL with the solvent mixture. Dilute 1 mL of this solution to 10 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of paroxetine impurity A CRS in the solvent mixture and dilute to 20 mL with the same solvent mixture.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m);
— temperature: 40 °C.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	80	20
30 - 50	80 \rightarrow 20	20 \rightarrow 80
50 - 60	20	80
60 - 65	20 \rightarrow 80	80 \rightarrow 20
65 - 70	80	20

— mobile phase A: trifluoroacetic acid R, tetrahydrofuran R, water R (5:100:900 V/V/V);
— mobile phase B: trifluoroacetic acid R, tetrahydrofuran R, acetonitrile R (5:100:900 V/V/V);

Flow rate 1 mL/min.

Detection Spectrophotometer at 295 nm.

Injection 20 μ L.

Relative retention With reference to paroxetine: impurity A = about 0.8.

System suitability: reference solution (b):
— resolution: minimum 3.5 between the peaks due to impurity C and paroxetine.

Limits:
— impurity A: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)
Maximum 20 ppm.

1.0 g complies with test C. Use a platinum crucible. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)
2.2 per cent to 2.7 per cent, determined on 0.300 g.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

ASSAY
Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of paroxetine hemihydrate CRS in water R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 5.0 mg of paroxetine hemihydrate CRS and 5 mg of paroxetine impurity A CRS in water R and dilute to 10.0 mL with the same solvent.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: trimethylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve 3.85 g of ammonium acetate R in water R, adjust to pH 5.5 with anhydrous acetic acid R and dilute to 600 mL with the same solvent; add 400 mL of acetonitrile R, slowly add, with stirring, 10 mL of triethylamine R and readjust to pH 5.5 with anhydrous acetic acid R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 295 nm.

Injection 10 μ L.

Run time Twice the retention time of paroxetine.

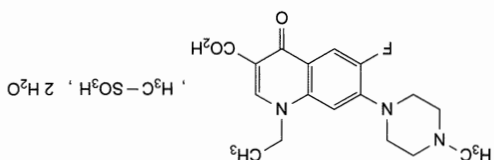
System suitability: reference solution (b):
— resolution: minimum 2 between the peaks due to paroxetine and impurity A.

Calculate the percentage content of paroxetine hydrochloride using the chromatogram obtained with reference solution (a).



Pefloxacin Mesilate

(Pefloxacin Mesilate Dihydrate,
Ph Eur monograph 1460)



$C_{18}H_{24}FN_3O_6S_2 \cdot 2H_2O$ 465.5 149676-40-4

Action and use
Antibacterial.

Ph Eur

DEFINITION

1-Ethyl-6-fluoro-7-(4-methylpiperazin-1-yl)-4-oxo-1,4-dihydroquinoline-3-carboxylic acid methanesulfonate dihydrate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

PRODUCTION

It is considered that alkylsulfonate esters are genotoxic and are potential impurities in pefloxacin mesilate dihydrate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods 2.5.37, Methyl, ethyl and isopropyl methanesulfonate in methanesulfonate in active substances and 2.5.39, methanesulfonate in active substances and 2.5.39, assist manufacturers.

CHARACTERS

Appearance

Fine, white or almost white powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Preparation Dissolve 0.1 g in 10 mL of water R. Add 5 mL of 1 M sodium hydroxide. Adjust to pH 7.4 ± 0.1 with phosphoric acid R and shake with 2 quantities, each of 30 mL, of methylene chloride R. Combine the organic layers and dry over anhydrous sodium sulfate R. Evaporate to dryness. Examine the residue as a disc of potassium bromide R.
Comparison Repeat the operations using 0.1 g of pefloxacin mesilate dihydrate CRS.

B. Thin-layer chromatography (2.2.27).
Test solution Dissolve 40 mg in water R and dilute to 1 mL with the same solvent.
Reference solution Dissolve 60 mg of methanesulfonic acid R in water R and dilute to 10 mL with the same solvent.

Plate TLC silica gel plate R.
Mobile phase water R, ammonia R, butanol R, acetone R (5:10:20:65 V/V/V/V).

Application 10 µL.
Development Over a path of 15 cm.
Drying In air.
Detection Spray with a 0.4 g/L solution of bromocresol purple R in ethanol (50 per cent V/V) R, adjusted to pH 10 using 1 M sodium hydroxide.

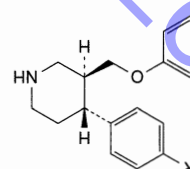
STORAGE

Protected from light.

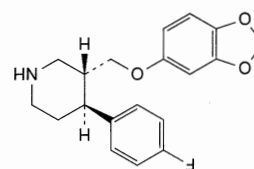
IMPURITIES

Specified impurities A, D, G.

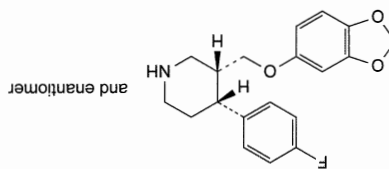
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10, Control of impurities in substances for pharmaceutical use): B, C, E, F.



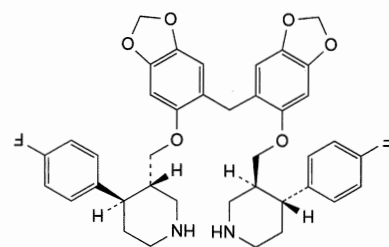
A, R = H: (3S,4R)-3-[(1,3-benzodioxol-5-yl)oxy]methyl-4-(4-fluorophenyl)piperidine (desfluoroparoxetine),
B, R = OCH₃: (3S,4R)-3-[(1,3-benzodioxol-5-yl)oxy]methyl-4-(4-methoxyphenyl)piperidine,
C, R = OC₂H₅: (3S,4R)-3-[(1,3-benzodioxol-5-yl)oxy]methyl-4-(4-ethoxyphenyl)piperidine.



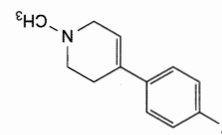
D, (3R,4S)-3-[(1,3-benzodioxol-5-yl)oxy]methyl-4-(4-fluorophenyl)piperidine ((+)-trans-paroxetine), and enantiomer



E, (3R,4R)-3-[(1,3-benzodioxol-5-yl)oxy]methyl-4-(4-fluorophenyl)piperidine (cis-paroxetine),



F, 3,3'-(methylenebis(1,3-benzodioxol-6,5-diyl)oxy)methyl-4-(4-fluorophenyl)piperidine],



G, 4-(4-fluorophenyl)-1-methyl-1,2,3,6-tetrahydropyridine.

Ph Eur

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution.

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 10.0 mL with the same solvent.

Appearance of solution

Examined within 1 h after its preparation, solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than intensity 3 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

pH (2.2.3)

3.5 to 4.5.

Dilute 1 mL of solution S to 10 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of pefloxacin impurity B CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. In 2.0 mL of this solution, dissolve the contents of a vial of pefloxacin impurity C CRS.

Reference solution (b) Dissolve 10.0 mg of norfloxacin impurity A CRS (impurity F) in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:
— size: $l = 0.15$ m, $\phi = 6$ mm;
— stationary phase: octadecylsilyl vinyl polymer for chromatography R (5 μ m).

Mobile phase Mix 30 volumes of acetonitrile R, 70 volumes of a solution containing 2.70 g/L of cetyltrimethylammonium bromide R and 6.18 g/L of boric acid R (exactly adjusted to pH 8.30 with 1 M sodium hydroxide), and 0.2 volumes of rhodithylene glycol R.

Detection Spectrophotometer at 258 nm and at 273 nm.

Injection 20 μ L.

Run time 4 times the retention time of pefloxacin (about 60 min).

Relative retentions and correction factors:

Impurity	Approximate relative retention	Correction factor
Impurity E	0.2	—
Impurity D	0.3	—
Impurity A	0.5	—
Impurity G	0.8	1.4
Pefloxacin	1	—
Impurity C	1.7	2.4
Impurity B	1.8	—
Impurity H	2.4	1.8
Impurity F	3.5	—

System suitability Reference solution (a) at 273 nm: — resolution: minimum 1.5 between the peaks due to impurities C and B.

From the chromatogram obtained at 258 nm with the test solution, calculate the percentage content of impurities C, F, G and H using the area of the principal peak in the chromatogram obtained at 258 nm with reference solution (b) (external standardisation) taking into account the correction factors indicated in the table.

From the chromatogram obtained at 273 nm with the test solution, calculate the percentage content of impurities A, B, D and E and of any other impurity from the areas of the peaks in the chromatogram obtained with the test solution by the normalisation procedure.

Limits:
— impurities A, B, D, E and any other impurity at 273 nm and impurities C, F, G, H at 258 nm: for each impurity, maximum 0.5 per cent and not more than 3 impurities have a content between 0.2 per cent and 0.5 per cent;
— total: maximum 1.0 per cent;
— disregard limit at 273 nm: 0.0005 times the area of the principal peak in the chromatogram obtained with the test solution (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

1.0 g complies with test E. Prepare the reference solution using 10.0 mL of lead standard solution (1 ppm Pb) R.

Water (2.5.12)

7.0 per cent to 8.5 per cent, determined on 50.0 mg using a mixture of 10 volumes of methanol R and 50 volumes of methylene chloride R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 15.0 mL of anhydrous acetic acid R and add 75.0 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

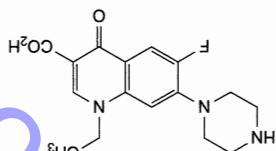
1 mL of 0.1 M perchloric acid is equivalent to 21.48 mg of $C_{18}H_{24}FN_3O_6S$.

STORAGE

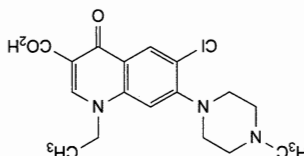
In an airtight container, protected from light.

IMPURITIES

Specified impurities A, B, C, D, E, F, G, H



A. 1-ethyl-6-fluoro-4-oxo-7-(piperazin-1-yl)-1,4-dihydroquinoline-3-carboxylic acid (demethylated pefloxacin or norfloxacin),

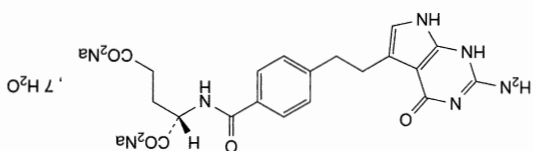


B. 6-chloro-1-ethyl-7-(4-methylpiperazin-1-yl)-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (chlorinated homologue of pefloxacin),



Pemetrexed Disodium Heptahydrate

(Ph. Eur. monograph 2637)



$C_{20}H_{19}N_5Na_2O_6 \cdot 7H_2O$ 597.5 357166-29-1

Action and use

Thymidylate synthetase inhibitor; cytostatic.

DEFINITION

Disodium (2S)-2-[[4-[2-(2-amino-4-oxo-4,7-dihydro-1H-pyrido[2,3-d]pyrimidin-5-yl)ethyl]benzoyl]amino]pentanedioate heptahydrate.

CHARACTERS

Appearance
White or almost white powder.

Solubility

Freely soluble in water, very slightly soluble in anhydrous ethanol, practically insoluble in methylene chloride.

IDENTIFICATION

Carry out either tests A, C, D, E or tests B, C, D, E.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison pemetrexed sodium heptahydrate CRS.

B. Nuclear magnetic resonance spectrometry (2.2.33).

Preparation 25–50 mg/mL solution in deuterium oxide R.

Comparison Solution of equal concentration of pemetrexed sodium heptahydrate CRS in deuterium oxide R.

Results The 1H NMR spectrum obtained is qualitatively similar to the 1H NMR spectrum obtained with pemetrexed sodium heptahydrate CRS; disregard the peak located at approximately 5.0 ppm for the comparison.

C. It gives reaction (a) of sodium (2.3.1).

D. Enantiomeric purity (see Tests).

E. Water (see Tests).

TESTS

Solution S

Dissolve 0.56 g in carbon dioxide-free water R and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution GY₄ or Y₄ (2.2.2, Method II).

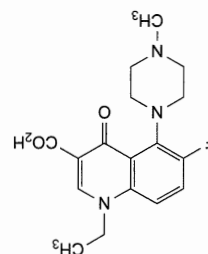
pH (2.2.3)

7.5 to 8.4 for solution S.

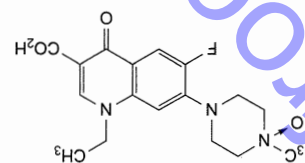
Enantiomeric purity

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or store them at 2–8 °C for not more than 24 h.

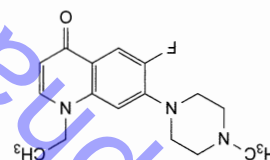
Solution A Dissolve 8 g of β-cyclodextrin R in 900 mL of water for chromatography R. Add 15 mL of triethylamine R then 6 mL of phosphoric acid R and adjust to pH 6.0 with



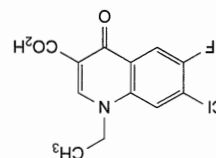
C. 1-ethyl-6-fluoro-5-(4-methylpiperazin-1-yl)-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (isopemfloxacin).



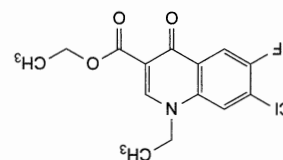
D. 4-(3-carboxy-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinolin-7-yl)-1-methylpiperazine 1-oxide (N-oxide of pemfloxacin).



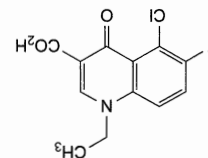
E. 1-ethyl-6-fluoro-7-(4-methylpiperazin-1-yl)quinoline-4(1H)-one (decarboxylated pemfloxacin).



F. 7-chloro-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (N-ethyl acid) (morfloxacin impurity A).



G. ethyl 7-chloro-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylate (N-ethyl ester).



H. 5-chloro-1-ethyl-6-fluoro-4-oxo-1,4-dihydroquinoline-3-carboxylic acid (iso-N-ethyl acid).

Ph Eur

Solution A 1.45 g/L solution of ammonium formate R in water for chromatography R, adjusted to pH 3.5 with anhydrous formic acid R.

Test solution Dissolve 20 mg of the substance to be examined in water for chromatography R and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 12 mg of the substance to be examined in water for chromatography R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 6 mg of pemretexed for system suitability CRS (containing impurity E) in water for chromatography R and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with water for chromatography R. Dilute 3.0 mL of this solution to 10.0 mL with water for chromatography R.

Column: — size: $l = 0.25$ m, $\varnothing = 4.6$ mm; — stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m) with a pore size of 12 nm; — temperature: 40 °C.

Mobile phase acetonitrile R, solution A (5:95 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 50 μ L.

Run time 1.5 times the retention time of pemretexed.

Relative retention With reference to pemretexed (retention time = about 30 min): impurity E = about 0.94.

System suitability:

— symmetry factor: maximum 2.0 for the principal peak in the chromatogram obtained with reference solution (b); — peak-to-valley ratio: minimum 5.0, where H_p = height above the baseline of the peak due to impurity E and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to pemretexed in the chromatogram obtained with reference solution (a).

Calculation of percentage contents:

— for impurity E, use the concentration of pemretexed disodium heptahydrate in reference solution (b).

Limit:

— impurity E: maximum 0.3 per cent.

Column rinse: the following program is given for information only.

Use a gradient column rinse before column storage or after 30 sample injections to avoid build-up on the column. If a drifting baseline is observed, allow additional time for equilibration with the mobile phase. If a blank chromatogram exhibits broad humps, perform a gradient column rinse.

Rinsing solution A water for chromatography R.

Rinsing solution B acetonitrile R1.

Time (min)	Mobile phase (per cent V/V)	Rinsing solution A (per cent V/V)	Rinsing solution B (per cent V/V)
0 - 4	100 \rightarrow 0	0 \rightarrow 50	0 \rightarrow 50
4 - 9	0	50	50
9 - 14	0	50 \rightarrow 10	50 \rightarrow 90
14 - 54	0	10	90
54 - 69	0	10 \rightarrow 95	90 \rightarrow 5
69 - 100	0	95	5

Related substances
Liquid chromatography (2.2.29). Prepare the solutions immediately before use or store them at 2-8 °C for not more than 24 h.

Mobile phase: — mobile phase A: acetonitrile R, solution A (5:95 V/V); — mobile phase B: acetonitrile R, solution A (30:70 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	100	0
3 - 45	100 \rightarrow 0	0 \rightarrow 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram supplied with pemretexed impurity mixture CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and D; use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and C.

Relative retention With reference to pemretexed (retention time = about 26 min): impurity A = about 0.82;

impurity B = about 0.87; impurity C = about 0.88;

impurity D = about 0.90.

System suitability: reference solution (b):

— peak-to-valley ratio: minimum 1.5, where H_p = height above the baseline of the peak due to impurity B and

H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to

impurity C.

Calculation of percentage contents:

— for each impurity, use the concentration of pemretexed disodium heptahydrate in reference solution (a).

Limits:

— impurities A, D: for each impurity, maximum

0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.6 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

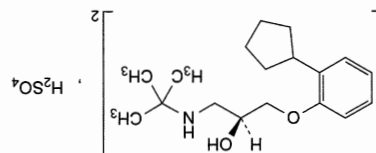
Solvent mixture acetone R, water R (40:60 V/V).

E. (2*R*)-2-[[4-(2-amino-4-oxo-4,7-dihydro-1*H*-pyrrolo[2,3-*d*]pyrimidin-5-yl)ethyl]benzoyl]amino]pentanedioic acid.

Penbutolol Sulfate

Penbutolol Sulphate

(Ph. Eur. monograph 1461)

C₂₀H₂₇N₂O₈S

681

38363-32-5

Action and use

Beta-adrenoceptor antagonist.

Ph. Eur.

DEFINITION

Dl[(2S)-1-(2-cyclopentylphenoxy)-3-[(1,1-dimethylethyl)amino]propan-2-yl] sulfate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, soluble in methanol, practically insoluble in cyclohexane.

IDENTIFICATION

First identification A, C, D

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison penbutolol sulfate CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 40 mg of the substance to be examined in 1 mL of methanol R.

Reference solution Dissolve 40 mg of penbutolol sulfate CRS in 1 mL of methanol R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase glacial acetic acid R, water R, butanol R, ethyl acetate R (10:20:35:35 V/V/V/V).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve 50 mg in a mixture of 5 mL of water R and 1 mL of 0.1 M hydrochloric acid. The solution gives reaction (a) of sulfates (2.3.1).

D. Specific optical rotation (see Tests).

TESTS

Solution S

Dissolve 1.00 g in methanol R and dilute to 20.0 mL with the same solvent.

Acidity or alkalinity

To 4 mL of solution S add 4 mL of carbon dioxide-free water R. Add 0.1 mL of methyl red solution R and 0.2 mL of

0.01 M sodium hydroxide; the solution is yellow. Add 0.4 mL

of 0.01 M hydrochloric acid; the solution is red.

Specific optical rotation (2.2.7)

–23 to –25 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase B, mobile phase A (40:60 V/V).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 4.0 mg of the substance to be examined and 1.0 mg of penbutolol impurity A CRS in 5.0 mL of the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5.0 mg of penbutolol impurity A CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

size: l = 0.25 m, Ø = 4.6 mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase:

mobile phase A: acetonitrile for chromatography R,

methanol R (39:61 V/V);

mobile phase B: dissolve 11 g of sodium heptanesulfonate R in 1000 mL of water R, add 5.0 mL of triethylamine R

and adjust to pH 2.7 with phosphoric acid R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	60	40
15 - 35	60 → 80	40 → 20

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 10 µL.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the 2 principal peaks.

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (d)

(0.5 per cent);

— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— sum of impurities other than A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1 per cent);

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

1.0 g complies with test F. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.500 g in 40 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

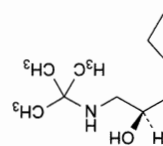
1 mL of 0.1 M perchloric acid is equivalent to 68.10 mg of $C_{12}H_{11}NO_2S$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A

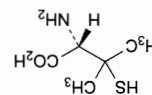


A. (2S)-1-[(2-cyclopentyl-1-enyl)phenoxy]-3-[(1,1-dimethylethyl)amino]propan-2-ol.

Ph Eur

Penicillamine

(Ph. Eur. monograph 0566)



$C_5H_{11}NO_2S$

149.2

52-67-5

Action and use

Disease-modifying antirheumatic drug; chelating agent; treatment of Wilson's disease; heavy metal poisoning; cystinuria.

Preparation

Penicillamine Tablets

Ph Eur

DEFINITION

(2S)-2-Amino-3-methyl-3-sulfanylbutananoic acid.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol

IDENTIFICATION

First identification A, B, D

Second identification A, C, D

A. Dissolve 0.5 g in a mixture of 0.5 mL of hydrochloric acid R and 4 mL of warm acetone R, cool in iced water and initiate crystallisation by scratching the wall of the tube with a glass rod. A white precipitate is formed. Filter with the aid of vacuum, wash with acetone R and dry with suction. A 10 g/L solution of the precipitate is dextrorotatory.

B. Examine the chromatograms obtained in the test for impurity A.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and approximate size to the principal peak in the chromatogram obtained with reference solution (a).

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in 4 mL of water R.

Reference solution Dissolve 10 mg of penicillamine CRS in 4 mL of water R.

Plate TLC silica gel G plate R.

Mobile phase glacial acetic acid R, water R, butanol R (18:18:72 V/V/V).

Application 2 μ L.

Development Over a path of 10 cm.

Drying At 100-105 °C for 5-10 min.

Detection Expose to iodine vapour for 5-10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution.

D. Dissolve 40 mg in 4 mL of water R and add 2 mL of phosphomungstic acid solution R. Allow to stand for 5 min.

A blue colour develops.

TESTS**Solution S**

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

pH (2.2.3)

4.5 to 5.5.

Dilute 1 mL of solution S to 10 mL with carbon dioxide-free water R.

Specific optical rotation (2.2.7)

−61.0 to −65.0 (dried substance).

Dissolve 0.500 g in 1 M sodium hydroxide and dilute to 10.0 mL with the same solvent.

Ultraviolet-absorbing substances

Maximum 0.5 per cent of penillic acid.

Dissolve 0.100 g in water R and dilute to 50.0 mL with the same solvent. The absorbance (2.2.25) of the solution at 268 nm is not greater than 0.07.

Impurity A

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 40 mg of penicillamine CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 20.0 mg of penicillamine disulfide CRS (impurity A) in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm.

— stationary phase: octylsilyl silica gel for chromatography R

(10 µm).

Mobile phase Solution containing 0.1 g/L of sodium edetate R and 2 g/L of methanesulfonic acid R.

Flow rate 1.7 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Relative retention With reference to penicillamine (retention time = about 6 min); impurity A = about 1.8.

System suitability: reference solution (a):

— resolution: minimum 4.0 between the peaks due to

penicillamine and impurity A.

Limit:

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (1 per cent).

Impurity B

Maximum 0.1 ppm.

Carry out all the operations in a penicillin-free atmosphere and with equipment reserved for this test. Sterilise the equipment at 180 °C for 3 h and the buffer solutions at 121 °C for 20 min before use.

Test solution (a) Dissolve 1.000 g in 8 mL of buffer solution pH 2.5 R and add 8 mL of ether R. Shake vigorously for 1 min. Repeat the extraction and combine the ether layers.

Add 8 mL of buffer solution pH 2.5 R. Shake for 1 min, allow to settle and quantitatively separate the upper layer, taking care to eliminate the aqueous phase completely (penicillin is unstable at pH 2.5; carry out operations at this pH within 6–7 min). Add 8 mL of phosphate buffer solution pH 6.0 R2 to the ether phase, shake for 5 min, allow to settle, then separate the aqueous layer and check that the pH is 6.0.

Test solution (b) To 2 mL of test solution (a) add 20 µL of penicillinase solution R and incubate at 37 °C for 1 h.

Reference solution (a) Dissolve 5 mg of benzylpenicillin sodium R in 500 mL of phosphate buffer solution pH 6.0 R2. Dilute 0.25 mL of the solution to 200.0 mL with buffer solution pH 2.5 R. Carry out the extraction using 8 mL of this solution as described for test solution (a).

Reference solution (b) To 2 mL of reference solution (a) add 20 µL of penicillinase solution R and incubate at 37 °C for 1 h.

Blank solution Prepare the solution as described for test solution (a) but omitting the substance to be examined.

Liquify a suitable nutrient medium such as that described below and inoculate it at a suitable temperature with a culture of *Kocuria rhizophila* (ATCC 9341), to give 5×10^4 micro-organisms per millilitre or a different quantity if necessary to obtain the required sensitivity and formation of clearly defined inhibition zones of suitable diameter.

Immediately pour the inoculated medium into 5 Petri dishes 10 cm in diameter to give uniform layers 2–5 mm deep. The medium may alternately consist of 2 layers, only the upper layer being inoculated. Store the dishes so that no appreciable growth or death of the micro-organisms occurs before use and so that the surface of the agar is dry at the time of use. In each dish, place 5 stainless steel hollow cylinders 6 mm in diameter on the surface of the agar evenly spaced on a circle with a radius of about 25 mm and concentric with the dish. For each dish, place in separate cylinders 0.15 mL of test solutions (a) and (b), reference solutions (a) and (b) and the blank solution. Maintain at 30 °C for at least 24 h. Measure the diameters of the inhibition zones with a precision of at least 0.1 mm. The test

is valid if reference solution (a) gives a clear inhibition zone and if reference solution (b) and the blank solution give no inhibition zone. If test solution (a) gives an inhibition zone, this is caused by penicillin if test solution (b) gives no inhibition zone. If this is so, the average diameter of the inhibition zones given by test solution (a) for the 5 Petri dishes is less than the average diameter of the inhibition zones given by reference solution (a) measured in the same conditions.

Nutrient medium (pH 6.0)

Peptone	5 g
Yeast extract	1.5 g
Meat extract	1.5 g
Sodium chloride	3.5 g
Agar	15 g
Distilled water R	1000 mL

Mercury

Maximum 10 ppm.

Test solution To 1.00 g add 10 mL of water R and 0.15 mL of perchloric acid R and swirl until dissolution is complete. Add 1.0 mL of a 10 g/L solution of ammonium pyroindinedithiocarbamate R which has been washed immediately before use 3 times, each time with an equal volume of methyl isobutyl ketone R. Mix and add 2.0 mL of methyl isobutyl ketone R and shake for 1 min. Dilute to 25.0 mL with water R and allow the 2 layers to separate; use the methyl isobutyl ketone layer.

Reference solutions Dissolve a quantity of mercuric oxide R equivalent to 0.108 g of HgO in the smallest necessary volume of dilute hydrochloric acid R and dilute to 100.0 mL with water R (100 ppm Hg). Prepare the reference solutions in the same manner as the test solution but using instead of the substance to be examined suitable volumes of the solution containing 100 ppm of Hg.

Source Mercury hollow-cathode lamp.

Wavelength 254 nm.

Atomisation device Air-acetylene flame.

Set the zero of the instrument using a methyl isobutyl ketone layer obtained as described for the test solution but omitting the substance to be examined.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying over diposphorus pentoxide R at 60 °C at a pressure not exceeding 0.67 kPa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

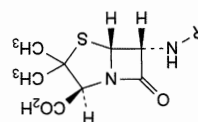
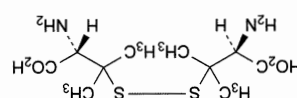
Dissolve 0.1000 g in 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 14.92 mg of $C_5H_{11}NO_2S$.

IMPURITIES

Specified impurities: A, B.

A. 3,3'-(disulfanediyl)bis[(2S)-2-amino-3-methylbutanoic acid (penicillamine disulfide)],

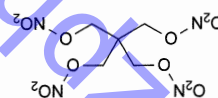


B. penicillin.

Ph Eur

Diluted Pentaerythrityl Tetranitrate

(Ph. Eur. monograph 1335)



78-11-5

Action and use
Vasodilator.

$C_5H_8N_4O_{12}$

Ph Eur

DEFINITION

Dry mixture of 2,2-bis(hydroxymethyl)propane-1,3-diol tetranitrate (pentaerythrityl tetranitrate) and Lactose monohydrate (0187) or Mannitol (0559).

Content

95.0 per cent *m/m* to 105.0 per cent *m/m* of the declared content of pentaerythrityl tetranitrate.

CAUTION: undiluted pentaerythrityl tetranitrate may explode if subjected to percussion or excessive heat. Appropriate precautions must be taken and only very small quantities handled.

CHARACTERS

Appearance of pentaerythrityl tetranitrate

White or slightly yellowish powder.

Solubility of pentaerythrityl tetranitrate

Practically insoluble in water, soluble in acetone, slightly soluble in ethanol (96 per cent).

The solubility of diluted pentaerythrityl tetranitrate depends on the diluent and its concentration.

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Separately shake a quantity of the substance to be examined and a quantity of the reference substance, each

corresponding to 25 mg of pentaerythrityl tetranitrate, with 10 mL of acetone R for 5 min; filter, evaporate to dryness at a temperature below 40 °C, and dry the residue over

diphosphorus pentoxide R at a pressure of 0.7 kPa for 16 h.

Examine the residues prepared as discs.

Comparison diluted pentaerythrityl tetranitrate CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Shake a quantity of the substance to be examined corresponding to 10 mg of pentaerythrityl

tetranitrate with 10 mL of ethanol (96 per cent) R for 5 min

and filter.

Reference solution Shake a quantity of diluted pentaerythrityl

tetranitrate CRS corresponding to 10 mg of pentaerythrityl tetranitrate with 10 mL of ethanol (96 per cent) R for 5 min

and filter.

Plate TLC silica gel plate R.

Mobile phase ethyl acetate R, toluene R (20:80 V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with freshly prepared potassium iodide and starch solution R, expose to ultraviolet light at 254 nm for

15 min and examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the

reference solution.

C. Thin-layer chromatography (2.2.27).

Test solution Shake a quantity of the substance to be examined corresponding to 0.10 g of lactose or mannitol

with 10 mL of water R. Filter if necessary.

Reference solution (a) Dissolve 0.10 g of lactose R in water R

and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 0.10 g of mannitol R in water R

and dilute to 10 mL with the same solvent.

Reference solution (c) Mix equal volumes of reference

solutions (a) and (b).

Plate TLC silica gel G plate R.

Mobile phase water R, methanol R, anhydrous acetic acid R,

ethylene chloride R (10:15:25:50 V/V/V/V). Measure the

volumes accurately since a slight excess of water produces

cloudiness.

Application 1 µL; thoroughly dry the points of application.

Development A Over 2/3 of the plate.

Drying A In a current of warm air.

Development B Immediately, over 2/3 of the plate, after

renewing the mobile phase.

Drying B In a current of warm air.

Detection Spray with 4-aminobenzoic acid solution R, dry in a

current of cold air until the acetone is removed, then heat at

100 °C for 15 min; allow to cool, spray with a 2 g/L solution

of sodium periodate R, dry in a current of cold air and heat at

100 °C for 15 min.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

solution (a) for lactose or to the principal spot in the

chromatogram obtained with reference solution (b) for

mannitol.

TESTS

Impurity A

Thin-layer chromatography (2.2.27).

Test solution Shake a quantity of the substance to be

examined corresponding to 0.10 g of pentaerythrityl

tetranitrate with 5 mL of ethanol (96 per cent) R and filter.

Reference solution Dissolve 10 mg of potassium nitrate R in

1 mL of water R and dilute to 100 mL with ethanol

(96 per cent) R.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, acetone R, toluene R

(15:30:60 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of air until the acetic acid is completely removed.

Detection Spray copiously with freshly prepared potassium

iodide and starch solution R, expose the plate to ultraviolet

light at 254 nm for 15 min and examine in daylight.

Limit:

— nitrate: any spot due to nitrate is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent, calculated as potassium nitrate).

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Sonicate for 15 min a quantity of the substance to be examined corresponding to 25.0 mg of pentaerythrityl tetranitrate in 20 mL of the mobile phase and dilute to 25.0 mL with the mobile phase. Filter.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the mobile phase.

Reference solution (a) Sonicate for 15 min a quantity of diluted pentaerythrityl tetranitrate CRS corresponding to 25.0 mg of pentaerythrityl tetranitrate in 20 mL of the mobile phase and dilute to 25.0 mL with the mobile phase. Filter.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 0.3 mL of reference solution (b) to 10.0 mL with the mobile phase.

Reference solution (d) Dilute 200 µL of glyceryl trinitrate solution CRS to 25.0 mL with the mobile phase.

Reference solution (e) To 1 mL of reference solution (b) add 1 mL of reference solution (d) and dilute to 10 mL with the mobile phase.

Reference solution (f) Dilute 1.0 mL of reference solution (a) to 20.0 mL with the mobile phase. Dilute 0.5 mL of this solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 µm).

Mobile phase water R, acetonitrile R (35:65 V/V).

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of test solution (a) and reference solutions (c), (e) and (f).

Run time 5 times the retention time of pentaerythrityl tetranitrate.

Relative retention With reference to pentaerythrityl tetranitrate (retention time = about 2.4 min): impurity B = about 0.7;

impurity C = about 3.0.

System suitability: reference solution (e):

— resolution: minimum 3.0 between the peaks due to glyceryl trinitrate and pentaerythrityl tetranitrate.

Limits:

— impurities C, D: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (f) (0.10 per cent);

— unspecified impurities: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (f) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.6 per cent);

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (f) (0.05 per cent).

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (b).

Calculate the percentage content of $C_5H_8N_4O_{12}$ from the declared content of diluted pentaerythrityl tetranitrate CRS.

STORAGE

Protected from light and heat.

LABELLING

The label states:

— the percentage content of pentaerythrityl tetranitrate;

— the diluent used.

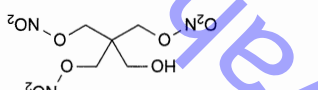
IMPURITIES

Specified impurities A, C, D

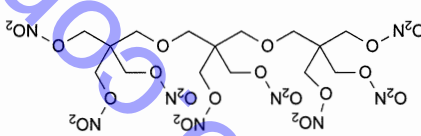
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B.



A. NO_3^- : inorganic nitrates,

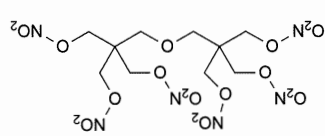


B. 2,2-bis(hydroxymethyl)propane-1,3-diol trinitrate (pentaerythritol trinitrate),



C. 2,2-bis[[3-hydroxy-2,2-bis(hydroxymethyl)propoxy]methyl]propane-1,3-diol

octanitrate (tripentaerythrityl octanitrate),



D. 2,2'-(oxybis(methylene)bis[2-(hydroxymethyl)propane-1,3-diol] hexanitrate (dipentaerythrityl hexanitrate).

Pentagastin

Boc-βAla-L-Trp-Met-Asp-Phe

Boc-tert-butyloxycarbonyl

C₃₇H₄₉N₇O₉S

767.9

5534-95-2

Action and use
Gastrin analogue.

Preparation

Pentagastin Injection

DEFINITION

Pentagastin is a pentapeptide that stimulates gastric secretion of acid. It contains not less than 97.0% and not more than 103.0% of C₃₇H₄₉N₇O₉S, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder.

Practically insoluble in water; soluble in 5M ammonia and in dimethylformamide; slightly soluble in ethanol (96%).

IDENTIFICATION

A. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.010% w/v solution in 0.01M ammonia exhibits maxima at 280 nm and 288 nm and an inflection at 275 nm.

B. Carry out the method for thin-layer chromatography, Appendix III A, using silica gel G as the coating substance on each of three plates and the following solvent systems as the mobile phases: (A) a mixture of 75 volumes of butan-2-ol and 25 volumes of a 3% v/v solution of 13.5M ammonia, (B) the upper layer produced by shaking together 50 volumes of water, 40 volumes of butan-1-ol and 10 volumes of glacial acetic acid and allowing to separate and (C) a mixture of 50 volumes of glacial acetic acid, 25 volumes of ether and 25 volumes of water. Apply separately to each plate 2 µL of each of two solutions in 0.01M ammonia containing (1) 0.5% w/v of the substance being examined and (2) pentagastin BPCRS containing 0.5% w/v of pentagastin.

After removal of the plates, allow them to dry in air, heat at 100° for 2 minutes, spray with a 1.0% w/v solution of 4-dimethylaminobenzaldehyde in a mixture of 3 volumes of methanol and 1 volume of hydrochloric acid and heat at 100° until purple spots are produced (about 2 minutes). Examine by transmitted light. On each plate, the principal spot in the chromatogram obtained with solution (1) corresponds to that in the chromatogram obtained with solution (2).

Light absorption

Ratio of the absorbance of a 0.010% w/v solution in 0.01M ammonia at the maximum at 280 nm to that at the maximum at 288 nm, 1.12 to 1.22, Appendix II B.

Specific optical rotation

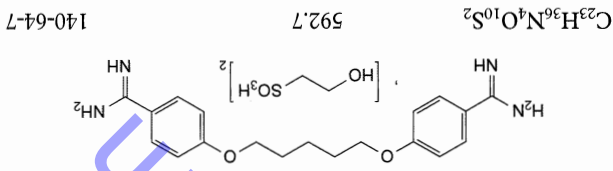
In a 1% w/v solution in dimethylformamide, -25.0 to -29.0, calculated with reference to the dried substance,

Amino acids

β-Alanine 11.0 to 12.2%, aspartic acid 16.4 to 18.2%, methionine 18.4 to 20.4%, phenylalanine, 20.4 to 22.6%, when determined by the following method. Examine using an amino acid analyser according to the manufacturer's instructions. Standardise the apparatus with 1 mg of an equimolar mixture of β-alanine, aspartic acid, methionine and phenylalanine. Place 1 mg of the substance being

Pentamidine Isetonate

(Pentamidine Disulfonate, Ph Eur monograph 1137)



DEFINITION

4,4'-[Pentane-1,5-diylbis(oxy)]dibenzamidine di(2-hydroxyethanesulfonate).

Ph Eur

Pentamidine Injection

Preparation

Antiprotocol.

Action and use

140-64-7

STORAGE

Pentagastin should be protected from light. Dissolve 5 mg in sufficient 0.01M ammonia to produce 100 mL and measure the absorbance at the maximum at 280 nm, Appendix II B. Calculate the content of C₃₇H₄₉N₇O₉S taking 70.0 as the value of A(1%, 1 cm) at the maximum at 280 nm.

ASSAY

When dried over phosphorus pentoxide at a pressure not exceeding 0.7 kPa for 24 hours, loses not more than 0.5% of its weight. Use 1 g.

Loss on drying

chromatogram obtained with solution (2). Any secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2). Produced (about 2 minutes). Examine by transmitted light. hydrochloric acid and heat at 100° until purple spots are mixture of 3 volumes of methanol and 1 volume of 1.0% w/v solution of 4-dimethylaminobenzaldehyde in a mixture of 3 volumes of water, 40 volumes of butan-1-ol and 10 volumes of glacial acetic acid and allowing to separate and (C) a mixture of 50 volumes of glacial acetic acid, 25 volumes of ether and 25 volumes of water. Apply separately to the plate 10 µL of each of two solutions of the substance being examined in a mixture of 24 volumes of methanol and 1 volume of 13.5M ammonia containing (1) 0.50% w/v and (2) 0.010% w/v. After removal of the plate, allow it to dry in air, heat at 100° for 2 minutes, spray with a

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using silica gel G as the coating substance and a mixture of 100 volumes of ether, 20 volumes of glacial acetic acid and 10 volumes of water as the mobile phase. Apply separately to the plate 10 µL of each of two solutions of the substance being examined in a mixture of 24 volumes of methanol and 1 volume of 13.5M ammonia containing (1) 0.50% w/v and (2) 0.010% w/v. After removal of the plate, allow it to dry in air, heat at 100° for 2 minutes, spray with a mixture of 3 volumes of methanol and 1 volume of 1.0% w/v solution of 4-dimethylaminobenzaldehyde in a mixture of 3 volumes of water, 40 volumes of butan-1-ol and 10 volumes of glacial acetic acid and allowing to separate and (C) a mixture of 50 volumes of glacial acetic acid, 25 volumes of ether and 25 volumes of water. Apply separately to the plate 10 µL of each of two solutions of the substance being examined in a mixture of 24 volumes of methanol and 1 volume of 13.5M ammonia containing (1) 0.50% w/v and (2) 0.010% w/v. After removal of the plate, allow it to dry in air, heat at 100° for 2 minutes, spray with a

Content

98.5 per cent to 101.5 per cent (dried substance).

PRODUCTION

The production method must be evaluated to determine the potential for formation of alkyl 2-hydroxyethanesulfonates, which is particularly likely to occur if the reaction medium contains lower alcohols. Where necessary, the production method is validated to demonstrate that alkyl 2-hydroxyethanesulfonates are not detectable in the final product.

CHARACTERS**Appearance**

White or almost white powder or colourless crystals,

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison pentamidine diseminate CRS.

B. Dissolve about 40 mg in 5 mL of water R and add dropwise with shaking 1 mL of a 10 g/L solution of sodium chloride R. Allow to stand for 5 min. The mixture remains clear.

C. Treat 0.15 g by the oxygen-flask method (2.5.10). Use 10 mL of dilute hydrogen peroxide solution R to absorb the combustion products. The solution gives reaction (a) of sulfates (2.3.1).

TESTS**Appearance of solution**

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent.

pH (2.2.3)

4.5 to 6.5.

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) To 0.1 g in a conical flask, add 40 mL of water R and glass beads. Adjust to pH 10.5 with dilute sodium hydroxide solution R and boil under reflux for 20 min. Cool and dilute to 50 mL with water R. Dilute 1 mL of the solution to 50 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 65 volumes of methanol R and 35 volumes of a 30 g/L solution of ammonium acetate R previously adjusted to pH 7.5 using triethylamine R.

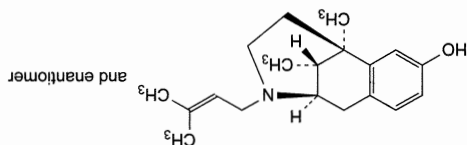
Flow rate 1 mL/min.

Preparation

Opioid receptor agonist; analgesic.

Action and use

C₁₉H₂₇NO 285.4 359-83-1



(Ph. Eur. monograph 1462)

Pentazocine

A. 4-[[5-(4-amidinophenoxy)pentyl]oxy]benzenecarboxamide.

**IMPURITIES**

In an airtight container.

STORAGE

29.63 mg of C₂₃H₃₆N₄O₁₀S₂.

1 mL of 0.1 M tetraabutylammonium hydroxide is equivalent to blank titration.

until the colour of the indicator changes to blue. Carry out a tetraabutylammonium hydroxide, under a current of nitrogen R, Add 0.25 mL of thymol blue solution R. Titrate with 0.1 M Dissolve 0.250 g in 50 mL of dimethylformamide R.

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 4.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

using 2 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test C. Prepare the reference solution

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.02 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.1 times the area of the principal peak in

(0.4 per cent),

the chromatogram obtained with reference solution (a)

— total: not more than twice the area of the principal peak in

reference solution (a) (0.2 per cent),

the principal peak in the chromatogram obtained with

— any impurity: for each impurity, not more than the area of

the chromatogram obtained shows 2 principal peaks,

— resolution: minimum 2.0 between the 2 principal peaks.

System suitability: reference solution (b):

Run time 3.5 times the retention time of pentamidine.

Injection 10 μ L.

Detection Spectrophotometer at 265 nm.

1 mL of 0.1 M perchloric acid is equivalent to 28.54 mg of $C_{19}H_{27}NO$.

STORAGE

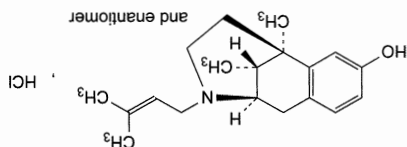
Store protected from light.

Ph Eur



Pentazocine Hydrochloride

(Ph. Eur. monograph 1463)



$C_{19}H_{28}ClNO$ 321.9 64024-15-3

Action and use

Opioid receptor agonist; analgesic.

Preparations

Pentazocine Capsules

Pentazocine Tablets

Ph Eur

DEFINITION

Pentazocine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (2RS,6RS,11RS)-6,11-dimethyl-3-(3-methylbut-2-enyl)-1,2,3,4,5,6-hexahydro-2,6-methano-3-benzazocin-8-ol hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white powder, sparingly soluble in water, soluble in ethanol (96 per cent) and sparingly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the Ph. Eur. reference spectrum of pentazocine hydrochloride.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

pH (2.2.3)

Dissolve 0.1 g in 10 mL of carbon dioxide-free water R.

The pH of the solution is 4.0 to 6.0.

Absorbance (2.2.25)

Dissolve 0.100 g in a mixture of 20 mL of water R and 1 mL of 1 M hydrochloric acid, and dilute to 100.0 mL with water R. To 10.0 mL add 1 mL of 1 M hydrochloric acid and dilute to 100.0 mL with water R. The absorbance at the absorption maximum at 278 nm is 0.59 to 0.63, calculated with reference to the dried substance.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel F_{254} plate R.

Test solution Dissolve 0.20 g in 3 mL of methanol R and dilute to 10 mL with methylene chloride R.

Reference solution (a) Dilute 1 mL of the test solution to 100 mL with methylene chloride R.

Reference solution (b) Dilute 5 mL of reference solution (a) to 10 mL with methylene chloride R.

DEFINITION

Pentazocine contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (2RS,6RS,11RS)-6,11-dimethyl-3-(3-methylbut-2-enyl)-1,2,3,4,5,6-hexahydro-2,6-methano-3-benzazocin-8-ol, calculated with reference to the dried substance.

CHARACTERS

A white or almost white powder, practically insoluble in water, freely soluble in methylene chloride and soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

Examine by infrared absorption spectrophotometry (2.2.24), comparing with the Ph. Eur. reference spectrum for pentazocine (form A).

TESTS

Absorbance (2.2.25)
Dissolve 0.100 g in a mixture of 20 mL of water R and 1 mL of 1 M hydrochloric acid, and dilute to 100.0 mL with water R. To 10.0 mL add 1 mL of 1 M hydrochloric acid and dilute to 100.0 mL with water R. The absorbance at the absorption maximum at 278 nm is 0.67 to 0.71, calculated with reference to the dried substance.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel F_{254} plate R.
Test solution Dissolve 0.20 g of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.
Reference solution (a) Dilute 1 mL of the test solution to 100 mL with methylene chloride R.
Reference solution (b) Dilute 5 mL of reference solution (a) to 10 mL with methylene chloride R.
Reference solution (c) Dilute 5 mL of reference solution (a) to 20 mL with methylene chloride R.

Apply to the plate 10 μ L of each solution. Develop over a path corresponding to two thirds of the plate height using a mixture of 3 volumes of isopropylamine R, 3 volumes of methanol R and 94 volumes of methylene chloride R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Heat the plate at 100–105 °C for 15 min, allow to cool, expose to iodine vapour and re-examine under ultraviolet light at 254 nm. By each method of visualisation: any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot obtained with reference solution (a) (1 per cent); not more than one such spot is more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent) and not more than 4 such spots are more intense than the spot in the chromatogram obtained with reference solution (c) (0.25 per cent).

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying at 60 °C at a pressure not exceeding 0.7 kPa for 4 h.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

Solubility
Sparingly soluble in water, freely soluble in methanol, slightly soluble in methylene chloride.

IDENTIFICATION
Infrared absorption spectrophotometry (2.2.4).
Comparison Ph. Eur. reference spectrum of pentazocine lactate.

TESTS
pH (2.2.3)
5.5 to 6.5.

Absorbance (2.2.25)
Dissolve 0.1 g in 10 mL of carbon dioxide-free water R.

0.50 to 0.54, determined at the absorption maximum at 278 nm.

Dissolve 0.10 g in 10.0 mL of 1 M hydrochloric acid and dilute to 100.0 mL with water R. Dilute 10.0 mL of the solution to 100.0 mL with water R.

Related substances

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.20 g of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 100 mg of the substance to be examined in acetic anhydride R and dilute to 5 mL with the same solvent. Heat at 80 °C for 10 min. Dilute 1 mL of the solution to 10 mL with methanol R. Mix 1 mL of this solution with 1 mL of the test solution.

Reference solution (b) Dilute 1 mL of the test solution to 100 mL with methylene chloride R. Dilute 2 mL of this solution to 10 mL with methylene chloride R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase isopropylamine R, methanol R, methylene chloride R (3:3:94 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm. Heat at 100-105 °C for 15 min. Allow to cool. Expose to iodine vapour and re-examine in ultraviolet light at 254 nm.

System suitability: reference solution (a):

— the chromatogram shows 2 clearly separated spots.

Limits By each method of detection:

— any impurity: any spots, apart from the principal spot, are not more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 30 mL of anhydrous acetic acid R and add 30 mL of dioxan R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 37.55 mg of C₂₂H₃₃NO₄.

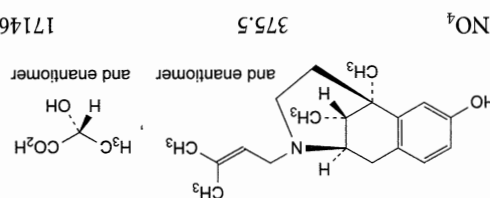
STORAGE

Protected from light.

Ph Eur

Pentazocine Lactate

(Ph. Eur. monograph 2000)



Ph Eur

Store protected from light.

STORAGE

C₁₉H₂₈ClNO.

1 mL of 0.1 M sodium hydroxide is equivalent to 32.19 mg of infection.

hydroxide. Read the volume added between the 2 points of

potentiometric titration (2.2.20), using 0.1 M sodium

Add 5 mL of 0.01 M hydrochloric acid. Carry out a

Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R.

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

drying at 60 °C at a pressure not exceeding 0.7 kPa for 4 h.

Not more than 0.5 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

(0.25 per cent).

chromatogram obtained with reference solution (c)

more than 4 such spots are more intense than the spot in the

obtained with reference solution (b) (0.5 per cent); and not

such spot is more intense than the spot in the chromatogram

with reference solution (a) (1 per cent); not more than 1

the principal spot, is not more intense than the spot obtained

the chromatogram obtained with the test solution, apart from

light at 254 nm. By each method of visualisation: any spot in

expose to iodine vapour and re-examine under ultraviolet

Heat the plate at 100-105 °C for 15 min, allow to cool,

plate to dry in air and examine in ultraviolet light at 254 nm.

mixture of 3 volumes of isopropylamine R, 3 volumes of

methanol R and 94 volumes of methylene chloride R. Allow the

path corresponding to two-thirds of the plate height using a

Apply to the plate 10 µL of each solution. Develop over a

20 mL with methylene chloride R.

Reference solution (c) Dilute 5 mL of reference solution (a) to

20 mL with methylene chloride R.

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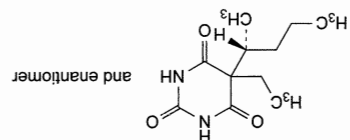
20 mL with methylene chloride R.

Reference solution (c) Dilute 5 mL of reference solution (a) to

20 mL with methylene chloride R.

Pentobarbital

(Ph. Eur. monograph 0200)



$C_{11}H_{18}N_2O_3$ 226.3

76-74-4

Action and use

Barbiturate.

Ph Eur

DEFINITION

Pentobarbital contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 5-ethyl-5-[(1*R*,5*S*)-1-methylbutyl]pyrimidine-2,4,6-trione, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, very slightly soluble in water, freely soluble in ethanol. It forms water-soluble compounds with alkali hydroxides and carbonates and with ammonia.

IDENTIFICATION

A. Determine the melting point (2.2.14) of the substance to be examined. Mix equal parts of the substance to be examined and *pentobarbital CRS* and determine the melting point of the mixture. The difference between the melting points (which are about 133 °C) is not greater than 2 °C.

B. Examine by thin-layer chromatography (2.2.27), using silica gel *GF₂₅₄* R as the coating substance.

Test solution Dissolve 0.1 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent.

Reference solution Dissolve 0.1 g of *pentobarbital CRS* in *alcohol R* and dilute to 100 mL with the same solvent.

Apply to the plate 10 µL of each solution. Develop over a path of 18 cm using the lower layer of a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine immediately in ultraviolet light at 254 nm. The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. To about 10 mg add about 10 mg of *vanillin R* and 2 mL of *sulfuric acid R*. Mix and heat on a water-bath for 2 min. A reddish-brown colour develops. Cool and add cautiously 5 mL of *ethanol R*. The colour becomes violet and then blue.

TESTS

Appearance of solution

Dissolve 1.0 g in a mixture of 4 mL of *dilute sodium hydroxide solution R* and 6 mL of *water R*. The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Acidity

Boil 1.0 g with 50 mL of *water R* for 2 min, allow to cool and filter. To 10 mL of the filtrate add 0.15 mL of *methyl red solution R*. The solution is orange-yellow. Not more than 0.1 mL of 0.1 M *sodium hydroxide* is required to produce a pure yellow colour.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel *GF₂₅₄* R as the coating substance.

Pentobarbital Tablets

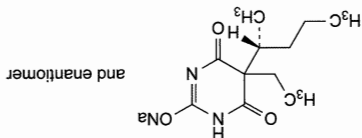
Preparation

Barbiturate.

Action and use

$C_{11}H_{17}N_2NaO_3$ 248.3

57-33-0



(Ph. Eur. monograph 0419)

Pentobarbital Sodium



Ph Eur

11.31 mg of $C_{11}H_{18}N_2O_3$.
1 mL of 0.1 M *ethanolic sodium hydroxide* is equivalent to titration.

Dissolve 0.100 g in 5 mL of *pyridine R*. Add 0.5 mL of *thymolphthalein solution R* and 10 mL of *silver nitrate solution in pyridine R*. Titrate with 0.1 M *ethanolic sodium hydroxide* until a pure blue colour is obtained. Carry out a blank titration.

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

Dissolve 0.3 g in 5 mL of a 50 g/L solution of *anhydrous sodium carbonate R*, heating slightly if necessary. Add a solution of 0.3 g of *nitrobenzyl chloride R* in 10 mL of *alcohol R* and heat under a reflux condenser for 30 min. Cool to 25 °C, filter and wash the precipitate with five quantities, each of 5 mL, of *water R*. In a small flask, heat the precipitate with 25 mL of *alcohol R* under a reflux condenser until dissolved (about 10 min). Cool to 25 °C, if necessary scratching the wall of the flask with a glass rod to induce crystallisation, and filter. The precipitate, washed with two quantities, each of 5 mL, of *water R* and dried at 100 °C to 105 °C for 30 min, melts (2.2.14) at 136 °C to 148 °C.

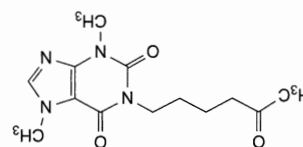
Isomer

(0.5 per cent).

Test solution Dissolve 1.0 g of the substance to be examined in *alcohol R* and dilute to 100 mL with the same solvent. *Reference solution* Dilute 0.5 mL of the test solution to 100 mL with *alcohol R*. Apply to the plate 20 µL of each solution. Develop over a path of 15 cm using the lower layer of a mixture of 5 volumes of *concentrated ammonia R*, 15 volumes of *alcohol R* and 80 volumes of *chloroform R*. Examine immediately in ultraviolet light at 254 nm. Spray with *diphenylcarbazone mercuric reagent R*. Allow the plate to dry in air and spray with freshly prepared *alkaline potassium dichromate solution R* diluted 1 in 5 with *aldehyde-free alcohol R*. Heat at 100 °C to 105 °C for 5 min and examine immediately. When examined in ultraviolet light and after spraying, any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution.

Pentoxifylline

(Ph. Eur. monograph 0851)

C₁₃H₁₈N₄O₃ 278.3 6493-05-6

Action and use

Vasodilator.

DEFINITION

3,7-Dimethyl-1-(5-oxohexyl)-3,7-dihydro-1H-purine-2,6-dione.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, freely soluble in methylene chloride, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Melting point (2.2.14): 103 °C to 107 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison pentoxifylline CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of pentoxifylline CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase methanol R, ethyl acetate R (15:85 V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives the reaction of xanthines (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

A 40 per cent V/V solution of solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Acidity

To 8 mL of solution S add 12 mL of carbon dioxide-free water R and 0.05 mL of bromothymol blue solution R1.

The solution is green or yellow. Not more than 0.2 mL of

0.01 M sodium hydroxide is required to change the colour of the indicator to blue.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture A mixture of equal volumes of a 5.44 g/L solution of potassium dihydrogen phosphate R and methanol R.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 10.0 mL of reference solution (a) to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of caffeine R (impurity F) and 2 mg of theophylline R (impurity C) in the solvent mixture, add 1 mL of the test solution and dilute to 10 mL with the solvent mixture.

Reference solution (d) Dissolve 5.0 mg of caffeine R (impurity F), 5.0 mg of theobromine R (impurity A) and 5.0 mg of theophylline R (impurity C) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute

1.0 mL to 25.0 mL with the solvent mixture.

Column: — temperature: 30 °C.

— chromatography R (5 µm),

— stationary phase: base-deactivated octylsilyl silica gel for

phosphate R;

70 volumes of a 5.44 g/L solution of potassium dihydrogen

phosphate R;

mobile phase B: mix 30 volumes of a 5.44 g/L solution of

potassium dihydrogen phosphate R and 70 volumes of

methanol R;

Time

(min)

(per cent V/V)

Mobile phase A

Mobile phase B

(per cent V/V)

Flow rate 1 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 10 µL.

Relative retention With reference to pentoxifylline (retention time = about 12 min): impurity A = about 0.3;

impurity C = about 0.4; impurity F = about 0.5;

impurity J = about 1.6.

System suitability: reference solution (c):

— retention time: impurity F = 4 min to 7 min;

pentoxifylline = 9 min to 13 min; if necessary adapt the

mixing ratio of the mobile phases;

— resolution: minimum 4 between the peaks due to

impurity C and impurity F.

Limits:

— impurities A, C, F: for each impurity, not more than the

area of the corresponding peak in the chromatogram

obtained with reference solution (d) (0.1 per cent),

- *impurity f*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
- *disregard limit*: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.02 per cent).

Chlorides (2.4.4)

Maximum 100 ppm.

Place 20 mL of solution S in a separating funnel and shake with 2 quantities, each of 20 mL, of 2-methylpropanol R. Dilute 10 mL of the aqueous layer to 15 mL with water R.

Sulfates (2.4.13)

Maximum 200 ppm, determined on 15 mL of solution S.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with limit test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying over diphosphorus pentoxide R at 60 °C at a pressure not exceeding 700 Pa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g. Dissolve 0.200 g in 5 mL of anhydrous acetic acid R. Add 20 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 27.83 mg of $C_{13}H_{18}N_4O_3$.

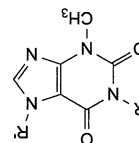
STORAGE

Protected from light.

IMPURITIES

Specified impurities A, C, F, J

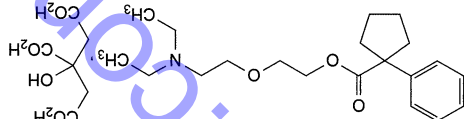
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, D, E, G, H, I, K.



- A. R = H, R' = CH₃: theobromine,
 B. R = R' = H: 3-methyl-3,7-dihydro-1H-purine-2,6-dione,
 C. R = CH₃, R' = H: theophylline,
 D. R = CH₂-CH₂-OH, R' = CH₃:
 1-(3-hydroxypropyl)-3,7-dimethyl-3,7-dihydro-1H-purine-2,6-dione,
 F. R = R' = CH₃: caffeine,

Pentoxifyverine Citrate

(Pentoxifyverine Hydrogen Citrate, Ph Eur monograph 1621)

C₂₆H₃₉NO₁₀

525.6

23142-01-0

Action and use
Cough suppressant.**DEFINITION**

2-[2-(Diethylamino)ethoxy]ethyl 1-phenylcyclopentanecarboxylate dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content

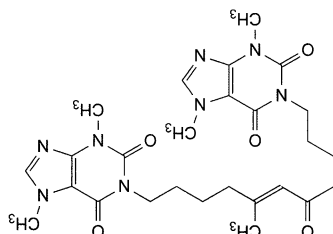
98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

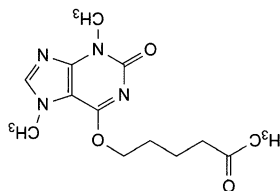
White or almost white, crystalline powder.



Ph Eur

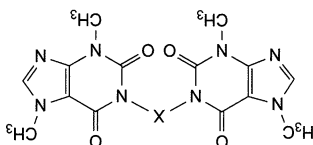


G. 3,7-dimethyl-6-(5-oxohexyloxy)-3,7-dihydro-2H-purin-2-one,



K. X = CH₂-CH₂-CH₂: 1,1'-(propane-1,3-diyl)bis(3,7-dimethyl-3,7-dihydro-1H-purine-2,6-dione),

E. X = CH₂: 1,1'-methylenebis(3,7-dimethyl-3,7-dihydro-1H-purine-2,6-dione),



I. R = CH₂-C₆H₅, R' = CH₃: 1-benzyl-3,7-dimethyl-3,7-dihydro-1H-purine-2,6-dione,
 bis(5-oxohexyl)-3,7-dihydro-1H-purine-2,6-dione,
 H. R = R' = CH₂-[CH₂]₃-CO-CH₃: 3-methyl-1,7-

Solubility

Freely soluble in water, very soluble in glacial acetic acid, freely soluble in methanol, soluble in alcohol and in methylene chloride.

mp

About 93 °C.

IDENTIFICATION

- A. Infrared absorption spectrophotometry (2.2.24).
Comparison Ph. Eur. reference spectrum of pentoxifyverine hydrate citrate.
- B. Dissolve 0.25 g in 5 mL of water R. The solution gives the reaction of citrates (2.3.1).

TESTS

Solution S
Dissolve 5.0 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

pH (2.2.3)

3.3 to 3.7 for solution S.

Related substances

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution Introduce 5.0 mg of pentoxifyverine impurity A CRS and 5.0 mg of pentoxifyverine impurity B CRS in a conical flask, add 5.0 mL of the test solution and dilute to 100.0 mL with the mobile phase. Dilute 3.0 mL of the solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm,
— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m) with a pore size of 10 nm and a carbon loading of 12 per cent,
— temperature: 50 °C.

Mobile phase Mix 35 volumes of acetonitrile R and 65 volumes of a 1.5 g/L solution of sodium heptanesulfonate R adjusted to pH 3.0 with dilute sulfuric acid R.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L.

Run time 3 times the retention time of pentoxifyverine.

Relative retention With reference to pentoxifyverine (retention time = about 6 min): impurity B = about 0.8;

impurity A = about 1.5.

System suitability: reference solution:

— **resolution:** minimum of 5.0 between the peaks due to

pentoxifyverine and to impurity A,

— **signal-to-noise ratio:** minimum of 100 for the peak due to

pentoxifyverine,

— **symmetry factor:** maximum of 2.0 for the peak due to

pentoxifyverine.

Limits:

— **impurity A:** not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.3 per cent),

— **impurity B:** not more than the area of the corresponding peak in the chromatogram obtained with the reference

solution (0.3 per cent),

(96 per cent).

Soluble in water, practically insoluble in ethanol

Solubility

hygroscopic.

White or slightly yellow, crystalline or amorphous powder.

Appearance**CHARACTERS**

consumption.

The animals from which pepsin powder is derived must fulfil the requirements for the health of animals suitable for human

PRODUCTION

Activity Not less than 0.5 Ph. Eur. U./mg (dried substance).

(pH 1 to 5).

Powder prepared from the gastric mucosa of pigs, cattle or sheep. It contains gastric proteinases, active in acid medium

DEFINITION

Ph Eur

Action and use
Proteolytic enzyme.

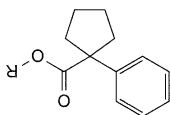
9001-75-6

(Pepsin Powder, Ph Eur monograph 0682)

Pepsin

Ph Eur

A. R = H; 1-phenylcyclopentanecarboxylic acid,
B. R = $\text{CH}_2\text{-CH}_2\text{-N}(\text{CH}_2\text{-CH}_3)_2$; 2-(diethylamino)ethyl 1-phenylcyclopentanecarboxylate (caramiphen).

**IMPURITIES**

Protected from light.

STORAGE

$\text{C}_{26}\text{H}_{39}\text{NO}_{10}$

1 mL of 0.1 M perchloric acid is equivalent to 52.56 mg of

potentiometrically (2.2.20).

with 0.1 M perchloric acid, determining the end-point

Dissolve 0.400 g in 70 mL of anhydrous acetic acid R. Titrate

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

in vacuo at 60 °C for 4 h.

Maximum 0.5 per cent, determined on 1.000 g by drying

Loss on drying (2.2.32)

with a retention time less than or equal to 2.5 min.

reference solution (0.03 per cent); disregard any peak

pentoxifyverine in the chromatogram obtained with the

— **disregard limit:** 0.1 times the area of the peak due to

with the reference solution (0.3 per cent),

peak due to pentoxifyverine in the chromatogram obtained

— **total of any other impurity:** not more than the area of the

obtained with the reference solution (0.1 per cent),

the peak due to pentoxifyverine in the chromatogram

— **any other impurity:** not more than one-third of the area of

Table 0682.-I

Tubes									
	B	T _p	T	S _{3b}	S ₃	S _{2b}	S ₂	S _{1b}	S ₁
Dilute hydrochloric acid R2 (mL)	0.5	0.5	0.25		0.25	0.25	0.25	0.5	0.5
Reference solution (mL)	0.5	0.5	0.75	1.0	1.0	0.75	0.75	0.5	0.5
Test solution (mL)			0.75						
Trichloroacetic acid solution R (mL)				10.0	10.0	10.0	10.0	10.0	10.0
Mix				+	+	+	+	+	+
Water bath at 25 °C				+	+	+	+	+	+
Haemoglobin solution R (mL)				5.0	5.0	5.0	5.0	5.0	5.0
Mix				+	+	+	+	+	+
Haemoglobin solution R (mL)				5.0	5.0	5.0	5.0	5.0	5.0
Mix				+	+	+	+	+	+
Haemoglobin solution R (mL)				5.0	5.0	5.0	5.0	5.0	5.0
Mix				+	+	+	+	+	+
Water bath at 25 °C, 10 min				+	+	+	+	+	+
Trichloroacetic acid solution R (mL)				10.0	10.0	10.0	10.0	10.0	10.0
Mix				+	+	+	+	+	+
Trichloroacetic acid solution R (mL)				10.0	10.0	10.0	10.0	10.0	10.0
Filter				+	+	+	+	+	+

The solution in water may be slightly opalescent with a weak acidic reaction.

IDENTIFICATION

In a mortar, pound 30 mg of fibrin blue R. Suspend in 20 mL of dilute hydrochloric acid R2. Filter the suspension on a filter paper and wash with dilute hydrochloric acid R2 until a

colourless filtrate is obtained. Perforate the filter paper and wash the fibrin blue R through it into a conical flask using 20 mL of dilute hydrochloric acid R2. Shake before use.

Dissolve a quantity of the substance to be examined, equivalent to not less than 20 Ph. Eur. U, in 2 mL of dilute hydrochloric acid R2 and adjust to pH 1.6 ± 0.1. Add 1 mL of this solution to a test-tube containing 4 mL of the fibrin

blue suspension, mix and place in a water-bath at 25 °C with gentle shaking. Prepare a blank solution at the same time and in the same manner using 1 mL of water R. After 15 min of incubation the blank solution is colourless and the test

TESTS

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 0.500 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 670 Pa for 4 h.

Microbial contamination

TAMC: acceptance criterion 10⁴ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

ASSAY

The activity of pepsin powder is determined by comparing the quantity of peptides, non-precipitable by trichloroacetic acid solution R and assayed using the phosphomolybdotungstic reagent R, which are released per minute from a substrate of haemoglobin solution R, with the quantity of such peptides released by pepsin powder BRP from the same substrate in the same conditions.

For the test solution and the reference solution, prepare the solution and carry out the dilution at 0 °C to 4 °C.

Avoid shaking and foaming during preparation of the test and reference solutions.

Test solution Immediately before use, prepare a solution of the substance to be examined expected to contain 0.5 Ph. Eur. U/mL in dilute hydrochloric acid R2; before dilution to volume, adjust to pH 1.6 ± 0.1, if necessary, using 1 M hydrochloric acid.

Reference solution Less than 15 min before use, prepare a solution of pepsin powder BRP containing 0.5 Ph. Eur. U/mL in dilute hydrochloric acid R2; before dilution to volume, adjust to pH 1.6 ± 0.1, if necessary, using 1 M hydrochloric acid.

Designate tubes in duplicate T, T_p, S₁, S_{1b}, S₂, S_{2b}, S₃, S_{3b}; designate a tube B.

Add dilute hydrochloric acid R2 to the tubes as follows:

B: 1.0 mL

S₁ and S_{1b}: 0.5 mL

S₂, S_{2b} and T and T_p: 0.25 mL

Add the reference solution to the tubes as follows:

S₁ and S_{1b}: 0.5 mL

S₂ and S_{2b}: 0.75 mL

S₃ and S_{3b}: 1.0 mL

Add 0.75 mL of the test solution to tubes T and T_p.

Add 10.0 mL of trichloroacetic acid solution R to tubes S_{1b}, S_{2b}, S_{3b}, T_p and B. Mix by shaking.

Place the tubes and haemoglobin solution R in a water bath at 25 ± 0.1 °C. When temperature equilibrium is reached, add 5.0 mL of haemoglobin solution R to tubes B, S_{1b}, S_{2b}, S_{3b} and T_p. Mix.

At time zero add 5.0 mL of haemoglobin solution R successively and at intervals of 30 s to tubes S₁, S₂, S₃ and T.

Mix immediately after each addition.

Exactly 10 min after adding the haemoglobin solution R, stop the reaction by adding, at intervals of 30 s, 10.0 mL of trichloroacetic acid solution R to tubes S₁, S₂, S₃ and T (the use of a fast-flowing or blow-out pipette is recommended).

Filter the contents of each tube (samples and blanks) twice through the same suitable filter paper previously washed with a 50 g/L solution of trichloroacetic acid R, then with water R and dried. Discard the first 5 mL of filtrate. Place 3.0 mL of

each filtrate separately in a tube containing 20 mL of water R. Mix.

A suitable filter paper complies with the following test. Filter 5 mL of a 50 g/L solution of trichloroacetic acid R through a 7 cm disc of white filter paper: the absorbance (2.2.25) of the filtrate, measured at 275 nm using unfiltered trichloroacetic acid R solution as the compensation liquid, is less than 0.04. Add to each tube 1.0 mL of sodium hydroxide solution R and 1.0 mL of phosphomolybdovanstic reagent R, beginning with the blanks and then the samples of each set, in a known order.

A schematic presentation of the above operations is shown in Table 0682-I.

After 15 min measure the absorbance (2.2.25) of solutions S_1 , S_2 , S_3 , S_{1b} , S_{2b} , S_{3b} and T at 540 nm using the filtrate obtained from tube B as the compensation liquid. Correct the average absorbance values for the filtrates obtained from tubes S_1 , S_2 and S_3 by subtracting the average values obtained for the filtrates from tubes S_{1b} , S_{2b} , S_{3b} respectively. Draw a calibration curve of the corrected values against volume of reference solution used. Determine the activity of the substance to be examined using the corrected absorbance for the test solution ($T - T_p$) together with the calibration curve and taking into account the dilution factors.

STORAGE

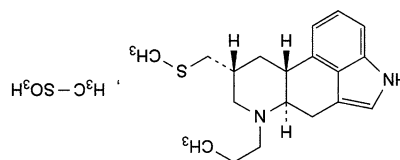
Store in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

LABELLING

The label states the activity in European Pharmacopoeia Units per milligram.

Pergolide Mesilate

(Ph. Eur. monograph I555)



$C_{20}H_{30}N_2O_3S_2$ 410.6

66104-23-2

Action and use
Dopamine receptor agonist; treatment of Parkinson's disease.

DEFINITION

(6aR,9R,10aR)-9-[(Methanysulfanylmethyl)-7-propyl-4,6,6a,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline monomethanesulfonate.

Content

97.5 per cent to 102.0 per cent (dried substance).

PRODUCTION

It is considered that alkylsulfonate esters are genotoxic and are potential impurities in pergolide mesilate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods 2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid, 2.5.38. Methyl, ethyl and isopropyl

methanesulfonate in active substances and 2.5.39. Methanesulfonfyl chloride in methanesulfonic acid are available to assist manufacturers.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent) and in methylene chloride, very slightly soluble in acetone.

IDENTIFICATION

A. Specific optical rotation (2.2.7): -23 to -17 (dried substance).

Dissolve 0.25 g in dimethylformamide R and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison pergolide mesilate CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 30.0 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (b) Dissolve 10 mg of 4,4'-dimethoxybenzophenone R in methanol R and dilute to 10 mL with the same solvent. To 1 mL of the solution add 2 mL of the test solution and dilute to 100 mL with methanol R. Dilute 1 mL of this solution to 10 mL with methanol R.

Column:

methanol R.

size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase:

mobile phase A: mix 5.0 mL of morpholine for chromatography R with 995 mL of water R and adjust to pH 7.0 with phosphoric acid R; use within 24 h;

mobile phase B: acetonitrile R, methanol R, tetrahydrofuran R (1:1:1 V/V/V);

Time

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 35	70 \rightarrow 0	30 \rightarrow 100
35 - 40	0 \rightarrow 70	100 \rightarrow 30
40 - 50	70	30

Flow rate

1 mL/min.

Detection

Spectrophotometer at 280 nm.

Injection

20 μ L.

System suitability:

reference solution (b):

— resolution: minimum 2.0 between the peaks due to 4,4'-dimethoxybenzophenone (1st peak) and pergolide (2nd peak).

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 105 °C for 1 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29).

Solution A Dissolve 5.0 mg of DL-methionine R in 500 mL of 0.01 M hydrochloric acid. Add 500 mL of methanol R and mix.

Test solution Dissolve 65.0 mg of the substance to be examined in solution A and dilute to 100.0 mL with solution A. Dilute 10.0 mL of this solution to 100.0 mL with solution A.

Reference solution Dissolve 65.0 mg of pergolide mesilate CRS in solution A and dilute to 100.0 mL with solution A. Dilute 10.0 mL of this solution to 100.0 mL with solution A.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated octylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase Mix 1 volume of acetonitrile R, 1 volume of methanol R and 2 volumes of a mixture prepared as follows:

dissolve 2.0 g of sodium octanesulfonate R in water R, add 1.0 mL of anhydrous acetic acid R and dilute to 1000 mL with water R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Retention time Pergolide = about 9 min.

System suitability: reference solution:

— *symmetry factor*: maximum 1.5 for the peak due to pergolide.

Calculate the percentage content of $C_{20}H_{30}N_2O_5S_2$ from the assigned content of pergolide mesilate CRS.

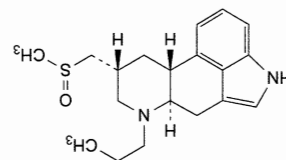
STORAGE

Protected from light.

IMPURITIES

Specified impurities A

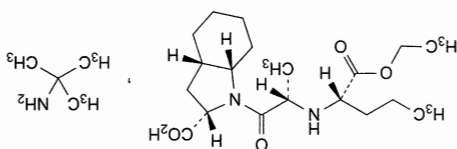
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: B.



A. (6aR,9R,10aR)-9-[(methanesulfonyl)methyl]-7-propyl-4,6,7,8,9,10,10a-octahydroindolo[4,3-fg]quinoline (pergolide sulfoxide);

Perindopril Erbumine

(Perindopril *tert*-Butylamine, Ph Eur monograph 2019)



$C_{23}H_{43}N_3O_5$ 441.6 107133-36-8

Action and use

Angiotensin converting enzyme inhibitor.

Preparation

Perindopril Erbumine Tablets

Ph Eur

DEFINITION

2-Methylpropan-2-amine (2S,3aS,7aS)-1-[(2S)-2-[[[(1S)-1-(ethoxycarbonyl)butyl]amino]propanoyl]octahydro-1H-indole-2-carboxylate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), soluble or sparingly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Specific optical rotation (2.2.7): –69 to –66 (anhydrous substance).
Dissolve 0.250 g in ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.
B. Infrared absorption spectrophotometry (2.2.24).
Comparison perindopril tert-butylamine CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in methylene chloride R, evaporate to dryness and record new spectra using the residues.

C. Examine the chromatograms obtained in the test for impurity A.

Results In the chromatogram obtained with the test solution a spot is observed with the same R_F as the spot with the higher R_F in the chromatogram obtained with reference solution (c) (*tert*-butylamine).

TESTS

Impurity A

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.20 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of perindopril impurity A CRS in methanol R and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 20.0 mL with methanol R.

Reference solution (c) To 5 mL of reference solution (a) add 5 mL of a 20 g/L solution of 1,1-dimethylethylamine R in methanol R.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, toluene R, methanol R (1:40:60 V/V/V).

Application 10 µL of the test solution and reference solutions (b) and (c).

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Expose to iodine vapour for at least 20 h.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated spots.

Limit:

— **impurity A:** any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent).

Stereochemical purity

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with ethanol (96 per cent) R. Dilute 1.0 mL of this solution to 10.0 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of perindopril for stereochemical purity CRS (containing impurity I) in ethanol (96 per cent) R and dilute to 5.0 mL with the same solvent.

Column:

— **size:** $l = 0.25$ m, $\phi = 4.6$ mm;

— **stationary phase:** spherical octadecylsilyl silica gel for chromatography R (5 µm);

— **temperature:** 50 °C for the column and the tubing preceding the column (the method has been developed with a temperature of 50 °C for at least 30 cm of the tubing preceding the column).

Mobile phase Mix, in the following order, 21.7 volumes of acetonitrile R, 0.3 volumes of pentanol R, and 78 volumes of 1.50 g/L solution of sodium heptanesulfonate R previously adjusted to pH 2.0 with a mixture of equal volumes of perchloric acid R and water R.

Detection Spectrophotometer at 215 nm.

Equilibration Minimum 4 h.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with perindopril for stereochemical purity CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity I.

Run time 1.5 times the retention time of perindopril.

Relative retention With reference to perindopril (retention time = about 100 min): impurity I = about 0.9.

System suitability:

— the chromatogram obtained with reference solution (b) is similar to the chromatogram supplied with perindopril for stereochemical purity CRS;

— **signal-to-noise ratio:** minimum 3 for the principal peak in the chromatogram obtained with reference solution (a);

— **peak-to-valley ratio:** minimum 3, where H_p = height above the baseline of the peak due to impurity I and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to perindopril in the chromatogram obtained with reference solution (b).

Limits:

— **impurity I:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **disregard limit:** disregard any peak with a relative retention with reference to perindopril of less than 0.6 or more than 1.4.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or maintain them at a temperature below 10 °C.

Test solution Dissolve 60 mg of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dissolve 3 mg of perindopril for peak identification CRS (containing impurities B, E, F, H and K) in 1 mL of mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with mobile phase A.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with mobile phase A.

Column:

— **size:** $l = 0.15$ m, $\phi = 4$ mm;

— **stationary phase:** spherical end-capped octylsilyl silica gel for chromatography R (5 µm) with a pore size of 15 nm;

— **temperature:** 60 °C for the column and the tubing preceding the column.

Mobile phase:

— **mobile phase A:** water R adjusted to pH 2.5 with a mixture of equal volumes of perchloric acid R and water R;

— **mobile phase B:** 0.03 per cent V/V solution of perchloric acid R in acetonitrile R₁;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - (5 - t)	95	5
(5 - t) - (60 - t)	95 → 40	5 → 60
(60 - t) - (65 - t)	40 → 95	60 → 5

The isocratic step is described for a chromatographic system with a dwell volume (D) of 2 mL. If D is different from 2 mL, correct the gradient times with the value t_r calculated using the following expression:

$$\frac{\text{flow rate}}{D - 2}$$

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Identification of impurities Use the chromatogram supplied

with perindopril for peak identification *C_{RS}* and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, E, F, H and K.

Relative retention With reference to perindopril (retention time = about 25 min): impurity B = about 0.68; impurity K = about 0.72; impurity E = about 1.2; impurity F = about 1.6; impurity H = about 1.8 (impurity H may be eluted as 1 or 2 peaks).

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 3, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity K.

Limits:

— **impurity E:** not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— **impurity B:** not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— **impurities F, H:** for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— **disregard limit:** the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.50 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.160 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 22.08 mg of $C_{23}H_{43}N_3O_5$.

STORAGE

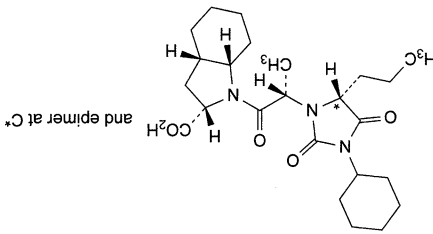
In an airtight container.

IMPURITIES

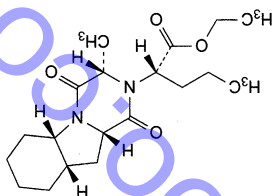
Specified impurities A, B, E, F, H, I.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: C, D, G, J, K, L, M, N, O, P, Q, R, S, T, U, V, W, X, Y, Z, AA, BB, CC.

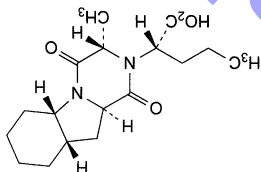
G. (2S,3aS,7aS)-1-[(2S)-2-[(5R)-3-cyclohexyl-2,4-dioxo-5-propylimidazolidin-1-yl]propanoyl]octahydro-1H-indole-2-carboxylic acid,



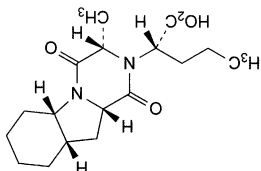
F. ethyl (2S)-2-[(3S,5aS,9aS,10aS)-3-methyl-1,4-dioxodecahydropyrazino[1,2-a]indol-2(1H)-yl]pentanoate,



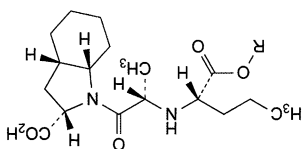
D. (2S)-2-[(3S,5aS,9aS,10aR)-3-methyl-1,4-dioxodecahydropyrazino[1,2-a]indol-2(1H)-yl]pentanoic acid,



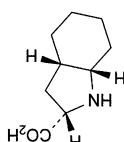
C. (2S)-2-[(3S,5aS,9aS,10aS)-3-methyl-1,4-dioxodecahydropyrazino[1,2-a]indol-2(1H)-yl]pentanoic acid,

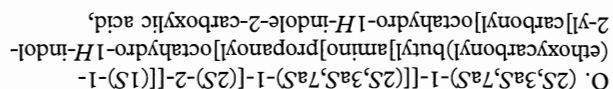


B. R = H: (2S,3aS,7aS)-1-[(2S)-2-[(1S)-1-carboxybutyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid (perindoprilat),
E. R = $CH(CH_3)_2$: (2S,3aS,7aS)-1-[(2S)-2-[(1S)-1-[[1-methylethoxy]carbonyl]butyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid,

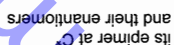


A. (2S,3aS,7aS)-octahydro-1H-indole-2-carboxylic acid,





(Ph Eur monograph 1762)


$$\text{C}_{21}\text{H}_{20}\text{Cl}_2\text{O}_3$$
Undefined
cis/trans ratio: [52645-53-1]

Action and use

Insecticide

DEFINITION

Ph Eur -

Mixture of the *cis* and *trans* isomers of

3,3-dimethylcyclopropane-1-carboxylate.

Content

—total: 98.0 per cent to 102.0 per cent (anhydrous substance);

— *cis* isomer: 23.0 per cent to 27.0 per cent (anhydrous substance);

— *trans* isomer: 73.0 per cent to 77.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Colourless or slightly brownish viscous liquid, semi-solid or crystalline solid.

Solubility

Practically insoluble in water, very soluble in heptane, freely soluble in anhydrous ethanol, sparingly soluble in ethylene glycol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison *permethrin* (25:75) CRS.

B. Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time and in *cis/trans* ratio to the 2 principal peaks in the chromatogram obtained with the reference solution (*cis/trans* ratio = about 0.3).

TESTS

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution Warm the substance to be examined at 70–85 °C for 20 min. Dissolve 1.0 g of this substance in *heptane* R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of *permethrin* for system suitability CRS (containing impurities B, C and G) in *heptane* R and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *heptane* R. Dilute 1.0 mL of this solution to 20.0 mL with *heptane* R.

Column:

— material: fused silica;

— size: $l = 15$ m, $\varnothing = 0.53$ mm;

— stationary phase: poly(*dimethyl*)siloxane R (film thickness 1.5 μ m).

Carrier gas nitrogen for chromatography R.

Flow rate 12.0 mL/min.

Split ratio 1:10.

Temperature:

Time (min)	Temperature (°C)
0 - 2	45
2 - 26.5	290
26.5 - 40.5	250
40.5 - 45	290
45 - 290	290

Detection Flame ionisation.

Injection 1 μ L.

Identification of impurities Use the chromatogram supplied with *permethrin* for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B, C and G.

Relative retention With reference to *permethrin* (retention time = about 24 min): impurity B = about 0.5;

impurity C = about 0.7; impurity G = about 0.9.

ASSAY

Liquid chromatography (2.2.29).

Test solution Warm the substance to be examined at 70–85 °C for 20 min. Dissolve 0.100 g of this substance in the mobile phase using sonication and dilute to 50.0 mL with the mobile phase.

Reference solution Warm *permethrin* (25:75) CRS at 70–85 °C for 20 min. Dissolve 0.100 g of this substance in the mobile phase using sonication and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50 mg of *permethrin* for system suitability CRS (containing impurities B, C and G) in *heptane* R and dilute to 1.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *heptane* R. Dilute 1.0 mL of this solution to 20.0 mL with *heptane* R.

Mobile phase *dioxan* R, *heptane* R (1.1:98.9 V/V). Temperature: 30 °C.

Stationary phase silica gel for chromatography R (5 μ m); size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Run time Twice the retention time of *cis*-*permethrin*.

Retention time *cis*-*permethrin* = about 6 min.

Blution order *cis*-*permethrin*, *trans*-*permethrin*.

System suitability: reference solution:

— resolution: minimum 2.0 between the peaks due to *cis*-*permethrin* and *trans*-*permethrin*.

Calculation of percentage contents:

— calculate the percentage content of *cis*-*permethrin* and *trans*-*permethrin* by normalisation;

— calculate the percentage content of $C_{21}H_{20}Cl_2O_3$ taking into account the assigned content of *permethrin* (25:75) CRS.

STORAGE

Protected from light.

IMPURITIES

Specified impurities B, C

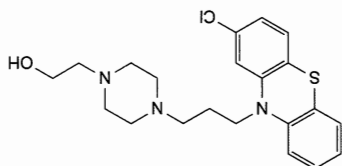
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, D, E, F, G, H.



(Ph. Eur. monograph 0629)

Perphenazine


 $C_{21}H_{26}ClN_3OS$ 404.0 58-39-9

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Perphenazine Tablets

Ph Eur

DEFINITION

2-[4-[3-(2-chloro-10H-phenothiazin-10-yl)propyl]piperazin-1-yl]ethanol.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in ethanol (96 per cent). It dissolves in dilute solutions of hydrochloric acid.

IDENTIFICATION

A. Melting point (2.2.14): 96 °C to 100 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison perphenazine CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1).

Dissolve 0.20 g in 10 mL of methanol R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use. Carry out the test protected from light.

Test solution Dissolve 20 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 2 mg of perphenazine for system suitability CRS (containing impurities A and B) in 1.0 mL of mobile phase A.

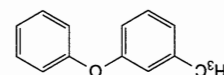
Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;— stationary phase: spherical base-deactivated octylsilyl silica gel for chromatography R (4 μ m);

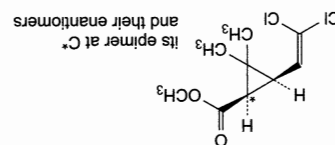
— temperature: 30 °C.

Mobile phase:

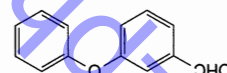
— mobile phase A: mix 35 volumes of acetonitrile R and 65 volumes of a 7 g/L solution of sodium dihydrogen phosphate R;



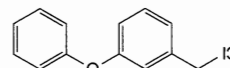
A. 1-methyl-3-phenoxymethylbenzene,



B. methyl (1R,2R)-2-(2-(2-dichloroethenyl)-3,3-dimethylcyclopropyl)-1-carboxylate,

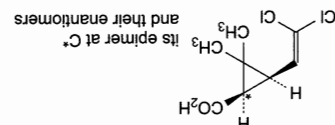


C. (3-phenoxymethyl)phenylmethanol,

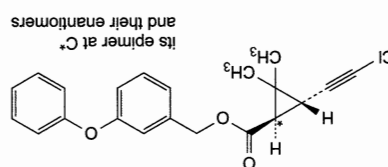


D. 3-phenoxymethylbenzaldehyde,

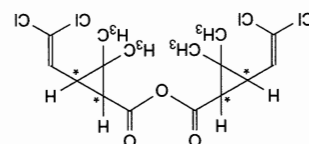
E. 1-(chloromethyl)-3-phenoxymethylbenzene,



F. (1R,2R)-2-(2-(2-dichloroethenyl)-3,3-dimethylcyclopropyl)-1-carboxylic acid,



G. 3-(phenoxymethyl)phenyl (1R,2R)-2-(chloroethenyl)-3,3-dimethylcyclopropyl-1-carboxylate,



H. 2-(2-(2-dichloroethenyl)-3,3-dimethylcyclopropyl)-1-carboxylic anhydride.

Ph Eur

— mobile phase B: acetonitrile R₃

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 5	100	0
5 - 10	100 → 80	0 → 20
10 - 33	80 → 30	20 → 70
33 - 48	30 → 100	70 → 0

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with perphenazine for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A and B.

Relative retention With reference to perphenazine (retention

time = about 12 min): impurity A = about 0.3;

impurity B = about 0.8.

System suitability: reference solution (b):

— resolution: minimum 4.0 between the peaks due to

impurity B and perphenazine.

Limits:

— correction factor: for the calculation of content, multiply the

peak area of impurity A by 0.6;

— impurity A: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.2 per cent);

— impurity B: not more than 5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.5 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 10 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (1.0 per cent);

— disregard limit: 0.5 times the area of the principal peak in

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 65 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 25 mL of anhydrous acetic acid R. Titrate

with 0.1 M perchloric acid determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 20.20 mg of

C₂₁H₂₆ClNO₃OS.

STORAGE

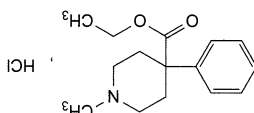
Protected from light.

IMPURITIES

Specified impurities A, B

Pethidine Hydrochloride

(Ph. Eur. monograph 0420)



283.8

C₁₅H₂₂ClNO₂

50-13-5

Action and use

Opioid receptor agonist; analgesic.

Preparations

Pethidine Injection

Pethidine Tablets

DEFINITION

Ethyl 1-methyl-4-phenylpiperidine-4-carboxylate hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

If intended for use in the manufacture of parenteral preparations, the manufacturing process is validated to show that the content of impurity B is not more than 0.1 ppm.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, freely soluble in alcohol.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D

A. Melting point (2.2.14): 187 °C to 190 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of pethidine hydrochloride.

C. Dissolve 0.1 g in 10 mL of ethanol R and add 10 mL of picric acid solution R. A crystalline precipitate is formed which, when washed with water R and dried at 100-105 °C, melts (2.2.14) at 186 °C to 193 °C. Mix equal quantities of the precipitate and the substance to be examined and determine the melting point of the mixture. The melting point is at least 20 °C lower than that of the precipitate.

D. To 5 mL of solution S (see Tests) add 5 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.2 mL of methyl red solution R and 0.2 mL of 0.01 M sodium hydroxide. The solution is yellow. Add 0.3 mL of 0.01 M hydrochloric acid. The solution is red.

Impurity B

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 0.100 g of the substance to be examined in a mixture of 20 volumes of acetonitrile R and 80 volumes of water R and dilute to 25.0 mL with the same mixture of solvents.

Test solution (b) Dissolve 0.125 g of the substance to be examined in a mixture of 20 volumes of acetonitrile R and 80 volumes of water R and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (a) Dilute 0.5 mL of test solution (a) to 100.0 mL with a mixture of 20 volumes of acetonitrile R and 80 volumes of water R.

Reference solution (b) Dissolve 10.0 mg of pethidine impurity A CRS in a mixture of 20 volumes of acetonitrile R and 80 volumes of water R and dilute to 100.0 mL with the same mixture of solvents.

Reference solution (c) Dissolve 12.5 mg of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine R in a mixture of 20 volumes of acetonitrile R and 80 volumes of water R and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (d) Dilute 5.0 mL of reference solution (b) and 1.0 mL of reference solution (c) to 100.0 mL with a mixture of 20 volumes of acetonitrile R and 80 volumes of water R.

Stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 µm) with a specific surface area of 340 m²/g, a pore size of 10 nm and a carbon loading of 19 per cent.

Mobile phase: mix equal volumes of a 42.0 g/L solution of sodium perchlorate R and of a 11.6 g/L solution of phosphoric acid R, adjust to pH 2.0 with triethylamine R, — mobile phase B: acetonitrile R,

Column: size: l = 0.25 m, Ø = 4.0 mm,

Flow rate: 1.0 mL/min.

Detection: Spectrophotometer at 210 nm.

Injection: 50 µL; inject test solution (b) and reference solution (d).

Relative retention: With reference to pethidine (retention time = about 2.4 min): impurity B = about 0.66; impurity A = about 0.68.

System suitability: Reference solution (d):

— **signal-to-noise ratio:** minimum 10 for the first peak,

— **peak-to-valley ratio:** minimum 4, where H_p = height above the baseline of the peak due to impurity B, and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity A.

Limit:

— **impurity B:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (10 ppm) if intended for non-parenteral administration.

Related substances

Liquid chromatography (2.2.29) as described in the test for impurity B with the following modifications.

Injection: 20 µL; inject test solution (a) and reference solution (a).

Limit:

— **any impurity:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent),

— **disregard limit:** 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.220 g in 50 mL of alcohol R. Add 5.0 mL of 0.01 M hydrochloric acid. Titrate with 0.1 M sodium hydroxide determining the end-point potentiometrically (2.2.20). Read the volume added between the 2 points of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 28.38 mg of C₁₅H₂₂ClNO₂.

STORAGE

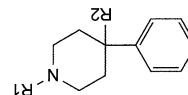
In an airtight container, protected from light.

LABELLING

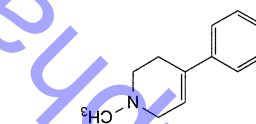
The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	80 → 75	20 → 25
15 - 31	75 → 55	25 → 45
31 - 40	55	45

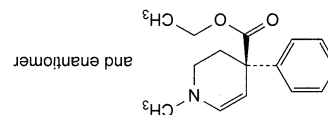
IMPURITIES



- A. $R_1 = CH_3$, $R_2 = H$: 1-methyl-4-phenylpiperidine (MP), carboxylic acid,
 C. $R_1 = CH_3$, $R_2 = CO_2H$: 1-methyl-4-phenylpiperidine-4-carboxylic acid,
 D. $R_1 = CH_3$, $R_2 = CO_2-CH_3$: methyl 1-methyl-4-phenylpiperidine-4-carboxylate,
 E. $R_1 = H$, $R_2 = CO_2-CH_2-CH_3$: ethyl 4-phenylpiperidine-4-carboxylate,
 F. $R_1 = CH_2-C_6H_5$, $R_2 = CO_2H$: 1-benzyl-4-phenylpiperidine-4-carboxylic acid,
 G. $R_1 = CH_3$, $R_2 = CO_2-CH(CH_3)_2$: 1-methylethyl 1-methyl-4-phenylpiperidine-4-carboxylate,
 H. $R_1 = CH_2-C_6H_5$, $R_2 = CO_2-CH_2-CH_3$: ethyl 1-benzyl-4-phenylpiperidine-4-carboxylate,
 J. $R_1 = CH_2-CH_3$, $R_2 = CO_2-CH_2-CH_3$: ethyl 1-ethyl-4-phenylpiperidine-4-carboxylate,



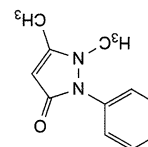
- B. 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP),



- I. ethyl (4R)-1-methyl-4-phenyl-1,2,3,4-tetrahydropyridine-4-carboxylate.

Phenazone

(Ph. Eur. monograph 0421)



$C_{11}H_{12}N_2O$ 188.2 60-80-0

Action and use

Analgesic; used to test hepatic drug-metabolizing activity.

Ph Eur

DEFINITION

1,5-Dimethyl-2-phenyl-1,2-dihydro-3H-pyrazol-3-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Very soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Melting point (2.2.14): 109 °C to 113 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison phenazone CRS.

C. To 1 mL of solution S (see Tests) add 4 mL of water R and 0.25 mL of dilute sulfuric acid R. Add 1 mL of sodium nitrite solution R; a green colour develops.
 D. To 1 mL of solution S add 4 mL of water R and 0.5 mL of ferric chloride solution R2. A red colour develops which is discharged on the addition of dilute sulfuric acid R.

TESTS

Solution S
 Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of phenolphthalein solution R; the solution is colourless. Add 0.2 mL of 0.01 M sodium hydroxide; the solution is red. Add 0.25 mL of methyl red solution R and 0.4 mL of 0.01 M hydrochloric acid; the solution is red or yellowish-red.

Related substances

Liquid chromatography (2.2.29).
 Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of phenazone impurity A CRS in the mobile phase, add 10 mL of the test solution and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Dissolve 5.0 mg of phenazone impurity A CRS in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 6.0$ mm;

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve 6.8 g of potassium dihydrogen phosphate R in water R and dilute to 1000 mL with the same solvent. Add 2 mL of triethylamine R and adjust to pH 7.0 with sodium hydroxide solution R. Add 430 mL of methanol R. Flow rate 1.0 mL/min.

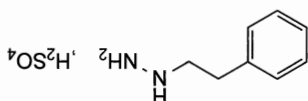
Detection Spectrophotometer at 254 nm.

Injection 10 μ L.

Run time 3 times the retention time of phenazone.
 Relative retention With reference to phenazone (retention time = about 13 min): impurity A = about 0.8.
 System suitability: reference solution (b):
 — resolution: minimum 3.0 between the peaks due to impurity A and phenazone.

Phenelzine Sulfate

Phenelzine Sulfate

 $C_8H_{12}N_2H_2SO_4$

234.3

156-51-4

Action and use

Monoamine oxidase inhibitor; antidepressant.

Preparation

Phenelzine Tablets

DEFINITION

Phenelzine Sulfate is phenethylhydrazine hydrogen sulfate. It contains not less than 98.0% and not more than 100.5% of $C_8H_{12}N_2H_2SO_4$, calculated with reference to the dried substance.

CHARACTERISTICS

A white powder or pearly platelets. Freely soluble in water; practically insoluble in ethanol (96%) and in ether.

IDENTIFICATION

A. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.1% w/v solution in 0.05M sulfuric acid exhibits three well-defined maxima, at 252, 258 and 263 nm. The absorbances at the maxima are about 0.62, about 0.77 and about 0.58, respectively.

B. Dissolve 0.1 g in 5 mL of water, make alkaline with 5M sodium hydroxide and add 1 mL of cupri-tartrate solution R1. A red precipitate is produced.

C. Yields reaction A characteristic of sulfates, Appendix VI.

TESTS

Melting point

164° to 168°, Appendix V A.

Loss on drying

When dried over phosphorus pentoxide at a pressure not exceeding 0.7 kPa for 24 hours, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out the method for liquid chromatography, Appendix III D, using the following solutions.

(1) 0.026% w/v of the substance to be examined in the mobile phase.

(2) 0.026% w/v phenelzine sulfate BPGRS in the mobile phase.

CHROMATOGRAPHIC CONDITIONS

(a) A stainless steel column (150 × 3.9 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Waters Symmetry C18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1.0 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 210 nm.

(f) Inject 20 µL of each solution.

Limits:

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.05 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— the chromatogram obtained with reference solution (a) — disregard limit: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Chlorides (2.4.4)

Maximum 100 ppm.

Sulfates (2.4.13)

Maximum 100 ppm.

Dissolve 1.5 g in distilled water R and dilute to 15 mL with the same solvent.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in vacuum at 60 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 20 mL of water R. Add 2 g of sodium acetate R and 25.0 mL of 0.05 M iodine. Allow to stand protected from light for 30 min. Add 25 mL of methylene chloride R and shake until the precipitate dissolves. Titrate with 0.1 M sodium thiosulfate, using 1 mL of starch solution R, added towards the end of the titration, as indicator. Carry out a blank titration.

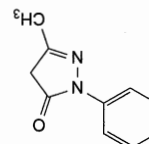
1 mL of 0.05 M iodine is equivalent to 9.41 mg of $C_{11}H_{12}N_2O$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A



A. 5-methyl-2-phenyl-2,4-dihydro-3H-pyrazol-3-one.

Ph Eur

MOBILE PHASE

40 volumes of methanol R2 and 60 volumes of a solution of 0.216% w/v sodium octanesulfonate and 0.68% w/v potassium dihydrogen phosphate previously adjusted to pH 3.0 with orthophosphoric acid.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (2), the symmetry factor of the principal peak is between 0.8 and 2.0.

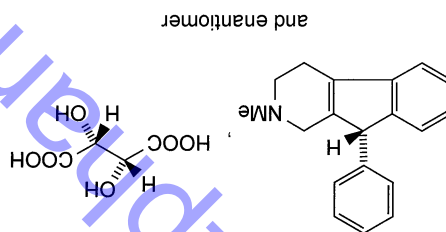
DETERMINATION OF CONTENT

Calculate the content of $C_8H_{12}N_2H_2SO_4$ in the substance being examined using the declared content of $C_8H_{12}N_2H_2SO_4$ in phenelzine sulfate BPCRS.

STORAGE

Phenelzine Sulfate should be protected from light.

Phenindamine Tartrate



$C_{19}H_{19}N_3C_4H_6O_6$

411.5

569-59-5

Action and use

Histamine H_1 receptor antagonist; antihistamine.

DEFINITION

Phenindamine Tartrate is (R,S)-2,3,4,9-tetrahydro-2-methyl-9-phenyl-1H-indeno[2,1-c]pyridine hydrogen (2R,3R)-

tartrate. It contains not less than 98.5% and not more than 101.0% of $C_{19}H_{19}N_3C_4H_6O_6$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, voluminous powder.

Sparingly soluble in water; slightly soluble in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

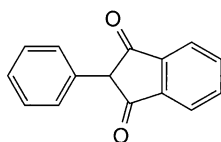
A. The light absorption, Appendix II B, in the range 230 to 300 nm of a 0.004% w/v solution exhibits a maximum only at 259 nm. The absorbance at the maximum is about 0.88.

B. Dissolve 25 mg in 5 mL of sulfuric acid. An orange-brown colour is produced which is discharged when the solution is carefully added to 20 mL of water.

C. Dissolve 0.5 g in 15 mL of hot water, add a slight excess of 5M sodium hydroxide, filter and neutralise the filtrate to litmus paper with 2M hydrochloric acid. The solution yields reaction B characteristic of tartrates, Appendix VI.

D. Melting point, 160° to 162°, Appendix V A. When heated to 163° it resolidifies and it melts again at about 168°, with decomposition.

Phenindione



222.2

83-12-5

Action and use

Oral anticoagulant (indanedione).

Preparation

Phenindione Tablets

DEFINITION

Phenindione is 2-phenylindane-1,3-dione. It contains not less than 98.0% and not more than 100.5% of $C_{15}H_{10}O_2$, calculated with reference to the dried substance.

CHARACTERISTICS

Soft, white or creamy white crystals.

Very slightly soluble in water; slightly soluble in ethanol (96%) and in ether. Solutions are yellow to red.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of phenindione (RS 268).

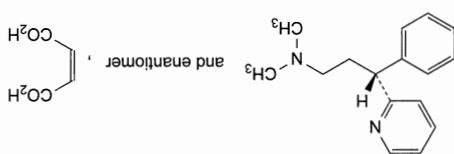
B. Dissolve 0.1 g in 30 mL of ethanol (96%) with the aid of heat, cool and add sufficient ethanol (96%) to produce 50 mL. Dilute 10 mL to 250 mL with 0.1M sodium hydroxide and further dilute 5 mL to 100 mL with 0.1M sodium hydroxide. The absorbance of the resulting solution at the maximum at 278 nm is about 0.54 and at the maximum at 330 nm is about 0.16, Appendix II B.

C. To 1 g add 50 mL of ethanol (96%) and 0.5 mL of aniline, heat gently under a reflux condenser for 3 hours, cool in ice and filter. The melting point of the residue, after washing with 2 mL of ethanol (96%) and recrystallising from chloroform, is about 225°, Appendix V A.



Pheniramine Maleate

(Ph. Eur. monograph 1357)



$C_{20}H_{24}N_2O_4$ 356.4 132-20-7

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Ph. Eur.

DEFINITION

(3*R*,5)-*N*,*N*-Dimethyl-3-phenyl-3-(pyridin-2-yl)propan-1-amine (Z)-butenedioate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent), in methanol and in methylene chloride.

IDENTIFICATION

First identification C

Second identification A, B, D

A. Melting point (2.2.14): 106 °C to 109 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 40.0 mg in 0.1 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 5.0 mL of the solution to 50.0 mL with 0.1 M hydrochloric acid.

Spectral range 220-320 nm.

Absorption maximum At 265 nm.

Shoulder At 261 nm.

Specific absorbance at the absorption maximum 200 to 220.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison pheniramine maleate CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 65 mg of maleic acid R in methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 0.10 g of pheniramine maleate CRS in methanol R and dilute to 5.0 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase water R, anhydrous formic acid R, methanol R, di-isopropyl ether R (3:7:20:70 V/V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Detection Examine in ultraviolet light at 254 nm.

Results The chromatogram obtained with the test solution shows 2 clearly separated spots; the upper spot is similar in position and size to the spot in the chromatogram obtained with reference solution (a); the lower spot is similar in

TESTS

Melting point

148° to 151°, Appendix V A.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in dichloromethane and develop immediately.

(1) 1.0% w/v of the substance being examined.

(2) Dilute 2 volumes of solution (1) to 100 volumes.

(3) Dilute 1 volume of solution (2) to 4 volumes.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel F₂₅₄ (Merck silica gel 60 F₂₅₄ plates are suitable).

(b) Use the mobile phase as described below.

(c) Allow the solvent front to ascend 4 cm, remove the plate and dry it in a current of cold air for 1 minute. Immediately apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, allow it to dry in a current of warm air and examine under ultraviolet light (254 nm).

MOBILE PHASE

0.02% w/v of butylated hydroxytoluene in a mixture of 4 volumes of glacial acetic acid, 20 volumes of ethyl acetate and 80 volumes of toluene.

LIMITS

The principal spot in the chromatogram obtained with solution (1) corresponds in position and colour to that in the chromatogram obtained with solution (2).

In the chromatogram obtained with solution (1):

any secondary spot is not more intense than the spot in the chromatogram obtained with solution (2) (2%);

not more than one secondary spot is more intense than the spot in the chromatogram obtained with solution (3) (0.5%).

Loss on drying

When dried at 105° for 2 hours, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

To 0.3 g add 50 mL of ethanol (96%) and warm until solution is effecied. Cool to room temperature, add 10 mL of a 10% v/v solution of bromine in ethanol (96%) and allow to stand for 10 minutes, shaking occasionally. Add 1 g of 2-naphthol and shake until the colour of the bromine is discharged. Remove any vapour of bromine in the flask with a current of air, add 50 mL of water and 10 mL of dilute potassium iodide solution and titrate the liberated iodine with 0.1M sodium thiosulfate VS using starch mucilage as indicator. Each mL of 0.1M sodium thiosulfate VS is equivalent to 11.11 mg of C₁₅H₁₀O₂.

position and size to the lower spot in the chromatogram obtained with reference solution (b).

TESTS

Solution S

Dissolve 2.0 g in water R and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)

4.5 to 5.5.

Dissolve 0.20 g in 20 mL of carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R₃, mobile phase A (10:90 V/V).

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 10.0 mg of phenitramine impurity A CRS and 10 mg of 4-benzylpyridine R (impurity B) in 10.0 mL of the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the solvent mixture.

Reference solution (d) Dilute 1.0 mL of the test solution to 10.0 mL with reference solution (a). Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.30$ m, $\varnothing = 3.9$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

Mobile phase:

— mobile phase A: dissolve 5.056 g of sodium

heptanesulfonate R in 900 mL of water R, adjust to pH 2.5 with dilute phosphoric acid R and dilute to 1000 mL with water R;

— mobile phase B: acetonitrile R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	90	10
2 - 37	90 → 62	10 → 38

Flow rate 1 mL/min.

Detection Spectrophotometer at 264 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram obtained

with reference solution (a) to identify the peaks due to

impurities A and B.

Relative retention With reference to phenitramine (retention

time = about 31 min): maleic acid = about 0.1;

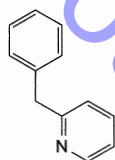
impurity A = about 0.9; impurity B = about 0.97.

System suitability Reference solution (d):

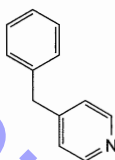
— resolution: minimum 1.5 between the peaks due to

impurity B and phenitramine.

A. 2-benzylpyridine,



B. 4-benzylpyridine.



IMPURITIES

Protected from light.

STORAGE

of C₂₀H₂₄N₂O₄.

1 mL of 0.1 M perchloric acid is equivalent to 17.82 mg

potentiometrically (2.2.20).

Dissolve 0.130 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 3 h.

Loss on drying (2.2.32)

reference solution using lead standard solution (2 ppm Pb) R.

12 mL of solution S complies with test A. Prepare the

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.05 per cent); disregard the peak due to maleic acid.

the chromatogram obtained with reference solution (b)

— disregard limit: 0.5 times the area of the principal peak in

— total: maximum 1.0 per cent;

with reference solution (b) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

solution (b) (0.2 per cent);

peak in the chromatogram obtained with reference

— impurity B: not more than twice the area of the principal

solution (c) (0.2 per cent);

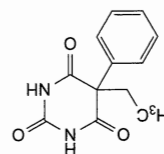
peak in the chromatogram obtained with reference

— impurity A: not more than the area of the corresponding

Limits:

Phenobarbital

(Ph. Eur. monograph 0201)

 $C_{12}H_{12}N_2O_3$

232.2

50-06-6

Action and use

Barbiturate.

Preparations

Phenobarbital Elixir

Phenobarbital Tablets

Ph Eur

DEFINITION

5-Ethyl-5-phenylpyrimidine-2,4,6-(1*H*,3*H*,5*H*)-trione.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals.

Solubility

Very slightly soluble in water, freely soluble in ethanol

(96 per cent).

It forms water-soluble compounds with alkali hydroxides,

IDENTIFICATION

First identification A, B

Second identification A, C, D.

A. Determine the melting point (2.2.14) of the substance to

be examined. Mix equal parts of the substance to be

examined and phenobarbital CRS and determine the melting

point of the mixture. The difference between the melting

points (which are about 176 °C) is not greater than 2 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison phenobarbital CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 10 mg of phenobarbital CRS in

ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase concentrated ammonia R, ethanol (96 per cent) R, methylene chloride R (5:15:80 V/V/V); use the lower layer.

Application 10 µL.

Development Over 2/3 of the plate.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

D. It gives the reaction of non-nitrogen substituted

barbiturates (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 1.0 g in a mixture of 4 mL of dilute sodium hydroxide

solution R and 6 mL of water R.

Acidity

Boil 1.0 g with 50 mL of water R for 2 min, allow to cool and filter. To 10 mL of the filtrate add 0.15 mL of methyl red

solution R. The solution is orange-yellow. Not more than

0.1 mL of 0.1 M sodium hydroxide is required to produce a

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.125 g of the substance to be examined in 5.0 mL of methanol R and dilute to 25.0 mL

with the mobile phase.

Reference solution (a) Mix 1.0 mL of the test solution and

20.0 mL of methanol R and dilute to 100.0 mL with the

mobile phase. Mix 1.0 mL of this solution with 2.0 mL of

methanol R and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of phenobarbital

impurity A CRS and 5.0 mg of phenobarbital impurity B CRS

in 2.0 mL of methanol R and dilute to 10.0 mL with the

mobile phase. Mix 1.0 mL of this solution with 20.0 mL of

methanol R and dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 µm).

Mobile phase Dissolve 6.60 g of sodium acetate R in 900 mL of

water R, add 3 mL of glacial acetic acid R, adjust to pH 4.5

with glacial acetic acid R and dilute to 1000 mL with water R.

Mix 60 volumes of this solution with 40 volumes of

methanol R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Run time 2.1 times the retention time of phenobarbital.

Identification of impurities Use the chromatogram obtained

with reference solution (b) to identify the peaks due to

impurities A and B.

Relative retention With reference to phenobarbital (retention

time = about 14 min): impurity A = about 0.2;

impurity B = about 0.3.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to

impurities A and B.

Limits:

— impurity A: not more than 1.5 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— impurity B: not more than 1.5 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— total: not more than twice the area of the principal peak in

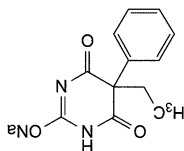
the chromatogram obtained with reference solution (a)

(0.2 per cent);



Phenobarbital Sodium

(Ph. Eur. monograph 0630)



C₁₂H₁₁N₂NaO₃ 254.2 57-30-7

Action and use

Barbiturate.

Preparations

Phenobarbital Injection
Paediatric Phenobarbital Oral Solution
Phenobarbital Sodium Tablets

Ph Eur

DEFINITION

Phenobarbital sodium contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of the sodium derivative of 5-ethyl-5-phenylpyrimidine-2,4,6(1H,3H,5H)-trione, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, hygroscopic, freely soluble in carbon dioxide-free water (a small fraction may be insoluble), soluble in alcohol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification A, B, E.
Second identification A, C, D, E.

A. Acidity 10 mL of solution S (see Tests) with dilute hydrochloric acid R and shake with 20 mL of ether R. Separate the ether layer, wash with 10 mL of water R, dry over anhydrous sodium sulfate R and filter. Evaporate the filtrate to dryness and dry the residue at 100 °C to 105 °C. Determine the melting point (2.2.14) of the test residue. Mix equal parts of the residue and of phenobarbital CRS and determine the melting point of the mixture. The difference between the two melting points (which are about 176 °C) is not greater than 2 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing the residue obtained during identification test A with the spectrum obtained with phenobarbital CRS. If the spectra obtained in the solid state show differences, dissolve the test residue and the reference substance separately in ethanol R, evaporate to dryness and record the spectra again.

C. Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.10 g of the substance to be examined in alcohol (50 per cent V/V) R and dilute to 100 mL with the same solvent.

Reference solution Dissolve 90 mg of phenobarbital CRS in alcohol (50 per cent V/V) R and dilute to 100 mL with the same solvent.

Apply separately to the plate 10 µL of each solution. Develop over a path of 18 cm using the lower layer from a mixture of 5 volumes of concentrated ammonia R, 15 volumes of alcohol R and 80 volumes of chloroform R. Examine immediately in

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 40 mL of ethanol (96 per cent) R and add

20 mL of water R. Titrate with 0.1 M sodium hydroxide,

determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 23.22 mg of

C₁₂H₁₁N₂O₃.

IMPURITIES

Specified impurities A, B

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

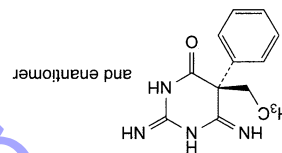
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

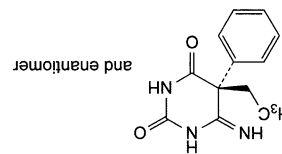
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

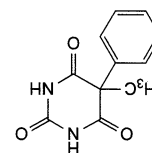
Control of impurities in substances for pharmaceutical use): C.



A. (5R,5S)-5-ethyl-2,6-diimino-5-phenyltetrahydropyrimidin-4(1H)-one,



B. (5R,5S)-5-ethyl-6-imino-5-phenylidihydropyrimidine-2,4(1H,3H)-dione,

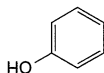


C. 5-methyl-5-phenylpyrimidine-2,4,6(1H,3H,5H)-trione.

Ph Eur



(Ph. Eur. monograph 0631)

C₆H₅O

94.1

108-95-2

Action and use

Antiseptic; antimicrobial preservative; antipruritic.

Preparations

Aqueous Phenol Injection

Oily Phenol Injection

Phenol and Glycerol Injection

DEFINITION**Content**

99.0 per cent to 100.5 per cent.

CHARACTERS**Appearance**

Colourless or faintly pink or faintly yellowish, crystals or

Solubility

Soluble in water, very soluble in ethanol (96 per cent), in

IDENTIFICATION

A. Dissolve 0.5 g in 2 mL of concentrated ammonia R.

The substance dissolves completely. Dilute to about 100 mL

with water R. To 2 mL of this solution add 0.05 mL of strong

sodium hypochlorite solution R. A blue colour develops and

becomes progressively more intense.

B. To 1 mL of solution S (see Tests) add 10 mL of water R

and 0.1 mL of ferric chloride solution R₁. A violet colour is

produced which disappears on addition of 5 mL of

2-propanol R.

C. To 1 mL of solution S add 10 mL of water R and 1 mL

of bromine water R. A white precipitate is formed.

TESTS**Solution S**

Dissolve 1.0 g in water R and dilute to 15 mL with the same

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution B₆ (2.2.2, Method II).**Acidity**

To 2 mL of solution S add 0.05 mL of methyl orange

solution R. The solution is yellow.

Freezing point (2.2.18)

Minimum 39.5 °C.

Residue on evaporation

Maximum 0.05 per cent.

Evaporate 5.000 g to dryness on a water-bath and dry at

100-105 °C for 1 h. The residue weighs a maximum of

2.5 mg.

ASSAY

Dissolve 2.000 g in water R and dilute to 1000.0 mL with the same solvent. Transfer 25.0 mL of the solution to a ground-glass-stoppered flask and add 50.0 mL of 0.0167 M bromide-bromate and 5 mL of hydrochloric acid R, close the

ultraviolet light at 254 nm. The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives the reaction of non-nitrogen substituted barbiturates (2.3.1).

E. It gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 5.0 g in alcohol (50 per cent V/V) R and dilute to

50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution Y₇ (2.2.2, Method II).**pH (2.2.3)**

Dissolve 5.0 g as completely as possible in carbon dioxide-free

water R and dilute to 50 mL with the same solvent. The pH

of the solution is not greater than 10.2.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica

gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 1.0 g of the substance to be examined

in alcohol (50 per cent V/V) R and dilute to 100 mL with the

same solvent.

Reference solution Dilute 0.5 mL of the test solution to

100 mL with alcohol (50 per cent V/V) R.

Apply separately to the plate 20 µL of each solution. Develop

over a path of 15 cm using the lower layer from a mixture of

5 volumes of concentrated ammonia R, 15 volumes of alcohol R

and 80 volumes of chloroform R. Examine immediately in

ultraviolet light at 254 nm. Spray with diphenylcarbazone

mercuric reagent R. Allow the plate to dry in air and spray

with freshly prepared alcoholic potassium hydroxide solution R

diluted 1 in 5 with aldehyde-free alcohol R. Heat at 100 °C to

105 °C for 5 min and examine immediately. When examined

in ultraviolet light and after spraying, any spot in the

chromatogram obtained with the test solution, apart from the

principal spot, is not more intense than the spot in the

chromatogram obtained with the reference solution

(0.5 per cent). Disregard any spot at the point of application.

Loss on drying (2.2.32)

Not more than 7.0 per cent, determined on 0.500 g by

drying in an oven at 150 °C for 4 h.

ASSAY

Dissolve 0.150 g in 2 mL of water R and add 8 mL of

0.05 M sulfuric acid. Heat to boiling and cool. Add 30 mL of

methanol R and shake until dissolution is complete. Carry out

a potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. After the first point of inflexion, interrupt the

addition of sodium hydroxide, add 10 mL of pyridine R, mix

and continue the titration. Read the volume added between

the two points of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 25.42 mg of

C₁₂H₁₁N₂NaO₃.**STORAGE**

Store in an airtight container.

Ph Eur

Liquefied Phenol

Action and use

Antiseptic; antimicrobial preservative; antipruritic. When Phenol is to be mixed with colloidal, fixed oils or paraffins, melted Phenol should be used and not Liquefied Phenol.

DEFINITION

Liquefied Phenol contains not less than 77.0% and not more than 81.5% w/w of Phenol in Purified Water. It is prepared by adding Purified Water to melted Phenol.

CHARACTERISTICS

A colourless or faintly coloured liquid; caustic. Soluble in water; miscible with ethanol (96%), with ether and with glycerol.

IDENTIFICATION

A. Dissolve 0.6 g in 2 mL of 13.5M ammonia and dilute to 100 mL with water. To 2 mL of the resulting solution add 0.05 mL of sodium hypochlorite solution. A blue colour is produced which becomes progressively more intense. B. Dilute 1 mL of a 15% w/v solution to 10 mL and add 0.1 mL of iron(III) chloride solution R1. A violet colour is produced which is discharged on the addition of propan-2-ol. C. To 1 mL of a 15% w/v solution add 10 mL of water and 1 mL of bromine water. A white or yellowish white precipitate is produced.

TESTS

Acidity

To 2 mL of a 15% w/v solution add 0.05 mL of methyl orange solution. The solution is yellow.

Clarity and colour of solution

A solution of 1.0 mL in 14 mL of water, at 20°, is clear, Appendix IV A, and not more intensely coloured than reference solution R₇ or B₇, Appendix IV B, Method II.

Weight per mL

1.055 to 1.060 g, Appendix V G.

Non-volatile matter

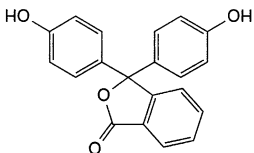
When heated on a water bath and dried at 105°, leaves not more than 0.05% w/v of residue.

ASSAY

Dissolve 2.5 g in sufficient water to produce 1000 mL. Transfer 25 mL to a 500 mL glass-stoppered flask and add 50 mL of 0.05M bromine VS and 5 mL of hydrochloric acid, stopper, swirl occasionally over a period of 30 minutes and allow to stand for 15 minutes. Add 5 mL of a 20% w/v solution of potassium iodide taking care to avoid loss of

Phenolphthalein

(Ph. Eur. monograph 1584)



C₂₀H₁₄O₄

318.3

77-09-8

Action and use

Stimulant laxative.

DEFINITION

Phenolphthalein contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of 3,3-bis(4-hydroxyphenyl)isobenzofuran-1(3H)-one, calculated with reference to the dried substance.

CHARACTERS

A white or almost white powder, practically insoluble in water, soluble in alcohol. It melts at about 260 °C.

IDENTIFICATION

A. Dissolve 25.0 mg in alcohol R and dilute to 100.0 mL with the same solvent (solution A). To 2.0 mL of solution A add 5.0 mL of 1 M hydrochloric acid and dilute to 50.0 mL with alcohol R (solution A₁). To 10.0 mL of solution A add 5.0 mL of 1 M hydrochloric acid and dilute to 50.0 mL with alcohol R (solution A₂). To 2.0 mL of solution A add 5.0 mL of 1 M sodium hydroxide and dilute to 50.0 mL with alcohol R. Examined between 220 nm and 250 nm (solution B). Examined between 220 nm and 250 nm (2.2.25), solution A₁ shows an absorption maximum at 229 nm. The specific absorbance at the maximum at 229 nm is 922 to 1018. Examined between 250 nm and 300 nm, solution A₂ shows an absorption maximum at 276 nm. The specific absorbance at the maximum at 276 nm is 142 to 158. Examined between 230 nm and 270 nm, solution B shows an absorption maximum at 249 nm. The specific absorbance at the maximum at 249 nm is 744 to 822. B. Dissolve about 10 mg in alcohol R. Add 1 mL of dilute sodium hydroxide solution R. The solution is red. Add 5 mL of dilute sulfuric acid R. The colour disappears.

LABELLING

The label states 'contains 80% phenol'. Liquefied Phenol should be protected from light. Liquefied Phenol may congeal or deposit crystals if stored at a temperature below 4°. It should be completely melted before use.

STORAGE

bromine, shake thoroughly and titrate with 0.1M sodium thiosulfate VS until only a faint yellow colour remains. Add 0.1 mL of starch mucilage and 10 mL of chloroform and complete the titration with vigorous shaking. Repeat the operation without the material being examined. The difference between the titrations represents the amount of bromine required. Each mL of 0.05M bromine VS is equivalent to 1.569 mg of C₆H₆O.



TESTS**Solution S**

To 2.0 g add 40 mL of distilled water R and heat to boiling. Cool and filter.

Appearance of solution

Dissolve 0.20 g in 5 mL of alcohol R. The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.15 mL of bromothymol blue solution R1. Add 0.05 mL of 0.01 M hydrochloric acid, the solution is yellow. Add 0.10 mL of 0.01 M sodium hydroxide, the solution is blue.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel F₂₅₄ plate R.

Test solution Dissolve 0.5 g of the substance to be examined in alcohol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dilute 1 mL of the test solution to 10 mL with alcohol R. Dilute 5 mL of this solution to 100 mL with alcohol R.

Reference solution (b) Dissolve 25 mg of fluorene R in alcohol R, add 0.5 mL of the test solution and dilute to 10 mL with alcohol R.

Apply to the plate 5 µL of the test solution and 5 µL of each of the reference solutions. Develop over a path corresponding to two-thirds of the plate height using a mixture of 50 volumes of acetone R and 50 volumes of methylene chloride R. Allow the plate to dry in air. Examine in ultraviolet light at 254 nm and re-examine after exposure to ammonia vapour. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (b) shows 2 clearly separated spots.

Chlorides (2.4.4)

Dilute 10 mL of solution S to 15 mL with water R. The solution complies with the limit test for chlorides (100 ppm).

Sulfates (2.4.13)

15 mL of solution S complies with the limit test for sulfates (200 ppm).

Heavy metals (2.4.8)

Heat 3 g with 50 mL of dilute hydrochloric acid R on a water-bath for 5 min and filter. Evaporate the filtrate almost to dryness and dissolve the residue in 30 mL of water R. 12 mL of this solution complies with test A for heavy metals (10 ppm). Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

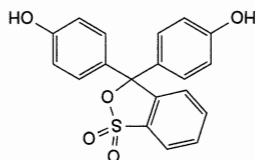
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 5 mL of dimethylformamide R. Add 5 mL of sodium carbonate solution R, 10 mL of sodium hydrogen carbonate solution R, 35 mL of water R and 50.0 mL of 0.05 M iodine. Add 10 mL of methylene chloride R and 20 mL of dilute sulfuric acid R. Titrate the excess of iodine with 0.1 M sodium thiosulfate, using 0.3 mL of starch solution R as indicator.

Phenolsulfonphthalein

(Ph. Eur. monograph 0242)



(Ph. Eur. monograph 0242)

C₁₉H₁₄O₅S

354.4

143-74-8

Action and use

Measurement of renal function; marker in drug absorption studies.

DEFINITION

Phenolsulfonphthalein (phenol red) contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of 3,3-bis(4-hydroxyphenyl)-3H-2,1-benzoxathiole 1,1-dioxide, calculated with reference to the dried substance.

CHARACTERS

A bright-red to dark-red, crystalline powder, very slightly soluble in water, slightly soluble in alcohol.

IDENTIFICATION

A. Dissolve 10 mg in a 10 g/L solution of sodium carbonate R and dilute to 200.0 mL with the sodium carbonate solution. Dilute 5.0 mL of the solution to 100.0 mL with a 10 g/L solution of sodium carbonate R. Examine between 400 nm and 630 nm (2.2.25), the solution shows an absorption maximum at 558 nm. The specific absorbance at the maximum is 1900 to 2100.

B. Dissolve about 10 mg in 1 mL of dilute sodium hydroxide solution R and add 9 mL of water R. The solution is deep red. To 5 mL of the solution add a slight excess of dilute sulfuric acid R. The colour becomes orange.

C. To 5 mL of the solution prepared for identification test B add 1 mL of 0.0167 M bromide-bromate and 1 mL of dilute hydrochloric acid R, shake and allow to stand for 15 min. Make alkaline with dilute sodium hydroxide solution R. An intense violet-blue colour is produced.

TESTS**Related substances**

Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance. **Test solution** Dissolve 0.1 g of the substance to be examined in 0.1 M sodium hydroxide and dilute to 5 mL with the same solvent. **Reference solution** Dilute 0.5 mL of the test solution to 100 mL with 0.1 M sodium hydroxide.

than 101.0% of $C_{18}H_{22}ClNO_2 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder.

Sparsely soluble in water; freely soluble in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of phenoxymbenzamine hydrochloride (RS 271).

B. Dissolve 0.5 g in 50 mL of ethanol-free chloroform and extract with three 20-mL quantities of 0.01M hydrochloric acid. Filter the chloroform layer through absorbent cotton and dilute 5 mL of the filtrate to 250 mL with ethanol-free chloroform. The light absorption of the resulting solution, Appendix II B, in the range 250 to 350 nm exhibits two maxima, at 272 nm and 279 nm. The absorbances at the maxima are about 1.1 and about 0.90, respectively.

C. Yields the reactions characteristic of chlorides, Appendix VI.

TESTS

Melting point

137.5° to 140°, Appendix V A.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in acetonitrile. (1) 0.08% w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 100 volumes.

(3) Dilute 1 volume of solution (2) to 10 volumes.

(4) To 10 volumes of solution (1) add 0.5 volume of 0.1M sodium hydroxide.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (15 cm × 4.6 mm) packed with octylsilyl silica gel for chromatography (5µm) (HiChrom Kromasil is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1 mL per minute.

(d) Use a column temperature of 25°.

(e) Use a detection wavelength of 268 nm.

(f) Inject 10 µL of each solution.

(g) Allow the chromatography to proceed for three times the retention time of phenoxymbenzamine.

MOBILE PHASE

45 volumes of a 0.22% w/v solution of anhydrous sodium dihydrogen orthophosphate, previously adjusted to pH 3.0 with orthophosphoric acid and 55 volumes of acetonitrile. When the chromatograms are recorded under the prescribed conditions, the relative retention with reference to phenoxymbenzamine (retention time = about 11 minutes) is: phenoxymbenzamine tertiary amine, about 0.2.

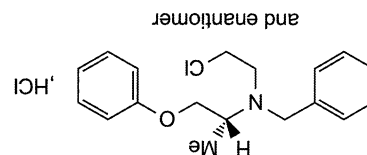
SYSTEM STABILITY

The test is not valid unless, in the chromatogram obtained with solution (4), the resolution factor between the peaks due to phenoxymbenzamine and the secondary peak with a relative retention of about 1.3 is at least 4.0.

LIMITS

In the chromatogram obtained with solution (1): the area of any peak corresponding to phenoxymbenzamine tertiary amine is not greater than the area of the principal

Phenoxymbenzamine Hydrochloride



$C_{18}H_{22}ClNO_2 \cdot HCl$

340.3

63-92-3

Sterile Phenoxymbenzamine Concentrate

Phenoxymbenzamine Capsules

Preparations

Alpha-adrenoceptor antagonist.

Action and use

Phenoxymbenzamine Hydrochloride is (RS)-benzyl(2-chloroethyl)-1-methyl-2-phenoxylethylamine hydrochloride. It contains not less than 98.5% and not more

of $C_{19}H_{14}O_5S$.

1 mL of 0.0167 M potassium bromate is equivalent to 4.43 mg

starch solution R as indicator.

Immediately with 0.1 M sodium thiosulfate, using 0.1 mL of

10 mL of a 100 g/L solution of potassium iodide R and titrate

acid R. Allow to stand protected from light for 15 min, add

solution of potassium bromide R and 5 mL of hydrochloric

20.0 mL of 0.0167 M potassium bromate, 5 mL of a 100 g/L

in a glass-stoppered flask add 25 mL of glacial acetic acid R,

dilute to 250.0 mL with water R. To 10.0 mL of the solution

Dissolve 0.900 g in 15 mL of 1 M sodium hydroxide and

ASSAY

Not more than 0.2 per cent, determined on 0.5 g.

Sulfated ash (2.4.14)

105 °C.

powdered substance to be examined by drying in an oven at

Not more than 1.0 per cent, determined on 1.00 g of the

Loss on drying (2.2.32)

(0.5 per cent).

105 °C. The residue weighs not more than 5 mg

carbonate R and then 25 mL of water R. Dry at 100 °C to

residue with 25 mL of a 10 g/L solution of sodium hydrogen

3000 g, for 30 min, decant the supernatant and wash the

water R and allow to stand for 15 h. Centrifuge at 2000 g to

stand for 1 h, shaking frequently. Dilute to 100 mL with

add 12 mL of sodium hydrogen carbonate solution R. Allow to

To 1.0 g of the finely powdered substance to be examined

Insoluble matter

(0.5 per cent).

the chromatogram obtained with the reference solution

solution and this spot is not more intense than the spot in

spot, appears in the chromatogram obtained with the test

254 nm. Not more than one spot, apart from the principal

concentrated ammonia R. Examine in ultraviolet light at

has evaporated and expose the plate to the vapour from

penyl alcohol R. Allow the plate to dry in air until the solvent

acetic acid R, 25 volumes of water R and 100 volumes of ter-

over a path of 15 cm using a mixture of 25 volumes of glacial

Apply separately to the plate 10 µL of each solution. Develop

CHARACTERS**Appearance**

Colourless, slightly viscous liquid.

Solubility

Slightly soluble in water, miscible with acetone, with ethanol (96 per cent) and with glycerol, slightly soluble in arachis oil and in olive oil.

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Refractive index (2.2.6): 1.537 to 1.539.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 80.0 mg in water R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with water R.

Spectral range 240–350 nm.

Absorption maxima At 269 nm and 275 nm.

Specific absorbances at the absorption maxima:

— at 269 nm: 95 to 105;

— at 275 nm: 75 to 85.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison phenoxyethanol CRS.

D. Shake 2 mL with a mixture of 4 g of potassium

permanganate R, 5.4 g of sodium carbonate R and 75 mL of

water R for 30 min. Add 25 g of sodium chloride R and stir

continuously for 60 min, filter and acidify with hydrochloric

acid R to about pH 1.7. The melting point of the precipitate,

after recrystallisation from water R, is 96 °C to 99 °C (2.2.14).

TESTS

Relative density (2.2.5)

1.105 to 1.110.

Related substances

Gas chromatography (2.2.28).

Internal standard solution Dissolve 1.25 g of methyl laurate R in methyl chloride R and dilute to 25 mL with the same

solvent.

Test solution (a) Dissolve 5.0 g of the substance to be

examined in methyl chloride R and dilute to 10.0 mL with

the same solvent.

Test solution (b) Dissolve 5.0 g of the substance to be

examined in methyl chloride R, add 1.0 mL of the internal

standard solution and dilute to 10.0 mL with methyl chloride R.

Reference solution To 1.0 mL of test solution (a) add 10.0 mL of the internal standard solution and dilute to 100.0 mL with methyl chloride R.

Column:

methylene chloride R.

— *material:* glass;

— *size:* l = 1.5 m, Ø = 4 mm,

— *stationary phase:* silanised diatomaceous earth for gas chromatography R (150–180 µm) impregnated with

3 per cent m/m of polydimethylsiloxane R.

Carrier gas nitrogen for chromatography R.

Flow rate 30 mL/min.

Temperature:

— *column:* 130 °C;

— *injection port and detector:* 200 °C.

Detection Flame ionisation.

Injection 1 µL.

peak in the chromatogram obtained with solution (2)

(1.0 %);

the area of any other *secondary peak* is not greater than

4 times the area of the principal peak in the chromatogram

obtained with solution (3) (0.4%);

the sum of the areas of all the *secondary peaks* is not greater

than 1.5 times the area of the principal peak in the

chromatogram obtained with solution (2) (1.5%).

Disregard any peak with an area less than the area of the

principal peak in the chromatogram obtained with solution

(3) (0.1%).

When dried over phosphorus pentoxide at a pressure not

exceeding 0.7 kPa for 24 hours, loses not more than 0.5% of

its weight. Use 1 g.

Sulphated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out the method for liquid chromatography,

Appendix III D, using the following solutions in acetonitrile.

(1) 0.02% w/v of the substance being examined.

(2) 0.02% w/v of phenoxybenzamine hydrochloride BPCRS.

(3) To 10 mL of solution (1) add 0.5 mL of 0.1M sodium

hydroxide.

CHROMATOGRAPHIC CONDITIONS

The chromatographic conditions described under Related

substances may be used.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained

with solution (3), the resolution factor between the peaks due

to phenoxybenzamine and the secondary peak with a relative

retention of about 1.3 is at least 4.0.

DETERMINATION OF CONTENT

Calculate the content of C₁₈H₂₂CINO₂HCl using the

declared content of C₁₈H₂₂CINO₂HCl in phenoxybenzamine

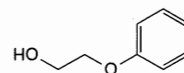
IMPURITIES

The impurities limited by the requirements of this

monograph include phenoxybenzamine tertiary amine.

Phenoxyethanol

(Ph. Eur. monograph 0781)



C₈H₁₀O₂

138.2

122-99-6

Action and use

Antimicrobial preservative; also used topically in treatment of bacterial infections.

Ph Eur

DEFINITION

2-Phenoxyethanol.

Content

99.0 per cent m/m to 100.5 per cent m/m.

Substance produced by the growth of certain strains of *Penicillium notatum* or related organisms on a culture medium containing an appropriate precursor, or obtained by any other means.

Content

95.0 per cent to 102.0 per cent for the sum of the percentage contents of phenoxymethylpenicillin and 4-hydroxyphenoxymethylpenicillin (anhydrous substance).

CHARACTERS

Appearance

White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Very slightly soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B

Second identification A, C, D

A. pH (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of acetone R.

Reference solution (a) Dissolve 25 mg of phenoxymethylpenicillin CRS in 5 mL of acetone R.

Reference solution (b) Dissolve 25 mg of benzylpenicillin potassium CRS and 25 mg of phenoxymethylpenicillin potassium CRS in 5 mL of water R.

Plate TLC silanised silica gel plate R.

Mobile phase Mix 30 volumes of acetone R and 70 volumes of a 154 g/L solution of ammonium acetate R adjusted to pH 5.0 with glacial acetic acid R.

Application 1 µL.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of water R and add 2 mL of sulfuric acid-formaldehyde reagent R. Mix the contents of the tube by swirling; the solution is reddish-brown. Place the test-tube on a water-bath for 1 min; a dark reddish-brown colour develops.

TESTS

pH (2.2.3)

2.4 to 4.0.

Suspend 50 mg in 10 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7)

+ 186 to + 200 (anhydrous substance).

Dissolve 0.250 g in butanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Run time 5 times the retention time of phenoxymethylpenicillin.

Elution order Phenoxymethylpenicillin, methyl laurate.

Retention time Phenoxymethylpenicillin = about 5 min.

System suitability:

— resolution: minimum 12 between the peaks due to phenoxymethylpenicillin and methyl laurate in the chromatogram

obtained with the reference solution;

— in the chromatogram obtained with test solution (a) there is no peak with the same retention time as the internal standard.

Limit:

— total: calculate the ratio (R) of the area of the peak due to phenoxymethylpenicillin to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with test solution (b), calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard; this ratio is not greater than R (1.0 per cent).

Phenol

Maximum 0.1 per cent.

Dissolve 1.00 g in 50 mL of methylene chloride R, add 1 mL of dilute sodium hydroxide solution R and 10 mL of water R.

Shake. Wash the upper layer with 2 quantities, each of 20 mL, of methylene chloride R and dilute to 100.0 mL with water R. The absorbance (2.2.25) of the solution measured at the absorption maximum at 287 nm is not greater than 0.27.

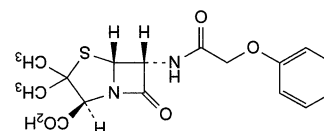
ASSAY

To 2.000 g in an acetylation flask fitted with an air condenser, add 10.0 mL of freshly prepared acetic anhydride solution R1 and heat with frequent shaking in a water-bath for 45 min. Cool and carefully add 10 mL of water R. Heat for a further 2 min. Cool, add 10 mL of butanol R, shake vigorously and titrate the excess of acetic acid with 1 M sodium hydroxide using 0.2 mL of phenolphthalein solution R as indicator. Repeat the procedure without the substance to be examined. The difference between the volumes used in the titrations represents the amount of acetic anhydride required for the acetylation of the substance to be examined.

1 mL of 1 M sodium hydroxide is equivalent to 0.1382 g of C₁₆H₁₈N₂O₅.

Phenoxymethylpenicillin

(Ph. Eur. monograph 0148)



C₁₆H₁₈N₂O₅S

350.4

87-08-1

Action and use

Penicillin antibacterial.

DEFINITION

(2S,5R,6R)-3,3-Dimethyl-7-oxo-6-[(phenoxymethyl)amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid.

- **signal-to-noise ratio:** minimum 3 for the principal peak in the chromatogram obtained with reference solution (d);
- **mass distribution ratio:** 5.0 to 7.0 for the peak due to phenoxyethylpenicillin (2^{nd} peak) in the chromatogram obtained with reference solution (c).

Limits:

- **any impurity:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (1 per cent);

— **disregard limit:** disregard the peak due to

4-hydroxyphenoxyethylpenicillin.

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile

phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (b).

Limit:

— 4-hydroxyphenoxyethylpenicillin: maximum 4.0 per cent

(anhydrous substance).

Calculate the percentage content by multiplying, if necessary, by the correction factor supplied with the CRS.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.000 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile

phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solutions (a) and (b).

System suitability: reference solution (a):

repeatability: maximum relative standard deviation of

1.0 per cent after 6 injections.

Calculate the percentage content of phenoxyethylpenicillin

by multiplying the percentage content of

phenoxyethylpenicillin potassium by 0.902. Calculate the

percentage content of 4-hydroxyphenoxyethylpenicillin by

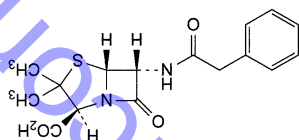
multiplying, if necessary, by the correction factor supplied

with the CRS.

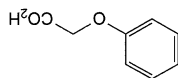
STORAGE

In an airtight container.

IMPURITIES



A. (2S,5R,6R)-3,3-dimethyl-7-oxo-6-[(phenylacetyl)amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (benzylpenicillin),



B. phenoxyacetic acid,

Dissolution mixture To 250 mL of 0.2 M potassium dihydrogen phosphate R add 500 mL of water R, adjust to pH 6.5 with an 8.4 g/L solution of sodium hydroxide R and dilute to 1000 mL with water R.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the dissolution mixture and dilute to 50.0 mL

with the dissolution mixture.

Test solution (b) Prepare immediately before use Dissolve

80.0 mg of the substance to be examined in the dissolution

mixture and dilute to 20.0 mL with the dissolution mixture.

Reference solution (a) Dissolve 55.0 mg of

phenoxyethylpenicillin potassium CRS in the dissolution

mixture and dilute to 50.0 mL with the dissolution mixture.

Reference solution (b) Dissolve 4.0 mg of

4-hydroxyphenoxyethylpenicillin potassium CRS in the

dissolution mixture and dilute to 10.0 mL with the

dissolution mixture. Dilute 5.0 mL of this solution to

100.0 mL with the dissolution mixture.

Reference solution (c) Dissolve 10 mg of phenoxyethylpenicillin

potassium CRS and 10 mg of benzylpenicillin sodium CRS

(impurity A) in the dissolution mixture and dilute to 50 mL

with the dissolution mixture.

Reference solution (d) Dilute 1.0 mL of reference solution (a)

to 20 mL with the dissolution mixture. Dilute 1.0 mL of this

solution to 50 mL with the dissolution mixture.

Reference solution (e) Dilute 1.0 mL of reference solution (a)

to 25.0 mL with the dissolution mixture.

Column:

— **size:** $l = 0.25$ m, $\varnothing = 4.6$ mm;

— **stationary phase:** octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase:

— **mobile phase A:** phosphate buffer solution pH 3.5 R,

methanol R (10:30:60 V/V/V);

— **mobile phase B:** phosphate buffer solution pH 3.5 R, **water R**,

methanol R (10:35:55 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - t_R	60	40
$t_R - (t_R + 20)$	60 \rightarrow 0	40 \rightarrow 100
$(t_R + 20) - (t_R + 35)$	0	100
$(t_R + 35) - (t_R + 50)$	0 \rightarrow 60	100 \rightarrow 40

t_R = retention time of phenoxyethylpenicillin determined with reference solution (d)

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of reference solutions (c), (d) and (e) with

isocratic elution at the initial mobile phase composition and

20 μ L of test solution (b) according to the elution gradient

described under Mobile phase; inject the dissolution mixture

as a blank according to the elution gradient described under

Mobile phase.

System suitability:

— **resolution:** minimum 6.0 between the peaks due to

impurity A and phenoxyethylpenicillin in the

chromatogram obtained with reference solution (c);

if necessary, adjust the ratio A:B of the mobile phases;

Content
95.0 per cent to 102.0 per cent for the sum of the percentage contents of phenoxyethylpenicillin potassium and 4-hydroxyphenoxyethylpenicillin potassium (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, D

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison phenoxyethylpenicillin potassium CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of water R.

Reference solution (a) Dissolve 25 mg of phenoxyethylpenicillin potassium CRS in 5 mL of water R.

Reference solution (b) Dissolve 25 mg of benzylpenicillin potassium CRS and 25 mg of phenoxyethylpenicillin potassium CRS in 5 mL of water R.

Plate TLC silanised silica gel plate R.

Mobile phase Mix 30 volumes of acetone R and 70 volumes of a 154 g/L solution of ammonium acetate R adjusted to pH 5.0 with glacial acetic acid R.

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability: reference solution (b):

the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of water R and add 2 mL of sulfuric acid-formaldehyde reagent R. Mix the contents of the tube by swirling; the solution is reddish-brown. Place the test-tube in a water-bath for 1 min; a dark reddish-brown colour develops.

D. It gives reaction (a) of potassium (2.3.1).

TESTS

pH (2.2.3)

5.5 to 7.5.

Dissolve 50 mg in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Specific optical rotation (2.2.7)

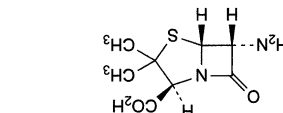
+215 to +230 (anhydrous substance).

Dissolve 0.250 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Related substances

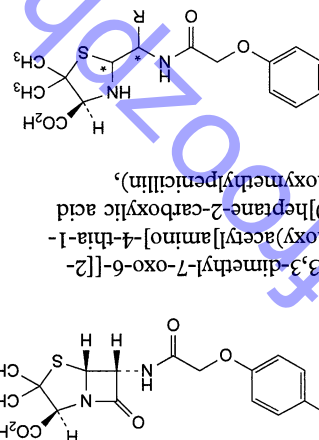
Liquid chromatography (2.2.29).

Dissolution mixture T to 250 mL of 0.2 M potassium dihydrogen phosphate R add 500 mL of water R and adjust to pH 6.5



C. (2S,5R,6R)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),

D. (2S,5R,6R)-3,3-dimethyl-7-oxo-6-[[2-(4-hydroxyphenoxy)acetyl]amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (4-hydroxyphenoxyethylpenicillin),



E. R = CO₂H: (4S)-2-

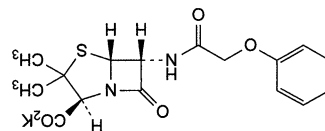
[carboxy][(phenoxycarbonyl)amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of phenoxyethylpenicillin),

F. R = H: (2R,4S)-5,5-dimethyl-2-

[(phenoxycarbonyl)amino]methyl]thiazolidine-4-carboxylic acid (penicilloic acids of phenoxyethylpenicillin).

Potassium Phenoxyethylpenicillin

(Ph. Eur. monograph 0149)



C₁₆H₁₇KN₂O₅S

388.5

132-98-9

Action and use

Penicillin antibacterial.

Preparations

Phenoxyethylpenicillin Oral Solution

Phenoxyethylpenicillin Tablets

DEFINITION

Potassium salt of (2S,5R,6R)-3,3-dimethyl-7-oxo-6-[(phenoxycarbonyl)amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid. Substance produced by the growth of certain strains of *Penicillium notatum* or related organisms on a culture medium containing an appropriate precursor, or obtained by any other means.

— *mass distribution ratio*: 5.0 to 7.0 for the peak due to phenoxyethylpenicillin (2nd peak) in the chromatogram obtained with reference solution (c).

Limits:

— *any impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (1 per cent);

— *disregard limit*: disregard the peak due to 4-hydroxyphenoxyethylpenicillin.

4-Hydroxyphenoxyethylpenicillin potassium

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (b).

Limit:

— 4-hydroxyphenoxyethylpenicillin potassium: maximum 4.0 per cent (anhydrous substance).

Calculate the percentage content by multiplying, if necessary, by the correction factor supplied with the CRS.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.000 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

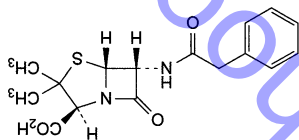
Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solutions (a) and (b). *System suitability*: reference solution (a);

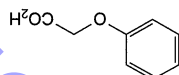
— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of phenoxyethylpenicillin potassium and of 4-hydroxyphenoxyethylpenicillin

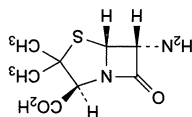
IMPURITIES



A. (2S,5R,6R)-3,3-dimethyl-7-oxo-6-[(phenylacetyl)amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (benzylpenicillin),



B. phenoxyacetic acid,



C. (2S,5R,6R)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),

with an 8.4 g/L solution of sodium hydroxide R. Dilute to 1000 mL with water R.

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the dissolution mixture and dilute to 50.0 mL with the dissolution mixture.

Test solution (b) Prepare immediately before use. Dissolve 80.0 mg of the substance to be examined in the dissolution mixture and dilute to 20.0 mL with the dissolution mixture.

Reference solution (a) Dissolve 50.0 mg of phenoxyethylpenicillin potassium CRS in the dissolution mixture and dilute to 50.0 mL with the dissolution mixture.

Reference solution (b) Dissolve 4.0 mg of 4-hydroxyphenoxyethylpenicillin potassium CRS in the dissolution mixture and dilute to 10.0 mL with the dissolution mixture. Dilute 5.0 mL of this solution to 100.0 mL with the dissolution mixture.

Reference solution (c) Dissolve 10 mg of phenoxyethylpenicillin potassium CRS and 10 mg of benzylpenicillin sodium CRS (impurity A) in the dissolution mixture and dilute to 50 mL with the dissolution mixture.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 20 mL with the dissolution mixture. Dilute 1.0 mL of this solution to 50 mL with the dissolution mixture.

Reference solution (e) Dilute 1.0 mL of reference solution (a) to 25.0 mL with the dissolution mixture.

Column:

— *size*: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— *stationary phase*: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

— *mobile phase A*: phosphate buffer solution pH 3.5 R,

methanol R, water R (10:30:60 V/V/V);

— *mobile phase B*: phosphate buffer solution pH 3.5 R, water R, *methanol* R (10:35:55 V/V/V);

t _R = retention time of phenoxyethylpenicillin determined with reference solution (d)		
Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - t _R	60	40
t _R - (t _R + 20)	60 → 0	40 → 100
(t _R + 20) - (t _R + 35)	0	100
(t _R + 35) - (t _R + 50)	0 → 60	100 → 40

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of reference solutions (c), (d) and (e) with isocratic elution at the initial mobile phase composition and 20 μ L of test solution (b) according to the elution gradient described under *Mobile phase*;

as a blank according to the elution gradient described under *Mobile phase*.

System suitability:

— *resolution*: minimum 6.0 between the peaks due to impurity A and phenoxyethylpenicillin in the chromatogram obtained with reference solution (c);

if necessary, adjust the ratio A:B of the mobile phases; — *signal-to-noise ratio*: minimum 3 for the principal peak in the chromatogram obtained with reference solution (d);

CHARACTERS**Appearance**

White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification C, E

Second identification A, B, D, E.

A. Melting point (2.2.14): 178 °C to 182 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 60.0 mg in water R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with water R.

Spectral range 230-350 nm.

Absorption maximum At 278 nm.

Specific absorbance at the absorption maximum 220 to 245.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison phentolamine mesilate CRS.

D. Dissolve 0.5 g in a mixture of 5 mL of ethanol (96 per cent) R and 5 mL of a 10 g/L solution of hydrochloric acid R and add 0.5 mL of a 5 g/L solution of ammonium vanadate R. A light green precipitate is produced.

E. Mix 50 mg with 0.2 g of sodium hydroxide R, heat to fusion and continue heating for a few seconds. Allow to cool and add 0.5 mL of warm water R. Acidity with dilute hydrochloric acid R and heat. Sulfur dioxide is evolved, which turns moistened starch iodate paper R blue.

TESTS

Acidity Dissolve 0.1 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent. Add 0.1 mL of methyl red solution R. If the solution is red, not more than 0.05 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to yellow.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of phentolamine for system suitability CRS (containing impurities A and C) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: phenylsilyl silica gel for chromatography R1 (5 μ m);

— temperature: 30 °C.

Mobile phase Mix 33 volumes of acetonitrile R1 and 67 volumes of a 0.5 g/L solution of ammonium acetate R previously adjusted to pH 5.9 with dilute acetic acid R.

Flow rate 1.5 mL/min.

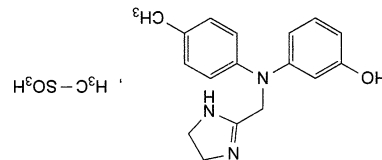
Detection Spectrophotometer at 230 nm.

Injection 10 μ L.

Run time 1.5 times the retention time of phentolamine.

Phentolamine Mesilate

(Ph. Eur. monograph 1138)



$C_{18}H_{23}N_3O_4S$ 377.5 65-28-1

Preparation

Phentolamine Injection

Action and use

Alpha-adrenoreceptor antagonist.

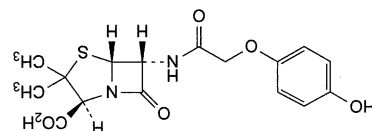
Content

98.0 per cent to 101.0 per cent (dried substance).

PRODUCTION

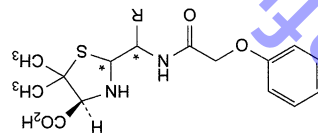
It is considered that alkylsulfonate esters are genotoxic and are potential impurities in phentolamine mesilate.

The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods 2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid, 2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances and 2.5.39. Methanesulfonyl chloride in methanesulfonic acid are available to assist manufacturers.



D. (2S,5R,6R)-3,3-dimethyl-7-oxo-6-[[2-(4-hydroxyphenoxy)acetyl]amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (4-hydroxyphenoxy)methylpenicillin),

E. R = CO₂H: (4S)-2-[carboxy[[[phenoxyacetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of phenoxy)methylpenicillin), F. R = H: (2R,5,4S)-5,5-dimethyl-2-[[[phenoxyacetyl]amino]methyl]thiazolidine-4-carboxylic acid (penicilloic acids of phenoxy)methylpenicillin).



Identification of impurities Use the chromatogram supplied with phenylalanine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and C.

Relative retention With reference to phenylalanine (retention time = about 15 min): impurity A = about 0.7; impurity C = about 1.2.

System suitability: reference solution (b):
— **resolution:** minimum 3.0 between the peaks due to phenylalanine and impurity C.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity A by 1.7;

— **impurity A:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a)

— **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

— **total:** not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.5 per cent);
— **disregard limit:** 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).
Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 100 mL of 2-propanol R1. Titrate under a stream of nitrogen with 0.1 M tetrabutylammonium hydroxide in 2-propanol. Determine the end-point potentiometrically (2.2.20), using a glass indicator electrode and a calomel reference electrode containing a saturated solution of

tetramethylammonium chloride R in 2-propanol R1. Carry out a blank titration.

1 mL of 0.1 M tetrabutylammonium hydroxide in 2-propanol is equivalent to 37.75 mg of $C_{18}H_{23}N_3O_4S$.

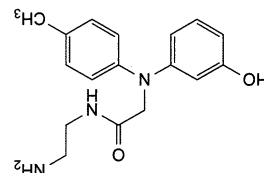
STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A

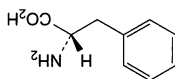
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C.



A. N-(2-aminoethyl)-2-[(3-hydroxyphenyl)(4-methylphenyl)amino]acetamide,

Phenylalanine

(Ph. Eur. monograph 0782)



$C_9H_{11}NO_2$ 165.2

63-91-2

Action and use

Amino acid.

Ph. Eur.

DEFINITION

(2S)-2-Amino-3-phenylpropanoic acid.

Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder, or shiny, white flakes.

Solubility

Sparsely soluble in water, very slightly soluble in ethanol (96 per cent). It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

C. Comparison phenylalanine CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of equal volumes of glacial acetic acid R and water R and dilute to 50 mL with the same mixture of solvents.

Reference solution. Dissolve 10 mg of phenylalanine CRS in a mixture of equal volumes of glacial acetic acid R and water R and dilute to 50 mL with the same mixture of solvents.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the Test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To about 10 mg add 0.5 g of potassium nitrate R and 2 mL of sulfuric acid R. Heat on a water-bath for 20 min. Allow to cool. Add 5 mL of a 50 g/L solution of hydroxylamine hydrochloride R and allow to stand in iced water for 10 min. Add 9 mL of strong sodium hydroxide solution R. A violet-red or violet-brown colour develops.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Dissolve 0.5 g in a 103 g/L solution of hydrochloric acid R and dilute to 10 mL with the same solution.

Specific optical rotation (2.2.7)

–35.5 to –33.0 (dried substance).
Dissolve 0.50 g in water R and dilute to 25.0 mL with the same solvent.

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.
Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of proline R in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of ammonium standard solution (100 ppm NH₄) R to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of isoleucine R and 30 mg of leucine R in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability: Reference solution (d):
— resolution: minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of phenylalanine in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b);
if a peak is above the reporting threshold at both

Limits:

— any ninhydrin-positive substance: for each impurity, maximum 0.2 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances

(Table 2034.-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Chlorides (2.4.4)

Dissolve 0.25 g in 3 mL of dilute nitric acid R and dilute to 15 mL with water R. The solution, without further addition of nitric acid, complies with the test.

Sulfates (2.4.13)

Maximum 300 ppm.
Dissolve 0.5 g in a mixture of 5 volumes of dilute hydrochloric acid R and 25 volumes of distilled water R and dilute to 15 mL with the same mixture of solvents.

Ammonium

Amino acid analysis (2.2.56) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:
— ammonium at 570 nm: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (2.4.9)

Maximum 10 ppm.
In a separating funnel, dissolve 1.0 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.
2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 3 mL of anhydrous formic acid R. Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 16.52 mg of C₉H₁₁NO₂.

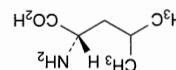
STORAGE

Protected from light.

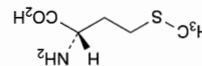
IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph). They are limited by the general

acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: A, B, C, D.

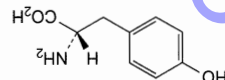


A. (2S)-2-amino-4-methylpentanoic acid (leucine),

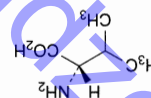


B. (2S)-2-amino-4-(methylsulfonyl)butanoic acid

(methionine),



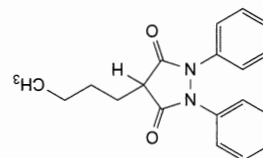
C. (2S)-2-amino-3-(4-hydroxyphenyl)propanoic acid (tyrosine),



D. (2S)-2-amino-3-methylbutanoic acid (valine).

Phenylbutazone

(Ph. Eur. monograph 0422)



$C_{19}H_{20}N_2O_2$

308.4

50-33-9

Action and use
Cyclo-oxygenase inhibitor; pyrazolone analgesic.

DEFINITION

4-Butyl-1,2-diphenylpyrazolidine-3,5-dione.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in alcohol.

It dissolves in alkaline solutions.

IDENTIFICATION

First identification A, C.

Second identification A, B, D.

A. Melting point (2.2.14): 104 °C to 107 °C.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in a mixture of equal volumes of ethanol R and methylene chloride R and dilute to 25 mL with the same mixture of solvents.

Reference solution Dissolve 25 mg of phenylbutazone CRS in a mixture of equal volumes of ethanol R and methylene chloride R and dilute to 25 mL with the same mixture of solvents.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase acetone R, methylene chloride R (20:80 V/V).

Application 5 µL.

Development Over a path of 10 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison phenylbutazone CRS.

D. To 0.1 g add 1 mL of glacial acetic acid R and 2 mL of

hydrochloric acid R and heat the mixture under a reflux condenser for 30 min. Cool, add 10 mL of water R and filter.

To the filtrate add 3 mL of a 7 g/L solution of sodium nitrite R. A yellow colour is produced. To 1 mL of the

solution add a solution of 10 mg of β-naphthol R in 5 mL of sodium carbonate solution R. A brownish-red to violet-red

precipitate is formed.

TESTS

Solution S

Dissolve 1.0 g with shaking in 20 mL of dilute sodium hydroxide solution R and maintain the solution at 25 °C for

3 h.

Appearance of solution

Solution S is clear (2.2.1).

Acidity or alkalinity

Heat to boiling 1.0 g in 50 mL of water R, cool with shaking

in a closed flask and filter. To 25 mL of the filtrate add 0.5 mL of phenolphthalein solution R. The solution is

colourless. Not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator.

Add 0.6 mL of 0.01 M hydrochloric acid and 0.1 mL of methyl red solution R; the solution is red or orange.

Absorbance (2.2.25)

Maximum 0.20 for solution S at 420 nm in a 4 cm cell.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Test solution Dissolve 100.0 mg of the substance to be examined in acetone/nitrite R and dilute to 10.0 mL with the

same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with acetone/nitrite R. Dilute 1.0 mL to 10.0 mL with

acetone/nitrite R.

Reference solution (b) Dissolve 5 mg of phenylbutazone

impurity B CRS and 5 mg of 1,2-diphenylhydrazine R in

acetone/nitrite R, add 0.5 mL of the test solution and dilute to

50 mL with acetone/nitrite R. Dilute 2.5 mL to 10 mL with

acetone/nitrite R.

Reference solution (c) Dissolve 1.0 mg of benzidine R in acetone/nitrite R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL to 100.0 mL with acetone/nitrite R. Dilute 5.0 mL

to 10.0 mL with acetone/nitrite R.

Column: size: l = 0.125 m, Ø = 4.0 mm,

- stationary phase: octadecylsilyl silica gel for chromatography R
- temperature: 30 °C.
- Mobile phase:
 - mobile phase A: dissolve 1.36 g of sodium acetate R in water R, adjust to pH 5.2 with a 52.5 g/L solution of citric acid R and dilute to 1000 mL with water R,
 - mobile phase B: acetonitrile R,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	70	30
10 - 20	70 → 40	30 → 60
20 - 35	40	60
35 - 40	40 → 70	60 → 30

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 240 nm.

Injection 20 µL; inject the test solution and reference solutions (a) and (b).

Relative retentions With reference to phenylbutazone (retention time = about 13 min): impurity B = about 0.2; impurity A = about 0.5; impurity B = about 1.2; impurity C = about 1.3; impurity D = about 1.7.

System suitability: reference solution (b):
— resolution: minimum 2.0 between the peaks due to phenylbutazone and to impurity B.

Limits:
— correction factor for the calculation of content, multiply the peak area of impurity C by 0.55,
— impurities A, B: for each impurity, not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent),
— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— any other impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),
— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.025 per cent); disregard any peak due to impurity E.

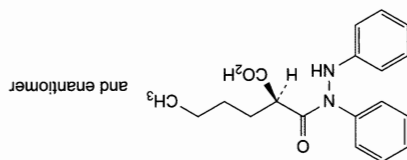
Impurity E
Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.
Detection Spectrophotometer at 280 nm.

Injection Test solution and reference solution (c).
System suitability: reference solution (c):
— signal-to-noise ratio: minimum 10 for the principal peak.

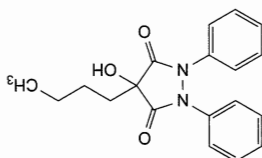
Limits:
— impurity E: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (5 ppm).

Heavy metals (2.4.8)
Maximum 20 ppm.

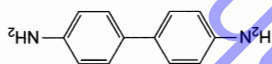
1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.



A. (2RS)-2-[(1,2-diphenyldiazany)carbonyl]hexanoic acid,



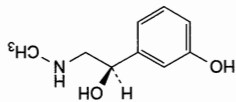
- B. 4-benzyl-4-hydroxy-1,2-diphenylpyrazolidine-3,5-dione, C. C₆H₅-NH-NH-C₆H₅; 1,2-diphenyldiazane (1,2-diphenylhydrazine), D. C₆H₅-N=N-C₆H₅; 1,2-diphenyldiazene,



E. biphenyl-4,4'-diamine (benzidine).

Phenylephrine

(Ph. Eur. monograph 1035)



C₉H₁₃NO₂

167.2

59-42-7

Action and use
Alpha-adrenoceptor agonist.

DEFINITION

(1R)-1-(3-Hydroxyphenyl)-2-(methylamino)ethanol.

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent). It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

mp

About 174 °C.

IDENTIFICATION

First identification A, B.

Second identification A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison phenylephrine CRS.

C. Thin-layer chromatography (2.2.27).

Solvent mixture A: mixture of equal volumes of methylene chloride R and methanolic hydrochloric acid (hydrochloric acid R diluted 10-fold with methanol R).

Test solution Dissolve 0.1 g of the substance to be examined in the solvent mixture and dilute to 5 mL with the solvent mixture.

Reference solution Dissolve 20 mg of phenylephrine CRS in the solvent mixture and dilute to 1 mL with the solvent mixture.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, methanol R, methylene chloride R (0.5:25:70 V/V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In a current of cold air.

Detection Examine in ultraviolet light at 254 nm; spray with a 1 g/L solution of fast red B salt R in a 50 g/L solution of sodium carbonate R and examine in daylight.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 10 mg in 1 mL of 1 M hydrochloric acid, add 0.05 mL of copper sulfate solution R and 1 mL of a 200 g/L solution of sodium hydroxide R. A violet colour develops. Add 1 mL of ether R and shake. The upper layer remains colourless.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Dissolve 1 g in 1 M hydrochloric acid and dilute to 10 mL with the same acid.

Specific optical rotation (2.2.7)

— 53 to –57 (dried substance).

Dissolve 1.250 g in 1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture dilute hydrochloric acid R, mobile phase B, mobile phase A (5:200:800 V/V/V).

Buffer solution pH 2.8 Dissolve 3.25 g of sodium octanesulfonate monohydrate R in 1000 mL of water R by stirring for 30 min and adjust to pH 2.8 with dilute phosphoric acid R.

Test solution Dissolve 41.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of phenylephrine hydrochloride for peak identification CRS (containing impurities C and E) in 2.0 mL of the solvent mixture.

Column:

— size: l = 0.055 m, Ø = 4.0 mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 µm);

— temperature: 45 °C.

Mobile phase:

— mobile phase A: acetonitrile R₁, buffer solution pH 2.8 (10:90 V/V);

— mobile phase B: buffer solution pH 2.8, acetonitrile R₁ (10:90 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	93	7
3 - 13	93 → 70	7 → 30
13 - 14	70 → 93	30 → 7

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 µL.

Relative retention With reference to phenylephrine (retention time = about 2.8 min): impurity C = about 1.3;

impurity E = about 3.6.

System suitability:

— symmetry factor: maximum 1.9 for the principal peak in the chromatogram obtained with the test solution;

— peak-to-valley ratio: minimum 5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to phenylephrine in the chromatogram obtained with reference solution (b).

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.5;

impurity E = 0.5;

— impurities C, E: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 60 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 16.72 mg

of $C_9H_{13}NO_2$.

STORAGE

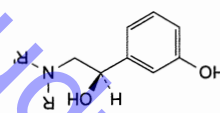
In an airtight container, protected from light.

IMPURITIES

Specified impurities C, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, D.



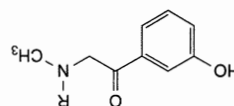
A, R = R' = H: (1R)-2-amino-1-(3-hydroxyphenyl)ethanol

(norphenylephrine),

D, R = $CH_2-C_6H_5$, R' = CH_3 : (1R)-2-

(benzylmethylamino)-1-(3-hydroxyphenyl)ethanol

(benzylphenylephrine),



C, R = H: 1-(3-hydroxyphenyl)-2-(methylamino)ethanone

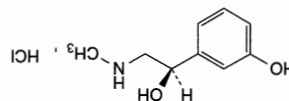
(phenylephrone),

E, R = $CH_2-C_6H_5$; 2-(benzylmethylamino)-1-

(3-hydroxyphenyl)ethanone (benzylphenylephrone).

Phenylephrine Hydrochloride

(Ph. Eur. monograph 0632)



$C_9H_{11}ClNO_2$

203.7

61-76-7

Action and use

Alpha-adrenoceptor agonist.

Preparations

Phenylephrine Eye Drops

Phenylephrine Injection

Phenylephrine Intracameral Injection

DEFINITION
(1R)-1-(3-Hydroxyphenyl)-2-(methylamino)ethanol hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in ethanol (96 per cent).

mp: about 143 °C.

IDENTIFICATION

First identification A, C, E.

Second identification A, B, D, E.

A. Specific optical rotation (see Tests).

B. Melting point (2.2.14): 171 °C to 176 °C.

Dissolve 0.3 g in 3 mL of water R, add 1 mL of dilute ammonia R1 and initiate crystallisation by scratching the wall of the tube with a glass rod. Wash the crystals with iced water R and dry at 105 °C for 2 h.

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison phenylephrine hydrochloride CRS.

D. Dissolve about 10 mg in 1 mL of water R and add 0.05 mL of a 125 g/L solution of copper sulfate R and 1 mL of a 200 g/L solution of sodium hydroxide R. A violet colour is produced. Add 1 mL of ether R and shake; the upper layer remains colourless.

E. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S Dissolve 2.00 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of methyl red solution R and 0.2 mL of 0.01 M sodium hydroxide. The solution is yellow. Not more than 0.4 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to red.

Specific optical rotation (2.2.7)

−43 to −47 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase B, mobile phase A (20:80 V/V).

Buffer solution pH 2.8 Dissolve 3.25 g of sodium octanesulfonate monohydrate R in 1000 mL of water R by stirring for 30 min and adjust to pH 2.8 with dilute phosphoric acid R.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

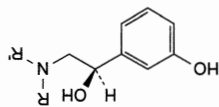
Reference solution (b) Dissolve the contents of a vial of phenylephrine hydrochloride for peak identification CRS (containing impurities C and E) in 2.0 mL of the solvent mixture.

1 mL of 0.1 M ethanolic sodium hydroxide is equivalent to 20.37 mg of $C_{10}H_{14}ClNO_2$.

IMPURITIES

Specified impurities C, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D.



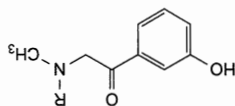
A, R = R' = H: (1R)-2-amino-1-(3-hydroxyphenyl)ethanol

(norphenylephrine),

D, R = $CH_2-C_6H_5$, R' = CH_3 : (1R)-2-

(benzylmethylethylamino)-1-(3-hydroxyphenyl)ethanol

(benzylphenylephrine),



C, R = H: 1-(3-hydroxyphenyl)-2-(methylamino)ethanol

(phenylephrine),

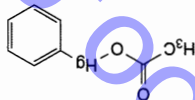
E, R = $CH_2-C_6H_5$; 2-(benzylmethylethylamino)-1-

(3-hydroxyphenyl)ethanol (benzylphenylephrine).

Ph Eur

Phenylmercuric Acetate

(Ph. Eur. monograph 2042)



$C_{10}H_{14}HgO_2$

336.7

62-38-4

Action and use

Antiseptic, antimicrobial preservative.

Ph Eur

DEFINITION

Content

98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish, crystalline powder or small, colourless crystals.

Solubility

Slightly soluble in water, soluble in acetone and in alcohol.

IDENTIFICATION

First identification A

Second identification B, C

Column: — size: $l = 0.055$ m, $\phi = 4.0$ mm; — stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m); — temperature: 45 °C. — Mobile phase: — mobile phase A: acetonitrile R1, buffer solution pH 2.8 (10:90 V/V); — mobile phase B: buffer solution pH 2.8, acetonitrile R1 (10:90 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0-3	93	7
3-13	93 \rightarrow 70	7 \rightarrow 30
13-14	70 \rightarrow 93	30 \rightarrow 7

Flow rate 1.5 mL/min. Detection Spectrophotometer at 215 nm. Injection 10 μ L.

Relative retention With reference to phenylephrine (retention time = about 2.8 min): impurity C = about 1.3; impurity E = about 3.6.

System suitability: — symmetry factor: maximum 1.9 for the principal peak in the chromatogram obtained with the test solution; — peak-to-valley ratio: minimum 5, where H_p = height above the baseline of the peak due to impurity C and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to phenylephrine in the chromatogram obtained with reference solution (b).

Limits:

— the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.5; impurity E = 0.5; — impurities C, E: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent); — unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent); — total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent); — disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Sulfates (2.4.13)

Maximum 500 ppm, determined on solution S.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in a mixture of 0.5 mL of 0.1 M hydrochloric acid and 80 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20) using 0.1 M ethanolic sodium hydroxide. Read the volume added between the 2 points of inflexion.

A. Infrared absorption spectrophotometry (2.2.24).
 Comparison Ph. Eur. reference spectrum of phenylmercuric acetate.
 B. To 5 mL of solution S (see Tests) add 5 mL of water R
 and 0.1 mL of sodium sulfide solution R. A white precipitate is
 formed that darkens slowly on heating.
 C. To 10 mL of solution S add 2 mL of potassium iodide
 solution R and shake vigorously. Filter. The filtrate gives
 reaction (b) of acetates (2.3.1).

TESTS

Solution S
 Dissolve 0.250 g in 40 mL of water R by heating to boiling.
 Allow to cool and dilute to 50 mL with water R. Prepare the
 solution immediately before use.

Appearance of solution

Solution S is not more opalescent than reference
 suspension II (2.2.1) and is colourless (2.2.2, Method II).

Ionised mercury

Maximum 0.2 per cent.

To 2 mL of solution S add 8 mL of water R, 2 mL of
 potassium iodide solution R and 3 mL of dilute hydrochloric
 acid R. Filter. The filtrate is not more coloured than the
 3 mL of water R. Combine the filtrate and the washings, add
 2 mL of dilute sodium hydroxide solution R and dilute to
 20 mL with water R. 12 mL of this solution complies with
 test A for heavy metals (2.4.8). Prepare the reference solution
 using lead standard solution (1 ppm Pb) R.

Polymercuric benzene compounds

Maximum 1.5 per cent.

Shake 0.2 g with 10 mL of acetone R. Filter. Wash the
 residue twice with 5 mL of acetone R. Dry the residue at
 105 °C for 1 h. The residue weighs a maximum of 3 mg.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 0.500 g by drying in
 an oven at 45 °C for 15 h.

ASSAY

Dissolve with heating 0.300 g in 100 mL of water R. Cool
 and add 3 mL of nitric acid R. Titrate with 0.1 M ammonium
 thiocyanate using 2 mL of ferric ammonium sulfate solution R2
 as indicator, until a persistent reddish-yellow colour is
 obtained.

1 mL of 0.1 M ammonium thiocyanate is equivalent to
 33.67 mg of phenylmercuric acetate.

STORAGE

Protected from light.

Phenylmercuric Borate

(Ph. Eur. monograph 0103)

Action and use

Antiseptic; antimicrobial preservative.

DEFINITION

Compound consisting of equimolecular proportions of
 phenylmercuric orthoborate and phenylmercuric hydroxide
 ($C_{12}H_{13}BHg_2O_4$; 633) or of the dehydrated form
 (metaborate, $C_{12}H_{11}BHg_2O_3$; 615) or of a mixture of the
 2 compounds.



Content

— mercury (Hg; A_1 200.6): 64.5 per cent to 66.0 per cent
 (dried substance),
 — borates expressed as Hg_3BO_3 : 9.8 per cent to 10.3 per cent
 (dried substance).

CHARACTERS

Appearance
 White or slightly yellowish, crystalline powder or colourless,
 shiny crystals.

Solubility

Slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
 Preparation Discs.
 Comparison Ph. Eur. reference spectrum of phenylmercuric borate.
 B. To 2 mL of solution S (see Tests) add 8 mL of water R
 and 0.1 mL of sodium sulfide solution R. A white precipitate is
 formed that darkens slowly on heating.
 C. Dissolve about 20 mg in 2 mL of methanol R.
 The solution is clear and colourless. Ignite; the solution
 burns with a green-edged flame.

TESTS

Solution S
 Dissolve 0.25 g by sprinkling it on the surface of 25 mL of
 boiling water R, cool and dilute to 25 mL with water R.
Appearance of solution
 Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Ionised mercury

Maximum 0.01 per cent.

To 10 mL of solution S add 2 mL of potassium iodide
 solution R and 3 mL of dilute hydrochloric acid R. Filter.
 The filtrate is colourless. Wash the precipitate with 3 mL of
 water R. Combine the filtrate and the washings, add 2 mL of
 dilute sodium hydroxide solution R and dilute to 20 mL with
 water R. 12 mL of this solution complies with test A for
 heavy metals (2.4.8). Prepare the reference solution using a
 mixture of 2.5 mL of lead standard solution (2 ppm Pb) R and
 7.5 mL of water R.

Loss on drying (2.2.32)

Maximum 3.5 per cent, determined on 0.50 g by drying in
 an oven at 45 °C for 15 h ± 30 min.

ASSAY

Dissolve 0.300 g in 100 mL of water R and add 3 mL of
 nitric acid R. Titrate with 0.1 M ammonium thiocyanate, using
 2 mL of ferric ammonium sulfate solution R2 as indicator, until
 a persistent reddish-yellow colour is obtained.
 1 mL of 0.1 M ammonium thiocyanate is equivalent to
 20.06 mg of Hg.

Borates

Dissolve 0.600 g with heating in 25 mL of water R. Dissolve
 10 g of sorbitol R in the hot solution and cool. Titrate with
 0.1 M sodium hydroxide, using 0.5 mL of phenolphthalein
 solution R as indicator, until a persistent pink colour is
 obtained. Carry out a blank titration.
 1 mL of 0.1 M sodium hydroxide is equivalent to 6.18 mg of
 Hg_3BO_3 .

STORAGE

Protected from light.

Phenylmercuric Nitrate

(Ph. Eur. monograph 0783)

Action and use

Antiseptic; antimicrobial preservative.

DEFINITION

Mixture of phenylmercuric nitrate ($C_6H_5HgNO_3$; 339.7) and phenylmercuric hydroxide (C_6H_5HgOH ; 294.7).

Content

62.5 per cent to 64.0 per cent of Hg (A_1 , 200.6) (dried substance).

CHARACTERS

Appearance

White or pale yellow powder.

Solubility

Very slightly soluble in water and in ethanol (96 per cent), slightly soluble in hot water. It dissolves in glycerol and in fatty oils.

IDENTIFICATION

A. To 5 mL of solution S (see Tests) add 8 mL of water R and 0.1 mL of sodium sulfide solution R. A white precipitate is formed that darkens slowly on heating.

B. To 1 mL of a saturated solution of the substance to be examined add 1 mL of dilute hydrochloric acid R. A white, flocculent precipitate is formed.

C. To 5 mL of solution S add 1 mL of dilute hydrochloric acid R, 2 mL of methylene chloride R and 0.2 mL of diethyleneglycol R. Shake. The lower layer is orange-yellow.

D. About 10 mg gives the reaction of nitrates (2.3.1).

TESTS

Solution S

To 0.1 g add 45 mL of water R and heat to boiling with shaking. Cool, filter and dilute to 50 mL with water R.

Appearance of solution

Solution S is colourless (2.2.2, Method II).

Inorganic mercuric compounds

Maximum 0.1 per cent.

To 10 mL of solution S add 2 mL of potassium iodide solution R and 3 mL of dilute hydrochloric acid R. Filter. The filtrate is colourless. Wash the precipitate with 2 mL of water R. Combine the filtrate and washings, add 2 mL of dilute sodium hydroxide solution R and dilute to 20 mL with water R. 12 mL of the solution complies with test A for heavy metals (2.4.8). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in *vacuo* for 24 h.

ASSAY

Dissolve 0.150 g in a mixture of 10 mL of dilute nitric acid R and 90 mL of water R, heating to boiling. Cool to 15–20 °C. Titrate with 0.1 M ammonium thiocyanate using 2 mL of ferric ammonium sulfate solution R2 as indicator, until a persistent reddish-yellow colour is obtained. Carry out a blank titration. 1 mL of 0.1 M ammonium thiocyanate is equivalent to 20.06 mg of Hg.

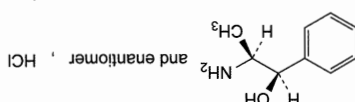
STORAGE

Protected from light.

Ph Eur

Phenylpropanolamine Hydrochloride

(Ph. Eur. monograph 0683)



$C_9H_{11}ClNO$ 187.7

154-41-6

Action and use

Adrenoceptor agonist.

Ph Eur

DEFINITION

Phenylpropanolamine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.5 per cent of (1R,2S)-2-amino-1-phenylpropan-1-ol hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, freely soluble in water and in alcohol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification B, E.

Second identification A, C, D, E.

A. Melting point (2.2.14): 194 °C to 197 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with

phenylpropanolamine hydrochloride CRS. Examine the substances prepared as discs without recrystallisation.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour

and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve 50 mg in 5 mL of water R, add 0.2 mL of copper sulfate solution R and 0.3 mL of dilute sodium hydroxide solution R. A violet colour develops. Add 2 mL of ether R and shake. A violet precipitate is formed between the two layers.

E. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 1.25 g in water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of methyl red solution R and 0.2 mL of 0.01 M sodium hydroxide. The solution is yellow. Add 0.4 mL of 0.01 M hydrochloric acid. The solution is red.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel H R as the coating substance.

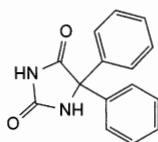
Test solution (a) Dissolve 0.20 g of the substance to be examined in alcohol R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with alcohol R.



Phenytoin

(Ph. Eur. monograph 1253)

 $C_{15}H_{12}N_2O_2$

252.3

57-41-0

Action and use

Antiepileptic.

Preparation

Phenytoin Oral Suspension

DEFINITION

5,5-Diphenylhydantoin-2,4-dione.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison phenytoin CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Dissolve 1.0 g in a mixture of 5 mL of 1 M sodium hydroxide and 20 mL of water R.

Acidity or alkalinity

To 1.0 g add 45 mL of water R and boil for 2 min. Allow to cool and filter. Wash the filter with carbon dioxide-free water R and dilute the combined filtrate and washings to 50 mL with the same solvent. To 10 mL of the solution add 0.15 mL of methyl red solution R. Not more than 0.5 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to red. To 10 mL of the solution add 0.15 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to blue.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve 2 mg of 2,2-diphenylglycine R (impurity C) in 100.0 mL of the mobile phase.
Reference solution (c) Dissolve 10 mg of phenytoin for system suitability CRS (containing impurities D and E) in the mobile

Reference solution (a) Dissolve 20 mg of phenylpropanolamine hydrochloride CRS in alcohol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of reference solution (a) to 10 mL with alcohol R.

Reference solution (c) Dissolve 20 mg of norpseudoephedrine hydrochloride CRS in alcohol R, add 1 mL of test solution (a) and dilute to 10 mL with alcohol R.

Reference solution (d) Dissolve 60 mg of ammonium chloride R in methanol R and dilute to 10 mL with the same solvent.

Before applying the solutions, spray the plate with a 20 g/L solution of disodium tetraborate R, using 8 mL for a plate 100 mm by 200 mm and dry in a stream of cold air for 30 min. Apply separately to the plate as bands about 10 mm by 3 mm 10 µL of each solution. Develop over a path of 10 cm using a mixture of 6 volumes of concentrated ammonia R, 24 volumes of alcohol R and 70 volumes of butanol R. Dry the plate in a current of warm air until the solvents have evaporated, allow to cool, spray with a 2 g/L solution of ninhydrin R in alcohol R and heat at 110 °C for 15 min. Any spot in the chromatogram obtained with test solution (a) apart from the principal spot and the spot corresponding to ammonium chloride is not more intense than the spot in the chromatogram obtained with reference solution (b) (1.0 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated spots.

Phenylpropanolamine

Dissolve 1.0 g in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid. The absorbance (2.2.25) of the solution measured at 283 nm is not greater than 0.10.

Heavy metals (2.4.8)

12 mL of solution S complies with test A for heavy metals (20 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

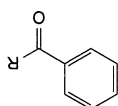
Dissolve 0.1500 g in a mixture of 5 mL of 0.01 M hydrochloric acid and 50 mL of alcohol R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the two points of inflexion.
1 mL of 0.1 M sodium hydroxide is equivalent to 18.77 mg of C_9H_9ClNO .

Ph Eur

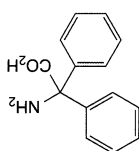
IMPURITIES

Specified impurities C, D, E.

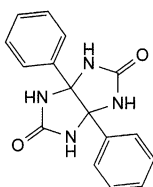
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, F.



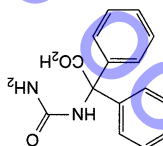
A. R = C₆H₅; diphenylmethanone (benzophenone),
B. R = CO-C₆H₅; diphenylethanedione (benzil),



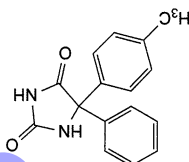
C. amino(diphenyl)acetic acid (2,2-diphenylglycine),



D. 3a,6a-diphenyltetrahydroimidazo[4,5-d]imidazole-2,5(1H,3H)-dione,



E. (carbamoylamino)(diphenyl)acetic acid,



F. 5-(4-methylphenyl)-5-phenylimidazolidine-2,4-dione.

phase, add 1.0 mL of reference solution (b) and dilute to 10.0 mL with the mobile phase.
Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).
Mobile phase Mix 20 volumes of methanol R2, 35 volumes of acetonitrile R1 and 45 volumes of a 5.75 g/L solution of ammonium dihydrogen phosphate R adjusted to pH 2.5 with phosphoric acid R.
Flow rate 1.5 mL/min.
Detection Spectrophotometer at 220 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (c).

Run time 4 times the retention time of phenytoin.

Identification of impurities Use the chromatogram supplied with phenytoin for system suitability CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities C, D and E.

Relative retention With reference to phenytoin (retention

time = about 4 min): impurity C = about 0.5;

impurity D = about 0.6; impurity E = about 0.8.

System suitability: reference solution (c):

— resolution: minimum 3.5 between the peaks due to impurities D and E.

Limits:

— correction factors: for the calculation of content, multiply the peaks areas of the following impurities by the

corresponding correction factor: impurity D = 1.7;

impurity E = 1.4;

— impurity E: not more than 3 times the area of the principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.2 per cent);

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

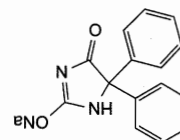
ASSAY

Dissolve 0.200 g in 50 mL of dimethylformamide R. Titrate with 0.1 M sodium methoxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium methoxide is equivalent to 25.23 mg of C₁₅H₁₂N₂O₂.

Phenytoin Sodium

(Ph Eur monograph 0521)

 $C_{15}H_{11}N_2NaO_2$

274.3

630-93-3

Action and use

Antiepileptic.

Preparations

Phenytoin Capsules

Phenytoin Injection

Phenytoin Tablets

Ph Eur

DEFINITION

Sodium 4-oxo-5,5-diphenyl-4,5-dihydro-1H-imidazol-2-olate.

Content

98.5 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder, slightly

hygroscopic.

Solubility

Soluble in water and in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification A, C.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Prepare Suspension 0.1 g in 20 mL of water R. Acidify with

dilute hydrochloric acid R and shake with 3 quantites, each of

30 mL, of ethyl acetate R. Wash the combined ethyl acetate

layers with water R, evaporate to dryness and dry the residue

at 100–105 °C (test residue). Repeat the operations using

0.1 g of phenytoin sodium CRS (reference residue). Examine

as discs prepared using potassium bromide R.

Comparison phenytoin sodium CRS.

B. Thin-layer chromatography (2.2.27).

Solvent mixture acetone R, methanol R (50:50 V/V).

Test solution Dissolve 20 mg of the substance to be examined

in the solvent mixture and dilute to 10 mL with the solvent

mixture.

Reference solution Dissolve 20 mg of phenytoin sodium CRS in

the solvent mixture and dilute to 10 mL with the solvent

mixture.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, toluene R, 2-propanol R

(10:40:50 V/V/V).

Application 10 µL as bands of 8 mm.

Development Over 2/3 of the plate.

Drying At 80 °C for 5 min; allow to cool.

Detection Examine in ultraviolet light at 254 nm.

Results The principal zone in the chromatogram obtained

with the test solution is similar in position and size to the

principal zone in the chromatogram obtained with the

reference solution.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of 2,2-diphenylglycine R (impurity C) in 100.0 mL of the mobile phase.

Reference solution (c) Dissolve 10 mg of phenytoin for system suitability CRS (containing impurities D and E) in the mobile phase, add 1.0 mL of reference solution (b) and dilute to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 µm).

Mobile phase Mix 20 volumes of methanol R₂, 35 volumes of acetonitrile R₁ and 45 volumes of a 5.75 g/L solution of ammonium dihydrogen phosphate R adjusted to pH 2.5 with phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL of the test solution and reference solutions (a) and (c).

Run time 4 times the retention time of phenytoin.

Identification of impurities Use the chromatogram supplied

with phenytoin for system suitability CRS and the chromatogram obtained with reference solution (c) to identify

the peaks due to impurities C, D and E.

Relative retention With reference to phenytoin (retention

time = about 4 min): impurity C = about 0.5;

impurity D = about 0.6; impurity E = about 0.8.

System suitability: reference solution (c):

— resolution: minimum 3.5 between the peaks due to

impurities D and E.

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity D = 1.7;

impurity E = 1.4;

impurity F: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

impurity C: not more than twice the area of the principal

peak in the chromatogram obtained with

reference solution (a) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

- *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Free phenytoin

Dissolve 0.30 g in 10 mL of a mixture of equal volumes of pyridine R and water R. Add 0.5 mL of phenolphthalein solution R and 3 mL of silver nitrate solution in pyridine R.

Not more than 1.0 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to pink.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.000 g.

ASSAY

Suspend 0.180 g in 2 mL of water R. Add 8.0 mL of 0.05 M

sulfuric acid and heat gently for 1 min. Add 30 mL of

methanol R and cool. Carry out a potentiometric titration

(2.2.20), using 0.1 M sodium hydroxide. After reaching the 1st

point of inflexion, interrupt the addition of 0.1 M sodium

hydroxide, add 5 mL of silver nitrate solution in pyridine R, mix

and continue the titration. Read the volume of 0.1 M sodium

hydroxide added between the 2 points of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 27.43 mg of

C₁₅H₁₁N₂NaO₂.

STORAGE

In an airtight container.

IMPURITIES

Specified impurities C, D, E

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

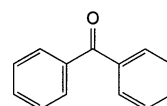
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

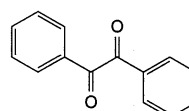
impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, B,

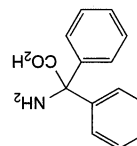
F.



A. diphenylmethanone (benzophenone),



B. diphenylethanedione (benzil),



C. amino(diphenyl)acetic acid (2,2-diphenylglycine),

Action and use

Antispasmodic.

Ph Eur

DEFINITION

Benzene-1,3,5-triol.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Sparingly soluble in water, freely soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

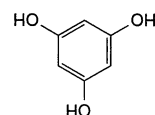
A. Infrared absorption spectrophotometry (2.2.24).

Comparison anhydrous phloroglucinol CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.20 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 0.20 g of anhydrous phloroglucinol CRS in methanol R and dilute to 10 mL with the same solvent.



126.1

C₆H₆O₃

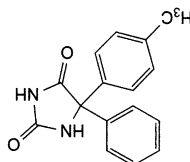
108-73-6

Anhydrous Phloroglucinol

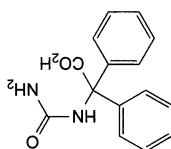
(Ph. Eur. monograph 2301)

Ph Eur

F. 5-(4-methylphenyl)-5-phenylimidazolidine-2,4-dione.



E. (carbamoylamino)(diphenyl)acetic acid,



D. 3a,6a-diphenyltetrahydroimidazo[4,5-d]imidazole-2,5(1H,3H)-dione,

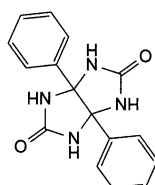


Plate TLC silica gel F₂₅₄ plate R.
Mobile phase anhydrous formic acid R, hexane R, ethyl acetate R (2:37.5:62.5 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Loss on drying (see Tests).

TESTS

Solution S
Dissolve 2.50 g in ethanol (96 per cent) R and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

pH (2.2.3)
4.0 to 6.0.

Dilute 10 mL of solution S to 100 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and protect from light.

Solvent mixture Mobile phase B, mobile phase A (10:90 V/V).
Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 6 mg each of resorcinol R (impurity B), phloroglucinol R (impurity D) and pyrogallol R (impurity A) in 10 mL of the solvent mixture, add 2 mL of the test solution and dilute to 20.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 4 mg each of pyrogallol R (impurity A), phloroglucinol R (impurity D), benzene-1,2,4-triol R (impurity E), 2,6-dichlorophenol R (impurity I), 4-chlororesorcinol R (impurity K) and 3,5-dichloroaniline R (impurity L) in 10 mL of the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (d) Dissolve 10 mg of the substance to be examined in 10 mL of the solvent mixture, add 1.0 mL of reference solution (c) and dilute to 20.0 mL with the solvent mixture.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography with embedded polar groups R (5 µm).
Mobile phase:
— mobile phase A: 1.36 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 3.0 with phosphoric acid R;
— mobile phase B: acetonitrile R.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 9	100	0
9 - 15	100 → 50	0 → 50
15 - 25	50 → 20	50 → 80
25 - 30	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 20 µL of the test solution and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, D, E, I, K and L.

Relative retention With reference to phloroglucinol (retention time = about 12 min): impurity B = about 0.7; impurity A = about 0.9; impurity D = about 1.3; impurity B = about 1.35; impurity K = about 1.5; impurity I = about 1.8; impurity L = about 2.0.

System suitability: reference solution (b):
— resolution: minimum 2.5 between the peaks due to impurity A and phloroglucinol; minimum 4.0 between the peaks due to impurities D and B.
Limits:
— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6; impurity D = 0.2; impurity E = 0.7; impurity I = 0.6; impurity K = 0.6; impurity L = 0.4;
— impurities A, D, E, I, K, L: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (2.4.4)
Maximum 200 ppm.
Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)
Maximum 500 ppm.
Dilute 3 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)
Maximum 20 ppm.
Solvent mixture water R, ethanol (96 per cent) R (10:90 V/V).
0.500 g complies with test H. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.500 g in 50 mL of water R. Titrate with 1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).
1 mL of 1 M sodium hydroxide is equivalent to 63.05 mg of $C_6H_6O_3$.

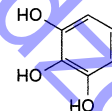
STORAGE

Protected from light.

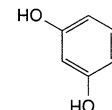
IMPURITIES

Specified impurities: A, D, E, I, K, L.

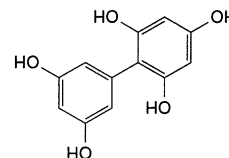
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, O.



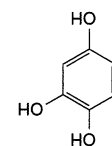
A. benzene-1,2,3-triol (pyrogallol),



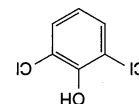
B. benzene-1,3-diol (resorcinol),



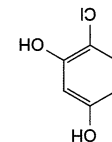
D. 2,3',4,5',6-biphenylpentol (phloroglucide),



E. benzene-1,2,4-triol,



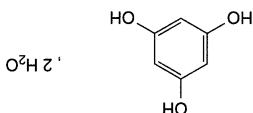
I. 2,6-dichlorophenol,



K. 4-chlorobenzene-1,3-diol (4-chlororesorcinol),

Phloroglucinol Dihydrate

(Ph. Eur. monograph 2302)



60999-90-7

Action and use
Antispasmodic.

Ph Eur

DEFINITION

Benzene-1,3,5-triol dihydrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Sparingly soluble in water, freely soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Previously dry the substance to be examined in an oven at 105 °C.

Comparison anhydrous phloroglucinol CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.20 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 0.20 g of anhydrous phloroglucinol CRS in methanol R and dilute to 10 mL with

the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase anhydrous formic acid R, hexane R, ethyl acetate R (2:37.5:62.5 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal



Ph Eur

spot in the chromatogram obtained with the reference

solution.

C. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 2.50 g in ethanol (96 per cent) R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

pH (2.2.3)

4.0 to 6.0.

Dilute 10 mL of solution S to 100 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and protect from light.

Solvent mixture Mobile phase B, mobile phase A (10:90 V/V).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 6 mg each of resorcinol R

(impurity B), phloroglucinol R (impurity D) and pyrogallol R

(impurity A) in 10 mL of the solvent mixture, add 2 mL of

the test solution and dilute to 20.0 mL with the solvent

mixture. Dilute 1.0 mL of this solution to 50.0 mL with the

solvent mixture.

Reference solution (c) Dissolve 4 mg each of pyrogallol R

(impurity A), phloroglucinol R (impurity D), benzene-1,2,4-

triol R (impurity E), 2,6-dichlorophenol R (impurity I), 4-

chlororesorcinol R (impurity K) and 3,5-dichloroaniline R

(impurity L) in 10 mL of the solvent mixture and dilute to

20.0 mL with the solvent mixture.

Reference solution (d) Dissolve 10 mg of the substance to be

examined in 10 mL of the solvent mixture, add 1.0 mL of

reference solution (c) and dilute to 20.0 mL with the solvent

mixture.

Column: size: $l = 0.25$ m, $\phi = 4.0$ mm;

stationary phase: end-capped octadecylsilyl silica gel for

chromatography with embedded polar groups R (5 μ m).

Mobile phase: mobile phase A: 1.36 g/L solution of potassium dihydrogen

phosphate R previously adjusted to pH 3.0 with phosphoric

acid R; mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 9	100	0
9 - 15	100 \rightarrow 50	0 \rightarrow 50
15 - 25	50 \rightarrow 20	50 \rightarrow 80
25 - 30	20	80

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 20 μ L of the test solution and reference solutions (a), (b) and (d).

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, D, E, I, K and L.

Relative retention With reference to phloroglucinol (retention

time = about 12 min): impurity E = about 0.7;

impurity A = about 0.9; impurity D = about 1.3;

impurity B = about 1.35; impurity K = about 1.5;

impurity I = about 1.8; impurity L = about 2.0.

System suitability: reference solution (b):

— resolution: minimum 2.5 between the peaks due to

impurity A and phloroglucinol; minimum 4.0 between the

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity A = 0.6;

impurity D = 0.2; impurity E = 0.7; impurity I = 0.6;

impurity K = 0.6; impurity L = 0.4;

— impurities A, D, E, I, K, L: for each impurity, not more

than 1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 500 ppm.

Dilute 3 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent mixture water R, ethanol (96 per cent) R (10:90 V/V).

0.500 g complies with test H. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

20.0 per cent to 23.0 per cent, determined on 1.000 g by

drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.600 g in 50 mL of water R. Titrate with 1 M

sodium hydroxide, determining the end-point

potentiometrically (2.2.20).

1 mL of 1 M sodium hydroxide is equivalent to 63.05 mg

of C₆H₆O₃.

STORAGE

Protected from light.

IMPURITIES

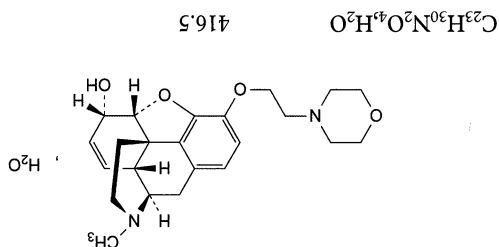
Specified impurities A, D, E, I, K, L

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use



Pholcodine

(Ph. Eur. monograph 0522)



Action and use
Opioid receptor agonist; cough suppressant.

Preparations

Pholcodine Linctus
Strong Pholcodine Linctus

Ph Eur

DEFINITION

7,8-Didehydro-4,5 α -epoxy-17-methyl-3-[2-(morpholin-4-yl)ethoxy]morphinan-6 α -ol monohydrate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Sparsely soluble in water, freely soluble in acetone and in ethanol (96 per cent). It dissolves in dilute mineral acids.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison pholcodine CRS.

TESTS

Specific optical rotation (2.2.7)

–98 to –94 (dried substance).

Dissolve 1.000 g in ethanol (96 per cent) R and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

0.02 M phosphate buffer solution To 80.0 mL of 0.2 M sodium hydroxide add 100.0 mL of 0.2 M potassium dihydrogen phosphate R and dilute to 1000.0 mL with water R.

Solvent mixture Dilute 80 mL of acetonitrile R to 1000 mL

with the 0.02 M phosphate buffer solution.

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 50 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of codeine R

(impurity B) in the solvent mixture and dilute to 10 mL with the solvent mixture. To 0.5 mL of this solution add 0.5 mL

of the test solution and dilute to 50 mL with the solvent mixture.

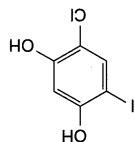
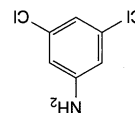
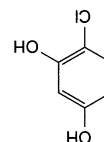
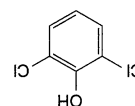
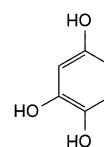
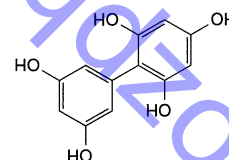
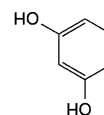
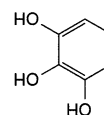
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of pholcodine for peak

identification CRS (containing impurities A, B and D) in the solvent mixture and dilute to 5 mL with the solvent mixture.

(2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, O.



Ph Eur

Column:
— size: $l = 0.075$ m, $\varnothing = 4.6$ mm;
— stationary phase: spherical end-capped phenylhexylsilica
— gel for chromatography R (3 μ m) with a specific surface area of 400 m²/g and a pore size of 10 nm;
— temperature: 35 °C.

Mobile phase To 50 mL of tetrahydrofuran for chromatography R add 75 mL of acetonitrile R and dilute to 1000 mL with the 0.02 M phosphate buffer solution; adjust to pH 7.9 \pm 0.05 with 0.2 M sodium hydroxide; the pH must not exceed 8.0.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 238 nm.

Injection 20 μ L.

Run time 5 times the retention time of pholcodine.

Identification of impurities Use the chromatogram supplied with pholcodine for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and D.

Relative retention With reference to pholcodine (retention time = about 10 min): impurity A = about 0.4;

impurity B = about 0.8; impurity D = about 2.3.

System suitability: reference solution (a):

— resolution: minimum 3 between the peaks due to

impurity B and pholcodine.

Limits:

— impurities A, B, D: for each impurity, not more than twice

the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 7 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.7 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

Loss on drying (2.2.32)

3.9 per cent to 4.5 per cent, determined on 0.500 g by

drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.180 g in 50 mL of anhydrous acetic acid R,

warming gently. Titrate with 0.1 M perchloric acid,

determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 19.93 mg

of C₂₃H₃₀N₂O₄.

IMPURITIES

Specified impurities A, B, D

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

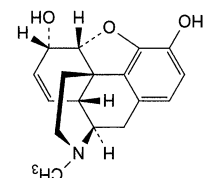
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

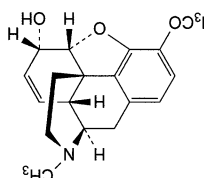
Control of impurities in substances for pharmaceutical use: C, E,

F.

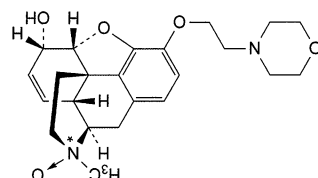
A, 7,8-didehydro-4,5 α -epoxy-17-methylmorphinan-3,6 α -diol (morphine),



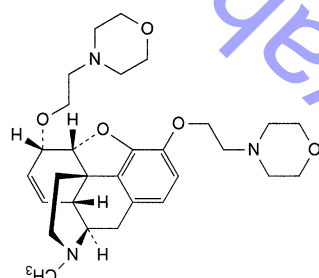
B, 7,8-didehydro-4,5 α -epoxy-3-methoxy-17-methylmorphinan-6 α -ol (codeine),



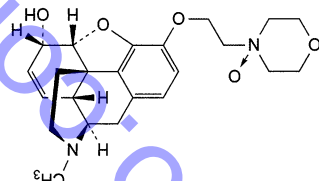
and epimer at N^o



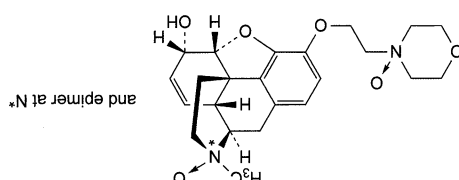
C, (17R)-7,8-didehydro-4,5 α -epoxy-17-methyl-3-[2-(morpholin-4-yl)ethoxy]morphinan-6 α -ol 17-oxide (pholcodine N^o-oxide),



D, 7,8-didehydro-4,5 α -epoxy-17-methyl-3,6 α -bis[2-(morpholin-4-yl)ethoxy]morphinan,



E, 7,8-didehydro-4,5 α -epoxy-17-methyl-3-[2-(4-oxidomorpholin-4-yl)ethoxy]morphinan-6 α -ol (pholcodine N^o-oxide),



F, (17R)-7,8-didehydro-4,5 α -epoxy-17-methyl-3-[2-(4-oxidomorpholin-4-yl)ethoxy]morphinan-6 α -ol 17-oxide (pholcodine N^o,N^o-dioxide).

Phosphoric Acid

(Concentrated Phosphoric Acid,
Ph Eur monograph 0004)

H₃PO₄

98.0

7664-38-2

Preparation

Phosphate Oral Solution

DEFINITION

Content

84.0 per cent *m/m* to 90.0 per cent *m/m*.

CHARACTERS

Appearance

Clear, colourless, syrupy liquid, corrosive.

Solubility

Miscible with water and with ethanol (96 per cent).

When stored at a low temperature it may solidify into a mass of colourless crystals which do not melt at a temperature below 28 °C.

Relative density

About 1.7.

IDENTIFICATION

A. Dilute with water R. The solution is strongly acid (2.2.4).

B. Solution S (see Tests) neutralised with dilute sodium

hydroxide solution R gives the reactions of phosphates (2.3.1).

TESTS

Solution S

Dilute 10.0 g to 150 mL with water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Substances precipitated with ammonia

To 10 mL of solution S add 8 mL of dilute ammonia R1.

Any opalescence in the solution is not more intense than that

in a mixture of 10 mL of solution S and 8 mL of water R.

Hypophosphorous acid and phosphorous acid

To 5 mL of solution S add 2 mL of silver nitrate solution R2

and heat on a water-bath for 5 min. The solution shows no

change in appearance.

Chlorides (2.4.4)

Maximum 50 ppm, determined on solution S.

Sulfates (2.4.13)

Maximum 100 ppm.

Dilute 1.5 g to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 7.5 mL of solution S.

Iron (2.4.9)

Maximum 50 ppm.

Dilute 3 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

To 2.5 g add 4 mL of dilute ammonia R1 and dilute to

25 mL with water R. 12 mL of the solution complies with

test A. Prepare the reference solution using lead standard

solution (1 ppm Pb) R.

ASSAY

To 1.000 g add a solution of 10 g of sodium chloride R in 30 mL of water R. Titrate with 1 M sodium hydroxide, using phenolphthalein solution R as indicator.

Dilute Phosphoric Acid

(Ph. Eur. monograph 0005)



DEFINITION

Content

9.5 per cent *m/m* to 10.5 per cent *m/m* of H₃PO₄ (98.0).

PREPARATION

To 885 g of water R add 115 g of concentrated phosphoric acid and mix.

IDENTIFICATION

A. It is strongly acid (2.2.4).

B. Solution S (see Tests), neutralised with dilute sodium

hydroxide solution R, gives the reactions of phosphates (2.3.1).

TESTS

Solution S

Dilute 86 g to 150 mL with water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Substances precipitated with ammonia

To 10 mL of solution S add 8 mL of dilute ammonia R1.

Any opalescence in the solution is not more intense than that

in a mixture of 10 mL of solution S and 8 mL of water R.

Hypophosphorous acid and phosphorous acid

To 5 mL of solution S add 2 mL of silver nitrate solution R2

and heat on a water-bath for 5 min. The solution shows no

change in appearance.

Chlorides (2.4.4)

Maximum 6 ppm, determined on solution S.

Sulfates (2.4.13)

Maximum 10 ppm, determined on the preparation to be

examined.

Arsenic (2.4.2, Method A)

Maximum 0.2 ppm, determined on 7.5 mL of solution S.

Iron (2.4.9)

Maximum 6 ppm.

Heavy metals (2.4.8)

Maximum 1 ppm.

To 20 g add 4 mL of dilute ammonia R1 and dilute to 25 mL

with water R. 12 mL of the solution complies with test A.

Prepare the reference solution using a mixture of 2 mL of

water R and 8 mL of lead standard solution (1 ppm Pb) R.

ASSAY

To 8.60 g add a solution of 10 g of sodium chloride R in

30 mL of water R. Titrate with 1 M sodium hydroxide, using

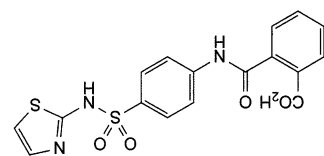
phenolphthalein solution R as indicator.

1 mL of 1 M sodium hydroxide is equivalent to 49.00 mg

of H₃PO₄.

Phthalylsulfathiazole

(Ph. Eur. monograph 0352)

 $C_{17}H_{13}N_3O_5S_2$

403.4

85-73-4

Action and use

Antibacterial.

Ph Eur

DEFINITION

2-[[4-(Thiazol-2-yl)sulfamoyl]phenyl]carbamoyl]benzoic acid.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in dimethylformamide, slightly soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

First identification A, B, E.

Second identification B, C, D, E.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison phthalylsulfathiazole CRS.

B. To 1 g add 8.5 mL of dilute sodium hydroxide solution R

and boil under a reflux condenser for 30 min. Cool and add

17.5 mL of dilute hydrochloric acid R. Shake vigorously and

filter. Neutralise the filtrate with dilute sodium hydroxide

solution R. Filter, wash the precipitate with water R,

recrystallise from water R and dry the crystals at 100-105 °C.

The crystals melt (2.2.14) at 200 °C to 203 °C.

C. To 0.1 g in a test-tube add 3 mL of dilute sulfuric acid R

and 0.5 g of zinc powder R. Fumes are evolved which

produce a black stain on lead acetate paper R.

D. To 0.1 g add 0.5 g of resorcinol R and 0.3 mL of sulfuric

acid R and heat on a water-bath until a homogeneous

mixture is obtained. Allow to cool. Add 5 mL of dilute sodium

hydroxide solution R. Dilute 0.1 mL of this brownish-red

mixture to 25 mL with water R. An intense green

fluorescence appears which disappears on acidification.

E. Dissolve about 10 mg of the crystals obtained in

identification test B in 200 mL of 0.1 M hydrochloric acid.

2 mL of the solution gives the reaction of primary aromatic

amines (2.3.1) with formation of an orange precipitate.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely

coloured than reference solution BY₅ (2.2.2, Method II).

Dissolve 1.0 g in 1 M sodium hydroxide and dilute to 20 mL

with the same solvent.

Acidity

To 2.0 g add 20 mL of water R, shake continuously for

30 min and filter. To 10 mL of the filtrate add 0.1 mL of

phenolphthalein solution R. Not more than 0.2 mL of 0.1 M

sodium hydroxide is required to change the colour of the

indicator.

Sulfathiazole and other primary aromatic amines

Maximum 2.0 per cent.

Dissolve 5 mg in a mixture of 3.5 mL of water R, 6 mL of

dilute hydrochloric acid R and 25 mL of ethanol (96 per cent) R,

previously cooled to 15 °C. Place immediately in iced water

and add 1 mL of a 2.5 g/L solution of sodium nitrite R. Allow

to stand for 3 min, add 2.5 mL of a 40 g/L solution of

sulfamic acid R and allow to stand for 5 min. Add 1 mL of a

4 g/L solution of naphthylethylenediamine dihydrochloride R and

dilute to 50 mL with water R. Measured at 550 nm, the

absorbance (2.2.25) is not greater than that of a standard

prepared at the same time and in the same manner using a

mixture of 1 mL of a 100 mL aqueous solution containing

10 mg of sulfathiazole R and 0.5 mL of hydrochloric acid R,

2.5 mL of water R, 6 mL of dilute hydrochloric acid R,

and 25 mL of ethanol (96 per cent) R.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 2 per cent, determined on 1.00 g by drying in an

oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 40 mL of dimethylformamide R. Titrate

with 0.1 M sodium hydroxide until the colour becomes blue.

Using 0.2 mL of thymolphthalein solution R as indicator. Carry

out a blank titration.

1 mL of 0.1 M sodium hydroxide is equivalent to 20.17 mg of

 $C_{17}H_{13}N_3O_5S_2$.

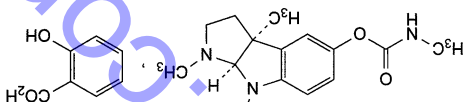
STORAGE

Protected from light.

Ph Eur

Physostigmine Salicylate

(Ph. Eur. monograph 0286)

 $C_{22}H_{27}N_3O_5$

413.5

57-64-7

Action and use

Cholinesterase inhibitor.

DEFINITION

Physostigmine salicylate contains not less than 98.5 per cent

and not more than the equivalent of 101.0 per cent of

(3a,5,8aR)-1,2,3,3a,8,8a-hexahydro-1,3a,8-

trimethylpyrrolo[2,3-b]indol-5-yl methylcarbamate salicylate,

calculated with reference to the dried substance.

CHARACTERS

Colourless or almost colourless crystals, sparingly soluble in water, soluble in alcohol. The crystals gradually become red when exposed to air and light; the colour develops more

quickly when the crystals are also exposed to moisture.

Aqueous solutions are unstable.

It melts at about 182 °C, with decomposition.

IDENTIFICATION

First identification A, B.

Second identification B, C, D.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with *physosugmine salicylate CRS*.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Heat about 10 mg in a porcelain dish with a few drops of *dilute ammonia RL*. An orange colour develops. Evaporate the solution to dryness. The residue dissolves in *alcohol R* giving a blue solution. Add 0.1 mL of *glacial acetic acid R*.

The colour becomes violet. Dilute with *water R*. An intense red fluorescence appears.

(2.3.1).

TESTS

Solution S

Dissolve 0.900 g, without heating, in 95 mL of carbon dioxide-free water *R* prepared from distilled water *R* and dilute to 100.0 mL with the same solvent. Prepare immediately before use.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, *Method II*).

pH (2.2.3)

The pH of solution S is 5.1 to 5.9.

Specific optical rotation (2.2.7)

–90 to –94, determined on solution S and calculated with reference to the dried substance.

Related substances

Examine by thin-layer chromatography (2.2.27), using *silica gel G R* as the coating substance.

Test solution (a) Dissolve 0.2 g of the substance to be examined in *alcohol R* and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 2.5 mL of test solution (a) to 50 mL with *alcohol R*.

Reference solution (a) Dissolve 10 mg of *physosugmine salicylate CRS* in *alcohol R* and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 2 mL of reference solution (a) to 20 mL with *alcohol R*.

Apply to the plate 20 µL of each solution. Develop over a path of 15 cm using a mixture of 2 volumes of *concentrated ammonia R*, 23 volumes of 2-propanol *R* and 100 volumes of *cyclohexane R*. Dry the plate in a current of cold air and carry out a second development in the same direction. Allow the plate to dry in air and spray with freshly prepared *potassium iodobismuthate solution R* and then with *dilute hydrogen peroxide solution R*. Examine the plate within 2 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Mixture of 2-methyl-3-[(2*E*,7*R*,11*R*)-3,7,11,15-tetramethylhexadec-2-enyl]naphthalene-1,4-dione (*trans*-phytomenadione), 2-methyl-3-[(2*Z*,7*R*,11*R*)-3,7,11,15-tetramethylhexadec-2-enyl]naphthalene-1,4-dione (*cis*-phytomenadione) and 2,3-epoxy-2-methyl-3-[(2*E*,7*R*,11*R*)-3,7,11,15-tetramethylhexadec-2-enyl]naphthalene-1,4-dione (*trans*-epoxyphytomenadione).

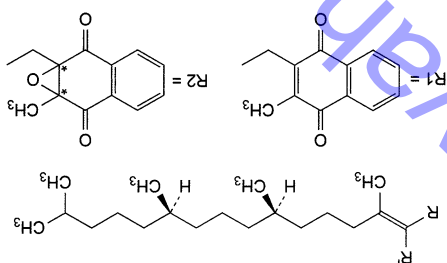
DEFINITION

Action and use
Vitamin K analogue.

Preparations
Phytomenadione Injection
Phytomenadione Tablets

When vitamin K1 is prescribed or demanded, Phytomenadione shall be dispensed or supplied.

Phytomenadione		R ¹	R ²	M _r
trans	R ¹	H	C ₃₁ H ₄₆ O ₂	450.7
cis	R ¹	H	C ₃₁ H ₄₆ O ₂	450.7
trans-epoxy	R ²	H	C ₃₁ H ₄₆ O ₃	466.7



(Ph. Eur. monograph 1036)

Phytomenadione



Ph Eur

STORAGE

Store in an airtight container, protected from light.

C₂₂H₂₇N₃O₅.

1 mL of 0.1 M *perchloric acid* is equivalent to 41.35 mg of (2.2.20).

Dissolve 0.350 g in 50 mL of a mixture of equal volumes of *anhydrous acetic acid R* and *chloroform R*. Titrate with 0.1 M *perchloric acid* determining the end-point potentiometrically (2.2.20).

ASSAY

obtained in the test for loss on drying.

Not more than 0.1 per cent, determined on the residue

Sulfated ash (2.4.14)

in an oven at 105 °C.

Not more than 1.0 per cent, determined on 1.00 g by drying

Loss on drying (2.2.32)

(0.1 per cent).

15 mL of solution S complies with the limit test for sulfates

Sulfates (2.4.13)

chloroform R within 1 min.

To 5 mL of solution S add a few crystals of *potassium iodate R*, 0.05 mL of *dilute hydrochloric acid R* and 2 mL of

Esclidine

Content

- *trans*-epoxyphytomenadione: maximum 4.0 per cent;
- *trans*-phytomenadione: minimum 75.0 per cent;
- sum of *trans*-phytomenadione, *cis*-phytomenadione and *trans*-epoxyphytomenadione: 97.0 per cent to 103.0 per cent.

CHARACTERS**Appearance**

Clear, intense yellow, viscous, oily liquid.

Solubility

Practically insoluble in water, sparingly soluble in ethanol (96 per cent), miscible with fatty oils.

It decomposes on exposure to actinic light.

Refractive index About 1.526.

IDENTIFICATION

Carry out all operations as rapidly as possible, avoiding exposure to actinic light.

A. Ultraviolet and visible absorption

spectrophotometry (2.2.25).

Test solution (a) Dissolve 10.0 mg of the substance to be examined in *triethylpentane R* and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 10.0 mL of test solution (a) to 50.0 mL with *triethylpentane R*.

Spectral range 275–340 nm for test solution (a); 230–280 nm for test solution (b).

Absorption maxima At 327 nm for test solution (a);

at 243 nm, 249 nm, 261 nm and 270 nm for test solution (b).

Absorption minimum At 285 nm for test solution (a). *Specific absorbance at the absorption maximum at 327 nm* 67 to 73 for test solution (a).

B. Examine the chromatograms obtained in the assay. *Results* The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (d).

C. Dissolve 50 mg in 10 mL of *methanol R* and add 1 mL of a 200 g/L solution of *potassium hydroxide R* in *methanol R*. A green colour is produced which becomes violet-red on heating in a water-bath at 40 °C and then reddish-brown on standing.

TESTS**Appearance of solution**

The solution is clear (2.2.1).

Dissolve 2.5 g in *triethylpentane R* and dilute to 25 mL with the same solvent.

Acid value (2.5.1)

Maximum 2.0, determined on 2.00 g.

Impurity A

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.40 g of the substance to be examined in *cyclohexane R* and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 4.0 mg of *menadione R* (impurity A) in *cyclohexane R* and dilute to 50.0 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase *cyclohexane R*, *toluene R* (20:80 V/V).

Application 10 µL.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and protect from light.

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of *phytomenadione CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2.0 mg of *trans-epoxyphytomenadione CRS* in the mobile phase, add 5.0 mL of reference solution (a) and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 25.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: spherical silica gel for chromatography R (5 µm).

Mobile phase *octanol R*, *di-isopropyl ether R*, *heptane R* (2:6:6:2000 V/V/V).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Equilibration With the mobile phase for at least 24 h.

Injection 50 µL of the test solution and reference solutions (b) and (c).

Run time 1.6 times the retention time of *trans-phytomenadione*.

Relative retention With reference to *trans-phytomenadione* (retention time = about 30 min):

trans-epoxyphytomenadione = about 0.6;

cis-phytomenadione = about 0.65.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to *trans-epoxyphytomenadione* and *cis-phytomenadione*;

minimum 4.0 between the peaks due to *cis-phytomenadione* and *trans-phytomenadione*.

Calculation of percentage contents:

— for each impurity, use the concentration of *trans-phytomenadione* in reference solution (c).

Limits:

— impurities eluting before *trans-epoxyphytomenadione*: for each impurity, maximum 0.15 per cent;

— sum of impurities eluting before *trans-epoxyphytomenadione*: maximum 0.2 per cent;

— impurities eluting between *cis-phytomenadione* and *trans-phytomenadione*: for each impurity, maximum 0.4 per cent.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the peak in the chromatogram obtained with reference solution (b).

TESTS

Solution S Dissolve 1.0 g in tetrahydrofuran R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Acidity or alkalinity

Shake 0.20 g with a mixture of 4.0 mL of ethyl acetate R and 10.0 mL of carbon dioxide-free water R for 3 min. Allow the layers to separate. To the aqueous layer add 0.1 mL of bromothymol blue solution R1. If the solution is yellow, not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour to yellow.

Specific optical rotation (2.2.7)

–15.0 to –28.0 (dried substance).
Dissolve 0.500 g in ethyl acetate R and dilute to 10.0 mL with the same solvent.

Acid value (2.5.1)

Maximum 1.0, determined on 2.0 g.

Peroxide value (2.5.5)

Maximum 10.0.

Saponification value (2.5.6)

Maximum 1.0.

Carry out the test using 2.50 g of the substance to be examined, 0.1 M alcoholic potassium hydroxide, 0.1 M hydrochloric acid, and a factor of 5.61 (instead of 28.05).

Other sterols

Examine the chromatogram obtained with the test solution in the assay (Figure 1911.-1).

Composition of the other sterols:

- cholesterol: maximum 0.5 per cent;
- brassicasterol: maximum 0.5 per cent;
- campesterol: maximum 15.0 per cent;
- campestanol: maximum 5.0 per cent;
- stigmasterol: maximum 5.0 per cent;
- sitosterol: maximum 15.0 per cent;
- Δ7-stigmastenol: maximum 5.0 per cent.

Loss on drying (2.2.32)

Maximum 4.0 per cent, determined on 0.250 g by drying in an oven at 105 °C for 2 h.

Total ash (2.4.16)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution Dissolve 0.100 g in tetrahydrofuran R and dilute to 10.0 mL with the same solvent. Introduce 100 µL of this solution into a 3 mL flask and evaporate to dryness under nitrogen R. Add 100 µL of a freshly prepared mixture of 50 µL of 1-methylimidazole R and 1.0 mL of heptafluoro-N-methyl-N-(trimethylsilyl)butanamide R, close the flask tightly and heat to 100 °C for 15 min. Allow to cool.

— sum of impurities eluting between cis-phytomenadione and trans-phytomenadione: maximum 0.5 per cent;

— impurities eluting after trans-phytomenadione: for each impurity, maximum 0.25 per cent;

— sum of impurities eluting after trans-phytomenadione: maximum 0.5 per cent;

— total: maximum 1.2 per cent;

— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (d).

— **repeatability**: maximum relative standard deviation of 1.0 per cent for the peak due to trans-phytomenadione after 6 injections.

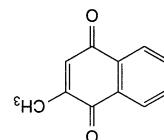
Calculate the percentage contents of trans-phytomenadione, cis-phytomenadione and trans-epoxyphytomenadione taking into account the assigned content of phytomenadione CRS.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A



A. 2-methylnaphthalene-1,4-dione (menadione).

Ph Eur



Phytosterol

(Ph. Eur. monograph 1911)

Ph Eur

DEFINITION

Natural mixture of sterols obtained from plants of the genera *Hypoxis*, *Pinus* and *Picea*.

Content

Minimum 70.0 per cent of β-sitosterol (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, soluble in tetrahydrofuran, sparingly soluble in ethyl acetate.

IDENTIFICATION

A. Mix 1 mL of acetic anhydride R with 0.5 mL of solution S (see Tests). After the addition of 0.1 mL of sulfuric acid R a red colour is produced, which changes rapidly to violet, then to blue and finally to green.
B. Examine the chromatograms obtained in the assay.

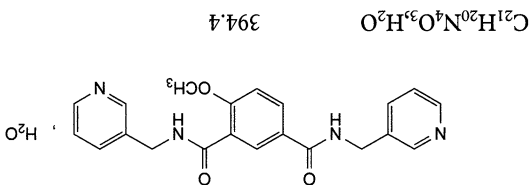
brassicasterol = about 0.77; campesterol = about 0.84; campestanol = about 0.86; stigmasterol = about 0.9; sitostanol = about 1.02; Δ^7 -stigmasterol = about 1.1.
— *resolution*: minimum 1.0 between the peaks due to β -sitosterol and sitostanol.
STORAGE

In an airtight container, protected from light.

Ph Eur

Picotamide Monohydrate

(Ph. Eur. monograph 1358)



Action and use
Thromboxane Synthetase inhibitor; thromboxane receptor antagonist; antiplatelet agent.

Ph Eur

DEFINITION

Picotamide monohydrate contains not less than 98.0 per cent and not more than 101.0 per cent of 4-methoxy- N,N' -bis(pyridin-3-ylmethyl)benzene-1,3-dicarboxamide, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, crystalline powder, slightly soluble in water, soluble in ethanol and in methylene chloride.
It dissolves in dilute mineral acids.
It shows polymorphism (5.9).

IDENTIFICATION

Examine by infrared spectrophotometry (2.2.24), comparing with the spectrum obtained with picotamide monohydrate CRS.
If the spectra obtained in the solid state shows differences, dissolve the substance to be examined and the reference substance separately in acetone R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution
Dissolve 2.5 g in methanol R and dilute to 50 mL with the same solvent. The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

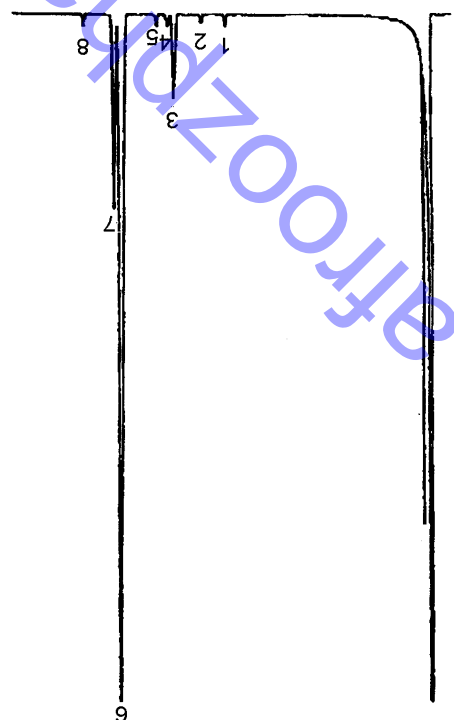
Related substances

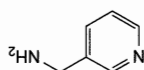
Examine by thin-layer chromatography (2.2.27), using a TLC silica gel F₂₅₄ plate R.
Test solution Dissolve 0.5 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.
Reference solution (a) Dilute 1 mL of the test solution to 10 mL with methanol R. Dilute 1 mL of the solution to 20 mL with methanol R.
Reference solution (b) Dilute 5 mL of reference solution (a) to 10 mL with methanol R.

Relative retentions With reference to β -sitosterol (retention time = about 16 min): cholesterol = about 0.7;
Injection 1 μ L.
Detection Flame ionisation.
— *injection port and detector*: 300 °C.
— *column*: 280 °C;
Temperature:
Split ratio 1:20.
Flow rate: 2 mL/min.
Carrier gas: hydrogen for chromatography R.
Stationary phase: poly(dimethyl)(diphenyl)(divinyl)siloxane R (1 μ m).

Reference solution (a) Dissolve 25 mg of β -sitosterol R and 25 mg of sitostanol R in tetrahydrofuran R and dilute to 10.0 mL with the same solvent. Introduce 100 μ L of this solution into a 3 mL flask and evaporate to dryness under nitrogen R. Add 100 μ L of a freshly prepared mixture of 50 μ L of 1-methylimidazole R and 1.0 mL of heptafluoro- N -(trimethylsilyl)butanamide R. Close the flask tightly and heat to 100 °C for 15 min. Allow to cool.
Reference solution (b) Dissolve 0.100 g of β -sitosterol R in tetrahydrofuran R and dilute to 10.0 mL with the same solvent. Introduce 100 μ L of this solution into a 3 mL flask and evaporate to dryness under nitrogen R. Add 100 μ L of a freshly prepared mixture of 50 μ L of 1-methylimidazole R and 1.0 mL of heptafluoro- N -(trimethylsilyl)butanamide R. Close the flask tightly and heat to 100 °C for 15 min. Allow to cool.

Figure 1911-1. - Chromatogram for the assay of picotamide monohydrate (trimethylsilyl derivatives)





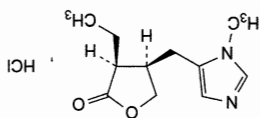
D. (pyridin-3-yl)methanamine.

Ph Eur



Pilocarpine Hydrochloride

(Ph. Eur. monograph 0633)



244.7

 $C_{11}H_{17}ClN_2O_2$

54-71-7

Action and use

Cholinocceptor agonist; treatment of glaucoma.

Preparation

Pilocarpine Hydrochloride Eye Drops

Ph Eur

DEFINITION

(3S,4R)-3-Ethyl-4-[(1-methyl-1H-imidazol-5-yl)methyl]dihydrofuran-2(3H)-one hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals, hygroscopic.

Solubility

Very soluble in water and freely soluble in ethanol

(96 per cent).

mp

About 203 °C.

IDENTIFICATION

First identification A, B, D

Second identification A, C, D

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison pilocarpine hydrochloride CRS.

If the substances are examined as discs, prepare them using

potassium chloride R.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in methanol R and dilute to 2 mL with the same solvent.

Reference solution Dissolve 10 mg of pilocarpine

hydrochloride CRS in methanol R and dilute to 2 mL with the

same solvent.

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R, methanol R, methylene

chloride R (1:14:85 V/V/V).

Application 2 µL.

Development Over a path of 15 cm.

Drying At 100-105 °C for 10 min, then allow to cool.

Detection Spray with dilute potassium iodobismuthate solution R.

IMPURITIES

$C_{21}H_{20}N_4O_3$.

1 mL of 0.1 M perchloric acid is equivalent to 18.82 mg of

(2.2.20).

perchloric acid, determining the end-point potentiometrically

acid R and 20 mL of acetic anhydride R. Titrate with 0.1 M

Dissolve 0.150 g in a mixture of 20 mL of anhydrous acetic

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

semi-micro determination of water.

4.5 per cent to 5.0 per cent, determined on 0.300 g by the

Water (2.5.12)

85 volumes of methanol R.

(100 ppm Pb) R with a mixture of 15 volumes of water R and

(1 ppm Pb) obtained by diluting lead standard solution

Prepare the reference solution using lead standard solution

solution complies with test B for heavy metals (20 ppm).

20 mL with the same mixture of solvents, 12 mL of the

of water R and 85 volumes of methanol R and dilute to

Dissolve 1.0 g by gently heating in a mixture of 15 volumes

Heavy metals (2.4.8)

limit test for chlorides (200 ppm).

and 12.5 mL of water R. The solution complies with the

Dissolve 0.25 g in a mixture of 2.5 mL of dilute nitric acid R

Chlorides (2.4.4)

shows two clearly separated principal spots.

unless the chromatogram obtained with reference solution (c)

reference solution (b) (0.25 per cent). The test is not valid

intense than the spot in the chromatogram obtained with

(0.5 per cent) and not more than one such spot is more

the chromatogram obtained with reference solution (a)

principal spot, is not more intense than the principal spot in

chromatogram obtained with the test solution, apart from the

examine in ultraviolet light at 254 nm. Any spot in the

8 volumes of butanol R. Allow the plate to dry in air and

acid R, 1 volume of water R, 2.5 volumes of methanol R and

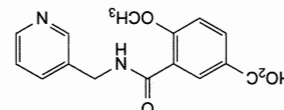
path of 15 cm using a mixture 0.8 volumes of glacial acetic

Apply to the plate 5 µL of each solution. Develop over a

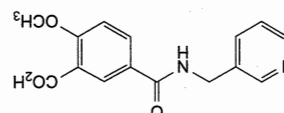
methanol R and dilute to 10 mL with the same solvent.

examined and 5 mg of picotamide impurity A CRS in

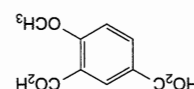
Reference solution (c) Dissolve 0.5 g of the substance to be



B. 2-methoxy-5-[(pyridin-3-



A. 4-methoxybenzene-1,3-dicarboxylic acid,



Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

pH (2.2.3)

3.5 to 4.5 for solution S.

Specific optical rotation (2.2.7)

+ 89 to + 93 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with water R. Dilute 2.0 mL of this solution to 20.0 mL with water R.

Reference solution (b) Dissolve 5.0 mg of pilocarpine nitrate for system suitability CRS (containing impurity A) in water R and dilute to 50.0 mL with the same solvent.

Reference solution (c) To 5 mL of the test solution, add 0.1 mL of ammonia R and heat the solution on a water-bath for 30 min, cool and dilute to 25 mL with water R. Dilute 3 mL of this solution to 25 mL with water R. Mainly pilocarpic acid (impurity B) is formed.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R₁ (5 μ m) with a pore size of 10 nm and a carbon loading of 19 per cent.

Mobile phase Mix 55 volumes of methanol R, 60 volumes of acetonitrile R and 85 volumes of a 0.679 g/L solution of tetrabutylammonium dihydrogen phosphate R previously adjusted to pH 7.7 with dilute ammonia R₂.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Run time Twice the retention time of pilocarpine.

Elution order Impurity B, impurity C, impurity A, pilocarpine.

Retention time Pilocarpine = about 20 min.

System suitability: reference solution (b):

— resolution: minimum 1.6 between the peaks due to impurity A and pilocarpine.

Limits:

— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);

— sum of impurities A and B: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);

— sum of impurities other than A and B: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent).

Iron (2.4.9)

Maximum 10 ppm, determined on solution S. Prepare the standard using 5 mL of iron standard solution (1 ppm Fe) R and 5 mL of water R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of ethanol (96 per cent) R and add 5 mL of 0.01 M hydrochloric acid. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). Read the volume added between the 2 points of inflexion.

1 mL of 0.1 M sodium hydroxide is equivalent to 24.47 mg of C₁₁H₁₇ClN₂O₂.

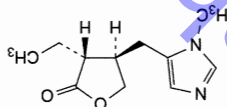
STORAGE

In an airtight container, protected from light.

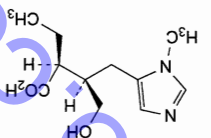
IMPURITIES

Specified impurities A, B

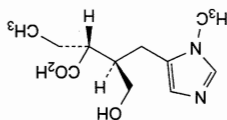
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2.0.34). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



A. (3R,4R)-3-ethyl-4-[(1-methyl-1H-imidazol-5-yl)methyl]dihydrofuran-2(3H)-one (isopilocarpine),



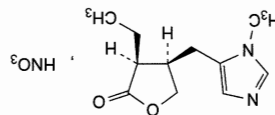
B. (2S,3R)-2-ethyl-3-(hydroxymethyl)-4-(1-methyl-1H-imidazol-5-yl)butanoic acid (pilocarpic acid),



C. (2R,3R)-2-ethyl-3-(hydroxymethyl)-4-(1-methyl-1H-imidazol-5-yl)butanoic acid (isopilocarpic acid).

Pilocarpine Nitrate

(Ph. Eur. monograph 0104)

 $C_{11}H_{17}N_3O_5$

271.3

148-72-1

Action and use
Cholinergic agonist; treatment of glaucoma.**Preparation**

Pilocarpine Nitrate Eye Drops

Ph Eur

DEFINITION

(3S,4R)-3-Ethyl-4-[(1-methyl-1H-imidazol-5-yl)methyl]dihydrofuran-2(3H)-one nitrate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless crystals, sensitive to light.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent).

m.p.

About 174 °C, with decomposition.

IDENTIFICATION

First identification A, B, D

Second identification A, C, D

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison pilocarpine nitrate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in water R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of pilocarpine nitrate CRS in water R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R, methanol R, methylene chloride R (1:14:85 V/V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying At 100-105 °C for 10 min and allow to cool.

Detection Spray with potassium iodobismuthate solution R.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives the reaction of nitrates (2.3.1).

TESTS**Solution S**

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent. Prepare immediately before use.

Appearance of solution
Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).**pH (2.2.3)**

3.5 to 4.5 for solution S.

Specific optical rotation (2.2.7)

+ 80 to + 83 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with water R. Dilute 2.0 mL of this solution to 20.0 mL with water R.

Reference solution (b) Dissolve 5.0 mg of pilocarpine nitrate for system suitability CRS (containing impurity A) in water R and dilute to 50.0 mL with the same solvent.

Reference solution (c) To 5 mL of the test solution, add 0.1 mL of ammonia R and heat the solution on a water-bath for 30 min, cool and dilute to 25 mL with water R. Dilute 3 mL of this solution to 25 mL with water R. Mainly pilocarpic acid (impurity B) is formed.

Column:

— size: l = 0.15 m, Ø = 4.6 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R₁ (5 µm) with a pore size of 10 nm and a carbon loading of 19 per cent.Mobile phase Mix 55 volumes of methanol R, 60 volumes of acetonitrile R and 885 volumes of a 0.679 g/L solution of tetrabutylammonium dihydrogen phosphate R previously adjusted to pH 7.7 with dilute ammonia R₂.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time Twice the retention time of pilocarpine.

Elution order Impurity B, impurity C, impurity A, pilocarpine.

Retention time Pilocarpine = about 20 min.

System suitability: reference solution (b):

— resolution: minimum 1.6 between the peaks due to impurity A and pilocarpine.

Limits:

— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent);

— sum of impurities A and B: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);

— sum of impurities other than A and B: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— with a relative retention with reference to pilocarpine of about 0.3.

Chlorides (2.4.4)

Maximum 70 ppm, determined on solution S.

Iron (2.4.9)

Maximum 10 ppm, determined on solution S. Prepare the standard using 5 mL of iron standard solution (1 ppm Fe) R and 5 mL of water R.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 27.13 mg of $C_{19}H_{18}N_4O_2$.

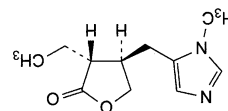
STORAGE

Protected from light.

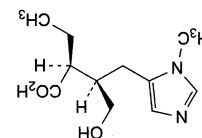
IMPURITIES

Specified impurities A, B

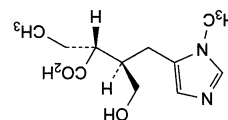
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



A. (3R,4R)-3-ethyl-4-[(1-methyl-1H-imidazol-5-yl)methyl]dihydrofuran-2(3H)-one (isopilocarpine).



B. (2S,3R)-2-ethyl-3-(hydroxymethyl)-4-(1-methyl-1H-imidazol-5-yl)butanoic acid (pilocarpic acid).

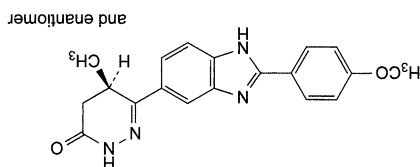


C. (2R,3R)-2-ethyl-3-(hydroxymethyl)-4-(1-methyl-1H-imidazol-5-yl)butanoic acid (isopilocarpic acid).

Ph Eur

Pimobendan

(Ph. Eur. monograph 2179)



$C_{19}H_{18}N_4O_2$ 334.4 74150-27-9

Action and use

Inhibitor of phosphodiesterase type III; calcium sensitizer.

Ph Eur

DEFINITION

(5R)-6-[2-(4-Methoxyphenyl)-1H-benzimidazol-5-yl]-5-methyl-4,5-dihydropyridazin-3(2H)-one.

CHARACTERS

Appearance

White or slightly yellowish powder, hygroscopic.

Solubility

Practically insoluble in water, freely soluble in dimethylformamide, slightly soluble in acetone and in methanol.

mp

About 242 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison pimobendan CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent. Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve the contents of a vial of pimobendan for system suitability CRS (impurities A and B) in 1.0 mL of methanol R.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm,

— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R1 (5 μ m),

— temperature: 45 °C.

Mobile phase:

— mobile phase A: dissolve 3.0 g of potassium dihydrogen phosphate R in 950 mL of water for chromatography R, adjust to pH 2.5 with dilute phosphoric acid R and dilute to 1000 mL with water for chromatography R,

— mobile phase B: acetonitrile for chromatography R,

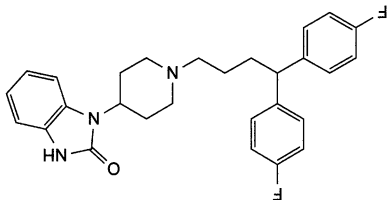
Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 6	85 → 80	15 → 20
6 - 20	80 → 20	20 → 80
20 - 20.1	20 → 85	80 → 15
20.1 - 30	85	15





Pimozide

(Ph Eur monograph 1254)



$C_{28}H_{29}F_2N_3O$ 461.6 2062-78-4

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Pimozide Tablets

Ph Eur

DEFINITION

1-[1-[4,4-Bis(4-fluorophenyl)butyl]piperidin-4-yl]-1,3-dihydro-2H-benzimidazol-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water; soluble in methylene chloride, sparingly soluble in methanol, slightly soluble in ethanol.

(96 per cent).

IDENTIFICATION

First identification B

Second identification A, C, D

A. Melting point (2.2.14): 216 °C to 220 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison pimozide CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 10 mL with the mobile phase.

phase.

Reference solution (a) Dissolve 30 mg of pimozide CRS in the mobile phase and dilute to 10 mL with the mobile phase.

Reference solution (b) Dissolve 30 mg of pimozide CRS and 30 mg of benperidol CRS in the mobile phase and dilute to 10 mL with the mobile phase.

Plate TLC silica gel plate R.

Mobile phase acetone R, methanol R (10:90 V/V).

Application 10 µL.

Development Over 3/4 of the plate.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear.

System suitability: reference solution (b):

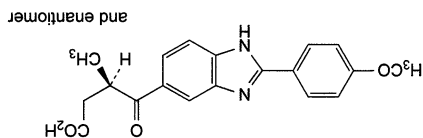
the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference

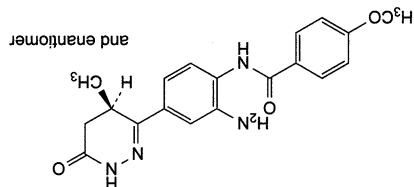
solution (a).

D. Mix about 5 mg with 45 mg of heavy magnesium oxide R and ignite in a crucible until an almost white residue is

Flow rate 1 mL/min.
Detection Spectrophotometer at 290 nm.
Injection 10 µL.
Relative retention With reference to pimobendan (retention time = about 8.3 min): impurity A = about 1.3; impurity B = about 1.4.
System suitability: reference solution (b):
— resolution: minimum 2.0 between the peaks due to impurity A and impurity B.
Limits:
— impurities A, B: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a)
— any other impurity: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a)
— total: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).
Heavy metals (2.4.8)
Maximum 10 ppm.
2.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.
Water (2.5.12)
Maximum 1.0 per cent, determined on 0.500 g.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.
Assay
Dissolve 0.250 g in 5 mL of anhydrous formic acid R. Add 10 mL of acetic anhydride R and 70 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 33.44 mg of $C_{19}H_{18}N_4O_2$.
In an airtight container.
IMPURITIES
Specified impurities: A, B.



A. (3RS)-4-[2-(4-methoxyphenyl)-1H-benzimidazol-5-yl]-3-methyl-4-oxobutanoic acid,



B. N-[2-amino-4-[(4RS)-4-methyl-6-oxo-1,4,5,6-tetrahydropyridazin-3-yl]phenyl]-4-methoxybenzamide.

Ph Eur

obtained (usually less than 5 min). Allow to cool, then add 1 mL of water R, 0.05 mL of phenolphthalein solution R1 and about 1 mL of dilute hydrochloric acid R to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of alizarin S solution R and 0.1 mL of zincanyl nitrate solution R, add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Dissolve 0.2 g in methanol R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of pimozide CRS and 2.0 mg of mebendazole CRS in methanol R and dilute to

100.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to

10.0 mL with methanol R.

Column:

— size: $l = 0.1$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(3 μ m).

Mobile phase:

— mobile phase A: solution containing 2.5 g/L of ammonium

acetate R and 8.5 g/L of tetrabutylammonium hydrogen

sulfate R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	80 \rightarrow 70	20 \rightarrow 30
10 - 15	70	30
15 - 20	80	20

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 μ L.

Relative retention With reference to pimozide (retention

time = about 8 min): impurity A = about 0.1;

mebendazole = about 0.88; impurity B = about 0.9;

impurity C = about 0.95; impurity D = about 1.1;

impurity E = about 1.3.

System suitability: reference solution (a):

— resolution: minimum 5.0 between the peaks due to

mebendazole and pimozide.

Limits:

— impurities A, B, C, D, E: for each impurity, not more than

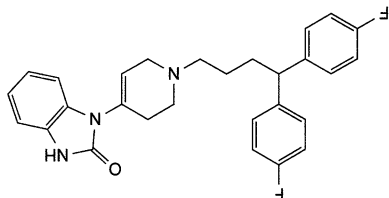
the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.5 per cent);

0.2 times the area of the principal peak in the

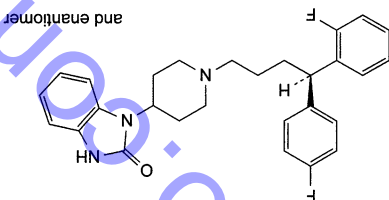
chromatogram obtained with reference solution (b)

(0.10 per cent);

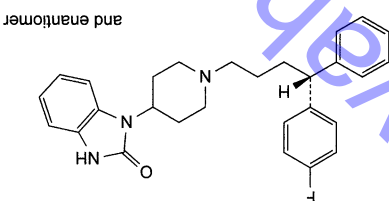


D. 1-[1-[1-[4,4-bis(4-fluorophenyl)butyl]-1,2,3,6-tetrahydropyridin-4-yl]-1,3-dihydro-2H-benzimidazol-2-one,

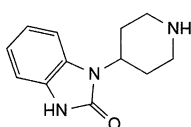
C. 1-[1-[1-[(4R)-4-(2-fluorophenyl)-4-(4-fluorophenyl)butyl]piperidin-4-yl]-1,3-dihydro-2H-benzimidazol-2-one,



B. 1-[1-[1-[(4R)-4-(4-fluorophenyl)-4-phenylbutyl]piperidin-4-yl]-1,3-dihydro-2H-benzimidazol-2-one,



A. 1-(piperidin-4-yl)-1,3-dihydro-2H-benzimidazol-2-one,



Specified impurities A, B, C, D, E

IMPURITIES

Protected from light.

STORAGE

of C₂₈H₂₉F₂N₃O.

1 mL of 0.1 M perchloric acid is equivalent to 46.16 mg

naphtholbenzenesulfonic acid as indicator.

Titrate with 0.1 M perchloric acid, using 0.2 mL of

anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R.

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of

ASSAY

crucible.

Maximum 0.1 per cent, determined on 1.0 g in a platinum

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

(0.05 per cent).

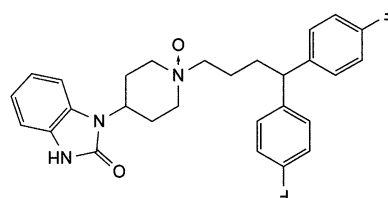
the chromatogram obtained with reference solution (b)

— disregard limit: 0.1 times the area of the principal peak in

solution (b) (0.75 per cent);

peak in the chromatogram obtained with reference

— total: not more than 1.5 times the area of the principal

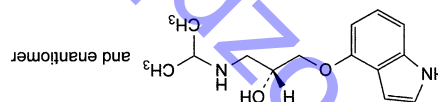


E. 1-[1-[4,4-bis(4-fluorophenyl)butyl]piperidin-4-yl] 1-oxide]-1,3-dihydro-2H-benzimidazol-2-one.

Ph Eur

Pindolol

(Ph. Eur. monograph 0634)



13523-86-9

Action and use

Beta-adrenoceptor antagonist.

Preparation

Pindolol Tablets

Ph Eur

DEFINITION

Pindolol contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (2*RS*)-1-(1*H*-indol-4-yloxy)-3-[(1-methylethyl)amino]propan-2-ol, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, practically insoluble in water, slightly soluble in methanol. It dissolves in dilute mineral acids.

IDENTIFICATION

First identification A, C.

Second identification A, B, D.

A. Melting point (2.2.14): 169 °C to 174 °C.

B. Dissolve 20.0 mg in a 0.085 per cent *V/V* solution of hydrochloric acid R in methanol R and dilute to 100.0 mL with the same solution. Dilute 10.0 mL of the solution to 100.0 mL with a 0.085 per cent *V/V* solution of hydrochloric acid R in methanol R. Examined between 230 nm and 320 nm (2.2.25), the solution shows two absorption maxima, at 264 nm and at 287 nm, and a shoulder at 275 nm. The specific absorbance at the maxima are 330 to 350 and 170 to 190, respectively.

C. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with pindolol CRS.

D. Examine in daylight the chromatograms on plate A obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

Store protected from light.

STORAGE

$C_{14}H_{20}N_2O_2$
1 mL of 0.1 M hydrochloric acid is equivalent to 24.83 mg of (2.2.20).
Dissolve 0.200 g in 80 mL of methanol R. Titrate with 0.1 M hydrochloric acid, determining the end-point potentiometrically (2.2.20).

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

drying in an oven at 105 °C.

Not more than 0.5 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

solution (10 ppm Pb) R.

Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Heavy metals (2.4.8)

1.0 g complies with test C for heavy metals (20 ppm).

Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R. Examine the chromatogram obtained with test solution (a), apart from the principal spot and the spots detected on plate A, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.3 per cent).

B. Apply separately 10 µL of each solution. Develop the plate without delay over a path of 10 cm using a freshly prepared mixture of 4 volumes of concentrated ammonia R, 50 volumes of ethyl acetate R and 50 volumes of methanol R. Dry the plate briefly in a current of cold air. Examine the plate without delay in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.3 per cent).

A. Apply separately 5 µL of each solution. Develop the plate without delay over a path of 10 cm using a freshly prepared mixture of 4 volumes of concentrated ammonia R, 50 volumes of ethyl acetate R and 50 volumes of methanol R. Dry the plate briefly in a current of cold air. Spray the plate without delay with dimethylaminobenzaldehyde solution R7 and heat to 50 °C for 20 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.3 per cent).

Test solution (a) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of methanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of methanol R.

Reference solution (a) Dissolve 20 mg of pindolol CRS in a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of solvents.

Reference solution (b) Dilute 1.5 mL of reference solution (a) to 50 mL with a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of methanol R.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance. Carry out all operations as rapidly as possible, protected from light.

Test solution (a) Dissolve 0.10 g of the substance to be examined in a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of solvents. Prepare immediately before use and apply this solution to the plate last.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of methanol R.

Reference solution (a) Dissolve 20 mg of pindolol CRS in a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of methanol R.

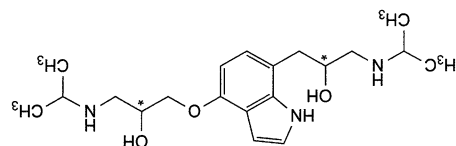
Reference solution (b) Dilute 1.5 mL of reference solution (a) to 50 mL with a mixture of 1 volume of anhydrous acetic acid R and 99 volumes of methanol R.

TESTS

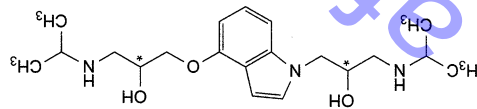
Appearance of solution

Dissolve 0.5 g in dilute acetic acid R and dilute to 10 mL with the same acid. The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₅ or B₅ (2.2.2, Method II).

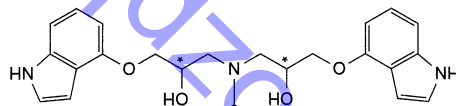
IMPUITIES



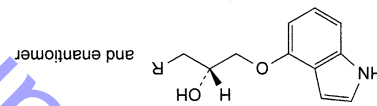
A. 1-[1-[7-[2-hydroxy-3-[(1-methylethyl)amino]propyl]-1H-indol-4-yl]oxy]-3-[(1-methylethyl)amino]propan-2-ol,



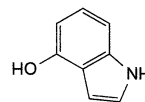
B. 1-[4-[2-hydroxy-3-[(1-methylethyl)amino]propoxy]-1H-indol-1-yl]-3-[(1-methylethyl)amino]propan-2-ol,



C. 1,1'-[(1-methylethyl)imino]bis[3-(1*H*-indol-4-yl)oxy]propan-2-ol],

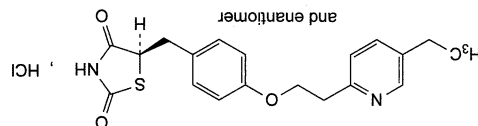


D, R = OH: (2*RS*)-3-(1*H*-indol-4-ylxy)propane-1,2-diol,
F, R = Cl: (2*RS*)-1-chloro-3-(1*H*-indol-4-ylxy)propan-2-ol,

E. 1*H*-indol-4-ol.

Pioglitazone Hydrochloride

(Ph. Eur. monograph 2601)



C₁₉H₂₁ClN₂O₃S 392.9 112529-15-4

Action and use

Peroxisome proliferator-activated receptor (PPAR)-gamma agonist; treatment of diabetes mellitus.

DEFINITION

Ph Eur —

Content
(3K5)-5-[4-[2-(5-ethylpyridin-2-yl)ethoxy]phenyl]methylthiazolidine-2,4-dione hydrochloride.
98.0 per cent to 102.0 per cent (anhydrous substance).

Content

CHARACTERS

Appearance

White or almost white crystals or crystalline powder.

Solubility

Practically insoluble in water, slightly soluble to soluble in methanol, very slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison pioglitazone hydrochloride CRS.

B. Dissolve 25 mg in 0.5 mL of *nitric acid R* and add 2 mL of *dilute nitric acid R*. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Specific optical rotation (2.2.7)

Dissolve 2.5 g in *dimethylformamide* R and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 20 mg of the substance to be examined in 20 mL of *methanol R* and dilute to 100.0 mL.

Test solution (b) Dissolve 50.0 mg of the substance to be examined in *methanol R* and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Suspend 5 mg of progesterone for system suitability CRS (containing impurities B and C) in 5 mL of methanol R. Heat at 60 °C for about 30 s, cool to room

Fluxes through a membrane filter (nominal pore size 0.45 μm).

Reference solution (c) Dissolve 50.0 mg of progesterone in methanol R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 20.0 mL with the mobile phase.

Column: size: $l = 0.15$ m, $\varnothing = 4.6$ mm; stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase glacial acetic acid R, acetonitrile R, 7:1 v/v

Solution of *aminomycin acetate* λ (1.25-2.5 μ m):
Flow rate 0.7 mL/min.
Detection Spectrophotometer at 269 nm.
Injection 40 μ L of test solution (a) and reference solutions (a) and (b).

Run time t times the retention time of progesterone. Identification of impurities Use the chromatogram supplied with progesterone for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and C.

– *Resolution*: minimum 5.0 between the peaks due to pioglitazone and impurity B.

Limits:

- **impurities B, C:** for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
- **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
- **total:** not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
- **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.32)

Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

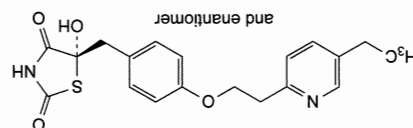
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection 20 µL of test solution (b) and reference solution (c). Calculate the percentage content of $C_{19}H_{21}ClN_5O_3$ taking into account the assigned content of pioglitazone hydrochloride CRS.

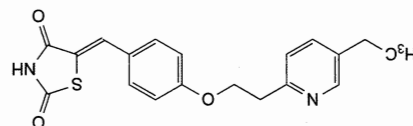
IMPURITIES

Specified impurities B, C.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D, E.



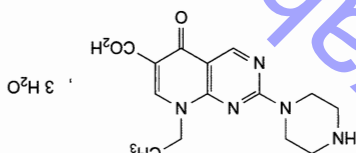
A. (5R,5'-S)-5'-[4-{2-(5-ethoxypyridin-2-yl)ethoxy}phenyl]methyl-5-hydroxythiazolidine-2,4-dione,



B. (5Z)-5'-[4-{2-(5-ethoxypyridin-2-yl)ethoxy}phenyl]methylthiazolidine-2,4-dione,

Pipemidic Acid Trihydrate

(Ph. Eur. monograph 1743)

 $C_{14}H_{17}N_5O_3 \cdot 3H_2O$

357.4

72571-82-5

Action and use

Antibacterial.

DEFINITION

8-Ethyl-5-oxo-2-(piperazin-1-yl)-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylic acid trihydrate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

Pale yellow or yellow, crystalline powder.

Solubility

Very slightly soluble in water and in methylene chloride, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of acids and of alkali hydroxides.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison pipemidic acid trihydrate CRS.

TESTS**Related substances**

Liquid chromatography (2.2.29).



Ph Eur

Test solution Dissolve 20 mg of the substance to be examined in 10 mL of the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 1.0 mg of ethyl parahydroxybenzoate R in 2.0 mL of the test solution and dilute to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R1 (5 μ m).

Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a solution containing 5.7 g/L

of citric acid R and 1.7 g/L of sodium decahydrogenate R.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 20 μ L.

Run time 2.5 times the retention time of pipemidic acid.

Relative retention With reference to pipemidic acid (retention time = about 15 min): ethyl parahydroxybenzoate = about 0.8.

System suitability: reference solution (b):
— resolution: minimum 4.0 between the peaks due to ethyl parahydroxybenzoate and pipemidic acid.

Limits:

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (a)

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

0.5 g complies with test G. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.3.2)

14.0 per cent to 16.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.240 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 30.33 mg of $C_{14}H_{17}N_5O_3$.

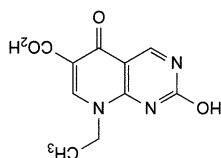
STORAGE

Protected from light.

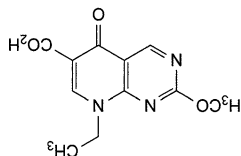
IMPURITIES

Other detectable impurities (the following substances would, if present in the monograph, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

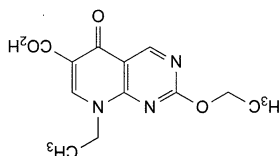
impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F.



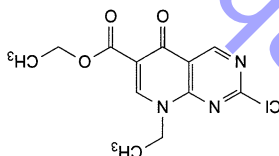
A. 8-ethyl-2-hydroxy-5-oxo-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylic acid,



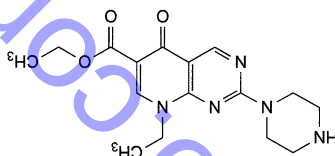
B. 8-ethyl-2-methoxy-5-oxo-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylic acid,



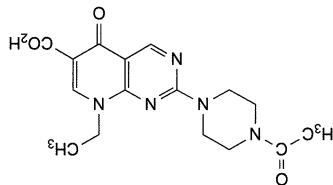
C. 2-ethoxy-8-ethyl-5-oxo-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylic acid,



D. ethyl 2-chloro-8-ethyl-5-oxo-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylate,



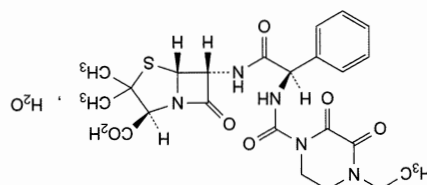
E. ethyl 8-ethyl-5-oxo-2-(piperazin-1-yl)-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylate,



F. 2-(4-acetylpiperazin-1-yl)-8-ethyl-5-oxo-5,8-dihydropyrido[2,3-d]pyrimidine-6-carboxylic acid (acetylpipemidic acid).

Piperacillin

(Ph. Eur. monograph 1169)


 $C_{23}H_{27}N_5O_7S_2 \cdot H_2O$ 535.6 66258-76-2

Action and use

Penicillin antibacterial.

Preparation

Piperacillin Infusion

Ph Eur

DEFINITION

(2S,5R,6R)-6-[[[(4R)-2-[[[(2R)-2,3-dioxopiperazin-1-yl]carbonyl]-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid

monohydrate.

Semi-synthetic product derived from a fermentation product.

Content

96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, freely soluble in methanol, slightly

soluble in ethyl acetate.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison piperacillin CRS.

TESTS

Solution S

Dissolve 2.50 g in sodium carbonate solution R and dilute to

25 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference

suspension II (2.2.1) and its absorbance (2.2.25) at 430 nm is not greater than 0.10.

Specific optical rotation (2.2.7)

+ 168 to + 178 (anhydrous substance).

Dissolve 0.250 g in methanol R and dilute to 25.0 mL with

the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, 31.2 g/L solution of sodium

dihydrogen phosphate R (25:75 V/V).

Test solution (a) Dissolve 25.0 mg of the substance to be

examined in the solvent mixture and dilute to 50.0 mL with

the solvent mixture.

Test solution (b) Prepare the solution immediately before use.

Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent

mixture.

Reference solution (a) Dissolve 25.0 mg of piperacillin CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 25.0 mL with the solvent mixture.

Reference solution (c) Dissolve 10.0 mg of piperacillin CRS and 10.0 mg of anhydrous ampicillin CRS (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

mobile phase A: mix 576 mL of water R, 200 mL of a

31.2 g/L solution of sodium dihydrogen phosphate R and 24 mL of an 80 g/L solution of tetrabutylammonium

hydroxide R; if necessary, adjust to pH 5.5 with dilute phosphoric acid R or dilute sodium hydroxide solution R;

add 200 mL of acetonitrile R;

mobile phase B: mix 126 mL of water R, 200 mL of a

31.2 g/L solution of sodium dihydrogen phosphate R and 24 mL of an 80 g/L solution of tetrabutylammonium

hydroxide R; if necessary, adjust to pH 5.5 with dilute phosphoric acid R or dilute sodium hydroxide solution R;

add 650 mL of acetonitrile R;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - t_R	88	12
$t_R - (t_R + 30)$	88 \rightarrow 0	12 \rightarrow 100
$(t_R + 30) - (t_R + 45)$	0 \rightarrow 88	100 \rightarrow 12
t_R = retention time of piperacillin determined with reference solution (b)		

If the mobile phase composition has been adjusted to achieve the required resolution, the adjusted composition will apply at time zero in the gradient and in the assay.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L of reference solutions (b), (c) and (d) withisocratic elution at the initial mobile phase composition and 20 μ L of test solution (b) according to the elution gradient

described under Mobile phase.

System suitability:

— resolution: minimum 10 between the peaks due to

impurity A and piperacillin in the chromatogram obtained with reference solution (c); if necessary, adjust the ratio

A:B of the mobile phase;

— signal-to-noise ratio: minimum 3 for the principal peak in

the chromatogram obtained with reference solution (d); — mass distribution ratio: 2.0 to 3.0 for the peak due to

piperacillin in the chromatogram obtained with reference

solution (c).

Limit:

— any impurity: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained

with reference solution (b) (2 per cent).

N,N-Dimethylamine (2.4.26, Method A)

Maximum 20 ppm.

Heavy metals (2.4.8)

Maximum 20 ppm.

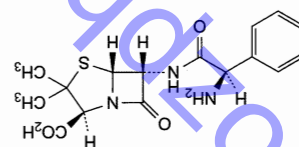
1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

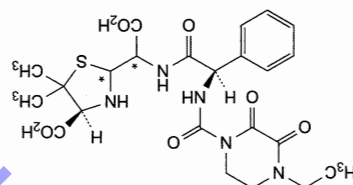
2.0 per cent to 4.0 per cent, determined on 0.500 g.

ASSAY

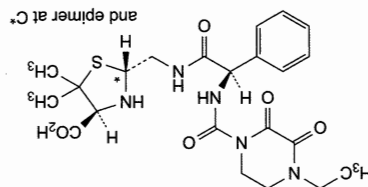
Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.*Injection* Test solution (a) and reference solution (a).*System suitability*: reference solution (a):— *repeatability*: maximum relative standard deviation of 1.0 per cent after 6 injections.**IMPURITIES**

A. (2S,5R,6R)-6-[[[(2R)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (ampicillin),



B. (4S)-2-[carboxy[[[(2R)-2-[[[(4-ethyl-2,3-dioxopiperazin-1-yl)carboxyl]amino]-2-phenylacetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of piperacillin),

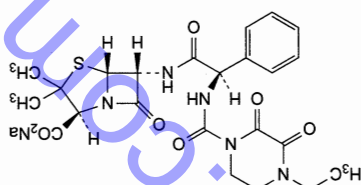


C. (2R,4S)-2-[[[(2R)-2-[[[(4-ethyl-2,3-dioxopiperazin-1-yl)carboxyl]amino]-2-phenylacetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of piperacillin),

Action and use
Penicillin antibacterial. $C_{23}H_{26}N_5NaO_7S$

539.5

59703-84-3

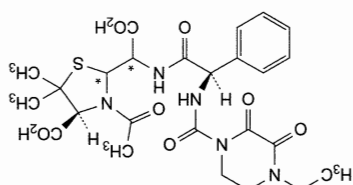


(Ph. Eur. monograph 1168)

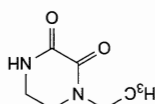
Piperacillin Sodium

Ph Eur

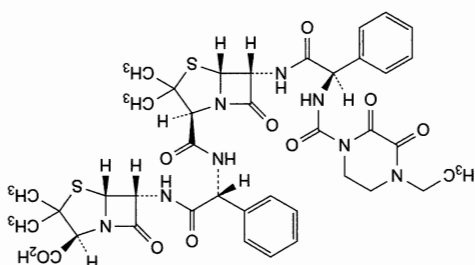
F. (4S)-3-acetyl-2-[carboxy[[[(2R)-2-[[[(4-ethyl-2,3-dioxopiperazin-1-yl)carboxyl]amino]-2-phenylacetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (acetylated penicilloic acids of piperacillin).



E. 1-ethylpiperazine-2,3-dione,



D. (2S,5R,6R)-6-[[[(2R)-2-[[[(2S,5R,6R)-6-[[[(2R)-2-[[[(4-ethyl-2,3-dioxopiperazin-1-yl)carboxyl]amino]-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (piperacillin)ampicillin),



CHARACTERS**Appearance**

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water and in methanol, practically insoluble in ethyl acetate.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 0.250 g in water R₃, add 0.5 mL of dilute hydrochloric acid R and 5 mL of ethyl acetate R₃, stir and allow to stand for 10 min in iced water. Filter the crystals through a small sintered-glass filter (40), applying suction. Wash with 5 mL of water R and 5 mL of ethyl acetate R₃, then dry in an oven at 60 °C for 60 min.

Comparison piperacillin CRS.

B. It gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and its absorbance (2.2.25) at 430 nm is not greater than 0.10.

pH (2.2.3)

5.0 to 7.0 for solution S.

Specific optical rotation (2.2.7)

+175 to +190 (anhydrous substance).

Dissolve 0.250 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R₃, 31.2 g/L solution of sodium dihydrogen phosphate R (25:75 V/V).

Test solution (a) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Test solution (b) Prepare the solution immediately before use.

Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 25.0 mg of piperacillin CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 25.0 mL with the solvent mixture.

Reference solution (c) Dissolve 10.0 mg of piperacillin CRS and 10.0 mg of anhydrous ampicillin CRS (impurity A) in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: mix 576 mL of water R₃, 200 mL of a 31.2 g/L solution of sodium dihydrogen phosphate R and 24 mL of an 80 g/L solution of tetrabutylammonium hydroxide R₃; if necessary, adjust to pH 5.5 with dilute

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - t_R	88	12
$t_R - (t_R + 30)$	88 \rightarrow 0	12 \rightarrow 100
$(t_R + 30) - (t_R + 45)$	0 \rightarrow 88	100 \rightarrow 12

add 200 mL of acetonitrile R₃; phosphoric acid R or dilute sodium hydroxide solution R₃; mobile phase B: mix 126 mL of water R₃, 200 mL of a 31.2 g/L solution of sodium dihydrogen phosphate R and 24 mL of an 80 g/L solution of tetrabutylammonium hydroxide R₃; if necessary, adjust to pH 5.5 with dilute phosphoric acid R or dilute sodium hydroxide solution R₃; add 650 mL of acetonitrile R₃;

Flow rate 1.0 mL/min. Detection Spectrophotometer at 220 nm. Injection 20 μ L of reference solutions (b), (c) and (d) with isocratic elution at the initial mobile phase composition and 20 μ L of test solution (b) according to the elution gradient described under Mobile phase.

System suitability: — resolution: minimum 10 between the peaks due to impurity A and piperacillin in the chromatogram obtained with reference solution (c); if necessary, adjust the ratio A:B of the mobile phase;

— signal-to-noise ratio: minimum 3 for the principal peak in the chromatogram obtained with reference solution (d); — mass distribution ratio: 2.0 to 3.0 for the peak due to piperacillin in the chromatogram obtained with reference solution (c).

Limit: — any impurity: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent).

N,N-Dimethylamine (2.4.26, Method A)

Maximum 20 ppm.

Heavy metals (2.4.8)

Maximum 20 ppm.

Water (2.5.12)

Maximum 2.0 per cent, determined on 0.500 g.

Bacterial endotoxins (2.6.14)

Less than 0.07 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Initial composition of the mixture of mobile phases A and B, adjusted where applicable.

Injection Test solution (a) and reference solution (a).

System suitability: reference solution (a): — repeatability: maximum relative standard deviation of 1.0 per cent after 6 injections.

Calculate the percentage content of piperacillin sodium, multiplying the result by 1.042.

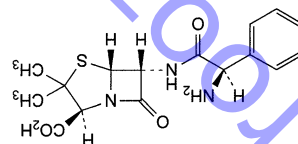
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

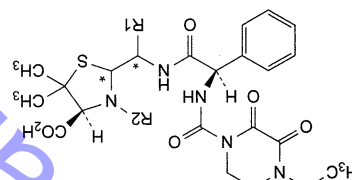
IMPURITIES

Specified impurities A, B, C, D, E, F, G.

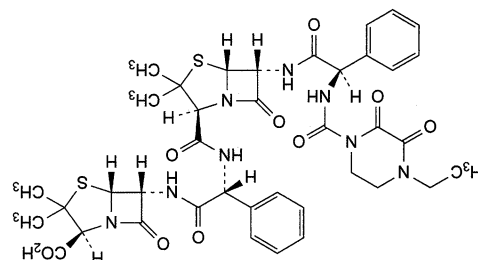
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): H.



A. (2S,5R,6R)-6-[[[(2R)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (ampicillin),



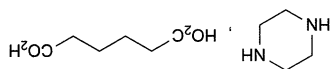
B. R1 = CO₂H, R2 = H: (4S)-2-[[carboxy[(2R)-2-[[[(4S)-3-acetyl-2-carboxy-1-yl]carboxyl]amino]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of piperacillin), R1 = CO₂H, R2 = CO-CH₃: (4S)-3-acetyl-2-[[carboxy[(2R)-2-[[[(4-ethyl-2,3-dioxopiperazin-1-yl]carboxyl]amino]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of piperacillin),



acids of piperacillin), dimethylthiazolidine-4-carboxylic acid (acetylated penicilloic

Piperazine Adipate

(Ph. Eur. monograph 0423)



C₁₀H₂₀N₂O₄ 232.3

I42-88-1

Action and use

Anthelmintic.

DEFINITION

Piperazine adipate contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of piperazine hexanedioate, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white crystalline powder, soluble in water, practically insoluble in alcohol. It melts at about 250 °C, with decomposition.

IDENTIFICATION

First identification A.

Second identification B, C.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with piperazine adipate CRS. Examine the substances prepared as discs.

B. Examine the chromatograms obtained in the test for related substances after spraying with the ninhydrin solutions. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 10 mL of solution S (see Tests) add 5 mL of hydrochloric acid R and shake with three quantities, each of 10 mL, of ether R. Evaporate the combined ether layers to dryness. The residue, washed with 5 mL of water R and dried at 100 °C to 105 °C, melts (2.2.14) at 150 °C to 154 °C.

TESTS

Solution S
Dissolve 2.5 g in water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₈ (2.2.2, Method II).

Related substances

Examine by thin-layer chromatography (2.2.27), using a suitable silica gel as the coating substance.

Test solution (a) Dissolve 1.0 g of the substance to be examined in 6 mL of concentrated ammonia R and dilute to 10 mL with ethanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R.

Reference solution (a) Dissolve 0.1 g of piperazine adipate CRS in a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 10 mL with the same mixture of solvents.

Reference solution (b) Dissolve 25 mg of ethylenediamine R in a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (c) Dissolve 25 mg of triethylenediamine R in a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (d) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (e) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (f) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (g) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (h) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (i) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (j) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (k) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (l) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (m) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (n) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (o) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (p) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (q) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (r) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (s) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (t) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (u) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

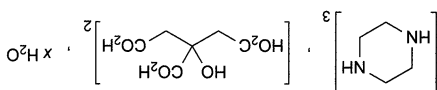
Reference solution (v) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (w) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (x) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Piperazine Citrate

(Ph. Eur. monograph 0424)



(anhydrous substance)

$C_{24}H_{46}N_6O_{14} \cdot xH_2O$

643

41372-10-5



Ph Eur

$C_{10}H_{20}N_2O_4$
1 mL of 0.1 M perchloric acid is equivalent to 11.61 mg of changes from brownish-yellow to green.

naphtholbenzenesulfonate R as indicator until the colour changes from brownish-yellow to green.
Titrant with 0.1 M perchloric acid using 0.25 mL of gentle heating and dilute to 70 mL with the same acid.

ASSAY

Dissolve 0.100 g in 10 mL of anhydrous acetic acid R with gentle heating and dilute to 70 mL with the same acid.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

Water (2.5.12)
Not more than 0.5 per cent, determined on 1.00 g by the semi-micro determination of water.

IDENTIFICATION

First identification A.

Second identification B, C.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with piperazine citrate CRS. Dry the substance to be examined and the reference substance at 120 °C for 5 h, powder the substances avoiding uptake of water, prepare discs and record the spectra without delay.

B. Examine the chromatograms obtained in the test for related substances after spraying with the ninhydrin solution. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.5 g in water R and dilute to 5 mL with the same solvent. The solution gives the reaction of citrates (2.3.1).

TESTS

Solution S
Dissolve 1.25 g in water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₈ (2.2.2, Method II).

Related substances

Examine by thin-layer chromatography (2.2.27), using a suitable silica gel as the coating substance.

Test solution (a) Dissolve 1.0 g of the substance to be examined in 6 mL of concentrated ammonia R and dilute to 10 mL with anhydrous ethanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a mixture of 2 volumes of anhydrous ethanol R and 3 volumes of concentrated ammonia R.

Reference solution (a) Dissolve 0.1 g of piperazine citrate CRS in a mixture of 2 volumes of anhydrous ethanol R and 3 volumes of concentrated ammonia R.

Reference solution (b) Dissolve 25 mg of ethylenediamine R in a mixture of 2 volumes of anhydrous ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (c) Dissolve 25 mg of triethylenediamine R in a mixture of 2 volumes of anhydrous ethanol R and 3 volumes of concentrated ammonia R and dilute to 100 mL with the same mixture of solvents.

Reference solution (d) Dissolve 12.5 mg of triethylenediamine R in 5.0 mL of test solution (a) and dilute to 50 mL with a mixture of 2 volumes of anhydrous ethanol R and 3 volumes of concentrated ammonia R.

Apply separately to the plate 5 µL of each solution. Develop over a path of 15 cm using a freshly prepared mixture of 20 volumes of concentrated ammonia R and 80 volumes of acetone R. Dry the plate at 105 °C and spray successively with a 3 g/L solution of ninhydrin R in a mixture of 3 volumes of anhydrous acetic acid R and 100 volumes of butanol R and a 1.5 g/L solution of ninhydrin R in anhydrous ethanol R. Dry the plate at 105 °C for 10 min. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b).

(0.25 per cent). Spray the plate with 0.05 M iodine and allow to stand for about 10 min. Any spot corresponding to triethylenediamine in the chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with reference solution (c).

(0.25 per cent). The test is not valid unless the chromatogram obtained with reference solution (d) shows 2 clearly separated spots. Disregard any spots remaining on the line of application.

Heavy metals (2.4.8)

12 mL of solution S complies with limit test A for heavy metals (20 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

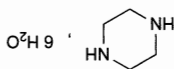
10.0 per cent to 14.0 per cent, determined on 0.300 g by the semi-micro determination of water.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

Piperazine Hydrate

(Ph. Eur. monograph 0425)



C₄H₁₀N₂·6H₂O

194.2

142-63-2

Action and use

Anthelmintic.

Ph. Eur.

DEFINITION

Piperazine hexahydrate.

Content

98.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

Colourless, deliquescent crystals.

Solubility

Freely soluble in water and in ethanol (96 per cent).

mp

About 43 °C.

IDENTIFICATION

First identification A.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24). Preparation Dry the substance to be examined and the reference substance over diphosphorus pentoxide R in vacuo for 48 h, powder the substances avoiding uptake of water, prepare discs and record the spectra without delay. Comparison piperazine hydrate CRS.

B. Examine the chromatograms obtained in the test for related substances after spraying with the ninhydrin solutions. Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.5 g in 5 mL of dilute sodium hydroxide solution R. Add 0.2 mL of benzoyl chloride R and mix. Continue to add benzoyl chloride R in portions of 0.2 mL until no further precipitate is formed. Filter and wash the precipitate with a total of 10 mL of water R added in small portions. Dissolve the precipitate in 2 mL of hot ethanol (96 per cent) R and pour the solution into 5 mL of water R. Allow to stand for 4 h, filter, wash the crystals with water R and dry at 100-105 °C. The crystals melt (2.2.14) at 191 °C to 196 °C.

ASSAY
Dissolve 0.100 g in 10 mL of anhydrous acetic acid R with gentle heating and dilute to 70 mL with the same acid. Titrate with 0.1 M perchloric acid using 0.25 mL of naphtholbenzenesulfonate solution R as indicator until the colour changes from brownish-yellow to green.
1 mL of 0.1 M perchloric acid is equivalent to 10.71 mg of C₄H₁₀N₂·6H₂O.
Ph. Eur.



ASSAY

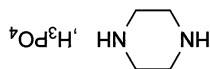
Dissolve 80.0 mg in 10 mL of anhydrous acetic acid R with gentle heating and dilute to 70 mL with the same acid. Titrate with 0.1 M perchloric acid using 0.25 mL of naphtholbenzenesulfonate solution R as indicator until the colour changes from brownish-yellow to green.

1 mL of 0.1 M perchloric acid is equivalent to 9.705 mg of $C_4H_{10}N_2H_3PO_4 \cdot 2H_2O$.

STORAGE

In an airtight container, protected from light.

Ph Eur

Piperazine Phosphate

$C_4H_{10}N_2H_3PO_4 \cdot 2H_2O$ 202.1 18534-18-4

Action and use

Anthelmintic.

Preparation

Chewable Piperazine Phosphate Tablets

Piperazine Phosphate Tablets

DEFINITION

Piperazine Phosphate contains not less than 98.5% and not more than 100.5% of $C_4H_{10}N_2H_3PO_4$ calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white, crystalline powder; odourless or almost odourless.

Sparingly soluble in water; practically insoluble in ethanol (96%).

IDENTIFICATION

A. Dissolve 0.1 g in 5 mL of water, add 0.5 g of sodium hexacyanoferrate(III) and 0.1 mL of mercury. Shake vigorously for 1 minute and allow to stand for 20 minutes. A reddish colour is produced slowly.

B. Dissolve 0.2 g in 5 mL of 2 M hydrochloric acid, add with stirring 1 mL of a 50% w/v solution of sodium nitrite and cool in ice for 15 minutes, stirring if necessary to induce crystallisation. The melting point of the crystals, after washing with 10 mL of iced water and drying at 105°, is about 159°.

Appendix V A.

C. A solution yields the reactions characteristic of phosphates, Appendix VI.

TESTS**Acidity**

pH of a 1% w/v solution, 6.0 to 6.5, Appendix V L.

Heavy metals

Dissolve 2.0 g in 20 mL of 2 M acetic acid. 12 mL of the

resulting solution complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (2 ppm Pb) to prepare the standard (20 ppm).

Water

8.0 to 9.5% w/w, Appendix IX C. Use 0.25 g.

TESTS**Solution S**

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₈ (2.2.2, Method II).

pH (2.2.3)

10.5 to 12.0 for solution S.

Related substances

Thin-layer chromatography (2.2.27).

Solvent mixture anhydrous ethanol R, concentrated ammonia R (40:60 V/V).

Test solution (a) Dissolve 1.0 g of the substance to be

examined in 6 mL of concentrated ammonia R and dilute to

10 mL with anhydrous ethanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL

with the solvent mixture.

Reference solution (a) Dissolve 0.1 g of piperazine hydrate CRS

in the solvent mixture and dilute to 10 mL with the solvent

mixture.

Reference solution (b) Dissolve 25 mg of ethylenediamine R in

the solvent mixture and dilute to 100 mL with the solvent

mixture.

Reference solution (c) Dissolve 12.5 mg of triethylenediamine R

in 5.0 mL of test solution (a) and dilute to 50 mL with the

solvent mixture.

Plate Suitable silica gel as the coating substance.

Mobile phase concentrated ammonia R, acetone R (20:80 V/V);

use a freshly prepared mixture.

Application 5 µL.

Development Over a path of 15 cm.

Drying At 105° C.

Detection A Spray successively with a 3 g/L solution of

minhydram R in a mixture of 3 volumes of anhydrous acetic

acid R and 100 volumes of butanol R and a 1.5 g/L solution

of minhydram R in anhydrous ethanol R and dry the plate at

105° C for 10 min.

Limits A Any spot in the chromatogram obtained with test

solution (a), apart from the principal spot, is not more

intense than the spot in the chromatogram obtained with

reference solution (b) (0.25 per cent).

Detection B Spray with 0.05 M iodine and allow to stand for

about 10 min.

Limits B Any spot corresponding to triethylenediamine in the

chromatogram obtained with test solution (a) is not more

intense than the spot in the chromatogram obtained with

reference solution (c) (0.25 per cent).

System suitability Reference solution (d):

— the chromatogram shows 2 clearly separated spots.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Sulfated ash (2.4.14)

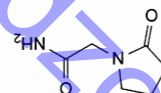
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.2 g in a mixture of 3.5 mL of 0.5M sulfuric acid and 10 mL of water. Add 100 mL of picric acid solution R₁, heat on a water bath for 15 minutes and allow to stand for 1 hour. Filter through a sintered-glass filter (ISO 4793, porosity grade 4, is suitable) and wash the residue with successive 10 mL quantities of a mixture of equal volumes of a saturated solution of picric acid and water until the washings are free from sulfate. Wash the residue with five 10 mL quantities of absolute ethanol and dry to constant weight at 100° to 105°. Each g of residue is equivalent to 0.3382 g of $C_4H_{10}N_2 \cdot H_3PO_4$.

Piracetam

(Ph. Eur. monograph 1733)

 $C_6H_{10}N_2O_2$ 142.2 7491-74-9**Action and use**

Nootropic; cortical myoclonus.

DEFINITION

2-(2-Oxopropylidino-1-yl)acetamide.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison piracetam CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in ethanol (96 per cent) R, evaporate to dryness on a water-bath and record new spectra using the residues.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 2.0 g in water R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in a mixture of 10 volumes of acetonitrile R₁ and 90 volumes of water R and dilute to 100.0 mL with the same mixture of solvents.Test solution (b) Dilute 10.0 mL of test solution (a) to 50.0 mL with a mixture of 10 volumes of acetonitrile R₁ and 90 volumes of water R.

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection Test solution (b) and reference solution (c).

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105°C.

Maximum 1.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

lead standard solution (1 ppm Pb) R.

Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution

Maximum 10 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

— the chromatogram obtained with reference solution (b) (0.3 per cent),
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— total: not more than 3 times the area of the principal peak with reference solution (b) (0.1 per cent),
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— impurities A, B, C, D: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),
— symmetry factor: maximum 2.0 for the peak due to piracetam.

System suitability: reference solution (a):
— resolution: minimum 3.0 between the peaks due to piracetam and impurity A,
— symmetry factor: maximum 2.0 for the peak due to piracetam.

Run time 8 times the retention time of piracetam.
Relative retention With reference to piracetam (retention time = about 4 min): impurity D = about 0.8; impurity A = about 1.15; impurity B = about 2.8; impurity C = about 6.3.
System suitability: reference solution (a):
— resolution: minimum 3.0 between the peaks due to piracetam and impurity A,
— symmetry factor: maximum 2.0 for the peak due to piracetam.

Detection Spectrophotometer at 205 nm.
Injection 20 µL of test solution (a) and reference solutions (a) and (b).

Flow rate 1.0 mL/min.

phosphate R₂ adjust to pH 6.0 with dilute phosphoric acid R.

90 volumes of a 1.0 g/L solution of dipotassium hydrogen

Mobile phase Mix 10 volumes of acetonitrile R₁ and

chromatography R (5 µm).

— stationary phase: end-capped octadecylsilyl silica gel for

— size: l = 0.25 m, Ø = 4.6 mm,

Column:

water R.

mixture of 10 volumes of acetonitrile R₁ and 90 volumes of

solvents. Dilute 10.0 mL of this solution to 50.0 mL with a

mixture of 10 volumes of acetonitrile R₁ and 90 volumes of

water R and dilute to 100.0 mL with the same mixture of

Reference solution (c) Dissolve 50.0 mg of piracetam CRS in a

90 volumes of water R.

50.0 mL with a mixture of 10 volumes of acetonitrile R₁ and

90 volumes of water R. Dilute 5.0 mL of this solution to

100.0 mL with a mixture of 10 volumes of acetonitrile R₁ and

Reference solution (b) Dilute 1.0 mL of test solution (a) to

dilute to 100.0 mL with the same mixture of solvents.

10 volumes of acetonitrile R₁ and 90 volumes of water R and

examined and 10 µL of 2-pyridone R in a mixture of

Reference solution (a) Dissolve 5 mg of the substance to be

STORAGE
Protected from light.

IMPURITIES
Specified impurities: A, B, C, D.

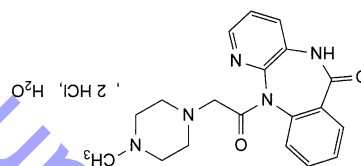


A. R = H: pyrrolidin-2-one (2-pyrrolidone),
B. R = CH₂-CO-O-CH₃: methyl (2-oxopyrrolidin-1-yl)acetate,
C. R = CH₂-CO-O-C₂H₅: ethyl (2-oxopyrrolidin-1-yl)acetate,
D. R = CH₂-CO₂H: (2-oxopyrrolidin-1-yl)acetic acid.

Ph Eur

Pirenzepine Hydrochloride

(Pirenzepine Dihydrochloride Monohydrate,
Ph Eur monograph 2001)



C₁₉H₂₃Cl₂N₅O₂·H₂O 442.3

Action and use
Muscarinic M₃ receptor antagonist.

Ph Eur

DEFINITION

1-[4-Methylpiperazin-1-yl]acetyl]-5,11-dihydro-6H-pyrido[2,3-b][1,4]benzodiazepin-6-one dihydrochloride monohydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or yellowish, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in methanol, very slightly soluble in ethanol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification B, D
Second identification A, C, D

A. Dissolve 30.0 mg in methanol R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with methanol R. Examined between 240 nm and 360 nm (2.2.25), the solution shows an absorption maximum at 283 nm. The specific absorbance at the maximum is 190 to 205 (anhydrous substance).
B. Infrared absorption spectrophotometry (2.2.24). Comparison pirenzepine dihydrochloride monohydrate CRS.

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution
Solution S is clear (2.2.1) and not more intensely coloured than reference solution GY₅ (2.2.2, Method II).

pH (2.2.3)
1.0 to 2.0 for solution S.

Impurity D
Thin-layer chromatography (2.2.27).

Test solution (a) To 0.10 g add 0.1 mL of concentrated ammonia R and dilute to 10 mL with methanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with methanol R.

Reference solution (a) To 0.1 g of pirenzepine dihydrochloride monohydrate CRS add 0.1 mL of concentrated ammonia R and dilute to 10 mL with methanol R.

Reference solution (b) Dissolve 25 mg of methylpiperazine R in methanol R and dilute to 25 mL with the same solvent.

Dilute 2.0 mL of the solution to 100 mL with methanol R.

Reference solution (c) Dilute 5 mL of test solution (a) to 100 mL with methanol R. Dilute 4 mL of this solution to 100 mL with methanol R. Mix 1 mL with 1 mL of reference solution (b).

Reference solution (d) Dilute 1 mL of reference solution (a) to 10 mL with methanol R.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, methanol R, ethyl acetate R, toluene R (7:25:28:40 V/V/V/V).

Application 20 µL as zones of 20 mm by 2 mm.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose the plate to iodine vapour until the zone in the chromatogram obtained with reference solution (b) is clearly visible (at most 60 min).

System suitability The test is not valid unless the chromatogram obtained with reference solution (c) shows 2 clearly separated zones.

Limit:

— **impurity D:** any zone corresponding to impurity D in the chromatogram obtained with test solution (a) is not more intense than the zone in the chromatogram obtained with reference solution (b) (0.2 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.30 g of the substance to be examined in water R and dilute to 10.0 mL with the same solvent. To 1.0 mL of the solution add 5 mL of methanol R and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 0.1 g of 1-phenylpiperazine R in methanol R and dilute to 10 mL with the same solvent. Mix 1 mL of the solution with 1 mL of the test solution, add 5 mL of methanol R and dilute to 10 mL with mobile phase A.

Column: size: $l = 0.125$ m, $\varnothing = 4.6$ mm, stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase: dissolve 2.0 g of sodium dodecyl sulfate R in water R, adjust to pH 3.2 with acetic acid R and dilute to 1000 mL with water R,

mobile phase B: methanol R, mobile phase C: acetonitrile R,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Mobile phase C (per cent V/V)
0 - 15	55 \rightarrow 25	30	15 \rightarrow 45
15 - 18	25 \rightarrow 20	30 \rightarrow 0	45 \rightarrow 80

Flow rate 1 mL/min.

Detection Spectrophotometer at 283 nm.

Injection 10 μ L.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to pirenzepine and 1-phenylpiperazine.

Limits:

— any impurity: not more than the peak area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),

— total: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent),

— disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.04 per cent).

Water (2.5.12)

3.5 per cent to 5.0 per cent, determined on 0.250 g. Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

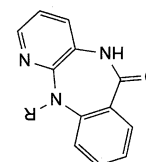
Dissolve 0.300 g in 50 mL of water R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume at the first point of inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 42.43 mg of $C_{19}H_{23}Cl_2N_5O_2$.

STORAGE

Protected from light.

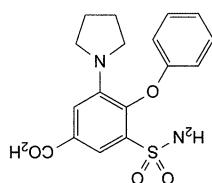
IMPURITIES



A. R = CO-CH₂-Cl; 1-(chloroacetyl)-5,11-dihydro-6H-pyrido[2,3-b][1,4]benzodiazepin-6-one, B. R = H; 5,11-dihydro-6H-pyrido[2,3-b][1,4]benzodiazepin-6-one,

Pirtetanide

(Ph. Eur. monograph 1556)



$C_{17}H_{18}N_2O_5S$

362.4

55837-27-9

Action and use
Thiazide diuretic.

DEFINITION

4-Phenoxy-3-(pyrrolidin-1-yl)-5-sulfamoylbenzoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellowish-white or yellowish powder.

Solubility

Very slightly soluble in water, sparingly soluble in anhydrous ethanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24):

Comparison pyrene CRs.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in acetone R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

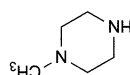
The solution is clear (2.2.1) and not more intensely coloured than reference solution GY₄ (2.2.2, Method II).

Dissolve 0.1 g in methanol R and dilute to 10 mL with the same solvent.

Related substances

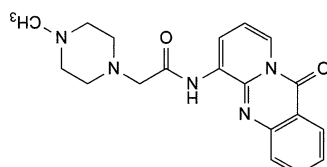
Liquid chromatography (2.2.29).

D. 1-methylpiperazine.



[b]quinazolin-1-one,

C. 6-[[[(4-methylpiperazin-1-yl)acetyl]amino]-1H-pyrido[2,1-



Ph Eur

Solvent mixture anhydrous ethanol R, acetonitrile R, water R (10:45:45 V/V/V).

Test solution Dissolve 20 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of pirtetamide for system suitability CRS (containing impurities A, B and C) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 0.3 mL of the test solution to 100.0 mL with the solvent mixture.

Column:
size: $l = 0.125$ m, $\varnothing = 4$ mm;
stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase A mixture of 35 volumes of acetonitrile R1 and 65 volumes of a solution prepared as follows: add 1 mL of trifluoroacetic acid R to 500 mL of water for chromatography R, add 1 mL of triethylamine R and dilute to 1000 mL with water for chromatography R.

Detection Spectrophotometer at 232 nm.
Injection 10 μ L.

Run time 5 times the retention time of pirtetamide.

Identification of impurities Use the chromatogram supplied with pirtetamide for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C.

Relative retention With reference to pirtetamide (retention time = about 10 min): impurity A = about 0.8; impurity B = about 3.1; impurity C = about 4.1.

System suitability: reference solution (a):
— resolution: minimum 2 between the peaks due to impurity A and pirtetamide.

Limits:

— impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— unspecified impurities: for each impurity, not more than 0.33 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 3.33 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— disregard limit: 0.17 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

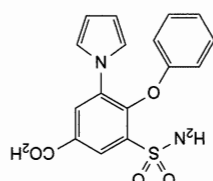
Dissolve 0.300 g in 25 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

IMPURITIES

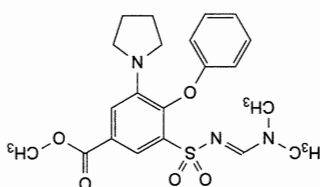
Protected from light.

STORAGE

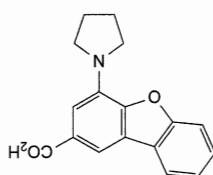
1 mL of 0.1 M perchloric acid is equivalent to 36.24 mg of $C_{17}H_{18}N_2O_5S$.



A. 4-phenoxy-3-(1H-pyrral-1-yl)-5-sulfamoylbenzoic acid,



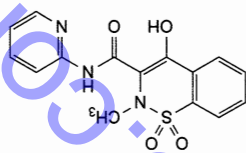
B. methyl-3-[[[(dimethylamino)methylidene]sulfamoyl]-4-phenoxy-5-(pyrralidin-1-yl)benzoate,



C. 4-(pyrralidin-1-yl)dibenzo[b,d]furan-2-carboxylic acid.

Piroxicam

(Ph. Eur. monograph 0944)



$C_{15}H_{13}N_3O_5S$

331.4

36322-90-4

Preparations

Piroxicam Capsules

Piroxicam Gel

DEFINITION

4-Hydroxy-2-methyl-N-(pyridin-2-yl)-2H-1,2-benzothiazine-3-carboxamide 1,1-dioxide.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or slightly yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in methylene chloride, slightly soluble in anhydrous ethanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison piroxicam CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of methylene chloride R₁ evaporate to dryness on a water-bath and record new spectra using the residues.

TESTS**Related substances**

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 75 mg of the substance to be examined in acetonitrile R₁, warming slightly if necessary, and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 7 mg of piroxicam for system suitability CRS (containing impurities A, B, D, G and J) in acetonitrile R₁ and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 10.0 mL with acetonitrile R₁. Dilute 1.0 mL of this solution to 50.0 mL with acetonitrile R₁.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 50 °C.

Mobile phase Mix 30 volumes of acetonitrile R₁ and

70 volumes of a 6.81 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 3.0 with phosphoric acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time 5 times the retention time of piroxicam.

Identification of impurities Use the chromatogram supplied with piroxicam for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, D, G and J.

Relative retention With reference to piroxicam (retention time = about 16 min): impurity A = about 0.1;

impurity D = about 0.6; impurity G = about 0.7;

impurity B = about 0.8; impurity J = about 1.8.

System suitability: reference solution (a):

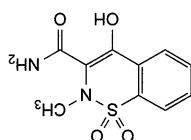
— resolution: minimum 1.5 between the peaks due to impurities G and B.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity A by 0.6;

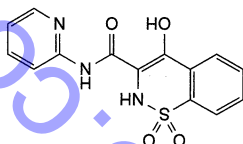
— **impurities A, B, D, G, J:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

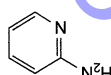


C. 4-hydroxy-2-methyl-1,2-benzothiazine-3-carboxamide 1,1-dioxide,

B. 4-hydroxy-N-(pyridin-2-yl)-2H-1,2-benzothiazine-3-carboxamide 1,1-dioxide,



A. pyridin-2-amine,



F, H, I, K, L.
Control of impurities in substances for pharmaceutical use: C, E, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these by the general monograph Substances for pharmaceutical use and/or acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of Other detectable impurities (the following substances would, if specified impurities A, B, D, G, J

IMPURITIES

Protected from light.

STORAGE

of C₁₅H₁₃N₃O₄S.
1 mL of 0.1 M perchloric acid is equivalent to 33.14 mg potentiometrically (2.2.20).
0.1 M perchloric acid, determining the end-point acetic anhydride R and anhydrous acetic acid R. Titrate with Dissolve 0.250 g in 60 mL of a mixture of equal volumes of

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in vacuo at 105 °C for 4 h.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying using 2 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test C. Prepare the reference solution Maximum 20 ppm.

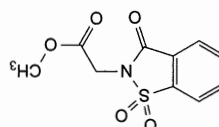
Heavy metals (2.4.8)

Maximum 20 ppm.

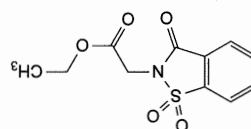
(0.05 per cent).

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

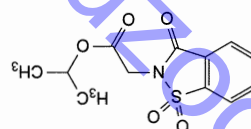
— **disregard limit:** 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).



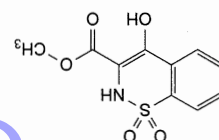
D. methyl (1,1-dioxido-3-oxo-1,2-benzisothiazol-2(3H)-yl)acetate,



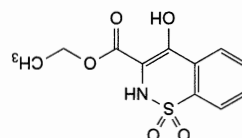
E. ethyl (1,1-dioxido-3-oxo-1,2-benzisothiazol-2(3H)-yl)acetate,



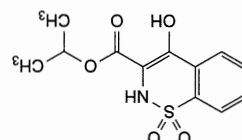
F. 1-methylethyl (1,1-dioxido-3-oxo-1,2-benzisothiazol-2(3H)-yl)acetate,



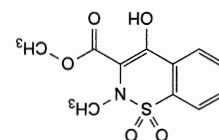
G. methyl 4-hydroxy-2H-1,2-benzothiazine-3-carboxylate, 1,1-dioxide,



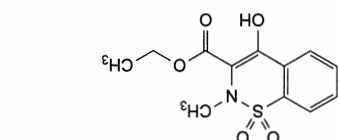
H. ethyl 4-hydroxy-2H-1,2-benzothiazine-3-carboxylate, 1,1-dioxide,



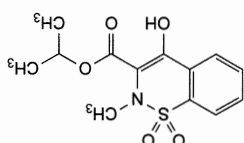
I. 1-methylethyl 4-hydroxy-2H-1,2-benzothiazine-3-carboxylate, 1,1-dioxide,



J. methyl 4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxylate, 1,1-dioxide,



K. ethyl 4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxylate, 1,1-dioxide,

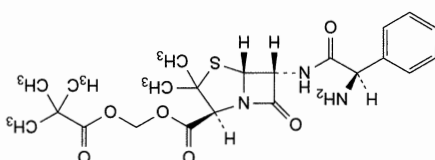


L. 1-methylethyl 4-hydroxy-2-methyl-2H-1,2-benzothiazine-3-carboxylate, 1,1-dioxide.



Pivampicillin

(Ph. Eur. monograph 0852)



$C_{22}H_{29}N_3O_8S$ 463.6 33817-20-8

Action and use

Antibacterial.

Ph. Eur.

DEFINITION

Methylene (2S,5R,6R)-6-[[[(2R)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate.

Semi-synthetic product derived from a fermentation product.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methanol, soluble in anhydrous ethanol. It dissolves in dilute acids.

IDENTIFICATION

First identification A

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison pivampicillin CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in 2 mL of methanol R.

Reference solution (a) Dissolve 10 mg of pivampicillin CRS in 2 mL of methanol R.

Reference solution (b) Dissolve 10 mg of bacampicillin hydrochloride CRS, 10 mg of pivampicillin CRS and 10 mg of talampicillin hydrochloride CRS in 2 mL of methanol R.

Plate TLC silanised silica gel plate R.

Mobile phase Mix 10 volumes of a 272 g/L solution of sodium acetate R adjusted to pH 5.0 with glacial acetic acid R, 40 volumes of water R and 50 volumes of ethanol (96 per cent) R.

Application 1 µL.

Development Over a path of 15 cm.

Drying In a current of warm air.

Detection Spray with ninhydrin solution R1 and heat at 60 °C for 10 min.

System suitability: reference solution (b):

- the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 150 mm long and 15 mm in diameter. Moisten with 0.05 mL of water R and add 2 mL of *sulfuric acid-formaldehyde reagent* R. Mix the contents of the tube by swirling; the solution is almost colourless. Place the test-tube in a water-bath for 1 min; a dark yellow colour develops.

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method J).

Dissolve 50 mg in 12 mL of 0.1 M hydrochloric acid.

Specific optical rotation (2.2.7)

Dissolve 0.100 g in 5.0 mL of ethanol (96 per cent) R and dilute to 10.0 mL with 0.1 M hydrochloric acid.

Triethanolamine

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.100 g of the substance to be examined in 1.0 mL of a mixture of 1 volume of water R and 9 volumes of acetonitrile R.

Reference solution Dissolve 5.0 mg of triethanolamine R in a mixture of 1 volume of water R and 9 volumes of acetonitrile R and dilute to 100 mL with the same mixture of solvents.

Plate TLC silica gel plate R.

Mobile phase methanol R, butanol R, phosphate buffer solution pH 5.8 R, glacial acetic acid R, butyl acetate R (5:15:24:40:80 V/V/V/V/V).

Application 10 µL.

Development Over a path of 12 cm.

Drying At 110 °C for 10 min and allow to cool.

Chlorine treatment Place at the bottom of a chromatographic tank an evaporating dish containing a mixture of 1 volume of hydrochloric acid R1, 1 volume of water R and 2 volumes of a 15 g/L solution of potassium permanganate R; close the tank and allow to stand for 15 min; place the dried plate in the tank and close the tank; leave the plate in contact with the chlorine vapour in the tank for 15–20 min; withdraw the plate and allow it to stand in air for 2–3 min.

Detection Spray with tetramethylammoniodiphenylmethane reagent R.

Limit:

— triethanolamine: any spot due to triethanolamine is not more intense than the corresponding spot in the chromatogram obtained with the reference solution (0.05 per cent).

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 50.0 mg of the substance to be examined in 10.0 mL of acetonitrile R and dilute to 20 mL with a 1 g/L solution of phosphoric acid R.

Reference solution Mix 2.0 mL of the test solution with 9.0 mL of acetonitrile R and 9.0 mL of a 1 g/L solution of phosphoric acid R.

Column:

- size: $l = 0.125$ m, $\varnothing = 4$ mm;
- stationary phase: end-capped octylsilyl silica gel for chromatography R.

Mobile phase:

- **mobile phase A:** mix 50 volumes of a 1.32 g/L solution of ammonium phosphate R, adjusted to pH 2.5 with a 100 g/L solution of phosphoric acid R, and 50 volumes of acetonitrile R;
- **mobile phase B:** mix 15 volumes of a 1.32 g/L solution of ammonium phosphate R, adjusted to pH 2.5 with a 100 g/L solution of phosphoric acid R, and 85 volumes of acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	100	0
10 - 12	0	100
12 - 17	100	0

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 50 µL.

Retention time Pivampicillin dimer = about 5 min.

System suitability: reference solution:

— **ratio of the mass distribution ratio:** minimum 12 for the peak due to pivampicillin dimer to that of the peak due to pivampicillin (principal peak).

Limits:

— **total:** not more than 0.3 times the area of the principal peak in the chromatogram obtained with the reference solution (3 per cent);

— **disregard limit:** 0.01 times the area of the principal peak in the chromatogram obtained with the reference solution (0.1 per cent).

N,N-Dimethylamine (2.4.26, Method B)

Maximum 20 ppm.

Test solution To 1.00 g of the substance to be examined in a ground-glass-stoppered tube add 10 mL of 0.5 M sulfuric acid. Heat the tube for 10 min in a water-bath, cool and add 15 mL of 1 M sodium hydroxide and 1.0 mL of the internal standard solution. Stopper the tube and shake vigorously for 1 min. Centrifuge if necessary and use the upper layer.

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.30 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29). Use the solutions within 2 h of preparation.

Test solution Dissolve 50.0 mg of the substance to be

examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to

50.0 mL with the mobile phase.

Reference solution (a) Dissolve 50.0 mg of pivampicillin CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 25.0 mg of propyl

parahydroxybenzoate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the

mobile phase. Mix 5.0 mL of the mobile phase and dilute to 50.0 mL with the mobile phase. Mix 5.0 mL of this solution with 5.0 mL of reference solution (a).

Column:

— size: $l = 0.125$ m, $\varnothing = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 40 volumes of acetonitrile R and 60 volumes of a 2.22 g/L solution of phosphoric acid R adjusted to pH 2.5 with triethylamine R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

System suitability:

— **resolution:** minimum 5.0 between the peaks due to pivampicillin (1st peak) and propyl parahydroxybenzoate (2nd peak) in the chromatogram obtained with reference solution (b);

— **symmetry factor:** maximum 2.0 for the peak due to pivampicillin in the chromatogram obtained with reference solution (b);

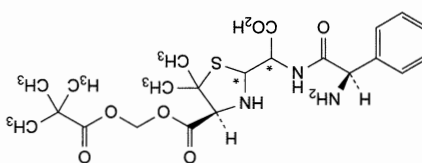
— **repeatability:** maximum relative standard deviation of 1.0 per cent after 6 injections of reference solution (a).

Calculate the percentage content of $C_{22}H_{29}N_3O_6S$ from the declared content of pivampicillin CRS.

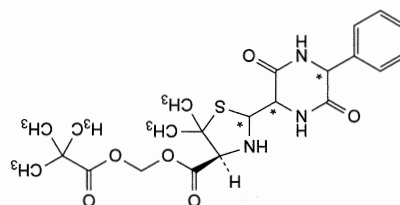
STORAGE

In an airtight container.

IMPURITIES



A. 2-[[[(2*R*)-2-amino-2-phenylacetyl]amino]-2-[(4*S*)-4-[[[(2*R*)-2-dimethylpropionoyl]oxy]methoxy]carbonyl]-5,5-dimethylthiazolidin-2-yl]acetic acid (penicilloic acids of pivampicillin),



B. methylene (4*S*)-5,5-dimethyl-2-(3,6-dioxo-5-phenylpiperazin-2-yl)thiazolidine-4-carboxylate 2,2-dimethylpropanoate (diketopiperazines of pivampicillin),

Action and use

Antibacterial.

DEFINITION

Methylene 2,2-dimethylpropanoate (2*S*,5*R*,6*R*)-6-[[[hexahydro-1*H*-azepin-1-yl]methylene]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate hydrochloride.

Semi-synthetic product derived from a fermentation product.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, in anhydrous ethanol and in methanol, slightly soluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison pivmecillinam hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution B₈ (2.2.2, Method I).

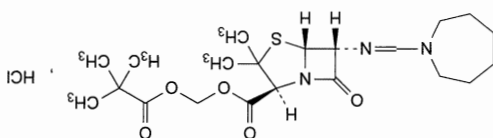
Dissolve 0.5 g in water R and dilute to 10 mL with the same solvent.

pH (2.2.3)

2.8 to 3.8.

Pivmecillinam Hydrochloride

(Ph. Eur. monograph 1359)



$C_{21}H_{34}ClN_3O_5S$

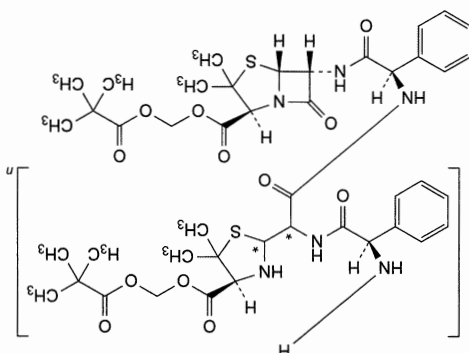
476.0

32887-03-9



Ph. Eur.

C. co-oligomers of pivampicillin and of penicilloic acids of pivampicillin.



Dissolve 1.0 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Solvent mixture To 45 volumes of acetonitrile R add

55 volumes of a 13.5 g/L solution of potassium dihydrogen

phosphate R previously adjusted to pH 3.0 with dilute

phosphoric acid R.

Test solution (a) Dissolve 20.0 mg of the substance to be

examined in the solvent mixture and dilute to 200.0 mL with

the solvent mixture.

Test solution (b) Dissolve 25.0 mg of the substance to be

examined in the solvent mixture and dilute to 25.0 mL with

the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of pivmecillinam

hydrochloride CRS in the solvent mixture and dilute to

200.0 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of reference solution (a)

to 50.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of pivmecillinam

hydrochloride CRS and 5 mg of pivmecillinam impurity C CRS

in the solvent mixture, and dilute to 50 mL with the solvent

mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Dissolve 0.55 g of tetraethylammonium hydrogen

sulfate R and 1.0 g of tetraethylammonium hydrogen sulfate R

in the solvent mixture and dilute to 1000 mL with the

solvent mixture.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L of test solution (b) and reference solutions (b)

and (c).

Run time 3 times the retention time of pivmecillinam.

System suitability: reference solution (c):

— resolution: minimum 3.5 between the peaks due to

pivmecillinam (1st peak) and impurity C (2nd peak).

Limits:

— any impurity: for each impurity, not more than 1.5 times

the area of the principal peak in the chromatogram

obtained with reference solution (b) (1.5 per cent);

total: not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(3 per cent);

— the chromatogram obtained with reference solution (b)

(0.1 per cent).

N,N-Dimethylamine (2.4.26, Method A)

Maximum 20 ppm.

Test solution Prepare as described in the general method but

heat at about 27 °C after the addition of strong sodium

hydroxide solution R, to dissolve the precipitate formed, then

add the trimethylpentane R.

Heavy metals (2.4.8)

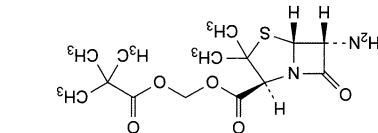
Maximum 20 ppm.

Dissolve 1.0 g in water R and dilute to 20 mL with the same

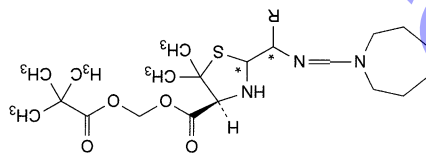
solvent. 12 mL of the solution complies with test A. Prepare

the reference solution using lead standard solution

(1 ppm Pb) R.

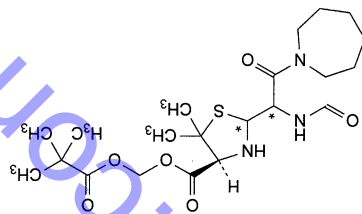


A. methyl 2,2,4,4-tetrahydro-1H-azepine-3-carboxylate
(2S,5R,6R)-6-amino-3,3-dimethyl-7-oxo-4-thia-
1-azabicyclo[3.2.0]heptane-2-carboxylate
2,2-dimethylpropionate (pivaloyloxymethyl
6-aminopenicillanate),



B. R = CO₂H; 2-[[[(hexahydro-1H-azepin-1-yl)methylene]amino]-2-[[[(4S)-4-[[[(2,2-dimethylpropoxy)methoxy]carbonyl]-5,5-dimethylthiazolidin-2-yl]acetic acid (penicilloic acids of pivmecillinam),

C. R = H; methyl 2,2-dimethylpropionate (2RS,4S)-2-[[[(hexahydro-1H-azepin-1-yl)methylene]amino]methyl]-5,5-dimethylthiazolidin-4-carboxylate,



D. methyl 2,2-dimethylpropionate (4S)-2-[[[(formylamino)-2-(hexahydro-1H-azepin-1-yl)-2-oxoethyl]-5,5-dimethylthiazolidin-4-carboxylate.

IMPURITIES

Protected from light, at a temperature of 2 °C to 8 °C.

STORAGE

Calculate the percentage content of C₂₁H₃₄N₃O₅S from the declared content of pivmecillinam hydrochloride CRS.

1.0 per cent after 6 injections.

— repeatability: maximum relative standard deviation of

System suitability: reference solution (a):

Injection Test solution (a) and reference solution (a).

related substances with the following modifications.

Liquid chromatography (2.2.29) as described in the test for

ASSAY

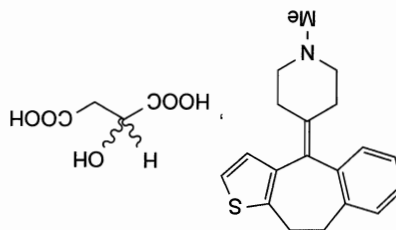
Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.00 g.

Water (2.5.12)

Pizotifen Malate



$C_{19}H_{21}NS, C_4H_6O_5$ 429.5 5189-11-7

Action and use

Serotonin (5HT₁) receptor partial agonist; prophylaxis of

migraine.

Preparation

Pizotifen Tablets

DEFINITION

Pizotifen Malate is 9,10-dihydro-4-(1-methylpiperidin-4-ylidene)-4H-benzo[4,5]cyclohepta[1,2-b]thiophene hydrogen malate. It contains not less than 98.5% and not more than 101.5% of $C_{19}H_{21}NS, C_4H_6O_5$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or slightly yellowish white, crystalline powder.

Very slightly soluble in water; slightly soluble in ethanol.

(96%); sparingly soluble in methanol.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is

concordant with the reference spectrum of pizotifen malate

(RS 277).

B. In the test for Related substances, the principal spot in the

chromatogram obtained with solution (2) corresponds to that

in the chromatogram obtained with solution (5).

C. Carry out the method for thin-layer chromatography,

Appendix III A, using silica gel G as the coating substance

and a mixture of 70 volumes of isopropyl ether, 25 volumes of

anhydrous formic acid and 5 volumes of water as the mobile

phase. Apply separately to the plate 5 µL of each of the

following solutions. For solution (1) dissolve 30 mg of the

substance being examined in 1 mL of ethanol (80%), heating

if necessary. Solution (2) contains 1% w/v of malic acid in

ethanol (80%). After removal of the plate, dry it at 100° for

30 minutes, cool, spray with 0.02M potassium permanganate

and dry in a current of warm air for about 1 minute.

The chromatogram obtained with solution (1) exhibits a spot

corresponding in position, colour and size to the spot in the

chromatogram obtained with solution (2).

TESTS

Clarity and colour of solution

A 1.0% w/v solution in a mixture of equal volumes of ethanol

(96%) and water is clear, Appendix IV A, and not more

intensely coloured than reference solution BY₆, Appendix IV B,

Method II.

Related substances

Carry out the method for thin-layer chromatography,

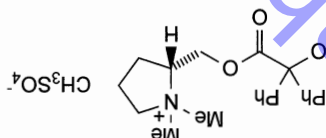
Appendix III A, using silica gel G as the coating substance

and a mixture of 100 volumes of toluene, 60 volumes of

butan-1-ol, 40 volumes of hexane and 3 volumes of

plate 5 µL of each of five solutions in a mixture of 9 volumes

Poldine Metilsulfate



and enantiomer

$C_{22}H_{29}NO_3S$ 451.5

545-80-2

Action and use

Anticholinergic.

Preparation

Poldine Tablets

DEFINITION

Poldine Metilsulfate is (RS)-2-benzoyloxy-1-methyl-1,1-

dimethylpyrrolidinium methyl sulfate. It contains not less

than 98.5% and not more than 100.5% of $C_{22}H_{29}NO_3S$,

calculated with reference to the dried substance.

CHARACTERISTICS

A white, crystalline powder.

Freely soluble in water; soluble in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is

concordant with the reference spectrum of poldine metilsulfate

(RS 278).

B. The light absorption, Appendix II B, in the range 250 to

350 nm of a 0.1% w/v solution exhibits maxima at 252 nm

and 258 nm and inflections at 262, 264 and 268 nm. The

absorbance at the maximum at 252 nm is about 0.85 and at

the maximum at 258 nm is about 0.99.

C. Dissolve 2 mg in 10 mL of water, add 20 mL of ammonium cobaltotrioximate solution and 5 mL of dichloromethane and shake well. The dichloromethane layer becomes blue.

TESTS

Acidity or alkalinity

pH of a 1% w/v solution, 5.0 to 7.0, Appendix V L.

Melting point

137° to 142°, Appendix V A.

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in a mixture of 5 volumes of methanol and 1 volume of formic acid.

(1) 1.0% w/v of the substance being examined.

(2) 0.010% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel (Merck silica gel 60 plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 20 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, spray with sulfuric acid (50%) and heat at 110° for 10 minutes.

MOBILE PHASE

5 volumes of formic acid, 5 volumes of water, 30 volumes of methanol and 60 volumes of dichloromethane.

LIMITS

Any pink secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (1.0%).

Loss on drying

When dried to constant weight at 80°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.5 g in 10 mL of water in a stoppered flask, add 0.05 mL of cresol red solution and, if necessary, sufficient 0.05M sodium hydroxide to produce a pale pink colour. Add 20 mL of 0.1M sodium hydroxide VS, stopper the flask and allow to stand for 10 minutes at 25°. Titrate with 0.1M hydrochloric acid VS using cresol red solution as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of alkali required. Each mL of 0.1M sodium hydroxide VS is equivalent to 45.15 mg of C₂₂H₂₉NO₇S.

Poloxamers

(Ph. Eur. monograph 1464)

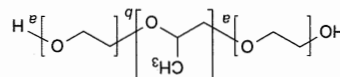
Action and use

Non-ionic surfactant.

Ph. Eur.

DEFINITION

Synthetic block copolymer of ethylene oxide and propylene oxide, represented by the following general formula:



CHARACTERS

A suitable antioxidant may be added.

Appearance

— poloxamer 124: colourless or almost colourless liquid; poloxamers 188, 237, 338, 407: white or almost white, waxy powder, microbeads or flakes.

Solubility

— poloxamers 124, 237, 338, 407: very soluble in water and in ethanol (96 per cent), practically insoluble in light petroleum (bp: 50-70 °C); poloxamer 188: soluble in water and in ethanol (96 per cent).

mp

About 50 °C for poloxamers 188, 237, 338 and 407.

IDENTIFICATION

First identification: A, B.

Second identification: B, C.

A. Infrared absorption spectrophotometry (2.2.24). Comparison Chemical reference substance of the Ph. Eur. corresponding to the type of poloxamer to be examined.

B. Average relative molecular mass (see Tests).

C. Oxypropylene:oxyethylene ratio (see Tests).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

pH (2.2.3)

5.0 to 7.5 for solution S.

Ethylene oxide, propylene oxide and dioxan

Head-space gas chromatography (2.2.28). Ethylene oxide stock solution Introduce 0.5 mL of ethylene oxide stock solution R2 into a vial and dilute to 50.0 mL with dimethyl sulfoxide R1. Mix carefully. Ethylene oxide solution Dilute 1.0 mL of the ethylene oxide stock solution to 250 mL with dimethyl sulfoxide R1.

Propylene oxide stock solution Introduce about 7 mL of methylene chloride R into a volumetric flask, add 0.500 g (m) of propylene oxide R and dilute to 10.0 mL with methylene chloride R. Dilute 0.5 mL of this solution to 50.0 mL with dimethyl sulfoxide R1. Mix carefully. Calculate the exact concentration of propylene oxide, in milligrams per millilitre, using the following expression:

$$\frac{m \times 1000 \times 0.5}{10 \times 50}$$

— propylene oxide: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (5 ppm);

— dioxan: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (10 ppm).

Average relative molecular mass

Weigh 15 g (*m*) of the substance to be examined into a 250 mL ground-glass-stoppered flask, add 25.0 mL of *phthalic anhydride solution R* and a few glass beads and swirl to dissolve. Boil gently under a reflux condenser for 1 h, allow to cool and add 2 quantities, each of 10 mL, of *pyridine R*, through the condenser. Add 10 mL of *water R*, mix and allow to stand for 10 min. Add 40.0 mL of 0.5 *M sodium hydroxide* and 0.5 mL of a 10 g/L solution of *sodium phenolphthalein R* in *pyridine R*. Titrate with 0.5 *M sodium hydroxide* to a light pink endpoint that persists for 15 s and record the volume of sodium hydroxide used (*S*). Prepare a blank. Record the volume of sodium hydroxide used (*B*). Calculate the average relative molecular mass using the following expression:

$$\frac{4000m}{B - S}$$

Oxypropylene:oxyethylene ratio

Nuclear magnetic resonance spectrometry (2.2.33). Use a 100 g/L solution of the substance to be examined in *deuterated chloroform R*. Record the average area of the doublet appearing at about 1.08 ppm due to the methyl groups of the oxypropylene units (*A*₁) and the average area of the composite band from 3.2 ppm to 3.8 ppm due to CH2O groups of both the oxyethylene and oxypropylene units and the CHO groups of the internal oxypropylene units (*A*₂) with reference to the internal standard.

Calculate the percentage of oxyethylene, by weight, in the sample being examined using the following expression:

$$\text{where } \alpha = \frac{A_2}{A_1} - 1$$

$$\frac{3300\alpha}{33\alpha + 58}$$

Water (2.5.12)
Maximum 1.0 per cent, determined on 1.000 g.

Total ash (2.4.16)
Maximum 0.4 per cent, determined on 1.0 g.

STORAGE

In an airtight container.

LABELLING

The label states the type of poloxamer.

Propylene oxide solution Dilute 1.0 mL of the propylene oxide stock solution to 50.0 mL with *dimethyl sulfoxide R1*. Calculate the exact concentration of propylene oxide, in micrograms per millilitre, using the following expression:

$$\frac{C \times 1000 \times 1}{50}$$

C = concentration of the propylene oxide stock solution, in milligrams per millilitre.

Dioxan solution Introduce 0.100 g (*m*) of *dioxan R* into a flask and dilute to 50.0 mL with *dimethyl sulfoxide R1*. Dilute 2.50 mL of this solution to 100.0 mL with *dimethyl sulfoxide R1*.

Calculate the exact concentration of dioxan, in micrograms per millilitre, using the following expression:

$$\frac{m \times 2.50 \times 1000 \times 1000}{50 \times 100}$$

Mixture solution Dilute a mixture of 6.0 mL of the ethylene oxide solution, 6.0 mL of the propylene oxide solution and 2.5 mL of the dioxan solution to 25.0 mL with *dimethyl sulfoxide R1*.

Test solution To 1.000 g of the substance to be examined in a head-space vial, add 4.0 mL of *dimethyl sulfoxide R1* and close the vial immediately.

Reference solution To 1.000 g of the substance to be examined in a head-space vial, add 2.0 mL of *dimethyl sulfoxide R1* and 2.0 mL of the mixture solution. Close the vial immediately.

Column:

— material: fused silica;
— size: $l = 50$ m, $\varnothing = 0.32$ mm;
— stationary phase: poly(*dimethyl*) (diphenyl) siloxane *R* (film thickness 5 µm).

Carrier gas helium for chromatography *R*.

Flow rate 1.4 mL/min.

Static head-space conditions:

— equilibrium temperature: 110 °C;

— equilibration time: 30 min;

— transfer-time temperature: 140 °C;

— pressurisation time: 1 min;

— injection time: 0.05 min.

Temperature:

Time (min)	Temperature (°C)
0 - 10	70
10 - 27	70 → 240
Injection port	250
Detector	250

Detection Flame ionisation.

Injection Inject a suitable volume of the gaseous phase, for example 1 mL.

Relative retention With reference to ethylene oxide (retention time = about 6 min): propylene oxide = about 1.3;

methylene chloride = about 1.6; dioxan = about 3.0;

dimethyl sulfoxide = about 3.7.

Limits:

— ethylene oxide: not more than 0.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (1 ppm);

Polyacrylate Dispersion (30 per cent)

(Ph Eur monograph 0733)

Action and use

Excipient.

DEFINITION

Dispersion in water of a copolymer of ethyl acrylate and methyl methacrylate having a mean relative molecular mass of about 800 000.

Content

28.5 per cent to 31.5 per cent (residue on evaporation). It may contain a suitable emulsifier.

CHARACTERS

Appearance

Opaque, white or almost white, slightly viscous liquid.

Solubility

Miscible with water, soluble in acetone, in anhydrous ethanol and in 2-propanol.

IDENTIFICATION

First identification A

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of polyacrylate.

B. To 1 g add 5 mL of water R and mix; the mixture remains opaque. Take 3 portions of 1 g and mix separately with 5 g of anhydrous ethanol R, 5 g of acetone R and 5 g of 2-propanol R. Transparent solutions are obtained.

C. To 1 g add 10 mL of 0.1 M sodium hydroxide.

The mixture remains opaque.

D. Appearance of a film (see Tests).

E. Dry 4 g in a Petri dish at 60 °C in an oven for 4 h and transfer the resulting clear film to a small test-tube

(100 mm × 12 mm). Heat over a flame and collect the fumes that evolve in a 2nd test-tube held over the mouth of the 1st tube. The condensate gives the reaction of esters

(2.3.1).

TESTS

Relative density (2.2.5)

1.037 to 1.047.

Viscosity (2.2.10)

Maximum 50 mPa.s, determined using a rotating viscometer at 20 °C and a shear rate of 10 s⁻¹.

Appearance of a film

Pour 1 mL on a glass plate and allow to dry. A clear elastic film is formed.

Particulate matter

Filter 100.0 g through a tared stainless steel sieve (90). Rinse to constant mass. The residue weighs not more than 0.500 g.

Residual monomers

Liquid chromatography (2.2.29).

Test solution Dissolve 1.00 g of the substance to be examined in tetrahydrofuran R and dilute to 50.0 mL with the same solvent. To 5.0 mL of a 35 g/L solution of sodium perchlorate R add 10.0 mL of the solution dropwise whilst stirring continuously. Centrifuge and filter the clear supernatant. Dilute 5.0 mL of this solution to 10.0 mL with water R.



Reference solution Dissolve 10 mg of ethyl acrylate R and 10 mg of methyl methacrylate R in tetrahydrofuran R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with tetrahydrofuran R. To 10.0 mL of the solution add 5.0 mL of a 35 g/L solution of sodium perchlorate R and mix. Dilute 5.0 mL of the mixture to 10.0 mL with water R.

Column: — size: $l = 0.12$ m, $\phi = 4.6$ mm; — stationary phase: octadecylsilyl silica gel for chromatography R (5–10 μ m).

Mobile phase acetonitrile R1, water for chromatography R (15:85 V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 205 nm.

Injection About 50 μ L.

Limit: — residual monomers: maximum 100 ppm.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Sulfated ash (2.4.14)

Maximum 0.4 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

ASSAY

Dry 1.000 g at 110 °C for 3 h and weigh the residue.

STORAGE

At a temperature of 5 °C to 25 °C, protected from freezing.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the mandatory-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for polyacrylate dispersion 30 per cent used as film former or matrix former in prolonged-release dosage forms.

Viscosity (see Tests).

Appearance of a film (see Tests).

Solubility of a film

Take a piece of the film obtained in the test for appearance of a film and place it in a flask containing a 10.3 g/L solution of hydrochloric acid R with stirring. It does not dissolve within 2 h. Take another piece of the film and place it in a flask containing 0.33 M phosphate buffer solution pH 7.5 R with stirring. It also does not dissolve within 2 h.

Polymyxin B Sulphate

Reference solution (a) Dissolve 20 mg of serine R in water R and dilute to 10 mL with the same solvent.

Carry out the following procedures protected from light

Application 5 μ l as bands of 10 mm, then place the plate in the chromatographic tank so that it is not in contact with the mobile phase, and allow it to become impregnated with the vapour of the mobile phase for at least 12 h.

pnase:
Drying At 100-105 °C.
Detection Spray with ninhydrin solution RI and heat at 110 °C for 5 min.

the test solution also shows a zone with a very low R_F value

b. Examine the chromatograms obtained in the assay. *Results* The peaks due to polymyxins B1, B2, B3 and B1-I in the chromatogram obtained with the test solution are similar in retention time to the corresponding peaks in the chromatogram obtained with reference solution (a).

a 100 g/L solution of *sodium hyaluronate K*. Shake and add dropwise 0.25 mL of a 10 g/L solution of *copper sulfate R₅*, shaking after each addition. A reddish-violet colour develops. D. It gives reaction (a) of sulfates (2.3.1).

Dissolve 0.2 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Dissolve 0.50 g in water *R* and dilute to 25.0 mL with the same solvent.

80 volumes of water *R* and dilute to 100.0 mL with the same mixture of solvents.

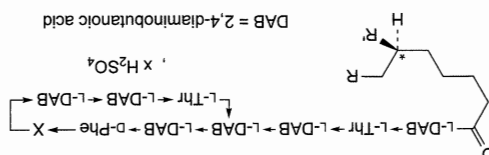
mixture of solvents.
Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with a mixture of 20 volumes of acetonitrile R and 80 volumes of water R.

- stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
- temperature: 30 °C.

Action and use
Antibacterial.

Preparations
Polymyxin and Bacitracin Ointment
Polymyxin and Bacitracin Eye Ointment

Polyimide	R	R'	X	Molecular formula	M _n
B1	CH ₃	CH ₃	L-Leu	C ₅₆ H ₉₈ N ₁₆ O ₁₃	1204
B2	H	CH ₃	L-Leu	C ₅₅ H ₉₆ N ₁₆ O ₁₃	1190
B3	CH ₃	H	L-Leu	C ₅₅ H ₉₆ N ₁₆ O ₁₃	1190
B1-I	CH ₃	CH ₃	L-Ile	C ₅₈ H ₉₈ N ₁₆ O ₁₃	1204



(Ph. Eur. monograph 0203)

sodium sulfate R in 900 mL of water R, adjust to pH 2.3 with dilute phosphoric acid R and dilute to 1000 mL with water R. Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 1.4 times the retention time of polymyxin B₁.

Relative retention With reference to polymyxin B₁ retention time = about 35 min; polymyxin B₂ = about 0.5; polymyxin B₃ = about 0.6; polymyxin B₁-I = about 0.8.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to polymyxin B₂ and polymyxin B₃.

Limits:

— any impurity: for each impurity, maximum 3.0 per cent;

— total: maximum 17.0 per cent;

— disregard limit: 0.7 times the area of the principal peak in the chromatogram obtained with reference solution (b).

Sulfate

15.5 per cent to 17.5 per cent (dried substance).

Dissolve 0.250 g in 100 mL of water R and adjust the solution to pH 11 with concentrated ammonia R.

Add 10.0 mL of 0.1 M barium chloride and about 0.5 mg of phthalate purple R. Titrate with 0.1 M sodium edetate, adding 50 mL of ethanol (96 per cent) R when the colour of the solution begins to change and continuing the titration until the violet-blue colour disappears.

1 mL of 0.1 M barium chloride is equivalent to 9.606 mg of SO₄.

Loss on drying (2.2.32)

Maximum 6.0 per cent, determined on 1.000 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 670 Pa for 3 h.

Sulfated ash (2.4.14)

Maximum 0.75 per cent, determined on 1.0 g.

Pyrogens (2.6.8)

If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of pyrogens, it complies with the test for pyrogens. Inject, per kilogram of the rabbit's mass, 1 mL of a solution in water for injections R containing 1.5 mg of the substance to be examined per millilitre.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of polymyxin B₃ of polymyxin B₁-I, and of the sum of polymyxins B₁, B₂, B₃ and B₁-I, using the following expression:

$$C_{B_i} = \frac{A_{B_i} \times m_2 \times D_{B_i}}{m_1 \times B_{B_i}}$$

where:

C_{B_i} = percentage content of polymyxin B_i in the chromatogram obtained with the test solution;

A_{B_i} = area of the peak due to polymyxin B_i in the chromatogram obtained with reference solution (a);

B_{B_i} = area of the peak due to polymyxin B_i in the chromatogram obtained with reference solution (a);

m₂ = mass of polymyxin B sulfate CRS in reference solution (a), in milligrams;

m₁ = mass of polymyxin B sulfate CRS in reference solution (a), in milligrams.

D_{B_i} = declared percentage content for polymyxin B_i in polymyxin B sulfate CRS.

STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

Ph Eur

DEFINITION

Mixture of ethers of polyoxypropylene with linear alcohols, mainly stearyl alcohol, obtained by the reaction of stearyl alcohol with propylene oxide. It may contain some free polyoxypropylene and various amounts of free stearyl alcohol. The number of moles of propylene oxide reacted per mole of stearyl alcohol is 11 (nominal value). A suitable antioxidant may be added.

CHARACTERS

Appearance

Colourless or pale yellow, clear or slightly turbid liquid.

Solubility

Practically insoluble in water, soluble in ethanol (96 per cent), in mineral oils and in 2-propanol, practically insoluble in propylene glycol and in glycerol.

Relative density

About 0.94 at 25 °C.

Refractive index About 1.448 at 25 °C.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison polyoxypropylene stearyl ether CRS.

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Viscosity (2.2.9): 89.0 mPa·s to 95.0 mPa·s.

TESTS

Acid value (2.5.1)

Maximum 2.0.

Hydroxyl value (2.5.3, Method A)

60 to 77.

Iodine value (2.5.4, Method A)

Maximum 3.0.

Peroxide value (2.5.5, Method A)

Maximum 5.0.

Saponification value (2.5.6)

Maximum 3.0.

Propylene oxide

Head-space gas chromatography (2.2.28).

Propylene oxide stock solution Weigh 45 g of cold macrogol 200 R1 and add 1.000 g (m) of propylene oxide R.

Mix carefully by swirling to ensure a homogeneous solution.

Add more macrogol 200 R1 until the total weight is 50.0 g

and mix again. [NOTE: the solution is stable for 1 month if stored at -20 °C]. Allow to reach room temperature. Dilute

0.50 g of this solution to 100.0 mL with water R.

Propylene oxide standard solution Dilute 10.0 mL of propylene oxide stock solution to 100.0 mL with water R.

Polyoxypropylene Stearyl Ether

(Ph. Eur. monograph 2602)

Ph Eur



Propionaldehyde stock solution Weigh 0.1 g of propionaldehyde R into a volumetric flask and dilute to 100.0 mL with water R.

Test solution (a) Weigh 1.00 g of the substance to be examined into a 20 mL head-space vial. Add 0.5 mL of cold water R and seal the vial immediately with a polytetrafluoroethylene-coated silicon membrane and an aluminium cap. Mix carefully.

Test solution (b) Weigh 1.00 g of the substance to be examined into a 20 mL head-space vial. Add 0.5 mL of cold propylene oxide standard solution and seal the vial immediately with a polytetrafluoroethylene-coated silicon membrane and an aluminium cap. Mix carefully.

Reference solution Introduce 50.0 mL of cold propylene oxide standard solution into a volumetric flask, add 0.5 mL of cold propionaldehyde stock solution and dilute to 100.0 mL with water R. Introduce 0.5 mL of this solution into a 20 mL head-space vial.

Column:

- material: fused silica;
- size: $l = 60$ m, $\varnothing = 0.32$ mm;
- stationary phase: poly(dimethyl)siloxane R (film thickness 5 μ m).

Carrier gas helium for chromatography R.

Flow rate 2.6 mL/min.

Split ratio 10:1.

Static head-space conditions that may be used:

- equilibration temperature: 90 °C;
- equilibration time: 45 min.

Temperature:

Time (min)	Temperature (°C)
0 - 5	50
5 - 31	50 \rightarrow 180
31 - 32.7	180 \rightarrow 230
32.7 - 37.7	230
Injection port	150
Detector	250

Detection Flame ionisation.

Injection A suitable volume, for example 1.0 mL, of the gaseous phase of test solutions (a) and (b) and of the reference solution.

Identification of peaks Use the chromatogram obtained with the reference solution to identify the peaks due to propylene oxide and propionaldehyde.

Relative retention With reference to propylene oxide (retention time = about 10.4 min): propionaldehyde = about 0.96.

System suitability: reference solution:

- resolution: minimum 1.5 between the peaks due to propionaldehyde and propylene oxide;
- signal-to-noise ratio: minimum 10 for the peak due to propylene oxide.

Calculate the content of propylene oxide in parts per million using the following expression:

$$A_1 \times m \times 5 \frac{(A_2 \times M_1) - (A_1 \times M_2)}{A_1 \times m \times 5}$$

A_1 = area of the peak due to propylene oxide in the chromatogram obtained with test solution (a);

Polysorbate 20

Polyoxyethylene 20 Sorbitan Monolaurate
(Ph. Eur. monograph 0426)

Action and use

Non-ionic surfactants.

Ph. Eur.

DEFINITION

Mixture of partial esters of fatty acids, mainly lauric (dodecanoic) acid, with sorbitol and its anhydrides ethoxylated with approximately 20 moles of ethylene oxide for each mole of sorbitol and sorbitol anhydrides.

CHARACTERS

Appearance

Oily, yellow or brownish-yellow, clear or slightly opalescent liquid.

Solubility

Soluble in water, in anhydrous ethanol, in ethyl acetate and in methanol, practically insoluble in fatty oils and in liquid paraffin.

Relative density

About 1.10.

Viscosity

About 400 mPa.s at 25 °C.

IDENTIFICATION

First identification A, D

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison: Ph. Eur. reference spectrum of polysorbate 20.

STORAGE

In an airtight container.

LABELLING

The label states the number of moles of propylene oxide reacted per mole of stearyl alcohol (nominal value).

Ph. Eur.



B. Hydroxyl value (see Tests).
C. Saponification value (see Tests).
D. Composition of fatty acids (see Tests).
E. Dissolve 0.1 g in 5 mL of methylene chloride R. Add 0.1 g of potassium thiocyanate R and 0.1 g of cobalt nitrate R. Stir with a glass rod. The solution becomes blue.

TESTS

Acid value (2.5.1)

Maximum 2.0.

Hydroxyl value (2.5.3, Method A)

96 to 108.

Peroxide value

Maximum 10.0.

Introduce 10.0 g into a 100 mL beaker and dissolve with

20 mL of glacial acetic acid R. Add 1 mL of saturated

potassium iodide solution R, mix and allow to stand for 1 min.

Add 50 mL of carbon dioxide-free water R and a magnetic

stirring bar. Titrate with 0.01 M sodium thiosulfate,

determining the end-point potentiometrically (2.2.20). Carry

out a blank titration. If the result of the blank determination

exceeds 0.1 mL of titration reagent, replace the reagents and

repeat the determination.

Determine the peroxide value using the following expression:

$$\frac{(n_1 - n_2) \times M \times 1000}{m}$$

n_1 = volume of 0.01 M sodium thiosulfate required for the

substance to be examined, in millilitres;

n_2 = volume of 0.01 M sodium thiosulfate required for the

blank titration, in millilitres;

M = molarity of the sodium thiosulfate solution, in moles

per litre;

m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)

40 to 50, determined on 4.0 g.

Use 15.0 mL of 0.5 M alcoholic potassium hydroxide and dilute

with 50 mL of ethanol (96 per cent) R before carrying out the

titration. Heat under reflux for 60 min.

Composition of fatty acids (2.4.22, Method C)

Prepare reference solution (a) as indicated in

Table 2.4.22-2.

Column:

— material: fused silica;

— size: $l = 30$ m, $\phi = 0.32$ mm;

— stationary phase: macrogol 20 000 R (film

thickness 0.5 μ m).

Carrier gas helium for chromatography R.

Linear velocity 50 cm/s.

Temperature

Time (min)	Temperature (°C)
0 - 14	80 \rightarrow 220
14 - 54	220
Injection port	250
Detector	250

Detection: Flame ionisation.

Injection 1 μ L.

Polysorbate 40

Polyoxyethylene 20 Sorbitan Monopalmitate

(Ph. Eur. monograph 1914)

Action and use

Non-ionic surfactants.

DEFINITION

Mixture of partial esters of fatty acids, mainly *Palmitic*

acid (1904), with sorbitol and its anhydrides ethoxylated with

approximately 20 moles of ethylene oxide for each mole of

sorbitol and sorbitol anhydrides.

CHARACTERS

Appearance

Oily, viscous, yellowish or brownish-yellow liquid.

Solubility

Miscible with water, with anhydrous ethanol, with ethyl

acetate and with methanol, practically insoluble in fatty oils

and in liquid paraffin.

Relative density

About 1.10.

Viscosity

About 400 mPa.s at 30 °C.

IDENTIFICATION

First identification A, D

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of polysorbate 40.

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Composition of fatty acids (see Tests).



Ph Eur

STORAGE

In an airtight container, protected from light.

Total ash (2.4.16)

Maximum 0.25 per cent, determined on 2.0 g.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.00 g.

using 2 mL of lead standard solution (10 ppm Pb) R.

2.0 g complies with test C. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

Maximum 1 ppm of ethylene oxide and 10 ppm of dioxan.

Ethylene oxide and dioxan (2.4.25, Method A)

— linoleic acid: maximum 3.0 per cent.

— oleic acid: maximum 11.0 per cent;

— stearic acid: maximum 7.0 per cent;

— palmitic acid: 7.0 per cent to 15.0 per cent;

— myristic acid: 14.0 per cent to 25.0 per cent;

— lauric acid: 40.0 per cent to 60.0 per cent;

— capric acid: maximum 10.0 per cent;

— caprylic acid: maximum 10.0 per cent;

— caproic acid: maximum 1.0 per cent;

Composition of the fatty-acid fraction of the substance:

Ethylene oxide and dioxan (2.4.25, Method A)
Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.
2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.00 g.

Total ash (2.4.16)

Maximum 0.25 per cent, determined on 2.0 g.

STORAGE

In an airtight container, protected from light.

Ph Eur



Polysorbate 60

Polyoxyethylene 20 Sorbitan Monostearate
(Ph. Eur. monograph 0427)

Action and use
Non-ionic surfactants

DEFINITION

Mixture of partial esters of fatty acids, mainly *Stearic acid* 50 (1474), with sorbitol and its anhydrides ethoxylated with approximately 20 moles of ethylene oxide for each mole of sorbitol and sorbitol anhydrides.

CHARACTERS

Appearance
Yellowish-brown gelatinous mass which becomes a clear liquid at temperatures above 25 °C.

Solubility

Soluble in water, in anhydrous ethanol, in ethyl acetate and in methanol, practically insoluble in fatty oils and in liquid paraffin.

Relative density

About 1.10.

Viscosity

About 400 mPa.s at 30 °C.

IDENTIFICATION

First identification A, D

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison: Ph. Eur. reference spectrum of polysorbate 60.

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Composition of fatty acids (see Tests).

E. Dissolve 0.1 g in 5 mL of methylene chloride R. Add 0.1 g of potassium thiocyanate R and 0.1 g of cobalt nitrate R. Stir with a glass rod. The solution becomes blue.

TESTS

Acid value (2.5.1)

Maximum 2.0.

Dissolve 5.0 g in 50 mL of the prescribed solvent mixture.

Hydroxyl value (2.5.3, Method A)

81 to 96.

E. Dissolve 0.1 g in 5 mL of methylene chloride R. Add 0.1 g of potassium thiocyanate R and 0.1 g of cobalt nitrate R. Stir with a glass rod. The solution becomes blue.

TESTS

Acid value (2.5.1)

Maximum 2.0.

Dissolve 5.0 g in 50 mL of the prescribed solvent mixture.

Hydroxyl value (2.5.3, Method A)

89 to 105.

Peroxide value

Maximum 10.0.

Introduce 10.0 g into a 100 mL beaker and dissolve with 20 mL of glacial acetic acid R. Add 1 mL of saturated potassium iodide solution R, mix and allow to stand for 1 min.

Add 50 mL of carbon dioxide-free water R and a magnetic stirring bar. Titrate with 0.01 M sodium thiosulfate, determining the end-point potentiometrically (2.2.20). Carry out a blank titration. If the result of the blank determination exceeds 0.1 mL of titration reagent, replace the reagents and repeat the determination.

Determine the peroxide value using the following expression:

$$\frac{(n_1 - n_2) \times M \times 1000}{m}$$

n_1 = volume of 0.01 M sodium thiosulfate required for the substance to be examined, in millilitres;
 n_2 = volume of 0.01 M sodium thiosulfate required for the blank titration, in millilitres;
 M = molarity of the sodium thiosulfate solution, in moles per litre;
 m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)

41 to 52, determined on 4.0 g.
Use 15.0 mL of 0.5 M alcoholic potassium hydroxide and dilute with 50 mL of ethanol (96 per cent) R before carrying out the titration. Heat under reflux for 60 min.

Composition of fatty acids (2.4.22, Method C)

Prepare reference solution (a) as indicated in

Table 2.4.22.-1.

Column:

— material: fused silica;

— size: $l = 30$ m, $\phi = 0.32$ mm;

— stationary phase: macrogol 20 000 R (film

thickness 0.5 μ m).

Carrier gas helium for chromatography R.

Linear velocity 50 cm/s.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 14	80 \rightarrow 220	14 - 54	250	250

Detection Flame ionisation.

Injection 1 μ L.

Composition of the fatty-acid fraction of the substance: — palmitic acid: minimum 92.0 per cent.

Peroxide value

Maximum 10.0.

Introduce 10.0 g into a 100 mL beaker and dissolve with

20 mL of *glacial acetic acid* R. Add 1 mL of *saturated*

potassium iodide solution R, mix and allow to stand for 1 min.

Add 50 mL of *carbon dioxide-free water* R and a magnetic

stirring bar. Titrate with 0.01 M *sodium thiosulfate*,

determining the end-point potentiometrically (2.2.20). Carry

out a blank titration. If the result of the blank determination

exceeds 0.1 mL of titration reagent, replace the reagents and

repeat the determination.

Determine the peroxide value using the following expression:

$$\frac{m}{(n_1 - n_2) \times M \times 1000}$$

n_1 = volume of 0.01 M *sodium thiosulfate* required for the

substance to be examined, in millilitres;

n_2 = volume of 0.01 M *sodium thiosulfate* required for the

blank titration, in millilitres;

M = molarity of the sodium thiosulfate solution, in moles

per litre;

m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)

45 to 55, determined on 4.0 g.

Use 15.0 mL of 0.5 M *alcoholic potassium hydroxide* and dilute

with 50 mL of *ethanol* (96 per cent) R before carrying out the

titration. Heat under reflux for 60 min.

Composition of fatty acids (2.4.22, Method C)

Prepare reference solution (a) as indicated in

Table 2.4.22.-1.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— stationary phase: macrogol 20 000 R (film thickness

0.5 μ m).

Carrier gas *helium* for chromatography R.

Linear velocity 50 cm/s.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
14 - 54	220	0 - 14	250	250
80 \rightarrow 220				

Detection: Flame ionisation.

Injection 1 μ L.

Composition of the fatty-acid fraction of the substance:

— stearic acid: 40.0 per cent to 60.0 per cent;

— sum of the contents of palmitic and stearic acids: minimum

90.0 per cent.

Ethylene oxide and dioxan (2.4.25, Method A)

Maximum 1 ppm of ethylene oxide and maximum 10 ppm

of dioxan.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution

using 2 mL of *lead standard solution* (10 ppm Pb) R.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.00 g.

Polysorbate 80

Polyoxyethylene 20 Sorbitan Mono-oleate

(Ph. Eur. monograph 0428)

Action and use

Non-ionic surfactant.

DEFINITION

Mixture of partial esters of fatty acids, mainly Oleic

acid (0799), with sorbitol and its anhydrides ethoxylated with

approximately 20 moles of ethylene oxide for each mole of

sorbitol and sorbitol anhydrides.

CHARACTERS

Appearance

Oily, colourless or brownish-yellow, clear or slightly

opalescent liquid.

Solubility

Dispersible in water, in anhydrous ethanol, in ethyl acetate

and in methanol, practically insoluble in fatty oils and in

liquid paraffin.

Relative density

About 1.10.

Viscosity About 400 mPa.s at 25 °C.

IDENTIFICATION

First identification A, D

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of polysorbate 80.

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Composition of fatty acids (see Tests).

E. Dissolve 0.1 g in 5 mL of *methylene chloride* R. Add 0.1 g

of *cobalt nitrate* R and 0.1 g of *potassium thiocyanate* R. Stir

with a glass rod. The solution becomes blue.

TESTS

Acid value (2.5.1)

Maximum 2.0.

Dissolve 5.0 g in 50 mL of the prescribed mixture of

solvents.

Hydroxyl value (2.5.3, Method A)

65 to 80.

Peroxide value

Maximum 10.0.

Introduce 10.0 g into a 100 mL beaker and dissolve with

20 mL of *glacial acetic acid* R. Add 1 mL of *saturated*

potassium iodide solution R, mix and allow to stand for 1 min.

Add 50 mL of *carbon dioxide-free water* R and a magnetic

stirring bar. Titrate with 0.01 M *sodium thiosulfate*,

determining the end-point potentiometrically (2.2.20). Carry

out a blank titration.

Determine the peroxide value using the following expression:



Ph. Eur.

STORAGE

Maximum 0.25 per cent, determined on 2.0 g.

In an airtight container, protected from light.

$$\frac{m}{(n_1 - n_2) \times M \times 1000}$$

n_1 = volume of 0.01 M sodium thiosulfate required for the

substance to be examined, in millilitres;

n_2 = volume of 0.01 M sodium thiosulfate required for the

blank titration, in millilitres;

M = molarity of the sodium thiosulfate solution, in

moles per litre;

m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)

45 to 55, determined on 4.0 g.

Use 30.0 mL of 0.5 M alcoholic potassium hydroxide, heat

under reflux for 60 min and add 50 mL of anhydrous

ethanol R before carrying out the titration.

Composition of fatty acids (2.4.22, Method C)

Use the mixture of calibrating substances in Table 2.4.22-3.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— stationary phase: macrogel 20 000 R (film thickness

0.5 μ m).

Carrier gas helium for chromatography R.

Linear velocity 50 cm/s.

Temperature:

Time	Temperature	Column	Injection port	Detector
(min)	(°C)	(min)		
0 - 14	80 \rightarrow 220	14 - 54	250	250

Detection Flame ionisation.

Injection 1 μ L.

Composition of the fatty-acid fraction of the substance:

— myristic acid: maximum 5.0 per cent;

— palmitic acid: maximum 16.0 per cent;

— palmitoleic acid: maximum 8.0 per cent;

— stearic acid: maximum 6.0 per cent;

— oleic acid: minimum 58.0 per cent;

— linoleic acid: maximum 18.0 per cent;

— linolenic acid: maximum 4.0 per cent.

Ethylene oxide and dioxan

Maximum 1 ppm of ethylene oxide and maximum 10 ppm

of dioxan.

Head-space gas chromatography (2.2.28).

Ethylene oxide stock solution Dilute 0.5 mL of a commercially

available solution of ethylene oxide in methylene chloride

(50 mg/mL) to 50.0 mL with water R. [NOTE: the solution

is stable for 3 months, if stored in vials with

polytetrafluoroethylene coated silicone membrane crimped

caps at -20°C]. Allow to reach room temperature. Dilute

1.0 mL of this solution to 250.0 mL with water R.

Dioxan stock solution Dilute 1.0 mL of dioxan R to 200.0 mL

with water R. Dilute 1.0 mL of this solution to 100.0 mL

with water R.

Acetaldehyde stock solution Weigh about 0.100 g of

acetaldehyde R into a 100 mL volumetric flask and dilute to

100.0 mL with water R. Dilute 1.0 mL of this solution to

$$\frac{m}{(n_1 - n_2) \times M \times 1000}$$

n_1 = volume of 0.01 M sodium thiosulfate required for the

substance to be examined, in millilitres;

n_2 = volume of 0.01 M sodium thiosulfate required for the

blank titration, in millilitres;

M = molarity of the sodium thiosulfate solution, in

moles per litre;

m = mass of the substance to be examined, in grams.

Saponification value (2.5.6)

45 to 55, determined on 4.0 g.

Use 30.0 mL of 0.5 M alcoholic potassium hydroxide, heat

under reflux for 60 min and add 50 mL of anhydrous

ethanol R before carrying out the titration.

Composition of fatty acids (2.4.22, Method C)

Use the mixture of calibrating substances in Table 2.4.22-3.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— stationary phase: macrogel 20 000 R (film thickness

0.5 μ m).

Carrier gas helium for chromatography R.

Linear velocity 50 cm/s.

Temperature:

Standard solution To 6.0 mL of ethylene oxide stock solution add 2.5 mL of dioxan stock solution and dilute to 25.0 mL with water R.

Test solution (a) Weigh 1.00 g of the substance to be examined into a 10 mL head-space vial. Add 2.0 mL of water R, seal the vial immediately with a polytetrafluoroethylene coated silicon membrane and an aluminium cap. Mix carefully.

Test solution (b) Weigh 1.00 g of the substance to be examined into a 10 mL head-space vial. Add 2.0 mL of standard solution, seal the vial immediately with a polytetrafluoroethylene coated silicon membrane and an aluminium cap. Mix carefully.

Reference solution Introduce 2.0 mL of acetaldehyde stock solution and 2.0 mL of ethylene oxide stock solution into a 10 mL head-space vial and seal the vial immediately with a polytetrafluoroethylene coated silicon membrane and an aluminium cap. Mix carefully.

Column:

— material: fused silica;

— size: $l = 50$ m, $\varnothing = 0.53$ mm;

— stationary phase: poly(dimethyl) (diphenyl) siloxane R (5 μ m).

Carrier gas helium for chromatography R.

Flow rate 4.0 mL/min.

Split ratio 1:3.5.

Static head-space conditions that may be used:

— equilibration temperature: 80°C ;

— equilibration time: 30 min.

Temperature:

Time	Temperature	Column	Injection port	Detector
(min)	(°C)	(min)		
0 - 18	70 \rightarrow 250	18 - 23	250	250

Detection Flame ionisation.

Injection 1.0 mL of test solutions (a) and (b) and of the

reference solution.

Relative retention With reference to ethylene oxide (retention

time = about 6.5 min): acetaldehyde = about 0.9;

dioxan = about 1.9.

System suitability: reference solution.

— resolution: minimum 2.0 between the peaks due to

acetaldehyde and ethylene oxide.

Calculate the content of ethylene oxide using the following

expression:

$$\frac{A_b - A_a}{2C_{EO} \times A_a}$$

C_{EO} = concentration of added ethylene oxide in test

solution (b), in micrograms per millilitre;

A_a = peak area of ethylene oxide in the chromatogram

obtained with test solution (a);

A_b = peak area of ethylene oxide in the chromatogram

obtained with test solution (b).

Calculate the content of dioxan using the following

expression:

$$2 \times 1.03 \times C_D \times A_{a'}$$

$$\frac{A_{b'} - A_{a'}}{A_{a'}}$$

- C_D = concentration of added dioxan in test solution
 (b), in microlitres per millilitre;
 1.03 = density of dioxan, in grams per millilitre;
 $A_{a'}$ = peak area of dioxan in the chromatogram
 obtained with test solution (a);
 $A_{b'}$ = peak area of dioxan in the chromatogram
 obtained with test solution (b).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 3.0 per cent, determined on 1.00 g.

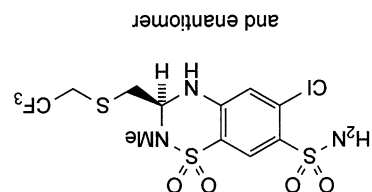
Total ash (2.4.16)

Maximum 0.25 per cent, determined on 2.0 g.

STORAGE

In an airtight container, protected from light.

Ph Eur

Polythiazide

$C_{11}H_{13}ClF_3N_3O_4S_3$ 439.9

346-18-9

DEFINITION

Polythiazide is (R,S)-3,4-dihydro-2-methyl-3-(2,2,2-trifluoroethylthiomethyl)-1,2,4-benzothiadiazine-7-sulfonamide 1,1-dioxide. It contains not less than 97.0% and not more than 102.0% of $C_{11}H_{13}ClF_3N_3O_4S_3$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder. Practically insoluble in water; sparingly soluble in ethanol (96%).

IDENTIFICATION

- A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of polythiazide (RS 280).
 B. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.002% w/v solution in methanol exhibits a maximum at 268 nm and a less well-defined maximum at 317 nm. The absorbance at 268 nm is about 1.0.
 C. Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in methanol.
 (1) 0.02% w/v of the substance being examined.

ASSAY

Not more than 0.1%, Appendix IX A.
 Dissolve 0.1 g in sufficient methanol to produce 250 mL, dilute 5 mL to 200 mL with methanol and measure the absorbance of the resulting solution at the maximum at 268 nm, Appendix II B. Calculate the content of $C_{11}H_{13}ClF_3N_3O_4S_3$ taking 500 as the value of A(1%, 1 cm) at the maximum at 268 nm.

Sulfated ash

1.0% of its weight. Use 1 g.
 When dried to constant weight at 105°, loses not more than

Loss on drying

The sum of the intensities so assessed does not exceed 3% and no such spot is more intense than the spot in the chromatogram obtained with 5 µL of solution (2) (2%).

LIMITS

10 volumes of methanol and 90 volumes of dichloromethane.

MOBILE PHASE

applications of solution (2).
 the spots in the chromatograms obtained with the the chromatogram obtained with solution (1) by reference to using Method I. Assess the intensity of any secondary spot in (e) After removal of the plate, dry in air and reveal the spots (d) Develop the plate to 15 cm.

and 5 µL of solution (2).
 (c) Apply 5 µL of solution (1) and quantities of 1, 2, 3, 4 (b) Use the mobile phase as described below.
 (a) Use as the coating silica gel G.

CHROMATOGRAPHIC CONDITIONS

(2) 0.020% w/v of the substance being examined.
 (1) 1.0% w/v of the substance being examined.
 Appendix III A, using the following solutions in acetone.

Related substances**TESTS**

The principal spot in the chromatogram obtained with solution (1) corresponds in position and colour to that in the chromatogram obtained with solution (2).

CONFIRMATION

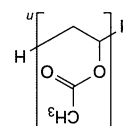
20 volumes of acetone, 30 volumes of ether and 50 volumes of toluene.

MOBILE PHASE

treat the plate by Method I and examine again.
 (e) After removal of the plate, dry in air until the solvent has evaporated, examine under ultraviolet light (254 nm) and then (d) Develop the plate to 15 cm.
 (c) Apply 20 µL of each solution.
 (b) Use the mobile phase as described below.

CHROMATOGRAPHIC CONDITIONS

(2) 0.02% w/v of polythiazide BPCRS.



DEFINITION

Poly(vinyl acetate) is a thermoplastic polymer obtained by polymerisation of vinyl acetate using a suitable starter, without solvent or with water or 2-propanol. The vast majority of the acetate moieties are attached to non-neighbouring carbon atoms of the chain.

The index n is about 100 - 17 000. The relative molecular mass lies between 10 000 and 1500 000. The viscosity is 4 to 250 mPa·s. The ester value, which characterises the degree of hydrolysis, is 615 to 675.

CHARACTERS

Appearance
White or almost white powder or colourless granules or beads.

Solubility
Practically insoluble in water, freely soluble in ethyl acetate, soluble in ethanol (96 per cent). It is hygroscopic and swells in water.

It softens at temperatures above 40-50 °C.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Discs prepared as follows: dissolve about 200 mg in 5 mL of acetone R, place 100 µL on a potassium bromide R disc and dry to evaporate the solvent. Alternatively, the spectrum may be recorded directly by attenuated total reflectance (ATR).

Comparison poly(vinyl acetate) CRS.

B. Viscosity (see Tests).

C. Saponity (2.5.6) 0.500 g in a mixture of 25.0 mL of 0.5 M alcoholic potassium hydroxide and 25.0 mL of water R. 0.15 mL of the solution obtained gives reaction (b) of acetates (2.3.1).

TESTS

Solution S

Suspend 50.0 g in 100 mL of ethyl acetate R in a borosilicate glass flask with a ground-glass neck. Heat under a reflux condenser with constant stirring for 30 min. Allow to cool. Filter through a sintered-glass filter (16) (2.1.2), wash the residue with 50.0 mL of ethyl acetate R and pour the filtrate into a 250 mL graduated flask. Dilute to 250 mL with ethyl acetate R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method 1).

Viscosity (2.2.49)

85 per cent to 115 per cent of the value stated on the label. Determine the viscosity immediately after preparation of solution S at 20 ± 0.1 °C by using a falling ball viscometer.

Acid value (2.5.1)

Maximum 2.0, determined on 5.0 g dissolved in 50 mL of ethanol (96 per cent) R by shaking for 3 h.

Ester value (2.5.2)

615 to 675.

Saponity (2.5.6) 0.500 g in a mixture of 25.0 mL of 0.5 M alcoholic potassium hydroxide and 25.0 mL of water R.

Residual peroxides

Maximum 100 ppm, calculated as hydrogen peroxide.

Place 0.85 g in a borosilicate glass flask with a ground-glass neck. Add 10 mL of ethyl acetate R and heat under a reflux condenser with constant agitation. Allow to cool. Replace the air in the container with oxygen-free nitrogen R and add a solution of 1 mL of glacial acetic acid R and 0.5 g of sodium iodide R in 40 mL of water R. Shake thoroughly and allow to stand protected from light for 20 min. Titrate with 0.005 M sodium thiosulfate until the yellow colour is discharged. Carry out a blank titration. The difference between the titration volumes is not greater than 1.0 mL.

Vinyl acetate

Head-space gas chromatography (2.2.28).

Test solution (a) Place 0.2000 g of the substance to be examined in a 20 mL vial and add 1.00 mL of

dimethylformamide R. Close the vial and secure the stopper. Shake, avoiding contact between the stopper and the liquid.

Test solution (b) Place 0.2000 g of the substance to be examined in a 20 mL vial and add 1.00 mL of the reference

solution. Close the vial and secure the stopper. Shake, avoiding contact between the stopper and the liquid.

Reference solution Place 15 mL of dimethylformamide R in a 20 mL vial, add 45 µL of vinyl acetate R and 50.0 µL of

butanal R and dilute to volume with dimethylformamide R. Dilute 1 mL of the solution to 10 mL with

dimethylformamide R.

Column:

— material: fused silica;

— size: $l = 25$ m, $\varnothing = 0.32$ mm;

— stationary phase: poly(dimethyl (diphenyl) siloxane R (film thickness 0.32 µm).

Carrier gas nitrogen for chromatography R.

Flow rate 20 mL/min.

Static head-space conditions that may be used:

— equilibration temperature: 60 °C;

— equilibration time: 20 min;

— transfer-time temperature: 120 °C;

— carrier gas: nitrogen for chromatography R.

Temperature:

— column: 155 °C;

— injection port: 120 °C;

— detector: 180 °C.

Detection Flame ionisation.

Injection 1.6 mL of the gaseous phase of test solutions (a) and (b).

System suitability: test solution (b):

— resolution: minimum 2.0 between the peaks due to vinyl acetate and butanal;

— signal-to-noise ratio: minimum 5 for the peak due to vinyl acetate.

Calculate the percentage content of vinyl acetate using the following expression:

$$S_1 = \frac{(m_1 S_2 - m_2 S_1) \times 2000}{V \times S_1 \times 0.931}$$

S_1 = area (or height) of the peak due to vinyl acetate in the chromatogram obtained with test solution (a);



Poly(vinyl acetate) Dispersion 30 per cent

(Ph. Eur. monograph 2152)

DEFINITION

Dispersion in water of poly(vinyl acetate) having a mean relative molecular mass of about 450 000. It may contain *Povidone* (0685) and a suitable surface-active agent, such as *Sodium laurylsulfate* (0098), as stabilisers.

Content

25.0 per cent to 30.0 per cent of poly(vinyl acetate).

CHARACTERS

Appearance

Opaque, white or almost white, slightly viscous liquid.

Solubility

Miscible with water and with ethanol (96 per cent). It is sensitive to spoilage by microbial contaminants.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dry 1 mL in *vacuo*, dissolve the residue in *acetone* R, and spread 1 drop of the solution between 2 sodium chloride R plates; remove 1 plate and allow the solvent to evaporate.

Comparison Repeat the operation using *poly(vinyl acetate)* dispersion 30 per cent CRS.

B. Place 3 mL on a glass plate and allow to dry. A clear film is formed.

C. 50 mg gives the reaction of acetyl (2.3.1).

TESTS

Agglomerates

Filter 100.0 g through a tared stainless steel sieve (90). Rinse with water R until a clear filtrate is obtained and dry to constant mass at 100–105 °C. The mass of the residue is not greater than 0.5 g.

Vinyl acetate

Test solution Introduce 0.250 g into a 10 mL volumetric flask and add about 1 mL of *methanol* R2. Sonicate. Add about 8 mL of water for chromatography R. Sonicate and dilute to 10.0 mL with water for chromatography R. Centrifuge for about 10 min and filter.

Reference solution (a) Dissolve 5.0 mg of *vinyl acetate* CRS in *methanol* R2 and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 20.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) To 5 mg of *vinyl acetate* R and 5 mg of *1-vinylpyrrolidin-2-one* R, add 10 mL of *methanol* R2 and sonicate. Dilute to 50 mL with mobile phase A. Dilute 1 mL of this solution to 20 mL with mobile phase A. A precolumn containing *octadecylsilyl silica* gel for chromatography R (5 µm) may be used if a matrix effect is observed.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: *octadecylsilyl silica* gel for chromatography R (5 µm);

— temperature: 30 °C.

S_2 = area (or height) of the peak due to vinyl acetate in the chromatogram obtained with test solution (b);
 m_1 = mass of the substance to be examined used to prepare test solution (a), in grams;
 m_2 = mass of the substance to be examined used to prepare test solution (b), in grams;
 ρ = density of vinyl acetate, in grams per millilitre;
 V = volume of vinyl acetate used to prepare the reference solution, in millilitres.

Limit:

— *vinyl acetate*: maximum 0.3 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

1.0 g complies with test D. Prepare the reference solution using 1 mL of *lead standard solution* (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

STORAGE

In an airtight container.

LABELLING

The label states:

— the nominal relative molecular mass;

— the viscosity.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for poly(vinyl acetate) used as matrix former in prolonged-release dosage forms or film former.

Viscosity

(see Tests).

Solubility of a film

Place 3 mL of solution S on a glass plate and dry. Place the film obtained in 50 mL of *phosphate buffer solution* pH 6.8 R whilst stirring continuously. The film does not dissolve within 30 min.

Ph Eur

Residue on evaporation
28.5 per cent to 31.5 per cent, determined on 1.000 g at 110 °C for 5 h.

Sulfated ash
Maximum 0.5 per cent, determined on 1.0 g.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the preparation to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 ± 25 °C, until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting with "Moisten the substance to be examined...".

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).
TYMC: acceptance criterion 10² CFU/g (2.6.12).

ASSAY

Determine the saponification value (2.5.6) on 1.5 g and calculate the percentage content of poly(vinyl acetate) using the following expression:

$$I_s \times 0.1534$$

$$I_s = \text{saponification value.}$$

STORAGE

At a temperature of 5 °C to 30 °C. Handle the substance so as to minimise microbial contamination.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for poly(vinyl acetate) dispersion 30 per cent used in the manufacture of modified-release dosage forms and to mask taste.

Solubility of a film

Place the film obtained in identification test B in 50 mL of phosphate buffer solution pH 6.8 R whilst stirring continuously. The film does not dissolve within 30 min.

Apparent viscosity (2.2.10)
Maximum 100 mPa·s, determined using a rotating viscometer at 20 °C and a shear rate of 100 s⁻¹.

Ph Eur

Mobile phase:

— mobile phase A: acetonitrile for chromatography R, (5:5:90 V/V/V);
— methanol R2, water for chromatography R (5:5:90 V/V/V);
— mobile phase B: methanol R2, acetonitrile for chromatography R, water for chromatography R (5:45:50 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	100	0
2 - 26	100 → 80	0 → 20
26 - 27	80 → 0	20 → 100
27 - 30	0 → 100	100 → 0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 10 µL.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to vinyl acetate and 1-vinylpyrrolidin-2-one.

Limit:

in the chromatogram obtained with reference solution (a) (100 ppm).

Povidone

Maximum 4.0 per cent.

Carry out the determination of nitrogen by sulfonic acid digestion (2.5.9), using 0.25 g. Calculate the percentage content of povidone using the following expression:

$$\frac{N}{0.126}$$

N = percentage content of nitrogen.

Acetic acid

Liquid chromatography (2.2.29).

Test solution Mix 0.200 g with water for chromatography R.

Sonicate for about 10 min and dilute to 10.0 mL with water

for chromatography R.

Reference solution Dissolve 30.0 mg of acetic acid R and 30 mg of citric acid R in the mobile phase. Shake gently to dissolve

and dilute to 100.0 mL with the mobile phase.

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase 0.005 M sulfonic acid.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 µL; after each injection, rinse the column with a mixture of equal volumes of acetonitrile for chromatography R and 0.005 M sulfonic acid.

Retention time Acetic acid = about 6 min; citric acid = about 8 min.

System suitability: reference solution:

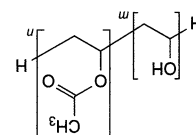
— resolution: minimum 2.0 between the peaks due to acetic acid and citric acid.

Limit:

— acetic acid: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (1.5 per cent).

Poly(vinyl alcohol)

(Ph. Eur. monograph 1961)



DEFINITION

Poly(vinyl alcohol) is obtained by polymerisation of vinyl acetate, followed by partial or almost complete hydrolysis of poly(vinyl acetate) in the presence of catalytic amounts of alkali or mineral acids.

Poly(vinyl alcohol) polymers comply with the following indices:

$$0 \leq \frac{m}{n} \leq 0.35$$

The mean relative molecular mass lies between 20 000 and 150 000. The viscosity is 3 to 70 mPa.s. The ester value, which characterises the degree of hydrolysis, is not greater than 280.

CHARACTERS

Appearance

Yellowish-white powder or translucent granules.

Solubility

Soluble in water, slightly soluble in ethanol, practically insoluble in acetone.

Various grades of poly(vinyl alcohol) are available. They differ in their degree of polymerisation and their degree of hydrolysis which determine the physical properties of the different grades. They are characterised by the viscosity and the ester value of the substance.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison poly(vinyl alcohol) CRS.

The intensities of the absorption bands at about 1720 cm⁻¹ and 1260 cm⁻¹ are inversely proportional to the degree of hydrolysis.

TESTS

B. Viscosity (see Tests).

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

pH (2.2.3)

4.5 to 6.5 for solution S.

Viscosity (2.2.49)

85 per cent to 115 per cent of the value stated on the label.

Determine the viscosity using a falling ball viscometer immediately after preparation of solution S at 20 ± 0.1 °C.

Acid value

Maximum 3.0.

Add 1 mL of phenolphthalein solution R to 50 mL of solution S and titrate with 0.05 M potassium hydroxide until the pink colour persists for 15 s. Calculate the acid value using the following expression:

$$\frac{2.805V}{2}$$

V = volume of 0.05 M potassium hydroxide used, in millilitres.

Ester value (2.5.2)

90 per cent to 110 per cent of the value stated on the label. Saponify (2.5.6) 1.00 g in a mixture of 25.0 mL of 0.5 M alcoholic potassium hydroxide and 25.0 mL of water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

1.0 g complies with test D. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 1.0 per cent, determined on 1.0 g.

LABELLING

The label states:

— the viscosity for a 40 g/L solution;

— the ester value.

Ph Eur

Potassium Acetate

(Ph. Eur. monograph 1139)

C₂H₃KO₂

98.1

127-08-2



Action and use

Used in solutions for dialysis.

DEFINITION

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals, deliquescent.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. It gives reaction (a) of acetates (2.3.1).

B. It gives reaction (a) of potassium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).



Potassium Bicarbonate

(Potassium Hydrogen Carbonate,
Ph Eur monograph 1141)

298-14-6

100.1

KHCO₃

Action and use

Excipient.

Ph Eur

DEFINITION

Content

99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, practically insoluble in ethanol

(96 per cent).

When heated in the dry state or in solution, it is gradually

converted to potassium carbonate.

IDENTIFICATION

A. To 5 mL of solution S (see Tests) add 0.1 mL of

phenolphthalein solution R. A pale pink colour is produced.

Heat; gas is evolved and the colour becomes red.

B. It gives the reaction of carbonates and bicarbonates

(2.3.1).

C. 1 mL of solution S gives reaction (b) of potassium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in 90 mL of carbon dioxide-free water R

prepared from distilled water R and dilute to 100 mL with the

same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Carbonates

The pH (2.2.3) of freshly prepared solution S is not greater

than 8.6.

Chlorides (2.4.4)

Maximum 150 ppm.

Dilute 7 mL of solution S to 15 mL with dilute nitric acid R.

Sulfates (2.4.13)

Maximum 150 ppm.

Dilute 10 mL of solution S to 15 mL with acetic acid R.

Prepare the standard using a mixture of 7.5 mL of sulfate

standard solution (10 ppm SO₄) R and 7.5 mL of distilled

water R.

Ammonium (2.4.1)

Maximum 20 ppm.

Dilute 10 mL of solution S to 15 mL with water R.

Calcium (2.4.3)

Maximum 100 ppm.

Dilute 10 mL of solution S to 15 mL with acetic acid R.

Prepare the standard using a mixture of 5 mL of calcium

standard solution (10 ppm Ca) R and 10 mL of distilled

water R.

Iron (2.4.9)

Maximum 20 ppm, determined on solution S.

Sodium

Maximum 0.5 per cent.

pH (2.2.3)
7.5 to 9.0.
Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Reducing substances

Dilute 10 mL of solution S to 100 mL with water R.

Add 5 mL of dilute sulfuric acid R and 0.5 mL of a 0.32 g/L

solution of potassium permanganate R. Mix and boil gently for

5 min. The solution remains pink.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 200 ppm.

Dilute 7.5 mL of solution S to 15 mL with distilled water R.

Aluminium (2.4.17)

Maximum 1 ppm, if intended for use in the manufacture of

peritoneal dialysis solutions, haemofiltration solutions or

haemodialysis solutions.

Prescribed solution Dissolve 2.0 g in 50 mL of water R and add

5 mL of acetate buffer solution pH 6.0 R.

Reference solution Mix 1 mL of aluminium standard solution

(2 ppm Al) R, 5 mL of acetate buffer solution pH 6.0 R and

49 mL of water R.

Blank solution Mix 5 mL of acetate buffer solution pH 6.0 R

and 50 mL of water R.

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Sodium

Maximum 0.5 per cent.

Atomic emission spectrometry (2.2.22, Method II).

Test solution Dissolve 1.00 g in water R and dilute to

100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using sodium

standard solution (200 ppm Na) R, diluted as necessary with

water R.

Wavelength 589 nm.

Heavy metals (2.4.8)

Maximum 4 ppm.

Dissolve 5.0 g in water R and dilute to 20 mL with the same

solvent. 12 mL of the solution complies with test A. Prepare

the reference solution using lead standard solution

(1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 3.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

ASSAY

Dissolve 80.0 mg in 20 mL of anhydrous acetic acid R.

Add 0.2 mL of naphtholbenzenesulfonic acid R. Titrate with 0.1 M

perchloric acid. Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 9.81 mg

of C₂H₃KO₂.

STORAGE

In an airtight container.

Ph Eur

Atomic emission spectrometry (2.2.22, Method II).
Test solution Dissolve 1.00 g in water R and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using sodium standard solution (200 ppm Na) R, diluted as necessary with water R.

Wavelength 589 nm.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in a mixture of 2 mL of hydrochloric acid R

and 18 mL of water R. 12 mL of the solution complies with

test A. Prepare the reference solution using lead standard

solution (1 ppm Pb) R.

ASSAY

Dissolve 0.800 g in 50 mL of carbon dioxide-free water R.

Add 0.1 mL of methyl orange solution R. Titrate with 1 M

hydrochloric acid until the yellow colour begins to change to

yellowish-pink. Heat cautiously and boil for at least 2 min.

The solution becomes yellow. Cool and titrate until a

yellowish-red colour is obtained.

1 mL of 1 M hydrochloric acid is equivalent to 0.1001 g

of KHCO_3 .

Potassium Bromide

(Ph. Eur. monograph 0184)

Ph Eur

DEFINITION

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals.

Solubility

Freely soluble in water and in glycerol, slightly soluble in

ethanol (96 per cent).

IDENTIFICATION

A. It gives reaction (a) of bromides (2.3.1).

B. Solution S (see Tests) gives the reactions of potassium

(2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R and dilute to

100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of bromothymol blue

solution R1. Not more than 0.5 mL of 0.01 M hydrochloric

acid or 0.01 M sodium hydroxide is required to change the

colour of the indicator.

Bromates

To 10 mL of solution S add 1 mL of starch solution R,
0.1 mL of a 100 g/L solution of potassium iodide R and
0.25 mL of 0.5 M sulfuric acid and allow to stand protected
from light for 5 min. No blue or violet colour develops.

Chlorides and sulfates

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 0.400 g of the substance to be

examined in 50 mL of water for chromatography R and dilute

to 100.0 mL with the same solvent.

Test solution (b) Dilute 25.0 mL of test solution (a) to

50.0 mL with water for chromatography R.

Reference solution (a) To 25.0 mL of test solution (a) add

1.0 mL of sulfate standard solution (10 ppm SO_4) R and

12.0 mL of chloride standard solution (50 ppm Cl) R and dilute

to 50.0 mL with water for chromatography R.

Reference solution (b) Dilute 10.0 mL of test solution (a) to

100.0 mL with water for chromatography R. To 2.0 mL of this

solution add 8.0 mL of chloride standard solution

(50 ppm Cl) R and dilute to 20.0 mL with water for

chromatography R.

Blank solution water for chromatography R.

Column:

— size: $l = 0.25$ m, $\varnothing = 2$ mm;

— stationary phase: strongly basic anion-exchange resin for

chromatography R (13 μm).

Mobile phase Dissolve 0.600 g of potassium hydroxide R in

same solvent.

Flow rate 0.4 mL/min.

Detection Conductivity detector equipped with a suitable ion

suppressor.

Injection 50 μL of test solution (b), reference solutions (a)

and (b) and the blank solution.

Run time 2.5 times the retention time of bromide.

Retention time Chloride = about 5 min; bromide = about

8 min; sulfate = about 16 min.

System suitability: reference solution (b):

— resolution: minimum 8.0 between the peaks due to

chloride and bromide.

Calculation of percentage contents:

— for chlorides, use the concentration of chloride in

reference solution (a); correct the area of the peak due to

chloride in the chromatogram obtained with reference

solution (a) by subtracting the area of the peak due to

chloride in the chromatogram obtained with test

solution (b);

— for sulfates, use the concentration of sulfate in reference

solution (a); correct the area of the peak due to sulfate in

the chromatogram obtained with reference solution (a) by

subtracting the area of the peak due to sulfate in the

chromatogram obtained with test solution (b).

Limits:

— chlorides: maximum 0.6 per cent;

— sulfates: maximum 0.01 per cent.

Iodides

To 5 mL of solution S add 0.15 mL of ferric chloride

solution R1 and 2 mL of methylene chloride R. Shake and allow

to separate. The lower layer is colourless (2.2.2, Method I).

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Magnesium and alkaline-earth metals (2.4.7)

Maximum 200 ppm, calculated as Ca.

10.0 g complies with the test for magnesium and alkaline-

earth metals. The volume of 0.01 M sodium edetate used does

not exceed 5.0 mL.

Dissolve 0.50 g in 10 mL of water R. Carefully add dropwise 1 mL of nitric acid R. Boil. Cool, add 5 mL of dilute nitric acid R and dilute to 15 mL with water R.

Sulfates (2.4.13)

Dilute 7.50 mL of solution S to 15 mL with distilled water R.

Calcium (2.4.3)

Maximum 100 ppm.

To 5 mL of solution S add 1 mL of concentrated ammonia R.

Boil. Cool. Dilute to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 10 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dilute 10 mL of solution S to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 0.300 g by drying in an oven at 120–125 °C for 5 h.

ASSAY

Dissolve 0.500 g in 50 mL of carbon dioxide-free water R.

Carry out a potentiometric titration (2.2.20), using 1 M hydrochloric acid. Read the volume added at the 2nd point of inflexion.

1 mL of 1 M hydrochloric acid is equivalent to 69.1 mg of K₂CO₃.

STORAGE

In an airtight container.

Ph Eur

Potassium Chloride

(Ph. Eur. monograph 0185)

KCl

74.6

7447-40-7



Action and use

Used in prevention and treatment of potassium deficiency and electrolyte imbalance.

Preparations

Bumetanide and Prolonged-release Potassium Tablets

Oral Rehydration Salts

Sterile Potassium Chloride Concentrate

Potassium Chloride and Glucose Intravenous Infusion

Potassium Chloride and Sodium Chloride Intravenous Infusion

Potassium Chloride, Sodium Chloride and Glucose Intravenous Infusion

Potassium Chloride Oral Solution

Prolonged-release Potassium Chloride Tablets

Ph Eur

DEFINITION

Content

99.0 per cent to 101.0 per cent of KCl (dried substance).

CHARACTERS

Appearance

White or almost white granular powder, hygroscopic.

Solubility

Freely soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. Dissolve 1 g in 10 mL of water R. The solution is strongly alkaline (2.2.4).

B. 2 mL of the solution prepared for identification test A gives the reaction of carbonates and bicarbonates (2.3.1).

C. 1 mL of the solution prepared for identification test A gives reaction (b) of potassium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in 25 mL of distilled water R. Slowly add 14 mL of hydrochloric acid R. When the effervescence has ceased, boil for a few minutes. Allow to cool and dilute to 50 mL with distilled water R.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Chlorides (2.4.4)

Maximum 100 ppm.

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, practically insoluble in anhydrous ethanol.

IDENTIFICATION

A. It gives the reactions of chlorides (2.3.1).
B. Solution S (see Tests) gives the reactions of potassium (2.3.1).

TESTS

Solution S
Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity
To 50 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Bromides
Maximum 0.1 per cent.

Dilute 1.0 mL of solution S to 50 mL with water R.
To 5.0 mL of the solution add 2.0 mL of phenol red solution R2 and 1.0 mL of chloramine solution R1 and mix immediately. After exactly 2 min add 0.15 mL of 0.1 M sodium thiosulfate, mix and dilute to 10.0 mL with water R.
The absorbance (2.2.25) of the solution measured at 590 nm, using water R as the compensation liquid, is not greater than that of a standard prepared at the same time and in the same manner using 5 mL of a 3.0 mg/L solution of potassium bromide R.

Iodides

Moisten 5 g by the dropwise addition of a freshly prepared mixture of 0.15 mL of sodium nitrite solution R, 2 mL of 0.5 M sulfuric acid, 25 mL of iodide-free starch solution R and 25 mL of water R. After 5 min, examine in daylight.
The substance shows no blue colour.

Sulfates

Maximum 300 ppm.
Dilute 5 mL of solution S to 15 mL with distilled water R.

Aluminium (2.4.17)
Maximum 1.0 ppm, if intended for use in the manufacture of haemodialysis solutions.

Prescribed solution Dissolve 4 g in 100 mL of water R and add 10 mL of acetate buffer solution pH 6.0 R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R.

Barium

To 5 mL of solution S add 5 mL of distilled water R and 1 mL of dilute sulfuric acid R. After 15 min, any opalescence of solution S and 6 mL of distilled water R.

Iron

Maximum 20 ppm.
Dilute 5 mL of solution S to 10 mL with water R.

Sodium

Maximum 200 ppm, calculated as Ca, determined on 10.0 g using 0.15 g of mordant black II titrate R. The volume of 0.01 M sodium edetate used does not exceed 5.0 mL.

Maximum 0.1 per cent, if intended for use in the manufacture of parenteral preparations or haemodialysis solutions.

Atomic emission spectrometry (2.2.22, Method I).

Test solution Dissolve 1.00 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions by diluting as required a solution containing 200 µg of Na per millilitre, prepared as follows: dissolve in water R 0.5084 g of sodium chloride R, previously dried at 105 °C for 3 h, and dilute to 1000.0 mL with the same solvent.

Heavy metals

Maximum 10 ppm.
12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

ASSAY

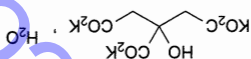
Dissolve 60.0 mg in water R, add 5 mL of dilute nitric acid R and dilute to 50 mL with water R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M silver nitrate is equivalent to 7.46 mg of KCl.

LABELLING

The label states:
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations;
— where applicable, that the substance is suitable for use in the manufacture of haemodialysis solutions.

Potassium Citrate

(Ph. Eur. monograph 0400)



$C_6H_5K_3O_7 \cdot H_2O$ 324.4

6100-05-6

Action and use

Alkalisiation of urine.

Preparation

Potassium Citrate Mixture

DEFINITION

Tripotassium 2-hydroxypropane-1,2,3-tricarboxylate monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).



CHARACTERS

Appearance
White or almost white, granular powder or transparent crystals, hygroscopic.

Solubility
Very soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. To 1 mL of solution S (see Tests) add 4 mL of water R. The solution gives the reaction of citrates (2.3.1).

B. 0.5 mL of solution S gives reaction (b) of potassium (2.3.1).

TESTS

Solution S
Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of phenolphthalein solution R. Not more than 0.2 mL of 0.1 M hydrochloric acid or 0.1 M sodium hydroxide is required to change the colour of the indicator.

Readily carbonisable substances

To 0.20 g of the powdered substance to be examined add 10 mL of sulfuric acid R and heat in a water-bath at

$90 \pm 1^\circ \text{C}$ for 60 min. Cool rapidly. The solution is not more intensely coloured than reference solution Y₂ or GY₂ (2.2.2, Method II).

Chlorides (2.4.4)

Maximum 50 ppm.

Dilute 10 mL of solution S to 15 mL with water R.

Oxalates

Maximum 300 ppm.

Dissolve 0.50 g in 4 mL of water R, add 3 mL of hydrochloric acid R and 1 g of zinc R in granules and heat on a water-bath for 1 min. Allow to stand for 2 min, decant the liquid into a test-tube containing 0.25 mL of a 10 g/L solution of

phenylhydrazine hydrochloride R and heat to boiling. Cool rapidly, transfer to a graduated cylinder and add an

equal volume of hydrochloric acid R and 0.25 mL of potassium ferricyanide solution R. Shake and allow to stand for 30 min.

Any pink colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 4 mL of a 0.05 g/L solution of oxalic acid R.

Sulfates (2.4.13)

Maximum 150 ppm.

To 10 mL of solution S add 2 mL of hydrochloric acid R1 and dilute to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Sodium

Maximum 0.3 per cent.

Atomic emission spectrometry (2.2.22, Method II).

Test solution To 10 mL of solution S add 1 mL of dilute hydrochloric acid R and dilute to 100 mL with distilled water R.

Reference solutions Prepare the reference solutions using a solution of sodium chloride R containing 1 mg of Na per millilitre diluted as necessary with distilled water R.

Wavelength 589 nm.

Water (2.5.12)

4.0 per cent to 7.0 per cent, determined on 0.250 g. Use a

mixture of 1 volume of formamide R and 2 volumes of methanol R as solvent. After adding the substance to be

examined, stir for 15 min before titrating.

ASSAY

Dissolve 0.150 g in 20 mL of anhydrous acetic acid R, heating to about 50°C . Allow to cool. Titrate with 0.1 M perchloric acid using 0.25 mL of naphtholbenzene solution R as indicator

until a green colour is obtained.

1 mL of 0.1 M perchloric acid is equivalent to 10.21 mg

of $\text{C}_8\text{H}_8\text{KNO}_5$.

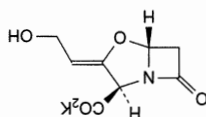
STORAGE

In an airtight container.

Ph Eur

Potassium Clavulanate

(Ph. Eur. monograph 1140)



237.3

$\text{C}_8\text{H}_8\text{KNO}_5$

61177-45-5

Action and use

Beta-lactamase inhibitor, potentiation of the action of amoxicillin and ticarcillin.

Preparations

Co-amoxiclav Injection

Co-amoxiclav Oral Suspension

Co-amoxiclav Tablets

Dispersible Co-amoxiclav Tablets

Ticarcillin and Clavulanic Acid Infusion

Ph Eur

DEFINITION

Potassium (2R,3Z,5R)-3-(2-hydroxyethylidene)-7-oxo-4-oxa-1-azabicyclo[3.2.0]heptane-2-carboxylate, the potassium salt of a substance produced by the growth of certain strains of *Streptomyces clavuligenus* or obtained by any other means.

Content

96.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder, hygroscopic.

Solubility

Freely soluble in water, slightly soluble in ethanol

(96 per cent), very slightly soluble in acetone.

PRODUCTION

The methods of production, extraction and purification are such that clavam-2-carboxylate is eliminated or present at a level not exceeding 0.01 per cent.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of potassium clavulanate.

B. It gives reaction (b) of potassium (2.3.1).

TESTS

Solution S

Dissolve 0.400 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

pH (2.2.3)

5.5 to 8.0.

Dilute 5 mL of solution S to 10 mL with carbon dioxide-free water R.

Specific optical rotation (2.2.7)

+ 53 to + 63 (anhydrous substance), determined on solution S.

Polymetric impurities and other impurities absorbing at 278 nm

Dissolve 50.0 mg in 0.1 M phosphate buffer solution pH 7.0 R and dilute to 50.0 mL with the same buffer solution.

Measure the absorbance immediately.

The absorbance (2.2.25) of the solution determined at 278 nm is not greater than 0.40.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 0.250 g of the substance to be examined in mobile phase A and dilute to 25.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 10 mg of lithium clavulanate CRS and 10 mg of amoxicillin trihydrate CRS in mobile phase A and dilute to 100 mL with mobile phase A.

Reference solution (c) Dissolve 2 mg of potassium clavulanate impurity G CRS in 20 mL of mobile phase A.

Column: size: $l = 0.10$ m, $\phi = 4.6$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase: mobile phase A: a 7.8 g/L solution of sodium dihydrogen phosphate R adjusted to pH 4.0 with phosphoric acid R;

mobile phase B: a mixture of equal volumes of methanol R and mobile phase A;

Time (min)

Mobile phase A (per cent V/V)

Mobile phase B (per cent V/V)

0 - 4 100 0

4 - 15 100 \rightarrow 50 0 \rightarrow 50

15 - 18 50 50

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Relative retention With reference to clavulanate (retention time = about 3 min): impurity E = about 2.3;

impurity G = about 3.6.

System suitability: reference solution (b):

— resolution: minimum 13 between the peaks due to clavulanate (1st peak) and amoxicillin (2nd peak).

Limits:

— impurities E, G: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— any other impurity: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (2.0 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Aliphatic amines

Gas chromatography (2.2.28).

The method shown below can be used to determine the following aliphatic amines: 1,1-dimethylethylamine;

tetramethylethylenediamine; 1,1,3,3-tetramethylbutylamine;

N,N' -diisopropylethylenediamine;

2,2'-oxybis(N,N' -dimethylethylamine).

Internal standard solution Dissolve 50 μ L of 3-methylpentan-2-one R in water R and dilute to 100.0 mL with the same solvent.

Test solution Weigh 1.00 g of the substance to be examined into a centrifuge tube. Add 5.0 mL of the internal standard solution, 5.0 mL of dilute sodium hydroxide solution R,

10.0 mL of water R, 5.0 mL of 2-methylpropanol R and 5 g of sodium chloride R. Shake vigorously for 1 min. Centrifuge to separate the layers.

Reference solution Dissolve 80.0 mg of each of the following amines: 1,1-dimethylethylamine R; tetramethylethylenediamine R; 1,1,3,3-tetramethylbutylamine R;

N,N' -diisopropylethylenediamine R and 2,2'-oxybis(N,N' -dimethylethylamine) R in dilute hydrochloric acid R and dilute to 200.0 mL with the same acid. Introduce 5.0 mL of this solution into a centrifuge tube. Add 5.0 mL of the internal standard solution, 10.0 mL of dilute sodium hydroxide

solution R, 5.0 mL of 2-methylpropanol R and 5 g of sodium chloride R. Shake vigorously for 1 min. Centrifuge to separate the layers.

Column: material: fused silica;

size: $l = 50$ m, $\phi = 0.53$ mm;

stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 5 μ m).

Carrier gas helium for chromatography R.

Flow rate 8 mL/min.

Split ratio 1:10.

Temperature:

Time (min)

Temperature (°C)

Column 0 - 7 35

Injection port 7 - 10.8 35 \rightarrow 150

Detector 10.8 - 25.8 150

200

250

Detection Flame ionisation.

Injection 1 μ L of the upper layers obtained from the test solution and the reference solution.

Relative retention With reference to 3-methylpentan-2-one (retention time = about 11.4 min):

impurity H = about 0.55; impurity J = about 1.07;

impurity K = about 1.13; impurity L = about 1.33;

impurity M = about 1.57.

Limit:

— total of aliphatic amines: maximum 0.2 per cent.

2-Ethylhexanoic acid (2.4.28)

Maximum 0.8 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.00 g.

Bacterial endotoxins (2.6.14)

Less than 0.03 IU/mg if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 50.0 mg of the substance to be examined in a 4.1 g/L solution of sodium acetate R previously adjusted to pH 6.0 with glacial acetic acid R, and dilute to 50.0 mL with the same solution.

Reference solution (a) Dissolve 50.0 mg of lithium

clavulanate CRS in a 4.1 g/L solution of sodium acetate R previously adjusted to pH 6.0 with glacial acetic acid R and dilute to 50.0 mL with the same solution.

Reference solution (b) Dissolve 10 mg of amoxicillin trihydrate CRS in 10 mL of reference solution (a).

Column:

— size: $l = 0.3$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (10 μ m).

Mobile phase Mix 5 volumes of methanol R1 and 95 volumes of a 15 g/L solution of sodium dihydrogen phosphate R previously adjusted to pH 4.0 with dilute phosphoric acid R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 μ L.

System suitability: reference solution (b):
— resolution: minimum 3.5 between the peaks due to clavulanate (1st peak) and amoxicillin (2nd peak).

1 mg of clavulanate ($C_{18}H_{19}NO_5$) is equivalent to 1.191 mg of $C_8H_8KNO_5$.

STORAGE

In an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, tamper-proof container.

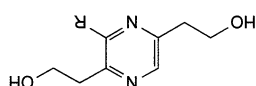
IMPURITIES

Specified impurities E, G, H, J, K, L, M.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, F.

By liquid chromatography A, B, C, D, E, F, G.

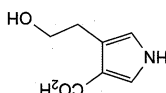
By gas chromatography H, J, K, L, M.



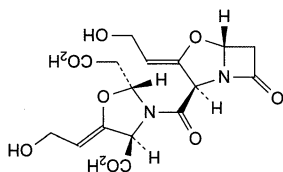
A. R = H; 2,2'-(pyrazine-2,5-diyl)diethanol,

B. R = $CH_2-CH_2-CO_2H$; 3-[3,6-bis(2-hydroxyethyl)pyrazin-2-yl]propanoic acid,

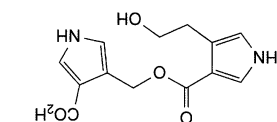
C. R = CH_2-CH_3 ; 2,2'-(3-ethylpyrazine-2,5-diyl)diethanol,



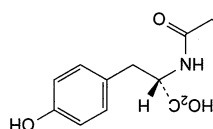
D. 4-(2-hydroxyethyl)-1H-pyrrrole-3-carboxylic acid,



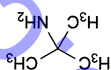
E. (2R,4R,5Z)-2-(carboxymethyl)-5-(2-hydroxyethylidene)-3-azabicyclo[3.2.0]hept-2-yl]carboxylate, 7-oxo-4-oxa-1-azabicyclo[3.2.0]hept-2-yl]carboxylate, 4-carboxylic acid,



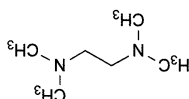
F. 4-[[[4-(2-hydroxyethyl)-1H-pyrrrole-3-yl]carboxyl]oxy]methyl]-1H-pyrrrole-3-carboxylic acid,



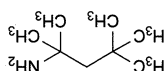
G. 4-[[[(1S)-1-carboxy-2-(4-hydroxyphenyl)ethyl]amino]-4-oxobutanoic acid (N-(hydrogensuccinyl)tyrosine),



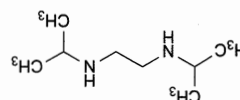
H. 2-methylpropan-2-amine (2-amino-2-methylpropane, tert-butylamine, ethyldimethylamine),



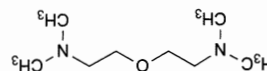
J. N,N,N',N'-tetraethylethane-1,2-diamine (1,2-bis(dimethylamino)ethane, N,N,N',N'-tetraethylethylenediamine),



K. 2,4,4-trimethylpentan-2-amine (2-amino-2,4,4-trimethylpentane, 1,1,3,3-tetramethylbutylamine),



L, N,N'-diisopropylethane-1,2-diamine (N,N'-bis(1-methylethyl)ethane-1,2-diamine),



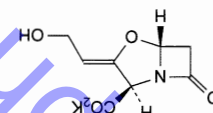
M, 2,2'-oxybis(N,N'-dimethylethanamine) (bis[2-(dimethylamino)ethoxy] ether, N,N,N',N'-tetramethyl(oxydiethylene)diamine).

Ph Eur

Diluted Potassium Clavulanate



(Ph. Eur. monograph 1653)

C₈H₈KNO₅ 237.3

Action and use

Beta-lactamase inhibitor, potentiation of the action of amoxicillin and ticarcillin.

Preparations

Co-amoxiclav Oral Suspension
Co-amoxiclav Tablets
Dispersible Co-amoxiclav Tablets

Ph Eur

DEFINITION

Dry mixture of Potassium clavulanate (1140) and Cellulose, microcrystalline (0316) or Silica, colloidal anhydrous (0434) or Silica, colloidal hydrated (0738).

Content

91.2 per cent to 107.1 per cent of the content of potassium clavulanate stated on the label.

CHARACTERS

Appearance of diluted potassium clavulanate White or almost white powder, hygroscopic.
Solubility of potassium clavulanate Freely soluble in water, slightly soluble in ethanol (96 per cent), very slightly soluble in acetone.
The solubility of the diluted product depends on the diluent and its concentration.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.
Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).
B. It gives reaction (b) of potassium (2.3.1).
C. Depending on the diluent used, carry out the corresponding identification test (a) or (b).

(a) A quantity of the substance to be examined, corresponding to 20 mg of cellulose, when placed on a watch-glass and dispersed in 4 mL of iodinated zinc chloride solution R, becomes violet-blue.
(b) It gives the reaction of silicates (2.3.1).
TESTS
pH (2.2.3) 4.8 to 8.0.
Suspend a quantity of the substance to be examined corresponding to 0.200 g of potassium clavulanate in 20 mL of carbon dioxide-free water R.
Polymetric impurities and other impurities absorbing at 278 nm
Disperse a quantity of the substance to be examined corresponding to 50.0 mg of potassium clavulanate in 10 mL of 0.1 M phosphate buffer solution pH 7.0 R, dilute to 50.0 mL with the same buffer solution and filter. Measure the absorbance immediately.
The absorbance (2.2.25) of the solution determined at 278 nm is not greater than 0.40.
Related substances
Liquid chromatography (2.2.29). Prepare the solutions immediately before use.
Test solution Disperse a quantity of the substance to be examined corresponding to 0.250 g of potassium clavulanate in 5 mL of mobile phase A, dilute to 25.0 mL with mobile phase A and filter.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.
Reference solution (b) Dissolve 10 mg of amoxicillin trihydrate CRS in 1 mL of the test solution and dilute to 100 mL with mobile phase A.
Reference solution (c) 2 mg of potassium clavulanate impurity G CRS in 20 mL of mobile phase A.
Column:
— size: $l = 0.10$ m, $\phi = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 40 °C.
— mobile phase A: a 7.8 g/L solution of sodium dihydrogen phosphate R adjusted to pH 4.0 with dilute phosphoric acid R;
— mobile phase B: a mixture of equal volumes of mobile phase A and methanol R;
Time (min)
Mobile phase A (per cent V/V) 100
Mobile phase B (per cent V/V) 0
15 - 18 4 - 15 0 - 4
50 100 $50 \rightarrow 50$
50 0 50

Flow rate 1 mL/min.
Detection Spectrophotometer at 230 nm.
Injection 20 μ L.
Relative retention With reference to clavulanate (retention time = about 3 min): impurity B = about 2.3; impurity G = about 3.6.
System suitability: reference solution (b):
— resolution: minimum 1.3 between the peaks due to clavulanate (1st peak) and amoxicillin (2nd peak).

— *impurities E, G*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— *any other impurity*: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.2 per cent);

— *total*: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a)

(2.0 per cent);

— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Water (2.5.12)

Maximum 2.5 per cent, determined on 1.000 g.

ASSAY

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Disperse a quantity of the substance to be examined corresponding to 50.0 mg of potassium clavulanate

in a 4.1 g/L solution of sodium acetate R previously adjusted to pH 6.0 with glacial acetic acid R, dilute to 50.0 mL with the same solution and filter.

Reference solution (a) Dissolve 50.0 mg of lithium

clavulanate CRS in a 4.1 g/L solution of sodium acetate R previously adjusted to pH 6.0 with glacial acetic acid R and dilute to 50.0 mL with the same solution.

Reference solution (b) Dissolve 10 mg of amoxicillin

trihydrate CRS in 10 mL of reference solution (a).

Column:

— *stationary phase*: octadecylsilyl silica gel for chromatography R

(10 µm).

Mobile phase Mix 5 volumes of methanol R1 and 95 volumes of a 15 g/L solution of sodium dihydrogen phosphate R

previously adjusted to pH 4.0 with dilute phosphoric acid R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

System suitability: reference solution (b):

— *resolution*: minimum 3.5 between the peaks due to clavulanate (1st peak) and amoxicillin (2nd peak).

1 mg of C₈H₉NO₅ is equivalent to 1.191 mg of C₈H₈KNO₅.

STORAGE

In an airtight container.

LABELLING

The label states the percentage content of potassium clavulanate and the diluent used to prepare the mixture.

IMPURITIES

Specified impurities E, G

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, B, C, D, F.

Potassium Dihydrogen Phosphate

(Ph. Eur. monograph 0920)
KH₂PO₄ 136.1
7778-77-0

Preparations
Sterile Potassium Dihydrogen Phosphate Concentrate
Potassium Dihydrogen Phosphate Oral Solution

DEFINITION

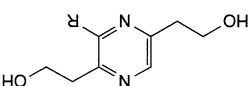
Content

98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

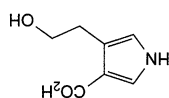
White or almost white, crystalline powder or colourless crystals.



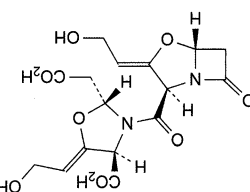
A. R = H: 2,2'-(pyrazine-2,5-diyl)diethanol,

B. R = CH₂-CH₂-CO₂H: 3-[3,6-bis(2-hydroxyethyl)pyrazin-2-yl]propanoic acid,

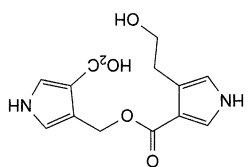
C. R = CH₂-CH₃: 2,2'-(3-ethylpyrazine-2,5-diyl)diethanol,



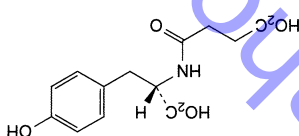
D. 4-(2-hydroxyethyl)-1H-pyrazole-3-carboxylic acid,



E. (2R,4R,5Z)-2-(carboxymethyl)-5-(2-hydroxyethylidene)-3-[[[(2R,3Z,5R)-3-(2-hydroxyethylidene)-7-oxo-4-oxa-1-azabicyclo[3.2.0]hept-2-ylidene]carbonyl]oxazolidine-4-carboxylic acid,



F. 4-[[[4-(2-hydroxyethyl)-1H-pyrazol-3-yl]carbonyl]oxy]methyl-1H-pyrazole-3-carboxylic acid,



G. 4-[[[4-(2-hydroxyethyl)-1H-pyrazol-3-yl]carbonyl]oxy]methyl-1H-pyrazole-3-carboxylic acid, (N-(hydrogensuccinyl)tyrosine).

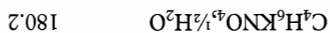
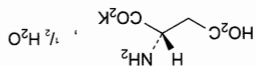
Ph Eur

Ph Eur



Potassium Hydrogen Aspartate Hemihydrate

(Ph. Eur. monograph 2076)



180.2

Action and use

Excipient.

Ph Eur

DEFINITION

Potassium hydrogen (2S)-2-aminobutanedioate hemihydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, powder or crystalline powder, or

Solubility

Very soluble in water, practically insoluble in alcohol and in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Examine the chromatograms obtained in the test for

ninhydrin-positive substances.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. It gives reaction (b) of potassium (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

6.0 to 7.5 for solution S.

Specific optical rotation (2.2.7)

+ 18.0 to + 20.5 (anhydrous substance).

Dissolve 0.50 g in a mixture of equal volumes of hydrochloric acid R and water R and dilute to 25.0 mL with the same mixture of solvents.

Ninhydrin-positive substances

Thin-layer chromatography (2.2.27).

Test solution (a) Solution S.

Test solution (b) Dilute 1.0 mL of solution S to 10.0 mL with water R.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is faintly acid (2.2.4).

B. Solution S gives reaction (b) of phosphates (2.3.1).

C. 0.5 mL of solution S gives reaction (b) of potassium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.2 to 4.5.

To 5 mL of solution S add 5 mL of carbon dioxide-free water R.

Reducing substances

To 5 mL of solution S add 5 mL of dilute sulfuric acid R and 0.25 mL of 0.02 M potassium permanganate. Heat on a water-bath for 5 min. The colour of the permanganate is not completely discharged.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.

To 5 mL of solution S add 0.5 mL of hydrochloric acid R and dilute to 15 mL with distilled water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Iron (2.4.9)

Maximum 10 ppm, determined on solution S.

Sodium

Maximum 0.1 per cent, if intended for use in the manufacture of parenteral preparations.

Atomic emission spectrometry (2.2.22, Method I).

Test solution Dissolve 1.00 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using the following solution, diluted as necessary with water R: dissolve 0.5084 g of sodium chloride R, previously dried at 100-105 °C for 3 h, in water R and dilute to 1000.0 mL with the same solvent (200 µg of Na per millilitre).

Wavelength 589 nm.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 2.0 per cent, determined on 1.000 g by drying in an oven at 125-130 °C.

ASSAY

Dissolve 1.000 g in 50 mL of carbon dioxide-free water R. Titrate with carbonate-free 1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

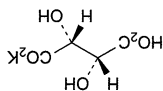
1 mL of 1 M sodium hydroxide is equivalent to 0.1361 g of KH_2PO_4 .



Potassium Hydrogen Tartrate

Cream of Tartar

(Ph. Eur. monograph 1984)



868-14-4

188.2

$C_4H_5KO_6$

Action and use

Excipient.

Ph Eur

DEFINITION

Potassium hydrogen (2R,3R)-2,3-dihydroxybutane-1,4-dioate.

Content

99.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Slightly soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute solutions of mineral acids and alkali hydroxides.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Suspend 0.5 g in 50 mL of water R and boil until dissolution is complete. Allow to cool (solution A). To 5 mL of solution A, add 0.1 mL of methyl red solution R.

The solution is red.

C. Solution A gives reaction (a) of tartrates (2.3.1).

D. Solution A gives reaction (b) of potassium (2.3.1).

TESTS

Specific optical rotation (2.2.7)

+ 8.0 to + 9.2 (dried substance).

Dissolve 2.50 g in 20 mL of 1 M hydrochloric acid with heating. Allow to cool. Dilute to 25.0 mL with water R.

Oxalic acid

Maximum 500 ppm.

Dissolve 0.43 g in 4 mL of water R. Add 3 mL of hydrochloric acid R and 1 g of zinc R in granules and boil for 1 min. Allow to stand for 2 min. Collect the liquid in a test-tube containing 0.25 mL of a 10 g/L solution of phenylhydrazine hydrochloride R and heat to boiling. Cool rapidly, transfer to a graduated cylinder and add an equal volume of hydrochloric acid R and 0.25 mL of a 50 g/L solution of potassium ferricyanide R. Shake and allow to stand for 30 min. Any pink colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 1 mL of water R and 3 mL of a 0.1 g/L solution of oxalic acid R.

Chlorides (2.4.4)

Maximum 500 ppm.

Dissolve 1.0 g with heating in a mixture of 3 mL of dilute nitric acid R and 50 mL of water R. Dilute to 100 mL with water R. Dilute 10 mL of the solution to 15 mL with

Reference solution (a) Dissolve 25 mg of potassium hydrogen aspartate hemihydrate CRS in water R and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of test solution (b) to 20.0 mL with water R.

Reference solution (c) Dissolve 10 mg of glutamic acid CRS and 10 mg of the substance to be examined in water R and dilute to 25 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 100-105 °C for 15 min.

System suitability: reference solution (c):

— the chromatogram shows 2 clearly separated principal spots.

Limits: test solution (a):

— any impurity: any spot, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

To 10 mL of solution S add 5 mL of water R.

Sulfates (2.4.13)

Maximum 500 ppm.

To 12 mL of solution S add 3 mL of distilled water R.

Ammonium (2.4.1, Method B)

Maximum 200 ppm, determined on 50 mg.

Prepare the standard using 0.1 mL of ammonium standard solution (100 ppm NH_4^+) R.

Iron (2.4.9)

Maximum 30 ppm.

In a separating funnel, dissolve 0.33 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. The aqueous layer complies with the limit test for iron.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution

Water (2.5.12)

4.0 per cent to 6.0 per cent, determined on 0.200 g.

Dissolve the substance to be examined in 10 mL of formamide R1 and add 10 mL of anhydrous methanol R.

ASSAY

Dissolve 70.0 mg in 5 mL of anhydrous formic acid R, add 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 8.56 mg of $C_4H_5KNO_4$.

Ph Eur

Sulfates (2.4.13)

Suspend 0.30 g in 3.0 mL of dilute hydrochloric acid R and dilute to 15 mL with distilled water R. Heat until dissolution is complete.

Barium

Suspend 0.50 g in a mixture of 1.5 mL of dilute hydrochloric acid R and 8.5 mL of water R. Heat until dissolution is complete. Allow to cool. Add 1 mL of dilute sulfuric acid R. The solution remains clear (2.2.1) on standing for 15 min.

Heavy metals (2.4.8)

Maximum 10 ppm.
2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.170 g in 100 mL of water R at 100 °C. Titrate the hot solution with 0.1 M sodium hydroxide, using 0.3 mL of phenolphthalein solution R as indicator.

1 mL of 0.1 M sodium hydroxide is equivalent to 18.82 mg of $C_4H_3KO_6$.

Potassium Hydroxide

Caustic Potash

(Ph. Eur. monograph 0840)

KOH

Potassium Ascorbate Eye Drops

Potassium Hydroxide Solution

Ph Eur

DEFINITION

Content
85.0 per cent to 100.5 per cent of total alkali, calculated as KOH.

CHARACTERS

Appearance
White or almost white, crystalline, hard masses, supplied as sticks, pellets or irregularly shaped pieces, deliquescent, hygroscopic, absorbing carbon dioxide.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).
A. pH (2.2.3): minimum 10.5.

IDENTIFICATION

Dissolve 0.1 g in 10 mL of water R (solution A used for identification test B). Dilute 1 mL of this solution to 100 mL with water R.
B. 1 mL of solution A prepared in identification test A gives reaction (b) of potassium (2.3.1).

TESTS

Solution S1
Dissolve 2.5 g in 10 mL of water R. Carefully add 2 mL of nitric acid R while cooling, and dilute to 25 mL with dilute nitric acid R.

Solution S2

Dissolve 10 g in 15 mL of distilled water R. Carefully add 12 mL of hydrochloric acid R while cooling, and dilute to 50 mL with dilute hydrochloric acid R.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Carbonates

Maximum 2.0 per cent, calculated as K_2CO_3 as determined in the assay.

Chlorides (2.4.4)

Maximum 50 ppm.

Dilute 10 mL of solution S1 to 15 mL with water R.

Phosphates (2.4.11)

Maximum 20 ppm.

Dilute 5 mL of solution S1 to 100 mL with water R.

Sulfates (2.4.13)

Maximum 50 ppm, determined on solution S2.

Aluminium (2.4.17)

Maximum 0.2 ppm, if intended for use in the manufacture of haemodialysis solutions.

Prescribed solution Dissolve 20 g in 100 mL of water R and add 10 mL of acetate buffer solution pH 6.0 R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R.

Iron (2.4.9)

Maximum 10 ppm.

Dilute 5 mL of solution S2 to 10 mL with water R.

Sodium

Maximum 1.0 per cent.

Atomic absorption spectrometry (2.2.23, Method II)

Test solution Dissolve 1.00 g in 50 mL of water R, add 5 mL of sulfuric acid R and dilute to 100.0 mL with water R. Dilute 1.0 mL of this solution to 10.0 mL with water R.

Reference solutions Prepare the reference solutions using sodium standard solution (200 ppm Na) R, diluted as necessary with water R.

Source Sodium hollow-cathode lamp.

Wavelength 589 nm.

Atomisation device Air-acetylene flame.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dilute 10 mL of solution S2 to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

Dissolve 2.000 g in 25 mL of carbon dioxide-free water R. Add 25 mL of freshly prepared barium chloride solution R1 and 0.3 mL of phenolphthalein solution R. Add slowly while shaking 25.0 mL of 1 M hydrochloric acid and continue the titration with 1 M hydrochloric acid until the colour changes from pink to colourless. Add 0.3 mL of bromophenol blue solution R and continue the titration with 1 M hydrochloric acid until the colour changes from violet-blue to yellow.

1 mL of 1 M hydrochloric acid used in the 2nd part of the titration is equivalent to 69.11 mg of K_2CO_3 .
1 mL of 1 M hydrochloric acid used in the combined titrations is equivalent to 56.11 mg of total alkali, calculated as KOH.
STORAGE
In an airtight, non-metallic container.
LABELLING
The label states, where applicable, that the substance is suitable for use in the manufacture of haemodialysis solutions.

Potassium Hydroxyquinoline Sulfate

Potassium Hydroxyquinoline Sulphate

Action and use

Used in treatment of acne.

Preparation

Potassium Hydroxyquinoline Sulfate and Benzoyl Peroxide Cream.

DEFINITION

Potassium Hydroxyquinoline Sulfate is an equimolecular mixture of quinolin-8-ol sulfate monohydrate,

$(C_9H_7NO)_2 \cdot H_2SO_4 \cdot H_2O$, and potassium sulfate, K_2SO_4 .

It contains not less than 50.6% and not more than 52.6% of quinolin-8-ol, C_9H_7NO , and not less than 29.5% and not more than 32.5% of K_2SO_4 , calculated with reference to the anhydrous substance.

CHARACTERISTICS

A pale yellow, microcrystalline powder. It partly liquefies between 172° and 184°.

Freely soluble in water; insoluble in ether. On extraction with hot absolute ethanol a residue of potassium sulfate and a solution of quinolin-8-ol sulfate are obtained.

IDENTIFICATION

A. To 5 mL of a 5% w/v solution add drop wise, with shaking, 5m sodium hydroxide until a heavy precipitate is produced. Filter, wash with water and dry at a pressure not exceeding 0.7 kPa for 3 hours. The infrared absorption spectrum of the residue, Appendix II A, is concordant with the reference spectrum of quinolin-8-ol (RS 310).

B. To 5 mL of a 5% w/v solution add 0.5 mL of iron(III) chloride solution R1. A dark green colour is produced.

C. Yields reaction A characteristic of potassium salts,

Appendix VI.

D. Yields reaction A characteristic of sulfates, Appendix VI.

TESTS

Water

Not more than 5.0% w/w, Appendix IX C. Use 0.5 g.

ASSAY

For quinolin-8-ol

Dissolve 0.35 g in 50 mL of water and 20 mL of hydrochloric acid, add 50 mL of 0.05M bromine VS, stopper the flask and shake for 15 minutes. Allow to stand for 15 minutes, add 80 mL of water and 10 mL of dilute potassium iodide solution and titrate with 0.1M sodium thiosulfate VS using starch mucilage, added towards the end of the titration, as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the amount

Potassium Iodate

KIO_3

7758-05-6

214.0

Action and use

Iodine supplement for emergency use.

Preparation

Potassium Iodate Tablets

DEFINITION

Potassium Iodate contains not less than 99.0% and not more than 101.0% of KIO_3 , calculated with reference to the dried substance.

CHARACTERISTICS

A white crystalline powder.

Slowly soluble in water; insoluble in ethanol (96%).

Dissolve 10 g of the substance being examined in sufficient water to produce 200 mL (solution S1).

Add 25 mL of hydrochloric acid to 6 g of the substance being examined, evaporate to dryness and repeat. Heat until iodine is removed. Dissolve the residue in 2.5 mL of a 25% v/v solution of hydrochloric acid and dilute to 50 mL with water (solution S2).

IDENTIFICATION

A. 1 mL of solution S1 yields reaction B characteristic of potassium salts, Appendix VI.

B. Dissolve 0.1 g in 5 mL of water. Add 1 mL of silver nitrate solution followed by 1 mL of sulfur dioxide solution. A yellow precipitate is produced immediately.

TESTS

Acidity or alkalinity

pH of solution S1, 5.0 to 8.0, Appendix V L.

Clarity and colour of solution

Solution S1 is clear, Appendix IV A, and colourless,

Appendix IV B, Method II.

Chloride, chlorate, bromide, bromate

Dilute 5 mL of solution S1 to 15 mL with water, add 20 mL of sulfur dioxide solution and heat on a water bath for 30 minutes. Heat to boiling, cool, add 10 mL of 18M ammonia and 20 mL of silver nitrate solution R2 and dilute to 70 mL with water. Filter, transfer 35 mL of the filtrate to a Nessler cylinder and acidify with 6 mL of nitric acid. After 5 minutes, any opalescence, when viewed vertically, is not greater than that produced by treating 5 mL of a 0.00165% w/v solution of sodium chloride at the same time and in the same manner (0.02%).

Iodide

Add 1 mL of 1.8M sulfuric acid to 25 mL of solution S1 and shake with 1 mL of chloroform. Any violet colour produced is not more intense than that of a solution prepared at the same

time and in the same manner but using 5 mL of solution S1 and 2 mL of iodide standard solution (10 ppm I) (20 ppm).

Sulfate

Add 1 mL of a 25% w/v solution of barium chloride to 1.5 mL of sulfate standard solution (10 ppm SO_4), shake and allow to stand for 1 minute. Add 12.5 mL of solution S2 diluted to 15 mL with distilled water and 0.5 mL of 5M acetic acid and allow to stand for 5 minutes. Any opalescence produced is not more intense than that of a standard prepared in the same manner but using 7.5 mL of sulfate standard solution (10 ppm SO_4) diluted to 15 mL with distilled water in place of the solution being examined (50 ppm).

Heavy metals

Adjust the pH of 20 mL of solution S2 to about 5 with 5M ammonia. The solution complies with limit test A for heavy metals, Appendix VII. Use 10 mL of lead standard solution (2 ppm Pb) to prepare the standard (20 ppm).
When dried at 130° for 1 hour, loses not more than 0.5% of its weight. Use 1 g.

ASSAY

To 1.5 g add sufficient water to produce 250 mL. To 25 mL of the resulting solution in an iodine flask add 3 g of potassium iodide, 100 mL of water and 10 mL of hydrochloric acid. Close the flask and stand in the dark for 5 minutes. Titrate the solution with 0.1M sodium thiosulfate VS to a light straw colour and then complete the titration to a colourless end point using starch mucilage as indicator. Each mL of 0.1M sodium thiosulfate VS is equivalent to 3.567 mg of KIO₃.

Potassium Iodide

(Ph. Eur. monograph 0186)

KI

166.0

7681-11-0



Action and use
Antithyroid.

DEFINITION

Content 99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder or colourless crystals.

Solubility

Very soluble in water, freely soluble in glycerol, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) gives the reactions of iodides (2.3.1).
B. Solution S gives the reactions of potassium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Potassium Metabisulfite

Potassium Metabisulfite

(Ph. Eur. monograph 2075)

K₂S₂O₅

222.3

16731-55-8



Action and use

Preservative.

DEFINITION

Potassium metabisulfite (potassium disulfite).

Content

95.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

White or almost white powder, or colourless crystals.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

STORAGE

Protected from light.

KI.

1 mL of 0.05 M potassium iodate is equivalent to 16.60 mg of KI.

until the chloroform layer is decolourised.
chloroform R and continue the titration, shaking vigorously, until the colour changes from red to yellow. Add 5 mL of hydrochloric acid R and titrate with 0.05 M potassium iodate same solvent. To 20.0 mL of the solution add 40 mL of

ASSAY

Dissolve 1.500 g in water R and dilute to 100.0 mL with the powdered substance by drying in an oven at 105° C for 3 h.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.00 g of previously reference solution using lead standard solution (1 ppm Pb) R.

12 mL of solution S complies with test A. Prepare the

Maximum 10 ppm.

Heavy metals (2.4.8)

Dilute 5 mL of solution S to 10 mL with water R.

Maximum 20 ppm.

Iron (2.4.9)

To 10 mL of solution S add 0.1 mL of starch solution R and 0.1 mL of 0.005 M iodine. A blue colour is produced.

Thiosulfates

Dilute 10 mL of solution S to 15 mL with distilled water R.

Maximum 150 ppm.

Sulfates (2.4.13)

develops.

stand protected from light for 2 min. No blue colour

solution R and 0.2 mL of dilute sulfuric acid R and allow to

To 10 mL of solution S add 0.25 mL of iodide-free starch

Iodates

acid is required to change the colour of the indicator.

solution R1. Not more than 0.5 mL of 0.01 M hydrochloric

To 12.5 mL of solution S add 0.1 mL of bromotymol blue

Alkalinity

IDENTIFICATION

- A. pH (see Tests).
 B. To 5 mL of solution S (see Tests), add 0.5 mL of 0.05 M iodine. The mixture is colourless and gives reaction (a) of sulfates (2.3.1).
 C. Solution S gives reaction (a) of potassium (2.3.1).

TESTS

Solution S
 Dissolve 5.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method I).
 pH (2.2.3)
 3.0 to 4.5 for solution S.

Thiosulfates

To 2.00 g add 25 mL of a 42.5 g/L solution of sodium hydroxide R and 75 mL of water R. Shake until dissolved and add 10 mL of formaldehyde R and 10 mL of acetic acid R. After 5 min, titrate with 0.05 M iodine using 1 mL of starch solution R. Carry out a blank titration. The difference between the volumes consumed in the 2 titrations is not more than 0.15 mL.

Iron

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method I).

Test solution Dilute 20 mL of solution S to 50 mL with water R.

Reference solutions Prepare the reference solutions using iron standard solution (20 ppm Fe) R, diluted as necessary with water R.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Selenium

Maximum 10 ppm.

To 3.0 g add 10 mL of formaldehyde R. Carefully add 2 mL of hydrochloric acid R in small portions. Heat on a water-bath for 20 min. Any pink colour in the solution is not more intense than that of a reference solution prepared at the same time in the same manner using 1.0 g of the substance to be examined to which 0.2 mL of selenium standard solution (100 ppm Se) R has been added.

Zinc

Maximum 25 ppm.

Atomic absorption spectrometry (2.2.23, Method I).

Test solution Dilute 20 mL of solution S to 50 mL with water R.

Reference solutions Prepare the reference solutions using zinc standard solution (100 ppm Zn) R, diluted as necessary with water R.

Source Zinc hollow-cathode lamp.

Wavelength 213.9 nm.

Atomisation device Air-acetylene flame.

Heavy metals (2.4.8)

Maximum 10 ppm.

Introduce 40 mL of solution S into a silica crucible, add 10 mL of hydrochloric acid R and evaporate to dryness.

Dissolve the residue in 19 mL of water R and add 1 mL of a 40 g/L solution of sodium fluoride R. The solution complies with test B. Prepare the reference solution using 20 mL of lead standard solution (1 ppm Pb) R.

Potassium Nitrate

(Ph. Eur. monograph 1465)

KNO₃ 101.1

7757-79-1

**DEFINITION****Content**

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, very soluble in boiling water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. It gives the reaction of nitrates (2.3.1).

B. Solution S (see Tests) gives the reactions of potassium (2.3.1).

TESTS**Solution S**

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.05 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Reducible substances

To 10 mL of solution S, add 0.5 mL of dilute sulfuric acid R and 2 mL of zinc iodide and starch solution R. The solution does not become blue within 2 min.

Chlorides (2.4.4)

Maximum 20 ppm, if intended for ophthalmic use.

Dissolve 2.5 g in water R and dilute to 15 mL with the same solvent.

Sulfates (2.4.13)

Maximum 150 ppm.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Ammonium (2.4.1)

Maximum 100 ppm, determined in 1 mL of solution S;

maximum 50 ppm if intended for ophthalmic use.

Calcium (2.4.3)

Maximum 100 ppm; maximum 50 ppm if intended for ophthalmic use.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 20 ppm; maximum 10 ppm if intended for

ophthalmic use.

Dilute 5 mL of solution S to 10 mL with water R.

Sodium

Maximum 0.1 per cent.

Atomic emission spectrometry (2.2.22, Method II).

Test solution Dissolve 1.00 g in water R and dilute to

100.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using sodium

standard solution (200 ppm Na) R, diluting with water R.

Wavelength 589 nm.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

ASSAY

Prepare a chromatography column 0.3 m long and 10 mm in

internal diameter and filled with 10 g of strongly acidic ion-

exchange resin R covered with carbon dioxide-free water R.

Maintain a 1 cm layer of liquid above the resin at all times.

Allow 100 mL of dilute hydrochloric acid R to run through the

column at a flow rate of about 5 mL/min. Wash the column

(with the tap completely open) with carbon dioxide-free

water R until neutral to blue litmus paper R. Dissolve 0.200 g

of the substance to be examined in 2 mL of carbon dioxide-

free water R in a beaker and transfer it to the column

reservoir, allow the solution to run through the column at a

flow rate of about 3 mL/min and collect the eluate. Wash the

beaker with 10 mL of carbon dioxide-free water R and transfer

this solution at the same flow rate to the column before it

runs dry. Finally wash the column with 200 mL of carbon

dioxide-free water R (with the tap completely open) until

neutral to blue litmus paper R. Titrate the combined eluate

and washings with 0.1 M sodium hydroxide, using 1 mL of

1 mL of 0.1 M sodium hydroxide is equivalent to 10.11 mg of

KNO₃.

LABELLING

The label states, where applicable, that the substance is

suitable for ophthalmic use.

Potassium Perchlorate

(Ph. Eur. monograph 1987)

KClO₄

138.6

7778-74-7



Action and use

Diagnostic aid; treatment of hyperthyroidism.

DEFINITION

Content

99.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

Solubility

Sparsely soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. Dissolve 0.1 g in 5 mL of water R. Add 5 mL of indigo

carmine solution R and heat to boiling. The colour of the

solution does not disappear.

B. Chlorates and chlorides (see Tests).

C. Heat 10 mg over a flame for 2 min. Dissolve the residue

in 2 mL of water R. The solution gives reaction (a) of

chlorides (2.3.1).

D. Dissolve 50 mg with heating in 5 mL of water R. Allow to

cool to room temperature. The solution gives reaction (a) of

TESTS

Solution S

Suspend 5.0 g in 90 mL of distilled water R and heat to

boiling. Allow to cool. Filter. Dilute the filtrate to 100 mL

with carbon dioxide-free water R.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dissolve 0.20 g in water R and dilute to 20 mL with the

same solvent.

Acidity or alkalinity

To 5 mL of solution S add 5 mL of water R and 0.1 mL of

phenolphthalein solution R. Not more than 0.25 mL of 0.01 M

sodium hydroxide is required to change the colour of the

indicator. To 5 mL of solution S, add 5 mL of water R and

0.25 mL of 0.01 M hydrochloric acid is required to change the

colour of the indicator.

Chlorates and chlorides (2.4.4)

Maximum 100 ppm (calculated as chlorides).

To 5 mL of solution S, add 5 mL of water R and heat to

boiling. Add 1 mL of nitric acid R and 0.1 g of sodium

nitrite R. Allow to cool to room temperature. Dilute to

15 mL with water R. The solution complies with the limit

test for chlorides. Prepare the standard using 5 mL of chloride

standard solution (5 ppm Cl) R and 10 mL of water R, and

adding only 1 mL of dilute nitric acid R.

Sulfates (2.4.13)

Maximum 100 ppm, determined on solution S.

Prepare the standard using a mixture of 7.5 mL of sulfate

standard solution (10 ppm SO₄) R and 7.5 mL of water R.

Calcium (2.4.3)

Maximum 100 ppm, determined on solution S.

Prepare the standard using a mixture of 7.5 mL of calcium

standard solution (10 ppm Ca) R, 1 mL of dilute acetic acid R

and 7.5 mL of distilled water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

Prepare a chromatography column 0.3 m long and 10 mm in

internal diameter and filled with 10 g of strongly acidic ion-

exchange resin R covered with carbon dioxide-free water R. Maintain a 1 cm layer of liquid above the resin throughout the determination. Allow 100 mL of dilute hydrochloric acid R to run through the column at a flow rate of about 5 mL/min. Wash the column (with the tap completely open) with carbon dioxide-free water R until the eluate is neutral to blue litmus paper R. Dissolve 0.100 g of the substance to be examined in 10 mL of carbon dioxide-free water R in a beaker and transfer it to the column reservoir, allow the solution to run through the column at a flow rate of about 3 mL/min and collect the eluate. Wash the beaker 3 times with 10 mL of carbon dioxide-free water R and transfer this solution at the same flow rate to the column before it runs dry. Finally, wash the column with 200 mL of carbon dioxide-free water R (with the tap completely open) until the eluate is neutral to blue litmus paper R. Titrate the combined eluate and washings with 0.1 M sodium hydroxide, using 1 mL of phenolphthalein solution R as indicator.

1 mL of 0.1 M sodium hydroxide is equivalent to 13.86 mg of KClO_4 .

Ph Eur

Potassium Permanganate

(Ph. Eur. monograph 0121)

KMnO_4 158.0 7722-64-7

Action and use
Antiseptic.

Content
99.0 per cent to 100.5 per cent.

CHARACTERS

Appearance

Dark purple or brownish-black, granular powder or dark purple or almost black crystals, usually having a metallic lustre.

Solubility

Soluble in cold water, freely soluble in boiling water.

It decomposes on contact with certain organic substances.

IDENTIFICATION

A. Dissolve about 50 mg in 5 mL of water R and add 1 mL of ethanol (96 per cent) R and 0.3 mL of dilute sodium hydroxide solution R. A green colour develops. Heat to boiling. A dark brown precipitate is formed.

B. Filter the mixture obtained in identification test A. The filtrate gives reaction (b) of potassium (2.3.1).

TESTS

Solution S

Dissolve 0.75 g in 25 mL of distilled water R, add 3 mL of ethanol (96 per cent) R and boil for 2-3 min. Cool, dilute to 30 mL with distilled water R and filter.

Appearance of solution

Solution S is colourless (2.2.2, Method II).

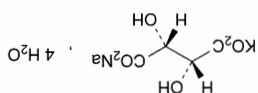
Substances insoluble in water

Maximum 1.0 per cent.

Dissolve 0.5 g in 50 mL of water R and heat to boiling. Filter through a tared sintered-glass filter (16) (2.1.2). Wash with water R until the filtrate is colourless and collect the residue

Potassium Sodium Tartrate

(Ph. Eur. monograph 1986)



$\text{C}_4\text{H}_4\text{KNaO}_6 \cdot 4\text{H}_2\text{O}$ 282.2

Action and use

Excipient.

Ph Eur

DEFINITION

Potassium sodium (+)-(2R,3R)-2,3-dihydroxybutanedioate tetrahydrate.

Content

98.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless, transparent crystals.

Solubility

Very soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. It gives reaction (b) of tartrates (2.3.1).

C. It gives reaction (b) of potassium (2.3.1).

D. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

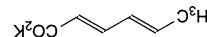
Dissolve 5.000 g in carbon dioxide-free water R, prepared from distilled water R, and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Potassium Sorbate

(Ph. Eur. monograph 0618)


 $C_6H_7KO_2$ 150.2 590-00-1

Action and use

Antimicrobial preservative.

DEFINITION

Potassium (E,E)-hexa-2,4-dienoate.

Ph Eur



To 0.100 g of finely powdered substance add 40 mL of anhydrous acetic acid R and 20 mL of acetic anhydride R. Titrate slowly with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 10.51 mg of $C_6H_7KNaO_6$.

ASSAY

Use 50 mL of anhydrous methanol R. Titrate slowly. 24.0 per cent to 26.5 per cent, determined on 50.0 mg.

Water (2.5.12)

(1 ppm Pb) R

the reference solution using lead standard solution
Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare
Maximum 10 ppm.

Heavy metals (2.4.8)

Dilute 10 mL of solution S to 15 mL with distilled water R.

Maximum 200 ppm.

Calcium (2.4.3)

of calcium sulfate solution R and 5 mL of distilled water R.
solution is not more intense than that in a mixture of 3 mL of calcium sulfate solution R. Allow to stand for 5 min. Any opalescence in the
To 5 mL of solution S, add 3 mL of calcium sulfate

Barium and oxalates

Maximum 40 ppm, determined on 5 mL of solution S.

Ammonium (2.4.1)

and 10 mL of distilled water R.
mixture of 5 mL of sulfate standard solution (10 ppm SO_4) R
the same solvent. Prepare the reference solution with a
Dissolve 1.0 g in distilled water R and dilute to 15 mL with
Maximum 50 ppm.

Sulfates (2.4.13)

Dilute 10 mL of solution S to 15 mL with water R.

Maximum 100 ppm.

Chlorides (2.4.4)

solution S.

+ 28.0 to + 30.0 (anhydrous substance), determined on

Specific optical rotation (2.2.7)

of the indicator.
or 0.01 M sodium hydroxide is required to change the colour
solution R. Not more than 0.5 mL of 0.01 M hydrochloric acid
To 5 mL of solution S, add 0.1 mL of phenolphthalein

Acidity or alkalinity

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder or granules.

Solubility

Very soluble in water, slightly soluble in ethanol

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in water R and dilute to

250.0 mL with the same solvent. Dilute 2.0 mL of this

solution to 200.0 mL with 0.1 M hydrochloric acid.

Spectral range 230-350 nm.

Absorption maximum At 264 nm.

Specific absorbance at the absorption maximum 1650 to 1900.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison potassium sorbate CRS.

C. Dissolve 1.0 g in 50 mL of water R, add 10 mL of dilute

hydrochloric acid R and shake. Filter the crystalline precipitate,

wash with water R and dry in vacuo over sulfuric acid R for

4 h. The residue obtained melts (2.2.14) at 132 °C to

136 °C.

D. Dissolve 0.2 g in 2 mL of water R and add 2 mL of dilute

acetic acid R. Filter. The solution gives reaction (b) of

potassium (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to

50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution Y₅ (2.2.2, Method II).

Acidity or alkalinity

To 20 mL of solution S add 0.1 mL of phenolphthalein

solution R. Not more than 0.25 mL of 0.1 M sodium hydroxide

or 0.1 M hydrochloric acid is required to change the colour of

Aldehydes

Maximum 0.15 per cent, expressed as C_2H_4O .

Dissolve 1.0 g in a mixture of 30 mL of water R and 50 mL

of 2-propanol R, adjust to pH 4 with 1 M hydrochloric acid

and dilute to 100 mL with water R. To 10 mL of the

solution add 1 mL of decolorised fuchsin solution R and allow

to stand for 30 min. Any colour in the solution is not more

intense than that in a standard prepared at the same time by

adding 1 mL of decolorised fuchsin solution R to a mixture of

1.5 mL of acetaldehyde standard solution (100 ppm C_2H_4O) R,

4 mL of 2-propanol R and 4.5 mL of water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

ASSAY

Dissolve 0.120 g in 20 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid using 0.1 mL of crystal violet solution R as indicator until the colour changes from violet to bluish-green.

1 mL of 0.1 M perchloric acid is equivalent to 15.02 mg of $C_6H_7KO_2$.

STORAGE

Protected from light.

Potassium Sulfate

Potassium Sulphate

(Ph. Eur. monograph 1622)

K_2SO_4 174.3

7778-80-5



Ph Eur

DEFINITION

Content 98.5 per cent to 101.0 per cent of K_2SO_4 (dried substance).

CHARACTERS

Appearance White or almost white, crystalline powder or colourless crystals.

Solubility

Soluble in water, practically insoluble in ethanol.

IDENTIFICATION

A. It gives the reactions of sulfates (2.3.1).
B. It gives the reactions of potassium (2.3.1).

TESTS

Solution S Dissolve 10.0 g in 90 mL of carbon dioxide-free water R and dilute to 100 mL with carbon dioxide-free water R prepared from distilled water R, heating gently. Allow to cool and dilute to 100 mL with distilled water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Chlorides (2.4.4)

Maximum 40 ppm.

Dilute 12.5 mL of solution S to 15 mL with water R.

Calcium (2.4.3)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 10 ppm, determined on 10 mL of solution S.

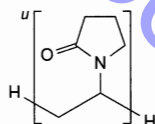
Magnesium

Maximum 20 ppm.

To 5 mL of solution S add 5 mL of water R, 1 mL of glycerol (85 per cent) R, 0.15 mL of titan yellow solution R, 0.25 mL of ammonium oxalate solution R and 5 mL of dilute sodium hydroxide solution R and shake. Any pink colour in the test solution is not more intense than that in a standard prepared at the same time and in the same manner using a mixture of

Povidone

(Ph Eur monograph 0685)



$C_{6m}H_{9m+2}N_mO_m$

Action and use

Disinfectant.

Preparations

Povidone-Iodine Eye Drops
Povidone-Iodine Mouthwash
Povidone-Iodine Solution

DEFINITION

It consists of linear polymers of 1-ethenylpyrrolidin-2-one. α -Hydro- ω -hydroxypoly[1-(2-oxopyrrolidin-1-yl)ethylene].

Content

11.5 per cent to 12.8 per cent of nitrogen (N; A_r 14.01)

(anhydrous substance).

The different types of povidone are characterised by their viscosity in solution expressed as a K-value.

CHARACTERS

Appearance White or yellowish-white, hygroscopic powder or flakes.



Ph Eur

ASSAY Dissolve 0.150 g in 40 mL of water R. Add 0.2 mL of 0.1 M hydrochloric acid and 80 mL of methanol R. Carry out a potentiometric titration (2.2.20), using 0.1 M lead nitrate and as indicator electrode a lead-selective electrode and as reference electrode a silver-silver chloride electrode.

1 mL of 0.1 M lead nitrate is equivalent to 17.43 mg of K_2SO_4 .

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 4 h.

Loss on drying (2.2.32)

reference solution using lead standard solution (2 ppm Pb) R.

12 mL of solution S complies with test A. Prepare the

Maximum 20 ppm.

Heavy metals (2.4.8)

Wavelength 589 nm.

millilitre). Dilute as required.

to 1000.0 mL with the same solvent (200 µg of Na per chloride R, previously dried at 100-105 °C for 3 h, and dilute in water R and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 1.00 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Atomic emission spectrometry (2.2.22, Method I).

Maximum 0.10 per cent.

Sodium

9 mL of water R.

1 mL of magnesium standard solution (10 ppm Mg) R and

Solubility

Freely soluble in water, in ethanol (96 per cent) and in methanol, very slightly soluble in acetone.

IDENTIFICATION

First identification: A, E.

Second identification: B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dry the substances beforehand at 105 °C for 6 h; record the spectra using 4 mg of substance.

Comparison povidone CRS.

B. To 0.4 mL of solution S1 (see Tests) add 10 mL of water R, 5 mL of dilute hydrochloric acid R and 2 mL of potassium dichromate solution R. An orange-yellow precipitate is formed.

C. To 1 mL of solution S1 add 0.2 mL of dimethylaminobenzaldehyde solution R1 and 0.1 mL of sulfuric acid R. A pink colour is produced.

D. To 0.1 mL of solution S1 add 5 mL of water R and 0.2 mL of 0.05 M iodine. A red colour is produced.

E. To 0.5 g add 10 mL of water R and shake. The substance dissolves.

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent. Add the substance to be examined to the water in small portions, stirring using a magnetic stirrer.

Solution S1

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent. Add the substance to be examined to the water in small portions, stirring using a magnetic stirrer.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₆, BY₆ or R₆ (2.2.2, Method II).

pH (2.2.3)

3.0 to 5.0 for solution S, for povidone having a stated K-value of not more than 30; 4.0 to 7.0 for solution S, for povidone having a stated K-value of more than 30.

Viscosity, expressed as K-value

For povidone having a stated value of 18 or less, use a 50 g/L solution. For povidone having a stated value of more than 18 and not more than 95, use a 10 g/L solution. For povidone having a stated value of more than 95, use a 1.0 g/L solution. Allow to stand for 1 h and determine the viscosity (2.2.9) of the solution at 25 °C, using a size no. 1 viscometer with a minimum flow time of 100 s. Calculate the K-value using the following expression:

$$K = \frac{1.5 \log_{10} \eta - 1}{\sqrt{300c \log_{10} \eta + (c + 1.5c \log_{10} \eta)^2}} + \frac{0.15 + 0.003c}{0.15c + 0.003c^2}$$

c = concentration of the substance to be examined, calculated with reference to the anhydrous substance, in grams per 100 mL;

η = kinematic viscosity of the solution relative to that of water R.

The K-value of povidone having a stated K-value of 15 or less is 85.0 per cent to 115.0 per cent of the stated value.

The K-value of povidone having a stated K-value or a stated K-value range with an average of more than 15 is examined equivalent to 2.0 g of the anhydrous substance in

Aldehydes

average of the stated range.

90.0 per cent to 108.0 per cent of the stated value or of the

Maximum 500 ppm, expressed as acetaldehyde.

Test solution Dissolve 1.0 g of the substance to be examined in phosphate buffer solution pH 9.0 R and dilute to 100.0 mL with the same solvent. Stopper the flask tightly and heat at 60 °C for 1 h. Allow to cool to room temperature.

Reference solution Dissolve 0.140 g of acetaldehyde ammonia trimer trihydrate R in water R and dilute to 200.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with phosphate buffer solution pH 9.0 R.

Into 3 identical spectrophotometric cells with a path length of 1 cm, introduce separately 0.5 mL of the test solution, 0.5 mL of the reference solution and 0.5 mL of water R (blank). To each cell add 2.5 mL of phosphate buffer solution pH 9.0 R and 0.2 mL of nicotinamide-adenine dinucleotide solution R. Mix and stopper tightly. Allow to stand at 22 ± 2 °C for 2–3 min and measure the absorbance (2.2.25) of each solution at 340 nm, using water R as the compensation liquid.

each solution at 340 nm using water R as the compensation stand at 22 ± 2 °C for 5 min. Measure the absorbance of dehydrogenase solution R, mix and stopper tightly. Allow to compensation liquid. To each cell add 0.05 mL of aldehyde solution R. Mix and stopper tightly. Allow to stand at 22 ± 2 °C for 2–3 min and measure the absorbance (2.2.25) of each solution at 340 nm, using water R as the compensation liquid.

Calculate the content of aldehydes using the following expression:

$$\frac{(A_{t2} - A_{t1}) - (A_{b2} - A_{b1})}{(A_{s2} - A_{s1}) - (A_{b2} - A_{b1})} \times \frac{100\,000}{m \times C}$$

A_{t1} = absorbance of the test solution before the addition of aldehyde dehydrogenase;

A_{t2} = absorbance of the test solution after the addition of aldehyde dehydrogenase;

A_{s1} = absorbance of the reference solution before the addition of aldehyde dehydrogenase;

A_{s2} = absorbance of the reference solution after the addition of aldehyde dehydrogenase;

A_{b1} = absorbance of the blank before the addition of aldehyde dehydrogenase;

A_{b2} = absorbance of the blank after the addition of aldehyde dehydrogenase;

m = mass of povidone calculated with reference to the anhydrous substance, in grams;

C = concentration of acetaldehyde in the reference solution, calculated from the weight of the acetaldehyde ammonia trimer trihydrate with the factor 0.172, in milligrams per millilitre.

Peroxides

Maximum 400 ppm, expressed as H₂O₂.

Dissolve a quantity of the substance to be examined equivalent to 4.0 g of the anhydrous substance in water R and dilute to 100.0 mL with the same solvent (stock solution). To 25.0 mL of the stock solution add 2.0 mL of titanium trichloride-sulfuric acid reagent R. Allow to stand for 30 min. The absorbance (2.2.25) of the solution, measured at 405 nm using a mixture of 25.0 mL of the stock solution and 2.0 mL of a 13 per cent V/V solution of sulfuric acid R as the compensation liquid, is not greater than 0.35.

Formic acid

Liquid chromatography (2.2.29).

Test solution Dissolve a quantity of the substance to be examined equivalent to 2.0 g of the anhydrous substance in

- water R and dilute to 100.0 mL with the same solvent (test stock solution). Transfer a suspension of strongly acidic ion-exchange resin R for column chromatography in water R to a column of about 0.8 cm in internal diameter to give a packing of about 20 mm in length and keep the strongly acidic ion-exchange resin layer constantly immersed in water R. Pour 5 mL of water R and adjust the flow rate so that the water drops at a rate of about 20 drops per minute. When the level of the water comes down to near the top of the strongly acidic ion-exchange resin layer, put the test stock solution into the column. After dropping 2 mL of the solution, collect 1.5 mL of the solution and use this solution as the test solution.
- Reference solution** Dissolve 0.100 g of anhydrous formic acid R in water R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with water R.
- Column:**
— size: $l = 0.25$ m, $\phi = 4.8$ mm;
— stationary phase: strongly acidic ion-exchange resin R for column chromatography (5–10 μm);
— temperature: 30 °C.
- Mobile phase** Dilute 5 mL of perchloric acid R to 1000 mL with water R.
- Flow rate** Adjusted so that the retention time of formic acid is about 11 min.
- Detection** Spectrophotometer at 210 nm.
- Injection** 50 μL .
- System suitability:** reference solution:
— **repeatability:** maximum relative standard deviation of 2.0 per cent after 6 injections.
Limit:
— **formic acid:** not more than 10 times the area of the principal peak in the chromatogram obtained with the reference solution (0.5 per cent).
- Hydrazine**
Thin-layer chromatography (2.2.27). Use freshly prepared solutions.
- Test solution** Dissolve a quantity of the substance to be examined equivalent to 2.5 g of the anhydrous substance in 25 mL of water R. Add 0.5 mL of a 50 g/L solution of salicylaldehyde R in methanol R, mix and heat in a water-bath at 60 °C for 15 min. Allow to cool, add 2.0 mL of toluene R, shake for 2 min and centrifuge. Use the upper layer of the mixture.
- Reference solution** Dissolve 90 mg of salicylaldehyde azine R in toluene R and dilute to 100 mL with the same solvent. Dilute 1 mL of the solution to 100 mL with toluene R.
- Plate** TLC silanised silica gel F₂₅₄ plate R.
- Mobile phase** water R, methanol R (1:2 V/V).
- Application** 10 μL .
- Development** Over 3/4 of the plate.
- Drying** In air.
- Detection** Examine in ultraviolet light at 365 nm.
- Retardation factor** Salicylaldehyde azine = about 0.3.
- Limit:**
— **hydrazine:** any spot due to salicylaldehyde azine is not more intense than the spot in the chromatogram obtained with the reference solution (1 ppm).
- Impurity A**
Liquid chromatography (2.2.29).
- Test solution** Dissolve a quantity of the substance to be examined equivalent to 0.250 g of the anhydrous substance
- in the mobile phase and dilute to 10.0 mL with the mobile phase.
- Reference solution (a)** Dissolve 50.0 mg of 1-vinylpyrrolidin-2-one R (impurity A) in methanol R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with methanol R. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.
- Reference solution (b)** Dissolve 10 mg of 1-vinylpyrrolidin-2-one R and 0.5 g of vinyl acetate R in methanol R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.
- Precolumn:**
— size: $l = 0.025$ m, $\phi = 4$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μm).
- Column:**
— size: $l = 0.25$ m, $\phi = 4$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μm);
— temperature: 40 °C.
- Mobile phase** acetonitrile R, water R (10:90 V/V).
- Flow rate** Adjusted so that the retention time of impurity A is about 10 min.
- Detection** Spectrophotometer at 235 nm.
- Injection** 50 μL ; after injection of the test solution, wait for about 2 min and wash the precolumn by passing the mobile phase through the column backwards for 30 min at the same flow rate as applied in the test.
- System suitability:**
— **resolution:** minimum 2.0 between the peaks due to impurity A and vinyl acetate in the chromatogram obtained with reference solution (b);
— **repeatability:** maximum relative standard deviation of 2.0 per cent after 6 injections of reference solution (a).
- Limit:**
— **impurity A:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (10 ppm).
- Impurity B**
Liquid chromatography (2.2.29).
- Test solution** Dissolve a quantity of the substance to be examined equivalent to 0.100 g of the anhydrous substance in water R and dilute to 50.0 mL with the same solvent.
- Reference solution** Dissolve 0.100 g of 2-pyrrolidone R (impurity B) in water R and dilute to 100.0 mL with the same solvent. Dilute 3.0 mL of the solution to 50.0 mL with water R.
- Precolumn:**
— size: $l = 0.025$ m, $\phi = 3$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μm).
- Column:**
— size: $l = 0.25$ m, $\phi = 3$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μm);
— temperature: 30 °C.
- Mobile phase** water R adjusted to pH 2.4 with phosphoric acid R.
- Flow rate** Adjusted so that the retention time of impurity B is about 11 min.
- Detection** Spectrophotometer at 205 nm.
- Injection** 50 μL ; after each injection of the test solution, wash away the polymeric material of povidone from the precolumn

by passing the mobile phase through the column backwards for about 30 min at the same flow rate as applied in the test.
System suitability: reference solution.
— repeatability: maximum relative standard deviation of 2.0 per cent after 6 injections.
Limit:

— impurity B: not more than the area of the principal peak in the chromatogram obtained with the reference solution (3.0 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2.0 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Place 0.100 g of the substance to be examined (*m* mg) in a combustion flask and add 5 g of a mixture of 1 g of copper sulfate R, 1 g of titanium dioxide R and 33 g of dipotassium sulfate R, and 3 glass beads. Wash any adhering particles from the neck into the flask with a small quantity of water R. Add 7 mL of sulfuric acid R, allowing it to run down the

insides of the flask. Heat the flask gradually until the solution has a clear, yellowish-green colour, and the inside wall of the flask is free from any carbonised material, and then heat for a further 45 min. After cooling, add cautiously 20 mL of water R, and connect the flask to the distillation apparatus which has been previously washed by passing steam through it. To the absorption flask add 30 mL of a 40 g/L solution of boric acid R, 3 drops of bromocresol green-methyl red solution R and sufficient water to immerse the lower end of the condenser tube. Add 30 mL of strong sodium hydroxide solution R through the funnel, rinse the funnel cautiously with 10 mL of water R, immediately close the clamp on the rubber tube, then start distillation with steam to obtain 80–100 mL of distillate. Remove the absorption flask from the lower end of the condenser tube, rinsing the end part with a small quantity of water R, and titrate the distillate with 0.025 *M* sulfuric acid until the colour of the solution changes from green through pale greyish blue to pale greyish reddish-purple. Carry out a blank determination.

1 mL of 0.025 *M* sulfuric acid is equivalent to 0.7004 mg of N.

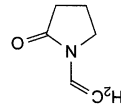
STORAGE

In an airtight container.

LABELLING

The label indicates the nominal *K*-value.

IMPURITIES



A. 1-ethenylpyrrolidin-2-one (1-vinylpyrrolidin-2-one),



B. pyrrolidin-2-one (2-pyrrolidone).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory related characteristics section since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.
The following characteristics may be relevant for povidone used as solvent and stabiliser in liquid dosage forms.

Viscosity (2.2.9)

Determine the dynamic viscosity using a capillary viscometer on a 10 per cent solution (dried substance) at 25 °C. Typical values are shown in Table 0685.-1.

Molecular mass

(see Viscosity, expressed as *K*-value). Typical values are shown in Table 0685.-1.

The following characteristic may be relevant for povidone used as binder in tablets and granules.

Molecular mass

(see Viscosity, expressed as *K*-value). Typical values are shown in Table 0685.-1.

Table 0685.-1. – Typical viscosity ranges and ranges for viscosity, expressed as *K*-value

Viscosity range (mPa.s)	Molecular mass: viscosity, expressed as <i>K</i> -value
Povidone K 12	1.3-2.3
Povidone K 17	1.5-3.5
Povidone K 25	3.5-5.5
Povidone K 30	5.5-8.5
Povidone K 90	300-700
	85-95

Iodinated Povidone

(Ph. Eur. monograph 1142)

Action and use

Antiseptic.

Preparations

Povidone-Iodine Eye Drops
Povidone-Iodine Mouthwash
Povidone-Iodine Solution

DEFINITION

Complex of iodine and povidone.

Content

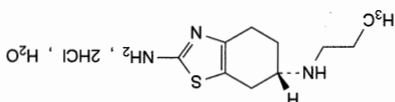
9.0 per cent to 12.0 per cent of available iodine (dried substance).





Pharmipexole Dihydrochloride Monohydrate

(Ph. Eur. monograph 2416)



$C_{10}H_{19}Cl_2N_3S \cdot H_2O$ 302.3 191217-81-9

Action and use

Dopamine receptor agonist; treatment of Parkinson's disease.

Ph. Eur.

DEFINITION

(6S)-6-N-Propyl-4,5,6,7-tetrahydro-1,3-benzothiazole-2,6-diamine dihydrochloride monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in methanol, sparingly soluble or slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

Carry out either tests B, C, D or tests A, B, D.

A. Specific optical rotation (2.2.7): -69.5 to -67.0 (anhydrous substance).

Dissolve 0.250 g in methanol R and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison prism: pexole dihydrochloride monohydrate CRS.

C. Enantiometric purity (see Tests).

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 0.1 g in water R and dilute to 10 mL with the same solvent.

pH (2.2.3)

2.8 to 3.4 .

Dissolve 0.4 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Buffer solution Dissolve 5 g of sodium octanesulfonate

monohydrate R and 9.1 g of potassium dihydrogen phosphate R in 900 mL of water R. Adjust to pH 3.0 with phosphoric acid R and dilute to 1000 mL with water R.

Solvent mixture acetonitrile R₁, buffer solution (20:80 V/V).

Test solution Dissolve 75 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

PRODUCTION

It is produced using povidone that complies with the monograph on Povidone (0685), except that the povidone used may contain not more than 2.0 per cent of formic acid and not more than 8.0 per cent of water.

CHARACTERS

Appearance

Yellowish-brown or reddish-brown, amorphous powder.

Solubility

Soluble in water and in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison iodinated povidone CRS.

B. Dissolve 10 mg in 10 mL of water R and add 1 mL of starch solution R. An intense blue colour is produced.

C. Dissolve 0.1 g in 5 mL of water R and add a 10 g/L solution of sodium sulfite R dropwise, until the solution becomes colourless. Add 2 mL of potassium dichromate solution R and 1 mL of hydrochloric acid R. A light brown precipitate is formed.

TESTS

pH (2.2.3)

1.5 to 5.0 .

Iodide

Dissolve 1.0 g in 10 mL of carbon dioxide-free water R.

Maximum 6.0 per cent (dried substance).

Dissolve 0.500 g in 100 mL of water R. Add sodium metabisulfite R until the colour of the iodine has disappeared.

Add 25.0 mL of 0.1 M silver nitrate, 10 mL of nitric acid R and 5 mL of ferric ammonium sulfate solution R₂. Titrate with 0.1 M ammonium thiocyanate. Carry out a blank titration.

1 mL of 0.1 M silver nitrate is equivalent to 12.69 mg of total iodine. From the percentage of total iodine, calculated with reference to the dried substance, subtract the percentage of available iodine as determined in the assay to obtain the percentage of iodide.

Loss on drying (2.2.32)

Maximum 8.0 per cent, determined on 0.500 g by drying in an oven at 105°C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Transfer 1.000 g into a ground-glass-stoppered flask containing 150 mL of water R and stir for 1 h. Add 0.1 mL of dilute acetic acid R and titrate with 0.1 M sodium thiosulfate using starch solution R as indicator.

1 mL of 0.1 M sodium thiosulfate is equivalent to 12.69 mg of available iodine.

STORAGE

Protected from light.

Ph. Eur.

Reference solution (b) Dissolve 7.5 mg of pramipexole for system suitability CRS (containing impurities A, B and C) in 5.0 mL of the solvent mixture.

Column:
— size: $l = 0.125$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 40 °C.

Mobile phase:
— mobile phase A: buffer solution;
— mobile phase B: acetonitrile R, buffer solution (50:50 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	60 \rightarrow 20	40 \rightarrow 80

Flow rate 1.5 mL/min.
Detection Spectrophotometer at 264 nm.

Injection 5 μ L.

Identification of impurities Use the chromatogram supplied with pramipexole for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B and C.

Relative retention With reference to pramipexole (retention time = about 6 min): impurity A = about 0.7;

impurity B = about 1.5; impurity C = about 1.7.

System suitability: reference solution (b):

— resolution: minimum 6.0 between the peaks due to

impurity A and pramipexole.

Limits:

— impurities A, B, C: for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— unspecified impurities: for each impurity, not more than

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

(0.05 per cent).

Enantiomeric purity

Liquid chromatography (2.2.29).

Test solution Dissolve 6 mg of the substance to be examined in 5 mL of anhydrous ethanol R and dilute to 20.0 mL with

the mobile phase.

Reference solution (a) Dissolve 2 mg of pramipexole

impurity D CRS in the mobile phase and dilute to 10 mL

with the mobile phase. To 1 mL of this solution add 1 mL of

the test solution and dilute to 20 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to

20.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: silica gel AD for chiral separation R.

Mobile phase diethylamine R, anhydrous ethanol R, hexane R

(0.1:15:85 V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 75 μ L.

Run time 1.5 times the retention time of pramipexole.

Relative retention With reference to pramipexole (retention

time = about 11 min): impurity D = about 0.5.

System suitability:

— resolution: minimum 5 between the peaks due to

impurity D and pramipexole in the chromatogram

obtained with reference solution (a);

— symmetry factor: maximum 2.4 for the peak due to

pramipexole in the chromatogram obtained with reference

solution (b).

Limit:

— impurity D: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.5 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent water R.

0.500 g complies with test H. Prepare the reference solution

using 0.5 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

5.0 per cent to 7.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.120 g in 150 mL of water R. Add 10 mL of

3 M nitric acid and titrate with 0.1 M silver nitrate,

determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M silver nitrate is equivalent to 14.213 mg of

$C_{10}H_{19}Cl_2N_3S$.

IMPURITIES

Specified impurities A, B, C, D

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

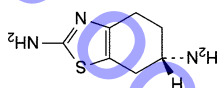
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

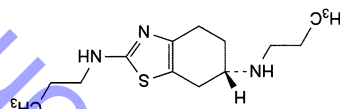
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: F.

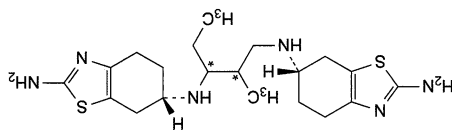


A. (6S)-4,5,6,7-tetrahydro-1,3-benzothiazole-2,6-diamine,



B. (6S)-N,N'-dipropyl-4,5,6,7-tetrahydro-1,3-benzothiazole-

2,6-diamine,



C. mixture of diastereoisomers of (6S)-6-N-[3-[[[(6S)-2-amino-4,5,6,7-tetrahydro-1,3-benzothiazol-6-yl]amino]-1-ethyl-2-methylpropyl]-4,5,6,7-tetrahydro-1,3-benzothiazole-2,6-diamine,

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in a mixture of 15 volumes of water R and

85 volumes of methanol R and dilute to 20 mL with the same

mixture of solvents. 12 mL of the solution complies with

test B. Prepare the reference solution using lead standard

solution (2 ppm Pb) obtained by diluting lead standard

solution (100 ppm Pb) R with a mixture of 15 volumes of

water R and 85 volumes of methanol R.

Water (2.5.12)

Maximum 4.0 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{23}H_{35}NaO_7$ using the

chromatogram obtained with reference solution (c) and the

declared content of pravasatin in pravasatin 1,1,3,3-

terramethylbutylamine CRS.

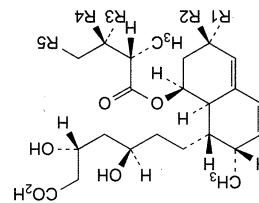
1 mg of pravasatin is equivalent to 1.052 mg of pravasatin

STORAGE

In an airtight container.

IMPURITIES

Specified impurities A, B, C, D, E, F, G



A. $R_1 = R_3 = R_4 = R_5 = OH$; $R_2 = OH$; $(3R,5R)$ -3,5-dihydroxy-7-[(1S,2S,6R,8S,8aR)-6-hydroxy-2-methyl-8-[[[2S]-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]heptanoic acid (6'-epipravasatin),

B. $R_1 = R_4 = OH$; $R_2 = R_3 = R_5 = H$; $(3R,5R)$ -3,5-dihydroxy-7-[(1S,2S,6S,8S,8aR)-6-hydroxy-8-[[[2S,3R]-3-

hydroxy-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]heptanoic acid (3'')-(R)-

hexahydronaphthalen-1-yl]heptanoic acid (3'')-(R)-

hydroxypravasatin),

C. $R_1 = OH$; $R_2 = R_3 = R_4 = H$; $R_5 = CH_3$; $(3R,5R)$ -3,5-dihydroxy-7-[(1S,2S,6S,8S,8aR)-6-hydroxy-2-methyl-8-[[[2S]-

2-methylpentanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]heptanoic acid,

E. $R_1 = R_3 = OH$; $R_2 = R_4 = R_5 = H$; $(3R,5R)$ -3,5-dihydroxy-7-[(1S,2S,6S,8S,8aR)-6-hydroxy-8-[[[2S,3S]-3-

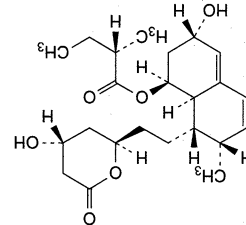
dihydroxy-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]heptanoic acid (3'')-(S)-

hydroxy-2-methylbutanoyl]oxy]-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]heptanoic acid (3'')-(S)-

hydroxypravasatin),

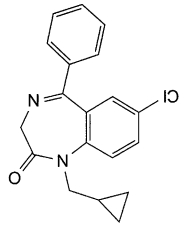
D. $(1S,3S,7S,8S,8aR)$ -3-hydroxy-8-[[2-[(2R,4R)-4-hydroxy-6-

oxotetrahydro-2H-pyran-2-yl]ethyl]-7-methyl-1,2,3,7,8,8a-hexahydronaphthalen-1-yl (2S)-2-methylbutanoate (pravasatin lactone),



Prazepam

(Ph. Eur. monograph 1466)



Action and use

Anxiolytic.

Ph. Eur.

DEFINITION

7-Chloro-1-(cyclopropylmethyl)-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in anhydrous ethanol.

mp

About 145 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison prazepam CRS.

TESTS

Related substances

Liquid chromatography (2.2.29).

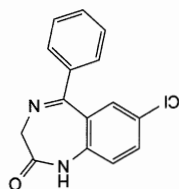
1 mL of 0.1 M perchloric acid is equivalent to 32.48 mg of $C_{19}H_{17}ClN_2O$.

STORAGE

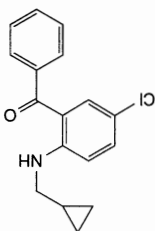
Protected from light.

IMPURITIES

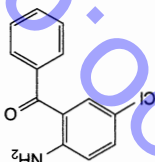
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by the general tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D.



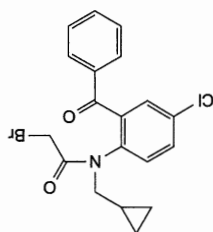
A. 7-chloro-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one (nordazepam),



B. [5-chloro-2-[(cyclopropylmethyl)amino]phenyl]phenylmethanone,



C. (2-amino-5-chlorophenyl)phenylmethanone (aminochlorobenzophenone),



D. N-(2-benzoyl-4-chlorophenyl)-2-bromo-N-(cyclopropylmethyl)acetamide.

Solvent mixture acetonitrile R₁, water for chromatography R (50:50 V/V).

Test solution Dissolve 50 mg of the substance to be examined in 25 mL of acetonitrile R₁ and dilute to 50.0 mL with water for chromatography R.

Reference solution (a) Dissolve 5 mg of

aminochlorobenzophenone R (impurity C) in acetonitrile R₁ and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of

the solution to 10.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Column: size: $l = 0.10$ m, $\varnothing = 4.6$ mm;

stationary phase: end-capped extra-dense bonded octadecylsilyl silica gel for chromatography R (1.8 μ m);

temperature: 55 °C.

Mobile phase: mobile phase A: methanol R₁, water for chromatography R (5:95 V/V);

mobile phase B: methanol R₁, acetonitrile R₁ (5:95 V/V);

Time

Mobile phase A

Mobile phase B

(min)

(per cent V/V)

0 - 0.5

0.5 - 6.5

6.5 - 8.5

15

45

45 - 85

85

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 2 μ L.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity C.

Relative retention With reference to prazepam (retention time = about 3 min): impurity C = about 0.9.

System suitability: reference solution (a):

— resolution: minimum 2.5 between the peaks due to impurity C and prazepam.

Calculation of percentage contents:

— for each impurity, use the concentration of prazepam in reference solution (b).

Limits:

— unspecified impurities: for each impurity, maximum 0.10 per cent;

— total: maximum 0.2 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

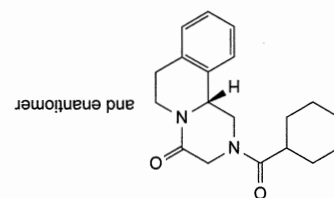
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 25 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

Praziquantel

(Ph. Eur. monograph 0855)



312.4

55268-74-1

Action and use
Anthelmintic.

Ph. Eur.

DEFINITION

(11bR5)-2-(cyclohexylcarbonyl)-1,2,3,6,7,11b-hexahydro-4H-pyrazino[2,1-a]isquinolin-4-one.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison praziquantel CRS.

If the spectra obtained show differences, dissolve 50 mg of the substance to be examined and 50 mg of the reference substance separately in 2 mL of methanol R. Evaporate and dry the residue at 60 °C at a pressure not exceeding 0.7 kPa. Record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 40.0 mg of praziquantel CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 2 mg of praziquantel for system suitability CRS (containing impurities A and B) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R₁, water for chromatography R (45:55 V/V).

Flow rate 1 mL/min.

Limits:

— **correction factor**: for the calculation of content, multiply the peak area of impurity B by 1.4;

— **impurities A, B**: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— **unspecified impurities**: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— **total**: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— **disregard limit**: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 50 °C over diphosphorus pentoxide R at a pressure not exceeding 0.7 kPa for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{19}H_{24}N_2O_2$ taking into account the assigned content of praziquantel CRS.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



IDENTIFICATION**First identification:** B, D.**Second identification:** A, C, D

A. Dissolve 50.0 mg in a 0.1 per cent *V/V* solution of hydrochloric acid R and dilute to 100.0 mL with the same acid solution. Dilute separately 1.0 mL and 5.0 mL of this solution to 100.0 mL with a 0.1 per cent *V/V* solution of hydrochloric acid R in methanol R (solution A and solution B, respectively). Examined between 220 nm and 280 nm (2.2.25), solution A shows an absorption maximum at 247 nm. The specific absorbance at the maximum is 1320 to 1400. Examined between 280 nm and 400 nm, solution B shows 2 absorption maxima, at 330 nm and 343 nm. The specific absorbances at the maxima are 260 to 280 and 240 to 265, respectively.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of potassium chloride R.

Comparison prazosin hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of 1 volume of diethylamine R, 10 volumes of methanol R and 10 volumes of methylene chloride R and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 10 mg of prazosin hydrochloride CRS in a mixture of 1 volume of diethylamine R, 10 volumes of methanol R and 10 volumes of methylene chloride R and dilute to 10 mL with the same mixture of solvents.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase diethylamine R, ethyl acetate R (5:95 *V/V*).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 2 mg in 2 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS**Related substances**

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 8 mg of metoprolamide hydrochloride CRS in 1 mL of the test solution and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm),

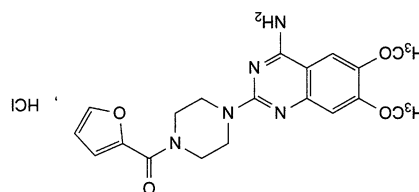
Mobile phase Mix 50 volumes of methanol R and 50 volumes of a solution containing 3.484 g/L of sodium

pentamethanesulfonate R and 3.64 g/L of tetramethylammonium hydroxide R adjusted to pH 5.0 with glacial acetic acid R.

Flow rate 1 mL/min.

Prazosin Hydrochloride

(Ph. Eur. monograph 0856)



C₁₉H₂₂ClN₅O₄ 419.9 19237-84-4

Preparation
Prazosin Tablets

Action and use Alpha₁-adrenoceptor antagonist.

DEFINITION
1-(4-Amino-6,7-dimethoxyquinoxalin-2-yl)-4-(furan-2-ylcarbonyl)piperazine hydrochloride.

Content 98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Very slightly soluble in water, slightly soluble in alcohol and in methanol, practically insoluble in acetone.

Detection Spectrophotometer at 254 nm.
Injection 20 µL.

Run time 6 times the retention time of prazosin.
Retention times Prazosin = about 9 min;

metoclopramide = about 5 min.

System suitability: reference solution (b):

— resolution: minimum 8 between the peaks due to metoclopramide and to prazosin.

Limits:

— any impurity: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.2 per cent),

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.5 per cent),

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Iron

Maximum 100 ppm.

Atomic absorption spectrometry (2.2.23, Method I).

Test solution To 1.0 g add dropwise about 1.5 mL of nitric

acid R. After fuming has subsided, evaporate on a water-bath

and ignite by gradually raising the temperature from 150 °C

to 1000 ± 50 °C, maintaining the final temperature for 1 h.

Cool, dissolve the residue in 20 mL of dilute hydrochloric

acid R, evaporate to about 5 mL and dilute to 25.0 mL with

Reference solutions Prepare the reference solutions using iron

standard solution (8 ppm Fe) R, diluted as necessary with

water R.

Source Iron hollow-cathode lamp.

Wavelength 248 nm.

Flame Air-acetylene.

Nickel

Maximum 50 ppm.

Atomic absorption spectrometry (2.2.23, Method I).

Test solution Use the test solution prepared in the test for

iron.

Reference solutions Prepare the reference solutions using nickel

standard solution (10 ppm Ni) R, diluted as necessary with

water R.

Source Nickel hollow-cathode lamp.

Wavelength 232 nm.

Flame Air-acetylene.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.000 g. Use a

mixture of equal volumes of methanol R and methylene

chloride R as the solvent.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix

thoroughly throughout and stop the titration immediately after the

end-point has been reached.

Dissolve 0.350 g in a mixture of 20 mL of anhydrous formic

acid R and 30 mL of acetic anhydride R. Titrate quickly with

0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

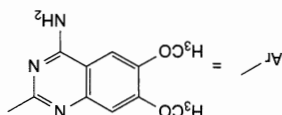
1 mL of 0.1 M perchloric acid is equivalent to 41.99 mg of

C₁₉H₂₂ClN₃O₄.

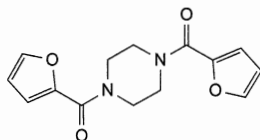
STORAGE

Protected from light.

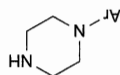
IMPURITIES



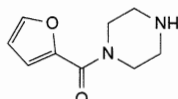
A. Ar-Cl: 2-chloro-6,7-dimethoxyquinazolin-4-amine,



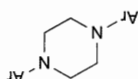
B. 1,4-bis(furan-2-ylcarbonyl)piperazine,



C. 6,7-dimethoxy-2-(piperazin-1-yl)quinazolin-4-amine,



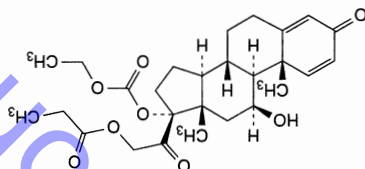
D. 1-(furan-2-ylcarbonyl)piperazine,



E. 2,2'-(piperazin-1,4-diyl)bis(6,7-dimethoxyquinazolin-4-amine).

Prednicarbate

(Ph. Eur. monograph 1467)



C₂₇H₃₆O₈ 488.6

73771-04-7

Action and use

Glucocorticoid.

Ph. Eur.

DEFINITION

1 β-Hydroxy-3,20-dioxopregna-1,4-diene-17,21-diyl

17-(ethylcarbonate) 21-propanoate.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone and in ethanol (96 per cent), sparingly soluble in propylene glycol. It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison prednicarbate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of ethanol (96 per cent) R, evaporate to dryness on a water-bath and record new spectra using the residues.

B. Thin-layer chromatography (2.2.27).

Solvent mixture: methanol R, methylene chloride R (10:90 V/V).

Test solution: Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a): Dissolve 10 mg of prednicarbate CRS in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b): Dissolve 5 mg of prednisolone acetate CRS in 5.0 mL of reference solution (a).

Plate: TLC silica gel F₂₅₄ plate R.

Mobile phase: Add a mixture of 1.2 volumes of water R and 8 volumes of methanol R to a mixture of 15 volumes of either R and 77 volumes of methylene chloride R.

Application: 5 µL.

Development: Over 3/4 of the plate.

Drying: In air.

Detection A: Examine in ultraviolet light at 254 nm.

Results A: The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B: Spray with alcoholic solution of sulfuric acid R, heat at 120 °C for 10 min or until the spots appear and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B: The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

TESTS**Specific optical rotation (2.2.7)**

Dissolve 0.250 g in ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution: Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a): Dissolve 3 mg of prednicarbate for system suitability A CRS (containing impurities B, C, D, E and F) in

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection: Test solution and reference solution (c).

Calculate the percentage content of C₂₇H₃₆O₈ taking into account the assigned content of prednicarbate CRS.

STORAGE

Protected from light.

IMPURITIES

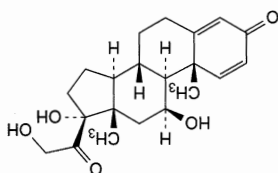
Specified impurities: B, C, D, E, F

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A.

Prednisolone

(Ph Eur monograph 0353)



$C_{21}H_{28}O_5$ 360.4

50-24-8

Action and use

Glucocorticoid.

Preparations

Prednisolone Tablets

Gastro-resistant Prednisolone Tablets

DEFINITION

11 β ,17,21-Trihydroxypregna-1,4-diene-3,20-dione.

Content

96.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline, hygroscopic powder.

Solubility

Very slightly soluble in water; soluble in ethanol (96 per cent) and in methanol, sparingly soluble in acetone, slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison prednisolone CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of acetone R , evaporate to dryness on a water-bath and record new spectra using the residues.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with test solution (b) is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (d).

TESTS

Specific optical rotation (2.2.7)

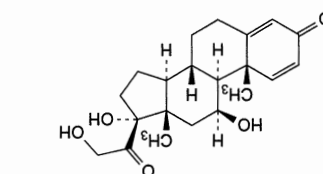
+113 to +119 (dried substance).
Dissolve 0.250 g in ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.

Related substances

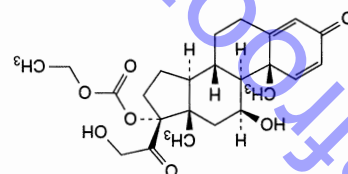
Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture acetonitrile R , water R (40:60 V/V).

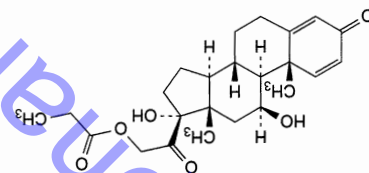
Test solution (a) Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.
Test solution (b) Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.



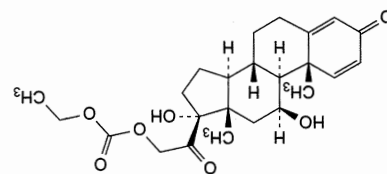
A. 11 β ,17,21-trihydroxypregna-1,4-diene-3,20-dione (prednisolone),



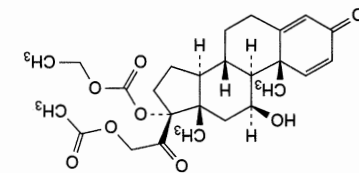
B. 11 β ,21-dihydroxy-3,20-dioxopregna-1,4-dien-17-yl ethyl carbonate (prednisolone 17-ethylcarbonate),



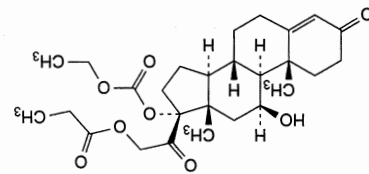
C. 11 β ,17-dihydroxy-3,20-dioxopregna-1,4-dien-21-yl propionate (prednisolone 21-propionate),



D. 11 β ,17-dihydroxy-3,20-dioxopregna-1,4-dien-21-yl ethyl carbonate (prednisolone 21-ethylcarbonate),



E. 11 β -hydroxy-3,20-dioxopregna-1,4-dien-17,21-diyl 21-acetate 17-(ethylcarbonate) (prednisolone 21-acetate 17-ethylcarbonate),



F. 11 β -hydroxy-3,20-dioxopregna-4-ene-17,21-diyl 17-(ethylcarbonate) 21-propionate (1,2-dihydroprednisolone).

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total*: not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (2.2.32)
Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d). Calculate the percentage content of $C_{21}H_{28}O_5$ from the declared content of prednisolone CRS.

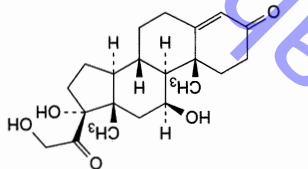
STORAGE

In an airtight container, protected from light.

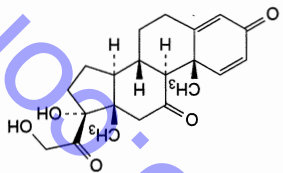
IMPURITIES

Specified impurities A, B, C, F, J

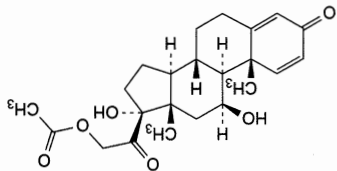
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, E, G, H, I.



A. 11β,17,21-trihydroxypregn-4-ene-3,20-dione (hydrocortisone),



B. 17,21-dihydroxypregn-1,4-diene-3,11,20-trione (prednisolone),



C. 11β,17-dihydroxy-3,20-dioxopregn-1,4-dien-21-yl acetate (prednisolone acetate),

Reference solution (a) Dissolve 5 mg of prednisolone for system suitability CRS (containing impurities A, B and C) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of prednisolone for peak identification CRS (containing impurities F and J) in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 25.0 mg of prednisolone CRS in the solvent mixture and dilute to 20.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Column:
size: $l = 0.15$ m, $\phi = 4.6$ mm;
stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m);
temperature: 40 °C.

Mobile phase:
mobile phase A: water R;
mobile phase B: acetonitrile R, methanol R (50:50 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 14	60	40
14 - 20	60 → 20	40 → 80
20 - 25	20	80

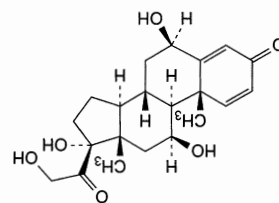
Detection Spectrophotometer at 254 nm.
Injection 10 μ L of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities Use the chromatogram supplied with prednisolone for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C; use the chromatogram supplied with prednisolone for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities F and J.

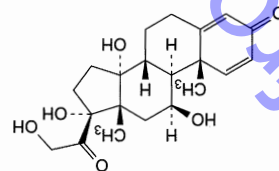
Relative retention With reference to prednisolone (retention time = about 12 min): impurity F = about 0.7; impurity B = about 0.9; impurity A = about 1.05; impurity J = about 1.5; impurity C = about 1.7.

System suitability: reference solution (a):
— *peak-to-valley ratio*: minimum 3, where H_p = height above the baseline of the peak due to impurity A and H_b = height above the baseline of the lowest point of the curve separating this peak from the peak due to prednisolone.

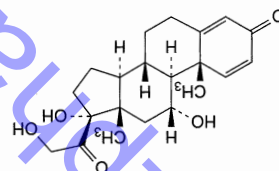
Limits:
— *impurity A*: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
— *impurity F*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
— *impurities B, C, J*: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);



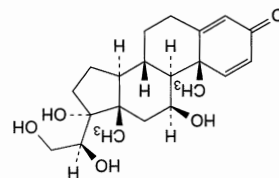
D. 6β,11β,17,21-tetrahydroxypregna-1,4-diene-3,20-dione (11-deoxyprednisolone),



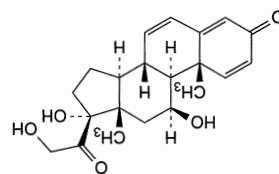
E. 11β,14α,17,21-tetrahydroxypregna-1,4-diene-3,20-dione (14α-hydroxyprednisolone),



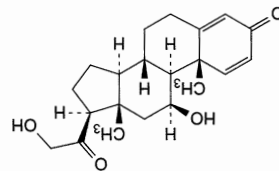
F. 11α,17,21-trihydroxypregna-1,4-diene-3,20-dione (11-epi-prednisolone),



G. 11β,17,20β,21-tetrahydroxypregna-1,4-dien-3-one (20β-hydroxyprednisolone),



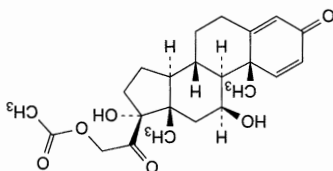
H. 11β,17,21-trihydroxypregna-1,4,6-triene-3,20-dione (Δ⁶-prednisolone),



I. 11β,21-dihydroxypregna-1,4-diene-3,20-dione (17-deoxyprednisolone),

Prednisolone Acetate

(Ph. Eur. monograph 0734)



C₂₃H₃₀O₆

402.5

52-21-1

Action and use

Glucocorticoid.

Preparation

Prednisolone Acetate Injection

Ph Eur

DEFINITION

11β,17-Dihydroxy-3,20-dioxopregna-1,4-dien-21-yl acetate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol

(96 per cent) and in methylene chloride.

IDENTIFICATION

First identification A, B

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison prednisolone acetate CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in a mixture of 1 volume of methanol R and 9 volumes of

methylene chloride R and dilute to 10 mL with the same

mixture of solvents.

Reference solution (a) Dissolve 20 mg of prednisolone

acetate CRS in a mixture of 1 volume of methanol R and

9 volumes of methylene chloride R and dilute to 20 mL with

the same mixture of solvents.

Reference solution (b) Dissolve 10 mg of prednisolone

pivalate CRS in reference solution (a) and dilute to 10 mL

with the same solution.

Plate TLC silica gel F₂₅₄ plate R.

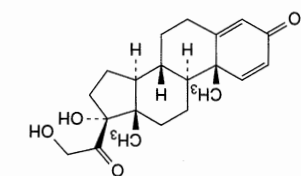
Mobile phase Add a mixture of 1.2 volumes of water R and

8 volumes of methanol R to a mixture of 15 volumes of

ether R and 77 volumes of methylene chloride R.



Ph Eur



J. 17,21-dihydroxypregna-1,4-diene-3,20-dione (11-deoxyprednisolone).

Injection 20 mL.

Run time 2.5 times the retention time of prednisolone acetate.

Identification of impurities Use the chromatogram supplied with prednisolone acetate for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to prednisolone acetate

(retention time = about 1.7 min): impurity B = about 0.4; impurity A = about 1.1; impurity C = about 2.0.

System suitability: reference solution (a):

— *resolution*: minimum 2.0 between the peaks due to prednisolone acetate and impurity A.

Limits:

— *impurities A, B*: for each impurity, not more than 5 times the area of the principal peak in the chromatogram

obtained with reference solution (b) (1.0 per cent);

— *impurity C*: not more than 2.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.5 per cent);

— *unspecified impurities*: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— *total*: not more than 10 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (2.0 per cent);

— *disregard limit*: 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

ASSAY

Dissolve 0.100 g in ethanol (96 per cent) R and dilute to

100.0 mL with the same solvent. Dilute 2.0 mL of this

solution to 100.0 mL with ethanol (96 per cent) R. Measure

the absorbance (2.2.25) at the absorption maximum at

243 nm.

Calculate the content of C₂₃H₃₀O₆ taking the specific

absorbance to be 370.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C.

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

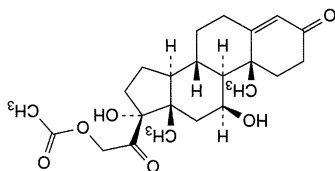
acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: D, E.



A. 11 β ,17-dihydroxy-3,20-dioxopregn-4-en-21-yl acetate (hydrocortisone acetate).

Application 5 μ L.
Development Over a path of 15 cm.
Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R. Heat

at 105 °C for 10 min or until the spots appear. Allow to

cool. Examine in daylight and in ultraviolet light at 365 nm.

System suitability: reference solution (b):

— the chromatogram obtained shows 2 clearly separated

spots.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight,

fluorescence in ultraviolet light at 365 nm and size to the

principal spot in the chromatogram obtained with reference

solution (a).

C. Add about 2 mg to 2 mL of sulfuric acid R and shake to

dissolve. Within 5 min, an intense red colour develops. When

examined in ultraviolet light at 365 nm, a reddish-brown

fluorescence is seen. Add the solution to 10 mL of water R

and mix. The colour fades and there is greenish-yellow

fluorescence in ultraviolet light at 365 nm.

D. About 10 mg gives the reaction of acetyl (2.3.1).

TESTS

Specific optical rotation (2.2.7)

+ 128 to + 137 (dried substance).

Dissolve 70.0 mg in methanol R2 and dilute to 20.0 mL with

the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Buffer solution pH 4 Mix 1 volume of dilute hydrochloric acid R,

5 volumes of a 68.1 g/L solution of sodium acetate R,

15 volumes of a 37.3 g/L solution of potassium chloride R and

79 volumes of water R.

Solvent mixture Mix equal volumes of acetonitrile R and buffer

solution pH 4.

Test solution Dissolve 25.0 mg of the substance to be

examined in methanol R and dilute to 10.0 mL with the same

solvent.

Reference solution (a) Dissolve 2 mg of prednisolone

acetate CRS and 2 mg of hydrocortisone acetate CRS

(impurity A) in the solvent mixture and dilute to 100.0 mL

with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 2.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of prednisolone acetate for

peak identification CRS (containing impurities A, B and C) in

the solvent mixture and dilute to 50 mL with the solvent

mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

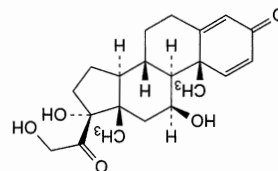
chromatography R (5 μ m);

— temperature: 40 °C.

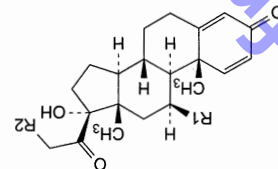
Mobile phase acetonitrile R, water R (350:650 V/V).

Flow rate 1 mL/min.

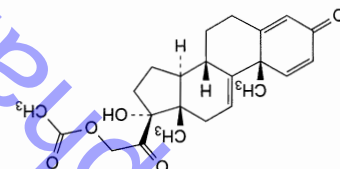
Detection Spectrophotometer at 254 nm.



B. 11 β ,17,21-trihydroxyprogesterone-1,4-diene-3,20-dione (prednisolone),



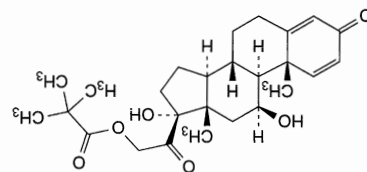
C. R1 = R2 = O-CO-CH₃; 17-hydroxy-3,20-dioxopregna-1,4-diene-11 β ,21-diyl diacetate (prednisolone 11,21-diacetate),
D. R1 = OH, R2 = H; 11 β ,17-dihydroxyprogesterone-1,4-diene-3,20-dione,



E. 17-hydroxy-3,20-dioxopregna-1,4,9(11)-trien-21-yl acetate.

Prednisolone Pivalate

(Ph. Eur. monograph 0736)



C₂₆H₃₆O₆

444.6

1107-99-9

Action and use
Glucocorticoid.

DEFINITION

11 β ,17-Dihydroxy-3,20-dioxopregna-1,4-dien-21-yl 2,2-dimethylpropanoate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent), soluble in methylene chloride.

mp: about 229 °C, with decomposition.

IDENTIFICATION

First identification B, C.

Second identification A, C, D.

A. Dissolve 10.0 mg in anhydrous ethanol R and dilute to 100.0 mL with the same solvent. Place 2.0 mL of this solution in a ground-glass-stoppered tube, add 10.0 mL of phenylhydrazine-sulfuric acid solution R, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (2.25) at the absorption maximum at 415 nm is 0.20 to 0.30.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison prednisolone pivalate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of ethanol (96 per cent) R, evaporate to dryness on a water-bath and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Solvent mixture methanol R, methylene chloride R (1:9 V/V).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 10 mg of prednisolone pivalate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of prednisolone acetate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Dilute 5 mL of this solution to 10 mL with the solvent mixture.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and 8 volumes of methanol R to a mixture of 15 volumes of ether R and 77 volumes of methylene chloride R.

Application 5 μ L.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R, heat at 120 °C for 10 min or until the spots appear, and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability: reference solution (b):

the chromatogram shows 2 clearly separated spots.

D. To 2 mL of sulfuric acid R, add about 2 mg and shake to dissolve. Within 5 min, an intense red colour develops. When examined in ultraviolet light at 365 nm, a reddish-brown fluorescence is seen. Add this solution to 10 mL of water R and mix. The colour fades and there is greenish-yellow fluorescence in ultraviolet light at 365 nm.

TESTS

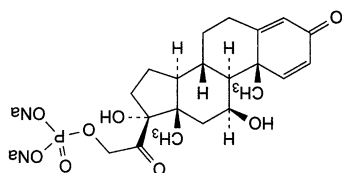
Specific optical rotation (2.2.7)

Dissolve 0.250 g in dioxan R and dilute to 25.0 mL with the same solvent.



Prednisolone Sodium Phosphate

(Ph. Eur. monograph 0735)



125-02-0

484.4

$C_{21}H_{27}Na_2O_8P$

Action and use

Glucocorticoid.

Preparations

Prednisolone Enema

Prednisolone Sodium Phosphate Ear Drops

Prednisolone Sodium Phosphate Eye Drops

Prednisolone Sodium Phosphate Oral Solution

Soluble Prednisolone Tablets

Ph Eur

DEFINITION

11 β ,17-Dihydroxy-3,20-dioxopregn-1,4-dien-21-yl disodium phosphate.

Content

96.0 per cent to 103.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B, C

Second identification A, C, D, E

A. Dissolve 10.0 mg in 5 mL of water R and dilute to 100.0 mL with anhydrous ethanol R. Place 2.0 mL of this solution in a ground-glass-stoppered tube, add 10.0 mL of phenylhydrazine-sulfuric acid solution R, mix and heat in a water-bath at 60 °C for 20 min. Cool immediately. The absorbance (2.2.25) at the absorption maximum at 415 nm is 0.10 to 0.20.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison prednisolone sodium phosphate CRS. If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of ethanol (96 per cent) R, evaporate to dryness on a water-bath and record new spectra using the residues. C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent. Reference solution (a) Dissolve 10 mg of prednisolone sodium phosphate CRS in methanol R and dilute to 10 mL with the same solvent. Reference solution (b) Dissolve 10 mg of dexamethasone sodium phosphate CRS in methanol R and dilute to 10 mL with the same solvent. Dilute 5 mL of this solution to 10 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Ph Eur

Protected from light.

STORAGE

Calculate the content of C₂₆H₃₀O₆ taking the specific absorbance to be 337. Dissolve 0.100 g in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 250.0 mL with ethanol (96 per cent) R. Measure the absorbance (2.2.25) at the absorption maximum at 243 nm.

ASSAY

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C. Loss on drying (2.2.32)

(0.05 per cent). in the chromatogram obtained with reference solution (b) disregard limit: 0.025 times the area of the principal peak solution (b) (2.5 per cent); peak in the chromatogram obtained with reference solution (b) not more than 1.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent); of the principal peak in the chromatogram obtained with one such peak has an area greater than 0.5 times the area reference solution (b) (2.0 per cent), and not more than the principal peak in the chromatogram obtained with any impurity: for each impurity, not more than the area of

Limits:

adjust the concentration of water in the mobile phase. prednisolone acetate and cortisone acetate; if necessary,

— resolution: minimum 2.5 between the peaks due to System suitability: reference solution (a):

pivalate = about 13 min.

cortisone acetate = about 4.5 min; prednisolone

Retention time Prednisolone acetate = about 3.5 min;

pivalate.

Run time 1.5 times the retention time of prednisolone

Injection 20 μ L.

Equilibration With the mobile phase for about 30 min.

Detection Spectrophotometer at 254 nm.

Flow rate 1 mL/min.

filter.

allow to equilibrate for 1 h and filter through a 0.45 μ m

monomethyl ether R, then add with 231 mL of water R, mix,

37 mL of tetrahydrofuran R and 213 mL of ethylene glycol

Mobile phase Carefully mix 19 mL of butyl acetate R1 with

(5 μ m).

— stationary phase: octadecylsilyl silica gel for chromatography R

— size: 1 = 0.15 m, ϕ = 4.6 mm;

Column:

50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to

solution to 25.0 mL with the mobile phase.

25.0 mL with the mobile phase. Dilute 1.0 mL of this

of water R and 4 volumes of tetrahydrofuran R and dilute to

prednisolone pivalate CRS in 2 mL of a mixture of 1 volume

acetate CRS, 25 mg of cortisone acetate CRS and 25 mg of

Reference solution (a) Dissolve 25 mg of prednisolone

the mobile phase.

4 volumes of tetrahydrofuran R and dilute to 25.0 mL with

examined in 2 mL of a mixture of 1 volume of water R and

Test solution Dissolve 62.5 mg of the substance to be

Liquid chromatography (2.2.29).

Related substances

Mobile phase: glacial acetic acid R, water R, butanol R

(20:20:60 V/V/V).

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R, heat at 120 °C for 10 min or until the spots appear, and allow to cool; examine in daylight and in ultraviolet light at 365 nm.

Results B The principal spot in the chromatogram obtained with the test solution is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with reference solution (a).

System suitability: reference solution (b): the chromatogram shows 2 spots which may, however, not be completely separated.

D. To 2 mL of sulfuric acid R add about 2 mg and shake to dissolve. Within 5 min, an intense red colour develops. When examined in ultraviolet light at 365 nm, a reddish-brown fluorescence is seen. Add this solution to 10 mL of water R and mix. The colour fades and there is a greenish-yellow fluorescence in ultraviolet light at 365 nm.

E. To about 40 mg add 2 mL of sulfuric acid R and heat gently until white fumes are evolved. Add nitric acid R dropwise, continue the heating until the solution is almost colourless, and cool. Add 2 mL of water R, heat until white fumes are again evolved, cool, add 10 mL of water R and neutralise to red litmus paper R with dilute ammonia R1.

The solution gives reaction (a) of sodium (2.3.1) and reaction (b) of phosphates (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

pH (2.2.3)

7.5 to 9.0 for solution S.

Specific optical rotation (2.2.7)

+ 94 to + 100 (anhydrous substance).

Dissolve 0.250 g in water R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 62.5 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 25 mg of prednisolone sodium phosphate CRS and 25 mg of prednisolone CRS in the mobile phase and dilute to 25.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 25.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase.

Column:

— size: l = 0.15 m, Ø = 4.6 mm;

Inorganic phosphate

Maximum 1 per cent.

Dissolve 50 mg in water R and dilute to 100 mL with the same solvent. To 10 mL of this solution add 5 mL of molybdovanadic reagent R, mix, and allow to stand for 5 min. Any yellow colour in the solution is not more intense than that in a standard prepared at the same time and in the same manner using 10 mL of phosphate standard solution (5 ppm PO₄) R.

Water (2.5.12)

Maximum 8.0 per cent, determined on 0.200 g.

ASSAY

Dissolve 0.100 g in water R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 250.0 mL with water R. Measure the absorbance (2.2.25) at the absorption maximum at 247 nm.

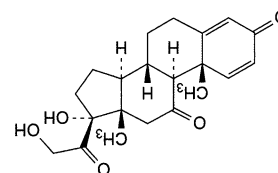
Calculate the content of C₂₁H₂₇Na₂O₈P taking the specific absorbance to be 312.

STORAGE

Protected from light.

Prednisone

(Ph Eur monograph 0354)



$C_{21}H_{26}O_5$

358.4

53-03-2

Action and use

Glucocorticoid.

Ph Eur

DEFINITION

17,21-Dihydroxypregna-1,4-diene-3,11,20-trione.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol

(96 per cent) and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

First identification A, B.

Second identification C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison prednisone CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in the minimum volume of acetone R,

evaporate to dryness on a water-bath and record new spectra

using the residues.

B. Thin-layer chromatography (2.2.27).

Solvent mixture methanol R, methylene chloride R (1:9 V/V).

Test solution Dissolve 10 mg of the substance to be examined

in the solvent mixture and dilute to 10 mL with the solvent

mixture.

Reference solution (a) Dissolve 20 mg of prednisone CRS in the

solvent mixture and dilute to 20 mL with the solvent

mixture.

Reference solution (b) Dissolve 10 mg of betamethasone CRS in

reference solution (a) and dilute to 10 mL with reference

solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and

8 volumes of methanol R to a mixture of 15 volumes of

ether R and 77 volumes of methylene chloride R.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained

with the test solution is similar in position and size to the

principal spot in the chromatogram obtained with reference

solution (a).

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 25 mg of the substance to be

examined in methanol R and dilute to 5 mL with the same

solvent (solution A). Dilute 2 mL of this solution to 10 mL

with methylene chloride R.

Test solution (b) Transfer 0.4 mL of solution A to a glass tube

100 mm long and 20 mm in diameter and fitted with a

ground-glass stopper or a polytetrafluoroethylene cap and

evaporate the solvent with gentle heating under a stream of

nitrogen R. Add 2 mL of a 15 per cent V/V solution of glacial

acetic acid R and 50 mg of sodium bismuthate R. Stopper the

tube and shake the suspension in a mechanical shaker,

protected from light, for 1 h. Add 2 mL of a 15 per cent V/V

solution of glacial acetic acid R and filter into a 50 mL

separating funnel, washing the filter with 2 quantities, each of

5 mL, of water R. Shake the clear filtrate with 10 mL of

methylene chloride R. Wash the organic layer with 5 mL of

1 M sodium hydroxide and 2 quantities, each of 5 mL, of

water R. Dry over anhydrous sodium sulfate R.

Reference solution (a) Dissolve 25 mg of prednisone CRS in

methanol R and dilute to 5 mL with the same solvent

(solution B). Dilute 2 mL of this solution to 10 mL with

methylene chloride R.

Reference solution (b) Transfer 0.4 mL of solution B to a glass

tube 100 mm long and 20 mm in diameter and fitted with a

ground-glass stopper or a polytetrafluoroethylene cap and

evaporate the solvent with gentle heating under a stream of

nitrogen R. Add 2 mL of a 15 per cent V/V solution of glacial

acetic acid R and 50 mg of sodium bismuthate R. Stopper the

tube and shake the suspension in a mechanical shaker,

protected from light, for 1 h. Add 2 mL of a 15 per cent V/V

solution of glacial acetic acid R and filter into a 50 mL

separating funnel, washing the filter with 2 quantities, each of

5 mL, of water R. Shake the clear filtrate with 10 mL of

methylene chloride R. Wash the organic layer with 5 mL of

1 M sodium hydroxide and 2 quantities, each of 5 mL, of

water R. Dry over anhydrous sodium sulfate R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and

8 volumes of methanol R to a mixture of 15 volumes of

ether R and 77 volumes of methylene chloride R.

Application 5 µL of test solution (a) and reference solution (a)

and 50 µL of test solution (b) and reference solution (b),

applying the latter 2 in small quantities in order to obtain

small spots.

Development Over a path of 15 cm.

Drying In air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in each of the chromatograms

obtained with the test solutions is similar in position and size

to the principal spot in the chromatogram obtained with the

corresponding reference solution.

Detection B Spray with alcoholic solution of sulfuric acid R. Heat at 120 °C for 10 min or until the spots appear. Allow to cool. Examine in daylight and in ultraviolet light at 365 nm. **Results B** The principal spot in each of the chromatograms obtained with the test solutions is similar in position, colour in daylight, fluorescence in ultraviolet light at 365 nm and size to the principal spot in the chromatogram obtained with the corresponding reference solution. The principal spots in the chromatograms obtained with test solution (b) and the chromatograms obtained with test solution (a) have an R_F value distinctly higher than that of the principal spots in the chromatograms obtained with test solution (a) and reference solution (a). **D.** Add about 2 mg to 2 mL of sulfuric acid R and shake to dissolve. Within 5 min, a yellow colour develops with a blue fluorescence in ultraviolet light at 365 nm. Add this solution to 10 mL of water R and mix. The colour fades but the blue fluorescence in ultraviolet light does not disappear.

TESTS

Specific optical rotation (2.2.7)
+ 167 to + 175 (dried substance).
Dissolve 0.125 g in dioxan R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 25.0 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of prednisone CRS and 2 mg of prednisolone CRS in methanol R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R.

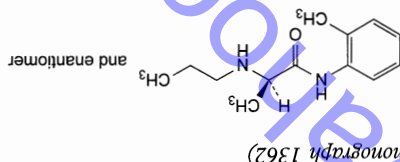
Column:
— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 45 °C.

Mobile phase:
— mobile phase A: in a 1000 mL volumetric flask mix 100 mL of acetonitrile R with 200 mL of methanol R and 650 mL of water R; allow to equilibrate; adjust to 1000 mL with water R and mix again;
— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 25	100	0
25 - 40	100 \rightarrow 40	0 \rightarrow 60
40 - 41	40 \rightarrow 0	60 \rightarrow 100
41 - 46	0	100
46 - 47	0 \rightarrow 100	100 \rightarrow 0
47 - 52	100	0

Flow rate 2.5 mL/min.
Detection Spectrophotometer at 254 nm.
Equilibration With mobile phase B for at least 30 min, and then with mobile phase A for 5 min. For subsequent chromatograms, use the conditions described from 40.0 min to 52.0 min.
Injection 20 μ L, inject methanol R as a blank.

Prilocaine



$C_{13}H_{20}N_2O$ 220.3
721-50-6

Action and use
Local anaesthetic.
Ph Eur

DEFINITION
(2RS)-N-(2-Methylphenyl)-2-(propylamino)propanamide.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS
Appearance
White or almost white, crystalline powder.

Solubility
Slightly soluble in water, very soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION
Infrared absorption spectrophotometry (2.2.24).
Preparation Prepare a film between 2 plates of sodium chloride R by heating at 40-45 °C until the substance has melted.

Retention time Prednisone = about 19 min; prednisolone = about 23 min.
— **resolution:** minimum 2.7 between the peaks due to prednisone and prednisolone; if necessary, adjust the concentration of acetonitrile in mobile phase A.
Limits:
— **any impurity:** for each impurity, not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);
— **total:** not more than 0.75 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.75 per cent);
— **disregard limit:** 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)
Maximum 1.0 per cent, determined on 0.500 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.100 g in ethanol (96 per cent) R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with ethanol (96 per cent) R. Measure the absorbance (2.2.25) at the absorption maximum at 238 nm.
Calculate the content of $C_{13}H_{20}N_2O$ taking the specific absorbance to be 425.

STORAGE
Protected from light.

Ph Eur

Comparison prilocaine CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dissolve 2.50 g in 15 mL of dilute hydrochloric acid R and dilute to 50.0 mL with water R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.5 mg of the substance to be examined and 3 mg of prilocaine impurity E CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 33.5 mg of prilocaine impurity B CRS in the mobile phase and dilute to 100.0 mL

with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 15 mg of prilocaine for peak identification CRS (containing impurity G) in the mobile phase and dilute to 5.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 26 volumes of acetonitrile R and 74 volumes of a solution prepared as follows: dissolve 0.180 g of sodium

dihydrogen phosphate monohydrate R and 2.89 g of disodium dihydrogen phosphate dihydrate R in 1000 mL of water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 μ L.

Run time Twice the retention time of prilocaine.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to

impurity B; use the chromatogram obtained with reference solution (a) to identify the peak due to impurity E; use the chromatogram obtained with reference solution (d) to

identify the peak due to impurity G.

Relative retention With reference to prilocaine (retention time = about 25 min): impurity B = about 0.3;

impurity G = about 0.8; impurity E = about 1.2.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to prilocaine and impurity E.

Limits:

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference

solution (c) (100 ppm);

— impurity G: not more than 1.5 times the area of the principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent ethanol (96 per cent) R.

1.0 g complies with test H. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

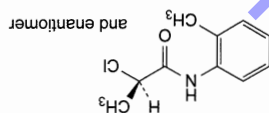
Dissolve 0.180 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 22.03 mg of $C_{13}H_{20}N_2O$.

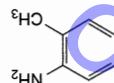
IMPURITIES

Specified impurities B, G

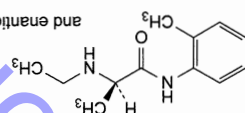
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, D, E, F.



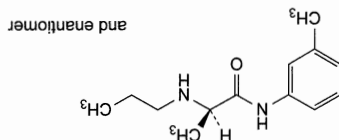
A. (2RS)-2-chloro-N-(2-methylphenyl)propanamide,



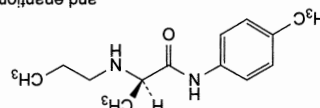
B. 2-methylbenzenamine (o-toluidine),



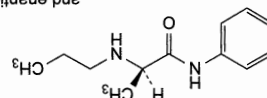
C. (2RS)-2-(ethylamino)-N-(2-methylphenyl)propanamide,



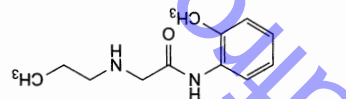
D. (2RS)-N-(3-methylphenyl)-2-(propylamino)propanamide,



E. (2R,5S)-N-(4-methylphenyl)-2-(propylamino)propanamide,



F. (2R,5S)-N-phenyl-2-(propylamino)propanamide,



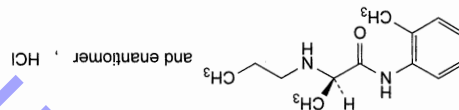
G. N-(2-methylphenyl)-2-(propylamino)acetamide.

Ph Eur



Prilocaine Hydrochloride

(Ph. Eur. monograph 1363)



$C_{13}H_{21}ClN_2O$ 256.8 1786-81-8

Action and use

Local anaesthetic.

Preparation

Prilocaine Injection

Ph Eur

DEFINITION

(2R,5S)-N-(2-Methylphenyl)-2-(propylamino)propanamide hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water and in ethanol (96 per cent), very slightly soluble in acetone.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Melting point (2.2.14): 168 °C to 171 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison prilocaine hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20.0 mg of the substance to be examined in ethanol (96 per cent) R and dilute to 5 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 30 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 3 mg of the substance to be examined and 3 mg of prilocaine impurity B CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 30.0 mg of prilocaine impurity B CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (d) Dissolve 15 mg of prilocaine for peak identification CRS (containing impurity G) in the mobile phase and dilute to 5.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 26 volumes of acetonitrile R and 74 volumes of a solution prepared as follows: dissolve 0.180 g of sodium

yellow.

Add 0.80 mL of 0.01 M hydrochloric acid; the solution is 0.40 mL of 0.01 M sodium hydroxide; the solution is blue. water R. Add 0.1 mL of bromocresol green solution R and dilute 4 mL of solution S to 10 mL with carbon dioxide-free

Acidity or alkalinity

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Appearance of solution

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

TESTS

D. It gives reaction (a) of chlorides (2.3.1).

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

— the chromatogram shows 2 clearly separated spots.

System suitability: reference solution (b):

Detection Examine in ultraviolet light at 254 nm.

Drying In air.

Development Over a path of 12 cm.

Application 10 μ L.

Mobile phase concentrated ammonia R, methanol R, 1,1-dimethylethyl methyl ether R (15:100 V/V/V).

Plate TLC silica gel F₂₅₄ plate R.

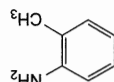
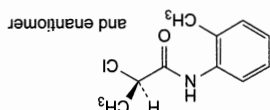
solvent.

Reference solution (a) Dissolve 20.0 mg of prilocaine hydrochloride CRS in ethanol (96 per cent) R and dilute to 5 mL with the same solvent.

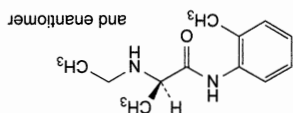
Reference solution (b) Dissolve 20.0 mg of lidocaine hydrochloride CRS and 20.0 mg of prilocaine hydrochloride CRS in ethanol (96 per cent) R and dilute to 5 mL with the same

Control of impurities in substances for pharmaceutical use): A, C, D, E, F.

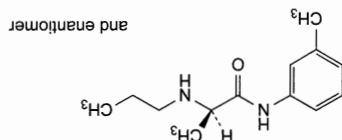
A. (2*RS*)-2-chloro-*N*-(2-methylphenyl)propanamide,



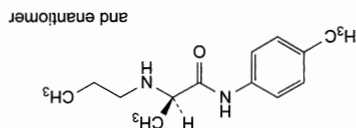
B. 2-methylbenzenamine (*o*-toluidine),



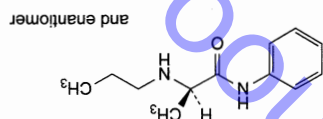
C. (2*RS*)-2-(ethylamino)-*N*-(2-methylphenyl)propanamide,



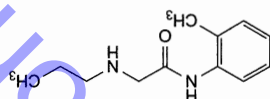
D. (2*RS*)-*N*-(3-methylphenyl)-2-(propylamino)propanamide,



E. (2*RS*)-*N*-(4-methylphenyl)-2-(propylamino)propanamide,



F. (2*RS*)-*N*-phenyl-2-(propylamino)propanamide,



G. *N*-(2-methylphenyl)-2-(propylamino)acetamide.

dihydrogen phosphate monohydrate R and 2.89 g of disodium hydrogen phosphate R in 1000 mL of water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 µL.

Run time Twice the retention time of prilocaine.

Identification of impurities Use the chromatogram obtained

with reference solution (c) to identify the peak due to

impurity B; use the chromatogram obtained with reference

solution (a) to identify the peak due to impurity E; use the

chromatogram obtained with reference solution (d) to

identify the peak due to impurity G.

Relative retention With reference to prilocaine (retention

time = about 25 min): impurity B = about 0.3;

impurity G = about 0.8; impurity E = about 1.2.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to

prilocaine and impurity E.

Limits:

— impurity B: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (c) (100 ppm);

— impurity G: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (b)

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

Maximum 20 ppm.

Solvent ethanol (96 per cent) R.

1.0 g complies with test H. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 5.0 mL of 0.01 M

hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry

out a potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 25.68 mg

of C₁₃H₂₁ClN₂O.

IMPURITIES

Specified impurities B, G

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by the general

acceptance criterion for other/unspecified impurities and/or

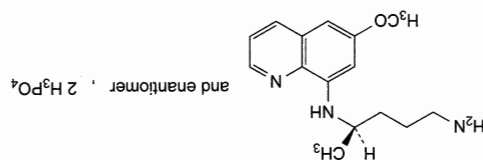
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Primaquine Phosphate

(Primaquine Diphosphate, Ph Eur monograph 0635)


 $C_{15}H_{27}N_3O_9P_2$ 455.3 63-45-6

Action and use

Antiprotosozal (malaria).

Ph Eur

DEFINITION

(4*RS*)-*N*'-(6-Methoxyquinolin-8-yl)pentane-1,4-diamine bisphosphate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

Orange crystalline powder.

Solubility

Soluble in water, practically insoluble in ethanol

(96 per cent).

mp: about 200 °C, with decomposition.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution (a) Dissolve 15 mg in 0.01 M hydrochloric acid

and dilute to 100.0 mL with the same acid.

Test solution (b) Dilute 5.0 mL of test solution (a) to

50.0 mL with 0.01 M hydrochloric acid.

Spectral range 310–450 nm for test solution (a); 215–310 nm

for test solution (b).

Absorption maxima At 332 nm and 415 nm for test

solution (a); at 225 nm, 265 nm and 282 nm for test

solution (b).

Specific absorbance at the absorption maxima:

— at 332 nm: 45 to 52 for test solution (a);

— at 415 nm: 27 to 35 for test solution (a);

— at 225 nm: 495 to 515 for test solution (b);

— at 265 nm: 335 to 350 for test solution (b);

— at 282 nm: 330 to 345 for test solution (b).

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Dissolve separately 0.1 g of the substance to be examined

and 0.1 g of the reference substance in 5 mL of water R, add

2 mL of dilute ammonia R2 and 5 mL of methylene chloride R,

then shake. Dry the methylene chloride layer over 0.5 g of

anhydrous sodium sulfate R. Prepare a blank disc using about

0.1 mL of the methylene chloride layer, allowing the

methylene chloride to evaporate between applications.

Dry the disc at 50 °C for 2 min.

Comparison primaquine diphosphate CRS.

C. Thin-layer chromatography (2.2.27). Carry out all operations as rapidly as possible, protected from light. Prepare the solutions immediately before use.

Test solution Dissolve 0.20 g of the substance to be examined in 5 mL of water R and dilute to 10 mL with methanol R. Dilute 1 mL of this solution to 10 mL with a mixture of equal volumes of methanol R and water R.

Reference solution Dissolve 20 mg of primaquine diphosphate CRS in 5 mL of water R and dilute to 10 mL with methanol R.

Plate TLC silica gel GF₂₅₄ plate R. Pretreatment Wash the plate with the mobile phase and allow to dry in air.

Mobile phase concentrated ammonia R, methanol R, methylene chloride R (1:40:60 V/V/V).

Application 5 µL. Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 50 mg in 5 mL of water R. Add 2 mL of dilute

sodium hydroxide solution R and shake with 2 quantities, each

of 5 mL, of methylene chloride R. The aqueous layer, acidified

by addition of nitric acid R, gives reaction (b) of phosphates

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined

in water R and dilute to 5.0 mL with the same solvent.

To 1.0 mL of this solution add 0.2 mL of concentrated

ammonia R and shake with 10.0 mL of the mobile phase.

Use the clear lower layer.

Reference solution (a) Dissolve 50 mg of primaquine

diphosphate CRS in water R and dilute to 5.0 mL with the

same solvent. To 1.0 mL of this solution add 0.2 mL of the

mobile phase. Use the clear lower layer.

Reference solution (b) Dilute 3.0 mL of the test solution to

100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to

10.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.2$ m, $\phi = 4.6$ mm;

— stationary phase: silica gel for chromatography R (10 µm).

Mobile phase concentrated ammonia R, methanol R, hexane R,

methylene chloride R (0.1:10:45:45 V/V/V/V).

Flow rate 3.0 mL/min.

Detection Spectrophotometer at 261 nm.

Injection 20 µL.

Run time At least twice the retention time of primaquine.

System suitability:

— the chromatogram obtained with reference solution (a)

shows just before the principal peak a peak whose area is

about 6 per cent of that of the principal peak;

— resolution: minimum 2.0 between the peak just before the

principal peak and the principal peak in the

chromatogram obtained with reference solution (a);

— **signal-to-noise ratio:** minimum 5 for the principal peak in the chromatogram obtained with reference solution (c).

Limits:
— **total:** not more than the area of the principal peak in the chromatogram obtained with reference solution (b)

— **disregard limit:** the area of the principal peak in the chromatogram obtained with reference solution (c)

(0.2 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.2000 g in 40 mL of anhydrous acetic acid R,

heating gently. Allow to cool and titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 22.77 mg

of $C_{15}H_{27}N_3O_4P_2$.

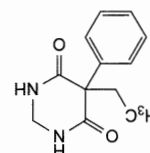
STORAGE

Protected from light.

Ph Eur

Primidone

(Ph. Eur. monograph 0584)



$C_{12}H_{14}N_2O_2$

218.3

125-33-7

Action and use

Antiepileptic.

Preparations

Primidone Oral Suspension

Primidone Tablets

Ph Eur

DEFINITION

5-Ethyl-5-phenylidihydropyrimidine-4,6-(1H,5H)-dione.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, slightly soluble in ethanol

(96 per cent). It dissolves in alkaline solutions.

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Use the solution prescribed for the assay. Examined between 240 nm and 300 nm (2.2.25), the solution shows 3 absorption maxima, at 252 nm, 257 nm and 264 nm, and 2 absorption minima, at 254 nm and 261 nm. The ratio of the absorbance measured at the absorption maximum at 257 nm to that measured at the absorption minimum at

261 nm is 2.00 to 2.20. The identification is valid if, in the test for resolution (2.2.25), the ratio of the absorbances is not less than 2.0.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of potassium bromide R.

Comparison primidone CRS.

C. Dissolve 0.1 g in 5 mL of a 5 g/L solution of chromotropic acid, sodium salt R in a mixture of 4 volumes of water R and 9 volumes of sulfuric acid R. A pinkish-blue colour develops on heating.

D. Mix 0.2 g and 0.2 g of anhydrous sodium carbonate R. Heat until the mixture melts. Ammonia is evolved which is detectable by its alkaline reaction (2.2.4).

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in methanol R1 and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R1. Dilute 1.0 mL of this solution to 10.0 mL with methanol R1.

Reference solution (b) Dissolve 5 mg of primidone for peak identification CRS (containing impurities A, B, C, D,

E and F) in methanol R1 and dilute to 5 mL with the same solvent.

Column:

— **size:** $l = 0.10$ m, $\varnothing = 4.6$ mm,

— **stationary phase:** monolithic octadecylsilyl silica gel for chromatography R.

Mobile phase:

— **mobile phase A:** 1.36 g/L solution of potassium dihydrogen phosphate R,

— **mobile phase B:** methanol R1,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 1	75	25
1 - 6	75 → 40	25 → 60
6 - 8	40	60
8 - 8.5	40 → 75	60 → 25
8.5 - 10	75	25

Flow rate 3.2 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with primidone for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks.

Relative retention With reference to primidone (retention time = about 2.2 min): impurity A = about 0.5;

impurity B = about 1.4; impurity C = about 1.6;

impurity D = about 1.75; impurity E = about 2.0;

impurity F = about 2.8.

System suitability: reference solution (b):

— **resolution:** minimum 2.5 between the peaks due to impurity B and impurity C.

Limits:

— **correction factors:** for the calculation of contents, multiply the peak areas of the following impurities by the

corresponding correction factor: impurity A = 1.5; impurity C = 1.5; impurity D = 1.4; impurity E = 1.3; impurity F: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent); impurities A, B, C, D, E: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent); any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent); *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent); *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with limit test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

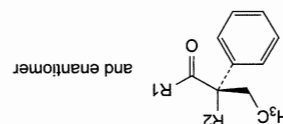
ASSAY

Dissolve 60.0 mg with heating in 70 mL of ethanol (96 per cent) R, cool and dilute to 100.0 mL with the same solvent. Prepare a reference solution in the same manner using 60.0 mg of *primidone* CRS. Measure the absorbance (2.2.25) of the 2 solutions at the absorbance maximum at 257 nm.

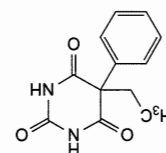
Calculate the content of $C_{12}H_{14}N_2O_2$ from the absorbances measured and the concentrations of the solutions.

IMPURITIES

Specified impurities A, B, C, D, E, F.

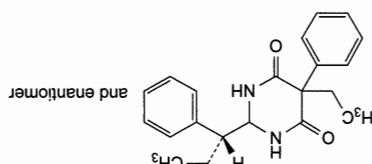


- A. R1 = NH₂, R2 = CO-NH₂, 2-ethyl-2-phenylpropionamide (ethylphenylalaninamide),
 C. R1 = NH₂, R2 = H: (2RS)-2-phenylbutanamide,
 D. R1 = NH₂, R2 = CN: (2RS)-2-cyano-2-phenylbutanamide,
 E. R1 = OH, R2 = H: (2RS)-2-phenylbutanoic acid,



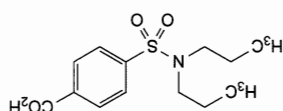
B. 5-ethyl-5-phenylpyrimidine-2,4,6-(1H,3H,5H)-trione (phenobarbital),

F. 5-ethyl-5-phenyl-2-[(1RS)-1-phenylpropyl]dihydropyrimidine-4,6-(1H,5H)-dione.



Probeneid

(Ph. Eur. monograph 0243)



$C_{13}H_{19}NO_4S$

285.4

57-66-9

Action and use

Anticancer drug.

Preparation

Probeneid Tablets

Ph. Eur.

DEFINITION

4-(Dipropylsulfonyl)benzoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or small crystals.

Solubility

Practically insoluble in water, soluble in acetone, sparingly soluble in anhydrous ethanol.

IDENTIFICATION

First identification: A, B, C.

Second identification: A, B, D.

A. Melting point (2.2.14): 197 °C to 202 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 20 mg in a mixture of 1 volume of 0.1 M hydrochloric acid and 9 volumes of ethanol (96 per cent) R and dilute to 100.0 mL with the same mixture of solvents. Dilute 5.0 mL of the solution to 100.0 mL with a mixture of 1 volume of 0.1 M hydrochloric acid and 9 volumes of ethanol (96 per cent) R.

Spectral range 220-350 nm.

Absorption maxima At 223 nm and 248 nm.

Specific absorbance at the absorption maximum at 248 nm 310 to 350.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison probeneid CRS.

D. Dissolve 0.2 g in the smallest necessary quantity of dilute ammonia R2 (about 0.6 mL). Add 3 mL of silver nitrate solution R2. A white precipitate is formed which dissolves in an excess of ammonia.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 1.0 g in 1 M sodium hydroxide and dilute to 10 mL with the same solvent.

Acidity

To 2.0 g add 100 mL of water R and heat on a water-bath for 30 min. Make up to the original volume with water R.

allow to cool to room temperature and filter. To 50 mL of the filtrate add 0.1 mL of phenolphthalein solution R.

Not more than 0.5 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Related substances

Liquid chromatography (2.2.29).

Solution A Mix 1 volume of glacial acetic acid R and 100 volumes of acetonitrile R.

Solution B Mix 1 volume of glacial acetic acid R and 100 volumes of a 6.9 g/L solution of sodium dihydrogen phosphate monohydrate R in water for chromatography R and adjust to pH 3.0 with dilute phosphoric acid R.

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of potassium 4-sulfobenzoate R (impurity A) in the mobile phase and dilute

to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped phenylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Solution A, solution B (50:50 V/V).

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 μ L.

Run time 4 times the retention time of probenecid.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to

impurity A.

Relative retention With reference to probenecid (retention time = about 7 min): impurity A = about 0.3.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to

impurity A and probenecid.

Calculation of percentage contents:

— correction factor: multiply the peak area of impurity A

by 0.4;

— for each impurity, use the concentration of probenecid in

Limits:

— impurity A: maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.2 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture water R, acetone R (20:80 V/V).

0.500 g complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R, shaking and heating slightly if necessary. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 28.54 mg of C₁₃H₁₉NO₄S.

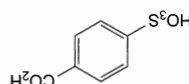
IMPURITIES

Specified impurities A

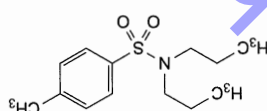
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C,

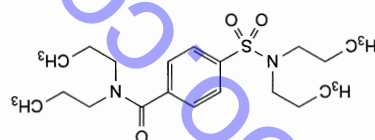
D.



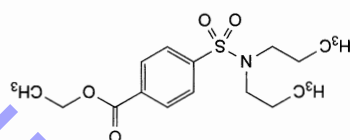
A. 4-sulfobenzoic acid,



B. 4-methyl-N,N-dipropylbenzenesulfonamide,



C. 4-(dipropylsulfamoyl)-N,N-dipropylbenzamide,

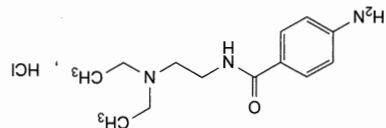


D. ethyl 4-(dipropylsulfamoyl)benzoate.

Ph Eur

Procainamide Hydrochloride

(Ph. Eur. monograph 0567)

 $C_{13}H_{22}ClN_3O$ 271.8 614-39-1

Action and use

Class I antiarrhythmic.

Preparations

Procainamide Injection

Procainamide Tablets

Ph Eur

DEFINITION

Procainamide hydrochloride contains not less than 98.0 per cent and not more than the equivalent of

(diethylamino)ethyl[benzamide hydrochloride, calculated with

reference to the dried substance.

CHARACTERS

A white or very slightly yellow, crystalline powder, hygroscopic, very soluble in water, freely soluble in alcohol, slightly soluble in acetone.

IDENTIFICATION

First identification C, D.

Second identification A, B, D, E.

A. Melting point (2.2.14): 166 °C to 170 °C.

B. Dissolve 10.0 mg in 0.1 M sodium hydroxide and dilute to

100.0 mL with the same solvent. Dilute 10.0 mL of the

solution to 100.0 mL with 0.1 M sodium hydroxide. Examine

between 220 nm and 350 nm (2.2.25), the solution shows an

absorption maximum at 273 nm. The specific absorbance at

the maximum is 580 to 610.

C. Examine by infrared absorption spectrophotometry

(2.2.24), comparing with the spectrum obtained with

procainamide hydrochloride CRS.

D. Dilute 1 mL of solution S to 5 mL with water R.

The solution gives reaction (a) of chlorides (2.3.1).

E. Dilute 1 mL of solution S (see Tests) to 2 mL with

water R. 1 mL of this solution gives the reaction of primary

aromatic amines (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to

25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution B₆ (2.2.2, Method II).

pH (2.2.3)

The pH of solution S is 5.6 to 6.3.

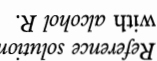
Related substances

Examine by thin-layer chromatography (2.2.27), using silica

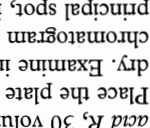
gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.10 g of the substance to be examined

in alcohol R and dilute to 10 mL with the same solvent.



(Ph. Eur. monograph 0567)

 $C_{13}H_{22}ClN_3O$ 271.8 614-39-1

Action and use

Class I antiarrhythmic.

Preparations

Procainamide Injection

Procainamide Tablets

Ph Eur

DEFINITION

Procainamide hydrochloride contains not less than

98.0 per cent and not more than the equivalent of

(diethylamino)ethyl[benzamide hydrochloride, calculated with

reference to the dried substance.

A white or very slightly yellow, crystalline powder, hygroscopic, very soluble in water, freely soluble in alcohol, slightly soluble in acetone.

IDENTIFICATION

First identification C, D.

Second identification A, B, D, E.

A. Melting point (2.2.14): 166 °C to 170 °C.

B. Dissolve 10.0 mg in 0.1 M sodium hydroxide and dilute to

100.0 mL with the same solvent. Dilute 10.0 mL of the

solution to 100.0 mL with 0.1 M sodium hydroxide. Examine

between 220 nm and 350 nm (2.2.25), the solution shows an

absorption maximum at 273 nm. The specific absorbance at

the maximum is 580 to 610.

C. Examine by infrared absorption spectrophotometry

(2.2.24), comparing with the spectrum obtained with

procainamide hydrochloride CRS.

D. Dilute 1 mL of solution S to 5 mL with water R.

The solution gives reaction (a) of chlorides (2.3.1).

E. Dilute 1 mL of solution S (see Tests) to 2 mL with

water R. 1 mL of this solution gives the reaction of primary

aromatic amines (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to

25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

than reference solution B₆ (2.2.2, Method II).

pH (2.2.3)

The pH of solution S is 5.6 to 6.3.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica

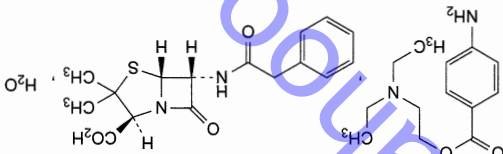
gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 0.10 g of the substance to be examined

in alcohol R and dilute to 10 mL with the same solvent.

Procaine Benzylpenicillin

(Ph. Eur. monograph 0115)

 $C_{29}H_{38}N_4O_6S_2H_2O$ 588.7

6130-64-9

Action and use

Penicillin antibacterial.

Ph Eur

DEFINITION

(2S,5R,6R)-3,3-Dimethyl-7-oxo-6-[(phenylacetyl)amino]-4-

thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid compound

with 2-(diethylamino)ethyl 4-aminobenzoate monohydrate.

Substance produced by the growth of certain strains of

Penicillium notatum or related organisms, or obtained by any

Content

— procaine benzylpenicillin: 96.0 per cent to 102.0 per cent

(anhydrous substance);

— procaine ($C_{13}H_{20}N_2O_2$; 236.3): 39.0 per cent to

42.0 per cent (anhydrous substance).

Dispersing or suspending agents (for example, lecithin and

poly sorbate 80) may be added.

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION**First identification A**

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison procaine benzylpenicillin CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of acetone R.

Reference solution Dissolve 25 mg of procaine benzylpenicillin CRS in 5 mL of acetone R.

Plate TLC silanised silica gel plate R.

Mobile phase Mix 30 volumes of acetone R and 70 volumes of a 154 g/L solution of ammonium acetate R previously adjusted to pH 7.0 with ammonia R.

Application 1 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Expose to iodine vapour until the spots appear and examine in daylight.

System suitability: reference solution:

— the chromatogram shows 2 clearly separated spots.

Results The 2 principal spots in the chromatogram obtained with the test solution are similar in position, colour and size to the 2 principal spots in the chromatogram obtained with the reference solution.

C. Place about 2 mg in a test-tube about 150 mm long and add 2 mL of sulfuric acid-formaldehyde reagent R. Mix the contents of the tube by swirling; the solution is practically colourless. Place the test-tube on a water-bath for 1 min; a reddish-brown colour develops.

D. Dissolve 0.1 g in 2 mL of dilute hydrochloric acid R and use the solution which may be turbid. The solution gives the reaction of primary aromatic amines (2.3.1).

TESTS

pH (2.2.3)

5.0 to 7.5.

Dissolve 50 mg in carbon dioxide-free water R and dilute to 15 mL with the same solvent. Shake until dissolution is complete.

Specific optical rotation (2.2.7)

+ 165 to + 180 (anhydrous substance).

Dissolve 0.250 g in a mixture of 2 volumes of water R and 3 volumes of acetone R, then dilute to 25.0 mL with the same mixture of solvents.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution (a) Dissolve 70.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dissolve 70.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

ASSAY

Procedure for the removal of bacterial endotoxins. of parenteral preparations without a further appropriate Less than 0.10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate

Bacterial endotoxins (2.6.14, Method B)

2.8 per cent to 4.2 per cent, determined on 0.500 g.

Water (2.5.12)

(1 per cent).

chromatogram obtained with reference solution (c)

area of the peak due to benzylpenicillin in the

— any other impurity: for each impurity, not more than the solution (c) (0.024 per cent);

peak in the chromatogram obtained with reference

— impurity A: not more than the area of the corresponding

Limits:

concentration of acetonitrile in the mobile phase.

impurity A and procaine; if necessary, adjust the

— resolution: minimum 2.0 between the peaks due to

System suitability: reference solution (b):

Elution order Impurity A, procaine, benzylpenicillin.

Run time 1.5 time the retention time of benzylpenicillin.

and (c).

Injection 10 µL of test solution (a) and reference solutions (b)

Detection Spectrophotometer at 225 nm.

Flow rate 1.75 mL/min.

mixture to pH 7.2 with dilute phosphoric acid R.

pH 7.0 with 1 M potassium hydroxide; if necessary, adjust the

potassium dihydrogen phosphate R and 6.5 g/L of

water R and 500 mL of a solution containing 14 g/L of

Mobile phase Mix 250 mL of acetonitrile R1, 250 mL of

(5 µm).

— stationary phase: octadecylsilyl silica gel for chromatography R

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

Column:

solution (a) and dilute to 100.0 mL with the mobile phase.

with water R. To 1.0 mL of this solution, add 1.0 mL with

the same solvent. Dilute 1.0 mL of the solution to 10.0 mL

acid R (impurity A) in water R and dilute to 50.0 mL with

Reference solution (c) Dissolve 16.8 mg of 4-aminobenzoic

with reference solution (a).

(impurity A) in reference solution (a) and dilute to 25 mL

Reference solution (b) Dissolve 4 mg of 4-aminobenzoic acid R

100.0 mL with the mobile phase.

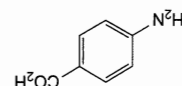
benzylpenicillin CRS in the mobile phase and dilute to

Reference solution (a) Dissolve 70.0 mg of procaine

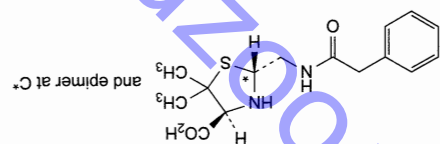
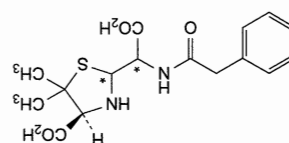
benzylpenicillin, procaine II-673

IMPURITIES

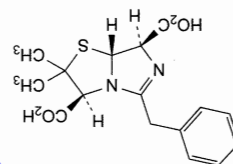
A. 4-aminobenzoic acid,



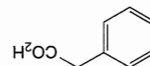
B. (4S)-2-[carboxy[(phenylacetyl)amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penicilloic acids of benzylpenicillin),



C. (2R,4S)-2-[[[(phenylacetyl)amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of benzylpenicillin),



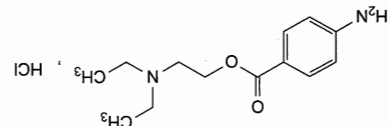
D. (3S,7R,7aR)-5-benzyl-2,2-dimethyl-3,7,7a-tetrahydroimidazo[5,1-b]thiazole-3,7-dicarboxylic acid (penillilic acid of benzylpenicillin),



E. phenylacetic acid.

Procaine Hydrochloride

(Ph. Eur. monograph 0050)

 $C_{13}H_{21}ClN_2O_2$

272.8

51-05-8

Action and use
Local anaesthetic.

DEFINITION

Procaine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 2-(diethylamino)ethyl 4-aminobenzoate hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, very soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B, E.

Second identification A, C, D, E, F.

A. Melting point (2.2.14): 154 °C to 158 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with procaine hydrochloride CRS.

C. To about 5 mg add 0.5 mL of fuming nitric acid R.

Evaporate to dryness on a water-bath, allow to cool and dissolve the residue in 5 mL of acetone R. Add 1 mL of

0.1 M alcoholic potassium hydroxide. Only a brownish-red colour develops.

D. To 0.2 mL of solution S (see Tests) add 2 mL of water R

1 g/L solution of potassium permanganate R. The colour is immediately discharged.

E. It gives reaction (a) of chlorides (2.3.1).

F. Dilute 1 mL of solution S to 100 mL with water R. 2 mL of this solution gives the reaction of primary aromatic amines (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

Dilute 4 mL of solution S to 10 mL with carbon dioxide-free water R. The pH of the solution is 5.0 to 6.5.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution Dissolve 1.0 g of the substance to be examined in water R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of 4-aminobenzoic acid R in water R and dilute to 100 mL with the same solvent. Dilute

1 mL of the solution to 10 mL with water R.

Apply separately to the plate 5 µL of each solution. Develop over a path of 10 cm using a mixture of 4 volumes of glacial acetic acid R, 16 volumes of hexane R and 80 volumes of

dibutyl ether R. Dry the plate at 100 °C to 105 °C for 10 min and examine in ultraviolet light at 254 nm. Any spot in the

chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the

chromatogram obtained with the reference solution (0.05 per cent). The principal spot in the chromatogram

obtained with the test solution remains on the point of application.

Heavy metals (2.4.8)

Dissolve 1.0 g in water R and dilute to 25.0 mL with the same solvent. Carry out the prefiltration. 10 mL of the

prefiltrate complies with test E (5 ppm). Prepare the reference solution using 5 mL of lead standard solution

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.00 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.400 g in 50 mL of dilute hydrochloric acid R. Carry out the determination of primary aromatic amino nitrogen (2.5.8).

1 mL of 0.1 M sodium nitrite is equivalent to 27.28 mg of $C_{13}H_{21}ClN_3O_2$.

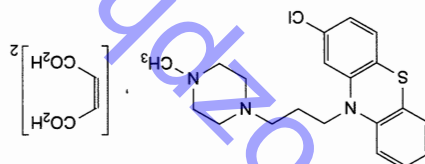
STORAGE

Store protected from light.

Ph Eur

Prochlorperazine Maleate

(Ph. Eur. monograph 0244)



$C_{28}H_{32}ClN_3O_8$ 606 84-02-6

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparations

Prochlorperazine Tablets
Prochlorperazine Buccal Tablets

Ph Eur

DEFINITION

2-Chloro-10-[3-(4-methylpiperazin-1-yl)propyl]-10H-phenothiazine bis[hydrogen (Z)-butenedioate].

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or pale-yellow, crystalline powder.

Solubility

Very slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification B, C, D

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25). Carry out the identification test protected from light and measure the absorbances immediately.

Test solution (a) Dissolve 50 mg in 0.1 M hydrochloric acid and dilute to 500.0 mL with the same acid.

Test solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with 0.1 M hydrochloric acid.

Spectral range 280-350 nm for test solution (a); 230-280 nm for test solution (b).

Absorption maximum At 305 nm for test solution (a); at 255 nm for test solution (b).

Specific absorbance at the absorption maximum at 255 nm 525 to 575 for test solution (b).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison prochlorperazine maleate CRS.

C. Identification test for phenothiazines by thin-layer chromatography (2.3.3) with the following modifications.

Test solution Dissolve 20 mg of the substance to be examined in a mixture of equal volumes of methanol R and methylene chloride R and dilute to 20 mL with the same mixture of solvents.

Reference solution Dissolve 20 mg of prochlorperazine maleate CRS in a mixture of equal volumes of methanol R and methylene chloride R and dilute to 20 mL with the same mixture of solvents.

Application 4 µL.

D. Triturate 0.2 g with a mixture of 1 mL of strong sodium hydroxide solution R and 3 mL of water R. Shake with 3 quantities, each of 5 mL, of ether R. To 0.1 mL of the aqueous layer add a solution of 10 mg of resorcinol R in 3 mL of sulfuric acid R. Heat in a water-bath for 15 min. No colour develops. To the remainder of the aqueous layer add 2 mL of brown solution R. Heat in a water-bath for 15 min and then heat to boiling. Cool. To 0.1 mL of the solution add a solution of 10 mg of resorcinol R in 3 mL of sulfuric acid R. Heat in a water-bath for 15 min. A blue colour develops.

TESTS

pH (2.2.3)

3.0 to 4.0 for a freshly prepared saturated solution in carbon dioxide-free water R.

Related substances

Thin-layer chromatography (2.2.27). Carry out the test protected from light.

Solvent mixture diethylamine R, methanol R (5:95 V/V).

Test solution Dissolve 0.2 g of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture. Prepare the solution immediately before use.

Reference solution Dilute 1 mL of the test solution to 200 mL with the solvent mixture.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase acetone R, diethylamine R, cyclohexane R (10:10:80 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Limit Any spot, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent); disregard any spots remaining at the points of application.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

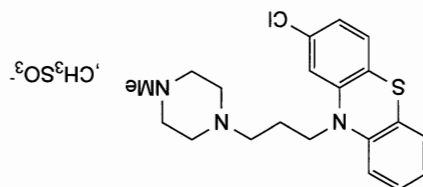
Dissolve 0.200 g of the powdered substance to be examined in 50 mL of anhydrous acetic acid R, warming on a water-bath. Allow to cool to room temperature. Titrate with 0.1 M perchloric acid determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 30.31 mg of $C_{28}H_{32}ClN_3O_8$.

STORAGE

Protected from light.

Prochlorperazine Mesilate


 $C_{20}H_{24}ClN_3S_2CH_4SO_3$ 566.2 5132-55-8

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparations

Prochlorperazine Injection
Prochlorperazine Oral Solution

DEFINITION

Prochlorperazine Mesilate is 2-chloro-10-[3-(4-methylpiperazin-1-yl)propyl]phenothiazine di(methanesulfonate). It contains not less than 98.0% and not more than 101.0% of $C_{20}H_{24}ClN_3S_2CH_4SO_3$ calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder.
Very soluble in water; sparingly soluble in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of prochlorperazine mesilate (RS 290).

B. Dissolve 5 mg in 2 mL of sulfuric acid and allow to stand for 5 minutes. A red colour is produced.

TESTS

Acidity

pH of a 2% w/v solution, 2.0 to 3.0, Appendix V L.

Related substances

Complies with the test for related substances in phenothiazines, Appendix III A, using mobile phase A and dissolving the substance being examined in methanol containing 0.5% v/v of 13.5M ammonia.

Loss on drying

When dried to constant weight at 100° at a pressure not exceeding 0.7 kPa, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

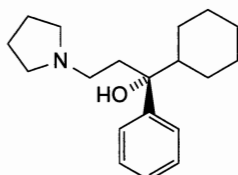
ASSAY

Dissolve 0.8 g in 10 mL of water, add 5 mL of 1M sodium hydroxide and extract by shaking with successive quantities of 50, 25, 25 and 25 mL of ether. Wash the combined ether extracts with 5 mL of water, shake the washings with 5 mL of ether, add the ether to the combined ether extracts and evaporate to dryness. Add 2 mL of absolute ethanol to the residue, evaporate to dryness and carry out Method I for non-aqueous titration, Appendix VIII A, using 1-naphtholbenzenesulfonic acid VS as indicator. Each mL of $C_{20}H_{24}ClN_3S_2CH_4SO_3$ is equivalent to 28.31 mg of

STORAGE

Prochlorperazine Mesilate should be protected from light.

Procyclidine Hydrochloride


 $C_{19}H_{29}NO_2HCl$ 323.9 1508-76-5

Action and use

Anticholinergic.

Preparations

Procyclidine Injection
Procyclidine Tablets

DEFINITION

Procyclidine Hydrochloride is (RS)-1-cyclohexyl-1-phenyl-3-pyrrolidin-1-ylpropan-1-ol hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{19}H_{29}NO_2HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white, crystalline powder.
Sparingly soluble in water; soluble in ethanol (96%); practically insoluble in acetone and in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of procyclidine hydrochloride (RS 292).

B. Dissolve 0.25 g in 10 mL of water, make alkaline with 5M ammonia and extract with three 10-mL quantities of ether. Dry the combined extracts over anhydrous sodium sulfate, filter, evaporate the ether and scratch the residue with a glass rod to induce solidification. The melting point of the residue is about 85°, Appendix V A.

Acidity

pH of a 1% w/v solution, 4.5 to 6.5, Appendix V L.

Related substances

A. Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in chloroform.
(1) 2.0% w/v of the substance being examined.
(2) 0.0040% w/v of 1-phenyl-3-pyrrolidinopropan-1-ol hydrochloride BPCRS.
(3) 0.010% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel F₂₅₄.
(b) Use the mobile phase as described below.
(c) Apply 5 µL of each solution.
(d) Develop the plate to 15 cm.
(e) After removal of the plate, dry it at 105° for 15 minutes and examine under ultraviolet light (254 nm). Spray the plate with dilute potassium iodobismuthate solution and examine.

MOBILE PHASE

1 volume of 13.5M ammonia and 100 volumes of ether.

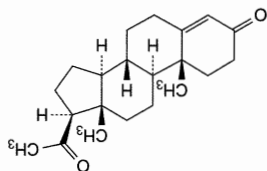
LIMITS

When examined under UV light:



(Ph. Eur. monograph 0429)

Progesterone



314.5

 $C_{21}H_{30}O_2$

57-83-0

Action and use

Progesterone

Progesterone Injection

Ph Eur

DEFINITION

Pregn-4-ene-3,20-dione.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

Solubility

Practically insoluble in water, freely soluble in anhydrous ethanol, sparingly soluble in acetone and in fatty oils.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison progesterone CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in anhydrous ethanol R, evaporate to

dryness and record new spectra using the residues.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of 1 volume of methanol R and 9 volumes of methylene chloride R and dilute to 10 mL with the same mixture of solvents.

Reference solution Dissolve 10 mg of progesterone CRS in a mixture of 1 volume of methanol R and 9 volumes of methylene chloride R and dilute to 10 mL with the same mixture of solvents.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ethyl acetate R, methylene chloride R (33:66 V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Specific optical rotation (2.2.7)

+ 186 to + 194 (dried substance).

any spot corresponding to 1-phenyl-3-pyrrolidinopropan-1-one in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.2%).

When examined after spraying:

any other secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (3) (0.5%).

B. Carry out the method for gas chromatography,

Appendix III B. Use a solution containing 0.06% w/v of triphenylethylene in ether as the internal standard.

(1) Add 5 mL of 1.25M sodium hydroxide to 20 mL of a

0.50% w/v solution of the substance being examined and

mix. Extract with two 20-mL quantities of ether, add to the

combined extracts 5 mL of the internal standard, shake with

anhydrous sodium sulfate and filter; evaporate the filtrate and

dissolve the residue in 1 mL of ether.

(2) Prepare in the same manner as solution (1) but omitting

the addition of the internal standard solution.

(3) Prepare in the same manner as solution (1) but using

20 mL of a 0.015% w/v solution of the substance being

examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use a fused silica column (15 m × 0.32 mm) bonded

with a 0.25 µm film of polyethylene glycol 20,000 (DB-Wax is

suitable).

(b) Use nitrogen as the carrier gas at 40 mL per minute.

(c) Use isothermal conditions maintained at 210°.

(d) Use an inlet temperature of 250°.

(e) Use a flame ionisation detector at a temperature of 250°.

(f) Inject 1 µL of each solution.

(g) Use a split ratio of 1:5.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with

solution (3) shows two clearly separated peaks.

LIMITS

The ratio of the sum of the areas of any secondary peaks to

the area of the peak due to the internal standard in the

chromatogram obtained with solution (1) is not more than

the ratio of the area of the principal peak to the area of the

internal standard peak in the chromatogram obtained with

solution (3) (3%).

Loss on drying

When dried to constant weight at 105°, loses not more than

0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for non-aqueous titration,

Appendix VIII A, using 0.7 g and crystal violet solution as

indicator. Each mL of 0.1M perchloric acid VS is equivalent to

32.39 mg of C₁₉H₂₉NO₂HCl.

Dissolve 0.250 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 20.0 mg of the substance to be examined in mobile phase B and dilute to 50.0 mL with mobile phase B.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase B. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase B.

Reference solution (b) Dissolve 2 mg of progesterone for system suitability CRS (containing impurities B, C, G, I and M) in mobile phase B and dilute to 5.0 mL with mobile phase B.

Reference solution (c) Dissolve 2 mg of progesterone for peak identification CRS (containing impurities D, E, J, K and L) in mobile phase B and dilute to 5.0 mL with mobile phase B.

Reference solution (d) Dissolve 10.0 mg of progesterone for impurity H identification CRS (with an assigned content of impurity H) in mobile phase B and dilute to 25.0 mL with mobile phase B.

Reference solution (e) Dissolve 20.0 mg of progesterone CRS in mobile phase B and dilute to 50.0 mL with mobile phase B.

Column:
 size: $l = 0.15$ m, $\phi = 4.6$ mm;
 stationary phase: end-capped extra-dense bonded octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:
 mobile phase A: acetonitrile R, water R (50:50 V/V);
 mobile phase B: water R, acetonitrile R (20:80 V/V);

Time (min)
 Mobile phase A (per cent V/V)
 Mobile phase B (per cent V/V)

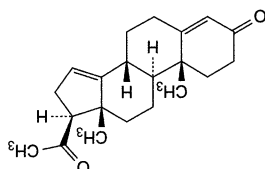
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 241 nm and, for impurity H, at 286 nm.

Injection 10 μ L of the test solution and reference solutions (a), (b), (c) and (d).

Identification of impurities Use the chromatogram supplied with progesterone for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B, C, G, I and M; use the chromatogram supplied with progesterone for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities D + E, J, K and L; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity H.

Relative retention With reference to progesterone (retention time = about 14 min): impurity B = about 0.60; impurity J = about 0.65; impurity H = about 0.82; impurity K = about 0.85; impurity C = about 0.93; impurity M = about 1.1; impurity L = about 1.90; impurity I = about 1.95; impurities D and E = about 2.05; impurity G = about 2.65.

System suitability: reference solution (b):
 peak-to-valley ratio: minimum 4.0, where H_p = height above the baseline of the peak due to impurity M and H_v = height above the baseline of the lowest point of the



A. pregna-4,14-diene-3,20-dione,

curve separating this peak from the peak due to progesterone.

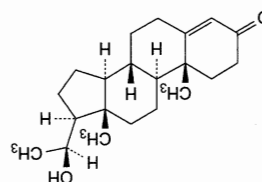
Limits:
 impurity I (sum of the 2 epimers): not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);
 impurity C: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);
 impurity B: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
 sum of impurities D and E: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
 impurities G, J, K, L, M: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
 sum of impurities other than H: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
 disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)
 Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C for 2 h.

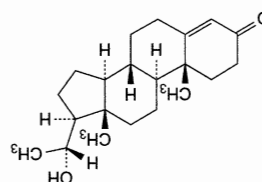
ASSAY
 Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection Test solution and reference solution (e).
 Calculate the percentage content of $C_{21}H_{30}O_2$ taking into account the assigned content of progesterone CRS.

STORAGE
 Protected from light.

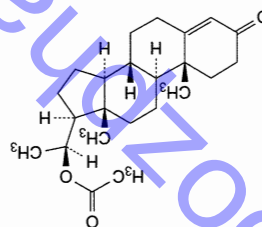
IMPURITIES
 Specified impurities B, C, D, E, G, H, I, J, K, L, M
 Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A.



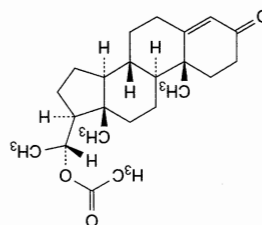
B. (20S)-20-hydroxypregnen-4-en-3-one,



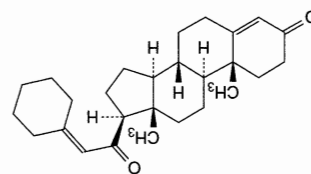
C. (20R)-20-hydroxypregnen-4-en-3-one,



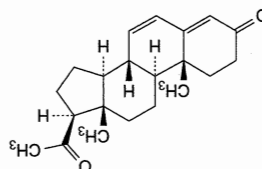
D. (20S)-3-oxopregnen-4-en-20-yl acetate,



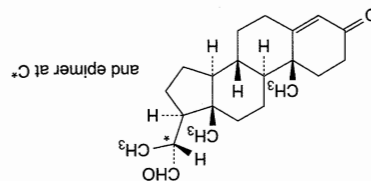
E. (20R)-3-oxopregnen-4-en-20-yl acetate,



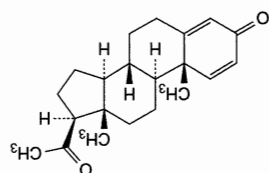
G. 21-(cyclohexylidene)pregnen-4-ene-3,20-dione,



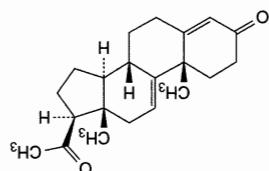
H. pregna-4,6-diene-3,20-dione (Δ6-progesterone),



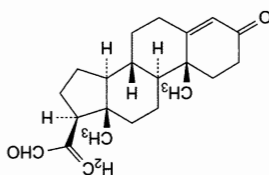
I. (20RS)-20-methyl-3-oxopregnen-4-en-21-al,



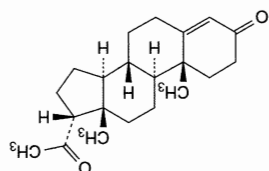
J. pregna-1,4-diene-3,20-dione,



K. pregna-4,9(11)-diene-3,20-dione,



L. 20-methylidene-3-oxopregnen-4-en-21-al,

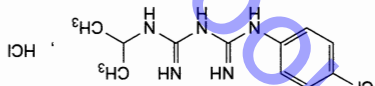


M. (17α)-pregnen-4-ene-3,20-dione.

Ph Eur

Proguanil Hydrochloride

(Ph. Eur. monograph 2002)

 $C_{11}H_{17}ClN_5$

290.2

637-32-1

Action and use

Antiprotozoal (malaria).

Ph Eur

DEFINITION

1-(4-Chlorophenyl)-5-(1-methylethyl)biguanide hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in ethanol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification A, D

Second identification B, C, D



A. Infrared absorption spectrophotometry (2.2.24).
Comparison Ph. Eur. reference spectrum of proguanil hydrochloride.

B. Dissolve 0.4 g in 50 mL of water R (solution A).
To 15 mL of solution A add 2 mL of dilute sodium hydroxide solution R. Extract with 20 mL of ethyl acetate R. Wash the organic layer with water R, evaporate to dryness and dry at 105 °C. The melting point (2.2.14) of the residue is 130 °C to 133 °C.

C. To 10 mL of solution A, add 1 drop of copper sulfate solution R and 2 mL of dilute ammonia R1. Add 5 mL of toluene R and stir. Allow to stand until separation of the layers is obtained. The upper layer is violet-red.
D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Acidity or alkalinity

To 35 mL of water R maintained at 60–65 °C, add 0.2 mL of methyl red mixed solution R. Neutralise to a grey colour with either 0.01 M sodium hydroxide or 0.01 M hydrochloric acid. Add 0.4 g of the substance to be examined and stir until completely dissolved. The solution is grey or green. Not more than 0.2 mL of 0.01 M hydrochloric acid is required to change the colour of the solution to reddish-violet.

Chloroaniline

Maximum 250 ppm.
Dissolve 0.10 g in 1 mL of 2 M hydrochloric acid R and dilute to 20 mL with water R. Cool to 5 °C. Add 1 mL of a 3.45 g/L solution of sodium nitrite R and allow to stand at 5 °C for 5 min. Add 2 mL of a 50 g/L solution of ammonium naphthylethylenediamine dihydrochloride solution R, dilute to 50 mL with water R and allow to stand for 30 min. Any red colour produced is not more intense than that of a standard prepared at the same time and in the same manner, using 20 mL of a 1.25 mg/L solution of chloroaniline R.

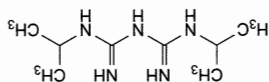
Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
Reference solution (b) Dissolve 5 mg of proguanil impurity C CRS in the mobile phase and dilute to 100 mL with the mobile phase. Dilute 0.1 mL to 10 mL with the mobile phase.
Reference solution (c) Dissolve 5 mg of proguanil impurity D CRS in the mobile phase and dilute to 100 mL with the mobile phase. Dilute 0.1 mL to 10 mL with the mobile phase.
Reference solution (d) Dilute 1 mL of the test solution to 200 mL with the mobile phase. To 1 mL add 1 mL of reference solution (c) and mix.

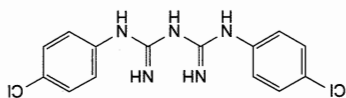
Column:

— size: $l = 0.125$ m, $\phi = 4.6$ mm,
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).
Mobile phase Dissolve 3.78 g of sodium hexanesulfonate R in a mixture of 10 volumes of glacial acetic acid R, 800 volumes of water R and 1200 volumes of methanol R.
Flow rate 1 mL/min.
Detection Spectrophotometer at 230 nm and 254 nm.

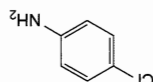
D. 1,5-bis(1-methylethyl)biguanide.



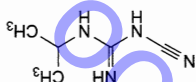
C. 1,5-bis(4-chlorophenyl)biguanide,



B. 4-chloroaniline,



A. 1-cyano-3-(1-methylethyl)guanidine,



IMPURITIES

Protected from light

STORAGE

$C_{11}H_{17}Cl_2N_5$.
1 mL of 0.1 M perchloric acid is equivalent to 14.51 mg of acid, determining the end-point potentiometrically (2.2.20).
Suspend 0.100 g in 20 mL of anhydrous acetic acid R, shake and heat at 50 °C for 5 min. Cool to room temperature and add 40 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

— total: the sum of the calculated percentage contents of known and unknown impurities is not greater than 1 per cent, considering each peak at the wavelength at which the peak shows the higher value,

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.2 per cent),

— any other impurity: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.2 per cent),

— impurity C: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

— impurity C: not more than 3.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

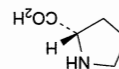
— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) at 230 nm (0.7 per cent),

Proline

(Ph. Eur. monograph 0785)



$C_5H_9NO_2$ 115.1 147-85-3

Action and use

Amino acid.

Ph. Eur.

DEFINITION

(2S)-Pyrrolidine-2-carboxylic acid.

Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance White or almost white, crystalline powder or colourless crystals.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B

Second identification A, C

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison proline CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a 1 per cent V/V solution of hydrochloric acid R and dilute to 50 mL with the same solution.

Reference solution Dissolve 10 mg of proline CRS in a 1 per cent V/V solution of hydrochloric acid R and dilute to 50 mL with the same solution.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).

Application 5 µL.

Development: Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Solution S

Dissolve 2.5 g in distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Specific optical rotation (2.2.7)

Dissolve 1.00 g in water R and dilute to 25.0 mL with the same solvent.

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method I.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dissolve 30.0 mg of alanine R (impurity A) in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 100.0 mL with solution A. Dilute 1.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of ammonium standard solution (100 ppm NH_4^+) R to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of isoleucine R and 30 mg of leucine R in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

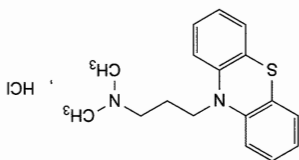
System suitability Reference solution (d):
— resolution: minimum 1.5 between the peaks due to isoleucine and leucine.Calculation of percentage contents:
— for any ninhydrin-positive substance detected at 570 nm, use the concentration of impurity A in reference solution (a);
— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b);
if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.Limits:
— any ninhydrin-positive substance, for each impurity, maximum 0.2 per cent;
— total: maximum 0.5 per cent;
— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Chlorides (2.4.4)
Maximum 200 ppm.Sulfates (2.4.13)
Dilute 5 mL of solution S to 15 mL with water R. Maximum 300 ppm.Ammonium
Dilute 10 mL of solution S to 15 mL with distilled water R. Amino acid analysis (2.2.56) as described in the test for ninhydrin-positive substances with the following modifications.

Promazine Hydrochloride

(Ph. Eur. monograph 1365)

C₁₇H₂₁ClN₂S 320.9 53-60-1

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparations

Promazine Injection

Promazine Tablets

DEFINITION

Promazine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 3-(10*H*-phenothiazin-10-yl)-*N,N*-dimethylpropan-1-amine hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, slightly hygroscopic, very soluble in water, in alcohol and in methylene chloride.

It melts at about 179 °C.

IDENTIFICATION

First identification A, B, D.

Second identification B, C, D.

A. Examine by infrared absorption spectrophotometry (2.2.4), comparing with the spectrum obtained with promazine hydrochloride CRS.

B. It complies with the identification test for phenothiazines by thin-layer chromatography (2.3.3). Use promazine hydrochloride CRS to prepare the reference solution.

C. Dissolve about 5 mg in 2 mL of sulfuric acid R and allow to stand for 5 min. An orange colour is produced.

D. It gives reaction (b) of chlorides (2.3.1).

TESTS

pH (2.2.3)

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent. The pH of the freshly prepared solution is 4.2 to 5.2.

Related substances

Carry out the test protected from bright light. Prepare the solutions immediately before use.

Examine by thin layer chromatography (2.2.27), using a TLC silica gel F₂₅₄ plate R.

Test solution Dissolve 0.10 g of the substance to be examined in a mixture of 5 volumes of diethylamine R and 95 volumes of methanol R and dilute to 10 mL with the same mixture of solvents.

Reference solution (a) Dilute 1 mL of the test solution to 200 mL with a mixture of 5 volumes of diethylamine R and 95 volumes of methanol R.

Reference solution (b) Dissolve 10 mg of chlorprothixene hydrochloride CRS in a mixture of 5 volumes of diethylamine R

— ammonium at 570 nm: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Limit:

solution.

Injection Test solution, reference solution (c) and blank

Iron (2.4.9)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R₁, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 3 mL of anhydrous formic acid R.

Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

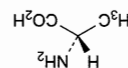
1 mL of 0.1 M perchloric acid is equivalent to 11.51 mg of C₅H₉NO₂.

STORAGE

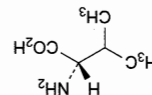
Protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B.



A. (2S)-2-aminopropanoic acid (alanine),



B. (2S)-2-amino-3-methylbutanoic acid (valine).

Ph Eur



DEFINITION
(2*RS*)-*N,N*-Dimethyl-1-[(10*H*-phenothiazin-10-yl)propan-2-amine hydrochloride.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or faintly yellowish, crystalline powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

mp: about 222 °C, with decomposition.

IDENTIFICATION

First identification A, B, D
Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison promethazine hydrochloride CRS.

B. It complies with the identification test for phenothiazines by thin-layer chromatography (2.3.3): use promethazine hydrochloride CRS to prepare the reference solution.

C. Dissolve 0.1 g in 3 mL of water R. Add dropwise 1 mL of nitric acid R. A precipitate is formed which rapidly dissolves to give a red solution, becoming orange and then yellow.

Heat to boiling. The solution becomes orange and an orange-red precipitate is formed.

D. It gives reaction (b) of chlorides (2.3.1).

TESTS

pH (2.2.3)

4.0 to 5.0, measured immediately after preparation.

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light and use freshly prepared solutions.

Solvent mixture methylaniline R, methanol R (1:1000 V/V).

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2.5 mg of promethazine for peak identification CRS (containing impurities A, B and C) in the solvent mixture and dilute to 5 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5.0 mg of promethazine impurity D CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1 mL of this solution to 100 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

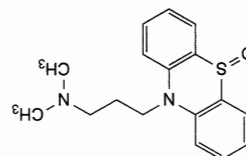
— stationary phase: end-capped octylsilyl silica gel for chromatography with polar incorporated groups R (5 μ m).

Mobile phase Mix 20 volumes of methanol R, 30 volumes of acetonitrile R and 50 volumes of a 3.4 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 7.0 with potassium hydroxide R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

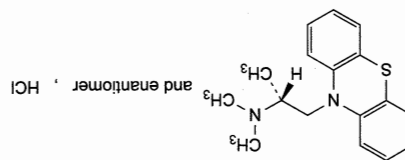
Injection 10 μ L.



A. 3-[(10*H*-phenothiazin-10-yl)-*N,N*-dimethylpropyl]-1-amine S-oxide (promazine sulfoxide).

Promethazine Hydrochloride

(Ph. Eur. monograph 0524)



$C_{17}H_{21}ClN_2S$ 320.9 58-33-3

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparations

Promethazine Injection

Promethazine Oral Solution

Promethazine Hydrochloride Tablets

Run time 2.5 times the retention time of promethazine. Identification of impurities Use the chromatogram supplied with promethazine for peak identification CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity D.

Relative retention With reference to promethazine (retention time = about 18 min): impurity D = about 0.2; impurity C = about 0.5; impurity B = about 1.4; impurity A = about 1.8.

System suitability: resolution: minimum 2.0 between the peaks due to impurities B and A in the chromatogram obtained with reference solution (a); the chromatogram obtained with reference solution (a) is similar to the chromatogram supplied with promethazine for peak identification CRS.

Limits: — correction factor for the calculation of content, multiply the peak area of impurity A by 0.5;

— impurity B: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity D: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 12 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 1.0 g in 5 mL of water R, then add 5 mL of acetone R and 5 mL of buffer solution pH 3.5 R. Carry out the prefiltration. The prefiltrate complies with test E. Prepare the reference solution using 5 mL of lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32) Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.

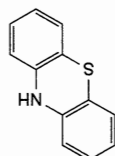
1 mL of 0.1 M sodium hydroxide is equivalent to 32.09 mg of $C_{17}H_{21}ClN_2S$.

STORAGE

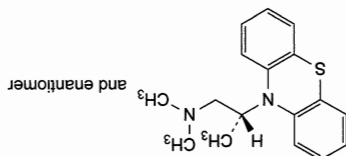
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IMPURITIES

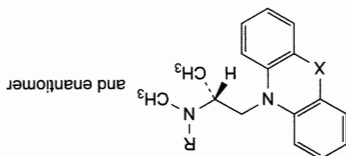
Specified impurities A, B, C, D



A. phenothiazine,



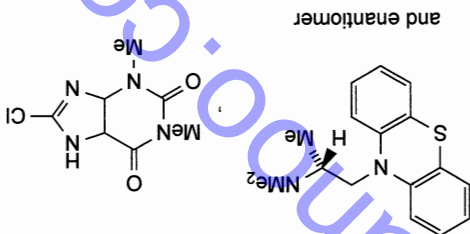
B. (2R,5R)-N,N'-dimethyl-2-(10H-phenothiazin-10-yl)propan-1-amine (isopromethazine),



C. R = H, X = S: (2R,5R)-N-methyl-1-(10H-phenothiazin-10-yl)propan-2-amine,

D. R = CH₃, X = SO: (2R,5R)-N,N'-dimethyl-1-(10H-phenothiazin-10-yl)propan-2-amine S-oxide.

Promethazine Teoclate



and enantiomer

$C_{17}H_{20}N_2S \cdot C_7H_7ClN_4O_2$ 499.0

17693-51-5

Action and use

Histamine H₁ receptor antagonist; antihistamine.

Preparation

Promethazine Teoclate Tablets

DEFINITION

Promethazine Teoclate is the (R,S)-dimethyl(2-phenothiazin-10-yl)propylamine salt of 8-chlorophenothiazine. It contains not less than 98.0% and not more than 101.0% of $C_{17}H_{20}N_2S \cdot C_7H_7ClN_4O_2$, calculated with reference to the dried substance.

CHARACTERISTICS

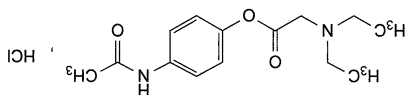
A white or almost white powder.

Very slightly soluble in water; sparingly soluble in ethanol (96%); practically insoluble in ether.



Propacetamol Hydrochloride

(Ph. Eur. monograph 1366)



$C_{14}H_{21}ClN_2O_3$ 300.8 66532-86-3

Action and use

Analgesic; antipyretic.

DEFINITION

4-(Acetylamino)phenyl (diethylamino)acetate hydrochloride.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in anhydrous ethanol, practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of propacetamol hydrochloride.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Prepare the solution immediately before use. Dissolve 1.75 g in water R and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ or BY₆ (2.2.2, Method II).

Absorbance

Maximum 0.05, determined at 390 nm on solution S.

Impurity B

Thin-layer chromatography (2.2.27).

Test solution Suspend 4.00 g of the substance to be examined in 8 mL of acetonitrile R. Shake for 30 min and filter. Dilute to 10 mL with acetonitrile R.

Reference solution (a) Dissolve 25 mg of 4-aminophenol R (impurity B) in acetonitrile R and dilute to 50 mL with the same solvent. Dilute 10 mL of this solution to 50 mL with acetonitrile R.

Reference solution (b) Dilute 5 mL of reference solution (a) to 50 mL with acetonitrile R.

Reference solution (c) Dilute 0.2 mL of reference solution (a) to 5 mL with the test solution.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase anhydrous formic acid R, water R, methanol R, methylene chloride R (3:4:30:64 V/V/V/V).

Application 50 µL of the test solution and of reference solutions (b) and (c).

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm. Spray with a 10 g/L solution of dimethylaminobenzaldehyde R in ethanol (96 per cent) R.

IDENTIFICATION

A. Shake 0.15 g with 2.5 mL of water, add 1 mL of 5M ammonia and extract with 30 mL of ether. Wash the ether extract with 10 mL of water, dry with anhydrous sodium sulfate and evaporate the ether to dryness. Dissolve the residue in 1 mL of chloroform IR. The infrared absorption spectrum of the resulting solution, Appendix II A, is concordant with the reference spectrum of promethazine (RS 297).

B. Dissolve 5 mg in 2 mL of sulfuric acid and allow to stand for 5 minutes. A red colour is produced.

C. Shake 0.4 g with 10 mL of water, add 4 mL of 5M ammonia, shake with two 30-mL quantities of ether and add 4 mL of hydrochloric acid to the aqueous solution. Filter the white precipitate, wash with water and dry at 105°.

Dissolve 10 mg of the residue in 1 mL of hydrochloric acid, add 0.1 g of potassium chlorate and evaporate to dryness.

A reddish residue remains which becomes purple on exposure to the vapour of ammonia.

TESTS

Chloride

Shake 0.3 g with 30 mL of water for 2 minutes and filter.

15 mL of the filtrate complies with the limit test for chlorides, Appendix VII, but using 2 mL of nitric acid in place of the 1 mL of dilute nitric acid (350 ppm).

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using a silica gel F₂₅₄ precoated plate (Merck silica gel 60 F₂₅₄ plates are suitable) and a mixture of 5 volumes of diethylamine, 10 volumes of acetone and 85 volumes of cyclohexane as the mobile phase. Pour the mobile phase into an unlined tank, immediately place the prepared plate in the tank, close the tank and allow the solvent front to ascend 12 cm above the line of application. Apply separately to the plate 10 µL of each of the following solutions in a mixture of 5 volumes of diethylamine and 95 volumes of methanol. Solution (1) contains 2% w/v of the substance being examined. Solution (2) contains 0.02% w/v of isopromethazine hydrochloride BCRS. For solution (3) dilute 1 volume of solution (1) to 200 volumes. For solution (4) dilute 1 volume of solution (1) to 500 volumes. Allow the plate to dry in air and examine under ultraviolet light (254 nm). In the chromatogram obtained with solution (1) any spot corresponding to isopromethazine is not more intense than the spot in the chromatogram obtained with solution (2) (1%), any other secondary spot is not more intense than the spot in the chromatogram obtained with solution (3) (0.5%) and not more than three such spots are more intense than the spot in the chromatogram obtained with solution (4) (0.2%). Disregard any spot remaining on the line of application.

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 1 g in 200 mL of acetone and carry out Method I for non-aqueous titration, Appendix VIII A, using 3 mL of a saturated solution of methyl orange in acetone as indicator.

Each mL of 0.1M perchloric acid VS is equivalent to 49.90 mg of C₁₇H₂₀N₂S, C₁₇H₁₉ClN₂O₂.

STORAGE

Promethazine Teoclate should be protected from light.

Identification of spots Reference solution (c) shows 2 spots, one visible in ultraviolet light due to propacetamol and the other one yellow, visible after spraying due to impurity B. An additional spot may appear in ultraviolet light and corresponds to impurity A.

System suitability: reference solution (c):

- the chromatogram shows 2 clearly separated spots.

Limit:

- **impurity B:** any yellow spot due to impurity B not visible in ultraviolet light is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (25 ppm).

Related substances

Liquid chromatography (2.2.29).

Solution A Dissolve 2.16 g of sodium octanesulfonate R in 900 mL of water R and dilute to 1000 mL with the same solvent. Adjust to pH 3.0 with acetic acid R.

Test solution Suspend 1.00 g of the substance to be examined in 10.0 mL of acetonitrile R. Shake for 10 min. Allow to stand. Take 3.0 mL of the supernatant solution and dilute to 10.0 mL with solution A. Inject immediately.

Reference solution (a) Dissolve 50 mg of paracetamol R (impurity A) in acetonitrile R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with acetonitrile R. Dilute 3.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 10 mg of paracetamol R (impurity A) and 0.100 g of 4-aminophenol R (impurity B) in acetonitrile R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with acetonitrile R. Dilute 3.0 mL of this solution to 10.0 mL with solution A.

Column:

- **stationary phase:** octadecylsilyl silica gel for chromatography R (5 µm).
- **size:** $l = 0.25$ m, $\phi = 4.6$ mm;

Flow rate 1 mL/min.

Detection Spectrophotometer at 246 nm.

Injection 20 µL.

Run time Twice the retention time of paracetamol.

Identification of impurities The chromatogram obtained with reference solution (b) shows a peak due to impurity A (1st peak) and a peak due to impurity B (2nd peak).

Relative retention With reference to impurity A:

- impurity B = about 1.6.

Limits:

- **impurity A:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (200 ppm);
- **unspecified impurities:** for each impurity, not more than 3.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent taking into account the response factor of paracetamol of 1.6);
- **total:** not more than 6.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3 ppm taking into account the relative response factor of paracetamol of 1.6).

— **disregard limit:** 0.01 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent taking into account the relative response factor of paracetamol of 1.6);

— **total:** not more than 6.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3 ppm taking into account the relative response factor of paracetamol of 1.6).

Methanol

Gas chromatography (2.2.28).

Internal standard solution Dilute 2.0 mL of propanol R to 20.0 mL with water R. Dilute 1.0 mL of the solution to 25.0 mL with water R.

Test solution Dissolve 2.00 g of the substance to be examined in water R, add 2.0 mL of the internal standard solution and dilute to 10.0 mL with water R.

Reference solution Dilute 0.8 mL of methanol R to 50.0 mL with water R.

Injection 1.0 mL of the solution to 25.0 mL of the water R. To 2.0 mL of this solution, add 2.0 mL of the internal standard solution and dilute to 10.0 mL with water R.

Column:

— **material:** glass;

— **size:** $l = 2$ m, $\phi = 2$ mm;

— **stationary phase:** carbon molecular sieve impregnated with 0.2 per cent of macrogol 1500.

Carrier gas nitrogen for chromatography R.

Temperature:

Temperature (°C)	Time (min)	Column
60	0 - 1.5	60 → 80
80	1.5 - 5.5	80
170	5.5 - 15.5	170
220		220

Detection Flame ionisation.

Injection 2 µL.

Limit:

— **methanol:** calculate the ratio (R) of the area of the peak due to methanol to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the ratio of the area of any peak due to methanol to the area of the peak due to the internal standard, this ratio is not greater than R (500 ppm).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 25 mL of anhydrous acetic acid R and 25 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

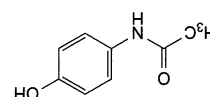
1 mL of 0.1 M perchloric acid is equivalent to 30.08 mg of $C_{14}H_{21}ClN_2O_3$.

STORAGE

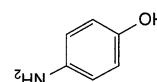
Protected from humidity.

IMPURITIES

Specified impurities A, B



A. N-(4-hydroxyphenyl)acetamide (paracetamol),

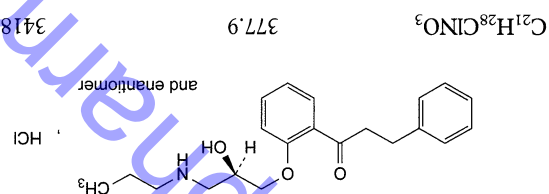


B. 4-aminophenol.

Ph Eur

Propafenone Hydrochloride

(Ph. Eur. monograph 2103)

**Action and use**
Class I antiarrhythmic.

Ph Eur

DEFINITION1-[2-[(2*R,S*)-2-Hydroxy-3-(propylamino)propoxy]phenyl]-3-phenylpropan-1-one hydrochloride.**CHARACTERS****Content**
99.0 per cent to 101.0 per cent (dried substance).**Appearance**

Colourless crystals or white or almost white powder.

Solubility

Slightly soluble in cold water, soluble in methanol and in hot water, practically insoluble in ethanol (96 per cent).

mp

About 173 °C.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison propafenone hydrochloride CRS.

B. To 5.0 mL of solution S (see Tests) add 2 drops of dilute nitric acid R. A precipitate is formed. After 10 min, filter. 2.0 mL of the clear filtrate gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

To 0.500 g in a 100 mL volumetric flask add 50 mL of water R, and heat to boiling for 5 min. Allow to cool to room temperature and dilute to 100.0 mL with carbon dioxide-free water R.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and is colourless (2.2.2, Method II).

Optical rotation (2.2.7)Dissolve 1.00 g in methanol R and dilute to 100.0 mL with the same solvent.
-0.05° to +0.05°.**Related substances**Liquid chromatography (2.2.29).
Solvent mixture Mobile phase B, mobile phase A (35:65 V/V).
Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.
Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.
Reference solution (c) Dissolve 5.0 mg of the substance to be examined and 5.0 mg of propafenone impurity B CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.Column:
mixture.
size: $l = 0.15$ m, $\varnothing = 4.6$ mm,
stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m) with a specific surface area of 320-350 m²/g and a pore size of 12-13 nm,
temperature: 30 °C.
Mobile phase:
— mobile phase A: 3.42 g/L solution of dipotassium hydrogen phosphate trihydrate R adjusted to pH 2.5 with phosphoric acid R,
mobile phase B: acetonitrile for chromatography R,

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	65	35
8 - 20	65 \rightarrow 30	35 \rightarrow 70
20 - 30	30	70

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Equilibration 60 min with the mobile phase at the initial composition, before each series of injections.

Injection 20 μ L of the test solution, reference solutions (a), (b) and (c) and of the solvent mixture as a blank.

Relative retention With reference to propafenone (retention time = about 5 min): impurity B = about 0.8;

impurity D = about 2.3; impurity G = about 3.6;

impurity C = about 4.1; impurity F = about 5.3.

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to impurity B and propafenone.

Limits:— impurities B, C, D, F, G: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent).

— disregard limit: 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in a mixture of 0.4 mL of acetic acid R and 15.0 mL of water R, heating on a water-bath. To the warm solution add 3 mL of buffer solution pH 3.5 R. After cooling to room temperature, filter through a sintered-glass filter (40) (2.1.2) and rinse with water R until 20.0 mL of filtrate is obtained. 12.0 mL of the filtrate complies with limit test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating during the titration, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.300 g in 2 mL of anhydrous formic acid R.

Add 50 mL of acetic anhydride R. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

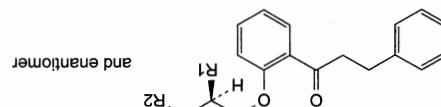
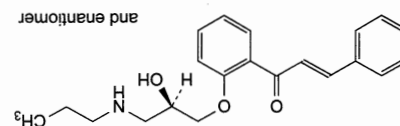
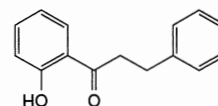
1 mL of 0.1 M perchloric acid is equivalent to 37.79 mg

of $C_{21}H_{28}ClNO_3$.

IMPURITIES

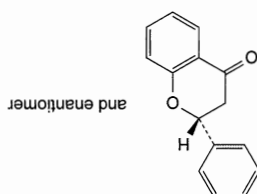
Specified impurities B, C, D, F, G.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, E, H.

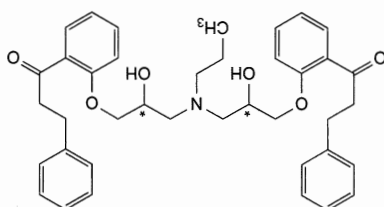


D, R1 = R2 = OH: 1-[2-[(2R)-2,3-dihydroxypropoxy]phenyl]-3-phenylpropan-1-one, E, R1 = OH, R2 = Cl: 1-[2-[(2R)-3-chloro-2-hydroxypropoxy]phenyl]-3-phenylpropan-1-one,

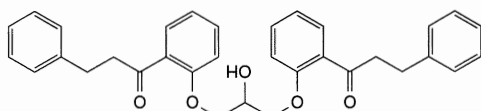
H. (2R)-2-phenyl-2,3-dihydro-4H-1-benzopyran-4-one.



G. 1,1'-[propylidenebis[(2-hydroxypropyl)oxy-2,1-phenylene]]bis(3-phenylpropan-1-one),



F. 1,1'-[2-hydroxypropyl-1,3-diyloxy-2,1-phenylene]]bis(3-phenylpropan-1-one),



DEFINITION

Propan-1-ol.

CHARACTERS

Appearance

Clear, colourless liquid.

Solubility

Miscible with water and with ethanol.

IDENTIFICATION

First identification C, B

Second identification A, B, D

A. Refractive index (2.2.6): 1.384 to 1.387.

B. Boiling point (2.2.12): 96 °C to 98 °C.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of propanol.

D. To 1.0 mL add 0.10 g of dinitrobenzoyl chloride R and

0.05 mL of sulfuric acid R. Boil under reflux for 30 min.

Evaporate until the excess of propanol is removed, add 5 mL

of heptane R to the residue and heat to boiling. Filter the hot

solution. Wash the crystals formed on cooling with heptane R

and dry in vacuum (2 kPa, at room temperature for 24 h).

The small, colourless, shiny plates melt (2.2.14) between

71 °C and 74 °C.

TESTS**Appearance**

The substance to be examined is clear (2.2.1) and colourless (2.2.2, Method II). Dilute 2 mL to 10 mL with water R. After 5 min, the solution is clear (2.2.1).

Acidity or alkalinity

To 10.0 mL of carbon dioxide-free water R add 0.1 mL of

phenolphthalein solution R and 0.01 M sodium hydroxide until

the solution becomes pale pink. After addition of 5.0 mL of

the substance to be examined the colour of the solution does

not become more intense. If the colour fades, add 0.2 mL of

0.01 M sodium hydroxide. The solution is pink.

Absorbance (2.2.25)

Measure the absorbance between 230 nm and 310 nm using water R as the compensation liquid. The absorbance *A* is not

greater than the following values.

Wavelength (nm)	Absorbance <i>A</i>
230	0.300
250	0.100
270	0.030
290	0.020
310	0.010

The absorption curve does not show any peaks.

Reducing substances

Place 10.0 mL in a test tube of about 20 mm in diameter in a water bath at 20 °C. Keep protected from actinic light and

add 1.0 mL of a freshly prepared 0.16 g/L solution of

potassium permanganate R. The mixture, maintained at 20 °C,

the test solution is not less intensely coloured (2.2.2,

Method II) than 10.0 mL of a reference solution prepared as

follows: to 5.5 mL of primary solution yellow, add 13.0 mL

of primary solution red and dilute to 100.0 mL with water R.

Related substances

Gas chromatography (2.2.28).

The substance to be examined.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with heptane R. Dilute 1.0 mL of this solution to

10.0 mL with heptane R.

Reference solution (b) Mix 0.1 mL of acetone R and 0.1 mL of

2-propanol R and dilute to 100 mL with the test solution.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.25$ mm;

— stationary phase:

poly[(cyanopropyl)(phenyl)]dimethylsiloxane R (film

thickness 1.4 μ m).

Carrier gas helium for chromatography R.

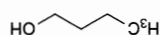
Linear velocity 25 cm/s.

Split ratio 1:200.

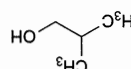
Temperature:

Time (min)	Temperature (°C)
0 - 12	40
12 - 28	40 \rightarrow 200
28 - 38	200
Injection port	240
Detector	240

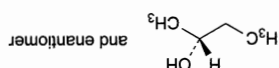
H. butan-1-ol (*n*-butanol),



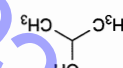
G. 2-methylpropan-1-ol (isobutanol),



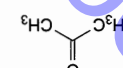
F. butan-2-ol (*sec*-butanol),



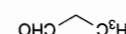
E. isopropyl alcohol (2-propanol),



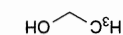
D. propanone (acetone),



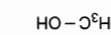
C. propanal,



B. ethanol,



A. methanol,

**IMPURITIES**

Protected from light.

STORAGE

Maximum 0.2 per cent, determined on 10 g.

Water (2.5.12)

Evaporate 50 mL of the substance to be examined to dryness at 100 °C and dry the residue in an oven at 100-105 °C to constant mass. The residue weighs a maximum of 2 mg.

Maximum 0.004 per cent.

Non-volatile matter

— any impurity: not more than the area of the peak due to propanol in the chromatogram obtained with reference solution (a) (0.1 per cent);
— total: not more than 3 times the area of the peak due to propanol in the chromatogram obtained with reference solution (a) (0.3 per cent);
— disregard limit: 0.1 times the area of the peak due to propanol in the chromatogram obtained with reference solution (a) (0.01 per cent).

Limits:

impurities D and E.

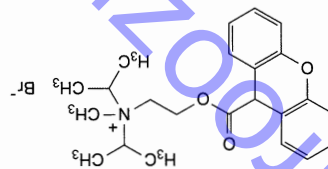
— resolution: minimum 2.0 between the peaks due to System suitability: reference solution (b):

Injection 1 μ L.

Detection Flame ionisation.

Propantheline Bromide

(Ph. Eur. monograph 0857)


 $C_{23}H_{30}BrNO_3$ 448.4 50-34-0

Action and use

Anticholinergic.

Preparation

Propantheline Tablets

Ph Eur

DEFINITION

N-Methyl-N,N-bis(1-methylethyl)-2-[(9H-xanthen-9-ylcarbonyl)oxy]ethanaminium bromide.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, slightly hygroscopic powder.

Solubility

Very soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 60 mg in methanol R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with methanol R.

Spectral range 230-350 nm.

Absorption maxima At 246 nm and 282 nm.

Specific absorbance at the absorption maxima:

— at 246 nm: 115 to 125;

— at 282 nm: 57 to 63.

B. Dissolve 0.2 g in 15 mL of water R and add 1 mL of strong sodium hydroxide solution R. Boil for 2 min and cool slightly. Add 7.5 mL of dilute hydrochloric acid R and filter.

Wash the residue with water R and recrystallise from ethanol (50 per cent V/V) R. Dry at 100-105 °C for 1 h. Dissolve about 10 mg of the residue in 5 mL of sulfuric acid R.

The solution has an intense yellow colour and shows an intense yellowish-green fluorescence when examined in ultraviolet light at 365 nm.

C. Dissolve 50 mg in 0.1 mL of water R in a 25 mL flask and add 1 mL of a saturated solution of potassium

permanganate R. Attach a fractionating column and a condenser, with the end of the delivery tube immersed in 1 mL of water R in a test-tube placed in a bath of iced water.

Distill fairly vigorously and continue heating for 1 min after a dry residue has been obtained in the flask. Prepare a blank

by introducing into an identical test-tube a volume of water R equal to that of the distillate. Place the tubes in a bath of iced water. To each tube, add 0.5 mL of a 20 per cent V/V solution of morpholine R and 0.5 mL of a freshly prepared

50 g/L solution of sodium nitroprusside R. Mix and allow to stand at 0 °C for 5 min, and then at room temperature for 3 min. No blue colour develops in either tube. Add 1 g of ammonium sulfate R, mix and allow to stand for 15 min.

A stable, intense pink colour develops in the test solution.

D. It gives reaction (a) of bromides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1).

Dissolve 0.6 g in water R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, water R (40:60 V/V).

Test solution (a) Dissolve 6 mg of the substance to be examined in the solvent mixture and dilute to 50 mL with the solvent mixture.

Test solution (b) Dissolve 6 mg of the substance to be examined in 30 mL of the solvent mixture. Add 5 mL of reference solution (b) and dilute to 50 mL with the solvent mixture.

Test solution (c) Dissolve 6 mg of xanthinol R1 and 6 mg of the substance to be examined in the solvent mixture, then dilute to 50 mL with the solvent mixture.

Reference solution (a) Dissolve 6 mg of xanthinol R1 in the solvent mixture and dilute to 50 mL with the solvent mixture.

Reference solution (b) Dilute 5 mL of reference solution (a) to 50 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mixture of equal volumes of acetonitrile R and of a solution containing 28 g/L of sodium perchlorate R and 11 g/L of phosphoric acid R, adjusted to pH 3.8 with strong sodium hydroxide solution R and then with 0.1 M sodium hydroxide.

Flow rate 1 mL/min.

Detection Spectrophotometer at 206 nm.

Injection 20 μ L of test solutions (a), (b), (c) and reference solution (a).

Run time Twice the retention time of propantheline.

System suitability: test solution (c):

— is no peak corresponding to the principal peak in the chromatogram obtained with reference solution (a);

— resolution: minimum 8.0 between the peaks due to propantheline and xanthinol.

Limits: test solution (b):

— any impurity: for each impurity, not more than the area of the peak due to xanthinol (1.0 per cent), and not more than one such peak has an area greater than or equal to

0.5 times the area of the peak due to xanthidrol
 — *disregard limit*: disregard any peak with a retention time relative to propantheline of less than 0.2 (bromide); disregard the peak due to xanthidrol.
Loss on drying (2.2.32)
 Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.
ASSAY
 Dissolve 0.400 g in 50 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
 1 mL of 0.1 M perchloric acid corresponds to 44.84 mg of $C_{23}H_{30}BrNO_3$.
STORAGE
 In an airtight container.
 Ph Eur

Chemical structure of Propofol: CC(C)C1=CC=C(C=C1)C(C)C
 178.3
 2078-54-8

Run time 7 times the retention time of propofol.
 Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities G and E.
 Relative retention With reference to propofol (retention time = about 3 min): impurity G = about 0.5; impurity I = about 0.6; impurity B = about 0.7; impurity N = about 2.3; impurity D = about 2.5; impurity P = about 2.9; impurity A = about 3.0; impurity C = about 3.4; impurity E = about 4.0; impurity F = about 5.8; impurity H = about 6.4.
 System suitability: reference solution (a):
 — *resolution*: minimum 4.0 between the peaks due to impurity J and propofol.
 Limits:
 — *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity E = 0.25; impurity G = 5.0;
 — *impurity G*: not more than twice the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.2 per cent);
 — *impurity E*: not more than 0.1 times the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.01 per cent);
 — *unspecified impurities*: for each impurity, not more than 0.5 times the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.3 per cent);
 — *disregard limit*: 0.3 times the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.03 per cent), except for impurity E.
Impurities J, K, L and O
 Gas chromatography (2.2.28).
 Test solution Dissolve 40.0 mg of the substance to be examined in methylene chloride R and dilute to 10.0 mL with the same solvent.

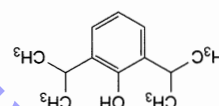
0.5 times the area of the peak due to xanthidrol
 — *disregard limit*: disregard any peak with a retention time relative to propantheline of less than 0.2 (bromide); disregard the peak due to xanthidrol.
Loss on drying (2.2.32)
 Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.
ASSAY
 Dissolve 0.400 g in 50 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
 1 mL of 0.1 M perchloric acid corresponds to 44.84 mg of $C_{23}H_{30}BrNO_3$.
STORAGE
 In an airtight container.
 Ph Eur

Chemical structure of Propofol: CC(C)C1=CC=C(C=C1)C(C)C
 178.3
 2078-54-8

Run time 7 times the retention time of propofol.
 Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities G and E.
 Relative retention With reference to propofol (retention time = about 3 min): impurity G = about 0.5; impurity I = about 0.6; impurity B = about 0.7; impurity N = about 2.3; impurity D = about 2.5; impurity P = about 2.9; impurity A = about 3.0; impurity C = about 3.4; impurity E = about 4.0; impurity F = about 5.8; impurity H = about 6.4.
 System suitability: reference solution (a):
 — *resolution*: minimum 4.0 between the peaks due to impurity J and propofol.
 Limits:
 — *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity E = 0.25; impurity G = 5.0;
 — *impurity G*: not more than twice the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.2 per cent);
 — *impurity E*: not more than 0.1 times the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.01 per cent);
 — *unspecified impurities*: for each impurity, not more than 0.5 times the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.3 per cent);
 — *disregard limit*: 0.3 times the area of the peak due to propofol in the chromatogram obtained with reference solution (c) (0.03 per cent), except for impurity E.
Impurities J, K, L and O
 Gas chromatography (2.2.28).
 Test solution Dissolve 40.0 mg of the substance to be examined in methylene chloride R and dilute to 10.0 mL with the same solvent.

Propofol

(Ph. Eur. monograph 1558)



$C_{12}H_{18}O$

Action and use
 Intravenous general anaesthetic.

Preparation
 Propofol Injection

Ph Eur

DEFINITION

2,6-Bis(1-methylethyl)phenol.

Content

98.0 per cent to 102.0 per cent.

This monograph applies to propofol prepared using distillation for purification.

CHARACTERS

Appearance

Colourless or very light yellow, clear liquid.

Solubility

Very slightly soluble in water, miscible with hexane and with methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison propofol CRS.

TESTS

Refractive index (2.2.6)

1.5125 to 1.5145.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 1.00 g of the substance to be examined in hexane R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methylene chloride R. Dilute 1.0 mL of this solution to 10.0 mL with methylene chloride R.

Reference solution (b) Dissolve 5 µL of propofol impurity J CRS (corresponding to 5 mg) in methylene chloride R and dilute to 100 mL with the same solvent. Dilute 1.0 mL of this solution to 25 mL with methylene chloride R.

Reference solution (c) Dissolve 4 mg of propofol CRS in reference solution (b) and dilute to 1 mL with the same solution.

Column:

material: fused silica;

size: $l = 30$ m, $\phi = 0.32$ mm;

stationary phase: polymethylphenylsiloxane R (film thickness 0.5 µm).

Carrier gas helium for chromatography R.

Flow rate 1.7 mL/min.

Split ratio 1:5.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 3	80			270
3 - 25	80 → 210			100
25 - 40	210			

Detection Flame ionisation.

Injection 1 µL of the test solution and reference solutions (a) and (c).

Relative retention With reference to propofol (retention

time = about 17 min): impurity K = about 0.76;

impurity L = about 0.81; impurity J = about 1.01;

impurity O = about 1.03.

System suitability: reference solution (c):

above the baseline of the peak due to impurity J, and

H_p = height above the baseline of the lowest point of the

curve separating this peak from the peak due to propofol.

Limits:

— impurities J, K, L, O: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.05 per cent).

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

Calculate the percentage content of $C_{12}H_{18}O$ using the

declared content of propofol CRS.

STORAGE

Protected from light under an inert gas.

IMPURITIES

Specified impurities E, G, J, K, L, O

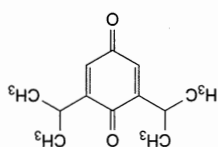
Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by the general

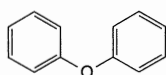
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

J. 2,6-bis(1-methylethyl)benzene-1,4-dione,



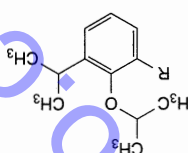
I. oxydibenzene,



K. R = H: 1-(1-methylethoxy)-2-(1-methylethyl)benzene,

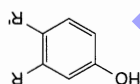
bis(1-methylethyl)benzene,

G. R = CH(CH₃)₂: 2-(1-methylethoxy)-1,3-

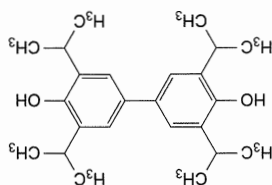


H. R = H, R' = CH(CH₃)₂: 4-(1-methylethyl)phenol,

F. R = CH(CH₃)₂, R' = H: 3-(1-methylethyl)phenol,



B. 3,3',5,5'-tetraakis(1-methylethyl)biphenyl-4,4'-diol,



I-methylethyl 4-hydroxy-3,5-bis(1-methylethyl)benzoate,

P. R1 = CO-O-CH(CH₃)₂, R2 = H, R3 = CH(CH₃)₂:

6-propylphenol,

O. R1 = R2 = H, R3 = CH₂-CH₂-CH₃: 2-(1-methylethyl)-

bis(1-methylethyl)benzoic acid,

N. R1 = CO₂H, R2 = H, R3 = CH(CH₃)₂: 4-hydroxy-3,5-

2,5-bis(1-methylethyl)phenol,

D. R1 = R3 = H, R2 = CH(CH₃)₂:

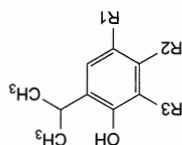
C. R1 = R2 = R3 = H: 2-(1-methylethyl)phenol,

6-(1-methylethyl)phenol,

B. R1 = R2 = H, R3 = C(CH₃)=CH₂: 2-(1-methylethyl)-

2,4-bis(1-methylethyl)phenol,

A. R1 = CH(CH₃)₂, R2 = R3 = H:

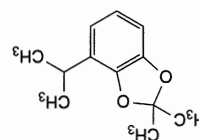


C, D, F, H, I, N, P.

Control of impurities in substances for pharmaceutical use: A, B,

impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these



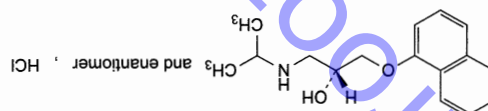
I. 2,2-dimethyl-4-(1-methylethyl)-1,3-benzodioxole.

Ph Eur



Propranolol Hydrochloride

(Ph. Eur. monograph 0568)



$C_{16}H_{22}ClNO_2$ 295.8 318-98-9

Action and use

Beta-adrenoceptor antagonist.

Preparations

Prolonged-release Propranolol Capsules

Propranolol Injection

Propranolol Tablets

Ph Eur

DEFINITION

(2RS)-1-[(1-Methylethyl)amino]-3-(naphthalen-1-yloxy)propan-2-ol hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Melting point (2.2.14): 163 °C to 166 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison propranolol hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in 1 mL of methanol R.

Reference solution Dissolve 10 mg of propranolol hydrochloride CRS in 1 mL of methanol R.

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R1, methanol R (1:99 V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying At 100-105 °C.

Detection Spray with anisaldehyde solution R and heat at 100-105 °C until the colour of the spots reaches maximum intensity (10-15 min).

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Dissolve 2.0 g in methanol R and dilute to 20 mL with the same solvent.

Acidity or alkalinity

Dissolve 0.20 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent. Add 0.2 mL of 0.01 M hydrochloric acid; the solution is red. Add 0.4 mL of 0.01 M sodium hydroxide; the solution is yellow.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of propranolol hydrochloride for performance test CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m; $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 1.6 g of sodium laurylsulfate R and 0.31 g of tetraethylammonium dihydrogen phosphate R in a mixture of 1 mL of sulfuric acid R, 450 mL of water R and 550 mL of acetonitrile R; adjust to pH 3.3 using dilute sodium hydroxide solution R.

Flow rate 1.8 mL/min.

Detection Spectrophotometer at 292 nm.

Equilibration For at least 30 min.

Injection 20 µL.

Run time 7 times the retention time of propranolol.

Identification of impurities Use the chromatogram supplied with propranolol hydrochloride for performance test CRS to identify the peak due to impurity A.

System suitability: reference solution (a):

— baseline separation is obtained between the peaks due to impurity A and propranolol.

Limits:

— any impurity: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.1 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in a mixture of 15 volumes of water R and 85 volumes of methanol R and dilute to 20 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (1 ppm Pb) prepared by diluting lead standard

solution (100 ppm Pb) R with a mixture of 15 volumes of

water R and 85 volumes of methanol R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

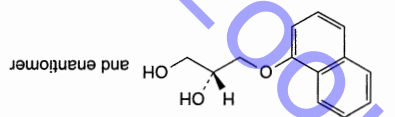
ASSAY

Dissolve 0.250 g in 25 mL of ethanol (96 per cent) R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

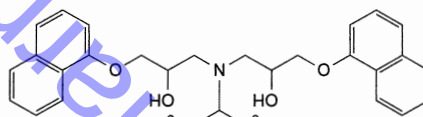
1 mL of 0.1 M sodium hydroxide is equivalent to 29.58 mg

of $C_{10}H_{12}O_5$.

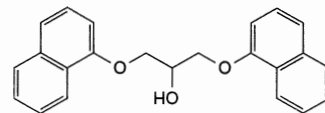
IMPURITIES



A. (2R,3R)-3-(naphthalen-1-yloxy)propane-1,2-diol (diol derivative),



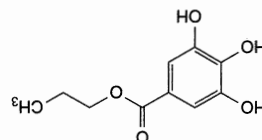
B. 1,1'-bis-[(1-methylethyl)imino]bis[3-(naphthalen-1-yloxy)propan-2-ol] (tertiary amine derivative),



C. 1,3-bis(naphthalen-1-yloxy)propan-2-ol (bis-ether derivative).

Propyl Gallate

(Ph. Eur. monograph 1039)



$C_{10}H_{12}O_5$

212.2

121-79-9

Action and use

Antioxidant.

DEFINITION

Propyl gallate contains not less than 97.0 per cent and not more than the equivalent of 103.0 per cent of propyl 3,4,5-trihydroxybenzoate, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, very slightly soluble in water, freely soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification B
Second identification A, C, D

A. Melting point (2.2.14): 148 °C to 151 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with propyl gallate CRS.

C. Examine the chromatograms obtained in the test for gallic acid. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 10 mg in 10 mL of water R by heating to about 70 °C. Cool and add 1 mL of bismuth subnitrate solution R. A bright yellow precipitate is formed.

TESTS

Appearance of solution
Dissolve 1.0 g in ethanol (96 per cent) R and dilute to 20 mL with the same solvent. The solution is clear (2.2.1 and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Gallic acid

Examine by thin-layer chromatography (2.2.27), using silica gel G R as the coating substance.

Test solution (a) Dissolve 0.20 g of the substance to be examined in acetone R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 20 mL with acetone R.

Reference solution (a) Dissolve 10 mg of propyl gallate CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 20 mg of gallic acid R in acetone R and dilute to 20 mL with the same solvent. Dilute 1 mL of the solution to 10 mL with acetone R.

Reference solution (c) Dilute 0.5 mL of test solution (b) to 5 mL with reference solution (b).

Apply separately to the plate 5 µL of each solution. Develop over a path of 8 cm using a mixture of 10 volumes of anhydrous formic acid R, 40 volumes of ethyl formate R and 50 volumes of toluene R. Allow the plate to dry in air for 10 min and spray with a mixture of 1 volume of ferric chloride solution R1 and 9 volumes of ethanol (96 per cent) R. Any spot due to gallic acid in the chromatogram obtained with test solution (a) is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows 2 clearly separated principal spots.

Total chlorine

Mix 0.5 g with 2 g of calcium carbonate R1. Dry and ignite at 700 ± 50 °C. Take up the residue with 20 mL of dilute nitric acid R and dilute to 30 mL with water R. 15 mL of the solution, without further addition of dilute nitric acid R, complies with the limit test for chlorides (2.4.4) (200 ppm).

Chlorides (2.4.4)

To 1.65 g add 50 mL of water R. Shake for 5 min. Filter. 15 mL of the filtrate complies with the limit test for chlorides (100 ppm).

Zinc

Not more than 25 ppm of Zn, determined by atomic absorption spectrometry (2.2.23, Method II).

Test solution To 2.5 mL of the solution obtained in the test for heavy metals, add 2.5 mL of water R.

Reference solutions Prepare the reference solutions using zinc standard solution (10 ppm Zn) R, diluted as necessary with water R.

Measure the absorbance at 213.9 nm using a zinc hollow-cathode lamp as the source of radiation and an air-acetylene flame.

Heavy metals (2.4.8)

2.0 g complies with limit test C for heavy metals (10 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in methanol R and dilute to 250.0 mL with the same solvent. Dilute 5.0 mL of the solution to 200.0 mL with methanol R. Measure the absorbance (2.2.25) at the absorption maximum at 275 nm.

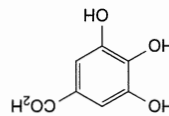
Calculate the content of C₁₀H₁₂O₃ taking the specific absorbance to be 503.

STORAGE

Protected from light.

IMPURITIES

Specified impurities: A.

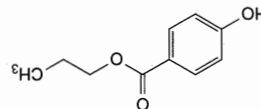


A. 3,4,5-trihydroxybenzoic acid (galllic acid).

Propyl Hydroxybenzoate

Propylparaben

(Propyl Parahydroxybenzoate, Ph Eur monograph 0431)



C₁₀H₁₂O₃

180.2

94-13-3

Action and use

Antimicrobial preservative.

DEFINITION

Propyl 4-hydroxybenzoate.

Content

98.0 per cent to 102.0 per cent.

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

First identification A, B

Second identification A, C

A. Melting point (2.2.14): 96 °C to 99 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g of the substance to be examined in acetone R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with acetone R.

Reference solution (a) Dissolve 10 mg of propyl parahydroxybenzoate CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of ethyl parahydroxybenzoate CRS in 1 mL of test solution (a) and dilute to 10 mL with acetone R.

Mobile phase glacial acetic acid R, water R, methanol R (1:30:70 V/V/V).

Application 2 µL of test solution (b) and reference solutions (a) and (b).

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b): — the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS**Solution S**

Dissolve 1.0 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Acidity

To 2 mL of solution S add 3 mL of ethanol (96 per cent) R₃, 5 mL of carbon dioxide-free water R and 0.1 mL of bromocresol green solution R. Not more than 0.1 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to blue.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of 4-hydroxybenzoic acid R (impurity A), 5 mg of ethyl parahydroxybenzoate R

(impurity C) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the

mobile phase.

Reference solution (b) Dissolve 50.0 mg of propyl

parahydroxybenzoate CRS in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of this

solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to

20.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 μ m).

Mobile phase 6.8 g/L solution of potassium dihydrogen phosphate R, methanol R (35:65 V/V).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 10 μ L of the test solution and reference solutions (a)

and (c).

Run time 2.5 times the retention time of propyl

parahydroxybenzoate.

Relative retention With reference to propyl

parahydroxybenzoate (retention time = about 4.5 min):

impurity A = about 0.3; impurity C = about 0.7.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to

impurity C and propyl parahydroxybenzoate.

Limits:

— correction factor: for the calculation of contents, multiply the

peak area of impurity A by 1.4;

— impurity A: not more than the area of the principal peak

in the chromatogram obtained with reference solution (c)

(0.5 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (c) (0.5 per cent);

— total: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (c)

(0.1 per cent).

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{10}H_{12}O_3$ from the

declared content of propyl parahydroxybenzoate CRS.

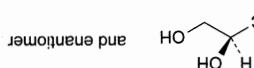
IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.

Propylene Glycol

(Ph. Eur. monograph 0430)



76.1

57-55-6

Action and use

Excipient.

Action and use

Propylene Glycol Solution

DEFINITION

Propylene glycol is (RS)-propane-1,2-diol.

CHARACTERS

A viscous, clear, colourless, hygroscopic liquid, miscible with

water and with ethanol (96 per cent).

IDENTIFICATION

A. Relative density (see Tests).

B. Refractive index (see Tests).

C. Boiling point (2.2.12): 184 °C to 189 °C.

D. To 0.5 mL add 5 mL of pyridine R and 2 g of finely

ground nitrobenzoyl chloride R. Boil for 1 min and pour into

15 mL of cold water R with shaking. Filter, wash the

precipitate with 20 mL of a saturated solution of sodium

hydrogen carbonate R and then with water R and dry. Dissolve

in boiling ethanol (80 per cent V/V) R and filter the hot

solution. On cooling, crystals are formed which, after drying

IDENTIFICATION

- A. Refractive index (2.2.6): 1.439 to 1.442.
B. Relative density (2.2.5): 0.910 to 0.930.
C. Viscosity (2.2.9): 9 mPa·s to 12 mPa·s.
D. Composition of fatty acids (see Tests).

TESTS

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Acid value (2.5.1)

Maximum 0.2.

Hydroxyl value (2.5.3, Method A)

Maximum 10.

Iodine value (2.5.4)

Maximum 1.0.

Peroxide value (2.5.5, Method A)

Maximum 1.0.

Saponification value (2.5.6)

320 to 340.

Unsaponifiable matter (2.5.7)

Maximum 0.3 per cent, determined on 5.0 g.

Alkaline impurities

Dissolve 2.00 g of the substance to be examined in a mixture of 1.5 mL of ethanol (96 per cent) R and 3.0 mL of ether R. Add 0.05 mL of bromophenol blue solution R. Not more than 0.15 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to yellow.

Composition of fatty acids

Gas chromatography (2.4.22, Method C). Prepare reference solution (a) as indicated in Table 2.4.22-2.

Column:

- material: fused silica,
- size: $l = 30$ m, $\varnothing = 0.32$ mm,
- stationary phase: macrogol 20 000 R (film thickness 0.5 μ m),

Carrier gas helium for chromatography R.

Flow rate 1.3 mL/min.

Split ratio 1:100.

Temperature:

Time (min)	Temperature (°C)
0 - 1	70
1 - 35	70 → 240
35 - 50	240
Injection port	250
Detector	250

Detection Flame ionisation.

Composition of the fatty acid fraction of the substance to be examined:

- caproic acid: maximum 2.0 per cent,
- caprylic acid: 50.0 per cent to 80.0 per cent,
- capric acid: maximum 3.0 per cent,
- lauric acid: maximum 1.0 per cent,
- myristic acid: maximum 1.0 per cent.

Water (2.5.12)

Maximum 0.1 per cent, determined on 5.00 g.

TESTS

Appearance

It is clear (2.2.1) and colourless (2.2.2, Method II).

Relative density (2.2.5)

1.035 to 1.040.

Refractive index (2.2.6)

1.431 to 1.433.

Acidity

To 10 mL add 40 mL of water R and 0.1 mL of bromothymol blue solution R. The solution is greenish-yellow. Not more than 0.05 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to blue.

Oxidising substances

To 10 mL add 5 mL of water R, 2 mL of potassium iodide solution R and 2 mL of dilute sulfuric acid R and allow to stand in a ground-glass-stoppered flask protected from light for 15 min. Titrate with 0.05 M sodium thiosulfate, using 1 mL of starch solution R as indicator. Not more than 0.2 mL of 0.05 M sodium thiosulfate is required.

Reducing substances

To 1 mL add 1 mL of dilute ammonia R1 and heat in a water-bath at 60 °C for 5 min. The solution is not yellow. Immediately add 0.15 mL of 0.1 M silver nitrate and allow to stand for 5 min. The solution does not change its appearance.

Heavy metals (2.4.8)

Mix 4 mL with 16 mL of water R. 12 mL of the solution complies with test A for heavy metals (5 ppm m/L). Prepare the reference solution using lead standard solution

(1 ppm Pb) R.

Water (2.5.12)

Not more than 0.2 per cent, determined on 5.00 g by the semi-micro determination of water.

Sulfated ash (2.4.14)

Heat 50 g until it burns and ignite. Allow to cool. Moistened the residue with sulfuric acid R and ignite; repeat the operations. The residue weighs not more than 5 mg (0.01 per cent).

STORAGE

Store in an airtight container.

Propylene Glycol Dicaprylocaprate

(Ph. Eur. monograph 2122)

Action and use

Excipient.

DEFINITION

Propylene glycol diesters of saturated fatty acids, mainly caprylic (octanoic) acid and capric (decanoic) acid, of vegetable origin.

CHARACTERS

Appearance

Almost colourless to light yellow, oily liquid.

Solubility

Practically insoluble in water, soluble in fatty oils and in light petroleum, slightly soluble in anhydrous ethanol.

Total ash (2.4.16)
Maximum 0.1 per cent, determined on 2.0 g.

STORAGE
Protected from light.

Ph Eur



Propylene Glycol Dilaurate

(Ph. Eur. monograph 2087)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of propylene glycol mono- and diesters of lauric (dodecanoic) acid.

Content

Minimum 70.0 per cent of diesters and maximum 30.0 per cent of monoesters.

CHARACTERS

Appearance

Clear, oily liquid at 20 °C, colourless or slightly yellow.

Solubility

Practically insoluble in water, very soluble in alcohol, in methanol and in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.1 g of the substance to be examined in methylene chloride R and dilute to 2 mL with the same solvent.

Reference solution Dissolve 0.1 g of propylene glycol dilaurate CRS in methylene chloride R and dilute to 2 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase hexane R, ether R (30:70 V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of rhodamine 6 G R in alcohol R. Examine in ultraviolet light at 365 nm.

Results The spots in the chromatogram obtained with the test solution are similar in position to those in the chromatogram obtained with the reference solution.

B. Composition of fatty acids (see Tests).

C. It complies with the assay (content of diesters).

TESTS

Acid value (2.5.1)

Maximum 4.0, determined on 5.00 g.

Iodine value (2.5.4, Method A)

Maximum 1.0.

Saponification value (2.5.6)

230 to 250.

Composition of fatty acids

Gas chromatography (2.4.22, Method C). Use the mixture of calibrating substances in Table 2.4.22.-2.

Composition of the fatty acid fraction of the substance:

— caprylic acid: maximum 0.5 per cent,
— capric acid: maximum 2.0 per cent,

— lauric acid: minimum 95.0 per cent,

— myristic acid: maximum 3.0 per cent,

— palmitic acid: maximum 1.0 per cent.

Free propylene glycol

Maximum 2.0 per cent, determined as prescribed under Assay.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.00 g.

Total ash (2.4.16)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Size-exclusion chromatography (2.2.30).

Stock solution Introduce 0.100 g of propylene glycol R into a flask and dilute to 25.0 mL with tetrahydrofuran R.

Test solution In a 15 mL flask, weigh 0.200 g (m).

Add 5.0 mL of tetrahydrofuran R and shake to dissolve.

Reweight the flask and calculate the total mass of solvent and substance (M).

Reference solutions Into four 15 mL flasks, introduce respectively 0.25 mL, 0.5 mL, 1.0 mL and 2.5 mL of stock solution and add 5.0 mL of tetrahydrofuran R. Weigh each flask and calculate the concentration of propylene glycol in milligrams per gram for each reference solution.

Column:

— size: l = 0.6 m, Ø = 7 mm,

— stationary phase: styrene-divinylbenzene copolymer R (5 µm) with a pore size of 10 nm.

Mobile phase tetrahydrofuran R.

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 µL.

Relative retention With reference to propylene glycol:

diesters = about 0.85; monoesters = about 0.90.

Calculations:

— free propylene glycol: from the calibration curve obtained with the reference solutions, determine the

concentration (C) in milligrams per gram in the test solution and calculate the percentage content in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— monoesters: calculate the percentage content of monoesters using the following expression:

$$\frac{A}{A+B} \times (100 - D)$$

A = area of the peak due to the monoesters,
B = area of the peak due to the diesters,

D = percentage content of free propylene glycol + percentage content of free fatty acids.

Calculate the percentage content of free fatty acids using the expression:

$$\frac{I_A \times 200}{561.1}$$

I_A = acid value.

— diesters: calculate the percentage content of diesters using the following expression:

$$\frac{A+B}{B} \times (100 - D)$$

Saponification value (2.5.6)
210 to 245 for propylene glycol monolaurate (type I) and 200 to 230 for propylene glycol monolaurate (type II).

Composition of fatty acids

Gas chromatography (2.4.22, Method C). Use the mixture of calibrating substances in Table 2.4.22.2.

Composition of the fatty acid fraction of the substance:

- caprylic acid: maximum 0.5 per cent,
- capric acid: maximum 2.0 per cent,
- lauric acid: minimum 95.0 per cent,
- myristic acid: maximum 3.0 per cent,
- palmitic acid: maximum 1.0 per cent.

Free propylene glycol

Maximum 5.0 per cent for propylene glycol monolaurate (type I) and maximum 1.0 per cent for propylene glycol monolaurate (type II), determined as prescribed under Assay.

Water

Maximum 1.0 per cent, determined on 1.00 g.

Total ash

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Size-exclusion chromatography (2.2.30).

Stock solution Introduce 0.100 g of propylene glycol R into a vial and dilute to 25.0 mL with tetrahydrofuran R.

Test solution In a 15 mL flask, weigh 0.200 g (m).

Add 5.0 mL of tetrahydrofuran R and shake to dissolve.

Reweight the flask and calculate the total mass of solvent and substance (M).

Reference solutions Into four 15 mL flasks, introduce respectively 0.25 mL, 0.5 mL, 1.0 mL and 2.5 mL of stock solution and add 5.0 mL of tetrahydrofuran R. In a fifth

15 mL flask, introduce 5.0 mL of stock solution. Weigh each flask and calculate the concentration of propylene glycol in milligrams per gram for each reference solution.

Column:

— size: $l = 0.6$ m, $\varnothing = 7$ mm,

— stationary phase: styrene-divinylbenzene copolymer R (5 μ m) with a pore size of 10 nm.

Mobile phase tetrahydrofuran R.

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 μ L.

Relative retention With reference to propylene glycol: diesters = about 0.85; monoesters = about 0.90.

Calculations:

— free propylene glycol: from the calibration curve obtained with the reference solutions, determine the concentration (C) in milligrams per gram in the test solution and calculate the percentage content in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— monoesters: calculate the percentage content of monoesters using the following expression:

$$\frac{A}{A+B} \times (100 - D)$$

A = area of the peak due to the monoesters,
B = area of the peak due to the diesters,

Propylene Glycol Monolaurate

(Ph. Eur. monograph 1915)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of propylene glycol mono- and diesters of lauric (dodecanoic) acid.

Content

— propylene glycol monolaurate (type I): 45.0 per cent to 70.0 per cent of monoesters and 30.0 per cent to 55.0 per cent of diesters,

— propylene glycol monolaurate (type II): minimum 90.0 per cent of monoesters and maximum 10.0 per cent of diesters.

CHARACTERS

Appearance

Clear, oily liquid at 20 °C, colourless or slightly yellow.

Solubility

Practically insoluble in water, very soluble in alcohol, in methanol and in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.1 g of the substance to be examined in methylene chloride R and dilute to 2 mL with the same solvent.

Reference solution Dissolve 0.1 g of propylene glycol monolaurate CRS in methylene chloride R and dilute to 2 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase hexane R, ether R (30:70 V/V).

Application 10 μ L.

Development Over a path of 15 cm.

Drying In air.

Detection Spray with a 0.1 g/L solution of rhodamine 6 G R in alcohol R. Examine in ultraviolet light at 365 nm.

Results The spots in the chromatogram obtained with the test solution are similar in position to those in the chromatogram obtained with the reference solution.

B. It complies with the test for composition of fatty acids (see Tests).

C. It complies with the assay (content of monoesters).

TESTS

Acid value (2.5.1)

Maximum 4.0, determined on 5.00 g.

Iodine value (2.5.4, Method A)

Maximum 1.0.

D = percentage content of free propylene glycol + percentage content of free fatty acids.
Calculate the percentage content of free fatty acids using the expression:

$$\frac{I_A \times 200}{561.1}$$

I_A = acid value.
— *diesters*: calculate the percentage content of diesters using the following expression:

$$\frac{A+B}{B} \times (100 - D)$$

STORAGE

Protected from moisture.

LABELLING

The label states the type of propylene glycol monolaurate (type I or type II).

Ph Eur

Propylene Glycol

Monopalmitostearate

Propylene Glycol Monostearate

(Ph. Eur. monograph 1469)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of propylene glycol mono- and diesters and of stearic (octadecanoic) and palmitic (hexadecanoic) acids, produced by the condensation of propylene glycol and stearic acid 50 of vegetable or animal origin (see *Stearic acid* (1474)).

Content

Minimum of 50.0 per cent of monoesters.

CHARACTERS

Appearance

White or almost white, waxy solid.

Solubility

Practically insoluble in water, soluble in acetone and in hot alcohol.

IDENTIFICATION

A. Melting point (see Tests).
B. Composition of fatty acids (see Tests).
C. It complies with the assay (monoesters content).

TESTS

Melting point (2.2.15)
33 °C to 40 °C.

Acid value (2.5.1)
Maximum 4.0, determined on 10.0 g.

Iodine value (2.5.4)
Maximum 3.0.

Saponification value (2.5.6)
170 to 185, determined on 2.0 g.

Composition of fatty acids (2.4.22, Method A)
The fatty acid fraction has the following composition:
— *stearic acid*: 40.0 per cent to 60.0 per cent,
— *sum of contents of palmitic acid and stearic acid*: minimum 90.0 per cent.

Free propylene glycol

Maximum 5.0 per cent, determined as prescribed under

Assay.

Total ash (2.4.16)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Size-exclusion chromatography (2.2.30).

Test solution In a 15 mL flask, weigh about 0.2 g (m), to the nearest 0.1 mg. Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Heat gently, if necessary. Reweigh the flask and calculate the total mass of solvent and substance (M).

Reference solutions In four 15 mL flasks, weigh, to the nearest 0.1 mg, about 2.5 mg, 5.0 mg, 10.0 mg and 20.0 mg of *propylene glycol R*. Add 5.0 mL of *tetrahydrofuran R* and shake to dissolve. Weigh the flasks again and calculate the concentration of propylene glycol in milligrams per gram for each reference solution.

Column:

— size: $l = 0.6$ m, $\phi = 7$ mm,

— stationary phase: styrene-divinylbenzene copolymer R (particle diameter 5 μ m, pore size 10 nm).

Mobile phase tetrahydrofuran R.

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 40 μ L.

Relative retention With reference to propylene glycol:

diesters = about 0.78, monoesters = about 0.84.

Limits:

— *free propylene glycol*: from the calibration curve obtained with the reference solutions, determine the concentration (C) in milligrams per gram in the test solution and calculate the percentage content in the substance to be examined using the following expression:

$$\frac{C \times M}{m \times 10}$$

— *monoesters*: calculate the percentage content of monoesters using the following expression:

$$\frac{A}{A+B} \times (100 - D)$$

A = area of the peak due to the monoesters,
 B = area of the peak due to the diesters,
 D = percentage content of free propylene glycol + percentage content of free fatty acids which is determined using the following expression:

$$\frac{I_A \times 270}{561.1}$$

I_A = acid value.

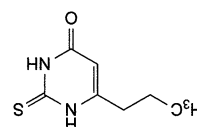
STORAGE

Protected from light.

Ph Eur

Propylthiouracil

(Ph. Eur. monograph 0525)



$C_7H_{10}N_2OS$

170.2

51-52-5

Action and use
Thiourea antithyroid drug.

Preparation

Propylthiouracil Tablets

Ph Eur

DEFINITION

Propylthiouracil contains not less than 98.0 per cent and not more than the equivalent of 100.5 per cent of 2,3-dihydro-6-propyl-2-thioxopyrimidin-4(1H)-one, calculated with reference to the dried substance.

CHARACTERS

White or almost white, crystalline powder or crystals, very slightly soluble in water, sparingly soluble in alcohol. It dissolves in solutions of alkali hydroxides.

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Melting point (2.2.14): 217 °C to 221 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with propylthiouracil CRS. Examine as discs prepared using 1 mg of substance and 0.3 g of potassium bromide R.

C. Examine the chromatograms obtained in the test for impurity A and related substances in ultraviolet light at 254 nm before exposure of the plate to iodine vapour. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To about 20 mg add 8 mL of bromine water R and shake for a few minutes. Boil until the mixture is decolourised, allow to cool and filter. To the filtrate add 2 mL of barium chloride solution R1. A white precipitate is formed whose colour does not become violet on the addition of 5 mL of dilute sodium hydroxide solution R.

TESTS

Impurity A and related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel GF₂₅₄ plate R.

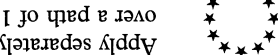
Test solution (a) Dissolve 0.1 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with methanol R.

Reference solution (a) Dissolve 10 mg of propylthiouracil CRS in methanol R and dilute to 10 mL with the same solvent.

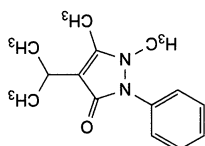
Reference solution (b) Dissolve 50 mg of thiourea R in methanol R and dilute to 100 mL with the same solvent. Dilute 1 mL of this solution to 100 mL with methanol R.

Reference solution (c) Dilute 1 mL of test solution (a) to 100 mL with methanol R.



Propyphenazone

(Ph. Eur. monograph 0636)



$C_{14}H_{18}N_2O$

230.3

479-92-5

Action and use

Pyrazolone analgesic.

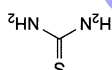
DEFINITION

1,5-Dimethyl-4-(1-methylethyl)-2-phenyl-1,2-dihydro-3H-pyrazol-3-one.

Ph Eur

Ph Eur

A. thiourea.



IMPURITIES

Store protected from light.

STORAGE

$C_7H_{10}N_2OS$.

1 mL of 0.1 M sodium hydroxide is equivalent to 8.511 mg of final titration.

The volume of 0.1 M sodium hydroxide used is equal to the sum of the volume added initially and the volume used in the determining the end-point potentiometrically (2.2.20).

for 5 min and cool. Titrate with 0.1 M sodium hydroxide, Add 50 mL of 0.1 M silver nitrate while stirring, boil gently

sodium hydroxide. Boil and shake until dissolution is complete. To 0.300 g add 30 mL of water R and 30.0 mL of 0.1 M

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

solution (10 ppm Pb) R.

Prepare the reference solution using 2 mL of lead standard

1.0 g complies with test F for heavy metals (20 ppm).

Heavy metals (2.4.8)

solution (c) (1.0 per cent).

spot in the chromatogram obtained with reference corresponding to impurity A is not more intense than the spot apart from the principal spot and any spot

obtained with reference solution (b) (0.05 per cent) and any is not more intense than the spot in the chromatogram

with test solution (a), any spot corresponding to impurity A iodine vapour for 10 min. In the chromatogram obtained

Examine in ultraviolet light at 254 nm. Expose the plate to 50 volumes of chloroform R. Allow the plate to dry in air.

glacial acetic acid R, 6 volumes of 2-propanol R and over a path of 15 cm using a mixture of 0.1 volumes of

Apply separately to the plate 10 µL of each solution. Develop



Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Melting point (2.2.14): 102 °C to 106 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 80 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution Dissolve 80 mg of propyphenazone CRS in methanol R and dilute to 5 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase butanol R, cyclohexane R, ethyl acetate R

(10:45:45 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In a current of hot air for 15 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 1 mL of solution S (see Tests) add 0.1 mL of ferric chloride solution R1. A brownish-red colour appears which becomes yellow on addition of 1 mL of dilute hydrochloric acid R.

TESTS

Solution S

Dissolve 2 g in a mixture of equal volumes of carbon dioxide-free water R and ethanol (96 per cent) R and dilute to 50 mL with the same mixture of solvents.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of phenolphthalein solution R. The solution is colourless. Add 0.2 mL of 0.01 M sodium hydroxide; the solution becomes pink. Add 0.4 mL of 0.01 M hydrochloric acid; the solution becomes colourless.

Add 0.2 mL of methyl red solution R. The solution becomes orange or red.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 1 mg of the substance to be examined and 1 mg of phenazone R (impurity A) in the mobile phase and dilute to 10.0 mL with the mobile phase.

IMPURITIES

Protected from light.

STORAGE

C₁₄H₁₈N₂O.

1 mL of 0.1 M perchloric acid is equivalent to 23.03 mg of acid, determining the end-point potentiometrically (2.2.20).

Dissolve 0.200 g in 10 mL of anhydrous acetic acid R and add 75 mL of ethylene chloride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

ASSAY

Maximum 0.1 per cent, determined on 0.5 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in vacuum at 60 °C for 4 h.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in vacuum at 60 °C for 4 h.

Sulfated ash (2.4.14)

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Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in

Solubility
Sparingly soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (2.2.7): –85 to –65 (dried substance).
Dissolve 1.000 g in 0.1 M hydrochloric acid and dilute to 100.0 mL with the same solvent.

B. In the conditions of the assay, protamine sulfate forms a precipitate.

C. To 0.5 mL of solution S (see Tests) add 4.5 mL of water R, 1.0 mL of a 100 g/L solution of sodium hydroxide R and 1.0 mL of a 0.2 g/L solution of α -naphthol R and mix. Cool the mixture to 5 °C. Add 0.5 mL of sodium hypobromite solution R. An intense red colour is produced.

D. Heat 2 mL of solution S in a water-bath at 60 °C, add 0.1 mL of mercuric sulfate solution R and mix. No precipitate is formed. Cool the mixture in iced water. A precipitate is formed.

E. It gives reaction (a) of sulfates (2.3.1).

TESTS

Solution S
Dissolve 0.20 g in water R and dilute to 10.0 mL with the same solvent.

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution BY₆ or Y₆ (2.2.2, Method II).

To 2.5 mL of solution S add 7.5 mL of water R.

Absorbance (2.2.25)

Maximum 0.1 between wavelengths of 260 nm and 280 nm. Dilute 2.5 mL of solution S to 5.0 mL with water R.

Sulfate

16 per cent to 24 per cent (dried substance).

Dissolve 0.150 g in 15 mL of distilled water R in a beaker. Add 5 mL of dilute hydrochloric acid R. Heat to boiling and slowly add to the boiling solution 10 mL of a 100 g/L solution of barium chloride R. Cover the beaker and heat on a water-bath for 1 h. Filter. Wash the precipitate several times with small quantities of hot water R. Dry and ignite the residue at 600 ± 50 °C to constant mass.
1.0 g of residue is equivalent to 0.4117 g of SO₄.

Iron (2.4.9)

Maximum 10 ppm.

Dissolve 1.0 g with heating in water R and dilute to 10 mL with the same solvent.

Mercury

Maximum 10 ppm.

Introduce 2.0 g of the substance to be examined into a 250 mL ground-glass-stoppered conical flask and add 20 mL of a mixture of equal volumes of nitric acid R and sulfuric acid R. Boil under a reflux condenser for 1 h, cool and cautiously dilute with water R. Boil until nitrous fumes are no longer seen. Cool the solution, cautiously dilute to 200.0 mL with water R, mix and filter. Transfer 50.0 mL of the filtrate to a separating funnel. Shake with successive small portions of chloroform R until the chloroform layer remains colourless. Discard the chloroform layers. To the aqueous layer add 25 mL of dilute sulfuric acid R, 115 mL of water R and 10 mL of a 200 g/L solution of hydroxylamine hydrochloride R. Titrate with diethylenetriamine solution R₂ after each addition, shake the mixture 20 times and towards the end of the titration allow

Protamine Sulfate

Protamine Sulphate

(Ph. Eur. monograph 0569)

9009-65-8



Preparation
Protamine Sulphate Injection

Antidote to heparin.

Action and use

Antidote to heparin.

DEFINITION

Protamine sulfate consists of the sulfates of basic peptides extracted from the sperm or roe of fish, usually species of *Salmonidae* and *Clupeidae*. It binds with heparin in solution, inhibiting its anticoagulant activity, in the conditions of the assay this binding gives rise to a precipitate. Calculated with reference to the dried substance, 1 mg of protamine sulfate precipitates not less than 100 IU of heparin.

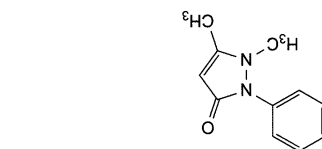
PRODUCTION

The animals from which protamine sulfate is derived must fulfil the requirements for the health of animals suitable for human consumption.
The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.
Abnormal toxicity (2.6.9)
Inject into each mouse 0.5 mg dissolved in 0.5 mL of water for injections R.

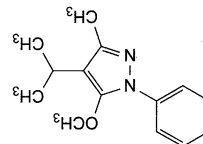
CHARACTERS

Appearance

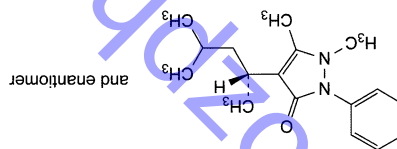
White or almost white, hygroscopic powder.



A. 1,5-dimethyl-2-phenyl-1,2-dihydro-3H-pyrazol-3-one (phenazone),



B. 5-methoxy-3-methyl-4-(1-methylethyl)-1-phenyl-1H-pyrazole,

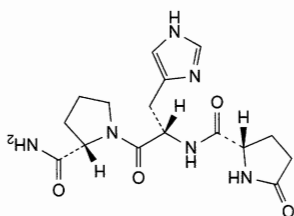


C. 4-[(1RS)-1,3-dimethylbutyl]-1,5-dimethyl-2-phenyl-1,2-dihydro-3H-pyrazol-3-one.

Ph Eur

Protirelin

(Ph. Eur. monograph 1144)

C₁₆H₂₂N₆O₄ 362.4 24305-27-9

Action and use

Thyrotrophin-releasing hormone.

DEFINITION

5-Oxo-L-prolyl-L-histidyl-L-prolinamide.

Synthetic tripeptide with the same sequence of amino acids as the natural hypothalamic neurohormone, which stimulates the release and synthesis of thyrotropin.

Content

97.0 per cent to 102.0 per cent (anhydrous and acetic acid-free substance).

CHARACTERS

Appearance

White or yellowish-white powder, hygroscopic.

Solubility

Very soluble in water, freely soluble in methanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison protirelin CRS.

B. Examine the chromatograms obtained in the assay. Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

TESTS

Appearance of solution

A 10 g/L solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Specific optical rotation (2.2.7)

–62 to –70 (anhydrous and acetic acid-free substance).

Related substances

Dissolve 10 mg in 1.0 mL of water R.

Liquid chromatography (2.2.29).

Test solution Dissolve 5.0 mg of the substance to be examined in mobile phase A and dilute to 5.0 mL with mobile phase A.

Reference solution (a) Dissolve the contents of a vial of D-His-proline CRS in an appropriate volume of mobile phase A to obtain a concentration of 1 mg/mL. Mix equal volumes of this solution and the test solution.

Reference solution (b) Dilute 0.2 mL of the test solution to 10.0 mL with mobile phase A.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm,

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m) with a pore size of 12 nm.

to separate and discard the chloroform layer. Titrate until a bluish-green colour is obtained. Calculate the content of millilitre of titrant, determined in the standardisation of the dithionite solution R2.

Nitrogen

21.0 per cent to 26.0 per cent (dried substance).

Carry out the determination of nitrogen by sulfuric acid digestion (2.5.9), using 10.0 mg and heating for 3–4 h.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Bacterial endotoxins (2.6.14)

Less than 7.0 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Test solution (a) Dissolve 15.0 mg of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 2.0 mL of test solution (a) to 3.0 mL with water R.

Test solution (c) Dilute 1.0 mL of test solution (a) to 3.0 mL with water R.

Use as titrant a 6-fold dilution of heparin sodium BRP in water R (for example, 1.7 mL diluted to 10.0 mL with water R). Titrate each test solution in duplicate as follows:

introduce an accurately measured volume of the solution to be titrated, for example 1.5 mL, into the cell of a suitable colorimeter and set the apparatus for measurement at a suitable wavelength (none is critical) in the visible range. Add the titrant in small volumes until there is a sharp increase in the absorbance and note the volume of titrant added.

Carry out 3 independent assays. For each individual titration, calculate the number of International Units of heparin in the volume of titrant added at the end-point per milligram of the substance to be examined. Calculate the potency of the substance as the average of the 18 values. Test the linearity of the response by the usual statistical methods (for example, 5.3). Calculate the 3 standard deviations for the results obtained with each of the 3 test solutions. Calculate the 3 standard deviations for the results obtained with each of the 3 independent assays. The assay is not valid unless each of the 6 standard deviations is less than 5 per cent of the average result.

STORAGE

In an airtight, tamper-proof container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

Ph Eur



STORAGE

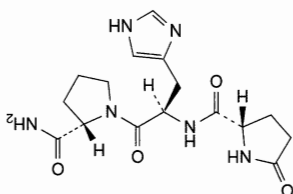
In an airtight container, protected from light at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

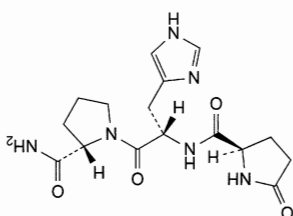
The label states the mass of peptide in the container.

IMPURITIES

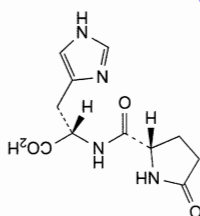
Specified impurities: A, B, C, D, E.



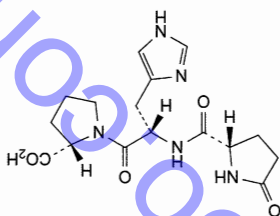
A. 5-oxo-L-prolyl-D-histidyl-L-prolinamide,



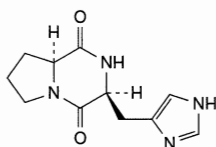
B. 5-oxo-D-prolyl-L-histidyl-L-prolinamide,



C. 5-oxo-L-prolyl-L-histidine,



D. 5-oxo-L-prolyl-L-histidyl-L-proline,



E. (3S,8aS)-3-(1H-imidazol-4-ylmethyl)hexahydropyrido[1,2-a]pyrazine-1,4-dione (cyclo-L-histidyl-L-prolyl-).

Mobile phase:
mobile phase A: a mixture of 100 mL of acetonitrile for chromatography R, 1900 mL of water R and 2.0 g of sodium octanesulfonate R, containing 2.5 mL/L of tetraethylammonium hydroxide solution R₃ adjust to pH 3.5 with phosphoric acid R₃
mobile phase B: a mixture of 300 mL of acetonitrile for chromatography R, 1700 mL of water R and 2.0 g of sodium octanesulfonate R, containing 2.5 mL/L of tetraethylammonium hydroxide solution R₃ adjust to pH 3.5 with phosphoric acid R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	74 → 41	26 → 59
30 - 35	41 → 74	59 → 26
35 - 50	74	26

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL.

Relative retention With reference to protirelin (retention time = about 18 min): impurity C = about 0.2;

impurity D = about 0.68; impurity A = about 0.91;

impurity B = about 0.95; impurity E = about 1.08.

System suitability: reference solution (a):

— **resolution:** minimum 2.5 between the peaks due to impurity A and protirelin,

— **symmetry factor:** 0.9 to 1.2 for the peak due to protirelin.

Limits:

— **any impurity:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (2 per cent),

— **total:** not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3 per cent),

— **disregard limit:** 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

Acetic acid (2.5.34)

Maximum 2.0 per cent.

Test solution Dissolve 40.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of solvents.

Water (2.5.12)

Maximum 7.0 per cent, determined on 0.200 g.

Bacterial endotoxins (2.6.14)

Less than 0.7 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

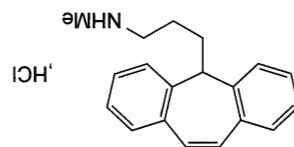
ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Reference solution Dissolve the contents of a vial of protirelin CRS in an appropriate volume of mobile phase A to obtain a concentration of 1.0 mg/mL.

Calculate the content of protirelin (C₁₆H₂₂N₆O₄) using the peak areas of the chromatograms obtained with the test solution and the reference solution and the declared content of C₁₆H₂₂N₆O₄ in protirelin CRS.

Protriptyline Hydrochloride



$C_{19}H_{21}N.HCl$ 299.8

1225-55-4

Action and use
Monoamine reuptake inhibitor; tricyclic antidepressant.

DEFINITION

Protriptyline Hydrochloride is 3-(5H-dibenzo[a,d]cyclohept-5-yl)propyl(methyl)amine hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{19}H_{21}N.HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white to yellowish white powder.
Freely soluble in water and in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

A. Dissolve 0.1 g in 10 mL of water, make alkaline with 1M sodium hydroxide, extract with 5 mL of chloroform, dry with anhydrous sodium sulfate and evaporate the solvent using a current of nitrogen. The infrared absorption spectrum of the oily residue, Appendix II A, is concordant with the reference spectrum of protriptyline (RS 301).

B. Yields reaction A characteristic of chlorides, Appendix VI.

TESTS

Acidity

pH of a 1% w/v solution, 5.0 to 6.5, Appendix V L.

Loss on drying

When dried to constant weight at 60° at a pressure not exceeding 0.7 kPa, loses not more than 0.5% of its weight. Use 1 g.

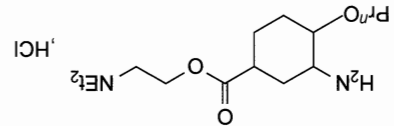
Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.7 g and crystal violet solution as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 29.98 mg of $C_{19}H_{21}N.HCl$.

Proxymetacaine Hydrochloride



$C_{16}H_{26}N_2O_3.HCl$ 330.9

5875-06-9

Action and use
Local anaesthetic.
Preparation
Proxymetacaine Eye Drops

DEFINITION

Proxymetacaine Hydrochloride is 2-diethylaminoethyl 3-amino-4-propoxybenzoate hydrochloride. It contains not less than 98.0% and not more than 102.0% of $C_{16}H_{26}N_2O_3.HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder.
Soluble in water; very soluble in absolute ethanol; practically insoluble in ether.

IDENTIFICATION

A. The light absorption, Appendix II B, in the range 220 to 350 nm of a 0.002% w/v solution exhibits three maxima, at 231, 268 and 310 nm. The absorbances at the maxima, at 268 nm and at 310 nm are about 0.58 and about 0.32, respectively.

B. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of proxymetacaine hydrochloride (RS 303).

C. A 5% w/v solution yields the reaction characteristic of primary aromatic amines and the reactions characteristic of chlorides, Appendix VI.

TESTS

Acidity

pH of a 1% w/v solution, 5.7 to 6.4, Appendix V L.

Related substances

A. Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions of the substance being examined in methanol.

(1) 2.0% w/v of the substance being examined.

(2) 0.020% w/v of the substance being examined.

(3) 0.010% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel GF₂₅₄.

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, heat at 105° for 10 minutes, allow to cool and examine under ultraviolet light (254 nm).

MOBILE PHASE

5 volumes of diethylamine, 30 volumes of ethyl acetate and 75 volumes of toluene.

LIMITS

Any secondary spot in the chromatogram obtained with solution (1);

is not more intense than the spot in the chromatogram obtained with solution (2) (1%);

not more than one such spot is more intense than the spot in the chromatogram obtained with solution (3) (0.5%).

Disregard any spot remaining on the line of application.

B. Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in methanol.

(1) 2.0% w/v of the substance being examined.

(2) 0.0050% w/v of 3-amino-4-propoxybenzoic acid BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel GF₂₅₄.

(b) Use the mobile phase as described below.

(c) Apply 10 µL of each solution.

(d) Develop the plate to 15 cm.
(e) After removal of the plate, dry in air, examine under ultraviolet light (254 nm).

MOBILE PHASE
4 volumes of glacial acetic acid, 20 volumes of cyclohexane and 80 volumes of 1,4-dioxan.

LIMITS
Any secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.25%).
The principal spot remains on or near the line of application. When dried at 105° for 3 hours, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash
Not more than 0.15%, Appendix IX A.

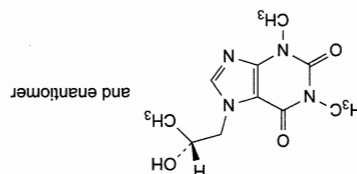
ASSAY
Carry out Method I for non-aqueous titration,

Appendix VIII A, using 0.25 g, 20 mL of mercury(II) acetate solution and 1-naphtholbenzenesulfonic acid as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 16.54 mg of $C_{16}H_{26}N_2O_3 \cdot HCl$.

STORAGE
Proxymetacaine Hydrochloride should be protected from light.

Proxiphylline

(Ph. Eur. monograph 0526)



$C_{10}H_{14}N_4O_3$ 238.2 603-00-9

Action and use
Non selective phospho-diesterase inhibitor; treatment of reversible airways obstruction.

DEFINITION

Proxiphylline contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of 7-[(2RS)-2-hydroxypropyl]-1,3-dimethyl-3,7-dihydro-1H-purine-2,6-dione, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder, very soluble in water, soluble in alcohol.

IDENTIFICATION

First identification B, C
Second identification A, C, D

A. Melting point (2.2.14): 134 °C to 136 °C.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with proxiphylline CRS. Examine the substances as discs prepared

using 0.5 mg to 1 mg of the substance to be examined in 0.3 g of potassium bromide R.
C. Dissolve 1 g in 5 mL of acetic anhydride R and boil under a reflux condenser for 15 min. Allow to cool and add 100 mL of a mixture of 20 volumes of ether R and 80 volumes of light petroleum R. Cool in iced water for at least 20 min, shaking from time to time. Filter, wash the precipitate with a mixture of 20 volumes of ether R and 80 volumes of light petroleum R, recrystallise from alcohol R and dry *in vacuo*. The crystals melt (2.2.14) at 87 °C to 92 °C.
D. It gives the reaction of xanthines (2.3.1).

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.25 mL of bromothymol blue solution R1. The solution is yellow or green. Not more than 0.4 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to blue.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel HF₂₅₄ R as the coating substance.
Test solution Dissolve 0.3 g of the substance to be examined in a mixture of 20 volumes of water R and 30 volumes of methanol R and dilute to 10 mL with the same mixture of solvents. Prepare immediately before use.
Reference solution (a) Dilute 1 mL of the test solution to 100 mL with methanol R.
Reference solution (b) Dilute 0.2 mL of the test solution to 100 mL with methanol R.
Reference solution (c) Dissolve 10 mg of theophylline R in methanol R, add 0.3 mL of the test solution and dilute to 10 mL with methanol R.

Apply separately to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 1 volume of concentrated ammonia R, 10 volumes of ethanol R and 90 volumes of chloroform R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (1 per cent) and at most one such spot is more intense than the spot in the chromatogram obtained with reference solution (b) (0.2 per cent). The test is not valid unless the chromatogram obtained with reference solution (c) shows two clearly separated spots.

Chlorides (2.4.4)

Dilute 2.5 mL of solution S to 15 mL with water R. The solution complies with the limit test for chlorides (400 ppm).

Heavy metals (2.4.8)

12 mL of solution S complies with test A for heavy metals (20 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.200 g in 3.0 mL of anhydrous formic acid R and add 50.0 mL of acetic anhydride R. Titrate with 0.1 M

perchloric acid determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 23.82 mg of

$C_{10}H_{14}N_4O_3$.

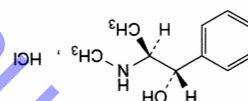
STORAGE

Store protected from light.

Ph Eur

Pseudoephedrine Hydrochloride

(Ph. Eur. monograph 1367)



$C_{10}H_{16}ClNO$ 201.7 345-78-8

Action and use

Adrenoceptor agonist.

Preparations

Pseudoephedrine Oral Solution

Pseudoephedrine Tablets

Ph Eur

DEFINITION

(1*S*,2*S*)-2-(Methylamino)-1-phenylpropan-1-ol hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water and in ethanol (96 per cent), sparingly soluble in methylene chloride.

IDENTIFICATION

First identification A, B, D.

Second identification A, C, D.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

C. Comparison pseudoephedrine hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of pseudoephedrine hydrochloride CRS in methanol R and dilute to 10 mL with the

same solvent.

Reference solution (b) Dissolve 10 mg of ephedrine hydrochloride CRS in reference solution (a) and dilute to 5 mL with reference solution (a).

Plate TLC silica gel plate R.

Mobile phase methylamine chloride R, concentrated ammonia R, 2-propanol R (5:15:80 V/V/V).

Application 10 μ L.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 110 °C for 5 min.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Solution S (see Tests) gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 1.25 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

Dilute 2 mL of solution S to 10 mL with carbon dioxide-free water R. Add 0.1 mL of methyl red solution R and 0.1 mL of 0.01 M sodium hydroxide; the solution is yellow. Add 0.2 mL of 0.01 M hydrochloric acid; the solution is red.

Specific optical rotation (2.2.7)

+ 61.0 to + 62.5 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dissolve 20.0 mg of ephedrine hydrochloride CRS (impurity A) in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Reference solution (c) Dissolve 10 mg of ephedrine hydrochloride CRS (impurity A) in 5 mL of the test solution and dilute to 100 mL with the mobile phase.

Column:

— size: 1 = 0.25 m, ϕ = 4.6 mm;

— stationary phase: phenylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 6 volumes of methanol R and 94 volumes of an 11.6 g/L solution of ammonium acetate R previously adjusted to pH 4.0 with glacial acetic acid R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 257 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of pseudoephedrine.

Relative retention With reference to pseudoephedrine

(retention time = about 18 min): impurity A = about 0.9.

System suitability: reference solution (c):
— resolution: minimum 2.0 between the peaks due to impurity A and pseudoecephedrine; if necessary, reduce the content of methanol in the mobile phase.

Limits:
— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— sum of impurities other than A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

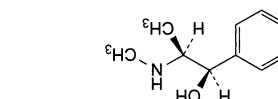
Dissolve 0.170 g in 30 mL of ethanol (96 per cent) R. Add 5.0 mL of 0.01 M hydrochloric acid. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.
1 mL of 0.1 M sodium hydroxide is equivalent to 20.17 mg of C₁₀H₁₆ClNO.

STORAGE

Protected from light.

IMPURITIES

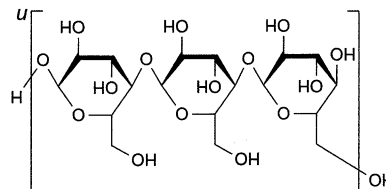
Specified impurities A.



A. (1R,2S)-2-(methylamino)-1-phenylpropan-1-ol (ephedrine).

Pullulan

(Ph. Eur. monograph 2603)



C_{18n}H_{30n+2}O_{15n+1}

Action and use
Excipient.

9057-02-7



Ph Eur

DEFINITION

Neutral polysaccharide produced by the growth of *Aureobasidium pullulans*. It has a chain structure consisting of α-(1→6)-linked maltotriose units, each of which is composed of three α-(1→4)-linked D-glucopyranose units. It may contain some maltotetraose units.

CHARACTERS

Appearance

White or almost white, very hygroscopic powder.

Solubility

Freely soluble in water, practically insoluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

A. Dissolve 10 g in 100 mL of water R, adding the substance to be examined in small quantities while stirring. A viscous solution is produced.

B. Mix 10 mL of the solution obtained in identification test A with 0.1 mL of a 10 units/mL solution of pullulanase R. Incubate at 25 °C for about 20 min.

A substantial decrease in viscosity is observed.

C. Dissolve 2 g in 100 mL of water R. To 10 mL of the solution, add 2 mL of macrogol 600 R. A white precipitate is formed immediately.

TESTS

pH (2.2.3)

4.5 to 6.5.

Dissolve 1.0 g in 10 mL of carbon dioxide-free water R.

Viscosity (2.2.9)

100 mm²·s⁻¹ to 180 mm²·s⁻¹.

Dissolve 10.0 g (dried substance) in water R to make 100 g of solution, adding the substance to be examined in small quantities while stirring. Carry out the test at 30 ± 0.1 °C.

Mono-, di- and oligosaccharides

Maximum 10.0 per cent for the sum of mono-, di- and oligosaccharides.

Stock solution Dissolve 0.80 g (dried substance) in water R, adding the substance to be examined in small quantities while stirring, and dilute to 100.0 mL with the same solvent.

Test solution To 1.0 mL of the stock solution, add 0.1 mL of a saturated solution of potassium chloride R (2 g in 5 mL).

Mix vigorously with 3 mL of methanol R. Centrifuge at 12 000-16 000 g for 10 min maintaining at 4 °C. Use the supernatant.

Reference solution Dilute 1.0 mL of the stock solution to 50.0 mL with water R.

Blank solution water R.

Transfer 0.2 mL of the test solution, the reference solution and the blank solution to separate test tubes, placed in a bath of iced water with each containing 5 mL of a 2 g/L solution of anthrone R in sulfuric acid R1. Stir immediately, then heat at 90 °C for 10 min. Cool the tubes under running water. Measure the absorbance (2.2.25) of the test solution, the reference solution and the blank solution at 620 nm, using water R as the compensation liquid.
Calculate the percentage content of mono-, di- and oligosaccharides using the following expression:

$$100 \times \frac{4.1}{50} \times \frac{A_1 - A_2}{A_3 - A_2}$$

4.1 = dilution factor for the test solution;

- 50 = dilution factor for the reference solution;
 A_1 = absorbance of the test solution;
 A_2 = absorbance of the blank solution;
 A_3 = absorbance of the reference solution.

Loss on drying (2.2.32)

Maximum 6.0 per cent, determined on 1.000 g by drying *in vacuo* at 90 °C for 6 h.

Sulfated ash (2.4.14)

Maximum 0.3 per cent, determined on 2.0 g.

Nitrogen (2.5.9)

Maximum 0.05 per cent (dried substance), determined on 4.0 g.

Carry out the determination of nitrogen using 40 mL of *sulfuric acid R* and heating until a green colour is obtained (about 1.5 h) to complete the decomposition and add strong *sodium hydroxide solution R* until a dark brown colour is obtained (about 90 mL).

Microbial contamination

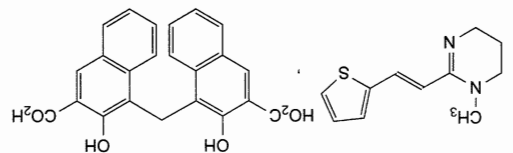
TAMC: acceptance criterion 10^2 CFU/g (2.6.12).

STORAGE

In an airtight container.

Pyrantel Embonate

(Ph. Eur. monograph 1680)



$C_{34}H_{30}N_2O_6S$ 594.7 22204-24-6

Action and use

Anthelmintic.

Ph Eur

DEFINITION

1-Methyl-2-[(E)-2-(thiophen-2-yl)ethenyl]-1,4,5,6-tetrahydropyrimidine hydrogen tetrahydrophthalene-2-carboxylate.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Pale yellow or yellow powder.

Solubility

Practically insoluble in water, soluble in dimethyl sulfoxide, practically insoluble in methanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison pyrantel embonate CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and strictly protect from light at all stages.

Solvent mixture Mix 5 volumes of glacial acetic acid *R* with 5 volumes of water *R* and add 2 volumes of diethylamine *R* with cooling.

Test solution Dissolve 80 mg in 7 mL of the solvent mixture and dilute to 100.0 mL with acetonitrile *R*.

Reference solution (a) Dissolve 10.0 mg of pyrantel impurity *A* CRS in the solvent mixture, add 2.5 mL of the test solution and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: silica gel for chromatography *R* (5 μ m).

Mobile phase Solvent mixture, acetonitrile for chromatography *R* (72.928 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 288 nm.

Injection 20 μ L.

Run time 4 times the retention time of pyrantel.

Relative retention With reference to pyrantel (retention time = about 11 min): embonic acid = about 0.5;

impurity *B* = about 1.3; impurity *A* = about 1.8 (impurity *A* also gives rise to an embonate peak).

System suitability: reference solution (a):

— resolution: minimum 4.0 between the peaks due to pyrantel and impurity *A*.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity *B* by 0.4;

— **impurity A:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— **impurity B:** not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **sum of impurities other than A and B:** not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— **disregard limit:** 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (2.4.4)

Maximum 360 ppm.

To 0.46 g add 10 mL of dilute nitric acid *R* and 30 mL of water *R*. Heat on a water-bath for 5 min. Cool, dilute to 50 mL with water *R*, mix well and filter.

Sulfates (2.4.13)

Maximum 0.1 per cent.

To 0.50 g add 2.5 mL of dilute nitric acid *R* and dilute to 50 mL with distilled water *R*. Heat on a water-bath for 5 min, shake for 2 min, cool and filter.

Iron (2.4.9)

Maximum 75 ppm.

Ignite 0.66 g at 800 ± 50 °C for 2 h. Dissolve the residue in 2.5 mL of dilute hydrochloric acid *R* with gentle heating for 10 min. Cool and dilute to 50 mL with water *R*.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution

using 2.0 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying

in vacuo at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

To 0.450 g add 10 mL of acetic anhydride R and 50 mL

glacial acetic acid R, heat at 50 °C and stir for 10 min. Allow

to cool (a clear solution is not obtained). Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2.20). Carry out a blank titration.

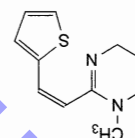
1 mL of 0.1 M perchloric acid is equivalent to 59.47 mg of

 $C_3H_3N_2O_5$.**STORAGE**

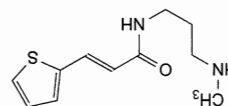
Protected from light.

IMPURITIES

Specified impurities A, B.



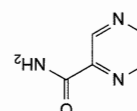
A. 1-methyl-2-[(Z)-2-(thiophen-2-yl)ethenyl]-1,4,5,6-tetrahydropyrimidine,



B. (E)-N-[3-(methylamino)propyl]-3-(thiophen-2-yl)prop-2-enamide.

Pyrazinamide

(Ph. Eur. monograph 0859)

 $C_5H_5N_3O$

123.1

98-96-4

Action and use

Antituberculous drug.

Preparations

Pyrazinamide Oral Suspension

Pyrazinamide Tablets

DEFINITION

Pyrazine-2-carboxamide.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Sparsingly soluble in water, slightly soluble in ethanol

(96 per cent) and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Melting point (2.2.14): 188 °C to 191 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution (a) Dissolve 50.0 mg in water R and dilute to

100.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to

10.0 mL with water R.

Test solution (c) Dilute 2.0 mL of test solution (a) to

100.0 mL with water R.

Spectral range 290-350 nm for test solution (b); 230-290 nm

for test solution (c).

Absorption maxima At 310 nm for test solution (b);

at 268 nm for test solution (c).

Specific absorbance at the absorption maximum at 268 nm

640 to 680 for test solution (c).

C. Infrared absorption spectrophotometry (2.2.24).

Comparison pyrazinamide CRS.

If the spectra obtained show differences, dissolve the

substance to be examined and the reference substance

separately in ethanol (96 per cent) R, evaporate to dryness and

record new spectra using the residues.

D. Dissolve 0.1 g in 5 mL of water R. Add 1 mL of ferrous

sulfate solution R2. The solution becomes orange. Add 1 mL

of dilute sodium hydroxide solution R. The solution becomes

dark blue.

TESTS**Solution S**

Dissolve 0.5 g in carbon dioxide-free water R and dilute to

50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 25 mL of solution S add 0.05 mL of phenolphthalein

solution R1 and 0.2 mL of 0.01 M sodium hydroxide.

The solution is red. Add 1.0 mL of 0.01 M hydrochloric acid.

The solution is colourless. Add 0.15 mL of methyl red

solution R. The solution is red.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Test solution Dissolve 50 mg of the substance to be examined

in water R and dilute to 25.0 mL with the same solvent.

Dilute 5.0 mL of the solution to 25.0 mL with water R.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with water R. Dilute 1.0 mL of this solution to

10.0 mL with water R.

Reference solution (b) Dissolve 10 mg of pyrazine-2-

carbonitrile R (impurity B) in water R and dilute to 50.0 mL

with the same solvent. Dilute 5.0 mL of the solution to

50.0 mL with water R. To 5.0 mL of this solution add

5.0 mL of the test solution and dilute to 25.0 mL with water R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 30 °C.

Mobile phase Dissolve 6.80 g of potassium dihydrogen phosphate R in 800 mL of water R, add 1.84 g of sodium hydroxide R, adjust to pH 3.0 with dilute phosphoric acid R and dilute to 1000 mL with water R; add 10.0 mL of acetonitrile R and 1.0 mL of tetrahydrofuran R.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 40 μ L.

Run time 4 times the retention time of pyrazinamide.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to pyrazinamide (retention time = about 5 min): impurity B = about 1.6.

System suitability: reference solution (b):

— resolution: minimum 4.0 between the peaks due to pyrazinamide and impurity B.

Limits:

— impurity B: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— disregard limit: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.03 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture water R, ethanol (96 per cent) R (50:50 V/V).

0.25 g complies with test H. Prepare the reference solution using 0.25 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 50 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 12.31 mg of $C_5H_5N_3O$.

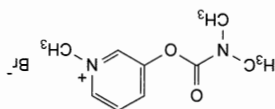
IMPURITIES

Specified impurities B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

Pyridostigmine Bromide

(Ph. Eur. monograph 1255)



$C_9H_{13}BrN_2O_2$

261.1

101-26-8

Action and use

Cholinesterase inhibitor.

Preparation

Pyridostigmine Tablets

Ph. Eur.

DEFINITION

3-[(Dimethylcarbamoyloxy)-1-methylpyridinium bromide.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline, deliquescent powder.

Solubility

Very soluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison pyridostigmine bromide CRS.

B. It gives reaction (a) of bromides (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

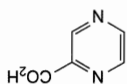
Acidity or alkalinity

To 40 mL of solution S add a few drops of methyl red solution R. To 20 mL of this solution add 0.2 mL of 0.02 M sodium hydroxide. The solution is yellow. To the other 20 mL add 0.2 mL of 0.02 M hydrochloric acid. The solution is red.

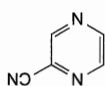
Related substances

Liquid chromatography (2.2.29).

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): 4.



A. pyrazine-2-carboxylic acid,



B. pyrazine-2-carbonitrile.

Ph. Eur.



ASSAY

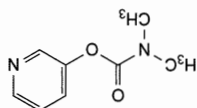
Dissolve 0.230 g in 10 mL of anhydrous acetic acid R. Add 40 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 26.11 mg of $C_8H_{13}BrN_2O_2$.

STORAGE

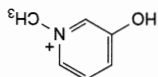
In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container, protected from light.

IMPURITIES

Specified impurities: A, B.



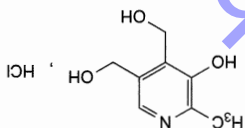
A. pyridin-3-yl dimethylcarbamate,



B. 3-hydroxy-1-methylpyridinium.

Pyridoxine Hydrochloride

(Ph. Eur. monograph 02455)



$C_8H_{12}ClNO_3$ 205.6

58-56-0

Action and use

Vitamin B₆.

Preparations

Pyridoxine Tablets

Vitamins B and C Injection

When vitamin B₆ is prescribed or demanded, Pyridoxine Hydrochloride shall be dispensed or supplied.

DEFINITION

(5-Hydroxy-6-methylpyridine-3,4-diyl)dimethanol hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

m.p.

About 205 °C, with decomposition.

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase at about 40 °C. Allow to cool and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 4 mg of pyridoxine bromide CRS, 4 mg of pyridoxine impurity A CRS and 4 mg of pyridoxine impurity B CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5–10 μ m).

Mobile phase Mix 30 volumes of acetonitrile R and 70 volumes of a 4.33 g/L solution of sodium dodecyl sulfate R previously adjusted to pH 2.0 with phosphoric acid R.

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Run time Twice the retention time of pyridoxine.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B.

Relative retention With reference to pyridoxine (retention time = about 3.2 min): impurity B = about 0.7;

impurity A = about 0.9.

System suitability: reference solution (a):

— resolution: minimum 1.5 between the peaks due to impurity A and pyridoxine.

Limits:

— impurities A, B: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

at most one such peak has an area greater than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.10 per cent);

— total: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (c)

(0.05 per cent); disregard the peak due to the bromide ion.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Solution A Dilute 1.0 mL of solution S (see Tests) to 50.0 mL with 0.1 M hydrochloric acid.

Solution B Dilute 1.0 mL of solution A to 100.0 mL with 0.1 M hydrochloric acid.

Solution C Dilute 1.0 mL of solution A to 100.0 mL with the potassium dihydrogen phosphate 0.025 M + disodium hydrogen phosphate 0.025 M solution described in chapter 2.2.3.

Spectral ranges 250–350 nm for solution B; 220–350 nm for solution C.

Absorption maxima 288–296 nm for solution B; 248–256 nm and 320–327 nm for solution C.

Specific absorbances at the absorption maxima:

- 425–445 for solution B at 288–296 nm;
- 175–195 for solution C at 248–256 nm;
- 345–365 for solution C at 320–327 nm.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison pyridoxine hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 1.0 g of the substance to be examined in water R and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 10 mL with water R.

Reference solution Dissolve 0.10 g of pyridoxine hydrochloride CRS in water R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R, methylene chloride R, tetrahydrofuran R, acetone R (9:13:13:65 V/V/V/V).

Application 2 µL.

Development In an unsaturated tank, over a path of 15 cm.

Drying In air.

Detection Spray with a 50 g/L solution of sodium carbonate R in a mixture of 30 volumes of ethanol (96 per cent) R and 70 volumes of water R; dry in a current of air, spray with a 1 g/L solution of dichloroguanmonochlorimide R in ethanol (96 per cent) R and examine immediately.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Solution S gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

pH (2.2.3)

2.4 to 3.0 for solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.

C₈H₁₂ClNO₃.

1 mL of 0.1 M perchloric acid is equivalent to 20.56 mg of

potentiometrically (2.2.20). Carry out a blank titration.

0.1 M perchloric acid, determining the end-point

Add 50 mL of acetic anhydride R. Titrate with

Dissolve 0.150 g in 5 mL of anhydrous formic acid R.

end-point has been reached.

In order to avoid overheating in the reaction medium, mix thoroughly throughout and stop the titration immediately after the

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

reference solution using lead standard solution (1 ppm Pb) R.

12 mL of solution S complies with test A. Prepare the

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

(0.2 per cent);

the chromatogram obtained with reference solution (a)

— total: not more than twice the area of the principal peak in

with reference solution (a) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

reference solution (a) (0.15 per cent);

principal peak in the chromatogram obtained with

peak area of impurity B by 1.5;

— correction factor: for the calculation of content, multiply the

Limits:

impurities A and B.

— resolution: minimum 1.5 between the peaks due to

System suitability: reference solution (b):

impurity B = about 1.9.

time = about 12 min; impurity A = about 1.7;

Relative retention With reference to pyridoxine (retention

impurities A and B.

Identification of impurities Use the chromatogram obtained

with reference solution (b) to identify the peaks due to

Run time 2.5 times the retention time of pyridoxine.

Injection 5 µL.

Detection Spectrophotometer at 210 nm.

Flow rate 1.0 mL/min.

Mobile phase Dissolve 2.72 g of potassium dihydrogen

phosphate R in 900 mL of water R, adjust to pH 3.0 with

silica gel for chromatography R (5 µm).

— stationary phase: base-deactivated end-capped octadecylsilyl

— size: l = 0.25 m, Ø = 4.6 mm;

Column:

10.0 mL with water R.

with the same solvent. Dilute 2.0 mL of this solution to

hydrochloride R (impurity B) in water R and dilute to 10.0 mL

impurity A CRS and 2.5 mg of 4-deoxypyridoxine

Reference solution (b) Dissolve 2.5 mg of pyridoxine

10.0 mL with water R.

Reference solution (a) Dilute 1.0 mL of the test solution to

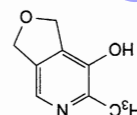
STORAGE

Protected from light.

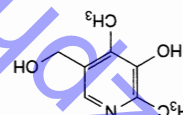
IMPURITIES

Specified impurities B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A.



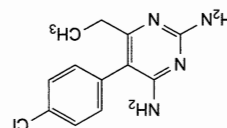
A. 6-methyl-1,3-dihydroxy-3,4-c[pyridin-7-yl]-5-(hydroxymethyl)-2,4-dimethylpyridin-3-ol.



B. 5-(hydroxymethyl)-2,4-dimethylpyridin-3-ol.

Pyrimethamine

(Ph. Eur. monograph 0288)



$C_{12}H_{13}ClN_4$

248.7

58-14-0

Action and use

Dihydrofolate reductase inhibitor; antiprotozoal (malaria).

Preparation

Pyrimethamine Tablets

Ph Eur

DEFINITION

Pyrimethamine contains not less than 99.0 per cent and more than the equivalent of 101.0 per cent of 5-(4-chlorophenyl)-6-ethylpyrimidine-2,4-diamine, calculated with reference to the dried substance.

CHARACTERS

An almost white, crystalline powder or colourless crystals, practically insoluble in water, slightly soluble in alcohol.

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Melting point (2.2.14): 239 °C to 243 °C.

B. Dissolve 0.14 g in ethanol R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with 0.1 M hydrochloric acid. Dilute 10.0 mL of this solution to 100.0 mL with 0.1 M hydrochloric acid.

Sulfates (2.4.13)

15 mL of solution S complies with the limit test for sulfates (80 ppm). Prepare the standard using a mixture of 2.5 mL of

(0.25 per cent).

chromatogram obtained with reference solution (b)

principal spot, is not more intense than the spot in the

chromatogram obtained with test solution (a), apart from the

Examine in ultraviolet light at 254 nm. Any spot in the

and 76 volumes of toluene R. Allow the plate to dry in air.

8 volumes of propanol R, 12 volumes of glacial acetic acid R,

path of 10 cm using a mixture of 4 volumes of chloroform R,

Apply to the plate 20 µL of each solution. Develop over a

9 volumes of chloroform R.

10 mL with a mixture of 1 volume of methanol R and

9 volumes of chloroform R. Dilute 1 mL of the solution to

100 mL with a mixture of 1 volume of methanol R and

Reference solution (b) Dilute 2.5 mL of test solution (a) to

solvents.

chloroform R and dilute to 100 mL with the same mixture of

a mixture of 1 volume of methanol R and 9 volumes of

Reference solution (a) Dissolve 0.1 g of pyrimethamine CRS in

chloroform R.

with a mixture of 1 volume of methanol R and 9 volumes of

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL

mixture of solvents.

9 volumes of chloroform R and dilute to 25 mL with the same

examined in a mixture of 1 volume of methanol R and

Test solution (a) Dissolve 0.25 g of the substance to be

immediately before use.

gel GF₂₅₄ R as the coating substance. Prepare the solutions

Examine by thin-layer chromatography (2.2.27), using silica

Related substances

The solution is red or orange.

hydrochloric acid and 0.05 mL of methyl red solution R.

colour of the indicator to pink. Add 0.4 mL of 0.01 M

0.2 mL of 0.01 M sodium hydroxide is required to change the

solution R1. The solution is colourless. Not more than

To 10 mL of solution S add 0.05 mL of phenolphthalein

Acidity or alkalinity

(2.2.2, Method II).

and not more intensely coloured than reference solution BY₆

the same mixture of solvents. The solution is clear (2.2.1)

3 volumes of methylene chloride R and dilute to 10 mL with

Dissolve 0.25 g in a mixture of 1 volume of methanol R and

Prepare the solution immediately before use.

Appearance of solution

filter.

Shake 1.0 g with 50 mL of distilled water R for 2 min and

Solution S**TESTS**

solution (a).

spot in the chromatogram obtained with reference

solution (b) is similar in position and size to the principal

The principal spot in the chromatogram obtained with test

related substances in ultraviolet light at 254 nm.

D. Examine the chromatograms obtained in the test for

pyrimethamine CRS.

(2.2.24), comparing with the spectrum obtained with

C. Examine by infrared absorption spectrophotometry

the maximum is 310 to 330.

absorption minimum at 261 nm. The specific absorbance at

solution shows an absorption maximum at 272 nm and an

Examined between 250 nm and 300 nm (2.2.25), the

sulfate standard solution (10 ppm SO_4) R and 12.5 mL of

distilled water R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 0.50 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 25 mL of *anhydrous acetic acid* R, heating gently. Cool. Titrate with 0.1 M *perchloric acid* determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M *perchloric acid* is equivalent to 24.87 mg of $\text{C}_{12}\text{H}_{13}\text{ClN}_4$.

STORAGE

Store protected from light.

Ph Eur

Pyroxylin

Cellulose Nitrate

Preparation

Flexible Collodion

DEFINITION

Pyroxylin is a nitrated cellulose obtained by the action of a mixture of nitric and sulfuric acids on wood pulp or cotton linters that have been freed from fatty matter. It must be damped with not less than 25% by weight of Isopropyl Alcohol or of Industrial Methylated Spirit.

CAUTION Compliance is required with the provisions of relevant legislation relating to the storage of, use of and containers used for Pyroxylin.

In the following tests, particular care should be exercised when drying pyroxylin. The material so obtained is explosive and sensitive to ignition by impact or friction and it should be handled as carefully as possible.

CHARACTERISTICS

White or almost white, cuboid granules or fibrous material, the latter resembling absorbent cotton but harsher to the touch and more powdery. Both the granules and the fibrous material appear moist. Highly flammable.

IDENTIFICATION

Add water to a solution in *acetone*. A white, viscid mass is readily precipitated.

TESTS

Clarity and colour of solution

A 10% w/v solution dissolves at 20° in a mixture of 1 volume of *ethanol* (90%) and 3 volumes of *ether* to produce an almost clear and colourless to pale straw-coloured solution.

Kinematic viscosity

1160 to 2900 mm^2s^{-1} when determined in the following manner. To 20 g, previously dried to constant weight by heating on a water-bath at 80° and allowing to cool in a desiccator over *silica gel*, add 200 mL of a mixture of 19 volumes of *acetone* and 1 volume of *water*. Shake vigorously to prevent the formation of aggregates of partly solvated pyroxylin and rotate suitably to obtain a homogeneous solution. Measure the viscosity of the solution within 48 hours using a falling sphere viscometer complying with British Standard 188: 1977 (Methods for the

Nitrogen

11.7 to 12.2%, calculated with reference to the material dried to constant weight by heating on a water-bath at 80° and allowing to cool in a desiccator over *silica gel*, when determined by the following method. Transfer 0.4 g of the dried substance to a 750 mL round-bottomed flask using (20 vol) and then add slowly, with shaking, 50 mL of a 30% w/v solution of *potassium hydroxide* and 5 mL of *ethanol* (96%). Agitate slowly for 3 hours or until the substance being examined has dissolved. Add 3 g of *Devarda's alloy* and immediately connect the flask to the spray trap of an ammonia-distillation apparatus, the receiver of which contains 50 mL of 0.1M *hydrochloric acid VS* and 0.15 mL of *methyl red solution*. Allow the reaction to proceed until no further evolution of gas occurs and then heat the contents of the flask to boiling and distil carefully until 30 mL remains in the round-bottomed flask. Rinse the delivery tube into the receiver with *water* and titrate the combined distillate and rinsings with 0.1M *sodium hydroxide VS*. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of acid required to neutralise the ammonia formed. Each mL of 0.1M *hydrochloric acid VS* is equivalent to 1.401 mg of nitrogen.

STORAGE

Pyroxylin should be loosely packed, protected from light and stored at a temperature not exceeding 15°, remote from fire. The container should be suitably designed to disrupt should the internal pressure reach or exceed 1400 kPa. The amount of damping fluid must not be allowed to fall below 25% w/w; should this happen, the material should be either rewetted or used immediately for the preparation of Collodion.

$$v = \frac{d^2 g (\delta - \rho)}{0.867 \times 0.187 \rho}$$

where d = the diameter of the sphere in cm,

δ = density of the sphere in g cm^{-3} ,

ρ = density of the liquid being tested in g cm^{-3} ,

v = velocity of fall in cm s^{-1} ,

g = local acceleration due to gravity in cm s^{-2} .

the expression:

determination of viscosity of liquids). Fill the fall tube with the solution being examined to about 10 mm above the 220-mm mark, place vertically in the bath and allow to stand for air bubbles to clear and for temperature equilibrium to be attained. Clean the sphere, immerse it in a portion of the liquid being examined maintained at a temperature of 19.9° to 20.1° and when it is at this temperature introduce it, without wiping, into the delivery tube. Observe the time for the lowest part of the sphere to pass through the planes of the tops of the 175-mm mark and the 25-mm mark, using a telescope or other suitable device to avoid errors due to parallax. The average of three readings concordant to within 0.5% is taken as the time of fall. Calculate the kinematic viscosity (ν) in square millimetres per second (mm^2s^{-1}) from the expression:

Pyrrolidone

(Ph. Eur. monograph 2180)



C₄H₇NO

85.1

616-45-5

Ph Eur

DEFINITION

Pyrrolidin-2-one.

CHARACTERS

Appearance Clear, colourless or slightly greyish liquid, or white or almost white crystals, or colourless crystal needles.

Solubility

Miscible with water, with ethanol (96 per cent) and with most common organic solvents.

mp

About 25 °C; the molten substance remains liquid at

temperatures below the melting point.

bp

About 245 °C.

IDENTIFICATION

First identification A.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison pyrrolidone CRS.

B. Relative density (2.2.5): 1.112 to 1.115.

C. Refractive index (2.2.6): 1.487 to 1.490.

TESTS

Use the molten substance for all tests.

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than intensity 7 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Alkalinity

To 100 mL of water R add 1.0 mL of bromothymol blue solution R1 and adjust to a green colour with 0.02 M potassium hydroxide or 0.02 M hydrochloric acid. To 50 mL of this solution add 20 mL of the substance to be examined and titrate with 0.02 M hydrochloric acid to the initial colour. Not more than 8.0 mL of 0.02 M hydrochloric acid is required.

Related substances

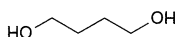
Gas chromatography (2.2.28): use the normalisation procedure.

Test solution The substance to be examined.

Reference solution (a) Dissolve 1 mL of the substance to be examined and 1 mL of *N*-methylpyrrolidone R (impurity C) in methylene chloride R and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 1.1 g of the substance to be the same solvent. Dilute 1.0 mL of this solution to 20.0 mL with methylene chloride R.

Reference solution (c) Dissolve 1 mL of butyrolactone R (impurity B) and 1 mL of butane-1,4-diol R (impurity A) in methylene chloride R and dilute to 20 mL with the same solvent.



A. butane-1,4-diol,

IMPURITIES

Protected from light.

STORAGE

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.00 g.

Water (2.5.32)

solution (2 ppm Pb) R.

Prepare the reference solution using lead standard

Dissolve 4.0 g in water R and dilute to 20.0 mL with the same solvent. 12 mL of the solution complies with test A.

Maximum 10 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

chromatogram obtained with reference solution (b)

— disregard limit: the area of the principal peak in the

total: maximum 0.7 per cent;

0.10 per cent;

— unspecified impurities: for each impurity, maximum

0.15 per cent;

— impurities A, C: for each impurity, maximum

— impurity B: maximum 0.5 per cent;

Limits:

impurity C and pyrrolidone.

— resolution: minimum 2.0 between the peaks due to

System suitability: reference solution (a):

identify the peak due to impurity C.

chromatogram obtained with reference solution (a) to

to identify the peaks due to impurities A and B; use the

Use the chromatogram obtained with reference solution (c)

impurity A = about 0.76; impurity C = about 0.97.

time = about 13 min): impurity B = about 0.73;

Relative retention With reference to pyrrolidone (retention

Injection 0.1 µL.

Detection Flame ionisation.

Temperature (°C)	Time (min)	Column	Injection port	Detector
100 → 250	0 - 18.75	18.75 - 30	250	250

Temperature

Split ratio 1:80.

Flow rate 1.3 mL/min.

Carrier gas nitrogen for chromatography R.

5 µm).

— stationary phase: poly(dimethyl)siloxane R (film thickness

— size: l = 30 m, Ø = 0.32 mm;

— material: fused silica;

Column:



B, dihydrofuran-2(3H)-one (γ -butyrolactone),

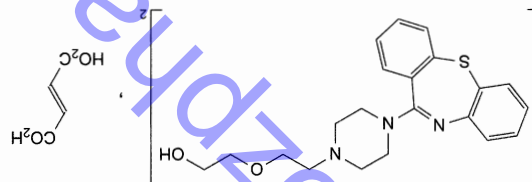


C, 1-methylpyrrolidin-2-one (N-methylpyrrolidone).

Ph Eur

Quetiapine Fumarate

(Ph. Eur. monograph 2541)



$C_{46}H_{54}N_6O_{12}$ 883 111974-72-2

Action and use
Dopamine receptor antagonist; neuroleptic

Ph Eur

DEFINITION

Bis[2-[2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]ethoxy]ethanol] (2E)-but-2-enedioate.

Content

— quetiapine fumarate ($C_{46}H_{54}N_6O_{12}$; M_r 883): 99.0 per cent to 101.0 per cent (dried substance);
— fumaric acid ($C_4H_4O_4$; M_r 116.1): 12.5 per cent to 13.8 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, in anhydrous ethanol and in methanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison quetiapine fumarate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R , evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R , water R (50:50 V/V).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of quetiapine for system suitability CRS (containing impurities G and N) in 1.0 mL of the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.10$ m, $\phi = 2.1$ mm;

— stationary phase: end-capped phenylsilyl silica gel for chromatography R (1.7 μ m);

— temperature: 50 °C.

Mobile phase:

— mobile phase A: mix 10 volumes of methanol R and 90 volumes of a 3.85 g/L solution of ammonium acetate R , previously adjusted to pH 9.0 with ammonia R ;

— mobile phase B: acetonitrile R ;

Time	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	80	20
8 - 14.50	80 \rightarrow 60	20 \rightarrow 40
14.50 - 22.60	60 \rightarrow 50	40 \rightarrow 50
22.60 - 26	50 \rightarrow 30	50 \rightarrow 70
26 - 29	30 \rightarrow 10	70 \rightarrow 90
29 - 30	10	90

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 3.0 μ L.

Identification of impurities Use the chromatogram supplied with quetiapine for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities G and N.

Relative retention

With reference to quetiapine (retention time = about 13 min): fumaric acid = about 0.05; impurity G = about 0.5; impurity N = about 1.04.

System suitability:

— signal-to-noise ratio: minimum 40 for the principal peak in the chromatogram obtained with reference solution (b);
— peak-to-valley ratio: minimum 5.0, where H_p = height above the baseline of the peak due to impurity N and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to quetiapine in the chromatogram obtained with reference solution (a).

Calculation of percentage contents:

— correction factors: multiply the peak areas of the following impurities by the corresponding correction factor:
impurity G = 0.5; impurity N = 2.0;
— for each impurity, use the concentration of quetiapine in reference solution (b).

Limits:

— impurities G, N: for each impurity, maximum 0.15 per cent;
— unspecified impurities: for each impurity, maximum 0.10 per cent;
— total: maximum 0.5 per cent;
— reporting threshold: 0.05 per cent; disregard any peak due to fumaric acid.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture methanol R , water R (50:50 V/V).

0.25 g complies with test H. Prepare the reference solution using 0.25 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Quetiapine fumarate

Dissolve 0.170 g in 40 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 22.08 mg of $C_{46}H_{54}N_{10}O_{10}S_2$.

Fumaric acid

Dissolve 0.350 g in 70 mL of a mixture of equal volumes of methanol R and water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 5.804 mg of $C_4H_4O_4$.

STORAGE

Protected from light.

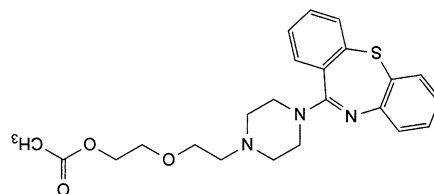
IMPURITIES

Specified impurities G, N

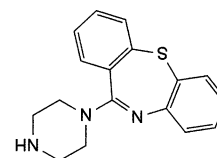
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

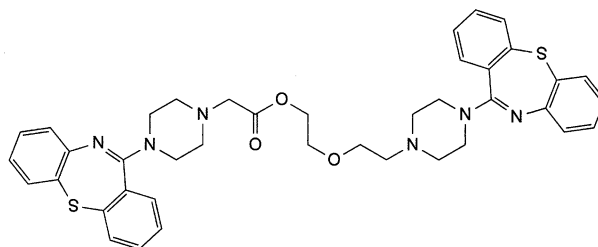
Control of impurities in substances for pharmaceutical use: A, B, C, D, E, F, H, I, J, K, L, O, P, Q, S, T, U, V, W.



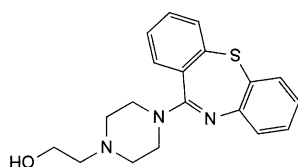
A. 2-[2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]ethoxy]ethyl acetate,



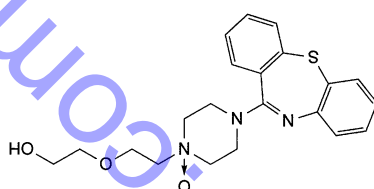
B. 1-(piperazin-1-yl)dibenzo[b,f][1,4]thiazepine,



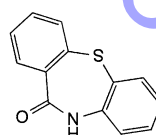
C. 2-[2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]ethoxy]ethyl 2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]piperazin-1-yl]ethoxy]ethyl acetate,



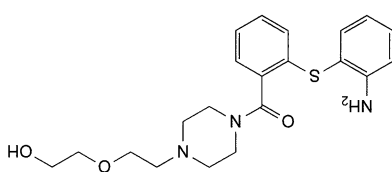
I. 2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]ethanol,



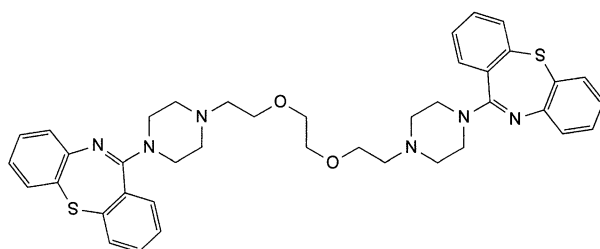
H. 2-[2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]ethoxy]ethyl 2-[4-(dibenzo[b,f][1,4]thiazepin-1-yl)piperazin-1-yl]ethoxy]ethyl acetate,



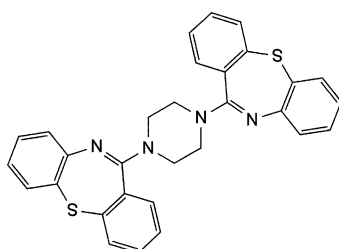
G. dibenzo[b,f][1,4]thiazepin-11(10H)-one,

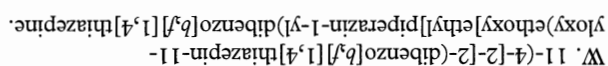


E. 1,1,1'-[ethylenebis(oxyethyl)enepiperazine-4,1'-diyl]bis(dibenzo[b,f][1,4]thiazepine),



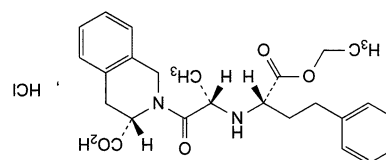
D. 1,1,1'-[piperazine-1,4-diyl]bis(dibenzo[b,f][1,4]thiazepine),





Quinapril Hydrochloride

(Ph. Eur. monograph 1763)


 $C_{25}H_{31}ClN_2O_5$ 475.0 82586-55-8

Action and use

Angiotensin converting enzyme inhibitor.

Ph. Eur.

DEFINITION

(3S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]-1,2,3,4-tetrahydroisquinoline-3-carboxylic acid hydrochloride.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white or slightly pink, hygroscopic powder.

Solubility

Freely soluble in water and in ethanol (96 per cent), very slightly soluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Comparison quinapril hydrochloride CRS.

B. Specific optical rotation (see Tests).

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

Specific optical rotation (2.2.7)

+ 14.4 to + 16.6 (anhydrous substance).

Dissolve 0.500 g in methanol R and dilute to 25.0 mL with the same solvent.

Diastereoisomers

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture Adjust 500 mL of the mobile phase to pH 6.5 with concentrated ammonia R.

Test solution Dissolve 100 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of quinapril for peak identification CRS (containing impurities G, H and I) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 25 °C.

Mobile phase Mix 260 mL of tetrahydrofuran R (non-stabilised) with 740 mL of a freshly prepared solution containing 1.08 g/L of sodium octanesulfonate R and 2.88 g/L

of ammonium dihydrogen phosphate R, previously adjusted to pH 4.5 with phosphoric acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Run time 3.5 times the retention time of quinapril.

Identification of impurities Use the chromatogram supplied with quinapril for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities G, H and I.

Relative retention With reference to quinapril (retention time = about 18 min): impurity G = about 0.9; impurity H = about 1.2; impurity I = about 1.3.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to impurity G and quinapril;

— peak-to-valley ratio: minimum 2.0, where H_p = height above the baseline of the peak due to impurity H and H_b = height above the baseline of the lowest point of the curve separating this peak from the peak due to quinapril.

Limits:

— impurities G, H, I: for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mix 40 volumes of acetonitrile R1 and 60 volumes of a 2.88 g/L solution of ammonium dihydrogen phosphate R previously adjusted to pH 6.5 with dilute ammonia R1.

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of quinapril for system suitability CRS (containing impurities A, C, D, E and G) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (c) In order to prepare impurity M *in situ*, dissolve 250 mg of the substance to be examined in methylene chloride R and dilute to 5.0 mL with the same solvent.

Expose this solution to a source of ultraviolet light for 2.5 h and evaporate the solvent. Dissolve 40 mg of the remaining substance in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Dissolve 100 mg of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m).

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m);

— column: 30 °C;

— autosampler: 5 °C.

Mobile phase acetonitrile R1, 5.77 g/L solution of sodium dodecyl sulfate R adjusted to pH 2.2 with phosphoric acid R (48:52 V/V).

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 10 μ L.

Run time 3 times the retention time of quinapril.

Identification of impurities Use the chromatogram supplied with quinapril for system suitability (b) to identify the peaks due to impurities A, C, D, E and G; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity M.

Relative retention With reference to quinapril (retention time = about 12 min): impurity A = about 0.1;

impurity C = about 0.3; impurity D = about 0.4;

impurity M = about 0.7; impurities G + H = about 0.9;

impurity E = about 2.3.

System suitability: reference solution (b):

— **resolution:** minimum 1.5 between the peaks due to impurities C and D; minimum 1.5 between the peaks due to impurity G and quinapril.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurity E by 1.5;

— **impurities C, D:** for each impurity, not more than 5 times the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.5 per cent);

— **impurity A:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— **impurities E, M:** for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);

— **unspecified impurities:** for each impurity, not more than area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **total:** not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to impurities G + H.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent dimethyl sulfoxide R.

1.0 g complies with test H. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R. If the substance precipitates after addition of buffer solution pH 3.5 R, dilute to 100 mL with dimethyl sulfoxide R; the substance re-dissolves completely. Treat the reference solution in the same way.

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 23.75 mg of $C_{25}H_{31}ClN_2O_5$.

STORAGE

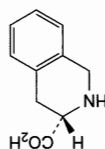
In an airtight container at a temperature of 2 °C to 8 °C.

IMPURITIES

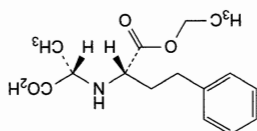
Specified impurities A, C, D, E, G, H, I, M

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of

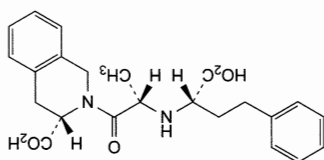
the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: B, F.



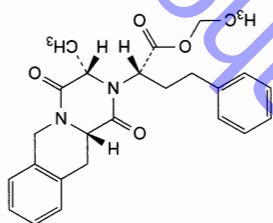
A. (3S)-1,2,3,4-tetrahydroisoquinoline-3-carboxylic acid,



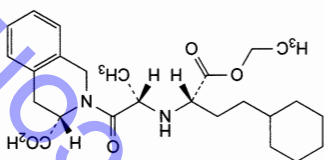
B. (2S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid,



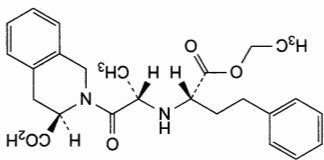
C. (3S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid,



D. ethyl (2S)-2-[(3S)-3-methyl-1,4-dioxo-1,3,4,6,11,11a-hexahydro-2H-pyrazino[1,2-b]isoquinolin-2-yl]-4-phenylbutanoate,



E. (3S)-2-[(2S)-2-[(1S)-3-cyclohexyl-1-(ethoxycarbonyl)propyl]amino]propanoic acid,



G. (3R)-2-[(2S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid,

CHARACTERISTICS

Colourless crystals; odourless or almost odourless. Freely soluble in *water* and in *ethanol* (96%); practically insoluble in *ether*.

IDENTIFICATION

A. Carry out the method for *thin-layer chromatography*, Appendix III A, using *silica gel G* as the coating substance and a mixture of 15 volumes of *diethylamine*, 36 volumes of *ether* and 60 volumes of *toluene* as the mobile phase. Apply separately to the plate 4 µL of each of three solutions in *methanol* containing (1) 1.0% w/v of the substance being examined, (2) 1.0% w/v of *quinidine sulfate BPGRS* and (3) 1.0% w/v each of *quinidine sulfate BPGRS* and *quinine sulfate BPGRS*. After removal of the plate, dry it in a current of air for 15 minutes and repeat the development. Dry the plate at 105° for 30 minutes, allow it to cool and spray with *iodoplatinate reagent*. The principal spot in the chromatogram obtained with solution (1) is similar in position, colour and size to that in the chromatogram obtained with solution (2). The test is not valid unless the chromatogram obtained with solution (3) shows two clearly separated spots.

B. Complies with the test for Acidity.

C. Yields the reactions characteristic of *sulfates*, Appendix VI.

TESTS**Acidity**

pH of a 1% w/v solution, 2.6 to 3.6, Appendix V L.

Specific optical rotation

In a 2% w/v solution in 0.1M *hydrochloric acid*, +246 to +258, Appendix V F, determined using a 2-dm layer and calculated with reference to the anhydrous substance.

Other cinchona alkaloids

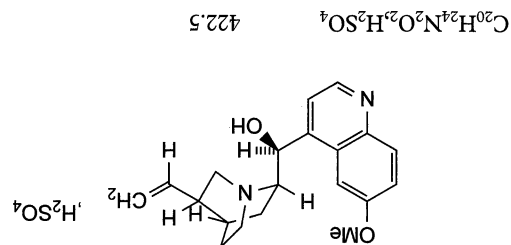
Carry out the method for *liquid chromatography*, Appendix III D, using the following solutions. For solution (1) dissolve 20 mg of the substance being examined, with gentle heating if necessary, in 5 mL of the mobile phase and dilute to 10 mL with the mobile phase. Prepare solutions (2) and (3) in the same manner using *quinine sulfate BPGRS* and *quinidine sulfate BPGRS* respectively in place of the substance being examined. Solution (4) is a mixture of equal volumes of solutions (2) and (3). For solution (5) dilute 1 volume of solution (2) to 10 volumes with the mobile phase and dilute (2) to 10 volumes with the mobile phase and dilute 1 volume of the resulting solution to 50 volumes with the mobile phase. Solution (6) contains 0.10% w/v of *thiourea* in the mobile phase.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with *octadecylsilyl silica gel for chromatography* (5 µm) (Hypersil ODS 5 µm is suitable), (b) as the mobile phase with a flow rate of 1.5 mL per minute a solution prepared by dissolving 6.8 g of *potassium dihydrogen orthophosphate* and 3.0 g of *hexylamine* in 700 mL of *water*, adjusting the pH to 2.8 with 1M *orthophosphoric acid*, adding 60 mL of *acetonitrile* and diluting to 1000 mL with *water* and (c) a detection wavelength of 250 nm for recording the chromatogram obtained with solution (6) and 316 nm for the other solutions.

Inject separately 10 µL of each of solutions (3) and (6). If necessary adjust the concentration of acetonitrile in the mobile phase so that in the chromatogram obtained with solution (3) the *capacity factor* of the peak due to quinidine is 3.5 to 4.5, *V₀* being calculated from the peak due to thiourea in the chromatogram obtained with solution (6). Inject 10 µL of each of solutions (2), (3), (4) and (5). The chromatogram obtained with solution (2) shows a substance.

Quinidine Bisulfate

Quinidine Bisulfate



50-54-4

422.5

C₂₀H₂₄N₂O₂·H₂SO₄**Action and use**

Class I antiarrhythmic.

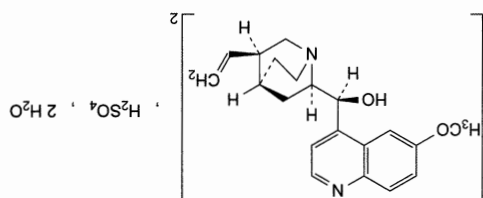
DEFINITION

Quinidine Bisulfate is (8*R*,9*S*)-6'-methoxycinchonan-9-ol hydrogen sulfate. It contains not less than 98.5% and not more than 101.5% of alkaloid hydrogen sulfates, calculated as C₂₀H₂₄N₂O₂·H₂SO₄ with reference to the anhydrous substance.



Quinidine Sulfate

Quinidine Sulphate
(Ph. Eur. monograph 0017)



$C_{40}H_{50}N_4O_8S_2 \cdot 2H_2O$ 783 6591-63-5

Action and use

Class I antiarrhythmic.

Preparation

Quinidine Sulfate Tablets

DEFINITION

99.0 per cent to 101.0 per cent of alkaloid monosulfates, expressed as bis[(S)-[(2R,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl](6-methoxyquinolin-4-yl)methanol] sulfate (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or silky, colourless needles.

Solubility

Slightly soluble in water, soluble in boiling water and in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 0.10 g of quinidine sulfate CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase diethylamine R, ether R, toluene R (10:24:40 V/V/V).

Application 5 µL.

Development Twice over a path of 15 cm, dry in a current of air for 15 min between the 2 developments.

Drying At 105 °C for 30 min and allow to cool.

Detection Spray with iodoplatinate reagent R.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Dissolve about 5 mg in 5 mL of water R. Add 0.2 mL of brownie water R and 1 mL of dilute ammonia R2. A green colour develops.

C. Dissolve 0.1 g in 3 mL of dilute sulfuric acid R and dilute to 100 mL with water R. When examined in ultraviolet light at 366 nm, an intense blue fluorescence appears which disappears almost completely on addition of 1 mL of hydrochloric acid R.

D. Dissolve about 50 mg in 5 mL of hot water R, cool, add 1 mL of silver nitrate solution R1 and stir with a glass rod.

principal peak due to quinidine and a peak due to dihydroquinine with a retention time relative to quinine of about 1.4. The chromatogram obtained with solution (3)

shows a principal peak due to quinidine and a peak due to dihydroquinidine, with a retention time relative to quinidine of about 1.2. The chromatogram obtained with solution (4) shows four peaks due to quinine, dihydroquinine, quinidine and dihydroquinidine which are identified by comparison of their retention times with those of the corresponding peaks in the chromatograms obtained with solutions (2) and (3).

The test is not valid unless (a) in the chromatogram obtained with solution (4) the resolution factor between the peaks due to quinine and quinidine is at least 1.5 and the resolution factor between the peaks due to dihydroquinidine and quinine is at least 1.0 and (b) the signal-to-noise ratio of the principal peak in the chromatogram obtained with solution (5) is at least 5.

Inject 10 µL of solution (1) and allow the chromatography to proceed for 2.5 times the retention time of the principal peak. Calculate the percentage content of related substances by normalisation, disregarding any peaks the areas of which are less than that of the peak in the chromatogram obtained with solution (5) (0.2%). The content of dihydroquinidine is not greater than 1.5%, the content of any related substance eluting before quinidine is not greater than 5% and the content of any other related substance is not greater than 2.5%.

Sulfated ash

Not more than 0.1%, Appendix IX A.

Water

Not more than 5.0% w/w, Appendix IX C. Use 1 g.

Titrateable cation

75.3 to 79.6%, calculated with reference to the anhydrous substance, when determined by the following method. To the combined aqueous solutions reserved in the Assay add 0.1 mL of phenolphthalein solution R1 and titrate with 0.1M hydrochloric acid VS. Each mL of 0.1M sodium hydroxide VS is equivalent to 16.32 mg of $[C_{20}H_{26}N_2O_2]^{2+}$.

ASSAY

Dissolve 0.45 g in 15 mL of water, add 25 mL of 0.1M sodium hydroxide VS and extract with three 25 mL quantities of chloroform. Wash each chloroform extract successively with the same 20 mL of water, combine the aqueous solution and reserve for the test for Titrateable cation. Dry the chloroform extracts with anhydrous sodium sulfate, evaporate to dryness at a pressure of 2 kPa and dissolve the residue in 50 mL of anhydrous acetic acid. Carry out Method I for non-aqueous titration, Appendix VIII A, using crystal violet solution as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 21.13 mg of $C_{20}H_{24}N_2O_2 \cdot H_2SO_4$.

STORAGE

Quinidine Bisulfate should be protected from light.

After a few minutes, a white precipitate is formed that dissolves on the addition of dilute nitric acid R.

F. It gives reaction (a) of sulfates (2.3.1).

F. pH (see Tests).

TESTS

Solution S

Dissolve 0.500 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution GY₆ (2.2.2, Method II).

pH (2.2.3)

Dissolve 0.10 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

+ 275 to + 290 (dried substance), determined on solution S.

Other cinchona alkaloids

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 20 mg of the substance to be examined in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.

Reference solution (a) Dissolve 20 mg of quinidine sulfate CRS (impurity A) in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.

Reference solution (b) Dissolve 20 mg of quinidine sulfate CRS in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.

Reference solution (c) To 1 mL of reference solution (a) add 1 mL of reference solution (b).

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (e) Dissolve 10 mg of thiourea R in the mobile phase and dilute to 10 mL with the mobile phase.

Column:

— size: $l = 0.15\text{--}0.25\text{ m}$, $\varnothing = 4.6\text{ mm}$;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5–10 μm).

Mobile phase Dissolve 6.8 g of potassium dihydrogen phosphate R and 3.0 g of hexylamine R in 700 mL of water R, adjust to pH 2.8 with dilute phosphoric acid R, add 60 mL of acetonitrile R and dilute to 1000 mL with water R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 250 nm for reference solution (e) and at 316 nm for the other solutions.

Injection 10 μL .

Run time 2.5 times the retention time of quinidine.

Identification of peaks Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurity A and dihydroquinidine; use the chromatogram obtained with reference solution (b) to identify the peaks due to quinidine and impurity C; the chromatogram obtained with reference solution (c) shows 4 peaks due to quinidine, impurity A, impurity C and dihydroquinidine which are identified by comparison of their retention times with those of the corresponding peaks in the chromatograms obtained with reference solutions (a) and (b).

STORAGE

Protected from light.

of $\text{C}_{40}\text{H}_{50}\text{N}_4\text{O}_8\text{S}$.

1 mL of 0.1 M perchloric acid is equivalent to 24.90 mg

solution R as indicator.

Dissolve 0.200 g in 20 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, using 0.15 mL of naphtholbenzenes

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

drying in an oven at 130 °C.

3.0 per cent to 5.0 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

solution.

The blank solution is yellow. Any red colour in the test

20 min add 25.0 mL of ethanol (96 per cent) R. Mix.

acetic acid R and 0.3 mL of sulfuric acid R. Mix and after

2.0 mL of a 3.75 g/L solution of curcumin R in anhydrous

Allow to stand for 6 min. To 1.0 mL of the lower layer, add

solution and to the blank solution, then shake for 1 min.

in methylene chloride R to the test solution, to the reference

Add 3.0 mL of a 100 g/L solution of 2-ethylhexane-1,3-diol R

water R.

Blank solution Add 0.5 mL of hydrochloric acid R to 4.0 mL of

0.5 mL of hydrochloric acid R.

To 1.0 mL of this solution add 3.0 mL of water R and

5.0 mL of the solution to 100.0 mL with water R.

Reference solution Dissolve 0.572 g of boric acid R in water R

and dilute to 1000.0 mL with the same solvent. Dilute

hydrochloric acid R and 4.0 mL of water R.

Test solution Dissolve 1.00 g in a mixture of 0.5 mL of

Maximum 5 ppm. Avoid where possible the use of glassware.

Boron

(0.2 per cent).

chromatogram obtained with reference solution (d)

— **disregard limit**: the area of the principal peak in the

2.5 per cent;

— **any other impurity**: for each impurity, maximum

maximum 5 per cent;

— **any impurity eluted before quinidine**: for each impurity,

— **impurity C**: maximum 15 per cent;

Limits:

acetonitrile in the mobile phase.

solution (e); if necessary, adjust the concentration of

thiourea in the chromatogram obtained with reference

solution (b), t_R being calculated from the peak due to

quinidine in the chromatogram obtained with reference

— **mass distribution ratio**: 3.5 to 4.5 for the peak due to

the chromatogram obtained with reference solution (d);

— **signal-to-noise ratio**: minimum 4 for the principal peak in

obtained with reference solution (c);

peaks due to impurities C and A in the chromatogram

impurity A and quinidine and minimum 2.0 between the

— **resolution**: minimum 3.0 between the peaks due to

System suitability:

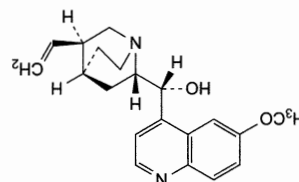
impurity C = about 1.5.

Relative retention With reference to quinidine:

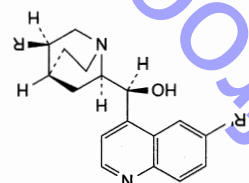
dihydroquinidine = about 1.4.

Relative retention With reference to impurity A:

IMPURITIES



A. (R)-[(2S,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl]-(6-methoxyquinolin-4-yl)methanol (quinine).



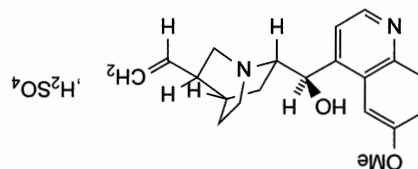
B. R = CH=CH₂, R' = H: (S)-[(2R,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl]-(quinolin-4-yl)methanol (cinchonine).

C. R = C₂H₅, R' = OCH₃: (S)-[(2R,4S,5R)-5-ethyl-1-azabicyclo[2.2.2]oct-2-yl]-(6-methoxyquinolin-4-yl)methanol (dihydroquinidine).

Ph Eur

Quinine Bisulfate

Quinine Bisulfate



$$\text{C}_{20}\text{H}_{24}\text{N}_2\text{O}_2 \cdot \text{H}_2\text{SO}_4 \cdot 7\text{H}_2\text{O} \quad 548.6$$

549-56-4

Preparation

Quinine Bisulfate Tablets

DEFINITION

Quinine Bisulfate is (8S,9R)-6'-methoxycinchonan-9-ol hydrogen sulfate heptahydrate. It contains not less than 98.5% and not more than 101.5% of alkaloid hydrogen sulfates, calculated as $\text{C}_{20}\text{H}_{24}\text{N}_2\text{O}_2 \cdot \text{H}_2\text{SO}_4$ with reference to the anhydrous substance.

CHARACTERISTICS

Colourless crystals or a white, crystalline powder; efflorescent in dry air.

Freely soluble in water; sparingly soluble in ethanol (96%).

IDENTIFICATION

A. Carry out the method for thin-layer chromatography, Appendix III A, using silica gel G as the coating substance and a mixture of 15 volumes of diethylamine, 36 volumes of ether and 60 volumes of toluene as the mobile phase. Apply separately to the plate 4 µL of each of three solutions in methanol containing (1) 1.0% w/v of the substance being

examined, (2) 1.0% w/v of quinine sulfate BPQRS and (3) 1.0% w/v each of quinine sulfate BPQRS and quinine sulfate BPQRS. After removal of the plate, dry it in a current of air for 15 minutes and repeat the development. Dry the plate at 105° for 30 minutes, allow it to cool and spray with iodoplatinate reagent. The principal spot in the chromatogram obtained with solution (1) is similar in position, colour and size to that in the chromatogram obtained with solution (2). The test is not valid unless the chromatogram obtained with solution (3) shows two clearly separated spots.

B. Compiles with the test for Acidity.

C. Yields the reactions characteristic of sulfates.

Appendix VI.

TESTS

Acidity

pH of a 1% w/v solution, 2.8 to 3.4, Appendix V L.

Specific optical rotation

In a 3% w/v solution in 0.1M hydrochloric acid, -208 to -216, calculated with reference to the anhydrous substance.

Appendix V F.

Other cinchona alkaloids

Carry out the method for liquid chromatography, Appendix III D, using the following solutions. For solution (1) dissolve 20 mg of the substance being examined, with gentle heating if necessary, in 5 mL of the mobile phase and dilute to 10 mL with the mobile phase. Prepare solutions (2) and (3) in the same manner using quinine sulfate BPQRS and quinine sulfate BPQRS respectively in place of the substance being examined. Solution (4) is a mixture of equal volumes of solutions (2) and (3). For solution (5) dilute 1 volume of solution (2) to 10 volumes with the mobile phase and dilute 1 volume of the resulting solution to 50 volumes with the mobile phase. Solution (6) contains 0.10% w/v of thiourea in the mobile phase.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Hypersil ODS 5 µm is suitable), (b) as the mobile phase with a flow rate of 1.5 mL per minute a solution prepared by dissolving 6.8 g of potassium dihydrogen orthophosphate and 3.0 g of hexylamine in 700 mL of water, adjusting the pH to 2.8 with 1M orthophosphoric acid, adding 60 mL of acetonitrile and diluting to 1000 mL with water and (c) a detection wavelength of 250 nm for recording the chromatogram obtained with solution (6) and 316 nm for the other solutions.

Inject separately 10 µL of each of solutions (3) and (6). If necessary adjust the concentration of acetonitrile in the mobile phase so that in the chromatogram obtained with solution (3) the capacity factor of the peak due to quinine is 3.5 to 4.5, 1% being calculated from the peak due to thiourea in the chromatogram obtained with solution (6). Inject 10 µL of each of solutions (2), (3), (4) and (5). The chromatogram obtained with solution (2) shows a principal peak due to quinine and a peak due to dihydroquinidine with a retention time relative to quinine of about 1.4. The chromatogram obtained with solution (3) shows a principal peak due to quinine and a peak due to dihydroquinidine, with a retention time relative to quinine of about 1.2. The chromatogram obtained with solution (4) shows four peaks due to quinine, dihydroquinidine, quinidine and dihydroquinidine which are identified by comparison of their retention times with those of the corresponding peaks in the chromatograms obtained with solutions (2) and (3).

more than 101.0% of alkaloid dihydrochlorides, calculated as $C_{20}H_{24}N_2O_2 \cdot 2HCl$, with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder.

Very soluble in *water*; soluble in *ethanol* (96%).

IDENTIFICATION

A. Carry out the method for *thin-layer chromatography*,

Appendix III A, using the following solutions in *methanol*.

(1) 1.0% w/v of the substance being examined.

(2) 1.0% w/v of *quinine sulfate BPGRS*.

(3) 1.0% w/v each of *quinidine sulfate BPGRS* and *quinine sulfate BPGRS*.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating *silica gel G*.

(b) Use the mobile phase as described below.

(c) Apply 4 μ L of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry it in a current of air for

15 minutes and repeat the development. Dry the plate at

105° for 30 minutes, allow to cool and spray with

iodoplatinate reagent.

MOBILE PHASE

15 volumes of *diethylamine*, 36 volumes of *ether* and

60 volumes of *toluene*.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with

solution (3) shows two clearly separated spots.

CONFIRMATION

The principal spot in the chromatogram obtained with

solution (1) corresponds in position, colour and size to that

in the chromatogram obtained with solution (2).

B. Complies with the test for Acidity.

C. Yields reaction A characteristic of *chlorides*, Appendix VI.

TESTS

Acidity

pH of a 3% w/v solution, 2.0 to 3.0, Appendix V L.

Specific optical rotation

In a 3% w/v solution in 0.1M *hydrochloric acid*, -223 to -229

calculated with reference to the dried substance,

Appendix V F.

Barium

To 15 mL of a 2.0% w/v solution add 1 mL of 1M *sulfuric*

acid. The solution remains clear for at least 15 minutes.

Sulfate

0.125 g complies with the limit test for *sulfates*, Appendix VII

(0.12%).

Other cinchona alkaloids

Carry out the method for *liquid chromatography*, and the

following solutions in the mobile phase.

(1) Dissolve 20 mg of the substance being examined, with

gentle heating if necessary, in 5 mL and dilute to 10 mL.

(2) Prepare as for solution (1) but using *quinine*

sulfate BPGRS in place of the substance being examined.

(3) Prepare as for solution (1) but using *quinidine*

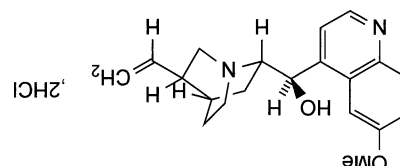
sulfate BPGRS in place of the substance being examined.

(4) Mix equal volumes of solutions (2) and (3).

(5) Dilute 1 volume of solution (2) to 10 volumes and dilute

1 volume of the resulting solution to 50 volumes.

Quinine Dihydrochloride



$C_{20}H_{24}N_2O_2 \cdot 2HCl$

397.3

60-93-5

DEFINITION

Quinine Dihydrochloride Infusion

Preparation

Antiprototozoal (malaria).

Action and use

Quinine Dihydrochloride is (8S,9R)-6'-methoxycinchonan-9-ol dihydrochloride. It contains not less than 99.0% and not

Quinine Bisulfate should be protected from light.

STORAGE

to 21.13 mg of $C_{20}H_{24}N_2O_2 \cdot H_2SO_4$.

as indicator. Each mL of 0.1M *perchloric acid VS* is equivalent

aqueous titration, Appendix VIII A, using crystal violet solution

50 mL of *anhydrous acetic acid*. Carry out method I for non-

dryness at a pressure of 2 kPa and dissolve the residue in

chloroform extracts with *anhydrous sodium sulfate*, evaporate

and reserve for the test for Titratable cation. Dry the

extracts with 20 mL of *water*, combine the aqueous solutions

quantities of *chloroform*. Wash the combined chloroform

0.1M *sodium hydroxide VS* and extract with three 25 mL

Dissolve 0.45 g in 15 mL of *water*. Add 25 mL of

VS is equivalent to 16.32 mg of $[C_{20}H_{26}N_2O_2]^{2+}$.

0.1M *hydrochloric acid VS*. Each mL of 0.1M *sodium hydroxide*

0.1 mL of *phenolphthalein solution RI* and titrate with

Add to the combined aqueous solutions reserved in the Assay

substance, when determined by the following method.

75.3 to 79.6%, calculated with reference to the anhydrous

Titratable cation

19.0 to 25.0% w/v, Appendix IX C. Use 0.2 g.

Water

Not more than 0.1%, Appendix IX A.

Sulfated ash

of any other related substances is not greater than 2.5%.

eluting before quinine is not greater than 5% and the content

not greater than 10%, the content of any related substances

with solution (5) (0.2%). The content of dihydroquinine is

are less than that of the peak in the chromatogram obtained

by *normalisation*, disregarding any peaks the areas of which

peak. Calculate the percentage content of related substances

proceed for 2.5 times the retention time of the principal

Inject 10 μ L of solution (1) and allow the chromatography to

least 5.

peak in the chromatogram obtained with solution (5) is at

is at least 1.0 and (b) the *signal-to-noise ratio* of the principal

factor between the peaks due to dihydroquinidine and quinine

to quinine and quinine is at least 1.5 and the *resolution*

with solution (4) the *resolution factor* between the peaks due

The test is not valid unless (a) in the chromatogram obtained

(6) 0.10% w/v of *thourea*.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Hypersil ODS 5 µm is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1.5 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 250 nm for recording the chromatogram obtained with solution (6) and a detection

wavelength of 316 nm for the other solutions.

(f) Inject 10 µL of each solution. Inject separately 10 µL of each of solutions (3) and (6).

(g) For solution (1) allow the chromatography to proceed for 2.5 times the retention time of the principal peak.

MOBILE PHASE

Dissolve 6.8 g of potassium dihydrogen orthophosphate and

3.0 g of hexylamine in 700 mL of water, adjust to pH 2.8

with 1M orthophosphoric acid, add 60 mL of acetonitrile and

dilute to 1000 mL with water. If necessary adjust the

concentration of acetonitrile in the mobile phase so that in

the chromatogram obtained with solution (3) the capacity

factor of the peak due to quinine is 3.5 to 4.5, V_0 being

calculated from the peak due to thiourea in the

chromatogram obtained with solution (6).

SYSTEM SUITABILITY

The test is not valid unless (a) in the chromatogram obtained

with solution (4) the resolution factor between the peaks due

to quinine and quinine is at least 1.5 and the resolution

factor between the peaks due to dihydroquinidine and quinine

is at least 1.0 and (b) the signal-to-noise ratio of the principal

peak in the chromatogram obtained with solution (5) is at

least 5.

The chromatogram obtained with solution (2) shows a

principal peak due to quinine and a peak due to

dihydroquinidine with a retention time relative to quinine of

about 1.4. The chromatogram obtained with solution (3)

shows a principal peak due to quinine and a peak due to

dihydroquinidine, with a retention time relative to quinine

of about 1.2. The chromatogram obtained with solution (4)

shows four peaks due to quinine, dihydroquinidine, quinine

and dihydroquinidine which are identified by comparison of

their retention times with those of the corresponding peaks in

the chromatograms obtained with solutions (2) and (3).

LIMITS

In the chromatogram obtained with solution (1):

the content of dihydroquinidine is not greater than 10%;

the content of any related substance eluting before quinine is

not more than 5%;

the content of any other related substance is not more than

2.5%.

Disregard any peak with an area less than that of the peak in

the chromatogram obtained with solution (5).

Loss on drying

When dried to constant weight at 105°, loses not more than 3.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

Titration

79.7 to 84.2%, calculated with reference to the dried substance, when determined by the following method. Dissolve 0.4 g in 10 mL of water, add 40 mL of methanol and titrate with 0.1M sodium hydroxide VS using phenolphthalein solution R1 as indicator. Each mL of 0.1M sodium hydroxide VS is equivalent to 16.32 mg of $[C_{20}H_{26}N_2O_2]^{2+}$.

ASSAY

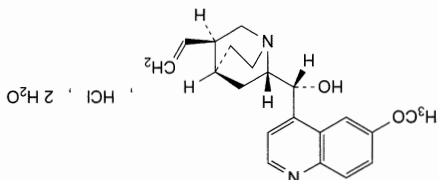
Dissolve 0.3 g in a mixture of 50 mL of anhydrous acetic acid and 20 mL of acetic anhydride, add 10 mL of mercury(II) acetate solution and carry out method I for non-aqueous titration, Appendix VIII A, using crystal violet solution as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 19.87 mg of $C_{20}H_{24}N_2O_2 \cdot 2HCl$.

STORAGE

Quinine Dihydrochloride should be protected from light.

Quinine Hydrochloride

(Ph. Eur. monograph 0018)



$C_{20}H_{25}ClN_2O_2 \cdot 2H_2O$ 396.9 6119-47-7

Action and use

Antiprotozoal (malaria).

DEFINITION

99.0 per cent to 101.0 per cent of alkaloid

monohydrochlorides, expressed as (R)-[(2S,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl](6-methoxyquinolin-4-yl)methanol hydrochloride (dried substance).

CHARACTERS

Appearance

White or almost white or colourless, fine, silky needles, often in clusters.

Solubility

Soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent. Reference solution Dissolve 0.10 g of quinine sulfate CRS in methanol R and dilute to 10 mL with the same solvent. Plate TLC silica gel G plate R.

Mobile phase diethylamine R, ether R, toluene R (10:24:40 V/V/V).

Application 5 µL.

Development Twice over a path of 15 cm, dry in a current of air for 15 min between the 2 developments.

Drying At 105° C for 30 min and allow to cool.

Detection Spray with iodoplatinate reagent R.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Dissolve about 10 mg in water R and dilute to 10 mL with the same solvent. To 5 mL of this solution add 0.2 mL of bromine water R and 1 mL of dilute ammonia R2. A green colour develops.

C. Dissolve 0.1 g in 3 mL of dilute sulfuric acid R and dilute to 100 mL with water R. When examined in ultraviolet light at 366 nm, an intense blue fluorescence appears which disappears almost completely on the addition of 1 mL of hydrochloric acid R.

D. It gives the reactions of chlorides (2.3.1).
E. pH (see Tests).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

pH (2.2.3)

Dilute 10 mL of solution S to 20 mL with carbon dioxide-free water R.
6.0 to 6.8.

Specific optical rotation (2.2.7)

—245 to –258 (dried substance).
Dissolve 0.500 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Other cinchona alkaloids

Test solution Dissolve 20 mg of the substance to be examined in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.
Reference solution (a) Dissolve 20 mg of quinine sulfate CRS in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.
Reference solution (b) Dissolve 20 mg of quinine sulfate CRS (impurity A) in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.
Reference solution (c) To 1 mL of reference solution (a) add 1 mL of reference solution (b).
Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.
Reference solution (e) Dissolve 10 mg of thiourea R in the mobile phase and dilute to 10 mL with the mobile phase.

Column:

— size: $l = 0.15\text{--}0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase = octadecylsilyl silica gel for chromatography R (5–10 μm).

Mobile phase Dissolve 6.8 g of potassium dihydrogen phosphate R and 3.0 g of hexylamine R in 700 mL of water R, adjust to pH 2.8 with dilute phosphoric acid R, add 60 mL of acetonitrile R and dilute to 1000 mL with water R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 250 nm for reference solution (e) and at 316 nm for the other solutions.

Injection 10 μL .

Run time 2.5 times the retention time of quinine.

Identification of peaks Use the chromatogram obtained with reference solution (a) to identify the peaks due to quinine and impurity C; use the chromatogram obtained with reference solution (b) to identify the peaks due to impurity A and dihydroquinidine; the chromatogram obtained with reference solution (c) shows 4 peaks due to impurity A, quinine, dihydroquinidine and impurity C, which are identified by comparison of their retention times with those of the corresponding peaks in the chromatograms obtained with reference solutions (a) and (b).
Relative retention With reference to quinine: impurity C = about 1.4.
Relative retention With reference to impurity A: dihydroquinidine = about 1.5.
System suitability:

— resolution: minimum 3.0 between the peaks due to quinine and impurity A and minimum 2.0 between the peaks due to dihydroquinidine and quinine in the chromatogram obtained with reference solution (c);
— signal-to-noise ratio: minimum 4 for the principal peak in the chromatogram obtained with reference solution (d);
— mass distribution ratio: 3.5 to 4.5 for the peak due to impurity A in the chromatogram obtained with reference solution (b), t_R being calculated from the peak due to thiourea in the chromatogram obtained with reference solution (e); if necessary, adjust the concentration of acetonitrile in the mobile phase.
Limits:

— impurity C: maximum 10 per cent;
— any impurity eluted before quinine: for each impurity, maximum 5 per cent;
— any other impurity: for each impurity, maximum 2.5 per cent;

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (d) (0.2 per cent).

Sulfates (2.4.13)

Maximum 500 ppm, determined on solution S.

Barium

To 15 mL of solution S add 1 mL of dilute sulfuric acid R. Allow to stand for 15 min. Any opalescence in the solution is not more intense than that in a mixture of 15 mL of solution S and 1 mL of distilled water R.

Loss on drying (2.2.32)

6.0 per cent to 10.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

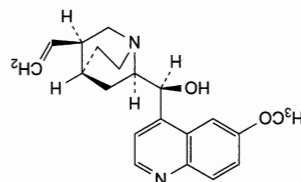
Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R and add 5.0 mL of 0.01 M hydrochloric acid. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). Read the volume added between the 2 inflexion points.

1 mL of 0.1 M sodium hydroxide is equivalent to 36.09 mg of $\text{C}_{20}\text{H}_{25}\text{ClN}_2\text{O}_2$

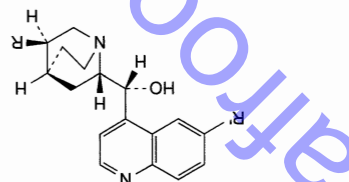
STORAGE

Protected from light.

IMPURITIES



A. (S)-[(2R,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl]-(6-methoxyquinolin-4-yl)methanol (quinidine),

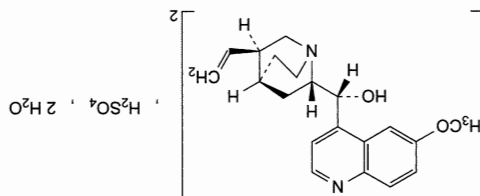


B. R = CH=CH₂, R' = H: (R)-[(2S,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl]-(quinolin-4-yl)methanol (cinchonidine),
C. R = C₂H₅, R' = OCH₃: (R)-[(2S,4S,5R)-5-ethyl-1-azabicyclo[2.2.2]oct-2-yl]-(6-methoxyquinolin-4-yl)methanol (dihydroquinine).

Quinine Sulfate

Quinine Sulphate

(Ph. Eur. monograph 0019)

C₄₀H₅₀N₄O₈S₂H₂O 783 6119-70-6

Action and use

Antiprotozoal (malaria).

Preparation

Quinine Sulfate Tablets

DEFINITION

Content

99.0 per cent to 101.0 per cent of alkaloid monosulfates, expressed as bis[(R)-[(2S,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl]-(6-methoxyquinolin-4-yl)methanol] sulfate (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or fine, colourless

needles.

Solubility

Slightly soluble in water, sparingly soluble in boiling water and in ethanol (96 per cent).

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent. Reference solution Dissolve 0.10 g of quinine sulfate CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase diethylamine R, ether R, toluene R (10:24:40 V/V/V).

Application 5 µL.

Development Twice over a path of 15 cm; dry in a current of air for 15 min between the 2 developments.

Drying At 105 °C for 30 min and allow to cool.

Detection Spray with iodoplatinate reagent R.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Dissolve about 5 mg in 5 mL of water R. Add 0.2 mL of bromine water R and 1 mL of dilute ammonia R2. A green colour develops.

C. Dissolve 0.1 g in 3 mL of dilute sulfuric acid R and dilute to 100 mL with water R. When examined in ultraviolet light at 366 nm, an intense blue fluorescence appears which disappears almost completely on the addition of 1 mL of hydrochloric acid R.

D. Dissolve about 45 mg in 5 mL of dilute hydrochloric acid R. The solution gives reaction (a) of sulfates (2.3.1).

E. pH (see Tests).

TESTS

Solution S

Dissolve 0.500 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution GY₆ (2.2.2, Method II).

pH (2.2.3)

5.7 to 6.6 for a 10 g/L suspension in water R.

Specific optical rotation (2.2.7)

-237 to -245 (dried substance); determined on solution S.

Other cinchona alkaloids

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 20 mg of the substance to be examined in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.

Reference solution (a) Dissolve 20 mg of quinine sulfate CRS in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.

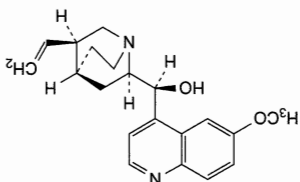
Reference solution (b) Dissolve 20 mg of quinine sulfate CRS (impurity A) in 5 mL of the mobile phase, with gentle heating if necessary, and dilute to 10 mL with the mobile phase.

Reference solution (c) To 1 mL of reference solution (a) add 1 mL of reference solution (b).

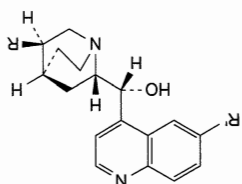
Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (e) Dissolve 10 mg of thionaea R in the mobile phase and dilute to 10 mL with the mobile phase.

IMPURITIES



A. (S)-[(2R,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl][(6-methoxyquinolin-4-yl)methanol (quinidine),

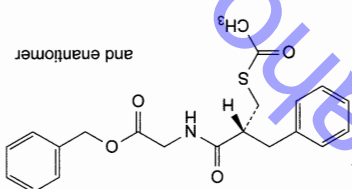


B. R = CH=CH₂, R' = H: (R)-[(2S,4S,5R)-5-ethenyl-1-azabicyclo[2.2.2]oct-2-yl][(6-methoxyquinolin-4-yl)methanol (cinchonidine),
C. R = C₂H₅, R' = OCH₃: (R)-[(2S,4S,5R)-5-ethyl-1-azabicyclo[2.2.2]oct-2-yl][(6-methoxyquinolin-4-yl)methanol (dihydroquinidine).

Ph Eur

Racacodotril

(Ph. Eur. monograph 2171)

C₂₁H₂₃NO₄S 385.5

Ph Eur

DEFINITION

Benzyl [(2R,5)-2-[(acetylsulfonyl)methyl]-3-phenylpropyl]amino]acetate.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methanol and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison racacodotril CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Column:

— size: l = 0.15-0.25 m, Ø = 4.6 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5-10 µm).

Mobile phase Dissolve 6.8 g of potassium dihydrogen

phosphate R and 3.0 g of hexylamine R in 700 mL of water R,

adjust to pH 2.8 with dilute phosphoric acid R, add 60 mL of

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 250 nm for reference solution (e) and at 316 nm for the other solutions.

Injection 10 µL.

Run time 2.5 times the retention time of quinine.

Identification of peaks Use the chromatogram obtained with

reference solution (a) to identify the peaks due to quinine

and impurity C; use the chromatogram obtained with

reference solution (b) to identify the peaks due to impurity A

and dihydroquinidine; the chromatogram obtained with

reference solution (c) shows 4 peaks due to impurity A,

quinine, dihydroquinidine and impurity C which are

identified by comparison of their retention times with those

of the corresponding peaks in the chromatograms obtained

with reference solutions (a) and (b).

Relative retention With reference to quinine:

impurity C = about 1.4.

Relative retention With reference to impurity A:

dihydroquinidine = about 1.5.

System suitability:

— resolution: minimum 3.0 between the peaks due to quinine

and impurity A and minimum 2.0 between the peaks due

to dihydroquinidine and quinine in the chromatogram

obtained with reference solution (c);

— signal-to-noise ratio: minimum 4 for the principal peak in

the chromatogram obtained with reference solution (d);

— mass distribution ratio: 3.5 to 4.5 for the peak due to

impurity A in the chromatogram obtained with reference

solution (b), *t_R* being calculated from the peak due to

thiourea in the chromatogram obtained with reference

solution (e); if necessary, adjust the concentration of

Limit:

— impurity C: maximum 10 per cent;

— any impurity eluted before quinine: for each impurity,

maximum 5 per cent;

— any other impurity: for each impurity, maximum

2.5 per cent;

— disregard limit: the area of the principal peak in the

chromatogram obtained with reference solution (d)

(0.2 per cent).

Loss on drying (2.2.32)

3.0 per cent to 5.0 per cent, determined on 1.000 g by

drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 10 mL of chloroform R and 20 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 24.90 mg of C₄₀H₅₀N₄O₈S.

STORAGE

Protected from light.

Dissolve 5.0 g in 10 mL of acetone R.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase A, mobile phase B (50:50 V/V).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Test solution (b) Dilute 5.0 mL of test solution (a) to 25.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) Prepare immediately before use. Dilute 500 µL of racecadotril impurity A CRS in acetonitrile R and dilute to 250.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture. Dilute

1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of racecadotril impurity G CRS in the solvent mixture and dilute to 50 mL with the solvent mixture. To 5 mL of this solution add 1 mL of test solution (b) and dilute to 100 mL with the solvent mixture.

Reference solution (d) Dissolve 50.0 mg of racecadotril CRS in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 25.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 25.0 mL with the solvent mixture.

Reference solution (e) Dissolve 2 mg of racecadotril for peak identification CRS (containing impurities C, E and F) in 1.0 mL of the solvent mixture.

Column: size: $l = 0.25$ m, $\phi = 4.0$ mm; stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm); temperature: 30 °C.

Mobile phase: mobile phase A: dissolve 1.0 g of potassium dihydrogen phosphate R in water R, adjust to pH 2.5 with phosphoric acid R and dilute to 1000 mL with water R; mobile phase B: acetonitrile R₁;

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of the solvent mixture, test solution (a) and reference solutions (a), (b), (c) and (e).

Identification of impurities Use the chromatogram supplied with racecadotril for peak identification CRS and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities C, E and F.

Relative retention With reference to racecadotril (retention time = about 16 min): impurity A = about 0.2; impurity C = about 0.3; impurity E = about 0.5; impurity F = about 0.9.

System suitability: reference solution (c):

— **resolution:** minimum 1.5 between the peaks due to impurity G and racecadotril.

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— **resolution:** minimum 1.5 between the peaks due to impurity G and racecadotril.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 1.4;

impurity E = 0.6; impurity F = 0.7;

impurities C, E, F: for each impurity, not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference

solution (b) (0.1 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in vacuo at 60 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (d).

Calculate the percentage content of C₂₁H₂₃NO₄S from the declared content of racecadotril CRS.

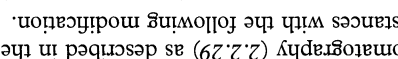
IMPURITIES

Specified impurities A, C, E, F

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, D, G, H.



A. ethanethioic acid (thioacetic acid),



B. [(2R,S)-2-benzyl-3-sulfanylpentanoyl]amino]acetic acid, and enantiomer

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 µL of the solvent mixture, test solution (a) and reference solutions (a), (b), (c) and (e).

Identification of impurities Use the chromatogram supplied with racecadotril for peak identification CRS and the chromatogram obtained with reference solution (e) to identify the peaks due to impurities C, E and F.

Relative retention With reference to racecadotril (retention time = about 16 min): impurity A = about 0.2; impurity C = about 0.3; impurity E = about 0.5; impurity F = about 0.9.

System suitability: reference solution (c):

— resolution: minimum 1.5 between the peaks due to impurity G and racecadotril.

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— system suitability: reference solution (c):

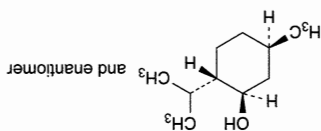
— resolution: minimum 1.5 between the peaks due to impurity G and racecadotril.

— system suitability: reference solution (c):



Racementhol

(*Racemic Menthhol*, Ph Eur monograph 0623)



$C_{10}H_{20}O$ 156.3

89-78-1

Preparation

Menthol and Benzoin Inhalation

Ph Eur

DEFINITION

Mixture of equal parts of (1*R*,2*S*,5*R*)-5-methyl-2-(1-methylethyl)cyclohexanol.

CHARACTERS

Appearance

Free-flowing or agglomerated, crystalline powder or prismatic or acicular, colourless, shiny crystals.

Solubility

Practically insoluble in water, very soluble in ethanol (96 per cent) and in light petroleum, freely soluble in fatty oils and in liquid paraffin, very slightly soluble in glycerol.

mp: about 34 °C.

IDENTIFICATION

First identification A, C.

Second identification B, D.

A. Optical rotation (see Tests).

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution Dissolve 25 mg of menthol CRS in methanol R and dilute to 5 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase ethyl acetate R, toluene R (5:95 V/V).

Application 2 µL.

Development Over a path of 15 cm.

Drying In air, until the solvents have evaporated.

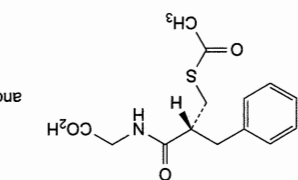
Detection Spray with anisaldehyde solution R and heat at 100-105 °C for 5-10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

C. Examine the chromatograms obtained in the test for related substances.

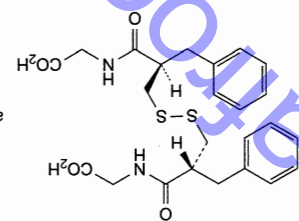
Results The principal peak in the chromatogram obtained with test solution (b) is similar in position and approximate dimensions to the principal peak in the chromatogram obtained with reference solution (c).

D. Dissolve 0.20 g in 0.5 mL of anhydrous pyridine R. Add 3 mL of a 150 g/L solution of dimethylbenzoyl chloride R in anhydrous pyridine R. Heat on a water-bath for 10 min. Add 7.0 mL of water R in small quantities with stirring and allow to stand in iced water for 30 min. A precipitate is formed. Allow to stand and decant the supernatant. Wash the precipitate with 2 quantities, each of 5 mL, of iced water R, recrystallise from 10 mL of acetone R, wash with iced acetone R and dry at 75 °C at a pressure not exceeding 2.7 kPa for 30 min. The crystals melt (2.2.14) at 130 °C to 131 °C.



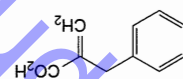
and enantiomer

C. [(2*R*,3*S*)-2-[(acetyl)sulfanylmethyl]-3-phenylpropionyl]amino]acetic acid,

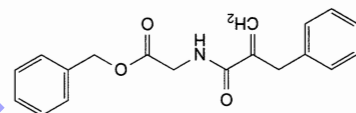


and stereoisomers

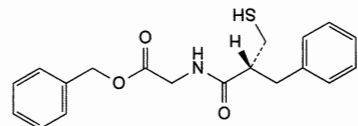
D. 5,10-dibenzyl-4,11-dioxo-7,8-dithia-3,12-diazatetradecanedioic acid,



E. 2-benzylprop-2-enoic acid (2-benzylacrylic acid),

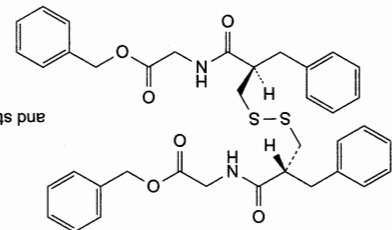


F. benzyl [(2-benzylprop-2-enoyl)amino]acetate,



and enantiomer

G. benzyl [(2*R*,3*S*)-2-benzyl-3-sulfanylpropionyl]amino]acetate,

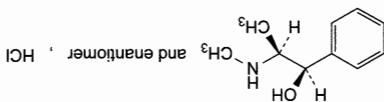


and stereoisomers

H. dibenzyl 5,10-dibenzyl-4,11-dioxo-7,8-dithia-3,12-diazatetradecanedioate.

Ph Eur

Racedephedrine Hydrochloride

(Racemic Ephedrine Hydrochloride,
Ph Eur monograph 0715)C₁₀H₁₆ClNO 201.7 134-71-4

Action and use

Adrenoceptor agonist.

DEFINITION

Racemic ephedrine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (1*R*,2*SR*)-2-(methylamino)-1-phenylpropan-1-ol hydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, crystalline powder or colourless crystals, freely soluble in water, soluble in ethanol (96 per cent).
It melts at about 188 °C.

IDENTIFICATION

First identification B, E.
Second identification A, C, D, E.

A. Optical rotation (see Tests).

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with racemic ephedrine hydrochloride CRS. Examine the substances prepared as discs.
C. Examine the chromatograms obtained in the test for related substances. The principal spot in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 0.1 mL of solution S (see Tests) add 1 mL of water R, 0.2 mL of copper sulfate solution R and 1 mL of strong sodium hydroxide solution R. A violet colour is produced. Add 2 mL of ether R and shake. The ether layer is purple and the aqueous layer is blue.
E. To 5 mL of solution S add 5 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 5.00 g in distilled water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of methyl red solution R and 0.1 mL of 0.01 M sodium hydroxide; the solution is yellow. Add 0.2 mL of 0.01 M hydrochloric acid; the solution is red.

Optical rotation (2.2.7)

+ 0.2° to – 0.2°, determined on solution S.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel G R as the coating substance.

TESTS

Solution S

Dissolve 2.50 g in 10 mL of ethanol (96 per cent) R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

Dissolve 1.0 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent. Add 0.1 mL of phenolphthalein solution R. The solution is colourless. Not more than 0.5 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to pink.

Optical rotation (2.2.7)

– 0.2° to + 0.2°, determined on solution S.

Related substances

Gas chromatography (2.2.28).

Test solution (a) Dissolve 0.20 g of the substance to be examined in methylene chloride R and dilute to 50.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with methylene chloride R.

Reference solution (a) Dissolve 40.0 mg of the substance to be examined and 40.0 mg of isomenthol R in methylene chloride R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dilute 0.10 mL of test solution (a) to 100.0 mL with methylene chloride R.

Reference solution (c) Dissolve 40.0 mg of menthol CRS in methylene chloride R and dilute to 100.0 mL with the same solvent.

Column:

material: glass;

size: 1 = 2.0 m, Ø = 2 mm;

stationary phase: diatomaceous earth for gas

chromatography R impregnated with 15 per cent m/m of macrogol 1500 R.

Carrier gas nitrogen for chromatography R.

Flow rate 30 mL/min.

Temperature:

column: 120 °C;

injection port: 150 °C;

detector: 200 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time Twice the retention time of menthol.

System suitability:

resolution: minimum 1.4 between the peaks due to menthol and isomenthol in the chromatogram obtained

with reference solution (a);

signal-to-noise ratio: minimum 5 for the principal peak in the chromatogram obtained with reference solution (b).

Limits: test solution (a):

total: not more than 1 per cent of the area of the principal

peak;

disregard limit: 0.05 per cent of the area of the principal

peak.

Residue on evaporation

Maximum 0.05 per cent.

Evaporate 2.00 g on a water-bath and heat in an oven at 100–105 °C for 1 h. The residue weighs not more than

1.0 mg.

Ph Eur

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS**Appearance**

Almost white or pale-yellow powder.

Solubility

Very slightly soluble or practically insoluble in water and in acetone, slightly soluble in ethanol (96 per cent V/V).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison raloxifene hydrochloride CRS.

B. Dissolve 20 mg of the substance to be examined in 2 mL of methanol R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS**Related substances**

Liquid chromatography (2.2.29).

Solvent mixture acetone/nitrite R, mobile phase A (30:70 V/V).

Test solution Dissolve 30 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) In order to produce impurity C *in situ*,

to 6 mg of the substance to be examined add 15 mL of

acetone/nitrite R, 3 mL of water R and 5 mL of stabilised strong

hydrogen peroxide solution R. Store at 30 °C for at least 6 h

then dilute to 50.0 mL with mobile phase A. To 1.0 mL of

this solution add 3 mg of the substance to be examined

dissolved in the solvent mixture and dilute to 10.0 mL with

the solvent mixture.

Reference solution (c) Dissolve 3 mg of raloxifene for peak

identification CRS (containing impurity A) in the solvent

mixture and dilute to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated octylsilyl silica gel for

chromatography R (5 μ m);

— temperature: 35 °C.

Mobile phase:

— mobile phase A: 9.0 g/L solution of potassium dihydrogen

phosphate R adjusted to pH 3.0 with phosphoric acid R;

— mobile phase B: acetone/nitrite R;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 9	75	25
9 - 40	75 \rightarrow 50	25 \rightarrow 50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 10 μ L.

Identification of impurity A Use the chromatogram supplied

chromatogram obtained with reference solution (c) to identify

the peak due to impurity A.

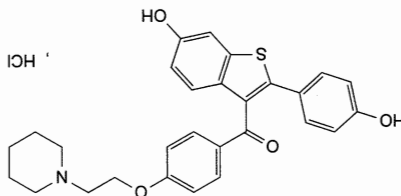
Relative retention With reference to raloxifene (retention

time = about 18 min): impurity A = about 0.7;

impurity C = about 1.2.

Raloxifene Hydrochloride

(Ph. Eur. monograph 2375)



Ph Eur

STORAGE

Store protected from light.

C₁₀H₁₆ClNO₄S

1 mL of 0.1 M sodium hydroxide corresponds to 20.17 mg of

inflixion.

hydroxide. Read the volume added between the two points of

potentiometric titration (2.2.20), using 0.1 M sodium

Add 5.0 mL of 0.01 M hydrochloric acid. Carry out a

Dissolve 0.170 g in 30 mL of ethanol (96 per cent) R.

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY**Sulfated ash (2.4.14)**

drying in an oven at 105 °C.

Not more than 0.5 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

(100 ppm).

15 mL of solution S complies with the limit test for sulfates

Sulfates (2.4.13)

lighter colour than the background.

reference solution (b) (0.5 per cent). Disregard any spot of

intense than the spot in the chromatogram obtained with

solution (a), apart from the principal spot, is not more

Any spot in the chromatogram obtained with test

Spray with ninhydrin solution R and heat at 110 °C for 5 min.

80 volumes of 2-propanol R. Allow the plate to dry in air.

chloroform R, 15 volumes of concentrated ammonia R and

over a path of 15 cm using a mixture of 5 volumes of

Apply separately to the plate 10 μ L of each solution. Develop

200 mL with methanol R.

Reference solution (b) Dilute 1 mL of test solution (a) to

same solvent.

hydrochloride CRS in methanol R and dilute to 10 mL with the

Reference solution (a) Dissolve 20 mg of racemic ephedrine

with methanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL

solvent.

examined in methanol R and dilute to 10 mL with the same

Test solution (a) Dissolve 0.20 g of the substance to be

[6-Hydroxy-2-(4-hydroxyphenyl)-1-benzothienyl]-[4-(2-(4-hydroxyphenyl)-1-benzothienyl)phenyl]methanone hydrochloride.

DEFINITION

Ph Eur

Action and use

Selective oestrogen receptor modulator.

C₂₈H₂₈ClNO₄S

510.0

82640-04-8

System suitability:

— resolution: minimum 3.0 between the peaks due to raloxifene and impurity C in the chromatogram obtained

— symmetry factor: maximum 1.8 for the principal peak in with reference solution (b);

— the chromatogram obtained with reference solution (a).

Limits:

— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29).

Buffer solution pH 2.5 7.2 g/L Solution of potassium dihydrogen phosphate R adjusted to pH 2.5 with phosphoric acid R.

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 50.0 mL

with the mobile phase. Dissolve 50.0 mg of raloxifene hydrochloride CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 5.0 mL of this

reference solution (a) Dissolve 50.0 mg of raloxifene hydrochloride CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 5.0 mL of this

reference solution (b) In order to produce impurity C *in situ*, to 6 mg of the substance to be examined add 15 mL of acetonitrile R, 3 mL of water R and 5 mL of stabilised strong hydrogen peroxide solution R. Store at 30 °C for at least 6 h, then dilute to 50.0 mL with buffer solution pH 2.5.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: base-deactivated octylsilyl silica gel for chromatography R (3.5 μ m);

— temperature: 35 °C.

Mobile phase acetonitrile R, buffer solution pH 2.5 (33:67 V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

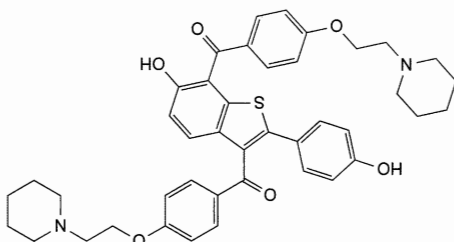
Injection 10 μ L.

Run time Twice the retention time of raloxifene.

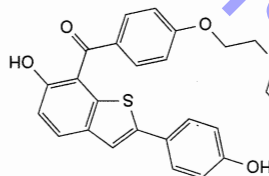
Relative retention With reference to raloxifene (retention time = about 3 min): impurity C = about 1.2.

System suitability:

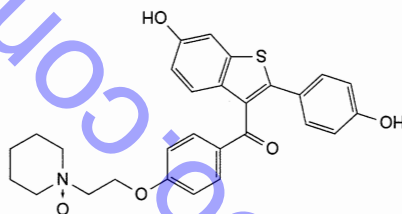
— resolution: minimum 2.0 between the peaks due to raloxifene and impurity C in the chromatogram obtained



A. [6-hydroxy-2-(4-hydroxyphenyl)-7-[4-[2-(piperidin-1-yl)ethoxy]benzoyl]-1-benzothiophen-3-yl][4-(2-piperidin-1-yl)ethoxy]phenylmethanone,



B. [6-hydroxy-2-(4-hydroxyphenyl)-1-benzothiophen-7-yl][4-(2-piperidin-1-yl)ethoxy]phenylmethanone,



C. [6-hydroxy-2-(4-hydroxyphenyl)-1-benzothiophen-3-yl][4-(2-piperidin-1-yl)ethoxy]phenylmethanone N-oxide.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C.

Specified impurities A

IMPURITIES

Calculate the percentage content of $C_{28}H_{28}ClNO_4$ from the declared content of raloxifene hydrochloride CRS. — symmetry factor: maximum 1.8 for the principal peak in the chromatogram obtained with reference solution (a). — concentration of acetonitrile in the mobile phase; with reference solution (b); if necessary, adjust the

Test solution Dissolve 0.200 g in a mixture of 0.3 volumes of *nitric acid R* and 99.7 volumes of *water R*, and dilute to 100.0 mL with the same mixture of solvents.

Reference solutions Use solutions containing 0.02 µg, 0.03 µg and 0.05 µg of palladium per millilitre, freshly prepared by dilution of *palladium standard solution (0.5 ppm Pd) R* with a mixture of 0.3 volumes of *nitric acid R* and 99.7 volumes of *water R*.

Modifier solution Dissolve 0.150 g of *magnesium nitrate R* in a mixture of 0.3 volumes of *nitric acid R* and 99.7 volumes of *water R*.

Injection 20 µL of the test solution and the reference solution, and 10 µL of the modifier solution.

Source Palladium hollow-cathode lamp using a transmission band and preferably of 1 nm and a graphite tube.

Wavelength 247.6 nm.

Loss on drying (2.2.32)
Maximum 0.2 per cent, determined on 1.000 g by drying in an oven under high vacuum at 60 °C for 4 h.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.300 g in 25 mL of *methanol R* and add 25 mL of *water R*. Titrate with 0.1 M *sodium hydroxide*, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M *sodium hydroxide* is equivalent to 41.65 mg of $C_{23}H_{32}N_2O_5$.

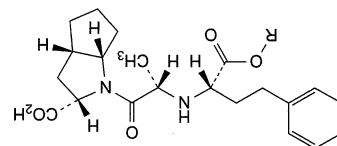
STORAGE

Protected from light.

IMPURITIES

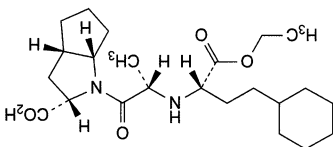
Specified impurities A, B, C, D

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10, *Control of impurities in substances for pharmaceutical use*: E, F, G, H, I, J, K, L, M, N, O.

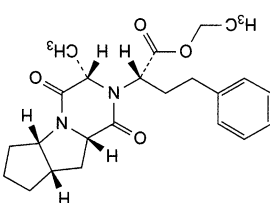


A. R = CH₃: (2S,3aS,6aS)-1-[(2S)-2-[(1S)-1-(methoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid (ramipril methyl ester).

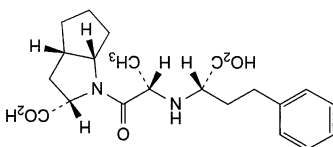
B. R = CH(CH₃)₂: (2S,3aS,6aS)-1-[(2S)-2-[(1S)-1-(1-methylethoxy)carbonyl]-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid (ramipril isopropyl ester).



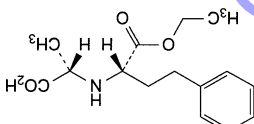
C. (2S,3aS,6aS)-1-[(2S)-2-[(1S)-3-cyclohexyl-(1-ethoxycarbonyl)-propyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid (hexahydrodroramipril).



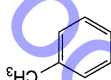
D. ethyl (2S)-2-[(3S,5S,8aS,9aS)-3-methyl-1,4-dioxodecahydro-2H-cyclopenta[4,5]pyrrolo[1,2-a]pyrazin-2-yl]-4-phenylbutanoate (ramipril dikeetopiperazine).



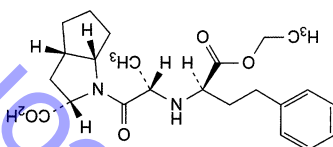
E. (2S,3aS,6aS)-1-[(2S)-2-[(1S)-1-carboxy-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid (ramipril diacid).



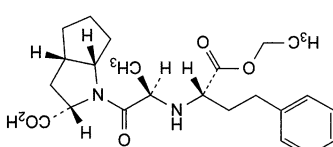
F. (2S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid.



G. methylbenzene (toluene).



H. (2S,3aS,6aS)-1-[(2S)-2-[(1R)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid ((R,S,S,S,S,S,S,S)-epimer of ramipril).

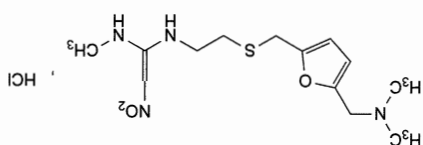


I. (2S,3aS,6aS)-1-[(2R)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid ((S,R,S,S,S,S,S,S)-epimer of ramipril).



Ranitidine Hydrochloride

(Ph. Eur. monograph 0946)



$C_{13}H_{23}ClN_4O_3S$ 350.9 66357-59-3

Action and use

Histamine H_2 receptor antagonist; treatment of peptic ulcer disease.

Preparations

Ranitidine Injection
Ranitidine Oral Solution
Ranitidine Tablets
Effervescent Ranitidine Tablets

DEFINITION

N -[2-[[[5-[(D -methylamino)methyl]furan-2-yl]methyl]sulfonyl]ethyl]- N' -methyl-2-nitroethene-1,1-diamine hydrochloride.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or pale yellow, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble or slightly soluble in anhydrous ethanol, very slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison ranitidine hydrochloride CRS.

If the spectra obtained in the solid state show differences, dissolve 10 mg of the substance to be examined and 10 mg of the reference substance separately in 0.5 mL of methanol *R* in an agate mortar. Evaporate to dryness under a stream of nitrogen *R*. Dry the residues under vacuum for 30 min. Add 3 drops of liquid paraffin *R* to the residues and triturate until the mull shows a milky appearance. Compare the mulls between 2 plates transparent to infrared radiation and record new spectra.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water *R* and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

pH (2.2.3)

4.5 to 6.0 for solution S.

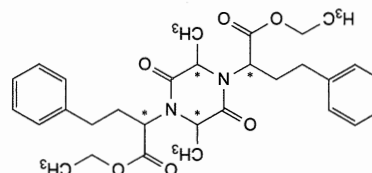
Related substances

Liquid chromatography (2.2.29).

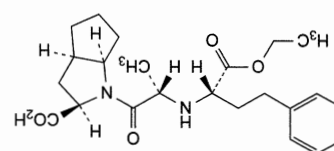
Buffer solution Dissolve 6.8 g of potassium dihydrogen phosphate *R* in 950 mL of water *R*. Adjust to pH 7.1 with

Ph Eur

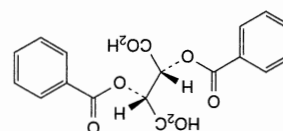
O, diethyl 2,2'-(2,5-dimethyl-3,6-dioxopiperazine-1,4-diyl)bis(4-phenylbutanoate).



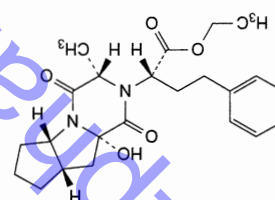
N. (2*R*,3*aR*,6*aR*)-1-[(2*S*)-2-[[[(1*S*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid ((*S*,*S*,*R*,*R*,*R*)-isomer of ramipril),



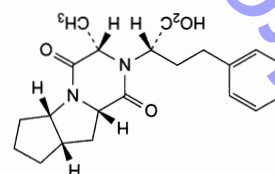
M. (2*R*,3*R*)-2,3-bis(benzoyloxy)butanedioic acid (dibenzoyltartaric acid),



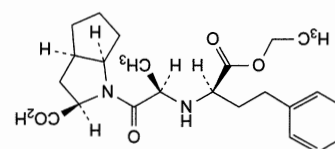
L. ethyl (2*S*)-2-[(3*S*,5*aS*,8*aS*,9*aS*)-9*a*-hydroxy-3-methyl-1,4-dioxodecahydro-2*H*-cyclopenta[4,5]pyrrolo[1,2-*a*]pyrazin-2-yl]-4-phenylbutanoate (ramipril hydroxydiketopiperazine),



K. (2*S*)-2-[(3*S*,5*aS*,8*aS*,9*aS*)-3-methyl-1,4-dioxodecahydro-2*H*-cyclopenta[4,5]pyrrolo[1,2-*a*]pyrazin-2-yl]-4-phenylbutanoic acid (ramipril diketopiperazine acid),



J. (2*R*,3*aR*,6*aR*)-1-[(2*R*)-2-[[[(1*R*)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydrocyclopenta[b]pyrrole-2-carboxylic acid (enantiomer of ramipril),



strong sodium hydroxide solution R and dilute to 1000 mL with water R.

Test solution Dissolve 13 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

Reference solution (a) Dissolve 6.5 mg of ranitidine for system suitability CRS (containing impurities A, D and H) in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A.

Reference solution (c) Dissolve the contents of a vial of ranitidine impurity J CRS in 1.0 mL of test solution.

Column:

— size: $l = 0.1$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl amorphous organosilica

— polymer R (3.5 μ m);

— temperature: 35 °C.

Mobile phase:

— mobile phase A: acetonitrile R, buffer solution (2:98 V/V);

— mobile phase B: acetonitrile R, buffer solution (22:78 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	100 \geq 0	0 \rightarrow 100
10 - 15	0	100

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 μ L of the test solution, reference solutions (a), (b) and (c) and mobile phase A as a blank.

Relative retention With reference to ranitidine (retention time = about 6.8 min): impurity H = about 0.1;

impurity G = about 0.2; impurity F = about 0.4;

impurity B = about 0.5; impurity C = about 0.6;

impurity E = about 0.7; impurity D = about 0.8;

impurity J = about 0.9; impurity I = about 1.3;

impurity A = about 1.7.

System suitability:

— resolution: minimum 1.5 between the peaks due to impurity J and ranitidine in the chromatogram obtained with reference solution (c);

— the chromatogram obtained with reference solution (a) is similar to the chromatogram supplied with ranitidine for system suitability CRS;

— the chromatogram obtained with the blank solution does not show any peak with the same relative retention as the peak due to impurity A in the chromatogram obtained with reference solution (a).

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity J by 2;

— impurity A: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurities B, C, D, E, F, G, H, I, J: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— sum of impurities other than A: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to the blank.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.75 per cent, determined on 1.000 g by drying under high vacuum at 60 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.280 g in 35 mL of water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 35.09 mg of $C_{13}H_{23}ClN_4O_3S$.

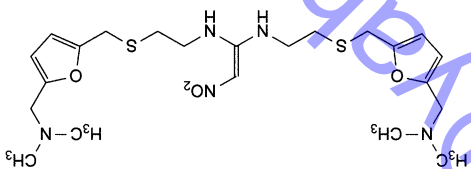
STORAGE

In airtight container, protected from light.

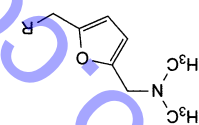
IMPURITIES

Specified impurities A, B, C, D, E, F, G, H, I, J

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): K.



A, N,N'-bis[2-[[[5-[(dimethylamino)methyl]furan-2-yl]methyl]sulfonyl]ethyl]-2-nitroethene-1,1-diamine,



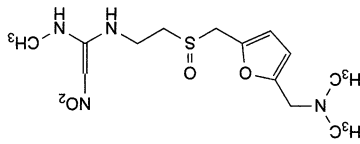
B, R = S-CH₂-CH₂-NH₂; 2-[[[5-

[(dimethylamino)methyl]furan-2-yl]methyl]sulfonyl]ethanamine,

D, R = S-CH₂-CH₂-NH-CO-CH₂-NO₂; N-[2-[[[5-

[(dimethylamino)methyl]furan-2-yl]methyl]sulfonyl]ethyl]-2-nitroacetamide,

F, R = OH; [5-[(dimethylamino)methyl]furan-2-yl]methanol,



C, N-[2-[[[5-[(dimethylamino)methyl]furan-2-yl]methyl]sulfonyl]ethyl]-N'-methyl-2-nitroethene-1,1-diamine,

two used in the manufacturing process is referred to as the host-vector system.

PRODUCTION

Production is based on a validated seed-lot system using a host-vector combination that has been shown to be suitable to the satisfaction of the competent authority. The seed-lot system uses a master cell bank and a working cell bank derived from the master seed lot of the host-vector combination. A detailed description of cultivation, extraction and purification steps and a definition of the production batch shall be established.

Where products of rDNA technology are manufactured using materials of human or animal origin, the requirements of chapter 5.1.7. *Viral safety* apply.

The determination of the suitability of the host-vector combination and the validation of the seed-lot system include the following elements.

CLONING AND EXPRESSION

The suitability of the host-vector system, particularly as regards microbiological purity, is demonstrated by: *Characterisation of the host cell, including source, phenotype and genotype, and of the cell-culture media;*

Documentation of the strategy for the cloning of the gene and characterisation of the recombinant vector, including:

- i. the origin and characterisation of the gene;
- ii. nucleotide-sequence analysis of the cloned gene and the flanking control regions of the expression vector; the cloned sequences are kept to a minimum and all relevant expressed sequences are clearly identified and confirmed at the RNA level; the DNA sequence of the cloned gene is normally confirmed at the seed-lot stage, up to and beyond the normal level of population doubling for full-scale fermentation;

in certain systems, for example, where multiple copies of the gene are inserted into the genome of a continuous cell line, it may be inappropriate to sequence the cloned gene at the production level; under these circumstances, Southern blot analysis of total cellular DNA or sequence analysis of the messenger RNA (mRNA) may be helpful, particular attention being paid to the characterisation of the expressed protein;

- iii. the construction, genetics and structure of the complete expression vector.

Characterisation of the host-vector system, including:

- i. mechanism of transfer of the vector into the host cells;
- ii. copy number, physical state and stability of the vector inside the host cell;
- iii. measures used to promote and control the expression.

CELL-BANK SYSTEM

The master cell bank Is a homogeneous suspension of the original cells already transformed by the expression vector containing the desired gene, distributed in equal volumes into individual containers for storage (for example, in liquid nitrogen). In some cases it may be necessary to establish separate master cell banks for the expression vector and the host cells.

The working cell bank Is a homogeneous suspension of the material derived from the master cell bank(s) at a finite passage level, distributed in equal volumes into individual containers for storage (for example, in liquid nitrogen). In both cell banks, all containers are treated identically during storage and, once removed from storage, the containers are not returned to the cell stock.

The cell bank may be used for production at a finite passage level or for continuous-culture production.

Production at a finite passage level

This cultivation method is defined by a limited number of passages or population doublings which must not be exceeded during production. The maximum number of doublings, or passage levels, during which the manufacturing process routinely meets the criteria described below must be stated.

Continuous-culture production

By this cultivation method the number of passages or population doublings is not restricted from the beginning of production. Criteria for the harvesting as well as for the termination of production have to be defined by the manufacturer. Monitoring is necessary throughout the life of the culture; the required frequency and type of monitoring will depend on the nature of the production system and the product.

Information is required on the molecular integrity of the gene being expressed and on the phenotypic and genotypic characteristics of the host cell after long-term cultivation. The acceptance of harvests for further processing must be clearly linked to the schedule of monitoring applied and a clear definition of a 'batch' of product for further processing is required.

VALIDATION OF THE CELL BANKS

Validation of the cell banks includes:

- i. stability by measuring viability and the retention of the vector;
- ii. identity of the cells by phenotypic features;
- iii. where appropriate, evidence that the cell banks are free from potentially oncogenic or infective adventitious agents (viral, bacterial, fungal or mycoplasma); special attention has to be given to viruses that can commonly contaminate the species from which the cell line has been derived; certain cell lines contain endogenous viruses, for example, retroviruses, which may not readily be eliminated; the expression of these organisms, under a variety of conditions known to cause their induction, shall be tested for;
- iv. for mammalian cells, details of the tumorigenic potential of the cell bank shall be obtained.

CONTROL OF THE CELLS

The origin, form, storage, use and stability at the anticipated rate of use must be documented in full for all cell banks under conditions of storage and recovery. New cell banks must be fully validated.

VALIDATION OF THE PRODUCTION PROCESS

Extraction and purification

The capacity of each step of the extraction and purification procedure to remove and/or inactivate contaminating substances derived from the host cell or culture medium, including, in particular, virus particles, proteins, nucleic acids and excipients, must be validated.

Validation studies are carried out to demonstrate that the production process routinely meets the following criteria:

- exclusion of extraneous agents from the product; studies including, for example, viruses with relevant physico-chemical features are undertaken, and a reduction capacity for such contaminants at each relevant stage of purification is established;
- adequate removal of vector, host-cell, culture medium and reagent-derived contaminants from the product; the reduction capacity for DNA is established by spiking;

- the reduction of proteins of animal origin can be determined by immunochemical methods;
- maintenance within stated limits of the yield of product from the culture;
- adequate stability of any intermediate of production and/or manufacturing when it is intended to use intermediate storage during the process.

Characterisation of the substance

The identity, purity, potency and stability of the final bulk product are established initially by carrying out a wide range of chemical, physical, immunochemical and biological tests. Prior to release, each batch of the product is tested by the manufacturer for identity and purity and an appropriate assay is carried out.

Production consistency

Suitable tests for demonstrating the consistency of the production and purification are performed. In particular, the tests include characterisation tests, in-process controls and final-product tests as exemplified below.

AMINO-ACID COMPOSITION

Partial amino-acid sequence analysis The sequence data permit confirmation of the correct N-terminal processing and detection of loss of the C-terminal amino acids.

Peptide mapping Peptide mapping using chemical and/or enzymatic cleavage of the protein product and analysis by a suitable method such as two-dimensional gel electrophoresis, capillary electrophoresis or liquid chromatography must show no significant difference between the test protein and the reference preparation. Peptide mapping can also be used to demonstrate correct disulphide bonding.

DETERMINATION OF MOLECULAR MASS

Cloned-gene retention The minimum percentage of cells containing the vector or the cloned gene after cultivation is approved by the relevant authority.

Total protein The yield of protein is determined. **Chemical purity** The purity of the protein product is analysed in comparison with a reference preparation by a suitable method such as liquid chromatography, capillary electrophoresis or sodium dodecyl sulfate polyacrylamide gel electrophoresis.

Host-cell-derived proteins Host-cell-derived proteins are detected by immunochemical methods, using, for example, polyclonal antisera raised against protein components of the host-vector system used to manufacture the product, unless otherwise prescribed. The following types of procedure may be used: liquid-phase displacement assays (for example, radio-binding assays using antigens immobilised on nitrocellulose (or similar) membranes (for example, dot-immunoblot assays, Western blots). General requirements for the validation of immunoassay procedures are given under 2.7.1. *Immunochemical Methods*. In addition, immunoassay methods for host-cell contaminants meet the following criteria.

— **Antigen preparations.** Antisera are raised against a preparation of antigens derived from the host organism, into which has been inserted the vector used in the manufacturing process that lacks the specific gene coding for the product. This host cell is cultured, and proteins are extracted, using conditions identical to those used for culture and extraction in the manufacturing process. Partly purified preparations of antigens, using some of the purification steps in the manufacturing process, may also be used for the preparation of antisera.

Hybridisation analysis

DNA in the test sample is denatured to give single-stranded DNA, immobilised on a nitrocellulose or other suitable filter and hybridised with labelled DNA prepared from the host-vector manufacturing system (DNA probes). Although a wide variety of experimental approaches is available, hybridisation methods for measurement of host-vector DNA meet the following criteria.

— **DNA probes.** Purified DNA is obtained from the host-vector system grown under the same conditions as those used in the manufacturing process. Host chromosomal DNA and vector DNA may be separately prepared and used as probes.

— **Calibration and standardisation.** Quantitative data are obtained by comparison with responses obtained using standard preparations. Chromosomal DNA probes and vector DNA probes are used with chromosomal DNA and vector DNA standards, respectively. Standard preparations are calibrated by spectroscopic measurements and stored in a state suitable for use over an extended period of time.

— **Hybridisation conditions.** The stringency of hybridisation conditions is such as to ensure specific hybridisation between probes and standard DNA preparations and the drug substances must not interfere with hybridisation at the concentrations used.

Sequence-independent techniques

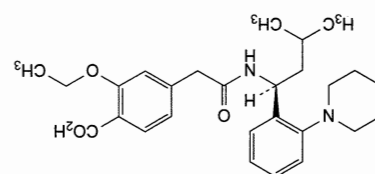
Suitable procedures include: detection of sulfonated cytosine residues in single-stranded DNA where DNA is immobilised on a filter and cytosines are derivatised *in situ*, before detection and quantitation using an antibody directed against the sulfonated group); detection of single-stranded DNA using a fragment of single-stranded DNA bound to a protein and an antibody of this protein. Neither procedure requires the use of specific host or vector DNA as an assay standard. However, the method used must be validated to ensure parallelism with the DNA standard used, linearity of response and non-interference of either the drug substance or excipients of the formulation at the dilutions used in the assay.

IDENTIFICATION, TESTS AND ASSAY

The requirements with which the final product (bulk material or dose form) must comply throughout its period of validity, as well as specific test methods, are stated in the individual monograph.

Repaglinide

(Ph. Eur. monograph 2135)

C₂₇H₃₆N₂O₄

432.6

135062-02-1

Action and use

Stimulates insulin release; treatment of diabetes mellitus.

Ph. Eur.

DEFINITION

2-Ethoxy-4-[[2-[(1S)-3-methyl-1-[2-(piperidin-1-yl)phenyl]butyl]amino]-2-oxoethyl]benzoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methanol and

in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Specific optical rotation (2.2.7): + 6.3 to + 7.7.

Dissolve 1.00 g in methanol R and dilute to 20.0 mL with the

same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison repaglinide CRS.

If the spectra obtained in the solid state show differences,

dissolve the substance to be examined and the reference

substance separately in anhydrous ethanol R, evaporate to

dryness and record new spectra using the residues.

TESTS

Enantiomeric purity

Liquid chromatography (2.2.29). Prepare the solutions in amber

flasks and vials.

Test solution Dissolve 10.0 mg of the substance to be

examined in methanol R and dilute to 10.0 mL with the same

solvent.

Reference solution (a) Dissolve 5.0 mg of repaglinide

impurity B CRS in methanol R and dilute to 50.0 mL with the

same of solvent.

Reference solution (b) Dilute 2.0 mL of reference solution (a)

to 100.0 mL with methanol R.

Reference solution (c) Mix 1.0 mL of the test solution and

10 mL of reference solution (a) and dilute to 50.0 mL with

methanol R.

Column:

— size: l = 0.1 m, Ø = 4.0 mm,

— stationary phase: silica gel AGP for chiral chromatography R

(5 µm).

Mobile phase:

— mobile phase A: 1.0 g/L solution of potassium dihydrogen

phosphate R adjusted to pH 4.7 with dilute sodium

hydroxide solution R;

— mobile phase B: acetonitrile R;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 4	80 → 60	20 → 40
4 - 6	60	40

Equilibration after installation of the column for use Using

water R, slowly increase the flow rate from 0.2 mL/min to

0.5 mL/min. Maintain the flow rate at 0.5 mL/min for

5 min. The column must be washed for 1 h at a flow rate of

1 mL/min with water R and for 1 h with the mobile phase at

the initial composition prior to the 1st analysis.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL of the test solution and reference solutions (b)

and (c).

Retention time Repaglinide = about 3.3 min;

impurity E = about 5.0 min.

System suitability: reference solution (c):

— resolution: minimum 1.5 between the peaks due to

repaglinide and impurity E.

Limit:

— impurity E: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.2 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 30.0 mg of the substance to be

examined in acetonitrile R and dilute to 10.0 mL with the

same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to

100.0 mL with acetonitrile R. Dilute 2.0 mL of this solution

to 100.0 mL with acetonitrile R.

Reference solution (b) With the aid of an ultrasonic bath,

dissolve the contents of 1 vial of repaglinide for system

suitability CRS in 2.0 mL of acetonitrile R.

Column:

— size: l = 0.15 m, Ø = 4.6 mm,

— stationary phase: silica gel for chromatography, alkyl-bonded

for use with highly aqueous mobile phases R (5 µm),

— temperature: 45 °C.

Mobile phase:

— mobile phase A: 4.0 g/L solution of potassium dihydrogen

phosphate R adjusted to pH 3.2 with dilute phosphoric

acid R;

— mobile phase B: mobile phase A, acetonitrile R

(300:700 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 20	50 → 7	50 → 93
20 - 30	7	93

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Relative retention With reference to repaglinide (retention

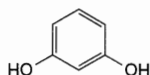
time = about 10 min): impurity A = about 0.2;

impurity B = about 0.3; impurity C = about 0.4;

impurity D = about 1.5.

Resorcinol

(Ph. Eur. monograph 0290)



110.1

C₆H₆O₂

108-46-3

Action and use

Keratolytic.

Ph Eur

DEFINITION

Resorcinol contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of benzene-1,3-diol, calculated with reference to the dried substance.

CHARACTERS

A colourless or slightly pinkish-grey, crystalline powder or crystals, turning red on exposure to light and air, very soluble in water and in alcohol.

IDENTIFICATION

A. Melting point (2.2.14): 109 °C to 112 °C.
B. Dissolve 0.1 g in 1 mL of water R, add 1 mL of strong sodium hydroxide solution R and 0.1 mL of chloroform R, heat and allow to cool. An intense, deep-red colour develops which becomes pale yellow on the addition of a slight excess of hydrochloric acid R.
C. Thoroughly mix about 10 mg with about 10 mg of potassium hydrogen phthalate R, both finely powdered. Heat over a naked flame until an orange-yellow colour is obtained. Cool and add 1 mL of dilute sodium hydroxide solution R and 10 mL of water R and shake to dissolve. The solution shows an intense green fluorescence.

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₅ or B₅ (2.2.2, Method II) and remains so when heated in a water-bath for 5 min.

Acidity or alkalinity

To 10 mL of solution S add 0.05 mL of 0.1 M bromophenol blue solution R₂. Not more than 0.05 mL of 0.1 M hydrochloric acid or 0.1 M sodium hydroxide is required to change the colour of the indicator.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel G R as the coating substance.

Test solution Dissolve 0.5 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.
Reference solution Dilute 0.1 mL of the test solution to 20 mL with methanol R.

Apply separately to the plate 2 µL of each solution. Develop over a path of 15 cm using a mixture of 40 volumes of ethyl acetate R and 60 volumes of hexane R. Allow the plate to dry in air for 15 min and expose it to iodine vapour. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison reserpine CRS.

C. To about 1 mg add 0.1 mL of a 1 g/L solution of sodium molybdate R in sulfuric acid R. A yellow colour is produced which becomes blue within 2 min.

D. To about 1 mg add 0.2 mL of a freshly prepared 10 g/L solution of vanillin R in hydrochloric acid R. A pink colour develops within 2 min.

E. Mix about 0.5 mg with 5 mg of dimethylaminobenzaldehyde R and 0.2 mL of glacial acetic acid R and add 0.2 mL of sulfuric acid R. A green colour is produced. Add 1 mL of glacial acetic acid R. The colour becomes red.

TESTS

Specific optical rotation (2.2.7)

–116 to –128 (dried substance).
Dissolve 0.250 g in chloroform R and dilute to 25.0 mL with the same solvent. Examine immediately.

Oxidation products

Dissolve 20 mg in glacial acetic acid R and dilute to 100.0 mL with the same acid. The absorbance (2.2.25) measured immediately at the absorption maximum at 388 nm is not greater than 0.10.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 0.500 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 667 Pa for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 0.5 g.

ASSAY

Total alkaloids

Dissolve 0.500 g in a mixture of 6 mL of acetic anhydride R and 40 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 60.9 mg of total alkaloids.

Reserpine

Protect the solutions from light. Moisten 25.0 mg with 2 mL of 10 mL of ethanol (96 per cent) R, add 2 mL of 0.25 M sulfuric acid and dissolve. Cool and dilute to 100.0 mL with ethanol (96 per cent) R. Dilute 5.0 mL of this solution to 50.0 mL with ethanol (96 per cent) R. Prepare a reference solution in the same manner using 25.0 mg of reserpine CRS. Place 10.0 mL of each solution separately in 2 boiling-tubes, add 2.0 mL of 0.25 M sulfuric acid and 2.0 mL of a freshly prepared 3 g/L solution of sodium nitrite R. Mix and heat in a water-bath at 55 °C for 35 min. Cool, add 1.0 mL of a freshly prepared 50 g/L solution of sulfamic acid R and dilute to 25.0 mL with ethanol (96 per cent) R. Measure the absorbance (2.2.25) of each solution at the absorption maximum at 388 nm, using as the compensation liquid 10.0 mL of the same solution prepared at the same time in the same manner, but omitting the sodium nitrite.

Calculate the content of C₃₃H₄₀N₂O₉ from the absorbances measured and the concentrations of the solutions.

STORAGE

Protected from light.

Ph Eur

Pyrocatechol

To 2 mL of solution S add 1 mL of ammonium molybdate solution R2 and mix. Any yellow colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using 2 mL of a 0.1 g/L solution of pyrocatechol R.

Loss on drying (2.2.32)

Not more than 1.0 per cent, determined on 1.00 g of powdered substance by drying in a desiccator for 4 h.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.500 g in water R and dilute to 250.0 mL with the same solvent. To 25.0 mL of the solution in a ground-glass-stoppered flask add 1.0 g of potassium bromide R, 50.0 mL of 0.0167 M potassium bromate, 15 mL of chloroform R and 15.0 mL of hydrochloric acid R1. Stopper the flask, shake and allow to stand in the dark for 15 min, shaking occasionally. Add 10 mL of a 100 g/L solution of potassium iodide R, shake thoroughly, allow to stand for 5 min and titrate with 0.1 M sodium thiosulfate, using 1 mL of starch solution R as indicator. 1 mL of 0.0167 M potassium bromate is equivalent to 1.835 mg of $C_{30}H_{50}O_2$.

STORAGE

Store protected from light.

Synthetic Retinol Concentrate (Oily Form)

(Vitamin A Concentrate (Oily Form) Synthetic, Ph Eur monograph 0219)

Action and use

Vitamin A.

Ph Eur

DEFINITION

Oily concentrate prepared from synthetic retinol ester (0217) as is or by dilution with a suitable vegetable fatty oil.

Content

95.0 per cent to 110.0 per cent of the vitamin A content stated on the label, which is not less than 500 000 IU/g. It may contain suitable stabilisers such as antioxidants.

CHARACTERS**Appearance**

Yellow or brownish-yellow, oily liquid.

Solubility

Practically insoluble in water, soluble or partly soluble in anhydrous ethanol, miscible with organic solvents. Partial crystallisation may occur in highly concentrated solutions.

IDENTIFICATION

Thin-layer chromatography (2.2.27).

Test solution Prepare a solution containing about 3.3 IU of vitamin A per micro litre in cyclohexane R containing 1 g/L of butylhydroxytoluene R.

Reference solution Prepare a 10 mg/mL solution of retinol esters CRS (i.e. 3.3 IU of each ester per micro litre) in cyclohexane R containing 1 g/L of butylhydroxytoluene R.

cyclohexane R containing 1 g/L of butylhydroxytoluene R.

Liquid chromatography (2.2.29).

Method B

If one or more of the ratios A_{326}/A_{326} exceeds the values given, or if the wavelength of the absorption maximum does not lie between 325 nm and 327 nm, use Method B.

A_{326} = absorbance at 326 nm,
 m = mass of the preparation to be examined, in grams,
 V = total volume to which the preparation to be examined is diluted to give 10–15 IU/mL,
 = factor to convert the specific absorbance of esters of retinol into International Units per gram.

$$\frac{A_{326} \times V \times 1900}{100 \times m}$$

Verify that the absorption maximum of the solution lies between 325 nm and 327 nm and measure the absorbances at 300 nm, 326 nm, 350 nm and 370 nm. Repeat the readings at each wavelength and take the mean values. Calculate the ratio A_{326}/A_{326} for each wavelength. If the ratios do not exceed: 0.60 at 300 nm, 0.54 at 350 nm, 0.14 at 370 nm, calculate the content of vitamin A in International Units per gram using the following expression:

Method A

Dissolve 25–100 mg, weighed with an accuracy of 0.1 per cent, in 5 mL of pentane R and dilute with 2-propanol R1 to a presumed concentration of 10–15 IU/mL. Carry out the assay as rapidly as possible, avoiding exposure to actinic light, air, oxidising agents, oxidation catalysts (e.g. copper, iron), acids and prolonged heat; use freshly prepared solutions. If partial crystallisation has occurred, homogenise the material at a temperature of about 65 °C, but avoid prolonged heating. Carry out the assay according to Method A. If the assay is not shown to be valid, use Method B.

ASSAY

The thresholds indicated under Related substances (Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Related substances

Maximum 10.0.
Peroxide value (2.5.5, Method A)
 Maximum 2.0, determined on 2.0 g.
Acid value (2.5.1)

TESTS

obtained with the reference solution.
 Results The composition of the test solution is confirmed by the correspondence of the principal spot or spots with those obtained with the reference solution.

top is: retinol acetate, retinol propionate and retinol palmitate.
 corresponding esters. The elution order from bottom to top is: retinol acetate, retinol propionate and retinol palmitate.

Detection Examine in ultraviolet light at 254 nm.

Drying In air.

Development Immediately, over a path of 15 cm.

Application 3 µL.

Mobile phase ether R, cyclohexane R (20:80 V/V).

Plate TLC silica gel F₂₅₄ plate R.

Synthetic Retinol Concentrate

(Powder Form)

(Vitamin A Concentrate (Powder Form), Synthetic, Ph Eur monograph 0218)

Action and use

Vitamin A.

DEFINITION

Powder concentrate obtained by dispersing a synthetic retinol ester (0217) in a matrix of *Gelatin* (0330) or *Acacia* (0307) or other suitable material.

Content

95.0 per cent to 115.0 per cent of the vitamin A content stated on the label, which is not less than 250 000 IU/g. It may contain suitable stabilisers such as antioxidants.

CHARACTERS

Appearance

Yellowish powder usually in the form of particles of almost uniform size.

Solubility

Practically insoluble in water, swells or forms an emulsion, depending on the formulation.

IDENTIFICATION

Thin-layer chromatography (2.2.27).

Test solution Introduce a quantity of the preparation to be examined containing about the equivalent of 17 000 IU of vitamin A into a 20 mL glass-stoppered test tube. Add about 20 mg of *bromelains* R, 2 mL of *water* R and about 150 µL of 2-propanol R, swirling gently for 2–5 min in a water-bath at 60–65 °C. Cool to below 30 °C and add 5 mL of 2-propanol R containing 1 g/L of *butylhydroxytoluene* R. Shake vigorously for 1 min, allow to stand for a few minutes and use the supernatant solution.

Reference solution Prepare a 10 mg/mL solution of *retinol* esters CRS (i.e. 3.3 IU of each ester per micro litre) in 2-propanol R containing 1 g/L of *butylhydroxytoluene* R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ether R, *cyclohexane* R (20:80 V/V).

Application 3 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution:

— the chromatogram shows the individual spots of the corresponding esters. The elution order from bottom to top is: retinol acetate, retinol propionate and retinol palmitate.

Results The composition of the test solution is confirmed by the correspondence of the principal spot or spots with those obtained with the reference solution.

TESTS

Related substances

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

ASSAY

Carry out the assay as rapidly as possible, avoiding exposure to actinic light, air, oxidising agents, oxidation catalysts (e.g. copper, iron), acids and prolonged heat.

Test solution (a) Introduce 0.100 g of the preparation to be examined into a 100 mL volumetric flask and dissolve immediately in 5 mL of *pentane* R. Add 40 mL of 0.1 M *tetrabutylammonium hydroxide* in 2-propanol. Swirl gently and let the mixture react for 10 minutes at 60–65 °C for hydrolysis, swirling occasionally. Allow to cool to room temperature, dilute to 100.0 mL with 2-propanol R containing 1 g/L *butylhydroxytoluene* R, and homogenise carefully to avoid air-bubbles.

Test solution (b) Dilute test solution (a) with 2-propanol R to a final concentration of 100 IU/mL.

Reference solution (a) Introduce about 0.100 g of *retinol*

acetate CRS into a 100 mL volumetric flask and proceed as described for test solution (a).

Reference solution (b) Introduce into a 50 mL volumetric flask 5.0 mL of reference solution (a) and dilute to 50.0 mL with 2-propanol R. Homogenise carefully to avoid air-bubbles.

Column:

— size: $l = 0.125$ m, $\phi = 4$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase *water* R, *methanol* R (5:95 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 325 nm.

Injection 10 µL of test solution (b) and reference solution (b).

Run time 1.5 times the retention time of retinol.

Retention time Retinol = about 3 min.

Calculate the content of vitamin A in International Units per gram using the following expression:

$$\frac{A_1 \times C \times m_2}{A_2 \times m_1}$$

A_1 = area of the peak due to retinol in the chromatogram

obtained with test solution (b),

A_2 = area of the peak due to retinol in the chromatogram

obtained with reference solution (b),

C = concentration of *retinol acetate* CRS in International

Units per gram, determined by method A; the

absorption ratios A_1/A_{326} must conform,

m_1 = mass of the substance to be examined in test

solution (a), in milligrams,

m_2 = mass of *retinol acetate* CRS in reference solution (a), in milligrams.

STORAGE

In an airtight container, protected from light.

Once the container has been opened, its contents are to be used as soon as possible; any part of the contents not used once should be protected by an atmosphere of inert gas.

LABELLING

The label states:

- the number of International Units per gram,
- the name of the ester or esters,
- the name of any added stabilisers,
- the method of restoring the solution if partial crystallisation has occurred.

Ph Eur



If the ratios do not exceed: 0.60 at 300 nm, 0.54 at 350 nm, 0.14 at 370 nm, calculate the content of vitamin A in International Units per gram using the following expression:

$$A_{326} \times V \times 1900$$

$$100 \times m$$

A_{326} = absorbance at 326 nm,
 m = mass of retinol acetate CRS, in grams,
 V = total volume to which the retinol acetate CRS is diluted to give 10-15 IU/mL,
 1900 = factor to convert the specific absorbance of esters of retinol into International Units per gram.

The absorbance ratios A_1/A_{326} must conform.

STORAGE

In an airtight container, protected from light.
 Once the container has been opened, its contents are to be used as soon as possible; any part of the contents not used at once should be protected by an atmosphere of inert gas.

LABELLING

The label states:
 — the number of International Units per gram,
 — the name of the ester or esters,
 — the name of the principal excipient or excipients used and the name of any added stabilisers.

Ph Eur



Synthetic Retinol Concentrate, Solubilisate/Emulsion

Synthetic Retinol Concentrate (Water-dispersible Form)

(Vitamin A Concentrate, (Solubilisate/Emulsion), Synthetic, Ph Eur monograph 0220)

Ph Eur

DEFINITION

Liquid concentrate (water is generally used as solvent) of a synthetic retinol ester (0217) and a suitable solubiliser.

Content

95.0 per cent to 115.0 per cent of the vitamin A content stated on the label, which is not less than 100 000 IU/g. It may contain suitable stabilisers such as antimicrobial preservatives and antioxidants.

CHARACTERS

Appearance

Yellow or yellowish liquid of variable opalescence and viscosity. Highly concentrated solutions may become cloudy at low temperature or take the form of a gel. A mixture of 1 g with 10 mL of water R previously warmed to 50 °C gives, after cooling to 20 °C, a uniform, slightly opalescent and slightly yellow dispersion.

IDENTIFICATION

Thin-layer chromatography (2.2.27).

Test solution Introduce a quantity of the preparation to be examined containing about the equivalent of 17 000 IU of vitamin A into a 20 mL glass-stoppered test-tube. Add 5 mL of 2-propanol R containing 1 g/L of butyllithoxytoluene R and mix thoroughly.

Liquid chromatography (2.2.29).

Test solution (a) Introduce 0.200 g of the preparation to be examined into a 100 mL volumetric flask. Add 20-30 mg of bromelain R, 5.0 mL of water R and 0.15 mL of

2-propanol R. Heat gently in a water-bath at 60 °C for about 5 min, swirling occasionally. Add 40 mL of 0.1 M tetrabutylammonium hydroxide in 2-propanol. Swirl gently and let the mixture react for 10 min at 60-65 °C for hydrolysis, swirling occasionally. Ensure that all sample material is wetted. Allow to cool to room temperature, dilute to 100.0 mL with 2-propanol R containing 1 g/L butyllithoxytoluene R, and homogenise carefully to avoid air-bubbles. The solution may be turbid.

Test solution (b) Dilute test solution (a) with 2-propanol R to a final concentration of 100 IU/mL. Filter before injection.

Reference solution (a) Introduce about 0.100 g of retinol acetate CRS into a 100 mL volumetric flask and dissolve immediately in 5 mL of pentane R. Add 40 mL of 0.1 M tetrabutylammonium hydroxide in 2-propanol. Swirl gently and let the mixture react for 10 min at 60-65 °C for hydrolysis, swirling occasionally. Allow to cool to room temperature, dilute to 100.0 mL with 2-propanol R containing 1 g/L butyllithoxytoluene R, and homogenise carefully to avoid air-bubbles.

Reference solution (b) Introduce into a 50 mL volumetric flask 5.0 mL of reference solution (a) and dilute to 50.0 mL with 2-propanol R. Homogenise carefully to avoid air-bubbles.

Column:

— size: $l = 0.125$ m, $\varnothing = 4$ mm,
 — stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase water R, methanol R (5:95 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 325 nm.

Injection 10 µL of test solution (b) and reference solution (b).

Run time 1.5 times the retention time of retinol.

Retention time Retinol = about 3 min.

Calculate the content of vitamin A using the following expression:

$$\frac{A_1 \times C \times m_2}{A_2 \times m_1}$$

A_1 = area of the peak due to retinol in the chromatogram obtained with test solution (b),
 A_2 = area of the peak due to retinol in the chromatogram obtained with reference solution (b),
 C = concentration of retinol acetate CRS in International Units per gram, determined by the method below,
 m_1 = mass of the substance to be examined in test solution (a), in milligrams,
 m_2 = mass of retinol acetate CRS in reference solution (a), in milligrams.

The exact concentration of retinol acetate CRS is assessed by ultraviolet absorption spectrophotometry (2.2.25). Dissolve 25-100 mg of retinol acetate CRS, weighed with an accuracy of 0.1 per cent, in 5 mL of pentane R and dilute with 2-propanol R to a presumed concentration of 10-15 IU/mL. Verify that the absorption maximum of the solution lies between 325 nm and 327 nm and measure the absorbances at 300 nm, 326 nm, 350 nm and 370 nm. Repeat the readings at each wavelength and take the mean values.

Calculate the ratio A_1/A_{326} for each wavelength.

Reference solution Prepare a 10 mg/mL solution of retinol

esters CRS (i.e. 3.3 IU of each ester per micro litre) in 2-propanol R containing 1 g/L of butylhydroxytoluene R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ether R, cyclohexane R (20:80 V/V).

Application 3 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution:

the chromatogram shows the individual spots of the

corresponding esters. The elution order from bottom to

top is: retinol acetate, retinol propionate and retinol

palmitate.

Results The composition of the test solution is confirmed by

the correspondence of the principal spot or spots with those

obtained with the reference solution.

TESTS

Related substances

The thresholds indicated under Related substances

(Table 2034-1) in the general monograph Substances for

pharmaceutical use (2034) do not apply.

ASSAY

Carry out the assay as rapidly as possible, avoiding exposure to actinic light, air, oxidising agents, oxidation catalysts (e.g. copper, iron), acids and prolonged heat.

Liquid chromatography (2.2.29).

Test solution (a) Introduce 0.200 g of the preparation to be

examined into a 100 mL volumetric flask. Add 40 mL of

0.1 M tetrabutylammonium hydroxide in 2-propanol. Swirl into

dispersion and let the mixture react for 10 min at 60–65 °C

for hydrolysis, swirling occasionally. Ensure that all sample

material is wetted. Allow to cool to room temperature, dilute

to 100.0 mL with 2-propanol R containing 1 g/L

butylhydroxytoluene R and homogenise carefully to avoid air-

bubbles. Residue of the matrix may make the solution more

or less cloudy.

Test solution (b) Dilute test solution (a) with 2-propanol R to a

final concentration of 100 IU/mL. Filter before injection.

Reference solution (a) Introduce about 0.100 g of retinol

acetate CRS into a 100 mL volumetric flask and dissolve

immediately in 5 mL of pentane R. Add 40 mL of 0.1 M

tetrabutylammonium hydroxide in 2-propanol. Swirl gently and

let the mixture react for 10 min at 60–65 °C for hydrolysis,

swirling occasionally. Allow to cool to room temperature,

dilute to 100.0 mL with 2-propanol R containing 1 g/L

butylhydroxytoluene R, and homogenise carefully to avoid air-

bubbles.

Reference solution (b) Introduce into a 50 mL volumetric flask

5.0 mL of reference solution (a) and dilute to 50.0 mL with

2-propanol R. Homogenise carefully to avoid air-bubbles.

Column:

— size: $l = 0.125$ m, $\phi = 4$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 µm).

Mobile phase water R, methanol R (5:95 V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 325 nm.

Injection 10 µL of test solution (b) and reference solution (b).

Run time 1.5 times the retention time of retinol.

Retention time Retinol = about 3 min.

expression:

$$A_1 \times C \times m_2$$

$$A_2 \times m_1$$

Calculate the content of vitamin A using the following

A_1 = area of the peak due to retinol in the chromatogram

obtained with test solution (b),

A_2 = area of the peak due to retinol in the chromatogram

obtained with reference solution (b),

C = concentration of retinol acetate CRS in International

Units per gram, determined by the method below,

m_1 = mass of the substance to be examined in test

solution (a), in milligrams,

m_2 = mass of retinol acetate CRS in reference solution (a),

in milligrams.

The exact concentration of retinol acetate CRS is assessed by

ultraviolet absorption spectrophotometry (2.2.25). Dissolve

25–100 mg of retinol acetate CRS, weighed with an accuracy

of 0.1 per cent, in 5 mL of pentane R and dilute with 2-

propanol R to a presumed concentration of 10–15 IU/mL.

Verify that the absorption maximum of the solution lies

between 325 nm and 327 nm and measure the absorbances

at 300 nm, 326 nm, 350 nm and 370 nm. Repeat the

readings at each wavelength and take the mean values.

Calculate the ratio A_{326}/A_{370} for each wavelength.

If the ratios do not exceed: 0.60 at 300 nm, 0.54 at 350 nm,

0.14 at 370 nm, calculate the content of vitamin A in

International Units per gram using the following expression:

$$\frac{A_{326} \times V \times 1900}{100 \times m}$$

A_{326} = absorbance at 326 nm,

m = mass of retinol acetate CRS, in grams,

V = total volume to which the retinol acetate CRS is

diluted to give 10–15 IU/mL,

1900 = factor to convert the specific absorbance of esters

of retinol into International Units per gram.

The absorbance ratios A_{326}/A_{370} must conform.

STORAGE

In an airtight container, protected from light, at the

temperature stated on the label.

Once the container has been opened, its contents are to be

used as soon as possible; any part of the contents not used at

once should be protected by an atmosphere of inert gas.

LABELLING

The label states:

— the number of International Units per gram,

— the name of the ester or esters,

— the name of the principal solvent or solvents used

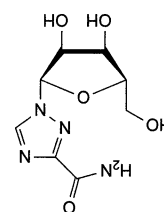
and the name of any added stabilisers,

— the storage temperature.

Ph Eur

Ribavirin

(Ph Eur monograph 2109)

 $C_8H_{12}N_4O_5$

244.2

36791-04-5

Action and use

Antiviral (hepatitis C, respiratory syncytial virus).

Preparation

Ribavirin Powder for Nebuliser Solution

DEFINITION

1-β-D-Ribofuranosyl-1H-1,2,4-triazole-3-carboxamide.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent), slightly soluble or very slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison ribavirin CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methylene chloride R, evaporate to dryness and record new spectra using the residues.

TESTS

pH (2.2.3)

4.0 to 6.5.

Dissolve 0.200 g in carbon dioxide-free water R and dilute to 10.0 mL with the same solvent.

Specific optical rotation (2.2.7)

-33 to -37 (dried substance).

Dissolve 0.250 g in water R and dilute to 25.0 mL with the same solvent. Determine the specific optical rotation within 10 min of preparing the solution.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in water for chromatography R and dilute to 100.0 mL with the same solvent.

Reference solution (a) In order to produce impurity A *in situ*, mix 5.0 mL of the test solution and 5.0 mL of a 42 g/L solution of sodium hydroxide R and allow to stand for 90 min. Neutralise with 5.0 mL of a 103 g/L solution of hydrochloric acid R and mix well.

solution of sodium hydroxide R and allow to stand for 90 min. Neutralise with 5.0 mL of a 103 g/L solution of hydrochloric acid R and mix well.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with water for chromatography R. Dilute 1.0 mL of this solution to 10.0 mL with water for chromatography R.

Reference solution (c) Dissolve 50.0 mg of ribavirin CRS in water for chromatography R and dilute to 100.0 mL with the same solvent.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (3 μ m) suitable for use with highly aqueous mobile phases;

— temperature: 25 °C.

Mobile phase:

— mobile phase A: dissolve 1.0 g of anhydrous sodium sulfate R in 950 mL of water for chromatography R, add 2.0 mL of a 5 per cent V/V solution of phosphoric acid R, adjust to pH 2.8 with a 5 per cent V/V solution of phosphoric acid R

and dilute to 1000 mL with water for chromatography R; — mobile phase B: acetonitrile R1, mobile phase A (5:95 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 15	100	0
15 - 25	100 → 0	0 → 100
25 - 35	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 5 μ L of the test solution and reference solutions (a) and (b).

Relative retention With reference to ribavirin (retention time = about 6 min): impurity A = about 0.8.

System suitability: reference solution (a):

— resolution: minimum 4.0 between the peaks due to impurity A and ribavirin.

Limits:

— correction factor: for the calculation of contents, multiply the peak area of impurity A by 2.3;

— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 4.0 g in 20 mL of water R, with heating if necessary.

12 mL of the solution complies with test A. Prepare the reference solution using 10 mL of lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 5 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

Calculate the percentage content of $C_{8}H_{12}N_4O_5$ from the declared content of riboflavin CRS.

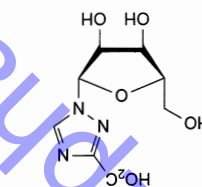
STORAGE

Protected from light.

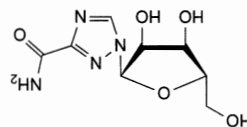
IMPURITIES

Specified impurities A.

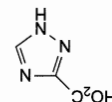
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, F, G.



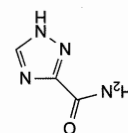
A. 1-β-D-ribofuranosyl-1H-1,2,4-triazole-3-carboxylic acid.



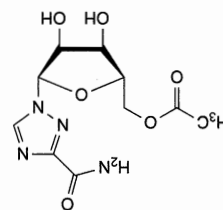
B. 1-α-D-ribofuranosyl-1H-1,2,4-triazole-3-carboxamide (anomer),



C. 1H-1,2,4-triazole-3-carboxylic acid.



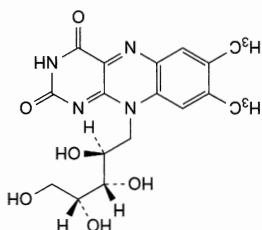
D. 1H-1,2,4-triazole-3-carboxamide,



F. 1-(5-O-acetyl-β-D-ribofuranosyl)-1H-1,2,4-triazole-3-carboxamide (5'-O-acetylriboflavin),

Riboflavin

(Ph. Eur. monograph 0292)



$C_{17}H_{20}N_4O_6$

376.4

83-88-5

Action and use
Vitamin B₂.

Ph. Eur.

DEFINITION

7,8-Dimethyl-10-[(2S,3S,4R)-2,3,4,5-tetrahydroxy-2,4-pyridinediyl]benzo[g]pteridine-2,4(3H,10H)-dione. This monograph applies to riboflavin produced by fermentation.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow or orange-yellow, crystalline powder.

Solubility

Very slightly soluble in water, practically insoluble in ethanol (96 per cent). Solutions deteriorate on exposure to light, especially in the presence of alkali. It shows polymorphism (5.9).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Thin-layer chromatography (2.2.27).

Test solution Suspend 25 mg of the substance to be examined in 10 mL of water R, shake for 5 min and filter the suspension to remove the undissolved material.

Reference solution Suspend 25 mg of riboflavin CRS in 10 mL of water R, shake for 5 min and filter the suspension to remove the undissolved material.

Plate TLC silica gel plate R (2-10 μm).

Mobile phase water R.

Application As follows, drying in a current of cold air after each individual application:

— 1st application: 2 μL of methylene chloride R then 2 μL of the test solution;

— 2nd application: 2 μL of methylene chloride R then 2 μL of the reference solution.

Development Over a path of 6 cm.

Drying In a current of cold air.

Detection Examine in ultraviolet light at 365 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. Dissolve about 1 mg in 100 mL of water R. The solution has, by transmitted light, a pale greenish-yellow colour, and, by reflected light, an intense yellowish-green fluorescence which disappears on the addition of mineral acids or alkalis.

TESTS

Specific optical rotation (2.2.7)

–115 to –135 (dried substance).

Dissolve 50.0 mg in 0.05 M sodium hydroxide free from carbonate and dilute to 10.0 mL with the same alkaline solution. Measure the optical rotation within 30 min of dissolution.

Absorbance (2.2.25)

Test solution Dilute the final solution prepared for the assay with an equal volume of water R.

Absorption maxima At 223 nm, 267 nm, 373 nm and 444 nm.

Absorbance ratios:

– $A_{267}/A_{297} = 0.31$ to 0.33;

– $A_{444}/A_{267} = 0.36$ to 0.39.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and protect from light.

Solution A 13.6 g/L solution of sodium acetate R.

Test solution With the aid of ultrasound, dissolve 0.120 g of the substance to be examined in 10 mL of 0.1 M sodium hydroxide and dilute to 100 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (b) With the aid of ultrasound, dissolve the contents of a vial of riboflavin for peak identification CRS (containing impurities C and D) in 1.0 mL of a mixture of 1 volume of mobile phase B and 9 volumes of mobile phase A.

Reference solution (c) In order to prepare *in situ* impurities A and B, dissolve 10 mg of the substance to be examined in 1 mL of 0.5 M sodium hydroxide. Expose to daylight for 1.5 h. Add 0.5 mL of acetic acid R and dilute to 100 mL with water R.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: phosphoric acid R, water R (1:1000 V/V);

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	90	10
5 - 20	90 \rightarrow 80	10 \rightarrow 20
20 - 25	80	20
25 - 35	80 \rightarrow 50	20 \rightarrow 50
35 - 45	50	50

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 267 nm.

Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with riboflavin for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C and D.

Relative retention With reference to riboflavin (retention time = about 16 min): impurity C = about 0.2;

impurity D = about 0.5; impurity A = about 1.4;

impurity B = about 1.9.

System suitability:

— resolution: minimum 5 between the peaks due to impurities A and B in the chromatogram obtained with reference solution (c);

— the chromatogram obtained with reference solution (b) is similar to the chromatogram supplied with riboflavin for peak identification CRS.

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.7;

impurity B = 1.4; impurity C = 2.3; impurity D = 1.4;

impurity A: not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.025 per cent);

impurities B, C, D: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit for peaks other than those due to impurity A: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 1.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on the residue obtained in the test for loss on drying.

ASSAY

Carry out the assay protected from light.

In a brown-glass 500 mL volumetric flask, suspend 65.0 mg in 5 mL of water R ensuring that it is completely wetted and dissolve in 5 mL of dilute sodium hydroxide solution R. As soon as dissolution is complete, add 100 mL of water R and 2.5 mL of glacial acetic acid R and dilute to 500.0 mL with water R. Place 20.0 mL of this solution in a 200 mL brown-glass volumetric flask, add 3.5 mL of a 14 g/L solution of sodium acetate R and dilute to 200.0 mL with water R.

Measure the absorbance (2.2.25) at the absorption maximum at 444 nm.

Calculate the content of $C_{17}H_{20}N_4O_6$ taking the specific absorbance to be 328.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A, B, C, D

Content
73.0 per cent to 79.0 per cent of riboflavin ($C_{17}H_{20}N_4O_6$; 376.4) (dried substance).
It contains a variable quantity of water.

CHARACTERS

Appearance
Yellow or orange-yellow, crystalline, hygroscopic powder.

Solubility

Soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 50.0 mg in phosphate buffer solution pH 7.0 R and dilute to 100.0 mL with the same buffer solution. Dilute 2.0 mL of this solution to 100.0 mL with phosphate buffer solution pH 7.0 R.

Spectral range 230–350 nm.

Absorption maximum At 266 nm.

B. Examine the chromatograms obtained in the test for specific absorbance at the absorption maximum 580 to 640.

Results The principal peak in the chromatogram obtained with the test solution is similar in position and approximate size to the principal peak in the chromatogram obtained with reference solution (b).

C. Dissolve about 10 mg in dilute sodium hydroxide solution R and dilute to 100 mL with the same solution. Expose 1 mL of this solution to ultraviolet light at 254 nm for 5 min, add sufficient acetic acid R to make the solution acidic to blue litmus paper R and shake with 2 mL of methylene chloride R. The lower layer shows yellow fluorescence.

D. To 0.5 g add 10 mL of nitric acid R and evaporate the mixture to dryness on a water-bath. Ignite the residue until it becomes white, dissolve the residue in 5 mL of water R and filter. The filtrate gives reaction (a) of sodium and reaction (b) of phosphates (2.3.1).

TESTS

pH (2.2.3)

5.0 to 6.5.

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Specific optical rotation (2.2.7)

Dissolve 0.300 g in 18.2 mL of hydrochloric acid R1 and dilute to 25.0 mL with water R.

Impurity E

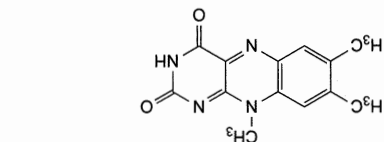
To about 35 mg add 10 mL of methylene chloride R, shake for 5 min and filter. The filtrate is not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Related substances

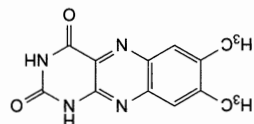
Liquid chromatography (2.2.29). Carry out the test protected from actinic light.

Test solution Dissolve 0.100 g of the substance to be examined in 50 mL of water R and dilute to 100.0 mL with the mobile phase. Dilute 8.0 mL of this solution to 50.0 mL with the mobile phase.

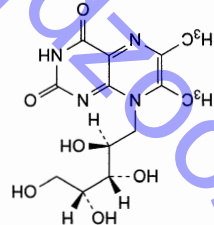
Reference solution (a) Dissolve 60 mg of riboflavin CRS (impurity D) in 1 mL of hydrochloric acid R and dilute to 250.0 mL with water R. Dilute 4.0 mL of this solution to 100.0 mL with the mobile phase.



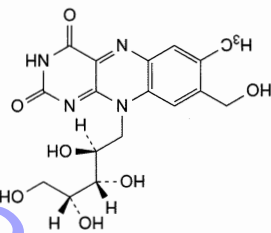
A. 7,8,10-trimethylbenzo[g]pteridine-2,4(3H,10H)-dione (lumiflavin),



B. 7,8-dimethylbenzo[g]pteridine-2,4(1H,3H)-dione,



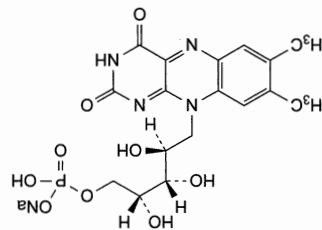
C. 6,7-dimethyl-8-[(2S,3S,4R)-2,3,4,5-tetrahydroxypentyl]pteridine-2,4(3H,8H)-dione,



D. 8-(hydroxymethyl)-7-methyl-10-[(2S,3S,4R)-2,3,4,5-tetrahydroxypentyl]benzo[g]pteridine-2,4(3H,10H)-dione.

Riboflavin Sodium Phosphate

(Ph. Eur. monograph 0786)



$C_{17}H_{20}N_4NaO_6P$

478.3

130-40-5

Vitamin B₂.

Preparation

Vitamins B and C Injection

DEFINITION

Mixture containing riboflavin 5'-(sodium hydrogen phosphate) as the main component and other riboflavin sodium monophosphates.

Reference solution (b) Dissolve 0.100 g of riboflavin sodium phosphate CRS in 50 mL of water R and dilute to 100.0 mL with the mobile phase. Dilute 8.0 mL of this solution to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase methanol R, 7.35 g/L solution of potassium

dihydrogen phosphate R (150:850 V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 266 nm.

Injection 100 μ L.

Run time Until the peak due to riboflavin can be clearly

evaluated.

Relative retention With reference to riboflavin

5'-monophosphate (retention time = about 20 min):

impurity A = about 0.2; impurity B = about 0.3;

impurity C = about 0.5; riboflavin

4'-monophosphate = about 0.7; riboflavin

3'-monophosphate = about 0.9; impurity D = about 2.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to

riboflavin 4'-monophosphate and riboflavin

5'-monophosphate.

Calculate the percentage content of free riboflavin

(impurity D) and of riboflavin in the form of the

diphosphates of riboflavin (impurities A, B, C) from the areas

solution and the amount of free riboflavin in reference

solution (a).

Limits:

— impurity D: maximum 6.0 per cent (dried substance);

— sum of impurities A, B and C: maximum 6.0 per cent

(dried substance).

The thresholds indicated under Related substances

(Table 2034.-1) in the general monograph Substances for

pharmaceutical use (2034) do not apply.

Inorganic phosphate

Maximum 1.5 per cent.

Dissolve 0.10 g in water R and dilute to 100 mL with the

same solvent. To 5 mL of this solution, add 10 mL of

water R, 5 mL of buffered copper sulfate solution pH 4.0 R,

2 mL of a 30 g/L solution of ammonium molybdate R, 1 mL

of a freshly prepared solution containing 20 g/L of

4-methylaminophenol sulfate R and 50 g/L of sodium

metabisulfite R, and 1 mL of a 3 per cent V/V solution of

perchloric acid R. Dilute to 25.0 mL with water R and

measure, within 15 min of its preparation, the absorbance

(2.2.25) of the solution at 800 nm, using as the

compensation liquid a solution prepared in the same manner

but without the substance to be examined. The absorbance is

not greater than that of a solution prepared as follows: to

15 mL of phosphate standard solution (5 ppm PO_4) R, add

5 mL of buffered copper sulfate solution pH 4.0 R, 2 mL of a

30 g/L solution of ammonium molybdate R, 1 mL of a freshly

prepared solution containing 20 g/L of 4-methylaminophenol

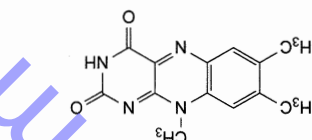
sulfate R and 50 g/L of sodium metabisulfite R, and 1 mL of a

3 per cent V/V solution of perchloric acid R; dilute to 25.0 mL

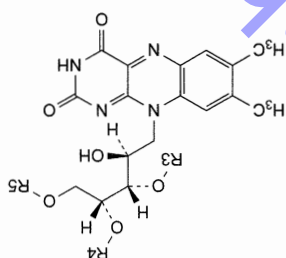
with water R.

Heavy metals (2.4.8)

Maximum 10 ppm.



- A. R3 = R4 = PO_3H_2 , R5 = H: riboflavin 3',4'-diphosphate;
 B. R3 = R5 = PO_3H_2 , R4 = H: riboflavin 3',5'-diphosphate;
 C. R3 = H, R4 = R5 = PO_3H_2 : riboflavin 4',5'-diphosphate;
 D. R3 = R4 = R5 = H: riboflavin.



Specified impurities A, B, C, D, E

IMPURITIES

In an airtight container, protected from light.

STORAGE

absorbance to be 328.

Calculate the content of $\text{C}_{17}\text{H}_{20}\text{N}_4\text{O}_6$ taking the specific

at 444 nm.

Measure the absorbance (2.2.25) at the absorption maximum

of sodium acetate R and dilute to 50.0 mL with water R.

To 10.0 mL of this solution add 3.5 mL of a 14 g/L solution

acetic acid R and dilute to 100.0 mL with water R.

Dissolve 0.100 g in 150 mL of water R, add 2 mL of glacial

Carry out the assay protected from light

ASSAY

5 h.

an oven at 105 °C at a pressure not exceeding 0.7 kPa for

Maximum 8.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

standard solution (1 ppm Pb) R.

test A. Prepare the reference solution using 10 mL of lead

to 20 mL with water R. 12 mL of the solution complies with

Dissolve the residue in 2 mL of dilute acetic acid R and dilute

hydrochloric acid R and evaporate the extracts to dryness.

the cooled residue with 2 quantities, each of 2 mL, of

cautiously until white fumes are evolved and ignite. Extract

dropwise, followed by 0.25 mL of sulfuric acid R. Heat

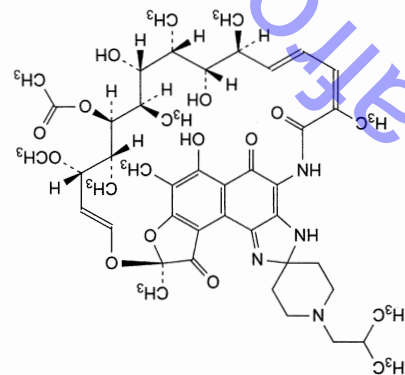
To 2.0 g in a silica crucible add 2 mL of nitric acid R,

E. 7,8,10-trimethylbenzo[g]pteridine-2,4(3H,10H)-dione

(lumiflavin).

Rifabutin

(Ph. Eur. monograph 1657)



C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

Action and use

72559-06-9

847

Plate TLC silica gel F₂₅₄ plate R.
Mobile phase acetone R, light petroleum R (23:77 V/V).
Application 10 µL.
Development Over 2/3 of the plate.
Drying In air.
Detection Expose the plate to iodine vapour for about 5 min, then spray with potassium iodide and starch solution R and allow to stand for 5 min.
Limit:
— impurity A: any spot corresponding to impurity A is not more intense than the spot in the chromatogram obtained with the reference solution (0.3 per cent).
Related substances
Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (a) Dissolve 50.0 mg of rifabutin CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.
Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.
Reference solution (c) Dissolve about 10 mg of rifabutin CRS in 2 mL of methanol R, add 1 mL of dilute sodium hydroxide solution R and allow to stand for about 4 min. Add 1 mL of dilute hydrochloric acid R and dilute to 50 mL with the mobile phase.
Column:
— size: l = 0.110 m, Ø = 4.6 mm,
— stationary phase: octylsilyl silica gel for chromatography R (5 µm).
Mobile phase Mix equal volumes of acetonitrile R and a 13.6 g/L solution of potassium dihydrogen phosphate R adjusted to pH 6.5 with dilute sodium hydroxide solution R.
Flow rate 1 mL/min.
Detection Spectrophotometer at 254 nm.
Injection 20 µL.
Run time 2.5 times the retention time of rifabutin.
Relative retention With reference to rifabutin (retention time = about 9 min): impurity E = about 0.5; impurity B = about 0.6; impurity D = about 0.9; impurity C = about 1.3.
System suitability: reference solution (c):
— resolution: minimum 2.0 between the second peak of the 3 peaks due to degradation products and the peak due to rifabutin.
Limits:
— any impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent); not more than 1 such peak has an area greater than half the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (3.0 per cent);
— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).
Water (2.5.12)
Maximum 2.5 per cent, determined on 0.200 g.
Sulfated ash (2.4.14)
Maximum 0.3 per cent, determined on 1.0 g.

Slightly soluble in water, soluble in methanol, slightly soluble in alcohol.

Solubility

Reddish-violet amorphous powder.

Appearance

CHARACTERS

96.0 per cent to 102.0 per cent (anhydrous substance).

Content

Semi-synthetic product derived from a fermentation product acetate.

furo[2,3':7,8]naphtho[1,2-d']imidazole-2,4'-piperidine-1,6-yl]tetrahydrospiro[9,4]-(epoxypentadeca[1,1,1,1,3]trienimin)-2H-heptamethyl-1'-(2-methylpropyl)-5,10,12-trioxo-3,5,9,10-6,18,20-trihydroxy-14-methoxy-7,9,15,17,19,21,25-(9S,12E,14S,15R,16S,17R,18R,19R,20S,21S,22E,24Z)-

DEFINITION

Ph Eur

Rifamycin antituberculosis drug.

72559-06-9

847

C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

72559-06-9

847

C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

72559-06-9

847

C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

72559-06-9

847

C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

72559-06-9

847

C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

72559-06-9

847

C₄₀H₆₂N₄O₁₁

Rifamycin antituberculosis drug.

72559-06-9

847

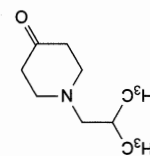
C₄₀H₆₂N₄O₁₁

ASSAY

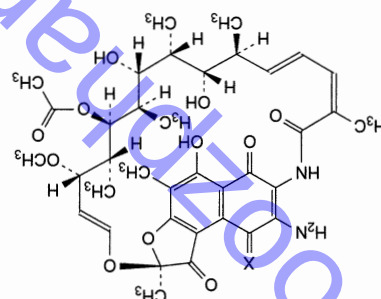
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of ribabutin.

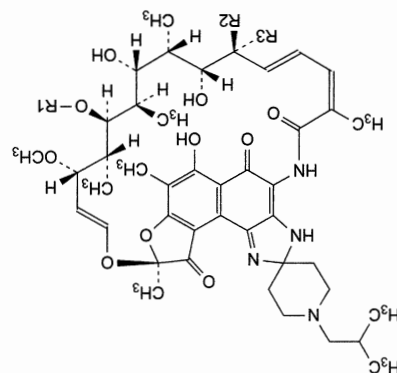
IMPURITIES

A. 1-(2-methylpropyl)piperidin-4-one,



B. X = O: 3-aminorifampicin S,

D. X = NH: 3-amino-4-imidorifampicin S,



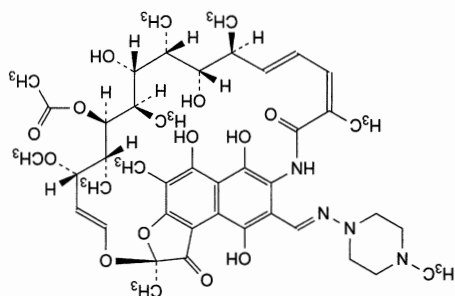
C. R1 = CO-CH₃, R2 + R3 = CH₂: 21,31-

E. R1 = R3 = H, R2 = CH₃: 16-deacetylribabutin.

Ph Eur

Rifampicin

(Ph. Eur. monograph 0052)



C₄₃H₅₈N₄O₁₂

823

13292-46-1

Action and use

Rifampicin antituberculosis drug.

Preparations

Rifampicin Capsules

Rifampicin Oral Suspension

Ph Eur

DEFINITION

(2S,12Z,14E,16S,17S,18R,19R,20R,21S,22R,23S,24E)-

5,6,9,17,19-Pentahydroxy-2,3-methoxy-2,4,12,16,18,20,22-heptamethyl-8-[[[(4-methylpiperazin-1-yl)imino]methyl]-1,11-dioxo-1,2-dihydro-2,7-

acetate.
(epoxypentadeca[1,11,13]trienimino)naphtho[2,1-b]furan-21-yl

Semisynthetic antibiotic obtained from rifamycin SV.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS**Appearance**

Reddish-brown or brownish-red, crystalline powder.

Solubility

Slightly soluble in water, soluble in methanol, slightly soluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50 mg in 50 mL of methanol R. Dilute 1 mL of this solution to 50 mL with phosphate buffer solution pH 7.4 R.

Spectral range 220-500 nm.

Absorption maxima At 237 nm, 254 nm, 334 nm and 475 nm.

Absorbance ratio A_{334}/A_{475} = about 1.75.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Mulls in liquid paraffin R.

Comparison rifampicin CRS.

C. Suspend about 25 mg in 25 mL of water R, shake for

5 min and filter. To 5 mL of the filtrate add 1 mL of a

100 g/L solution of ammonium persulfate R in phosphate buffer solution pH 7.4 R and shake for a few minutes. The colour

changes from orange-yellow to violet-red and no precipitate is formed.

TESTS**pH (2.2.3)**

4.5 to 6.5 for a 10 g/L suspension in carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29). Prepare the test solution and the reference solution immediately before use.

Solvent mixture To 10 volumes of a 210.1 g/L solution of citric acid R add 23 volumes of a 136.1 g/L solution of potassium dihydrogen phosphate R, 77 volumes of a 174.2 g/L solution of dipotassium hydrogen phosphate R, 250 volumes of acetonitrile R and 640 volumes of water R.

Test solution Dissolve 20.0 mg of the substance to be examined in acetonitrile R and dilute to 10.0 mL with the same solvent. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture.

Reference solution Dissolve 20.0 mg of rifampicin quinone CRS (impurity A) in acetonitrile R and dilute to 100.0 mL with the same solvent. To 1.0 mL of this solution add 1.0 mL of the test solution and dilute to 100.0 mL with the solvent

mixture.

Column:

size: $l = 0.12$ m, $\varnothing = 4.6$ mm;

stationary phase: octylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Mix 35 volumes of acetonitrile R and 65 volumes of a solution containing 0.1 per cent V/V of phosphoric acid R, 1.9 g/L of sodium perchlorate R, 5.9 g/L of citric acid R and 20.9 g/L of potassium dihydrogen phosphate R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Run time Twice the retention time of rifampicin.

System suitability: reference solution:

— **resolution:** minimum 4.0 between the peaks due to rifampicin and impurity A; if necessary, adjust the

concentration of acetonitrile in the mobile phase.

Limits:

— **impurity A:** not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with the reference solution (1.5 per cent);

— **any other impurity:** for each impurity, not more than the area of the peak due to rifampicin in the chromatogram

obtained with the reference solution (1.0 per cent);

— **sum of impurities other than A:** not more than 3.5 times the area of the peak due to rifampicin in the chromatogram

obtained with the reference solution (3.5 per cent);

— **disregard limit:** 0.05 times the area of the peak due to rifampicin in the chromatogram obtained with the

reference solution (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

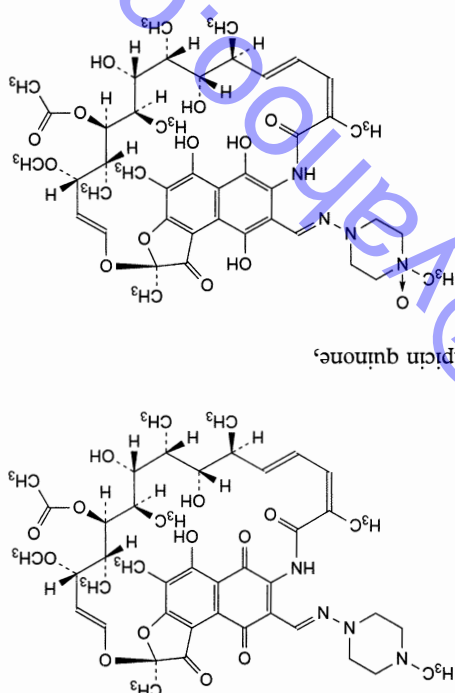
Maximum 1.0 per cent, determined on 1.000 g by drying at 80 °C at a pressure not exceeding 0.67 kPa for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 2.0 g.

ASSAY

Dissolve 0.100 g in methanol R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL



B. rifampicin N-oxide.

A. rifampicin quinone.

Specified impurities A
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B.

IMPURITIES

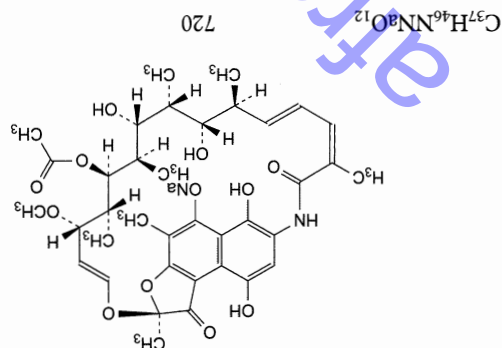
Under nitrogen in an airtight container, protected from light, at a temperature not exceeding 25 °C.

STORAGE

Calculate the content of $C_{43}H_{58}N_4O_{12}$, taking the specific absorbance to be 187.
liquid.
using phosphate buffer solution pH 7.4 R as the compensation absorbance (2.2.25) at the absorption maximum at 475 nm, with phosphate buffer solution pH 7.4 R. Measure the

Rifamycin Sodium

(Ph Eur monograph 0432)



Action and use

Rifamycin antituberculosis drug.

Ph Eur

DEFINITION

Sodium

(2S,12Z,14E,16S,17S,18R,19R,20R,21S,22R,23S,24E)-21-(epoxypentadeca[1,11,13]trienimino)naphtho[2,1-b]furan-5-yl-2,4,12,16,18,20,22-heptamethyl-1,11-dioxo-1,2-dihydro-2,7-(acetyloxy)-6,9,17,19-tetrahydroxy-2,3-methoxy-

Monosodium salt of rifamycin SV, obtained by chemical transformation of rifamycin B, which is produced during the growth of certain strains of *Amycolatopsis mediterranei*. Rifamycin SV may also be obtained directly from certain *A. mediterranei* mutants.

Potency

Minimum 900 IU/mg (anhydrous substance).

PRODUCTION

It is produced by methods of manufacture designed to minimise or eliminate substances lowering blood pressure. The manufacturing process is validated to demonstrate that the product, if tested, would comply with the following test.

Abnormal toxicity (2.6.9)

Inject into each mouse 4 mg dissolved in 0.5 mL of water for

injections R.

CHARACTERS

Appearance

Fine or slightly granular, red powder.

Solubility

Soluble in water, freely soluble in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of potassium bromide R.

Comparison rifamycin sodium CRS.

B. Suspend 70 mg of the substance to be examined in 0.5 mL of water R. Add 1.5 mL of methoxyphenylacetic reagent R to obtain a clear red solution. Cool in ice-water for 30 min. A precipitate is formed. Place in water at 20 °C and stir for 5 min. The precipitate does not disappear. Add 1 mL of dilute ammonia R1. The precipitate dissolves completely. Add 1 mL of ammonium carbonate solution R. No precipitate is formed.

TESTS

pH (2.2.3)

6.5 to 8.0.

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Absorbance (2.2.25)

Dissolve 20.0 mg in 5 mL of methanol R and dilute to 100.0 mL with freshly prepared phosphate buffer solution pH 7.0 R1 to which 1 g/L of ascorbic acid R has been added immediately before use. Dilute 5.0 mL of this solution to 50.0 mL with the same phosphate buffer solution containing ascorbic acid. Allow to stand for 30 min. The solution shows an absorption maximum at 445 nm. The specific absorbance at this absorption maximum is 190 to 210 (anhydrous substance).

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture Mix 50 volumes of acetonitrile R and 50 volumes of a 3.9 g/L solution of sodium dihydrogen phosphate R previously adjusted to pH 3.0 with phosphoric acid R.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture and dilute to 200.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture. (b) Dissolve 25 mg of the substance to be examined and 8 mg of rifamycin S CRS in the solvent mixture and dilute to 250.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: mix 10 volumes of acetonitrile R and 90 volumes of a 3.9 g/L solution of sodium dihydrogen phosphate R adjusted to pH 7.5 with dilute sodium hydroxide solution R;

— mobile phase B: mix 30 volumes of a 3.9 g/L solution of sodium dihydrogen phosphate R adjusted to pH 7.5 with dilute sodium hydroxide solution R and 70 volumes of acetonitrile R;

— temperature: minimum 20 °C;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 40	80 \rightarrow 20	20 \rightarrow 80
40 - 45	20	80
45 - 47	20 \rightarrow 80	80 \rightarrow 20

Flow rate 1 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Elution order Impurity A, rifamycin SV, impurity B.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to rifamycin SV and impurity B.

Limits:

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (2 per cent);

- **impurity A**: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
- **sum of impurities other than A and B**: not more than the area of the peak due to impurity B in the chromatogram obtained with reference solution (a) (2 per cent);
- **disregard limit**: 0.05 times the area of the peak due to impurity B in the chromatogram obtained with reference solution (a) (0.1 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.
2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

12.0 per cent to 17.0 per cent, determined on 0.200 g.

Bacterial endotoxins (2.6.14)

Less than 0.50 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

ASSAY

Carry out the microbiological assay of antibiotics (2.7.2).

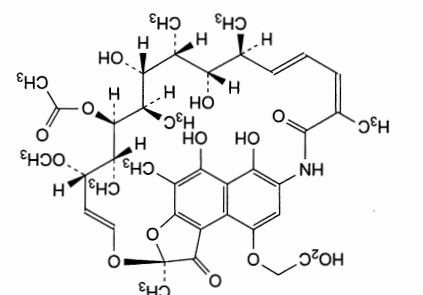
STORAGE

In an airtight container, protected from light at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

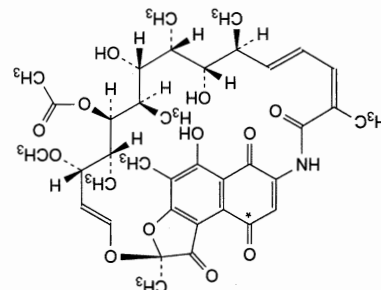
IMPURITIES

Specified impurities A, B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



A. rifamycin B,

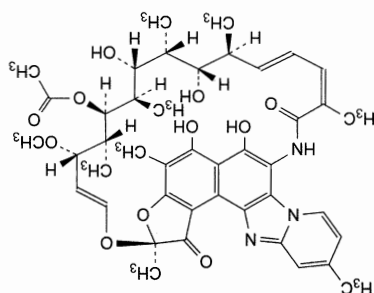


and epimer at C*

B. rifamycin S,

Rifaximin

(Ph. Eur. monograph 2362)



C₄₃H₅₁N₃O₁₁

786

80621-81-4

Action and use
Antibacterial; treatment of infective diarrhoea.

DEFINITION

(2S,16Z,18E,20S,21S,22R,23R,24R,25S,26R,27S,28E) 5,6,21,23-Tetrahydroxy-27-methoxy-2,4,11,16,20,22,24,26-octamethyl-1,1,5-dioxo-1,2-dihydro-2,7-(epoxypentadeca[1,1,1,3]trieniminol)[1]benzofuro[4,5-e]pyrido[1,2-a]benzimidazol-25-yl acetate.
Semi-synthetic product derived from a fermentation product. 97.0 per cent to 102.0 per cent (anhydrous substance).

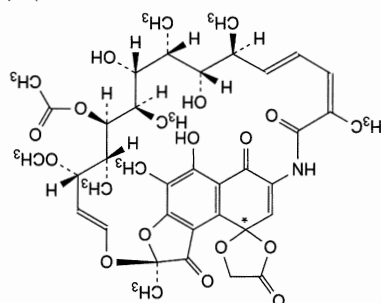
CHARACTERS

Appearance
Red-orange, hygroscopic, crystalline powder.
Solubility
Practically insoluble in water, soluble in acetone and in methanol.
IDENTIFICATION
It shows polymorphism (5.9).
Infrared absorption spectrophotometry (2.2.24).
Comparison rifaximin CRS.
If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in ethanol R, evaporate to dryness and record new spectra using the residues.

Ph Eur

C. rifamycin O.

and epimer at C*



TESTS**Related substances**

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R, water R (40:60 V/V).

Test solution (a) Dissolve 0.100 g of the substance to be examined in 8 mL of acetonitrile R and dilute to 20 mL with water R.

Test solution (b) Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 50.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of rifaximin for system suitability CRS (containing impurity H) in 4 mL of the solvent mixture.

Reference solution (c) Dissolve 40.0 mg of rifaximin CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 5.0 mL of the solution to 50.0 mL with the solvent mixture.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase Mix 37 volumes of a 3.16 g/L solution of ammonium formate R adjusted to pH 7.2 with dilute ammonia R1 and 63 volumes of a mixture of equal volumes of acetonitrile R and methanol R.

Flow rate 1.4 mL/min.

Detection Spectrophotometer at 276 nm.

Injection 20 μ L of test solution (a) and reference solutions (a) and (b).

Run time 3 times the retention time of rifaximin.

Relative retention With reference to rifaximin (retention time = about 12 min): impurities D and H = about 0.7.

System suitability: reference solution (b):

— **resolution:** minimum 3.0 between the peaks due to impurities D + H and rifaximin.

Limits:

— **sum of impurities D and H:** not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— **unspecified impurities:** for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **total:** not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— **disregard limit:** 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 20 mL of lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Maximum 4.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{43}H_{51}N_3O_{11}$ using the chromatogram obtained with reference solution (c) and the declared content of rifaximin CRS.

STORAGE

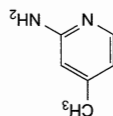
In an airtight container, protected from light.

IMPURITIES

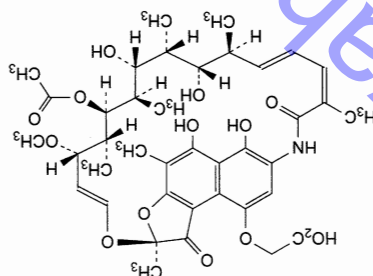
Specified impurities D, H

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

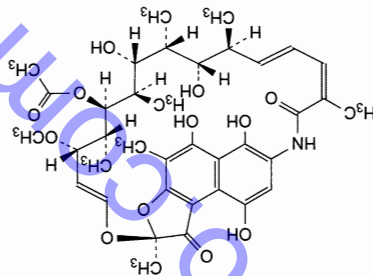
Control of impurities in substances for pharmaceutical use: A, B, C, E, F, G.



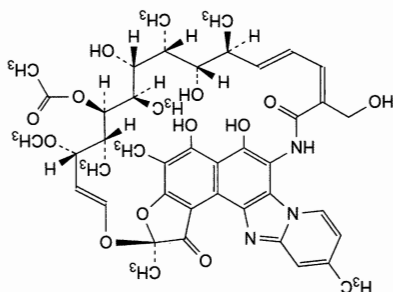
A. 4-methylpyridin-2-amine,



B. rifamycin B,



C. rifamycin SV,



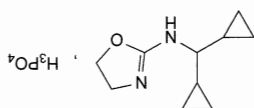
H. (2S,16Z,18E,20S,21S,22R,23R,24R,25S,26R,27S,28E)-5,6,21,23-tetrahydroxy-16-(hydroxymethyl)-27-methoxy-2,4,11,20,22,24,26-heptamethyl-1,15-dioxo-1,2-dihydro-7-(16-(hydroxymethyl)rifaximin).

Ph Eur



Rilmenidine Dihydrogen Phosphate

(Ph. Eur. monograph 2020)



85409-38-7

278.2

C₁₀H₁₉N₂O₅P

Action and use

Antihypertensive.

Ph Eur

DEFINITION

N-(Dicyclopropylmethyl)-4,5-dihydro-oxazol-2-amine dihydrogen phosphate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, slightly soluble in alcohol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of rilmenidine dihydrogen phosphate.

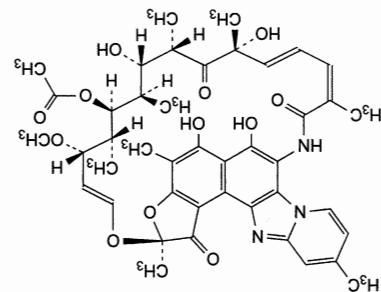
B. Dissolve 10 mg in water R and dilute to 1 mL with the same solvent. The solution gives reaction (b) of phosphates (2.3.1).

TESTS

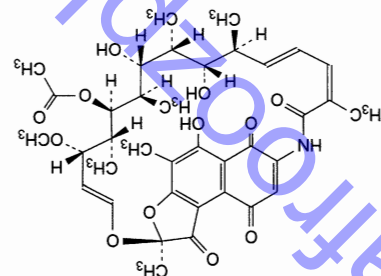
Related substances

Liquid chromatography (2.2.29).

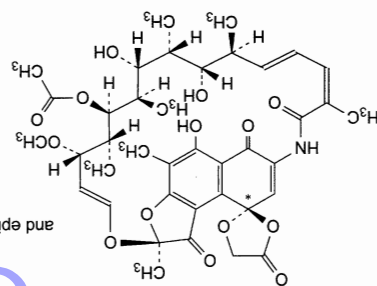
Test solution Dissolve 60.0 mg of the substance to be examined in water R and dilute to 20.0 mL with the same solvent.



D. rifaximin Y,



E. rifamycin S,



F. rifamycin O,

G. (2S,17Z,16Z,18E,20S,21S,22R,23R,24R,25S,26R,27S,28E)-5,21,23-trihydroxy-27-methoxy-2,4,11,16,20,22,24,26-octamethyl-1,6,15-dioxo-1,2,6,7-tetrahydro-2,7-(epoxypentadeca[1,11,13]trienonitro)[1]benzofuro[4,5-e]pyrido[1,2-a]benzimidazol-25-yl acetate (6-O,14-dihydrodrorifaximin).

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with water R and dilute 10.0 mL of this solution to 50.0 mL with the same solvent.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 20.0 mL with water R.

Reference solution (c) Dissolve 15.0 mg of rilmenidine for system suitability CRS in water R and dilute to 5.0 mL with the same solvent.

Column:

— size: $l = 0.15$ m, $\varnothing = 3$ mm,

— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5 μ m) with a pore size of 10 nm and a carbon loading of 25 per cent,

— temperature: 40 °C.

Mobile phase:

— mobile phase A: dissolve 3 g of sodium heptanesulfonate R in water R and dilute to 860 mL with the same solvent;

add 130 mL of methanol R₂, 10 mL of tetrahydrofuran for chromatography R and 1.0 mL of phosphoric acid R₃

— mobile phase B: dissolve 3 g of sodium heptanesulfonate R in water R and dilute to 600 mL with the same solvent;

add 350 mL of acetonitrile for chromatography R₃, 50 mL of tetrahydrofuran for chromatography R and 1.0 mL of phosphoric acid R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 14	100 → 0	0 → 100
14 - 15	0 → 100	100 → 0
15 - 30	100	0

Flow rate 1 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L.

Relative retention With reference to rilmenidine (retention time = about 13 min): impurity A = about 0.6; impurity B = about 0.9; impurity C = about 1.4.

With these conditions the inflexion of the baseline, corresponding to the beginning of the gradient, appears on the recorder after a minimum time t of 5 min. If this is not the case ($t > 5$ min) modify the chromatographic sequence by adding an isocratic elution with 100 per cent of mobile phase A for a time corresponding to (5- t) min before the linear gradient.

System suitability: reference solution (c):

— peak-to-valley ratio: minimum 3, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to rilmenidine.

Limits:

— any impurity: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent),

— disregard limit: area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

IDENTIFICATION

Solubility

White or almost white, crystalline powder.

Soluble in water, practically insoluble in methanol. It dissolves in dilute solutions of alkali hydroxides and mineral acids.

CHARACTERS

Content

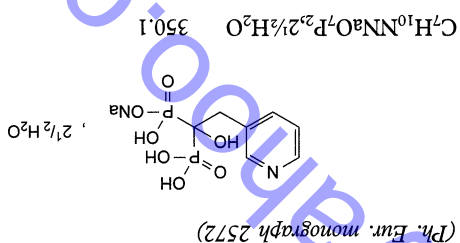
99.0 per cent to 101.0 per cent (anhydrous substance).

DEFINITION

Sodium hydrogen [1-hydroxy-1-phosphono-2-(pyridin-3-yl)ethyl]phosphonate hemipentahydrate.

Action and use

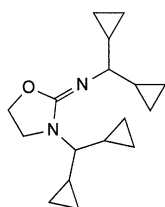
Bisphosphonate; treatment of osteoporosis, Paget's disease.



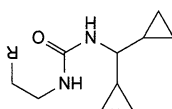
Risedronate Sodium 2.5-Hydrate



C, N,3-bis(dicyclopropylmethyl)oxazolidin-2-imine.



A, R = OH: 1-(dicyclopropylmethyl)-3-(2-hydroxyethyl)urea, B, R = Cl: 1-(2-chloroethyl)-3-(dicyclopropylmethyl)urea,



Specified impurities: A, B, C.

IMPURITIES

$C_{10}H_{19}N_2O_5P$.

1 mL of 0.1 M perchloric acid is equivalent to 27.82 mg of potentiometrically (2.2.20).

Dissolve 0.200 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point

ASSAY

2 h.

an oven in vacuo at 50 °C over diphenylpicrylhydrazide R for

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

Comparison risedronate sodium 2.5-hydrate CRS.
 B. It gives reaction (a) of sodium (2.3.1). Dissolution of the substance to be examined is achieved after the addition of the 150 g/L solution of potassium carbonate R.

C. Water (see Tests).

TESTS

pH (2.2.3)

4.0 to 5.0.
 Dissolve 0.10 g in carbon dioxide-free water R with the aid of an ultrasonic bath and dilute to 10 mL with the same solvent.

Related substances

A. Liquid chromatography (2.2.29).
Buffer solution Dissolve 0.410 g of sodium edetate R, 1.7 g of dipotassium hydrogen phosphate R and 1.7 g of tetrabutylammonium dihydrogen phosphate R in 900 mL of water R, adjust to pH 7.5 with 1 M sodium hydroxide and dilute to 1000 mL with water R.

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase by gentle swirling and heating for 5-10 min and dilute to 20.0 mL with the mobile phase.

Reference solution (a) To 2.0 mL of the test solution add 5 mg of risedronate impurity B CRS and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Blank solution Dissolve 100 mg of sodium chloride R in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
 — stationary phase: octadecylsilyl silica gel for chromatography R (3 μ m);
 — temperature: 40 °C.

Mobile phase acetonitrile R, buffer solution (10:90 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 263 nm.

Injection 20 μ L.

Run time Twice the retention time of risedronate.

Relative retention With reference to risedronate (retention time = about 17 min): impurity E = about 0.95.

System suitability: reference solution (a):

— resolution: minimum 3.0 between the peaks due to impurity E and risedronate.

Limits:

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent); disregard any peak due to the blank.

B. Liquid chromatography (2.2.29) as described in test A for related substances, with the following modifications.

Reference solution (a) Dissolve 5.0 mg of the substance to be examined in 50.0 mL of the mobile phase by gentle swirling and heating for 5-10 min, using an ultrasonic bath if necessary.

D, E.

Control of impurities in substances for pharmaceutical use: B, C, impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these by the general monograph *Substances for pharmaceutical use* acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of **Other detectable impurities** (the following substances would, if specified impurities A.

IMPURITIES

$C_7H_{10}NNaO_7P_2$.

1 mL of 0.1 M sodium hydroxide is equivalent to 15.26 mg of end-point potentiometrically (2.2.20).

Titrate with 0.1 M sodium hydroxide, determining the Dissolve 0.125 g in 50 mL of carbon dioxide-free water R.

ASSAY

11.9 per cent to 13.9 per cent, determined on 0.100 g.

Water (2.5.12)

using 1 mL of lead standard solution (10 ppm Pb) R. 0.500 g complies with test H. Prepare the reference solution

Solvent water R.

Maximum 20 ppm.

Heavy metals (2.4.8)

the blank.

solution (c) (0.05 per cent); disregard any peak due to in the chromatogram obtained with reference

— **disregard limit:** 0.5 times the area of the principal peak solution (c) (0.2 per cent);

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);

— **impurity A:** not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);

Limits:

— **resolution:** minimum 10.0 between the peaks due to risedronate and impurity A.

System suitability Reference solution (d): time = about 4 min: impurity A = about 2.2.

Relative retention With reference to risedronate (retention solution (b) to identify the peak due to impurity A.

Identification Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.

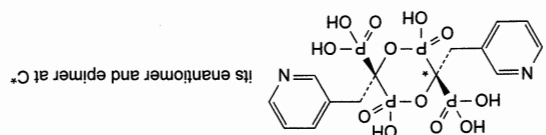
Run time 8 times the retention time of risedronate. solutions (b), (c) and (d).

Injection 10 μ L of the test solution and reference and 1.0 mL of reference solution (b) and dilute to 20.0 mL with the mobile phase.

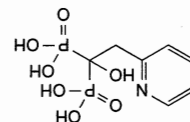
Reference solution (a) Mix 1.0 mL of reference solution (a) to 20.0 mL with the mobile phase.

Reference solution (c) Dilute 0.5 mL of reference solution (b) and dilute to 50.0 mL with the same solvent.

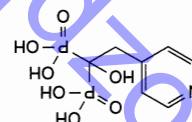
Reference solution (b) Dissolve 5.0 mg of risedronate impurity A CRS in the mobile phase by gentle swirling and heating for 5-10 min, using an ultrasonic bath if necessary.



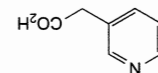
A. [(3*R*,6*R*S and 3*R*,6*S*-*meso*)-2,5-dihydroxy-2,5-dioxo-3,6-bis[(pyridin-3-yl)methyl]-1,4,2λ³,5λ⁵-dioxadiphosphinane-3,6-diyl]bis(phosphonic acid),



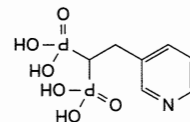
B. [1-hydroxy-2-(pyridin-2-yl)ethylidene]bis(phosphonic acid),



C. [1-hydroxy-2-(pyridin-4-yl)ethylidene]bis(phosphonic acid),



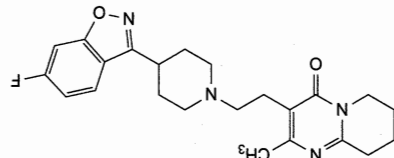
D. 2-(pyridin-3-yl)acetic acid,



E. [2-(pyridin-3-yl)ethylidene]bis(phosphonic acid).



Risperidone
(Ph. Eur. monograph 1559)



C₂₃H₂₇N₄O₂ 410.5 106266-06-2

Action and use
Dopamine D₂ receptor antagonist; serotonin 5HT₂ receptor antagonist; neuroleptic.

Preparations

Risperidone Oral Solution
Risperidone Tablets
Dispersible Risperidone Tablets

DEFINITION
3-[2-[4-(6-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4*H*-pyrido[1,2-*a*]pyrimidin-4-one.

Content
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in ethanol (96 per cent).

It dissolves in dilute acid solutions.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison *risperidone* CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance

separately in *acetone* R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.1 g in a 7.5 g/L solution of *tartaric acid* R and dilute to 100 mL with the same acid solution.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in *methanol* R and dilute to 10.0 mL with the same

solvent.

Reference solution (a) Dissolve 10 mg of *risperidone* for system suitability CRS (containing impurities A, B, C, D and E) in 1.0 mL of *methanol* R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with *methanol* R. Dilute 5.0 mL of this solution to 25.0 mL with *methanol* R.

Reference solution (c) Dissolve the contents of a vial of *risperidone* impurity K CRS in 1.0 mL of *methanol* R.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (3 μ m).

Mobile phase:

— mobile phase A: 5 g/L solution of *ammonium acetate* R;

— mobile phase B: *methanol* R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	70	30
2 - 17	70 \rightarrow 30	30 \rightarrow 70
17 - 22	30	70

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with *risperidone* for system suitability CRS and the

chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D and E; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity K.

Relative retention With reference to risperidone (retention time = about 12 min): impurity A = about 0.7;

impurity B = about 0.75; impurity C = about 0.8;

impurity K = about 0.9; impurity D = about 0.94;

impurity E = about 1.1.

System suitability: reference solution (a):

the chromatogram obtained is similar to the

chromatogram supplied with risperidone for system

suitability CRS;

peak-to-valley ratio: minimum 1.5, where H_p = height

above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

risperidone.

Limits:

impurities A, B, C, D, E: for each impurity, not more than

the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent);

impurity K: not more than 0.75 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

total: not more than 1.5 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.3 per cent);

disregard limit: 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum

crucible.

ASSAY

Dissolve 0.160 g in 70 mL of a mixture of 1 volume of

anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R

and titrate with 0.1 M perchloric acid. Determine the

end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 20.53 mg

of $C_{23}H_{27}FN_4O_2$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C, D, E, K

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

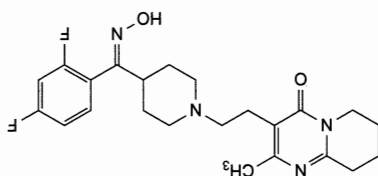
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

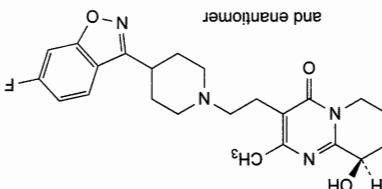
Control of impurities in substances for pharmaceutical use: F, H,

I, J, L, M.

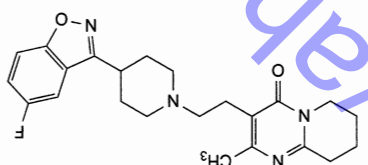
A. 3-[2-[4-[(E)-(2,4-difluorophenyl)(hydroxyimino)methyl]piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



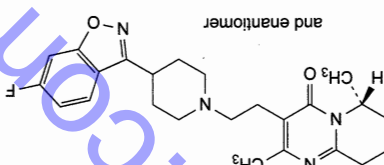
B. 3-[2-[4-[(Z)-(2,4-difluorophenyl)(hydroxyimino)methyl]piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



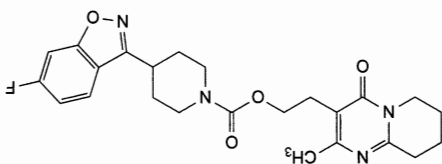
C. (9R,3S)-3-[2-[4-(6-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-yl]ethyl]-9-hydroxy-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



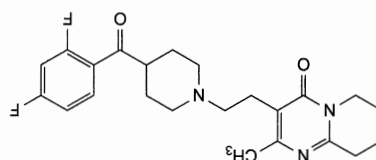
D. 3-[2-[4-(5-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



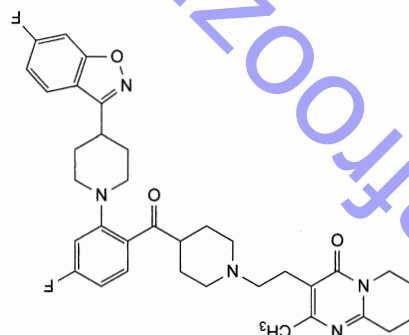
E. (6R,3S)-3-[2-[4-(6-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-yl]ethyl]-2,6-dimethyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



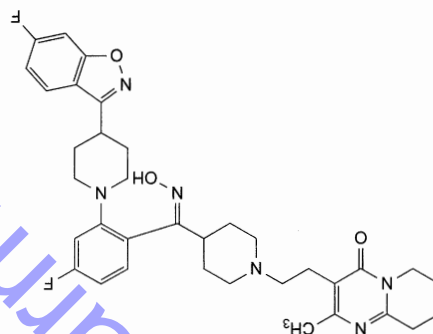
F. 2-[2-methyl-4-oxo-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-3-yl]ethyl 4-(6-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-carboxylate,



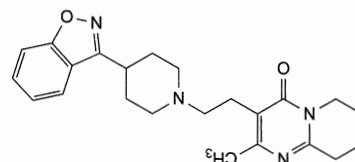
H. 3-[2-[4-(2,4-difluorobenzoyl)piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



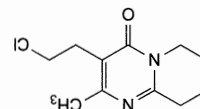
I. 3-[2-[4-[4-fluoro-2-[4-(6-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-yl]benzoyl]piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,



J. 3-[2-[4-(Z)-[4-fluoro-2-[4-(6-fluoro-1,2-benzisoxazol-3-yl)piperidin-1-yl]phenyl](hydroxyimino)methyl]piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one,

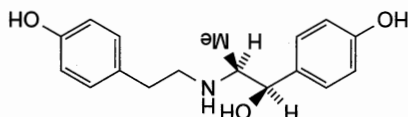


K. 3-[2-[4-(1,2-benzisoxazol-3-yl)piperidin-1-yl]ethyl]-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one (desfluoro ritodrine),



L. 3-(2-chloroethyl)-2-methyl-6,7,8,9-tetrahydro-4H-pyrido[1,2-a]pyrimidin-4-one (piperidopyrimidinone intermediate),

Ritodrine Hydrochloride



and enantiomer

$C_{17}H_{21}NO_3 \cdot HCl$

323.8

23239-51-2

Ritodrine Tablets

Ritodrine Injection

Preparations

Beta₂-adrenoceptor agonist; bronchodilator.

Action and use

Ritodrine Hydrochloride is *rac*-4-[(1*R*,2*S*)-1-hydroxy-2-[(2-(4-hydroxyphenyl)ethyl)amino]propyl]phenol hydrochloride. It contains not less than 97.0% and not more than 103.0% of $C_{17}H_{21}NO_3 \cdot HCl$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder.

Freely soluble in water; soluble in absolute ethanol; practically insoluble in acetone and in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of ritodrine hydrochloride (RS 313).

B. A 1% w/v solution yields reaction A characteristic of chlorides, Appendix VI.

TESTS

Acidity
pH of a 2% w/v solution, 4.5 to 6.0, Appendix V L.

Heavy metals
12 mL of a 10.0% w/v solution complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (2 ppm Pb) to prepare the standard (20 ppm).

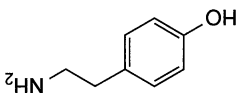
Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions. Solution (1) contains 0.10% w/v of the substance being examined in the mobile phase. For solution (2) dilute 1 volume of solution (1) to 100 volumes with the mobile phase. For solution (3) dissolve about 20 mg of the substance being examined in 50 mL of the mobile phase, add 5.6 mL of sulfuric acid and sufficient of the mobile phase to produce 100 mL, mix and heat at a temperature of 85° for 2 hours. Cool to room temperature, carefully mix 10 mL of the cooled solution with 8 mL of a 10% w/v solution of sodium hydroxide and allow to cool (generation of *threo*-diastereoisomer).

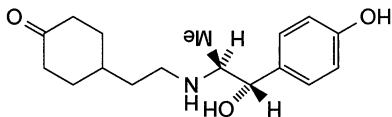
STORAGE

Ritodrine Hydrochloride should be kept in an airtight container.

IMPURITIES

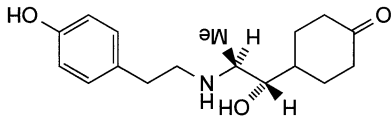


A. tyramine,



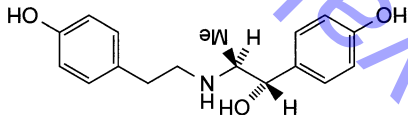
and enantiomer

B. *rac*-4-[(1*R*,2*R*)-2-{[1-hydroxy-1-(4-hydroxyphenyl)propan-2-yl]amino}cyclohexan-1-one (hexahydroketone I),



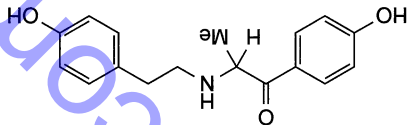
and enantiomer

C. *rac*-4-[(1*R*,2*R*)-1-hydroxy-2-{2-(4-hydroxyphenyl)ethyl]amino}propyl]cyclohexan-1-one (hexahydroketone II),



and enantiomer

D. *rac*-4-[(1*R*,2*R*)-1-hydroxy-2-{2-(4-hydroxyphenyl)ethyl]amino}propyl]phenol (threo-diastereoisomer),



E. *rac*-1-(4-hydroxyphenyl)-3-{2-(4-hydroxyphenyl)ethyl]amino}propan-1-one (amino ketone).

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with end-capped octylsilyl silica gel for chromatography (7 µm) (Zorbax C8 is suitable), (b) as the mobile phase with a flow rate of 2 mL per minute a mixture of a solution containing 6.6 g of ammonium hydrogen orthophosphate and 1.1 g of sodium heptanesulfonate in 700 mL of water and 300 mL of methanol; the mixture adjusted to pH 3.0 with an 85% v/v solution of orthophosphoric acid and (c) a detection wavelength of 214 nm.

The test is not valid unless, in the chromatogram obtained with solution (3), the resolution factor between the two principal peaks (ritodrine and the threo-diastereoisomer) is at least 1.5.

In the chromatogram obtained with solution (1), identify any peaks corresponding to tyramine (relative retention time to ritodrine, 0.3), hexahydroketone II (0.65), hexahydroketone I (0.85), threo-diastereoisomer (1.15) and amino ketone (2.3). In the chromatogram obtained with solution (1), (a) the area of any peak corresponding to hexahydroketone II is not greater than 0.1 times the area of the principal peak in the chromatogram obtained with solution (2) (0.3% [response relative to ritodrine 0.3]), (b) the areas of any peaks corresponding to tyramine and amino ketone are not greater than 0.2 times the area of the principal peak in the chromatogram obtained with solution (2) (0.2% each), (c) the area of any peak corresponding to hexahydroketone I is not greater than 0.2 times the area of the principal peak in the chromatogram obtained with solution (2) (0.4% [response relative to ritodrine 0.5]), (d) the area of any peak corresponding to the threo-diastereoisomer is not greater than 0.4 times the area of the principal peak in the chromatogram obtained with solution (2) (0.4%) and (e) the area of any other secondary peak is not greater than 0.1 times the area of the principal peak in the chromatogram obtained with solution (2) (0.1%).

Loss on drying

When dried at 105° for 2 hours, loses not more than 1.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.2%, Appendix IX A.

ASSAY

Carry out the method for liquid chromatography,

Appendix III D, using the following solutions. Solution (1) contains 0.02% w/v of the substance being examined in the mobile phase. Solution (2) contains 0.02% w/v of ritodrine hydrochloride BPCRS in the mobile phase. For solution (3) dissolve about 20 mg of the substance being examined in 50 mL of the mobile phase, add 5.6 mL of sulfuric acid and sufficient of the mobile phase to produce 100 mL, mix and heat at a temperature of 85° for 2 hours. Cool to room temperature, carefully mix 10 mL of the cooled solution with 8 mL of a 10% w/v solution of sodium hydroxide and allow to cool (generation of threo-diastereoisomer).

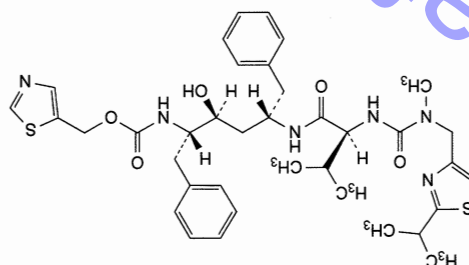
The chromatographic procedure described under Related substances may be used.

The test is not valid unless, in the chromatogram obtained with solution (3), the resolution factor between the two principal peaks (ritodrine and the threo-diastereoisomer) is at least 1.5.

Calculate the content of C₁₇H₂₁NO₃·HCl in the substance being examined from the chromatograms obtained and from the declared content of C₁₇H₂₁NO₃·HCl in ritodrine hydrochloride BPCRS.

Ritonavir

(Ph. Eur. monograph 2136)

C₃₇H₄₈N₆O₅S₂ 721 155213-67-5

Action and use

Protease inhibitor; antiviral (HIV).

DEFINITION

Thiazol-5-ylmethyl [(1*S*,2*S*,4*S*)-1-benzyl-2-hydroxy-4-[[[(2*S*)-3-methyl-2-[[methyl]-(2-(1-methylthiazol-4-yl)methyl]carbamoyl]amino]butanoyl]amino]-5-phenylpentyl]carbamate.

PRODUCTION

The production method is validated to demonstrate suitable enantiomeric purity of the final product.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methanol and in methylene chloride, very slightly soluble in acetonitrile.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison with reference CRS.

If the spectra obtained in the solid state show differences dissolve the substance to be examined and the reference substance separately in methylene chloride *R*, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mix equal volumes of acetonitrile *R* and a 4.1 g/L solution of potassium dihydrogen phosphate *R*.

Test solution (a) Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Sonicate if necessary.

Test solution (b) Dilute 5.0 mL of test solution (a) to 10.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5.0 mg of ritonavir for peak identification CRS (containing impurities E, F, L, O and T) in the solvent mixture and dilute to 5.0 mL with the solvent mixture. Sonicate if necessary.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dissolve 10.0 mg of ritonavir CRS in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Sonicate if necessary. Dilute 5.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

stationary phase: end-capped butylsilyl silica gel for chromatography *R* (3 μ m);

temperature: 60 °C.

Mobile phase:

mobile phase A: mix 5 volumes of butanol *R*, 8 volumes of tetrahydrofuran *R*, 18 volumes of acetonitrile *R* and 69 volumes of a 4.1 g/L solution of potassium dihydrogen phosphate *R* filtered through a 0.45 μ m nylon membrane;

mobile phase B: mix 5 volumes of butanol *R*, 8 volumes of tetrahydrofuran *R*, 40 volumes of a 4.1 g/L solution of potassium dihydrogen phosphate *R* filtered through a 0.45 μ m nylon membrane and 47 volumes of acetonitrile *R*;

Flow rate 1.0 mL/min.
Detection Spectrophotometer at 240 nm.
Injection 50 μ L of test solution (a) and reference solutions (a) and (b).

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 60	100	0
60 - 120	100 \rightarrow 0	0 \rightarrow 100
120 - 120.1	0 \rightarrow 100	100 \rightarrow 0
120.1 - 155	100	0

Identification of impurities Use the chromatogram supplied with ritonavir for peak identification CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities E, F, L, O and T. **Relative retention** With reference to ritonavir (retention time = about 3.4 min): impurity E = about 0.39; impurity F = about 0.40; impurity L = about 0.8; impurity O = about 1.1; impurity T = about 2.6. **System suitability:** reference solution (a): H_p = height above the baseline of the peak due to impurity E and H_p = height above the baseline of the peak due to impurity L; curve separating this peak from the peak due to impurity F.

Limits:

correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity F = 1.4; impurity L = 1.9; impurity T = 1.4; **impurities E, O:** for each impurity, not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent); **impurity T:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent); **impurities F, L:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent); **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 1.0 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c). Calculate the percentage content of $C_{37}H_{48}N_6O_{5.2}$ from the declared content of *ribovir CRS*.

STORAGE

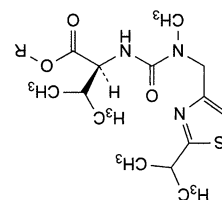
Protected from light.

IMPURITIES

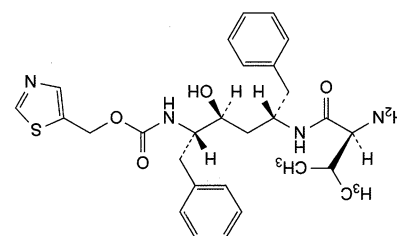
Specified injuries E, F, L, O, T.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

C, D, G, H, I, J, K, M, N, P, Q, R, S, U.

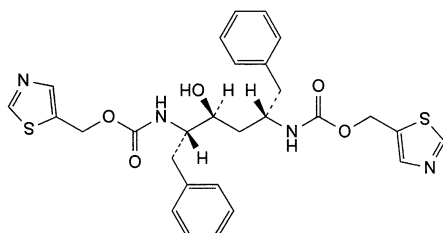


M. R = CH₃-CH(CH₃)₂: 2-methylpropyl (2S)-3-methyl-2-[[[methyl(1-methylethyl)thiazol-4-yl]methoxy]amino]butanoic acid,
H: (2S)-3-methyl-2-[[[methyl(1-methylethyl)thiazol-4-yl]methoxy]amino]butanoic acid,
Y: [methyl(carbamoyl)]amino]butanoate,

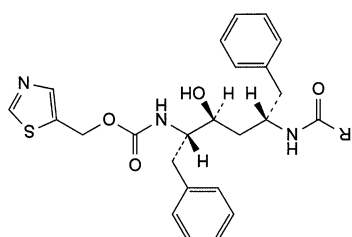


B. thiazol-5-ylmethyl [(1*S*,2*S*,4*S*)-4-[[[(2*S*)-2-amino-3-methylbutanoyl]amino]-1-benzyl-2-hydroxy-5-phenylpentyl]carbamate,

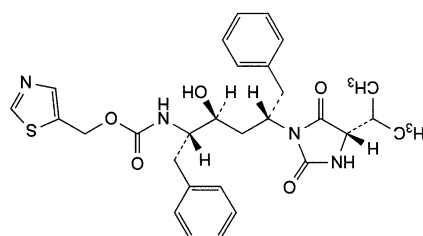
D, thiazol-5-ylmethyl [(1S,2S,4S)-1-benzyl-2-hydroxy-5-phenyl-4-[[[(thiazol-5-ylmethoxy)carbonyl]amino]pentyl]carbamate,



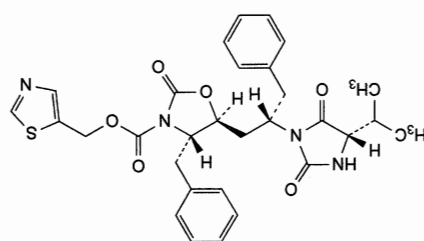
C, R = CH₃: thiazol-5-ylmethyl [1*S*,2*S*,4*S*]-4-(acetylamino)-1-benzyl-2-hydroxy-5-phenylpentyl]carbamate, J, R = O-C(CH₃)₃: thiazol-5-ylmethyl [1*S*,2*S*,4*S*]-1-benzyl-4-[[1,1-dimethylethoxy]carbonyl]amino]-2-hydroxy-5-phenylpentyl]carbamate, K, R = O-CH₂-CH(CH₃)₂: thiazol-5-ylmethyl [1*S*,2*S*,4*S*]-1-benzyl-2-hydroxy-4-[[2-methylpropoxy]carbonyl]amino]-5-phenylpentyl]carbamate,



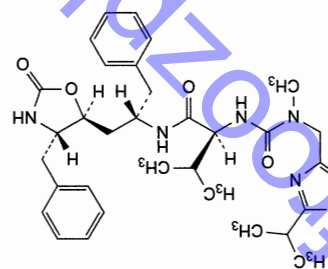
E. R = OH, R' = CH₃; thiazol-5-ylmethyl [(1S,2S,4S)-1-benzyl-2-hydroxy-4-[[[(2S)-2-(1-hydroxy-1-methylethyl)thiazol-4-yl]methyl]methoxy]amino]-3-methylbutanoyl]amino]-5-phenylpentyl]carbamate,
G. R = OOH, R' = CH₃; thiazol-5-ylmethyl [(1S,2S,4S)-1-benzyl-4-[[[(2S)-2-(1-hydroperoxy-1-methylethyl)thiazol-4-yl]methyl]methoxy]amino]-3-methylbutanoyl]amino]-2-hydroxy-5-phenylpentyl]carbamate,
I. R = H; thiazol-5-ylmethyl [(1S,2S,4S)-1-benzyl-4-[[[(2S)-2-(2-ethylthiazol-4-yl)methyl]methoxy]amino]-3-methylbutanoyl]amino]-2-hydroxy-5-phenylpentyl]carbamate,



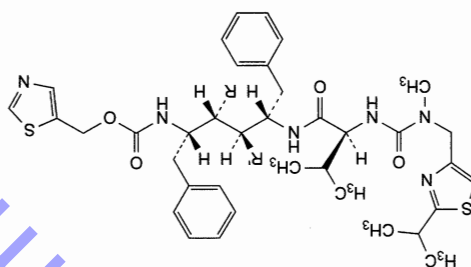
F, thiazol-5-ylmethyl [(1*S*,2*S*,4*S*)-1-benzyl-4-[[[(2*S*)-1-benzyl-2-hydroxy-4-[(4*S*)-4-(1-methylethyl)-2,5-dioximidazolidin-1-yl]-5-phenylpentyl]carbamate,



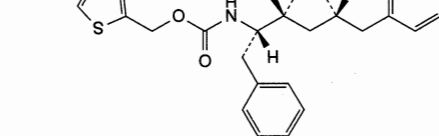
H. thiazol-5-ylmethyl] (4S,5S)-4-benzyl-5-[(2S)-2-[(4S)-4- (1-methylethyl)-2,5-dioximidazolidin-1-yl]-3-phenylpropyl]- 2-oxooxazolidine-3-carboxylate,



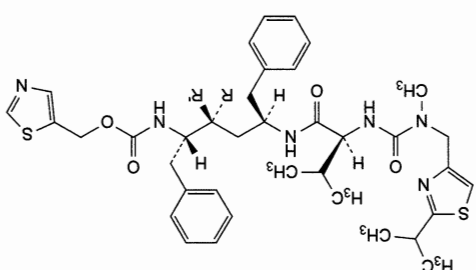
L. (4S,5S)-4-benzyl-5-[(2S)-2-[(2S)-3-methyl-2-[[methyl]2- (1-methylethyl) thiazol-4-yl]methyl]carbamoyl]amino]butanoyl]amino]-3-phenylpropyl] oxazolidin-2-one,



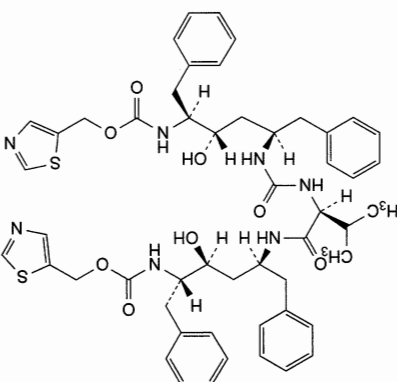
N. R = OH, R' = H; thiazol-5-ylmethyl] [(1S,3S,4S)-1-benzyl-3-hydroxy-4-[[[(2S)-3-methyl-2-[[methyl]2- (1-methylethyl) thiazol-4-yl]methyl]carbamoyl]amino]butanoyl]amino]-5-phenylpropyl]carbamate,



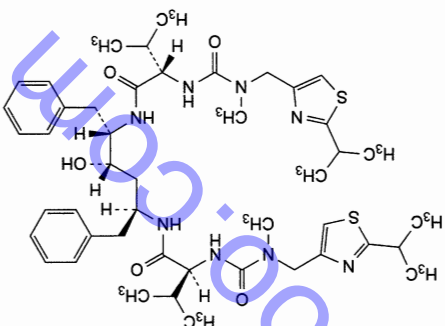
P. bis(thiazol-5-ylmethyl) [carbonylbis[imino[(2S,3S,5S)-3-hydroxy-1,6-diphenylhexane-5,2-diy]]]]dicarbamate,



Q. R = OH, R' = H; thiazol-5-ylmethyl] [(1S,2R,4R)-1-benzyl-2-hydroxy-4-[[[(2S)-3-methyl-2-[[methyl]2- (1-methylethyl) thiazol-4-yl]methyl]carbamoyl]amino]butanoyl]amino]-5-phenylpropyl]carbamate,



S. thiazol-5-ylmethyl] [(1S,2S,4S)-1-benzyl-4-[[[(2S)-2-[[[(1S,3S,4S)-1-benzyl-3-hydroxy-5-phenyl-4-[[[(thiazol-5-yl)methoxy]carbonyl]amino]penyl]carbamoyl]amino]-3-methylbutanoyl]amino]-2-hydroxy-5-phenylpropyl]carbamate,



T. (2S)-N-[(1S,2S,4S)-1-benzyl-2-hydroxy-4-[[[(2S)-3-methyl-2-[[methyl]2-[(1-methylethyl) thiazol-4-yl]methyl]carbamoyl]amino]butanoyl]amino]-5-phenylpropyl]carbamoyl]amino]butanamide,

vigorously for 2 min. Allow the layers to separate. Filter the upper organic layer through anhydrous sodium sulfate R. Evaporate the filtrate under reduced pressure at a temperature not exceeding 60 °C to obtain a residue. Record the reference spectrum using this residue.

C. Enantiomeric purity (see Tests).

TESTS

Enantiomeric purity

Solution A Solution containing 1.78 g/L of disodium hydrogen phosphate dihydrate R and 1.38 g/L of sodium dihydrogen phosphate monohydrate R. Adjust to pH 6.0 with phosphoric acid R.

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 5.0 mL of the solution to 20.0 mL

Reference solution (a) Dissolve 2.0 mg of rivastigmine impurity D CRS in the mobile phase and dilute to 200.0 mL with the mobile phase.

Reference solution (b) Dissolve 1 mg of rivastigmine hydrogen tartrate CRS in reference solution (a) and dilute to 10 mL with reference solution (a).

Column:
— size: $l = 0.10$ m, $\varnothing = 4.0$ mm;
— stationary phase: silica gel AGP for chiral chromatography R (5 μ m).

Mobile phase Mix 205 μ L of *N,N*-dimethylethylamine R and 20.0 mL of acetonitrile R1 and dilute to 1000 mL with solution A.

Detection Spectrophotometer at 200 nm.
Injection 20 μ L.

Run time Twice the retention time of rivastigmine. **Relative retention** With reference to rivastigmine (retention time = about 9 min): impurity D = about 0.8.

System suitability: reference solution (b):
— H_p = height above the baseline of the peak due to impurity D and H_b = height above the baseline of the lowest point of the curve separating this peak from the peak due to rivastigmine.

Calculation of percentage content:
— use the concentration of impurity D in reference solution (a).
Limit:
— impurity D: maximum 0.3 per cent.

Related substances

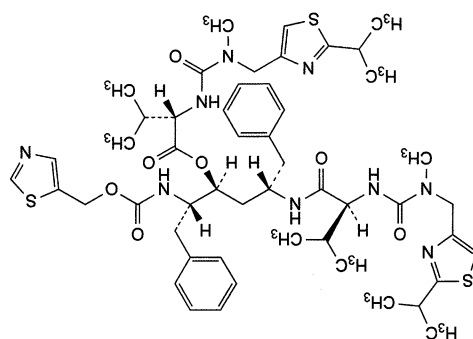
Liquid chromatography (2.2.29). Carry out the test protected from light.

Test solution Dissolve 62.5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of rivastigmine for system suitability CRS (containing impurities A, B and C) in 1.0 mL of the mobile phase.

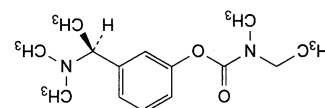
U. (1*S*,3*S*)-3-[[[(2*S*)-3-methyl-2-[[methyl]-(1-methylethyl)thiazol-4-yl]methoxy]carbamoyl]amino]-4-phenyl-1-[[methyl]-(1*S*)-2-phenyl-1-[[[(thiazol-5-yl)methoxy]carbamoyl]amino]ethyl]butyl (2*S*)-3-methyl-2-[[methyl]-(1-methylethyl)thiazol-4-yl]methoxy]carbamoyl]amino]butanoate.



Ph Eur

Rivastigmine

(Ph. Eur. monograph 2629)

C₁₄H₂₂N₂O₂ 250.3 123441-03-2

Action and use
Cholinesterase inhibitor; treatment of dementia in Alzheimer's disease and Parkinson's disease

Ph Eur

DEFINITION

3-[(1*S*)-1-(Dimethylamino)ethyl]phenyl ethyl(methyl)carbamate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Viscous, clear, colourless or yellow or very slightly brown, hygroscopic liquid.

Solubility

Sparsely soluble in water, very soluble in anhydrous ethanol and in heptane.

IDENTIFICATION

Carry out either tests A, B or tests B, C.

A. Specific optical rotation (2.2.7): -44.0 to -38.0

(anhydrous substance). Prepare the solution immediately before use.

Dissolve 0.300 g in ethyl acetate R and dilute to 50.0 mL

with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Film.

Comparison rivastigmine hydrogen tartrate CRS, treated as follows: dissolve 0.100 g in 30 mL of buffer solution pH 11 R, then add 30 mL of 1,1-dimethylethyl methyl ether R and shake

Reference solution (c) Dissolve 50.0 mg of rivastigmine hydrogen tartrate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase Mix 42 volumes of an 8.9 g/L solution of disodium hydrogen phosphate dihydrate R previously adjusted to pH 7.0 with phosphoric acid R and 58 volumes of methanol R1.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (b).

Run time Twice the retention time of rivastigmine.

Identification of impurities Use the chromatogram supplied with rivastigmine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention With reference to rivastigmine (retention time = about 10 min): impurity A = about 0.4;

impurity C = about 0.6; impurity B = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurities C and B.

Calculation of percentage contents:

— for each impurity, use the concentration of rivastigmine in reference solution (a).

Limits:

— impurity A: maximum 0.3 per cent;

— impurity B: maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum 0.10 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent mixture water R, acetone R (20:80 V/V).

0.250 g complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

Maximum 0.5 per cent, determined on 1.000 g.

Change the solvent after standardisation of the titrant and after every 3rd sample.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (c).

System suitability: reference solution (c):

— symmetry factor: maximum 2.5 for the peak due to rivastigmine.

Calculate the percentage content of $C_{14}H_{22}N_2O_2$ taking into account the assigned content of rivastigmine hydrogen tartrate CRS and a conversion factor of 0.625.

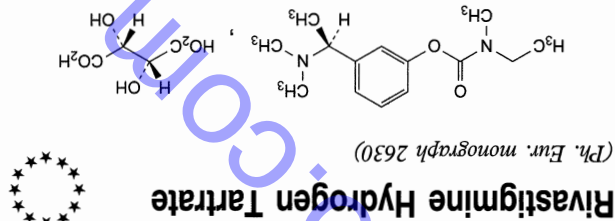
Under an inert gas, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

STORAGE

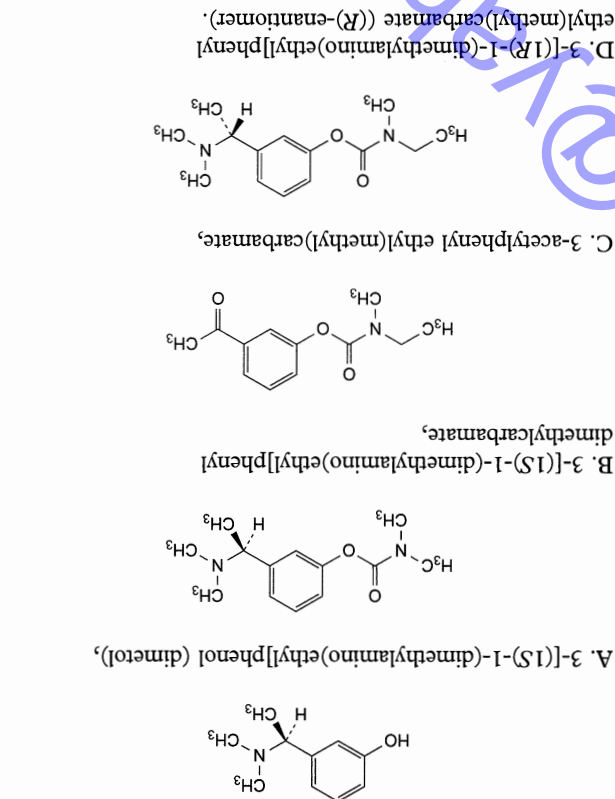
Under an inert gas, in an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

Content

98.0 per cent to 102.0 per cent (dried substance).



Ph Eur



IMPURITIES

Specified impurities: A, B, D.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.

CHARACTERS**Appearance**

White or almost white, very hygroscopic, crystalline or fine crystalline powder.

Solubility

Very soluble in water, soluble in methanol, very slightly soluble in ethyl acetate.

It shows polymorphism (5.9).

IDENTIFICATION

Carry out either tests A, B or tests B, C.

A. Specific optical rotation (2.2.7): + 4.2 to + 5.1.

Dissolve 0.600 g in methanol R and dilute to 20.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison rivastigmine hydrogen tartrate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

C. Enantiomeric purity (see Tests).

TESTS**Enantiomeric purity**

Liquid chromatography (2.2.29).

Solution A Solution containing 1.78 g/L of disodium hydrogen phosphate dihydrate R and 1.38 g/L of sodium dihydrogen phosphate monohydrate R, adjusted to pH 6.0 with phosphoric acid R.

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.0 mg of rivastigmine impurity D CRS in the mobile phase and dilute to 200.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 1 mg of rivastigmine hydrogen tartrate CRS in reference solution (a) and dilute to 10.0 mL with the same solution.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.0$ mm;

— stationary phase: α -1-acid-glycoprotein silica gel for chiral separation R (5 μ m).

Mobile phase Mix 205 μ L of *N,N*-dimethylethylamine R and 20.0 mL of acetonitrile R1 and dilute to 1000 mL with solution A.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 200 nm.

Injection 20 μ L.

Run time Twice the retention time of rivastigmine.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity D.

Relative retention With reference to rivastigmine (retention time = about 9 min): impurity D = about 0.8.

System suitability: reference solution (b):

— peak-to-valley ratio: minimum 2.5, where H_p = height above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to rivastigmine.

Calculation of percentage content:

— for impurity D, use the concentration of impurity D in reference solution (a).

Limit:

— impurity D: maximum 0.3 per cent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of rivastigmine for system suitability CRS (containing impurities A, B and C) in 1.0 mL of the mobile phase.

Reference solution (c) Dissolve 50.0 mg of rivastigmine hydrogen tartrate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase Mix 42 volumes of an 8.9 g/L solution of disodium hydrogen phosphate dihydrate R previously adjusted to pH 7.0 with phosphoric acid R and 58 volumes of methanol R1.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (b).

Run time Twice the retention time of rivastigmine.

Identification of impurities Use the chromatogram supplied with rivastigmine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention with reference to rivastigmine (retention time = about 10 min): impurity A = about 0.4; impurity C = about 0.6; impurity B = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurities C and B.

Calculation of percentage contents:

— for each impurity, use the concentration of rivastigmine in reference solution (a).

Limits:

— impurity A: maximum 0.3 per cent;

— unspecified impurities: for each impurity, maximum 0.10 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent; disregard the peak due to tartaric acid.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution

(1 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection Test solution and reference solution (c).

System suitability: reference solution (c):

— *symmetry factor*: maximum 2.5 for the peak due to

rivastigmine.

Calculate the percentage content of $C_{18}H_{28}N_2O_8$ taking into account the assigned content of rivastigmine hydrogen tartrate CRS.

STORAGE

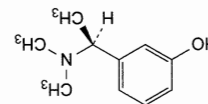
In an airtight container, protected from light.

IMPURITIES

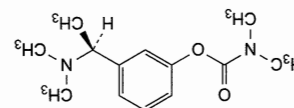
Specified impurities: A, D.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

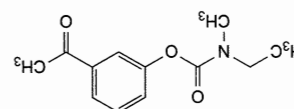
Control of impurities in substances for pharmaceutical use: B, C, E, F, G, H.



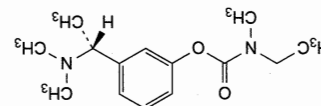
A. 3-[(1S)-1-(dimethylamino)ethyl]phenol,



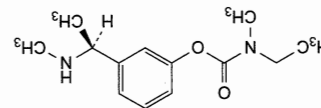
B. 3-[(1S)-1-(dimethylamino)ethyl]phenyl dimethylcarbamate,



C. 3-acetylphenyl ethyl(methyl)carbamate,



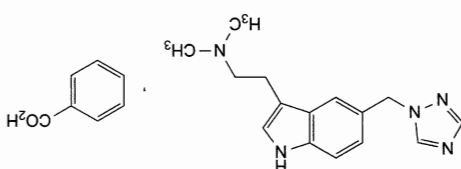
D. 3-[(1R)-1-(dimethylamino)ethyl]phenyl ethyl(methyl)carbamate,



E. 3-[(1S)-1-(methylamino)ethyl]phenyl ethyl(methyl)carbamate,

Rizatriptan Benzoate

(Ph. Eur. monograph 2585)



$C_{22}H_{25}N_2O_2$

391.5

145202-66-0

Action and use

Serotonin 5HT₁ receptor agonist; treatment of migraine.

Preparations

Orodispersible Rizatriptan Tablets

Rizatriptan Tablets

Ph Eur

DEFINITION

N,N' -Dimethyl-2-[5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethanamine benzoate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder or crystals.

Solubility

Soluble in water, sparingly soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison rizatriptan benzoate CRS.

If the spectra obtained show differences, dissolve the

substance to be examined and the reference substance

separately in methanol R, evaporate to dryness and record

new spectra using the residues.

B. Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time and size to the 2 principal peaks in the chromatogram obtained with reference solution (a).

TESTS

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure. Use silanised glass autosampler vials and freshly prepared solutions.

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 50.0 mg of rizatriptan benzoate CRS in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of rizatriptan for system suitability CRS (containing impurity C) in mobile phase A and dilute to 5.0 mL with mobile phase A.

Reference solution (c) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 20.0 mL with mobile phase A.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: phenylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: mix 160 mL of acetonitrile R and 840 mL of water R, add 1.0 mL of trifluoroacetic acid R and mix;
— mobile phase B: to 1000 mL of acetonitrile R add 1.0 mL of trifluoroacetic acid R and mix;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 8	100	0
8 - 17	100 \rightarrow 70	0 \rightarrow 30
17 - 20	70	30
20 - 20.1	70 \rightarrow 100	30 \rightarrow 0
20.1 - 23	100	0

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with rizatriptan for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to rizatriptan (retention time = about 5 min): impurity C = about 1.3; benzoic acid = about 2.1.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to rizatriptan and impurity C.

Limits:

— unspecified impurities: for each impurity, maximum 0.10 per cent;

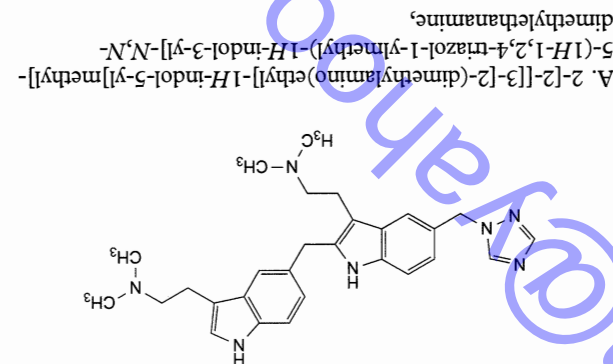
— total: maximum 0.3 per cent;

— disregard limit: the area of the peak due to rizatriptan in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard any peak due to benzoic acid.

Heavy metals (2.4.8)
Maximum 10 ppm.
Solvent water R.
0.50 g complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.
Water (2.5.12)
Maximum 0.5 per cent, determined on 0.500 g.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.
ASSAY
Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.
Injection Test solution and reference solution (a).
System suitability: reference solution (a):
— symmetry factor: maximum 3.5 for the peak due to rizatriptan.
Calculate the percentage content of $C_{22}H_{25}N_5O_2$ from the declared content of rizatriptan benzoate CRS.

IMPURITIES

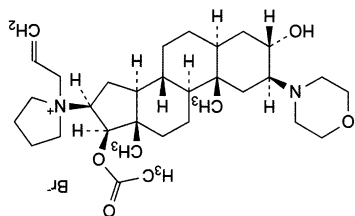
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, G, H, I.





Rocuronium Bromide

(Ph. Eur. monograph 1764)



$C_{32}H_{53}BrN_2O_4$

610

119302-91-9

Action and use

Non-depolarizing neuromuscular blocker.

DEFINITION

1-[17β-Acetoxy-3α-hydroxy-2β-(morpholin-4-yl)-5α-androstan-16β-yl]-1-(prop-2-enyl)pyrrolidinium bromide.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Almost white or pale yellow, slightly hygroscopic powder.

Solubility

Freely soluble in water, very soluble in methylene chloride, freely soluble in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison rocuronium bromide CRS.

B. Solution S (see Tests) gives reaction (a) of bromides (2.3.1).

TESTS

Solution S

Dissolve 0.10 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Specific optical rotation (2.2.7)

+28.5 to +32.0 (anhydrous substance). Dissolve 0.250 g in a 5.15 g/L solution of hydrochloric acid R and dilute to 25.0 mL with the same solution.

pH (2.2.3)

8.9 to 9.5 for solution S.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture water R, acetonitrile R1 (10:90 V/V).

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of rocuronium for peak

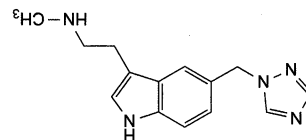
identification CRS (containing impurities A, B, C, F, G

and H) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

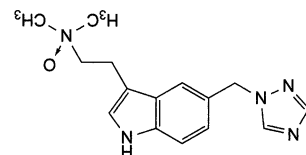
Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

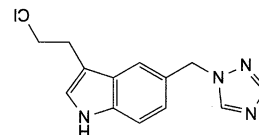
I. N-methyl-2-[5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethanamine.



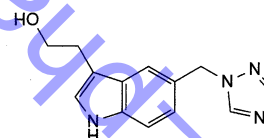
H. N,N-dimethyl-2-[5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethanamine N-oxide,



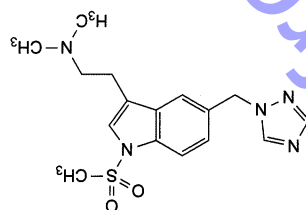
G. 3-(2-chloroethyl)-5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indole,



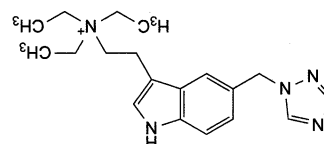
F. 2-[5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethanol,



E. N,N-dimethyl-2-[1-(methylsulfonyl)-5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethanamine,



D. N,N,N-triethyl-2-[5-(1H-1,2,4-triazol-1-ylmethyl)-1H-indol-3-yl]ethanaminium,



— stationary phase: silica gel for chromatography R (5 µm);
— temperature: 30 °C.
Mobile phase Mix 10 volumes of a 4.53 g/L solution of tetramethylammonium hydroxide R adjusted to pH 7.4 with phosphoric acid R and 90 volumes of acetonitrile R1.
Flow rate 2.0 mL/min.
Detection Spectrophotometer at 210 nm.

Injection 5 µL.
Run time 2.5 times the retention time of rocuronium.

Identification of impurities Use the chromatogram supplied with rocuronium for peak identification CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, F, G and H. Relative retention With reference to rocuronium (retention time = about 9 min): impurity A = about 0.2; impurity G = about 0.4; impurity F = about 0.75; impurity B = about 0.80; impurity H = about 0.95; impurity C = about 1.2.

System suitability: reference solution (b):
— peak-to-valley ratio: minimum 3.0, where H_p = height above the baseline of the peak due to impurity H and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to rocuronium.
Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.5; impurity F = 1.3; impurity G = 0.4; impurity H = 0.4; impurities A, B, C: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent); impurities F, G, H: for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.15 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— total: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peaks due to the blank and any peak eluting before impurity A.

Chlorides

Liquid chromatography (2.2.29).
Test solution Dissolve 20.0 mg of the substance to be examined in water R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 0.644 g of sodium bromide R and dilute to 1000.0 mL with the same solvent.
Reference solution (b) Dissolve 0.824 g of sodium chloride R in solution to 50.0 mL with water R.
1000.0 mL with the same solvent. Dilute 1.0 mL of the solution to 50.0 mL with water R.
Blank solution water R.
Precolumn: — size: $l = 0.05$ m, $\varnothing = 4.0$ mm;
— stationary phase: anion-exchange resin R (13 µm).

Column: — size: $l = 0.25$ m, $\varnothing = 4.0$ mm;
— stationary phase: anion-exchange resin R (13 µm).
Mobile phase A solution containing 0.063 g/L of sodium hydroxide R and 0.212 g/L of anhydrous sodium carbonate R.
Flow rate 2.0 mL/min.
Detection Conductivity detector set at 100 µS/V and maintained at 30 °C.
Use a self-regenerating anion suppressor.
Injection 25 µL.
Retention times Chloride = about 1.7 min; bromide = about 2.8 min.
System suitability: reference solution (a):
— resolution: minimum 2.5 between the peaks due to chloride and bromide.
Limit:

— chlorides: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

2-Propanol (2.4.24, System A)

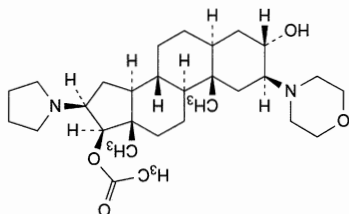
Maximum 1.0 per cent.
Heavy metals (2.4.8)
Maximum 20 ppm.
Solvent water R.
1.0 g complies with test H. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.
Water (2.5.12)
Maximum 4.5 per cent, determined on 0.400 g.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.
ASSAY
Dissolve 0.400 g in 40 mL of glacial acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 60.97 mg of $C_{22}H_{33}BrN_2O_4$.

STORAGE

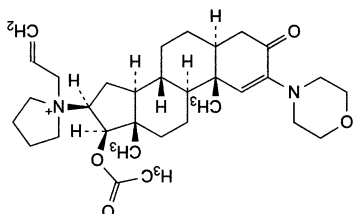
In an airtight container, protected from light, at a temperature below –15 °C.

IMPURITIES

Specified impurities A, B, C, F, G, H.
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, E.



A. 3 α -hydroxy-2 β -(morpholin-4-yl)-16 β -(pyrrolidin-1-yl)-5 α -androstane-17 β -yl acetate,



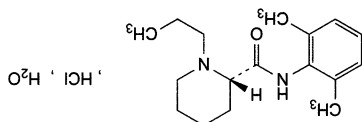
H. 1-[17β-acetoxy-2-(morpholin-4-yl)-3-oxo-5α-androst-1-en-16β-yl]-1-(prop-2-enyl)pyrrolidinium.

Ph Eur



Ropivacaine Hydrochloride Monohydrate

(Ph. Eur. monograph 2335)



$C_{17}H_{27}ClN_2O_5 \cdot H_2O$ 328.9 132112-35-7

Action and use

Local anaesthetic.

Ph Eur

DEFINITION

(-)-(2*S*)-*N*-(2,6-Dimethylphenyl)-1-propylpiperidine-2-carboxamide hydrochloride monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water and in ethanol (96 per cent), slightly soluble in methylene chloride.

IDENTIFICATION

Carry out either tests A, C, D or tests A, B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison ropivacaine hydrochloride monohydrate CRS.

B. Specific optical rotation (2.2.7): -74.0 to -64.0

(anhydrous substance).

Mix 2 mL of a 200 g/L solution of sodium hydroxide R and

30 mL of water R and dilute to 100.0 mL with ethanol

(96 per cent) R (solution A). Dissolve 0.500 g of the

substance to be examined in solution A and dilute to

50.0 mL with solution A.

C. It gives reaction (a) of chlorides (2.3.1).

D. Enantiomeric purity (see Tests).

TESTS

Solution S

Dissolve 0.50 g in carbon dioxide-free water R and dilute to

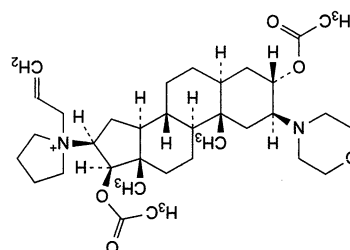
25.0 mL with the same solvent.

Appearance of solution

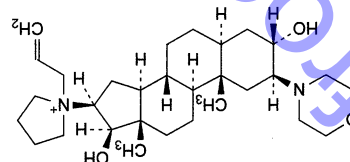
Solution S is clear (2.2.1).

pH (2.2.3)

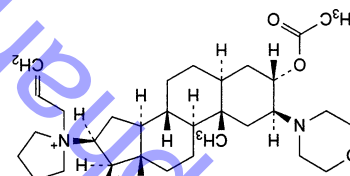
4.5 to 6.0 for solution S.



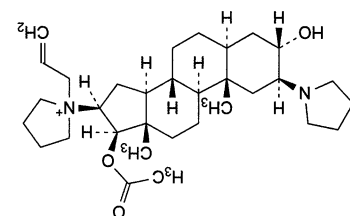
B. 1-[3α,17β-diacetoxy-2β-(morpholin-4-yl)-5α-androstan-16β-yl]-1-(prop-2-enyl)pyrrolidinium,



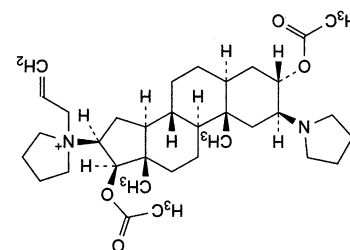
C. 1-[3α,17β-dihydroxy-2β-(morpholin-4-yl)-5α-androstan-16β-yl]-1-(prop-2-enyl)pyrrolidinium,



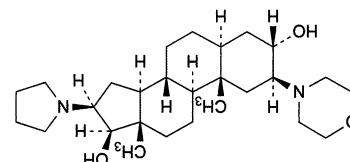
D. 1-[3α-acetoxy-17β-hydroxy-2β-(morpholin-4-yl)-5α-androstan-16β-yl]-1-(prop-2-enyl)pyrrolidinium,



E. 1-[17β-acetoxy-3α-hydroxy-2β-(pyrrolidin-1-yl)-5α-androstan-16β-yl]-1-(prop-2-enyl)pyrrolidinium,



F. 1-[3α,17β-acetoxy-2β-(pyrrolidin-1-yl)-5α-androstan-16β-yl]-1-(prop-2-enyl)pyrrolidinium,



G. 2β-(morpholin-4-yl)-16β-(pyrrolidin-1-yl)-5α-androstan-3α,17β-diol,

Absorbance (2.2.25)

Maximum 0.030 at 405 nm and maximum 0.025 at 436 nm, determined on solution S prepared immediately before use, with a path length of 5 cm and using water R as the compensation liquid.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 55 mg of the substance to be examined in the mobile phase and dilute to 20 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of bupivacaine hydrochloride CRS in the mobile phase and dilute to 5 mL with the mobile phase. Dilute 1 mL of this solution to 100 mL with the mobile phase.

Column:
— size: 1 = 0.15 m, Ø = 3.9 mm;
— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (4 µm).

Mobile phase Mix 1.3 mL of a 138 g/L solution of sodium dihydrogen phosphate R and 32.5 mL of an 89 g/L solution of disodium hydrogen phosphate R and dilute to 1000 mL with water R; mix equal volumes of this solution (pH 8.0) and acetonitrile R.

Flow rate 1.0 mL/min.
Injection 20 µL.
Detection Spectrophotometer at 240 nm.

Run time 2.5 times the retention time of ropivacaine. **Relative retention** With reference to ropivacaine (retention time = about 6 min): impurity A = about 1.6.

System suitability: reference solution (b):
— resolution: minimum 6.0 between the peaks due to ropivacaine and impurity A.

Limits:
— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Impurity H

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution Dissolve 13.0 mg of 2,6-dimethylaniline hydrochloride R in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase.

Retention time Impurity H = about 2-3 min.

Enantiomeric purity

Capillary electrophoresis (2.2.47) use the normalisation procedure.

Test solution Dissolve 50 mg of the substance to be examined in water R and dilute to 25 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with water R.

Reference solution (b) Dissolve 1.5 mg of the substance to be examined and 1.5 mg of ropivacaine impurity G CRS in water R and dilute to 100 mL with the same solvent.

Capillary:
— material: fused silica;
— size: effective length = about 72 cm, total length = 80 cm, Ø = 50 µm.

Temperature 30 °C.
CZE buffer Prepare a 13.3 g/L solution of dimethyl-β-cyclodextrin R in an 11.5 g/L solution of phosphoric acid R previously adjusted to pH 3.0 with triethanolamine R.

The CZE buffer is prepared and filtered through a membrane filter (nominal pore size 0.45 µm) immediately before use.

Detection Spectrophotometer at 206 nm.

Preconditioning of the capillary Rinse the capillary at 100 kPa with water R for 1 min, with 0.1 M sodium hydroxide for 10 min and with water R for 3 min. If the capillary is new or dry, increase the sodium hydroxide rinse to 30 min.

Between-run rinsing Rinse the capillary at 100 kPa with water R for 1 min, with 0.1 M sodium hydroxide for 4 min, with water R for 1 min and with the CZE buffer for 4 min.

Injection Under pressure (5 kPa) for 5 s.
Migration Apply a field strength of 375 V/cm with an initial ramp of 500 V/s and a positive polarity corresponding to a current of 40-45 µA.

Run time 30 min.
System suitability:
— resolution: minimum 3.7 between the peaks due to impurity G (1st peak) and (S)-ropivacaine in the electropherogram obtained with reference solution (b);
if necessary, increase the dimethyl-β-cyclodextrin concentration in the CZE buffer or vary the pH between 2.9 and 3.1 or lower the temperature;

— signal-to-noise ratio: minimum 10 for the principal peak in the electropherogram obtained with reference solution (a). **Limit:**
— impurity G: maximum 0.5 per cent.

Heavy metals (2.4.8)

Dissolve 2.0 g in a mixture of 15 volumes of water R and 85 volumes of methanol R and dilute to 20 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (100 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with a mixture of 15 volumes of water R and 85 volumes of methanol R.

Water (2.5.12)

5.0 per cent to 6.0 per cent, determined on 0.100 g.

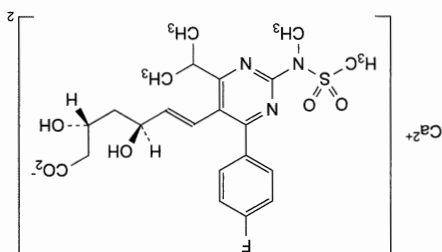
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.



Rosuvastatin Calcium

(Ph. Eur. monograph 2631)



C₄₄H₅₄CaF₂N₆O₁₂S₂ 1001 147098-20-2

Action and use

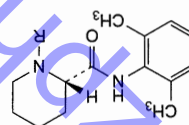
HMG Co-A reductase inhibitor; lipid-regulating drug.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E, F.

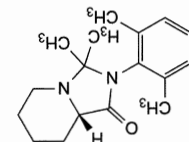
Specified impurities A, G, H

IMPURITIES

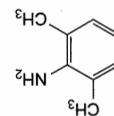
of C₁₇H₂₇ClN₂O.
1 mL of 0.1 M sodium hydroxide is equivalent to 31.09 mg between the 2 points of inflexion.
using 0.1 M sodium hydroxide. Read the volume added hydrochloric acid. Carry out a potentiometric titration (2.2.20).
40 mL of ethanol (96 per cent) R. Add 5.0 mL of 0.01 M Dissolve 0.250 g in a mixture of 10 mL of water R and



A. R = [CH₂]₃-CH₃, (S)-bupivacaine,
B. R = H: (-)-(2S)-N-(2,6-dimethylphenyl)piperidine-2-carboxamide,
C. R = CH₃: (-)-(2S)-N-(2,6-dimethylphenyl)-1-methylpiperidine-2-carboxamide ((S)-mepivacaine),
D. R = C₂H₅: (-)-(2S)-N-(2,6-dimethylphenyl)-1-ethylpiperidine-2-carboxamide,
E. R = CH(CH₃)₂: (-)-(2S)-N-(2,6-dimethylphenyl)-1-(1-methylethyl)piperidine-2-carboxamide,



F. (8aS)-2-(2,6-dimethylphenyl)-3,3-dimethylhexahydroindolizino[1,5-a]pyridin-1(5H)-one (acetone adduct),
G. (+)-(2R)-N-(2,6-dimethylphenyl)-1-propylpiperidine-2-carboxamide ((R)-ropivacaine),
H. 2,6-dimethylaniline.



Liquid chromatography (2.2.29). Carry out the test protected from light.
Solvent mixture acetonitrile for chromatography R, water for chromatography R (25:75 V/V).
Test solution Dissolve 35.0 mg of the substance to be examined in 12 mL of acetonitrile for chromatography R and dilute to 50.0 mL with water for chromatography R.
Reference solution (a) Dissolve 35.0 mg of rosuvastatin calcium CRS in 12 mL of acetonitrile for chromatography R and dilute to 50.0 mL with water for chromatography R.
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 10.0 mL with the solvent mixture.
Reference solution (c) Dilute 7 mg of rosuvastatin for system suitability CRS (containing impurities A, B and C) in 2.5 mL of acetonitrile for chromatography R and dilute to 10 mL with water for chromatography R.
Column:
— size: l = 0.15 m, Ø = 3.0 mm;
— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (3 µm);
— temperature: 40 °C.

TESTS

A. Infrared absorption spectrophotometry (2.2.24).
Comparison rosuvastatin calcium CRS.
B. Enantiomeric purity (see Tests).
C. It gives reaction (b) of calcium (2.3.1).

IDENTIFICATION

White or almost white, hygroscopic powder.
Slightly soluble in water, freely soluble in methylene chloride, practically insoluble in anhydrous ethanol.

Solubility

Appearance

CHARACTERS

97.0 per cent to 102.0 per cent (anhydrous substance).

Content

Calcium bis[(3R,5S,6E)-7-[4-(4-fluorophenyl)-6-(1-methylethyl)-2-[methyl(methylsulfonyl)amino]pyrimidin-5-yl]-3,5-dihydroxyhept-6-enoate].

Mobile phase:
— mobile phase A: 1 per cent V/V solution of trifluoroacetic acid R, acetonitrile for chromatography R, water for chromatography R (1:29:70 V/V/V);
— mobile phase B: 1 per cent V/V solution of trifluoroacetic acid R, water for chromatography R, acetonitrile for chromatography R (1:24:75 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	100	0
30 - 50	100 ÷ 60	0 ÷ 40
50 - 60	60 ÷ 0	40 ÷ 100
60 - 70	0	100

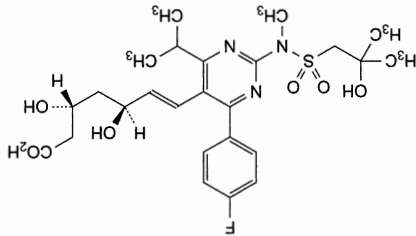
Detection Spectrophotometer at 242 nm.
Injection 10 µL of the test solution and reference solutions (b) and (c).
Identification of impurities Use the chromatogram supplied with rosuvastatin for system suitability CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B and C.

Relative retention With reference to rosuvastatin (retention time = about 25 min): impurity A = about 0.9; impurity B = about 1.1; impurity C = about 1.5.
System suitability: reference solution (c):
— resolution: minimum 2.0 between the peaks due to rosuvastatin and impurity B.
Calculation of percentage contents:
— correction factor: multiply the peak area of impurity C by 1.4;
— for each impurity, use the concentration of rosuvastatin in reference solution (b).

Limits:
— impurity C: maximum 0.8 per cent;
— impurity B: maximum 0.5 per cent;
— impurity A: maximum 0.2 per cent;
— unspecified impurities: for each impurity, maximum 0.10 per cent;
— total: maximum 1.2 per cent;
— reporting threshold: 0.05 per cent.

Enantiomeric purity
Liquid chromatography (2.2.29). Carry out the test protected from light.
Solvent mixture acetonitrile for chromatography R, water for chromatography R (25:75 V/V).

Test solution Dissolve 25.0 mg of the substance to be examined in 6 mL of acetonitrile for chromatography R and dilute to 25.0 mL with water for chromatography R.
Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.
Reference solution (b) Dissolve 5 mg of rosuvastatin impurity G CRS in a mixture of 12 mL of acetonitrile for chromatography R and 10 mL of water for chromatography R using sonication and dilute to 50.0 mL with water for chromatography R.
Reference solution (c) To 25 mg of the substance to be examined add 1.0 mL of reference solution (b), 6 mL of acetonitrile for chromatography R and dissolve using sonication; dilute to 25 mL with water for chromatography R.



A. (3R,5S,6E)-7-[4-(4-fluorophenyl)-2-[[[(2-hydroxy-2-methylpropyl)sulfonyl](methyl)amino]-6-(1-methylethyl)pyrimidin-5-yl]-3,5-dihydroxyhept-6-enoic acid,

IMPURITIES
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.
Specified impurities A, B, C, G
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, E, F, J.

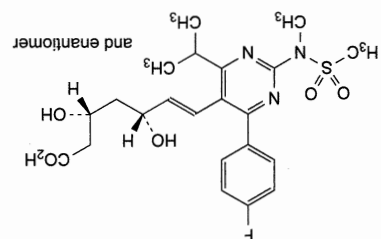
STORAGE
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.
IMPURITIES
In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.
Specified impurities A, B, C, G
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, E, F, J.

ASSAY
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection Test solution and reference solution (a).
Calculation Calculate the percentage content of C₄₄H₅₄CaF₂N₆O₁₂S₂ taking into account the assigned content of rosuvastatin calcium CRS.

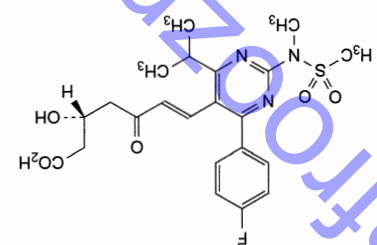
Water (2.5.12)
Maximum 6.1 per cent, determined on 0.100 g.
Limit:
— impurity G: maximum 0.1 per cent.
Calculation of percentage content:
— use the concentration of rosuvastatin in reference solution (a).
Relative retention With reference to rosuvastatin (retention time = about 29 min): impurity G = about 0.9.
System suitability: reference solution (c):
— resolution: minimum 1.5 between the peaks due to impurity G and rosuvastatin.
Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity G.
Run time 2.6 times the retention time of rosuvastatin.
Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity G.

Detection Spectrophotometer at 242 nm.
Injection 10 µL of the test solution and reference solutions (a) and (c).
Flow rate 0.5 mL/min.

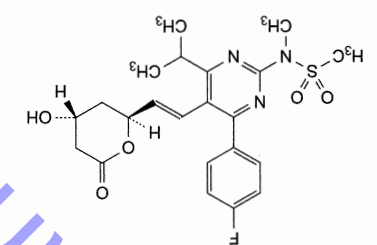
Mobile phase acetonitrile for chromatography R, trifluoroacetic acid R (25:75 V/V).
Flow rate 0.5 mL/min.
Detection Spectrophotometer at 242 nm.
Injection 10 µL of the test solution and reference solutions (a) and (c).
Run time 2.6 times the retention time of rosuvastatin.
Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity G.
Relative retention With reference to rosuvastatin (retention time = about 29 min): impurity G = about 0.9.
System suitability: reference solution (c):
— resolution: minimum 1.5 between the peaks due to impurity G and rosuvastatin.
Calculation of percentage content:
— use the concentration of rosuvastatin in reference solution (a).
Limit:
— impurity G: maximum 0.1 per cent.
Water (2.5.12)
Maximum 6.1 per cent, determined on 0.100 g.



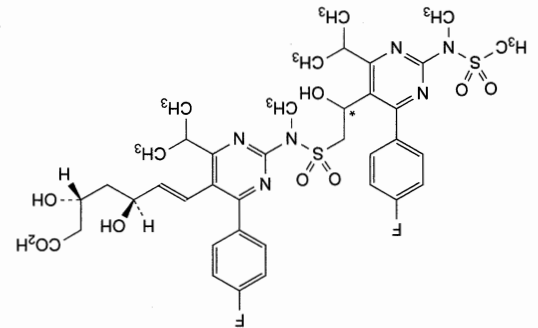
B. (3*R*,5*R*,6*E*)-7-[4-(4-fluorophenyl)-6-(1-methylethyl)-2-[methylethyl(methylsulfonyl)amino]pyrimidin-5-yl]-3,5-dihydroxyhept-6-enoic acid,



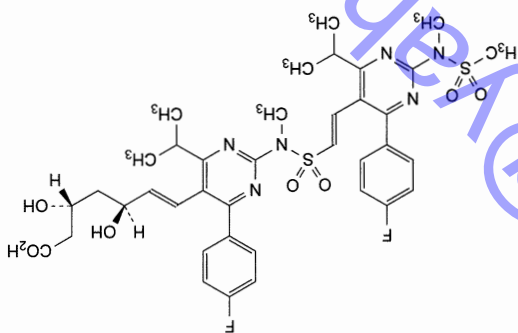
C. (3*R*,6*E*)-7-[4-(4-fluorophenyl)-6-(1-methylethyl)-2-[methylethyl(methylsulfonyl)amino]pyrimidin-5-yl]-3-hydroxy-5-oxohept-6-enoic acid,



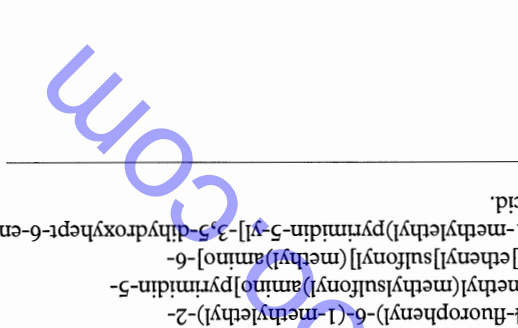
D. *N*-[4-(4-fluorophenyl)-5-[(*E*)-2-[(2*S*,4*R*)-4-hydroxy-6-oxotetrahydro-2*H*-pyran-2-yl]ethenyl]-6-(1-methylethyl)pyrimidin-2-yl]-*N*-methylmethanesulfonamide,



E. (3*R*,5*S*,6*E*)-7-[4-(4-fluorophenyl)-2-[[[2-(4-fluorophenyl)-6-(1-methylethyl)-2-[methylethyl(methylsulfonyl)amino]pyrimidin-5-yl]-3,5-dihydroxyhept-6-enoic acid,



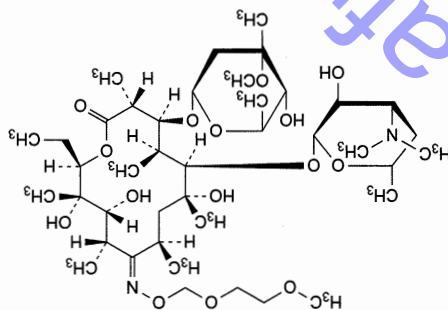
I. (3*R*,5*S*,6*E*)-7-[4-(4-fluorophenyl)-2-[[[2-(4-fluorophenyl)-6-(1-methylethyl)-2-[methylethyl(methylsulfonyl)amino]pyrimidin-5-yl]ethenyl]sulfonyl] (methyl)amino]-6-(1-methylethyl)pyrimidin-5-yl]-3,5-dihydroxyhept-6-enoic acid.



Ph Eur

Roxithromycin

(Ph. Eur. monograph 1146)

C₄₁H₇₆N₂O₁₅ 837

80214-83-1

Action and use
Antibacterial.

Ph Eur

DEFINITION

(3R,4S,5S,6R,7R,9R,11S,12R,13S,14R)-4-[(2,6-dideoxy-3-C-methyl-3-O-methyl-α-L-rhamno-hexopyranosyl)oxy]-14-ethyl-7,12,13-trihydroxy-10-

[(B)-[(2-methoxyethoxy)methoxy]imino]-3,5,7,9,11,13-hexamethyl-6-[[3,4,6-trideoxy-3-(dimethylamino)-β-D-

xylohexopyranosyl]oxy]oxacyclotetradecan-2-one(erythronycin 9-(B)-[O-

[(2-methoxyethoxy)methyl]oxime).
Semi-synthetic product derived from a fermentation product.

96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in acetone, in

alcohol and in methylene chloride. It is slightly soluble in

dilute hydrochloric acid.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison roxithromycin CRS.

If the spectra obtained shows differences, prepare further

spectra using 90 g/L solutions in methylene chloride R.

B. Examine the chromatograms obtained in the assay.

The principal peak in the chromatogram obtained

with the test solution is similar in retention time and size to

the principal peak in the chromatogram obtained with

reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dissolve 0.2 g in methanol R and dilute to 20 mL with the

same solvent.

Specific optical rotation (2.2.7)

-93 to -96 (anhydrous substance).

Limits:

— impurity G: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent),

Dissolve 0.500 g in acetone R and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solution A: Mix 30 volumes of acetonitrile R and 70 volumes

of a 48.6 g/L solution of ammonium dihydrogen phosphate R,

adjusted to pH 5.3 with dilute sodium hydroxide solution R.

Test solution: Dissolve 50.0 mg of the substance to be

examined in solution A and dilute to 25.0 mL with

solution A.

Reference solution (a): Dissolve 50.0 mg of roxithromycin CRS

in solution A and dilute to 25.0 mL with solution A.

Reference solution (b): Dilute 1.0 mL of reference solution (a)

to 100.0 mL with solution A.

Reference solution (c): Dissolve 2.0 mg of roxithromycin for

system suitability CRS in solution A and dilute to 1.0 mL with

solution A.

Reference solution (d): Dilute 1.0 mL of toluene R to 100.0 mL

with acetonitrile R. Dilute 0.2 mL of this solution to

200.0 mL with solution A.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm,

— stationary phase: spherical end-capped octadecylsilyl silica gel

for chromatography R (5 μ m) with a 10 nm pore size and a

carbon loading of about 19 per cent,

— temperature: 15 °C.

Mobile phase:

— mobile phase A: mix 26 volumes of acetonitrile R and

74 volumes of a 59.7 g/L solution of ammonium

dihydrogen phosphate R, adjusted to pH 4.3 with dilute

sodium hydroxide solution R,

— mobile phase B: water R, acetonitrile R (30:70 V/V),

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 50	100	0
50 - 51	100 → 90	0 → 10
51 - 80	90	10
80 - 81	90 → 100	10 → 0
81 - 100	100	0

Flow rate 1.1 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 20 μ L, using an injector maintained at 8 °C, of the

test solution and reference solutions (b), (c) and (d).

Relative retention: With reference to roxithromycin (retention

time = about 22 min): impurity A = about 0.28;

impurity B = about 0.31; impurity C = about 0.33;

impurity D = about 0.62; impurity E = about 0.67;

impurity F = about 0.83; impurity G = about 1.15;

impurity K = about 1.7; impurity H = about 1.85;

impurity J = about 2.65; impurity I = about 3.1.

System suitability: reference solution (c):

— peak-to-valley ratio: minimum 2.0, where H_p = height

above the baseline of the peak due to impurity G and

 H_p = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

roxithromycin.

Absorption maxima At 257 nm and 358 nm.

Specific absorbance at the absorption maximum at 358 nm

305 to 330 (anhydrous substance).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison rutoside trihydrate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 25 mg of rutoside trihydrate CRS in methanol R and dilute to 10.0 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase butanol R, anhydrous acetic acid R, water R, methyl ethyl ketone R, ethyl acetate R

(5:10:10:30:50 V/V/V/V/V).

Application 10 µL.

Development Over a path of 10 cm.

Drying In air.

Detection Spray with a mixture of 2.5 mL of ferric chloride solution R1 and 7.5 mL of a 10 g/L solution of potassium

ferricyanide R and examine for 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve 10 mg in 5 mL of ethanol (96 per cent) R, add 1 g of zinc R and 2 mL of hydrochloric acid R1. A red colour develops.

TESTS

Light-absorbing impurities

The absorbance (2.2.25) is not greater than 0.10 at wavelengths between 450 nm and 800 nm.

Dissolve 0.200 g in 40 mL of 2-propanol R. Stir for 15 min, dilute to 50.0 mL with 2-propanol R and filter.

Substances insoluble in methanol

Maximum 3 per cent.

Shake 2.5 g of the substance to be examined for 15 min in 50 mL of methanol R at 20-25 °C. Filter under reduced pressure through a sintered-glass filter (1.6) (2.1.2) previously dried for 15 min at 100-105 °C, allowed to cool in a desiccator and tared. Wash the filter 3 times with 20 mL of methanol R. Dry the filter for 30 min at 100-105 °C. Allow to cool and weigh. The residue weighs a maximum of 75 mg.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.10 g of the substance to be examined in 20 mL of methanol R and dilute to 100.0 mL with mobile phase B.

Reference solution (a) Dissolve 10.0 mg of rutoside trihydrate CRS in 2.0 mL of methanol R and dilute to 10.0 mL with mobile phase B.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 50.0 mL with mobile phase B.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: octylsilyl silica gel for chromatography R

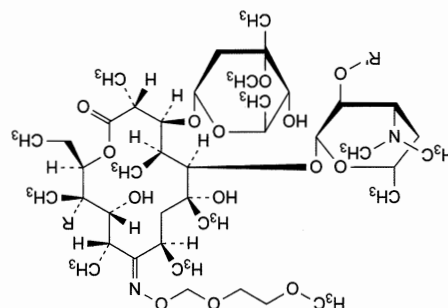
(5 µm);

— temperature: 30 °C.

Mobile phase:

— mobile phase A: mix 5 volumes of tetrahydrofuran R and 95 volumes of a 15.6 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R;

Ph Eur



H, R = R' = H: 12-deoxyerythrinyol-9-(E)-[O-[(2-methoxyethoxy)methyl]oxime],
I, R = OH, R' = CH₂-O-CH₂-CH₂-OCH₃; 2'-O-[(2-methoxyethoxy)methyl]erythrinyol-9-(E)-[O-[(2-methoxyethoxy)methyl]oxime].

(Ph. Eur. monograph 1795)



Rutoside Trihydrate

$C_{27}H_{30}O_{16} \cdot 3H_2O$

665

250249-75-3

Action and use
Bioflavonoid.

DEFINITION

3-[[6-O-(6-Deoxy- α -L-mannopyranosyl)- β -D-glucopyranosyl]oxy]-2-(3,4-dihydroxyphenyl)-5,7-dihydroxy-4H-1-benzopyran-4-one trihydrate.

Content

95.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Yellow or greenish-yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in methanol, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride. It dissolves in solutions of alkali hydroxides.

IDENTIFICATION

First identification: B.

Second identification: A, C, D.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in methanol R, dilute to 250.0 mL with the same solvent and filter if necessary. Dilute 5.0 mL of the solution to 50.0 mL with methanol R.

Spectral range 210-450 nm.

— *mobile phase B*: mix 40 volumes of tetrahydrofuran R and 60 volumes of a 15.6 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	50 → 0	50 → 100
10 - 15	0	100
15 - 16	0 → 50	100 → 50
16 - 20	50	50

Flow rate 1 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Relative retention With reference to rutoside (retention

time = about 7 min): impurity B = about 1.1;

impurity A = about 1.2; impurity C = about 2.5.

System suitability: reference solution (a):

— *resolution*: minimum 2.5 between the peaks due to

rutoside and impurity B.

Limits Locate the impurities by comparison with the

chromatogram provided with *rutoside trihydrate CRS*:

— *correction factors*: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity A = 0.8;

impurity C = 0.5;

— *impurities A, B, C*: For each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (2.0 per cent);

— *total*: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (b)

(4.0 per cent);

— *disregard limit*: 0.05 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

Water (2.5.12)

7.5 per cent to 9.5 per cent, determined on 0.100 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 20 mL of dimethylformamide R. Titrate

with 0.1 M tetrabutylammonium hydroxide, determining the

end-point potentiometrically (2.2.20).

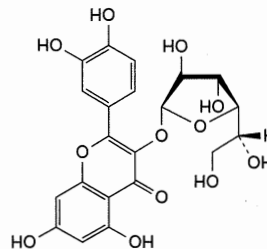
1 mL of 0.1 M tetrabutylammonium hydroxide is equivalent to

30.53 mg of C₂₇H₃₀O₁₆.

STORAGE

Protected from light.

IMPURITIES



A. 2-(3,4-dihydroxyphenyl)-3-(β-D-glucopyranosyloxy)-5,7-dihydroxy-4H-1-benzopyran-4-one (isouercitrinose),

Comparison saccharin CRS.

Preparation Discs.

C. Infrared absorption spectrophotometry (2.2.24).

B. Melting point (2.2.14): 226 °C to 230 °C. ◇

litmus paper R red.

A. A saturated solution, prepared without heating, turns blue

◇ Second identification: A, B, D, E.

First identification C.

IDENTIFICATION

dilute solutions of alkali hydroxides and carbonates. ◇
(96 per cent), slightly soluble in cold water. It dissolves in
sparingly soluble in boiling water and in ethanol

Solubility

crystals.

White or almost white, crystalline powder or colourless

Appearance

◆ CHARACTERS

99.0 per cent to 101.0 per cent (dried substance).

Content

1,2-Benzisothiazol-3(2H)-one 1,1-dioxide.

DEFINITION

Ph Eur

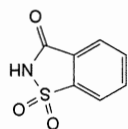
Sweetening agent.

Action and use

81-07-2

183.2

C₇H₅NO₃S

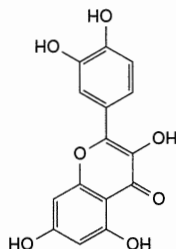


(Ph. Eur. monograph 0947)

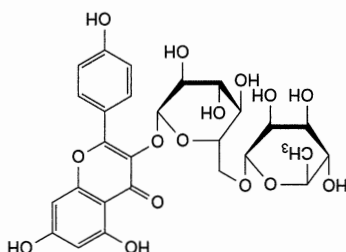
Saccharin (1)

Ph Eur

C. 2-(3,4-dihydroxyphenyl)-3,5,7-trihydroxy-4H-1-benzopyran-4-one (quercetin).



B. 3-[[6-O-(6-deoxy-α-L-mannopyranosyl)-β-D-glucopyranosyl]oxy]-5,7-dihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran-4-one (kaempferol 3-rutinoside),



◇D. Mix about 10 mg with about 10 mg of resorcinol R, add 0.25 mL of *sulfuric acid* R and carefully heat the mixture over a naked flame until a dark green colour is produced. Allow to cool, add 10 mL of *water* R and *dilute sodium hydroxide solution* R until an alkaline reaction is produced. An intense green fluorescence develops.

E. To 0.2 g add 1.5 mL of *dilute sodium hydroxide solution* R, evaporate to dryness and heat the residue carefully until it melts, avoiding carbonisation. Allow to cool, dissolve the mass in about 5 mL of *water* R, add *dilute hydrochloric acid* R until a weak acid reaction is produced and filter, if necessary. To the filtrate add 0.2 mL of *ferrous chloride solution* R2. A violet colour develops.

TESTS

Solution S

Dissolve 5.0 g in 20 mL of a 200 g/L solution of *sodium acetate* R and dilute to 25 mL with the same solution.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

o- and p-Toluenesulfonamide

Gas chromatography (2.2.28).

Internal standard solution Dissolve 25 mg of *caffeine* R in *methylene chloride* R and dilute to 100 mL with the same solvent.

Test solution Suspend 10.0 g of the substance to be examined in 20 mL of *water* R and dissolve using 5-6 mL of strong *sodium hydroxide solution* R. If necessary adjust the solution to pH 7-8 with 1 M *sodium hydroxide* or 1 M *hydrochloric acid* and dilute to 50 mL with *water* R. Shake the solution with 4 quantities, each of 50 mL, of *methylene chloride* R. Combine the lower layers, dry over *anhydrous sodium sulfate* R and filter. Wash the filter and the sodium sulfate with 10 mL of *methylene chloride* R. Combine the solution and the washings and evaporate almost to dryness in a water-bath at a temperature not exceeding 40 °C. Using a small quantity of suitable 10 mL tube, evaporate to dryness in a current of nitrogen and dissolve the residue in 1.0 mL of the internal standard solution.

Blank solution Evaporate 200 mL of *methylene chloride* R to dryness in a water-bath at a temperature not exceeding 40 °C. Dissolve the residue in 1 mL of *methylene chloride* R and 20.0 mg of *o-toluenesulfonamide* R and 20.0 mg of *p-toluenesulfonamide* R in *methylene chloride* R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 50.0 mL with *methylene chloride* R. Evaporate 5.0 mL of the final solution to dryness in a current of nitrogen. Dissolve the residue in 1.0 mL of the internal standard solution.

Column:
— *material:* fused silica,
— *size:* l = 10 m, Ø = 0.53 mm,
— *stationary phase:* polymethylphenylsiloxane R (film thickness 2 µm).
Carrier gas nitrogen for chromatography R.
Flow rate 10 mL/min.
Split ratio 1:2.
Temperature:
— *column:* 180 °C,
— *injection port and detector:* 250 °C.
Detection Flame ionisation.
Injection 1 µL.
Order of elution o-toluenesulfonamide, p-toluenesulfonamide, caffeine.

System suitability:
— *resolution:* minimum 1.5 between the peaks due to o-toluenesulfonamide and p-toluenesulfonamide in the chromatogram obtained with the reference solution, the chromatogram obtained with the blank solution does not show any peak with the same retention times as the internal standard, o-toluenesulfonamide and p-toluenesulfonamide.
Limits:
— o-toluenesulfonamide: the ratio of its area to that of the internal standard is not greater than the corresponding ratio in the chromatogram obtained with the reference solution (10 ppm),
— p-toluenesulfonamide: the ratio of its area to that of the internal standard is not greater than the corresponding ratio in the chromatogram obtained with the reference solution (10 ppm).
◆

Readily carbonisable substances

Dissolve 0.20 g in 5 mL of *sulfuric acid* R and keep at 48-50 °C for 10 min. When viewed against a white background, the solution is not more intensely coloured than a solution prepared by mixing 0.1 mL of red primary solution, 0.1 mL of blue primary solution and 0.4 mL of yellow primary solution (2.2.2) with 4.4 mL of *water* R.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dilute 10 mL of solution S to 20 mL with *water* R. 12 mL of the solution complies with test A. Prepare the reference solution using *lead standard solution* (2 ppm Pb) R. ◆

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

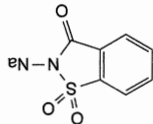
Dissolve 0.500 g in 40 mL of *ethanol* (96 per cent) R. Add 40 mL of *water* R. Titrate with 0.1 M *sodium hydroxide*, using a 10 g/L solution of *phenolphthalein* R in *ethanol* (96 per cent) R as indicator. Carry out a blank titration. 1 mL of 0.1 M *sodium hydroxide* is equivalent to 18.32 mg of C₇H₅NO₃S.

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.

Saccharin Sodium

Soluble Saccharin

(Ph. Eur. monograph 0787)



C₇H₄NNaO₃S

205.2

128-44-9

Action and use

Sweetening agent.

Ph Eur

DEFINITION

2-Sodio-1,2-benzisothiazol-3(2H)-one 1,1-dioxide.

Content
99.0 per cent to 101.0 per cent (anhydrous substance).
It may contain a variable quantity of water.

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless

crystals, efflorescent in dry air.

Solubility

Freely soluble in water, sparingly soluble in ethanol

(96 per cent).

IDENTIFICATION

First identification B, E.

Second identification A, C, D, E.

A. Melting point (2.2.14): 226 °C to 230 °C.

To 5 mL of solution S (see Tests) add 3 mL of dilute

hydrochloric acid R. A white precipitate is formed. Filter and

wash with water R. Dry the precipitate at 100-105 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs; dry the substances at 100-105 °C before

use.

Comparison saccharin sodium CRS.

C. Mix about 10 mg with about 10 mg of resorcinol R, add

0.25 mL of sulfuric acid R and carefully heat the mixture over

a naked flame until a dark green colour is produced. Allow to

cool, add 10 mL of water R and dilute sodium hydroxide

solution R until an alkaline reaction is produced. An intense

green fluorescence develops.

D. To 0.2 g add 1.5 mL of dilute sodium hydroxide solution R,

evaporate to dryness and heat the residue carefully until it

melts, avoiding carbonisation. Allow to cool, dissolve the

mass in about 5 mL of water R, add dilute hydrochloric acid R

until a weak acid reaction is produced and filter, if necessary.

To the filtrate add 0.2 mL of ferric chloride solution R2.

A violet colour develops.

E. Solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S
Dissolve 5.0 g in carbon dioxide-free water R and dilute to

50.0 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dissolve 5.0 g in 25 mL of carbon dioxide-free water R.

Acidity or alkalinity

To 10 mL of solution S add about 0.05 mL of a 10 g/L

solution of phenolphthalein R in ethanol (96 per cent) R.

The solution is not pink. Add 0.1 mL of 0.1 M sodium

hydroxide. The solution becomes pink.

o- and p-Toluenesulfonamide

Gas chromatography (2.2.28).

Internal standard solution Dissolve 25 mg of caffeine R in

methylene chloride R and dilute to 100 mL with the same

solvent.

Test solution Dissolve 10.0 g of the substance to be examined

in 50 mL of water R. If necessary adjust the solution to

pH 7-8 by addition of 1 M sodium hydroxide or 1 M

hydrochloric acid. Shake the solution with 4 quantities, each of

50 mL, of methylene chloride R. Combine the lower layers, dry

over anhydrous sodium sulfate R and filter. Wash the filter and

the sodium sulfate with 10 mL of methylene chloride R.

Combine the solution and the washings and evaporate almost

to dryness in a water-bath at a temperature not exceeding

40 °C. Using a small quantity of methylene chloride R,

quantitatively transfer the residue into a suitable 10 mL tube,

evaporate to dryness in a current of nitrogen R and add

1.0 mL of the internal standard solution.

Blank solution Evaporate 200 mL of methylene chloride R to

dryness in a water-bath at a temperature not exceeding

40 °C. Dissolve the residue in 1 mL of methylene chloride R.

Reference solution Dissolve 20.0 mg of o-toluenesulfonamide R

and 20.0 mg of toluenesulfonamide R in methylene chloride R

and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL

of the solution to 50.0 mL with methylene chloride R.

Evaporate 5.0 mL of the final solution to dryness in a current

of nitrogen R. Take up the residue using 1.0 mL of the

internal standard solution.

Column:

— material: fused silica,

— size: $l = 10$ m, $\varnothing = 0.53$ mm,

— stationary phase: polymethylphenylsiloxane R (film thickness

2 μ m).

Carrier gas nitrogen for chromatography R.

Flow rate 10 mL/min.

Split ratio 1:2.

Temperature:

— column: 180 °C,

— injection port and detector: 250 °C.

Detection Flame ionisation.

Injection 1 μ L.

Elution order o-toluenesulfonamide, p-toluenesulfonamide,

caffeine.

System suitability:

— resolution: minimum 1.5 between the peaks due to

o-toluenesulfonamide and p-toluenesulfonamide in the

chromatogram obtained with the reference solution,

— the chromatogram obtained with the blank solution does

not show any peak with the same retention times as the

internal standard, o-toluenesulfonamide and

p-toluenesulfonamide.

Limits:

— o-toluenesulfonamide: the ratio of its area to that of the

internal standard is not greater than the corresponding

ratio in the chromatogram obtained with the reference

solution (10 ppm).

Readily carbonisable substances

Dissolve 0.20 g in 5 mL of sulfuric acid R and keep at

48-50 °C for 10 min. When viewed against a white

background, the solution is not more intensely coloured than

a solution prepared by mixing 0.1 mL of red primary

solution, 0.1 mL of blue primary solution and 0.4 mL of

yellow primary solution (2.2.2) with 4.4 mL of water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (2 ppm Pb) R.

Water (2.5.12)

Maximum 15.0 per cent, determined on 0.200 g.

ASSAY

Dissolve 0.150 g in 50 mL of anhydrous acetic acid R, with

slight heating if necessary. Titrate with 0.1 M perchloric acid,

Refined Safflower Oil
(Ph. Eur. monograph 2088)

(Ph. Eur. monograph 2088)

Fatty oil obtained from seeds of *Carthamus tinctorius* L. (type I) or from seeds of hybrids of *Carthamus tinctorius* L. (type II), by expression and/or extraction followed by re-refining. Type II refined safflower oil is rich in oleic (*cis*-9-octadecenoic) acid. A suitable antioxidant may be added.

Appearance
Clear, viscous, yellow or pale yellow liquid.

Practically insoluble in ethanol (96 per cent), miscible with light petroleum (bp: 40-60 °C).

Relative density	about 0.922	about 0.914
Refractive index	about 1.476	about 1.472

First identification *B*
Second identification *A*
A. Identification of fatty oils by thin-layer chromatography (2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram for type I or type II shown in Figure 2,3,2.-1.

Peroxide value (2.5.5, *Method A*)
Maximum 0.5,
Maximum 10.0, or maximum 5.0 if intended for use in the
manufacture of parenteral preparations.

Alkaline impurities (2.4.19)
It complies with the test.

Composition of fatty acids (2.4.22, Method A)
Use the mixture of calibrating substances in Table 2.4.22-3.

Composition of the fatty-acid fraction of type 1 refined safflower oil:

—	saturated fatty acids of chain length less than C14; maximum 0.2 per cent;
—	myristic acid: maximum 0.2 per cent;
—	palmitic acid: 4.0 per cent to 10.0 per cent;
—	stearic acid: 1.0 per cent to 5.0 per cent.

[illegible]

Water (2.5.32)
Maximum 0.1 per cent, determined on 1.00 g.

Maximum 0.1 per cent, determined on 1.00 g.

Brassicasterol (2.4.23)
Maximum 0.3 per cent in the sterol fraction of the oil.

0.2 per cent;
myristic acid: maximum 0.2 per cent;
palmitic acid: 3.6 per cent to 6.0 per cent;
stearic acid: 1.0 per cent to 5.0 per cent;
oleic acid: 70.0 per cent to 84.0 per cent;
linolic acid: 7.0 per cent to 23.0 per cent;
linolenic acid: maximum 0.5 per cent;
arachidic acid: maximum 1.0 per cent;
tricosenoic acid: maximum 1.0 per cent;
behenic acid: maximum 1.2 per cent.

oil: Composition of the fatty-acid fraction of type II refined safflower — saturated fatty acids of chain length less than C14: maximum

— *linoleic acid*: 68.0 per cent to 83.0 per cent;
— *oleic acid*: 8.0 per cent to 21.0 per cent;
— *linolenic acid*: maximum 0.5 per cent;
— *arachidic acid*: maximum 0.5 per cent;
— *stearic acid*: maximum 0.5 per cent;
— *behenic acid*: maximum 1.0 per cent.

oleic acid: 8.0 per cent to 21.0 per cent;
linoleic acid: 68.0 per cent to 83.0 per cent;
linolenic acid: maximum 0.5 per cent;
arachidic acid: maximum 0.5 per cent;
eicosenoic acid: maximum 0.5 per cent;

- *oleic acid*: 8.0 per cent to 21.0 per cent;
- *linoleic acid*: 68.0 per cent to 83.0 per cent;
- *linolenic acid*: maximum 0.5 per cent;

— *oleic acid*: 8.0 per cent to 21.0 per cent;
— *linoleic acid*: 68.0 per cent to 83.0 per cent;

2016

IDENTIFICATION**First identification: B.****Second identification: A, C, D****A.** Ultraviolet and visible absorption spectrophotometry (2.2.25).**Test solution** Dissolve 80.0 mg in a 10 g/L solution of**hydrochloric acid R** and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of the solution to 100.0 mL with a10 g/L solution of **hydrochloric acid R**.**Spectral range** 230-350 nm.**Absorption maximum** At 276 nm.**Specific absorbance at the absorption maximum** 66 to 75.**B.** Infrared absorption spectrophotometry (2.2.24).**Comparison salbutamol CRS.****C.** Thin-layer chromatography (2.2.27).**Test solution** Dissolve 10 mg of the substance to be examined in **methanol R** and dilute to 50 mL with the same solvent.**Reference solution** Dissolve 10 mg of **salbutamol CRS** in**methanol R** and dilute to 50 mL with the same solvent.**Plate** TLC silica gel plate **R**.**Mobile phase** concentrated **ammonia R**, **water R**, **ethyl acetate R**,**2-propanol R**, **methyl isobutyl ketone R**

(3:18:35:45:50 V/V/V/V/V).

Application 5 µL.**Development** Over 3/4 of the plate.**Drying** In air.**Detection** Spray with a 1 g/L solution of **methylbenzothiazolone****hydrazone R** in a 90 per cent V/V solution of**methanol R**, followed by a 20 g/L solution of **potassium****ferricyanide R** in a mixture of 1 volume of concentrated**ammonia R** and 3 volumes of **water R**, followed by a furtherspraying with a 1 g/L solution of **methylbenzothiazolone****hydrazone R** in a 90 per cent V/V solution of**methanol R**. The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

reference solution.

D. Dissolve about 10 mg in 50 mL of a 20 g/L solution of**disodium tetraborate R**. Add 1 mL of a 30 g/L solution of**aminopyrazolone R**, 10 mL of **methylene chloride R** and 10 mLof a 20 g/L solution of **potassium ferricyanide R**. Shake and

allow to separate. An orange-red colour develops in the

TESTS**Solution S**Dissolve 0.50 g in **methanol R** and dilute to 25.0 mL with the

same solvent.

Appearance of solutionSolution **S** is clear (2.2.1) and not more intensely colouredthan reference solution **BY₅** (2.2.2, *Method II*).**Optical rotation** (2.2.7)-0.10° to + 0.10°, determined on solution **S**.**Related substances****Liquid chromatography** (2.2.29).

examined in the mobile phase and dilute to 50.0 mL with

the mobile phase.

Reference solution (a) Dissolve 2.0 mg of **salbutamol CRS**,2 mg of **salbutamol impurity B CRS**, 3.0 mg of **salbutamol****impurity D CRS**, 3.0 mg of **salbutamol impurity F CRS** and3.0 mg of **salbutamol impurity G CRS** in the mobile phase and

dilute to 10.0 mL with the mobile phase. Dilute 2.0 mL of

this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of**salbutamol impurity I CRS** in 1.0 mL of the mobile phase.**Reference solution (c)** Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

Column:

solution to 20.0 mL with the mobile phase.

— **size:** $l = 0.15$ m, $\varnothing = 3.9$ mm;— **stationary phase:** spherical end-capped octylsilyl silica gel for**chromatography R** (5 µm) with a specific surface area of335 m²/g, a pore size of 10 nm and a carbon loading of

1.7 per cent.

Mobile phase Mix 22 volumes of **acetonitrile R** and78 volumes of a solution containing 2.87 g/L of **sodium****heptanesulfonate R** and 2.5 g/L of **potassium dihydrogen****phosphate R** previously adjusted to pH 3.65 with **dilute****phosphoric acid R**.**Flow rate** 1 mL/min.**Detection** Spectrophotometer at 220 nm.**Injection** 20 µL.**Run time** 25 times the retention time of **salbutamol**.**Identification of impurities** Use the chromatogram obtained

with reference solution (a) to identify the peaks due to

impurities **B**, **D**, **F** and **G**; use the chromatogram obtained

with reference solution (b) to identify the peak due to

impurity I.**Relative retention** With reference to **salbutamol** (retentiontime = about 2 min): **impurity B** = about 1.3;**impurity A** = about 1.7; **impurity C** = about 2.0;**impurity D** = about 2.7; **impurity H** = about 3.0;**impurity E** = about 3.1; **impurity G** = about 4.1;**impurity F** = about 6.2; **impurity I** = about 23.2.**System suitability:** reference solution (a):— **resolution:** minimum 3.0 between the peaks due to**salbutamol** and **impurity B**.**Limits:**— **impurity D:** not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (a) (0.3 per cent);

— **impurity F:** not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (a) (0.3 per cent);

— **impurity G:** not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (a) (0.3 per cent);

— **impurities A, B, C, E, H, I:** for each impurity, not morethan 1.5 times the area of the peak due to **salbutamol** in

the chromatogram obtained with reference solution (a)

— **unspecified impurities:** for each impurity, not more than0.5 times the area of the peak due to **salbutamol** in the

chromatogram obtained with reference solution (c)

— **total:** maximum 1.0 per cent;— **disregard limit:** the area of the principal peak in the

chromatogram obtained with reference solution (c)

Impurity J

Maximum 0.2 per cent.

Dissolve 50.0 mg in a 1 g/L solution of **hydrochloric acid R**

and dilute to 25.0 mL with the same solvent.

The absorbance (2.2.25) of the solution measured at 310 nm is not greater than 0.10.

Boron

Maximum 50 ppm.

Test solution To 50 mg of the substance to be examined add 5 mL of a solution containing 13 g/L of anhydrous sodium carbonate R and 17 g/L of potassium carbonate R. Evaporate to dryness on a water-bath and dry at 120 °C. Ignite the residue rapidly until the organic matter has been destroyed, allow to cool and add 0.5 mL of water R and 3.0 mL of a freshly prepared 1.25 g/L solution of curcumin R in glacial acetic acid R. Warm gently to effect solution, allow to cool and add 3.0 mL of a mixture prepared by adding 5 mL of sulfuric acid R, slowly and with stirring, to 5 mL of glacial acetic acid R. Mix and allow to stand for 30 min. Dilute to 100.0 mL with ethanol (96 per cent) R, filter and use the filtrate.

Reference solution Dissolve 0.572 g of boric acid R in 1000.0 mL of water R. Dilute 1.0 mL of the solution to 100.0 mL with water R. To 2.5 mL of this solution add 5 mL of a solution containing 13 g/L of anhydrous sodium carbonate R and 17 g/L of potassium carbonate R, and treat this mixture in the same manner as the test solution. Measure the absorbance (2.2.25) of the test solution and of the reference solution at the absorption maximum at about 555 nm. The absorbance of the test solution is not greater than that of the reference solution.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

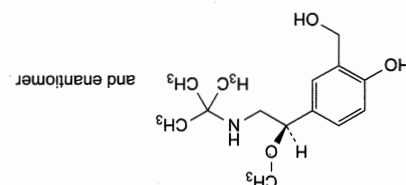
Dissolve 0.200 g in 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 23.93 mg of $C_{13}H_{21}NO_3$.

STORAGE

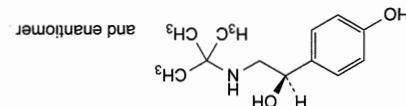
Protected from light.

IMPURITIES

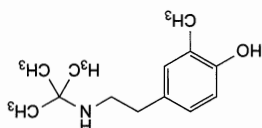
Specified impurities: A, B, C, D, E, F, G, H, I, J.



A. 5-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-methoxyethyl]-2-hydroxyphenylmethanol,

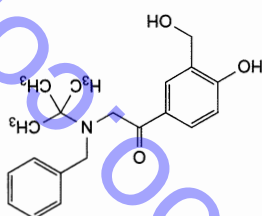


B. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxyphenyl)ethanol,

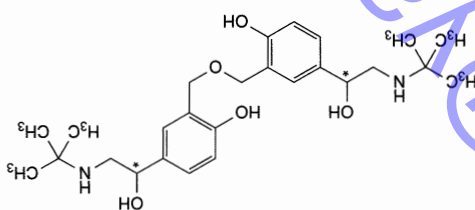


H. 4-[2-[(1,1-dimethylethyl)amino]ethyl]-2-methylphenol,

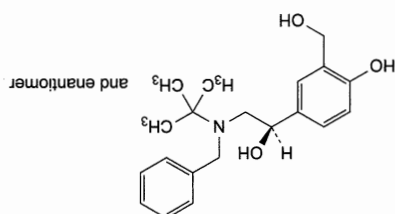
G. 2-[benzyl(1,1-dimethylethyl)amino]-1-[4-hydroxy-3-(hydroxymethyl)phenyl]ethanone,



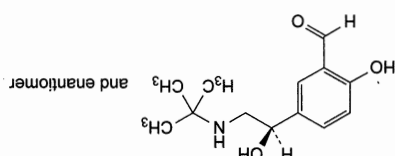
F. 1,1'-[oxybis(methylene(4-hydroxy-1,3-phenylene))]bis[2-[(1,1-dimethylethyl)amino]ethanol],



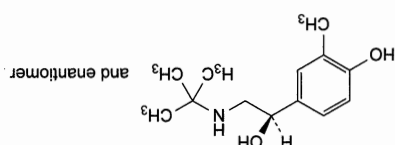
E. (1R,2S)-2-[benzyl(1,1-dimethylethyl)amino]-1-[4-hydroxy-3-(hydroxymethyl)phenyl]ethanol,



D. 5-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-hydroxyethyl]-2-hydroxybenzaldehyde,



C. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxy-3-methylphenyl)ethanol,



IDENTIFICATION

First identification B, E.

Second identification A, C, D, E

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 80.0 mg in a 10 g/L solution of hydrochloric acid R and dilute to 100.0 mL with the same acid. Dilute 10.0 mL of the solution to 100.0 mL with a 10 g/L solution of hydrochloric acid R.

Spectral range 230-350 nm.

Absorption maximum At 276 nm.

Specific absorbance at the absorption maximum 55 to 64.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison salbutamol sulfate CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in anhydrous ethanol R. Dry the residues and record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 12 mg of the substance to be examined in water R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 12 mg of salbutamol sulfate CRS in water R and dilute to 10 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, water R, ethyl acetate R, 2-propanol R, methyl isobutyl ketone R

(3:18:35:45:50 V/V/V/V/V).

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 1 g/L solution of methylbenzothiazolone hydrazone hydrochloride R in a 90 per cent V/V solution of methanol R, followed by a 20 g/L solution of potassium ferricyanide R in a mixture of 1 volume of concentrated ammonia R1 and 3 volumes of water R, followed by a further spraying with a 1 g/L solution of methylbenzothiazolone hydrazone hydrochloride R in a 90 per cent V/V solution of methanol R.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Dissolve about 10 mg in 50 mL of a 20 g/L solution of disodium tetraborate R. Add 1 mL of a 30 g/L solution of aminopyrazolone R, 10 mL of methylene chloride R and 10 mL of a 20 g/L solution of potassium ferricyanide R. Shake and allow to separate. An orange-red colour develops in the methylene chloride layer.

E. It gives reaction (a) of sulfates (2.3.1).

TESTS**Solution S**

Dissolve 0.250 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

Acidity or alkalinity

To 10 mL of solution S add 0.15 mL of methyl red solution R and 0.2 mL of 0.01 M sodium hydroxide. The solution is

It shows polymorphism (5.9).

Freely soluble in water, practically insoluble or very slightly soluble in ethanol (96 per cent) and in methylene chloride.

Solubility

White or almost white, crystalline powder.

Appearance**CHARACTERS**

98.0 per cent to 101.0 per cent (dried substance).

Content

Bis[(1R,5)-2-[(1,1-dimethylethyl)amino]-1-[4-hydroxy-3-(hydroxymethyl)phenyl]ethanol] sulfate.

DEFINITION

Ph Eur

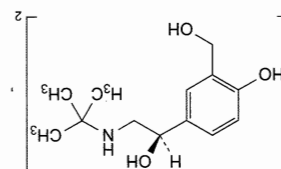
Salbutamol Inhalation Powder, pre-dispensed
Salbutamol Inhalation Powder
Salbutamol Injection
Salbutamol Nebuliser Solution
Salbutamol Oral Solution
Salbutamol Pressurised Inhalation
Salbutamol Tablets
Prolonged-release Salbutamol Capsules
Prolonged-release Salbutamol Tablets

Preparations

Beta₂-adrenoreceptor agonist; bronchodilator.

Action and use

C₂₆H₄₄N₂O₁₀S 576.7 51022-70-9



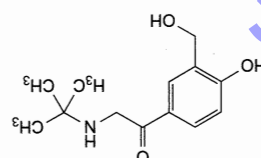
(Ph Eur monograph 0687)

Salbutamol Sulphate

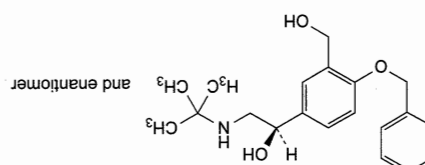
Salbutamol Sulfate

Ph Eur

1, 2-[(1,1-dimethylethyl)amino]-1-[4-hydroxy-3-(hydroxymethyl)phenyl]ethanol (salbutamone).



1, (1R,5)-2-[(1,1-dimethylethyl)amino]-1-[4-(benzyloxy)-3-(hydroxymethyl)phenyl]ethanol,



yellow. Not more than 0.4 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to red.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in mobile phase A and dilute to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 3.0 mg of salbutamol impurity D CRS and 3.0 mg of salbutamol impurity F CRS in

mobile phase A and dilute to 50.0 mL with mobile phase A.

Dilute 2.0 mL of the solution to 100.0 mL with mobile phase A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this

solution to 10.0 mL with mobile phase A.

Reference solution (c) With the aid of ultrasound, dissolve the contents of a vial of salbutamol impurity J CRS in 1.0 mL of the test solution.

Reference solution (d) Dissolve 1 mg of salbutamol impurity D CRS in mobile phase A and dilute to 100.0 mL

with mobile phase A.

Reference solution (e) Dissolve 4 mg of salbutamol sulfate for system suitability CRS (containing impurities C, F, N and O) in mobile phase A, add 0.4 mL of reference solution (d) and

dilute to 10.0 mL with mobile phase A.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: spherical end-capped octylsilyl silica gel for chromatography R (3 μ m);

— temperature: 30 °C.

Mobile phase:

— mobile phase A: dissolve 3.45 g of sodium dihydrogen phosphate monohydrate R in 900 mL of a

0.05 per cent V/V solution of triethylamine R, adjust to

pH 3.0 with dilute phosphoric acid R and dilute to

1000 mL with a 0.05 per cent V/V solution of

triethylamine R;

— mobile phase B: methanol R, acetonitrile R (35:65 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	95	5
5 - 30	95 \rightarrow 10	5 \rightarrow 90

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 μ l of the test solution and reference solutions (a), (b), (c) and (e).

Relative retention With reference to salbutamol (retention

time = about 7 min): impurity J = about 0.9; impurity

C = about 1.6; impurity N = about 1.67;

impurity D = about 1.68; impurity F = about 1.77;

impurity O = about 1.82.

Identification of impurities Use the chromatogram supplied

with salbutamol sulfate for system suitability CRS and the

chromatogram obtained with reference solution (e) to identify

the peaks due to impurities C, D, F, N and O; use the

chromatogram obtained with reference solution (c) to identify

the peak due to impurity J.

System suitability:

— peak-to-valley ratio: minimum 1.2, where H_p = height

above the baseline of the peak due to impurity N and

H_o = height above the baseline of the lowest point of the

curve separating this peak from the peak due to impurity D in the chromatogram obtained with reference solution (e); minimum 2.0, where H_p = height above the baseline of the peak due to impurity J and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to salbutamol in the chromatogram obtained with reference solution (c).

Limits:

— impurities D, F: for each impurity, not more than the area

of the corresponding peak in the chromatogram obtained

with reference solution (a) (0.3 per cent);

— impurities C, N, O: for each impurity, not more than twice

the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: maximum 0.9 per cent;

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Boron

Maximum 50 ppm.

Test solution To 50 mg of the substance to be examined add

5 mL of a solution containing 13 g/L of anhydrous sodium

carbonate R and 17 g/L of potassium carbonate R. Evaporate to

dryness on a water-bath and dry at 120 °C. Ignite the residue

rapidly until the organic matter has been destroyed, allow to

cool and add 0.5 mL of water R and 3.0 mL of a freshly

prepared 1.25 g/L solution of curcumin R in glacial acetic

acid R. Warm gently to effect solution, allow to cool and add

3.0 mL of a mixture prepared by adding 5 mL of sulfuric

acid R, slowly and with stirring, to 5 mL of glacial acetic

acid R. Mix and allow to stand for 30 min. Dilute to

100.0 mL with ethanol (96 per cent) R, filter and use the

filtrate.

Reference solution Dissolve 0.572 g of boric acid R in

1000.0 mL of water R. Dilute 1.0 mL of the solution to

100.0 mL with water R. To 2.5 mL of this solution add

5 mL of a solution containing 13 g/L of anhydrous sodium

carbonate R and 17 g/L of potassium carbonate R, and treat

this mixture in the same manner as the test solution.

Measure the absorbance (2.2.25) of the test solution and of

the reference solution at the absorption maximum at about

555 nm. The absorbance of the test solution is not greater

than that of the reference solution.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 5 mL of anhydrous formic acid R and add

35 mL of anhydrous acetic acid R. Titrate with 0.1 M

(2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 57.67 mg

of $C_{26}H_{44}N_{2}O_{10}S$.

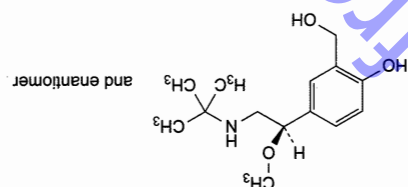
STORAGE

Protected from light.

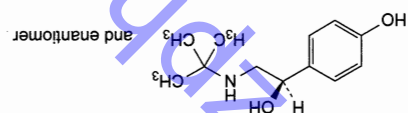
IMPURITIES

Specified impurities C, D, F, N, O

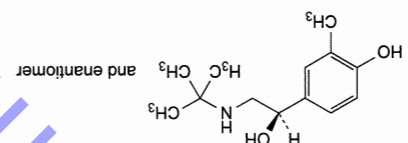
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, E, G, I, J, K, L, M.



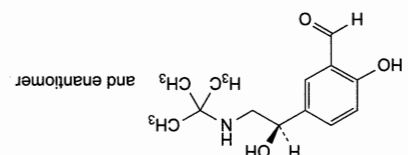
A. [5-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-methoxyethyl]-2-hydroxyphenyl]methanol,



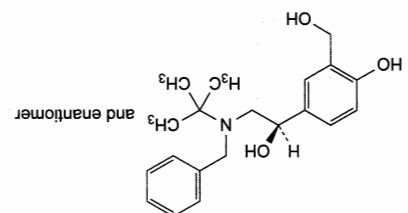
B. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxyphenyl)ethanol,



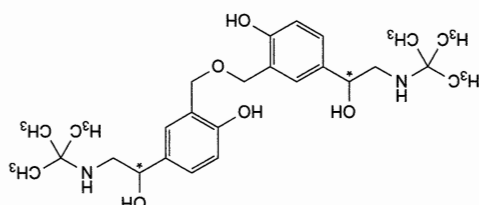
C. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxy-3-methylphenyl)ethanol,



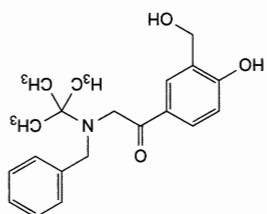
D. 5-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-hydroxyethyl]-2-hydroxybenzaldehyde,



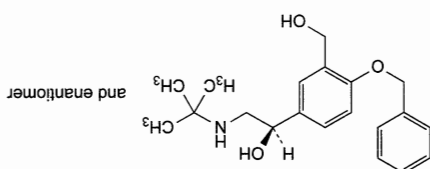
E. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxy-3-phenyl)ethanol,



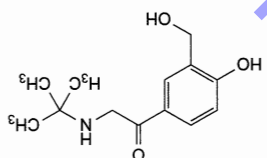
F. 1,1'-[oxybis(methylene(4-hydroxy-1,3-phenylene))]bis[2-[(1,1-dimethylethyl)amino]ethanol],



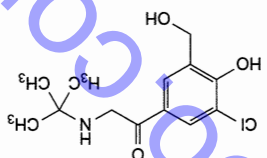
G. 2-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxy-3-phenyl)ethanol],



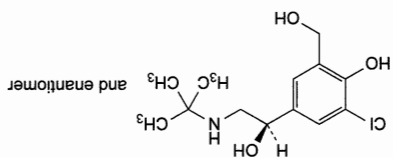
I. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-(benzyloxy)-3-hydroxyphenyl)ethanol,



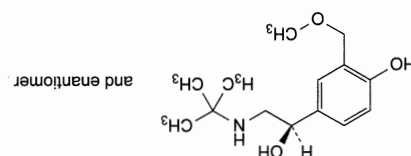
J. 2-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(4-hydroxy-3-phenyl)ethanol],



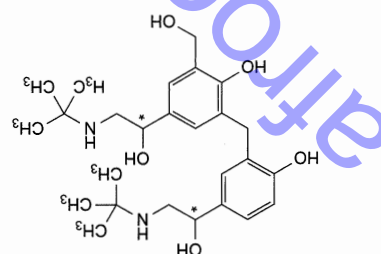
K. 2-[(1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(3-chloro-4-hydroxy-5-phenyl)ethanol],



L. (1R,2S)-2-[(1,1-dimethylethyl)amino]-1-(3-chloro-4-hydroxy-5-phenyl)ethanol,



M. (1R,2S)-2-[(1S,1-dimethylethyl)phenyl]-1-[4-hydroxy-3-(methoxymethyl)phenyl]ethanol,

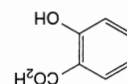


N, 2-[(1,1-dimethylethyl)amino]-1-[3-[[5-[2-[(1,1-dimethylethyl)amino]-1-hydroxy-5-hydroxyphenyl]methoxy]phenyl]ethanol, (hydroxymethyl)phenyl]ethanol, O, unknown structure.

Ph Eur

Salicylic Acid

(Ph. Eur. monograph 0366)



138.1

69-72-7

C₇H₆O₃

Action and use

Keratolytic.

Preparations

Coal Tar and Salicylic Acid Ointment
Dithranol and Salicylic Acid Ointment
Salicylic Acid Colloidion
Salicylic Acid Cream
Salicylic Acid Ointment
Zinc and Salicylic Acid Paste

DEFINITION

2-Hydroxybenzenecarboxylic acid.

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or white or colourless, acicular crystals.

Solubility

Slightly soluble in water, freely soluble in ethanol (96 per cent), sparingly soluble in methylene chloride.

IDENTIFICATION

First identification A, B

Second identification A, C

A. Melting point (2.2.14): 158 °C to 161 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison salicylic acid CRS.

C. Dissolve about 30 mg in 5 mL of 0.05 M sodium hydroxide, neutralise if necessary and dilute to 20 mL with water R. 1 mL of the solution gives reaction (a) of salicylates (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in 50 mL of boiling distilled water R, cool and filter.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 1 g in 10 mL of ethanol (96 per cent) R.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.50 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 10 mg of phenol R (impurity C) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of salicylic acid impurity B CRS in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 50 mg of 4-hydroxybenzoic acid R (impurity A) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (e) Dilute a mixture of 1.0 mL of each of reference solutions (a), (b) and (c) to 10.0 mL with the mobile phase.

Reference solution (f) Dilute a mixture of 0.1 mL of each of reference solutions (a), (b) and (c) to 10.0 mL with the mobile phase.

Column: mobile phase.

Flow rate 0.5 mL/min.
(1:40:60 V/V/V).

Detection Spectrophotometer at 270 nm.
Injection 10 µL of the test solution and reference solutions (d), (e) and (f).

Relative retention With reference to impurity C: impurity A = about 0.70; impurity B = about 0.90. System suitability: reference solution (e): the 3rd peak in the chromatogram corresponds to the peak due to phenol in the chromatogram obtained with reference solution (d);

resolution: minimum 1.0 between the peaks due to impurities B and C; if necessary, adjust the quantity of acetic acid in the mobile phase.

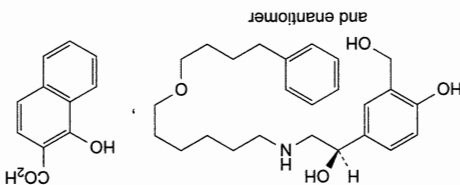
Limits:

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (0.1 per cent);



Salmeterol Xinafoate

(Ph. Eur. monograph 1765)



604

C₃₆H₄₅NO₇

94749-08-3

Action and use

Beta₂-adrenoceptor agonist; bronchodilator.

Preparations

Fluticasone & Salmeterol Inhalation Powder, pre-dispensed
Fluticasone & Salmeterol Pressurised Inhalation, suspension

DEFINITION

(1R,5S)-1-[4-Hydroxy-3-(hydroxymethyl)phenyl]-2-[[6-(4-phenylbutoxy)hexyl]amino]ethanol 1-hydroxynaphthalene-2-carboxylate.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, soluble in methanol, slightly soluble in anhydrous ethanol, practically insoluble in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison salmeterol xinafoate CRS.

TESTS

Related substances

Liquid chromatography (2.2.29). Protect the solutions from light.

Solvent mixture acetonitrile R, water R (50:50 V/V).

Test solution (a) Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 100.0 mL with mobile phase A.

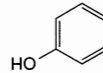
Reference solution (a) Dissolve 11 mg of salmeterol xinafoate for system suitability CRS (containing impurities E and G) in the solvent mixture and dilute to 2.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of Test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

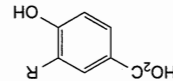
Reference solution (c) Dissolve 5 mg of salmeterol xinafoate for peak identification CRS (containing impurity D) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Reference solution (d) Dissolve 25.0 mg of salmeterol xinafoate CRS in mobile phase A and dilute to 100.0 mL with mobile phase A.

C. phenol.



A. R = H: 4-hydroxybenzoic acid,
B. R = CO₂H: 4-hydroxyisophthalic acid,



Specified impurities: A, B, C.

IMPURITIES

Protected from light.

STORAGE

C₇H₆O₃.

1 mL of 0.1 M sodium hydroxide is equivalent to 13.81 mg of 0.1 mL of phenol red solution R as indicator.

Dissolve 0.120 g in 30 mL of ethanol (96 per cent) R and add 20 mL of water R. Titrate with 0.1 M sodium hydroxide, using

ASSAY

Maximum 0.1 per cent, determined on 2.0 g.

Sulfated ash (2.4.14)

a desiccator.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

15 volumes of ethanol (96 per cent) R.

(100 ppm Pb) R with a mixture of 5 volumes of water R and (2 ppm Pb) prepared by diluting lead standard solution

Prepare the reference solution using lead standard solution 5 mL of water R. 12 mL of the solution complies with test B.

Dissolve 2.0 g in 15 mL of ethanol (96 per cent) R and add

Maximum 20 ppm.

Heavy metals (2.4.8)

water R and 5 mL of dimethylformamide R.

25 per cent m/m solution of barium chloride R, 3 mL of

0.2 mL of dilute hydrochloric acid R, 0.5 mL of a

to 2 mL of sulfate standard solution (100 ppm SO₄) R add not more intense than that in a standard prepared as follows:

chloride R. After 15 min any opalescence in the solution is acid R and 0.5 mL of a 25 per cent m/m solution of barium

of water R. Mix thoroughly. Add 0.2 mL of dilute hydrochloric acid R and 4 mL

Dissolve 1.0 g in 5 mL of dimethylformamide R and add 4 mL

Maximum 200 ppm.

Sulfates

Dilute 10 mL of solution S to 15 mL with water R.

Maximum 100 ppm.

Chlorides (2.4.4)

the chromatogram obtained with reference solution (f). — disregard limit: 0.01 times the area of the principal peak in

solution (f) (0.2 per cent);

impurity A in the chromatogram obtained with reference

total: not more than twice the area of the peak due to

obtained with reference solution (f) (0.05 per cent);

area of the peak due to impurity B in the chromatogram

any other impurity: for each impurity, not more than the

solution (f) (0.02 per cent);

peak in the chromatogram obtained with reference

impurity C: not more than the area of the corresponding

solution (f) (0.05 per cent);

peak in the chromatogram obtained with reference

impurity B: not more than the area of the corresponding

Reference solution (e) Dilute 1.0 mL of reference solution (a) to 20.0 mL with mobile phase A.

Column:
— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:
— mobile phase A: mix 24 volumes of a 7.71 g/L solution of ammonium acetate R and 24 volumes of a 28.84 g/L solution of sodium dodecyl sulfate R and adjust to pH 2.7 with glacial acetic acid R; mix with 52 volumes of acetonitrile R;
— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 16	100	0
16 - 36	100 \rightarrow 30	0 \rightarrow 70
36 - 45	30	70

Flow rate 2 mL/min.

Detection Spectrophotometer at 278 nm.

Injection 20 μ L Of test solution (a) and reference solutions (a), (b) and (c).

Identification of impurities: use the chromatogram supplied with salmeterol xinafoate for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities E and G; use the chromatogram supplied with salmeterol xinafoate for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peak due to impurity D.

Relative retention With reference to salmeterol (retention time = about 13 min): xinafoic acid = about 0.2; impurity D = about 0.8; impurity E = about 0.9; impurity G = about 2.7.

System suitability: reference solution (a):
— peak-to-valley ratio: minimum 10, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to salmeterol.
Calculation of percentage contents:
— for each impurity, use the concentration of salmeterol in reference solution (b).

Limits:
— impurities D, G: for each impurity, maximum 0.2 per cent; unspecified impurities: for each impurity, maximum 0.10 per cent;
— total: maximum 0.5 per cent;
— reporting threshold: 0.05 per cent; disregard the peak due to xinafoic acid.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.00 g.

Sulfated ash (2.4.14)

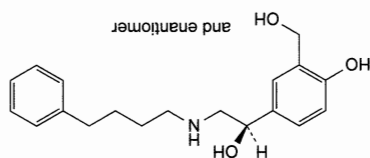
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

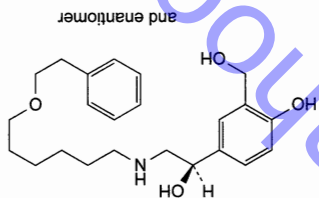
Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase A.

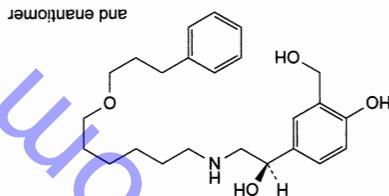
Injection 20 μ L of test solution (b) and reference solutions (d) and (e).



A. (1RS)-1-[4-hydroxy-3-(hydroxymethyl)phenyl]-2-[[6-[(4-phenylbutyl)amino]ethoxy]phenyl]-2-[[6-[(2-phenylethoxy)hexyl]amino]ethanol], and enantiomer



B. (1RS)-1-[4-hydroxy-3-(hydroxymethyl)phenyl]-2-[[6-[(2-phenylethoxy)hexyl]amino]ethoxy]phenyl]-2-[[6-[(2-phenylethoxy)hexyl]amino]ethanol], and enantiomer



C. (1RS)-1-[4-hydroxy-3-(hydroxymethyl)phenyl]-2-[[6-[(3-phenylpropoxy)hexyl]amino]ethoxy]phenyl]-2-[[6-[(3-phenylpropoxy)hexyl]amino]ethanol], and enantiomer

Specified impurities D, G.
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, E, F.

STORAGE

Protected from light.

IMPURITIES

Calculate the percentage content of $C_{36}H_{45}NO_7$ using the chromatogram obtained with reference solution (d) and taking into account the assigned content of salmeterol xinafoate CRS.

Run time Until complete elution of the peak due to salmeterol (about 16 min).
System suitability: reference solution (e):
— peak-to-valley ratio: minimum 10, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to salmeterol.

Content
— sum of the contents of EPA and DHA (expressed as triglycerides): 10.0 per cent to 28.0 per cent.

A suitable antioxidant may be added.

PRODUCTION

The fish shall only be given feed with a composition that is in accordance with the relevant European Union or other applicable regulations.

The content of dioxins and dioxin-like PCBs

(polychlorinated biphenyls) is controlled using methods and limits in accordance with the requirements set in the European Union or other applicable regulations.

The oil is produced by mechanical expression of fresh raw materials, either from the whole fish, or fish where the fillets have been removed, at a temperature not exceeding 100 °C, and without using solvents. After centrifugation, solid substances may be removed from the oil by cooling and filtering (winterisation).

CHARACTERS

Appearance

Pale pink liquid.

Solubility

Practically insoluble in water, very soluble in acetone and in heptane, slightly soluble in anhydrous ethanol.

IDENTIFICATION

Examine the ^{13}C NMR spectra obtained in the assay for positional distribution ($\beta(2)$ -acyl) of fatty acids. The spectra contain peaks between 172 ppm and 173 ppm with shifts similar to those in the type spectrum (Figure 1910-2). The oil to be examined complies with the limits of this assay.

TESTS

Absorbance (2.2.25)

Minimum 0.10, measured at the absorption maximum between 470 nm and 480 nm.

Dissolve 5.0 mL in 5.0 mL of trimethylpentane R.

Acid value (2.5.1)

Maximum 2.0.

Anisidine value (2.5.36)

Maximum 10.0.

Peroxide value (2.5.5, Method A)

Maximum 5.0.

Unsaponifiable matter (2.5.7)

Maximum 1.5 per cent, determined on 5.0 g.

Linoleic acid (2.4.29)

Maximum 11.0 per cent.

Identify the peak due to linoleic acid using the chromatogram in Figure 1910-1. Determine the percentage content by normalisation.

ASSAY

Positional distribution ($\beta(2)$ -acyl) of fatty acids

Apparatus High resolution FT-NMR spectrometer operating at minimum 300 MHz.
Test solution Dissolve 190-210 mg of fresh salmon oil in 500 μL of deuterated chloroform R. Prepare at least 3 samples and examine within 3 days.

Acquisition of ^{13}C NMR spectra The following parameters may be used:
— **sweep width:** 200 ppm (–5 to 195 ppm);
— **irradiation frequency offset:** 95 ppm;
— **time domain:** 64 K;

Farmed Salmon Oil

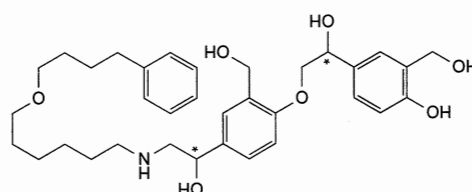
(Ph. Eur. monograph 1910)



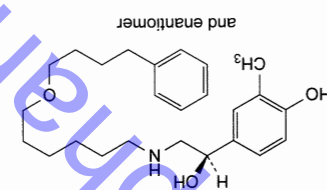
DEFINITION

Purified fatty oil obtained from fresh farmed *Salmo salar*. The positional distribution ($\beta(2)$ -acyl) is 60-70 per cent for ceronic (docosahexaenoic acid (C22:6 n-3; DHA), 25-35 per cent for timnodonic (eicosapentaenoic acid (C20:5 n-3; EPA) and 40-55 per cent for morocic acid (C18:4 n-3).

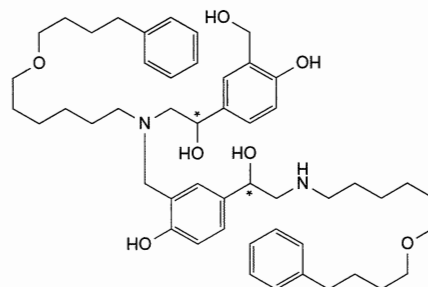
D. 1-[4-[2-hydroxy-2-[4-hydroxy-3-(hydroxymethyl)phenyl]ethoxy]-3-(hydroxymethyl)phenyl]-2-[[6-(4-phenylbutoxy)hexyl]amino]ethanol,



E. 1-[4-hydroxy-3-(hydroxymethyl)phenyl]-2-[[6-(1-methyl-3-phenylpropoxy)hexyl]amino]ethanol,



F. (1R,5S)-1-(4-hydroxy-3-methylphenyl)-2-[[6-(4-phenylbutoxy)hexyl]amino]ethanol,



G. 1-[4-hydroxy-3-[[[2-hydroxy-2-[4-hydroxy-3-(hydroxymethyl)phenyl]ethyl]-6-(4-phenylbutoxy)hexyl]amino]methyl]phenyl]-2-[[6-(4-phenylbutoxy)hexyl]amino]ethanol.

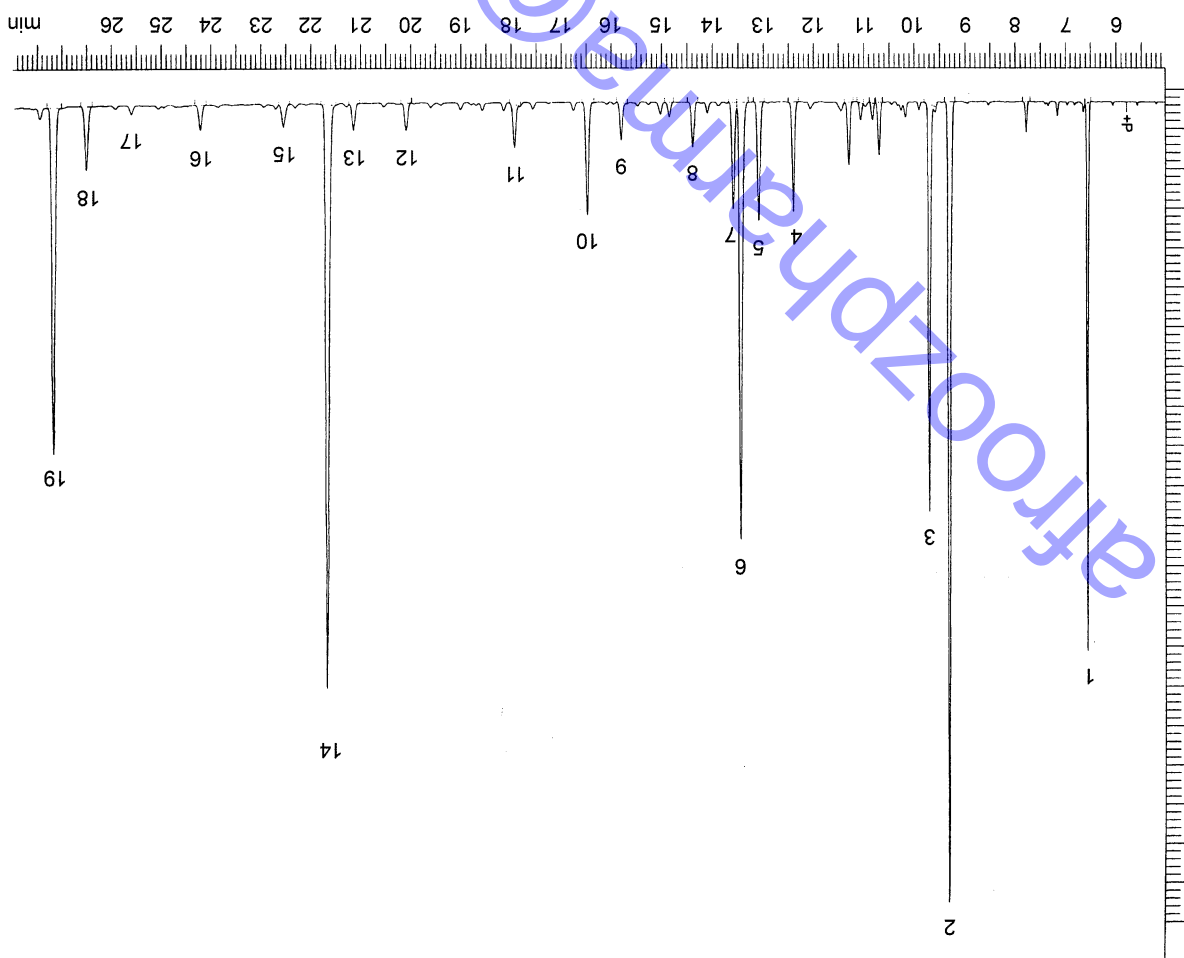


Figure 1910.-1. - Chromatogram for the composition of fatty acids in farmed salmon oil

— pulse delay: 2 s;

— pulse program: zgpg 30 (inverse gated, 30° excitation

— pulse);

— dummy scans: 4;

— number of scans: 4096.

Processing and plotting The following parameters may be used:

— size: 64 K (zero-filling);

— window multiplication: exponential;

— Lorentzian broadening factor: 0.2 Hz.

Use the CDCl₃ signal for shift referencing. The shift of the

central peak of the 1:1:1 triplet is set to 77.16 ppm.

Plot the spectral region δ 171.5-173.5 ppm. Compare the

spectrum with the reference spectrum in Figure 1910.-2.

The shift values lie within the ranges given in Table 1910.-1.

System suitability:

— signal-to-noise ratio: minimum 5 for the smallest relevant

peak corresponding to α C18:4 signal (in the range

δ 172.95-172.99 ppm);

— peak width at half-height maximum 0.02 ppm for the

central CDCl₃ signal (at δ 77.16 ppm).

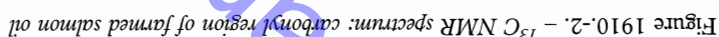
α = peak area of the corresponding α-carbonyl peak;
β = peak area of β-carbonyl peak from C22:6 n-3,
C20:5 n-3 or C18:4 n-3, respectively.

$$\frac{\alpha + \beta}{\beta} \times 100$$

following expression:

Calculation of positional distribution (β(2)-acyl) Use the

Signal	Shift range (ppm)
β DHA	172.05 - 172.09
α DHA	172.43 - 172.47
β EPA	172.52 - 172.56
α EPA	172.90 - 172.94
β C18:4	172.56 - 172.60
α C18:4	172.95 - 172.99



hydroxypropyl]-2-[(quinolin-2-ylcarbonyl)amino]butanediamide methanesulfonate.

97.5 per cent to 102.0 per cent (anhydrous substance).

consideration the principal risk management strategy for quality of starting together with considerations of starting

methods 2.5.37. Methyl, ethyl and isopropyl methanesulfonate in methanesulfonic acid, 2.5.38. Methyl, ethyl and isopropyl methanesulfonate in active substances and 2.5.39. Methanesulfonyl chloride in methanesulfonic acid are available to assist manufacturers.

White or almost white, slightly hygroscopic powder.

Practically insoluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent).

A. Specific optical rotation (see Tests).
B. Infrared absorption spectrophotometry (2.2.24).

Specific optical rotation (2.2.7)

Liquid chromatography (2.2.29).

Dissolve 0.25 g in *anhydrous methanol* R and dilute to 50.0 mL with the same solvent.

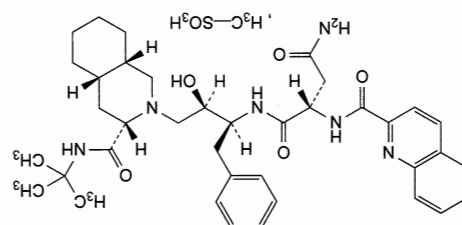
(2S)-N^t-[[(1S,2R)-1-Benzyl-3-[(3S,4aS,8aS)-3-[[[1,1-dimethylethyl]carbamoyl]octahydroisquinolin-2(1H)-yl]-2-

Protease inhibitor; antiviral (HIV).

$$\text{C}_{39}\text{H}_{54}\text{N}_6\text{O}_8\text{S}$$

197

149845-06-7



(Ph. Eur. monograph 2267)

Saquinavir Mesilate



Ph Eur

In an airtight, well-filled container, protected from light,

under inert gas.

See Figure 1910.-1.

EPA and DHA (2.4.29)

— *cervonic* (docosahexaenoic) acid (C22:6 n-3; DHA): 60 per cent to 70 per cent.
— *timnodonic* (eicosapentaenoic) acid (C20:5 n-3; EPA): 25 per cent to 35 per cent;
— *moronic* acid (C18:4 n-3): 40 per cent to 55 per cent.

Limits:

Solvent mixture water for chromatography R, acetonitrile R1 (47:53 V/V).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture, using sonication, and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of saquinavir for system suitability CRS (containing impurities A, B, C and D) in 1.0 mL of the solvent mixture and sonicate for 2 min.

Reference solution (c) Dissolve 30.0 mg of saquinavir mesilate CRS in the solvent mixture, using sonication, and dilute to 100.0 mL with the same solvent.

Column:

- size: $l = 0.15$ m, $\phi = 4.6$ mm;
- stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (3.5 μ m).

Mobile phase:

- mobile phase A: to 2.5 mL of strong sodium hydroxide solution R add 900 mL of water for chromatography R, adjust to pH 1.8 with perchloric acid R and dilute to 1000 mL with water for chromatography R;
- mobile phase B: mobile phase A, acetonitrile R1 (38:62 V/V);

Time (min)	Mobile phase A (per cent)	Mobile phase B (per cent)
0 - 1	50	50
1 - 31	50 \rightarrow 0	50 \rightarrow 100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 10 μ L of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied

with saquinavir for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B, C and D.

Relative retention With reference to saquinavir (retention

time = about 17 min): impurity A = about 0.2;

impurity B = about 0.3; impurity C = about 0.5;

impurity D = about 0.9.

System suitability: reference solution (b):

— *peak-to-valley ratio*: minimum 3, where H_p = height above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to

saquinavir.

— *correction factors*: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity A = 0.5;

impurity B = 0.5; impurity C = 2.5;

— *impurities A, B, C*: for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

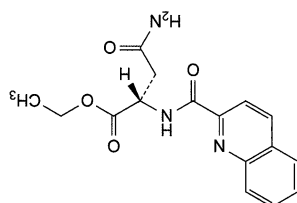
(0.15 per cent);

— *unspecified impurities*: for each impurity, not more than

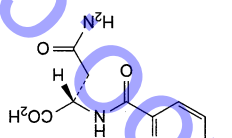
0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.05 per cent);



A. (2S)-4-amino-4-oxo-2-[(quinolin-2-yl)carbonyl]amino]butanoic acid,



F, G, H.

Control of impurities in substances for pharmaceutical use: D, E,

impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph *Substances for pharmaceutical use*

acceptance criterion for other/unspecified impurities and/or

the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities A, B, C

IMPURITIES

In an airtight container, protected from light.

STORAGE

the assigned content of saquinavir mesilate CRS.

Calculate the percentage content of saquinavir mesilate from

Injection 10 μ L of the test solution and reference solution (c).

related substances with the following modification.

Liquid chromatography (2.2.29) as described in the test for

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 1.0 per cent, determined on 0.250 g.

Water (2.5.12)

size 0.45 μ m).

Filter the solutions through a membrane filter (nominal pore

The solution may become yellow again after pH-adjustment.

using 0.5 mL of lead standard solution (10 ppm Pb) R.

0.50 g complies with test G. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

reference solution (a) (0.03 per cent).

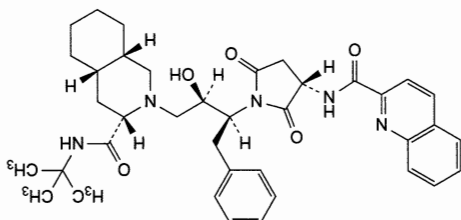
principal peak in the chromatogram obtained with

— *disregard limit*: not more than 0.3 times the area of the

(0.5 per cent);

in the chromatogram obtained with reference solution (a)

— *total*: not more than 5 times the area of the principal peak



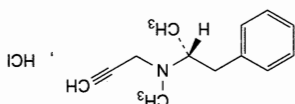
H. N-[(3S)-1-[(1S,2R)-1-benzyl-3-[(3S,4aS,8aS)-3-[(1,1-dimethylethyl)carbamoyl]octahydroisoquinolin-2(1H)-yl]-2-hydroxypropyl]-2,5-dioxopropylidene-3-yl]quinoline-2-carboxamide.

Ph Eur



(Ph Eur monograph 1260)

Selegiline Hydrochloride



C₁₃H₁₈ClN 223.7 14611-52-0

Action and use

Monoamine oxidase type B inhibitor; treatment of Parkinson's disease.

Preparations

Selegiline Oral Solution

Selegiline Tablets

Ph Eur

DEFINITION

N-Methyl-N'-[(1R)-1-methyl-2-phenylethyl]prop-2-yn-1-amine hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in methanol, slightly soluble in acetone and in ethyl acetate.

mp

About 143 °C.

IDENTIFICATION

Carry out either tests A, B, D or tests B, C, D.

A. Specific optical rotation (2.2.7): -12.0 to -10.0 (dried substance).

Dissolve 2.00 g in carbon dioxide-free water R and dilute to 20.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison selegiline hydrochloride CRS.

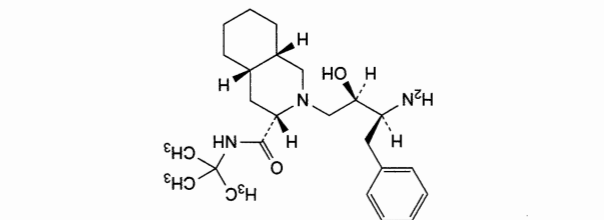
C. Enantiomeric purity (see Tests).

D. It gives reaction (a) of chlorides (2.3.1).

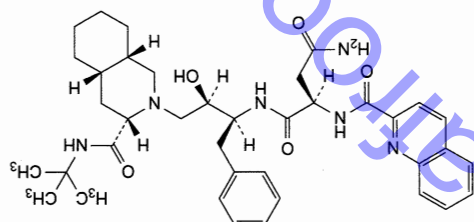
TESTS

pH (2.2.3)

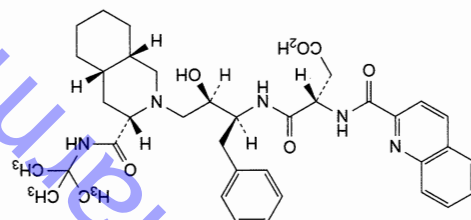
3.5 to 4.5.



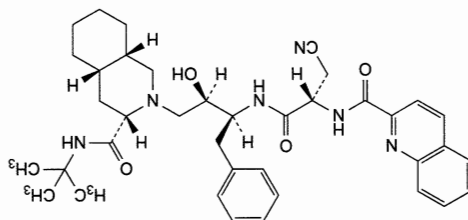
C. (3S,4aS,8aS)-2-[(2R,3S)-3-amino-2-hydroxy-4-phenylbutyl]-N-(1,1-dimethylethyl)decahydroisoquinoline-3-carboxamide,



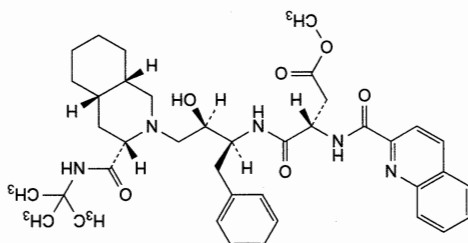
D. (2R)-N'-[(1S,2R)-1-benzyl-3-[(3S,4aS,8aS)-3-[(1,1-dimethylethyl)carbamoyl]octahydroisoquinolin-2(1H)-yl]-2-hydroxypropyl]-2-[(quinolin-2-yl)carbamoyl]butanediamide (2-epi-saquinavir),



E. (3S)-4-[[[(1S,2R)-1-benzyl-3-[(3S,4aS,8aS)-3-[(1,1-dimethylethyl)carbamoyl]octahydroisoquinolin-2(1H)-yl]-2-hydroxypropyl]amino]-4-oxo-3-[(quinolin-2-yl)carbamoyl]butanoic acid,



F. N-[(1S)-2-[(1S,2R)-1-benzyl-3-[(3S,4aS,8aS)-3-[(1,1-dimethylethyl)carbamoyl]octahydroisoquinolin-2(1H)-yl]-2-hydroxypropyl]amino]-1-(cyanomethyl)-2-oxoethyl]quinoline-2-carboxamide,



G. methyl (3S)-4-[[[(1S,2R)-1-benzyl-3-[(3S,4aS,8aS)-3-[(1,1-dimethylethyl)carbamoyl]octahydroisoquinolin-2(1H)-yl]-2-hydroxypropyl]amino]-4-oxo-3-[(quinolin-2-yl)carbamoyl]butanoate,

Dissolve 0.20 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Enantiomeric purity

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in a mixture of 10 µL of butylamine R and 1 mL of 2-propanol R and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 8.0 mg of (RS)-selegiline hydrochloride CRS in a mixture of 10 µL of butylamine R and 1 mL of 2-propanol R and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 0.5 mL of reference solution (a) to 20.0 mL with the mobile phase.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: cellulose derivative of silica gel for chiral separation R.

Mobile phase 2-propanol R, cyclohexane R (0.2:99.8 V/V).

Flow rate 1 mL/min.
Detection Spectrophotometer at 220 nm.

Injection 20 µL.
Relative retention With reference to (R)-selegiline (retention time = about 6 min): impurity E = about 0.9.

System suitability: reference solution (a):
— resolution: minimum 1.5 between the peaks due to impurity E and (R)-selegiline; if necessary, adjust the concentration of 2-propanol in the mobile phase.

Limit:
— impurity E: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.5 per cent).

Related substances
Liquid chromatography (2.2.29).

Butylammonium acetate buffer solution Dilute 4 mL of butylamine R in 900 mL of water R, adjust to pH 6.5 with acetic acid R and dilute to 1000.0 mL with water R.

Test solution Dissolve 20 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 50 mg of the substance to be examined and 10 mg of butyl parahydroxybenzoate R in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 µm).

Mobile phase acetonitrile R1, butylammonium acetate buffer solution (50:50 V/V).

Flow rate 1 mL/min.
Detection Spectrophotometer at 215 nm.

Injection 20 µL.
Run time 1.7 times the retention time of selegiline.

Relative retention With reference to selegiline (retention time = about 14 min): butyl parahydroxybenzoate = about 0.8.

System suitability: reference solution (a):
— resolution: minimum 3.0 between the peaks due to butyl parahydroxybenzoate and selegiline.

Limits:
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)
Maximum 20 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb).

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying at 60 °C at a pressure not exceeding 0.5 kPa for 3 h.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.180 g in 50 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 22.37 mg of $C_{13}H_{18}ClN$.

STORAGE
Protected from light.

IMPURITIES
Specified impurities E.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, B, C, D, G.

A. (2R)-N-methyl-1-phenylpropan-2-amine ((R)-metamfetamine),

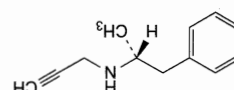


B. (2R)-1-phenylpropan-2-amine (amfetamine),

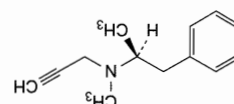


C. (1R,2S)-2-amino-1-phenylpropan-1-ol (phenylpropolanine),

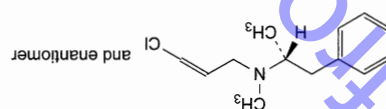




D, N-[(1R)-1-methyl-2-phenylethyl]prop-2-yn-1-amine (desmethylelegriline),



E, N-methyl-N-[(1S)-1-methyl-2-phenylethyl]prop-2-yn-1-amine,



G, (2E)-3-chloro-N-methyl-N-[(1R,S)-1-methyl-2-phenylethyl]prop-2-en-1-amine.

Ph Eur

Selenium Sulfide



Selenium Sulphide

(Selenium Disulfide, Ph Eur monograph 1147)

SeS₂ 143.1 7488-56-4

Action and use

Used in treatment of dandruff and seborrhoeic dermatitis of

the scalp.

Preparation

Selenium Sulfide Scalp Application

DEFINITION

Content 52.0 per cent to 55.5 per cent of Se.

CHARACTERS

Appearance

Bright orange or reddish-brown powder.

Solubility

Practically insoluble in water.

IDENTIFICATION

A. Gently boil about 50 mg with 5 mL of nitric acid R for 30 min. Dilute to 50 mL with water R and filter. To 5 mL of the filtrate add 10 mL of water R and 5 g of urea R. Heat to boiling, cool and add 1.5 mL of potassium iodide solution R. A yellow or orange colour is produced which darkens rapidly on standing. This solution is used in identification test B.

B. Allow the coloured solution obtained under identification A to stand for 10 min and filter through filter paper for chromatography R. 5 mL of the filtrate gives reaction (a) of sulfates (2.3.1).

TESTS

Soluble selenium compounds

Maximum 5 ppm, calculated as Se.

To 10 g add 100 mL of water R, mix well, allow to stand for 1 h with frequent shaking and filter. To 10 mL of the filtrate add 2 mL of a 115 g/L solution of anhydrous formic acid R.

Action and use

Amino acid.

Ph Eur

DEFINITION

(2S)-2-Amino-3-hydroxypropionic acid.

Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison serine CRS.

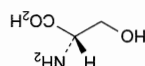
C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a 1 per cent V/V solution of hydrochloric acid R and dilute to 50 mL with the same solution.

Serine



(Ph. Eur. monograph 0788)



105.1

56-45-1

Ph Eur

Se.

To 0.100 g add 25 mL of fuming nitric acid R and heat on a water-bath for 1 h; a small insoluble residue may remain. Cool and dilute to 100.0 mL with water R. To 25.0 mL of this solution add 50 mL of water R and 5 g of urea R and heat to boiling. Cool, add 7 mL of potassium iodide solution R and 3 mL of starch solution R. Titrate immediately with 0.1 M sodium thiosulfate. Carry out a blank titration. 1 mL of 0.1 M sodium thiosulfate is equivalent to 1.974 mg of Se.

ASSAY

dilute to 50 mL with water R and adjust to pH 2.0-3.0 with an 115 g/L solution of anhydrous formic acid R. Add 2 mL of a 5 g/L solution of 3,3'-diaminobenzidine tetrahydrochloride R. Allow to stand for 45 min and then adjust to pH 6.0-7.0 with dilute ammonia R1. Shake the solution for 1 min with 10 mL of toluene R and allow the phases to separate. The absorbance (2.2.25) of the upper layer measured at 420 nm is not greater than that of a standard prepared at the same time and in the same manner beginning at the words "add 2 mL of an 115 g/L solution of anhydrous formic acid R" and using 5 mL of selenium standard solution (1 ppm Se) R instead of 10 mL of the filtrate.

Reference solution Dissolve 10 mg of serine CRS in a 1 per cent V/V solution of hydrochloric acid R and dilute to 50 mL with the same solution.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. To 1 mL of a 10 g/L solution of the substance to be examined in a test tube, add 5 mL of a 20 g/L solution of sodium periodate R. Heat on a water-bath and collect the vapour on glass wool moistened with water R and inserted in the opening of the test tube. After heating for 5 min, transfer the glass wool to a test tube containing 1 mL of a 15 g/L solution of chromotropic acid, sodium salt R and 3 mL of sulphuric acid R. Heat on a water-bath for 10 min. A violet-red colour is produced.

TESTS

Solution S

Dissolve 2.5 g in distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Specific optical rotation (2.2.7)

+ 14.0 to + 16.0 (dried substance).

Dissolve 2.50 g in dilute hydrochloric acid R and dilute to 25.0 mL with the same acid.

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method I.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of proline R in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of ammonium standard solution (100 ppm NH₄) R to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of isoleucine R and 30 mg of leucine R in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):

— **resolution:** minimum 1.5 between the peaks due to isoleucine and leucine.

Calculation of percentage contents:

— for any ninhydrin-positive substance detected at 570 nm, use the concentration of serine in reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b);

— if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:

— any ninhydrin-positive substance: for each impurity, maximum 0.2 per cent;

— **total:** maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Ammonium

Amino acid analysis (2.2.56) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

— **ammonium at 570 nm:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (2.4.9)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL of methyl isobutyl ketone R1, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 3 mL of anhydrous formic acid R. Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 10.51 mg of

$C_7H_7NO_3$.

STORAGE

Protected from light.

IMPURITIES

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

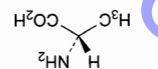
the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities. It is

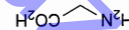
therefore not necessary to identify these impurities for

demonstration of compliance. See also 5.10. Control of

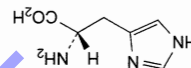
impurities in substances for pharmaceutical use): A, B, C.



A. (2S)-2-aminopropanoic acid (alanine),



B. 2-aminoacetic acid (glycine),

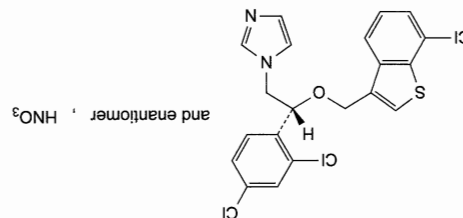


C. (2S)-2-amino-3-(imidazol-4-yl)propanoic acid (histidine).

Ph Eur

Sertaconazole Nitrate

(Ph. Eur. monograph 1148)



$C_{20}H_{16}Cl_3N_3O_4S$

500.8

99592-39-9

Action and use

Antifungal.

Ph Eur

DEFINITION

(RS)-1-[2-[(7-chloro-1-benzothio-3-yl)methoxy]-2-(2,4-dichlorophenyl)ethyl]-1H-imidazole nitrate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, soluble in methanol, sparingly soluble in ethanol (96 per cent) and in methylene chloride.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II). Dissolve 0.1 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase. Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of sertaconazole nitrate CRS in the mobile phase and dilute to 20.0 mL with the mobile phase.

TESTS

E. About 1 mg gives the reaction of nitrates (2.3.1).

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a). The chromatogram shows 2 clearly separated spots. — the chromatogram shows 2 clearly separated spots.

System suitability: reference solution (b):

Detection Expose to iodine vapour for 30 min.

Drying In a current of air for 15 min.

Development Over a path of 15 cm.

Application 5 µL.

(1:40:60 V/V/V).

Mobile phase concentrated ammonia R, toluene R, dioxan R

Plate TLC silica gel G plate R.

reference solution (a).

Reference solution (b) Dissolve 20 mg of sertaconazole nitrate CRS in reference solution (a) and dilute to 5 mL with the solvent mixture.

Reference solution (a) Dissolve 40 mg of sertaconazole nitrate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Test solution Dissolve 40 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

(10:90 V/V).

Solvent mixture concentrated ammonia R, methanol R

D. Thin-layer chromatography (2.2.27).

Comparison sertaconazole nitrate CRS.

examine as discs of potassium bromide R.

Preparation Dry the substances at 100-105 °C for 2 h and

C. Infrared absorption spectrophotometry (2.2.24).

Absorbance ratio $A_{302}/A_{293} = 1.16$ to 1.28 .

Absorption maxima At 260 nm, 293 nm and 302 nm.

Spectral range 240-320 nm.

to 100 mL with the same solvent. Dilute 10 mL of this solution

Test solution Dissolve 0.1 g in methanol R and dilute to 100 mL with the same solvent. Dilute 10 mL of this solution

(2.2.25).

B. Ultraviolet and visible absorption spectrophotometry

A. Melting point (2.2.14): 156 °C to 161 °C.

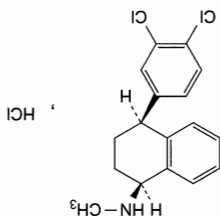
Second identification A, B, D, E.

First identification A, C.

IDENTIFICATION

Sertaline Hydrochloride

(Ph. Eur. monograph 1705)

C₁₇H₁₈Cl₂N 342.7 79559-97-0

Action and use
Selective serotonin reuptake inhibitor; antidepressant.

Preparation
Sertaline Tablets

DEFINITION

(1S,4S)-4-(3,4-Dichlorophenyl)-N-methyl-1,2,3,4-tetrahydronaphthalen-1-amine hydrochloride.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble or slightly soluble in anhydrous ethanol, slightly soluble in acetone and in 2-propanol.

It shows polymorphism (5.9).

IDENTIFICATION

Carry out either tests A, B, C or tests B, C, D.

A. Specific optical rotation (2.2.7): + 38.8 to + 43.0 (anhydrous substance), measured at 25 °C.

Solvent mixture Dilute 1 volume of a 103 g/L solution of hydrochloric acid R to 20 volumes with methanol R.

Dissolve 0.250 g in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison serratine hydrochloride CRS.

If the spectra obtained in the solid state show differences, record new spectra using 10 g/L solutions in methylene chloride R.

C. Dissolve 10 mg in 5 mL of anhydrous ethanol R and add 5 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

D. Enantiomeric purity (see Tests).

TESTS

Enantiomeric purity

Liquid chromatography (2.2.29). Prepare the test solution immediately before use.

Solvent mixture diethylamine R, hexane R, 2-propanol R (1:40:60 V/V/V).

Test solution Dissolve 60.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Dilute 1.0 mL of this solution to 50.0 mL with the mobile

Column:
— size: l = 0.25 m, Ø = 4.0 mm;
— stationary phase: nitrile silica gel for chromatography R1 (10 µm).

Mobile phase acetonitrile R1, 1.5 g/L solution of sodium dihydrogen phosphate R (37:63 V/V).

Flow rate 1.6 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 µL.

Run time 1.3 times the retention time of sertaconazole.

Retention time Nitrate ion = about 1 min;

miconazole = about 17 min; sertaconazole = about 19 min.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to miconazole and sertaconazole.

Limits:

— impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.25 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard the peak due to the nitrate ion.

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.50 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 50 mL of a mixture of equal volumes of anhydrous acetic acid R and methyl ethyl ketone R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

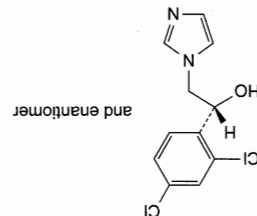
1 mL of 0.1 M perchloric acid is equivalent to 50.08 mg of C₂₀H₁₆Cl₂N₂O₄S.

STORAGE

Protected from light.

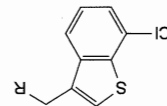
IMPURITIES

Specified impurities A, B, C



A. (1R,2S)-1-(2,4-dichlorophenyl)-2-(1H-imidazol-1-yl)ethanol,

B. R = Br: 3-(bromomethyl)-7-chloro-1-benzothiophen,
C. R = OH: (7-chloro-1-benzothiophen-3-yl)methanol.



Relative retention With reference to sertaline (retention time = about 18 min): impurity E = about 0.2; benzoic acid = about 0.3.

System suitability: reference solution (b):

— **resolution:** minimum 5.0 between the peaks due to impurity E and benzoic acid.

Limit:

— **impurity E:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent).

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution Introduce 0.250 g of the substance to be examined into a 15 mL stoppered centrifuge tube, add 2.0 mL of methanol R and 0.20 mL of a 25 per cent solution of potassium carbonate R and mix in a vortex mixer for 30 s. Add 8.0 mL of methylene chloride R, stopper the tube and mix in a vortex mixer for 60 s. Add 1 g of anhydrous sodium sulfate R, mix well and then centrifuge for about 5 min.

Reference solution (a) Dissolve the contents of a vial of sertaline for peak identification CRS (containing impurities A, B, C and F) in 0.2 mL of methylene chloride R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with methylene chloride R. Dilute 1.0 mL of this solution to 20.0 mL with methylene chloride R.

Column:

— **material:** fused silica;

— **size:** $l = 30$ m, $\varnothing = 0.53$ mm;

— **stationary phase:** polymethylphenylsiloxane R (film thickness 1.0 μ m).

Carrier gas helium for chromatography R.

Flow rate 9 mL/min.

Split ratio 1:10.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 1	200	1 - 31	260	280
31 - 39	200 → 260			
250				

Detection Flame ionisation.

Injection 1 μ L.

Identification of impurities Use the chromatogram supplied with sertaline for peak identification CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C and F.

Relative retention With reference to sertaline (retention time = about 24 min): impurity B = about 0.5; impurities C and D = about 0.7; impurity A = about 1.05; impurity F = about 1.1.

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 15, where H_p = height above the baseline of the peak due to impurity A and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to sertaline.

Limits:

— **sum of impurities C and D:** maximum 0.8 per cent;

Reference solution (a) Dissolve the contents of a vial of sertaline for system suitability CRS (containing impurity G) in 1.0 mL of the solvent mixture.

Reference solution (b) Dilute 0.5 mL of the test solution to 100.0 mL with the solvent mixture.

Column:

— **size:** $l = 0.25$ m, $\varnothing = 4.6$ mm;

— **stationary phase:** silica gel AD for chiral separation R (5 μ m).

Mobile phase Mix 30 volumes of hexane R and 70 volumes of a mixture of 1 volume of diethylamine R, 25 volumes of 2-propanol R and 975 volumes of hexane R.

Flow rate 0.4 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 20 μ L.

Run time 30 min.

Elution order Sertaline, impurity G.

System suitability:

— **resolution:** minimum 1.5 between the peaks due to sertaline and impurity G in the chromatogram obtained with reference solution (a);

— **signal-to-noise ratio:** minimum 10 for the peak due to sertaline in the chromatogram obtained with reference solution (b).

Limit:

— **impurity G:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent).

Impurity E

Liquid chromatography (2.2.29).

Solvent mixture Mobile phase A, mobile phase B (50:50 V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5.0 mg of sertaline impurity E CRS (mandelic acid) in the solvent mixture and dilute to 25.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of benzoic acid R and 20 mg of mandelic acid R (impurity E) in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute

1.0 mL of this solution to 50.0 mL with the solvent mixture.

Column:

— **size:** $l = 0.25$ m, $\varnothing = 4.6$ mm;

— **stationary phase:** end-capped octadecylsilyl silica gel for chromatography R (3 μ m).

Mobile phase:

— **mobile phase A:** dissolve 1.0 g of sodium laurylsulfate R in 800 mL of water R and add 200 mL of acetonitrile R1;

— **mobile phase B:** dissolve 1.0 g of sodium laurylsulfate R in 100 mL of water R and add 900 mL of acetonitrile R1;

add 1.0 mL of phosphoric acid R and mix;

Flow rate 1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L.

- impurities A, B, F: for each impurity, maximum 0.2 per cent;
- unspecified impurities: for each impurity, maximum 0.10 per cent;
- total: maximum 1.5 per cent;
- disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in ethanol (96 per cent) R and dilute to 20.0 mL with the same solvent. 12 mL of the solution

complies with test B. Prepare the reference solution using

lead standard solution (1 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with ethanol (96 per cent) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29).

Buffer solution T: To 28.6 mL of glacial acetic acid R slowly add, while stirring and cooling, 34.8 mL of triethylamine R, and dilute to 100 mL with water R. Dilute 10 mL of this solution

to 1000 mL with water R.

Test solution: Dissolve 55.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 5.0 mL of this solution to

100.0 mL with the mobile phase.

Reference solution: Dissolve 55.0 mg of serraline hydrochloride CRS in the mobile phase and dilute to 50.0 mL

with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\phi = 3.9$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(4 μ m);

— temperature: 30 °C.

Mobile phase: methanol R, buffer solution, acetonitrile R

(15:40:45 V/V/V).

Flow rate 1.8 mL/min.

Detection: Spectrophotometer at 254 nm.

Injection 20 μ L.

Run time: Twice the retention time of serraline.

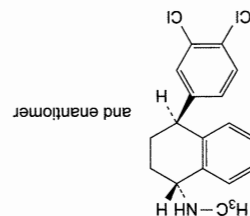
Calculate the percentage content of $C_{17}H_{18}Cl_3N$ taking into account the assigned content of serraline hydrochloride CRS.

STORAGE

Protected from light.

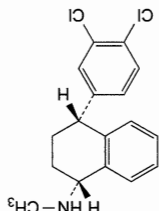
IMPURITIES

Specified impurities A, B, C, D, E, F, G

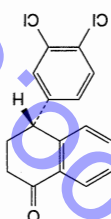


A. (1*RS*,4*SR*)-4-(3,4-dichlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine,

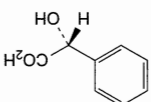
G. (1*R*,4*R*)-4-(3,4-dichlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine (serraline enantiomer).



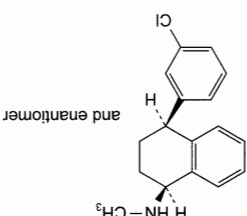
F. (4*R*)-4-(3,4-dichlorophenyl)-3,4-dihydronaphthalen-1(2*H*)-one,



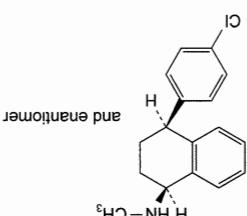
E. (2*R*)-hydroxyphenylacetic acid ((*R*)-mandelic acid),



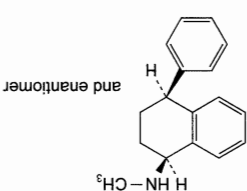
D. (1*RS*,4*RS*)-4-(3-chlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine,



C. (1*RS*,4*RS*)-4-(4-chlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine,



B. (1*RS*,4*RS*)-*N*-methyl-4-phenyl-1,2,3,4-tetrahydronaphthalen-1-amine,



Refined Sesame Oil

Sesame Oil

(Ph. Eur. monograph 0433)

Ph Eur

DEFINITION

Fatty oil obtained from the ripe seeds of *Sesamum indicum* L. by expression or extraction. It is then refined. Improved colour and odour may be obtained by further refining. It may contain a suitable antioxidant.

CHARACTERS

Appearance
Clear, light yellow liquid, almost colourless.

Solubility
Practically insoluble in ethanol (96 per cent), miscible with light petroleum.

Relative density
About 0.919.

Refractive index About 1.473.

It solidifies to a butter-like mass at about -4°C .

IDENTIFICATION

First identification A

Second identification B

- A. Composition of triglycerides (see Tests).
(2.3.2).
B. Identification of fatty oils by thin-layer chromatography

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3-1.

TESTS

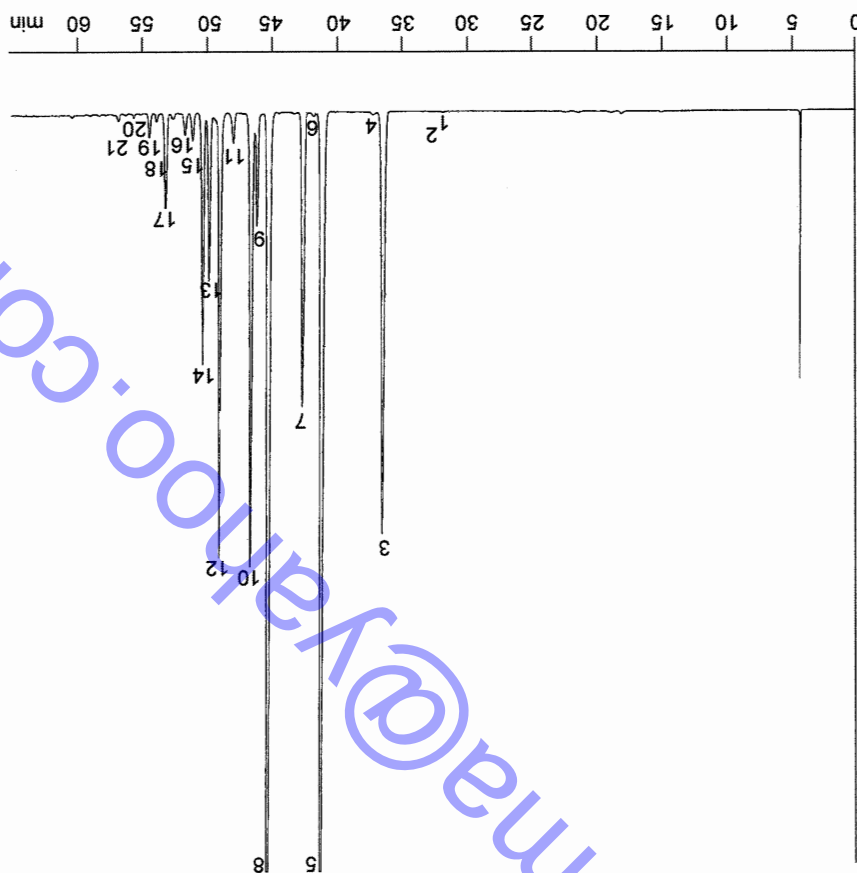
Acid value (2.5.1)
Maximum 0.5, determined on 10.0 g; maximum 0.3 if intended for use in the manufacture of parenteral preparations.

Unsaponifiable matter (2.5.7)
Maximum 2.0 per cent, determined on 5.0 g.

Alkaline impurities (2.4.19)
It complies with the test for alkaline impurities in fatty oils.

Cottonseed oil

Mix 5 mL in a test-tube with 5 mL of a mixture of equal volumes of pentanol R and a 10 g/L solution of sulfur R in carbon disulfide R. Warm the mixture carefully until the carbon disulfide is expelled, and immerse the tube to 1/3 of its depth in boiling saturated sodium chloride solution R. No reddish colour develops within 15 min.



1. LLLn	4. OLLn	7. PLL	10. POL	13. SOL	16. PPO	19. SSL
2. OLLn	5. OLL	8. OOL	11. PPL	14. POO	17. SOO	20. PPS
3. LLL	6. OOLLn	9. SIL	12. OOO	15. PSL	18. PSO	21. SSO

Figure 0433.-1. – Chromatogram for the composition of triglycerides in refined sesame oil

Composition of triglycerides

Test solution Dilute 50.0 mg of the substance to be examined to 10.0 mL with a mixture of equal volumes of acetone R and methyl chloride R.

Reference solutions Dissolve 80.0 mg of triolein R in a mixture of equal volumes of acetone R and methyl chloride R and dilute to 50.0 mL with the same mixture of solvents. Prepare 5 reference solutions by dilution of this solution so as to cover concentrations ranging from the disregard limit (0.5 per cent) to the upper limit for OIL (30.0 per cent). Plot the logarithm of the area of the peak due to triolein against the logarithm of the concentration of triolein in the reference solution.

Column 2 columns coupled in series:
— size of each column: $l = 0.25$ m, $\varnothing = 4$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (4 μ m).
Mobile phase:
— mobile phase A: acetone R, methylene chloride R, acetonitrile R (5:15:80 V/V/V);
— mobile phase B: acetone R, acetonitrile R, methylene chloride R (20:20:60 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	100 \rightarrow 75	0 \rightarrow 25
15 - 25	75	25
25 - 70	75 \rightarrow 0	25 \rightarrow 100
70 - 75	0 \rightarrow 100	100 \rightarrow 0
75 - 80	100	0

Detection Evaporative light-scattering detector; the following settings have been found to be suitable; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criterion:

— carrier gas: nitrogen R;
— flow rate: 0.7 L/min;
— evaporator temperature: 85 °C;
— nebuliser temperature: 45 °C.

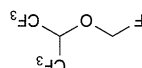
Identification of peaks Use the chromatograms obtained with the reference solutions to identify the peak due to triolein; identify the other peaks using the chromatogram shown in Figure 0433-1. The fatty acids are designated as linoleic (Ln), linoleic (L), oleic (O), palmitic (P) and stearic (S).

System suitability Test solution:
— resolution: minimum 1.5 between the peaks due to OOO (triolein) and SOL.
Using the calibration curve obtained with the reference solutions, determine the percentage content of each peak with an area greater than that of the peak corresponding to the disregard limit (0.5 per cent). Assuming that the sum of these percentage contents is 100 per cent, normalise the percentage content of each of the 8 triglycerides specified below.

Composition of triglycerides:
— LLL: 7.0 per cent to 19.0 per cent;
— OLL: 13.0 per cent to 30.0 per cent;
— PLL: 5.0 per cent to 9.0 per cent;

Sevoflurane

(Ph. Eur. monograph 2269)



200.1

28523-86-6

Action and use
General anaesthetic.

Ph Eur

DEFINITION
1,1,1,3,3,3-Hexafluoro-2-(fluoromethoxy)propane.

CHARACTERS

Appearance
Clear, colourless, volatile liquid.

Solubility
Slightly soluble in water, miscible with ethanol (96 per cent).

Relative density
About 1.52.

bp
About 59 °C.

It is non-flammable.

It decomposes in the presence of Lewis acids; this decomposition is inhibited by water in sufficient quantity.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Examine the substance in the gaseous state or in the liquid state.

Comparison sevoflurane CRS.

TESTS

Acidity or alkalinity

Introduce 20.0 mL of the substance to be examined and 20 mL of carbon dioxide-free water R into a separating funnel, shake for 3 min and allow to stand. Collect the aqueous

The label states:
— whether the oil is obtained by expression or extraction;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations;
— where applicable, the name of the inert gas used.

Ph Eur

LABELLING

at once is protected by an atmosphere of an inert gas. used as soon as possible. Any part of the contents not used When the container has been opened, its contents are to be preparations store under an inert gas in an airtight container. if intended for use in the manufacture of parenteral

STORAGE

Maximum 0.1 per cent, determined on 1.00 g.

Water (2.5.32)

— OOL: 12.0 per cent to 23.0 per cent;
— POL: 6.0 per cent to 14.0 per cent;
— OOO: 5.0 per cent to 14.0 per cent;
— SOL: 2.0 per cent to 8.0 per cent;
— POO: 2.0 per cent to 10.0 per cent.

upper layer and add 0.2 mL of bromocresol purple solution R. Not more than 0.10 mL of 0.01 M sodium hydroxide or not more than 0.60 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator.

Refractive index (2.2.6)

1.2745 to 1.2760.

Related substances

Gas chromatography (2.2.28).

Internal standard methylal R.

Test solution Introduce 20.0 mL of the substance to be examined into a vial and seal with a cap and septum. Using a microsyringe, add 5 µL of the internal standard and mix thoroughly.

Reference solution (a) Introduce 2.0 mL of ethylene chloride R into a screw-cap vial and immediately seal with a cap and septum. Using a microsyringe, add about 20 µL of the substance to be examined. Record the quantity added, in milligrams, of the substance to be examined (M_2). Then, using a microsyringe, add about 20 µL of the internal standard. Record the quantity added, in milligrams, of the

Reference solution (b) sevoflurane CRS (containing impurities A and B).

Reference solution (c) Introduce 20.0 mL of ethylene chloride R into a vial and seal with a cap and septum. Using a microsyringe, add 20 µL of the substance to be examined and mix thoroughly. Dilute 0.5 mL of this solution to 100.0 mL with ethylene chloride R.

Column:

— **material:** fused silica;

— **size:** $l = 30$ m, $\varnothing = 0.32$ mm;

— **stationary phase:**

poly[(cyanopropyl)(phenyl)dimethylsiloxane R (film thickness 3 µm).

Carrier gas helium for chromatography R.

Flow rate 1.0 mL/min.

Split ratio 1:20.

Temperature:

Time (min)	Temperature (°C)
0 - 10	40
10 - 26	40 → 200
26 - 40	200
Injection port	200
Detector	225

Detection Flame ionisation.

Injection 2 µL.

Rinse the syringe with a solution containing ethylene chloride R before the injection of the reference solutions. Rinse the syringe with the substance to be examined before the injection of the test solution.

Identification of impurities Use the chromatogram supplied with sevoflurane CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B.

Relative retention With reference to sevoflurane (retention time = about 6.6 min): impurity A = about 0.78; impurity B = about 0.83; internal standard = about 1.35.

Fluorides

Maximum 2 µg/mL.

Potentiometry (2.2.36, Method D). Use plastic utensils throughout this test.

Buffer solution Dissolve 0.5 g of sodium citrate R and 55 g of sodium chloride R in 350 mL of water R. Carefully add 75 g of sodium hydroxide R and shake to dissolve. Cool to room

temperature and carefully add 225 mL of glacial acetic acid R while stirring. Cool and add 300 mL of isopropyl alcohol R. Dilute with water R to 1000.0 mL. The apparent pH of this solution is between 5.0 and 5.5.

Test solution Introduce 50.0 mL of the substance to be examined and 50.0 mL of water R into a separating funnel, shake vigorously for 3 min and allow the layers to separate completely. Dilute 25.0 mL of the aqueous upper layer to 50.0 mL with the buffer solution.

Fluoride standard solution (1000 ppm F) Dissolve 221.0 mg of sodium fluoride R, previously dried at 150 °C for 4 h, in water R. Add 1.0 mL of 0.01 M sodium hydroxide and dilute to 100.0 mL with water R.

Reference stock solutions Dilute the fluoride standard solution (1000 ppm F) diluted with water R to obtain solutions having

$$\frac{M_1 \times R}{M_2}$$

M_1 = mass of the internal standard in reference solution (a), in milligrams;
 M_2 = mass of the substance to be examined in reference solution (a), in milligrams;
 R = ratio of the area of the peak due to sevoflurane to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (a).

Calculate the quantity of each impurity in the substance to be examined, in parts per million, using the following expression:

$$\frac{0.859 \times R_1 \times 250}{1.52 \times F_1}$$

0.859 = relative density of the internal standard;
 1.52 = relative density of sevoflurane;
 R_1 = ratio of the area of the peak due to the impurity to the area of the peak due to the internal standard from the chromatogram obtained with the test solution;
 F_1 = relative response factor for reference solution (a).

Limits:

— **impurity A:** maximum 25 ppm;
 — **impurity B:** maximum 100 ppm;
 — **unspecified impurities:** for each impurity, maximum 100 ppm;

— **total:** maximum 300 ppm;
 — **disregard limit:** the area of the peak due to sevoflurane in the chromatogram obtained with reference solution (c)

(5 ppm).

(Ph. Eur. monograph 1149)

Excipient.

DEFINITION

Purified material obtained from the resinous secretion of the female insect *Kemma lacca* (Kerr) Lindinger (*Laccifer lacca* Kerr). There are 4 types of shellac depending on the nature of the treatment of crude secretion (seedlac): wax-containing shellac, bleached shellac, dewaxed shellac and bleached,

Purified material obtained from the resinous secretion of the female insect *Kermia lacca* (Kerr) Lindinger (*Laccifer lacca* Kerr). There are 4 types of shellac depending on the nature of the treatment of crude secretion (seedac): wax-containing shellac, bleached shellac, dewaxed shellac and bleached,

Wax-containing shellac is obtained from seedlac; it is purified by filtration of the molten substance and/or by hot extraction using a suitable solvent.

de-waxed shellac is obtained from wax containing shellac or seedlac by treatment with a suitable solvent and removal of the insoluble wax by filtering.

Bleached, de-waxed shellac is obtained from wax-containing shellac or seedlac by treatment with sodium hypochlorite after dissolution in a suitable alkaline solution; the insoluble wax is removed by filtration. It is precipitated by dilute acid and dried.

CHARACTERS

Appearance

Solubility
Brownish-orange or yellow, shining, translucent, hard or brittle, more or less thin flakes (wax-containing shellac and dewaxed shellac), or a creamy white or brownish-yellow powder (bleached shellac and bleached, dewaxed shellac).

Solubility

Practically insoluble in water, gives a more or less opalescent solution (wax containing shellac and bleached shellac) or a clear solution (dewaxed shellac and bleached, dewaxed shellac) in anhydrous ethanol. When warmed it is sparingly soluble or soluble in alkaline solutions.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).
Test solution Heat 0.25 g of the powdered substance (500) (2.9.12) on a water-bath with 2 mL of dilute sodium hydroxide solution R for 5 min. Cool, add 5 mL of ethyl acetate R and slowly, with stirring, 2 mL of dilute acetic acid R. Shake and

filter the upper layer through anhydrous sodium sulfate R.
Reference solution Dissolve 6.0 mg of alcurtic acid R in 1.0 mL
of methanol R, heating slightly if necessary.
Plate TLC silica gel F₂₅₄ plate R.
Mobile phase acetic acid R, methanol R, methylène chloride R.

Application 10 μ L, as bands.
Development Twice over a path of 15 cm.
Drying In air.

Detection Spray with *anisaldehyde solution* R_f heat at 100-105 °C for 5-10 min and examine in daylight.

Results The chromatogram obtained with the test solution shows several coloured zones, one of which is similar in position and colour to the zone in the chromatogram obtained with the reference solution. Above this zone the

Reference solutions Dilute 25.0 mL of each reference stock solution to 50.0 mL with the buffer solution.

Indicator electrode Fluoride-selective.

Reference electrode Glass-sleeved calomel.

Apparatus Voltmeter capable of a minimum reproducibility of ± 0.2 mV.

polytetrafluoroethylene-coated magnetic stirring bar, and immerse the electrodes. Allow the system to sit on a magnetic stirrer with an air motor until equilibrium is attained (about 2–3 min), and record the potential. Rinse the electrodes with the buffer solution and dry, taking care to avoid damaging

Calculate the concentration of fluorides using the calibration curve.

Water (2.5.12)
2 h. The residue weighs a maximum of 1.0 mg.
Transfer 10.0 mL to a tared evaporating dish, evaporate to dryness on a water-bath and dry the residue at 105 °C for

STORAGE
In an airtight, stainless-steel container, protected from light.

Specified impurities A, B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use*

(2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: C.

A. 1,1,3,3,3-pentafluoro-2-(fluoromethoxy)prop-1-ene,

B. 1,1,1,3,3,3-hexafluoro-2-methoxypropane,

C, 1,1,1,3,3,3-hexafluoropropan-2-ol.

Ph Eur

chromatogram obtained with the test solution shows a pink zone and below it several violet zones. Below the zone due to alevuric acid, there is a light blue zone (shelloilic acid) accompanied by zones of the same colour but of lower intensity. Other faint grey and violet zones may be visible. B. Examine the chromatograms obtained in the test for colophony.

Results For wax-containing shellac, in the chromatogram obtained with the test solution, a more or less strong bluish-grey zone is visible, just above the zone due to thymolphthalein in the chromatogram obtained with the reference solution; for dewaxed shellac, no such zone is visible just above the zone due to thymolphthalein in the chromatogram obtained with the reference solution.

TESTS

Acid value (2.5.1) 65 to 95 (dried substance).

Examine 1.00 g of the coarsely ground substance. Determine the end-point potentiometrically (2.2.20).

Colophony

Thin-layer chromatography (2.2.27) as described under identification test A with the following modifications.

Test solution Dissolve 50 mg of the powdered substance (500) (2.9.12), with heating, in a mixture of 0.5 mL of methylene chloride R and 0.5 mL of methanol R.

Reference solution Dissolve 2.0 mg of thymolphthalein R in 1.0 mL of methanol R.

Detection Examine in ultraviolet light at 254 nm; mark the quenching zones in the chromatogram obtained with the test solution that have similar R_f values to that of the quenching zone due to thymolphthalein in the chromatogram obtained with the reference solution; spray with *anisaldehyde solution* R, heat at 100-105 °C for 5-10 min and examine in daylight.

Results The chromatogram obtained with the reference solution shows a principal zone with a reddish-violet colour (thymolphthalein). None of the quenching zones in the chromatogram obtained with the test solution that have an R_f value similar to the zone due to thymolphthalein in the reference solution show a more or less strong violet or brownish colour (colophony). Disregard any faint violet zone at this level that does not show quenching before spraying and heating.

Arsenic (2.4.2, Method A)

Maximum 3 ppm.

Introduce 0.33 g of the substance to be examined and 5 mL of *sulfuric acid* R into a combustion flask. Carefully add a few millilitres of *strong hydrogen peroxide solution* R and heat to boiling until a clear, colourless solution is obtained. Continue heating to eliminate the water and as much sulfuric acid as possible and dilute to 25 mL with *water* R.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2 mL of *lead standard solution* (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 2.0 per cent for unbleached shellac and maximum 6.0 per cent for bleached shellac, determined on 1.000 g of the powdered substance (500) (2.9.12) by drying in an oven at 40-45 °C for 24 h.

STORAGE

Protected from light. Store bleached shellac and bleached, dewaxed shellac at a temperature not exceeding 15 °C.

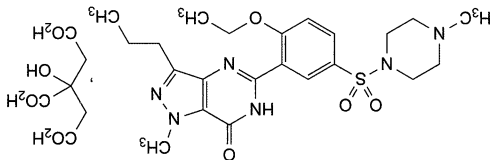
LABELLING

The label indicates the type of shellac.

Ph Eur

Sildenafil Citrate

(Ph. Eur. monograph 2270)



C₂₈H₃₈N₆O₁₁S 667 171599-83-0

Action and use

Selective inhibitor of cyclic GMP-specific phosphodiesterase (Type V) with vasodilator action; treatment of erectile dysfunction.

Ph Eur

DEFINITION

5-[2-Ethoxy-5-[(4-methylpiperazin-1-yl)sulfonyl]phenyl]-1-methyl-3-propyl-1,6-dihydro-7H-pyrazolo[4,3-d]pyrimidin-7-one dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Slightly soluble in water and in methanol, practically insoluble in hexane.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison *sildenafil citrate* CRS.

TESTS

Impurity E

Thin-layer chromatography (2.2.27).

Solvent mixture concentrated ammonia R, water R, methanol R (5:25:75 V/V/V).

Test solution Dissolve 35.0 mg of the substance to be examined in 2.0 mL of the solvent mixture, with the aid of ultrasound if necessary.

Reference solution (a) Dissolve 7.0 mg of *imidazole* CRS (impurity E) in the solvent mixture and dilute to 20.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Reference solution (c) Mix 1 mL of the test solution and 1 mL of reference solution (a).

Plate TLC silica gel F₂₅₄ plate R (2-10 µm).

Mobile phase concentrated ammonia R, ethanol (96 per cent) R, ethyl acetate R, methylene chloride R (1:20:30:50 V/V/V/V).

Application 10 µL of the test solution and reference solutions (b) and (c) as bands of 6 mm.

Development Over 2/3 of the plate.

Drying At 100 °C for about 15 min.

Detection Expose to iodine vapour until the plate is light brown and examine under ultraviolet light at 254 nm.

Retention factors Citrate = about 0.5;

impurity E = about 0.25; sildenafil = about 0.4.

System suitability: reference solution (c):

the chromatogram shows 2 clearly separated zones.

Limit:

— impurity E: any zone due to impurity E is not more intense than the principal zone in the chromatogram

obtained with reference solution (b) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 35.0 mg of the substance to be examined in the mobile phase, with the aid of ultrasound if

necessary, and dilute to 50.0 mL with the mobile phase.

Test solution (b) Dilute 2.0 mL of test solution (a) to

50.0 mL with the mobile phase.

Reference solution (a) Dissolve 35.0 mg of sildenafil citrate CRS

in the mobile phase, with the aid of ultrasound if necessary,

and dilute to 50.0 mL with the mobile phase. Dilute 2.0 mL

of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of test solution (b) to

100.0 mL with the mobile phase.

Reference solution (c) In order to prepare impurity B *in situ*,

dissolve 70 mg of the substance to be examined in 1 mL of a

mixture of 1 volume of anhydrous formic acid R and

2 volumes of stabilised strong hydrogen peroxide solution R.

Allow to stand for at least 10 min and dilute to 250 mL with

the mobile phase.

Reference solution (d) Dissolve 3 mg of sildenafil

impurity A CRS in the mobile phase and dilute to 20.0 mL

with the mobile phase. Dilute 1 mL of the solution to

20.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 μ m);

— temperature: 30 °C.

Mobile phase Mix 17 volumes of acetonitrile R, 25 volumes of

methanol R and 58 volumes of a 0.7 per cent V/V solution of

methanesulphonic acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 290 nm.

Injection 20 μ L of test solution (a) and reference

solutions (b), (c) and (d).

Run time 3 times the retention time of sildenafil.

Identification of impurities Use the chromatogram obtained

with reference solution (d) to identify the peak due to

impurity A.

Retention With reference to sildenafil (retention

time = about 7 min): impurity B = about 1.2;

impurity A = about 1.7.

System suitability: reference solution (c):

— resolution: minimum 2.5 between the peaks due to

sildenafil and impurity B.

Calculation of percentage contents:

— for each impurity, use the concentration of sildenafil in

reference solution (b).

— impurity A: maximum 0.3 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— sum of unspecified impurities: maximum 0.3 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 2.5 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 0.5 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution (b) and reference solution (a).

Calculate the percentage content of $C_{28}H_{35}N_6O_{11}$ taking

into account the assigned content of sildenafil citrate CRS.

STORAGE

In airtight container.

IMPURITIES

Specified impurities A, E

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

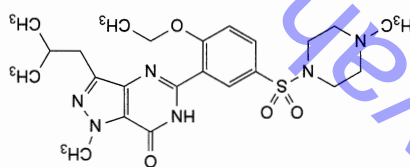
acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

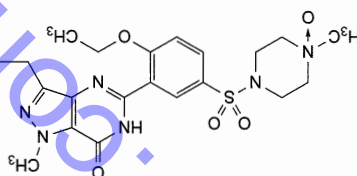
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

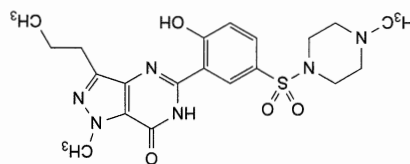
Control of impurities in substances for pharmaceutical use: B, C,



A. 5-[2-ethoxy-5-[(4-methylpiperazin-1-yl)sulfonyl]phenyl]-1-methyl-3-(2-methylpropyl)-1,6-dihydro-7H-pyrazolo[4,3-d]pyrimidin-7-one,



B. 5-[2-ethoxy-5-[(4-methyl-4-oxidopiperazin-1-yl)sulfonyl]phenyl]-1-methyl-3-(2-methylpropyl)-1,6-dihydro-7H-pyrazolo[4,3-d]pyrimidin-7-one,



C. 5-[2-ethoxy-5-[(4-methylpiperazin-1-yl)sulfonyl]phenyl]-1-methyl-3-(2-methylpropyl)-1,6-dihydro-7H-pyrazolo[4,3-d]pyrimidin-7-one,

of the solution add 50 mg of ascorbic acid R and 1 mL of concentrated ammonia R. Neutralise with dilute ammonia R2. Dilute to 25 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on ignition
Maximum 5.0 per cent, determined on 0.200 g by ignition in a platinum crucible at $900 \pm 50^\circ\text{C}$ for 2 h. Allow to cool in a desiccator before weighing.

ASSAY

To the residue obtained in the test for loss on ignition add 0.2 mL of sulfuric acid R and sufficient ethanol (96 per cent) R to moisten the residue completely. Add 6 mL of hydrofluoric acid R and evaporate to dryness on a hot-plate at $95\text{--}105^\circ\text{C}$, taking care to avoid loss from spattering. Wash down the sides of the dish with 6 mL of hydrofluoric acid R and evaporate to dryness. Ignite at $900 \pm 50^\circ\text{C}$, allow to cool in a desiccator and weigh.

The difference between the mass of the final residue and the mass of the residue obtained in the test for loss on ignition gives the amount of SiO_2 in the quantity of the substance to be examined used.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are also to be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for colloidal anhydrous silica used as glidant in tablets and capsules.

Specific surface area (2.9.26, Method J)
Determine the specific surface area in the P/P_0 range of 0.05 to 0.30.

Sample outgassing 20 min at 160°C .

Ph Eur



Colloidal Hydrated Silica

(Ph. Eur. monograph 0738)

63231-67-4

Action and use
Excipient.

Ph Eur

DEFINITION

Colloidal hydrated silica produced by precipitation or gelation process.

Content

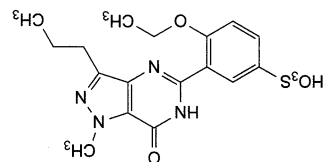
98.0 per cent to 100.5 per cent of SiO_2 (M_r 60.1) (ignited substance).

CHARACTERS

Appearance

White or almost white, light, fine, amorphous powder.

D. 4-ethoxy-3-(1-methyl-7-oxo-3-propyl-6,7-dihydro-1H-pyrazolo[4,3-d]pyrimidin-5-yl)benzenesulfonic acid,



E. 1H-imidazole.

Ph Eur

Colloidal Anhydrous Silica

(Ph. Eur. monograph 0434)

SiO_2 60.1

7631-86-9



Action and use

Excipient.

Ph Eur

DEFINITION

99.0 per cent to 100.5 per cent of SiO_2 (ignited substance).

CHARACTERS

Appearance

White or almost white, light, fine, amorphous powder, with a particle size of about 15 nm.

Solubility

Practically insoluble in water and in mineral acids except hydrofluoric acid. It dissolves in hot solutions of alkali hydroxides.

IDENTIFICATION

About 20 mg gives the reaction of silicates (2.3.1).

TESTS

pH (2.2.3)

3.5 to 5.5.

Shake 1.0 g with 30 mL of carbon dioxide-free water R.

Chlorides

Maximum 250 ppm. To 1.0 g add a mixture of 20 mL of dilute nitric acid R and 30 mL of water R and heat on a water-bath for 15 min, shaking frequently. Dilute to 50 mL with water R if necessary, filter and cool. Dilute 10 mL of the filtrate to 15 mL with water R.

Heavy metals

Maximum 25 ppm.

Suspend 2.5 g in sufficient water R to produce a semi-fluid slurry. Dry at 140°C . When the dried substance is white, break up the mass with a glass rod. Add 25 mL of 1 M hydrochloric acid and boil gently for 5 min, stirring frequently with the glass rod. Centrifuge for 20 min and filter the supernatant through a membrane filter. To the residue in the centrifuge tube add 3 mL of dilute hydrochloric acid R and 9 mL of water R and boil. Centrifuge for 20 min and filter the supernatant through the same membrane filter. Wash the residue with small quantities of water R, combine the filtrates and washings and dilute to 50 mL with water R. To 20 mL

mass of the final residue and that of the mass obtained in the test for loss on ignition corresponds to the mass of SiO_2 in the test sample.

Ph Eur



Silica Hydrophobic Colloidal Anhydrous

(Ph. Eur. monograph 2208)

Ph Eur

DEFINITION

Colloidal silicon dioxide partly alkylated for hydrophobation.

Content

99.0 per cent to 101.0 per cent SiO_2 (ignited substance).

CHARACTERS

Appearance

White or almost white, light, fine, amorphous powder, not wettable by water.

Solubility

Practically insoluble in water and mineral acids except hydrofluoric acid. It dissolves slowly in hot solutions of alkali hydroxides.

IDENTIFICATION

A. About 25 mg ignited in a platinum crucible at $900 \pm 50^\circ\text{C}$ for 2 h gives the reaction of silicates (2.3.1).
B. Water-dispersible fraction (see Tests).

TESTS

Chlorides (2.4.4)

Maximum 250 ppm.

To 1.0 g add 30 mL of methanol R and 20 mL of dilute nitric acid R. Heat on a water-bath for 15 min stirring frequently. Cool, dilute to 50 mL with water R and filter. Dilute 10 mL of the filtrate to 15 mL with water R.

Heavy metals (2.4.8)

Maximum 25 ppm.

Suspend 2.5 g in 30 mL of methanol R, stir and add 30 mL of dilute ammonia R1. With frequent stirring evaporate on a water-bath and dry the residue in an oven at 140°C . When the dried substance is white, break up the mass with a glass rod. Reduce the residue to a powder and add 15 mL of methanol R and 25 mL of 1 M hydrochloric acid. Boil gently for 5 min, stirring frequently with the glass rod. Centrifuge for 20 min and filter the supernatant through a membrane filter. To the residue in the centrifuge tube add 3 mL of dilute hydrochloric acid R and 9 mL of water R and boil. Centrifuge for 20 min and filter the supernatant through the same membrane filter. Wash the residue with small quantities of water R, combine the filtrates and washings and dilute to 50 mL with water R. To 20 mL of this solution add 50 mg of ascorbic acid R and 1 mL of concentrated ammonia R. Neutralise with dilute ammonia R2. Dilute to 25 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water-dispersible fraction

Maximum 3.0 per cent.

Place 0.400 g in a 500 mL separating funnel, add 100 mL of water R and shake for 1 min. Allow to stand for 1 h. Allow filtration into a suitable dish dried at 140°C and cooled in a

About 20 mg gives the reaction of silicates (2.3.1).

IDENTIFICATION

alkali hydroxides.

exception of hydrofluoric acid. It dissolves in hot solutions of

Solubility

To 2.5 g add 50 mL of hydrochloric acid R and mix. Heat on a water-bath for 30 min, stirring from time to time. Maintain the original volume by adding dilute hydrochloric acid R.

Evaporate to dryness. Add to the residue a mixture of 8 mL of dilute hydrochloric acid R and 24 mL of water R. Heat to boiling and filter under reduced pressure through a sintered-glass filter (16) (2.1.2). Wash the residue on the filter with a hot mixture of 3 mL of dilute hydrochloric acid R and 9 mL of water R. Wash with small quantities of water R, combine the filtrate and washings and dilute to 50 mL with water R.

pH (2.2.3)

4.0 to 7.0.

Suspend 1.0 g in 30 mL of a 75 g/L solution of potassium chloride R.

Water-absorption capacity

In a mortar, triturate 5 g with 5 mL of water R, added drop by drop. The mixture remains powdery.

Substances soluble in hydrochloric acid

Maximum 2.0 per cent.

In a platinum dish, evaporate to dryness 10.0 mL of solution S and dry to constant mass at $100\text{--}105^\circ\text{C}$. The mass of the residue is not more than 10 mg.
Chlorides (2.4.4)
Maximum 0.1 per cent.
Heat 0.5 g with 50 mL of water R on a water-bath for 15 min. Dilute to 100 mL with water R and centrifuge at 1500 g for 5 min. Dilute 10 mL of the supernatant to 15 mL with water R.
Sulfates (2.4.13)
Maximum 1 per cent.
Dilute 2 mL of solution S to 100 mL with distilled water R.

Iron (2.4.9)

Maximum 300 ppm.

To 2 mL of solution S add 28 mL of water R.

Heavy metals (2.4.8)

To 20 mL of solution S add 50 mg of hydroxylamine hydrochloride R and 1 mL of concentrated ammonia R. Adjust to pH 3.5 by adding dilute ammonia R2, monitoring the pH potentiometrically. Dilute to 25 mL with water R. 12 mL of the solution complies with test A (25 ppm). Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on ignition

Maximum 20.0 per cent, determined on 0.200 g in a platinum crucible by heating at $100\text{--}105^\circ\text{C}$ for 1 h and then at $900 \pm 50^\circ\text{C}$ for 2 h.

ASSAY

To the residue obtained in the test for loss on ignition add 0.2 mL of sulfuric acid R and a quantity of ethanol

(96 per cent) R sufficient to moisten the residue completely. Add 6 mL of hydrofluoric acid R and evaporate to dryness at $95\text{--}105^\circ\text{C}$, taking care to avoid loss from spattering. Wash the inside of the dish with 6 mL of hydrofluoric acid R and evaporate to dryness again. Ignite at $900 \pm 50^\circ\text{C}$, allow to cool in a desiccator and weigh. The difference between the

IDENTIFICATION

About 20 mg gives the reaction of silicates (2.3.1).

TESTS

Solution S

To 2.5 g add 50 mL of *hydrochloric acid R* and mix. Heat on a water-bath for 30 min, stirring from time to time. Evaporate to dryness. Add to the residue a mixture of 8 mL of *dilute hydrochloric acid R* and 24 mL of *water R*. Heat to boiling and filter under reduced pressure through a sintered-glass filter (16) (2.1.2). Wash the residue on the filter with a hot mixture of 3 mL of *dilute hydrochloric acid R* and 9 mL of *water R*. Wash with small quantities of *water R*, combine the washings and the filtrate, and dilute to 50 mL with *water R*.

pH (2.2.3)

3.2 to 8.9.

Suspend 5 g in a mixture of 5 mL of a 7.46 g/L solution of *potassium chloride R* and 90 mL of *carbon dioxide-free water R*.

Chlorides

Liquid chromatography (2.2.29) as described in the test for sulfates.

Retention time Chlorides = about 4 min.

Limit:

— chlorides: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.3 per cent).

Sulfates

Liquid chromatography (2.2.29).

Test solution To 0.625 g of the substance to be examined add 30 mL of *water R* and boil for 2 h. Allow to cool and quantitatively transfer to a 50 mL graduated flask. Dilute to 50.0 mL with *water R*. Dilute 5.0 mL of the supernatant to 50.0 mL with *water R* and filter through a membrane filter (nominal pore size 0.45 µm).

Reference solution Dissolve 0.50 g of *anhydrous sodium sulfate R* and 0.062 g of *sodium chloride R* in *water R* and dilute to 1000.0 mL with *water R*. Dilute 5.0 mL of the solution to 50.0 mL with *water R*.

Column:

— material: non-metallic;

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: suitable anion-exchange resin

(30–50 µm).

Mobile phase Dissolve 0.508 g of *sodium carbonate R* and 0.05 g of *sodium hydrogen carbonate R* in *water R* and dilute to 1000 mL with the same solvent.

Flow rate 1.2 mL/min.

Detection Conductivity detector.

Injection 25 µL.

Retention time Sulfates = about 8 min.

Limit:

— sulfates: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (4.0 per cent, expressed as sodium sulfate).

Iron (2.4.9)

Maximum 400 ppm.

Dilute 2 mL of solution S to 40 mL with *water R*.

Heavy metals (2.4.8)

Maximum 25 ppm.

To 20 mL of solution S, add 50 mg of *hydroxylamine hydrochloride R* and 1 mL of *concentrated ammonia R*. Adjust to pH 3.5 with *dilute ammonia R2*, monitoring the pH potentiometrically. Dilute to 25 mL with *water R*. 12 mL of

Dental-type Silica

(Ph. Eur. monograph 1562)

Action and use

Excipient.

Ph Eur

DEFINITION

Amorphous silica (precipitated, gel or obtained by flame hydrolysis).

Content

94.0 per cent to 100.5 per cent of SiO_2 (ignited substance).

CHARACTERS

Appearance

White or almost white, light, fine, amorphous powder.

Solubility

Practically insoluble in water and in mineral acids. It dissolves in hydrofluoric acid and hot solutions of alkali hydroxides.



Ph Eur

Sample outgassing 20 min at 160 °C.

0.05 to 0.30.

Determine the specific surface area in the P/P_0 range of

Specific surface area (2.9.26, Method I)

colloidal silica used as glidant in tablets and capsules.

The following characteristic may be relevant for hydrophobic

reported, the control method must be indicated.

also be used. Wherever results for a particular characteristic are recognised as being suitable for the purpose, but other methods can product during use. Where control methods are cited, they are of the manufacturing process and the performance of the medicinal to the quality of a medicinal product by improving the consistency compliance. Control of these characteristics can however contribute and it is not necessary to verify the characteristics to demonstrate 5.15). This section is a non-mandatory part of the monograph functions of the substance when used as an excipient (see chapter recognised as being relevant control parameters for one or more This section provides information on characteristics that are

FUNCTIONALITY-RELATED CHARACTERISTICS

be examined.

To the residue obtained in the test for loss on ignition add sufficient *ethanol* (96 per cent) *R* to moisten the residue completely and 0.2 mL of *sulfuric acid R*. Add 6 mL of *hydrofluoric acid R* and evaporate to dryness on a hot-plate at about 100 °C, taking care to avoid loss from spattering. Wash down the sides of the platinum crucible with 6 mL of *hydrofluoric acid R* and evaporate to dryness. Ignite at 900 ± 50 °C, allow to cool in a desiccator and weigh. The difference between the mass of the residue obtained in the test for loss on ignition and the mass of the final residue gives the amount of SiO_2 in the quantity of the substance to be examined.

ASSAY

oven. Allow to cool in a desiccator before weighing. place the crucible in a cold oven and then to heat up the a platinum crucible at 900 ± 50 °C for 2 h. It is advisable to Maximum 6.0 per cent, determined on 0.200 g by ignition in Loss on ignition

The residue weighs a maximum of 12 mg.

temperature to avoid splashing. Cool in a desiccator.

desiccator. Evaporate to dryness at 140 °C, starting at a low

the solution complies with test A. Prepare the reference solution using *lead standard solution* (1 ppm Pb) R.

Loss on ignition

Maximum 25.0 per cent, determined on 0.200 g by heating in a platinum crucible at 100-105 °C for 1 h and then at 1000 ± 50 °C for 2 h.

ASSAY

To the residue obtained in the test for loss on ignition add 0.2 mL of *sulfuric acid* R and a quantity of *ethanol* (96 per cent) R sufficient to moisten the residue completely.

Add 6 mL of *hydrofluoric acid* R and evaporate to dryness at 95-105 °C, taking care to avoid loss from spattering. Wash

the inside of the crucible with 6 mL of *hydrofluoric acid* R and evaporate to dryness again. Ignite at 900 ± 50 °C, allow

to cool in a desiccator and weigh. The difference between the mass of the final residue and that of the mass obtained in the test for loss on ignition corresponds to the mass of SiO₂ in

the test sample.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate

compliance. Control of these characteristics can however contribute

to the quality of a medicinal product by improving the consistency

of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristic may be relevant for dental type silica

used as abrasive.

Specific surface area (2.9.26, Method I)

Determine the specific surface area in the *P/P₀* range of

0.05 to 0.30.

Sample outgassing 60 min at 160 °C.

Colloidal Silver for External Use

(Ph. Eur. monograph 2281)

9015-51-4



Action and use

Antibacterial.

Ph. Eur.

DEFINITION

Colloidal metallic silver containing protein.

Content

70.0 per cent to 80.0 per cent of Ag (dried substance).

CHARACTERS

Appearance

Green or bluish-black metallic shiny flakes or powder,

hygroscopic.

Solubility

Freely soluble or soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. To 5 mL of the filtrate obtained in the test for alkalinity (see Tests) add 0.05 mL of *copper sulfate solution* R and 1 mL of *dilute sodium hydroxide solution* R. Shake. A violet colour

appears within 15 min.

B. To 1 mL of solution S (see Tests), add 2 mL of *sodium*

chloride solution R. A precipitate is formed which dissolves in

an excess of water.

C. Ignite 0.05 g of the substance to be examined. Dissolve

the residue in 10 mL of *nitric acid* R. The filtrate gives the

reaction of silver (2.3.1).

TESTS

Solution S

Dissolve 1.25 g of the substance to be examined in *carbon*

dioxide-free water R and dilute to 50.0 mL with the same

solvent. Allow to stand for 5 min then shake vigorously.

Filter through a tared sintered-glass filter (16) (2.1.2) after

Alkalinity

To 40.0 mL of solution S add 10.0 mL of 0.05 M *sulfuric*

acid and 2.0 g of *anhydrous sodium sulfate* R. Shake and filter

several times if necessary. To 25.0 mL of the clear and

colourless solution add 0.1 mL of *phenolphthalein solution* R.

Not less than 1.5 mL of 0.1 M *sodium hydroxide* is required

to change the colour of the indicator to pink.

Silver ions

To 0.50 g of the substance to be examined add 5 mL of

anhydrous ethanol R. Shake for 1 min, filter and add 2 mL of

hydrochloric acid R to the filtrate. No precipitate is formed.

Sensitivity to electrolytes

Dissolve 0.1 g of the substance to be examined in 100 mL of

water R. Transfer a part of the solution into a test tube.

When examined viewing horizontally the solution appears

clear and reddish-brown. When examined viewing vertically,

fluorescence. To 5 mL of the solution add 5 mL of a

solution of 0.50 g/L *sodium chloride* R and mix by shaking for

1 minute. When examined viewing horizontally the solution

remains clear and reddish-brown.

Water insoluble substances

Maximum 1.0 per cent

Wash the residue obtained on the filter during preparation of

solution S 5 times with 10 mL of *water* R. Dry the filter to

constant mass at 100-105 °C. The residue weighs a

maximum of 12.5 mg.

Loss on drying (2.2.32)

Maximum 8.0 per cent, determined on 0.500 g by drying in

an oven at 80 °C.

ASSAY

Ignite 0.200 g of the substance to be examined at

650 ± 50 °C until the residue is white. Allow to cool, add

10 mL of a mixture of equal volumes of *nitric acid* R and

water R and boil for 1 min. Transfer the contents of the

crucible into a flask and titrate with 0.1 M *ammonium*

thiocyanate using 50 mg *ferric sulfate* R as indicator, until a

reddish-brown colour appears.

1 mL of 0.1 M *ammonium thiocyanate* is equivalent to

10.79 mg of Ag.

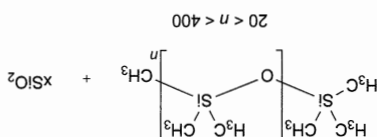
STORAGE

In an airtight container.

Ph. Eur.

Simeticone

(Ph. Eur. monograph 1470)



Action and use

Silicon dioxide analogue; defoaming agent.

8050-81-5



Silver Nitrate

(Ph. Eur. monograph 0009)

AgNO₃ 169.9 7761-88-8



Action and use

Antiseptic.

Preparations

Silver Nitrate Solution

Sterile Silver Nitrate Solution

Ph. Eur.

DEFINITION

99.0 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or transparent, colourless crystals.

Solubility

Very soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. 10 mg gives the reaction of nitrates (2.3.1).

B. 10 mg gives the reaction of silver (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in *water* R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 2 mL of solution S add 0.1 mL of bromocresol green solution R. The solution is blue. To 2 mL of solution S add 0.1 mL of phenol red solution R. The solution is yellow.

Foreign salts

Maximum 0.3 per cent.

To 30 mL of solution S, add 7.5 mL of dilute hydrochloric acid R, shake vigorously, heat for 5 min on a water-bath and filter. Evaporate 20 mL of the filtrate to dryness on a water-bath and dry at 100-105 °C. The residue weighs a maximum of 2 mg.

Aluminium, lead, copper and bismuth

Dissolve 1.0 g in a mixture of 4 mL of concentrated ammonia R and 6 mL of *water* R. The solution is clear (2.2.1) and colourless (2.2.2, Method II).

ASSAY

Dissolve 0.300 g in 50 mL of *water* R, add 2 mL of dilute nitric acid R and 2 mL of ferric ammonium sulfate solution R2.

Titrate with 0.1 M ammonium thiocyanate until a reddish-yellow colour is obtained.

1 mL of 0.1 M ammonium thiocyanate is equivalent to 16.99 mg of AgNO₃.

STORAGE

In a non-metallic container, protected from light.

Ph. Eur.

TESTS

Acidity

To 2.0 g add 25 mL of a mixture of equal volumes of anhydrous ethanol R and ether R, previously neutralised to 0.2 mL of bromothymol blue solution R1, and shake. Not more than 3.0 mL of 0.01 M sodium hydroxide is required to change the colour of the solution to blue.

Defoaming activity

Foaming solution Dissolve 5.0 g of docosate sodium R in 1 L of *water* R, warm to 50 °C if necessary.

Defoaming solution To 50 mL of methyl ethyl ketone R add 0.250 g of the substance to be examined, warm to not more than 50 °C with shaking.

Into a 250 mL cylindrical tube about 5 cm in diameter introduce 100 mL of foaming solution and 1 mL of defoaming solution. Close tightly and fix the tube on a suitable oscillating shaker that complies with the following conditions:

- 250-300 oscillations per minute;
- angle of oscillation of about 10°;
- oscillation radius of about 10 cm.

Shake for 10 s and record the time between the end of the shaking and the instant the 1st portion of foam-free liquid surface appears.

This duration is not longer than 15 s.

Mineral oils

Place 2.0 g in a test-tube and examine in ultraviolet light at 365 nm. The fluorescence is not more intense than that of a solution containing 0.1 ppm of quinine sulfate R in 0.005 M sulfuric acid examined in the same conditions.

Phenylated compounds

The corrected absorbance (2.2.25) is not greater than 0.2.

Test solution Dissolve 5.0 g with shaking in 10.0 mL of cyclohexane R.

Spectral range 200-350 nm.

Calculate the corrected absorbance using the following expression:

$$B - C$$

B = absorbance at the absorption maximum between 250 nm and 270 nm;

C = absorbance at 300 nm.

Heavy metals

Maximum 5 ppm.

Mix 1.0 g with methylene chloride R and dilute to 20 mL with the same solvent. Add 1.0 mL of a freshly prepared 0.02 g/L solution of diethylenetriamine R in methylene chloride R, 0.5 mL of water R and 0.5 mL of a mixture of 1 volume of dilute ammonia R2 and 9 volumes of a 2 g/L solution of hydroxylamine hydrochloride R. At the same time, prepare the reference solution as follows: to 20 mL of methylene chloride R add 1.0 mL of a freshly prepared 0.02 g/L solution of diethylenetriamine R in methylene chloride R, 0.5 mL of lead standard solution (10 ppm Pb) R and 0.5 mL of a mixture of 1 volume of dilute ammonia R2 and 9 volumes of a 2 g/L solution of hydroxylamine hydrochloride R. Immediately shake each solution vigorously for 1 min. Any red colour in the test solution is not more intense than that in the reference solution.

Volatile matter

Maximum 1.0 per cent, determined on 1.00 g by heating in an oven at 150 °C for 2 h. Carry out the test using a dish 60 mm in diameter and 10 mm deep.

ASSAY

Silica

Heat not less than 20.0 mg to 800 °C increasing the temperature by 20 °C/min under a current of nitrogen R at a flow rate of 200 mL/min and weigh the residue (silica).

Poly(dimethylsiloxane)

Infrared absorption spectrophotometry (2.2.24).

Test solution Place about 50 mg (E) in a screw-capped 125 mL cylindrical tube, add 25.0 mL of toluene R, swirl manually to disperse and add 50 mL of dilute hydrochloric

$$A_M = \frac{A_E \times E}{25 \times C \times A_M \times 100}$$

A_M = absorbance of the test solution;

A_E = absorbance of the reference solution;

C = concentration of the reference solution, in milligrams per millilitre;

E = mass of the substance to be examined, in milligrams.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.1.5). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related

characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for simeticone used as defoaming agent.

Defoaming activity

See Tests.

Simeticone for Oral Use

Action and use

Silicon dioxide analogue; defoaming agent.

Preparation

Simeticone Suspension for Infants

When activated dimethicone is prescribed or demanded, Simeticone for Oral Use shall be dispensed or supplied.

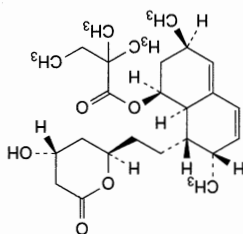
DEFINITION

Simeticone for Oral Use contains not less than 4.5% and not more than 8.0% w/w of silica, SiO₂, and not less than 91.0% and not more than 97.0% w/w of Dimeticone 1000. It is



Simvastatin

(Ph. Eur. monograph 1563)



418.6

$C_{25}H_{38}O_5$

79902-63-9

Action and use

HMG Co-A reductase inhibitor; lipid-regulating drug.

Preparation

Simvastatin Tablets

Simvastatin Oral Suspension

Ph Eur

DEFINITION

(1S,3R,7S,8S,8aR)-8-[2-[(2R,4R)-4-Hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,7,8,8a-hexahydronaphthalen-1-yl 2,2-dimethylbutanoate.

Content

97.0 per cent to 102.0 per cent (dried substance).
A suitable antioxidant may be added.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, very soluble in methylene chloride, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison *simvastatin* CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Dissolve 0.20 g in *methanol* R and dilute to 20 mL with the same solvent.

Specific optical rotation (2.2.7)

+ 285 to + 300 (dried substance).

Dissolve 0.125 g in *acetonitrile* R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture Mix 40 volumes of a 1.4 g/L solution of *potassium dihydrogen phosphate* R, adjusted to pH 4.0 with *phosphoric acid* R, and 60 volumes of *acetonitrile* R. Filter.

Test solution Dissolve 75.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 1.0 mg of *simvastatin* CRS and 1.0 mg of *lovastatin* CRS (impurity E) in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

TESTS

Acidity

Dissolve 15.0 g in a mixture of 15 mL of *toluene* and 15 mL of *butan-1-ol*, previously neutralised to a 0.5% w/v solution of *bromophenol blue* in *ethanol* (96%), and titrate with 0.1M *ethanolic potassium hydroxide* VS using the *bromophenol blue* solution as indicator. Not more than 0.15 mL is required to change the colour of the solution.

Refractive index

Of the supernatant liquid obtained in Identification test A, 1.4050 to 1.4080, Appendix V E.

Viscosity

Of the supernatant liquid obtained in Identification test A, 950 to 1050 mm² s⁻¹, Appendix V H, Method I, using a U-tube viscometer (size G).

Weight per mL

Of the supernatant liquid obtained in Identification test A, 0.965 to 0.980 g, Appendix V G.

Heavy metals

1.0 g complies with limit test F for heavy metals, Appendix VII (20 ppm). Use 2 mL of lead standard solution (10 ppm Pb) to prepare the standard.

ASSAY

For silica

To 1 g add 50 mL of *toluene*, mix well and filter through an ignited silica crucible. Wash the residue thoroughly with *toluene*, dry at 105° and ignite to constant weight at 500°.

For dimeticone

Place 40 mg in a stoppered centrifuge tube, add 20 mL of *toluene* and shake for 20 minutes. Filter and record the infrared absorption spectrum, Appendix II A, of a 0.5-mm layer of the filtrate over the range 1330 to 1180 cm⁻¹ (7.52 to 8.47 μm). Measure the absorbance of the CH₃Si stretching band at the maximum at 1261 cm⁻¹ (7.93 μm). Repeat the operation using *dimeticone* EP CRS in place of the preparation being examined.

Dimeticone 1000.
prepared by the addition of finely powdered silica to

Reference solution (b) Dilute 0.5 mL of the test solution to 100.0 mL with the solvent mixture.
Reference solution (c) Dissolve 75.0 mg of simvastatin CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.
Reference solution (d) Dissolve 5 mg of simvastatin for peak identification CRS (containing impurities A, B, C, D, E, F and G) in 5.0 mL of the solvent mixture.

Column:
 — size: $l = 0.033$ m, $\varnothing = 4.6$ mm;
 — stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 μ m).
Mobile phase:
 — mobile phase A: mix 50 volumes of acetonitrile R and 50 volumes of a 0.1 per cent V/V solution of phosphoric acid R;
 — mobile phase B: 0.1 per cent V/V solution of phosphoric acid R in acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 4.5	100	0
4.5 - 4.6	100 \rightarrow 95	0 \rightarrow 5
4.6 - 8.0	95 \rightarrow 25	5 \rightarrow 75
8.0 - 11.5	25	75

Detection Spectrophotometer at 238 nm.

Injection 5 μ L of the test solution and reference solutions (a), (b) and (d).

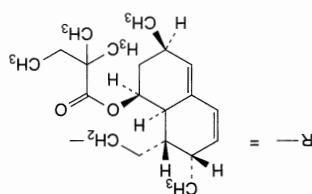
Identification of impurities Use the chromatogram supplied with simvastatin for peak identification CRS and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A, B, C, D, E + F and G.

Relative retention With reference to simvastatin (retention time = about 2.6 min): impurity A = about 0.5; impurities E + F = about 0.6; impurity G = about 0.8; impurities B and C = about 2.4; impurity D = about 3.8.

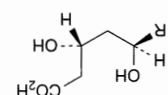
System suitability: reference solution (a):
 — resolution: minimum 4.0 between the peaks due to impurity E and simvastatin.

Limits:

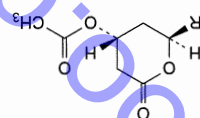
— sum of impurities E and F: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
 — impurities A, D, G: for each impurity, not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.8 per cent);
 — unspecified impurities: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
 — sum of impurities other than E and F: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
 — disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).



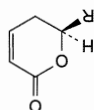
IMPURITIES
 Specified impurities A, B, C, D, E, F, G.



A. (3R,5R)-7-[(1S,2S,6R,8S,8aR)-8-[(2,2-dimethylbutanoyl)oxy]-2,6-dimethyl-1,2,6,7,8,8a-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoic acid (tenivastatin).



B. (1S,3R,7S,8S,8aR)-8-[(2R,4R)-4-(acetyloxy)-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,7,8,8a-hexahydronaphthalen-1-yl 2,2-dimethylbutanoate,



C. (1S,3R,7S,8S,8aR)-3,7-dimethyl-8-[(2R)-6-oxo-3,6-dihydro-2H-pyran-2-yl]ethyl]-1,2,3,7,8,8a-hexahydronaphthalen-1-yl 2,2-dimethylbutanoate,

ASSAY
 Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection Test solution and reference solution (c).
 Calculate the percentage content of simvastatin from the declared content of simvastatin CRS.
STORAGE
 Protected from light. If no antioxidant is present, store under nitrogen, in an airtight container.

IMPURITIES
 Specified impurities A, B, C, D, E, F, G.
Loss on drying (2.2.32)
 Maximum 0.5 per cent, determined on 1.000 g by drying in a desiccator under high vacuum at 60 °C for 3 h.
Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.
Heavy metals (2.4.8)
 Maximum 20 ppm.
 1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

0.1M sodium hydroxide VS; not more than 0.2 mL is required. If the solution is pink add, in a thin stream, 5 mL of hot barium chloride solution previously neutralised to phenolphthalein solution R1, mix thoroughly and titrate with 0.1M hydrochloric acid VS until the pink colour disappears; not more than 1.0 mL is required.

Total free alkali

To 100 mL of the neutral ethanol prepared as described in the test for Free fatty acid or alkali hydroxide add 10 g of the substance being examined and dissolve it as quickly as possible by heating under a reflux condenser. Add immediately 3 mL of 0.5M sulfuric acid VS and boil under a reflux condenser on a water bath for at least 10 minutes. If the solution is not pink, cool to 70° and titrate with 1M sodium hydroxide VS until a pink colour is produced. The volume of 0.5M sulfuric acid VS neutralised by the substance being examined is not more than 1.0 mL.

Unsapönifiable matter and unsapönified neutral fat

Dissolve 5 g in 80 mL of a mixture of 50 mL of ethanol (96%) and 100 mL of water, without heating more than is necessary, and transfer to a separating funnel, washing the vessel with the remaining 70 mL of the mixture. Extract with 100 mL of ether while still slightly warm, run off the ethanolic soap layer into a second separating funnel and extract with 50 mL of ether. Repeat the extraction with 50 mL of ether and pour the three ether extracts into a separating funnel without violent shaking and, after allowing the liquids to separate, run off the water. Repeat the washing with water in the same manner until the separated washings are not more than faintly turbid when acidified. Wash the ether solution twice by shaking vigorously with 20 mL of 0.5M potassium hydroxide, each washing with alkali being immediately followed by washing with 20 mL of water, shaking vigorously each time. Acidify the last alkali washing after separation and, if the liquid becomes turbid, repeat the washing with 0.5M potassium hydroxide and water until the alkali washing remains clear on acidification. Finally wash with successive 20 mL quantities of water until the washings do not give a pink colour with phenolphthalein solution R1. Transfer the ether solution to a tared flask and remove the ether. When nearly all the ether has evaporated, add 3 mL of acetone. With the aid of a gentle current of air remove the solvent completely from the flask, which is preferably almost entirely immersed in boiling water, held obliquely and rotated. Repeat the last operation until the weight of the residue is constant. The residue weighs not more than 40 mg.

CHARACTERISTICS OF THE FATTY ACIDS OBTAINED IN THE ASSAY

Acid value

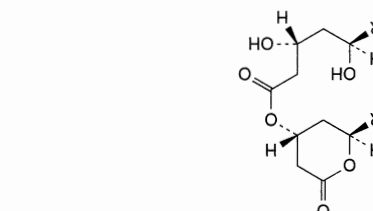
Not more than 205, Appendix X B, when determined on 2 to 3 g of the fatty acids, using 0.5M potassium hydroxide VS and substituting 2.805 for 5.610 in the formula.

Iodine value

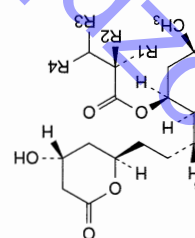
Not less than 83 (iodine monochloride method), Appendix X E. Solidifying point

Not above 31°, Appendix V B, with the following

modifications. Where the determination is made at 15° to 20° the 1000 mL beaker and cooling liquid need not be used. Where the room temperature falls outside this range the 1000 mL beaker should contain water maintained at 15° and the level of this water should not be below the level of the sample in the inner tube.



D. (2R,4R)-2-[2-[(1S,2S,6R,8S,8AR)-8-[(2,2-dimethylbutanoyl)oxy]-2,6-dimethyl-1,2,6,7,8,8A-hexahydronaphthalen-1-yl]ethyl]-6-oxotetrahydro-2H-pyran-4-yl (3R,5R)-7-[(1S,2S,6R,8S,8AR)-8-[(2,2-dimethylbutanoyl)oxy]-2,6-dimethyl-1,2,6,7,8,8A-hexahydronaphthalen-1-yl]-3,5-dihydroxyheptanoate,



E. R1 = R4 = CH₃, R2 = R3 = H; lovastatin,
F. R1 = R3 = H, R2 = R4 = CH₃; (1S,3R,7S,8S,8AR)-8-[2-[(2R,4R)-4-hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,7,8,8A-hexahydronaphthalen-1-yl (2R)-2-methylbutanoate (epilovastatin),
G. R1 = R2 = CH₃, R3 + R4 = CH₂; (1S,3R,7S,8S,8AR)-8-[2-[(2R,4R)-4-hydroxy-6-oxotetrahydro-2H-pyran-2-yl]ethyl]-3,7-dimethyl-1,2,3,7,8,8A-hexahydronaphthalen-1-yl
2,2-dimethylbut-3-enoate.

Soft Soap

Preparation

Soap Spirit

DEFINITION

Soft Soap is soap made by the interaction of potassium hydroxide or sodium hydroxide with a suitable vegetable oil or oils or with fatty acids derived there from. It yields not less than 44.0% of fatty acids. It may be coloured with chlorophyll or not more than 0.015% of a suitable green soap dye.

CHARACTERISTICS

A yellowish white to green or brown, unctuous substance. Soluble in water and in ethanol (96%).

TESTS

Chlorides and other ethanol-insoluble substances

Dissolve 5 g in 100 mL of hot ethanol (96%) previously neutralised to phenolphthalein solution R1, filter through a dried and tared filter, wash the residue thoroughly with hot neutralised ethanol (96%) and dry to constant weight at 105°. The residue weighs not more than 0.15 g.

Free fatty acid or alkali hydroxide

Boil 250 mL of ethanol (96%) to remove carbon dioxide, add 0.5 mL of phenolphthalein solution R1, allow to cool to 70° and neutralise, if necessary, with 0.1M sodium hydroxide VS or 0.05M sulfuric acid VS. To 100 mL of the neutral ethanol add 10 g of the substance being examined and dissolve it as quickly as possible by heating under a reflux condenser. Cool to 70° and, if the solution is not pink, titrate at 70° with

8-87-9008

Used to absorb carbon dioxide.

Soda Lime is a mixture of sodium hydroxide, or sodium hydroxide and potassium hydroxide, with calcium hydroxide.

White or greyish white granules, or it may be coloured with an indicator to show when its absorptive capacity is exhausted. It absorbs about 20% of its weight of carbon

A. When moistened with *hydrochloric acid* and introduced on a platinum wire into a flame, imparts a yellow colour to the

Hardness of granules

Shake 200 g on a sieve no. 2000 for 3 minutes using a mechanical sieve shaker that reproduces in a uniform manner the circular and tapping motion given to sieves in manual use and has a frequency of oscillation of 282 to 288 cycles per minute. Place 50 g of the retained material in a hardness pan

20 cm in diameter having a concave brass bottom, 7.9 mm thick at the circumference, 3.2 mm thick at the centre and with an inside spherical radius of curvature of 109 cm. Add 15 steel balls, 7.9 mm in diameter, and shake on the mechanical sieve shaker for 30 minutes. Remove the steel balls, transfer the contents of the pan to a sieve no. 2000 and again shake on the mechanical sieve shaker for 3 minutes. The material retained by the sieve weighs not less than

When dried to constant weight at 105°, loses 14.0 to 21.0% of its weight. Use 1 g.

Place 10 g in an open glass dish about 50 mm in diameter and 30 mm high in a desiccator over *sulfuric acid* (14%) and allow it to remain for 24 hours. The increase in weight is not more than 7.5%.

I am active in my leisure time and I am interested in many things by the following method.

Use a vertically-clamped tube of glass or other suitable transparent material about 25 cm long and 29 to 31 mm internal diameter with closely fitting rubber bungs at each end; the bungs are bored to receive polythene or glass tubing of 8 mm external diameter, the tubing being flush with

the inner ends of the bungs. With the lower bung in position, place sufficient nylon support on top of the bung to produce a bed of mesh about 10 cm deep and press a closely fitting disc of stainless steel (size of nominal mesh aperture

about 500 μm on top of the nylon mesh so that its surface is at right angles to the axis of the tube. Introduce 59.8 to 60.3 g of the substance being examined onto the steel gauze

portion. Place a second disc of steel gauze on top, followed by a sufficient quantity of nylon mesh such that the soda

The exit tube is connected to a condenser, consisting of two L-shaped tubes packed with calcium chloride and then a carbon dioxide analysis efficiently

A katharometer, calibrated for carbon dioxide and preferably sensitive to detect 0.2% v/v of carbon dioxide, is used in conjunction with a chart recorder, is suitable. Using the gas analyser in accordance with the manufacturer's

insistencies, accurately determine the carbon dioxide content, p , as a percentage V/V , of a nominal 5% carbon dioxide mixture, the balance gas being oxygen at or

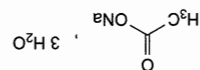
nitrogen as appropriate to the type of gas analyser being used. Suitable compressed gas mixtures are available commercially. Assemble the apparatus described above and pass the gas mixture downwards into the absorption tube at a

rate of 900 cm³ per minute until such time that the gas analyser shows the content of carbon dioxide in the effluent gas to have risen to 0.2% v/v. Steps should be taken to vent the effluent gas if an oxygen-carbon dioxide mixture is being

used. Record the time taken, t_1 in minutes. The activity of the soda lime is given, by the expression tp/c .

Sodium Acetate Trihydrate

(Ph. Eur. monograph 0411)



$C_2H_3NaO_3 \cdot 3H_2O$ 136.1 6131-90-4

Action and use

Used in solutions for dialysis; excipient.

Preparation

Sterile Sodium Acetate Concentrate

DEFINITION

Sodium ethanoate trihydrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance Colourless crystals.

Solubility

Very soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. 1 mL of solution S (see Tests) gives reaction (b) of acetates (2.3.1).

B. 1 mL of solution S gives reaction (a) of sodium (2.3.1).

C. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

7.5 to 9.0.

Dilute 5 mL of solution S to 10 mL with carbon dioxide-free water R.

Reducing substances

Dissolve 5.0 g in 50 mL of water R, then add 5 mL of dilute sulfuric acid R and 0.5 mL of 0.002 M potassium permanganate. The pink colour persists for at least 1 h.

Prepare a blank in the same manner but without the substance to be examined.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 200 ppm.

Dilute 7.5 mL of solution S to 15 mL with distilled water R.

Aluminium (2.4.17)

Maximum 0.2 ppm, if intended for use in the manufacture of dialysis solutions.

Prescribed solution Dissolve 20 g in 100 mL of water R and adjust to pH 6.0 by the addition of 1 M hydrochloric acid (about 10 mL).

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Calcium and magnesium

Maximum 50 ppm, calculated as Ca.

To 200 mL of water R add 10 mL of ammonium chloride buffer solution pH 10.0 R, 0.1 g of mordant black 11 triturate R, 2.0 mL of 0.05 M zinc chloride and, dropwise, 0.02 M sodium edetate until the colour changes from violet to blue. Add to the solution 10.0 g of the substance to be examined and shake to dissolve. Titrate with 0.02 M sodium edetate until the blue colour is restored. Not more than 0.65 mL of 0.02 M sodium edetate is required.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Iron (2.4.9)

Maximum 10 ppm, determined on 10 mL of solution S.

Loss on drying (2.2.32)

39.0 per cent to 40.5 per cent, determined on 1.000 g by drying in an oven at 130 °C. Introduce the substance to be examined into the oven while the latter is cold.

ASSAY

Dissolve 0.250 g in 50 mL of anhydrous acetic acid R, add 5 mL of acetic anhydride R, mix and allow to stand for 30 min. Using 0.3 mL of naphtholbenzoin solution R as indicator, titrate with 0.1 M perchloric acid until a green colour is obtained.

1 mL of 0.1 M perchloric acid is equivalent to 8.20 mg of $C_2H_3NaO_3$.

STORAGE

In an airtight container.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of dialysis solutions.

Sodium Acid Citrate

Sodium Hydrogen Citrate

$C_6H_6Na_2O_7 \cdot 1\frac{1}{2}H_2O$

263.1

144-33-2

Action and use

Anticoagulant.

DEFINITION

Sodium Acid Citrate contains not less than 98.0% and not more than 104.0% of $C_6H_6Na_2O_7 \cdot 1\frac{1}{2}H_2O$.

CHARACTERISTICS

A white powder.

Freely soluble in water, practically insoluble in ethanol (96%).

IDENTIFICATION

Yields the reactions characteristic of sodium salts and of citrates, Appendix VI.

TESTS

Acidity

pH of a 3% w/v solution, 4.9 to 5.2, Appendix V L.

Arsenic
0.50 g dissolved in 25 mL of water complies with the *limit test* for arsenic, Appendix VII (2 ppm).

Heavy metals

Dissolve 1.0 g in 20 mL of water. 12 mL of the resulting

solution complies with *limit test A* for heavy metals,

Appendix VII. Use *lead standard solution* (1 ppm Pb) to

prepare the standard (20 ppm).

Chloride

Dissolve 1.0 g in 100 mL of water. 15 mL of the resulting

solution complies with the *limit test* for chlorides, Appendix VII

(330 ppm).

Oxalate

Dissolve 1.0 g in 4 mL of water, add 3 mL of hydrochloric

acid and 1 g of granulated zinc and heat on a water bath for

1 minute. Allow to stand for 2 minutes, decant the liquid

into a test tube containing 0.25 mL of a 1% w/v solution of

phenylhydrazine hydrochloride and heat to boiling. Cool

rapidly, transfer to a graduated measuring cylinder, add an

equal volume of hydrochloric acid and 0.25 mL of potassium

hexacyanoferrate(III) solution, shake and allow to stand for

30 minutes. Any red colour produced is not more intense

than that produced by treating in the same manner 4 mL of

a 0.005% w/v solution of oxalic acid (150 ppm, calculated as

anhydrous oxalic acid).

Sulfate

Dissolve 0.50 g in 57 mL of water and add 3 mL of

2M hydrochloric acid. 15 mL of the resulting solution complies

with the *limit test* for sulfates, Appendix VII (0.12%).

Readily carbonisable substances

Heat 1.0 g, in powder, with 10 mL of sulfuric acid for

30 minutes in a water bath protected from light. Not more

than a pale brown colour is produced.

ASSAY

Heat 2 g until carbonised, cool and boil the residue with

50 mL each of water and 0.5M hydrochloric acid VS. Filter,

wash the filter with water and titrate the excess of acid in the

filtrate and washings with 0.5M sodium hydroxide VS using

methyl orange solution as indicator. Each mL of

0.5M hydrochloric acid VS is equivalent to 65.78 mg of

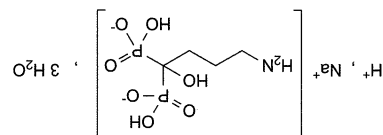
$C_6H_6Na_2O_7 \cdot 1\frac{1}{2}H_2O$.

Sodium Alendronate Trihydrate



Sodium Alendronate

(Ph. Eur. monograph 1564)



$C_4H_{12}NNaO_7P_2 \cdot 3H_2O$ 325.1

121268-17-5

DEFINITION

Monosodium trihydrogen (4-amino-1-hydroxybutylidene)bisphosphonate trihydrate.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, Practically insoluble in methanol and in

ethylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium alendronate trihydrate CRS.

B. Loss on drying (see Tests).

C. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 0.5 g in carbon dioxide-free water R prepared from

distilled water R and dilute to 50 mL with the same solvent.

pH (2.2.3)

4.0 to 5.0 for solution S.

Related substances

Liquid chromatography (2.2.29): use the normalisation

procedure.

Solution A Dissolve 29.4 g of sodium citrate R in water R and

dilute to 1.0 L with the same solvent.

Solution B Dissolve 19.1 g of disodium tetraborate R in water R

and dilute to 1.0 L with the same solvent.

Solution C Prepare immediately before use. Dissolve 0.200 g of

(9-fluorenyl)methyl chloroformate R in acetonitrile R and dilute

to 50.0 mL with the same solvent.

Buffer solution Dissolve 2.84 g of anhydrous disodium hydrogen

phosphate R and 5.88 g of sodium citrate R in 1.9 L of

water R, adjust to pH 8.0 with either phosphoric acid R or a

42 g/L solution of sodium hydroxide R and dilute to 2.0 L

with water R.

Test solution Dissolve 30 mg of the substance to be examined

in solution A and dilute to 50 mL with solution A. Transfer

5 mL of the solution to a 50 mL polypropylene screw-cap

centrifuge tube containing 5 mL of solution B. Add 5 mL of

acetonitrile R and 5 mL of solution C. Shake for 45 s and

allow to stand at room temperature for 30 min. Add 20 mL

of methylene chloride R and shake vigorously for 1 min.

Centrifuge for 5-10 min and use the clear upper layer.

Reference solution (a) Dissolve 15 mg of 4-aminobutanoic

acid R (impurity A) in solution A and dilute to 100 mL with

solution A. Dilute 10 mL of the solution to 50 mL with

solution A. Take 5 mL of this solution and proceed as

described for the test solution, starting from 'to a 50 mL

polypropylene screw-cap centrifuge tube'.

Reference solution (b) Dissolve 3 mg of sodium alendronate for

system suitability CRS (containing impurity D) in solution A

and dilute to 5 mL with solution A. Take this solution and

proceed as described for the test solution, starting from 'to a

50 mL polypropylene screw-cap centrifuge tube'.

Blank solution Take 5 mL of solution A and proceed as

described for the test solution, starting from 'to a 50 mL

polypropylene screw-cap centrifuge tube'.

Column:

polypropylene screw-cap centrifuge tube'.

size: $l = 0.25$ m, $\phi = 4.1$ mm;

stationary phase: styrene-divinylbenzene copolymer R

(10 μ m);

temperature: 45 °C.

Mobile phase:

mobile phase A: acetonitrile R, buffer solution (15:85 V/V);

mobile phase B: buffer solution, acetonitrile R (30:70 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	100	0
3 - 18	100 → 50	0 → 50
18 - 28	50 → 0	50 → 100

Flow rate 1.8 mL/min.

Detection Spectrophotometer at 266 nm.

Injection 20 µL of the test solution, reference solutions (a), (b) and the blank solution.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to

alendronate for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity D.

Relative retention With reference to alendronate (retention time = about 7 min); impurity D = about 1.4;

impurity A = about 1.9.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to alendronate and impurity D.

Limits:

— correction factor for the calculation of content, multiply the peak area of impurity A by 0.4;

— impurity A: maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum 0.10 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

Impurities B and C

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in water R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of sodium alendronate trihydrate CRS in water R and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dissolve 3.0 g of phosphoric acid R in water R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with water R.

Reference solution (c) Dissolve 2.5 g of phosphorous acid R in water R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 100.0 mL with water R.

Reference solution (d) Mix 2.0 mL of reference solution (b) with 2.0 mL of reference solution (c) and dilute to 50.0 mL with water R.

Mobile phase Mix 0.2 mL of anhydrous formic acid R with 1 L of water R; adjust to pH 3.5 with 2 M sodium hydroxide R.

Flow rate 1.2 mL/min.

Detection Differential refractometer.

Injection 100 µL Of the test solution and reference solutions (b), (c) and (d).

Run time Twice the retention time of alendronate.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity C.

Appearance White or pale yellowish-brown powder.

Sodium Alginate

(Ph. Eur. monograph 0625)

Action and use

Excipient.

Preparations

Alginate Raft-forming Oral Suspension

Compound Alginate Antacid Oral Suspension

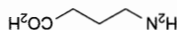
DEFINITION

Sodium alginate consists mainly of the sodium salt of alginic acid, which is a mixture of polyuronic acids $[(C_6H_8O_6)_n]$ composed of units of D-mannuronic acid and L-guluronic acid. Sodium alginate is obtained mainly from algae belonging to the Phaeophyceae.

CHARACTERS

Appearance

White or pale yellowish-brown powder.



Specified impurities A, B, C
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D.

ASSAY
Liquid chromatography (2.2.29) as described in the test for impurities B and C with the following modification.
Injection Test solution and reference solution (a).
Calculate the percentage content of $C_4H_7NNaO_7P_2$ taking into account the assigned content of sodium alendronate trihydrate CRS.

Loss on drying (2.2.32)
16.1 per cent to 17.1 per cent, determined on 1.000 g by drying in an oven at 140-145 °C.

Heavy metals (2.4.8)
Maximum 20 ppm.

Relative retention With reference to alendronate (retention time = about 16 min); impurity B = about 1.3; impurity C = about 1.6.
Limits:
— impurities B, C: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.5 per cent).



Solubility

Slowly soluble in water forming a viscous, colloidal solution, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 0.2 g with shaking in 20 mL of water R.
To 5 mL of this solution add 1 mL of calcium chloride solution R. A voluminous gelatinous mass is formed.
B. To 10 mL of the solution prepared in identification test A add 1 mL of dilute sulfuric acid R. A gelatinous mass is formed.

C. To 5 mg add 5 mL of water R, 1 mL of a freshly prepared 10 g/L solution of 1,3-dihydroxynaphthalene R in ethanol (96 per cent) R and 5 mL of hydrochloric acid R. Boil for 3 min, cool, add 5 mL of water R, and shake with 15 mL of di-isopropyl ether R. Carry out a blank test. The upper layer obtained with the substance to be examined exhibits a deeper bluish-red colour than that obtained with the blank.
D. Sulfated ash (see Tests). The residue obtained, dissolved in 2 mL of water R, gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 0.10 g in water R with constant stirring, dilute to 30 mL with the same solvent and allow to stand for 1 h.

Appearance of solution

The solution is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Dilute 1 mL of solution S to 10 mL with water R.

Chlorides

Maximum 1.0 per cent.

To 2.50 g add 50 mL of dilute nitric acid R, shake for 1 h and dilute to 100.0 mL with dilute nitric acid R. Filter. To 50.0 mL of the filtrate add 10.0 mL of 0.1 M silver nitrate and 5 mL of toluene R. Titrate with 0.1 M ammonium thiocyanate, using 2 mL of ferric ammonium sulfate solution R2 as indicator and shaking vigorously towards the end point. 1 mL of 0.1 M silver nitrate is equivalent to 3.545 mg of Cl.

Calcium

Maximum 1.5 per cent.

Atomic absorption spectrometry (2.2.23, Method II).
Test solution Dissolve 0.10 g in 50 mL of dilute ammonia R2, heating on a water-bath. Allow to cool and dilute to 100.0 mL with distilled water R (solution (a)). Dilute 3.0 mL of solution (a) to 100.0 mL with distilled water R.
Reference solutions Prepare 3 reference solutions in the same manner as the test solution but add 0.75 mL, 1.0 mL and 1.5 mL respectively of calcium standard solution (100 ppm Ca) R to the 3.0 mL of solution (a).
Set the zero of the instrument using a mixture of 1.5 volumes of dilute ammonia R2 and 98.5 volumes of distilled water R.
Source Calcium hollow-cathode lamp.
Wavelength 422.7 nm.
Atomisation device Air-acetylene flame.

Heavy metals (2.4.8)

Maximum 20 ppm.

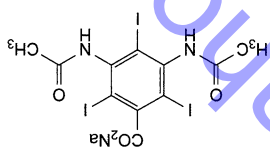
1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 15.0 per cent, determined on 0.1000 g by drying in an oven at 105 °C for 4 h.

Sodium Amidotrizoate

(Ph Eur monograph 1150)



C₁₁H₈I₃N₂NaO₄

636

737-31-5

Action and use

Iodinated contrast medium.

DEFINITION

Sodium amidotrizoate contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of sodium 3,5-bis(acetylamino)-2,4,6-tri-iodobenzoate, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white powder, freely soluble in water, slightly soluble in alcohol, practically insoluble in acetone. It melts at about 261 °C with decomposition.

IDENTIFICATION

First identification A, D
Second identification B, C, D

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with sodium amidotrizoate CRS. Dry both the substance to be examined and the reference substance at 100 °C to 105 °C for 3 h.

Ph Eur



add 3 mL of dilute nitric acid R. Filter and wash the precipitate with 5 mL of water R. Collect the filtrate and washings. Add 1 mL of strong hydrogen peroxide solution R and 1 mL of methylene chloride R. Shake. The lower layer is not more intensely coloured than a reference solution prepared simultaneously and in the same manner, using a mixture of 5 mL of iodide standard solution (10 ppm I) R, 3 mL of dilute nitric acid R and 15 mL of water R.

Heavy metals (2.4.8)

Dilute 4 mL of solution S to 20 mL with water R. 12 mL of this solution complies with test A for heavy metals (20 ppm). Prepare the reference solution using lead standard solution

(2 ppm Pb) R.

Water (2.5.12)

Not more than 11.0 per cent, determined on 0.400 g by the semi-micro determination of water.

ASSAY

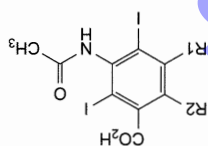
To 0.150 g in a 250 mL round-bottomed flask add 5 mL of strong sodium hydroxide solution R, 20 mL of water R, 1 g of zinc powder R and a few glass beads. Boil under a reflux condenser for 30 min. Allow to cool and rinse the condenser with 20 mL of water R, adding the rinsings to the flask. Filter through a sintered-glass filter (2.1.2) and wash the filter with several quantities of water R. Collect the filtrate and washings. Add 40 mL of dilute sulfuric acid R and titrate immediately with 0.1 M silver nitrate. Determine the end-point potentiometrically (2.2.20), using a suitable electrode system such as silver-mercurous sulfate.

1 mL of 0.1 M silver nitrate is equivalent to 21.20 mg of $C_{11}H_{13}N_2NaO_4$.

STORAGE

Store protected from light.

IMPURITIES



A. R1 = NH_2 , R2 = 1: 3-acetylamino-5-amino-2,4,6-tri-

iodobenzoic acid,

B. R1 = $NHCOCH_3$, R2 = H: 3,5-bis(acetylamino)-2,4-di-

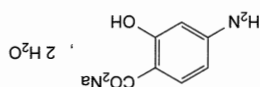
iodobenzoic acid.

Ph Eur



Sodium Aminosalicylate Dihydrate

(Ph. Eur. monograph 1993)



$C_{11}H_{13}N_2NaO_4 \cdot 2H_2O$

211.2

6018-19-5

Action and use

Antituberculous.

Ph Eur

DEFINITION

Sodium 4-amino-2-hydroxybenzoate dihydrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (b).

C. Heat 50 mg gently in a small porcelain dish over a naked flame. Violet vapour is evolved.

D. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 10 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Dilute 1 mL of solution S to 10 mL with water R. The solution is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

The pH of solution S is 7.5 to 9.5.

Related substances

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel GF₂₅₄ plate R. Prepare the solutions in subdued light and develop the chromatograms protected from light.

Test solution (a) Dissolve 0.50 g of the substance to be examined in a 3 per cent V/V solution of ammonia R in methanol R and dilute to 10 mL with the same solution.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with a 3 per cent V/V solution of ammonia R in methanol R.

Reference solution (a) Dilute 1 mL of test solution (b) to 50 mL with a 3 per cent V/V solution of ammonia R in methanol R.

Reference solution (b) Dissolve 50 mg of sodium amidoacetate CRS in 3 per cent V/V solution of ammonia R and dilute to 10 mL with the same solution.

Apply separately to the plate 2 µL of each solution. Develop over a path of 15 cm using a mixture of 20 volumes of anhydrous formic acid R, 25 volumes of methyl ethyl ketone R and 60 volumes of toluene R. Allow the plate to dry until the solvents have evaporated and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.2 per cent).

Free aromatic amines

Maintain the solutions and reagents in iced water, protected from light. To 0.50 g in a 50 mL volumetric flask add 15 mL of water R. Shake and add 1 mL of dilute sodium hydroxide solution R. Cool in iced water, add 5 mL of a freshly prepared 5 g/L solution of sodium nitrite R and 12 mL of exactly 2 min after adding the hydrochloric acid. Add 10 mL of a 20 g/L solution of ammonium sulfamate R. Allow to stand for 5 min, shaking frequently, and add 0.15 mL of a 100 g/L solution of α-naphthol R in alcohol R. Shake and allow to mix and dilute to 50.0 mL with water R. The absorbance (2.2.25), measured within 20 min at 485 nm using as the compensation liquid a solution prepared at the same time and in the same manner but omitting the substance to be examined, is not greater than 0.30.

Free iodine and iodides

Not more than 50 ppm. Dissolve 1.0 g in distilled water R and dilute to 10 mL with the same solvent. Add dropwise dilute nitric acid R until the precipitation is complete, then

CHARACTERS

Appearance

White or almost white crystalline powder or crystals, slightly

hygroscopic.

Solubility

Freely soluble in water, sparingly soluble in ethanol

(96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification A, E

Second identification B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium aminosallylate dihydrate CRS.

B. Introduce 0.3 g into a porcelain crucible. Cautiously heat on a small flame until vapour is evolved. Cover the crucible with a watch glass and collect the white sublimate.

The melting point (2.2.14) of the sublimate is 120 °C to

124 °C.

C. To 0.1 mL of solution S (see Tests) add 5 mL of water R

and 0.1 mL of ferric chloride solution R1. A reddish-brown

colour develops.

D. 2 mL of solution S gives the reaction of primary aromatic

amines (2.3.1).

E. 0.5 mL of solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 0.50 g in carbon dioxide-free water R and dilute to

25 mL with the same solvent.

Appearance of solution

The freshly prepared solution is clear (2.2.1) and not more

intensely coloured than reference solution B₅ (2.2.2,

Method II).

Dissolve 2.5 g in water R and dilute to 25 mL with the same

solvent.

pH (2.2.3)

6.5 to 8.5 for solution S.

Related substances

Liquid chromatography (2.2.29). Use freshly prepared solutions

and mobile phases.

Test solution Dissolve 50.0 mg of the substance to be

examined in water R and dilute to 50.0 mL with the same

solvent.

Reference solution (a) Dissolve 5.0 mg of 3-aminophenol R

(impurity A) in water R and dilute to 100.0 mL with the

same solvent.

Reference solution (b) Dissolve 5.0 mg of mesalazine CRS

(impurity B) in water R and dilute to 100.0 mL with the

same solvent. To 10.0 mL of this solution add 1.0 mL of

reference solution (a) and dilute to 50.0 mL with water R.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm,

— stationary phase: spherical base-deactivated octylsilyl silica gel

for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: dissolve 2.2 g of perchloric acid R and 1.0 g

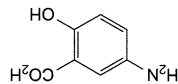
of phosphoric acid R in water R and dilute to 1000.0 mL

with the same solvent,

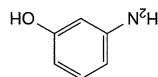
— mobile phase B: dissolve 1.7 g of perchloric acid R and 1.0 g

of phosphoric acid R in acetonitrile R and dilute to

1000.0 mL with the same solvent,



A. 3-aminophenol,



Specified impurities: A, B.

IMPURITIES

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

STORAGE

of $C_7H_6NNaO_3$.

1 mL of 0.1 M sodium nitrite is equivalent to 17.52 mg

end-point potentiometrically (2.2.20).

continue the titration with the same titrant, determining the

acidic acid R. Add 5 mL of 0.1 M sodium nitrite rapidly and

500 g/L solution of sodium bromide R and 25 mL of glacial

Dissolve 0.150 g in 20 mL of water R. Add 10 mL of a

ASSAY

drying in an oven at 105 °C.

16.0 per cent to 17.5 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

using 2 mL of lead standard solution (10 ppm Pb) R.

2.0 g complies with test C. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

solution (b) (0.05 per cent).

impurity B in the chromatogram obtained with reference

— disregard limit: 0.05 times the area of the peak due to

solution (b) (1.0 per cent),

impurity B in the chromatogram obtained with reference

— total: not more than the area of the peak due to

(0.10 per cent),

chromatogram obtained with reference solution (b)

0.1 times the area of the peak due to impurity B in the

— unspecified impurities: for each impurity, not more than

solution (b) (1.0 per cent),

peak in the chromatogram obtained with reference

— impurity B: not more than the area of the corresponding

reference solution (b) (0.15 per cent),

corresponding peak in the chromatogram obtained with

— impurity A: not more than 1.5 times the area of the

Limits:

impurity A and impurity B.

— resolution: minimum 4.0 between the peaks due to

System suitability: reference solution (b):

impurity B = about 0.37.

(retention time = about 12 min): impurity A = about 0.30;

Relative retention With reference to 4-aminosallylate

Injection 10 μ L of the test solution and reference solution (b).

Detection Spectrophotometer at 220 nm.

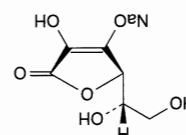
Flow rate 1.25 mL/min.

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 15	100	0
15 - 30	100 \rightarrow 40	0 \rightarrow 60

B. 5-amino-2-hydroxybenzoic acid (mesalazine).

Sodium Ascorbate

(Ph. Eur. monograph 1791)

C₆H₇NaO₆

198.1

134-03-2

Action and use

Excipient.

Ph. Eur.

DEFINITION

Sodium (2*R*)-2-[(1*S*)-1,2-dihydroxyethyl]-4-hydroxy-5-oxo-2,5-dihydrofuran-3-olate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish, crystalline powder or crystals.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Specific optical rotation (2.2.7) (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium ascorbate CRS.

C. To 1 mL of solution S (see Tests) add 0.2 mL of dilute nitric acid R and 0.2 mL of silver nitrate solution R2. A grey precipitate is formed.

D. 1 mL of solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ or BY₆ (2.2.2, Method II); examine the colour immediately after preparation of the solution.

pH (2.2.3)

7.0 to 8.0 determined on freshly prepared solution S.

Specific optical rotation (2.2.7)

+103 to +108 (dried substance), determined on freshly prepared solution S.

Impurity E

Maximum 0.3 per cent.

Test solution Dissolve 0.25 g in 5 mL of water R. Add 1 mL of dilute acetic acid R and 0.5 mL of calcium chloride solution R.

Reference solution Dissolve 70 mg of oxalic acid R in water R and dilute to 500 mL with the same solvent; to 5 mL of the solution add 1 mL of dilute acetic acid R and 0.5 mL of calcium chloride solution R.

Allow the solutions to stand for 1 h. Any opalescence in the test solution is not more intense than that in the reference solution.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Phosphate buffer solution Dissolve 6.8 g of potassium dihydrogen phosphate R in water R and dilute to about 175 mL with the same solvent. Filter through a membrane filter (nominal pore size 0.45 µm) and dilute to 1000 mL with water R.

Test solution Dissolve 0.500 g of the substance to be examined in the phosphate buffer solution and dilute to 10.0 mL with the phosphate buffer solution.

Reference solution (a) Dissolve 10.0 mg of ascorbic acid impurity C CRS in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of ascorbic acid impurity D CRS and 5.0 mg of ascorbic acid CRS in the mobile phase, add 2.5 mL of reference solution (a) and dilute to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase. Mix 1.0 mL of this solution with 1.0 mL of reference solution (a).

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

stationary phase: aminopropylsilyl silica gel for chromatography R (5 µm);

temperature: 45 °C.

Mobile phase Phosphate buffer solution, acetonitrile R1 (25:75 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Run time 2.5 times the retention time of ascorbic acid.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities C and D.

Relative retention With reference to ascorbic acid (retention time = about 1.1 min); impurity D = about 0.4;

impurity C = about 1.7.

System suitability:

— resolution: minimum 3.0 between the peaks due to ascorbic acid and impurity C in the chromatogram

obtained with reference solution (c);

— signal-to-noise ratio: minimum 20 for the peak due to impurity C in the chromatogram obtained with reference solution (b).

Limits:

— impurities C, D: for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (b)

chromatogram obtained with reference solution (b) (0.10 per cent);

— unspecified impurities: for each impurity, not more than the area of the peak due to ascorbic acid in the chromatogram obtained with reference solution (b)

(0.10 per cent);


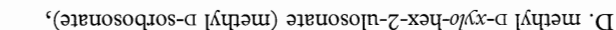
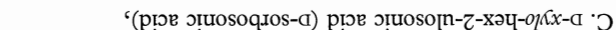
— total of impurities other than C and D: not more than twice the area of the peak due to ascorbic acid in the chromatogram obtained with reference solution (b) (0.2 per cent);

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, F, G, H.

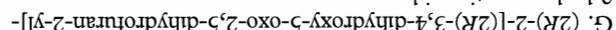
impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, F,

(2034). It is therefore not necessary to identify these

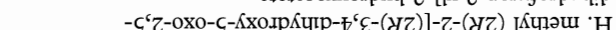
impurities for demonstration of compliance. See also 5.10.

 \mathcal{H}, \mathcal{G} 

7C(H)-α



2-hydroxyacetic acid,



diethylpropan-2-yl]-7-oxoheptanoate.

Maximum 150 ppm.

To 10 mL of solution S add 2 mL of hydrochloric acid R1 and

Copper

Atomic absorption spectrometry (2.2.23, *Method I*).

25.0 mL with the same acid.

0.4 ppm and 0.6 ppm) by diluting copper standard solution

Source: Copper hollow-cathode lamp.

Atomisation device Air-acetylene flame.

Maximum 2 ppm.

Test solution Dissolve 5.0 g in 0.1 M nitric acid and dilute to

Reference solutions Prepare the reference solutions (0.2 ppm,

(20 ppm Fe) R with 0.1 M nitric acid.

Wavelength 248.3 nm.



Atomic absorption spectrometry (2.2.23, Method I).

25.0 mL with the same acid.

0.4 ppm and 0.6 ppm) by diluting nickel standard solution

Source Nickel hollow-cathode lamp.

Atomisation device Air-acetylene flame.

Maximum 10 ppm.

solvent. 12 mL of the solution complies with test A. Prepare

(1 ppm Pb) R.

Maximum 0.25 per cent, determined on 1.000 g by drying in

AVSSV

and 80 mL of carbon dioxide-free water R. Add 1 mL of starch

blue colour is obtained.

 $C_6H_7N_3O_6$

In a non-metallic container, protected from light.

Sodium Aurothiomalate

(Ph. Eur. monograph 1994)

39377-38-3



Action and use
Used in the treatment of rheumatoid arthritis.

Preparation
Sodium Aurothiomalate Injection

Ph. Eur.

DEFINITION

Mixture of monosodium and disodium salts of (2R,5S)-2-(aurothiylamyl)butanedioic acid.

Content

— *gold* (Au, 4, 197.0): 44.5 per cent to 46.0 per cent (dried substance);
— *sodium* (Na, 4, 22.99): 10.8 per cent to 11.8 per cent (dried substance).

CHARACTERS

Appearance
Fine, pale yellow, hygroscopic powder.

Solubility

Very soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Dissolve 20 mg in *water* R and dilute to 2 mL with the same solvent. Add 2 mL of *strong hydrogen peroxide solution* R and 1 mL of *sodium hydroxide solution* R. Carefully heat to boiling and boil for 30 s. A precipitate is produced that appears brownish-black by reflected light and bluish-green by transmitted light.
B. To 20 mg add 2 mL of *water* R. The solution gives reaction (a) of sodium (2.3.1).
C. Ignite 100 mg, dissolve the residue in *hydrochloric acid* R and dilute to 10 mL with the same acid. Allow to stand. 5 mL of the clear supernatant gives reaction (a) of sulfates (2.3.1).

TESTS**Appearance of solution**

Dissolve 1.0 g in *water* R and dilute to 10 mL with the same solvent. Filter, seal in an ampoule and heat at 100 °C for 1 h. Cool and dilute the contents of the ampoule to 100 mL with *water* R. The solution remains clear and is not more intensely coloured than a 0.100 g/L solution of *potassium ferricyanide* R.
pH (2.2.3)
6.0 to 7.0.
Dissolve 1.0 g in *carbon dioxide-free water* R and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.
Test solution Dissolve 0.200 g of the substance to be examined in *water* R and dilute to 100.0 mL with the same solvent.
Reference solution (a) Dissolve 10.0 mg of *fumonic acid* R and 100.0 mg of *thiomalic acid* R in *water* R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 25.0 mL with *water* R.
Reference solution (b) Dissolve 12.0 mg of *thiomalic acid* R in *water* R and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dissolve 10.0 mg of *maleic acid* R in *water* R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 25.0 mL with *water* R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— *stationary phase*: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 90 volumes of a 10.5 g/L solution of *phosphoric acid* R, 100 volumes of *methanol* R2 and 810 volumes of *water* R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 205 nm.

Injection 10 μ L.

Run time Twice the retention time of impurity C.
Relative retention With reference to impurity C (retention time = about 8 min): impurity A = about 0.4; impurity B = about 0.6. Aurothiomalate does not elute under the chromatographic conditions described.

System suitability: reference solution (a):
— *resolution*: minimum 5.0 between the peaks due to impurities B and C.

Limits:

— *impurity A*: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
— *impurity B*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— *impurity C*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (6.0 per cent).

Glycerol

Test solution Dissolve 0.50 g of the substance to be examined in *water* R and dilute to 10 mL with the same solvent.
Reference solution (a) Dilute 0.80 g of *glycerol* R to 100.0 mL with *water* R.
Reference solution (b) To 2.5 mL of reference solution (a) add 7.5 mL of *water* R.
Reference solution (c) To 5.0 mL of reference solution (a) add 5.0 mL of *water* R.
Reference solution (d) To 7.5 mL of reference solution (a) add 2.5 mL of *water* R.
Blank solution 10 mL of *water* R.

To the test solution, reference solutions (b), (c) and (d) and the blank solution, add 2.5 mL of a freshly prepared 235 g/L solution of *sodium hydroxide* R and mix. Add dropwise in 0.2 mL increments a 38.0 g/L solution of *cupric chloride* R, shaking vigorously after each addition, until the solutions become slightly turbid. Then add 0.2 mL of the 38.0 g/L solution of *cupric chloride* R. Stopper and shake vigorously for 1 min. Dilute to 25.0 mL with *water* R and mix. Centrifuge for 2 min. Measure the absorbance (2.2.25) of the supernatant solution of a 1 cm layer at 635 nm. Use the solution prepared from the blank solution as the compensation liquid. Draw a calibration curve and calculate the content of glycerol in the sample.

Loss on drying (2.2.32)

Maximum 2.0 per cent, determined on 1.000 g by drying over *diphosphorus pentoxide* R at a pressure not exceeding 0.7 kPa for 24 h.

ASSAY

Gold
Heat 0.2 g with 10 mL of *sulfuric acid R* and continue to boil gently until a clear, pale yellow liquid is produced. Cool, add about 1 mL of *nitric acid R* dropwise and boil for 1 h. Cool, dilute to 70 mL with *water R*, boil for 5 min and filter. Wash the residue of gold with *water R* at 60 °C. Dry and ignite at a temperature of at least 600 °C for 3 h. Weigh the residue and calculate the percentage content of Au.

Sodium
Evaporate to dryness the filtrate and washings obtained in the assay for gold, moisten with *sulfuric acid R* and ignite at 600 ± 50 °C for 3 h.

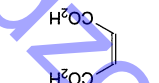
1.000 g of residue is equivalent to 0.324 g of Na.

STORAGE

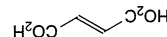
In an airtight container.

IMPURITIES

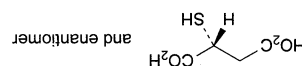
Specified impurities A, B, C.



A. (Z)-butenedioic acid (maleic acid),



B. (E)-butenedioic acid (fumaric acid),



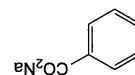
C. (2R,3S)-2-sulfanylbutanedioic acid (thiomalic acid).

Ph Eur

Sodium Benzoate



(Ph. Eur. monograph 0123)



C₇H₅NaO₂ 144.1 532-32-1 Ph Eur

DEFINITION

Sodium benzenecarboxylate.

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline or granular powder or flakes, slightly hygroscopic.

Solubility

Freely soluble in water, sparingly soluble in ethanol

IDENTIFICATION

A. It gives reactions (b) and (c) of benzoates (2.3.1).

B. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S
Dissolve 10.0 g in carbon dioxide-free water *R* and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 10 mL of carbon dioxide-free water *R* and 0.2 mL of phenolphthalein solution *R*. Not more than 0.2 mL of 0.1 M sodium hydroxide or 0.1 M hydrochloric acid is required to change the colour of the indicator.

Halogenated compounds

Maximum 200 ppm for ionised chlorine and maximum 300 ppm for total chlorine.

All glassware used must be chloride-free and may be prepared by soaking overnight in a 500 g/L solution of nitric acid *R*, rinsed with water *R* and stored full of water *R*. It is recommended that glassware be reserved exclusively for this test.

Test solution To 20.0 mL of solution S add 5 mL of water *R* and dilute to 50.0 mL with ethanol (96 per cent) *R*.

Determination of ionised chlorine

In three 25 mL volumetric flasks, prepare the following solutions.

Solution (a) To 4.0 mL of the test solution add 3 mL of dilute sodium hydroxide solution *R* and 3 mL of ethanol

(96 per cent) *R*. This solution is used to prepare solution A.

Solution (b) To 3 mL of dilute sodium hydroxide solution *R* add 2 mL of water *R* and 5 mL of ethanol (96 per cent) *R*. This

solution is used to prepare solution B.

Solution (c) To 4.0 mL of chloride standard solution

(8 ppm Cl) *R* add 6.0 mL of water *R*. This solution is used to prepare solution C.

In a fourth 25 mL volumetric flask, place 10 mL of water *R*.

To each flask add 5 mL of ferric ammonium sulfate

solution *R*5, mix and add dropwise and with swirling 2 mL of nitric acid *R* and 5 mL of mercuric thiocyanate solution *R*.

Shake. Dilute the contents of each flask to 25.0 mL with water *R* and allow the solutions to stand in a water-bath at

20 °C for 15 min. Measure at 460 nm in a 2 cm cell the absorbance (2.2.25) of solution A using solution B as the

compensation liquid, and the absorbance of solution C using the solution obtained with 10 mL of water *R* as the

compensation liquid. The absorbance of solution A is not greater than that of solution C.

Determination of total chlorine

Solution (a) To 10.0 mL of the test solution add 7.5 mL of dilute sodium hydroxide solution *R* and 0.125 g of nickel-

aluminium alloy *R* and heat on a water-bath for 10 min.

Allow to cool to room temperature, filter into a 25 mL volumetric flask and wash the filter with 3 quantities, each of

2 mL, of ethanol (96 per cent) *R* (a slight precipitate may form that disappears on acidification). Dilute the filtrate and

washings to 25.0 mL with water *R*. This solution is used to prepare solution A.

Solution (b) In the same manner, prepare a similar solution replacing the test solution by a mixture of 5 mL of ethanol

(96 per cent) *R* and 5 mL of water *R*. This solution is used to prepare solution B.

Solution (c) To 6.0 mL of chloride standard solution

(8 ppm Cl) *R* add 4.0 mL of water *R*. This solution is used to prepare solution C.

prepare solution C.

B. It gives the reaction of carbonates and bicarbonates (2.3.1).
C. Solution S gives reaction (a) of sodium (2.3.1).
TESTS
Solution S
Dissolve 5.0 g in 90 mL of carbon dioxide-free water R and dilute to 100.0 mL with the same solvent.
Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).
Carbonates
The pH (2.2.3) of freshly prepared solution S is not greater than 8.6.
Chlorides (2.4.4)
Maximum 150 ppm.
To 7 mL of solution S add 2 mL of nitric acid R and dilute to 15 mL with water R.
Sulfates (2.4.13)
Maximum 150 ppm.
To a suspension of 1.0 g in 10 mL of distilled water R add hydrochloric acid R until neutral and about 1 mL in excess. Dilute to 15 mL with distilled water R.
Ammonium (2.4.1)
Maximum 20 ppm.
Dilute 10 mL of solution S to 15 mL with water R. Prepare the standard using a mixture of 5 mL of water R and 10 mL of ammonium standard solution (1 ppm NH₄) R.
Arsenic (2.4.2, Method A)
Maximum 2 ppm, determined on 0.5 g.
Calcium (2.4.3)
Maximum 100 ppm.
To a suspension of 1.0 g in 10 mL of distilled water R add hydrochloric acid R until neutral and dilute to 15 mL with distilled water R.
Iron (2.4.9)
Maximum 20 ppm.
Dissolve 0.5 g in 5 mL of dilute hydrochloric acid R and dilute to 10 mL with water R.
Heavy metals (2.4.8)
Maximum 10 ppm.
Dissolve 2.0 g in a mixture of 2 mL of hydrochloric acid R and 18 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.
ASSAY
Dissolve 1.500 g in 50 mL of carbon dioxide-free water R. Titrate with 1 M hydrochloric acid, using 0.2 mL of methyl orange solution R as indicator.
1 mL of 1 M hydrochloric acid is equivalent to 84.0 mg of NaHCO₃.

Ph Eur

Sodium Bicarbonate

(Sodium Hydrogen Carbonate,
Ph Eur monograph 0195)

NaHCO₃

84.0

144-55-8

Ph Eur

In four 25 mL volumetric flasks, place separately 10 mL of solution (a), 10 mL of solution (b), 10 mL of solution (c) and 10 mL of water R. To each flask add 5 mL of ferric ammonium sulfate solution R₅, mix and add dropwise and with swirling 2 mL of nitric acid R and 5 mL of mercuric thiocyanate solution R. Shake. Dilute the contents of each flask to 25.0 mL with water R and allow the solutions to stand in a water-bath at 20 °C for 15 min. Measure at 460 nm in a 2 cm cell the absorbance (2.2.25) of solution A using solution B as the compensation liquid, and the absorbance of solution C using the solution obtained with 10 mL of water R as the compensation liquid. The absorbance of solution A is not greater than that of solution C.
Heavy metals (2.4.8)
Maximum 10 ppm.
2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.
Loss on drying (2.2.32)
Maximum 2.0 per cent, determined on 1.00 g by drying in an oven at 105 °C.
Dissolve 0.250 g in 20 mL of anhydrous acetic acid R, heating to 50 °C if necessary. Cool. Using 0.05 mL of naphtholbenzoin solution R as indicator, titrate with 0.1 M perchloric acid until a green colour is obtained.
1 mL of 0.1 M perchloric acid is equivalent to 14.41 mg of C₇H₅NaO₂.

Action and use
Antacid; used in treatment of electrolyte deficiency.

Preparations

Sodium Bicarbonate Ear Drops
Sodium Bicarbonate Eye Lotion
Sodium Bicarbonate Infusion
Sodium Bicarbonate Oral Solution
Compound Sodium Bicarbonate Tablets

DEFINITION

Content
99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, practically insoluble in ethanol

(96 per cent).

When heated in the dry state or in solution, it gradually

changes into sodium carbonate.

IDENTIFICATION

A. To 5 mL of solution S (see Tests) add 0.1 mL of phenolphthalein solution R. A pale pink colour is produced. Heat; gas is evolved and the solution becomes red.

Sodium Bromide

(Ph. Eur. monograph 0190)

NaBr

102.9

7647-15-6



DEFINITION

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, granular powder or small, colourless, transparent or opaque crystals, slightly hygroscopic.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. It gives reaction (a) of bromides (2.3.1).

B. Solution S (see Tests) gives the reactions of sodium

(2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Bromates

To 10 mL of solution S add 1 mL of starch solution R, 0.1 mL of a 100 g/L solution of potassium iodide R and 0.25 mL of 0.5 M sulfuric acid and allow to stand protected from light for 5 min. No blue or violet colour develops.

Chlorides and sulfates

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 0.400 g of the substance to be examined in 50 mL of water for chromatography R and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 25.0 mL of test solution (a) to 50.0 mL with water for chromatography R.

Reference solution (a) To 25.0 mL of test solution (a) add 1.0 mL of sulfate standard solution (10 ppm SO₄) R and 12.0 mL of chloride standard solution (50 ppm Cl) R and dilute to 50.0 mL with water for chromatography R.

Reference solution (b) Dilute 10.0 mL of test solution (a) to 100.0 mL with water for chromatography R. To 2.0 mL of this solution add 8.0 mL of chloride standard solution (50 ppm Cl) R and dilute to 20.0 mL with water for chromatography R.

Blank solution water for chromatography R.

Column:

— size: $l = 0.25$ m, $\varnothing = 2$ mm;— stationary phase: strongly basic anion-exchange resin for chromatography R (13 μ m).

Mobile phase Dissolve 0.600 g of potassium hydroxide R in water for chromatography R and dilute to 1000.0 mL with the same solvent.

Flow rate 0.4 mL/min.

Detection Conductivity detector equipped with a suitable ion

suppressor.

Injection 50 μ L of test solution (b), reference solutions (a)

and (b) and the blank solution.

Run time 2.5 times the retention time of bromide.

Retention time Chloride = about 5 min; bromide = about

8 min; sulfate = about 16 min.

System suitability: reference solution (b):

— resolution: minimum 8.0 between the peaks due to

chloride and bromide.

Calculation of percentage contents:

— for chlorides, use the concentration of chloride in reference solution (a); correct the area of the peak due to

chloride in the chromatogram obtained with reference

solution (a) by subtracting the area of the peak due to

chloride in the chromatogram obtained with test

solution (b);

— for sulfates, use the concentration of sulfate in reference

solution (a); correct the area of the peak due to sulfate in

the chromatogram obtained with reference solution (a) by

subtracting the area of the peak due to sulfate in the

chromatogram obtained with test solution (b).

Limits:

— chlorides: maximum 0.6 per cent;

— sulfates: maximum 0.01 per cent.

Iodides

To 5 mL of solution S add 0.15 mL of ferric chloride

solution R1 and 2 mL of methylene chloride R. Shake and allow

to separate. The lower layer is colourless (2.2.2, Method I).

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Magnesium and alkaline-earth metals (2.4.7)

Maximum 200 ppm, calculated as Ca.

10.0 g complies with the test for magnesium and alkaline-

earth metals. The volume of 0.01 M sodium edetate used does

not exceed 5.0 mL.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 3.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

ASSAY

Dissolve 85.0 mg in water R, add 5 mL of dilute nitric acid R

and dilute to 50 mL with water R. Titrate with 0.1 M silver

nitrate, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M silver nitrate is equivalent to 10.29 mg of

NaBr.

Calculate the percentage content of NaBr using the following

expression:

$$a - 2.902b$$

a = percentage content of NaBr and NaCl obtained in the

assay and calculated as NaBr;

b = percentage content of Cl obtained in the test for

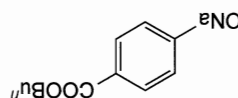
chlorides.

STORAGE

In an airtight container.

Sodium Butyl Hydroxybenzoate

Sodium Butylparaben



$C_{11}H_{13}NaO_3$ 216.2 36457-20-2

Action and use

Antimicrobial preservative.

DEFINITION

Sodium Butyl Hydroxybenzoate is the sodium salt of butyl 4-hydroxybenzoate. It contains not less than 99.0% and not more than 102.0% of $C_{11}H_{13}NaO_3$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white powder; hygroscopic.
Freely soluble in water and in ethanol (96%).

IDENTIFICATION

A. Dissolve 0.5 g in 5 mL of water and acidity to litmus paper with hydrochloric acid. A white precipitate is produced. Wash the precipitate with water and dry. The infrared absorption spectrum of the precipitate, Appendix II A, is concordant with the reference spectrum of butyl hydroxybenzoate (RS 036).

B. The residue on ignition yields the reactions characteristic of sodium salts, Appendix VI.

TESTS

Alkalinity

pH of a 0.1% w/v solution, 9.5 to 10.5, Appendix V L.

Clarity of solution

Dissolve 1.0 g in 10 mL of water. The solution is clear.

Chloride

Dissolve 1.0 g in 100 mL of water, add 1 mL of nitric acid and filter. 15 mL of the filtrate complies with the limit test for chlorides, Appendix VII (330 ppm).

Sulfate

Dissolve 0.50 g in 40 mL of water, add 3.5 mL of 2M hydrochloric acid, dilute to 60 mL with water and filter. 15 mL of the filtrate complies with the limit test for sulfates, Appendix VII (0.12%).

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions.

(1) Dilute a 2.0% w/v solution of the substance being examined in water with an equal volume of acetone.
(2) Dilute 1 volume of solution (1) to 25 volumes with a mixture of equal volumes of acetone and water.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel F₂₅₄ the surface of which has been modified with chemically-bonded octadecylsilyl groups (Whatman KCI8F plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 2 μ L of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air and examine under ultraviolet light (254 nm).

MOBILE PHASE
1 volume of glacial acetic acid, 30 volumes of water and 70 volumes of methanol.

LIMITS

Any secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2).

Water

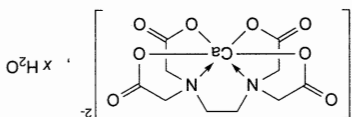
Not more than 5.0% w/w, Appendix IX C. Use 1 g.

ASSAY

Gently boil 0.1 g under a reflux condenser with 25 mL of 1.25M sodium hydroxide for 30 minutes. Allow to cool, add 25 mL of 0.0333M potassium bromate VS, 5 mL of a 12.5% w/v solution of potassium bromide and 10 mL of hydrochloric acid and immediately stopper the flask. Shake for 15 minutes and allow to stand for 15 minutes. Add 25 mL of the liberated iodine with 0.1M sodium thiosulfate VS using starch mucilage, added towards the end of the titration, as indicator. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of potassium bromate required. The volume of 0.0333M potassium bromate VS used is equivalent to half of the volume of 0.1M sodium thiosulfate VS required for the titration. Each mL of 0.0333M potassium bromate VS is equivalent to 7.207 mg of $C_{11}H_{13}NaO_3$.

Sodium Calcium Edetate

(Ph. Eur. monograph 0231)



$C_{10}H_{12}CaN_2Na_2O_8 \cdot 2H_2O$ 374.3

(anhydrous substance)

Action and use

Chelating agent.

Preparation

Sodium Calcium Edetate Infusion

Ph Eur

DEFINITION

Disodium [(ethylenedinitrilo)tetraacetato]calcate(2-).

Content

98.0 per cent to 102.0 per cent (anhydrous substance).
It contains a variable quantity of water of crystallisation.

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water, practically insoluble in ethanol

IDENTIFICATION

First identification: A, C, D.

Second identification: B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison sodium calcium edetate CRS.

B. Dissolve 2 g in 10 mL of water R, add 6 mL of lead nitrate solution R, shake and add 3 mL of potassium iodide solution R. No yellow precipitate is formed. Make alkaline to red litmus paper R by the addition of dilute ammonia R2 and add 3 mL of ammonium oxalate solution R. A white precipitate is formed.

C. Ignite. The residue gives reaction (b) of calcium (2.3.1). D. Dissolve 0.5 g in 10 mL of water R and add 10 mL of potassium pyroantimonate solution R. A white, crystalline precipitate is formed. The formation of the precipitate is accelerated by rubbing the wall of the tube with a glass rod.

TESTS

Solution S

Dissolve 5.0 g in water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

6.5 to 8.0.

Dissolve 5.0 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Impurity A

Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture Dissolve 10.0 g of ferric sulfate pentahydrate R in 20 mL of 0.5 M sulfuric acid and add 780 mL of water R.

Adjust to pH 2.0 with 1 M sodium hydroxide and dilute to 1000 mL with water R.

Test solution Dissolve 0.100 g of the substance to be examined in the solvent mixture and dilute to 25.0 mL with the solvent mixture.

Reference solution Dissolve 40.0 mg of nitrotriacetic acid R (impurity A) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. To 1.0 mL of this solution add 0.1 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Column:

— size: $l = 0.10$ m, $\varnothing = 4.6$ mm;

— stationary phase: spherical graphitised carbon for chromatography R1 (5 μ m) with a specific surface area of 120 m²/g and a pore size of 25 nm.

Mobile phase Dissolve 50.0 mg of ferric sulfate pentahydrate R in 50 mL of 0.5 M sulfuric acid and add 750 mL of water R, adjust to pH 1.5 with 0.5 M sulfuric acid or 1 M sodium hydroxide, add 20 mL of ethylene glycol R and dilute to 1000 mL with water R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 273 nm.

Injection 20 μ L; filter the solutions and inject immediately.

Run time 4 times the retention time of the iron complex of impurity A.

Retention time Iron complex of impurity A = about 5 min; iron complex of edetic acid = about 10 min.

System suitability: reference solution:

— resolution: minimum 7 between the peaks due to the iron complex of impurity A and the iron complex of edetic acid;

— signal-to-noise ratio: minimum 50 for the peak due to impurity A.

Limit:

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (0.1 per cent).

Disodium edetate

Maximum 1.0 per cent.

Dissolve 5.0 g in 250 mL of water R. Add 10 mL of ammonium chloride buffer solution pH 10.0 R and about 50 mg of mordant black 11 triturate R. Not more than 1.5 mL of 0.1 M magnesium chloride is required to change the colour of the indicator to violet.

Chlorides

Maximum 0.1 per cent.

Dissolve 0.7 g in water R and dilute to 20 mL with the same solvent. Add 30 mL of dilute nitric acid R, allow to stand for 30 min and filter. Dilute 10 mL of the filtrate to 50 mL with water R. Use this solution as the test solution. Prepare the reference solution using 0.40 mL of 0.01 M hydrochloric acid, add 6 mL of dilute nitric acid R and dilute to 50 mL with water R. Filter both solutions if necessary. Add 1 mL of silver nitrate solution R2 to the test solution and to the reference solution and mix. After standing for 5 min protected from light, any opalescence in the test solution is not more intense than that in the reference solution.

Iron (2.4.9)

Maximum 80 ppm.

Dilute 2.5 mL of solution S to 10 mL with water R. Add 0.25 g of calcium chloride R to the test solution and the standard before the addition of the thioglycolic acid R.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

5.0 per cent to 13.0 per cent, determined on 0.200 g.

ASSAY

Dissolve 0.500 g in water R and dilute to 200 mL with the same solvent. To 20.0 mL of this solution, add 80 mL of water R and adjust to pH 2 with dilute nitric acid R. Titrate with 0.01 M bismuth nitrate, using 0.1 mL of a 1 g/L solution of xylenol orange R as indicator. The colour of the solution changes from yellow to red.

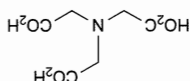
1 mL of 0.01 M bismuth nitrate is equivalent to 3.74 mg of C₁₀H₁₂CaN₂Na₂O₈.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A

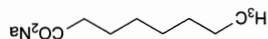


A. nitrotriacetic acid.

Sodium Caprylate

Sodium Octanoate

(Ph. Eur. monograph 1471)

 $\text{C}_8\text{H}_{15}\text{NaO}_2$

166.2

1984-06-1

Action and use

Excipient.

Ph Eur

DEFINITION

Sodium octanoate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble or freely soluble in water, freely soluble in acetic acid, sparingly soluble in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for

related substances.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

B. To 0.2 mL of solution S (see Tests) add 0.3 mL of water R. The solution gives reaction (b) of sodium (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

8.0 to 10.5 for solution S.

Related substances

Gas chromatography (2.2.28): use the normalisation

procedure.

Test solution Dissolve 0.116 g in water R and dilute to 5 mL with the same solvent. Add 1 mL of a 2.8 per cent V/V solution of sulfuric acid R and shake with 10 mL of ethyl acetate R. Separate the organic layer and dry over anhydrous sodium sulfate R.

Reference solution (a) Dissolve 0.10 g of caprylic acid CRS in ethyl acetate R and dilute to 10 mL with the same solvent.

Reference solution (b) Dilute 1 mL of the test solution to 100 mL with ethyl acetate R. Dilute 5 mL of this solution to 50 mL with ethyl acetate R.

Column:

material: fused silica;

size: $l = 30$ m, $\varnothing = 0.25$ mm;

stationary phase: macrogol 20 000 2-nitroterephthalate R (film thickness 0.25 μm).

Carrier gas helium for chromatography R.

Flow rate 1.5 mL/min.

Split ratio 1:100.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 1	100			
1 - 25	100 → 220			
25 - 35				
250				
250				

Detection Flame ionisation.

Injection 1 μL .

System suitability: reference solution (b):

— signal-to-noise ratio: minimum 5 for the principal peak.

Limits:

— any impurity: for each impurity, maximum 0.3 per cent;

— total: maximum 0.5 per cent;

— the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in glacial acetic acid R and dilute to 10 mL with the same acid. Add 10 mL of ethanol (96 per cent) R.

12 mL of the solution complies with test B. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R and 9 mL of a mixture of equal volumes of glacial acetic acid R and ethanol (96 per cent) R.

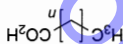
Water (2.5.12)

Maximum 3.0 per cent, determined on 1.000 g.

ASSAY

Dissolve 0.150 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 16.62 mg of $\text{C}_8\text{H}_{15}\text{NaO}_2$.

IMPURITIES

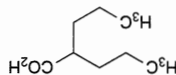


A. $n = 4$: hexanoic acid,

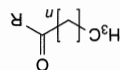
B. $n = 5$: heptanoic acid,

C. $n = 7$: nonanoic acid,

D. $n = 8$: decanoic acid,



E. 2-propylpentanoic acid (valproic acid),

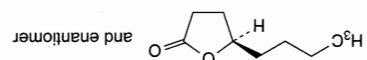


F. $R = \text{OCH}_3$, $n = 6$: methyl octanoate,

G. $R = \text{OC}_2\text{H}_5$, $n = 6$: ethyl octanoate,

H. $R = \text{OCH}_3$, $n = 8$: methyl decanoate,

I. $R = \text{CH}_3$, $n = 8$: undecan-2-one,



J. (R,S)-5-butyltetrahydrofuran-2-one (γ -hydroxyoctanoic acid lactone).

Ph Eur

Anhydrous Sodium Carbonate



(Ph. Eur. monograph 0773)

Na₂CO₃ 106.0 497-19-8

Ph Eur

DEFINITION

Content 99.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance White or almost white, slightly granular powder, hygroscopic.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 1 g in water R and dilute to 10 mL with the same solvent. The solution is strongly alkaline (2.2.4).

B. The solution prepared for identification test A gives the reaction of carbonates (2.3.1).

C. The solution prepared for identification test A gives reaction (a) of sodium (2.3.1).

D. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 2.0 g in portions in a mixture of 5 mL of hydrochloric acid R and 25 mL of distilled water R. Heat to boiling and cool. Add dilute sodium hydroxide solution R until the solution is neutral and dilute to 50 mL with distilled water R.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method I).

Dissolve 2.0 g in 10 mL of water R.

Alkali hydroxides and bicarbonates

Dissolve 0.4 g in 20 mL of water R. Add 20 mL of barium chloride solution R1 and filter. To 10 mL of the filtrate add 0.1 mL of phenolphthalein solution R. The solution does not become red. Boil the remainder of the filtrate for 2 min. The solution remains clear (2.2.1).

Chlorides (2.4.4)

Dissolve 0.4 g in water R, add 4 mL of dilute nitric acid R and dilute to 15 mL with water R.

Sulfates (2.4.13)

Maximum 250 ppm, determined on solution S.

Arsenic (2.4.2, Method A)

Maximum 5 ppm, determined on 5 mL of solution S.

Iron (2.4.9)

Maximum 50 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 50 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying at 300 ± 15 °C.

ASSAY

Dissolve 1.000 g in 25 mL of water R. Add 0.2 mL of methyl orange solution R as indicator. Titrate with 1 M hydrochloric acid until the colour changes from yellow to red. 1 mL of 1 M hydrochloric acid is equivalent to 52.99 mg of Na₂CO₃.

STORAGE

In an airtight container.

Ph Eur

Sodium Carbonate Decahydrate



(Ph. Eur. monograph 0191)

Na₂CO₃·10H₂O

286.1

6132-02-1

Ph Eur

DEFINITION

Content

36.7 per cent to 40.0 per cent of Na₂CO₃.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless, transparent crystals, efflorescent.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 1 g in water R and dilute to 10 mL with the same solvent. The solution is strongly alkaline (2.2.4).

B. The solution prepared for identification test A gives the reaction of carbonates (2.3.1).

C. The solution prepared for identification test A gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in portions in a mixture of 5 mL of hydrochloric acid R and 25 mL of distilled water R. Heat the solution to boiling and cool. Add dilute sodium hydroxide solution R until the solution is neutral and dilute to 50 mL with distilled water R.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method I).

Dissolve 4.0 g in 10 mL of water R.

Alkali hydroxides and bicarbonates

Dissolve 1.0 g in 20 mL of water R, add 20 mL of barium chloride solution R1 and filter. To 10 mL of the filtrate add 0.1 mL of phenolphthalein solution R. The solution does not become red. Heat the remainder of the filtrate to boiling for 2 min. The solution remains clear (2.2.1).

Chlorides (2.4.4)

Maximum 50 ppm.

Dissolve 1.0 g in water R, add 4 mL of dilute nitric acid R and dilute to 15 mL with water R.

Sulfates (2.4.13)

Maximum 100 ppm, determined on solution S.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 5 mL of solution S.

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

ASSAY

Dissolve 2.000 g in 25 mL of water R. Titrate with 1 M

hydrochloric acid, using 0.2 mL of methyl orange solution R as

indicator.

1 mL of 1 M hydrochloric acid is equivalent to 52.99 mg

of Na₂CO₃.

STORAGE

In an airtight container.

Ph Eur

Sodium Carbonate Monohydrate

(Ph. Eur. monograph 0192)

Na₂CO₃·H₂O

124.0

5968-11-6

DEFINITION

Content

83.0 per cent to 87.5 per cent of Na₂CO₃.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals.

Solubility

Freely soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. Dissolve 1 g in water R and dilute to 10 mL with the

same solvent. The solution is strongly alkaline (2.2.4).

B. The solution prepared for identification test A gives the

reaction of carbonates (2.3.1).

C. The solution prepared for identification test A gives

reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 2.0 g in portions in a mixture of 5 mL of

hydrochloric acid R and 25 mL of distilled water R. Heat the

solution to boiling and cool. Add dilute sodium hydroxide

with distilled water R.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured

than reference solution Y₆ (2.2.2, Method I).

Dissolve 2.0 g in 10 mL of water R.

Sodium Cetostearyl Sulfate

Sodium Cetostearyl Sulphate

(Ph. Eur. monograph 0847)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of sodium cetyl sulfate (C₁₆H₃₃NaO₄S; 344.5) and

sodium stearyl sulfate (C₁₈H₃₇NaO₄S; 372.5). A suitable

buffer may be added.

Content

— sodium cetostearyl sulfate: minimum 90.0 per cent

(anhydrous substance);

— sodium cetyl sulfate: minimum 40.0 per cent (anhydrous

substance).

CHARACTERS

Appearance

White or pale yellow, amorphous or crystalline powder.

Solubility

Soluble in hot water giving an opalescent solution, practically

insoluble in cold water, partly soluble in ethanol

IDENTIFICATION

First identification B, D, F

Second identification A, C, D, E, F



1 mL of 0.01 M lead nitrate is equivalent to 1.420 mg

of Na_2SO_4 .

Free cetostearyl alcohol

Maximum 4.0 per cent.

From the chromatogram obtained with test solution (a) in

the assay, calculate the percentage content of free cetostearyl

alcohol in the substance to be examined using the following

expression and taking into account the assigned content of

the chemical reference substances:

$$\frac{A_1}{A_2} \times \frac{m_1}{m_2} \times \frac{100}{2} \times 100$$

A_1 = sum of the areas of the peaks due to cetyl alcohol

and stearyl alcohol in the chromatogram obtained

with test solution (a);

A_2 = area of the peak due to the internal standard in the

chromatogram obtained with test solution (a);

m_1 = mass of the substance to be examined in test

solution (a), in milligrams;

m_2 = mass of the internal standard in the internal standard

solution, in milligrams.

Water (2.5.12)

Maximum 1.5 per cent, determined on 5.00 g.

ASSAY

Gas chromatography (2.2.28).

Internal standard solution Dissolve 0.200 g of 1-

nonadecanol CRS in anhydrous ethanol R and dilute to

100.0 mL with the same solvent.

Test solution (a) Dissolve 0.300 g of the substance to be

examined in 50.0 mL of anhydrous ethanol R and add 2.0 mL

of the internal standard solution and 48.0 mL of water R.

Shake with 4 quantities, each of 25 mL, of pentane R, adding

sodium chloride R, if necessary, to facilitate the separation of

the layers. Combine the upper layer. Reserve the lower layer

for the preparation of test solution (b). Wash the combined

upper layers with 2 quantities, each of 30 mL, of water R,

dry over anhydrous sodium sulfate R and filter.

Test solution (b) Transfer 25.0 mL of the lower layer obtained

in the preparation of test solution (a) to a 200 mL flask that

can be fitted with a reflux condenser. Add 10.0 mL of the

internal standard solution and 20 mL of hydrochloric acid R.

Boil under a reflux condenser for 2 h. Allow to cool and

shake with 4 quantities, each of 20 mL, of pentane R.

Combine the upper layers, wash with 2 quantities, each of

20 mL, of water R, dry over anhydrous sodium sulfate R and

filter.

Reference solution (a) Dissolve 0.100 g of cetyl alcohol CRS in

25.0 mL of the internal standard solution. Add 25 mL of

water R and shake with 4 quantities, each of 25 mL, of

pentane R, adding sodium chloride R, if necessary, to facilitate

the separation of the layers. Combine the upper layers, wash

with 2 quantities, each of 30 mL, of water R, dry over

anhydrous sodium sulfate R and filter.

Column:

— material: fused silica;

— size: $l = 25$ m, $\varnothing = 0.25$ mm;

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined

in 10 mL of ethanol (70 per cent V/V) R, heating on a water-

bath.

Reference solution Dissolve 50 mg of sodium cetostearyl

sulfate CRS in 10 mL of ethanol (70 per cent V/V) R, heating

on a water-bath.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase water R, acetone R, methanol R

(20:40:40 V/V/V).

Application 10 μL .

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 50 g/L solution of phosphomolybdic

acid R in ethanol (96 per cent) R; heat at 120 °C until spots

appear (about 5 min).

Results The principal spots in the chromatogram obtained

with the test solution are similar in position and colour to the

principal spots in the chromatogram obtained with the

reference solution.

B. Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained

with test solution (b) are similar in retention time to the

2 principal peaks in the chromatograms obtained with

reference solutions (a) and (b).

C. Dissolve 0.1 g in 10 mL of water R and shake. A foam is

formed.

D. It gives a yellow colour to a non-luminous flame.

E. To 0.1 mL of the solution prepared for identification

test C add 0.1 mL of a 1 g/L solution of methylene blue R and

2 mL of dilute sulfuric acid R. Add 2 mL of methylene

chloride R and shake. The methylene chloride layer has an

intense blue colour.

F. Mix about 10 mg with 10 mL of anhydrous ethanol R.

Heat to boiling on a water-bath, shaking frequently. Filter

immediately and evaporate to dryness. Dissolve the residue in

7 mL of water R, add 3 mL of dilute hydrochloric acid R and

evaporate the solution to half its volume. Allow to cool.

Filter. To the filtrate add 1 mL of barium chloride solution R1.

A white, crystalline precipitate is formed.

TESTS

Acidity or alkalinity

Dissolve 0.5 g with heating in a mixture of 10 mL of water R

and 15 mL of ethanol (90 per cent V/V) R. Add 0.1 mL of

phenolphthalein solution R1. The solution is colourless.

Add 0.1 mL of 0.1 M sodium hydroxide. The solution is red.

Sodium chloride and sodium sulfate

Maximum 8.0 per cent for the sum of the percentage

contents.

Sodium chloride Dissolve 5.00 g in 50 mL of water R, add

dilute nitric acid R dropwise until the solution is neutral to

blue litmus paper R. Titrate with 0.1 M silver nitrate

determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of

NaCl.

Sodium sulfate Dissolve 0.500 g in 20 mL of water R,

warming gently if necessary, and add 1 mL of a 0.5 g/L

solution of diethylenetriamine R. If the solution is red, add

1 M nitric acid dropwise until a bluish-green colour is

obtained. Add 2.0 mL of dichloroacetic acid solution R and

80 mL of acetone R. Titrate with 0.01 M lead nitrate until a

persistent orange-red colour is obtained.

— stationary phase: poly(dimethyl)siloxane R (film thickness 0.25 µm).
Carrier gas helium for chromatography R.
Flow rate 1 mL/min.
Split ratio 1:100.
Temperature:

Time (min)	Temperature (°C)
Column 0 - 20	150 → 250
Injection port 250	
Detector 250	

Detection Flame ionisation.

Injection 1 µL.

Elution order Cetyl alcohol, stearyl alcohol, 1-nonadecanol.

Calculate the percentage content of sodium cetyl sulfate in the substance to be examined using the following expression and taking into account the assigned content of cetyl alcohol CRS:

$$A_x \times \frac{A_1}{A_2} \times \frac{A_{xy}}{m_{xy}} \times \frac{m}{1} \times 100 \times 4 \times \frac{2.5}{1} \times F_x$$

A_x = area of the peak due to cetyl alcohol in the chromatogram obtained with test solution (b);
 A_{xy} = area of the peak due to cetyl alcohol CRS in the chromatogram obtained with reference solution (a);
 A_1 = area of the peak due to the internal standard in the chromatogram obtained with test solution (b);
 A_2 = area of the peak due to the internal standard in the chromatogram obtained with reference solution (a);
 F_x = conversion factor from cetyl alcohol to sodium cetyl sulfate (1.421);
 m = mass of the substance to be examined in test solution (a), in milligrams;
 m_{xy} = mass of cetyl alcohol CRS in reference solution (a), in milligrams.

Calculate the percentage content of sodium stearyl sulfate in the substance to be examined using the following expression and taking into account the assigned content of stearyl alcohol CRS:

$$A_z \times \frac{A_1}{A_3} \times \frac{A_{zy}}{m_{zy}} \times \frac{m}{1} \times 100 \times 4 \times \frac{2.5}{1} \times F_z$$

A_z = area of the peak due to stearyl alcohol in the chromatogram obtained with test solution (b);
 A_{zy} = area of the peak due to stearyl alcohol CRS in the chromatogram obtained with reference standard in the test solution (b);
 A_1 = area of the peak due to the internal standard in the chromatogram obtained with test solution (b);
 A_3 = area of the peak due to the internal standard in the chromatogram obtained with reference solution (b);
 F_z = conversion factor from stearyl alcohol to sodium stearyl sulfate (1.377);
 m = mass of the substance to be examined in test solution (a), in milligrams;
 m_{zy} = mass of stearyl alcohol CRS in reference solution (b), in milligrams.

The percentage content of sodium cetostearyl sulfate corresponds to the sum of the percentage contents of sodium cetyl sulfate and sodium stearyl sulfate.

LABELLING

The label states, where appropriate, the name and concentration of any added buffer.

Ph Eur



Sodium Chloride¹

(Ph. Eur. monograph 0193)

NaCl 58.44

7647-14-5

Action and use

Used in treatment of electrolyte deficiency.

Preparations

Compound Glucose, Sodium Chloride and Sodium Citrate Oral Solution
Oral Rehydration Salts
Potassium Chloride and Sodium Chloride Infusion
Potassium Chloride, Sodium Chloride and Glucose Infusion
Sodium Chloride Eye Drops
Sodium Chloride Eye Lotion
Sodium Chloride Eye Ointment
Sodium Chloride Infusion
Sodium Chloride and Glucose Infusion
Sodium Chloride Irrigation Solution
Compound Sodium Chloride Mouthwash
Sodium Chloride Nebuliser Solution
Sodium Chloride Oral Solution
Sodium Chloride Solution
Soluble Sodium Chloride Tablets

DEFINITION

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals or white or almost white pearls.

Solubility

Freely soluble in water, practically insoluble in anhydrous ethanol.◆

IDENTIFICATION

A. It gives reaction (a) of chlorides (2.3.1).

B. It gives the reactions of sodium (2.3.1).

TESTS

◆ If the substance is in the form of pearls, crush before use.◆

Solution S

Dissolve 20.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent.

◆ Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method ID).◆

Acidity or alkalinity

To 20 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Bromides

Maximum 100 ppm.

To 0.5 mL of solution S add 4.0 mL of water R, 2.0 mL of phenol red solution R2 and 1.0 mL of 0.1 g/L solution of chloramine R and mix immediately. After exactly 2 min, add 0.15 mL of 0.1 M sodium thiosulfate, mix and dilute to 10.0 mL with water R. The absorbance (2.2.25) of the solution measured at 590 nm, using water R as the compensation liquid, is not greater than that of a standard prepared at the same time and in the same manner, using 5.0 mL of a 3.0 mg/L solution of potassium bromide R.

Ferrocyanides

Dissolve 2.0 g in 6 mL of water R. Add 0.5 mL of a mixture of 5 mL of a 10 g/L solution of ferric ammonium sulfate R in a 2.5 g/L solution of sulfuric acid R and 95 mL of a 10 g/L solution of ferrous sulfate R. No blue colour develops within 10 min.

Iodides

Moisten 5 g by the dropwise addition of a freshly prepared mixture of 0.15 mL of sodium nitrite solution R, 2 mL of 0.5 M sulfuric acid, 25 mL of iodide-free starch solution R and 25 mL of water R. After 5 min, examine in daylight. The mixture shows no blue colour.

Nitrites

To 10 mL of solution S add 10 mL of water R. The absorbance (2.2.25) is not greater than 0.01 at 354 nm.

Phosphates (2.4.11)

Maximum 25 ppm.
Dilute 2 mL of solution S to 100 mL with water R.

Sulfates (2.4.13)

Maximum 200 ppm.
Dilute 7.5 mL of solution S to 30 mL with distilled water R.

Aluminium (2.4.17)

Maximum 0.2 ppm, if intended for use in the manufacture of peritoneal dialysis solutions, haemodialysis solutions or haemofiltration solutions.

Prescribed solution Dissolve 20.0 g in 100 mL of water R and add 10 mL of acetate buffer solution pH 6.0 R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of water R.

Arsenic (2.4.2, Method A)

Maximum 1 ppm, determined on 5 mL of solution S. ♦

Barium

To 5 mL of solution S add 5 mL of distilled water R and 2 mL of dilute sulfuric acid R. After 2 h, any opalescence in the solution is not more intense than that in a mixture of 5 mL of solution S and 7 mL of distilled water R.

Iron (2.4.9)

Maximum 2 ppm, determined on solution S.
Prepare the standard using a mixture of 4 mL of iron standard solution (1 ppm Fe) R and 6 mL of water R.

Maximum 100 ppm, calculated as Ca and determined on 10.0 g.

Use 0.150 g of mordant black 11 mixture R. The volume of 0.01 M sodium edetate used is not more than 2.5 mL.

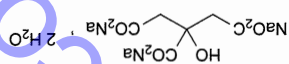
Potassium

Maximum 500 ppm, if intended for use in the manufacture of parenteral preparations or haemodialysis, haemofiltration or peritoneal dialysis solutions.

Sodium Citrate

Trisodium Citrate

(Ph. Eur. monograph 0412)



$C_6H_5Na_3O_7 \cdot 2H_2O$ 294.1

6132-04-3

Action and use

Systemic alkalinising substance.

Preparations

Compound Glucose, Sodium Chloride and Sodium Citrate Oral Solution
Sodium Citrate Eye Drops
Sodium Citrate Irrigation Solution

DEFINITION

Trisodium 2-hydroxypropane-1,2,3-tricarboxylate dihydrate. Content 99.0 per cent to 101.0 per cent (anhydrous substance).

LABELLING

The label states:
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations;
— where applicable, that the substance is suitable for use in haemodialysis solutions or haemofiltration solutions. ♦
1 This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.

ASSAY

Dissolve 50.0 mg in water R and dilute to 50 mL with the same solvent. Titrate with 0.1 M silver nitrate determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of NaCl.

Bacterial endotoxins (2.6.14)

Less than 5 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins. ♦
an oven at 105 °C for 2 h.
Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

Maximum 5 ppm.
12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R. ♦

Heavy metals (2.4.8)

Wave length 766.5 nm.
millilitre). Dilute as required.
Previously dried at 100–105 °C for 3 h, in water R and dilute to 1000.0 mL with the same solvent (600 µg of K per millilitre). Dissolve 1.144 g of potassium chloride R, 100.0 mL with the same solvent.
Test solution Dissolve 1.00 g in water R and dilute to Atomic emission spectrometry (2.2.22, Method I).



CHARACTERS**Appearance**

White or almost white, crystalline powder or white or almost white, granular crystals, slightly deliquescent in moist air.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. To 1 mL of solution S (see Tests) add 4 mL of water R. The solution gives the reaction of citrates (2.3.1).

B. 1 mL of solution S gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of phenolphthalein solution R. Not more than 0.2 mL of 0.1 M hydrochloric acid or 0.1 M sodium hydroxide is required to change the colour of the indicator.

Readily carbonisable substances

To 0.20 g of the powdered substance to be examined add 10 mL of sulfuric acid R and heat in a water-bath at

90 ± 1 °C for 60 min. Cool rapidly. The solution is not more intensely coloured than reference solution Y₂ or GY₂ (2.2.2, Method II).

Chlorides (2.4.4)

Maximum 50 ppm.

Dilute 10 mL of solution S to 15 mL with water R.

Oxalates

Maximum 300 ppm.

Dissolve 0.50 g in 4 mL of water R, add 3 mL of hydrochloric acid R and 1 g of zinc R in granules and heat on a water-bath for 1 min. Allow to stand for 2 min, decant the liquid into a

test-tube containing 0.25 mL of a 10 g/L solution of phenylhydrazine hydrochloride R and heat to boiling. Cool

rapidly, transfer to a graduated cylinder and add an equal volume of hydrochloric acid R and 0.25 mL of potassium

ferrioxalate solution R. Shake and allow to stand for 30 min. Any pink colour in the solution is not more intense than that in a standard prepared at the same time in the same manner

using 4 mL of a 50 mg/L solution of oxalic acid R.

Sulfates (2.4.13)

Maximum 150 ppm.

To 10 mL of solution S add 2 mL of hydrochloric acid R1 and dilute to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Use as the solvent a mixture of 20 mL of methanol R, 30 mL of formamide R and 5 g of salicylic acid R.

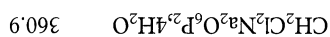
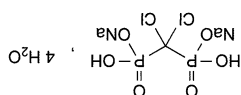
Pyrogens (2.6.8)

If intended for use in the manufacture of large-volume parenteral preparations, the competent authority may require that it comply with the test for pyrogens. Inject per kilogram

of the rabbit's mass 10 mL of a freshly prepared solution in

Sodium Clodronate Tetrahydrate

(Clodronate Disodium Tetrahydrate,
Ph Eur monograph 1777)



Ph Eur

DEFINITION

Disodium (dichloromethylene)bis(hydrogen phosphonate) tetrahydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent), slightly soluble in methanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Comparison clodronate disodium tetrahydrate CRS.

B. Dissolve 0.5 g in 10 mL of water R. The solution gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

3.0 to 4.5, for solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.125 g of the substance to be examined in 30 mL of water R, sonicate for 10 min and

dilute to 50.0 mL with water R (test stock solution). Dilute 10.0 mL of the test stock solution to 20.0 mL of water R. Dilute 1.0 mL of the test solution to

50.0 mL with water R.



Ph Eur

In an airtight container.

STORAGE

of C₆H₅Na₃O₇.

1 mL of 0.1 M perchloric acid is equivalent to 8.602 mg

until a green colour is obtained.

acid, using 0.25 mL of naphtholbenzenesulfonate solution R as indicator

to about 50 °C. Allow to cool. Titrate with 0.1 M perchloric

Dissolve 0.150 g in 20 mL of anhydrous acetic acid R, heating

ASSAY

chloride R.

substance to be examined and 7.5 mg of pyrogen-free calcium

water for injections R containing per millilitre 10.0 mg of the

Reference solution (b) Dissolve 1 mg of clodronate impurity D CRS in 10 mL of water R₃, sonicate for 10 min and dilute to 20.0 mL with water R. Mix 2.0 mL of this solution with 10.0 mL of the test stock solution and dilute to 20.0 mL with water R.

Reference solution (c) Dilute 1.0 mL of a 0.3 g/L solution of phosphoric acid R (impurity B) to 100.0 mL with water R.

Precolumn:
— size: 1 = 0.05 m, Ø = 4 mm;
— stationary phase: anion-exchange resin R₃
— particle size: 9 µm.

Column:
— size: 1 = 0.25 m, Ø = 4 mm;
— stationary phase: anion-exchange resin R₃
— particle size: 9 µm.

Mobile phase:
— mobile phase A: 0.21 g/L solution of sodium hydroxide R in carbon dioxide-free water R₃; close immediately, mix and use under helium pressure;
— mobile phase B: 4.2 g/L solution of sodium hydroxide R in carbon dioxide-free water R₃; close immediately, mix and use under helium pressure;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	90 → 60	10 → 40
10 - 22	60 → 50	40 → 50
22 - 23	50 → 20	50 → 80
23 - 25	20	80

Flow rate 1 mL/min.
Detection Conductivity detector. Use a self-regenerating anion suppressor.

Injection 20 µL.
Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B.

Relative retention With reference to clodronate (retention time = about 13 min): impurities A and B = about 0.7; impurity D = about 1.1.

System suitability: reference solution (b):
— peak-to-valley ratio: minimum 3, where H_p = height above the baseline of the peak due to impurity D and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to clodronate.

Limits:
— sum of impurities A and B: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— total: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)
Maximum 20 ppm.

0.5 g complies with test G. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)
18.5 per cent to 21.0 per cent, determined on 0.100 g.

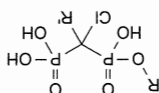
ASSAY
Dissolve 0.140 g in 10 mL of water R. Add 10 mL of strong sodium hydroxide solution R and some glass beads. Boil until the solution is completely decolourised (about 10 min). Cool in an ice-bath and add 30 mL of water R and 10 mL of nitric acid R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M silver nitrate is equivalent to 14.44 mg of $\text{CH}_2\text{Cl}_2\text{N}_2\text{O}_6\text{P}_2$.

IMPURITIES

Specified impurities A, B

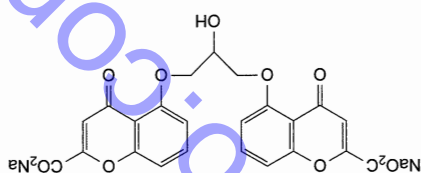
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D.



A. R = $\text{CH}(\text{CH}_3)_2$, R' = Cl: [dichloro]hydroxy (1-methylethoxy)phosphinoyl[methyl]phosphonic acid, D. R = R' = H: (chloromethylene)bis(phosphonic acid), B. H_3PO_4 : phosphoric acid.

Sodium Cromoglicate

(Ph. Eur. monograph 0562)



$\text{C}_{23}\text{H}_{14}\text{Na}_2\text{O}_{11}$ 512.3 15826-37-6

Action and use

Trough axis of allergic conditions.

Preparations

Sodium Cromoglicate Eye Drops
Sodium Cromoglicate Inhalation Powder, hard capsule

DEFINITION

Disodium 5,5'-[(2-hydroxypropyl)bis(4-oxo-4H-1-benzopyran-2-carboxylate)].

Content

98.0 per cent to 101.0 per cent (dried substance).



CHARACTERS**Appearance**

White or almost white, hygroscopic, crystalline powder.

Solubility

Soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 10.0 mg in phosphate buffer solution

pH 7.4 R and dilute to 100.0 mL with the same buffer

solution. Dilute 10.0 mL of this solution to 100.0 mL with

phosphate buffer solution pH 7.4 R.

Spectral range 230-350 nm.

Absorption maxima At 239 nm and 327 nm.

Absorbance ratio $A_{327}/A_{239} = 0.25$ to 0.30.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium cromoglicate CRS.

C. Dissolve about 5 mg in 0.5 mL of methanol R. Add 3 mL

of 5 g/L solution of aminopyrazolone R in methanol R

containing 1 per cent V/V of hydrochloric acid R. Allow to

stand for 5 min. An intense yellow colour develops.

D. It gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 0.5 g in carbon dioxide-free water R and dilute to

25 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference

suspension II (2.2.1) and not more intensely coloured than

reference solution BY₂ (2.2.2, Method ID).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of phenolphthalein

solution R. The solution is colourless. Add 0.2 mL of 0.01 M

sodium hydroxide. The solution is pink. Add 0.4 mL of

0.01 M hydrochloric acid. The solution is colourless.

Add 0.25 mL of methyl red solution R. The solution is red.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture water R, acetonitrile R (40:60 V/V).

Test solution Dissolve 0.100 g of the substance to be

examined in the solvent mixture and dilute to 50.0 mL with

the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 7 mg of sodium cromoglicate for

system suitability CRS (containing impurity C) in the solvent

mixture and dilute to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: base-deactivated end-capped octadecylsilyl

silica gel for chromatography R (3 μ m).

Mobile phase:

— mobile phase A: acetonitrile R, 10 g/L solution of

tetrabutylammonium hydrogen sulfate R (5:95 V/V);

— mobile phase B: acetonitrile R, 10 g/L solution of

tetrabutylammonium hydrogen sulfate R (50:50 V/V);

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 15	100 \rightarrow 0	0 \rightarrow 100
15 - 20	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 330 nm.

Injection 10 μ L.

Relative retention With reference to sodium cromoglicate

(retention time = about 11 min); impurity C = about 1.1.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to sodium

cromoglicate and impurity C.

Limits:

— impurity C: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

Oxalates

Maximum 0.35 per cent.

Dissolve 0.10 g in 20 mL of water R, add 5.0 mL of iron

salicylate solution R and dilute to 50.0 mL with water R.

Measure the absorbance (2.2.25) at 480 nm. The absorbance

is not less than that of a reference solution prepared in the

same manner using 0.35 mg of oxalic acid R instead of the

substance to be examined.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 10.0 per cent, determined on 1.000 g by drying at

105 °C and at a pressure of 0.3-0.6 kPa.

ASSAY

Dissolve 0.200 g with heating in a mixture of 5 mL of 2-

propanol R and 25 mL of ethylene glycol R. Cool and add a

mixture of 6 mL of tetrahydrofuran R and 24 mL of

acetonitrile R. Titrate with 0.1 M perchloric acid, determining

the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 25.62 mg

of $C_{23}H_{14}Na_2O_{11}$.

STORAGE

In an airtight container, protected from light.

IMPURITIES**Specified impurities C**

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

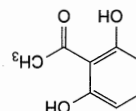
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

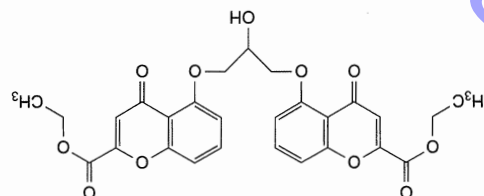
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use): A, B.



A. 1-(2,6-dihydroxyphenyl)ethanone,



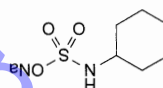
B. diethyl 5,5'-[(2-hydroxypropane-1,3-diyl)dioxy]bis(4-oxo-4H-1-benzopyran-2-carboxylate),

C. unknown structure.

Ph Eur

Sodium Cyclamate

(Ph. Eur. monograph 0774)



$C_6H_{12}NNaO_3S$ 201.2 139-05-9

Action and use
Sweetening agent.

Ph Eur

DEFINITION

Sodium N-cyclohexylsulfamate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless crystals.

Solubility
Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, E
Second identification B, C, D, E
A. Infrared absorption spectrophotometry (2.2.24).
Comparison sodium cyclamate CRS.

B. Examine the chromatograms obtained in the test for impurity A.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 1 mL of solution S (see Tests), add 1 mL of water R and 2 mL of silver nitrate solution R1, then shake. A white, crystalline precipitate is formed.
D. To 1 mL of solution S add 5 mL of water R, 2 mL of dilute hydrochloric acid R and 4 mL of barium chloride

TESTS

Solution S
Dissolve 5 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.
Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).
pH (2.2.3)
5.5 to 7.5 for solution S.
Absorbance (2.2.25)
Maximum 0.10, determined at 270 nm on solution S.

Impurity A

Thin-layer chromatography (2.2.27).

Test solution (a) Solution S.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with water R.

Reference solution (a) Dissolve 0.10 g of sodium cyclamate CRS in water R and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 10 mg of sulfamic acid R (impurity A) in water R and dilute to 100 mL with the same solvent.

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R, water R, ethyl acetate R, propanol R (10:10:20:70 V/V/V/V).

Application 2 µL.

Development Over a path of 12 cm.

Drying In a current of warm air, then heat at 105 °C for 5 min.

Detection Spray the hot plate with strong sodium hypochlorite solution R diluted to a concentration of 5 g/L of active chlorine. Place in a current of cold air until an area of coating below the points of application gives at most a faint blue colour with a drop of potassium iodide and starch solution R; avoid prolonged exposure to cold air. Spray with potassium iodide and starch solution R and examine the chromatograms within 5 min.

Limit: test solution (a):

— impurity A: any spot due to impurity A is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (0.1 per cent).

Impurities B, C and D

Gas chromatography (2.2.28).

Internal standard solution Dissolve 2 µL of tetradecane R in methylcyclohexane R and dilute to 100 mL with the same solvent.

Test solution Dissolve 2.00 g of the substance to be examined in 20 mL of water R, add 0.5 mL of strong sodium hydroxide solution R and shake with 30 mL of toluene R. Shake 20 mL of the upper layer with 4 mL of a mixture of equal volumes of dilute acetic acid R and water R. Separate the lower layer, add 0.5 mL of strong sodium hydroxide solution R and 0.5 mL of the internal standard solution and shake. Use the lower layer immediately after separation.

Reference solution Dissolve 10.0 mg (about 12 µL) of cyclohexylamine R (impurity C), 1.0 mg (about 1 µL) of dicyclohexylamine R (impurity D) and 1.0 mg (about 1 µL) of aniline R (impurity B) in water R, then dilute to 1000 mL

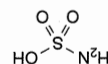
with the same solvent. Dilute 10.0 mL of this solution to 100.0 mL with water R (solution A). To 20.0 mL of solution A, add 0.5 mL of strong sodium hydroxide solution R and extract with 30 mL of toluene R. Shake 20 mL of the upper layer with 4 mL of a mixture of equal volumes of dilute acetic acid R and water R. Separate the lower layer, add 0.5 mL of strong sodium hydroxide solution R and 0.5 mL of the internal standard solution and shake. Use the lower layer immediately after separation.

Column:
— material: fused silica;
— size: $l = 25$ m, $\varnothing = 0.32$ mm;
— stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 0.51 μ m).
Carrier gas helium for chromatography R.
Flow rate 1.8 mL/min.
Temperature:

Time (min)	Temperature (°C)
0 - 1	85
1 - 9	85 \rightarrow 150
9 - 13	150
250	250
270	270

Detection Flame ionisation.
Injection 1.5 μ L; use a split vent at a flow rate of 20 mL/min.
Relative retention With reference to impurity C (retention time = about 2.3 min): impurity B = about 1.4; tetradecane = about 4.3; impurity D = about 4.5.
Limits:
— impurity C: maximum 10 ppm;
— impurities B, D: for each impurity, maximum 1 ppm.
Sulfates (2.4.13)
Maximum 0.1 per cent.
Dilute 1.5 mL of solution S to 15 mL with distilled water R.
Heavy metals (2.4.8)
Maximum 10 ppm.
12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.
Loss on drying (2.2.32)
Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.
ASSAY
Dissolve without heating 0.150 g in 60 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 20.12 mg of $C_6H_{12}NNaO_3S$.

Specified impurities A, B, C, D



A. sulfamic acid,

Heavy metals
12 mL of solution S complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (1 ppm Pb) to prepare the standard (10 ppm).

Arsenic
0.5 g complies with the limit test for arsenic, Appendix VII (2 ppm).

Clarity and colour of solution
Solution S is clear, Appendix IV A, and colourless, Appendix IV B, Method II.

Acidity
pH of a mixture of 5 mL of solution S and 5 mL of carbon dioxide-free water, 4.2 to 4.5, Appendix V L.

TESTS
D. Complies with the test for Loss on drying.
Acidity
pH of a mixture of 5 mL of solution S and 5 mL of carbon dioxide-free water, 4.2 to 4.5, Appendix V L.
Clarity and colour of solution
Solution S is clear, Appendix IV A, and colourless, Appendix IV B, Method II.
Arsenic
0.5 g complies with the limit test for arsenic, Appendix VII (2 ppm).

IDENTIFICATION
A. Solution S is faintly acid, Appendix V K.
B. Solution S yields the reactions characteristic of phosphates, Appendix VI.
C. Solution S neutralised with a 10% w/v solution of potassium hydroxide yields reaction A characteristic of sodium salts, Appendix VI.
D. Complies with the test for Loss on drying.

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

DEFINITION
Anhydrous Sodium Dihydrogen Phosphate contains not less than 98.0% and not more than 100.5% of NaH_2PO_4 , calculated with reference to the dried substance.

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

DEFINITION
Anhydrous Sodium Dihydrogen Phosphate contains not less than 98.0% and not more than 100.5% of NaH_2PO_4 , calculated with reference to the dried substance.

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

CHARACTERISTICS
White, slightly deliquescent crystals or granules.
Very soluble in water; very slightly soluble in ethanol (96%).
Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S).
From distilled water to produce 100 mL (solution S).

Iron 10 mL of solution S complies with the limit test for iron, Appendix VII (10 ppm).	Chloride Dilute 2.5 mL of solution S to 15 mL with water. The resulting solution complies with the limit test for chlorides, Appendix VII (200 ppm).	Sulfate To 5 mL of solution S add 0.5 mL of hydrochloric acid and dilute to 15 mL with distilled water. The resulting solution complies with the limit test for sulfates, Appendix VII (300 ppm).	Reducing substances To 5 mL of solution S add 0.25 mL of 0.02M potassium permanganate and 5 mL of 1M sulfuric acid and heat in a water bath for 5 minutes. The solution retains a slight red colour.	Loss on drying When dried to constant weight at 130°, loses not more than 1.0% of its weight. Use 1 g.	ASSAY Dissolve 2 g in 50 mL of water and titrate with carbonate-free 1M sodium hydroxide VS determining the end point potentiometrically. Each mL of 1M sodium hydroxide VS is equivalent to 0.120 g of NaH_2PO_4 .	Sodium Dihydrogen Phosphate Dihydrate Sodium Acid Phosphate $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$ 156.0 13472-35-0 (Ph Eur monograph 0194) Preparation Phosphates Enema Ph Eur	DEFINITION Content 98.0 per cent to 100.5 per cent (dried substance). CHARACTERS Appearance White or almost white powder or colourless crystals. Solubility Very soluble in water, very slightly soluble in ethanol (96 per cent).	IDENTIFICATION A. Solution S (see Tests) is slightly acid (2.2.4). B. Solution S gives the reactions of phosphates (2.3.1). C. Solution S previously neutralised using a 100 g/L solution of potassium hydroxide R gives reaction (a) of sodium (2.3.1). TESTS Solution S Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.	Appearance of solution Solution S is clear (2.2.1) and colourless (2.2.2, Method II). pH (2.2.3) 4.2 to 4.5.	To 5 mL of solution S add 5 mL of carbon dioxide-free water R.	Reducing substances To 5 mL of solution S add 0.25 mL of 0.02 M potassium permanganate and 5 mL of dilute sulfuric acid R and heat in a water-bath for 5 min. The colour of the permanganate is not completely discharged.	Chlorides (2.4.4) Maximum 200 ppm. Dilute 2.5 mL of solution S to 15 mL with water R.	Sulfates (2.4.13) Maximum 300 ppm. To 5 mL of solution S add 0.5 mL of hydrochloric acid R and dilute to 15 mL with distilled water R.	Arsenic (2.4.2, Method A) Maximum 2 ppm, determined on 0.5 g. Iron (2.4.9) Maximum 10 ppm, determined on solution S.	Heavy metals (2.4.8) Maximum 10 ppm. 12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.	Loss on drying (2.2.32) 21.5 per cent to 24.0 per cent, determined on 0.50 g by drying in an oven at 130 °C. ASSAY Dissolve 2.500 g in 40 mL of water R. Titrate with carbonate-free 1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). 1 mL of 1 M sodium hydroxide is equivalent to 0.120 g of NaH_2PO_4 . Ph Eur	Monohydrate $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ 138.0 10049-21-5 Action and use Excipient. Preparation Phosphates enema	DEFINITION Sodium Dihydrogen Phosphate Monohydrate contains not less than 98.0% and not more than 100.5% of NaH_2PO_4 , calculated with reference to the dried substance. CHARACTERISTICS Colourless crystals or a white powder. Very soluble in water, very slightly soluble in ethanol (96%). Dissolve 10.0 g in sufficient carbon dioxide-free water prepared from distilled water to produce 100 mL (solution S). IDENTIFICATION A. Solution S is faintly acid, Appendix V K. B. Solution S yields the reactions characteristic of phosphates, Appendix VI. C. Solution S neutralised with a 10% w/v solution of sodium potassium hydroxide yields reaction A characteristic of sodium salts, Appendix VI.
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D. Complies with the test for Loss on drying.

TESTS

Acidity

pH of a mixture of 5 mL of solution S and 5 mL of carbon dioxide-free water, 4.2 to 4.5, Appendix V L.

Clarity and colour of solution

Solution S is clear, Appendix IV A, and colourless, Appendix IV B, Method II.

Arsenic

0.5 g complies with the limit test for arsenic, Appendix VII (2 ppm).

Heavy metals

12 mL of solution S complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (1 ppm Pb) to prepare the standard (10 ppm).

Iron

10 mL of solution S complies with the limit test for iron, Appendix VII (10 ppm).

Chloride

Dilute 2.5 mL of solution S to 15 mL with water.

The resulting solution complies with the limit test for chlorides, Appendix VII (200 ppm).

Sulfate

To 5 mL of solution S add 0.5 mL of hydrochloric acid and dilute to 15 mL with distilled water. The resulting solution

complies with the limit test for sulfates, Appendix VII (300 ppm).

Reducing substances

To 5 mL of solution S add 0.25 mL of 0.02M potassium permanganate and 5 mL of 1M sulfuric acid and heat in a water bath for 5 minutes. The solution retains a slight red colour.

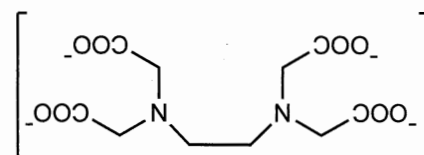
Loss on drying

When dried to constant weight at 130°, loses 11.5 to 14.5% of its weight. Use 0.5 g.

ASSAY

Dissolve 2.2 g in 50 mL of water and titrate with carbonate-free 1M sodium hydroxide VS determining the end point potentiometrically. Each mL of 1M sodium hydroxide VS is equivalent to 0.120 g of NaH_2PO_4 .

Sodium Feredetate



$\text{C}_{10}\text{H}_{12}\text{N}_2\text{O}_8\text{FeNa}_3\cdot 3\text{H}_2\text{O}$ 421.1 15708-41-5 (anhydrous)

Action and use

Source of iron.

Preparation

Sodium Feredetate Oral Solution

DEFINITION

Sodium Feredetate is iron(III) sodium ethylenediaminetetraacetate trihydrate. It contains not less than 98.0% and not

more than 102.0% of $\text{C}_{10}\text{H}_{12}\text{N}_2\text{O}_8\text{FeNa}_3$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A yellow or yellowish brown, crystalline powder.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of sodium feredetate (RS 378).

B. Ignite 0.5 g and allow to cool. Dissolve the residue in 2 mL of hydrochloric acid, add sufficient water to produce 20 mL and filter. The filtrate yields reaction C characteristic of iron salts, Appendix VI. Dilute 1 volume of the filtrate to 10 volumes with water; the solution yields reaction B characteristic of iron salts, Appendix VI.

C. Dissolve 2 g in 30 mL of water, slowly add 6.5 mL of a 20% w/v solution of potassium hydroxide, shake and filter the resulting suspension. Evaporate 8 mL of the filtrate to dryness, ignite and dissolve the residue in 4 mL of water. The solution yields reaction B characteristic of sodium salts, Appendix VI.

TESTS

Acidity

pH of a 1% w/v solution, 3.5 to 5.5, Appendix V L.

Free iron

Not more than 500 ppm when determined by the following method. Dissolve 0.2 g in sufficient water to produce 20 mL and filter. Label three tubes A, B and C. Place 5 mL of the filtrate into each of tubes A and B and 4 mL of water and 1 mL of an iron standard solution prepared in the following manner in tube C. For the iron standard solution dilute 25 volumes of a 0.1726% w/v solution of ammonium iron(III) sulfate in 0.05M sulfuric acid to 200 volumes with water.

(25 ppm of Fe(III)). Add 1 mL of a 1.0% w/v solution of disodium catechol-3,5-disulfonate into each of tubes A and C and 1 mL of water to tube B. Measure the absorbance of solution A at 670 nm, Appendix II B, using solution B in the reference cell and of solution C using water in the reference cell. The absorbance of solution A is not greater than that of solution C.

Heavy metals

1 g complies with limit test D for heavy metals, Appendix VII (20 ppm). Use 2 mL of lead standard solution (10 ppm Pb) to prepare the standard solution.

Free sodium edetate

To 4 mL of a 1.0% w/v solution add 2 mL of ferric iron standard solution (50 ppm) and 1 mL of a 1.0% w/v solution of disodium catechol-3,5-disulfonate and mix. Prepare a standard in the same manner using 4 mL of a 0.010% w/v solution of disodium edetate in place of the solution of the substance being examined. Measure the absorbance of the solutions at 670 nm, Appendix II B, using water in the reference cell. The absorbance of the solution of the substance being examined is not less than that of the standard solution (1%).

Nitritotriacetic acid

Carry out the method for liquid chromatography, Appendix III D, protected from light, using the following solutions.

Solvent A To 10 volumes of a 50% w/v solution of iron(III) sulfate pentahydrate in 0.5M sulfuric acid add 390 volumes of water, adjust to a pH of 2.0 with 1M sodium hydroxide and dilute to 500 volumes with water.

Sodium Fluoride

(Ph. Eur. monograph 0514)

NaF

41.99

7681-49-4



Action and use

Used in prevention of dental caries.

Preparations

Sodium Fluoride Mouthwash

Sodium Fluoride Oral Drops

Sodium Fluoride Oral Solution

Sodium Fluoride Tablets

Ph Eur

DEFINITION

Content

98.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder or colourless crystals.

Solubility

Soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. To 2 mL of solution S (see Tests) add 0.5 mL of calcium

chloride solution R. A gelatinous white precipitate is formed

that dissolves on adding 5 mL of ferric chloride solution R1.

B. To about 4 mg add a mixture of 0.1 mL of alizarin S

solution R and 0.1 mL of zincanyl nitrate solution R and mix.

The colour changes from red to yellow.

C. Solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R without heating

and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

Dissolve 2.5 g of potassium nitrate R in 40 mL of solution S

and dilute to 50 mL with carbon dioxide-free water R. Cool to

0 °C and add 0.2 mL of phenolphthalein solution R. If the

solution is colourless, not more than 1.0 mL of 0.1 M sodium

hydroxide is required to produce a red colour that persists for

at least 15 s. If the solution is red, not more than 0.25 mL of

0.1 M hydrochloric acid is required to change the colour of the

indicator.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 10 mL of solution S to 15 mL with water R.

Fluorosulfates

Heat to boiling the neutralised solution obtained in the test

for acidity or alkalinity and titrate whilst hot. Not more than

0.75 mL of 0.1 M sodium hydroxide is required to change the

colour of the indicator to red.

Sulfates (2.4.13)

Maximum 200 ppm.

Dissolve 0.25 g in 10 mL of a saturated solution of boric

acid R in distilled water R. Add 5 mL of distilled water R and

mixing 0.6 mL of hydrochloric acid R1, 5 mL of sulfate

(1) 0.4% w/w of the substance being examined in solvent A.

(2) 0.0004% w/v of nitrotriacetic acid in solvent A.

(3) 0.0004% w/v of nitrotriacetic acid in solution (1).

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (10 cm × 4.6 mm) packed with porous graphitic carbon (5 µm) (Hypersorb is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 1.0 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 273 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

To 50 volumes of a 0.01% w/v solution of iron(III) sulfate

pentahydrate in 0.5M sulfuric acid, add 750 volumes of water.

Adjust to a pH of 1.5 with 0.5M sulfuric acid or 1M sodium

hydroxide and add 20 volumes of ethylene glycol. Dilute to

1000 volumes with water.

When the chromatograms are recorded under the prescribed conditions the retention time relative to nitrotriacetic acid

(retention time about 5 minutes) is: sodium feredetate,

about 2.0.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained

with solution (3), the resolution between the peaks due to

nitrotriacetic acid and sodium feredetate is at least 4.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to nitrotriacetic acid is

not greater than the area of the principal peak in the

chromatogram obtained with solution 2 (0.1%);

Chlorides

Dissolve 1 g in 100 mL of water and filter. 15 mL of the

filtrate complies with the limit test for chlorides, Appendix VII

Sulfates

Dissolve 2 g in 40 mL of water, add 5 mL of a 20% w/v

solution of sodium hydroxide and sufficient water to produce

50 mL, shake and filter. Evaporate 6.2 mL of this solution to

dryness and ignite until no trace of carbon remains. Cool and

dissolve the residue in 10 mL of distilled water. Neutralise the

solution with 2M hydrochloric acid using litmus paper as

external indicator and add 2 mL in excess. Boil the solution,

cool, dilute to 15 mL with distilled water and filter.

Appendix VII (600 ppm).

Water

12.0% to 13.5% w/w, Appendix IX C Method I. Use 0.1 g.

ASSAY

Dissolve 1 g in 40 mL of water in an iodine flask, add 20 mL

of hydrochloric acid and 3 g of potassium iodide, close the flask

and allow to stand for 5 minutes. Titrate the liberated iodine

with 0.1M sodium thiosulfate VS using starch mucilage as

indicator. Repeat the procedure without the substance being

examined. The difference between the titrations represents

the amount of sodium thiosulfate required. Each mL of

0.1M sodium thiosulfate VS is equivalent to 36.71 mg of

 $C_{10}H_{12}O_8N_2FeNa$.

standard solution (10 ppm SO_4) R and 10 mL of a saturated solution of boric acid R in distilled water R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 130 °C for 3 h.

ASSAY

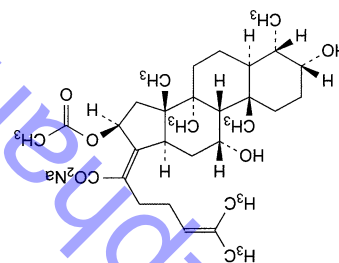
Dissolve 0.100 g in water R and dilute to 60 mL with the same solvent. Titrate with 0.1 M lanthanum nitrate, determining the end-point potentiometrically (2.2.20) using a fluoride-selective indicator electrode and a silver-silver chloride reference electrode.

1 mL of 0.1 M lanthanum nitrate is equivalent to 12.60 mg of NaF .

Ph Eur

Sodium Fusidate

(Ph. Eur. monograph 0848)



$\text{C}_{31}\text{H}_{47}\text{NaO}_6$

538.7

751-94-0

Action and use

Antibacterial.

Preparation

Sodium Fusidate Ointment

Ph Eur

DEFINITION

Sodium *ent*-(17Z)-16 α -(acetyloxy)-3 β ,11 β -dihydroxy-4 β ,8,14-trimethyl-18-nor-5 β ,10 α -cholesta-17(20),24-dien-21-oate. Antimicrobial substance produced by fermentation of certain strains of *Fusidium coccineum* or by any other means.

Content

97.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder, slightly hygroscopic.

Solubility

Freely soluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium fusidate CRS.

B. Ignite 1 g. The residue gives reaction (a) of sodium (2.3.1).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Dissolve 1.5 g in 10 mL of water R.

pH (2.2.3) 7.5 to 9.0.
Dissolve 0.125 g in 10 mL of carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture methanol R, 5 g/L solution of phosphoric acid R, acetonitrile R (10:40:50 V/V/V).

Test solution Dissolve 25 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of fusidic acid for peak identification CRS (containing impurities A, B, C, D, F, G, H and N) in the solvent mixture and dilute to 1.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve the contents of a vial of fusidic acid impurity mixture CRS (containing impurities I, K, L and M) in 1.0 mL of the solvent mixture.

Column: — size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3.5 μm);
— temperature: 30 °C.

Mobile phase: — mobile phase A: methanol R, acetonitrile R, 5 g/L solution of phosphoric acid R (20:40:40 V/V/V);
— mobile phase B: 5 g/L solution of phosphoric acid R, methanol R, acetonitrile R (10:20:70 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	100	0
3 - 28	100 \rightarrow 0	0 \rightarrow 100
28 - 33	0	100

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 235 nm.

Injection 20 μL .

Identification of impurities Use the chromatogram supplied with fusidic acid for peak identification CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, F, G, H and N; use the chromatogram supplied with fusidic acid impurity mixture CRS and the chromatogram obtained with reference solution (d) to identify the peaks due to impurities I, K, L and M.

Relative retention With reference to fusidic acid (retention time = about 18 min): impurity A = about 0.4;

impurity B = about 0.5; impurity C = about 0.6; impurity D = about 0.63; impurity N = about 0.65;

impurity F = about 0.7; impurity G = about 0.82;

impurity H = about 0.85; impurity I = about 0.96;

impurity K = about 1.18; impurity L = about 1.23;

impurity M = about 1.4.

System suitability: reference solution (a):

— resolution: minimum 1.5 between the peaks due to impurities G and H.

Limits:
— *correction factors:* for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.7; impurity D = 0.7; impurity F = 0.3; impurity I = 0.6; impurity K = 0.6;

— *impurity M:* not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— *impurity G:* not more than 0.7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);

— *impurity L:* not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *impurity B:* not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.4 per cent);

— *impurity A:* not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

— *impurities C, D, F, I, K, M:* for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— *unspecified impurities:* for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— *total:* not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— *disregard limit:* 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Water (2.5.12)

Maximum 2.0 per cent, determined on 0.500 g.

ASSAY

Dissolve 0.400 g in 30 mL of water R and add 40 mL of ethanol (96 per cent) R. Titrate with 0.1 M hydrochloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M hydrochloric acid is equivalent to 53.87 mg of $C_{31}H_{47}NaO_6$.

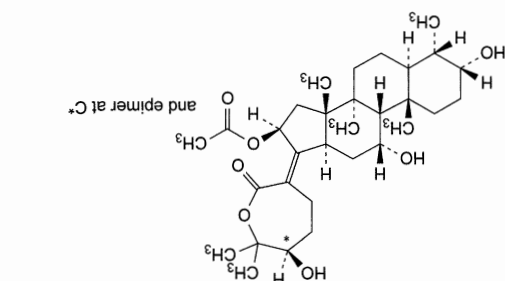
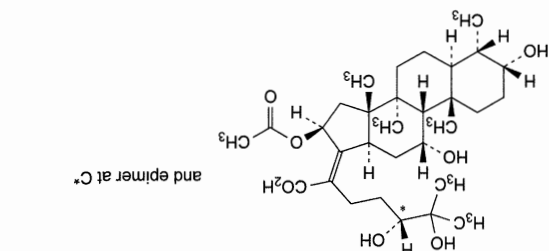
STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

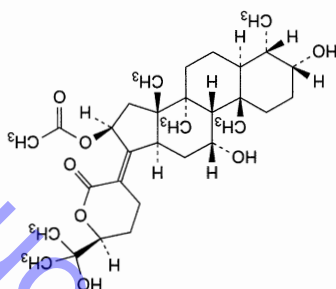
IMPURITIES

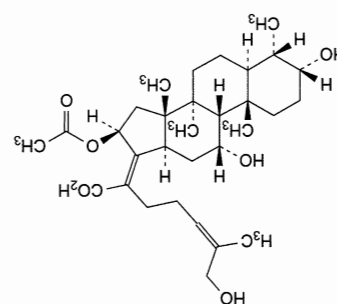
Specified impurities A, B, C, D, F, G, I, K, L, M, N

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*: E, H, J, O.

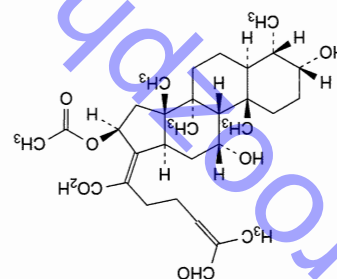


C. *ent*-(17*Z*)-3β,11β-dihydroxy-17-[(6*S*)-6-(1-hydroxy-1-methylethyl)-2-oxodihydro-2*H*-pyran-3(4*H*)-ylidene]-4β,8,14-trimethyl-18-nor-5β,10α-androstan-16α-yl acetate ((24*R*)-24,25-dihydro-24,25-dihydroxyfusidic acid 21,24-lactone),

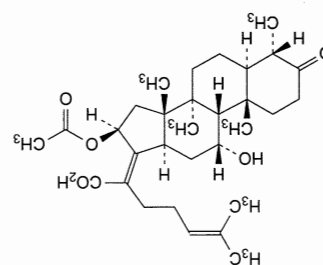




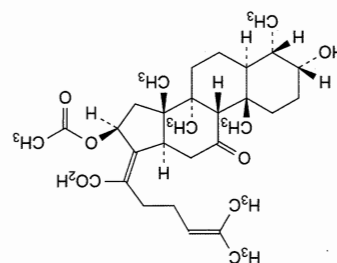
E. *ent*-(17Z,24EZ)-16α-(acetyloxy)-3β,11β,26-trihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (26-hydroxyfusidic acid),



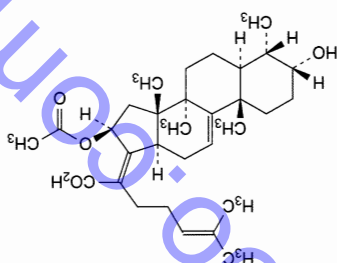
F. *ent*-(17Z,24EZ)-16α-(acetyloxy)-3β,11β-dihydroxy-4β,8,14-trimethyl-26-oxo-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (26-oxofusidic acid),



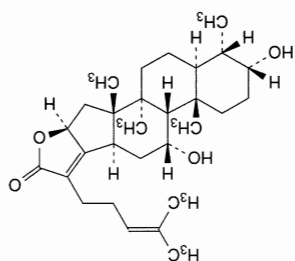
G. *ent*-(17Z)-16α-(acetyloxy)-11β-hydroxy-4β,8,14-trimethyl-3-oxo-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (3-didehydrofusidic acid),



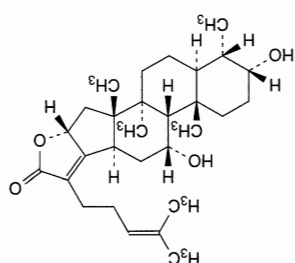
H. *ent*-(17Z)-16α-(acetyloxy)-3β-hydroxy-4β,8,14-trimethyl-11-oxo-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (11-didehydrofusidic acid),



L. *ent*-(17Z)-16α-(acetyloxy)-3β-hydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-9(11),17(20),24-trien-21-oic acid (9,11-anhydrofusidic acid),

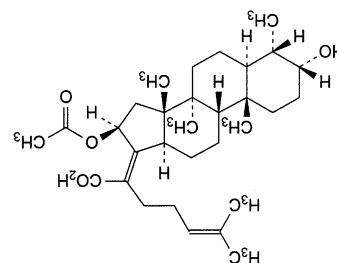


K. *ent*-(17Z)-3β,11β-dihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dieno-21(16α)-lactone (deacetylifusidic acid 21,16-lactone),



J. *ent*-(17Z)-3β,11β-dihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dieno-21(16β)-lactone (16-*epi*-deacetylifusidic acid 21,16-lactone),

I. *ent*-(17Z)-3β,11β,16β-trihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid (16-*epi*-deacetylifusidic acid),

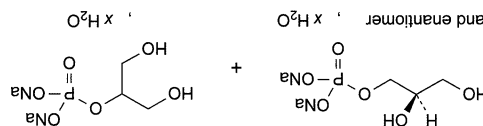


M. *ent*-(17Z)-16α-(acetyloxy)-3β-hydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid
(11-deoxyfusidic acid),
N. unknown structure,

O. *ent*-(17Z)-3β,11β,16α-trihydroxy-4β,8,14-trimethyl-18-nor-5β,10α-cholesta-17(20),24-dien-21-oic acid
(deacetylfusidic acid).

Hydrated Sodium Glycerocephosphate

(Ph. Eur. monograph 1995)



$C_3H_7Na_2O_6P, xH_2O$
216.0
(anhydrous substance)

DEFINITION

Mixture of variable proportions of sodium (2*RS*)-2,3-dihydroxypropyl phosphate and sodium 2-hydroxy-1-(hydroxymethyl)ethyl phosphate. The mixture may contain various amounts of other glycerophosphate esters. The degree of hydration is 4 to 6.

Content

98.0 per cent to 105.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or crystals.

Solubility

Freely soluble in water, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) gives reaction (a) of sodium (2.3.1).

B. To 0.1 g add 5 mL of dilute nitric acid R. Heat to boiling and boil for 1 min. Cool. The solution gives reaction (b) of phosphates (2.3.1).
C. In a test-tube fitted with a glass tube, mix 0.1 g with 5 g of potassium hydrogen sulfate R. Heat strongly and direct the white vapour into 5 mL of decolourised fuchsin solution R. A violet-red colour develops which becomes violet upon heating for 30 min on a water-bath.

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Alkalinity

To 10 mL of solution S add 0.2 mL of phenolphthalein solution R. Not more than 1.0 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator (*n*₂).

Glycerol and ethanol (96 per cent)-soluble substances

Maximum 1.0 per cent.
Shake 1.000 g with 25 mL of ethanol (96 per cent) R for 10 min. Filter. Evaporate the filtrate on a water-bath and dry the residue at 70 °C for 1 h. The residue weighs not more than 10 mg.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 2.5 mL of solution S to 15 mL with water R.

Phosphates (2.4.11)

Maximum 0.1 per cent.

Dilute 1 mL of solution S to 10 mL with water R. Dilute 1 mL of this solution to 100 mL with water R.

Sulfates (2.4.13)

Maximum 500 ppm.

Dilute 3 mL of solution S to 15 mL with water R.

Iron (2.4.9)

Maximum 20 ppm.

Dilute 5 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dilute 10 mL of solution S to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Water (2.5.12)

25.0 per cent to 35.0 per cent, determined on 0.100 g.

ASSAY

Dissolve 0.250 g in 30 mL of water R. Titrate with 0.05 M sulfuric acid, determining the end-point potentiometrically (2.2.20), (*n*₁).

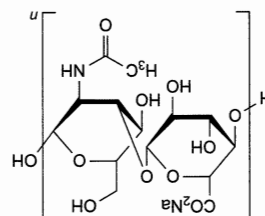
Calculate the percentage content of sodium glycerophosphate (anhydrous substance) using the following expression:

$$\frac{216.0}{n_1 - \frac{n_2}{2}} \left(\frac{m}{100} - a \right)$$

a = percentage content of water;
*n*₁ = volume of 0.05 M sulfuric acid used in the assay, in millilitres;
*n*₂ = volume of 0.1 M hydrochloric acid used in the test for alkalinity, in millilitres;
m = mass of the substance to be examined, in grams.

Sodium Hyaluronate

(Ph. Eur. monograph 1472)



Intrinsic viscosity

Sodium hyaluronate is very hygroscopic and must be protected from moisture during weighing.

Buffer solution (0.15 M sodium chloride in 0.01 M phosphate buffer solution pH 7.0). Dissolve 0.78 g of sodium dihydrogen phosphate R and 4.50 g of sodium chloride R in water R and dilute to 500.0 mL with the same solvent (solution A). Dissolve 1.79 g of disodium hydrogen phosphate R and 4.50 g of sodium chloride R in water R and dilute to 500.0 mL with the same solvent (solution B). Mix solutions A and B until a pH of 7.0 is reached. Filter through a sintered-glass filter (4) (2.1.2).

Test solution (a) Weigh 0.200 g (m_{op}) (NOTE: this value is only indicative and should be adjusted after an initial measurement of the viscosity of test solution (a)) of the substance to be examined and dilute with 50.0 g (m_{os}) of buffer solution at 4 °C. Mix the solution by shaking at 4 °C during 24 h. Weigh 5.00 g (m_{ip}) of the solution and dilute with 100.0 g (m_{is}) of buffer solution at 25 °C. Mix this solution by shaking for 20 min. Filter the solution through a sintered-glass filter (100) (2.1.2), and discard the first 10 mL.

Test solution (b) Weigh 30.0 g (m_{2p}) of test solution (a) and dilute with 10.0 g (m_{2s}) of buffer solution at 25 °C. Mix this solution by shaking for 20 min. Filter the solution through a sintered-glass filter (100) (2.1.2) and discard the first 10 mL.

Test solution (c) Weigh 20.0 g (m_{3p}) of test solution (a) and dilute with 20.0 g (m_{3s}) of buffer solution at 25 °C. Mix this solution by shaking for 20 min. Filter the solution through a sintered-glass filter (100) (2.1.2) and discard the first 10 mL.

Test solution (d) Weigh 10.0 g (m_{4p}) of test solution (a) and dilute with 30.0 g (m_{4s}) of buffer solution at 25 °C. Mix this solution by shaking for 20 min. Filter the solution through a sintered-glass filter (100) (2.1.2) and discard the first 10 mL.

Determine the flow-times (2.2.9) for the buffer solution (t_0) and for the 4 test solutions (t_1 , t_2 , t_3 and t_4), at 25.00 ± 0.03 °C. Use an appropriate suspended level viscometer (specifications: viscometer constant about 0.005 mm²/s²; kinematic viscosity of 1–5 mm²/s, internal diameter of tube R 0.53 mm, volume of bulb C 5.6 mL, internal diameter of tube N 2.8–3.2 mm) with a funnel-shaped lower capillary end. Use the same viscometer for all measurements; measure all outflow times in triplicate.

The test is not valid unless the results do not differ by more than 0.35 per cent from the mean and if the flow time t_1 is not less than 1.6 and not more than 1.8 times t_0 . If this is not the case, adjust the value of m_{op} and repeat the procedure.

Calculation of the relative viscosities Since the densities of the sodium hyaluronate solutions and of the solvent are almost equal, the relative viscosities η_r (being η_{r1} , η_{r2} , η_{r3} and η_{r4}) can be calculated from the ratio of the flow times for the respective solutions t_i (being t_1 , t_2 , t_3 and t_4) to the flow time of the solvent t_0 , but taking into account the kinetic energy correction factor for the capillary ($B = 30\,800\,s^3$), using the following expression:

$$\eta_r = \frac{t_i - \frac{B}{t_i}}{t_0 - \frac{B}{t_0}}$$

Calculation of the concentrations Calculate the concentration c_i (expressed in kg/m³) of sodium hyaluronate in test solution (a) using the following expression:

(C₁₄H₂₀NNaO₁₁)_n

9067-32-7

Action and use
High viscosity mucopolysaccharide.

Ph. Eur.

DEFINITION

Sodium salt of hyaluronic acid, a glycosaminoglycan consisting of D-glucuronic acid and N-acetyl-D-glucosamine disaccharide units. It is extracted from cocks' combs or obtained by fermentation from *Streptococcus*, *Lancefield* Groups A and C.

Content

95.0 per cent to 105.0 per cent (dried substance).
90 per cent to 120 per cent of the value stated on the label.

PRODUCTION

Where applicable, the animals from which sodium hyaluronate is derived must fulfil the requirements for the health of animals suitable for human consumption.

When produced by fermentation of gram-positive bacteria, the process must be shown to reduce or eliminate pyrogenic or inflammatory components of the cell wall.

CHARACTERS

Appearance
White or almost white, very hygroscopic powder or fibrous aggregate.

Solubility

Sparsely soluble or soluble in water, practically insoluble in acetone and in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison Ph. Eur. reference spectrum of sodium hyaluronate.
B. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S
Weigh a quantity of the substance to be examined equivalent to 0.10 g of the dried substance and add 30.0 mL of a 9 g/L solution of sodium chloride R. Mix gently on a shaker until dissolved (about 12 h).

Appearance of solution

Solution S is clear (2.2.1) and its absorbance (2.2.25) at 600 nm is not greater than 0.01.

pH (2.2.3)

5.0 to 8.5.

Dissolve the substance to be examined in carbon dioxide-free water R to obtain a solution containing a quantity equivalent to 5 mg of the dried substance per millilitre.

$$\frac{m_{op} \times x \times (100 - h) \times m_{ip} \times p_{25}}{100 \times 100 \times (m_{op} + m_{os}) \times (m_{ip} + m_{is})}$$

x = percentage content of sodium hyaluronate as

determined under Assay;

h = percentage loss on drying;

$p_{25} = 1005 \text{ kg/m}^3$ (density of the test solution at 25 °C).

Calculate the concentration c_2 (expressed in kg/m^3) of sodium hyaluronate in test solution (b) using the following expression:

$$c_1 \times \frac{m_{2s} + m_{2p}}{m_{2p}}$$

Calculate the concentration c_3 (expressed in kg/m^3) of sodium hyaluronate in test solution (c) using the following expression:

$$c_1 \times \frac{m_{3s} + m_{3p}}{m_{3p}}$$

Calculate the concentration c_4 (expressed in kg/m^3) of sodium hyaluronate in test solution (d) using the following expression:

$$c_1 \times \frac{m_{4s} + m_{4p}}{m_{4p}}$$

Calculation of the intrinsic viscosity Calculate the intrinsic viscosity $[\eta]$ by linear least-squares regression analysis using the Martin equation:

$$\log \left(\frac{c}{c_1 - 1} \right) = \log [\eta] + k [\eta] c$$

The decimal antilogarithm of the intercept is the intrinsic viscosity expressed in m^3/kg .

Sulfated glycosaminoglycans

Maximum 1 per cent, if the product is extracted from cocks'

combs.

Appropriate safety precautions are to be taken when handling

perchloric acid at elevated temperature.

Test solution Introduce a quantity of the substance to be examined equivalent to 50.0 mg of the dried substance into a

test-tube 150 mm long and 16 mm in internal diameter and

dissolve in 1.0 mL of perchloric acid R.

Reference solution Dissolve 0.149 g of anhydrous sodium

sulfate R in water R and dilute to 100.0 mL with the same

solvent. Dilute 10.0 mL of this solution to 100.0 mL with

water R. Evaporate 1.0 mL in a test-tube 150 mm long and

16 mm in internal diameter in a heating block at 90–95 °C,

and dissolve the residue in 1.0 mL of perchloric acid R.

Plug each test-tube with a piece of glass wool. Place the test-

tubes in a heating block or a silicone oil bath maintained at

180 °C and heat until clear, colourless solutions are obtained

(about 12 h). Remove the test-tubes and cool to room

temperature. Add to each test-tube 3.0 mL of a 33.3 g/L

solution of *barium chloride* R, cap and shake vigorously. Allow

the test-tubes to stand for 30 min. Shake each test-tube once

again, and determine the absorbance (2.2.25) at 660 nm,

using water R as a blank.

The absorbance obtained with the test solution is not greater

than the absorbance obtained with the reference solution.

Nucleic acids

The absorbance (2.2.25) of solution S at 260 nm is

maximum 0.5.

Protein

Maximum 0.3 per cent; maximum 0.1 per cent, if intended

for use in the manufacture of parenteral preparations.

Test solution (a) Dissolve the substance to be examined in

water R to obtain a solution containing a quantity equivalent

to about 10 mg of the dried substance per millilitre.

Test solution (b) Mix equal volumes of test solution (a) and

water R.

Reference solutions Prepare a 0.5 mg/mL stock solution of

bovine albumin R in water R. Prepare 5 dilutions of the stock

solution containing between 5 µg/mL and 50 µg/mL of bovine

albumin R.

Add 2.5 mL of freshly prepared cupri-tartrate solution R3 to

test-tubes containing 2.5 mL of water R (blank), 2.5 mL of

the test solutions (a) or (b) or 2.5 mL of the reference

solutions. Mix after each addition. After about 10 min, add

to each test-tube 0.50 mL of a mixture of equal volumes of

phosphomolybdic reagent R and water R prepared

immediately before use. Mix after each addition. After

30 min, measure the absorbance (2.2.25) of each solution at

750 nm against the blank. From the calibration curve

obtained with the 5 reference solutions determine the content

of protein in the test solutions.

Chlorides (2.4.4)

Maximum 0.5 per cent.

Dissolve 67 mg in 100 mL of water R.

Iron

Maximum 80 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve a quantity of the substance to be

examined equivalent to 0.25 g of the dried substance in

1 mL of nitric acid R by heating on a water-bath. Cool and

dilute to 10.0 mL with water R.

Reference solutions Prepare 2 reference solutions in the same

manner as the test solution, adding 1.0 mL and 2.0 mL

respectively of non standard solution (10 ppm Fe) R to the

dissolved substance to be examined.

Source Iron hollow-cathode lamp using a transmission band

of 0.2 nm.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Heavy metals (2.4.8)

Maximum 20 ppm; maximum 10 ppm if intended for use in

the manufacture of parenteral preparations.

1.0 g complies with test F. Prepare the reference solution

using 2.0 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 20.0 per cent, determined on 0.500 g by drying at

100–110 °C over diphosphorus pentoxide R for 6 h.

Microbial contamination

TAMC: acceptance criterion 10^2 CFU/g (2.6.12). Use 1 g of

the substance to be examined.

Bacterial endotoxins (2.6.14)

Less than 0.5 IU/mg, if intended for use in the manufacture

of parenteral preparations without a further appropriate

procedure for the removal of bacterial endotoxins; less than

0.05 IU/mg, if intended for use in the manufacture of intra-

ocular preparations or intra-articular preparations without a

further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Determine the glucuronic acid content by reaction with

carbazole as described below.

Reagent A Dissolve 0.95 g of disodium tetraborate R in

100.0 mL of sulfuric acid R.

Reagent B Dissolve 0.125 g of carbazole R in 100.0 mL of

anhydrous ethanol R.

Test solution Prepare this solution in triplicate. Dissolve

0.170 g of the substance to be examined in water R and

dilute to 100.0 g with the same solvent. Dilute 10.0 g of this

solution to 200.0 g with water R.

Reference stock solution Dissolve 0.100 g of D-glucuronic acid R,

previously dried to constant mass in vacuum over

diphosphorus pentoxide R (2.2.32), in water R and dilute to

100.0 g with the same solvent.

Reference solutions Prepare 5 dilutions of the reference stock

solution containing between 6.5 µg/g and 65 µg/g of D-

glucuronic acid R.

Place 25 test-tubes, numbered 1 to 25, in iced water.

Add 1.0 mL of the 5 reference solutions in triplicate to the

test-tubes 1 to 15 (reference tubes), 1.0 mL of the 3 test

solutions in triplicate to the test-tubes 16 to 24 (sample

tubes), and 1.0 mL of water R to test-tube 25 (blank).

Add to each test-tube 5.0 mL of freshly prepared reagent A,

previously cooled in iced water. Tightly close the test-tubes

with plastic caps, shake the contents, and place on a water

bath for exactly 15 min. Cool in iced water, and add to each

test tube 0.20 mL of reagent B. Recap the tubes, shake, and

put them again on a water-bath for exactly 15 min. Cool to

room temperature and measure the absorbance (2.2.25) of

the solutions at 530 nm, against the blank.

From the calibration curve obtained with the mean

absorbances read for each reference solution, determine the

mean concentrations of D-glucuronic acid in the test

solutions.

Calculate the percentage content of sodium hyaluronate

using the following expression:

$$\frac{c_g}{c_s} \times Z \times \frac{100}{100 - h} \times \frac{194.1}{401.3}$$

c_g = mean of concentrations of D-glucuronic acid in

the test solutions, in milligrams per gram;

c_s = mean of concentrations of the substance to be

examined in the test solutions, in milligrams

per gram;

Z = determined percentage content of $C_6H_{10}O_7$ in

D-glucuronic acid R;

h = percentage loss on drying;

401.3 = relative molecular mass of the disaccharide

fragment;

194.1 =

STORAGE

In an airtight container, protected from light and humidity.

If the substance is sterile, store in a sterile, airtight, tamper-

proof container.

LABELLING

The label states:

— the intrinsic viscosity;

— the origin of the substance;

— the intended use of the substance;

Sodium Hydroxide

Caustic Soda

(Ph. Eur. monograph 0677)

NaOH

40.00

1310-73-2

Ph Eur

Ph Eur



- where applicable, that the substance is suitable for parenteral administration other than intra-articular administration;
- where applicable, that the substance is suitable for parenteral administration, including intra-articular administration;
- where applicable that the material is suitable for intra-ocular use.

Appearance of solution
The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Carbonates
Maximum 2.0 per cent, calculated as Na_2CO_3 , as determined in the assay.

Chlorides (2.4.4)
Maximum 50 ppm.
Dissolve 1.0 g in 5 mL of water R, acidify the solution with about 4 mL of nitric acid R and dilute to 15 mL with water R. The solution, without addition of dilute nitric acid R, complies with the test.

Sulfates (2.4.13)
Maximum 50 ppm.
Dissolve 3.0 g in 6 mL of distilled water R, adjust to pH 7 with hydrochloric acid R (about 7.5 mL) and dilute to 15 mL with distilled water R.

Iron (2.4.9)
Maximum 10 ppm, determined on solution S.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

ASSAY

Dissolve 2.000 g in about 80 mL of carbon dioxide-free water R. Add 0.3 mL of phenolphthalein solution R and titrate with 1 M hydrochloric acid. Add 0.3 mL of methyl orange solution R and continue the titration with 1 M hydrochloric acid.

1 mL of 1 M hydrochloric acid used in the 2nd part of the titration is equivalent to 0.1060 g of Na₂CO₃.

1 mL of 1 M hydrochloric acid used in the combined titrations is equivalent to 40.00 mg of total alkali, calculated as NaOH.

STORAGE

In an airtight, non-metallic container.

Sodium Iodide

(Ph. Eur. monograph 0196)

NaI

Preparation
Sodium Iodide Injection

Ph Eur

DEFINITION

Content

99.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals, hygroscopic.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) gives the reactions of iodides (2.3.1).

B. Solution S gives the reactions of sodium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Alkalinity

To 12.5 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.7 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator.

Iodates

To 10 mL of solution S add 0.25 mL of iodide-free starch solution R and 0.2 mL of dilute sulfuric acid R and allow to stand protected from light for 2 min. No blue colour develops.

Sulfates (2.4.13)

Maximum 150 ppm.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Thiosulfates

To 10 mL of solution S add 0.1 mL of starch solution R and 0.1 mL of 0.005 M iodine. A blue colour is produced.

Sodium Lactate Solution

(Ph. Eur. monograph 1151)

Action and use

Systemic alkalinising agent.

Ph Eur

DEFINITION

Solution of a mixture of the enantiomers of sodium 2-hydroxypropionate in approximately equal proportions.

Content

Minimum declared content 50 per cent *m/m* of sodium 2-hydroxypropionate (C₃H₅NaO₃; 112.1); 96.0 per cent to 104.0 per cent of the content of sodium lactate stated on the label.

CHARACTERS

Appearance

Clear, colourless, slightly syrupy liquid.

Solubility

Miscible with water and with ethanol (96 per cent).

IDENTIFICATION

A. To 0.1 mL add 10 mL of water R. 5 mL of the solution gives the reaction of lactates (2.3.1).

B. It gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dilute a quantity of the substance to be examined corresponding to 40.0 g of sodium lactate to 200 mL with distilled water R.

Appearance of solution

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

pH (2.2.3)

6.5 to 9.0 for the substance to be examined.

Reducing sugars and sucrose

To 5 mL of the substance to be examined add 0.2 mL of copper sulfate solution R and 2 mL of dilute sodium hydroxide solution R. The solution is clear and blue and remains so on boiling. Add to the hot solution 4 mL of hydrochloric acid R. Boil for 1 min. Add 6 mL of strong sodium hydroxide solution R and heat to boiling again. The solution is clear and blue.

Methanol

Gas chromatography (2.4.24).

Limit:

— methanol: maximum 50 ppm, calculated with reference to sodium lactate, if intended for use in the manufacture of parenteral preparations, dialysis, haemodialysis or haemofiltration solutions.

Chlorides (2.4.4)

Maximum 50 ppm calculated with reference to sodium lactate.

Dilute 5 mL of solution S to 15 mL with water R. The solution complies with the test for chlorides.

Oxalates and phosphates

To 1 mL of the substance to be examined add 15 mL of ethanol (96 per cent) R and 2 mL of calcium chloride solution R. Heat at 75 °C for 5 min. Any opalescence in the solution is not more intense than that of a standard prepared at the same time and in the same manner using a mixture of 1 mL of the substance to be examined, 15 mL of ethanol (96 per cent) R and 2 mL of water R.

Sulfates (2.4.13)

Maximum 100 ppm calculated with reference to sodium lactate.

To 7.5 mL of solution S, add 1.9 mL of hydrochloric acid R1 and dilute to 15 mL with distilled water R. The solution complies with the test for sulfates without addition of 0.5 mL of acetic acid R. Acidify the standard solution with 0.05 mL of hydrochloric acid R1 instead of 0.5 mL of acetic acid R.

Aluminium

Maximum 0.1 ppm, if intended for use in the manufacture of parenteral preparations, dialysis, haemodialysis or haemofiltration solutions.

Atomic absorption spectrometry (2.2.23, Method I). For the preparation of the solutions, use equipment that is aluminium-free or that will not release aluminium under the conditions of use (glass, polyethylene, etc).

Modifier solution Dissolve 100.0 g of ammonium nitrate R in a mixture of 4 mL of nitric acid R and 50 mL of water R and dilute to 200 mL with water R.

Blank solution Dilute to 2.0 mL of the modifier solution to 25.0 mL with water R.

Test solution To 5.0 g add 2.0 mL of the modifier solution and dilute to 25.0 mL with water R.

Reference solutions Prepare the reference solutions immediately before use (0.010 ppm to 0.050 ppm of aluminium) using aluminium standard solution (200 ppm Al) R.

Source Aluminium hollow-cathode lamp.

Wavelength 309.3 nm.

Atomisation device A graphite furnace.

Carrier gas argon R.

Conditions The device is equipped with a non-specific absorption correction system. Heat the oven to 120 °C for as many seconds as there are microlitres of solution introduced into the apparatus, then heat at 1000 °C for 30 s and finally at 2700 °C for 6 s.

Sodium (S)-Lactate Solution

(Ph. Eur. monograph 2033)

**DEFINITION****Content**

Minimum 50.0 per cent *m/m* of sodium (S)-2-hydroxypropionate ($C_3H_5NaO_3$; 112.1); 96.0 per cent to 104.0 per cent of the content of sodium lactate stated on the label, not less than 95.0 per cent of which is the (S)-enantiomer.

CHARACTERS**Appearance**

Clear, colourless, slightly syrupy liquid.

Solubility

Miscible with water and with ethanol (96 per cent).

IDENTIFICATION

A. To 0.1 mL add 10 mL of water R. 5 mL of the solution gives the reaction of lactates (2.3.1).

LABELLING

The label states:

- where applicable, that the substance is suitable for use in the manufacture of dialysis, haemodialysis and haemofiltration solutions,
- where applicable, that the substance is suitable for use in the manufacture of parenteral preparations,
- the declared content of sodium lactate.

ASSAY

Dissolve a quantity of the substance to be examined corresponding to 75.0 mg of sodium lactate in a mixture of 10 mL of glacial acetic acid R and 20 mL of acetic anhydride R. Allow to stand for 15 min. Add 1 mL of naphtholbenzenesulfonate solution R and titrate with 0.1 M perchloric acid. 1 mL of 0.1 M perchloric acid is equivalent to 11.21 mg of $C_3H_5NaO_3$.

Bacterial endotoxins (2.6.14)

Less than 5 IU/g, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Heavy metals (2.4.8)

The solution complies with the test for iron. Dilute 5 mL of solution S to 10 mL with water R.

Iron (2.4.9)

Maximum 10 ppm calculated with reference to sodium lactate. Maximum 10 ppm calculated with reference to sodium lactate.

To 10 mL of solution S add 10 mL of calcium sulfate solution R. Allow to stand for 30 min. Any opalescence (2.2.1) in the solution is not more intense than that of a standard prepared at the same time and in the same manner using a mixture of 10 mL of solution S and 10 mL of distilled water R.

B. It gives reaction (a) of sodium (2.3.1).
C. It complies with the limits of the assay.

TESTS

Solution S

Dilute a quantity of the substance to be examined corresponding to 40.0 g of sodium lactate to 200 mL with distilled water R.

Appearance of solution

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

pH (2.2.3)

6.5 to 9.0 for the substance to be examined.

Reducing sugars and sucrose

To 5 mL of the substance to be examined add 2 mL of dilute sodium hydroxide solution R and 0.2 mL of copper sulfate solution R. The solution is clear and blue and remains so on boiling. Add to the hot solution 4 mL of hydrochloric acid R. Boil for 1 min. Add 6 mL of strong sodium hydroxide solution R and heat to boiling again. The solution is clear and blue.

Methanol

Gas chromatography (2.4.24).

Limit:

— methanol: maximum 50 ppm, calculated with reference to sodium lactate, if intended for use in the manufacture of parenteral preparations, dialysis, haemodialysis or haemofiltration solutions.

Chlorides (2.4.4)

Maximum 50 ppm calculated with reference to sodium lactate.

Dilute 5 mL of solution S to 15 mL with water R.

Oxalates and phosphates

To 1 mL of the substance to be examined add 15 mL of ethanol (96 per cent) R and 2 mL of calcium chloride solution R. Heat at 75 °C for 5 min. Any opalescence in the solution is not more intense than that of a standard prepared at the same time and in the same manner using a mixture of 1 mL of the substance to be examined, 15 mL of ethanol (96 per cent) R and 2 mL of water R.

Sulfates (2.4.13)

Maximum 100 ppm calculated with reference to sodium lactate.

To 7.5 mL of solution S, add 1.9 mL of hydrochloric acid R1 and dilute to 15 mL with distilled water R. The solution complies with the limit test for sulfates without addition of 0.5 mL of acetic acid R. Acidify the standard solution with 0.05 mL of hydrochloric acid R1 instead of 0.5 mL of acetic acid R.

Aluminium

Maximum 0.1 ppm, if intended for use in the manufacture of parenteral preparations, dialysis, haemodialysis or haemofiltration solutions.

Atomic absorption spectrometry (2.2.23, Method I). For the preparation of the solutions, use equipment that is aluminium-free or that will not release aluminium under the conditions of use (glass, polyethylene, etc).

Modifier solution Dissolve 100.0 g of ammonium nitrate R in a mixture of 50 mL of water R and 4 mL of nitric acid R and dilute to 200 mL with water R.

Blank solution Dilute 2.0 mL of the modifier solution to 25.0 mL with water R.

Test solution To 1.25 g add 2.0 mL of the modifier solution and dilute to 25.0 mL with water R.

Reference solutions Prepare the reference solutions immediately before use (0.010 ppm to 0.050 ppm of aluminium) using aluminium standard solution (200 ppm Al) R.

Source Aluminium hollow-cathode lamp.

Wavelength 309.3 nm.

Atomisation device A graphite furnace.

Carrier gas argon R.

Conditions The device is equipped with a non-specific absorption correction system. Heat the oven to 120 °C for as many seconds as there are microlitres of solution introduced into the apparatus, then heat at 1000 °C for 30 s and finally at 2700 °C for 6 s.

Barium

To 10 mL of solution S add 10 mL of calcium sulfate solution R. Allow to stand for 30 min. Any opalescence (2.2.1) in the solution is not more intense than that of a standard prepared at the same time and in the same manner using a mixture of 10 mL of solution S and 10 mL of distilled water R.

Iron (2.4.9)

Maximum 10 ppm calculated with reference to sodium lactate.

Dilute 5 mL of solution S to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 10 ppm calculated with reference to sodium lactate.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Bacterial endotoxins (2.6.14)

Less than 5 IU/g if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Dissolve a quantity of the substance to be examined corresponding to 75.0 mg of sodium lactate in a mixture of 10 mL of glacial acetic acid R and 20 mL of acetic anhydride R. Allow to stand for 15 min. Add 1 mL of naphtholbenzenesulfonic acid R and titrate with 0.1 M perchloric acid. 1 mL of 0.1 M perchloric acid is equivalent to 11.21 mg of C₃H₅NaO₃.

(S)-enantiomer

Transfer a quantity of the substance to be examined corresponding to 2.50 g of sodium lactate into a 50 mL volumetric flask, dilute with about 30 mL of water R and add 5.0 g of ammonium molybdate R. Dissolve and dilute with water R to 50.0 mL. Measure the angle of optical rotation (2.2.7). Calculate the percentage content of (S)-enantiomer using the expression:

$$50 + \left(24.04 \times \alpha \times \frac{m}{5.0} \times \frac{c}{50} \right)$$

α = angle of optical rotation (absolute value),
 m = mass of the substance to be examined, in grams,
 c = percentage content of C₃H₅NaO₃ in the substance to be examined.

The complex of sodium (S)-lactate formed under these test conditions is levorotatory.

LABELLING
The label states:
— where applicable, that the substance is suitable for use in the manufacture of dialysis, haemodialysis and haemofiltration solutions,
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations,
— the declared content of sodium lactate.

Ph Eur



Sodium Dodecyl Sulfate; Sodium Lauryl Sulphate
(Sodium Laurylsulfate, Ph Eur monograph 0098)

Action and use
Antionic emulsifying agent.

Ph Eur

DEFINITION
Mixture of sodium alkyl sulfates consisting chiefly of sodium dodecyl sulfate ($C_{12}H_{25}NaO_4S$; 288.4).

Content
— sodium alkyl sulfates: minimum 85.0 per cent, expressed as $C_{12}H_{25}NaO_4S$.

CHARACTERS

Appearance
White or pale yellow, powder or crystals.

Solubility

Freely soluble in water giving an opalescent solution, partly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Dissolve 0.1 g in 10 mL of water R and shake. A copious foam is formed.

B. To 0.1 mL of the solution prepared for identification test A, add 0.1 mL of a 1 g/L solution of methylene blue R and 2 mL of dilute sulfuric acid R. Add 2 mL of methylene chloride R and shake. An intense blue colour develops in the methylene chloride layer.

C. Mix about 10 mg with 10 mL of anhydrous ethanol R.

Heat to boiling on a water-bath, shaking frequently. Filter immediately and evaporate the ethanol. Dissolve the residue in 8 mL of water R, add 3 mL of dilute hydrochloric acid R, evaporate the solution to half its volume and allow to cool. Separate the congealed fatty alcohols by filtration. To the filtrate add 1 mL of barium chloride solution R1. A white, crystalline precipitate is formed.

D. Ignite 0.5 g. The residue gives reaction (a) of sodium (2.3.1).

TESTS

Alkalinity

Dissolve 1.0 g in 100 mL of carbon dioxide-free water R and add 0.1 mL of phenol red solution R. Not more than 0.5 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator.

Non-esterified alcohols

Maximum 4 per cent.

Dissolve 10 g in 100 mL of water R, add 100 mL of ethanol (96 per cent) R and shake the solution with 3 quantities, each of 50 mL, of pentane R, adding sodium chloride R, if necessary, to promote separation of the 2 layers. Wash the combined organic layers with 3 quantities, each of 50 mL, of

Sodium Metabisulfite

Ph Eur



Sodium Metabisulfite; Sodium Pyrosulphite
(Ph. Eur. monograph 0849)

$Na_2S_2O_5$

190.1

7681-57-4

Action and use
Antioxidant.

Ph Eur

DEFINITION

Sodium metabisulfite also called sodium disulfite.

Content

95.0 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, slightly soluble in ethanol

IDENTIFICATION

A. pH (see Tests).

B. To 0.4 mL of iodinated potassium iodide solution R add 8 mL of distilled water R and 1 mL of solution S diluted 1 to

ASSAY
Dissolve 1.15 g in water R, warming if necessary, and dilute to 1000.0 mL with the same solvent. To 20.0 mL of the solution add 15 mL of chloroform R and 10 mL of dimidium bromide-sulfan blue mixed solution R. Titrate with 0.004 M benzethonium chloride, shaking vigorously and allowing the layers to separate before each addition, until the pink colour of the chloroform layer is completely discharged and a greyish-blue colour is obtained.
1 mL of 0.004 M benzethonium chloride is equivalent to 1.154 mg of sodium alkyl sulfates, expressed as $C_{12}H_{25}NaO_4S$.

ASSAY
Dissolve 0.500 g in 20 mL of water R, warming gently if necessary, then add 1 mL of a 0.5 g/L solution of dihydrozone R1 in acetone R. If the solution is red, add 1 M nitric acid, dropwise, until the solution becomes bluish-green. Add 2.0 mL of dichloroacetic acid solution R and 80 mL of acetone R. Titrate with 0.01 M lead nitrate until a persistent violet-red or orange-red colour is obtained. Carry out a blank titration.
1 mL of 0.01 M lead nitrate is equivalent to 1.420 mg of Na_2SO_4 .

ASSAY
Dissolve 0.500 g in 20 mL of water R, warming gently if necessary, then add 1 mL of a 0.5 g/L solution of dihydrozone R1 in acetone R. If the solution is red, add 1 M nitric acid, dropwise, until the solution becomes bluish-green. Add 2.0 mL of dichloroacetic acid solution R and 80 mL of acetone R. Titrate with 0.01 M lead nitrate until a persistent violet-red or orange-red colour is obtained. Carry out a blank titration.
1 mL of 0.01 M lead nitrate is equivalent to 1.420 mg of Na_2SO_4 .

Sodium chloride and sodium sulfate
Maximum 8.0 per cent for the total percentage content.
Sodium chloride Dissolve 5.00 g in 50 mL of water R, add dilute nitric acid R dropwise until the solution is neutral to blue litmus paper R, add 2 mL of potassium chromate solution R and titrate with 0.1 M silver nitrate.
1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of NaCl.

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Dissolve 0.5 g in 50 mL of water R. Immediately add 5 mL of hydrochloric acid R1. Filter and wash the precipitate with water R. Dry in vacuo at 80 °C for 2 h. The precipitate melts (2.2.14) at 125 °C to 128 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Precipitate obtained in identification test A.

Comparison methyl parahydroxybenzoate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g of the substance to be examined in 10 mL of water R. Immediately add 2 mL of hydrochloric acid R and shake with 50 mL of 1,1-dimethylethyl methyl ether R. Evaporate the upper layer to dryness and take up the residue with 10 mL of acetone R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with acetone R.

Reference solution (a) Dissolve 10 mg of methyl parahydroxybenzoate CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of ethyl parahydroxybenzoate CRS in 1 mL of test solution (a) and dilute to 10 mL with acetone R.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase glacial acetic acid R, water R, methanol R (1:30:70 V/V/V).

Application 5 µL of test solution (b) and reference solutions (a) and (b).

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 1 mL of solution S (see Tests) add 1 mL of water R. The solution gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S examined immediately after preparation is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)

Dilute 1 mL of solution S to 100 mL with carbon dioxide-free water R. The pH of the solution is 9.5 to 10.5.

Related substances

Liquid chromatography (2.2.29).

10 in distilled water R. The solution is colourless and gives reaction (a) of sulfates (2.3.1).

C. Solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

3.5 to 5.0 for solution S.

Thiosulfates

To 5 mL of solution S add 5 mL of dilute hydrochloric acid R. The solution remains clear (2.2.1) for at least 15 min.

Arsenic (2.4.2, Method A)

Maximum 5 ppm.

Mix 0.20 g with 2 mL of water R in a dish. Add, drop by drop, 1.5 mL of nitric acid R. Evaporate the mixture to dryness on a water-bath. Heat over a flame until no more vapour is evolved. Take up the residue in 25 mL of water R.

Iron (2.4.9)

Maximum 20 ppm, determined on solution S.

Heavy metals (2.4.8)

Maximum 20 ppm.

To 40 mL of solution S in a silica crucible, add 10 mL of hydrochloric acid R and evaporate to dryness. Dissolve the residue in 19 mL of water R and add 1 mL of a 40 g/L solution of sodium fluoride R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

ASSAY

Dissolve 0.200 g in 50.0 mL of 0.05 M iodine and add 5 mL of dilute hydrochloric acid R. Titrate the excess of iodine with 0.1 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator.

1 mL of 0.05 M iodine is equivalent to 4.753 mg of Na₂S₂O₅.

STORAGE

Protected from light.

Ph Eur

(Sodium Methyl Parahydroxybenzoate, Ph Eur monograph 1262)

Sodium Methylparaben

Antimicrobial preservative.

Ph Eur

174.1

5026-62-0

C₈H₇NaO₃

Sodium 4-(methoxycarbonyl)phenolate.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

Ph Eur

DEFINITION

Ph Eur

Ph Eur

Ph Eur

Ph Eur

Ph Eur

Ph Eur

Ph Eur

Ph Eur

Ph Eur

Ph Eur

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Ph Eur

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of 4-hydroxybenzoic acid R (impurity A) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 50.0 mg of methyl parahydroxybenzoate CRS in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

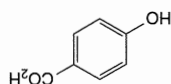
Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;
 — stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).
 Mobile phase 6.8 g/L solution of potassium dihydrogen phosphate R, methanol R (35:65 V/V).
 Flow rate 1.3 mL/min.
 Detection Spectrophotometer at 272 nm.
 Injection 10 μ L of the test solution and reference solutions (a) and (c).

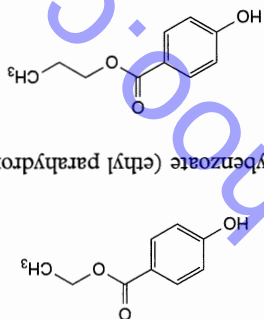
Run time 5 times the retention time of methyl parahydroxybenzoate.
 Relative retention With reference to methyl parahydroxybenzoate (retention time = about 2.3 min):
 impurity A = about 0.6.
 System suitability: reference solution (a):
 — resolution: minimum 2.0 between the peaks due to impurity A and methyl parahydroxybenzoate.
Limits:
 — correction factor: for the calculation of content, multiply the peak area of impurity A by 1.4;
 — impurity A: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (c) (3.0 per cent);
 — unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
 — sum of impurities other than A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);
 — disintegration limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Chlorides (2.4.4)

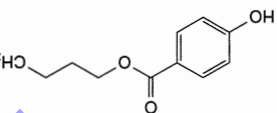
Maximum 350 ppm.
 To 10 mL of solution S, add 30 mL of water R and 1 mL of nitric acid R and dilute to 50 mL with water R. Shake and filter. Dilute 10 mL of the filtrate to 15 mL with water R. Prepare the standard using 14 mL of chloride standard solution (5 ppm Cl) R to which 1 mL of water R has been added.
Sulfates (2.4.13)
 Maximum 300 ppm.
 To 25 mL of solution S, add 5 mL of distilled water R and 10 mL of hydrochloric acid R and dilute to 50 mL with distilled water R. Shake and filter. Dilute 10 mL of the filtrate to 15 mL with distilled water R.



A. 4-hydroxybenzoic acid,



B. ethyl 4-hydroxybenzoate (ethyl parahydroxybenzoate),



C. propyl 4-hydroxybenzoate (propyl parahydroxybenzoate),

D. butyl 4-hydroxybenzoate (butyl parahydroxybenzoate),

Specified impurities A.
 Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.

IMPURITIES

In an airtight container.

STORAGE

multiply by a correction factor of 1.145.
 declared content of methyl parahydroxybenzoate CRS and Calculate the percentage content of $C_8H_7NaO_3$ using the Injection Test solution and reference solution (b).
 related substances with the following modification.
 Liquid chromatography (2.2.29) as described in the test for

ASSAY

Maximum 5.0 per cent, determined on 0.500 g.

Water (2.5.12)

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Heavy metals (2.4.8)

Maximum 10 ppm.

Sodium Molybdate Dihydrate



(Ph. Eur. monograph 1565)

$\text{MoNa}_2\text{O}_4 \cdot 2\text{H}_2\text{O}$

241.9

10102-40-6

Ph Eur

DEFINITION

Content

98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder or colourless crystals.

Solubility

Freely soluble in water.

IDENTIFICATION

A. Loss on drying (see Tests).

B. Dissolve 0.2 g in 5 mL of a mixture of equal volumes of nitric acid R and water R and add 0.1 g of ammonium chloride R. Add 0.3 mL of disodium hydrogen phosphate solution R and heat slowly at 50–60 °C. A yellow precipitate is formed.

C. Dissolve 0.15 g in 2 mL of water R, the solution gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Chlorides

Maximum 50 ppm.

To 10 mL of a mixture of equal volumes of nitric acid R and water R add 10 mL of solution S with shaking. Add 1 mL of 0.1 M silver nitrate. Any opalescence in the solution is not more intense after 5 min than that of a standard solution prepared at the same time in the same manner with 2 mL of chloride standard solution (50 ppm Cl) R.

Phosphates

Maximum 200 ppm.

Dissolve 2.0 g by heating in 13 mL of water R. In the still-hot solution, dissolve 8.0 g of ammonium nitrate R1. Add this solution to 27 mL of a mixture of equal volumes of nitric acid R and water R. Any yellow colour or opalescence in the solution is not more intense within 3 h than that in a standard solution prepared at the same time in the same manner as follows: dissolve 1.0 g in 12 mL of water R and add 1 mL of phosphate standard solution (200 ppm PO_4) R.

Maximum 10 ppm, determined on 0.10 g.

Ammonium (2.4.1, Method B)

Prepare the standard using 1 mL of ammonium standard solution (1 ppm NH_4) R.

Heavy metals

Maximum 10 ppm.

To 10 mL of solution S, add 2 mL of water R, 6 mL of a 168 g/L solution of sodium hydroxide R and 2 mL of concentrated ammonia R (solution A). To 0.5 mL of thioacetamide reagent R add a mixture of 15 mL of solution A and 5 mL of water R. Any coloration of the solution is not more intense after 2 min than that of a reference solution prepared at the same time as follows: to 0.5 mL of thioacetamide reagent R add a mixture of 5 mL of solution A, prepared at the same time as follows: to 0.5 mL of

Sodium Nitrite

(Ph. Eur. monograph 1996)

NaNO_2

69.0

7632-00-0



Action and use

Used in treatment of cyanide poisoning.

Ph Eur

DEFINITION

Content

98.5 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

Colourless crystals or mass or yellowish rods, hygroscopic.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Dilute 1 mL of solution S1 (see Tests) to 25 mL with water R. To 0.1 mL of the solution add 1 mL of sulfanilic acid solution R1. Allow to stand for 2–3 min. Add 1 mL of β-naphthol solution R and 1 mL of dilute sodium hydroxide solution R. An intense red colour develops.

B. To 1 mL of the solution prepared for identification test A add 3 mL of a 20 g/L solution of phenazone R and 0.4 mL of dilute sulfuric acid R. An intense green colour develops.

C. To 0.15 mL of solution S1, add 0.35 mL of water R. The solution gives reaction (b) of sodium (2.3.1).

TESTS

Solution S1

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Solution S2


Dissolve 3 g in distilled water R. Cautiously add 10 mL of nitric acid R and evaporate to dryness. Dissolve the residue in 10 mL of distilled water R, neutralise with dilute sodium hydroxide solution R and dilute to 30 mL with distilled water R.

Appearance of solution

Solution S1 is clear (2.2.1) and not more intensely coloured than reference solution B6 (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S1, add 0.05 mL of phenol red solution R. Add 0.1 mL of 0.01 M sodium hydroxide. The solution is red. Add 0.3 mL of 0.01 M hydrochloric acid. The solution is yellow.

<p>Absorption maximum At 395 nm.</p> <p>Shoulder At about 510 nm.</p> <p>Absorption minimum At 370 nm.</p> <p>Specific absorbance at the absorption maximum 0.65 to 0.80.</p> <p>B. Dissolve about 20 mg in 2 mL of water R and add 0.1 mL of sodium sulfide solution R. A deep violet-red colour is produced.</p> <p>C. Dissolve 50 mg in 1 mL of water R and acidify the solution by the addition of hydrochloric acid R. Place a drop of the solution in an oxidising flame. A persistent yellow colour is produced.</p> <p>TESTS</p> <p>Insoluble matter</p> <p>Maximum 100 ppm.</p> <p>Dissolve 10 g without heating in 50 mL of water R. Allow to stand for 30 min and filter through a sintered-glass filter (16) (2.1.2). Wash the filter with cold water R until the filtrate is colourless. Dry the residue on the filter at 105 °C.</p> <p>The residue weighs a maximum of 1 mg.</p> <p>Chlorides (2.4.4)</p> <p>Maximum 200 ppm.</p> <p>In a metallic crucible (nickel) mix 1.0 g with 8 mL of a 200 g/L solution of sodium hydroxide R. Heat slowly and evaporate carefully to dryness over a small flame, then heat to a dull red colour for 30 min. Allow to cool and transfer the solid residue with 3 quantities, each of 8 mL, of dilute sulfuric acid R. Filter the sulfuric acid extracts on a filter paper and collect the filtrates. Render the filtrate acid to litmus paper R by adding, if necessary, a few drops of dilute sulfuric acid R. Wash the crucible and the filter paper with 3 quantities, each of 10 mL, of water R, add the washings to the main sulfuric acid solution and dilute to 60 mL with water R. Mix.</p> <p>Ferriyanides</p> <p>Maximum 200 ppm.</p> <p>Dissolve 1.25 g in acetate buffer solution pH 4.6 R and dilute to 50.0 mL with the same buffer solution. Use three 50 mL volumetric flasks (4, B, C). To flask B add 1.0 mL of ferriyanide standard solution (50 ppm Fe(CN)₆³⁻) R. To flasks A and B add 1 mL of sulfate R. To the 3 flasks add 10.0 mL of the solution of the substance to be examined. Dilute the contents of each flask to 50.0 mL with water R. Allow to stand for 30 min. The absorbance (2.2.25) of the solution in flask A measured at 720 nm using the solution in flask C as the compensation liquid is not greater than the absorbance of the solution in flask B measured at 720 nm using the solution in flask A as the compensation liquid.</p> <p>Ferrocyanides</p> <p>Maximum 200 ppm.</p> <p>Dissolve 4.0 g in water R and dilute to 100.0 mL with the same solvent. Use three 50 mL volumetric flasks (4, B, C). To flask B add 2.0 mL of ferrocyanide standard solution (100 ppm Fe(CN)₆⁴⁻) R. To flasks A and B add 1 mL of ferric chloride solution R2. To the 3 flasks add 25.0 mL of the solution of the substance to be examined. Dilute the contents of each flask to 50.0 mL with water R. Allow to stand for 30 min. The absorbance (2.2.25) of the solution in flask A measured at 695 nm using the solution in flask C as the compensation liquid is not greater than the absorbance of the solution in flask B measured at 695 nm using the solution in flask A as the compensation liquid.</p>	<p>Chlorides (2.4.4)</p> <p>Maximum 50 ppm.</p> <p>Dilute 10 mL of solution S2 to 15 mL with distilled water R.</p> <p>Heavy metals (2.4.8)</p> <p>Maximum 200 ppm.</p> <p>Dilute 7.5 mL of solution S2 to 15 mL with distilled water R.</p> <p>Sulfates (2.4.13)</p> <p>Maximum 200 ppm.</p> <p>Dilute 7.5 mL of solution S2 to 15 mL with distilled water R.</p> <p>Loss on drying (2.2.32)</p> <p>Maximum 1.0 per cent, determined on 1.000 g by drying <i>in vacuo</i>.</p> <p>ASSAY</p> <p>Dissolve 0.400 g in 100.0 mL of water R. Introduce 20.0 mL of the solution, while stirring continuously and keeping the tip of the pipette below the surface of the liquid, into a conical flask containing 30.0 mL of 0.1 M cerium sulfate. Immediately stopper the flask and allow to stand for 2 min. Add 10 mL of a 200 g/L solution of potassium iodide R and 2 mL of starch solution R.</p> <p>While stirring continuously, titrate with 0.1 M sodium thiosulfate until the blue colour disappears. Carry out a blank titration.</p> <p>1 mL of 0.1 M cerium sulfate is equivalent to 3.45 mg of NaNO₂.</p> <p>STORAGE</p> <p>In an airtight container.</p> <p>Ph Eur</p> <p></p> <p>13755-38-9</p> <p>Ph Eur</p> <p>Sodium Nitroprusside</p> <p>(Ph. Eur. monograph 0565)</p> <p>Na₂[Fe(CN)₅(NO)]·2H₂O 298.0</p> <p>Action and use</p> <p>Vasodilator.</p> <p>Preparation</p> <p>Sodium Nitroprusside Infusion</p> <p>Ph Eur</p> <p>DEFINITION</p> <p>Sodium pentacyanonitrosylferrate (III) dihydrate.</p> <p>99.0 per cent to 100.5 per cent (anhydrous substance).</p> <p>CHARACTERS</p> <p>Appearance</p> <p>Reddish-brown powder or crystals.</p> <p>Solubility</p> <p>Freely soluble in water, slightly soluble in ethanol (96 per cent).</p> <p>IDENTIFICATION</p> <p>A. Ultraviolet and visible absorption spectrophotometry (2.2.25).</p> <p>Test solution Dissolve 0.700 g in water R and dilute to 100.0 mL with the same solvent. Examine the solution immediately after preparation.</p> <p>Spectral range 350–600 nm.</p>
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Sulfates

Maximum 100 ppm.

Test solution Dissolve 3.6 g in 120 mL of distilled water R, add with mixing 4 mL of sulfate standard solution (10 ppm SO₄) R and 20 mL of a 250 g/L solution of cupric chloride R and allow to stand for 15.0 mL with distilled water R. Allow to stand for 16 h and filter or centrifuge until a clear light-blue solution is obtained.

Reference solution To 40 mL of sulfate standard solution (10 ppm SO₄) R add 80 mL of distilled water R and 12-13 mL of a 250 g/L solution of cupric chloride R. Dilute to 150.0 mL with distilled water R. The volume of cupric chloride solution added is such that the colour of the final solution matches that of the test solution.

Allow the solutions to stand. Filter both solutions, discarding the first 25 mL of filtrate. To 100 mL of each filtrate, add 0.5 mL of acetic acid R. Mix and add 2 mL of a 250 g/L solution of barium chloride R and mix again. The test solution is not more opalescent than the reference solution.

Water (2.5.12)

9.0 per cent to 15.0 per cent, determined on 0.250 g.

ASSAY

Dissolve 0.250 g in 100 mL of water R and add 0.1 mL of dilute sulfuric acid R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20) with a silver-mercurous sulfate electrode system.

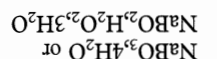
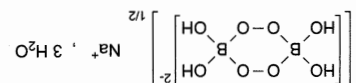
1 mL of 0.1 M silver nitrate is equivalent to 13.10 mg of Na₂[Fe(CN)₅(NO)].

STORAGE

Protected from light.

Sodium Perborate

(Hydrated Sodium Perborate, Ph Eur monograph 1997)



153.9

7632-04-4

Action and use
Antiseptic.

DEFINITION

Content
96.0 per cent to 103.0 per cent.

CHARACTERISTICS

Appearance

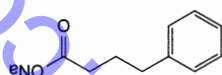
Colourless, prismatic crystals or white or almost white powder, stable in the crystalline form.

Solubility

Sparingly soluble in water, with slow decomposition. It dissolves in dilute mineral acids.

Sodium Phenylbutyrate

(Ph. Eur. monograph 2183)



C₁₀H₁₁NaO₂

186.2

1716-12-7

Action and use

Glutamine conjugate; treatment of hyperammonaemia.

DEFINITION

Sodium 4-phenylbutanoate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERISTICS

Appearance

White or yellowish-white powder.

Solubility

Freely soluble in water and in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Dissolve 20 mg in a mixture of 1 mL of dilute sulfuric acid R and 1 mL of water R. Add 1 mL of potassium iodide solution R. A reddish-brown colour appears.

B. The mixture obtained by treating about 100 mg with 0.1 mL of sulfuric acid R and 5 mL of methanol R burns with a greenish flame when ignited.

C. It gives reaction (a) of sodium (2.3.1).

TESTS

Chlorides (2.4.4)

Maximum 330 ppm.

Dissolve 0.15 g in 15 mL of water R.

Sulfates (2.4.13)

Maximum 1.2 per cent.

Dissolve 0.13 g in 150 mL of distilled water R.

Iron (2.4.9)

Maximum 20 ppm.

Dissolve 2.5 g in 10 mL of dilute hydrochloric acid R with heating, evaporate to dryness, with stirring, and dissolve the residue in 25 mL of hot water R. Dilute 5 mL of the obtained solution to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of the solution obtained in the test for iron complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

Dissolve 0.300 g in 50.0 mL of water R. Dilute 10.0 mL of the solution to 50 mL with water R and add 10 mL of dilute sulfuric acid R. Titrate with 0.02 M potassium permanganate. 1 mL of 0.02 M potassium permanganate is equivalent to 7.693 mg of NaH₈BO₇.

STORAGE

In an airtight container.

Ph Eur

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison sodium phenylbutyrate CRS.
B. Dissolve 0.15 g in 2 mL of water R. The solution gives reaction (a) of sodium (2.3.1).

TESTS

pH (2.2.3)

6.5 to 7.5.

Dissolve 0.20 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Impurity C

Gas chromatography (2.2.28).

Silylation solution To 2 mL of N₂O-

bis(trimethylsilyl)trifluoroacetamide R add 0.04 mL of

chlorotrimethylsilane R and mix.

Test solution Dissolve 50.0 mg of the substance to be

examined in 3 mL of water R and add 0.5 mL of methylene

chloride R. Extract with 2 quantities, each of 5 mL, of methylene

chloride R. Evaporate the combined methylene chloride

extracts to dryness in a vial with a screw cap and add 0.5 mL

of the silylation solution. Seal the vial and heat at 70 ± 5 °C

for 20 min.

Reference solution (a) Dissolve 5.0 mg of sodium phenylbutyrate

impurity C CRS in methylene chloride R and dilute to 10.0 mL

with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a)

to 10.0 mL with methylene chloride R. Place 1.0 mL of this

solution in a vial with a screw cap, evaporate to dryness and

add 0.5 mL of the silylation solution. Seal the vial and heat

at 70 ± 5 °C for 20 min.

Reference solution (c) Dissolve 10 mg of the substance to be

examined in 25 mL of water R. To 3 mL of this solution add

0.1 mL of hydrochloric acid R. Extract with 2 quantities, each

of 5 mL, of methylene chloride R. Combine the methylene

chloride extracts in a vial with a screw cap and add 2 mL of

reference solution (a). Evaporate to dryness and add 0.5 mL

of the silylation solution. Seal the vial and heat at 70 ± 5 °C

for 20 min.

Column:

— material: fused silica;

— size: l = 25 m, Ø = 0.25 mm;

— stationary phase: poly(dimethyl) (diphenyl) siloxane R (film

thickness 1.0 µm).

Carrier gas helium for chromatography R.

Flow rate 0.9 mL/min.

Split ratio 1:100.

Temperature:

Time (min)	Temperature (°C)
0 - 5	50
5 - 27	50 → 270
27 - 32	270
Injection port	270
Detector	270

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to phenylbutyrate (retention time = about 20 min): impurity C = about 0.98.

System suitability: reference solution (c):
— resolution: minimum 3.0 between the peaks due to
impurity C and phenylbutyrate.
Limit:
— impurity C: not more than the area of the corresponding
peak in the chromatogram obtained with reference
solution (b) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.20 g of the substance to be examined

in 10 mL of methanol R and dilute to 50.0 mL with water R.

Reference solution (a) Dissolve 4.0 mg of α-tetralone R

(impurity B) in 10 mL of methanol R and dilute to 200.0 mL

with the same solvent.

Reference solution (b) Dissolve 0.20 g of the substance to be

examined in 10 mL of methanol R, add 1 mL of reference

solution (a) and dilute to 50 mL with water R.

Reference solution (c) Dilute 1.0 mL of reference solution (a)

to 50.0 mL with water R.

Reference solution (d) Dissolve 5.0 mg of 3-benzoylpropionic

acid R (impurity A) in 2.5 mL of methanol R and dilute to

50.0 mL with the same solvent. Dilute 1.0 mL of this

solution to 50.0 mL with water R.

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: base-deactivated end-capped octadecylsilyl

silica gel for chromatography R (5 µm).

Mobile phase: glacial acetic acid R, methanol R, water R

(1:49:50 V/V/V).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 245 nm.

Injection 20 µL of the test solution and reference

solutions (b), (c) and (d).

Run time Twice the retention time of phenylbutyrate.

Relative retention With reference to phenylbutyrate (retention

time = about 17 min): impurity A = about 0.3;

impurity B = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 6 between the peaks due to

impurity B and phenylbutyrate.

Limits:

— impurity A: not more than twice the area of the

corresponding peak in the chromatogram obtained with

reference solution (d) (0.1 per cent);

— impurity B: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (c) (0.01 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (d) (0.05 per cent);

— total: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (d)

(0.1 per cent);

— disregard limit of impurities other than B: 0.6 times the area

of the principal peak in the chromatogram obtained with

reference solution (d) (0.03 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in a mixture of 25 volumes of water R and

75 volumes of ethanol (96 per cent) R and dilute to 20 mL

with the same mixture of solvents. 12 mL of the solution

complies with test B. Prepare the reference solution using

lead standard solution (1 ppm Pb) obtained by diluting lead

standard solution (100 ppm Pb) R with a mixture of 25 volumes of water R and 75 volumes of ethanol (96 per cent) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

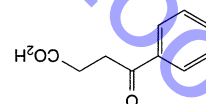
ASSAY

Disperse 0.150 g in 50 mL of anhydrous acetic acid R. The opalescence of the solution disappears during the titration. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

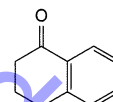
1 mL of 0.1 M perchloric acid is equivalent to 18.62 mg of $C_{18}H_{13}NNa_2O_8 \cdot H_2O$.

IMPURITIES

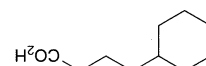
Specified impurities A, B, C.



A. 4-oxo-4-phenylbutanoic acid (3-benzoylpropionic acid),



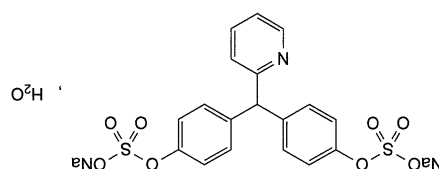
B. 3,4-dihydronaphthalen-1(2H)-one (α -tetralone),



C. 4-cyclohexylbutanoic acid.

Sodium Picosulfate

(Ph. Eur. monograph 1031)



$C_{18}H_{13}NNa_2O_8 \cdot H_2O$ 499.4

10040-45-6

(anhydrous)

Action and use

Stimulant laxative.

Preparations

Compound Sodium Picosulfate Powder for Oral Solution

Sodium Picosulfate Oral Solution

Ph. Eur.

DEFINITION

4,4'-[(Pyridin-2-yl)methylene]diphenyl bis(sodium sulfate) monohydrate.

Content

98.5 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, D.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium picosulfate CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution Dissolve 20 mg of sodium picosulfate CRS in methanol R and dilute to 5 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase anhydrous formic acid R, water R, methanol R, ethyl acetate R (2.5:12.5:25:60 V/V/V/V).

Application 5 μ L.

Development Over 1/2 of the plate.

Drying In a current of warm air for 15 min.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

C. To 5 mL of solution S (see Tests) add 1 mL of dilute hydrochloric acid R and heat to boiling. Add 1 mL of barium chloride solution R1. A white precipitate is formed.

D. Solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution GY₇ (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.05 mL of phenolphthalein solution R. The solution is colourless. Not more than 0.25 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to pink.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 10.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of picosulfate for system suitability CRS (containing impurities A and B) in 1.0 mL of the mobile phase.

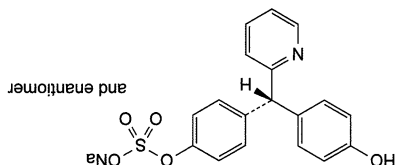
Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

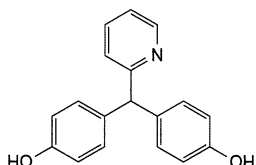
— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R1 (5 μ m);

— temperature: 40 °C.

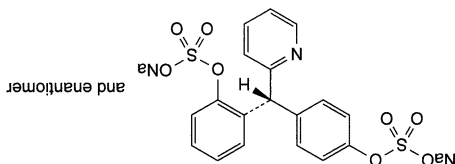
impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C.



A. 4-[(R,S)-(4-hydroxyphenyl)(pyridin-2-yl)methyl]phenyl sodium sulfate,



B. 4,4'-[(pyridin-2-yl)methylene]diphenol,



C. 2-[(R,S)-(pyridin-2-yl)[4-(sulfonatooxy)phenyl]methyl]phenyl disodium sulfate.

Ph Eur



Sodium Polystyrene Sulfonate

Sodium Polystyrene Sulfonate
(Ph. Eur. monograph 1909)

Action and use

Used in the treatment of hyperkalaemia.

Ph Eur

DEFINITION

Polystyrene sulfonate resin prepared in the sodium form. Exchange capacity 2.8 mmol to 3.4 mmol of potassium per gram (dried substance).

Content

9.4 per cent to 11.0 per cent of Na (dried substance).

CHARACTERS

Appearance

Almost white or light brown powder.

Solubility

Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs using finely ground substance. Comparison Ph. Eur. reference spectrum of sodium polystyrene sulfonate.

B. Suspend 0.1 g in water R, add 2 mL of a 150 g/L solution of potassium carbonate R, and heat to boiling. Allow to cool and filter. To the filtrate add 4 mL of potassium

Mobile phase Dissolve 2.3 g of disodium hydrogen phosphate dihydrate R in 800 mL of water for chromatography R, add 0.2 g of cetyltrimethylammonium bromide R, adjust to pH 7.5 with phosphoric acid R and dilute to 1000 mL with water for chromatography R; mix 550 mL of this solution and 450 mL of acetonitrile R (if necessary vary the buffer/acetonitrile proportion in 10 mL increments in order to fulfil the resolution requirement). Flow rate 1.0 mL/min. Detection Spectrophotometer at 263 nm. Injection 40 µL.

Run time Twice the retention time of picosulfate. Identification of impurities Use the chromatogram supplied with picosulfate for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and B. Relative retention With reference to picosulfate (retention time = about 7.4 min): impurity B = about 0.5; impurity A = about 0.7. System suitability: reference solution (b): resolution: minimum 4.0 between the peaks due to impurities B and A. Limits: correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.7; impurity B = 0.5; impurities A, B: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent); unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent); total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent); disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 400 ppm.

Dilute 7.5 mL of solution S to 15 mL with distilled water R.

Water (2.5.12)

3.0 per cent to 5.0 per cent, determined on 0.500 g.

ASSAY

Dissolve 0.400 g in 80 mL of methanol R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 48.14 mg of $C_{18}H_{13}NNa_2O_8S_2$.

IMPURITIES

Specified impurities A, B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

pyroantimonate solution R and heat to boiling. Allow to cool in iced water and if necessary rub the inside of the test-tube with a glass rod. A dense white precipitate is formed.

TESTS

Styrene

Liquid chromatography (2.2.29).

Test solution Shake 10.0 g of the substance to be examined with 10 mL of acetone R for 30 min, centrifuge and use the supernatant.

Reference solution Dissolve 10 mg of styrene R in acetone R and dilute to 100 mL with the same solvent. Dilute 1 mL of this solution to 100 mL with acetone R.

Column:

— size: $l = 0.25$ m, $\phi = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R, water R (1:1 V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Limit:

— styrene: not more than the area of the principal peak in the chromatogram obtained with the reference solution (1 ppm).

Calcium

Maximum 0.10 per cent.

Atomic emission spectrometry (2.2.22, Method D).

Test solution To 1.10 g add 5 mL of hydrochloric acid R, heat to boiling, cool and add 10 mL of water R. Filter, wash the filter and residue with water R and dilute the filtrate and washing to 25.0 mL with water R.

Reference solutions Prepare the reference solutions using calcium standard solution (400 ppm Ca) R, diluted as necessary with water R.

Wavelength 422.7 nm.

Potassium

Maximum 0.10 per cent.

Atomic emission spectrometry (2.2.22, Method D).

Test solution To 1.10 g add 5 mL of hydrochloric acid R, heat to boiling, cool and add 10 mL of water R. Filter, wash the filter and residue with water R and dilute the filtrate and washings to 25.0 mL with water R.

Reference solutions Prepare the reference solutions using potassium standard solution (100 ppm K) R, diluted as necessary with water R.

Wavelength 766.5 nm.

Heavy metals (2.4.8)

Maximum 10 ppm.

Treat 1.0 g as described in test F. After the addition of the buffer solution pH 3.5 R and of the thioacetamide reagent R, dilute to 50 mL with water R and continue as described in test E, beginning at the words "mix and allow to stand for 10 min...".

Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 7.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Microbial contamination (2.6.13)

Bile-tolerant gram-negative bacteria: acceptance criterion less than 10^2 CFU/g.

ASSAY

Sodium

Atomic emission spectrometry (2.2.22, Method D).

Test solution In a platinum crucible moisten 0.90 g with a few drops of sulfuric acid R, ignite very gently and allow to cool. Moistens with a few drops of sulfuric acid R again, ignite at 800 \pm 50 °C until a carbon-free ash is obtained and allow to cool.

Add 20 mL of water R to the crucible, warm gently on a water-bath until dissolution, cool, transfer quantitatively to a 100 mL graduated flask and dilute to 100.0 mL with water R. Dilute 5 mL of this solution to 100.0 mL with water R.

Reference solutions Prepare the reference solutions using sodium standard solution (200 ppm Na) R, diluted as necessary with water R.

Wavelength 589 nm.

Exchange capacity

Atomic emission spectrometry (2.2.22, Method D).

Solution A 9.533 g/L solution of potassium chloride R.

Test solution To 1.6 g of the substance to be examined in a dry 250 mL ground-glass-stoppered flask add 100 mL of solution A, stopper and shake for 15 min. Filter, discard the first 20 mL of the filtrate and dilute 4 mL of the filtrate to 1000 mL with water R.

Reference solutions Prepare the reference solutions by diluting 0, 1, 2, 3 and 4 mL of solution A respectively and 4, 3, 2, 1 and 0 mL of a 7.63 g/L solution of sodium chloride R to 1000 mL with water R.

Wavelength 766.5 nm. Prepare a calibration curve using the reference solutions and calculate the potassium exchange capacity of the substance to be examined in millimoles per gram taking the concentration of potassium in solution A as 128 mmol/L.

STORAGE

In an airtight container.

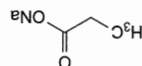
IMPURITIES

Specified impurities A

A. styrene.

Sodium Propionate

(Ph. Eur. monograph 2041)



96.1

$C_5H_9NaO_2$

Action and use

Antifungal.

DEFINITION

Sodium propionate.

Content

99.0 per cent to 101.0 per cent (dried substance).

137-40-6

Ph Eur



CHARACTERS

Appearance
Colourless crystals or, white or almost white powder, slightly hygroscopic.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: A, D.

Second identification: B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of sodium propionate.

B. Dissolve 0.1 g in a mixture of 2 mL of copper sulfate

solution R and 2 mL of methylene chloride R. Shake vigorously and allow to stand. Both the upper and the lower layer show a blue colour.

C. To 5 mL of solution S (see Tests) add 2 mL of 0.1 M silver nitrate. A white precipitate is formed.

D. Solution S gives reaction (a) of sodium (2.3.1).

TESTS**Solution S**

Dissolve 10 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

7.8 to 9.2.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.250 g of the substance to be examined in water R and dilute to 100 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of the substance to be examined and 10 mg of sodium acetate R in water R and dilute to 100 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100 mL with water R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dilute 1 mL of phosphoric acid R to 1000 mL with water R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L.

System suitability: reference solution (a):

— resolution: minimum 5 between the peaks due to sodium acetate and sodium propionate.

Limits:

— any impurity: not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent),

— total: not more than half the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent),

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

IDENTIFICATION

First identification: B, D.

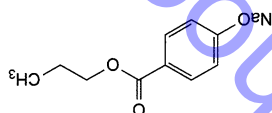
Second identification: A, C, D.

A. Dissolve 0.5 g in 50 mL of water R. Immediately add 5 mL of hydrochloric acid R1. Filter and wash the precipitate

Sodium Propyl Hydroxybenzoate

Sodium Propylparaben

(Sodium Propyl Parahydroxybenzoate, Ph. Eur monograph 1263)



$C_{10}H_{11}NaO_3$

202.2

35285-69-9

Action and use

Antimicrobial preservative.

DEFINITION

Sodium 4-(propoxycarbonyl)phenolate.

Content

94.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, hygroscopic, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification: B, D.

Second identification: A, C, D.

A. Dissolve 0.5 g in 50 mL of water R. Immediately add 5 mL of hydrochloric acid R1. Filter and wash the precipitate

STORAGE

In an airtight container.

Ph. Eur.

with water R. Dry in *vacuo* at 80 °C for 2 h. The precipitate melts (2.2.14) at 96 °C to 99 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Precipitate obtained in identification test A.

Comparison propyl parahydroxybenzoate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g of the substance to be examined in 10 mL of water R. Immediately add 2 mL of hydrochloric acid R and shake with 50 mL of 1,1-dimethylethyl methyl ether R. Evaporate the upper layer to dryness and take up the residue with 10 mL of acetone R.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with acetone R.

Reference solution (a) Dissolve 10 mg of propyl parahydroxybenzoate CRS in acetone R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of ethyl parahydroxybenzoate CRS in 1 mL of test solution (a) and dilute to 10 mL with acetone R.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase glacial acetic acid R, water R, methanol R (1:30:70 V/V/V).

Application 5 µL of test solution (b) and reference solutions (a) and (b).

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To 1 mL of solution S (see Tests) add 1 mL of water R. The solution gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in carbon dioxide-free water R prepared from distilled water R, and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S, examined immediately after preparation, is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

pH (2.2.3)

9.5 to 10.5.

Dilute 1 mL of solution S to 100 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 2.5 mL of methanol R and dilute to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of ethyl parahydroxybenzoate CRS (impurity C), 5 mg of 4-hydroxybenzoic acid R (impurity A) and 5 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 50.0 mg of propyl parahydroxybenzoate CRS in 2.5 mL of methanol R and dilute

to 50.0 mL with the mobile phase. Dilute 10.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of the test solution to 20.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column: size: $l = 0.15$ m, $\phi = 4.6$ mm; stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase 6.8 g/L solution of potassium dihydrogen phosphate R, methanol R (35:65 V/V).

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 10 µL of the test solution and reference solutions (a) and (c).

Run time 2.5 times the retention time of propyl parahydroxybenzoate.

Relative retention With reference to propyl parahydroxybenzoate (retention time = about 4.5 min):

impurity A = about 0.3; impurity C = about 0.7.

System suitability: reference solution (a):

— resolution: minimum 5.0 between the peaks due to impurity C and propyl parahydroxybenzoate.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity A by 1.4;

— impurity A: not more than 8 times the area of the principal peak in the chromatogram obtained with reference solution (c) (4.0 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

— sum of impurities other than A: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Chlorides (2.4.4)

Maximum 350 ppm.

To 10 mL of solution S, add 1 mL of nitric acid R and 30 mL of water R and dilute to 50 mL with water R. Shake and filter. Dilute 10 mL of the filtrate to 15 mL with water R. Prepare the standard using 14 mL of chloride standard solution (5 ppm Cl) R to which 1 mL of water R has been added.

Sulfates (2.4.13)

Maximum 300 ppm.

To 25 mL of solution S, add 5 mL of distilled water R and 10 mL of hydrochloric acid R and dilute to 50 mL with distilled water R. Shake and filter. Dilute 10 mL of the filtrate to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.500 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{10}H_{11}NaO_3$ from the declared content of *propyl parahydroxybenzoate CRS*, multiplied by a correction factor of 1.122.

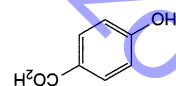
STORAGE

In an airtight container.

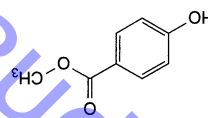
IMPURITIES

Specified impurities A.

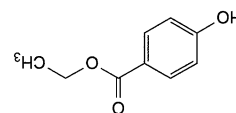
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.



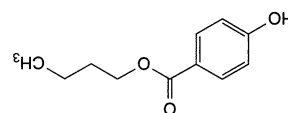
A. 4-hydroxybenzoic acid,



B. methyl 4-hydroxybenzoate (methyl parahydroxybenzoate),



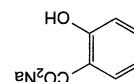
C. ethyl 4-hydroxybenzoate (ethyl parahydroxybenzoate),



D. butyl 4-hydroxybenzoate (butyl parahydroxybenzoate).

Sodium Salicylate

(Ph. Eur. monograph 0413)



$C_7H_5NaO_3$

160.1

54-21-7

Action and use

Anti-inflammatory; analgesic.

DEFINITION

Sodium 2-hydroxybenzenecarboxylate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or small, colourless crystals or shiny flakes.

Solubility

Freely soluble in water, sparingly soluble in ethanol

IDENTIFICATION

First identification A, C
Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium salicylate CRS.

B. Solution S (see Tests) gives the reactions of salicylates (2.3.1).

C. It gives reaction (b) of sodium (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Acidity

To 20 mL of solution S add 0.1 mL of phenol red solution R. The solution is yellow. Not more than 2.0 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to violet-red.

Chlorides (2.4.4)

Maximum 200 ppm.

To 5 mL of solution S add 5 mL of water R and 10 mL of dilute nitric acid R and filter. Dilute 10 mL of the filtrate to

15 mL with water R.

Sulfates (2.4.13)

Maximum 600 ppm.

Dilute 2.5 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.6 g in 16 mL of a mixture of 5 volumes of water R

and 10 volumes of ethanol (96 per cent) R. 12 mL of the solution complies with test B. Prepare the reference solution

using lead standard solution (2 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with a mixture of

5 volumes of water R and 10 volumes of ethanol

(96 per cent) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.00 g by drying in an oven at 105 °C.

ASSAY

Dissolve 0.130 g in 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 16.01 mg of $C_7H_5NaO_3$.

STORAGE

In an airtight container, protected from light.

Ph Eur

Sodium Selenite Pentahydrate



(Ph. Eur. monograph 1677)

$\text{Na}_2\text{SeO}_3 \cdot 5\text{H}_2\text{O}$

263.0

26970-82-1

Action and use

Used in treatment of selenium deficiency.

DEFINITION

Content

98.5 per cent to 101.5 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder, hygroscopic.

Solubility

Freely soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. Dissolve 50 mg in 5 mL of a mixture of equal volumes of

dilute hydrochloric acid R and water R and heat to boiling.

Add 0.05 g of ascorbic acid R; a red precipitate is formed

which may become black.

B. Dissolve 50 mg in a mixture of 1 mL of dilute hydrochloric

acid R and 5 mL of water R. Add 1 mL of barium chloride

solution R1; the solution remains clear.

C. It gives reaction (a) of sodium (2.3.1).

D. It complies with the limits of the assay.

TESTS

Solution S

Dissolve 5.0 g in carbon dioxide-free water R and dilute to

50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

9.8 to 10.8 for solution S.

Chlorides (2.4.4)

Maximum 50 ppm.

To 10 mL of solution S add 2 mL of nitric acid R and dilute

to 15 mL with water R.

Sulfates and selenates (2.4.13)

Maximum 300 ppm (determined as sulfates).

Dissolve 0.5 g in 10 mL of distilled water R. Add 0.5 mL of

hydrochloric acid R1 and dilute to 15 mL with distilled water R.

Iron

Maximum 50 ppm.

To 2 mL of solution S add 2 mL of a 200 g/L solution of

sulfosalicic acid R, 5 mL of concentrated ammonia R and

dilute to 10 mL with water R. The solution is not more

intensely coloured than a reference solution prepared in the

same manner using 1 mL of iron standard solution

ASSAY

Dissolve 0.120 g in 50 mL of water R, add 7 mL of glacial

acetic acid R, 25.0 mL of 0.1 M sodium thiosulfate and 0.5 g of

potassium iodide R. Titrate immediately with 0.05 M iodine

using starch solution R as indicator.

1 mL of 0.1 M sodium thiosulfate is equivalent to 6.575 mg of

$\text{Na}_2\text{SeO}_3 \cdot 5\text{H}_2\text{O}$.

STORAGE

In an airtight container.

Ph Eur

Sodium Starch Glycolate

(Type A)⁽¹⁾

(Ph. Eur. monograph 0983)

Action and use

Excipient.

DEFINITION

Sodium salt of a cross-linked partly O-carboxymethylated

starch.

Content

2.8 per cent to 4.2 per cent of Na (4.22.99) (substance

washed with ethanol (80 per cent V/V) and dried).

◆ CHARACTERS

Appearance

White or almost white, fine, free-flowing powder, very

hygroscopic.

Solubility

Practically insoluble in methylene chloride. It gives a

translucent suspension in water. ◆

IDENTIFICATION

◆ A. pH (see Tests).

B. Prepare with shaking and without heating a mixture of

4.0 g of the substance to be examined and 20 mL of carbon

dioxide-free water R. The mixture has the appearance of a gel.

Add 100 mL of carbon dioxide-free water R and shake.

A suspension forms that settles after standing. ◆

C. To an acidified solution, add iodinated potassium iodide

solution R1. The solution becomes blue or violet.

D. Solution S2 (see Tests) gives reaction (a) of sodium

(2.3.1).

Solution S1

Centrifuge the suspension obtained in identification test B at

2500 g for 10 min. Collect carefully the supernatant.

Solution S2

Place 2.5 g in a silica or platinum crucible and add 2 mL of

a 500 g/L solution of sulfuric acid R. Heat on a water-bath,

then cautiously over a naked flame, raising the temperature

progressively, then incinerate in a muffle furnace at

600 ± 25 °C. Continue heating until all black particles have

disappeared. Allow to cool, add a few drops of dilute sulfuric

acid R, heat and incinerate as above. Allow to cool, add a few

drops of ammonium carbonate solution R, evaporate to dryness

and incinerate cautiously. Allow to cool and dissolve the

◆ Appearance of solution S1

Solution S1 is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

5.5 to 7.5.

Disperse 1.0 g in 30 mL of carbon dioxide-free water R.

Sodium glycolate

Maximum 2.0 per cent. Carry out the test protected from light.

Test solution Place 0.20 g in a beaker. Add 5 mL of acetic

acid R and 5 mL of water R. Stir until dissolution is complete

(about 10 min). Add 50 mL of acetone R and 1 g of sodium

chloride R. Filter through a fast filter paper impregnated with

acetone R, rinse the beaker and filter with acetone R. Combine

the filtrate and washings and dilute to 100.0 mL with

acetone R. Allow to stand for 24 h without shaking. Use the

clear supernatant.





Sodium Starch Glycolate (Type B)⁽¹⁾

(Ph. Eur. monograph 0984)

Action and use

Excipient.

DEFINITION

Sodium salt of a cross-linked partly O-carboxymethylated starch.

Content

2.0 per cent to 3.4 per cent of Na (M_r 22.99) (substance washed with ethanol (80 per cent V/V) and dried).

CHARACTERS

Appearance

White or almost white, fine, free-flowing powder, very hygroscopic.

Solubility

Practically insoluble in methylene chloride. It gives a translucent suspension in water. ◆

IDENTIFICATION

◆ A. pH (see Tests).

B. Prepare with shaking and without heating a mixture of 4.0 g of the substance to be examined and 20 mL of carbon dioxide-free water R. The mixture has the appearance of a gel.

Add 100 mL of carbon dioxide-free water R and shake.

◆ A suspension forms that settles after standing.

C. To an acidified solution, add iodinated potassium iodide solution R1. The solution becomes blue or violet.

D. Solution S2 (see Tests) gives reaction (a) of sodium

TESTS

Solution S1

Centrifuge the suspension obtained in identification test B at 2500 g for 10 min. Collect carefully the supernatant.

Solution S2

Place 2.5 g in a silica or platinum crucible and add 2 mL of a 500 g/L solution of sulfuric acid R. Heat on a water-bath, then cautiously over a naked flame, raising the temperature progressively, and then incinerate in a muffle furnace at 600 ± 25 °C. Continue heating until all black particles have disappeared. Allow to cool, add a few drops of dilute sulfuric acid R and heat and incinerate as above. Allow to cool, add a few drops of ammonium carbonate solution R, evaporate to dryness and incinerate cautiously. Allow to cool and dissolve the residue in 50 mL of water R.

Appearance of solution S1

Solution S1 is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

3.0 to 5.0.

Sodium glycolate

Disperse 1.0 g in 30 mL of carbon dioxide-free water R.

Maximum 2.0 per cent. Carry out the test protected from light.

Test solution Place 0.20 g in a beaker. Add 5 mL of acetic acid R and 5 mL of water R. Stir until dissolution is complete (about 10 min). Add 50 mL of acetone R and 1 g of sodium chloride R. Filter through a fast filter paper impregnated with acetone R, rinse the beaker and filter with acetone R. Combine the filtrate and washings and dilute to 100.0 mL with acetone R. Allow to stand for 24 h without shaking. Use the clear supernatant.

Reference solution Dissolve 0.310 g of glycolic acid R, previously dried in vacuo over diphosphorus pentoxide R at room temperature overnight, in water R and dilute to 500.0 mL with the same solvent. To 5.0 mL of this solution add 5 mL of acetic acid R and allow to stand for about 30 min. Add 50 mL of acetone R and 1 g of sodium chloride R. Filter through a fast filter paper impregnated with acetone R, rinse the beaker and filter with acetone R. Combine the filtrate and washings and dilute to 100.0 mL with acetone R. Allow to stand for 24 h without shaking. Use the clear supernatant.

Heat 2.0 mL of the test solution on a water-bath for 20 min. Cool to room temperature and add 20.0 mL of 2,7-dihydroxynaphthalene solution R. Shake and heat in a water-bath for 20 min. Cool under running water, transfer to a volumetric flask and dilute to 25.0 mL with sulfuric acid R, maintaining the flask under running water. Within 10 min, measure the absorbance at 540 nm (2.2.25) using water R as the compensation liquid. The absorbance of the solution prepared with the test solution is not greater than that of a solution prepared at the same time and in the same manner with 2.0 mL of the reference solution.

◆ Sodium chloride Maximum 7.0 per cent.

Place 0.500 g in a beaker and suspend in 100 mL of water R. Add 1 mL of nitric acid R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20), using a silver-based indicator electrode and a double-junction reference electrode containing a 100 g/L solution of potassium nitrate R in the outer jacket and a standard filling solution in the inner jacket.

1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of NaCl.

Iron (2.4.9) Maximum 20 ppm, determined on 10 mL of solution S2.

◆ Heavy metals (2.4.8) Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R. ◆

Loss on drying (2.2.32) Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 1.5 h.

Microbial contamination It complies with the test for *Escherichia coli* and *Salmonella* (2.6.13).

ASSAY Shake 1.000 g with 20 mL of ethanol (80 per cent V/V) R, stir for 10 min and filter. Repeat the operation until chloride has been completely extracted and verify the absence of chloride using silver nitrate solution R2. Dry the residue at 105 °C to constant mass. To 0.700 g of the dried residue, add 80 mL of glacial acetic acid R and heat under a reflux condenser for 2 h. Cool the solution to room temperature. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 2.299 mg of Na.

◆ STORAGE In an airtight container, protected from light. ◆

Ph Eur

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.



Sodium Starch Glycolate (Type C)⁽¹⁾

(Ph. Eur. monograph 1566)

Action and use

Excipient.

DEFINITION

Sodium salt of a partly *O*-carboxymethylated starch, cross-linked by physical dehydration.

Content

2.8 per cent to 5.0 per cent of Na (M_r 44.01) (substance washed with ethanol (80 per cent V/V) and dried).

CHARACTERS

Appearance

White or almost white, fine, free-flowing powder, very hygroscopic.

Microscopic examination It is seen to consist of granules, irregularly shaped, ovoid or pear-shaped, 30–100 µm in size,

or rounded, 10–35 µm in size; compound granules consisting of 2–4 components occur occasionally; the granules have an eccentric hilum and clearly visible concentric striations; between crossed nicol prisms, the granules show a distinct black cross intersecting at the hilum; small crystals are visible at the surface of the granules. The granules show considerable swelling in contact with water.

Solubility

Soluble in water, practically insoluble in methylene chloride. It gives a translucent gel-like product in water.

IDENTIFICATION

A. pH (see Tests).

B. Prepare with shaking and without heating a mixture of 4.0 g of the substance to be examined and 20 mL of carbon dioxide-free water R. The mixture has the appearance of a gel, remains stable (difference from types A and B). Keep the gel for the tests for appearance of gel and pH.

C. To 5 mL of the gel obtained in identification test B add 0.05 mL of iodine solution R1. A dark blue colour is produced.

D. Solution S (see Tests) gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Place 2.5 g in a silica or platinum crucible and add 2 mL of a 500 g/L solution of sulfuric acid R. Heat on a water-bath, then cautiously over a naked flame, raising the temperature progressively, and then incinerate in a muffle furnace at 600 ± 25 °C. Continue heating until all black particles have disappeared. Allow to cool, add a few drops of sulfuric acid R, heat and incinerate as above. Allow to cool, add a few drops of ammonium carbonate solution R, evaporate to dryness and incinerate cautiously. Allow to cool and dissolve the residue in 50 mL of water R.

Appearance of gel

The gel obtained in identification test B is colourless (2.2.2,

pH (2.2.3)

Method II).

5.5 to 7.5 for the gel obtained in identification test B.

Reference solution Dissolve 0.310 g of glycolic acid R, previously dried *in vacuo* over diphenylphosphorus pentoxide R at room temperature overnight, in water R and dilute to 500.0 mL with the same solvent. To 5.0 mL of this solution add 5 mL of acetic acid R and allow to stand for about 30 min. Add 50 mL of acetone R and 1 g of sodium chloride R. Filter through a fast filter paper impregnated with acetone R, rinse the beaker and filter with acetone R. Combine the filtrate and washings and dilute to 100.0 mL with acetone R. Allow to stand for 24 h without shaking. Use the clear supernatant.

Heat 2.0 mL of the test solution on a water-bath for 20 min. Cool to room temperature and add 20.0 mL of 2,7-dihydroxynaphthalene solution R. Shake and heat in a water-bath for 20 min. Cool under running water, transfer quantitatively to a volumetric flask and dilute to 25.0 mL with sulfuric acid R, maintaining the flasks under running water. Within 10 min, measure the absorbance at 540 nm (2.2.25) using water R as the compensation liquid. The absorbance of the solution prepared with the test solution is not greater than that of a solution prepared at the same time and in the same manner with 2.0 mL of the reference solution.

Sodium chloride
Maximum 7.0 per cent.
Place 0.500 g in a beaker and suspend in 100 mL of water R. Add 1 mL of nitric acid R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20), using a silver-based indicator electrode and a double-junction reference electrode containing a 100 g/L solution of potassium nitrate R in the outer jacket and a standard filling solution in the inner jacket.
1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of NaCl.

Iron (2.4.9)
Maximum 20 ppm, determined on 10 mL of solution S2.

Heavy metals (2.4.8)
Maximum 20 ppm.
1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R. ♦

Loss on drying (2.2.32)
Maximum 10.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 1.5 h.

Microbial contamination
It complies with the test for *Escherichia coli* and *Salmonella* (2.6.13).

ASSAY
Shake 1.000 g with 20 mL of ethanol (80 per cent V/V) R, stir for 10 min and filter. Repeat the operation until chloride has been completely extracted and verify the absence of chloride using silver nitrate solution R2. Dry the residue at 105 °C to constant mass. To 0.700 g of the dried residue, add 80 mL of glacial acetic acid R and heat under a reflux condenser for 2 h. Cool the solution to room temperature. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.
1 mL of 0.1 M perchloric acid is equivalent to 2.299 mg of Na.

♦ STORAGE

In an airtight container, protected from light. ♦

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.

Ph Eur

1 mL of 0.1 M perchloric acid is equivalent to 2.299 mg of Na.

STORAGE

In an airtight container, protected from light.

Ph Eur

¹ This monograph has undergone pharmacopoeial harmonisation.

See chapter 5.8 Pharmacopoeial Harmonisation.

Sodium Stearate

(Ph. Eur. monograph 2058)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of sodium salts of different fatty acids consisting mainly of stearic (octadecanoic) acid $[C_{17}H_{35}COONa; M_r 306.5]$ and palmitic (hexadecanoic) acid $[C_{15}H_{31}COONa; M_r 278.4]$.

Content

— sodium: 7.4 per cent to 8.5 per cent (A_r 22.99) (dried substance);
— stearic acid in the fatty acid fraction: minimum 40 per cent; sum of stearic acid and palmitic acid in the fatty acid fraction: minimum 90 per cent.

CHARACTERS

Appearance

White or yellowish, fine powder, greasy to the touch.

Solubility

Slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification: C, D.

Second identification: A, B, D.

A. Freezing point (2.2.18): minimum 53 °C for the residue obtained in the preparation of solution S (see Tests).
B. Acid value (2.5.1): 195 to 210, determined on 0.200 g of the residue obtained in the preparation of solution S
C. Examine the chromatograms obtained in the assay of stearic acid and palmitic acid.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time and size to the 2 principal peaks in the chromatogram obtained with the reference solution.
D. Solution S gives reaction (b) of sodium (2.3.1).

TESTS

Solution S

To 10.0 g add 100 mL of peroxide-free ether R and 80 mL of acetic acid R. Boil under a reflux condenser until dissolution is complete. Allow to cool. In a separating funnel, separate the aqueous layer and shake the ether layer with 2 quantities, each of 8 mL, of acetic acid R. Combine the aqueous layers, wash with 30 mL of peroxide-free ether R and dilute to 100 mL with distilled water R (solution S). Evaporate the ether layers to dryness on a water-bath and dry the residue at 100-105 °C.

Sodium glycolate

Maximum 2.0 per cent. Carry out the test protected from light.

Test solution Place 0.20 g in a beaker. Add 5 mL of acetic acid R and 5 mL of water R. Stir until dissolution is complete (about 10 min). Add 50 mL of acetone R and 1 g of sodium chloride R. Filter through a fast filter paper impregnated with acetone R, rinse the beaker and filter with acetone R. Combine the filtrate and washings and dilute to 100.0 mL with acetone R. Allow to stand for 24 h without shaking. Use the clear supernatant.

Reference solution Dissolve 0.310 g of glycolic acid R, in previously dried *in vacuo* over diphosphorus pentoxide R, in water R and dilute to 500.0 mL with the same solvent.

To 5.0 mL of this solution, add 5 mL of acetic acid R and allow to stand for about 30 min. Add 50 mL of acetone R and 1 g of sodium chloride R and dilute to 100.0 mL with acetone R.

Hear 2.0 mL of the test solution on a water-bath for 20 min. Cool to room temperature and add 20.0 mL of 2,7-dihydroxynaphthalene solution R. Shake and heat on a water-bath for 20 min. Cool under running water, transfer to a volumetric flask and dilute to 25.0 mL with sulfuric acid R, maintaining the flask under running water. Within 10 min, measure the absorbance (2.2.25) at 540 nm using water R as the compensation liquid. The absorbance of the solution prepared with the test solution is not greater than that of a solution prepared at the same time and in the same manner with 2.0 mL of the reference solution.

Sodium chloride

Maximum 1 per cent.

Shake 1.00 g with 20 mL of ethanol (80 per cent V/V) R for 10 min and filter. Repeat the operation 4 times. Dry the residue to constant mass at 100 °C and set aside for the assay. Combine the filtrates. Evaporate to dryness, take up the residue with water R and dilute to 25.0 mL with the same solvent. To 10.0 mL of the solution add 30 mL of water R and 5 mL of dilute nitric acid R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20), using a silver-based indicator electrode and a double-junction reference electrode containing a 100 g/L solution of potassium nitrate R in the outer jacket and a standard filling solution in the inner jacket.

1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of NaCl.

Iron (2.4.9)

Maximum 20 ppm, determined on solution S.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 7.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Microbial contamination

It complies with the test for *Escherichia coli* and *Salmonella* (2.6.13).

ASSAY

To 0.500 g of the dried and crushed residue obtained in the test for sodium chloride add 80 mL of anhydrous acetic acid R and heat under a reflux condenser for 2 h. Cool the solution to room temperature. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank test.

Acidity or alkalinity

Suspend 2.0 g in 50 mL of previously neutralised ethanol (96 per cent) R. Heat under reflux to dissolve and add 3 drops of phenolphthalein solution R; the solution is colourless. Not less than 0.60 mL and not more than 0.85 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Chlorides (2.4.4)

Maximum 0.2 per cent.

Dilute 0.25 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 0.3 per cent.

Dilute 0.5 mL of solution S to 15 mL with distilled water R.

Nickel

Maximum 5 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Place 50.0 mg of the substance to be examined in a polytetrafluoroethylene digestion flask and add 0.5 mL of a mixture of 1 volume of heavy metal-free hydrochloric acid R and 5 volumes of heavy metal-free nitric acid R. Allow to digest at 170 °C for 5 h. Allow to cool. Dissolve the residue in water R and dilute to 5.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using nickel standard solution (10 ppm Ni) R, diluting as necessary with water R.

Source Nickel hollow-cathode lamp.

Wavelength 232.0 nm.

Atomisation device Furnace.

Loss on drying (2.2.32)

Maximum 5.0 per cent.

In a weighing glass introduce 1.0 g of previously washed sand R, dry at 105 °C and weigh. Add 0.500 g of the substance to be examined and 10 mL of ethanol (96 per cent) R. Evaporate at 80 °C and dry the residue at 105 °C for 4 h.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

ASSAY

Sodium
Dissolve 0.250 g with gentle heating in a mixture of 5 mL of acetic anhydride R and 20 mL of anhydrous acetic acid R. Cool and add 20 mL of dioxan R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 2.299 mg of Na.

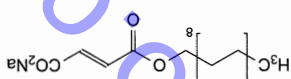
Stearic acid and palmitic acid

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution In a conical flask fitted with a reflux condenser, dissolve 0.10 g of the substance to be examined in 5 mL of boron trifluoride-methanol solution R. Boil under a reflux condenser for 10 min. Add 4 mL of heptane R through the condenser and boil again under a reflux condenser for 10 min. Allow to cool. Add 20 mL of saturated sodium chloride solution R. Shake and allow the layers to separate. Remove about 2 mL of the organic layer and dry over 0.2 g

Sodium Stearyl Fumarate

(Ph. Eur. monograph 1567)



C₂₂H₃₉NaO₄

390.5

4070-80-8

DEFINITION

Sodium octadecyl (E)-butenedioate.

Content

99.0 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, fine powder with agglomerates of flat, circular particles.

Solubility

Practically insoluble in water, slightly soluble in methanol, practically insoluble in acetone and in anhydrous ethanol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).
Comparison sodium stearyl fumarate CRS.

STORAGE

In an airtight container, protected from light.

Ph Eur

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to methyl stearate (retention time = about 40 min): methyl palmitate = about 0.88.

System suitability: reference solution:

— resolution: minimum 5.0 between the peaks due to methyl palmitate and methyl stearate.

Calculate the content of stearic acid and palmitic acid.

Temperature (°C)	Time (min)	Column	Injection port	Detector
70	0 - 2			
70 → 240	2 - 36			
240	36 - 41			
220				
260				

Temperature:

Flow rate 2.4 mL/min.

Carrier gas helium for chromatography R.

0.5 µm).

— stationary phase: macrogol 20 000 R (film thickness

— size: l = 30 m, Ø = 0.32 mm;

— material: fused silica;

Column:

substance to be examined.

manner as the test solution using 50.0 mg of palmitic acid CRS and 50.0 mg of stearic acid CRS instead of the

Reference solution Prepare the reference solution in the same

to 100.0 mL with heptane R.

of anhydrous sodium sulfate R. Dilute 1.0 mL of the solution

TESTS**Related substances**

Gas chromatography (2.2.28): use the normalisation

procedure.

Silylation solution To 2 mL of N_2O -bis(trimethylsilyl) trifluoroacetamide R add 0.02 mL of

chlorotrimethylsilane R and mix.

Test solution Introduce 15.0 mg of the substance to be

silylation solution. Seal the vial and heat at about 70 °C for

1 h. After the reaction a precipitate remains in the vial; filter

the solution through a nylon filter (pore size 0.45 µm).

Reference solution Introduce 1.0 mg of sodium stearyl

maleate CRS and 1.0 mg of sodium stearyl fumarate CRS into

a vial with a screw cap and add 1 mL of the silylation

solution. Seal the vial and heat at about 70 °C for 1 h.

Column:

— material: fused silica;

— size: $l = 15$ m, $\phi = 0.53$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness

0.15 µm).

Carrier gas helium for chromatography R.

Flow rate 2 mL/min.

Split ratio 1:25.

Temperature:

Time (min)	Temperature (°C)
0 - 1	180
1 - 21	180 → 320
21 - 26	320
Injection port	250
Detector	320

Detection Flame ionisation.

Injection 2 µL.

Relative retention With reference to stearyl trimethylsilyl

fumarate (retention time = about 9 min): stearyl

alcohol = 0.30; stearyl trimethylsilyl ether = 0.35; palmityl

trimethylsilyl fumarate = 0.80; heptadecyl trimethylsilyl

fumarate = 0.85; stearyl trimethylsilyl maleate = 0.90;

nonadecyl trimethylsilyl fumarate = 1.05; eicos-1-enyl

trimethylsilyl fumarate = 1.15; distearyl fumarate = 2.25.

System suitability:

— resolution: minimum 1.5 between the peaks in the

chromatogram obtained with the reference solution.

Limits:

— any impurity: maximum 0.5 per cent;

— total: maximum 5.0 per cent.

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.250 g.

ASSAY

Dissolve 0.250 g, accurately weighed, in 10 mL of methylene

chloride R and add 30 mL of anhydrous acetic acid R. Titrate

with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

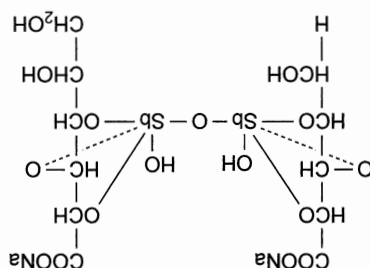
1 mL of 0.1 M perchloric acid is equivalent to 39.05 mg of

$C_{22}H_{39}NaO_4$.

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more

functions of the substance when used as an excipient (see chapter

Sodium Stibogluconate**Action and use**

Pentavalent antimony compound; antiprotozoal.

Preparation

Sodium Stibogluconate Injection

DEFINITION

Sodium Stibogluconate is mainly the disodium salt of μ -oxy-bis[gluconato(3-)-O⁻, O⁻, O⁻, O⁻-hydroxoantimony]. It contains

not less than 30.0% and not more than 34.0% of

antimony(V), calculated with reference to the dried and

methanol-free substance.

PRODUCTION

The method of manufacture is such as to ensure consistently

controlled reaction stoichiometry in order to yield sodium

stibogluconate that is satisfactory with regard to intrinsic

toxicity.

A colourless, mostly amorphous powder.

Very soluble in water; practically insoluble in ethanol (96%)

and in ether.

IDENTIFICATION

A. A solution is dextrorotatory.

B. Pass hydrogen sulfide into a 5% w/v solution for several

minutes. An orange precipitate is produced.

C. When heated, it chars without melting leaving a residue

which yields the reactions characteristic of antimony

compounds and the reactions characteristic of sodium salts,

Appendix VI.

TESTS

Stability and acidity of solution

Heat a solution containing the equivalent of 10% w/v of

antimony(V) in an autoclave at 115.5° and at a pressure of

Specific surface area (2.9.26, Method I)

Particle-size distribution (2.9.31)

The following characteristics may be relevant for sodium stearyl

fumarate used as a lubricant in tablets and capsules.

reported, the control method must be indicated.

also be used. Wherever results for a particular characteristic are

recognised as being suitable for the purpose, but other methods can

product during use. Where control methods are cited, they are

to the quality of a medicinal product by improving the consistency

compliance. Control of these characteristics can however contribute

and it is not necessary to verify the characteristics to demonstrate

5.15). This section is a non-mandatory part of the monograph

16037-91-5

70 kPa for 30 minutes. The resulting solution is colourless or almost colourless and has a pH of 5.0 to 5.6, Appendix V L.

Antimony(III)

Dissolve 2 g in 30 mL of water, add 15 mL of hydrochloric acid and titrate with 0.00833M potassium bromate VS using methyl orange solution as indicator. Not more than 1.3 mL of 0.00833M potassium bromate VS is required.

Chloride

Dissolve 2.5 g in 50 mL of water and add 2 mL of 2M nitric acid and 75 mL of acetate buffer pH 5.0. Titrate with 0.1M silver nitrate VS determining the end point

potentiometrically. Not more than 3.0 mL of 0.1M silver nitrate VS is required.

Methanol

Not more than 2.0% w/w when determined by the following method. Carry out the method for gas chromatography, Appendix III B, using the following solutions. For solution

(1) add 1 mL of a 1.0% v/v solution of methanol to 5 mL of a 0.2% v/v solution of absolute ethanol (internal standard).

For solution (2) add 5 mL of water to 0.5 g of the substance being examined and mix with the aid of ultrasound until solution is complete.

For solution (3) add 5 mL of a 0.2% v/v solution of the internal standard to 0.5 g of the substance being examined and mix with the aid of ultrasound until solution is complete.

The chromatographic procedure may be carried out using a glass column (1.5 m × 4 mm) packed with porous polymer beads (80 to 100 mesh) (Porapak Q and Chromosorb 101

are suitable) and maintained at 130°. Calculate the percentage w/w of methanol taking 0.792 g as its weight per mL at 20°.

When dried to constant weight at 130° at a pressure not exceeding 0.7 kPa, loses not more than 15.0% of its weight.

Loss on drying

Use 1 g.

ASSAY

Dissolve 0.16 g in 30 mL of hydrochloric acid, add 70 mL of orthophosphoric acid and stir carefully until completely mixed.

Titrate with 0.05M ammonium iron(II) sulfate VS prepared using sulfuric acid (1%) and determining the end point potentiometrically using a platinum electrode and a silver-silver chloride reference electrode. Each mL of

0.05M ammonium iron(II) sulfate VS is equivalent to 3.044 mg of antimony(V).

Anhydrous Sodium Sulfate



Anhydrous Sodium Sulphate

(Ph. Eur. monograph 0099)

Na₂SO₄

Action and use

Laxative.

DEFINITION

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder, hygroscopic.

Solubility

Freely soluble in water.

IDENTIFICATION

A. It gives the reactions of sulfates (2.3.1).

B. It gives the reactions of sodium (2.3.1).

C. Loss on drying (see Tests).

TESTS

Solution S

Dissolve 2.2 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Chlorides (2.4.4)

Maximum 450 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Calcium (2.4.3)

Maximum 450 ppm, if intended for use in the manufacture of parenteral preparations.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 90 ppm, if intended for use in the manufacture of parenteral preparations.

Dilute 5 mL of solution S to 10 mL with water R.

Magnesium

Maximum 200 ppm, if intended for use in the manufacture of parenteral preparations.

To 10 mL of solution S add 1 mL of glycerol (85 per cent) R, 0.15 mL of titan yellow solution R, 0.25 mL of ammonium oxalate solution R and 5 mL of dilute sodium hydroxide solution R and shake. Any pink colour in the test solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 5 mL of magnesium standard solution (10 ppm Mg) R and 5 mL of water R.

Heavy metals (2.4.8)

Maximum 45 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 130 °C.

ASSAY

Dissolve 0.100 g in 40 mL of water R. Add a mixture of 0.2 mL of 0.1 M hydrochloric acid and 80 mL of methanol R. Carry out a potentiometric titration (2.2.20), using 0.1 M lead nitrate and as indicator electrode a lead-selective electrode and as reference electrode a silver-silver chloride electrode.

1 mL of 0.1 M lead nitrate is equivalent to 14.20 mg of Na₂SO₄.

STORAGE

Store in an airtight container.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

Sodium Sulfate



Glauber's Salt

(Sodium Sulfate Decahydrate, Ph Eur monograph 0100)

 $\text{Na}_2\text{SO}_4 \cdot 10\text{H}_2\text{O}$

322.2

7727-73-3

Action and use

Laxative.

Ph Eur

DEFINITION

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder or colourless, transparent crystals.

Solubility

Freely soluble in water, practically insoluble in ethanol (96 per cent). It partly dissolves in its own water of crystallisation at about 33 °C.

IDENTIFICATION

A. It gives the reactions of sulfates (2.3.1).
B. It gives the reactions of sodium (2.3.1).

C. Loss on drying (see Tests).

TESTS

Solution S
Dissolve 5.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Calcium (2.4.3)

Maximum 200 ppm, if intended for use in the manufacture of parenteral preparations.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Iron (2.4.9)

Maximum 40 ppm, if intended for use in the manufacture of parenteral preparations.

Dilute 5 mL of solution S to 10 mL with water R.

Magnesium

Maximum 100 ppm, if intended for use in the manufacture of parenteral preparations.

To 10 mL of solution S add 1 mL of glycerol (85 per cent) R, 0.15 mL of nitric acid solution R, 0.25 mL of ammonium oxalate solution R and 5 mL of dilute sodium hydroxide solution R and shake. Any pink colour in the test solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 5 mL of magnesium standard solution (10 ppm Mg) R and 5 mL of water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Anhydrous Sodium Sulfite



Anhydrous Sodium Sulfite

(Ph. Eur. monograph 0775)

 Na_2SO_3

7757-83-7

Action and use

Antioxidant.

Ph Eur

DEFINITION

Content

95.0 per cent to 100.5 per cent of Na_2SO_3 .

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is slightly alkaline (2.2.4).
B. To 5 mL of solution S add 0.5 mL of 0.05 M iodine. The solution is colourless and gives reaction (a) of sulfates (2.3.1).
C. Solution S gives reaction (a) of sodium (2.3.1).
D. It complies with the limits of the assay.

TESTS

Solution S

Dissolve 5 g in water R and dilute to 100 mL with the same solvent.

Solution S1

To 10.0 g add 25 mL of water R. Shake until mostly dissolved, carefully and progressively add 15 mL of hydrochloric acid R. Heat to boiling. Cool and dilute to 100.0 mL with water R.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method I).

Thiosulfates

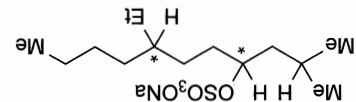
Maximum 0.1 per cent.

To 2.00 g add 100 mL of water R. Shake, add 10 mL of formaldehyde solution R and 10 mL of acetic acid R. Allow to

<p>stand for 5 min. Add 0.5 mL of starch solution R and titrate with 0.05 M iodine. Carry out a blank titration.</p> <p>The difference between the volumes used in the titrations is not more than 0.15 mL.</p> <p>Iron (2.4.9)</p> <p>Maximum 10 ppm, determined on solution S1.</p> <p>Selenium</p> <p>Maximum 10 ppm.</p> <p>To 3.0 g add 10 mL of formaldehyde solution R, carefully and progressively add 2 mL of hydrochloric acid R. Heat on a water-bath for 20 min. Any pink colour in the solution is not more intense than that of a standard prepared at the same time and in the same manner using 1.0 g of the substance to be examined to which 0.2 mL of selenium standard solution (100 ppm Se) R has been added.</p> <p>Zinc</p> <p>Maximum 25 ppm.</p> <p>Atomic absorption spectrometry (2.2.23, Method I).</p> <p>Test solution Dilute 2.0 mL of solution S1 to 10.0 mL with water R.</p> <p>Reference solutions Prepare the reference solutions using zinc standard solution (100 ppm Zn) R, diluting with water R.</p> <p>Source Zinc hollow-cathode lamp.</p> <p>Wavelength 213.9 nm.</p> <p>Atomisation device Air-acetylene flame.</p> <p>Heavy metals (2.4.8)</p> <p>Maximum 10 ppm.</p> <p>Evaporate 20 mL of solution S1 almost to dryness.</p> <p>Add 10 mL of water R, neutralise with concentrated ammonia R and dilute to 20 mL with water R. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.</p> <p>ASSAY</p> <p>Introduce 0.250 g into a 500 mL conical flask containing 50.0 mL of 0.05 M iodine. Shake until completely dissolved. Add 1 mL of starch solution R and titrate the excess of iodine with 0.1 M sodium thiosulfate. Carry out a blank titration.</p> <p>1 mL of 0.05 M iodine is equivalent to 6.30 mg of Na₂SO₃.</p>	<p>Ph Eur</p> <p>Sodium Sulphite Heptahydrate</p> <p>(Ph. Eur. monograph 0776)</p> <p>Na₂SO₃·7H₂O</p> <p>252.2</p> <p>10102-15-5</p> <p>Ph Eur</p> <p>Ph Eur</p> <p>IDENTIFICATION</p> <p>A. Solution S (see Tests) is slightly alkaline (2.2.4).</p> <p>B. To 5 mL of solution S add 0.5 mL of 0.05 M iodine. The solution is colourless and gives reaction (a) of sulfates (2.3.1).</p> <p>C. Solution S gives reaction (a) of sodium (2.3.1).</p> <p>D. It complies with the limits of the assay.</p> <p>TESTS</p> <p>Solution S</p> <p>Dissolve 10 g in water R and dilute to 100 mL with the same solvent.</p> <p>Solution S1</p> <p>To 20.0 g add 25 mL of water R. Shake until mostly dissolved, and carefully and progressively, add 15 mL of hydrochloric acid R. Heat to boiling. Cool and dilute to 100.0 mL with water R.</p> <p>Appearance of solution</p> <p>Solution S is clear (2.2.1) and colourless (2.2.2, Method I).</p> <p>Thiosulfates</p> <p>Maximum 0.05 per cent.</p> <p>To 4.00 g add 100 mL of water R. Shake to dissolve, add 10 mL of formaldehyde solution R and 10 mL of acetic acid R. Allow to stand for 5 min, then add 0.5 mL of starch solution R and titrate with 0.05 M iodine. Carry out a blank titration. The difference between the volumes used in the titrations is not more than 0.15 mL.</p> <p>Iron (2.4.9)</p> <p>Maximum 5 ppm, determined on solution S1.</p> <p>Selenium</p> <p>Maximum 5 ppm.</p> <p>To 6.0 g add 10 mL of formaldehyde solution R, carefully and progressively add 2 mL of hydrochloric acid R. Heat on a water-bath for 20 min. Any pink colour in the solution is not more intense than that of a standard prepared at the same time and in the same manner using 2.0 g of the substance to be examined to which 0.2 mL of selenium standard solution (100 ppm Se) R has been added.</p> <p>Zinc</p> <p>Maximum 12 ppm.</p> <p>Atomic absorption spectrometry (2.2.23, Method I).</p> <p>Test solution Dilute 2.0 mL of solution S1 to 10.0 mL with water R.</p> <p>Reference solutions Prepare the reference solutions using zinc standard solution (100 ppm Zn) R, diluting with water R.</p> <p>Source Zinc hollow-cathode lamp.</p> <p>Wavelength 213.9 nm.</p> <p>Atomisation device Air-acetylene flame.</p> <p>Heavy metals (2.4.8)</p> <p>Maximum 5 ppm.</p> <p>Evaporate 20 mL of solution S1 almost to dryness.</p> <p>Add 10 mL of water R, neutralise with concentrated ammonia R and dilute to 20 mL with water R. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.</p> <p>ASSAY</p> <p>Introduce 0.250 g into a 500 mL conical flask containing 50.0 mL of 0.05 M iodine. Shake until completely dissolved. Add 1 mL of starch solution R and titrate the excess of iodine with 0.1 M sodium thiosulfate. Carry out a blank titration.</p> <p>1 mL of 0.05 M iodine is equivalent to 6.30 mg of Na₂SO₃.</p> <p>Ph Eur</p> <p>CHARACTERS</p> <p>Content</p> <p>48.0 per cent to 52.5 per cent of Na₂SO₃.</p> <p>Appearance</p> <p>Colourless crystals.</p> <p>Solubility</p> <p>Freely soluble in water, very slightly soluble in ethanol (96 per cent).</p>
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Sodium Tetradecyl Sulfate Concentrate

Sodium Tetradecyl Sulphate Concentrate



$C_{14}H_{29}NaO_4S$ 316.4

139-88-8

Action and use

Sclerosant.

Preparation

Sodium Tetradecyl Sulfate Injection

DEFINITION

Sodium Tetradecyl Sulfate Concentrate is an aqueous gel containing sodium *all-rac-4-ethyl-1-isobutylsulfate*. It contains not less than 46.0% w/w and not more than 52.0% w/w of $C_{14}H_{29}NaO_4S$.

CHARACTERISTICS

A clear, colourless gel.

IDENTIFICATION

A. Carry out the method for gas chromatography, Appendix III B, using the following solutions. For solution (1) boil 0.2 g under a reflux condenser with 20 mL of 2M hydrochloric acid for 15 minutes, allow to cool, add 20 mL of ethanol (96%) and extract the mixture with two 10 mL quantities of *n*-pentane. Wash the combined pentane extracts with 20 mL of water and dry over anhydrous sodium sulfate.

Solution (2) contains 0.35% w/v of decan-1-ol and 0.7% w/v of dodecan-1-ol in *n*-pentane. The chromatographic procedure may be carried out using a glass column (1.5 m × 4 mm) packed with acid-washed, silanised diatomaceous support (80 to 100 mesh) coated with 3% w/w of polyethylene glycol (Carbowax 20M is suitable) and maintained at 120°.

The retention time of the principal peak in the chromatogram obtained with solution (1) is less than the retention time of the peak due to decan-1-ol and more than that of the peak due to decan-1-ol in the chromatogram obtained with solution (2).

B. Mix 0.1 mL of a 2% w/v solution with 0.1 mL of a 0.1% w/v solution of methylene blue and 2 mL of 1M sulfuric acid, add 2 mL of chloroform and shake. The chloroform layer is intensely blue.

C. Mix 20 mg with 10 mL of ethanol (96%) and heat to boiling on a water bath, shaking frequently. Filter immediately and evaporate the ethanol. Dissolve the residue in 8 mL of water, add 3 mL of 2M hydrochloric acid, evaporate the solution to half its volume and cool. Filter to remove the congealed fatty alcohols and add 1 mL of 0.25M barium chloride to the filtrate. A white, crystalline precipitate is produced.

D. Yields reaction B characteristic of sodium salts,

TESTS

Alkalinity

Dissolve 1 g in 100 mL of water and add 0.1 mL of methyl red solution; the solution is yellow. Not more than 0.5 mL of 0.1M hydrochloric acid VS is required to change the colour of the solution.

Non-ionic impurities

Not more than 3.0% w/w, with respect to the nominal content of Sodium Tetradecyl Sulfate, when determined by the method for gas chromatography, Appendix III B, using the following solutions. Dissolve 75 mg of dodecan-1-ol (internal standard) in sufficient *n*-hexane to produce 100 mL (solution A). For solution (1) extract 1 g of the concentrate being examined with 10 mL of solution A followed by two 10 mL quantities of *n*-hexane. Pass 10 mL of the combined extracts at a rate of about 1.5 mL per minute through a column, 1 cm in diameter, packed with 5 g of basic aluminium oxide (Brockmann grade II is suitable) and pre-washed with 25 mL of *n*-hexane. Elute with 20 mL of a mixture of equal volumes of *n*-hexane and ether, evaporate to dryness using a rotary evaporator and dissolve the residue in 2 mL of *n*-hexane. Prepare solution (2) in the same manner but using 10 mL of *n*-hexane in place of 10 mL of solution A.

The chromatographic procedure may be carried out using a glass column (1.5 m × 4 mm) packed with acid-washed, silanised diatomaceous support (80 to 100 mesh) coated with 3% w/w of polyethylene glycol (Carbowax 20M is suitable) and maintained at 120°.

Allow the chromatography to proceed for the retention time of the peak due to the internal standard. In the chromatogram obtained with solution (1) the sum of the areas of any secondary peaks is not greater than twice the area of the peak due to the internal standard.

Chloride

Dissolve 33 mg in sufficient water to produce 100 mL. 15 mL of the resulting solution complies with the limit test for chlorides, Appendix VII (1%).

Sulfated ash

10.3 to 13.3%, Appendix IX A.

ASSAY

Dissolve 2.4 g in sufficient water to produce 1000 mL. To 20 mL add 15 mL of chloroform and 10 mL of dimidium bromide-sulfan blue mixed solution and titrate with 0.004M benzethonium chloride VS, shaking vigorously and allowing the layers to separate after each addition, until the pink colour of the chloroform layer is completely discharged and a greyish blue colour is produced. Each mL of 0.004M benzethonium chloride VS is equivalent to 1.266 mg of $C_{14}H_{29}NaO_4S$.

STORAGE

Sodium Tetradecyl Sulfate Concentrate should be protected from light.

Sodium Thiosulfate

Sodium Thiosulphate

(Ph. Eur. monograph 0414)

$Na_2S_2O_3 \cdot 5H_2O$ 248.2

10102-17-7

Action and use

Used in treatment of cyanide poisoning.

Preparation

Sodium Thiosulfate Injection

DEFINITION

Content

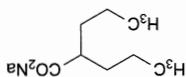
99.0 per cent to 101.0 per cent of $Na_2S_2O_3 \cdot 5H_2O$.





Sodium Valproate

(Ph. Eur. monograph 0678)


 $C_8H_{15}NaO_2$

166.2

1069-66-5

Action and use

Antiepileptic.

Preparations

Sodium Valproate Oral Solution

Sodium Valproate Tablets

Gastro-resistant Sodium Valproate Tablets

Prolonged-release Sodium Valproate Capsules

Prolonged-release Sodium Valproate Tablets

DEFINITION

Sodium 2-propylpentanoate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline, hygroscopic powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sodium valproate CRS.

If the spectra obtained in the solid state show differences, record new spectra using discs prepared by placing 50 µL of

a 100 g/L solution in methanol R on a disc of potassium

bromide R and evaporating the solvent *in vacuo*. Examine

immediately.

B. 2 mL of solution S (see Tests) gives reaction (a) of

sodium (2.3.1).

TESTS

Solution S

Dissolve 1.25 g in 20 mL of distilled water R in a separating

funnel, add 5 mL of dilute nitric acid R and shake. Allow the

mixture to stand for 12 h. Use the aqueous lower layer.

Appearance of solution

The solution is not more opalescent than reference

suspension II (2.2.1) and not more intensely coloured than

reference solution Y₆ (2.2.2, Method II).

Dissolve 2.0 g in water R and dilute to 10 mL with the same

Acidity or alkalinity

Dissolve 1.0 g in 10 mL of water R. Add 0.1 mL of

phenolphthalein solution R. Not more than 0.75 mL of 0.1 M

hydrochloric acid or 0.1 M sodium hydroxide is required to

change the colour of the indicator.

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 0.500 g of the substance to be

examined in 10 mL of water R. Add 5 mL of dilute sulfuric

acid R and shake with 3 quantities, each of 20 mL, of

CHARACTERS

Appearance

Transparent, colourless crystals, efflorescent in dry air.

Solubility

Very soluble in water, practically insoluble in ethanol

96 per cent. It dissolves in its water of crystallisation at about

49 °C.

IDENTIFICATION

A. It decolourises iodinated potassium iodide solution R.

B. To 0.5 mL of solution S (see Tests) add 0.5 mL of

water R and 2 mL of silver nitrate solution R2. A white

precipitate is formed which rapidly becomes yellowish and

then black.

C. To 2.5 mL of solution S add 2.5 mL of water R and

1 mL of hydrochloric acid R. A precipitate of sulfur is formed

and gas is evolved which gives a blue colour to starch iodate

paper R.

D. 1 mL of solution S gives reaction (a) of sodium (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from

distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

The freshly prepared solution is clear (2.2.1) and colourless

(2.2.2, Method II).

Dissolve 10.0 g in 50 mL of distilled water R, add 1 mL of

0.1 M sodium hydroxide and dilute to 100 mL with the same

pH (2.2.3)

6.0 to 8.4 for the freshly prepared solution S.

Sulfates and sulfites (2.4.13)

Maximum 0.2 per cent.

Dilute 2.5 mL of freshly prepared solution S to 10 mL with

distilled water R. To 3 mL of this solution first add 2 mL of

iodinated potassium iodide solution R and continue the addition

dropwise until a very faint persistent yellow colour appears.

Dilute to 15 mL with distilled water R.

Sulfides

To 10 mL of solution S add 0.05 mL of a freshly prepared

50 g/L solution of sodium nitroprusside R. The solution does

not become violet.

Heavy metals

Maximum 10 ppm.

To 10 mL of solution S add 0.05 mL of sodium sulfide

solution R. After 2 min, any brown colour in the solution is

not more intense than that in a reference solution prepared at

the same time and in the same manner using 10 mL of lead

standard solution (1 ppm Pb) R.

ASSAY

Dissolve 0.500 g in 20 mL of water R and titrate with

0.05 M iodine, using 1 mL of starch solution R, added towards

the end of the titration, as indicator.

1 mL of 0.05 M iodine is equivalent to 24.82 mg

of Na₂S₂O₃·5H₂O.

STORAGE

In an airtight container.

Ph Eur

ASSAY

Dissolve 0.150 g in 25 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 16.62 mg of $C_8H_{15}NaO_2$.

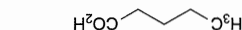
STORAGE

In an airtight container.

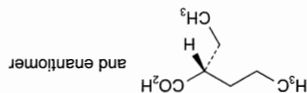
IMPURITIES

Specified impurities K

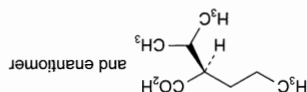
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, F, G, I, J, L.



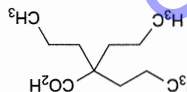
A. pentanoic acid (valeric acid),



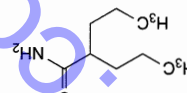
B. (2R,5S)-2-ethylpentanoic acid,



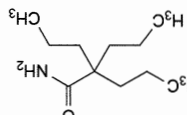
C. (2R,5S)-2-(1-methylethyl)pentanoic acid,



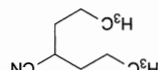
D. 2,2-dipropylpentanoic acid,



F. 2-propylpentanamide,



G. 2,2-dipropylpentanamide,



I. 2-propylpentanenitrile,

heptane R. Dilute the combined upper layers to 100.0 mL

with heptane R.

Reference solution (a) Dissolve 5 mg of valproic acid for system suitability CRS (containing impurity K) in 1.0 mL of

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with heptane R.

Column:

— material: wide-bore fused silica;

— size: $l = 30$ m, $\phi = 0.53$ mm;

— stationary phase: macrogol 20 000 2-nitroterephthalate R

Carrier gas helium for chromatography R.

Flow rate 8 mL/min.

Temperature:

Temperature	Time	Column
(°C)	(min)	
80	0 - 5	80
80 → 150	5 - 15	80 → 150
150 → 190	15 - 28.3	150 → 190
190	28.3 - 30	190
220		220
Detector		220

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to valproic acid (retention

time = about 17 min): impurity K = about 0.97.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to

impurity K and valproic acid.

Limits:

— impurity K: not more than 0.15 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than

0.05 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.05 per cent);

— total: not more than 0.2 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— disregard limit: 0.03 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.03 per cent).

Chlorides (2.4.4)

Maximum 200 ppm.

Sulfates (2.4.13)

Maximum 200 ppm, determined on solution S.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 2.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

glycine: 0.90 to 1.10; alanine: 0.90 to 1.10; phenylalanine: 2.7 to 3.3; serine: 0.7 to 1.05; threonine: 1.4 to 2.1; half-cystine: 1.4 to 2.2. Not more than traces of other amino acids are present.

TESTS

Specific optical rotation (2.2.7)

—47 to -37 (anhydrous and acetic acid-free substance). Dissolve 2.0 mg in 1.0 mL of a 1 per cent *V/V* solution of glacial acetic acid *R*.

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure. Prepare the solutions immediately before use.

Test solution Dissolve the substance to be examined in water *R* to obtain a concentration of 1.0 mg/mL.

Reference solution (a) Dissolve the contents of a vial of somatostatin CRS in water *R* to obtain a concentration of 1.0 mg/mL.

Reference solution (b) Dissolve the contents of a vial of somatostatin CRS in a 1 mg/mL solution of dihydroxythiitol *R* to obtain a concentration of 1.0 mg/mL.

room temperature for about 1 h. Mix 1 volume of this solution and 1 volume of reference solution (a).

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography *R* (5 μ m) with a pore size of 10 nm.

Mobile phase:

— mobile phase A: trifluoroacetic acid *R*, acetonitrile *R*, water *R* (0.1:20:80 *V/V/V*);
— mobile phase B: trifluoroacetic acid *R*, acetonitrile *R*, water *R* (0.1:45:55 *V/V/V*);

Time (min)	Mobile phase A (per cent <i>V/V</i>)	Mobile phase B (per cent <i>V/V</i>)
0 - 25	100 \rightarrow 0	0 \rightarrow 100
25 - 30	0	100
30 - 32	0 \rightarrow 100	100 \rightarrow 0

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 15 μ L of the test solution and reference solution (b).

Relative retention With reference to somatostatin (retention time = about 12 min): reduced somatostatin = about 1.09.

System suitability: reference solution (b):

— resolution: minimum 3.5 between the peak due to reduced somatostatin and the principal peak;

— number of theoretical plates: minimum 15 000, calculated for the principal peak;

— symmetry factor: maximum 2.7 for the principal peak.

Limits:

— any impurity: maximum 1 per cent;

— total: maximum 2 per cent;

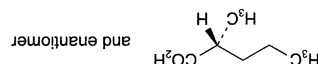
— disregard limit: 0.1 per cent.

Acetic acid (2.5.34)

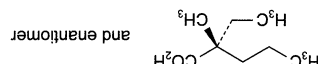
3.0 per cent to 15.0 per cent.

Test solution Dissolve 7.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A, then dilute to 10.0 mL with the same mixture of mobile phases.

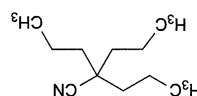
L. (2*R,S*)-2-methylpentanoic acid.



K. (2*R,S*)-2-ethyl-2-methylpentanoic acid,



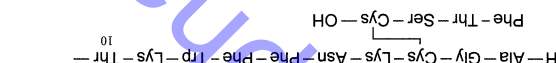
J. 2,2-dipropylpentanenitrile,



Ph Eur

Somatostatin

(Ph. Eur. monograph 0949)



C₇₆H₁₀₄N₁₈O₁₉S₂ 1638 38916-34-6

Action and use

Growth hormone release inhibiting hormone.

Ph Eur

DEFINITION

L-Alanylglutyl-L-cysteiny-L-lysyl-L-asparagyl-L-phenylalanyl-L-phenylalanyl-L-tyrosyl-L-threonyl-L-phenylalanyl-L-threonyl-L-seryl-L-cysteinyl-L-phenylalanyl-L-phenylalanyl-L-cysteine cyclic (3 \rightarrow 14)-disulfide.

Synthetic cyclic tetradecapeptide having the structure of the hypothalamic hormone that inhibits the release of human growth hormone. It contains a variable quantity of acetic acid. It is available in the freeze-dried form.

Content

95.0 per cent to 104.0 per cent (anhydrous and acetic acid-free substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, amorphous powder.

Solubility

Freely soluble in water and in acetic acid, practically insoluble in methylene chloride.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

B. Amino acid analysis (2.2.56). Method 1 for hydrolysis and method 1 for analysis are suitable.

Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids taking 1/8 of the sum of the number of moles of aspartic acid, alanine, lysine, glycine and phenylalanine as equal to 1. The values fall within the following limits: aspartic acid: 0.90 to 1.10;

Water (2.5.32)

Maximum 8.0 per cent, determined on 10.0 mg.

Bacterial endotoxins (2.6.14)

Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection 10 µL of the test solution and reference solution (a).

Calculate the content of somatostatin ($C_{76}H_{104}N_{18}O_{19}S_2$) taking into account the assigned content of

$C_{76}H_{104}N_{18}O_{19}S_2$ in somatostatin CRS.

STORAGE

In an airtight container, protected from light, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

Ph Eur

Somatropin

(Ph. Eur. monograph 0951)



FEETLSRLT	DNALRAHRL	HOLAFDTYOE	FEAVIPIKEO
KYSFLQNPQT	SLCESSEIPT	PSNREETQOK	SNLEILRISL
LIIGQSWTEBPV	QLTRSVFANS	LTVGASDSNV	YDLKDLLEG
IOTLMGRLED	GSPTGQIFK	QTYSKFDTS	HNDALTKNY
GLTYCFKDM	DKVETFLRIV	QCRSVEGSCG	F

 $C_{990}H_{1528}N_{262}O_{300}S_7$

22,125

12629-01-5

Somatropin Injection**Preparation**

Growth hormone.

Action and use

Protein having the structure (191 amino-acid residues) of the major component of growth hormone produced by the human pituitary.

Content

91.0 per cent to 105.0 per cent (anhydrous substance).

By convention, for the purpose of labelling somatropin preparations, 1 mg of anhydrous somatropin ($C_{990}H_{1528}N_{262}O_{300}S_7$) is equivalent to 3.0 IU of biological activity.

PRODUCTION

Somatropin is produced by a method based on recombinant DNA (rDNA) technology. During the course of product development, it must be demonstrated that the manufacturing process produces a product having a biological activity of not less than 2.5 IU/mg, using a validated bioassay based on growth promotion and approved by the competent authority.

Somatropin complies with the following additional requirements.

Host-cell-derived proteins

The limit is approved by the competent authority.

Host-cell- and vector-derived DNA

The limit is approved by the competent authority.

CHARACTERS**Appearance**

White or almost white powder.

IDENTIFICATION

A. Capillary electrophoresis (2.2.47) as described in the test for charged variants with the following modifications.

Injection Test solution (b); under pressure or vacuum, using the following sequence: sample injection for at least 3 s then CZE buffer injection for 1 s.

Results In the electropherogram obtained, only 1 principal peak, corresponding to somatropin, is detected: no doubling of this peak is observed.

B. Examine the chromatograms obtained in the test for related proteins.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

C. Peptide mapping (2.2.55).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Prepare a solution of the substance to be examined in 0.05 M tris-hydrochloride buffer solution pH 7.5 R

to obtain a solution containing 2.0 mg/mL of somatropin and transfer about 1.0 mL to a tube made from a suitable material such as polypropylene. Prepare a 1 mg/mL solution of trypsin for peptide mapping R in 0.05 M tris-hydrochloride buffer solution pH 7.5 R and add 30 µL to the solution of the substance to be examined. Cap the tube and place in a water-bath at 37 °C for 4 h. Remove from the water-bath and stop the reaction immediately, for example by freezing. If analysed immediately using an automatic injector, maintain at 2–8 °C.

Reference solution Prepare at the same time and in the same manner as for the test solution, but using somatropin CRS instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (2.2.29).

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: octylsilyl silica gel for chromatography R (5–10 µm) with a pore size of 30 nm;
— temperature: 30 °C.

Mobile phase:

— mobile phase A: dilute 1 mL of trifluoroacetic acid R to 1000 mL with water R;
— mobile phase B: to 100 mL of water R, add 1 mL of trifluoroacetic acid R and dilute to 1000 mL with acetonitrile for chromatography R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 20	100 → 80	0 → 20
20 - 40	80 → 75	20 → 25
40 - 65	75 → 50	25 → 50
65 - 70	50 → 20	50 → 80

Flow rate 1 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 100 µL.

System suitability The chromatograms obtained with the test solution and the reference solution are similar to the

chromatogram of somatropin digest supplied with

somatropin CRS.

Results The profile of the chromatogram obtained with the

test solution corresponds to that of the chromatogram

obtained with the reference solution.

D. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

TESTS

Related proteins

Liquid chromatography (2.2.29): use the normalisation

procedure. Maintain the solutions at 2–8 °C and use within

24 h. If an automatic injector is used, maintain it at 2–8 °C.

Test solution Prepare a solution of the substance to be

examined in 0.05 M tris-hydrochloride buffer solution pH 7.5 R,

containing 2.0 mg/mL of somatropin.

Reference solution Prepare a solution of somatropin CRS in

0.05 M tris-hydrochloride buffer solution pH 7.5 R, containing

2.0 mg/mL of somatropin.

Resolution solution Dissolve the contents of a vial of

somatropin/desamidatosomatropin mixture CRS in

0.05 M tris-hydrochloride buffer solution pH 7.5 R to obtain a

concentration of 2 mg/mL of somatropin.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: a suitable singly end-capped butylsilyl

silica gel, with a granulometry of 5 µm and a porosity of

30 nm; a silica saturation column is placed between the

pump and the injector valve;

— temperature: 45 °C.

Mobile phase propanol R, 0.05 M tris-hydrochloride buffer

solution pH 7.5 R (29:71 V/V).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 220 nm.

Preconditioning of the column Rinse with 200–500 mL of a

0.1 per cent V/V solution of trifluoroacetic acid R in a

50 per cent V/V solution of acetonitrile R; repeat as necessary,

to improve column performance.

Injection 20 µL.

Relative retention With reference to somatropin (retention

time = about 33 min; if necessary adjust the concentration of

propanol R in the mobile phase):

desamidatosomatropin = about 0.85.

System suitability Resolution solution:

— resolution: minimum 1.0 between the peaks due to

desamidatosomatropin and somatropin;

— symmetry factor: 0.9 to 1.8 for the peak due to somatropin.

Limit:

— total: maximum 6.0 per cent.

Dimer and related substances of higher molecular

mass

Size-exclusion chromatography (2.2.30)

Use the normalisation procedure.

Test solution Prepare a solution of the substance to be

examined in 0.025 M phosphate buffer solution pH 7.0 R,

containing 1.0 mg/mL of somatropin.

Reference solution Dissolve the contents of a vial of

somatropin CRS in 0.025 M phosphate buffer solution pH 7.0 R

and dilute with the same solution to obtain a concentration

of 1.0 mg/mL.

Resolution solution Place 1 vial of somatropin CRS in an oven

at 50 °C for a period sufficient to generate 1–2 per cent of

dimer (typically 12–24 h). Dissolve its contents in 0.025 M

phosphate buffer solution pH 7.0 R and dilute with the same

solution to obtain a concentration of 1.0 mg/mL.

Column:

— size: $l = 0.30$ m, $\varnothing = 7.8$ mm;

— stationary phase: hydrophilic silica gel for chromatography R

of a grade suitable for fractionation of globular proteins in

the relative molecular mass range of 5000 to 150 000.

Mobile phase 2-propanol R, 0.063 M phosphate buffer solution

pH 7.0 R (3:97 V/V); filter and degas.

Flow rate 0.6 mL/min.

Detection Spectrophotometer at 214 nm.

Injection 20 µL.

Relative retention With reference to somatropin monomer

(retention time = 12 min to 17 min): related substances of

higher molecular mass = about 0.65; somatropin

dimer = about 0.9.

System suitability Resolution solution:

— peak-to-valley ratio: minimum 2.5, where H_p = height

above the baseline of the peak due to the dimer and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to the

monomer.

Charged variants

Capillary electrophoresis (2.2.47).

Test solution (a) Prepare a solution of the substance to be

examined containing 1 mg/mL of somatropin.

Test solution (b) Mix equal volumes of test solution (a) and

the reference solution.

Reference solution Dissolve the contents of a vial of

somatropin CRS in water R and dilute with the same solvent

to obtain a concentration of 1 mg/mL.

Capillary:

— material: uncured fused silica;

— size: effective length = at least 70 cm, $\varnothing = 50$ µm.

Temperature 30 °C.

CZE buffer 13.2 g/L solution of ammonium phosphate R

adjusted to pH 6.0 with phosphoric acid R and filtered.

Detection Spectrophotometer at 200 nm.

Set the autosampler to store the samples at 4 °C during analysis.

Preconditioning of the capillary Rinse with 1 M sodium hydroxide

for 20 min, with water R for 10 min and with the CZE buffer

for 20 min.

Between-run rinsing Rinse with 0.1 M sodium hydroxide for

2 min and with the CZE buffer for 6 min.

Note: rinsing times may be adapted according to the length of the

capillary and the equipment used.

Injection Test solution (a) and the reference solution; under

pressure or vacuum, using the following sequence: sample

injection for at least 3 s then CZE buffer injection for 1 s.

The injection time and pressure may be adapted in order to

meet the system suitability criteria.

Migration Apply a field strength of 217 V/cm (20 kV for

capillaries of 92 cm total length) for 80 min, using the CZE

buffer as the electrolyte in both buffer reservoirs.

Relative migration With reference to somatropin: deamidated

forms = 1.02 to 1.11.

System suitability: reference solution: — the electropherogram obtained is similar to the electropherogram of somatropin supplied with principal peak and at least 2 peaks (I_1 , I_2) eluting prior to the principal peak and clearly visible.

Notes: peak I_2 corresponds to the cleaved form and peak I_4 corresponds to the deamidated forms, eluting as a doublet.

Limits:

— **deamidated forms:** maximum 5.0 per cent;
— **any other impurity:** for each impurity, maximum 2.0 per cent;
— **total:** maximum 10.0 per cent.

Water (2.5.32)

Maximum 10.0 per cent.

Bacterial endotoxins (2.6.14)

Less than 5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for removal of bacterial endotoxins.

ASSAY

Size-exclusion chromatography (2.2.30) as described in the test for dimer and related substances of higher molecular mass.

Calculate the content of somatropin ($C_{990}H_{1528}N_{262}O_{300}S_7$) from the declared content of $C_{990}H_{1528}N_{262}O_{300}S_7$ in somatropin CRS.

STORAGE

In an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, tamper-proof container.

Somatropin Concentrated Solution



Somatropin Bulk Solution
(Ph. Eur. monograph 0950)

$C_{990}H_{1528}N_{262}O_{300}S_7$ 22,125

FFTIPIISRLT	DNAMLRARHL	HQLAFDTYQE	FEAVYIPKEQ
KYSFLONPQT	STCFSESIPT	PSNRREETQOK	SNLEELRLST
LLIQSWLEBP	QFLRSVFANS	LVYGASDSNV	VDLTKDLEEG
IQTIMGRLED	GSPRTGQIEK	QTSKEDTNS	HNDALTKNY
GLLYCFERKDM	DKVETFLRIIV	QCRSVEGSCG	F

Action and use

Growth hormone.

Preparation

Somatropin Injection

DEFINITION

Solution containing a protein having the structure (191 amino-acid residues) of the major component of growth hormone produced by the human pituitary. It may contain buffer salts and other auxiliary substances.

Content

91.0 per cent to 105.0 per cent of the amount of somatropin stated on the label.
By convention, for the purpose of labelling somatropin preparations, 1 mg of anhydrous somatropin

($C_{990}H_{1528}N_{262}O_{300}S_7$) is equivalent to 3.0 IU of biological activity.

PRODUCTION

Somatropin concentrated solution is produced by a method based on recombinant DNA (rDNA) technology. During the course of product development, it must be demonstrated that the manufacturing process produces a product having a biological activity of at least 2.5 IU/mg, using a validated bioassay based on growth promotion and approved by the competent authority.

Host-cell-derived proteins

The limit is approved by the competent authority.

Host-cell- and vector-derived DNA

The limit is approved by the competent authority.

CHARACTERS

Appearance

Clear or slightly turbid, colourless solution.

IDENTIFICATION

A. Capillary electrophoresis (2.2.47) as described in the test for charged variants with the following modifications.

Injection Test solution (b); under pressure or vacuum, using the following sequence: sample injection for at least 3 s then CZE buffer injection for 1 s.

Results In the electropherogram obtained, only 1 principal peak, corresponding to somatropin, is detected: no doubling

B. Examine the chromatograms obtained in the test for related proteins.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

C. Peptide mapping (2.2.55).

SELECTIVE CLEAVAGE OF THE PEPTIDE BONDS

Test solution Dilute the solution to be examined with 0.05 M *tris-hydrochloride buffer solution pH 7.5* so that it contains 2.0 mg/mL of somatropin and transfer about 1.0 mL to a tube made from a suitable material such as polypropylene. Prepare a 1 mg/mL solution of trypsin for peptide mapping R in 0.05 M *tris-hydrochloride buffer solution pH 7.5* R and add 30 µL to the solution of the substance to be examined. Cap the tube and place in a water-bath at 37 °C for 4 h. Remove from the water-bath and stop the reaction immediately, for example by freezing. If analysed immediately using an automatic injector, maintain at 2–8 °C.

Note: If a 2 mg/mL somatropin concentration is not obtainable, a similar digest relationship (micrograms of trypsin per milligram of somatropin) may be used.

Reference solution Prepare at the same time and in the same manner as for the test solution, but using somatropin CRS instead of the substance to be examined.

CHROMATOGRAPHIC SEPARATION

Liquid chromatography (2.2.29).

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: octylsilyl silica gel for chromatography R (5–10 µm) with a pore size of 30 nm;
— temperature: 30 °C.

Mobile phase:
— **mobile phase A:** dilute 1 mL of trifluoroacetic acid R to 1000 mL with water R;
— **mobile phase B:** to 100 mL of water R, add 1 mL of trifluoroacetic acid R and dilute to 1000 mL with acetonitrile for chromatography R;
— **mobile phase:**

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 20	100 → 80	0 → 20
20 - 40	80 → 75	20 → 25
40 - 65	75 → 50	25 → 50
65 - 70	50 → 20	50 → 80

Detection Spectrophotometer at 214 nm.
Flow rate 1 mL/min.

System suitability The chromatograms obtained with the test solution and the reference solution are similar to the chromatogram of somatropin digest supplied with somatropin CRS.

Results The profile of the chromatogram obtained with the test solution corresponds to that of the chromatogram obtained with the reference solution.

D. Examine the chromatograms obtained in the assay. **Results** The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

TESTS

Related proteins
Liquid chromatography (2.2.29): use the normalisation procedure. Maintain the solutions at 2-8 °C and use within 24 h. If an automatic injector is used, maintain it at 2-8 °C. **Test solution** Dilute the solution to be examined in 0.05 M tris-hydrochloride buffer solution pH 7.5 R, so as to contain 2.0 mg/mL of somatropin. A weaker solution may be prepared, in which case the injection volume is adjusted accordingly.

Reference solution Prepare a solution of somatropin CRS in 0.05 M tris-hydrochloride buffer solution pH 7.5 R, containing 2.0 mg/mL of somatropin.

Resolution solution Dissolve the contents of a vial of somatropin/desamidosomatropin resolution mixture CRS in 0.05 M tris-hydrochloride buffer solution pH 7.5 R to obtain a concentration of 2 mg/mL of somatropin.

Column:
— **size:** $l = 0.25$ m, $\phi = 4.6$ mm;
— **stationary phase:** a suitable singly end-capped butylsilyl silica gel, with a granulometry of 5 µm and a porosity of 30 nm; a silica saturation column is placed between the pump and the injector valve;
— **temperature:** 45 °C.

Mobile phase propanol R, 0.05 M tris-hydrochloride buffer solution pH 7.5 R (29:71 V/V).
Flow rate 0.5 mL/min.

Detection Spectrophotometer at 220 nm.
Preconditioning of the column Rinse with 200-500 mL of a 0.1 per cent V/V solution of trifluoroacetic acid R in a 50 per cent V/V solution of acetonitrile R; repeat as necessary, to improve column performance.

Injection 20 µL.

Relative retention With reference to somatropin (retention time = about 33 min; if necessary adjust the concentration of propanol R in the mobile phase):
desamidosomatropin = about 0.85.

System suitability Resolution solution:
— **resolution:** minimum 1.0 between the peaks due to desamidosomatropin and somatropin;
— **symmetry factor:** 0.9 to 1.8 for the peak due to somatropin.

Limit:
— **total:** maximum 6.0 per cent.
Dimer and related substances of higher molecular mass

Size-exclusion chromatography (2.2.30)
Use the normalisation procedure.
Test solution Dilute the solution to be examined in 0.025 M phosphate buffer solution pH 7.0 R, so as to contain 1.0 mg/mL of somatropin.

Reference solution Dissolve the contents of a vial of somatropin CRS in 0.025 M phosphate buffer solution pH 7.0 R and dilute with the same solution to obtain a concentration of 1.0 mg/mL.

Resolution solution Place 1 vial of somatropin CRS in an oven at 50 °C for a period sufficient to generate 1-2 per cent of dimer (typically 12-24 h). Dissolve its contents in 0.025 M phosphate buffer solution pH 7.0 R and dilute with the same solution to obtain a concentration of 1.0 mg/mL.

Column:
— **size:** $l = 0.30$ m, $\phi = 7.8$ mm;
— **stationary phase:** hydrophilic silica gel for chromatography R of a grade suitable for fractionation of globular proteins in the relative molecular mass range of 5000 to 150 000.

Mobile phase 2-propanol R, 0.063 M phosphate buffer solution pH 7.0 R (3:97 V/V); filter and degas.
Flow rate 0.6 mL/min.

Detection Spectrophotometer at 214 nm.
Injection 20 µL.

Relative retention With reference to somatropin monomer (retention time = 12 min to 17 min): related substances of dimer = about 0.9.

System suitability Resolution solution:
— **peak-to-valley ratio:** minimum 2.5, where H_p = height above the baseline of the peak due to the dimer and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to the monomer.

Limit:
— **sum of the peaks with retention times less than that of the principal peak:** maximum 4.0 per cent.

Charged variants
Capillary electrophoresis (2.2.47).
Test solution (a) Dilute the solution to be examined so as to obtain a concentration of 1 mg/mL of somatropin.

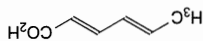
Test solution (b) Mix equal volumes of test solution (a) and the reference solution.
Reference solution Dissolve the contents of a vial of somatropin CRS in water R and dilute with the same solvent to obtain a concentration of 1 mg/mL.

Capillary:
— **material:** uncoated fused silica;
— **size:** effective length = at least 70 cm, $\phi = 50$ µm.



Sorbic Acid

(Ph. Eur. monograph 0592)



112.1

110-44-1

Action and use

Antimicrobial preservative.

Ph Eur

DEFINITION

(E,E)-Hexa-2,4-dienoic acid.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in ethanol

IDENTIFICATION

First identification A, C.

Second identification A, B, D.

A. Melting point (2.2.14): 132 °C to 136 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in water R and dilute to

250.0 mL with the same solvent. Dilute 2.0 mL of this

solution to 200.0 mL with 0.1 M hydrochloric acid.

Spectral range 230-350 nm.

Absorption maximum At 264 nm.

Specific absorbance at the absorption maximum 2150 to 2550.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison sorbic acid CRS.

D. Dissolve 0.2 g in 2 mL of ethanol (96 per cent) R and add

0.2 mL of bromine water R. The solution is decolorised.

TESTS

Solution S

Dissolve 1.25 g in ethanol (96 per cent) R and dilute to

25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Aldehydes

Maximum 0.15 per cent, calculated as C₂H₄O.

Dissolve 1.0 g in a mixture of 30 mL of water R and 50 mL

of 2-propanol R, adjust to pH 4 with 0.1 M hydrochloric acid

or 0.1 M sodium hydroxide and dilute to 100 mL with

water R. To 10 mL of this solution add 1 mL of decolorised

fuchsin solution R and allow to stand for 30 min. Any colour

in the solution is not more intense than that in a standard

prepared at the same time by adding 1 mL of decolorised

fuchsin solution R to a mixture of 1.5 mL of acetaldehyde

standard solution (100 ppm C₂H₄O) R, 4 mL of 2-propanol R

and 4.5 mL of water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test B. Prepare the

reference solution using 5 mL of lead standard solution

(1 ppm Pb) R and 5 mL of ethanol (96 per cent) R.

Temperature 30 °C.

CZE buffer 13.2 g/L solution of ammonium phosphate R

adjusted to pH 6.0 with phosphoric acid R and filtered.

Detection Spectrophotometer at 200 nm.

Set the autosampler to store the samples at 4 °C during analysis.

Preconditioning of the capillary Rinse with 1 M sodium hydroxide

for 20 min, with water R for 10 min and with the CZE buffer

for 20 min.

Between-run rinsing Rinse with 0.1 M sodium hydroxide for

2 min and with the CZE buffer for 6 min.

Note: rinsing times may be adapted according to the length of the

capillary and the equipment used.

Injection Test solution (a) and the reference solution; under

pressure or vacuum, using the following sequence: sample

injection for at least 3 s then CZE buffer injection for 1 s.

The injection time and pressure may be adapted in order to

meet the system suitability criteria.

Migration Apply a field strength of 217 V/cm (20 kV for

capillaries of 92 cm total length) for 80 min, using the CZE

buffer as the electrolyte in both buffer reservoirs.

Relative migration With reference to somatropin: deamidated

forms = 1.02 to 1.11.

System suitability: reference solution.

— the electropherogram obtained is similar to the

electropherogram of somatropin supplied with

somatropin CRS; 2 peaks (I₁, I₂) eluting prior to the

principal peak and at least 2 peaks (I₃, I₄) eluting after

the principal peak are clearly visible.

Note: peak I₂ corresponds to the cleaved form and peak I₄

corresponds to the deamidated forms, eluting as a doublet.

Limits:

— deamidated forms: maximum 5.0 per cent;

— any other impurity: for each impurity, maximum

2.0 per cent;

— total: maximum 10.0 per cent.

Bacterial endotoxins (2.6.14)

Less than 5 IU in the volume that contains 1 mg of

somatropin, if intended for use in the manufacture of

parenteral preparations without a further appropriate

procedure for removal of bacterial endotoxins.

ASSAY

Size-exclusion chromatography (2.2.30) as described in the

test for dimer and related substances of higher molecular

mass.

Calculate the content of somatropin (C₉₉₀H₁₅₂₈N₂₆₂O₃₀₀₅)

from the declared content of C₉₉₀H₁₅₂₈N₂₆₂O₃₀₀₅ in

somatropin CRS.

STORAGE

In an airtight container at a temperature of –20 °C. Avoid

repeated freezing and thawing. If the solution is sterile, store

in a sterile, airtight, tamper-proof container.

The label states:

— the content of somatropin in milligrams per millilitre;

— the name and concentration of any auxiliary substance.

Ph Eur

Water (2.5.12)

Maximum 1.0 per cent, determined on 2.000 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.1000 g in 20 mL of ethanol (96 per cent) R. Using 0.2 mL of phenolphthalein solution R as indicator, titrate with 0.1 M sodium hydroxide until a pink colour is obtained. 1 mL of 0.1 M sodium hydroxide is equivalent to 11.21 mg of $C_{18}H_{34}O_2$.

STORAGE

Protected from light.

Ph Eur

Sorbitan Laurate

(Ph. Eur. monograph 1040)

1338-39-2



Action and use

Non-ionic surfactant.

When sorbitan monolaurate is demanded, Sorbitan Laurate

shall be supplied.

Ph Eur

DEFINITION

Mixture usually obtained by partial esterification of sorbitol and its mono- and di-anhydrides with lauric (dodecanoic) acid.

CHARACTERS

Appearance

Brownish-yellow, viscous liquid.

Solubility

Practically insoluble, but dispersible in water, miscible with

alcohol.

Relative density

About 0.98.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Composition of fatty acids (see Tests).

TESTS

Acid value (2.5.1)

Maximum 7.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

330 to 358.

Iodine value (2.5.4)

Maximum 10.

Peroxide value (2.5.5)

Maximum 5.0.

Saponification value (2.5.6)

158 to 170.

Carry out the saponification for 1 h.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Prepare reference solution (a) as indicated in tables 2.4.22-1 and 2.4.22-2.

DEFINITION

Mixture usually obtained by esterification of 1 mole of sorbitol and its mono- and di-anhydrides per mole of oleic (cis-9-octadecenoic) acid. A suitable antioxidant may be added.

CHARACTERS

Appearance

Brownish-yellow, viscous liquid.

Solubility

Practically insoluble but dispersible in water, soluble in fatty

Relative density

About 0.99.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Composition of fatty acids (see Tests).

Margaric acid Maximum 0.2 per cent for oleic acid of

vegetable origin and maximum 4.0 per cent for oleic acid of

TESTS

Acid value (2.5.1)

Maximum 8.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

190 to 210.

Sorbitan Oleate

(Ph. Eur. monograph 1041)

1338-43-8



Action and use

Non-ionic surfactant.

When sorbitan mono-oleate is demanded, Sorbitan Oleate

shall be supplied.

Ph Eur

STORAGE

Protected from light.

Ph Eur

Total ash (2.4.16)

Maximum 0.5 per cent.

Water (2.5.12)

Maximum 1.5 per cent, determined on 1.00 g.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Heavy metals (2.4.8)

Maximum 10 ppm.

Composition of the fatty acid fraction of the substance:

- caproic acid: maximum 1.0 per cent,
- caprylic acid: maximum 10.0 per cent,
- capric acid: maximum 10.0 per cent,
- lauric acid: 40.0 per cent to 60.0 per cent,
- myristic acid: 14.0 per cent to 25.0 per cent,
- palmitic acid: 7.0 per cent to 15.0 per cent,
- stearic acid: maximum 7.0 per cent,
- oleic acid: maximum 11.0 per cent,
- linoleic acid: maximum 3.0 per cent.

Iodine value (2.5.4) 62 to 76.

Peroxide value (2.5.5) Maximum 10.0.

Saponification value (2.5.6) 145 to 160.

Carry out the saponification for 1 h.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Composition of the fatty acid fraction of the substance:

- myristic acid: maximum 5.0 per cent,
- palmitic acid: maximum 16.0 per cent,
- palmitoleic acid: maximum 8.0 per cent,
- stearic acid: maximum 6.0 per cent,
- oleic acid: 65.0 per cent to 88.0 per cent,
- linoleic acid: maximum 18.0 per cent,
- linolenic acid: maximum 4.0 per cent,
- fatty acids with chain length greater than C_{18} : maximum 4.0 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.5 per cent, determined on 1.000 g.

Total ash (2.4.16)

Maximum 0.5 per cent, determined on 1.5 g.

STORAGE

Protected from light.

LABELLING

The label states the origin of the oleic acid used (animal or vegetable).

Sorbitan Palmitate

(Ph. Eur. monograph 1042)

26266-57-9



Ph Eur

Action and use

Non-ionic surfactant.

When sorbitan monopalmitate is demanded, Sorbitan

Palmitate shall be supplied.

Ph Eur

DEFINITION

Mixture usually obtained by partial esterification of sorbitol

and its mono- and di-anhydrides with palmitic

(hexadecanoic) acid.

CHARACTERS

Appearance

Yellow or yellowish powder, waxy flakes or hard masses.

Solubility

Practically insoluble in water, soluble in fatty oils, slightly

soluble in alcohol.

IDENTIFICATION

A. Melting point (2.2.15): 44 °C to 51 °C.

Introduce the melted substance into the glass capillary tubes and allow to stand at a temperature below 10 °C for 24 h.

TESTS

C. Composition of fatty acids (see Tests).

B. Hydroxyl value (see Tests).

Acid value (2.5.1)

Maximum 8.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

270 to 305.

Peroxide value (2.5.5)

Maximum 5.0.

Saponification value (2.5.6)

140 to 155.

Carry out the saponification for 1 h.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Composition of the fatty acid fraction of the substance:

- palmitic acid: minimum 92.0 per cent,
- stearic acid: maximum 6.0 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.5 per cent, determined on 1.00 g.

Total ash (2.4.16)

Maximum 0.5 per cent.

STORAGE

Protected from light.

Ph Eur

Sorbitan Sesquioleate

(Ph. Eur. monograph 1916)

8007-43-0



Action and use

Non-ionic surfactant.

Ph Eur

DEFINITION

Mixture usually obtained by esterification of 2 moles of

sorbitol and its mono- and di-anhydrides per 3 moles of oleic

(*cis*-9-octadecenoic) acid. A suitable antioxidant may be

added.

CHARACTERS

Appearance

Pale yellow or slightly brownish-yellow paste, which becomes

a viscous, oily, brownish-yellow liquid at about 25 °C.

Solubility

Dispersible in water, soluble in fatty oils, slightly soluble in

ethanol.

Relative density

About 0.99.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Composition of fatty acids (see Tests).

Margaric acid Maximum 0.2 per cent for oleic acid of vegetable origin and maximum 4.0 per cent for oleic acid of animal origin.

TESTS

Acid value (2.5.1) Maximum 16.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A) 180 to 215.

Iodine value (2.5.4) 70 to 95.

Peroxide value (2.5.5) Maximum 10.0.

Saponification value (2.5.6) 145 to 166.

Carry out the saponification for 1 h.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Composition of the fatty acid fraction of the substance:

- myristic acid: maximum 5.0 per cent,
- palmitic acid: maximum 16.0 per cent,
- palmitoleic acid: maximum 8.0 per cent,
- stearic acid: maximum 6.0 per cent,
- oleic acid: 65.0 per cent to 88.0 per cent,
- linoleic acid: maximum 18.0 per cent,
- linolenic acid: maximum 4.0 per cent,
- fatty acids with chain length greater than C₁₈: maximum 4.0 per cent.

Heavy metals (2.4.8) Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12) Maximum 1.5 per cent, determined on 1.000 g.

Total ash (2.4.16) Maximum 0.5 per cent, determined on 1.5 g.

STORAGE

Protected from light.

LABELLING

The label states the origin of the oleic acid used (animal or vegetable).

Ph Eur

Sorbitan Stearate

(Ph. Eur. monograph 1043)

Action and use

Non-ionic surfactant.

When sorbitan monostearate is demanded, Sorbitan Stearate shall be supplied.

Ph Eur

DEFINITION

Mixture usually obtained by partial esterification of sorbitol and its mono- and di-anhydrides with Stearic acid 50 (1474) or Stearic acid 70 (1474).

CHARACTERS

Appearance

Pale yellow, waxy solid.

DEFINITION

Mixture usually obtained by esterification of 1 mole of sorbitol and its mono-anhydride per 3 moles of oleic (cis-9-octadecenoic) acid. A suitable antioxidant may be added.

Ph Eur

Action and use

Non-ionic surfactant.

(Ph. Eur. monograph 1044)

Sorbitan Trioleate

26266-58-0



Ph Eur

The label states the type of sorbitan stearate.

LABELLING

Protected from light.

STORAGE

Maximum 0.5 per cent.

Total ash (2.4.16) Maximum 1.5 per cent, determined on 1.00 g.

Water (2.5.12) Maximum 1.5 per cent, determined on 1.00 g.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Heavy metals (2.4.8) Maximum 10 ppm.

Type of fatty acid used	Composition of fatty acids
Sorbitan Stearic acid 50	Stearic acid: 40.0 per cent to 60.0 per cent, minimum 90.0 per cent, Sum of the contents of palmitic and stearic acids: (type I)
Sorbitan Stearic acid 70	Stearic acid: 60.0 per cent to 80.0 per cent, minimum 90.0 per cent, Sum of the contents of palmitic and stearic acids: (type II)

Composition of the fatty acid fraction of the substance:

Gas chromatography (2.4.22, Method C).

Composition of fatty acids

Carry out the saponification for 1 h.

147 to 157.

Saponification value (2.5.6) Maximum 5.0.

Peroxide value (2.5.5) 235 to 260.

Hydroxyl value (2.5.3, Method A) Maximum 10.0, determined on 5.0 g.

Acid value (2.5.1) Maximum 16.0, determined on 5.0 g.

TESTS

C. Composition of fatty acids (see Tests).

B. Hydroxyl value (see Tests).

Introduce the melted substance into the capillary tubes and allow to stand at a temperature below 10 °C for 24 h.

A. Melting point (2.2.15): 50 °C to 60 °C.

IDENTIFICATION

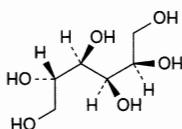
Practically insoluble, but dispersible in water, slightly soluble in alcohol.

Solubility



Sorbitol

(Ph. Eur. monograph 0435)



$C_6H_{14}O_6$ 182.2

50-70-4

Action and use

Used for parenteral nutrition.

DEFINITION

D-Glucitol (D-sorbitol).

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, practically insoluble in ethanol

It shows polymorphism (5.9).

IDENTIFICATION

First identification A

Second identification B, C, D

A. Examine the chromatograms obtained in the assay.

Results: The principal peak in the chromatogram obtained

with the test solution is similar in retention time and size to

the principal peak in the chromatogram obtained with

reference solution (a).

B. Dissolve 0.5 g with heating in a mixture of 0.5 mL of

pyridine R and 5 mL of acetic anhydride R. After 10 min, pour

the solution into 25 mL of water R and allow to stand in iced

water for 2 h. The precipitate, recrystallised from a

small volume of ethanol (96 per cent) R and dried in vacuo,

melts (2.2.14) at 98 °C to 104 °C.

C. Thin-layer chromatography (2.2.27).

Test solution: Dissolve 25 mg of the substance to be examined

in water R and dilute to 10 mL with the same solvent.

Reference solution (a): Dissolve 25 mg of sorbitol CRS in

water R and dilute to 10 mL with the same solvent.

Reference solution (b): Dissolve 25 mg of mannitol CRS and

25 mg of sorbitol CRS in water R and dilute to 10 mL with

the same solvent.

Plate: TLC silica gel G plate R.

Mobile phase: water R, ethyl acetate R, propanol R

(10:20:70 V/V/V).

Application: 2 µL.

Development: Over a path of 17 cm.

Drying: In air.

Detection: Spray with 4-aminobenzoic acid solution R; dry in a

current of cold air until the acetone is removed; heat at

100 °C for 15 min; allow to cool and spray with a 2 g/L

solution of sodium periodate R; dry in a current of cold air;

heat at 100 °C for 15 min.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

CHARACTERS

Appearance

Pale yellow, light yellowish or brown solid, which becomes a

viscous, oily, brownish-yellow liquid at about 25 °C.

Solubility

Practically insoluble but dispersible in water, soluble in fatty

oils, slightly soluble in alcohol.

Relative density

About 0.98.

IDENTIFICATION

A. Hydroxyl value (see Tests).

B. Iodine value (see Tests).

C. Composition of fatty acids (see Tests).

Marginal acid: Maximum 0.2 per cent for oleic acid of

vegetable origin and maximum 4.0 per cent for oleic acid of

animal origin.

TESTS

Acid value (2.5.1)

Maximum 16.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

55 to 75.

Iodine value (2.5.4)

76 to 90.

Peroxide value (2.5.5)

Maximum 10.0.

Saponification value (2.5.6)

170 to 190.

Carry out the saponification for 1 h.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Composition of the fatty acid fraction of the substance:

— myristic acid: maximum 5.0 per cent,

— palmitic acid: maximum 16.0 per cent,

— palmitoleic acid: maximum 8.0 per cent,

— stearic acid: maximum 6.0 per cent,

— oleic acid: 65.0 per cent to 88.0 per cent,

— linoleic acid: maximum 18.0 per cent,

— linolenic acid: maximum 4.0 per cent,

— fatty acids with chain length greater than C_{18} : maximum

4.0 per cent.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 1.5 per cent, determined on 1.000 g.

Total ash (2.4.16)

Maximum 0.5 per cent, determined on 1.5 g.

STORAGE

Protected from light.

LABELLING

The label states the origin of the oleic acid used (animal or

vegetable).

Ph Eur

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Specific optical rotation (2.2.7): + 4.0 to + 7.0 (anhydrous substance).

Dissolve 5.00 g of the substance to be examined and 6.4 g of disodium tetraborate R in 40 mL of water R. Allow to stand for 1 h, shaking occasionally, and dilute to 50.0 mL with water R. Filter if necessary.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 5 g in water R and dilute to 50 mL with the same solvent.

Conductivity (2.2.38)

Maximum 20 $\mu\text{S}\cdot\text{cm}^{-1}$

Dissolve 20.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution while gently stirring with a magnetic stirrer.

Reducing sugars

Maximum 0.2 per cent, expressed as glucose equivalent.

Dissolve 5.0 g in 6 mL of water R with the aid of gentle heat. Cool and add 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R and, when the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 5.0 g of the substance to be examined in 20 mL of water R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 0.50 g of sorbitol CRS in 2 mL of water R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with water R.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 100.0 mL with water R.

Reference solution (d) Dissolve 0.5 g of sorbitol R and 0.5 g of mannitol R (impurity A) in 5 mL of water R and dilute to 10.0 mL with the same solvent.

Column:

size: $l = 0.3$ m, $\phi = 7.8$ mm;

stationary phase: strong cation-exchange resin (calcium form) R (9 μm);

temperature: $85 \pm 1^\circ\text{C}$.

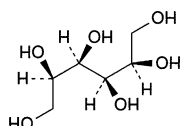
Mobile phase Degassed water R.

Flow rate 0.5 mL/min.

Detection Refractometer maintained at a constant temperature.

Injection 20 μL of the test solution and reference solutions (b), (c) and (d).

Run time 3 times the retention time of sorbitol.



A. D-mannitol.

IMPURITIES

— where applicable, the maximum concentration of bacterial endotoxins;

— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

The label states:

LABELLING

Calculate the percentage content of D-sorbitol from the declared content of sorbitol CRS.

Injection Test solution and reference solution (a).

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

ASSAY

concentration of 100 g/L or more of sorbitol.

— less than 2.5 IU/g for parenteral preparations having a concentration of less than 100 g/L of sorbitol;

— less than 4 IU/g for parenteral preparations having a removal of bacterial endotoxins;

preparations without a further appropriate procedure for the removal of bacterial endotoxins;

If intended for use in the manufacture of parenteral preparations (2.6.14)

Bacterial endotoxins (2.6.14)

— absence of *Salmonella* (2.6.13);

— absence of *Escherichia coli* (2.6.13);

— TYMC: acceptance criterion 10^3 CFU/g (2.6.12);

— TAMC: acceptance criterion 10^3 CFU/g (2.6.12);

preparations:

If not intended for use in the manufacture of parenteral preparations:

— TAMC: acceptance criterion 10^2 CFU/g (2.6.12).

If intended for use in the manufacture of parenteral preparations:

Microbial contamination

Maximum 1.5 per cent, determined on 1.00 g.

Water (2.5.12)

prescribed mixture of solvents.

Dissolve the substance to be examined in 150.0 mL of the

Maximum 1 ppm.

Nickel (2.4.15)

Maximum 0.5 ppm.

Lead (2.4.10)

(0.1 per cent).

chromatogram obtained with reference solution (c)

— disregard limit: the area of the principal peak in the

solution (b) (3 per cent);

peak in the chromatogram obtained with reference

— total: not more than 1.5 times the area of the principal

reference solution (b) (2 per cent);

the principal peak in the chromatogram obtained with

— any impurity: for each impurity, not more than the area of

Limits:

impurity A and sorbitol.

— resolution: minimum 2 between the peaks due to

System suitability Reference solution (d):

impurity A = about 0.8; impurity B = about 1.1.

time = about 27 min; impurity C = about 0.6;

Relative retention With reference to sorbitol (retention

CHARACTERS**Appearance**

Clear, colourless, syrupy liquid.

Solubility

Miscible with water, practically insoluble in mineral oils and vegetable oils.

IDENTIFICATION

Examine the chromatograms obtained in the assay.

Results The 2 principal peaks in the chromatogram obtained with the test solution are similar in retention time and size to the peaks in the chromatogram obtained with reference solution (a).

TESTS

solution (a).

Solution S

Dilute the substance to be examined with carbon dioxide-free water R prepared from distilled water R to obtain a solution containing 50.0 per cent *m/m* of anhydrous substance.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Conductivity (2.2.38)

Maximum 20 $\mu\text{S}\cdot\text{cm}^{-1}$.

Measure the conductivity of solution S, while gently stirring with a magnetic stirrer.

Reducing sugars

Maximum 0.3 per cent, calculated as glucose (anhydrous substance).

To an amount of the substance to be examined equivalent to 3.3 g of anhydrous substance, add 3 mL of water R, 20.0 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min. Maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent *V/V* solution of glacial acetic acid R and 20.0 mL of 0.025 *M* iodine. With continuous shaking, add 25 mL of a mixture of 6 mL of hydrochloric acid R and 94 mL of water R. When the precipitate has dissolved, titrate the excess of iodine with 0.05 *M* sodium thiosulfate using 2 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 *M* sodium thiosulfate is required.

Nickel (2.4.15)

Maximum 1 ppm (anhydrous substance).

Water (2.5.12)

15.0 per cent to 32.0 per cent, determined on 0.10 g.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (2.6.12).

TYMC: acceptance criterion 10^2 CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

ASSAY

Liquid chromatography (2.2.29).

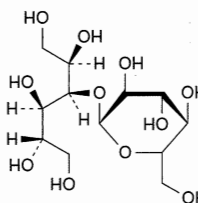
Test solution Dissolve 0.400 g of the substance to be examined in water R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dissolve 50.0 mg of sorbitol CRS and 20.0 mg of 1,4-sorbitan CRS in water R and dilute to 5.0 mL with the same solvent.

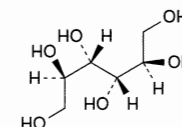
Reference solution (b) Dissolve 0.100 g of mannitol R and 0.100 g of sorbitol R in water R and dilute to 10.0 mL with the same solvent.

Column:

— size: $l = 0.3$ m, $\varnothing = 7.8$ mm;



B. D-iditol.



C. 4-O- α -D-glucopyranosyl-D-glucitol (D-maltitol).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph

and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute

to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal

product during use. Where control methods are cited, they are

recognised as being suitable for the purpose, but other methods can

also be used. Wherever results for a particular characteristic are

reported, the control method must be indicated.

The following characteristics may be relevant for sorbitol used as

filler and binder in tablets.

Particle-size distribution (2.9.31 or 2.9.38).**Powder flow (2.9.36)**

(Sorbitol, Liquid, Partially Dehydrated,
Ph Eur monograph 2048)

Action and use

Excipient.

Ph Eur

DEFINITION

Partially dehydrated liquid sorbitol is obtained by acid-catalysed partial internal dehydration of liquid sorbitol. It contains not less than 68.0 per cent *m/m* and not more than 85.0 per cent *m/m* of anhydrous substances, composed of a mixture of mainly D-sorbitol and 1,4-sorbitan, with mannitol, hydrogenated oligo- and disaccharides, and sorbitans.

Content

(nominal value):
— 1,4-sorbitan ($\text{C}_6\text{H}_{12}\text{O}_5$): minimum 15.0 per cent (anhydrous substance);
— D-sorbitol ($\text{C}_6\text{H}_{14}\text{O}_6$): minimum 25.0 per cent (anhydrous substance).
The contents of 1,4-sorbitan and D-sorbitol are within 95.0 per cent to 105.0 per cent of the nominal values.

— stationary phase: strong cation-exchange resin (calcium form) R (9 µm);
— temperature: 80 ± 5 °C.
Mobile phase Degassed water R.
Flow rate 0.5 mL/min.
Detection Refractometer maintained at a constant temperature of about 30-35 °C.
Injection 40 µL.
Relative retention With reference to D-sorbitol (retention time = about 25 min): 1,4-sorbitan = about 0.5; mannitol = about 0.8.
System suitability: reference solution (b):
— resolution: minimum 2.0 between the peaks due to mannitol and D-sorbitol.
Calculate the percentage contents of 1,4-sorbitan and D-sorbitol using the chromatogram obtained with reference solution (a) and the declared contents of 1,4-sorbitan CRS and of sorbitol CRS.

LABELLING

The label states the content of D-sorbitol and the content of 1,4-sorbitan (= nominal values).

Sorbitol Solution (70 per cent) (Crystallising)
(Ph. Eur. monograph 0436)



Liquid Sorbitol (Crystallising)

Ph Eur

DEFINITION

Aqueous solution of a hydrogenated, partly hydrolysed starch.

Content

— anhydrous substance: 68.0 per cent m/m to 72.0 per cent m/m,
— D-glucitol (D-sorbitol, C₆H₁₄O₆): 92.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, colourless, syrupy liquid, miscible with water.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).
B. To 7.0 g add 40 mL of water R and 6.4 g of disodium tetraborate R, allow to stand for 1 h, shaking occasionally, and dilute to 50.0 mL with water R. Filter if necessary. The angle of rotation (2.2.7) is 0° to +1.5°.
C. It is a clear, syrupy liquid at a temperature of 25 °C.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dilute 7.0 g to 50 mL with water R.

Conductivity (2.2.38)
Maximum 10 µS·cm⁻¹ measured on the undiluted liquid sorbitol (crystallising) while gently stirring with a magnetic stirrer.
Reducing sugars
Maximum 0.2 per cent calculated as glucose equivalent.
To 5.0 g add 6 mL of water R, 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R and, when the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.
Lead (2.4.10)
Maximum 0.5 ppm.
Nickel (2.4.15)
Maximum 1 ppm.
Water (2.5.12)
28.0 per cent to 32.0 per cent m/m, determined on 0.1 g.

ASSAY

Liquid chromatography (2.2.29).
Test solution Mix 1.00 g of the substance to be examined with 20 mL of water R and dilute to 50.0 mL with the same solvent.
Reference solution (a) Dissolve 65.0 mg of sorbitol CRS in 2 mL of water R and dilute to 5.0 mL with the same solvent.
Reference solution (b) Dissolve 65 mg of mannitol R and 65 mg of sorbitol R in 2 mL of water R and dilute to 5.0 mL with the same solvent.

Column:

— size: l = 0.3 m, Ø = 7.8 mm,
— stationary phase: strong cation-exchange resin (calcium form) R (9 µm),
— temperature: 85 ± 1 °C.

Mobile phase Degassed water R.

Flow rate 0.5 mL/min.

Detection Refractometer maintained at a constant temperature (for example, 35 °C).
Injection 20 µL.

Run time Twice the retention time of sorbitol.

Relative retention With reference to sorbitol (retention time = about 27 min): mannitol = about 0.8.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to mannitol and to sorbitol.
Calculate the percentage content of D-sorbitol taking into account the assigned content of sorbitol CRS.

Ph Eur

Liquid Sorbitol (Non-crystallising)

Sorbitol Solution (70 per cent) (Non-crystallising)

(Ph. Eur. monograph 0437)

Action and use

Excipient.

DEFINITION

Aqueous solution of a hydrogenated, partly hydrolysed starch.

Content

— anhydrous substance: 68.0 per cent *m/m* to

72.0 per cent *m/m*,

— D-glucitol (D-sorbitol, $C_6H_{14}O_6$): 72.0 per cent to 92.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, colourless, syrupy liquid, miscible with water.

IDENTIFICATION

A. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (a).

B. To 7.0 g add 40 mL of water R and 6.4 g of disodium

terborate R. Allow to stand for 1 h, shaking occasionally,

and dilute to 50.0 mL with water R. Filter if necessary.

C. It is a clear, syrupy liquid at 25 °C.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dilute 7.0 g to 50 mL with water R.

Conductivity (2.2.38)

Maximum $10 \mu S \cdot cm^{-1}$ measured on the undiluted liquid sorbitol (non crystallising) while gently stirring with a magnetic stirrer.

Reducing sugars

Maximum 0.2 per cent calculated as glucose equivalent.

To 5.0 g add 6 mL of water R, 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R and, when the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.

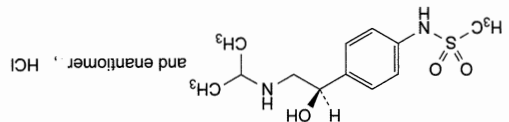
Reducing sugars after hydrolysis

Maximum 9.3 per cent calculated as glucose equivalent. To 6.0 g add 35 mL of water R, 40 mL of 1 M hydrochloric acid and a few glass beads. Boil under a reflux condenser for 4 h. Cool and neutralise with dilute sodium hydroxide solution R using 0.2 mL of bromothymol blue solution R1 as indicator. Cool and dilute to 100.0 mL with water R. To 3.0 mL of the solution add 5 mL of water R, 20 mL of cupri-citric solution R and a few glass beads. Heat so that



Sotalol Hydrochloride

(Ph. Eur. monograph 2004)


 $C_{12}H_{21}ClN_2O_3S$ 308.8 959-24-0

Action and use

Beta-adrenoceptor antagonist; class II and class III antiarrhythmic.

DEFINITION

N-[4-[(1*R*)-1-*H*-hydroxy-2-[(1-methylethyl)amino]ethyl]phenyl]methanesulfonamide hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, soluble in alcohol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison *sotalol hydrochloride CRS*.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in carbon dioxide-free water *R* and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension III (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

pH (2.2.3)

4.0 to 5.0.

Dilute 5.0 mL of solution S to 10.0 mL with carbon dioxide-free water *R*.

Optical rotation (2.2.7)

−0.10° to +0.10°.

Dilute 25.0 mL of solution S to 50.0 mL with water *R*.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 3.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 8.0 mg of *sotalol impurity B CRS* in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 100.0 mL with the mobile phase.

Reference solution (d) Dilute 1.5 mL of reference solution (b) to 100 mL with the mobile phase. To 1 mL of this solution add 1 mL of reference solution (a).

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography *R* (5 μ m).

Mobile phase Dissolve 2 g of sodium octanesulfonate *R* in 790 mL of water *R*. Adjust to pH 3.0 with phosphoric acid *R* and add 210 mL of acetonitrile *R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 228 nm.

Injection 10 μ L; inject the test solution and reference solutions (a), (c) and (d).

Run time 2.5 times the retention time of *sotalol*.

System suitability Reference solution (d):

— resolution: minimum 4.0 between the peaks due to *sotalol* and to *impurity B*.

Limits:

— *impurity B*: not more than 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent);

— any other *impurity*: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent), and not more than 1 such peak has an area greater than 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— total of other *impurities*: not more than 1.65 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— *disregard limit*: 0.17 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Palladium

Maximum 0.5 ppm.

Atomic absorption spectrometry (2.2.23, Method I).

Test solution Dissolve 1.00 g in a mixture of 0.25 volumes of nitric acid *R*, 0.75 volumes of hydrochloric acid *R* and 99.0 volumes of water *R* and dilute to 20.0 mL with the same mixture of solvents.

Reference solutions Use solutions containing 0.02 μ g, 0.03 μ g and 0.05 μ g of palladium per millilitre, freshly prepared by dilution of palladium standard solution (0.5 ppm Pd) *R* with a mixture of 0.25 volumes of nitric acid *R*, 0.75 volumes of hydrochloric acid *R* and 99.0 volumes of water *R*.

Source Palladium hollow-cathode lamp.

Wavelength 247.6 nm.

Use a graphite tube.

Heavy metals (2.4.8)

Maximum 20 ppm.

To 10 mL of solution S add 10 mL of water *R*. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) *R*.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

In order to avoid overheating during the titration, mix thoroughly throughout and stop the titration immediately after the end-point has been reached.

Dissolve 0.250 g in 10 mL of anhydrous formic acid R, if

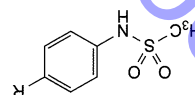
necessary with the aid of ultrasound. Add 40 mL of acetic anhydride R and titrate immediately with 0.1 M perchloric acid. Determine the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 30.88 mg of $C_{12}H_{21}ClN_2O_3S$.

STORAGE

Protected from light.

IMPURITIES



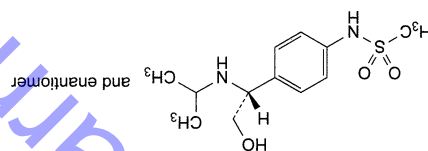
A. R = $CH_2-CH_2-NH-CH(CH_3)_2$; N-[4-[2-

[(1-methylethyl)amino]ethyl]phenyl]methanesulfonamide,

B. R = $CO-CH_2-NH-CH(CH_3)_2$; N-[4-

[(1-methylethyl)amino]acetyl]phenyl]methanesulfonamide,

C. R = CHO ; N-(4-formylphenyl)methanesulfonamide,



D. N-[4-[(1R)-2-hydroxy-1-

[(1-methylethyl)amino]ethyl]phenyl]methanesulfonamide.

Ph Eur

Hydrogenated Soya Oil

Hydrogenated Soyabean Oil

(Hydrogenated Soya-bean Oil, Ph Eur monograph 1265)

Action and use

Excipient.

Ph Eur

DEFINITION

Product obtained by refining, bleaching, hydrogenation and deodorisation of oil obtained from seeds of *Glycine max* (L.) Merr. (*G. hispida* (Moench) Maxim.). The product consists mainly of triglycerides of palmitic and stearic acids.

CHARACTERS

Appearance

White or almost white mass or powder which melts to a clear, pale yellow liquid when heated.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, in light petroleum (bp: 65–70 °C) after heating and in toluene, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Melting point (see Tests).

B. Composition of fatty acids (see Tests).

TESTS

Melting point (2.2.15)

66 °C to 72 °C.

Acid value (2.5.1)
Maximum 0.5.

Dissolve 10.0 g in 50 mL of a hot mixture of equal volumes of ethanol (96 per cent) R and toluene R, previously neutralised with 0.1 M potassium hydroxide using 0.5 mL of phenolphthalein solution R1 as indicator. Titrate the solution immediately while still hot.

Peroxide value (2.5.5, Method A)

Maximum 5.0.

Unsaponifiable matter (2.5.7)

Maximum 1.0 per cent, determined on 5.0 g.

Alkaline impurities (2.4.19)

Dissolve 2.0 g with gentle heating in a mixture of 1.5 mL of ethanol (96 per cent) R and 3 mL of toluene R. Add 0.05 mL of a 0.4 g/L solution of bromophenol blue R in ethanol

(96 per cent) R. Not more than 0.4 mL of 0.01 M hydrochloric acid is required to change the colour to yellow.

Composition of fatty acids (2.4.22, Method A)
Use the mixture of calibrating substances in Table 2.4.22.-3.

Column:

— material: fused silica;

— size: $l = 25$ m, $\varnothing = 0.25$ mm;

— stationary phase: poly(cyanopropyl)siloxane R (film thickness

0.2 μ m).

Carrier gas helium for chromatography R.

Flow rate 0.65 mL/min.

Split ratio 1:100.

Temperature:

— column: 180 °C for 20 min;

— injection port and detector: 250 °C.

Detection Flame ionisation.

Composition of the fatty-acid fraction of the oil:

— saturated fatty acids of chain length less than C_{14} : maximum

0.1 per cent;

— myristic acid: maximum 0.5 per cent;

— palmitic acid: 9.0 per cent to 16.0 per cent;

— stearic acid: 79.0 per cent to 89.0 per cent;

— oleic acid and isomers: maximum 4.0 per cent;

— linoleic acid and isomers: maximum 1.0 per cent;

— arachidic acid: maximum 1.0 per cent;

— behenic acid: maximum 1.0 per cent.

Nickel

Maximum 1 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Introduce 5.0 g into a platinum or silica crucible,

previously tared after calcination. Cautiously heat and

introduce into the substance a wick formed from twisted

ashless filter paper. Light the wick. When the substance is

alight stop heating. After combustion, ignite in a muffle

furnace at about 600 \pm 50 °C. Continue the ignition until

white ash is obtained. After cooling, take up the residue with

2 quantities, each of 2 mL, of dilute hydrochloric acid R and

transfer into a 25 mL graduated flask. Add 0.3 mL of nitric

acid R and dilute to 25.0 mL with water R.

Reference solutions Prepare 3 reference solutions by adding

1.0 mL, 2.0 mL and 4.0 mL of nickel standard solution

(0.2 ppm Ni) R to 2.0 mL of the test solution and diluting to

10.0 mL with water R.

Source Nickel hollow-cathode lamp.
Wavelength 232 nm.
Atomisation device Graphite furnace.
Carrier gas argon R.
STORAGE
Protected from light.

Ph Eur

Refined Soya Oil



Refined Soyabean Oil
(Refined Soya-bean Oil, Ph Eur monograph 1473)

Action and use
Excipient.
When Soya Oil, Soyabean Oil or Soya-bean Oil is demanded,

Refined Soya Oil shall be supplied.

When intended for use in the manufacture of a parenteral dosage form, Refined Soya Oil complying with the

requirement for Water below should be used.

DEFINITION

Fatty oil obtained from seeds of *Glycine max* (L.) Merr. (*Glycine hispida* (Moench) Maxim.) by extraction and subsequent refining. It may contain a suitable antioxidant.

CHARACTERS

Appearance
Clear, pale yellow liquid.

Solubility

Practically insoluble in ethanol (96 per cent), miscible with light petroleum (bp: 50-70 °C).

Relative density

About 0.922.
Refractive index About 1.475.

IDENTIFICATION

Identification of fatty oils by thin-layer chromatography (2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2-1.

TESTS

Acid value (2.5.1)
Maximum 0.5.

Peroxide value (2.5.5, Method A)
Maximum 10.0, and maximum 5.0 if intended for use in the

manufacture of parenteral preparations.

Unsaturation matter (2.5.7)
Maximum 1.5 per cent, determined on 5.0 g.

Alkaline impurities (2.4.19)
It complies with the test.

Composition of fatty acids (2.4.22, Method A)
Use the mixture of calibrating substances in Table 2.4.22-3.

Composition of the fatty-acid fraction of the oil:
— saturated fatty acids of chain length less than C₁₄: maximum 0.1 per cent;

— myristic acid: maximum 0.2 per cent;

— palmitic acid: 9.0 per cent to 13.0 per cent;

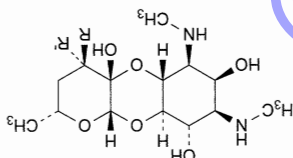
— palmitoleic acid: maximum 0.3 per cent;

— stearic acid: 2.5 per cent to 5.0 per cent;

Spectinomycin Dihydrochloride



(Ph. Eur. monograph 1152)



Compound	R	R'	Molec. Formula	M _r
spectinomycin	R + R' = O		C ₁₄ H ₂₆ Cl ₂ N ₂ O ₇ .5H ₂ O	495.4
(4R)-dihydro-spectinomycin	OH	H	C ₁₄ H ₂₈ Cl ₂ N ₂ O ₇ .5H ₂ O	497.4

Action and use

Aminocyclitol antibacterial.

Ph Eur

DEFINITION

Mixture of (2R,4aR,5aR,6S,7S,8R,9S,9aR,10aS)4a,7,9-trihydroxy-2-methyl-6,8-bis(methylamino)decahydro-4H-pyranol[2,3-b][1,4]benzodioxin-4-one dihydrochloride and of (2R,4R,4aS,5aR,6S,7S,8R,9S,9aR,10aS)-2-methyl-6,8-bis(methylamino)decahydro-2H-pyranol[2,3-b][1,4]benzodioxin-4,4a,7,9-tetrol dihydrochloride pentahydrate ((4R)-dihydro-spectinomycin dihydrochloride pentahydrate).
It is produced by *Streptomyces spectabilis* or by any other means.

Content

— (4R)-dihydro-spectinomycin dihydrochloride: maximum 9.0 per cent (anhydrous substance);
— sum of the contents of spectinomycin dihydrochloride and (4R)-dihydro-spectinomycin dihydrochloride: 93.0 per cent to 102.0 per cent (anhydrous substance).

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

STORAGE

In a well-filled container, protected from light, at a temperature not exceeding 25 °C.

Water

Maximum 0.1 per cent, determined on 1.00 g.

Brassicasterol

Maximum 0.3 per cent in the sterol fraction of the oil.

oleic acid

17.0 per cent to 30.0 per cent;

— linoleic acid: 48.0 per cent to 58.0 per cent;

— linolenic acid: 5.0 per cent to 11.0 per cent;

— arachidic acid: maximum 1.0 per cent;

— eicosenoic acid: maximum 1.0 per cent;

— behenic acid: maximum 1.0 per cent.

CHARACTERS**Appearance**

White or almost white, slightly hygroscopic powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison spectinomycin hydrochloride CRS.

B. Dilute 1.0 mL of solution S (see Tests) to 10 mL with water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2,

Method II).

Dilute 2.0 mL of solution S to 20.0 mL with water R.

pH (2.2.3)

3.8 to 5.6 for solution S.

Specific optical rotation (2.2.7)

+15.0 to +21.0 (anhydrous substance), determined on solution S within 20 min of preparation.

Related substances

Liquid chromatography (2.2.29). In order to avoid formation of anomers, prepare the solutions immediately before use.

Test solution Dissolve 15.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 3 mg of spectinomycin for system suitability CRS in the mobile phase and dilute to 20 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 3.0 mL of reference solution (b) to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R

(5 μ m);

— temperature: ambient and constant.

Mobile phase Dissolve 4.2 g of oxalic acid R and 2.0 mL of heptafluorobutyric acid R in water R and dilute to 1000 mL with water R; adjust to pH 3.2 with sodium hydroxide solution R; add 105 mL of acetonitrile R and mix; filter through a 0.45 μ m filter and degas with helium for chromatography R for 10 min.

Flow rate 1.0 mL/min.

Post-column solution carbonate-free sodium hydroxide solution R diluted with carbon dioxide-free water R to obtain a final concentration of NaOH of 21 g/L. Degas the solution with helium for chromatography R for 10 min before use. Add it pulse-less to the column effluent using a 375 μ L polymeric mixing coil.

Post-column flow rate 0.5 mL/min.

Detection Pulsed amperometric detection or equivalent with a gold indicator electrode having preferably a diameter of 1.4 mm or greater, a suitable reference electrode and a stainless steel counter electrode, held at +0.12 V detection, +0.70 V oxidation and -0.60 V reduction potentials

respectively, with pulse durations according to the instrument used. Keep the detection cell at ambient and constant temperature. Clean the gold indicator electrode with an eraser and damp precision wipe prior to start-up of the system to enhance the detector sensitivity and increase the signal-to-noise ratio.

Injection 20 μ L.

Run time 1.5 times the retention time of spectinomycin.

Identification of impurities Use the chromatogram supplied with spectinomycin for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, D and E.

Relative retention With reference to spectinomycin (retention time = 11 min to 20 min): impurity A = about 0.5;

impurity F = about 0.53; impurity G = about 0.6; impurity D = about 0.7; impurity E = about 0.9;

(4R)-dihydrospectinomycin = about 1.3;

impurity C = about 1.4.

System suitability: reference solution (a):

— resolution: minimum 1.5 between the peaks due to impurity E and spectinomycin.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity A by 0.4;

— impurities A, C, F, G: for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (b) (1.0 per cent);

— impurities D, E: for each impurity, not more than 4 times the area of the principal peak in the chromatogram

obtained with reference solution (b) (4.0 per cent);

— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— total: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (6.0 per cent);

— disregard limit: the area of the principal peak in the chromatogram obtained with reference solution (c) (0.3 per cent); disregard the peak due to (4R)-dihydrospectinomycin.

Water (2.5.12)

16.0 per cent to 20.0 per cent, determined on 0.100 g.

Sulfated ash (2.4.14)

Maximum 1.0 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 0.09 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins. Prepare the solutions using a 0.42 per cent *m/m* solution of sodium hydroxigen carbonate R.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Test solution Dissolve 40.0 mg of the substance to be examined in water R and dilute to 50.0 mL with the same solvent. Allow to stand for not less than 15 h and not more than 72 h (formation of anomers). Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution Dissolve 40.0 mg of spectinomycin hydrochloride CRS (containing (4R)-dihydrospectinomycin) in water R and dilute to 50.0 mL with the same solvent. Allow to stand for the same period of time as the test solution (formation of anomers). Dilute 5.0 mL of this solution to 50.0 mL with the mobile phase.

System suitability:

— *repeatability*: maximum relative standard deviation of 3.0 per cent for the principal peak after 6 injections of the reference solution.

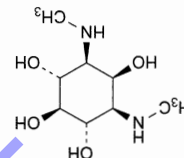
STORAGE

In an airtight container. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

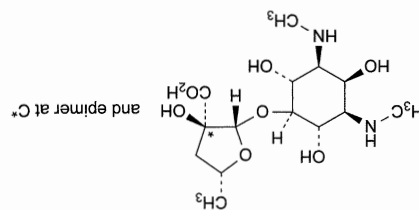
IMPURITIES

Specified impurities A, C, D, E, F, G

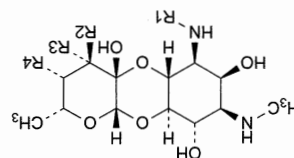
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. *Control of impurities in substances for pharmaceutical use*): B.



A. 1,3-dideoxy-1,3-bis(methylamino)-*myo*-inositol (actinamine),



B. (2*S*,3*R*,5*R*)-3-*hydroxy*-5-methyl-2-[[[(1*r*,2*R*,3*S*,4*r*,5*R*,6*S*)-2,4,6-trihydroxy-3,5-bis(methylamino)cyclohexyl]oxy]tetrahydrofuran-3-carboxylic acid (actinospectroic acid),

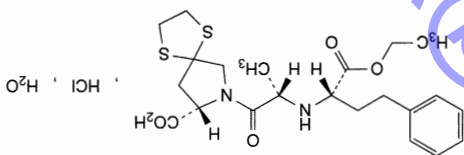


$(2R,4S,4aS,5aR,6S,7S,8R,9S,9aR,10aS)-2\text{-methyl-6,8-bis(methylamino)decahydrot-2H-pyranol[2,3-}b[1,4]\text{benzodioxine-4',4a,7,9-tetrol ((4S)-dihydrorespecinomycin)}$
 D. $R1 = CH_3$, $R2 = H$, $R3 = R4 = OH$:
 $(2R,4S,4aS,5aR,6S,7S,8R,9S,9aR,10aS)-2\text{-methyl-6,8-bis(methylamino)decahydrot-2H-pyranol[2,3-}b[1,4]\text{benzodioxine-4',4a,7,9-pentol(dihydroxyrespecinomycin)}$
 E. $R1 = R4 = H$, $R2 + R3 = O$:
 $(2R,4aR,5aR,6S,7R,8R,9S,9aR,10aS)-6\text{-amino-4a,7,9-}$

Spirapril Hydrochloride Monohydrate



(Ph. Eur. monograph 1766)


$$\text{C}_{22}\text{H}_{31}\text{ClN}_2\text{O}_5\text{S}_2\cdot\text{H}_2\text{O}$$

Action and use

Angiotensin-converting enzyme inhibitor.

DEFINITION

(8S)-7-[[[(2S)-2-[[[(1S)-1-(Ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]-1,4-dithia-7-azaspiro[4.4]nonane-8-carboxylic acid hydrochloride monohydrate.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, fine crystalline powder.

Solubility

Very slightly soluble in water, soluble in methanol, slightly soluble in acetonitrile, practically insoluble in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).
B. Infrared absorption spectrophotometry (2.2.24).
Preparation Discs of potassium bromide R.
Comparison *sphrapril hydrate monohydrate* CRS.
C. It gives the reactions of chlorides (2.3.1).

TESTS**Specific optical rotation (2.2.7)**

— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **disregard limit:** area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent); disregard any peak due to the blank (solvent mixture).

Water (2.5.12)

3.0 per cent to 4.0 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29).

Solvent mixture Mix equal volumes of acetonitrile R1 and water R.

Test solution Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of spirapril hydrochloride monohydrate CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 6.0 mg of spirapril for system suitability CRS (spirapril spiked with impurity B and impurity D) in a mixture of 2 volumes of acetonitrile R and 8 volumes of water R and dilute to 20 mL with the same mixture of solvents.

Solution A Dissolve 4.5 g of tetramethylammonium hydroxide R in 900 mL of water R, adjust to pH 1.75 with phosphoric acid R and add 100 mL of acetonitrile R1.

Solution B Dissolve 4.5 g of tetramethylammonium hydroxide R in 400 mL of water R, adjust to pH 1.75 with phosphoric acid R and add 600 mL of acetonitrile R1.

Column:

— **size:** $l = 0.125$ m, $\phi = 4.6$ mm;

— **stationary phase:** octadecylsilyl silica gel for chromatography R (5 μ m);

— **temperature:** 70 °C.

Mobile phase Solution A, solution B (45:55 V/V).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L.

Retention time Spirapril = 1.6 min to 2.9 min; impurity D = about 13 min. Adjust the proportion of solution B in the mobile phase if necessary.

System suitability: reference solution (b);

— **resolution:** minimum 1.5 between the peaks due to spirapril and impurity D.

Calculate the percentage content of $C_{22}H_{31}ClN_2O_5S_2$ from the chromatograms obtained with the test solution and reference solution (a) and the declared content of spirapril hydrochloride monohydrate CRS.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities: A, B, C, D.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R1, water R (2:8 V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 6 mg of spirapril for system suitability CRS (containing impurities B and D) in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 5.0 mL of this solution to 50.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with the solvent mixture.

Column:

— **size:** $l = 0.125$ m, $\phi = 4.6$ mm;

— **stationary phase:** octadecylsilyl silica gel for chromatography R (5 μ m);

— **temperature:** 70 °C.

Mobile phase:

— **mobile phase A:** dissolve 4.5 g of tetramethylammonium hydroxide R in 900 mL of water R, add 100 mL of acetonitrile R1 and adjust to pH 2.2 with phosphoric acid R;

— **mobile phase B:** dissolve 4.5 g of tetramethylammonium hydroxide R in 400 mL of water R, add 600 mL of acetonitrile R1 and adjust to pH 2.2 with phosphoric acid R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 4	90	10
4 - 14	90 \rightarrow 10	10 \rightarrow 90
14 - 20	10	90

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L.

Relative retention With reference to spirapril (retention time = about 10 min): impurity C = about 0.6;

impurity B = about 0.7; impurity A = about 1.26; impurity D = about 1.38.

System suitability: reference solution (a);

— **resolution:** minimum 3.5 between the peaks due to impurity B and spirapril, and minimum 5.5 between the peaks due to spirapril and impurity D.

Limits:

— **impurity D:** not more than 0.8 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— **impurity B:** not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— **impurities A, C:** for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— **unspecified impurities:** for each impurity, not more than 0.2 times the area of the principal peak in the

CHARACTERS

Appearance

White or yellowish-white powder.

Solubility

Practically insoluble in water, soluble in ethanol

It shows polymorphism (5.9).

IDENTIFICATION

First identification A.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison spironolactone CRS.

If the spectra obtained in the solid state shows differences,

dissolve the substance to be examined and the reference

substance separately in the minimum volume of methanol R,

evaporate to dryness and record new spectra using the

residues.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined

in methylene chloride R and dilute to 10 mL with the same

solvent.

Reference solution Dissolve 20 mg of spironolactone CRS in

methylene chloride R and dilute to 10 mL with the same

solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase water R, cyclohexane R, ethyl acetate R

(1:24:75 V/V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

C. To about 10 mg add 2 mL of a 50 per cent V/V solution

of sulfuric acid R and shake. An orange solution with an

intense yellowish-green fluorescence is produced. Heat the

solution gently; the colour becomes deep red and hydrogen

sulfide, which blackens lead acetate paper R, is evolved.

Add the solution to 10 mL of water R; a greenish-yellow

solution is produced, showing opalescence or a precipitate.

TESTS

Specific optical rotation (2.2.7)

Dissolve 0.100 g in ethanol (96 per cent) R and dilute to

10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use.

Solvent mixture acetonitrile R, water R (50:50 V/V).

Test solution (a) Dissolve 50.0 mg of the substance to be

examined in 2.5 mL of tetrahydrofuran R and dilute to

25.0 mL with the solvent mixture.

Test solution (b) Dilute 1.0 mL of test solution (a) to

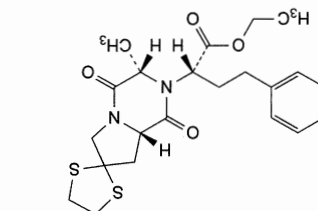
100.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of test solution (b) to

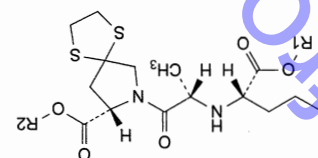
10.0 mL with the solvent mixture.

Reference solution (b) Dissolve with the aid of ultrasound the

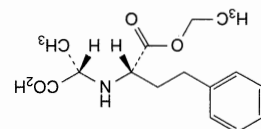
contents of a vial of spironolactone for system suitability CRS



A. ethyl (2S)-2-[(3'S,8'S)-3'-methyl-1',4'-dioxohexahydrospiro[1,3'-dithiolane-2,7'(6'H)-pyrrolo[1,2-a]pyrazin]-2'-yl]-4-phenylbutanoate,



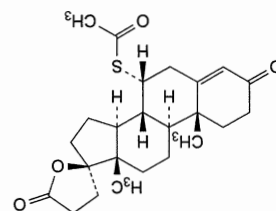
B. R1 = R2 = H: (8S)-7-[(2S)-2-[(1S)-1-carboxy-3-phenylpropyl]amino]propanoyl]-1,4-dithia-7-azaspiro[4.4]nonane-8-carboxylic acid (spiraprilat),
D. R1 = C₂H₅, R2 = CH(CH₃)₂: 1-methylethyl (8S)-7-[(2S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]-1,4-dithia-7-azaspiro[4.4]nonane-8-carboxylate,



C. (2S)-2-[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoic acid.

Spironolactone

(Ph. Eur. monograph 0688)



C₂₄H₃₂O₅

416.6

52-01-7



Ph Eur

Action and use

Aldosterone receptor antagonist; potassium-sparing diuretic.

Preparations

Spironolactone Oral Suspension

Spironolactone Tablets

Ph Eur

DEFINITION

(2'R)-7α-(Acetylsulfonyl)-3',4'-dihydro-5'H-spiro[androsta-4-ene-1,7,2'-furan]-3,5'-dione.

Content

97.5 per cent to 102.0 per cent (dried substance).

(containing impurities A, C, D, E and I) in 1.0 mL of the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of spiroinolactone CRS

in 2.5 mL of tetrahydrofuran R and dilute to 25.0 mL with

the solvent mixture. Dilute 1.0 mL of this solution to

100.0 mL with the solvent mixture.

Reference solution (d) Dissolve 5.0 mg of camphore CRS

(impurity F) in 2.5 mL of tetrahydrofuran R and dilute to

25.0 mL with the solvent mixture. Dilute 3.0 mL of this

solution to 100.0 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (3 μ m);

— temperature: 40 °C.

Mobile phase acetonitrile R, tetrahydrofuran R, methanol R1,

water R (15:20:42.5:54.0 V/V/V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of test solution (a) and reference solutions (a),

(b) and (d).

Run time 2.5 times the retention time of spiroinolactone.

Identification of impurities Use the chromatogram supplied

with spiroinolactone for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, C, D, E and I;

use the chromatogram obtained with reference solution (d) to

identify the peak due to impurity F.

Relative retention With reference to spiroinolactone (retention

time = about 26 min): impurity A = about 0.95;

impurity F = about 1.2; impurity C = about 1.5;

impurity D = about 1.6; impurity E = about 1.7;

impurity I = about 1.9.

System suitability: reference solution (b):

— peak-to-valley ratio: minimum 1.5, where H_p = height

above the baseline of the peak due to impurity A and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

spiroinolactone.

Limits:

— **correction factor:** for the calculation of content, multiply the

peak area of impurity F by 2.3;

— **impurity I:** not more than 5 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.5 per cent);

— **impurities E, F:** for each impurity, not more than 3 times

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.3 per cent);

— **impurities A, C:** for each impurity, not more than twice

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

— **impurity D:** not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.15 per cent);

— **unspecified impurities:** for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— **total:** not more than 7 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.7 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Free thiol compounds
To 2.0 g add 20 mL of water R, shake for 1 min and filter.
To 10 mL of the filtrate add 0.05 mL of 0.01 M iodine and
0.1 mL of starch solution R and mix. A blue colour develops.

Chromium

Maximum 50 ppm.

To 0.20 g in a platinum crucible add 1 g of potassium

carbonate R and 0.3 g of potassium nitrate R. Heat gently until

fused, and ignite at 600–650 °C until carbon is removed.

Cool, dissolve the residue as completely as possible in 10 mL

of water R with the aid of gentle heat, filter, and dilute to

20 mL with water R. To 10 mL of this solution add 0.5 g of

urea R, and add a 14 per cent V/V solution of sulfuric acid R

until the solution is just acid. When effervescence ceases, add

a further 1 mL of the 14 per cent V/V solution of sulfuric

acid R, dilute to 20 mL with water R and add 0.5 mL of

diphenylcarbazide solution R. The solution is not more

intensely coloured than a standard prepared by adding 1 mL

of a 14 per cent V/V solution of sulfuric acid R to 0.50 mL of

a freshly prepared 28.3 mg/L solution of potassium

dichromate R, diluting to 20 mL with water R and adding

0.5 mL of diphenylcarbazide solution R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{24}H_{32}O_4S$ from the

declared content of spiroinolactone CRS.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A, C, D, E, F, I

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

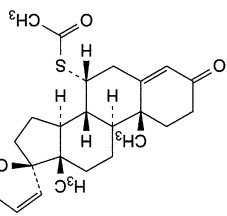
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

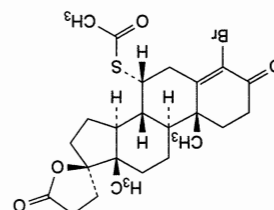
(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

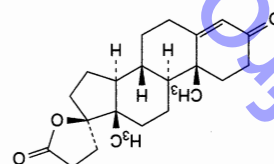
Control of impurities in substances for pharmaceutical use: B, G,



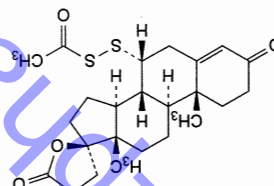
A. (2'R)-7 α -(acetylsulfonyl)-5'H-spiro[androst-4-ene-17,2'-furan]-3,5'-dione (Δ 20-spiroinolactone),



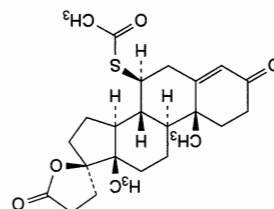
B. (2'R)-7α-(acetylsulfanyl)-4-bromo-3',4'-dihydro-5'H-spiro[androst-4-ene-17,2'-furan]-3,5'-dione (4-bromospironolactone),



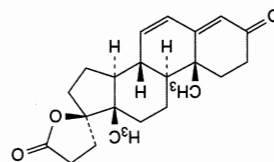
C. (2'R)-3',4'-dihydro-5'H-spiro[androst-4-ene-17,2'-furan]-3,5'-dione (aldone),



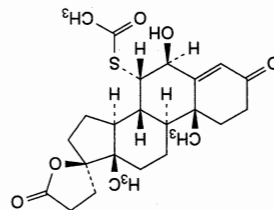
D. (2'R)-7α-(acetyldisulfanyl)-3',4'-dihydro-5'H-spiro[androst-4-ene-17,2'-furan]-3,5'-dione (disulfanyl-spiro[androst-4-ene-17,2'-furan]-3,5'-dione),



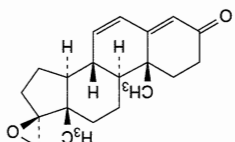
E. (2'R)-7β-(acetylsulfanyl)-3',4'-dihydro-5'H-spiro[androst-4-ene-17,2'-furan]-3,5'-dione (7β-spiro[androst-4-ene-17,2'-furan]-3,5'-dione),



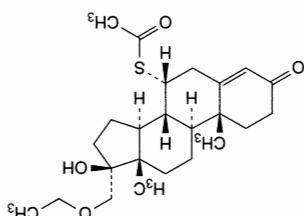
F. (2'R)-3',4'-dihydro-5'H-spiro[androst-4,6-diene-17,2'-furan]-3,5'-dione (canrenone),



G. (2'R)-7α-(acetylsulfanyl)-6β-hydroxy-3',4'-dihydro-5'H-spiro[androst-4-ene-17,2'-furan]-3,5'-dione (6β-hydroxy-spiro[androst-4-ene-17,2'-furan]-3,5'-dione),



H. (2'S)-spiro[androst-4,6-diene-17,2'-oxiran]-3-one,



I. S-[17α-(ethoxymethyl)-17-hydroxy-3-oxoandrost-4-en-7α-yl] ethanethioate.



Products with Risk of Transmitting Agents of Animal Spongiform Encephalopathies

(Ph. Eur. monograph 1483)

DEFINITION

Products with risk of transmitting agents of animal spongiform encephalopathies are those derived from tissues or secretions of animals susceptible to transmissible spongiform encephalopathies other than by experimental challenge. This monograph applies to all substances or preparations obtained from such animals and to all substances or preparations where products obtained from such animals are included as active substances or excipients or have been used during production, for example as raw or source materials, starting materials or reagents.

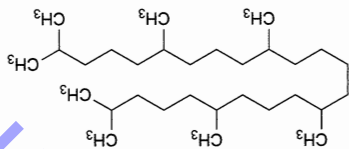
PRODUCTION

Production complies with chapter 5.2.8. Minimising the risk of transmitting animal spongiform encephalopathy agents via human and veterinary medicinal products.

Ph Eur

Squalane

(Ph. Eur. monograph 1630)



422.8

C₃₀H₆₂

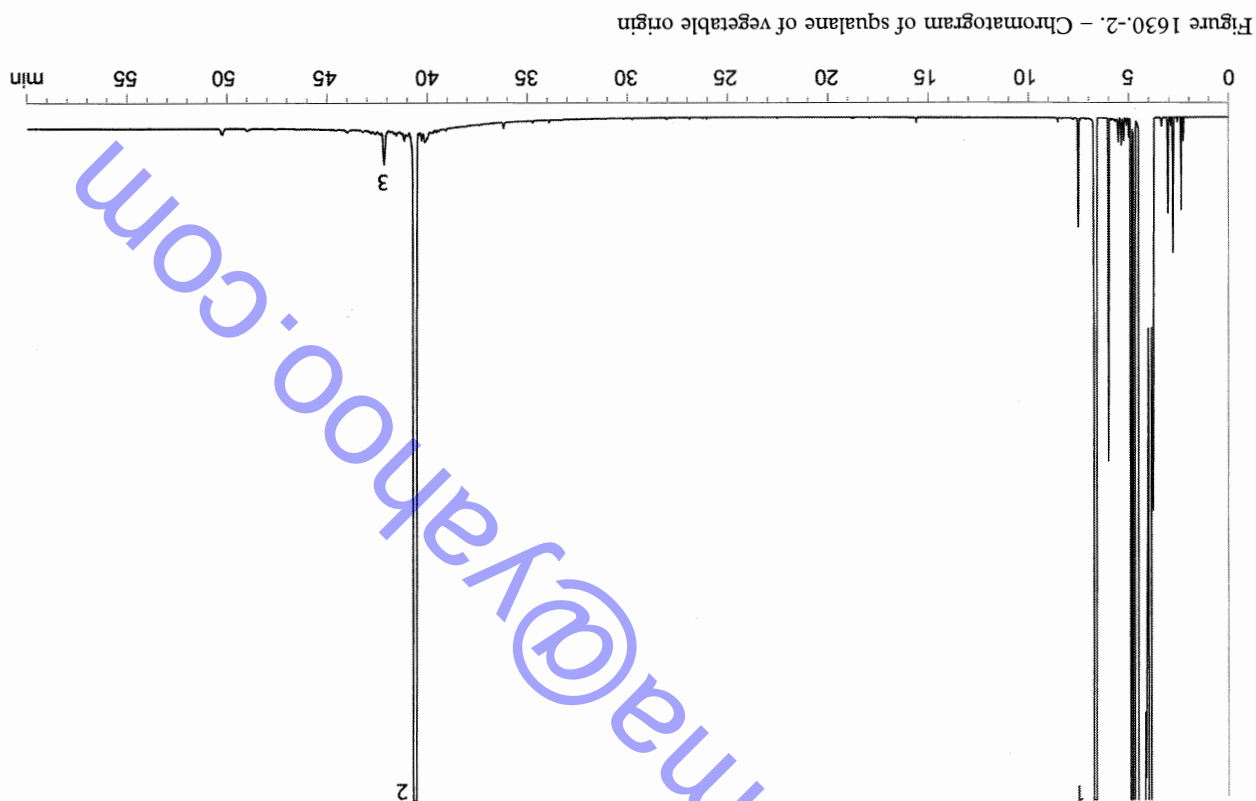
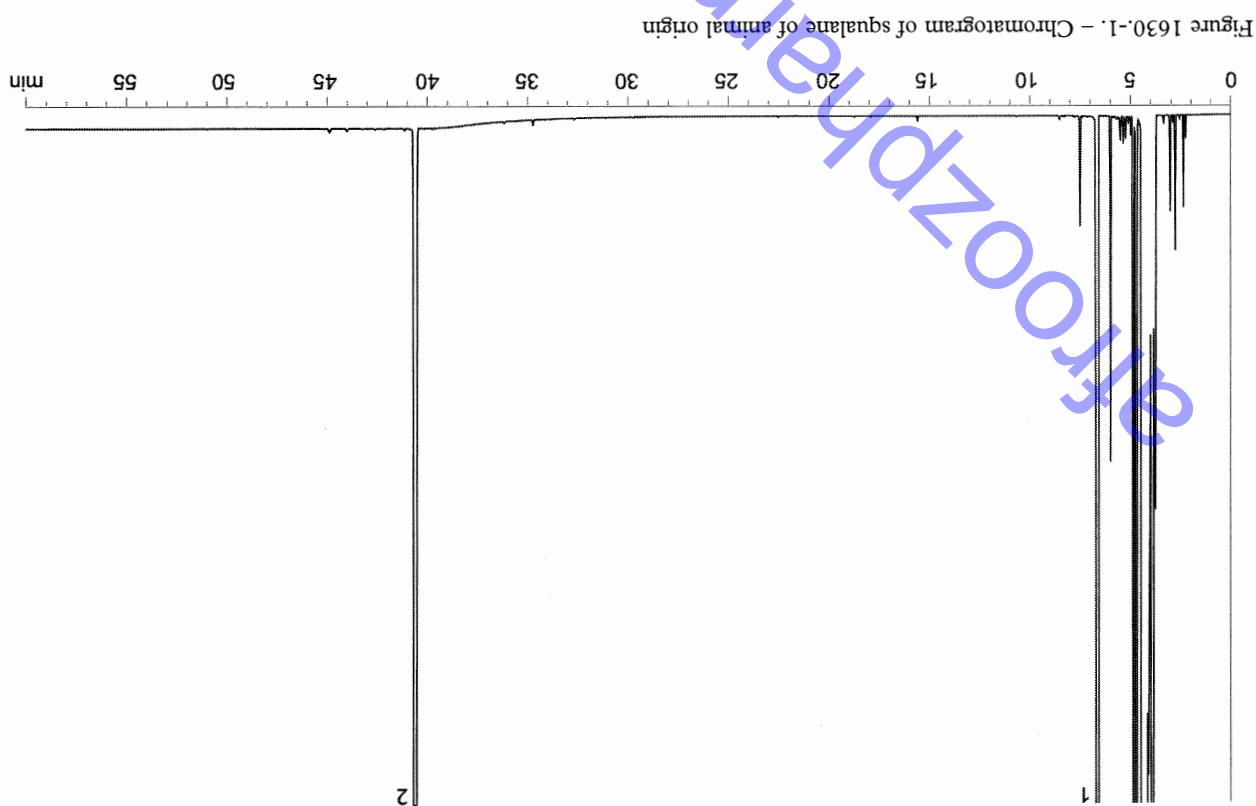
111-01-3

Action and use
Excipient; emollient.

Ph Eur

DEFINITION

2,6,10,15,19,23-Hexamethyltetracosane (perhydropentalene). It may be of vegetable (unsaponifiable matter of olive oil), animal (shark liver oil) or synthetic origin.

**CHARACTERS**

Content
96.0 per cent to 103.0 per cent.

Appearance
Clear, colourless, oily liquid.

Solubility

Practically insoluble in water, miscible with most fats and oils, freely soluble in acetone and in cyclohexane, practically insoluble in ethanol (96 per cent).

Relative density

About 0.815.

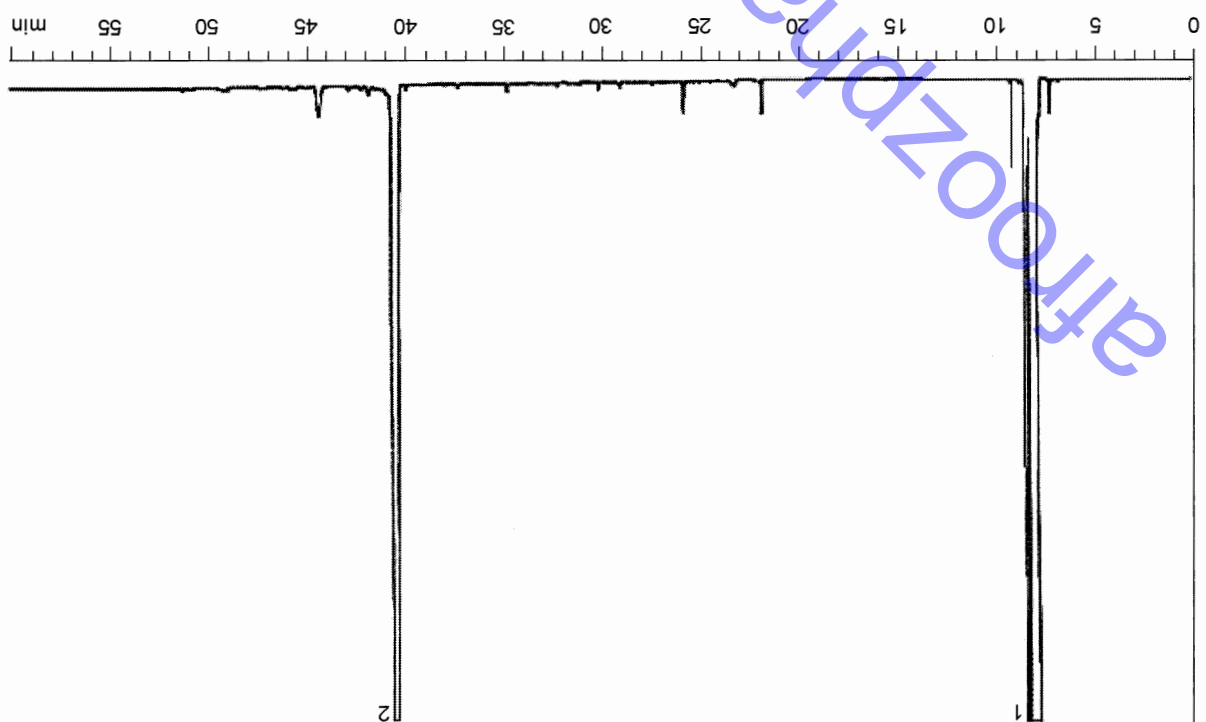


Figure 1630.-3. – Chromatogram of squalane of synthetic origin

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison squalane CRS.

B. Refractive index (see Tests).

C. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

The chromatogram obtained with squalane of animal origin does not show a peak due to cyclosqualane and does not show a peak with a relative retention of 1.1 with reference to squalane (Figure 1630.-1).

The chromatogram obtained with squalane of vegetable origin shows a peak due to cyclosqualane (Figure 1630.-2).

The chromatogram obtained with squalane of synthetic origin shows a peak with a relative retention of 1.1 with reference to squalane (Figure 1630.-3).

TESTS**Appearance**

The substance to be examined is clear (2.2.1) and colourless (2.2.2, Method II).

Refractive index (2.2.6)

1.450 to 1.454.

Acid value (2.5.1)

Maximum 0.2.

Iodine value (2.5.4, Method A)

Maximum 4.0.

Saponification value (2.5.6)

Maximum 3.0.

Nickel (2.4.31)

Maximum 1 ppm.

Time (min)	Temperature (°C)
0 - 39	60 - 290
39 - 50	290
Injection port	275
Detector	300

Temperature:

Split ratio 1:12.

Flow rate 1.7 mL/min.

Carrier gas helium for chromatography R.

1 µm).

— stationary phase: poly(dimethyl)siloxane R (film thickness

— size: l = 30 m, Ø = 0.32 mm;

— material: fused silica;

Column:

same solution.

internal standard solution and dilute to 25.0 mL with the

0.100 g of the substance to be examined, dissolve in the

Reference solution (b) To 0.1 mL of methyl erucate R add

same solution.

Reference solution (a) Dissolve 0.100 g of squalane CRS in the

internal standard solution and dilute to 25.0 mL with the

25.0 mL with the same solution.

Test solution Dissolve 0.100 g of the substance to be

examined in the internal standard solution and dilute to

(DMA), add 100.0 mL of heptane R.

Internal standard solution To 1.0 mL of dimethylacetamide R

Gas chromatography (2.2.28).

ASSAY

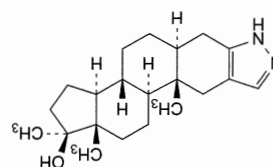
Maximum 0.5 per cent, determined on 1.000 g.

Total ash (2.4.16)

<p>Detection Flame ionisation.</p> <p>Injection 1 µL.</p> <p>Relative retention With reference to squalane (retention time = about 41 min): internal standard = about 0.2; methyl erucate = about 0.9; cyclosqualane = 1.05.</p> <p>System suitability: reference solution (b):</p> <p>— resolution: minimum 5 between the peaks due to methyl erucate and squalane.</p> <p>Calculate the percentage content of squalane taking into account the assigned content of <i>squalane CRS</i>.</p> <p>LABELLING</p> <p>The label states the origin of squalane (vegetable, animal or synthetic).</p> <p>Ph Eur</p> <p>Stannous Chloride Dihydrate</p> <p>(Ph. Eur. monograph 1266)</p> <p>$\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$</p> <p>225.6</p> <p>10025-69-1</p> <p>Ph Eur</p> <p>DEFINITION</p> <p>Content</p> <p>98.0 per cent to 102.0 per cent.</p> <p>CHARACTERS</p> <p>Appearance</p> <p>White or almost white, crystalline powder or colourless crystals, efflorescent in air.</p> <p>Solubility</p> <p>Freely soluble in water (the solution becomes cloudy after standing or on dilution), freely soluble in ethanol (96 per cent). It dissolves in dilute hydrochloric acid.</p> <p>IDENTIFICATION</p> <p>A. To 1 mL of solution S1 (see Tests) add 5 mL of water R and 0.05 mL of mercuric chloride solution R. A blackish-grey precipitate is formed.</p> <p>B. Dissolve 1.0 g in 3.0 mL of water R. Add 0.5 mL of dilute sodium hydroxide solution R to the cloudy solution; a yellowish flocculent precipitate is formed. Add 6.5 mL of water R. To 1.0 mL of the previously shaken suspension add 1.0 mL of strong sodium hydroxide solution R; the precipitate dissolves and the resulting solution is clear and colourless.</p> <p>C. Dissolve 10 mg in 2 mL of dilute nitric acid R. The solution gives reaction (a) of chlorides (2.3.1).</p> <p>TESTS</p> <p>Solution S1</p> <p>To 0.40 g add 1 mL of dilute hydrochloric acid R and dilute to 20 mL with distilled water R.</p> <p>Solution S2</p> <p>Dissolve 1.0 g in dilute hydrochloric acid R and dilute to 30 mL with the same acid. Heat to boiling. Add 30 mL of thioacetamide solution R and boil for 15 min (solution A). Take 5 mL, filter and heat to boiling. Add 5 mL of thioacetamide solution R and boil for 15 min. If a precipitate is formed, add the remainder of solution A (solution A') to the mixture. Add 10 mL of thioacetamide solution R and boil. Repeat the series of operations from "Take 5 mL" until a precipitate is no longer formed on addition of thioacetamide solution R to the filtrate obtained from the 5 mL of solution A (solution A', solution A'', etc. respectively). If no precipitate is formed or if no more precipitate is formed</p>	<p>combine the solution obtained with the remainder of solution A (solution A', solution A'', etc. respectively), filter and wash the precipitate with 10 mL of water R. Heat the filtrate until the resulting vapour no longer turns a moistened piece of lead acetate paper R blackish-grey. Allow to cool and dilute to 50 mL with water R.</p> <p>Appearance of solution</p> <p>The solution is clear (2.2.1) and colourless (2.2.2, Method II).</p> <p>Dissolve 10.0 g in dilute hydrochloric acid R and dilute to 20 mL with the same acid.</p> <p>Substances not precipitated by thioacetamide</p> <p>Maximum 0.2 per cent.</p> <p>Evaporate 25 mL of solution S2 to dryness and ignite at $600 \pm 50^\circ\text{C}$. The residue weighs a maximum of 1 mg.</p> <p>Sulfates (2.4.13)</p> <p>Maximum 500 ppm, determined on solution S1.</p> <p>Iron (2.4.9)</p> <p>Maximum 100 ppm.</p> <p>Dilute 5 mL of solution S2 to 10 mL with water R.</p> <p>Heavy metals</p> <p>Maximum 50 ppm.</p> <p>Dissolve 1.0 g in 2 mL of a mixture of 1 volume of nitric acid R and 3 volumes of hydrochloric acid R. Heat on a water-bath until nitrous vapour is no longer evolved. Dissolve the residue in water R and dilute to 25 mL with the same solvent. To 5 mL of this solution add 3 mL of strong sodium hydroxide solution R and 2 mL of water R. Heat until a clear solution is obtained, then cool and add 0.5 mL of thioacetamide reagent R. After 2 min, any colour in the solution is not more intense than that of a mixture of 1.0 mL of lead standard solution (10 ppm Pb) R, 6 mL of water R, 3 mL of strong sodium hydroxide solution R and 0.5 mL of thioacetamide reagent R.</p> <p>ASSAY</p> <p>Dissolve 0.100 g in 50 mL of water R, freed from oxygen by purging with carbon dioxide or nitrogen for 15 min. Add 1.5 mL of hydrochloric acid R1, 5 g of sodium potassium tartrate R, 10 g of sodium hydrogen carbonate R and 1 mL of starch solution R. Titrate immediately with 0.05 M iodine. Carry out a blank titration.</p> <p>1 mL of 0.05 M iodine is equivalent to 11.28 mg of $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$.</p> <p>STORAGE</p> <p>In an airtight container.</p> <p>Ph Eur</p>
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Stanazolol

(Ph. Eur. monograph 1568)



C₂₁H₃₂N₂O 328.5 10418-03-8

Action and use

Anabolic steroid; androgen.

Ph. Eur.

DEFINITION

17-Methyl-2'-H-5α-androst-2-eno[3,2-c]pyrazol-17β-ol.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Practically insoluble in water, soluble in dimethylformamide, slightly soluble in ethanol (96 per cent), very slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison stanazolol CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of methylene chloride R, evaporate at room temperature under an air-stream and record new spectra using the residues.

B. Liquid chromatography (2.2.29) as described in the test for related substances with the following modification. Injection Test solution and reference solution (c). Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

TESTS

Specific optical rotation (2.2.7)

Dissolve 60.0 mg in methanol R and dilute to 20.0 mL with the same solvent.

Impurities A and B

Thin-layer chromatography (2.2.27).

Solvent mixture methanol R₁, methylene chloride R (10:90 V/V).

Test solution Dissolve 20 mg of the substance to be examined in 1.0 mL of the solvent mixture.

Reference solution Dissolve 2 mg of stanazolol CRS, 2.0 mg of stanazolol impurity A CRS and 2.0 mg of stanazolol impurity B CRS in 1.0 mL of the solvent mixture. Dilute 0.1 mL of the solution to 2.0 mL with the solvent mixture.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, ethyl acetate R, cyclohexane R (2:48:50 V/V/V).

Application 10 μL.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with vanillin reagent R and heat at 120 °C.

System suitability: reference solution:

the chromatogram shows 3 clearly separated spots, due to stanazolol, impurity A and impurity B, in order of increasing R_F value.

Limits:

impurity A: any spot due to impurity A is not more intense than the corresponding spot in the chromatogram

obtained with the reference solution (0.5 per cent);

impurity B: any spot due to impurity B is not more intense than the corresponding spot in the chromatogram

obtained with the reference solution (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 15.0 mg of the substance to be examined in methanol R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (b) Dissolve 1 mg of stanazolol CRS and 1 mg of stanazolol impurity B CRS in methanol R and dilute to 20.0 mL with the same solvent.

Reference solution (c) Dissolve 15.0 mg of stanazolol CRS in methanol R and dilute to 5.0 mL with the same solvent.

Column:

size: l = 0.15 m, Ø = 4.6 mm;

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μm).

Mobile phase 1 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R, methanol R (30:70 V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 228 nm.

Injection 25 μL of the test solution and reference solutions (a) and (b).

Run time 3 times the retention time of stanazolol.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to stanazolol (retention time = about 12 min): impurity B = about 1.3.

System suitability: reference solution (b):

resolution: minimum 4.0 between the peaks due to stanazolol and impurity B.

Limits:

unspecific impurities: for each impurity, not more than area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying at 105 °C at a pressure not exceeding 0.7 kPa.

ASSAY

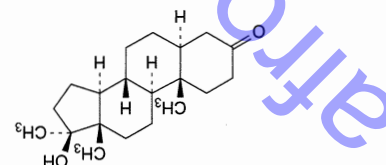
Dissolve 0.250 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 32.85 mg of $C_{21}H_{32}N_2O_7$.

STORAGE

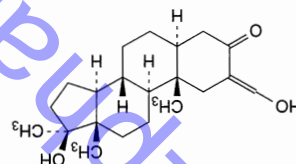
In an airtight container, protected from light.

IMPURITIES

Specified impurities A, B



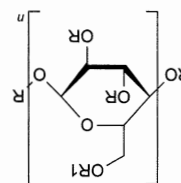
A. 17β-hydroxy-17-methyl-5α-androstan-3-one (mestanolone);



B. 17β-hydroxy-2-(hydroxymethyl)-17-methyl-5α-androstan-3-one (oxymetholone).

Hydroxyethyl Starches

(Ph. Eur. monograph 1785)



$$R = -[CH_2CH_2O]_nH \quad (n' = 0, 1, 2, \dots)$$

$$R1 = -[CH_2CH_2O]_{n'}H \quad (n'' = 0 \text{ or } 1) \text{ or glucose}$$

$$[C_6H_{10}O_5(C_2H_4O)_x]_n \text{ with } x = \text{molar substitution}$$

9005-27-0

DEFINITION

Hydroxyethyl starches are partially substituted poly(2-hydroxyethyl)ethers of waxy maize starch or potato starch, which primarily consist of amylopectine. The type of hydroxyethyl starch is defined by 2 numbers: the mean molecular weight (M_w) and the number of hydroxyethyl groups per anhydroglucose unit expressed as the molar substitution (MS). Hydroxyethyl starch is also characterised by the number of hydroxyethyl groups located at the C2 group over the number of hydroxyethyl groups located at the C6, expressed as the C2/C6 ratio. The parameters M_w , MS and C2/C6 ratio are determined by the reaction conditions of the production.

PRODUCTION

Hydroxyethyl starches are produced from waxy maize starch or potato starch by acidic hydrolysis and reaction with ethylene oxide and purified by ultrafiltration.

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Freely soluble in water and in dimethyl sulfoxide, practically insoluble in anhydrous ethanol.

Hydroxyethyl starches are hygroscopic until they reach a water content of about 12 per cent to 15 per cent.

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison medium M_w hydroxyethyl starch CRS.

Results The spectrum obtained shows the same absorption bands as the spectrum obtained with medium M_w

hydroxyethyl starch CRS. Due to the difference in the substitution of the substance, the intensity of some

absorption bands can vary.

B. To 5 mL of solution S (see Tests), add 0.1 mL of 0.05 M iodine. A reddish-brown or blue-violet colour appears.

C. Molecular weight (see Tests).

TESTS**Solution S**

Dissolve 5.0 g of the substance to be examined (dried substance) in carbon dioxide-free water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1).

pH (2.2.3)

4.5 to 7.0.

To 25 mL of solution S, add 0.2 mL of a saturated solution of potassium chloride R.

Absorbance (2.2.25)

Maximum 0.025, determined at 400 nm on solution S filtered through a 0.2 µm filter.

Molecular weight (M_w) and molecular weight distribution

Size-exclusion chromatography (2.2.30).

Buffer solution Dissolve 54.34 g of sodium acetate R in water R, add 100.0 mL of glacial acetic acid R and dilute to 1000.0 mL with water R.

Test stock solution Dissolve 2.0 g of the substance to be examined (dried substance) in water R and dilute to 50 mL with the same solvent. Add 10.0 mL of the buffer solution

and dilute to 100.0 mL with water R.

Reference solution (a) To prepare reference solution (a): — if the nominal M_w of the substance to be examined is below 300 000, use medium M_w hydroxyethyl starch CRS; — if the nominal M_w of the substance to be examined is above 300 000, use high M_w hydroxyethyl starch CRS.

Dissolve 0.4 g of medium M_w hydroxyethyl starch CRS or high M_w hydroxyethyl starch CRS in 10 mL of water R.

Add 2.0 mL of the buffer solution and dilute to 20.0 mL with water R.

Reference solution (b) Dilute 10.0 mL of reference solution (a) to 20.0 mL with the mobile phase.

- Reference solution (c) Dilute 10.0 mL of reference solution (b) to 20.0 mL with the mobile phase.
- Reference solution (d) Dilute 10.0 mL of reference solution (c) to 20.0 mL with the mobile phase.
- Column: — stationary phase: hydroxylated polymethacrylate gel R,
- 4 columns to be connected in series:

Length in m	Internal diameter in mm	Particle size in µm	Pore size in nm
0.30	7.5	17	> 100
0.30	7.5	17	100
0.30	7.5	10	20
0.30	7.5	10	12.5

Mobile phase Dilute 100.0 mL of the buffer solution to 1 L with water R.

Flow rate 0.5–1.0 mL/min.

Detection Multiple-angle laser light scattering detector and refractometer maintained at a constant temperature,

connected in series.

Injection volume 50 µL.

Determine the suitable working solution as follows: inject reference solutions (a) and (b), the mean *M_w* determined with reference solution (b) does not deviate by more than 3 per cent from the mean *M_w* determined with reference solution (a). If the deviation meets the requirement, use reference solution (a) to check the system suitability criterion. If the deviation is higher, inject reference solution (c) and determine the mean *M_w*. The mean *M_w* determined with reference solution (c) does not deviate by more than 3 per cent from the mean *M_w* determined with reference solution (b). If the deviation meets the requirement, use reference solution (b) to check the system suitability criterion. If the deviation is higher, inject reference solution (d) and determine the mean *M_w*. The mean *M_w* determined with reference solution (d) does not deviate by more than 3 per cent from the mean *M_w* determined with reference solution (c). If the deviation meets the requirement, use reference solution (c) to check the system suitability criterion.

System suitability: — mean *M_w*: within 5 per cent of the value assigned to the medium *M_w* hydroxyethyl starch CRS or high *M_w* hydroxyethyl starch CRS.

If necessary, dilute the test stock solution in order to have the same concentration as that of the reference solution used to check the system suitability.

Results Use a suitable integrator to determine the mean *M_w* and the *M_w* of the lowest and highest 10 per cent mass fraction.

Low <i>M_w</i> 2000 - 100 000	Medium <i>M_w</i> 100 000 - 300 000	High <i>M_w</i> 300 000 - 900 000
Determined <i>M_w</i> = nominal <i>M_w</i> ± 15 per cent		
<i>M_w</i> at 10 per cent lowest fraction > 10 per cent of nominal <i>M_w</i>	<i>M_w</i> at 10 per cent lowest fraction > 15 000	<i>M_w</i> at 10 per cent lowest fraction > 15 000
<i>M_w</i> at 10 per cent highest fraction < 300 per cent of nominal <i>M_w</i>	<i>M_w</i> at 10 per cent highest fraction < 300 per cent of nominal <i>M_w</i>	<i>M_w</i> at 10 per cent highest fraction < 500 per cent of nominal <i>M_w</i>

C2/C6 ratio
Gas chromatography (2.2.28).

Solution A Mix equal volumes of dilute sulfuric acid R and water R.

Test solution Introduce 0.18 g of the substance to be examined into a 5 mL vial. Add 3.0 mL of solution A, cap, seal the vial and shake until dissolution. Heat the vials for 4 h in a heating block already preheated to 100 °C, shaking them from time to time. Cool to room temperature. Open the vial and carefully add 0.9 g of barium carbonate R. Shake carefully and then centrifuge at about 9000 g for about 15 min. Test the clear supernatant for neutral pH with pH paper. If the solution is still acid, add more barium carbonate R in portions of 0.2 g until the solution is neutral. Filter the clear supernatant (pore size 0.45 µm). Introduce 0.5 mL of the filtrate into a autosampler vial and evaporate to dryness at 40 °C (several hours are usually needed). Take up the residue with 0.50 mL of pyridine R, 0.25 mL of *N₂O-bis(trimethylsilyl)acetamide* R and 25 µL of *chlorotrimethylsilane* R. Seal the vial and heat to 40 °C for 1 h shaking from time to time. Cool to room temperature. Place the vial into the autosampler and perform 3 injections from each vial. Prepare in duplicate.

Reference solution Prepare as prescribed for the test solution but using medium *M_w* hydroxyethyl starch CRS instead of the substance to be examined.

Column: — size: *l* = 15 m, ϕ = 0.32 mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 0.25 µm).

Carrier gas hydrogen for chromatography R at a constant pressure of 69 kPa.

Split ratio 1:20.

Temperature (°C)	Time (min)	Column (min)	Injection port	Detector
150	0.1	1.25	25 - 28	270
150 → 270				250
300				300

Detection Flame ionisation.

Injection 1 µL.

Identification of peaks Use the chromatogram supplied with medium *M_w* hydroxyethyl starch CRS and the chromatogram obtained with the reference solution to identify the peaks due to derivatised product 1, derivatised product 2, derivatised product 3, 2-*O*-hydroxyethyl- α -D-glucose, 6-*O*-hydroxyethyl- α -D-glucose, 2-*O*-hydroxyethyl- β -D-glucose and 6-*O*-hydroxyethyl- β -D-glucose.

System suitability: reference solution: — resolution: minimum 1.5 between the peaks due to 2-*O*-hydroxyethyl- β -D-glucose and 6-*O*-hydroxyethyl- β -D-glucose;

— symmetry factor: 0.6 to 1.5 for the peak due to derivatised product 1;

— repeatability: maximum relative standard deviation of 5.0 per cent for derivatised product 1 after 3 injections.

Calculate the C2/C6 ratio using the following expression:

$$A_1 + A_2 + A_3 + A_4 + A_5 \\ A_6 + A_7$$

- A_1 = area of the peak due to derivatised product 1;
 A_2 = area of the peak due to derivatised product 2;
 A_3 = area of the peak due to derivatised product 3;
 A_4 = area of the peak due to 2-O-hydroxyethyl- α -D-glucose;
 A_5 = area of the peak due to 2-O-hydroxyethyl- β -D-glucose;
 A_6 = area of the peak due to 6-O-hydroxyethyl- α -D-glucose;
 A_7 = area of the peak due to 6-O-hydroxyethyl- β -D-glucose.

Calculate the mean C2/C6 ratio from the values obtained with the 2 test solutions.
 The test is not valid unless the difference of the 2 values is not more than 5 per cent.

Limit Within 20.0 per cent of the nominal value.

Molar substitution (MS)

Gas chromatography (2.2.28).

The content of hydroxyethyl groups is determined after hydrolysis with hydriodic acid as iodoethane.

Internal standard solution Dilute 1.0 mL of toluene R to 200.0 mL with xylene R.

Test solution Introduce 50.0 mg of the substance to be examined and about 0.10-0.15 g of adipic acid R in a 5 mL

vial. Add 1.0 mL of the internal standard solution and 2.0 mL of hydriodic acid R. Tightly seal and cap the vial with a septum and an aluminium, centre tear-off seal. Prepare the test solution 5 times.

Reference solutions In each of 7 vials of 5 mL, introduce about 0.10-0.15 g of adipic acid R. To each vial add 1.0 mL of the

internal standard solution and 2.0 mL of hydriodic acid R. Tightly seal and cap the vials with a septum and an aluminium, centre tear-off seal. Weigh the vials with an accuracy of 0.01 mg. Introduce respectively 10 mg, 20 mg, 30 mg, 40 mg, 50 mg, 60 mg and 70 mg of iodoethane R with a 100 μ L syringe piercing the septa carefully. Weigh the vials again with an accuracy of 0.01 mg and calculate the exact amount of iodoethane R added.

Determine the mass of the vials to the nearest 1 mg. Place the vials for 10 h into a heating block already preheated to 150 °C. After cooling to room temperature, determine the mass of each vial to the nearest 1 mg. Disregard any vial with a loss in mass of more than 5 mg. From 4 vials of the test solution and 5 of the reference solutions, take-up 100 μ L of the upper layer. Introduce in an autosampler vial and dilute with 1.0 mL of xylene R. Seal immediately the vials and shortly shake.

Column:

— **material:** fused silica;

— **size:** $l = 30$ m, $\varnothing = 0.53$ mm;

— **stationary phase:**

poly[(cyanopropyl)(phenyl)dimethylsiloxane R (film thickness 3 μ m).

Carrier gas helium for chromatography R.

Flow rate 8 mL/min.

Split ratio 1:20.

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 4	50	4 - 16	230	280
16 - 20	50 → 230			

Detection Flame ionisation.

Injection volume 1 μ L; inject each solution twice.

Elution order Iodoethane, toluene.

System suitability Reference solutions:

— **resolution:** minimum 1.5 between the peaks due to iodoethane and toluene.

— calculate the ratio of the area of the peak due to iodoethane R to the area of the peak due to the internal

standard for each chromatogram. Calculate the linear regression curve plotting the ratios calculated for the

reference solutions against the quantity of iodoethane R added (in milligrams). The coefficient of determination

R^2 is not less than 0.990.

Results Calculate the quantity (T) of iodoethane in milligrams present in the test solution using the following expression:

$$\frac{M}{A - B}$$

A = ratio of the area of the peak due to iodoethane to the area of the peak due to the internal standard in the chromatogram obtained with the test solution;

B = y-intercept of the curve;

M = slope of the curve.

Then calculate the percentage content of ethylene oxide (C) using the following expression:

$$\frac{44.05 \times T \times 100}{155.97 \times m}$$

m = mass of the substance to be examined, in milligrams;

44.05 = molecular mass of ethylene oxide;

155.97 = molecular mass of iodoethane.

Then calculate the MS using the following expression:

$$\frac{C \times 162.14}{(100 - C) \times 44.05}$$

162.14 = molecular mass of anhydroglucose;

44.05 = molecular mass of ethylene oxide.

Calculate the mean MS from the values obtained with the 4 test solutions.

Limit 0.05 to 2.4, and within 8.0 per cent of the nominal value.

Ethylene glycol

Liquid chromatography (2.2.29).

Test solution Dissolve 1.0 g of the substance to be examined (dried substance) in water R and dilute to 50.0 mL with the same solvent.

Reference solution Dissolve 0.800 g of ethylene glycol R in

water R and dilute to 100.0 mL with the same solvent. Dilute

2.0 mL of this solution to 200.0 mL with water R.

Precolumn:
— size: $l = 0.01$ m, $\emptyset = 4.0$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).
Column:
— size: $l = 0.25$ m, $\emptyset = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 30 °C.
Mobile phase water R.
Flow rate 1.0 mL/min.
Post-column solution Dilute 750 mL of 2 M sodium hydroxide R to 1000 mL with water R.
Flow rate of post-column solution 0.2 mL/min.
Detection Pulsed amperometric detector.
Injection 20 μ L;
Run time 2.5 times the retention time of ethylene glycol.
Retention time Ethylene glycol = about 4 min.

System suitability: reference solution:

— **signal-to-noise ratio:** minimum 10 for the principal peak;
— **repeatability:** maximum relative standard deviation of 10.0 per cent after 6 injections.

After a maximum of 8 sample injections, wash the column using the following program.

Rinsing solution acetonitrile R, water R (20:80 V/V).

Time (min)	Mobile phase (per cent V/V)	Rinsing solution (per cent V/V)
0 - 15	75	25
15 - 20	75 \rightarrow 0	25 \rightarrow 100
20 - 25	0	100
25 - 30	0 \rightarrow 100	100 \rightarrow 0
30 - 100	100	0

Limit:

— **ethylene glycol:** not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (40 ppm).

2-Chloroethanol

Gas chromatography (2.2.28).
Solvent mixture methanol R, acetonitrile R (25:75 V/V).

Internal standard solution Dissolve 0.250 g of 2,6-dimethylanthiline R in the solvent mixture and dilute to 50.0 mL with the solvent mixture. Dilute 0.5 mL of this solution to 50.0 mL with the solvent mixture.
Test solution Introduce 1.0 g of the substance to be examined in a 20 mL vial. Add 10.0 mL of the solvent mixture. Close tightly. Treat in an ultrasonic bath for 3.5 h. Allow to cool at room temperature. To 1.0 mL of this solution, add 0.8 mL of the internal standard solution.

Reference solution Dissolve 0.250 g of 2-chloroethanol R in water R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with water R. Dilute 1.0 mL of this solution to 100.0 mL with water R. To 1.0 mL of this solution, add 0.8 mL of the internal standard solution.

Precolumn:

— **material:** fused silica;

— **size:** $l = 10$ m, $\emptyset = 0.53$ mm;

— **stationary phase:** polar-deactivated polyethyleneglycol R.

— **material:** quartz;

Column:

tightly. Prepare in duplicate.

Reference solution (b) Dissolve 1.0 g of the substance to be examined in 1.0 mL of reference solution (a). Close the vial

solution to 100.0 mL with water R. Use within 24 h.

Reference solution (a) Dilute 1.0 mL of the reference stock

solution to 100.0 mL with water R. Dilute 1.0 mL of this

solution to 100.0 mL with water R. Store in the refrigerator and use

within 4 weeks.

Test solution Introduce 80 mL of water R in a

100 mL volumetric flask. Cool at about 4 °C for at least

30 min. Place the flask on an analytical balance and slowly

introduce 1.0 g of ethylene oxide R. Determine the precise

quantity of ethylene oxide by differential weighing. Dilute to

100.0 mL with water R. Store in the refrigerator and use

within 4 weeks.

Test solution Dissolve 1.0 g of the substance to be examined

in 1.0 mL of water R. Close the vial tightly. Prepare in

duplicate.

Head-space gas chromatography (2.2.28).

Ethylene oxide

than R (5 ppm).

obtained with the test solution; this ratio is not greater

due to the internal standard from the chromatogram

of the peak due to 2-chloroethanol to the area of the peak

with the reference solution; calculate the ratio of the area

of the peak due to 2-chloroethanol to the area of the peak due

to the internal standard from the chromatogram obtained

with the reference solution; calculate the ratio (R) of the area of the

peak due to 2-chloroethanol to the area of the peak due

to the internal standard from the chromatogram obtained

with the reference solution; calculate the ratio of the area of the

peak due to 2-chloroethanol to the area of the peak due

to the internal standard from the chromatogram obtained

with the reference solution; calculate the ratio of the area of the

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peak due to 2-chloroethanol to the area of the peak due

to the internal standard from the chromatogram obtained

with the reference solution; calculate the ratio of the area of the

peak due to 2-chloroethanol to the area of the peak due

— size: $l = 30$ mm, $\varnothing = 0.32$ mm;
— stationary phase:
poly[(cyanopropyl)(phenyl)][dimethylsiloxane R (film
thickness 1.5 μ m).
Carrier gas helium for chromatography R at a pressure of
110.3 kPa.
Split ratio 1:35.
Static head-space conditions which may be used:

— equilibration temperature: 80 °C;

— equilibration time: 40 min;

— transfer-time temperature: 150 °C;

— pressurisation time: 2.0 min;

— injection time: 3 s.

Temperature:

Time (min)	Temperature (°C)
0-20	40
20-30	40 → 240
30-40	240
40	140
250	Detector

Detection Flame ionisation.

Injection Inject a suitable volume of the gaseous phase of the test solution and reference solution (b).

System suitability:

— signal-to-noise ratio: minimum 10 for the peak due to

ethylene oxide in the chromatogram obtained with reference solution (b).

Limit:

— ethylene oxide: not more than 0.5 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (1 ppm).

Sodium chloride

Maximum 0.1 per cent.

Test solution In a 250 mL conical flask, dissolve 10.0 g of the substance to be examined in 100 mL of water R. Add 2 mL of dilute nitric acid R and 5.0 mL of a 9 g/L solution of

Reference solution In a 250 mL conical flask, dilute 5.0 mL of a 9 g/L solution of sodium chloride R with 100 mL of water R.

Add 2 mL of dilute nitric acid R.

Carry out a potentiometric titration (2.2.20) with 0.1 M silver nitrate. Calculate the percentage content of sodium chloride using the following expression:

$$\frac{m}{(n_1 - n_2) \times 5.844 \times 100}$$

n_1 = volume of 0.1 M silver nitrate used for the test

solution, in millilitres;

n_2 = volume of 0.1 M silver nitrate used for the reference

solution, in millilitres;

m = mass of the substance to be examined in the test

solution, in milligrams.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2 g in water R and dilute to 20 mL with the same solvent. 12 mL complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Hydroxypropyl Starch

(Ph. Eur. monograph 2165)

9049-76-7



DEFINITION

Hydroxypropyl starch is a partially substituted 2-hydroxypropylether of *Maize starch* (0344), *Potato starch* (0355), *cassava starch*, *Rice starch* (0349) or *Pea starch* (2403) chemically modified by etherification with the reagent propylene oxide. In addition, this starch may be partially hydrolysed using acids or enzymes to obtain 'thinned starch' with reduced viscosity.

Content

hydroxypropyl groups: 0.5 per cent to 7.0 per cent.

PRODUCTION

The production of hydroxypropyl starch shall be in compliance with the requirements of the European legislation for food additives.
Mixing of starches from different botanical sources prior to chemical modification is not allowed.

CHARACTERS

Appearance

White or slightly yellowish powder.

Solubility

Practically insoluble in cold water and in ethanol

IDENTIFICATION

A. Examined under a microscope, using not less than $20 \times$ magnification and using a mixture of equal volumes of glycerol R and water R, it appears as follows according to the botanical source stated on the label.

— *Maize-based hydroxypropyl starch*: it presents either angular polyhedral granules of irregular sizes with diameters of about 2-23 μ m or rounded or spheroidal granules of irregular sizes with diameters of about 25-35 μ m; the central hilum consists of a distinct cavity or 2-to-5-rayed cleft and there are no concentric striations; between orthogonally orientated polarising plates or prisms, the starch granules show a distinct black cross intersecting at the hilum.
— *Potato-based hydroxypropyl starch*: it presents granules, either irregularly shaped, ovoid or pear-shaped, usually 30-100 μ m in size but occasionally exceeding 100 μ m, or rounded, 10-35 μ m in size; there are occasional compound granules having 2-4 components; the ovoid and pear-shaped granules have an eccentric hilum and the rounded granules a centric or slightly eccentric

Shake 1.0 g with 20 mL of dilute hydrochloric acid R. Filter.

The filtrate complies with the test for iron.

— For hydroxypropyl starch obtained from pea: maximum

50 ppm.

Shake 1.0 g with 50 mL of dilute hydrochloric acid R. Filter.

The filtrate complies with the test for iron.

Loss on drying (2.2.32)

Determined on 1.000 g by drying in an oven at 130 °C for

90 min:

— maximum 15.0 per cent for hydroxypropyl starch

obtained from maize, cassava, rice or pea;

Sulfated ash (2.4.14)

Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

ASSAY

Nuclear magnetic resonance spectrometry (2.2.33).

Internal standard solution. Disperse 50.0 mg of 3-trimethylsilyl-

1-propanesulfonic acid sodium salt CRS in about 5 g of

deuterium oxide R1, weighed to the nearest 0.1 mg. Store in a

sealed bottle.

Test solution. Disperse 20 g of the substance to be examined in

200.0 mL of carbon dioxide-free water R at room temperature.

Agitate for 15 min and filter. Repeat the operation twice.

If problems of poor dispersibility or slow filtration are

encountered, use cooled carbon dioxide-free water R for the

washing operation. Dry the washed starch for at least 4 h in

an oven *in vacuo* at 30 ± 5 °C. Determine the moisture

content (W) on 5 g of this washed and dried sample using

the test for loss on drying. Weigh 12.0 mg (dried substance)

of the washed and dried sample in a 5 mm NMR tube.

Add 0.1 mL of deuterium chloride solution R and 0.75 mL of

deuterium oxide R1. Cap the tube, mix, and place it in a

boiling water-bath until a clear solution is obtained (3 min to

1 h maximum). When a clear solution is obtained, allow to

cool to room temperature. Dry the exterior of the tube and

weigh to the nearest 0.1 mg. Add 0.05 mL of the internal

standard solution and weigh to the nearest 0.1 mg.

Determine the mass of the internal standard solution

introduced. Mix thoroughly.

Apparatus. FT-NMR spectrometer at minimum 300 MHz.

Acquisition of ¹H NMR spectra. The following parameters may

be used:

— sweep width: 8 ppm (–1.0 to + 7 ppm);

— irradiation frequency offset: none;

— time domain: 64 K at least;

— pulse width: 90°;

— pulse delay: 10 s;

— dummy scans: 0;

— number of scans: 8.

Use the CH₃ signal of the internal standard for shift

referencing. The shift of the singlet is set to 0 ppm.

Record the FID signal.

Call the integration sub-routine after phase corrections and

baseline correction between –0.5 ppm and + 6 ppm.

Measure the peak areas of the doublet from the methyl

groups of the hydroxypropyl function at + 1.2 ppm (4₂), and

hilum; all granules show clearly visible concentric

situations; between orthogonally orientated polarising

plates or prisms, the starch granules show a distinct

black cross intersecting at the hilum.

— Cassava-based hydroxypropyl starch: it presents spherical

granules with one truncated side, typically 5–35 µm in

diameter and having a circular or several-rayed central

cleft; some granules may also be egg-shaped or cap-

shaped; the hilum is centric, sometimes slightly

fissured; between orthogonally orientated polarising

plates or prisms, the starch granules show a distinct

black cross intersecting at the hilum.

— Rice-based hydroxypropyl starch: it presents polyhedral,

simple granules 1–10 µm, mostly 4–6 µm, in size; these

simple granules often gather in ellipsoidal, compound

granules 50–100 µm in diameter; the granules have a

poorly visible central hilum and there are no

concentric situations; between orthogonally orientated

polarising plates or prisms, the starch granules show a

distinct black cross intersecting at the hilum.

— Pea-based hydroxypropyl starch: it presents a majority of

large elliptical granules, 25–45 µm in size, sometimes

irregular or reniform; it also presents a minority of

small rounded granules, 5–8 µm in size; granules can

present cracks or irregularities; sometimes, granules

show barely visible concentric situations; exceptionally,

granules show a slit along the main axis; between

orthogonally orientated polarising plates or prisms, the

starch granules show a distinct black cross.

B. Suspend 1 g in 50 mL of water R, boil for 1 min and cool.

A translucent or clear mucilage is formed.

C. To 1 mL of the mucilage obtained in identification test B

add 0.05 mL of iodine solution R1. An orange-red or dark

blue colour is produced, which disappears on heating.

D. Introduce 0.1 g into a 100 mL volumetric flask and add

12.5 mL of dilute sulfuric acid R. Place the flask in a water-

bath and heat until the sample is dissolved. Cool and dilute

to 100 mL with water R. Introduce 1 mL of this solution into

a 25 mL graduated test-tube with glass stopper and, with the

tube immersed in cold water, add dropwise 8 mL of sulfuric

acid R. Mix well and place the tube in a boiling water-bath

for exactly 3 min. Immediately transfer the tube to an ice-

bath until the solution is chilled. Add 0.6 mL of ninhydrin

solution R2, carefully allowing the reagent to run down the

walls of the test-tube. Immediately shake well, and place the

tube in a water-bath at 25 °C for 100 min. Dilute to 25 mL

with sulfuric acid R and mix by inverting the tube several

times. Do not shake. A violet colour develops within 5 min.

TESTS

pH (2.2.3)

4.5 to 8.0.

Shake 5.0 g with 25.0 mL of carbon dioxide-free water R for

60 s. Allow to stand for 15 min.

Foreign matter

Examined under a microscope using a mixture of equal

volumes of glycerol R and water R, not more than traces of

matter other than starch granules are present.

Oxidising substances (2.5.30)

Maximum 20 ppm, calculated as H₂O₂.

Sulfur dioxide (2.5.29)

Maximum 50 ppm.

Iron (2.4.9)

— For hydroxypropyl starch obtained from maize, potato,

cassava or rice: maximum 20 ppm.

of the methyl groups at 0 ppm of the internal standard (A_1) without ^{13}C -satellites.
Results Measure the signal coming from the 3 protons of the methyl group in the hydroxypropyl function; calculate the hydroxypropyl groups content as a percentage *m/m* (dried substance) using the following expression:

$$3A_2 = \frac{A_1}{P} \times \frac{100}{W_1} \times \frac{218}{m_1} \times 59 \times \frac{m}{100} \times \frac{100}{100 - W}$$

3 = numerical value representing the 3 methyl groups in the internal standard;
 A_1 = area of the methyl groups in the internal standard;
 A_2 = area of the methyl groups of hydroxypropyl;

P = percentage content of 3-trimethylsilyl-*propanesulfonic acid sodium salt* CRS;
 W_1 = mass fraction of the internal standard in the internal standard solution, in milligrams per gram;

m_1 = mass of the internal standard solution in the NMR tube, in grams;

218 = molar mass of the internal standard, in grams per mole;
 59 = molar mass of the hydroxypropyl group, in grams per mole;

m = mass of the washed and dried sample in the NMR tube, in milligrams;

W = moisture content, as a percentage *m/m*.
 The label states the botanical source of the starch and the type of modification.

LABELLING

Pregelatinised Hydroxypropyl Starch

(Ph Eur monograph 2645)

Ph Eur

DEFINITION

Pregelatinised hydroxypropyl starch is prepared from Starch, hydroxypropyl (2165) by mechanical processing in the presence of water, with or without heat, to rupture all or part of the starch granules, and subsequent drying.

Content

— hydroxypropyl groups: 0.5 per cent to 7.0 per cent.

PRODUCTION

The production of pregelatinised hydroxypropyl starch shall be in compliance with the requirements of the European legislation for food additives.

CHARACTERS

Appearance

White or slightly yellowish powder.

IDENTIFICATION

A. It swells in cold water.
 B. Disperse 0.5 g in 2 mL of water R, without heating, and add 0.05 mL of iodine solution R1. A reddish-violet or greyish-blue colour is produced.
 C. To 0.1 g add 12.5 mL of dilute sulfuric acid R. Heat in a water-bath until the sample is dissolved. Cool and dilute to

100 mL with water R. Introduce 1 mL of the solution into a 25 mL graduated test-tube with a ground-glass stopper and, with the tube immersed in cold water, add dropwise 8 mL of sulfuric acid R. Mix well and place the tube in a boiling water-bath for exactly 3 min. Immediately transfer the tube to an ice-bath until the solution is chilled. Add 0.6 mL of ninhydrin solution R2, carefully allowing the reagent to run down the walls of the test-tube. Immediately shake well, and place the tube in a water-bath at 25 °C for 100 min. Dilute to 25 mL with sulfuric acid R and mix by inverting the tube several times. Do not shake. A violet colour develops within 5 min.

TESTS

pH (2.2.3)
 4.5 to 8.0.

Progressively add 3.0 g to 100.0 mL of carbon dioxide-free water R, stirring continuously. Determine the pH when a homogeneous solution is obtained.

Impurity A

Gas chromatography (2.2.28).

Internal standard solution Mix 50.0 mg of propane-1,3-diol R with anhydrous pyridine R and dilute to 100.0 mL with the same solvent.

Test solution To 0.200 g of the substance to be examined add 1.0 mL of the internal standard solution and 9.0 mL of anhydrous pyridine R. Heat under a reflux condenser for 20 min. Allow to cool. Transfer 1.0 mL of this solution to a 2 mL vial with a screw cap fitted with a septum. Add 0.1 mL of chlorotrimethylsilane R and 0.2 mL of hexamethyldisilazane R and allow to stand for 15 min.

Reference solution Mix 50.0 mg of propane-1,3-diol R and 50.0 mg of propylene glycol CRS (impurity A) with anhydrous pyridine R and dilute to 100.0 mL with the same solvent.

Transfer 0.1 mL of the solution to a 2 mL vial with a screw cap fitted with a septum. Add 0.1 mL of chlorotrimethylsilane R, 0.2 mL of hexamethyldisilazane R and 0.9 mL of anhydrous pyridine R. Close and mix. Allow to stand for 15 min.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 0.25 μm).

NOTE: the column must be desorbed regularly. Conditions:

temperature program of 70 °C to 300 °C at a rate of 7 °C/min. Maintain for 10 min at 300 °C.

Carrier gas helium for chromatography R.

Flow rate 3 mL/min.

Split ratio 1:30.

Temperature:

— column: 70 °C;

— injection port and detector: 250 °C.

Detection Flame ionisation.

Injection 1 μL .

Relative retention With reference to the trimethylsilyl derivative of propane-1,3-diol (retention time = about 8.5 min): trimethylsilyl derivative of propylene glycol = about 0.7.

System suitability: reference solution:

— resolution: minimum 5.0 between the peaks due to the trimethylsilyl derivative of propylene glycol and the trimethylsilyl derivative of propane-1,3-diol.

Calculation of percentage contents Use the internal standard

method.

Limit:

— impurity A: maximum 0.1 per cent.

Oxidising substances (2.5.30)

Maximum 20 ppm, calculated as H_2O_2 .

Use a mixture of equal volumes of methanol R and water R as solvent.

Sulfur dioxide (2.5.29)

Maximum 50 ppm.

Iron (2.4.9)

— For pregelatinised hydroxypropyl starch obtained from

maize, potato, cassava or rice: maximum 20 ppm.

Dissolve the residue obtained in the test for sulfated ash in

20 mL of dilute hydrochloric acid R and filter. The filtrate

complies with the test for iron.

— For pregelatinised hydroxypropyl starch obtained from

pea: maximum 50 ppm.

Dissolve the residue obtained in the test for sulfated ash in

50 mL of dilute hydrochloric acid R and filter. The filtrate

complies with the test for iron.

Loss on drying (2.2.32)

Determined on 1.000 g by drying in an oven at 130 °C for

90 min:

— maximum 15.0 per cent for pregelatinised hydroxypropyl

starch obtained from maize, cassava, rice or pea;

— maximum 20.0 per cent for pregelatinised hydroxypropyl

starch obtained from potato.

Sulfated ash (2.4.14)

Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10^3 CFU/g (2.6.12).

TYMC: acceptance criterion 10^2 CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

ASSAY

Nuclear magnetic resonance spectrometry (2.2.33).

Internal standard solution Disperse 50.0 mg of 3-trimethylsilyl-

1-propanesulfonic acid sodium salt CRS in about 5 g of

deuterium oxide R1, weighed to the nearest 0.1 mg. Store in a

sealed bottle.

Test solution Dry 5.000 g of the substance to be examined at

130 °C for 90 min. Weigh 12.0 mg of the dried substance in

a 5 mm NMR tube. Add 0.1 mL of deuterium chloride

solution R and 0.75 mL of deuterium oxide R1. Cap the tube,

mix, and place it in a boiling water-bath until a clear solution

is obtained (3 min to maximum 1 h). When a clear solution

is obtained, allow to cool to room temperature. Dry the

exterior of the tube and weigh to the nearest 0.1 mg.

Add 0.05 mL of the internal standard solution and weigh to

the nearest 0.1 mg. Determine the mass of the internal

standard solution introduced. Mix thoroughly.

Apparatus FT-NMR spectrometer operating at minimum

300 MHz.

Acquisition of 1H NMR spectra The following parameters may

be used:

— sweep width: 8 ppm (–1.0 to +7 ppm);

— irradiation frequency offset: none;

— time domain: at least 64 K;

— pulse width: 90°;

— pulse delay: 10 s;

— dummy scans: 0;

— number of scans: 8.

Use the CH_3 signal of the internal standard for shift

referencing. The shift of the singlet is set to 0 ppm.

Record the FID signal.

Call the integration sub-routine after phase corrections and

baseline correction between –0.5 ppm and +6 ppm.

Measure the peak areas of the doublet from the methyl

groups of the hydroxypropyl function at +1.2 ppm (A_2), and

of the methyl groups at 0 ppm of the internal standard (A_1)

without ^{13}C -satellites.

Results Measure the signal coming from the 3 protons of the

methyl group in the hydroxypropyl function; calculate the

percentage content of hydroxypropyl groups using the

following expression:

$$\frac{3A_2}{A_1} \times \frac{P}{W_1 \times m_1} \times \frac{100}{218} \times 59 \times \frac{m}{100}$$

= numerical value representing the 3 methyl groups

in the internal standard;

A_1 = area of the methyl groups in the internal

standard;

A_2 = area of the methyl groups of hydroxypropyl;

P = percentage content of 3-trimethylsilyl-1-

propanesulfonic acid sodium salt CRS;

W_1 = mass fraction of the internal standard in the

internal standard solution, in milligrams per

gram;

m_1 = mass of the internal standard solution in the

NMR tube, in grams;

218 = molar mass of the internal standard, in grams per

mole;

59 = molar mass of the hydroxypropyl group, in grams

per mole;

m = mass of the substance to be examined in the

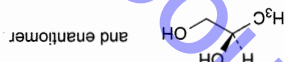
NMR tube, in milligrams.

LABELLING

The label states the botanical source of the starch and the

type of modification.

IMPURITIES



A. (2R,3S)-propane-1,2-diol (propylene glycol).

Ph Eur

Maize Starch¹

(Ph. Eur. monograph 0344)

Action and use

Excipient.

When Starch is specified and the type is not indicated, Maize

Starch, Potato Starch, Rice Starch, Wheat Starch or, in

tropical countries where these are not available, Tapioca

Starch may be supplied or used.

DEFINITION

Maize starch is obtained from the caryopsis of *Zea mays* L.

◆ **CHARACTERS**

Appearance

Mat, white to slightly yellowish, very fine powder that cracks

when pressed between the fingers.

Loss on drying (2.2.32)
Maximum 15.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 90 min.

Sulfated ash (2.4.14)
Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination
TAMC: acceptance criterion 10³ CFU/g (2.6.12).
TYMC: acceptance criterion 10² CFU/g (2.6.12).

◆ Absence of *Escherichia coli* (2.6.13).
◆ Absence of *Salmonella* (2.6.13).
◆ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.



Pea Starch

(Ph. Eur. monograph 2403)

Ph Eur

DEFINITION

Pea starch is obtained from the seeds of *Pisum sativum* L.

CHARACTERS

Appearance

White or almost white, very fine powder.

Solubility

Practically insoluble in cold water and in ethanol (96 per cent).

IDENTIFICATION

A. Microscopic examination (2.8.23) using a 50 per cent V/V solution of *glycerol* R. It presents a majority of large elliptical granules, 25–45 µm in size, sometimes irregular, or reniform. It also presents a minority of small rounded granules, 5–8 µm in size (Figure 2403.-1). Granules can present cracks or irregularities. Sometimes, granules show barely visible concentric striations. Exceptionally, granules show a slit along the main axis. Between orthogonally oriented polarising plates or prisms, the granules show a distinct black cross.

A. Microscope examination (2.8.23), using a 50 per cent V/V solution of *glycerol* R. It appears as either angular polyhedral granules of irregular sizes with diameters ranging from about 2 µm to about 23 µm or as rounded or spheroidal granules of irregular sizes with diameters ranging from about 25 µm to about 35 µm (Figure 0344.-1). The central hilum consists of a distinct cavity or 2- to 5-rayed cleft and there are no concentric striations. Between orthogonally orientated polarising plates or prisms, the starch granules show a distinct black cross intersecting at the hilum.

IDENTIFICATION

The presence of granules with cracks or irregularities on the edge is exceptional. ◆
(96 per cent).
Practically insoluble in cold water and in ethanol

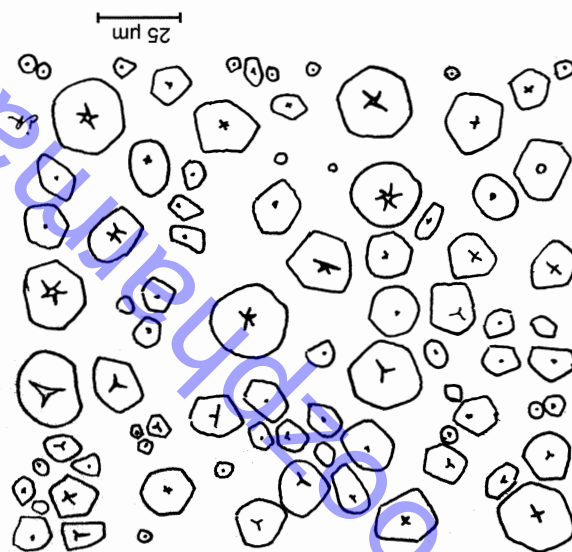


Figure 0344.-1. – Illustration for identification test A of maize starch

B. Suspend 1 g in 50 mL of *water* R, boil for 1 min and cool. A thin, cloudy mucilage is formed.
C. To 1 mL of the mucilage obtained in identification test B add 0.05 mL of *iodine solution* R1. An orange-red to dark blue colour is produced, which disappears on heating.

TESTS

pH (2.2.3)

4.0 to 7.0.

To 5.0 g add 25.0 mL of *carbon dioxide-free water* R. Agitate continuously at a moderate rate for 60 s. Stop the agitation and allow to stand for 15 min.

◆ Foreign matter

Examined under a microscope using a 50 per cent V/V solution of *glycerol* R, not more than traces of matter other than starch granules are present. No starch grains of any other origin are present. ◆

Oxidising substances (2.5.30)

Maximum 20 ppm, calculated as H₂O₂.

Sulfur dioxide (2.5.29)

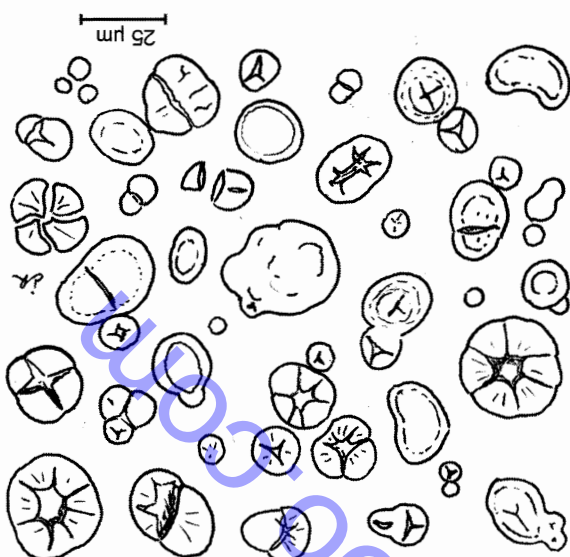
Maximum 50 ppm.

Iron (2.4.9)

Maximum 10 ppm.

Shake 1.5 g with 15 mL of *dilute hydrochloric acid* R. Filter. The filtrate complies with the test.

Figure 2403.-1. – Illustration for identification test A of pea starch



B. Suspend 1 g in 50 mL of water R, boil for 1 min and cool. A thin, cloudy mucilage is formed.
C. To 1 mL of the mucilage obtained in identification test B, add 0.05 mL of iodine solution R1. A dark blue colour is produced, which disappears on heating.

TESTS

pH (2.2.3) 5.0 to 8.0.

Shake 5.0 g with 25.0 mL of carbon dioxide-free water R for 60 s. Allow to stand for 15 min and shake again.

Foreign matter

Examined under a microscope using a 50 per cent V/V solution of glycerol R, not more than traces of matter other than starch granules are present. No starch granules of any other origin are present.

Oxidising substances (2.5.30)

Maximum 20 ppm, calculated as H₂O₂.

Sulfur dioxide (2.5.29)

Maximum 50 ppm.

Iron (2.4.9)

Maximum 50 ppm.

Shake 1.0 g with 50 mL of dilute hydrochloric acid R. Filter. The filtrate complies with the test for iron.

Loss on drying (2.2.32)

Maximum 16.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 90 min.

Sulfated ash (2.4.14)

Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

Ph Eur

Potato Starch¹

(Ph. Eur. monograph 0355)

Action and use

Excipient.

When Starch is specified and the type is not indicated, Maize

Starch, Potato Starch, Rice Starch, Wheat Starch or, in

tropical countries where these are not available, Tapioca

Starch may be supplied or used.

Ph Eur

DEFINITION

Potato starch is obtained from the tuber of *Solanum*

tuberosum L.

◆ CHARACTERS**Appearance**

Very fine, white or almost white powder which creaks when pressed between the fingers.

Solubility

Practically insoluble in cold water and in ethanol

(96 per cent).

Potato starch does not contain starch grains of any other origin. It may contain a minute quantity, if any, of tissue fragments of the original plant. ◆

pH (2.2.3) 5.0 to 8.0.

TESTS

blue colour is produced which disappears on heating.

add 0.05 mL of iodine solution R1. An orange-red to dark

C. To 1 mL of the mucilage obtained in identification test B,

A thick, opalescent mucilage is formed.

B. Suspend 1 g in 50 mL of water R, boil for 1 min and cool.

cross intersecting at the hilum.

visible concentric striations. Between orthogonally orientated

granules have an eccentric hilum and the rounded granules

components (Figure 0355.-1). The ovoid and pear-shaped

size. There are occasional compound granules having 2-4

occasionally exceeding 100 µm, or rounded, 10-35 µm in

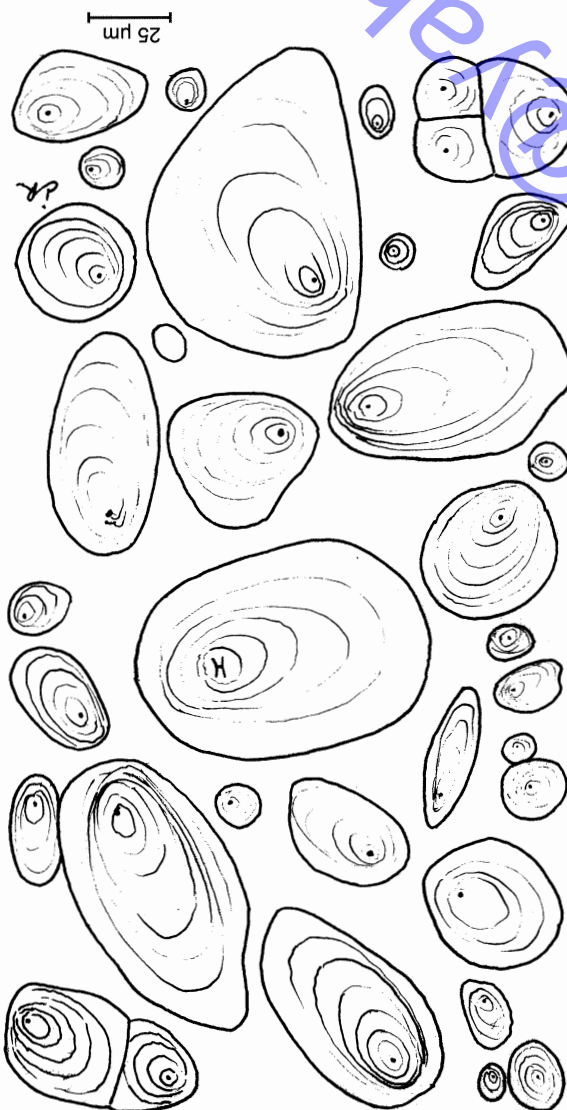
shaped, ovoid or pear-shaped, usually 30-100 µm in size but

solution of glycerol R. It presents granules, either irregularly

A. Microscopic examination (2.8.23) using a 50 per cent V/V

IDENTIFICATION

Figure 0355.-1. – Illustration for identification test A of potato starch



Shake 5.0 g with 25.0 mL of carbon dioxide-free water R for 60 s. Allow to stand for 15 min.

◆ Foreign matter

Examined under a microscope using a 50 per cent V/V solution of glycerol R, not more than traces of matter other than starch granules are present. No starch grains of any other origin are present. ◆

Oxidising substances (2.5.30)

Maximum 20 ppm, calculated as H₂O₂.

Sulfur dioxide (2.5.29)

Maximum 50 ppm.

Iron (2.4.9)

Maximum 10 ppm.

Shake 1.5 g with 15 mL of dilute hydrochloric acid R. Filter. The filtrate complies with the limit test for iron.

Loss on drying (2.2.32)

Maximum 20.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 90 min.

Sulfated ash (2.4.14)

Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13). ◆

Absence of *Salmonella* (2.6.13). ◆

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.

Pregelatinised Starch

Pregelatinised Maize Starch

(Ph. Eur. monograph 1267)

When Pregelatinised Starch is prepared *Zea mays*, the title Pregelatinised Maize Starch may be used.

DEFINITION

Pregelatinised starch is prepared from Maize starch (0344), Potato starch (0355) or Rice starch (0349) by mechanical processing in the presence of water, with or without heat, to rupture all or part of the starch granules, and subsequent drying. It contains no added substances but it may be modified to render it compressible and to improve its flow characteristics.

CHARACTERS

Appearance

White or yellowish-white powder.

IDENTIFICATION

A. Examined under a microscope using a mixture of equal volumes of glycerol R and water R it presents irregular, translucent, white or yellowish-white flakes or pieces with an uneven surface. Under polarised light (between crossed nicols), starch granules with a distinct black cross intersecting at the hilum may be seen.
B. Disperse 0.5 g in 2 mL of water R without heating and add 0.05 mL of iodine solution R1. A reddish-violet or blue colour is produced.

LABELLING

The label states the type of starch used as starting material.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.
The following characteristics may be relevant for pregelatinised starch used as filler, binder or disintegrant in tablets and in hard capsules.

Cold-water-soluble matter

Transfer 100 mL of water R at 25 ± 1 °C into a beaker and add 1.000–3.000 g of the substance to be examined while stirring. Continue to stir for 10 min. Transfer 35 mL of the dispersion to a centrifuge tube and centrifuge at 3000 g for 15 min. Transfer 25 mL of the supernatant to a crucible that has previously been dried in an oven at 120 ± 2 °C for 4 h and weighed to the nearest 0.1 mg. Evaporate to dryness on a water-bath, then place the crucible in an oven at 120 ± 2 °C for 4 h. Allow to cool in a desiccator. Weigh the crucible to the nearest 0.1 mg again.

TESTS

pH (2.2.3)

4.5 to 7.0.

Progressively add 3.0 g to 100.0 mL of carbon dioxide-free water R, stirring continuously. Determine the pH when a homogeneous solution is obtained.

Oxidising substances (2.5.30)

It complies with the test for oxidising substances. Use a mixture of equal volumes of methanol R and water R as solvent.

Sulfur dioxide (2.5.29)

Maximum 50 ppm.

Iron (2.4.9)

Maximum 20 ppm.

Dissolve the residue obtained in the test for sulfated ash in 20 mL of dilute hydrochloric acid R. Filter. The filtrate complies with the test.

Foreign matter

Examined under a microscope using a mixture of equal volumes of glycerol R and water R, not more than traces of matter other than starch granules are present.

Loss on drying (2.2.32)

Maximum 15.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 90 min.

Sulfated ash (2.4.14)

Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

LABELLING

The label states the type of starch used as starting material.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.
The following characteristics may be relevant for pregelatinised starch used as filler, binder or disintegrant in tablets and in hard capsules.

Cold-water-soluble matter

Transfer 100 mL of water R at 25 ± 1 °C into a beaker and add 1.000–3.000 g of the substance to be examined while stirring. Continue to stir for 10 min. Transfer 35 mL of the dispersion to a centrifuge tube and centrifuge at 3000 g for 15 min. Transfer 25 mL of the supernatant to a crucible that has previously been dried in an oven at 120 ± 2 °C for 4 h and weighed to the nearest 0.1 mg. Evaporate to dryness on a water-bath, then place the crucible in an oven at 120 ± 2 °C for 4 h. Allow to cool in a desiccator. Weigh the crucible to the nearest 0.1 mg again.

$$\frac{(B - A) \times \frac{100}{25} \times 100}{S \times \frac{100 - C}{100}}$$

A = initial crucible mass, in grams;
B = final crucible mass, in grams;
C = loss on drying, in per cent;
S = sample mass, in grams.

Particle-size distribution (2.9.31 or 2.9.38).
Powder flow (2.9.36)

Ph Eur



(Ph. Eur. monograph 0349)

Rice Starch

Ph Eur

DEFINITION

Rice starch is obtained from the caryopsis of *Oryza sativa* L.

◆ CHARACTERS

Appearance

Very fine, white or almost white powder, which creaks when pressed between the fingers.

Solubility

Practically insoluble in cold water and in ethanol (96 per cent).

Rice starch does not contain starch grains of any other origin. It may contain traces of, if any, fragments of the endosperm tissue of the fruit. ◆

IDENTIFICATION

A. Microscopic examination (2.8.23) using a 50 per cent *V/V* solution of *glycerol R*. It presents polyhedral, simple grains

1-10 µm (mostly 4-6 µm) in size (Figure 0349.-1). These simple grains often gather in ellipsoidal, compound grains 50-100 µm in diameter. The grains have a poorly visible central hilum and there are no concentric striations. Between grains show a distinct black cross intersecting at the hilum.

B. Suspend 1 g in 50 mL of *water R*, boil for 1 min and cool. *A* thin, cloudy mucilage is formed. **C.** To 1 mL of the mucilage obtained in identification test **B** add 0.05 mL of *iodine solution R1*. An orange-red to dark blue colour is produced, which disappears on heating.

TESTS

pH (2.2.3)

5.0 to 8.0.

Shake 5.0 g with 25.0 mL of *carbon dioxide-free water R* for 60 s. Allow to stand for 15 min.

◆ Foreign matter

Examined under a microscope using a 50 per cent *V/V* solution of *glycerol R*, not more than traces of matter other than starch granules are present. No starch grains of any other origin are present. ◆

Very fine powder which creaks when pressed between the fingers. Practically insoluble in cold *water* and in *ethanol* (96%).

CHARACTERISTICS

untissima Pohl.

Tapioca Starch is obtained from the rhizomes of *Manihot*

DEFINITION

When Starch is specified and the type is not indicated, Maize Starch, Potato Starch, Rice Starch, Wheat Starch or, in tropical and sub-tropical countries where these are not available, Tapioca Starch may be supplied or used.

Action and use

Excipient.

Cassava Starch

Tapioca Starch

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 *Pharmacopoeial Harmonisation*.

Ph Eur

◆ Absence of *Salmonella* (2.6.13). ◆

Absence of *Escherichia coli* (2.6.13).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

Microbial contamination

Maximum 0.6 per cent, determined on 1.0 g.

Sulfated ash

(2.4.14)

Maximum 15.0 per cent, determined on 1.00 g by drying in an oven at 130 °C for 90 min.

Loss on drying

(2.2.32)

The filtrate complies with the test.

Shake 1.5 g with 15 mL of *dilute hydrochloric acid R*. Filter.

Maximum 10 ppm.

Iron

(2.4.9)

Maximum 50 ppm.

Sulfur dioxide

(2.5.29)

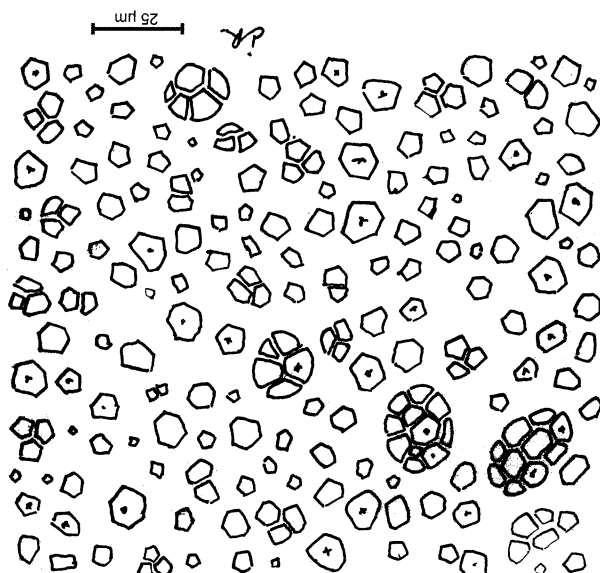
Maximum 20 ppm, calculated as H₂O₂.

Oxidising substances

(2.5.30)

starch

Figure 0349.-1. – Illustration for identification test *A* of rice



IDENTIFICATION

A. Principally simple granules, subspherical, muller-shaped or rounded polyhedral; smaller granules 5 to 10 µm, larger granules 20 to 35 µm in diameter; hilum, central, punctate, linear or triradiate; striations, faint, concentric; compound granules, few, of two to three unequal components.

B. Heat to boiling a suspension of 1 g in 50 mL of water for 1 minute and cool. A thin, cloudy mucilage is formed.

C. Mix 0.05 mL of iodine solution R1 with 1 mL of the mucilage obtained in test B. A dark blue colour is produced which disappears on heating and reappears on cooling.

TESTS

Add 10 g of the starch to 100 mL of ethanol (70%) previously neutralised to 0.5 mL of phenolphthalein solution, shake for 1 hour, filter and titrate 50 mL of the filtrate with 0.1M sodium hydroxide VS. Not more than 2.0 mL is required to change the colour of the solution.

Foreign matter

Not more than traces of cell membranes and protoplasm are present.

Loss on drying
When dried to constant weight at 100° to 105°, loses not more than 15.0% of its weight. Use 1 g.

Sulfated ash

Not more than 0.6%, Appendix IX A, Method II. Use 1 g.

Microbial contamination

1.0 g is free from *Escherichia coli*, Appendix XVI B1.

STORAGE

Tapioca Starch should be kept in an airtight container.

Wheat Starch¹

(Ph. Eur. monograph 0359)

Action and use

Excipient.

When Starch is specified and the type is not indicated, Maize Starch, Potato Starch, Rice Starch, Wheat Starch or, in tropical countries where these are not available, Tapioca Starch may be supplied or used.

Ph Eur

DEFINITION

Wheat starch is obtained from the caryopsis of *Triticum aestivum* L. (*T. vulgare* Vill.).

◆ CHARACTERS**Appearance**

Very fine, white or almost white powder that creaks when pressed between the fingers.

Solubility

Practically insoluble in cold water and in ethanol

(96 per cent).

Wheat starch does not contain starch grains of any other origin. It may contain a minute quantity, if any, of tissue fragments of the original plant. ◆

IDENTIFICATION

A. Microscopic examination (2.8.23) using a 50 per cent V/V solution of glycerol R. It presents large and small granules, and, very rarely, intermediate sizes (Figure 0359.-1).

The large granules, 10-60 µm in diameter, are discoid or, more rarely, reniform when seen face-on. The central hilum and striations are invisible or barely visible and the granules sometimes show cracks on the edges. Seen in profile, the granules are elliptical and fusiform and the hilum appears as a slit along the main axis. The small granules, rounded or polyhedral, are 2-10 µm in diameter. Between orthogonally orientated polarising plates or prisms, the granules show a distinct black cross intersecting at the hilum.

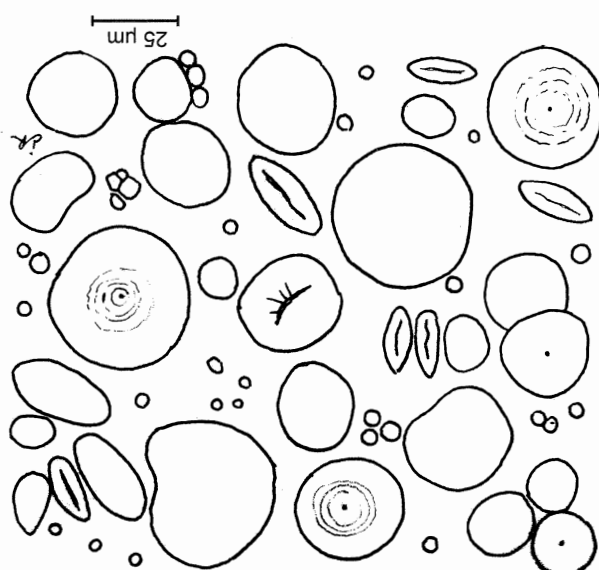


Figure 0359.-1. – Illustration for identification test A of wheat starch

B. Suspend 1 g in 50 mL of water R, boil for 1 min and cool. A thin, cloudy mucilage is formed.
C. To 1 mL of the mucilage obtained in identification test B add 0.05 mL of iodine solution R1. A dark blue colour is produced, which disappears on heating.

TESTS

pH (2.2.3)
4.5 to 7.0.

Shake 5.0 g with 25.0 mL of carbon dioxide-free water R for 60 s. Allow to stand for 15 min.

◆ Foreign matter

Examined under a microscope using a 50 per cent V/V solution of glycerol R, not more than traces of matter other than starch granules are present. No starch grains of any other origin are present. ◆

Total protein

Maximum 0.3 per cent of total protein (corresponding to 0.048 per cent N₂, conversion factor: 6.25), determined on 6.0 g by sulfuric acid digestion (2.5.9) modified as follows: wash any adhering particles from the neck into the flask with 25 mL of sulfuric acid R; continue the heating until a clear solution is obtained; add 45 mL of strong sodium hydroxide solution R.

Oxidising substances (2.5.30)

Maximum 20 ppm, calculated as H₂O₂.

Sulfur dioxide (2.5.29)

Maximum 50 ppm.

Iron (2.4.9)

Maximum 10 ppm.

Shake 1.5 g with 15 mL of dilute hydrochloric acid R. Filter.

The filtrate complies with the test.

Loss on drying (2.2.32)

Maximum 15.0 per cent, determined on 1.000 g by drying in an oven at 130 °C for 90 min.

Sulfated ash (2.4.14)

Maximum 0.6 per cent, determined on 1.0 g.

Microbial contamination

TAMC: acceptance criterion 10³ CFU/g (2.6.12).

TYMC: acceptance criterion 10² CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

◆ Absence of *Salmonella* (2.6.13).

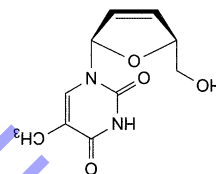
Ph Eur

¹ This monograph has undergone pharmacopoeial harmonisation.

See chapter 5.8 Pharmacopoeial Harmonisation.

Stavudine

(Ph. Eur. monograph 2130)



C₁₀H₁₂N₂O₄

224.2

3056-17-5

Action and use

Nucleoside reverse transcriptase inhibitor; antiviral (HIV).

Ph Eur

DEFINITION

1-(2,3-Dideoxy-β-D-erythro-pent-2-enofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Soluble in water, sparingly soluble in ethanol (96 per cent), slightly soluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Specific optical rotation (2.2.7): -45.9 to -39.5

Dissolve 0.100 g in water R and dilute to 10.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison stavudine CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in anhydrous ethanol R, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or store them at 2-8 °C until use. Test solution Dissolve 25.0 mg of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 0.5 mL of the test solution to 100.0 mL with water R.

Reference solution (b) Dilute 20.0 mL of reference solution (a) to 100.0 mL with water R.

Reference solution (c) Dissolve 5 mg of stavudine for system suitability CRS (containing impurities A, B, C, E and G) in water R and dilute to 10.0 mL with the same solvent.

Column: water R and dilute to 10.0 mL with the same solvent.

stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase: mobile phase A: mix 35 volumes of acetonitrile for chromatography R and 965 volumes of a 0.77 g/L solution of ammonium acetate R;

mobile phase B: mix 250 volumes of acetonitrile for chromatography R and 750 volumes of a 0.77 g/L solution of ammonium acetate R;

chromatography R and 750 volumes of a 0.77 g/L solution of ammonium acetate R;

Time (min) (per cent V/V) Mobile phase A 100 0

Mobile phase B 0 100 (per cent V/V)

Flow rate 2.0 mL/min. Detection Spectrophotometer at 254 nm.

Injection 10 µL. Identification of impurities Use the chromatogram supplied with stavudine for system suitability CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, E and G.

Relative retention With reference to stavudine (retention time = about 10 min): impurity A = about 0.3;

impurity B = about 0.50; impurity C = about 0.53; impurity E = about 1.1; impurity G = about 1.9.

System suitability: reference solution (c): peak-to-valley ratio: minimum 1.5; where H_p = height above the baseline of the peak due to impurity C and H_o = height separating this peak from the peak due to the curve separating this peak from the lowest point of the curve separating this peak from the peak due to stavudine.

Calculation of percentage contents: separating this peak from the peak due to stavudine.

Limit: impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent; impurity G: maximum 0.2 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 1.0 per cent;

— reporting threshold: 0.05 per cent.

Impurity I

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or store them at 2–8 °C until use.

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of stavudine impurity I CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 20.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 30 volumes of acetonitrile for chromatography R and 70 volumes of a 1.15 g/L solution of ammonium dihydrogen phosphate R previously adjusted to pH 6.8 with triethylamine R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 266 nm.

Injection 20 μ L.

Run time 7 times the retention time of stavudine.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peak due to impurity I.

Relative retention With reference to stavudine (retention time = about 3 min): impurity I = about 6.0.

System suitability: reference solution (b):

— signal-to-noise ratio: minimum 40 for the principal peak.

Calculation of percentage content:

— for impurity I, multiply the peak area of impurity I by 1.7;

— for impurity I, use the concentration of stavudine in reference solution (b).

Limit:

— impurity I: maximum 0.15 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or store them at 2–8 °C until use.

Test solution Dissolve 10.0 mg of the substance to be examined in water R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 50.0 mL with water R.

Reference solution (a) Dissolve 10.0 mg of stavudine CRS in water R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 50.0 mL with water R.

Reference solution (b) Dissolve 5 mg of thymine R (impurity A) and 7.5 mg of thymidine R (impurity C) in water R and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 50.0 mL with water R.

Column: — size: $l = 0.033$ m, $\phi = 4.0$ mm; — stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (3 μ m).

Mobile phase Mix 5 volumes of acetonitrile for chromatography R and 95 volumes of a 0.77 g/L solution of ammonium acetate R.

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 25 μ L.

Run time Twice the retention time of stavudine.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and C.

Relative retention With reference to stavudine (retention time = about 4 min): impurity A = about 0.4;

impurity C = about 0.6.

System suitability:

— resolution: minimum 3.5 between the peaks due to impurities A and C in the chromatogram obtained with reference solution (b);

— symmetry factor: maximum 1.6 for the peak due to stavudine in the chromatogram obtained with reference solution (a).

Calculate the percentage content of $C_{10}H_{12}N_2O_4$ using the chromatograms obtained with the test solution and reference solution (a) and taking into account the assigned content of stavudine CRS.

STORAGE

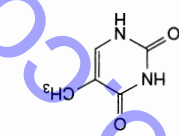
Protected from light and humidity.

IMPURITIES

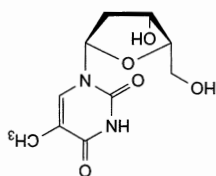
Specified impurities A, G, I

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

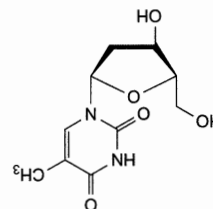
Control of impurities in substances for pharmaceutical use): B, C, D, E, F, H.



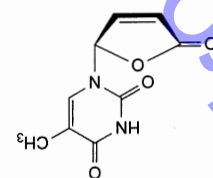
A. 5-methylpyrimidine-2,4(1H,3H)-dione (thymine).



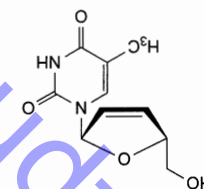
B. 1-(2-deoxy-β-D-threo-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione (3'-epithymidine).



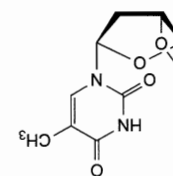
C. 1-(2-deoxy-β-D-erythro-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione (thymidine),



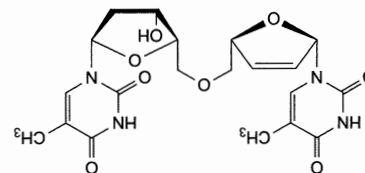
D. 1-[(2R)-5-oxo-2,5-dihydrofuran-2-yl]-5-methylpyrimidine-2,4(1H,3H)-dione,



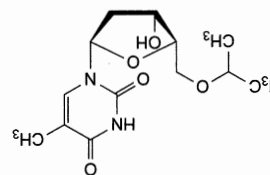
E. 1-(2,3-dideoxy-α-D-glycero-pent-2-enofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione (stavudine anomer α),



F. 1-(3,5-anhydro-2-deoxy-β-D-threo-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione,



G. 1-[2-deoxy-5-O-[(2S,5R)-5-[5-methyl-2,4-dioxo-3,4-dihydropyrimidine-1(2H)-yl]-2,5-dihydrofuran-2-yl]methyl]-β-D-threo-pentofuranosyl]-5-methylpyrimidine-2,4(1H,3H)-dione,



H. 1-[2-deoxy-5-O-(1-methyl-2,3-dideoxy-β-D-threo-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione,

Stearic Acid¹

(Ph. Eur. monograph 1474)

Action and use

Excipient.

Ph. Eur.

DEFINITION

Mixture consisting mainly of stearic (octadecanoic) acid ($C_{18}H_{36}O_2$; M_r 284.5) and palmitic (hexadecanoic) acid ($C_{16}H_{32}O_2$; M_r 256.4) obtained from fats or oils of vegetable or animal origin.

Content

Stearic acid 50	Stearic acid: 40.0 per cent to 60.0 per cent. Sum of the contents of stearic and palmitic acids: minimum 90.0 per cent.
Stearic acid 70	Stearic acid: 60.0 per cent to 80.0 per cent. Sum of the contents of stearic and palmitic acids: minimum 90.0 per cent.
Stearic acid 95	Stearic acid: minimum 90.0 per cent. Sum of the contents of stearic and palmitic acids: minimum 96.0 per cent.

◆ CHARACTERS

Appearance

White or almost white, waxy, flaky crystals, white or almost white hard masses, or white or yellowish-white powder.

Solubility

Practically insoluble in water, soluble in ethanol (96 per cent) and in light petroleum (bp: 50–70 °C). ◆

IDENTIFICATION

A. Freezing point (see Tests).

B. Acid value (2.5.1): 194 to 212, determined on 0.5 g.

C. Examine the chromatograms obtained in the assay.

Results The principal peaks in the chromatogram obtained with the test solution are similar in retention time to those in the chromatogram obtained with the reference solution.

TESTS

Appearance

Heat the substance to be examined to about 75 °C.

The resulting liquid is not more intensely coloured than reference solution Y₇ or BY₇ (2.2.2, Method D). ◇

Acidity

Melt 5.0 g, shake for 2 min with 10 mL of hot carbon

dioxide-free water R, cool slowly and filter. To the filtrate add 0.05 mL of methyl orange solution R. No red colour develops.

Iodine value (2.5.4)

See Table 1474-1.

Freezing point (2.2.18)
See Table 1474-1.

Type	Iodine value	Freezing point (°C)
Stearic acid 50	maximum 4.0	53 - 59
Stearic acid 70	maximum 4.0	57 - 64
Stearic acid 95	maximum 1.5	64 - 69

◆ **Nickel (2.4.31)**
Maximum 1 ppm. ◆

ASSAY
Gas chromatography (2.2.28): use the normalisation procedure.

Test solution In a conical flask fitted with a reflux condenser, dissolve 0.100 g of the substance to be examined in 5 mL of boron trifluoride-methanol solution R. Boil under reflux for 10 min. Add 4.0 mL of heptane R through the condenser and boil again under reflux for 10 min. Allow to cool. Add 20 mL of a saturated solution of sodium chloride R. Shake and allow the layers to separate. Remove about 2 mL of the organic layer and dry it over 0.2 g of anhydrous sodium sulfate R. Dilute 1.0 mL of this solution to 10.0 mL with heptane R.

Reference solution Prepare the reference solution in the same manner as the test solution using 50 mg of palmitic acid CRS and 50 mg of stearic acid CRS instead of the substance to be examined.

Column:
— material: fused silica;
— size: $l = 30$ m, $\varnothing = 0.32$ mm;
— stationary phase: macrogol 20 000 R (film thickness 0.5 µm).
Carrier gas helium for chromatography R.
Flow rate 2.4 mL/min.
Temperature:

Time (min)	Temperature (°C)	Column
0 - 2	70	36 - 41
2 - 36	70 → 240	
36 - 41	240	
41 - 260	220	
260	260	Detector

Detection Flame ionisation.

Injection 1 µL.

Relative retention With reference to methyl stearate: methyl

palmitate = about 0.9.

System suitability: reference solution:

— **resolution:** minimum 5.0 between the peaks due to methyl palmitate and methyl stearate;

— **repeatability:** maximum relative standard deviation of

3.0 per cent for the areas of the peaks due to methyl palmitate and methyl stearate, determined on 6 injections; maximum 1.0 per cent for the ratio of the areas of the peaks due to methyl palmitate to the areas of the peaks due to methyl stearate, determined on 6 injections.

LABELLING

The label states the type of stearic acid (50, 70, 95).

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for stearic acid used as a lubricant in tablets and capsules.

Particle-size distribution (2.9.31)

Specific surface area (2.9.26, Method I)

Ph Eur
1 This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.

Stearoyl Macrogolglycerides

(Ph. Eur. monograph 1268)

Action and use

Excipient.

DEFINITION

Mixtures of monoesters, diesters and triesters of glycerol and monoesters and diesters of macrogols with a mean relative molecular mass between 300 and 4000.

They are obtained by partial alcoholysis of saturated oils containing mainly triglycerides of stearic (octadecanoic) acid, using macrogol, or by esterification of glycerol and macrogol with saturated fatty acids, or by mixture of glycerol esters and condensates of ethylene oxide with the fatty acids of these hydrogenated oils.

The hydroxyl value is within 15 units of the nominal value. The saponification value is within 10 units of the nominal value.

CHARACTERS

Appearance

Pale yellow waxy solid.

Solubility

Dispersible in warm water and in warm liquid paraffin, freely soluble in methylene chloride, soluble in warm anhydrous ethanol.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 1.0 g of the substance to be examined in methylene chloride R and dilute to 20 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase hexane R, ether R (30:70 V/V).

Application 10 µL.

Development Over a path of 15 cm.



Ph Eur



Stearyl Alcohol

(Ph. Eur. monograph 0753)

Action and use

Excipient.

Ph Eur

DEFINITION

Mixture of solid alcohols, mainly octadecan-1-ol ($C_{18}H_{38}O$; M_r 270.5), of animal or vegetable origin.

Content

Minimum 95.0 per cent of $C_{18}H_{38}O$.

CHARACTERS

Appearance

White or almost white, unctuous flakes, granules or mass.

Solubility

Practically insoluble in water, soluble in ethanol (96 per cent). When melted, it is miscible with fatty oils, with liquid paraffin and with melted wool fat.

IDENTIFICATION

Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time to the principal peak in the chromatogram obtained with reference solution (b).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Dissolve 0.50 g in 20 mL of boiling ethanol (96 per cent) R. Allow to cool.

Melting point (2.2.14) 57 °C to 60 °C.

Acid value (2.5.1) Maximum 1.0.

Hydroxyl value (2.5.3, Method A) 197 to 217.

Iodine value (2.5.4, Method A) Maximum 2.0.

Dissolve 2.00 g in methylene chloride R, warming if necessary and dilute to 25 mL with the same solvent.

Saponification value (2.5.6) Maximum 2.0.

ASSAY

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution Dissolve 0.100 g of the substance to be examined in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Drying In air.

Detection Spray with a 0.1 g/L solution of rhodamine B R in ethanol (96 per cent) R and examine in ultraviolet light at 365 nm.

Results The chromatogram shows a spot due to triglycerides with an R_f value of about 0.9 (R_f 1) and spots due to 1,3-diglycerides (R_f 0.7), to 1,2-diglycerides (R_f 0.6), to monoglycerides (R_f 0.1) and to esters of macrogol (R_f 0).

B. Hydroxyl value (see Tests).

C. Saponification value (see Tests).

D. Fatty acid composition (see Tests).

TESTS

Acid value (2.5.1) Maximum 2.0, determined on 2.0 g.

Hydroxyl value (2.5.3, Method A) Within 15 units of the nominal value, determined on 1.0 g.

Peroxide value (2.5.5, Method A) Maximum 6.0, determined on 2.0 g.

Saponification value (2.5.6) Within 10 units of the nominal value, determined on 2.0 g.

Alkaline impurities

Into a test-tube introduce 5.0 g and carefully add a mixture, neutralised if necessary with 0.01 M hydrochloric acid or with 0.01 M sodium hydroxide, of 0.05 mL of a 0.4 g/L solution of bromophenol blue R in ethanol (96 per cent) R, 0.3 mL of water R and 10 mL of ethanol (96 per cent) R. Shake and allow to stand. Not more than 1.0 mL of 0.01 M hydrochloric acid is required to change the colour of the upper layer to yellow.

Free glycerol

Maximum 3.0 per cent.

Dissolve 1.20 g in 25.0 mL of methylene chloride R. Heat if necessary. After cooling, add 100 mL of water R. Shake and add 25.0 mL of peracetic acid solution R. Shake and allow to stand for 30 min. Add 40 mL of a 75 g/L solution of potassium iodide R. Allow to stand for 1 min. Add 1 mL of starch solution R. Titrate the iodine with 0.1 M sodium thiosulfate. Carry out a blank titration.

1 mL of 0.1 M sodium thiosulfate is equivalent to 2.3 mg of glycerol.

Composition of fatty acids

Gas chromatography (2.4.22, Method A).

Composition of the fatty-acid fraction of the substance:

— lauric acid: maximum 5.0 per cent;
— myristic acid: maximum 5.0 per cent;
— stearic acid and palmitic acid: different nominal amounts and minimum 90.0 per cent for the sum of $C_{18}H_{36}O_2$ and $C_{16}H_{32}O_2$.

Ethylene oxide and dioxan (2.4.25) Maximum 1 ppm of ethylene oxide and maximum 10 ppm of dioxan.

Heavy metals (2.4.8) Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12) Maximum 1.0 per cent, determined on 1.0 g. Use a mixture of 30 volumes of anhydrous methanol R and 70 volumes of methylene chloride R as solvent.

Total ash (2.4.16) Maximum 0.2 per cent.

CHARACTERS**Appearance**

Clear, colourless liquid.

IDENTIFICATION

A. Place 0.5 mL of citrated human plasma in a polystyrene tube maintained in a water-bath at 37 °C. Add 0.1 mL of a dilution of the preparation to be examined containing 10 000 IU of streptokinase activity per millilitre in phosphate buffer solution pH 7.2 R and 0.1 mL of a solution of human thrombin R containing 20 IU/mL in phosphate buffer solution pH 7.2 R. Mix immediately. A clot forms and lyses within 30 min. Repeat the procedure using citrated bovine plasma.

The clot does not lyse within 60 min.

B. Perform an immunochemical test using double immunodiffusion techniques (2.7.1). Place in the central cavity about 80 µL of goat or rabbit antistreptokinase serum containing about 10 000 units of antistreptokinase activity per millilitre; place in each of the surrounding cavities about 80 µL of a dilution of the preparation to be examined containing 125 000 IU of streptokinase activity per millilitre.

Allow the plates to stand in a humidified tank for 24 h. Only one precipitation arc appears and it is well defined.

TESTS

pH (2.2.3)

6.8 to 7.5.

Dilute the preparation to be examined in carbon dioxide-free water R to obtain a solution containing at least 1000 000 IU of streptokinase activity per millilitre.

Streptodornase

Maximum 10 IU of streptodornase activity per 100 000 IU of streptokinase activity.

Test solution Dilute the preparation to be examined in imidazole buffer solution pH 6.5 R to obtain a solution containing 150 000 IU of streptokinase activity per millilitre.

Reference solution Dissolve in imidazole buffer solution pH 6.5 R a reference preparation of streptodornase, calibrated in International Units against the International Standard of streptodornase, to obtain a solution containing 20 IU of streptodornase activity per millilitre. The equivalence in International Units of the International Standard is stated by the World Health Organization.

To each of 8 numbered centrifuge tubes, add 0.5 mL of a 1 g/L solution of sodium deoxyribonuclease R in imidazole buffer solution pH 6.5 R. To tube number 1 and tube number 2 add 0.25 mL of imidazole buffer solution pH 6.5 R, 0.25 mL of the test solution and, immediately, 3.0 mL of perchloric acid (25 g/L HClO₄). Mix, centrifuge at about 3000 g for 5 min and measure the absorbances (2.2.25) of the supernatant liquids at 260 nm, using as the compensation liquid a mixture of 1.0 mL of imidazole buffer solution pH 6.5 R and 3.0 mL of perchloric acid (25 g/L HClO₄) (absorbances A₁ and A₂). To the other 6 tubes (numbers 3 to 8) add 0.25 mL, 0.25 mL, 0.125 mL, 0.125 mL, 0 mL and 0 mL respectively of imidazole buffer solution pH 6.5 R; add to each tube 0.25 mL of the test solution and 0 mL, 0 mL, 0.125 mL, 0.125 mL, 0.25 mL and 0.25 mL respectively of the reference solution. Mix the contents of each tube and heat at 37 °C for 15 min. To each tube add 3.0 mL of perchloric acid (25 g/L HClO₄), mix and centrifuge. Measure the absorbances (2.2.25) of the supernatant liquids at 260 nm using the compensation liquid described above (absorbances A₃ to A₈). The absorbances comply with the following requirement:

Streptokinase Concentrated Solution

Streptokinase Bulk Solution
(Ph. Eur. monograph 0356)

Action and use
Plasminogen activator; fibrinolytic.

DEFINITION

Streptokinase concentrated solution is a preparation of a protein obtained from culture filtrates of certain strains of haemolytic *Streptococcus* group C; it has the property of combining with human plasminogen to form plasminogen activator. It may contain buffer salts and other excipients. The potency is not less than 510 IU per microgram of nitrogen.

PRODUCTION

The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.

Abnormal toxicity (2.6.9)
Inject into each mouse a quantity of the preparation to be examined (if necessary, dilute with water for injections R) equivalent to 50 000 IU of streptokinase activity; the injection lasting 15–20 s.

Temperature (°C)	Time (min)		Column	Injection port	Detector
	0–20	20–40			
150 → 250				250	250
250				250	250

Temperature:
Split ratio 1:100.
Flow rate 1 mL/min.
Carrier gas helium for chromatography R.
— stationary phase: poly(dimethyl)siloxane R (1 µm).
— size: l = 30 m, Ø = 0.32 mm,
— ethanol (96 per cent) R and dilute to 10 mL with solvent.
Reference solution (a) Dissolve 50 mg of cetyl alcohol R in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.
Reference solution (b) Dissolve 50 mg of stearyl alcohol CRS in ethanol (96 per cent) R and dilute to 5 mL with the same solvent.
Reference solution (c) Mix 1 mL of reference solution (a) and 1 mL of reference solution (b) and dilute to 10 mL with ethanol (96 per cent) R.



$$(A_3 + A_4) < \frac{2}{(A_5 + A_6 + A_7 + A_8) - (A_3 + A_4)}$$

Streptolysin

In a polystyrene tube, use a quantity of the preparation to be examined equivalent to 500 000 IU of streptokinase activity and dilute to 0.5 mL with a mixture of 1 volume of phosphate buffer solution pH 7.2 R and 9 volumes of a 9 g/L solution of sodium chloride R. Add 0.4 mL of a 23 g/L solution of sodium thioglycollate R. Heat in a water-bath at 37 °C for 10 min. Add 0.1 mL of a solution of a reference preparation of human antistreptolysin O containing 5 IU/mL. Heat at 37 °C for 5 min. Add 1 mL of rabbit erythrocyte suspension R. Heat at 37 °C for 30 min. Centrifuge at about 1000 g. In the same manner, prepare a polystyrene tube in which the solution of the preparation to be examined has been replaced by 0.5 mL of a mixture of 1 volume of phosphate buffer solution pH 7.2 R and 9 volumes of a 9 g/L solution of sodium chloride R. Measure the absorbances (2.2.25) of the supernatant liquids at 550 nm. The absorbance of the test solution is not more than 50 per cent greater than that of the reference solution.

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dilute the preparation to be examined with water R to obtain a concentration of about 0.5–1 g/L, depending on the chromatographic system used.

Reference solution Dilute 1 volume of streptokinase for system suitability CRS with 49 volumes of water R.

Column:

- size: $l = 0.10$ m, $\phi = 4.6$ mm;
- stationary phase: styrene-divinylbenzene copolymer R (10 μ m) with a pore size of 200 nm;
- temperature: 25 °C.

Mobile phase:

- mobile phase A: trifluoroacetic acid R, water for injections R (1:1000 V/V); degas;
- mobile phase B: trifluoroacetic acid R, acetonitrile for chromatography R (1:1000 V/V); degas;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0–1	68	32
1–4	68 \rightarrow 52	32 \rightarrow 48
4–5	52	48
5–7	0	100
7–10	68	32

The above conditions may be modified to improve the separation efficiency of the chromatographic system.

Flow rate 5 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L.

Retention time: streptokinase = 2.3 min to 2.8 min.

System suitability: reference solution:

- symmetry factor: maximum 1.9 for the peak due to streptokinase;
- peak-to-valley ratio: minimum 2, where H_p = height above the baseline of the 1st peak eluting after the principal peak and H_v = height above the baseline of the lowest point of the curve separating this peak from the 2nd peak eluting after the principal peak;

— the chromatogram obtained with the reference solution is similar to the chromatogram supplied with streptokinase for system suitability CRS.

Limit:

— total: maximum 5 per cent.

Bacterial endotoxins (2.6.14)

Less than 0.02 IU per 100 IU of streptokinase activity, if intended for use without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Nitrogen (2.5.9)

Potency

The potency of streptokinase is determined by comparing its capacity to activate plasminogen to form plasmin with the same capacity of a reference preparation of streptokinase calibrated in International Units; the formation of plasmin is determined using a suitable chromogenic substrate.

The International Unit is the activity of a stated amount of the International Standard for streptokinase. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Reference and test solutions

Prepare 2 independent series of at least 3 dilutions of each of the preparation to be examined and of the reference preparation of streptokinase in *tris(hydroxymethyl)aminomethane sodium chloride buffer solution* pH 7.4 R₁ in the linear range of the assay (a range of 0.5–4.0 IU/mL has been found suitable). Prepare and maintain all solutions at 37 °C.

Substrate solution

Mix 1.0 mL of *tris(hydroxymethyl)aminomethane buffer solution* pH 7.4 R with 1.0 mL of *chromogenic substrate* R₃. Add 5 μ L of a 100 g/L solution of *polyorbate* 20 R. Keep at 37 °C in a water-bath. Immediately before commencing the activation assay, add 45 μ L of a 1 mg/mL solution of *human plasminogen* R.

Analyse each streptokinase dilution, maintained at 37 °C, in duplicate. Initiate the activation reaction by adding 60 μ L of each dilution to 40 μ L of substrate solution. For blank wells, use 60 μ L of *tris(hydroxymethyl)aminomethane sodium chloride buffer solution* pH 7.4 R₁ instead of the reference and test solutions. Allow the reaction to proceed at 37 °C for 20 min and read the absorbance (2.2.25) at 405 nm. If a suitable thermostatted plate reader is available, this may be used to monitor the reaction. Alternatively, it may be necessary to stop the reaction after 20 min using 50 μ L of a 50 per cent V/V solution of *glacial acetic acid* R. Best results are obtained when the absorbance for the highest streptokinase concentration is between 0.1 and 0.2 (after blank subtraction). If necessary, adjust the time of incubation in order to reach this range of absorbances.

Calculate the regression of the absorbance on log concentrations of the solutions of the preparation to be examined and of the reference preparation of streptokinase using a suitable statistical method, for example the parallel-line assay (5.3).

The estimated potency is not less than 90 per cent and not more than 111 per cent of the stated potency.

The confidence limits ($P = 0.95$) are not less than 80 per cent and not more than 125 per cent of the estimated potency.

STORAGE

In an airtight container, protected from light and at a temperature of -20°C . If the preparation is sterile, store in a sterile, airtight, tamper-proof container.

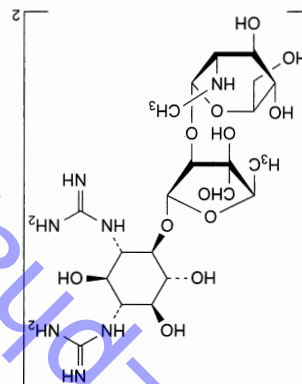
LABELLING

The label states:
— the number of International Units of streptokinase activity per milligram, calculated with reference to the dried preparation;
— that the preparation is suitable for use in the manufacture of parenteral preparations.

Ph Eur

Streptomycin Sulfate

Streptomycin Sulphate
(Ph. Eur. monograph 0053)



$\text{C}_{42}\text{H}_{84}\text{N}_{14}\text{O}_{36}\text{S}_3$ 1457 3810-74-0

Action and use
Aminoglycoside antibacterial; antituberculosis drug.

Preparation
Streptomycin Injection

Ph Eur

DEFINITION

Streptomycin sulfate is bis[N,N' -bis(aminomimethyl)-4-O-[(5-deoxy-2-O-[(2-deoxy-2-(methylamino)-α-L-glucopyranosyl]-3-C-formyl-α-L-lyxofuranosyl)-D-streptamino] tri]sulfate, a substance produced by the growth of certain strains of *Streptomyces griseus* or obtained by any other means. Stabilisers may be added. The potency is not less than 720 IU/mg, calculated with reference to the dried substance.

PRODUCTION

It is produced by methods of manufacture designed to eliminate or minimise substances lowering blood pressure. The method of manufacture is validated to demonstrate that the product if tested would comply with the following test:

Abnormal toxicity (2.6.9)

Inject into each mouse 1 mg of the substance to be examined dissolved in 0.5 mL of water for injections R.

CHARACTERS

A white or almost white powder, hygroscopic, very soluble in water, practically insoluble in ethanol.

IDENTIFICATION

A. Examine by thin-layer chromatography (2.2.27), using a plate coated with a 0.75 mm layer of the following mixture: mix 0.3 g of carbomer R with 240 mL of water R and allow to stand, with moderate shaking, for 1 h; adjust to pH 7 by the gradual addition, with continuous shaking, of dilute sodium hydroxide solution R and add 30 g of silica gel H R. Heat the plate at 110°C for 1 h, allow to cool and use immediately.

Test solution Dissolve 10 mg of the substance to be examined in water R and dilute to 10 mL with the same solvent.
Reference solution (a) Dissolve 10 mg of streptomycin sulfate CRS in water R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of kanamycin monosulfate CRS, 10 mg of neomycin sulfate CRS and 10 mg of streptomycin sulfate CRS in water R and dilute to 10 mL with the same solvent.

Apply separately to the plate 10 μL of each solution. Develop over a path of 12 cm using a 70 g/L solution of potassium dihydrogen phosphate R. Dry the plate in a current of warm air, and spray with a mixture of equal volumes of a 2 g/L solution of 1,3-dihydroxynaphthalene R in alcohol R and a 460 g/L solution of sulfuric acid R. Heat at 150°C for 5 min to 10 min. The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the spot in the chromatogram obtained with reference solution (a). The test is not valid unless the chromatogram obtained with reference solution (b) shows three clearly separated spots.

B. Dissolve 5 mg to 10 mg in 4 mL of water R and add 1 mL of 1 M sodium hydroxide. Heat in a water-bath for 4 min. Add a slight excess of dilute hydrochloric acid R and 0.1 mL of ferric chloride solution R1. A violet colour develops. C. Dissolve 0.1 g in 2 mL of water R, add 1 mL of α-naphthol solution R and 2 mL of a mixture of equal volumes of strong sodium hypochlorite solution R and water R. A red colour develops.

D. Dissolve about 10 mg in 5 mL of water R and add 1 mL of 1 M hydrochloric acid. Heat in a water-bath for 2 min. Add 2 mL of a 5 g/L solution of α-naphthol R in 1 M sodium hydroxide and heat in a water-bath for 1 min. A faint yellow colour develops. E. It gives the reactions of sulfates (2.3.1).

TESTS

Solution S
Dissolve 2.5 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Appearance of solution

Solution S is not more intensely coloured than intensity 3 of the range of reference solutions of the most appropriate colour (2.2.2, Method II). Allow to stand protected from light, at a temperature of about 20°C for 24 h. Solution S is not more opalescent than reference suspension II (2.2.1).

pH (2.2.3)

The pH of solution S is 4.5 to 7.0.

Methanol

Examine by gas chromatography (2.2.28).

Test solution Dissolve 1.00 g of the substance to be examined in water R and dilute to 25.0 mL with the same solvent.
Reference solution Dilute 12.0 mg of methanol R to 100 mL with water R.
The chromatographic procedure may be carried out using:

- a column 1.5 m to 2.0 m long and 2 mm to 4 mm in internal diameter, packed with ethylvinylbenzene-divinylbenzene copolymer R (150 µm to 180 µm), nitrogen for chromatography R as the carrier gas at a constant flow rate of 30 mL to 40 mL per minute, — a flame-ionisation detector.

Maintain the column at a constant temperature between 120 °C and 140 °C and the injection port and the detector at a temperature at least 50 °C higher than that of the column. Inject the test solution and the reference solution. The area of the peak due to methanol in the chromatogram obtained with the test solution is not greater than the area of the peak in the chromatogram obtained with the reference solution (0.3 per cent).

Streptomycin B

Examine by thin-layer chromatography (2.2.27), using silica gel G R as the coating substance.

Test solution Dissolve 0.2 g of the substance to be examined in a freshly prepared mixture of 3 volumes of sulfuric acid R and 97 volumes of methanol R and dilute to 5 mL with the same mixture of solvents. Heat under a reflux condenser for 1 h, cool, rinse the condenser with methanol R and dilute to 50 mL with the same solvent. Dilute 5 mL of the solution to 50 mL with methanol R (0.3 g/L solution expressed as strepto-mycin B; 1 mg of mannose R is equivalent to 4.13 mg of streptomycin B).

Reference solution Dissolve 36 mg of mannose R in a freshly prepared mixture of 3 volumes of sulfuric acid R and 97 volumes of methanol R and dilute to 5 mL with the same mixture of solvents. Heat under a reflux condenser for 1 h, cool, rinse the condenser with methanol R and dilute to 50 mL with the same solvent. Dilute 5 mL of the solution to 50 mL with methanol R (0.3 g/L solution expressed as strepto-mycin B; 1 mg of mannose R is equivalent to 4.13 mg of streptomycin B).

Apply separately to the plate 10 µL of each solution. Develop over a path of 13 cm to 15 cm using a mixture of 25 volumes of glacial acetic acid R, 25 volumes of methanol R and 50 volumes of toluene R. Allow the plate to dry in air and spray with a freshly prepared mixture of equal volumes of a 2 g/L solution of 1,3-dihydroxynaphthalene R in alcohol R and a 20 per cent V/V solution of sulfuric acid R and heat at 110 °C for 5 min. Any spot corresponding to streptomycin B in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with the reference solution (3.0 per cent).

Loss on drying (2.2.32)
Not more than 7.0 per cent, determined on 1.000 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 0.1 kPa for 24 h.

Sulfated ash (2.4.14)
Not more than 1.0 per cent, determined on 1.000 g.

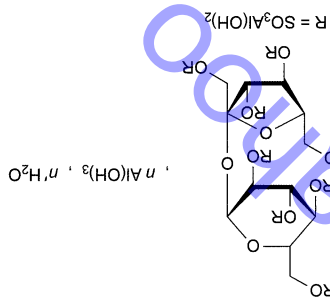
Sulfate
18.0 per cent to 21.5 per cent of sulfate (SO₄), calculated with reference to the dried substance. Dissolve 0.250 g in 100 mL of water R and adjust the solution to pH 11 using concentrated ammonia R. Add 10.0 mL of 0.1 M barium chloride and about 0.5 mg of phthalin purple R. Titrate with 0.1 M sodium edetate adding 50 mL of alcohol R when the colour of the solution begins to change and continue the titration until the violet-blue colour disappears.

1 mL of 0.1 M barium chloride is equivalent to 9.606 mg of sulfate (SO₄).

Colorimetric test
Dry the substance to be examined and streptomycin sulfate CRS at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 0.1 kPa for 24 h. Dissolve 0.100 g of

Sucralfate

(Ph. Eur. monograph 1796)



$C_{12}H_{30}Al_8O_{51}S_8[Al(OH)_3]_n[H_2O]_{n'}$
in which $n = 8$ to 10 and $n' = 22$ to 31

Action and use

Treatment of gastric and duodenal ulcers.

DEFINITION

β-D-Fructofuranosyl-α-D-glucopyranoside octakis(dihydroxyaluminum sulfate) with 8-10 molecules of aluminum hydroxide and 22-31 molecules of water.

Content

— β-D-fructofuranosyl-α-D-glucopyranoside octakis sulfate (C₁₂H₁₄O₃₅S₈; 975); 30.0 per cent to 36.0 per cent;

— aluminum (Al; A_r 26.98): 16.5 per cent to 18.5 per cent.

CHARACTERS

Appearance

White or almost white, amorphous powder.

Solubility

Practically insoluble in water, in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute solutions of mineral acids and alkali hydroxides.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sucralfate CRS.

B. To 2 g add 10 mL of 0.1 M hydrochloric acid and boil.

Cool and neutralise with 0.1 M sodium hydroxide. To 5 mL of

the solution add 0.15 mL of freshly prepared copper sulfate

solution R and 2 mL of freshly prepared dilute sodium

hydroxide solution R. The solution is blue and clear and

remains so after boiling. To the hot solution add 4 mL of

dilute sodium hydroxide solution R; an orange precipitate is

formed immediately.

C. Dissolve about 15 mg in a mixture of 0.5 mL of dilute

hydrochloric acid R and 2 mL of water R. The solution gives

the reaction of aluminium (2.3.1).

TESTS**Impurity A**

Liquid chromatography (2.2.29).

Test solution Dissolve 450.0 mg of the substance to be

examined in a mixture of equal volumes of an 88 g/L

solution of sodium hydroxide R and a 196.2 g/L solution of

sulfuric acid R and dilute to 10.0 mL with the same mixture

of solvents. Without delay, while shaking at a moderate rate,

add a volume (V), accurately measured in millilitres, of a

4 g/L solution of sodium hydroxide R to adjust the solution to

approximately pH 2.3. Dilute the solution with (15.0

–V) mL of water R. Shake for 1 min. If the pH is not

between 2.3 and 3.5, repeat the test using a different volume

of 4 g/L solution of sodium hydroxide R.

Reference solution (a) Dissolve 40.0 mg of potassium sucrose

octasulfate CRS in the mobile phase and dilute to 5.0 mL

with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a)

to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: aminopropylsilyl silica gel for

chromatography R (5 μ m).

Mobile phase 70 g/L solution of ammonium sulfate R, adjusted

to pH 3.5 with phosphoric acid R.

Flow rate 1 mL/min.

Detection Differential refractometer.

Injection 50 μ L of the test solution and reference solution (b).

Relative retention With reference to sucrose octasulfate

(retention time = about 6 min): impurity A = about 0.6.

System suitability: reference solution (b):

— number of theoretical plates: minimum 400;

— symmetry factor: maximum 4.0.

Limit:

— impurity A: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (5.0 per cent).

Neutralising capacity

Disperse 0.25 g in 100.0 mL of 0.1 M hydrochloric acid,

previously heated at 37 °C, stir continuously for 1 h in a

solution with 0.1 M sodium hydroxide to pH 3.5; not more

than 14.0 mL of 0.1 M sodium hydroxide is required.

Chlorides (2.4.4)

Maximum 0.50 per cent.

Dissolve 0.10 g in 5 mL of dilute nitric acid R and dilute to

50 mL with water R. Dilute 5 mL of this solution to 15 mL

Arsenic (2.4.2, Method A)

Maximum 4 ppm.

Introduce 0.25 g of the substance to be examined and 5 mL

of sulfuric acid R into a combustion flask. Carefully add a few

millilitres of strong hydrogen peroxide solution R and heat to

boiling until a clear, colourless solution is obtained. Continue

heating to eliminate the water and as much sulfuric acid as

possible and dilute to 25 mL with water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test F. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

ASSAY**Aluminium**

Disperse 1.0 g in 10 mL of 6 M hydrochloric acid R. Heat

with continuous stirring in a water-bath at 70 °C for 5 min.

Cool to room temperature, transfer quantitatively to a

volumetric flask, dilute to 250.0 mL with water R, and mix.

Filter the solution, discarding the 1st portion of the filtrate.

To 10.0 mL of the solution, add 10.0 mL of 0.1 M sodium

edate and 30 mL of a mixture of equal volumes of

ammonium acetate solution R and dilute acetic acid R. Heat in a

water-bath at 70 °C for 5 min, then cool. Add 25 mL of

ethanol (96 per cent) R and 1 mL of a freshly prepared

0.25 g/L solution of diethyzone R in ethanol (96 per cent) R.

Titrate the excess of sodium edetate with 0.1 M zinc sulfate

until the colour changes to pink.

1 mL of 0.1 M sodium edetate is equivalent to 2.698 mg of

Al.

Sucrose octasulfate

Liquid chromatography (2.2.29) as described in the test for

impurity A with the following modifications.

Mobile phase 132 g/L solution of ammonium sulfate R,

adjusted to pH 3.5 with phosphoric acid R.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{12}H_{14}O_{35}S_8$ from the

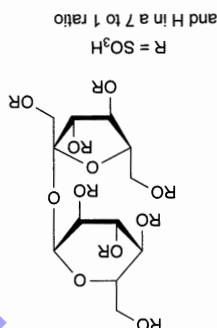
declared content of potassium sucrose octasulfate CRS and by

multiplying the potassium sucrose octasulfate content by

0.757.

IMPURITIES

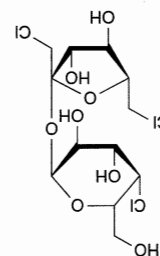
Specified impurities A



A. β -D-fructofuranosyl- α -D-glucopyranoside heptakis (hydrogensulfate).

Sucralose

(Ph Eur monograph 2368)



$C_{12}H_{19}Cl_3O_8$ 397.6 56038-13-2

Action and use

Sweetening agent.

Ph Eur

DEFINITION

1,6-Dichloro-1,6-dideoxy- β -D-fructofuranosyl 4-chloro-4-deoxy- α -D-galactopyranoside.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in anhydrous ethanol, slightly soluble in ethyl acetate.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison sucralose CRS.

TESTS

Specific optical rotation (2.2.7)
+ 84.0 to + 87.5 (anhydrous substance).

Dissolve 2.50 g in water R and dilute to 25.0 mL with the same solvent.

Impurities H and I

Test solution Dissolve 2.5 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 1.0 g of mannitol R in water R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dissolve 1.0 g of mannitol R and 4.0 mg of fructose R in water R and dilute to 10.0 mL with the same solvent.

Plate TLC silica gel plate R.

Application 5 μ L by applying the solution slowly in 1 μ L aliquots and allowing the plate to dry between applications; the 3 spots must be of a similar size.

Detection Spray with a solution prepared as follows: dissolve 1.23 g of *p*-anisidine R and 1.66 g of phthalic acid R in 100 mL of methanol R; store the solution in darkness and in a refrigerator to prevent it becoming discoloured; discard if the solution becomes discoloured; heat the plate at $100 \pm 2^\circ\text{C}$ for 15 min and examine immediately against a dark background.

System suitability The spot due to mannitol obtained with reference solution (a) is colourless; darkening of the mannitol

spot indicates that the plate has been held for too long in the oven and a 2nd plate has to be prepared.

Limit:

— sum of impurities H and I: any spot is not more intense than the spot due to fructose obtained with reference solution (b) (0.1 per cent).

Related substances

Thin-layer chromatography (2.2.27).

Test solution Dissolve 1.0 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 0.5 mL of the test solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve the contents of a vial of sucralose impurity B CRS in 1.0 mL of the test solution.

Plate TLC octadecylsilyl silica gel plate R.

Mobile phase acetonitrile R, 50 g/L solution of sodium chloride R (30:70 V/V).

Application 5 μ L.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with a 15 per cent V/V solution of sulfuric acid R in methanol R and heat at 125°C for 10 min.

Retention factors Impurity A = about 0.2;

impurity B = about 0.3; sucralose = about 0.4;

impurity F = about 0.67; impurity E = about 0.71;

impurity G = about 0.73; impurity D = about 0.8.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots due to impurity B and sucralose.

Limits:

— impurities A, B, D, E, F, G: any spot, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (a)

(0.5 per cent).

Heavy metals

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution

(1 ppm Pb) R.

Water

(2.5.12)

Maximum 2.0 per cent, determined on 0.500 g.

Sulfated ash

(2.4.14)

Maximum 0.7 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29).

Test solution Dissolve 0.25 g of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution Dissolve 0.25 g of sucralose CRS in the mobile phase and dilute to 25.0 mL with the mobile phase.

Column:

— size: $l = 0.10$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R, water R (15:85 V/V).

Flow rate 1.5 mL/min.

Detection Refractometer maintained at a constant temperature.

Injection 20 μ L.

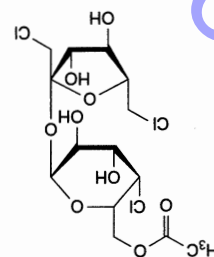
Retention time Sucralose = about 3 min.

System suitability: reference solution:
— symmetry factor: maximum 2.0 for the peak due to sucralose.

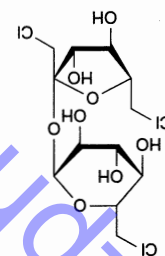
Calculate the percentage content of $C_{12}H_{19}Cl_3O_8$ from the declared content of sucralose CRS.

IMPURITIES

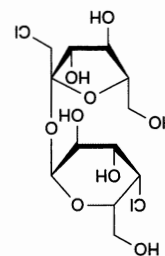
Specified impurities A, B, D, E, F, G, H, I



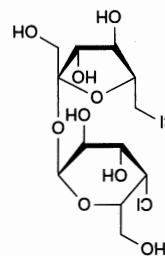
A. 1,6-dichloro-1,6-dideoxy-β-D-fructofuranosyl 6-O-acetyl-4-chloro-4-deoxy-α-D-galactopyranoside (6-O-acetylsucralose),



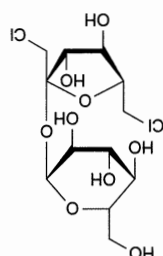
B. 1,6-dichloro-1,6-dideoxy-β-D-fructofuranosyl 6-chloro-6-deoxy-α-D-glucopyranoside (1',6,6'-trichloro-1',6,6'-trideoxysucrose),



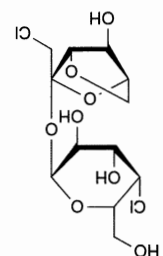
D. 1-chloro-1-deoxy-β-D-fructofuranosyl 4-chloro-4-deoxy-α-D-galactopyranoside (1',4-dichloro-1',4-dideoxygalactosucrose),



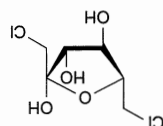
E. 6-chloro-6-deoxy-β-D-fructofuranosyl 4-chloro-4-deoxy-α-D-galactopyranoside (4,6'-dichloro-4,6'-dideoxygalactosucrose),



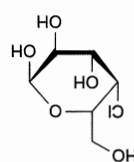
F. 1,6-dichloro-1,6-dideoxy-β-D-fructofuranosyl α-D-glucopyranoside (1',6'-dichloro-1',6'-dideoxysucrose),



G. 3,6-anhydro-1-chloro-1-deoxy-β-D-fructofuranosyl 4-chloro-4-deoxy-α-D-galactopyranoside (3',6'-anhydro-1',4'-dichloro-1',4'-dideoxygalactosucrose),

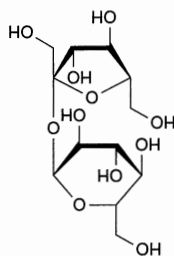


H. 1,6-dichloro-1,6-dideoxy-β-D-fructofuranose,



I. 4-chloro-4-deoxy-α-D-galactopyranose.

Sucrose¹



$C_{12}H_{22}O_{11}$

342.3

57-50-1

DEFINITION

β-D-Fructofuranosyl α-D-glucopyranoside.
It contains no additives.



◆ CHARACTERS

Appearance

White or almost white, crystalline powder, or lustrous, colourless or white or almost white crystals.

Solubility

Very soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in anhydrous ethanol.

IDENTIFICATION

First identification A

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sucrose CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a mixture of 2 volumes of water R and 3 volumes of methanol R and dilute to 20 mL with the same mixture of solvents.

Reference solution (a) Dissolve 10 mg of sucrose CRS in a mixture of 2 volumes of water R and 3 volumes of methanol R and dilute to 20 mL with the same mixture of solvents.

Reference solution (b) Dissolve 10 mg each of fructose CRS, glucose CRS, lactose CRS and sucrose CRS in a mixture of 2 volumes of water R and 3 volumes of methanol R and dilute to 20 mL with the same mixture of solvents.

Reference solution (c) Dissolve 10 mg each of fructose CRS, glucose CRS, lactose CRS and sucrose CRS in a mixture of 2 volumes of water R and 3 volumes of methanol R and dilute to 20 mL with the same mixture of solvents.

Plate TLC silica gel G plate R.

Mobile phase cold saturated boric acid solution R, 60 per cent V/V solution of glacial acetic acid R, ethanol R, acetone R, ethyl acetate R (10:15:20:60:60 V/V/V/V/V).

Application 2 µL.

Development In an unsaturated tank over a path of 15 cm. Drying In a current of warm air.

Detection Spray with a solution of 0.5 g of thymol R in a mixture of 5 mL of sulfuric acid R and 95 mL of alcohol R. Heat the plate at 130 °C for 10 min.

System suitability The chromatogram obtained with reference solution (b) shows 4 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dilute 1 mL of solution S (see Tests) to 100 mL with water R. To 5 mL of the solution add 0.15 mL of freshly prepared copper sulfate solution R and 2 mL of freshly prepared dilute sodium hydroxide solution R. The solution is blue and clear and remains so after boiling. To the hot solution add 4 mL of dilute hydrochloric acid R and boil for 1 min. Add 4 mL of dilute sodium hydroxide solution R. An orange precipitate is formed immediately. ◆

TESTS

Solution S

Dissolve 50.0 g in water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1).

Conductivity (2.2.38)

Maximum 35 µS·cm⁻¹ at 20 °C.

Dissolve 31.3 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent. Measure the conductivity of the solution (C₁), while gently stirring with a magnetic stirrer, and that of the water used for preparing the solution (C₂). The readings must be stable

within 1 per cent over a period of 30 s. Calculate the conductivity of the solution of the substance to be examined from the following expression:

$$C_1 - 0.35 C_2$$

Specific optical rotation (2.2.7)

+ 66.3 to + 67.0.

Dissolve 26.0 g in water R and dilute to 100.0 mL with the same solvent.

◆ Colour value

Maximum 45.

Dissolve 50.0 g in 50.0 mL of water R. Mix, filter (diameter of pores 0.45 µm) and degas. Measure the absorbance (2.2.25) at 420 nm, using a minimum path length of 4 cm (a path length of 10 cm or more is preferred).

Calculate the colour value using the following expression:

$$A \times 1000 \frac{b \times c}{b \times c}$$

A = absorbance measured at 420 nm;

b = path length in centimetres;

c = concentration of the solution, in grams per millilitre, calculated from the refractive index (2.2.6) of the solution; use Table 0204-1 and interpolate the values if necessary.

Table 0204-1

n_D^{20}	c (g/mL)
1.4138	0.570
1.4159	0.585
1.4179	0.600
1.4200	0.615
1.4221	0.630
1.4243	0.645
1.4264	0.661

System suitability:

— repeatability: the absolute difference between 2 results is not greater than 3. ◆

Dextrins

If intended for use in the manufacture of large-volume parenteral preparations, it complies with the test for dextrins.

To 2 mL of solution S add 8 mL of water R, 0.05 mL of dilute hydrochloric acid R and 0.05 mL of 0.05 M iodine.

The solution remains yellow.

Reducing sugars

To 5 mL of solution S in a test-tube about 150 mm long and 16 mm in diameter add 5 mL of water R, 1.0 mL of 1 M sodium hydroxide and 1.0 mL of a 1 g/L solution of methylene blue R. Mix and place in a water-bath. After exactly 2 min, take the tube out of the bath and examine the solution immediately. The blue colour does not disappear completely. Ignore any blue colour at the air/solution interface.

Sulfites

Maximum 10 ppm, calculated as SO₂.

Determine the sulfites content by a suitable enzymatic method based on the following reactions. Sulfite is oxidised by sulfite oxidase to sulfate and hydrogen peroxide which in

turn is reduced by nicotinamide-adenine dinucleotide-peroxidase in the presence of reduced nicotinamide-adenine dinucleotide (NADH). The amount of NADH oxidised is proportional to the amount of sulfite.

Test solution Dissolve 4.0 g of the substance to be examined in freshly prepared distilled water R and dilute to 10.0 mL with the same solvent.

Reference solution Dissolve 4.0 g of the substance to be examined in freshly prepared distilled water R, add 0.5 mL of sulfite standard solution (80 ppm SO₂) R and dilute to 10.0 mL with freshly prepared distilled water R.

Blank solution Freshly prepared distilled water R.

Separately introduce 2.0 mL each of the test solution, the reference solution and the blank in 10 mm cuvettes and add the reagents as described in the instructions in the kit for sulfite determination. Measure the absorbance (2.2.25) at the absorption maximum at about 340 nm before and at the end of the reaction time and subtract the value obtained with the blank.

The absorbance difference of the test solution is not greater than half the absorbance difference of the reference solution.

Loss on drying (2.2.32)

Maximum 0.1 per cent, determined on 2.000 g by drying in an oven at 105 °C for 3 h.

Bacterial endotoxins (2.6.14)

Less than 0.25 IU/mg, if intended for use in the manufacture of large-volume parenteral preparations.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of large-volume parenteral preparations.

¹ This monograph has undergone pharmacopoeial harmonisation. See chapter 5.8 Pharmacopoeial Harmonisation.



Sucrose Monopalmitate

(Ph. Eur. monograph 2319)

DEFINITION

Mixture of sucrose monoesters, mainly sucrose monopalmitate, obtained by transesterification of palmitic acid methyl esters of vegetable origin with Sucrose (0204).

The manufacture of the fatty acid methyl esters includes a distillation step.

It contains variable quantities of mono-, di-, tri- and polyesters.

Content

— **monoesters**: minimum 55.0 per cent;

— **diesters**: maximum 40.0 per cent;

— **sum of triesters and polyesters**: maximum 20.0 per cent.

CHARACTERS

Appearance

White or almost white, unctuous powder.

Solubility

Very slightly soluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Composition of fatty acids (see Tests).

B. It complies with the limits of the assay.

TESTS

Acid value (2.5.1)

Maximum 6.0, determined on 3.00 g.

Use a freshly neutralised mixture of 1 volume of water R and 2 volumes of 2-propanol R as solvent and heat gently.

Composition of fatty acids (2.4.22, Method C)

Use the mixture of calibrating substances in Table 2.4.22-1.

Composition of the fatty-acid fraction of the substance:

— **lauric acid**: maximum 3.0 per cent;

— **myristic acid**: maximum 3.0 per cent;

— **palmitic acid**: 70.0 per cent to 85.0 per cent;

— **stearic acid**: 10.0 per cent to 25.0 per cent;

— **sum of the contents of palmitic acid and stearic acid**: minimum 90.0 per cent.

Free sucrose

Liquid chromatography (2.2.29).

Solvent mixture water for chromatography R, tetrahydrofuran for chromatography R (12.5:87.5 V/V).

Test solution Dissolve 0.200 g of the substance to be examined in the solvent mixture and dilute to 4.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of sucrose CRS in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) In 4 volumetric flasks, introduce respectively 5.0 mg, 10.0 mg, 20.0 mg and 25.0 mg of sucrose CRS, dissolve in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Column:
— **size**: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— **stationary phase**: spherical aminopropylsilyl silica gel for chromatography R (4 μ m).

Mobile phase:

— **mobile phase A**: 0.01 g/L solution of ammonium acetate R in acetonitrile for chromatography R;

— **mobile phase B**: 0.01 g/L solution of ammonium acetate R in a mixture of 10 volumes of water for chromatography R and 90 volumes of tetrahydrofuran for chromatography R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Flow rate (mL/min)
------------	-------------------------------	-------------------------------	--------------------

0 - 1 100 0 1.0

1 - 9 100 \rightarrow 0 0 \rightarrow 100 1.0

9 - 16 0 0 100 1.0

16 - 16.01 0 0 100 1.0 \rightarrow 2.5

16.01 - 32 0 100 2.5

32 - 33 0 \rightarrow 100 100 \rightarrow 0 2.5

33 - 36 100 0 2.5 \rightarrow 1.0

Detection Evaporative light-scattering detector; the following settings have been found to be suitable; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criterion:

— **carrier gas**: nitrogen R;

— **flow rate**: 1.0 mL/min;

— **evaporator temperature**: 45 °C;

— **nebuliser temperature**: 40 °C.

Injection 20 μ L.

Retention time About 26 min.

System suitability: reference solution (a):

— **signal-to-noise ratio**: minimum 10.



Sucrose Stearate

(Ph. Eur. monograph 2318)

Ph Eur

DEFINITION

Mixture of sucrose esters, mainly sucrose stearate, obtained by transesterification of stearic acid methyl esters of vegetable origin with sucrose (0204). The manufacture of the fatty acid methyl esters includes a distillation step.

It contains variable quantities of mono-, di-, tri- and polyesters.

Content

Sucrose stearate type I:
— monoesters: minimum 50.0 per cent;
— diesters: maximum 40.0 per cent;
— sum of triesters and polyesters: maximum 25.0 per cent;

Sucrose stearate type II:

— monoesters: 20.0 per cent to 45.0 per cent;
— diesters: 30.0 per cent to 40.0 per cent;
— sum of triesters and polyesters: maximum 30.0 per cent;

Sucrose stearate type III:

— monoesters: 15.0 per cent to 25.0 per cent;
— diesters: 30.0 per cent to 45.0 per cent;
— sum of triesters and polyesters: 35.0 per cent to 50.0 per cent.

CHARACTERS

Appearance

White or almost white, unctuous powder.

Solubility

Very slightly soluble in water, sparingly soluble in ethanol (96 per cent).

IDENTIFICATION

A. Composition of fatty acids (see Tests).

B. It complies with the limits of the assay.

TESTS

Acid value (2.5.1)

Maximum 6.0, determined on 3.00 g.

Use a freshly neutralised mixture of 1 volume of water R and 2 volumes of 2-propanol R as solvent and heat gently.

Composition of fatty acids (2.4.22, Method C)

Use the mixture of calibrating substances in Table 2.4.22-1. Composition of the fatty-acid fraction of the substance:

— lauric acid: maximum 3.0 per cent;
— myristic acid: maximum 3.0 per cent;
— palmitic acid: 25.0 per cent to 40.0 per cent;
— stearic acid: 55.0 per cent to 75.0 per cent;
— sum of the contents of palmitic acid and stearic acid: minimum 90.0 per cent.

Free sucrose

Liquid chromatography (2.2.29).

Solvent mixture water for chromatography R, tetrahydrofuran for chromatography R (12.5:87.5 V/V).

Test solution Dissolve 0.200 g of the substance to be examined in the solvent mixture and dilute to 4.0 mL with the solvent mixture.

Reference solution (a) Dissolve 20.0 mg of sucrose CRS in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (b) In 4 volumetric flasks, introduce

respectively 5.0 mg, 10.0 mg, 20.0 mg and 25.0 mg of

normalisation procedure.
Size-exclusion chromatography (2.2.30): use the

ASSAY

Maximum 1.5 per cent.

Total ash (2.4.16)

Maximum 4.0 per cent, determined on 0.20 g.

Water (2.5.12)

Limit Maximum 4.0 per cent.

— size: $l = 0.6$ m, $\phi = 7$ mm;
— stationary phase: styrene-divinylbenzene copolymer R (5 μ m)
with a pore size of 10 nm.
Mobile phase: tetrahydrofuran R.

Flow rate 1.2 mL/min.

Detection Differential refractometer.

Injection 20 μ L.

Relative retention With reference to monoesters (retention time = about 10 min): diesters = about 0.92; triesters and polyesters = about 0.90.

Calculations:

— disregard limit: disregard the peaks having a signal-to-noise ratio less than 10;
— free fatty acids (D): calculate the percentage content of free fatty acids, using the following expression:

$$I_A \times 256 / 561.1$$

= acid values;

— monoesters: calculate the percentage content of monoesters using the following expression:

$$A \times (100 - D - S - E) / 100$$

— diesters: calculate the percentage content of diesters using the following expression:

$$B \times (100 - D - S - E) / 100$$

— sum of triesters and polyesters: calculate the sum of the percentage contents of triesters and polyesters using the following expression:

$$C \times (100 - D - S - E) / 100$$

A = percentage content of monoesters determined by the

normalisation procedure;

S = percentage content of free sucrose (see Tests);

E = percentage content of water (see Tests);

B = percentage content of diesters determined by the

normalisation procedure;

C = sum of the percentage contents of triesters and

polyesters determined by the normalisation

procedure.

STORAGE

Protected from humidity.

Ph Eur

sucrose CRS, dissolve in the solvent mixture and dilute to

10.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: spherical aminopropylsilyl silica gel for chromatography R (4 μ m).

Mobile phase:

— mobile phase A: 0.01 g/L solution of ammonium acetate R in acetonitrile for chromatography R;
— mobile phase B: 0.01 g/L solution of ammonium acetate R in a mixture of 10 volumes of water for chromatography R and 90 volumes of tetrahydrofuran for chromatography R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Flow rate (mL/min)
0 - 1	100	0	1.0
1 - 9	100 \rightarrow 0	0 \rightarrow 100	1.0
9 - 16	0	100	1.0
16 - 16.01	0	100	1.0 \rightarrow 2.5
16.01 - 32	0	100	2.5
32 - 33	0 \rightarrow 100	100 \rightarrow 0	2.5
33 - 36	100	0	2.5 \rightarrow 1.0

Detection Evaporative light-scattering detector; the following settings have been found to be suitable; if the detector has different setting parameters, adjust the detector settings so as to comply with the system suitability criterion:

— carrier gas: nitrogen R;
— flow rate: 1.0 mL/min;
— evaporator temperature: 45 °C;
— nebuliser temperature: 40 °C.

Injection 20 μ L.

Retention time About 26 min.

System suitability: reference solution (a):

— signal-to-noise ratio: minimum 10.

Limit:

— sucrose: maximum 4.0 per cent.

Water (2.5.12)

Maximum 4.0 per cent, determined on 0.20 g.

Total ash (2.4.16)

Maximum 1.5 per cent.

ASSAY

Size-exclusion chromatography (2.2.30): use the

normalisation procedure.

Test solution Dissolve 60.0 mg of the substance to be

examined in tetrahydrofuran R and dilute to 4.0 mL with the

same solvent.

Column:

— size: $l = 0.6$ m, $\varnothing = 7$ mm;

— stationary phase: styrene-divinylbenzene copolymer R (5 μ m)

with a pore size of 10 nm.

Mobile phase tetrahydrofuran R.

Flow rate 1.2 mL/min.

Detection Differential refractometer.

Injection 20 μ L.

Relative retention With reference to monoesters (retention

time = about 10 min): diesters = about 0.92; triesters and

polyesters = about 0.90.

Calculations: — disregard limit: disregard the peaks having a signal-to-noise ratio less than 10;

— free fatty acids (D): calculate the percentage content of

$$I_A \times 284.5$$

$$561.1$$

= acid value;

— monoesters: calculate the percentage content of monoesters using the following expression:

$$A \times (100 - D - S - E)$$

$$100$$

— diesters: calculate the percentage content of diesters using the following expression:

$$B \times (100 - D - S - E)$$

$$100$$

— sum of triesters and polyesters: calculate the sum of the percentage contents of triesters and polyesters using the following expression:

$$C \times (100 - D - S - E)$$

$$100$$

A = percentage content of monoesters determined by the

normalisation procedure;

S = percentage content of free sucrose (see Tests);

E = percentage content of water (see Tests);

B = percentage content of diesters determined by the

normalisation procedure;

C = sum of the percentage contents of triesters and

polyesters determined by the normalisation

LABELLING

The label states the type of sucrose stearate (type I, II or III).

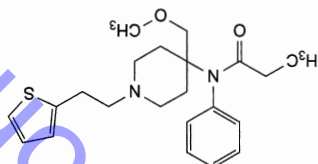
STORAGE

Protected from humidity.

Ph Eur

Sufentanil

(Ph. Eur. monograph 1569)



$C_{22}H_{30}N_2O_2S$

386.6

56030-54-7

Action and use

Opioid receptor agonist; analgesic.

Ph Eur

DEFINITION

N-[4-(Methoxymethyl)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]-*N*-phenylpropionamide.

Content

99.0 per cent to 101.0 per cent (dried substance).



CHARACTERS**Appearance**

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in ethanol (96 per cent) and in methanol.

mp

About 98 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of sufentanil.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.10 g in methanol R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) In order to produce impurity E *in situ*, dissolve 10 mg of the substance to be examined in 10.0 mL of dilute hydrochloric acid R. Heat on a water-bath under a reflux condenser for 4 h. Add 10.0 mL of dilute sodium hydroxide solution R. Evaporate to dryness on a water-bath. Cool and take up the residue in 10 mL of methanol R. Filter.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 5.0 mL of this solution to 20.0 mL with methanol R.

Column:

— size: $l = 0.1$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(3 μ m).

Mobile phase:

— mobile phase A: 5 g/L solution of ammonium carbonate R in a mixture of 10 volumes of tetrahydrofuran R and

90 volumes of water R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	90 → 40	10 → 60
15 - 20	40	60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L.

Relative retention With reference to sufentanil (retention time = about 13 min): impurity D = about 0.85;

impurity E = about 0.9; impurity F = about 0.95;

impurity H = about 1.1.

System suitability: reference solution (a):

— resolution: minimum 4.0 between the peaks due to

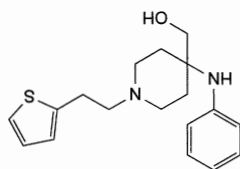
impurity E and sufentanil.

Limits:

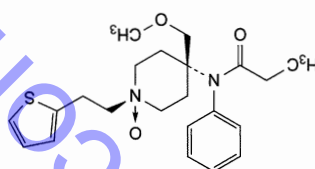
— impurities D, F, H: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (b) (0.25 per cent);

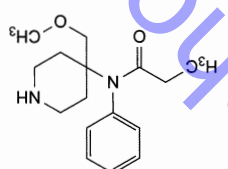
C. [4-(phenylamino)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]methanol,



B. *cis*-4-(methoxymethyl)-4-(phenylpropanoylamino)-1-[2-(thiophen-2-yl)ethyl]piperidine 1-oxide,



A. *N*-[4-(methoxymethyl)piperidin-4-yl]-*N*-phenylpropanamide,



C, E, G, I.

Control of impurities in substances for pharmaceutical use): A, B,

impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph Substances for pharmaceutical use

acceptance criterion for other/unspecified impurities and/or

the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities D, F, H

IMPURITIES

Protected from light.

STORAGE

of $C_{22}H_{30}N_2O_2S$.

1 mL of 0.1 M perchloric acid is equivalent to 38.66 mg

naphtholbenzenesulfonate solution R as indicator.

and titrate with 0.1 M perchloric acid, using 0.2 mL of

anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R

Dissolve 0.300 g in 50 mL of a mixture of 1 volume of

ASSAY

in vacuo at 60 °C for 2 h.

Maximum 0.5 per cent, determined on 1.000 g by drying

Loss on drying (2.2.32)

(0.05 per cent).

the chromatogram obtained with reference solution (b)

— disregard limit: 0.2 times the area of the principal peak in

(0.5 per cent);

the chromatogram obtained with reference solution (b)

— total: not more than twice the area of the principal peak in

(0.10 per cent);

chromatogram obtained with reference solution (b)

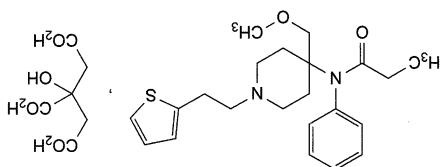
0.4 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than



Sufentanil Citrate

(Ph. Eur. monograph 1269)



$C_{28}H_{38}N_2O_9$ 578.7 60561-17-3

Action and use

Opioid receptor agonist; analgesic.

DEFINITION

N-[4-(Methoxymethyl)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]-N-phenylpropanamide citrate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Soluble in water, freely soluble in methanol, soluble in ethanol (96 per cent).

mp

About 140 °C, with decomposition.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of sufentanil citrate.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless.

(2.2.2, Method II).

Dissolve 0.2 g in water R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) In order to produce impurity E *in situ*, dissolve 10 mg of the substance to be examined in 10.0 mL of dilute hydrochloric acid R. Heat on a water-bath under a reflux condenser for 4 h. Add 10.0 mL of dilute sodium hydroxide solution R. Evaporate to dryness on a water-bath. Cool and take up the residue in 10 mL of methanol R. Filter.

Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Column:

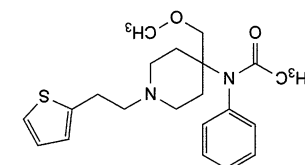
— size: $l = 0.1$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

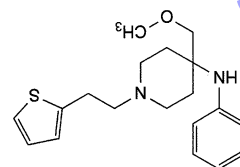
Mobile phase:

— mobile phase A: 5 g/L solution of ammonium carbonate R in a mixture of 10 volumes of tetrahydrofuran R and 90 volumes of water R;

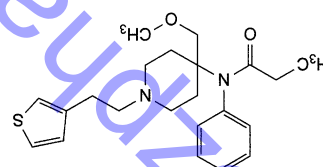
— mobile phase B: acetonitrile R;



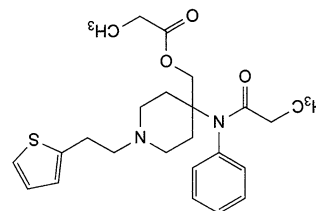
D. N-[4-(methoxymethyl)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]-N-phenylacetamide,



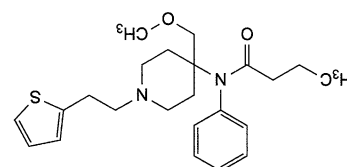
E. 4-(methoxymethyl)-N-phenyl-1-[2-(thiophen-2-yl)ethyl]piperidin-4-amine,



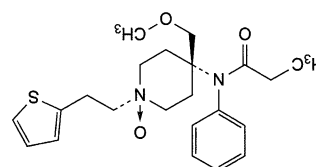
F. N-[4-(methoxymethyl)-1-[2-(thiophen-3-yl)ethyl]piperidin-4-yl]-N-phenylpropanamide,



G. [4-(phenylpropionylamino)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]methyl propanoate,



H. N-[4-(methoxymethyl)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]-N-phenylbutanamide,



I. *trans*-4-(methoxymethyl)-4-(phenylpropionylamino)-1-[2-(thiophen-2-yl)ethyl]piperidine 1-oxide.

Ph Eur

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 15	90 ÷ 40	10 ÷ 60
15 - 20	40	60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 µL.

Relative retention With reference to sufentanil (retention

time = about 13 min): impurity A = about 0.3;

impurity B = about 0.4; impurity I = about 0.45;

impurity C = about 0.7; impurity D = about 0.85;

impurity E = about 0.9; impurity F = about 0.95;

impurity G = about 1.05; impurity H = about 1.1.

System suitability: reference solution (a):

— resolution: minimum 4.0 between the peaks due to

impurity E and sufentanil.

Limits:

— impurities A, B, C, D, E, F, G, H, I: for each impurity,

not more than the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.5 per cent);

— unspecified impurities: for each impurity, not more than

0.2 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(1 per cent);

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent); disregard any peak with a retention time

relative to sufentanil of 0.05 or less.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 60 °C.

ASSAY

Dissolve 0.400 g in 50 mL of a mixture of 1 volume of

anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R

and titrate with 0.1 M perchloric acid, using 0.2 mL of

naphtholbenzene solution R as indicator.

1 mL of 0.1 M perchloric acid is equivalent to 57.87 mg

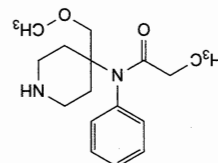
of C₂₈H₃₈N₂O₉S.

STORAGE

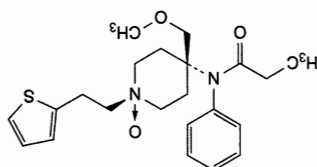
Protected from light.

IMPURITIES

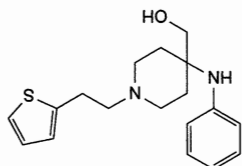
Specified impurities A, B, C, D, E, F, G, H, I



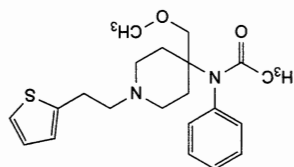
A. N-[4-(methoxymethyl)piperidin-4-yl]-N-phenylpropionamide,



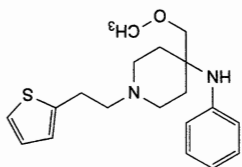
B. *cis*-4-(methoxymethyl)-4-(phenylpropionamido)-1-[2-(thiophen-2-yl)ethyl]piperidine-1-oxide,



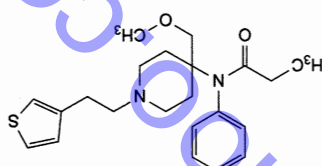
C. 4-(phenylamino)-1-[2-(thiophen-2-yl)ethyl]piperidine-4-ylmethanol,



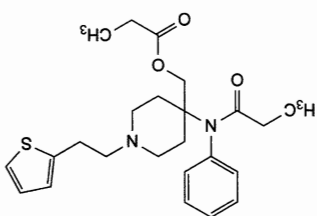
D. N-[4-(methoxymethyl)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]-N-phenylacetamide,



E. 4-(methoxymethyl)-N-phenyl-1-[2-(thiophen-2-yl)ethyl]piperidin-4-amine,



F. N-[4-(methoxymethyl)-1-[2-(thiophen-3-yl)ethyl]piperidin-4-yl]-N-phenylpropionamide,



G. 4-(phenylpropionamido)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-ylmethyl propionate,

solution. Dissolve 50 mg in 0.5 mL of lead-free nitric acid in a polyfluorocarbon-lined digestion bomb and heat at 150° for 5 hours. Allow to cool and dilute to 5 mL with water. Measure the absorbance at 283.3 nm, maintaining the drying temperature of the furnace at 110°, the ashing temperature at 600° and the atomising temperature at 2100°.

Chloride

2 mL of solution S diluted to 15 mL with water complies with the limit test for chlorides, Appendix VII (125 ppm).

Sulfate

7.5 mL of solution S diluted to 15 mL with water complies with the limit test for sulfates, Appendix VII (100 ppm).

Loss on drying

When dried at 105° for 4 hours, loses 0.25 to 1.0% of its weight. Use 1 g.

ASSAY

To 26 g of the substance being examined, previously dried, add 0.3 mL of a saturated aqueous solution of lead(II) acetate and 90 mL of water, shake, dilute to 100 mL with water and mix. Distribute evenly on the surface of a sheet of medium-fast filter paper 8 g of chromatographic siliceous earth and filter the solution with the aid of vacuum, discarding the first 20 mL of the filtrate. Transfer 25 mL of the filtrate into each of two 50 mL flasks. Slowly add 6 mL of 2M hydrochloric acid to one flask while rotating it, add 10 mL of water, mix, place the flask in a water bath at 60°, continuously shake for 3 minutes and allow the flask to stand in the water bath for a further 7 minutes. Immediately cool to 20° and dilute the solution with water to 50 mL and mix. Cool the contents of the second flask to 20°, dilute with water to volume and mix. Maintain both flasks at 20° for 30 minutes. Determine the specific optical rotation, Appendix V F, of each solution at 20°. Calculate the percentage of C₁₂H₂₂O₁₁ using the expression $100(a_0 - a_1)/88.3$ where a_0 and a_1 are the specific optical rotations of the uninverted and acid-inverted solutions respectively.

Sugar Spheres

(Ph. Eur. monograph 1570)

Action and use

Excipient.

Ph Eur

DEFINITION

Sugar spheres contain not more than 92 per cent of sucrose, calculated on the dried basis. The remainder consists of maize starch and may also contain starch hydrolysates and colour additives. The diameter of sugar spheres varies usually from 200 µm to 2000 µm and the upper and lower limits of the size of the sugar spheres are stated on the label.

IDENTIFICATION

A. Examine by thin-layer chromatography (2.2.27), using a TLC silica gel G plate R.
Solvent mixture water R, methanol R (2:3 V/V).
Test solution Mix 2 mL of solution S (see Tests) with 3 mL of methanol R and dilute to 20 mL with the solvent mixture.
Reference solution (a) Dissolve 10 mg of sucrose CRS in the solvent mixture and dilute to 20 mL with the solvent mixture.

Compressible Sugar

Action and use

Excipient.

DEFINITION

Compressible Sugar contains Sucrose and maltodextrin or dried glucose syrup. It contains 95.0 to 98.0% of Sucrose and 2 to 5% of dried glucose syrup or maltodextrin. It may contain a suitable lubricant, invert sugar or suitable colouring matter.

CHARACTERISTICS

Dry free-flowing powder or microcrystalline agglomerates. Very soluble in water.
Dissolve 20 g in 80 mL of water, dilute to 100 mL with water and filter (solution S).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of compressible sugar (RS 401).
B. The specific optical rotation of the uninverted solution obtained in the Assay is not less than 62.6 and the acid-inverted solution is laevorotatory.

TESTS

Conductivity

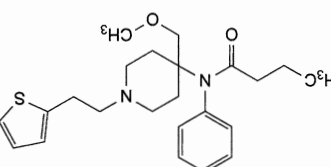
Not more than 35 µS·cm⁻¹, Appendix V O. Dissolve 31.3 g in carbon dioxide-free water prepared from distilled water and dilute to 100 mL with the same solvent. Measure the conductivity of the solution (C₁), while gently stirring with a magnetic stirrer, and that of the water used for preparing the solution (C₂). The readings must be stable within 1% over a period of 30 seconds. Calculate the conductivity of the solution of the substance being examined from the expression $C_1 - 0.35C_2$.

Calcium

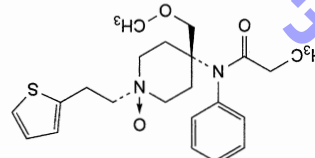
Add 1 mL of ammonium oxalate to 5 mL of solution S. The solution remains clear for not less than 1 minute.

Lead

Not more than 0.5 ppm of Pb when determined by atomic absorption spectrometry, Appendix II D, Method II, using an apparatus equipped with a graphite furnace and the following



H. N-[4-(methoxymethyl)-1-[2-(thiophen-2-yl)ethyl]piperidin-4-yl]-N-phenylbutanamide,



I. trans-4-(methoxymethyl)-4-(phenylpropylamino)-1-[2-(thiophen-2-yl)ethyl]piperidine 1-oxide.

Ph Eur



Reference solution (b) Dissolve 10 mg of fructose CRS, 10 mg of glucose CRS, 10 mg of lactose CRS and 10 mg of sucrose CRS in the solvent mixture and dilute to 20 mL with the solvent mixture.

Apply to the plate 2 µL of each solution and thoroughly dry the points of application. Develop over a path of 15 cm using a mixture of 10 volumes of water R, 15 volumes of methanol R, 25 volumes of anhydrous acetic acid R and 50 volumes of ethylene chloride R, measured accurately as a slight excess of water causes cloudiness of the solution. Dry the plate in a current of warm air. Repeat the development immediately after renewing the mobile phase. Dry the plate in a current of warm air and spray evenly with a 5 g/L solution of rhymol R in a mixture of 5 volumes of sulfuric acid R and 95 volumes of ethanol (96 per cent) R. Heat at 130 °C for 10 min. The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a). The test solution (b) shows 4 clearly separated spots.

B. To a water slurry of the insoluble portion obtained in the assay, add 0.05 mL of iodine solution R1. A dark-blue colour is produced, which disappears on heating.

C. To 5 mL of solution S add 0.15 mL of freshly prepared copper sulfate solution R and 2 mL of freshly prepared dilute sodium hydroxide solution R. The solution is blue and clear and remains so after boiling. To the hot solution add 4 mL of dilute hydrochloric acid R and boil for 1 min. Add 4 mL of dilute sodium hydroxide solution R. An orange precipitate is formed immediately.

TESTS

Solution S
To 0.5 g in a 100 mL volumetric flask add 80 mL of water R and shake until the sucrose is dissolved. Dilute to 100.0 mL with water R. Filter under vacuum to obtain a clear solution.

Fineness (2.9.35)
Not less than 90 per cent *m/m* of the sugar spheres are between the lower and the upper limits of the size of the sugar spheres stated on the label.

Heavy metals (2.4.8)
Maximum 5 ppm.

2.0 g complies with test C. Prepare the reference solution using 1.0 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)
Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14)
Maximum 0.2 per cent, determined on 2 g.

Microbial contamination
TAMC: acceptance criterion 10³ CFU/g (2.6.12).
TYMC: acceptance criterion 10² CFU/g (2.6.12).
Absence of *Escherichia coli* (2.6.13).
Absence of *Salmonella* (2.6.13).

ASSAY

Sucrose content
Weigh 10.000 g of ground sugar spheres in a 100 mL flask and make up to 100.0 mL with water R. Stir and decant. Filter under vacuum to obtain a clear solution (the insoluble portion is used for identification test B). Measure the angle of optical rotation (2.2.7) and calculate the sucrose percentage content using the following expression:

$$10^6 \times \frac{66.5 \times l \times m \times (100 - H)}{100 \times \alpha}$$

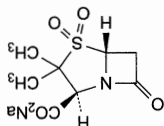
α = angle of rotation;
 l = length of the polarimeter tube, in decimetres;
 m = exact mass of the sample, in grams;
 H = loss on drying.

LABELLING
The label states the upper and the lower limits of the size of the sugar spheres.

Ph Eur

Sulbactam Sodium

(Ph. Eur. monograph 2209)



255.2

69388-84-7

Action and use
Beta-lactam antibacterial.

DEFINITION

Sodium (2S,5R)-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate 4,4-dioxide. Semi-synthetic product derived from a fermentation product.

Content
97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, hygroscopic, crystalline powder.

Solubility
Freely soluble in water, sparingly soluble in ethyl acetate, very slightly soluble in ethanol (96 per cent). It is freely soluble in dilute acids.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison substance CRS.

B. It gives reaction (a) of sodium (2.3.1).

TESTS

Appearance of solution
The solution is clear (2.2.1).
Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent.

Absorbance (2.2.25)
Maximum 0.10 at 430 nm.

Dissolve 1.0 g in water R and dilute to 100.0 mL with the same solvent.

pH (2.2.3)
4.5 to 7.2; if the substance is sterile: 5.2 to 7.2.

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Specific optical rotation (2.2.7)
+ 219 to + 233 (anhydrous substance).

Dissolve 0.500 g in water R and dilute to 50.0 mL with the

same solvent.

Related substances

Liquid chromatography (2.2.29).

Solution A 2.72 g/L solution of potassium dihydrogen phosphate R adjusted to pH 4.0 with dilute phosphoric acid R.

Solution B Dilute 2 mL of acetonitrile R1 to 100.0 mL with

solution A.

Test solution Suspend 77.0 mg of the substance to be

examined in 2 mL of acetonitrile R1, sonicate for about 5 min

and dilute to 100.0 mL with solution A.

Reference solution (a) Suspend 70.0 mg of sulbactam CRS in

2 mL of acetonitrile R1, sonicate for about 5 min and dilute

to 100.0 mL with solution A.

Reference solution (b) Dilute 1.0 mL of reference solution (a)

to 100.0 mL with solution B. Dilute 1.0 mL of this solution

to 10.0 mL with solution B.

Reference solution (c) Dissolve 15.0 mg of 6-aminopenicillanic

acid R (impurity B) in solution A and dilute to 50.0 mL with

1 mL of reference solution (c) and dilute to 25.0 mL with

solution B.

Reference solution (e) Dissolve 8 mg of sulbactam for peak

identification CRS (containing impurities A, C, D, E and F)

in 1 mL of acetonitrile R1, sonicate for about 5 min and

dilute to 10 mL with solution B.

Column:

— size: $l = 0.10$ m, $\phi = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (3.0 μ m);

— temperature: 40 °C.

Mobile phase:

— **mobile phase A:** 5.44 g/L solution of potassium dihydrogen

phosphate R adjusted to pH 4.0 with dilute phosphoric

acid R;

— **mobile phase B:** mobile phase A, acetonitrile R1

(40:60 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2.0	98	2
2.0 - 9.5	98 \rightarrow 50	2 \rightarrow 50
9.5 - 12.0	50	50

Flow rate: 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 μ L of the test solution, solution B and reference

solutions (b), (d) and (e).

Identification of impurities Use the chromatogram supplied

with sulbactam for peak identification CRS and the

chromatogram obtained with reference solution (e) to identify

the peaks due to impurities A, C, D, E and F; use the

chromatogram obtained with reference solution (d) to

identify the peak due to impurity B.

Relative retention With reference to sulbactam (retention

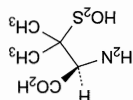
time = about 3 min): impurity A = about 0.4;

impurity B = about 0.7; impurity C = about 2.3;

impurity D = about 3.1; impurity E = about 3.3;

impurity F = about 3.9.

A. (2S)-2-amino-3-methyl-3-sulfinobutanoic acid,



Control of impurities in substances for pharmaceutical use: G. impurities for demonstration of compliance. See also 5.10. (2034). It is therefore not necessary to identify these

by the general monograph *Substances for pharmaceutical use* acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities A, B, C, D, E, F

IMPURITIES

In an airtight container. If the substance is sterile, store in a

sterile, the container is also sterile and tamper-proof.

STORAGE

conversion factor of 1.094.

into account the assigned content of sulbactam CRS and a

Calculate the percentage content of $C_8H_{10}NNaO_5$ taking

Injection Test solution and reference solution (a).

related substances with the following modification.

Liquid chromatography (2.2.29) as described in the test for

ASSAY

procedure for the removal of bacterial endotoxins.

of parenteral preparations without a further appropriate

Less than 0.17 IU/mg, if intended for use in the manufacture

Bacterial endotoxins (2.6.14, Method A)

Maximum 1.0 per cent, determined on 1.00 g.

Water (2.5.12)

(1 ppm Pb) R.

the reference solution using lead standard solution

solvent. 12 mL of the solution complies with test A. Prepare

Dissolve 1.0 g in water R and dilute to 20 mL with the same

Maximum 20 ppm.

Heavy metals (2.4.8)

Maximum 0.5 per cent m/m.

2-Ethylhexanoic acid (2.4.28)

— reporting threshold: 0.05 per cent.

— total: maximum 1.0 per cent;

0.10 per cent;

— unspecified impurities: for each impurity, maximum

0.1 per cent;

— impurities B, D, F: for each impurity, maximum

— impurities C, E: for each impurity, maximum 0.2 per cent;

— impurity A: maximum 0.5 per cent;

Limits:

— for each impurity, use the concentration of sulbactam in

reference solution (b).

impurity F = 0.6;

impurity A = 0.6; impurity B = 0.5; impurity D = 0.5;

impurities by the corresponding correction factor:

— correction factors: multiply the peak areas of the following

Calculation of percentage contents:

impurity B and sulbactam.

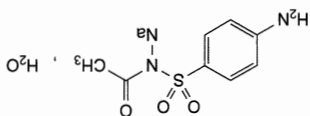
— resolution: minimum 5.0 between the peaks due to

System suitability Reference solution (d):

Sulfacetamide Sodium

Soluble Sulfacetamide

(Ph. Eur. monograph 0107)



$C_8H_9N_2NaO_3S \cdot H_2O$ 254.2

Action and use

Sulfonamide antibacterial.

DEFINITION

Sodium acetyl[(4-aminophenyl)sulfonyl]azanide.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in anhydrous ethanol.

IDENTIFICATION

First identification B, F

Second identification A, C, E, F.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 0.1 g in phosphate buffer solution

pH 7.0 R and dilute to 100.0 mL with the same buffer

phosphate buffer solution pH 7.0 R.

Spectral range 230-350 nm.

Absorption maximum At 255 nm.

Specific absorbance at the absorption maximum 660 to 720

(anhydrous substance).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison sulfacetamide sodium CRS.

C. Melting point (2.2.14): 181 °C to 185 °C.

Dissolve 1 g in 10 mL of water R, add 6 mL of dilute acetic

acid R and filter. Wash the precipitate with a small quantity

of water R and dry at 100-105 °C for 4 h.

E. Dissolve about 1 mg of the precipitate obtained in

identification C, with heating, in 1 mL of water R.

The solution gives the reaction of primary aromatic amines

(2.3.1) with formation of an orange-red precipitate.

F. Solution S (see Tests) gives the reactions of sodium

TESTS

Solution S

Dissolve 1.25 g in carbon dioxide-free water R and dilute to

25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

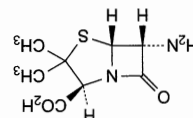
than reference solution GY₄ (2.2.2, Method II).

pH (2.2.3)

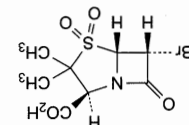
8.0 to 9.5 for solution S.

Related substances

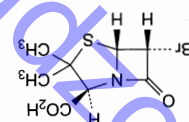
Liquid chromatography (2.2.29). Prepare the solutions immediately before use and carry out the test protected from light.



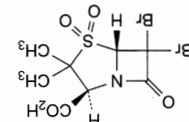
B. (2S,5R,6R)-6-amino-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-aminopenicillanic acid),



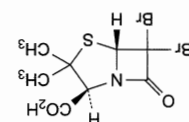
C. (2S,5R,6R)-6-bromo-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid 4,4-dioxide (6-bromopenicillanic acid sulfone),



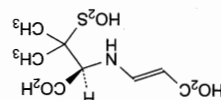
D. (2S,5R,6R)-6-bromo-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6-bromopenicillanic acid),



E. (2S,5R,6R)-6,6-dibromo-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid 4,4-dioxide (6,6-dibromopenicillanic acid sulfone),



F. (2S,5R,6R)-6,6-dibromo-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (6,6-dibromopenicillanic acid),



G. (2S)-2-[[[(2S)-2-carboxyethenyl]amino]-3-methyl-3-sulfimobutanoic acid.

Ph Eur



Test solution Dissolve 0.200 g of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of sulfacetamide sodium CRS and 5 mg of sulfanilamide R (impurity A) in 1.0 mL of the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.125$ m, $\varnothing = 4$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase glacial acetic acid R, methanol R, water for chromatography R (1:10:89 V/V/V).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 μ L.

Run time 7 times the retention time of sulfacetamide.

Relative retention With reference to sulfacetamide (retention time = about 5 min): impurity A = about 0.5.

System suitability: reference solution (a):

— resolution: minimum 5.0 between the peaks due to impurity A and sulfacetamide.

Limits:

— correction factor: for the calculation of the content, multiply the peak area of impurity A by 0.5;

— impurity A: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent).

Sulfates (2.4.13)

Maximum 200 ppm.

Dissolve 2.5 g in distilled water R and dilute to 25 mL with the same solvent. Add 25 mL of dilute acetic acid R, shake for 30 min and filter. 15 mL of the filtrate complies with the limit test for sulfates.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of the filtrate obtained in the test for sulfates complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

6.0 per cent to 8.0 per cent, determined on 0.200 g.

ASSAY

Dissolve 0.500 g in a mixture of 50 mL of water R and 20 mL of dilute hydrochloric acid R. Cool the solution in a bath of iced water and carry out the determination of primary aromatic amino-nitrogen (2.5.8), determining the end-point electrometrically.

1 mL of 0.1 M sodium nitrite is equivalent to 23.62 mg of $C_8H_9N_2NaO_3S$.

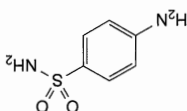
STORAGE

Protected from light.

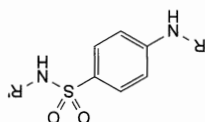
IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.

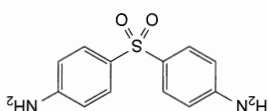


A. 4-amino-2-naphthylsulfonamide (sulfanilamide),



B, R = $CO-CH_3$, R' = H: N-(4-sulfamoylphenyl)acetamide,

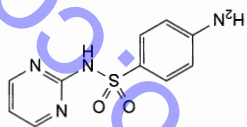
C, R = R' = $CO-CH_3$, N-[[4-(acetamido)phenyl]sulfonyl]acetamide,



D. 4,4'-sulfonyldianiline (dapsone).

Sulfadiazine

(Ph Eur monograph 0294)



$C_{10}H_{10}N_4O_2S$

250.3

68-35-9

Action and use

Sulfonamide antibacterial.

Preparation

Sulfadiazine Injection

DEFINITION

4-Amino-N-(pyrimidin-2-yl)benzenesulfonamide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White, yellowish-white or pinkish-white, crystalline powder or crystals.

Solubility

Practically insoluble in water, slightly soluble in acetone, very slightly soluble in ethanol (96 per cent). It dissolves in solutions of alkali hydroxides and in dilute mineral acids.

IDENTIFICATION

First identification A.

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison sulfadiazine CRS.

B. Thin-layer chromatography (2.2.27).

Solvent mixture concentrated ammonia R, methanol R

(4:96 V/V)

Test solution Dissolve 20 mg of the substance to be examined in 3 mL of the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution Dissolve 20 mg of sulfadiazine CRS in 3 mL of the solvent mixture and dilute to 5.0 mL with the solvent mixture.

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Reference solution Dissolve 20 mg of sulfadiazine CRS in 3 mL of the solvent mixture and dilute to 5.0 mL with the solvent mixture.

In the solvent mixture and dilute to 10.0 mL with water R.

Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase. Dilute 3.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of acetylsulfadiazine CRS (impurity E) in 1 mL of the mobile phase.

Reference solution (d) Dissolve 5 mg of sulfadiazine for identification of impurity F CRS in the solvent mixture and dilute to 10.0 mL with water R.

Column: size: $l = 0.25$ m, $\phi = 4.6$ mm; stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase acetonitrile R, 2.8 g/L solution of phosphoric acid R (10:90 V/V).

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 μ L.

Run time 7 times the retention time of sulfadiazine.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and B; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity E; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity F.

Relative retention With reference to sulfadiazine (retention time = about 8.5 min): impurity A = about 0.26; impurity B = about 0.30; impurity E = about 2.1; impurity F = about 6.0.

System suitability: reference solution (a): resolution: minimum 2.0 between the peaks due to impurities A and B.

Limits: peak area of impurity E by 0.7;

impurities A, B: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

impurity E: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

solution (b) (0.15 per cent);

reference solution (b) (0.15 per cent);

unspectified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent);

total: maximum 0.5 per cent;

disregard limit: 0.3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

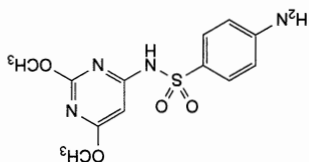
Solvent dimethyl sulfoxide R.

1.0 g complies with test H. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.



Sulfadimethoxine

(Ph. Eur. monograph 2741)



$C_{12}H_{14}N_4O_4S$ 310.3 122-11-2

Action and use

Sulfonamide antibacterial.

Ph Eur

DEFINITION

4-Amino-N-(2,6-dimethoxyphenyl)pyrimidin-4-yl)benzene-1-sulfonamide.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in methylene chloride, very slightly soluble in ethanol (96 per cent).

It is freely soluble in dilute sodium hydroxide and sparingly soluble in dilute hydrochloric acid.

mp: 197 °C to 202 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison sulfadimethoxine CRS.

TESTS

Acidity

Suspend 0.5 g in 25 mL of carbon dioxide-free water R. Heat the suspension at 70 °C for 5 min, cool quickly to room temperature and filter. Not more than 0.1 mL of 0.1 M sodium hydroxide is required to reach pH 7.0.

Related substances

Solution A Dissolve 6.0 g of sodium dihydrogen phosphate R in 950 mL of water R, adjust to pH 7.0 with dilute sodium hydroxide solution R and dilute to 1 L with water R.

Test solution Dissolve 20.0 mg of the substance to be examined in 25 mL of methanol R and dilute to 100.0 mL with solution A.

Reference solution (a) Dissolve 20.0 mg of sulfadimethoxine CRS in 25 mL of methanol R and dilute to 100.0 mL with solution A.

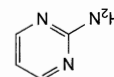
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (c) Dissolve 4 mg of sulfadimethoxine for peak identification CRS (containing impurities A and F) in 5 mL of methanol R and dilute to 20 mL with solution A.

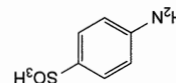
Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl amorphous organosilica polymer R (5 μ m);
— temperature: 25 °C.

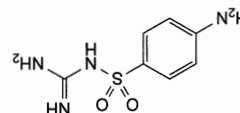
Specified impurities A, B, E, F
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C, D.



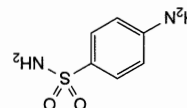
A. pyrimidin-2-amine,



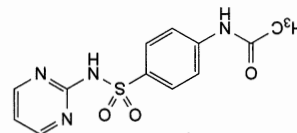
B. 4-aminoobenzenesulfonic acid (sulfanilic acid),



C. [(4-aminophenyl)sulfonyl]guanidine (sulfaguanidine),



D. 4-aminobenzenesulfonamide (sulfanilamide),



E. N-[4-(pyrimidin-2-yl)sulfonyl]phenyl]acetamide (acetylsulfadiazine),

F. unknown structure.

Ph Eur

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in a mixture of 20 mL of dilute hydrochloric acid R and 50 mL of water R. Cool the solution in iced water. Carry out the determination of primary aromatic amino-nitrogen (2.5.8), determining the end-point electrometrically.

1 mL of 0.1 M sodium nitrite is equivalent to 25.03 mg of $C_{12}H_{14}N_4O_4S$.

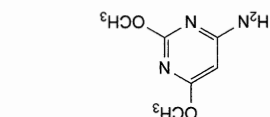
STORAGE

Protected from light.

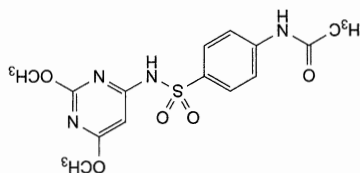
IMPURITIES

IMPURITIES

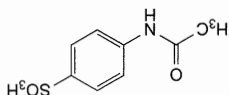
Specified impurities A, F
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E.



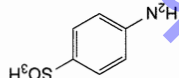
A. 2,6-dimethoxy-4-aminopyrimidin-4-amine,



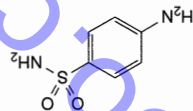
B. N-[4-[(2,6-dimethoxy-4-aminopyrimidin-4-



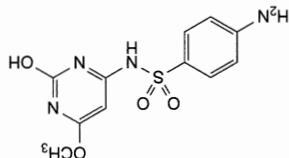
C. 4-(acetylamino)benzene-1-sulfonic acid,



D. 4-aminobenzenesulfonic acid (sulfanilic acid),



E. 4-aminobenzenesulfonamide (sulfanilamide),



F. 4-amino-N-(2-hydroxy-6-methoxy-4-yl)benzenesulfonamide.

Ph Eur

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 µL of the test solution and reference solutions (b)

and (c).

Identification of impurities Use the chromatogram supplied

with *sulfadimethoxine* for peak identification CRS and the

chromatogram obtained with reference solution (c) to identify

the peaks due to impurities A and F.

Relative retention With reference to *sulfadimethoxine*

(retention time = about 11 min): impurity F = about 0.4;

impurity A = about 1.2.

System suitability:

— resolution: minimum 2.5 between the peaks due to

sulfadimethoxine and impurity A in the chromatogram

obtained with reference solution (c);

— signal-to-noise ratio: minimum 40 for the principal peak in

the chromatogram obtained with reference solution (b).

Calculation of percentage contents:

— correction factors: multiply the peak areas of the following

impurities by the corresponding correction factor:

impurity A = 1.4; impurity F = 1.7;

— for each impurity, use the concentration of

sulfadimethoxine in reference solution (b).

Limits:

— impurities A, F: for each impurity,

maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum

0.10 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent mixture methanol R, water R, acetone R

(10:15:75 V/V/V).

0.5 g complies with test H. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{12}H_{14}N_4O_4S$ taking

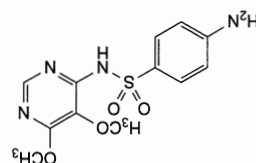
into account the assigned content of *sulfadimethoxine* CRS.

STORAGE

Protected from light.

Sulfadoxine

(Ph. Eur. monograph 0740)



$C_{12}H_{14}N_4O_4S$ 310.3 2447-57-6

Action and use
Sulfonamide antibacterial.

DEFINITION

Sulfadoxine contains not less than 99.0 per cent and not

more than the equivalent of 101.0 per cent of 4-amino-N-(5,6-dimethoxy-2-pyrimidin-4-yl)benzenesulfonamide, calculated with reference to the dried substance.

CHARACTERS

White or yellowish-white crystalline powder or crystals, very slightly soluble in water, slightly soluble in alcohol and in methanol. It dissolves in solutions of alkali hydroxides and in dilute mineral acids.

IDENTIFICATION

It melts at about 198 °C, with decomposition.

First identification A, C.
Second identification B, C, D.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with sulfadoxine CRS. Examine the substances prepared as discs.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.5 g in 1 mL of a 40 per cent V/V solution of

sulfuric acid R, heating gently. Continue heating until a crystalline precipitate appears (about 2 min). Allow to cool and add 10 mL of dilute sodium hydroxide solution R. Cool

again. Add 25 mL of ether R and shake for 5 min. Separate the ether layer, dry over anhydrous sodium sulfate R and filter. Evaporate the solvent by heating in a water-bath. The residue

melts (2.2.14) at 80 °C to 82 °C or at 90 °C to 92 °C. D. Dissolve about 5 mg in 10 mL of 1 M hydrochloric acid. Dilute 1 mL of the solution to 10 mL with water R. The solution, without further acidification, gives the reaction of primary aromatic amines (2.3.1).

TESTS

Appearance of solution

Dissolve 1.0 g in a mixture of 5 mL of dilute sodium hydroxide

solution R and 5 mL of water R. The solution is not more intensely coloured than reference solution Y₅, BY₅ or GY₅ (2.2.2, Method II).

Acidity

To 1.25 g, finely powdered, add 25 mL of carbon dioxide-free

water R. Heat at 70 °C for 5 min. Cool in a bath of iced water for about 15 min and filter. To 20 mL of the filtrate add 0.1 mL of bromothymol blue solution R1. Not more than 0.2 mL of 0.1 M sodium hydroxide is required to change the

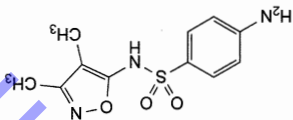
colour of the indicator.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Sulfafurazole

(Ph. Eur. monograph 0741)



$C_{11}H_{13}N_3O_3S$ 267.3 127-69-5

Action and use
Sulfonamide antibacterial.

DEFINITION

Sulfafurazole contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 4-amino-N-(3,4-dimethylisoxazol-5-yl)benzenesulfonamide, calculated with reference to the dried substance.

STORAGE

Store protected from light.

$C_{12}H_{14}N_4O_4S$.
1 mL of 0.1 M sodium nitrite is equivalent to 31.03 mg of

end-point electrometrically. Carry out the determination of primary aromatic amino-nitrogen (2.5.8), using 0.250 g and determining the

ASSAY

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Not more than 0.5 per cent, determined on 1.000 g by

loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by

loss on drying (2.2.32)

Prepare the reference solution using 2 mL of lead standard

solution (10 ppm Pb) R.

1.0 g complies with test D for heavy metals (20 ppm).

Heavy metals (2.4.8)

reference solution (b) (0.5 per cent).

Any spot in the chromatogram obtained with test

solution (a), apart from the principal spot, is not more

intense than the spot in the chromatogram obtained with

reference solution (b) (0.5 per cent).

Apply to the plate 5 µL of each solution. Develop over a

path of 15 cm using a mixture of 3 volumes of dilute

ammonia R1, 5 volumes of water R, 40 volumes of

nitromethane R and 50 volumes of dioxan R. Dry the plate at

100 °C to 105 °C and examine in ultraviolet light at 254 nm.

Test solution (a) Dissolve 0.10 g of the substance to be

examined in 3 mL of a mixture of 2 volumes of concentrated

ammonia R and 48 volumes of methanol R.

Test solution (b) Dilute 1 mL of test solution (a) to 5 mL

with the same mixture of solvents.

Test solution (c) Dilute 1 mL of test solution (a) to 5 mL

with a mixture of 2 volumes of concentrated ammonia R and

48 volumes of methanol R.

Reference solution (a) Dissolve 20 mg of sulfadoxine CRS in

3 mL of a mixture of 2 volumes of concentrated ammonia R

and 48 volumes of methanol R and dilute to 5 mL with the

same mixture of solvents.

Reference solution (b) Dilute 1 mL of test solution (a) to 5 mL

with the same mixture of solvents.

Test solution (c) Dissolve 0.10 g of the substance to be

examined in 3 mL of a mixture of 2 volumes of concentrated

ammonia R and 48 volumes of methanol R and dilute to 5 mL

CHARACTERS

White or yellowish-white, crystalline powder or crystals, practically insoluble in water, sparingly soluble in alcohol, slightly soluble in methylene chloride. It dissolves in solutions of alkali hydroxides and in dilute mineral acids.

IDENTIFICATION

First identification A, C.
Second identification B, C, D.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with *sulfafurazazole* CRS. Examine the substances prepared as discs. B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. To 0.5 g add 1 mL of a 40 per cent V/V solution of *sulfuric acid* R and heat over a low flame to dissolve. Continue heating until a crystalline precipitate appears (about 2 min). Allow to cool and add 10 mL of *dilute sodium hydroxide solution* R. Cool. Shake the solution for 5 min with 25 mL of *ether* R. Separate the ether layer, dry over *anhydrous sodium sulfate* R and filter. Evaporate the solvent by heating on a water-bath. The residue melts (2.2.14) at 119 °C to 123 °C. D. Dissolve about 5 mg in 10 mL of 1 M *hydrochloric acid*. Dilute 1 mL of the solution to 10 mL with *water* R. The solution, without further acidification, gives the reaction of primary aromatic amines (2.3.1).

TESTS

Appearance of solution

Dissolve 0.4 g in a mixture of 5 mL of *dilute sodium hydroxide solution* R and 5 mL of *water* R, with gently warming if necessary. The solution is not more intensely coloured than reference solution Y₆, BY₆ or GY₆ (2.2.2, Method II).

Acidity

To 1.25 g, finely powdered, add 25 mL of *carbon dioxide-free water* R. Heat at 70 °C for 5 min. Cool in iced water for about 15 min and filter. To 20 mL of the filtrate add 0.1 mL of *bromothymol blue solution* R1. Not more than 0.2 mL of 0.1 M *sodium hydroxide* is required to change the colour of the indicator.

Related substances

Examine by thin-layer chromatography (2.2.27), using *silica gel* GF₂₅₄ R as the coating substance. **Test solution (a)** Dissolve 0.10 g of the substance to be examined in 3 mL of a mixture of 2 volumes of *concentrated ammonia* R and 48 volumes of *methanol* R and dilute to 5 mL with the same mixture of solvents.

Test solution (b) Dilute 1 mL of test solution (a) to 5 mL with a mixture of 2 volumes of *concentrated ammonia* R and 48 volumes of *methanol* R.

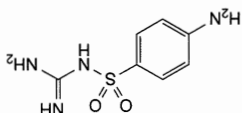
Reference solution (a) Dissolve 20 mg of *sulfafurazazole* CRS in 3 mL of a mixture of 2 volumes of *concentrated ammonia* R and 48 volumes of *methanol* R and dilute to 5 mL with the same mixture of solvents.

Reference solution (b) Dilute 1.25 mL of test solution (b) to 50 mL with a mixture of 2 volumes of *concentrated ammonia* R and 48 volumes of *methanol* R.

Apply to the plate 5 µL of each solution. Develop over a path of 15 cm using a mixture of 1 volume of *concentrated ammonia* R, 25 volumes of *methanol* R and 75 volumes of

Sulfaguanidine

(Ph. Eur. monograph 1476)



C₇H₁₀N₄O₂S

214.3

57-67-0

Action and use

Sulfonamide antibacterial.

DEFINITION

Sulfaguanidine contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of (4-aminophenylsulfonamido)guanidine, calculated with reference to the dried substance.

CHARACTERS

A white or almost white, fine crystalline powder, very slightly soluble in water, slightly soluble in acetone, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride. It dissolves in dilute solutions of mineral acids.

IDENTIFICATION

First identification A, B

Second identification A, C, D, E

A. Melting point (2.2.14): 189 °C to 193 °C, determined on the dried substance.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with *sulfaguanidine* CRS.

STORAGE

Store protected from light.

ASSAY

Dissolve 0.200 g in 50 mL of *acetone* R. Titrate with 0.1 M *tetrabutylammonium hydroxide* using a 4 g/L solution of *thymol blue* R in *methanol* R as indicator. 1 mL of 0.1 M *tetrabutylammonium hydroxide* is equivalent to 26.73 mg of C₇H₁₀N₄O₂S.

STORAGE

Store protected from light.

STORAGE

Store protected from light.

STORAGE

Store protected from light.

C. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 5 mg in 10 mL of 1 M hydrochloric acid. Dilute 1 mL of the solution to 10 mL with water R.

The solution, without further acidification, gives the reaction of primary aromatic amines (2.3.1).

E. Suspend 0.1 g in 2 mL of water R, add 1 mL of α -naphthol solution R and 2 mL of a mixture of equal volumes of water R and strong sodium hypochlorite solution R. A red colour develops.

TESTS

Solution S
To 2.5 g, add 40 mL of carbon dioxide-free water R. Heat at about 70 °C for 5 min. Cool while stirring in iced water for about 15 min, filter and dilute to 50 mL with carbon dioxide-free water R.

Acidity
To 20 mL of solution S, add 0.1 mL of bromothymol blue solution R1. Not more than 0.2 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Related substances

Examine by thin layer chromatography (2.2.27), using a TLC silica gel GF₂₅₄ plate R.

Test solution (a) Dissolve 50 mg of the substance to be examined in acetone R and dilute to 5 mL with the same solvent.

Test solution (b) Dilute 2 mL of test solution (a) to 10 mL with acetone R.

Reference solution (a) Dissolve 10 mg of sulfaguanidine CRS in acetone R.

Reference solution (b) Dilute 5 mL of test solution (b) to 200 mL with acetone R.

Reference solution (c) Dilute 5 mL of reference solution (b) to 10 mL with acetone R.

Reference solution (d) Dissolve 10 mg of sulfamizole R in test solution (b) and dilute to 5 mL with the same solution.

Apply to the plate 10 μ L of each solution. Develop over a path of 15 cm using a mixture of 10 volumes of anhydrous formic acid R, 20 volumes of methanol R and 70 volumes of methylene chloride R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent) and at most one such spot is more intense than the spot in the chromatogram obtained with reference solution (c) (0.25 per cent). The test is not valid unless the chromatogram obtained with reference solution (d) shows two clearly separated principal spots.

Heavy metals (2.4.8)

1.0 g complies with test F (20 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 8.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

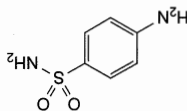
ASSAY
Dissolve 0.175 g in 50 mL of dilute hydrochloric acid R. Cool the solution in iced water. Carry out the determination of primary aromatic amino-nitrogen (2.5.8), determining the end-point electrometrically.

1 mL of 0.1 M sodium nitrite is equivalent to 21.42 mg of C₇H₁₀N₄O₂S.

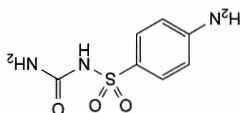
STORAGE

Store protected from light.

IMPURITIES



A. 4-aminobenzenesulfonamide (sulfanilamide),

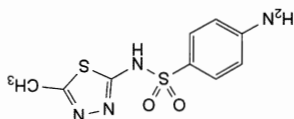


B. N-[(4-aminophenyl)sulfonyl]urea (sulfacarbamide).

Ph Eur

Sulfamethizole

(Ph. Eur. monograph 0637)



C₇H₁₀N₄O₂S

270.3

144-82-1

Action and use

Sulfonamide antibacterial.

Ph Eur

DEFINITION

Sulfamethizole contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 4-amino-N-(5-methyl-1,3,4-thiadiazol-2-yl)benzenesulfonamide, calculated with reference to the dried substance.

CHARACTERS

White or yellowish-white crystalline powder or crystals, very slightly soluble in water, soluble in acetone, sparingly soluble in alcohol. It dissolves in dilute solutions of alkali hydroxides and in dilute mineral acids.

It melts at about 210 °C.

IDENTIFICATION

First identification A, B.

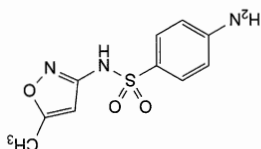
Second identification B, C, D.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with sulfamethizole CRS. Examine the substances prepared as discs.

B. Examine the chromatograms obtained in the test for related substances. The principal spot in the chromatogram

Sulfamethoxazole

(Ph. Eur. monograph 0108)



C₁₀H₁₁N₃O₃S 253.3 723-46-6

Action and use

Sulfonamide antibacterial.

Preparations

Co-trimoxazole Oral Suspension
Co-trimoxazole Tablets
Dispersible Co-trimoxazole Tablets
Paediatric Co-trimoxazole Tablets

Ph Eur

DEFINITION

4-Amino-N-(5-methylisoxazol-3-yl)benzenesulfonamide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone, sparingly soluble in ethanol (96 per cent). It dissolves in dilute solutions of sodium hydroxide and in dilute acids.

IDENTIFICATION

First identification A, B

Second identification A, C, D.

A. Melting point (2.2.14): 169 °C to 172 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Comparison sulfamethoxazole CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in 3 mL of a mixture of 2 volumes of concentrated ammonia R and 48 volumes of methanol R and dilute to 5 mL with the same mixture of solvents.

Reference solution Dissolve 20 mg of sulfamethoxazole CRS in 3 mL of a mixture of 2 volumes of concentrated ammonia R and 48 volumes of methanol R and dilute to 5 mL with the same mixture of solvents.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase dilute ammonia R1, water R, nitromethane R, dioxan R (3:5:41:51 V/V/V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying At 100-105 °C.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 50 mg in 4 mL of methanol R and add 0.2 mL of a 40 g/L solution of copper acetate R. A flocculent, yellowish-green precipitate is formed, changing to dark green.

D. Dissolve about 5 mg in 1 M hydrochloric acid and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 10 mL with water R. The solution, without further acidification, gives the reaction of primary aromatic amines (2.3.1).

TESTS

Appearance of solution

Dissolve 1.0 g in a mixture of 5 mL of dilute sodium hydroxide solution R and 5 mL of water R. The solution is not more intensely coloured than reference solution Y₅, BY₅ or GY₅ (2.2.2, Method II).

Acidity

To 1.25 g add 25 mL of carbon dioxide-free water R and heat at 70 °C for 5 min. Cool for about 15 min in iced water and filter. To 20 mL of the filtrate add 0.1 mL of bromothymol blue solution R1. Not more than 0.5 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator. Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Related substances

Test solution (a) Dissolve 0.30 g of the substance to be examined in acetone R and dilute to 10 mL with the same solvent. Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with acetone R.

Reference solution (a) Dissolve 30 mg of sulfamethoxazole CRS in acetone R and dilute to 10 mL with the same solvent. Reference solution (b) Dilute 1 mL of test solution (b) to 20 mL with acetone R.

Apply to the plate 2 µL of each solution. Develop over a path of 15 cm using a mixture of 15 volumes of methanol R and 80 volumes of chloroform R. Dry the plate at 100 °C to 105 °C and examine in ultraviolet light at 254 nm. Any spot from the principal spot, is not more intense than the spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).

Heavy metals (2.4.8)

1.0 g complies with test D for heavy metals (20 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Carry out the determination of primary aromatic amino-nitrogen (2.5.8), using 0.2500 g and determining the end-point electrometrically.

1 mL of 0.1 M sodium nitrite is equivalent to 27.03 mg of C₉H₁₀N₄O₃S₂.

STORAGE

Store protected from light.

Ph Eur

— *total*: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent),
 — *disregard limit*: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.025 per cent).

Heavy metals (2.4.8)
 Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)
 Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
 Maximum 0.1 per cent, determined on 1.0 g.

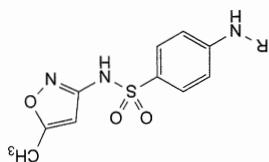
ASSAY
 Carry out the assay of primary aromatic amino-nitrogen (2.5.8), using 0.200 g and determining the end-point electrometrically.
 1 mL of 0.1 M sodium nitrite is equivalent to 25.33 mg of $C_{10}H_{11}N_3O_3S$.

STORAGE

Protected from light.

IMPURITIES

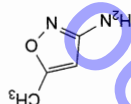
Specified impurities: A, B, C, D, E, F.



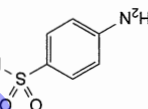
A. R = CO-CH₃; N-[4-[(5-methylisoxazol-3-yl)sulfamoyl]phenyl]acetamide,

B. R = SO₂-C₆H₄-p-NH₂;

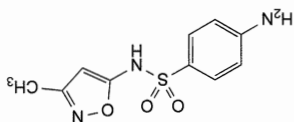
4-[[[(4-aminophenyl)sulfonyl]amino]-N-(5-methylisoxazol-3-yl)benzenesulfonamide,



C. 5-methylisoxazol-3-amine,



D. R = OH: 4-aminobenzenesulfonic acid (sulfanilic acid),
 E. R = NH₂: 4-aminobenzenesulfonamide (sulfanilamide),



F. 4-amino-N-(3-methylisoxazol-5-yl)benzenesulfonamide.

D. Dissolve about 5 mg in 10 mL of 1 M hydrochloric acid. Dilute 1 mL of the solution to 10 mL with water R. The solution, without further acidification, gives the reaction of primary aromatic amines (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅, BY₅ or GY₅ (2.2.2, Method II).

Dissolve 1.0 g in a mixture of 5 mL of dilute sodium hydroxide solution R and 5 mL of water R.

Acidity

To 1.25 g, finely powdered, add 25 mL of water R. Heat at 70 °C for 5 min. Cool in iced water for about 15 min and filter. To 20 mL of the filtrate add 0.1 mL of bromothymol blue solution R1. Not more than 0.3 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in 45 mL of the mobile phase, sonicate at about 45 °C for 10 min, cool and dilute to 50.0 mL with the mobile phase.
Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 1 mg of the substance to be examined and 1 mg of sulfamethoxazole impurity A CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.
Reference solution (c) Dissolve 1.0 mg of sulfamethoxazole impurity F CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:

— size: 1 = 0.25 m, Ø = 4.0 mm,
 — stationary phase: octylsilyl silica gel for chromatography R

(5 µm),

— temperature: 30 °C.

Mobile phase Mix 35 volumes of methanol R2 and 65 volumes of a 13.6 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 5.3 with a 20 g/L solution of potassium hydroxide R.

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Run time 3 times the retention time of sulfamethoxazole.

Relative retention With reference to sulfamethoxazole

(retention time = about 10 min): impurity D = about 0.3;

impurity E = about 0.35; impurity F = about 0.45;

impurity C = about 0.5; impurity A = about 1.2;

impurity B = about 2.0.

System suitability: reference solution (b):

— *resolution*: minimum 3.5 between the peaks due to sulfamethoxazole and impurity A.

Limits:

— *impurities* A, B, C, D, E: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent),
 — *impurity* F: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent),
 — *any other impurity*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent).

— *disregard limit*: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (2.4.4)

Maximum 140 ppm.

To 1.25 g add 50 mL of *distilled water* R. Heat at about

70 °C for 5 min. Cool and filter. To 20 mL of the filtrate

add 1 mL of *nitric acid* R, allow to stand for 5 min and filter

to obtain a clear solution.

Sulfates (2.4.13)

Maximum 400 ppm.

To 20 mL of the filtrate prepared for the test for chlorides

add 1 mL of *dilute hydrochloric acid* R, allow to stand for

5 min and filter.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution

using 2 mL of *lead standard solution* (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 0.1 M *sodium hydroxide* and dilute to

100.0 mL with the same solvent. Transfer 5.0 mL of this

solution to a 1000 mL volumetric flask containing about

750 mL of *water* R. Add 20.0 mL of 0.1 M *acetic acid* and

dilute to 1000.0 mL with *water* R. Prepare a standard

solution at the same time and in the same manner using

0.150 g of *sulfasalazine CRS*. Measure the absorbance

(2.2.25) of the 2 solutions at the absorption maximum at

359 nm.

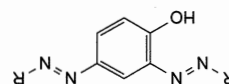
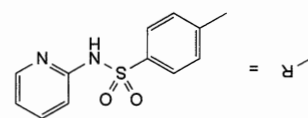
Calculate the content of C₁₈H₁₄N₄O₅ from the absorbances

STORAGE

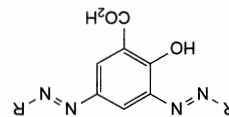
Protected from light.

IMPURITIES

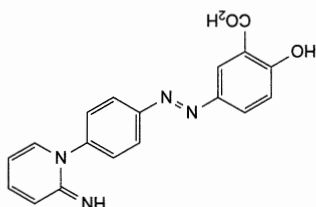
Specified impurities A, B, C, D, E, F, G, H, I, J



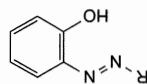
A. 4,4'-bis((4-hydroxy-3-phenylene)bis(diazenediyl))bis[N-(pyridin-2-yl)benzenesulfonamide],



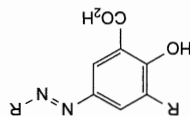
B. 2-hydroxy-3,5-bis[2-[4-(pyridin-2-ylsulfamoyl)phenyl]diazenyl]benzoic acid,



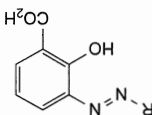
C. 2-hydroxy-5-[2-[4-(2-iminopyridin-1(2H)-yl)phenyl]diazenyl]benzoic acid,



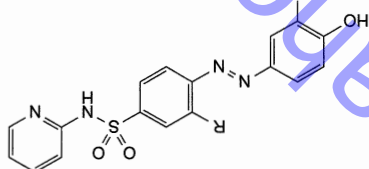
D. 4-[2-(2-hydroxyphenyl)diazenyl]-N-(pyridin-2-yl)benzenesulfonamide,



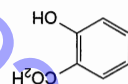
E. 2-hydroxy-4'-(pyridin-2-ylsulfamoyl)-5-[2-[4-(pyridin-2-ylsulfamoyl)phenyl]diazenyl]biphenyl-3-carboxylic acid,



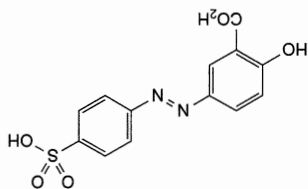
F. 2-hydroxy-3-[2-[4-(pyridin-2-ylsulfamoyl)phenyl]diazenyl]benzoic acid,



G. 5-[2-[4',5-bis(pyridin-2-ylsulfamoyl)biphenyl-2-yl]diazenyl]-2-hydroxybenzoic acid,



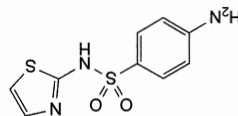
H. 2-hydroxybenzenecarboxylic acid (salicylic acid),



I. 2-hydroxy-5-[2-(4-sulphophenyl)diazenyl]benzoic acid, J. H₂N-R: 4-amino-N-(pyridin-2-yl)benzenesulfonamide (sulfapyridine).

Sulfathiazole

(Ph. Eur. monograph 0742)



$C_9H_9N_3O_2S_2$ 255.3 72-14-0

Action and use

Sulfonamide antibacterial.

Ph Eur

DEFINITION

Sulfathiazole contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 4-amino-N-(thiazol-2-yl)benzenesulfonamide, calculated with reference to the dried substance.

CHARACTERS

A white or slightly yellowish, crystalline powder, practically insoluble in water, slightly soluble in alcohol, practically insoluble in methylene chloride. It dissolves in dilute solutions of alkali hydroxides and in dilute mineral acids.

IDENTIFICATION

First identification A, B.
Second identification A, C, D, E.

Related substances
Examine by thin-layer chromatography (2.2.27), using silica gel H R as the coating substance.
Test solution (a) Dissolve 0.10 g of the substance to be examined in a mixture of 1 volume of concentrated ammonia R and 9 volumes of alcohol R and dilute to 10 mL with the same mixture of solvents.
Test solution (b) Dilute 1 mL of test solution (a) to 5 mL with a mixture of 1 volume of concentrated ammonia R and 9 volumes of alcohol R.
Reference solution (a) Dissolve 20 mg of sulfathiazole CRS in a mixture of 1 volume of concentrated ammonia R and 9 volumes of alcohol R and dilute to 10 mL with the same mixture of solvents.
Reference solution (b) Dissolve 50 mg of sulfanilamide R in a mixture of 1 volume of concentrated ammonia R and 9 volumes of alcohol R and dilute to 100 mL with the same mixture of solvents. Dilute 1 mL of this solution to 10 mL with the same mixture of solvents.
Apply to the plate 10 µL of each solution. Develop over a path of 15 cm using a mixture of 18 volumes of ammonia R and 90 volumes of butanol R. Dry the plate at 100 °C to 105 °C for 10 min and spray with a 1 g/L solution of dimethylaminobenzaldehyde R in alcohol R containing 1 per cent V/V of hydrochloric acid R. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent).
Heavy metals (2.4.8)
1.0 g complies with test C for heavy metals (20 ppm). Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.
Loss on drying (2.2.32)
Not more than 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
Not more than 0.1 per cent, determined on 1.0 g.
ASSAY
Carry out the determination of primary aromatic amino-nitrogen (2.5.8), using 0.200 g, determining the end-point electrometrically.
1 mL of 0.1 M sodium nitrite is equivalent to 25.53 mg of $C_9H_9N_3O_2S_2$.
STORAGE
Store protected from light.

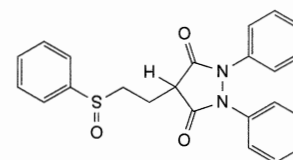
Ph Eur

TESTS

Appearance of solution
Dissolve 1.0 g in 10 mL of 1 M sodium hydroxide. The solution is clear (2.2.1) and not more intensely coloured than reference solution GY₄ (2.2.2, Method II).
Acidity
To 1.0 g add 50 mL of carbon dioxide-free water R. Heat to 70 °C for 5 min. Cool rapidly to 20 °C and filter. To 25 mL of the filtrate add 0.1 mL of bromothymol blue solution R1. Not more than 0.1 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.
TESTS
reaction of primary aromatic amines (2.3.1).
The solution, without further addition of acid, gives the Dilute 1 mL of the solution to 10 mL with water R.
E. Dissolve about 5 mg in 10 mL of 1 M hydrochloric acid. Dissolve about 10 mg in a mixture of 10 mL of water R and 2 mL of 0.1 M sodium hydroxide and add 0.5 mL of copper sulfate solution R. A greyish-blue or purple precipitate is formed.
D. Dissolve about 10 mg in a mixture of 10 mL of water R with reference solution (a).
and size to the principal spot in the chromatogram obtained with test solution (b) is similar in position, colour related substances. The principal spot in the chromatogram C. Examine the chromatograms obtained in the test for record the spectra again using the residues.
separately in alcohol R, evaporate to dryness *in vacuo* and substance to be examined and the reference substance If the spectra obtained show differences, dissolve the sulfathiazole CRS. Examine the substances prepared as discs. B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with second melting between 200 °C and 203 °C.
occur at about 175 °C, followed by solidification and a A. Melting point (2.2.14): 200 °C to 203 °C. Melting may

Sulfapyrazone

(Ph. Eur. monograph 0790)

 $C_{23}H_{20}N_2O_3S$

404.5

57-96-5

Action and use

Uricosuric.

Preparation

Sulfapyrazone Tablets

Ph. Eur.

DEFINITION

1,2-[2-phenyl-4-[2-(phenylsulfonyl)ethyl]pyrazol-5-ylidene]-3,5-dione.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly soluble in water, sparingly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification A, C.

Second identification A, B, D.

A. Melting point (2.2.16): 131 °C to 135 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 30.0 mg in 0.01 M sodium hydroxide and dilute to 100.0 mL with the same alkaline solution. Dilute 1.0 mL of this solution to 20.0 mL with 0.01 M sodium hydroxide.

Spectral range 230-350 nm.

Absorption maximum At 260 nm.

Specific absorbance at the absorption maximum 530 to 580.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison sulfapyrazone CRS.

D. Dissolve about 10 mg in 3 mL of acetone R and add a mixture of 0.2 mL of ferric chloride solution R2 and 3 mL of water R. A red to violet colour develops.

TESTS

Appearance of solution in acetone

The solution is clear (2.2.1) and its absorbance (2.2.25) at 420 nm using a path length of 4 cm is not greater than 0.10.

Dissolve 1.25 g in acetone R and dilute to 25 mL with the same solvent.

Appearance of solution in 1 M sodium hydroxide

The solution is clear (2.2.1) and its absorbance (2.2.25) at 420 nm using a path length of 4 cm is not greater than 0.15.

Dissolve 1.25 g, heating gently if necessary, in 25 mL of 1 M sodium hydroxide.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Solvent mixture water R, acetonitrile R (10:40 V/V).

Test solution Dissolve 10.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5.0 mg of sulfapyrazone impurity A CRS and 5.0 mg of sulfapyrazone impurity B CRS in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 100.0 mL with the solvent mixture.

Reference solution (d) Dissolve the contents of a vial of sulfapyrazone for system suitability CRS (containing impurity C) in 1.0 mL of the solvent mixture.

Reference solution (e) Dilute 1.0 mL of reference solution (a) to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.125$ m, $\varnothing = 4.6$ mm;— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase tetrahydrofuran for chromatography R, acetonitrile R, 0.3 per cent V/V solution of phosphoric acid R (7:35:58 V/V/V).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 255 nm.

Injection 20 μ L.

Run time 7 times the retention time of sulfapyrazone.

Identification of impurities Use the chromatogram supplied with sulfapyrazone for system suitability CRS and the chromatogram obtained with reference solution (d) to

identify the peak due to impurity C; use the chromatogram obtained with reference solution (b) to identify the peaks due

to impurities A and B.

Relative retention With reference to sulfapyrazone (retention time = about 3.5 min): impurity C = about 0.8; impurity A = about 1.6; impurity B = about 4.8.

System suitability Reference solution (d):

— resolution: minimum 2.0 between the peaks due to impurity C and sulfapyrazone.

Limits:

— impurities A, B: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (1.0 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (e) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (e) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (2.0 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (e) (0.05 per cent).

Heavy metals (2.4.8)

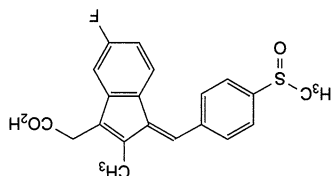
Maximum 10 ppm.

Solvent mixture acetone R, ethanol (96 per cent) R (50:50 V/V).



(Ph. Eur. monograph 0864)

Sulindac

C₂₀H₁₇FO₃S 356.4 38194-50-2

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Preparation

Sulindac Tablets

Ph Eur

DEFINITION

(Z)-[5-Fluoro-2-methyl-1-[4-(methylsulfinyl)benzylidene]-1H-inden-3-yl]acetic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Very slightly soluble in water, soluble in methylene chloride, sparingly soluble in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

It shows polymorphism (5.9).

IDENTIFICATION

First identification: C.

Second identification: A, B, D, E

A. Melting point (2.2.14): 182 °C to 186 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 50 mg in a 0.3 per cent V/V solution of hydrochloric acid R and dilute to 100 mL with the same acid solution. Dilute 2 mL of this solution to 50 mL with a 0.3 per cent V/V solution of hydrochloric acid R in methanol R.

Spectral range 230-350 nm.

Absorption maxima At 284 nm and 327 nm.

Shoulder At about 258 nm.

Absorbance ratio $A_{284}/A_{327} = 1.10$ to 1.20 .

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison sulindac CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of hot methanol R, evaporate to dryness and record new spectra using the residues.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

IMPURITIES

Specified impurities A, B, C.

STORAGE

Protected from light.

C₂₃H₂₀N₂O₃S.

Dissolve 0.300 g in 25 mL of acetone R. Add 0.5 mL of bromothymol blue solution R1. Titrate with 0.1 M sodium hydroxide until the colour changes from yellow to blue.

1 mL of 0.1 M sodium hydroxide is equivalent to 40.45 mg of

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.5 per cent, determined on 1.000 g by drying in

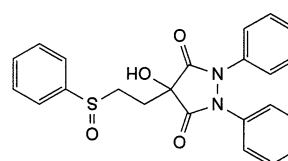
Loss on drying (2.2.32)

solution pH 3.5 R.

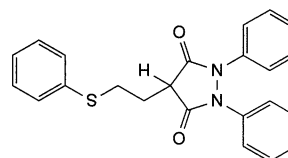
To each solution, add 10 mL of water R and 2 mL of buffer

using 2.5 mL of lead standard solution (1 ppm Pb) R.

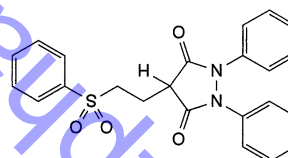
0.250 g complies with test H. Prepare the reference solution



B. 1,2-diphenyl-4-[2-(phenylsulfonyl)ethyl]pyrazolidine-3,5-dione,



A. 1,2-diphenyl-4-[2-(phenylsulfonyl)ethyl]pyrazolidine-3,5-dione,



C. 4-hydroxy-1,2-diphenyl-4-[2-(phenylsulfonyl)ethyl]pyrazolidine-3,5-dione.

Ph Eur

Reference solution (a) Dissolve 10 mg of *sulindac CRS* in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of *diffusional CRS* in methylene chloride R and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 2 mL with reference solution (a).

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase glacial acetic acid R, methylene chloride R, acetone R (1:49:50 V/V/V).

Application 5 µL.

Development Over a path of 15 cm.

Drying In a current of warm air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b): — the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

E. Mix about 5 mg with 45 mg of heavy magnesium oxide R and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of water R, 0.05 mL of phenolphthalein solution R1 and about 1 mL of dilute hydrochloric acid R to render the solution colourless. Filter. Add 1.0 mL of the filtrate to a freshly prepared mixture of 0.1 mL of alizarin S solution R and 0.1 mL of zirconyl nitrate solution R. Mix, allow to stand for 5 min and compare the colour of the solution with that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.10 g of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 20.0 mg of *sulindac CRS* (which has an assigned content of (*E*)-isomer) in the mobile phase and dilute to 10.0 mL with the mobile phase.

Column: — size: $l = 0.25$ m, $\phi = 4.6$ mm; — stationary phase: silica gel for chromatography R (10 µm).
Mobile phase glacial acetic acid R, ethanol (96 per cent) R, ethyl acetate R, ethanol-free chloroform R (1:4:100:400 V/V/V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Run time Twice the retention time of *sulindac*.

Identification of peaks The chromatogram obtained with reference solution (b) shows a principal peak due to *sulindac* and a peak due to the (*E*)-isomer.

Relative retention With reference to *sulindac*: (*E*)-isomer = about 1.75.

From the chromatograms obtained with the test solution and reference solution (b), determine the percentage content of (*E*)-isomer, taking into account the assigned content of this isomer in *sulindac CRS*.

Limits: — (*E*)-isomer: maximum 0.5 per cent; — impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent); — total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C at a pressure not exceeding 0.7 kPa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of methanol R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

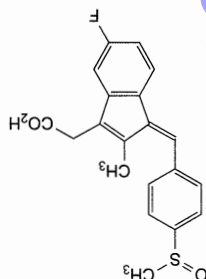
1 mL of 0.1 M sodium hydroxide is equivalent to 35.64 mg of C₂₀H₁₇FO₃S.

STORAGE

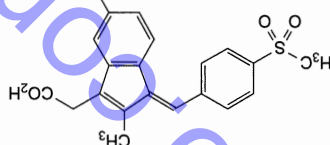
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IMPURITIES

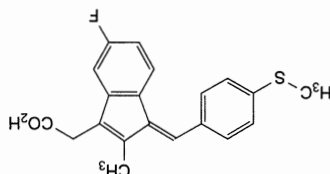
Specified impurities A, B, C



A. (*E*)-[5-fluoro-2-methyl-1-[4-(methylsulfonyl)benzylidene]-1*H*-inden-3-yl]acetic acid.



B. (*Z*)-[5-fluoro-2-methyl-1-[4-(methylsulfonyl)benzylidene]-1*H*-inden-3-yl]acetic acid.



C. (*Z*)-[5-fluoro-2-methyl-1-[4-(methylsulfonyl)benzylidene]-1*H*-inden-3-yl]acetic acid.

Sulfur for External Use

Sulphur for External Use
(Ph. Eur. monograph 0953)
32.07
7704-34-9

Action and use
Keratolytic.

DEFINITION

Content
99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance
Yellow powder.

Solubility

Practically insoluble in water, soluble in carbon disulfide, slightly soluble in vegetable oils.

mp: about 120 °C.

The size of most of the particles is not greater than 20 µm and that of almost all the particles is not greater than 40 µm.

IDENTIFICATION

A. Heated in the presence of air, it burns with a blue flame, emitting sulfur dioxide which changes the colour of moistened blue litmus paper R to red.

B. Heat 0.1 g with 0.5 mL of bromine water R until decolourised. Add 5 mL of water R and filter. The solution gives reaction (a) of sulfates (2.3.1).

TESTS

Solution S
To 5 g add 50 mL of carbon dioxide-free water R prepared from distilled water R. Allow to stand for 30 min with frequent shaking and filter.

Appearance of solution

Solution S is colourless (2.2.2, Method II).

Odour (2.3.4)

It has no perceptible odour of hydrogen sulfide.

Acidity or alkalinity

To 5 mL of solution S add 0.1 mL of phenolphthalein solution R1. The solution is colourless. Add 0.2 mL of 0.01 M sodium hydroxide. The solution is red. Add 0.3 mL of 0.01 M hydrochloric acid. The solution is colourless.

Add 0.15 mL of methyl red solution R. The solution is orange-red.

Chlorides (2.4.4)

Maximum 100 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 100 ppm, determined on solution S.

Sulfides

To 10 mL of solution S add 2 mL of buffer solution pH 3.5 R and 1 mL of a freshly prepared 1.6 g/L solution of lead nitrate R in carbon dioxide-free water R. Shake. After 1 min any colour in the solution is not more intense than that in a reference solution prepared at the same time using 1 mL of lead standard solution (10 ppm Pb) R, 9 mL of carbon dioxide-free water R, 2 mL of buffer solution pH 3.5 R and 1.2 mL of thioacetamide reagent R.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

Sulfuric Acid

Sulphuric Acid
(Ph. Eur. monograph 1572)
H₂SO₄
98.1
7664-93-9

Preparation

Dilute Sulfuric Acid

DEFINITION

Content

95.0 per cent *m/m* to 100.5 per cent *m/m*.

CHARACTERS

Appearance

Colourless, oily liquid, very hygroscopic.

Solubility

Miscible with water and with ethanol (96 per cent) producing intense heat.

Relative density

About 1.84.

IDENTIFICATION

A. Carefully add 1 mL to 100 mL of water R. The solution is strongly acid (2.2.4).
B. The solution obtained in identification test A gives reaction (a) of sulfates (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Carefully pour, while cooling, 5 mL into 30 mL of water R and dilute to 50 mL with the same solvent.

Chlorides (2.4.4)

Maximum 50 ppm.

Mix carefully, while cooling, 3.3 g with 30 mL of water R. Neutralise with ammonia R and dilute to 50 mL with water R.

Nitrates

Add 5 mL to 5 mL of water R. Cool to room temperature and add 0.5 mL of indigo carmine solution R. The blue colour persists for at least 1 min.

Arsenic (2.4.2, Method A)

Maximum 1 ppm.

Mix, while cooling, 1 g with 20 mL of water R and dilute to 25 mL with the same solvent.

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in methanol, slightly soluble in ethanol (96 per cent) and in methylene chloride. It dissolves in dilute solutions of mineral acids and alkali hydroxides.

IDENTIFICATION**First identification B.****Second identification A, C, D**

A. Melting point (2.2.14): 177 °C to 181 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Examine the chromatograms obtained in the test for

impurity A.

Detection Examine in ultraviolet light at 254 nm. **Results** The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. To about 1 mg in a porcelain dish, add 0.5 mL of *sulfuric acid R* and 0.05 mL of *formaldehyde solution R*. Examine in ultraviolet light at 365 nm, the solution shows blue fluorescence.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method J).

Dissolve 1.0 g in *dilute acetic acid R* and dilute to 10 mL with the same acid.

Impurity A.

Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.20 g of the substance to be

examined in *methanol R*, sonicate until dissolution is

complete and dilute to 10.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to

10.0 mL with *methanol R*.

Reference solution (a) Dissolve 20 mg of *sulpiride CRS* in

methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dissolve 5.0 mg of *sulpiride*

impurity A CRS in *methanol R* and dilute to 25.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with

methanol R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, dioxan R, *methanol R*, *methylene chloride R* (2:10:14:90 V/V/V/V).

Application 10 µL.

Development Over 1/2 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm for identification test C and then spray with *ninhydrin solution R*, heat at 100-105 °C for 15 min and examine in daylight.

Limit: test solution (a):

— **impurity A:** any spot due to impurity A is not more

intense than the corresponding spot in the chromatogram obtained with reference solution (b) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Iron (2.4.9)

Maximum 25 ppm.

Cautiously evaporate 10.0 g and ignite to dull redness.

Dissolve the ignition residue in 1 mL of *dilute hydrochloric acid R* with gentle heating and dilute to 25 mL with *water R*.

Dilute 1 mL of this solution to 10 mL with *water R*.

Heavy metals (2.4.8)

Maximum 5 ppm.

4.0 g complies with test F. Prepare the reference solution

using 2 mL of *lead standard solution* (10 ppm Pb) R.

ASSAY

Weigh accurately a ground-glass-stoppered flask containing

30 mL of *water R*. Introduce 0.2 mL, cool and weigh again.

Titrate with 1 M *sodium hydroxide*, determining the end-point potentiometrically (2.2.20).

1 mL of 1 M *sodium hydroxide* is equivalent to 49.04 mg of H₂SO₄.

STORAGE

In an airtight container.

Dilute Sulfuric Acid

Dilute Sulphuric Acid

DEFINITION

Dilute Sulfuric Acid is prepared by adding 104 g of Sulfuric

Acid to 896 g of Purified Water with constant stirring and

cooling. It contains not less than 9.5% and not more than

10.5% w/w of H₂SO₄.

Weight per mL

1.062 g to 1.072 g, Appendix V G.

ASSAY

To 10 g add 40 mL of *water* and titrate with 1 M *sodium*

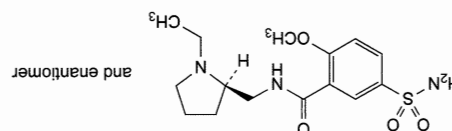
hydroxide VS using *methyl orange solution* as indicator. Each

mL of 1 M *sodium hydroxide VS* is equivalent to 49.04 mg of

H₂SO₄.

Sulpiride

(Ph. Eur. monograph 1045)



C₁₅H₂₃N₃O₄S 341.4 15676-16-1

Preparation

Dopamine receptor antagonist; neuroleptic.

Sulpiride Tablets

Ph. Eur.

DEFINITION

N-[(2*RS*)-1-Ethylpyrrolidin-2-yl]methyl]-2-methoxy-5-

sulfamoylbenzamide.

Content

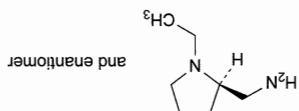
98.5 per cent to 101.0 per cent (dried substance).

1 mL of 0.1 M perchloric acid is equivalent to 34.14 mg of $C_{15}H_{23}N_3O_4S$.

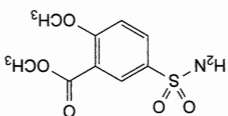
IMPURITIES

Specified impurities A

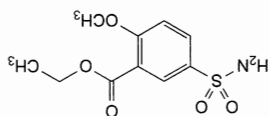
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E, F, G.



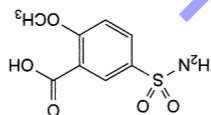
A. [(2R,S)-1-ethylpyrrolidin-2-yl]methanamine,



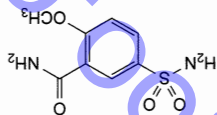
B. methyl 2-methoxy-5-sulfamoylbenzoate,



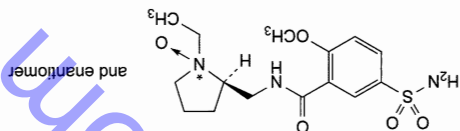
C. ethyl 2-methoxy-5-sulfamoylbenzoate,



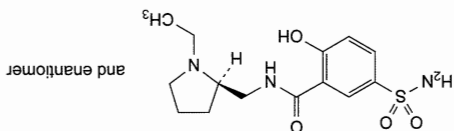
D. 2-methoxy-5-sulfamoylbenzoic acid,



E. 2-methoxy-5-sulfamoylbenzamide,



F. N-[(2R,S)-1-ethyl-1-oxidopyrrolidin-2-yl]methyl-2-methoxy-5-sulfamoylbenzamide,



G. N-[(2R,S)-1-ethylpyrrolidin-2-yl]methyl-2-hydroxy-5-sulfamoylbenzamide.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of the substance to be examined and 5 mg of sulpiride impurity B CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:
— size: $l = 0.25$ m, $\phi = 4.6$ mm;
— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 10 volumes of acetonitrile R, 10 volumes of methanol R and 80 volumes of a solution containing 6.8 g/L of potassium dihydrogen phosphate R and 1 g/L of sodium octanesulfonate R, previously adjusted to pH 3.3 with phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 μ L.

Run time: twice the retention time of sulpiride.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity B.

Relative retention With reference to sulpiride (retention time = about 15 min): impurity B = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 2.5 between the peaks due to impurity B and sulpiride.

Calculation of percentage contents:

— for each impurity, use the concentration of sulpiride in reference solution (a).

Limits:

— unspecified impurities: for each impurity, maximum 0.10 per cent;

— total: maximum 0.3 per cent;

— reporting threshold: 0.05 per cent.

Chlorides (2.4.4)

Maximum 100 ppm.

Shake 1.0 g with 20 mL of water R. Filter through a sintered-glass filter (40) (2.1.2). To 10 mL of the filtrate add 5 mL of water R.

Iron (2.4.9)

Maximum 10 ppm.

Ignite 1.0 g in a silica crucible. To the residue add 1 mL of 1 M hydrochloric acid, 3 mL of water R and 0.1 mL of nitric acid R. Heat on a water-bath for about 5 min. Place the solution in a test-tube. Rinse the crucible with 4 mL of water R. Collect the rinsings in the test-tube and dilute to 10 mL with water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

1.0 g complies with test C. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

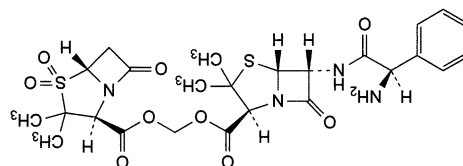
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 80 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

Sultamicillin

(Ph. Eur. monograph 2211)


 $C_{25}H_{30}N_4O_{12}$ 594.7 76497-13-7

Action and use
Beta-lactamase inhibitor.

Ph. Eur.

DEFINITION

Methylene (2S,5R,6R)-6-[[[(2R)-aminophenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate (2S,5R)-3,3-dimethyl-4,4,7-trioxo-4 λ^6 -thia-1-azabicyclo[3.2.0]heptane-2-carboxylate.

Content
Semi-synthetic product derived from a fermentation product.

96.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Practically insoluble in water, very slightly soluble in methanol, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison sultamicillin CRS.

TESTS

Specific optical rotation (2.2.7)

+ 190 to + 210 (anhydrous substance).

Dissolve 0.500 g in dimethylformamide R and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or keep at 2–8 °C for not more than 6 h.

Solution A methanol R1, acetonitrile R1 (20:80 V/V).

Solution B Dissolve 1.56 g of sodium dihydrogen phosphate R in 900 mL of water R. Add 7.0 mL of phosphoric acid R and dilute to 1000 mL with water R.

Blank solution Solution B, solution A (30:70 V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in 35 mL of solution A and sonicate for about 1 min. Add 13 mL of solution B, mix and sonicate for about 1 min. Dilute to 50.0 mL with solution B and mix.

Reference solution (a) Dissolve 70.0 mg of sultamicillin CRS in 35 mL of solution A and sonicate for about 1 min. Add 13 mL of solution B, mix and sonicate for about 1 min. Dilute to 50.0 mL with solution B and mix.

Reference solution (b) Suspend 15 mg of sultamicillin CRS in 20 mL of a 0.4 g/L solution of sodium hydroxide R and sonicate in an ultrasonic bath for about 5 min. Add 20 mL of a 0.36 g/L solution of hydrochloric acid R and dilute to 100 mL with water R.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the blank solution.

Reference solution (d) Dissolve 17.3 mg of ampicillin trihydrate CRS (impurity C) and 15.0 mg of subactam CRS (impurity A) in water R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of this solution to 100.0 mL with water R.

Reference solution (e) Dissolve 5 mg of sultamicillin for peak identification CRS (containing impurity G) in 7.0 mL of solution A and sonicate for about 1 min. Dilute to 10.0 mL with solution B, mix and sonicate for about 1 min.

Column: size: $l = 0.10$ m, $\varnothing = 4.6$ mm; stationary phase: octadecylsilyl silica gel for chromatography R (3.5 μ m); temperature: 25 °C.

Mobile phase: mobile phase A: 4.68 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R; mobile phase B: acetonitrile R1;

Flow rate 1.0 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	95 \rightarrow 30	5 \rightarrow 70
15 - 16	30	70
16 - 16.5	30 \rightarrow 95	70 \rightarrow 5
16.5 - 20	95	5

Detection Spectrophotometer at 215 nm.

Injection 5 μ L of the blank solution, the test solution and reference solutions (b), (c), (d) and (e).

Identification of impurities Use the chromatogram supplied with sultamicillin for peak identification CRS and the chromatogram obtained with reference solution (e) to identify the peak due to impurity G.

Relative retention With reference to sultamicillin (retention time = about 9.3 min): impurity A = about 0.41; ampicillin penicilloic acid = about 0.47; impurity B = about 0.50; impurity C = about 0.55; impurity D = about 0.94; impurity E = about 1.09; impurity F = about 1.26; impurity G = about 1.42.

System suitability: reference solution (b):

— resolution: minimum 2.5 between the peaks due to ampicillin penicilloic acid and impurity B and minimum 2.5 between the peaks due to impurities B and C.

Limits:

— impurity G: not more than the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (1.0 per cent);

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.3 per cent);

— impurity C: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.3 per cent);

— impurities D, E, F: for each impurity, not more than 0.3 times the area of the peak due to sultamicillin in the

chromatogram obtained with reference solution (c) (0.3 per cent); for each impurity, not more than 0.3 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (0.3 per cent); not more than 3 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (3.0 per cent); disregard limit: 0.1 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (0.1 per cent).

Ethyl acetate

Head-space gas chromatography (2.2.28).
Test solution: Dissolve 0.200 g in 7.0 mL of a mixture of 1 volume of water R and 99 volumes of dimethylformamide R. Reference solution: Dissolve 0.200 g of ethyl acetate R in 240 mL of a mixture of 1 volume of water R and 99 volumes of dimethylformamide R and dilute to 250.0 mL with the same mixture of solvents. Dilute 5.0 mL of this solution to 7.0 mL with a mixture of 1 volume of water R and 99 volumes of dimethylformamide R. Shake to obtain a homogeneous solution.

Close the vials immediately with a tight rubber membrane stopper coated with polytetrafluoroethylene and secure with an aluminium crimped cap. Shake to obtain a homogeneous solution.

Column:

— material: fused silica;
— size: $l = 50$ m, $\phi = 0.32$ mm;
— stationary phase: poly(dimethyl)siloxane R (film thickness: 1.8 μ m or 3 μ m).
Carrier gas: helium for chromatography R.
Linear velocity: 35 cm/s.

Split ratio: 1:5.

Static head-space conditions that may be used:
— equilibration temperature: 105 °C;
— equilibration time: 45 min;
— transfer-line temperature: 110 °C;
— pressurisation time: 30 s.

Temperature:

Time (min)	Temperature (°C)
0 - 6	70
6 - 16	70 → 220
16 - 18	220
Injection port	140
Detector	250

Detection: Flame ionisation.

Injection: 1 mL.

Relative retention: With reference to dimethylformamide

(retention time = about 14 min): ethyl acetate = about 0.7.

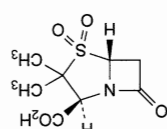
Limit:

— ethyl acetate: maximum 2.5 per cent.

Heavy metals (2.4.8)

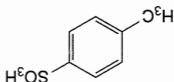
Maximum 20 ppm.

Dissolve 2.0 g in a mixture of 40 volumes of methanol R and 60 volumes of acetonitrile R and dilute to 20.0 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (2 ppm Pb) obtained by diluting lead

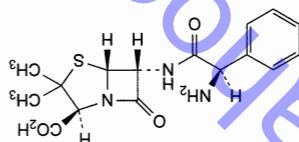


Specified impurities A, B, C, D, E, F, G

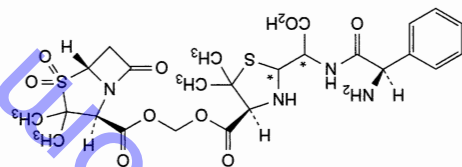
A. (2S,5R,6R)-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid 4,4-dioxide (sulbactam),



B. 4-methylbenzenesulfonic acid (*p*-toluenesulfonic acid),



C. (2S,5R,6R)-6-[[[(2R)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (ampicillin),



D. [[(2R)-aminophenylacetyl]amino] [(4S)-4-[[[(2S,5R)-3,3-dimethyl-4,7-oxo-4 λ^6 -thia-1-azabicyclo[3.2.0]hept-2-yl]carbonyl]oxy]methoxy]carbonyl]-5,5-dimethylthiazolidin-2-yl]acetic acid (penicillanic acids of sultamicillin),

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Injection: Test solution and reference solution (a).

Calculate the percentage content of sultamicillin ($C_{25}H_{30}N_4O_9S_2$) from the declared content of $C_{25}H_{30}N_4O_9S_2$ in sultamicillin tosylate CRS and by multiplying the sultamicillin tosylate content by 0.7752.

STORAGE

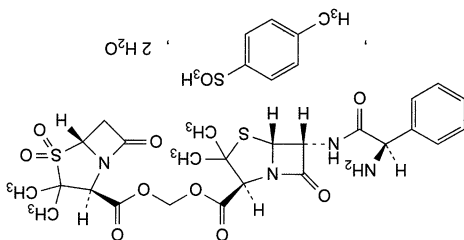
In an airtight container.

IMPURITIES



Sultamicillin Tosilate Dihydrate

(Ph. Eur. monograph 2212)



$C_{32}H_{38}N_4O_{12}S_3 \cdot 2H_2O$ 803 83105-70-8

Action and use

Beta-lactamase inhibitor.

Ph Eur

DEFINITION

4-Methylbenzenesulfonate of methylene (2S,5R,6R)-6-[[

[(2R)-amino]phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-

1-azabicyclo[3.2.0]heptane-2-carboxylate (2S,5R)-3,3-

dimethyl-4,4,7-trioxo-4λ⁶-thia-1-azabicyclo[3.2.0]heptane-2-

carboxylate dihydrate.

Semi-synthetic product derived from a fermentation product.

Content 95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in ethanol

(96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison sultamicillin tosilate CRS.

TESTS

Specific optical rotation (2.2.7)

+ 178 to + 195 (anhydrous substance).

Dissolve 1.000 g in dimethylformamide R and dilute to

50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or keep at 2–8 °C for not more than 6 h.

Solution A methanol R1, acetonitrile R1 (20:80 V/V).

Solution B Dissolve 1.56 g of sodium dihydrogen phosphate R in 900 mL of water R. Add 7.0 mL of phosphoric acid R and dilute to 1000 mL with water R.

Blank solution Solution B, solution A (30:70 V/V).

Test solution Dissolve 70.0 mg of the substance to be examined in 35 mL of solution A and sonicate for about

1 min. Add 13 mL of solution B, mix and sonicate for about

1 min. Dilute to 50.0 mL with solution B and mix.

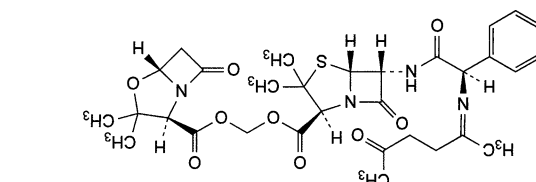
Reference solution (a) Dissolve 70.0 mg of sultamicillin tosilate CRS in 35 mL of solution A and sonicate for about

1 min. Add 13 mL of solution B, mix and sonicate for about

1 min. Dilute to 50.0 mL with solution B and mix.

Reference solution (b) Suspend 15 mg of the substance to be examined in 20 mL of a 0.4 g/L solution of sodium

hydroxide R and sonicate in an ultrasonic bath for about

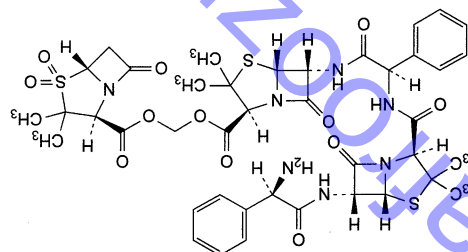


E. methylene (2S,5R,6R)-3,3-dimethyl-6-[[

[(2R)-amino]phenylacetyl]amino]-7-oxo-4-thia-1-

azabicyclo[3.2.0]heptane-2-carboxylate (2S,5R)-3,3-dimethyl-

7-oxo-4-oxa-1-azabicyclo[3.2.0]heptane-2-carboxylate,



F. methylene (2S,5R,6R)-6-[[

[(2R)-amino]-3,3-dimethyl-7-oxo-4-thia-1-

aminophenylacetyl]amino]-3,3-dimethyl-7-oxo-4-

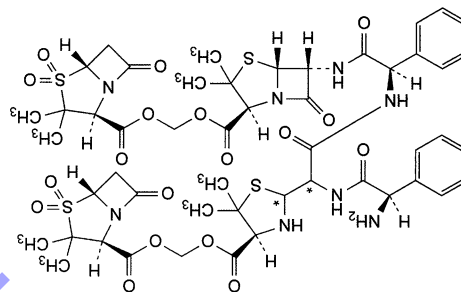
azabicyclo[3.2.0]hept-2-

yl]carbonyl]amino]phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-

thia-1-azabicyclo[3.2.0]heptane-2-carboxylate (2S,5R)-3,3-

dimethyl-4,4,7-trioxo-4λ⁶-thia-1-azabicyclo[3.2.0]heptane-2-

carboxylate (ampicillin sultamicillin amide),



G. methylene (2S,5R,6R)-6-[[

[(2R)-amino]phenylacetyl]amino]-7-oxo-4-thia-1-

azabicyclo[3.2.0]hept-2-

yl]carbonyl]oxy]methoxy]carbonyl]-5,5-dimethylthiazolidin-2-

4,4,7-trioxo-4λ⁶-thia-1-azabicyclo[3.2.0]hept-2-

yl]carbonyl]amino]phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-

thia-1-azabicyclo[3.2.0]heptane-2-carboxylate (2S,5R)-3,3-

dimethyl-4,4,7-trioxo-4λ⁶-thia-1-azabicyclo[3.2.0]heptane-2-

carboxylate (sultamicillin dimer).

Ph Eur

5 min. Add 20 mL of a 0.36 g/L solution of hydrochloric acid R and dilute to 100.0 mL with water R.

Reference solution (c) Dissolve 0.200 g of the substance to be examined in 70.0 mL of solution A and sonicate for about 1 min. Add 25.0 mL of solution B, mix and sonicate for about 1 min. Dilute to 100.0 mL with solution B and mix.

Dilute 1.0 mL of this solution to 100.0 mL with the blank solution.

Reference solution (d) Dissolve 32.3 mg of ampicillin trihydrate CRS (impurity B) and 7.0 mg of sulbactam CRS (impurity A) in water R and dilute to 1000 mL with the same solvent.

Column:
— size: $\varnothing = 0.10$ m, $\varnothing = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (3.5 μ m);
— temperature: 25 °C.

Mobile phase:
— mobile phase A: 4.68 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R;
— mobile phase B: acetonitrile R₁;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 15	95 \rightarrow 30	5 \rightarrow 70
15 - 16	30	70
16 - 16.5	30 \rightarrow 95	70 \rightarrow 5
16.5 - 20	95	5

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 5 μ L of the blank solution, the test solution and reference solutions (b), (c) and (d).

Relative retention With reference to sultamicillin (retention time = about 9.3 min): impurity A = about 0.41; ampicillin penicilloic acid = about 0.47; tosilate = about 0.50; impurity B = about 0.55; impurity C = about 0.94; impurity D = about 1.09; impurity F = about 1.23; impurity E = about 1.26; impurity G = about 1.42.

System suitability: reference solution (b):
— resolution: minimum 2.5 between the peaks due to ampicillin penicilloic acid and tosilate and minimum 2.5 between the peaks due to tosilate and impurity B.
Limits:
— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (2.0 per cent);
— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (d) (0.5 per cent);
— impurities C, D, E, F, G: for each impurity, not more than 0.5 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (0.5 per cent);
— any other impurity: for each impurity, not more than 0.5 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (0.5 per cent);
— total: not more than 4 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (4.0 per cent);

— disregard limit: 0.1 times the area of the peak due to sultamicillin in the chromatogram obtained with reference solution (c) (0.1 per cent).

Ethyl acetate

Head space gas chromatography (2.2.28).

Test solution Dissolve 0.200 g in 7.0 mL of a mixture of 1 volume of water R and 99 volumes of dimethylformamide R. **Reference solution** Dissolve 0.200 g of ethyl acetate R in 240 mL of a mixture of 1 volume of water R and 99 volumes of dimethylformamide R and dilute to 250.0 mL with the same mixture of solvents. Dilute 5.0 mL of this solution to 7.0 mL with a mixture of 1 volume of water R and 99 volumes of dimethylformamide R.

Immediately close the vials with a tight rubber membrane stopper coated with polytetrafluoroethylene and secure with an aluminium crimped cap. Shake to obtain a homogeneous solution.

Column:

— material: fused silica;
— size: $\varnothing = 50$ m, $\varnothing = 0.32$ mm;
— stationary phase: poly(dimethyl)siloxane R (film thickness: 1.8 μ m or 3 μ m).

Carrier gas helium for chromatography R.

Linear velocity 35 cm/s.

Split ratio 1:5.

Static head-space conditions that may be used:
— equilibration temperature: 105 °C;
— equilibration time: 45 min;
— transfer-line temperature: 110 °C;
— pressurisation time: 30 s.

Temperature:

Time (min)	Temperature (°C)
0 - 6	70
6 - 16	70 \rightarrow 220
16 - 18	220
Injection port	140
Detector	250

Detection Flame ionisation.

Injection 1 mL.

Relative retention With reference to dimethylformamide (retention time = about 14 min): ethyl acetate = about 0.7.

Limit:

— ethyl acetate: maximum 2.0 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in a mixture of 40 volumes of methanol R and 60 volumes of acetonitrile R and dilute to 20.0 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (2 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with a mixture of 40 volumes of methanol R and 60 volumes of acetonitrile R.

Water (2.5.12)

4.0 per cent to 6.0 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

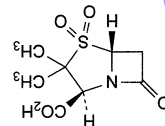
Calculate the percentage content of sumatriptan in tosilate ($C_{37}H_{38}N_4O_{12}S_3$) from the declared content of sumatriptan in tosilate CRS.

STORAGE

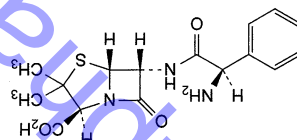
In an airtight container.

IMPURITIES

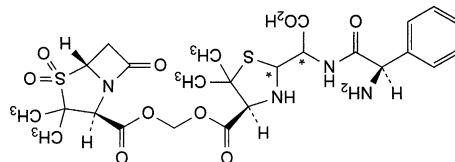
Specified impurities A, B, C, D, E, F, G



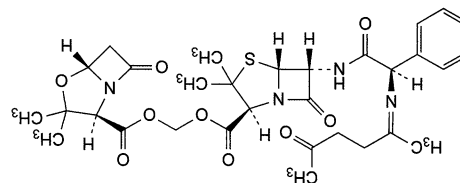
A. (2S,5R)-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid 4,4-dioxide (sulbactam),



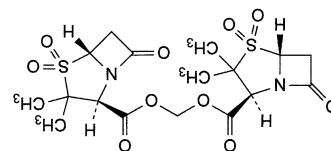
B. (2S,5R,6R)-6-[[[(2R)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (ampicillin),



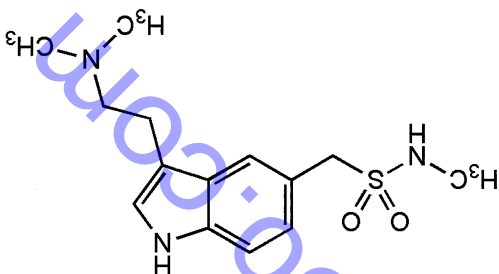
C. [(2R)-aminophenylacetyl]amino)-4-[[[(2S,5R)-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carboxyl]oxy]methoxy]carboxyl]-5,5-dimethylthiazolidin-2-yl]acetic acid (penicillotic acids of sulamcillin),



D. methylene bis[(2S,5R,6R)-3,3-dimethyl-6-[[[(2R)-[(1-methyl-4-oxopentylidene)amino]phenylacetyl]amino]-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate] (2S,5R)-3,3-dimethyl-7-oxo-4-oxa-1-azabicyclo[3.2.0]heptane-2-carboxylate,



E. methylene bis[(2S,5R)-3,3-dimethyl-4,4,7-trioxo-4λ⁶-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate] (sulbactam methylene ester),

Sumatriptan

$C_{14}H_{21}N_3O_2S$ 295.4 103628-46-2

DEFINITION

Sumatriptan is 3-(2-dimethylaminoethyl)indol-5-yl-N-

methylethanesulfonamide. It contains not less than 97.5% and not more than 102.0% of $C_{14}H_{21}N_3O_2S$, calculated with reference to the anhydrous substance.

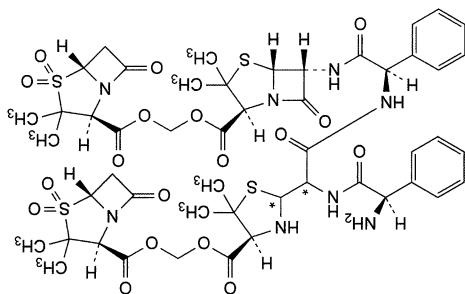
Preparation

Sumatriptan Nasal Spray

Action and use

Serotonin 5HT₁ receptor agonist; treatment of migraine.

F. methylene (2S,5R,6R)-6-[[[(2R)-2-amino-2-phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]hept-2-yl]carboxyl]-5,5-dimethylthiazolidin-2-yl]acetyl]amino]phenylacetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate (2S,5R)-3,3-dimethyl-4,4,7-trioxo-4λ⁶-thia-1-azabicyclo[3.2.0]heptane-2-carboxylate (ampicillin amide),



carboxylate (sulamcillin dimer).

CHARACTERISTICS

A white to pale yellow powder. Very slightly soluble in water.

IDENTIFICATION

The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of sumatriptan (RS 414).

TESTS

The total impurity content in the test for Impurities A and H and the test for Related substances is not greater than 1.5%.

Impurities A and H

Carry out the method for liquid chromatography,

Appendix III D. Prepare the solutions in a mixture

containing 3 volumes of 0.025M sodium dihydrogen

orthophosphate, the pH of which has been adjusted to 6.5, and

1 volume of acetonitrile (solution A).

(1) 0.2% w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 100 volumes.

(3) Dilute the contents of a vial of sumatriptan for system

suitability EPORS to 1 mL with 1M hydrochloric acid.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed

with silica gel for chromatography (5 µm) (Spherisorb silica

S5W is suitable).

(b) Use isocratic elution and the mobile phase described

below.

(c) Use a flow rate of 2.0 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 282 nm.

(f) Inject 20 µL of each solution.

(g) For solution (1) allow the chromatography to proceed for

5 times the retention time of the principal peak.

MOBILE PHASE

10 volumes of 10M ammonium acetate and 90 volumes of

methanol.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with

solution (3) resembles that supplied with sumatriptan for

impurity A and sumatriptan is at least 1.5.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to impurity A is not

greater than the area of the principal peak in the

chromatogram obtained with solution (2) (0.6% taking into

account the correction factor of 0.6 for impurity A);

the area of any peak due to impurity H is not greater than

0.3 times the area of the principal peak in the chromatogram

obtained with solution (2) (0.3%).

Related substances

Carry out the method for liquid chromatography,

Appendix III D. Prepare the solutions in a mixture

containing 3 volumes of 0.025M sodium dihydrogen

orthophosphate, the pH of which has been adjusted to 6.5, and

1 volume of acetonitrile (solution A).

(1) 0.2% w/v of the substance being examined.

(2) Dilute 1 volume of solution (1) to 100 volumes and

further dilute 1 volume of the resulting solution to

10 volumes.

(3) Dilute the contents of a vial of sumatriptan

impurity mixture EPORS to 1 mL with 0.1M hydrochloric acid.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed

with octadecylsilyl silica gel for chromatography (5 µm)

(Spherisorb ODS 1 is suitable).

(b) Use isocratic elution and the mobile phase described

below.

(c) Use a flow rate of 1.5 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 282 nm.

(f) Inject 20 µL of each solution.

MOBILE PHASE

Mix 25 volumes of acetonitrile with 75 volumes of a solution

containing 0.97 g of dibutylamine, 0.735 g of orthophosphoric

acid and 2.93 g of sodium dihydrogen orthophosphate in

750 mL water, adjust to pH 6.5 with 10M sodium hydroxide

and dilute to 1000 mL with water.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained

with solution (3), the resolution factor between the peaks due

to impurity C and sumatriptan is at least 1.5.

LIMITS

In the chromatogram obtained with solution (1):

the areas of any peaks corresponding to impurities B, C and

D are not greater than 5 times the area of the principal peak

in the chromatogram obtained with solution (2) (0.5% of

each);

the area of any peak corresponding to impurity E is not

greater than the area of the principal peak in the

chromatogram obtained with solution (2) (0.1%);

the area of any other secondary peak is not greater than the

area of the principal peak in the chromatogram obtained with

solution (2) (0.1%).

Disregard any peak with an area less than 0.5 times the area

of the principal peak in the chromatogram obtained with

solution (2) (0.05%).

Water

Not more than 1.0% w/w, Appendix IX C. Use 1.0 g.

ASSAY

Carry out the method for liquid chromatography,

Appendix III D. Prepare solutions (1) and (2) in a mixture

containing 3 volumes of 0.025M sodium dihydrogen

orthophosphate, the pH of which has been adjusted to 6.5, and

1 volume of acetonitrile (solution A).

(1) 0.01% w/v of the substance being examined.

(2) 0.014% w/v of sumatriptan succinate BPCS.

(3) Dilute the contents of a vial of sumatriptan

impurity mixture EPORS to 1 mL with 0.1M hydrochloric acid.

CHROMATOGRAPHIC CONDITIONS

The chromatographic conditions described under Related

substances may be used.

SYSTEM SUITABILITY

The Assay is not valid unless, in the chromatogram obtained

with solution (3), the resolution factor between the peaks due

to sumatriptan and impurity C is at least 1.5.

DETERMINATION OF CONTENT

Calculate the content of $C_{14}H_{21}N_3O_2S$ using the declared

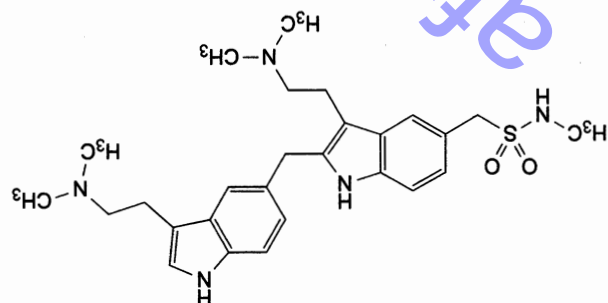
content of $C_{14}H_{21}N_3O_2S \cdot C_4H_6O_4$ in sumatriptan

succinate BPCS. Each 1 mg of $C_{14}H_{21}N_3O_2S$ is equivalent

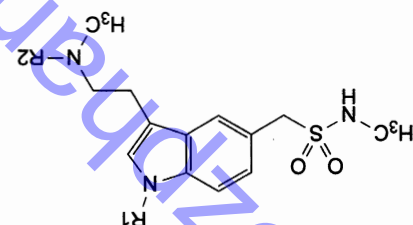
to 1.4 mg of $C_{14}H_{21}N_3O_2S \cdot C_4H_6O_4$.

IMPURITIES

Sumatriptan should be protected from light.

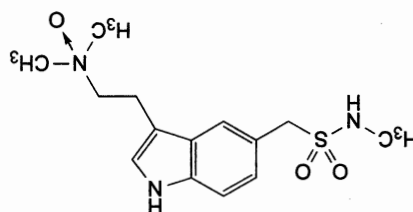
STORAGE

A. [3-[2-(dimethylamino)ethyl]-2-[[3-[2-(dimethylamino)ethyl]-1H-indol-5-yl]methyl]-1H-indol-5-yl]-N-methylmethanesulfonamide,

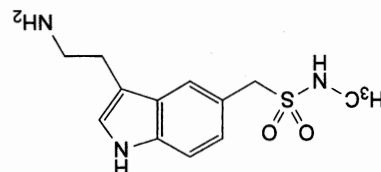


B. R1 = R2 = H: N-methyl[3-[2-(methylamino)ethyl]-1H-indol-5-yl]methanesulfonamide,

C. R1 = CH₂-OH, R2 = CH₂: [3-[2-(dimethylamino)ethyl]-1-(hydroxymethyl)-1H-indol-5-yl]-N-methylmethanesulfonamide,



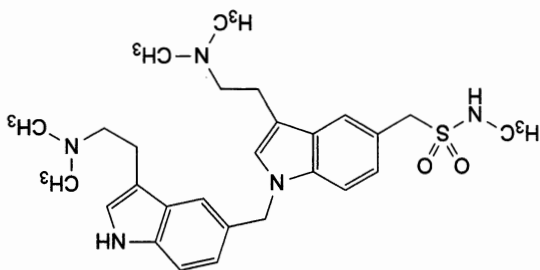
D. N,N'-dimethyl-2-[5-[(methylsulfonyl)methyl]-1H-indol-3-yl]ethanamine N-oxide,



E. [3-(2-aminoethyl)-1H-indol-5-yl]-N-methylmethanesulfonamide,

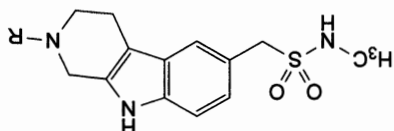
Sumatriptan Succinate

(Ph. Eur. monograph 1573)



F. R = H: N-methyl(2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-6-yl)methanesulfonamide,

G. R = CH₃: N-methyl(2-methyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-6-yl)methanesulfonamide,



H. [3-[2-(dimethylamino)ethyl]-1-[[3-[2-(dimethylamino)ethyl]-1H-indol-5-yl]methyl]-1H-indol-5-yl]-N-methylmethanesulfonamide.

103628-48-4

413.5

C₁₈H₂₇N₃O₆S

Action and use
Serotonin 5HT₁ receptor agonist; treatment of migraine.

Preparations
Sumatriptan Injection
Sumatriptan Tablets

Ph Eur

DEFINITION

[3-[2-(Dimethylamino)ethyl]-1H-indol-5-yl]-N-methylmethanesulfonamide hydrogen butanedioate.

CHARACTERS

Appearance
White or almost white powder.

Solubility

Freely soluble in water, sparingly soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison sumatriptan succinate CRS.

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 25.0 mL with the same solvent.

pH (2.2.3)

4.5 to 5.3.

Dilute 2.5 mL of solution S to 10 mL with carbon dioxide-free water R.

Absorbance (2.2.25)

Maximum 0.10, determined at 440 nm on solution S.

Impurities A and H

Liquid chromatography (2.2.29).

Test solution D Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of sumatriptan for system suitability CRS (containing impurities A and H) in the mobile phase and dilute to 1 mL with the mobile phase.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 282 nm.

Injection 20 µL.

Run time 5 times the retention time of sumatriptan.

Relative retention With reference to sumatriptan.

time = about 2 min; impurity A = about 1.8;

impurity H = about 2.6.

System suitability: reference solution (b):

the chromatogram obtained is similar to the chromatogram supplied with sumatriptan for system suitability CRS;

resolution: minimum 3.0 between the peaks due to impurities A and H.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity A by 0.6;

— impurity A: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);

— impurity H: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent).

Related substances

Liquid chromatography (2.2.29).

Solution A Dissolve 2.925 g of sodium dihydrogen phosphate R in 600 mL of water R, adjust to pH 6.5 with strong sodium hydroxide solution R, dilute to 750 mL with water R, add 250 mL of acetonitrile R and mix.

Test solution (a) Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Test solution (b) Dissolve 15.0 mg of the substance to be examined in solution A and dilute to 100.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of sumatriptan impurity mixture CRS (containing impurities B, C, D and E) in the mobile phase and dilute to 1 mL with the mobile phase.

Reference solution (c) Dissolve 15.0 mg of sumatriptan succinate CRS in solution A and dilute to 100.0 mL with solution A.

Column:

— size: $l = 0.25$ m, $\phi = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 25 volumes of acetonitrile R with 75 volumes of a solution prepared as follows: dissolve 0.970 g of dibutylamine R, 0.735 g of phosphoric acid R and 2.93 g of sodium dihydrogen phosphate R in 750 mL of water R, adjust to pH 6.5 with strong sodium hydroxide solution R and dilute to 1000 mL with water R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 282 nm.

Injection 10 µL of test solution (a) and reference solutions (a) and (b).

Run time 4 times the retention time of sumatriptan.

Identification of impurities Use the chromatogram obtained with reference solution (b) and the chromatogram supplied with sumatriptan impurity mixture CRS to identify the peaks due to impurities B, C, D and E.

Relative retention With reference to sumatriptan (retention time = about 7 min): impurity E = about 0.5;

impurity B = about 0.6; impurity D = about 0.7;

impurity C = about 0.8.

System suitability: reference solution (b):

resolution: minimum 1.5 between the peaks due to impurity C and sumatriptan;

the chromatogram shows 5 clearly separated peaks.

Limits:

— impurities B, C, D: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— impurity E: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.6 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (c).

Calculate the percentage content of $C_{18}H_{27}N_3O_6S$ from the declared content of *sunnampian succinate CRS*.

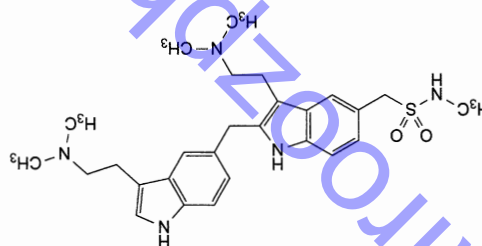
STORAGE

Protected from light.

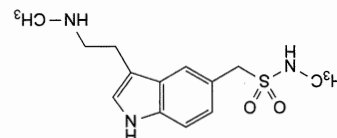
IMPURITIES

Specified impurities A, B, C, D, E, H

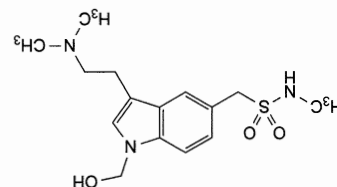
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: F, G.



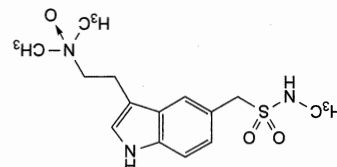
A. [3-[2-(dimethylamino)ethyl]-2-[[3-[2-(dimethylamino)ethyl]-1H-indol-5-yl]methyl]-1H-indol-5-yl]methanesulfonamide,



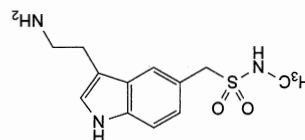
B. N-methyl[3-[2-(methylamino)ethyl]-1H-indol-5-yl]methanesulfonamide,



C. [3-[2-(dimethylamino)ethyl]-1-(hydroxymethyl)-1H-indol-5-yl]-N-methylmethanesulfonamide,



D. N,N-dimethyl-2-[5-[(methylsulfonyl)methyl]-1H-indol-3-yl]ethanamine N-oxide,



E. [3-(2-aminoethyl)-1H-indol-5-yl]-N-methylmethanesulfonamide,

DEFINITION

Fatty oil obtained from the seeds of *Helianthus annuus* L. by mechanical expression or by extraction. It is then refined. A suitable antioxidant may be added.

CHARACTERS

Appearance

Clear, light yellow liquid.

Solubility

Practically insoluble in water and in ethanol (96 per cent), miscible with light petroleum (bp: 40–60 °C).

Relative density

About 0.921.

Refractive index

About 1.474.

IDENTIFICATION

Identification of fatty oils by thin-layer chromatography

(2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

TESTS

Acid value (2.5.1)

Maximum 0.5, determined on 10.0 g.

Peroxide value (2.5.5, Method A)

Maximum 10.0.

Unaponifiable matter (2.5.7)

Maximum 1.5 per cent, determined on 5.0 g.

Refined Sunflower Oil

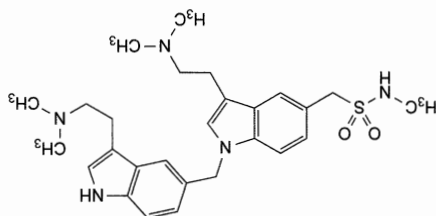
(Ph. Eur. monograph 1371)

Ph. Eur.

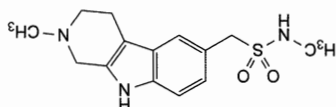


Ph. Eur.

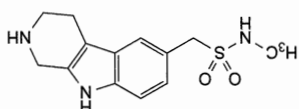
H. [3-[2-(dimethylamino)ethyl]-1-[[3-[2-(dimethylamino)ethyl]-1H-indol-5-yl]methyl]-1H-indol-5-yl]-N-methylmethanesulfonamide.



G. N-methyl(2-methyl-2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-6-yl)methanesulfonamide,



F. N-methyl(2,3,4,9-tetrahydro-1H-pyrido[3,4-b]indol-6-yl)methanesulfonamide,



Alkaline impurities (2.4.19)

It complies with the test.

Composition of fatty acids (2.4.22, Method A)

Use the mixture of calibrating substances in Table 2.4.22-3.

Composition of the fatty-acid fraction of the oil:

— *palmitic acid*: 4.0 per cent to 9.0 per cent,

— *stearic acid*: 1.0 per cent to 7.0 per cent,

— *oleic acid*: 14.0 per cent to 40.0 per cent,

— *linoleic acid*: 48.0 per cent to 74.0 per cent.

Water (2.5.32)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

In an airtight, well-filled container, protected from light.

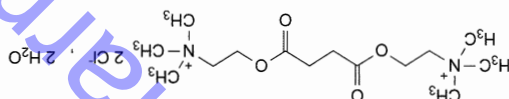
LABELLING

The label states whether the oil is obtained by mechanical

expression or by extraction.

Suxamethonium Chloride

(Ph. Eur. monograph 0248)



$C_{14}H_{30}Cl_2N_2O_4 \cdot 2H_2O$ 397.3 6101-15-1

Action and use
Depolarizing neuromuscular blocker.

Preparation

Suxamethonium Chloride Injection

DEFINITION

Suxamethonium chloride contains not less than 98.0 per cent and not more than the equivalent of 102.0 per cent of 2,2'-[butanedioylbis(oxy)]bis(*N,N,N*-trimethylethanaminium) dichloride, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, crystalline powder, hygroscopic, freely soluble in water, slightly soluble in alcohol. It melts at about 160 °C, determined without previous drying.

IDENTIFICATION

First identification A, D

Second identification B, C, D

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with *suxamethonium chloride CRS*. Examine the substances prepared as discs.

B. To 1 mL of solution S (see Tests) add 9 mL of water R, 10 mL of dilute sulfuric acid R and 30 mL of ammonium *reineckate solution R*. A pink precipitate is formed. Allow to stand for 30 min, filter, wash with water R, with alcohol R and then with ether R and dry at 80 °C. The melting point (2.2.14) of the precipitate is 180 °C to 185 °C.

C. Dissolve about 25 mg in 1 mL of water R and add 0.1 mL of a 10 g/L solution of cobalt chloride R and 0.1 mL

of potassium ferrocyanide solution R. A green colour is produced.

D. About 20 mg gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1). Dilute 4 mL of solution S to 10 mL with water R. The solution is colourless (2.2.2, Method II).

pH (2.2.3)

Dilute 1 mL of solution S to 10 mL with carbon dioxide-free water R. The pH of the solution is 4.0 to 5.0.

Choline chloride

Examine by thin-layer chromatography (2.2.27), using

cellulose for chromatography R1 as the coating substance.

Test solution Dissolve 0.4 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 0.4 g of *suxamethonium chloride CRS* and 2 mg of choline chloride R in methanol R and

dilute to 10 mL with the same solvent.

Apply to the plate 5 µL of each solution. Prepare the mobile phase as follows: shake together for 10 min, 10 volumes of anhydrous formic acid R, 40 volumes of water R and

50 volumes of butanol R; allow to stand and use the upper layer. Develop over a path of 15 cm. Dry the plate in a

current of air and spray with potassium iodobismuthate

solution R. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more

intense than the spot corresponding to choline chloride in the

chromatogram obtained with the reference solution

(0.5 per cent). The test is not valid unless the chromatogram

obtained with the reference solution shows two clearly

separated spots.

Water (2.5.12)

8.0 per cent to 10.0 per cent, determined on 0.30 g by the

semi-micro determination of water.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 50 mL of acetic anhydride R. Titrate with

0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 18.07 mg of

$C_{14}H_{30}Cl_2N_2O_4$.

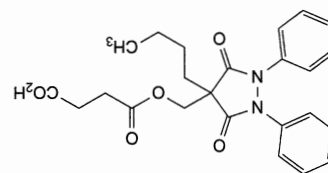
STORAGE

Store in an airtight container, protected from light.

Ph Eur

Suxibuzone

(Ph. Eur. monograph 1574)

 $C_{24}H_{26}N_2O_6$

438.5

27470-51-5

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph. Eur.

DEFINITION

4-[(4-butyl-3,5-dioxo-1,2-diphenylpyrazolidin-4-yl)methoxy]-4-oxobutanoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone, soluble in ethanol (96 per cent), practically insoluble in cyclohexane.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison suxibuzone CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 1 g in anhydrous ethanol R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.10 g of the substance to be examined in acetonitrile R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 2.8 mg of phenylbutazone CRS (impurity A), 2.8 mg of suxibuzone impurity B CRS and 2.8 mg of suxibuzone impurity C CRS in acetonitrile R and dilute to 10.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with acetonitrile R.

Reference solution (b) Dissolve 4 mg of phenylbutazone CRS (impurity A) in acetonitrile R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with acetonitrile R.

Reference solution (c) Dissolve 10 mg of phenylbutazone CRS (impurity A) in acetonitrile R and dilute to 25.0 mL with the same solvent. Mix 10.0 mL of this solution with 1.0 mL of the test solution and dilute the mixture to 25.0 mL with acetonitrile R.

Column:

acetonitrile R.

Reference solution (c) Dissolve 10 mg of phenylbutazone CRS (impurity A) in acetonitrile R and dilute to 25.0 mL with the same solvent. Mix 10.0 mL of this solution with 1.0 mL of the test solution and dilute the mixture to 25.0 mL with acetonitrile R.

size: $l = 0.125$ m, $\phi = 4.0$ mm;(5 μ m).

Mobile phase Mix 44 volumes of acetonitrile R and 56 volumes of a solution prepared as follows: dissolve 6.7 g of citric acid in 100 mL of water R, adjust to pH 3.0 with citric acid R and dilute to 1000 mL with water R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 250 nm.

Injection 10 μ L.

Relative retention With reference to suxibuzone (retention time = about 7 min): impurity C = 0.7; impurity A = 1.4; impurity B = 3.3.

System suitability: reference solution (c):

— resolution: minimum of 2.0 between the peaks due to suxibuzone and impurity A.

Limits:

— impurity A: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.7 per cent);

— impurity B: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.7 per cent);

— impurity C: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.7 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— disregard limit: 0.1 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (a) (0.07 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven *in vacuo* at 60 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in previously neutralised anhydrous ethanol R and dilute to 10 mL with the same solvent. Carry out a potentiometric titration (2.2.20) using 0.1 M sodium hydroxide.

1 mL of 0.1 M sodium hydroxide is equivalent to 43.85 mg of $C_{24}H_{26}N_2O_6$.

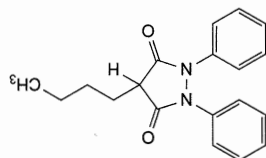
STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C

A. 4-butyl-1,2-diphenylpyrazolidine-3,5-dione (phenylbutazone),



TESTS

Impurities A, B and C

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R1, hexane R, 2-propanol R1 (20:40:40 V/V/V).

Solution A Dissolve 27 g of *tetrabutylammonium hydroxide* R in *methanol* R and dilute to 100.0 mL with the same solvent.

Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (a) Dissolve 25.0 mg of *tadalafil* CRS in the solvent mixture and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) In order to prepare impurity A *in situ*, dissolve 25 mg of the substance to be examined in 40 mL of the solvent mixture. Add 1 mL of solution A, mix well and allow to stand for 20 min. Add 1 mL of *trifluoroacetic acid* R and dilute to 100.0 mL with the solvent mixture.

Reference solution (d) To 1.0 mL of the test solution add 1.0 mL of reference solution (c) and dilute to 50.0 mL with the solvent mixture.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: silica gel AD for chiral separation R (10 μ m);
— temperature: 30 °C.

Mobile phase hexane R, 2-propanol R1 (50:50 V/V).
Flow rate 0.75 mL/min.

Detection Spectrophotometer at 222 nm.
Injection 20 μ L of the test solution and reference solutions (b) and (d).

Run time 2.2 times the retention time of *tadalafil*.
Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peak due to impurity A.

Relative retention With reference to *tadalafil* (retention time = about 11 min): impurity A = about 0.8.

System suitability Reference solution (d)
— resolution: minimum 2.0 between the peaks due to impurity A and *tadalafil*.

Limits:
— impurity A: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent).

Related substances

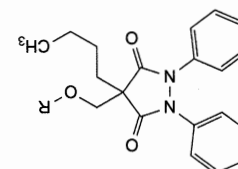
Liquid chromatography (2.2.29). Do not use sonication during the preparation of the solutions.

Solvent mixture acetonitrile R, 2-propanol R (50:50 V/V).

Solution A Dissolve 27 g of *tetrabutylammonium hydroxide* R in *methanol* R and dilute to 100.0 mL with the same solvent.

Test solution (a) Dissolve 40 mg of the substance to be examined in 50 mL of *acetonitrile* R and dilute to 100.0 mL with mobile phase A.

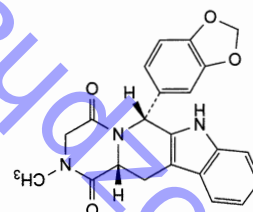
Test solution (b) Dissolve 50.0 mg of the substance to be examined in 50 mL of *acetonitrile* R and dilute to 100.0 mL with mobile phase A. To 10.0 mL of this solution add



B. R = CO-CH₂-CH₂-CO-O-CH₂-CH₃; (4-butyl-3,5-dioxo-1,2-diphenylpyrazolidin-4-yl)methyl ethyl butanedioate,
C. R = H; 4-butyl-4-(hydroxymethyl)-1,2-diphenyl-1,2-dihydro-4H-pyrazole-3,5-dione.

Ph Eur

Tadalafil
(Ph Eur monograph 2606)



C₂₂H₁₉N₃O₄ 389.4 171596-29-5

Action and use
Selective inhibitor of cyclic GMP-specific phosphodiesterase (Type V) with vasodilator action; treatment of erectile dysfunction.

Ph Eur

DEFINITION

(6R,12aR)-6-(1,3-Benzodioxol-5-yl)-2-methyl-2,3,6,7,12,12a-hexahydropyrazino[1',2':1,6]-pyrido[3,4-b]indole-1,4-dione.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in dimethyl sulfoxide, slightly soluble in methylene chloride.

IDENTIFICATION

Carry out either tests A, B or tests A, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison *tadalafil* CRS.

B. Liquid chromatography (2.2.29) as described in the test for impurities A, B and C with the following modification.

Injection Test solution and reference solution (a).

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).

C. Specific optical rotation (2.2.7): + 78.0 to + 84.0 (dried substance).

Dissolve 0.250 g in *dimethyl sulfoxide* R and dilute to 25.0 mL with the same solvent.

25.0 mL of acetonitrile R and dilute to 50.0 mL with mobile phase A.

Reference solution (a) To 1.0 mL of test solution (a) add 50 mL of acetonitrile R and dilute to 100.0 mL with mobile phase A. To 1.0 mL of this solution add 5 mL of

acetonitrile R and dilute to 10.0 mL with mobile phase A.

Reference solution (b) In order to prepare impurity A *in situ*,

dissolve 4.0 mg of the substance to be examined in 50 mL of the solvent mixture. Add 1 mL of solution A, mix, and allow

to stand for 40 min. Add 1 mL of trifluoroacetic acid R and

dilute to 100.0 mL with the solvent mixture.

Reference solution (c) Dilute 1 mL of reference solution (b) to

50.0 mL with test solution (a).

Reference solution (d) Dissolve 50.0 mg of tadalafil CRS in

50 mL of acetonitrile R and dilute to 100.0 mL with mobile

phase A. To 10.0 mL of this solution add 25.0 mL of

acetonitrile R and dilute to 50.0 mL with mobile phase A.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R

(5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: mix 1.0 mL of trifluoroacetic acid R with

water R and dilute to 1000 mL with the same solvent;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	85	15
3 - 30	85 \rightarrow 5	15 \rightarrow 95
30 - 33	5	95

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 285 nm.

Injection 20 μ L of test solution (a) and reference solutions (a)

and (c).

Identification of impurities Use the chromatogram obtained

with reference solution (c) to identify the peak due to

impurities A + C.

Relative retention With reference to tadalafil (retention

time = about 16 min): impurities A and C = about 1.03.

System suitability: reference solution (c):

— peak-to-valley ratio: minimum 3.3, where H_p = height

above the baseline of the peak due to impurities A + C

and H_v = height above the baseline of the lowest point of

the curve separating this peak from the peak due to

tadalafil.

Limits:

— **unspecified impurities:** for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— **total:** not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.3 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

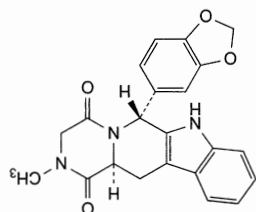
(0.05 per cent); disregard any peak due to impurity A

and/or C.

Loss on drying (2.2.32)

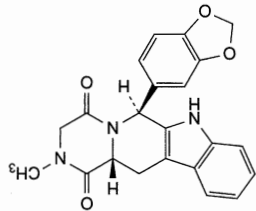
Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo at 105 °C for 3 h.



A. (6*R*,12*aS*)-6-(1,3-benzodioxol-5-yl)-2-methyl-2,3,6,7,12,12a-hexahydropyrazino[1',2':1,6]pyrido[3,4-*b*]indole-1,4-dione,

B. (6*S*,12*aS*)-6-(1,3-benzodioxol-5-yl)-2-methyl-2,3,6,7,12,12a-hexahydropyrazino[1',2':1,6]pyrido[3,4-*b*]indole-1,4-dione,



C. (6*S*,12*aR*)-6-(1,3-benzodioxol-5-yl)-2-methyl-2,3,6,7,12,12a-hexahydropyrazino[1',2':1,6]pyrido[3,4-*b*]indole-1,4-dione,

D, E, F, G, H, I.

Control of impurities in substances for pharmaceutical use: B, C, impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph Substances for pharmaceutical use

acceptance criterion for other/unspecified impurities and/or

the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities A

IMPURITIES

declared content of tadalafil CRS.

Calculate the percentage content of $C_{22}H_{19}N_3O_4$ from the

time = about 4.5 min).

Run time Twice the retention time of tadalafil (retention

Injection Test solution (b) and reference solution (d).

Flow rate 1.5 mL/min.

Mobile phase acetonitrile R, mobile phase A (45:55 V/V).

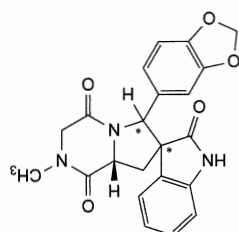
related substances with the following modifications.

Liquid chromatography (2.2.29) as described in the test for

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)



I. (8a'R)-6'-(1,3-benzodioxol-5-yl)-2'-methyl-2',3',8',8a'-tetrahydro-6'H-spiro[indole-3,7'-pyrrolo[1,2-a]pyrazine]-1',2,4'(1H)-trione.

Ph Eur



14807-96-6

(Talc, Ph Eur monograph 0438)

Purified Talc

Preparation

Ph Eur

DEFINITION

Powdered, selected, natural, hydrated magnesium silicate. Pure talc has the formula $Mg_3Si_4O_{10}(OH)_2$ (379.3). It may contain variable amounts of associated minerals among which chlorites (hydrated aluminum and magnesium silicates), magnesite (magnesium carbonate), calcite (calcium carbonate) and dolomite (calcium and magnesium carbonate) are predominant.

PRODUCTION

Talc derived from deposits that are known to contain associated asbestos is not suitable for pharmaceutical use. The manufacturer is responsible for demonstrating by the test for amphiboles and serpentine that the product is free from asbestos. The presence of amphiboles and of serpentine is revealed by X-ray diffraction or by infrared spectrophotometry (see A and B). If detected, the specific morphological criteria of asbestos are investigated by a suitable method of optical microscopy to determine whether tremolite asbestos or chrysotile is present, as described below.

A. Infrared absorption spectrophotometry (2.2.24).

In the range 740 cm^{-1} to 760 cm^{-1} using scale expansion, any absorption band at $758 \pm 1\text{ cm}^{-1}$ may indicate the presence of tremolite or of chrysotile. If the absorption band remains after ignition of the substance to be examined at $850 \pm 50^\circ\text{C}$ for at least 30 min, it indicates the presence of the tremolite. In the range 600 cm^{-1} to 650 cm^{-1} using scale expansion, any absorption band or shoulder may indicate the presence of serpentine.

B. X-ray diffraction.

Preparation Place the sample on the sample holder; pack and smooth its surface with a polished glass microscope slide.

Radiation Cu K α monochromatic, 40 kV, 24-30 mA.

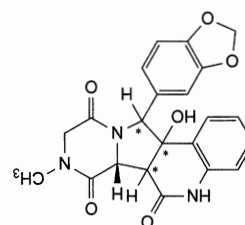
Incident slit 1° .

Detection slit 0.2° .

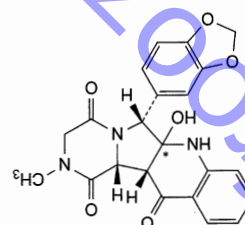
Goniometer speed $1/10^\circ$ $2\theta/\text{min}$.

Scanning range $10\text{--}13^\circ$ 2θ and $24\text{--}26^\circ$ 2θ .

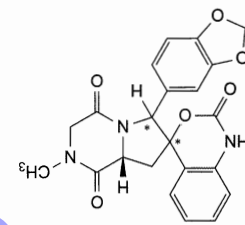
Sample Not oriented.



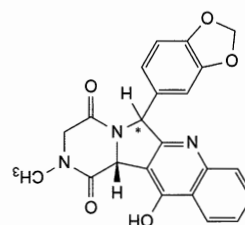
D. (6b'R)-12-(1,3-benzodioxol-5-yl)-12a-hydroxy-8-methyl-6a,6b,8,9,12,12a-hexahydropyrazino[1',2':1,2]pyrrolo[3,4-b]quinoline-6,7,10(5H)-trione.



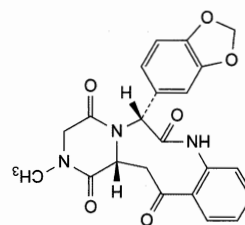
E. (6R,12aR,12bR)-6-(1,3-benzodioxol-5-yl)-6a-hydroxy-2-methyl-2,3,6a,7,12a,12b-hexahydropyrazino[1',2':1,5]pyrrolo[3,4-b]quinoline-1,4,12(6H)-trione.



F. (8a'R)-6'-(1,3-benzodioxol-5-yl)-2'-methyl-2',3',8',8a'-tetrahydro-6'H-spiro[3,1-benzoxazine-4,7'-pyrrolo[1,2-a]pyrazine]-1',2,4'(1H)-trione.



G. (12b'R)-6-(1,3-benzodioxol-5-yl)-12-hydroxy-2-methyl-2,3,6,12b-tetrahydropyrazino[1',2':1,5]pyrrolo[3,4-b]quinoline-1,4-dione.



H. (6R,14aR)-6-(1,3-benzodioxol-5-yl)-2-methyl-2,3,14,14a-tetrahydropyrazino[1',2'-d][1,4]benzodiazonine-1,4,7,13(6H,8H)-trione.

Results The presence of amphiboles is detected by a diffraction peak at $10.5 \pm 0.1^\circ 2\theta$, the presence of serpentine is detected by diffraction peaks at $24.3 \pm 0.1^\circ 2\theta$ and at $12.1 \pm 0.1^\circ 2\theta$.

If, by one of the 2 methods, amphiboles and/or serpentine are detected, examine by a suitable method of optical microscopy to determine the asbestos character.

The presence of asbestos is shown if the following 2 criteria are met:

— a range of length to width ratios of 20:1 to 100:1, or higher for fibres longer than $5 \mu\text{m}$;

— capability of splitting into very thin fibrils;

and if at least 2 of the following 4 criteria are met:

— parallel fibres occurring in bundles;

— fibre bundles displaying frayed ends;

— fibres in the form of thin needles;

— matted masses of individual fibres and/or fibres showing curvature.

CHARACTERS

Appearance

Light, homogeneous, white or almost white powder, greasy to the touch (non abrasive).

Solubility

Practically insoluble in water, in ethanol (96 per cent) and in dilute solutions of acids and alkali hydroxides.

IDENTIFICATION

First identification A.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs of potassium bromide R.

Absorption bands At $3677 \pm 2 \text{ cm}^{-1}$, $1018 \pm 2 \text{ cm}^{-1}$ and $669 \pm 2 \text{ cm}^{-1}$.

B. In a platinum crucible, melt a mixture of 0.2 g of anhydrous sodium carbonate R and 2.0 g of potassium carbonate R. To the melted mass add 0.1 g of the substance to be examined and heat until the mixture is completely melted. Allow to cool and transfer the melted mass into an evaporating dish with 50 mL of hot water R. Add hydrochloric acid R until effervescence ceases. Add 10 mL of hydrochloric acid R and evaporate to dryness on a water-bath. Allow to cool. Add 20 mL of water R, heat to boiling and filter (the residue is used for identification test C). To 5 mL of the filtrate add 1 mL of ammonia R and 1 mL of ammonium chloride solution R and filter. To the filtrate add 1 mL of disodium hydrogen phosphate solution R. A white, crystalline precipitate is formed.

reaction of silicates (2.3.1).

TESTS

Solution S1

Weigh 10.0 g into a conical flask fitted with a reflux condenser, gradually add 50 mL of 0.5 M hydrochloric acid while stirring and heat on a water-bath for 30 min. Allow to cool. Transfer the mixture to a beaker and allow the undissolved material to settle. Filter the supernatant through medium-speed filter paper into a 100 mL volumetric flask, retaining as much as possible of the insoluble material in the beaker. Wash the residue and the beaker with 3 quantities, each of 10 mL, of hot water R. Wash the filter with 15 mL of hot water R, allow the filtrate to cool and dilute to 100.0 mL with the same solvent.

Solution S2

Perchlorates mixed with heavy metals are known to be explosive. Take proper precautions while performing this procedure. Weigh 0.5 g in a 100 mL polytetrafluoroethylene dish, add 5 mL of hydrochloric acid R, 5 mL of lead-free nitric acid R and 5 mL of perchloric acid R. Stir gently then add 35 mL of hydrofluoric acid R and evaporate slowly to dryness on a hot plate. To the residue, add 5 mL of hydrochloric acid R, cover with a watch-glass, heat to boiling and allow to cool. Rinse the watch-glass and the dish with water R. Transfer into a volumetric flask, rinse the dish with water R and dilute to 50.0 mL with the same solvent.

Acidity or alkalinity

Boil 2.5 g with 50 mL of carbon dioxide-free water R under reflux. Filter *in vacuo*. To 10 mL of the filtrate add 0.1 mL of bromothymol blue solution R1; not more than 0.4 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to green. To 10 mL of the filtrate add 0.1 mL of phenolphthalein solution R1; not more than 0.3 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to pink.

Water-soluble substances

Maximum 0.2 per cent.
To 10.0 g add 50 mL of carbon dioxide-free water R, heat to boiling and maintain boiling under a reflux condenser for 30 min. Allow to cool, filter through a medium-speed filter paper and dilute to 50.0 mL with carbon dioxide-free water R. Take 25.0 mL of the filtrate, evaporate to dryness and heat at 105°C for 1 h. The residue weighs a maximum of 10 mg.
Aluminum 2.0 per cent.
Maximum 2.0 per cent.
Atomic absorption spectrometry (2.2.23, Method I).

Test solution To 5.0 mL of solution S2 add 10 mL of a 25.34 g/L solution of caesium chloride R, 10.0 mL of hydrochloric acid R and dilute to 100.0 mL with water R.
Reference solutions Into 4 identical volumetric flasks, each containing 10.0 mL of hydrochloric acid R and 10 mL of a 25.34 g/L solution of caesium chloride R, introduce respectively 5.0 mL, 10.0 mL, 15.0 mL and 20.0 mL of aluminum standard solution (100 ppm Al) R and dilute to 100.0 mL with water R.
Source Aluminum hollow-cathode lamp.
Wavelength 309.3 nm.

Atomisation device Nitrous oxide-acetylene flame.

Calcium

Maximum 0.9 per cent.
Atomic absorption spectrometry (2.2.23, Method I).
Test solution To 5.0 mL of solution S2 add 10.0 mL of hydrochloric acid R, 10 mL of lanthanum chloride solution R and dilute to 100.0 mL with water R.
Reference solutions Into 4 identical volumetric flasks, each containing 10.0 mL of hydrochloric acid R and 10 mL of lanthanum chloride solution R, introduce respectively 1.0 mL, 2.0 mL, 3.0 mL and 5.0 mL of calcium standard solution (100 ppm Ca) R1 and dilute to 100.0 mL with water R.
Source Calcium hollow-cathode lamp.
Wavelength 422.7 nm.

Atomisation device Nitrous oxide-acetylene flame.

Iron

Maximum 0.25 per cent.
Atomic absorption spectrometry (2.2.23, Method I).

of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for a particular characteristic are also used. Wherever results for a particular characteristic are reported, the control method must be indicated. The following characteristics may be relevant for use as an antiadhesive agent or glidant in tablets and capsules or as antistatic agent in coated and film-coated tablets.

Particle-size distribution (2.9.31)

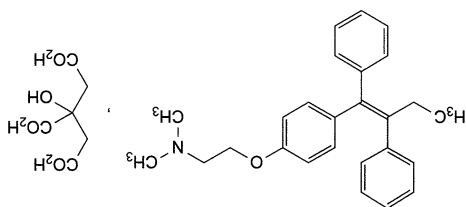
Specific surface area (2.9.26)

Ph Eur



Tamoxifen Citrate

(Ph. Eur. monograph 1046)



C₃₂H₃₇NO₈ 563.6 54965-24-1

Action and use
Selective estrogen receptor modulator.

Preparation
Tamoxifen Tablets

DEFINITION

2-[4-[(Z)-1,2-Diphenylbut-1-enyl]phenoxy]-N,N-dimethylethanamine dihydrogen 2-hydroxypropane-1,2,3-tricarboxylate.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, soluble in methanol, slightly soluble in acetone.

It shows polymorphism (5.9).

IDENTIFICATION

First identification B.

Second identification A, C.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 20 mg in methanol R and dilute to 50.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with methanol R.

Spectral range 220-350 nm.

Absorption maxima At 237 nm and 275 nm.

Absorption ratio $A_{237}/A_{275} = 1.45$ to 1.65.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison tamoxifen citrate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference

Test solution To 2.5 mL of solution S1, add 50.0 mL of 0.5 M hydrochloric acid and dilute to 100.0 mL with water R.

Reference solutions Into 4 identical volumetric flasks, each containing 50.0 mL of 0.5 M hydrochloric acid, introduce respectively 2.0 mL, 2.5 mL, 3.0 mL and 4.0 mL of iron standard solution (250 ppm Fe) R and dilute to 100.0 mL with water R.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Correction Deuterium lamp.

Lead

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method D).

Test solution Use solution S1.

Reference solutions Into 4 identical volumetric flasks, each containing 50.0 mL of 0.5 M hydrochloric acid, introduce respectively 5.0 mL, 7.5 mL, 10.0 mL and 12.5 mL of lead standard solution (10 ppm Pb) R1 and dilute to 100.0 mL with water R.

Source Lead hollow-cathode lamp.

Wavelength 217.0 nm.

Atomisation device Air-acetylene flame.

Magnesium

17.0 per cent to 19.5 per cent.

Atomic absorption spectrometry (2.2.23, Method D).

Test solution Dilute 0.5 mL of solution S2 to 100.0 mL with water R. To 4.0 mL of the solution, add 10.0 mL of hydrochloric acid R, 10 mL of lanthanum chloride solution R and dilute to 100.0 mL with water R.

Reference solutions Into 4 identical volumetric flasks, each containing 10.0 mL of hydrochloric acid R and 10 mL of lanthanum chloride solution R, introduce respectively 2.5 mL, 3.0 mL, 4.0 mL and 5.0 mL of magnesium standard solution (10 ppm Mg) R1 and dilute to 100.0 mL with water R.

Source Magnesium hollow-cathode lamp.

Wavelength 285.2 nm.

Atomisation device Air-acetylene flame.

Loss on ignition

Maximum 7.0 per cent, determined on 1.00 g by ignition to constant weight at 1050-1100 °C.

Microbial contamination

If intended for cutaneous administration:

— TAMC: acceptance criterion 10² CFU/g (2.6.12).

If intended for oral administration:

— TAMC: acceptance criterion 10³ CFU/g (2.6.12);

— TYMC: acceptance criterion 10² CFU/g (2.6.12).

LABELLING

The label states, where applicable, that the substance is suitable for oral or cutaneous administration.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency

substance separately in acetone R₃ evaporate to dryness and

record new spectra using the residues.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of tamoxifen citrate CRS in methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of clomifene citrate CRS and 10 mg of tamoxifen citrate CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase methylamine R, toluene R (10:90 V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference

solution (a).

TESTS

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use and protect from light.

Test solution Dissolve 15 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

phase.

Reference solution (a) Dissolve 3 mg of tamoxifen citrate for performance test CRS (containing impurities A and F) in the mobile phase and dilute to 2.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 40 volumes of acetonitrile R and 60 volumes of water R containing 0.9 g/L of sodium dihydrogen phosphate R and 4.8 g/L of *N,N*-dimethylethylamine R₃ adjust to pH 3.0 with phosphoric acid R.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time Twice the retention time of tamoxifen.

Identification of impurities Use the chromatogram supplied with tamoxifen citrate for performance test CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and F.

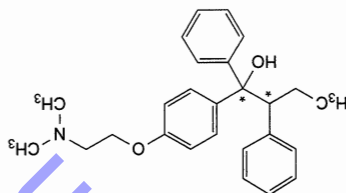
Relative retention With reference to tamoxifen (retention time = about 20 min): impurity A = about 0.8;

impurity F = about 0.9.

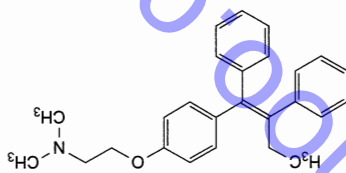
System suitability: reference solution (a):

— baseline separation between the peaks due to impurity F and tamoxifen;

— resolution: minimum 3.0 between the peaks due to impurities A and F.



A. 2-[4-[(E)-1,2-diphenylbut-1-en-1-yl]phenoxy]-N,N-dimethylethanamine ((E)-isomer)



B. 1-[4-[(2-(dimethylamino)ethoxy]phenyl]-1,2-diphenylbutan-1-ol

D, E, G, H.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C,

IMPURITIES

Specified impurities A, F

1 mL of 0.1 M perchloric acid is equivalent to 56.36 mg of C₂₂H₂₇NO₈.

solution R as indicator.

Dissolve 0.400 g in 75 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid using 0.1 mL of naphtholbenzene

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in vacuo at 65 °C for 4 h.

Loss on drying (2.2.32)

Maximum 0.5 per cent; disregard any peak due to the citrate.

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— impurity F: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— impurity A: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

— impurity F: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

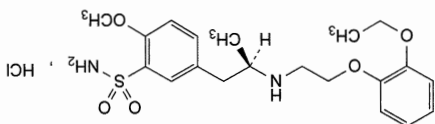
— impurity A: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.3 per cent);

Limits:



Tamsulosin Hydrochloride

(Ph. Eur. monograph 2131)



$C_{20}H_{29}ClN_2O_5$ 445.0 106463-17-6

Action and use

Alpha-1-adrenoceptor antagonist.

Preparations

Prolonged-release Tamsulosin Capsules

Prolonged-release Tamsulosin Tablets

DEFINITION

5-[(2*R*)-2-[(2-ethoxyphenoxy)ethyl]amino]propyl]-2-methoxybenzenesulfonamide hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Slightly soluble in water, freely soluble in formic acid, slightly soluble in anhydrous ethanol.

mp

About 230 °C.

IDENTIFICATION

Carry out either tests A, C, D or tests A, B, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tamsulosin hydrochloride CRS.

B. Specific optical rotation (2.2.7): -20.5 to -17.5 (dried substance).

Dissolve with heating 0.15 g in water *R* and dilute to 20.0 mL with the same solvent.

C. Enantiomeric purity (see Tests).

D. Dissolve with heating 0.75 g in water *R* and dilute to 100.0 mL with the same solvent. Take 5 mL of the solution and cool in an ice-bath. Add 3 mL of dilute nitric acid *R* and shake. Allow to stand at room temperature for 30 min and filter. The filtrate gives reaction (a) of chlorides (2.3.1).

TESTS

Related substances

A. Impurities eluting before tamsulosin. Liquid chromatography (2.2.29).

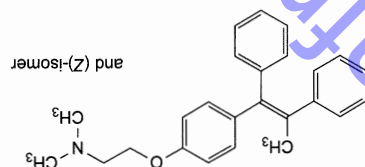
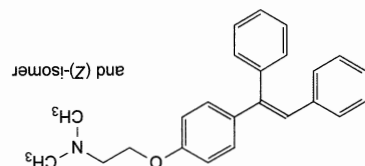
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

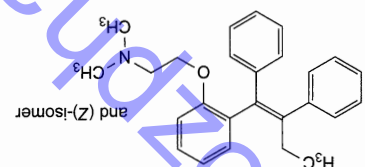
Reference solution (b) Dissolve 4 mg of tamsulosin impurity *D* CRS and 4 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 4 mg of tamsulosin impurity *H* CRS and 4 mg of the substance to be examined in the mobile phase.

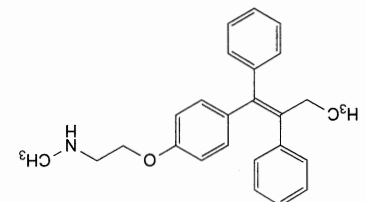
C. 2-[4-[(*EZ*)-1,2-diphenylethenyl]phenoxy]-*N,N*-dimethylethanamine,



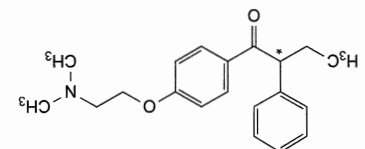
D. 2-[4-[(*EZ*)-1,2-diphenylprop-1-enyl]phenoxy]-*N,N*-dimethylethanamine,



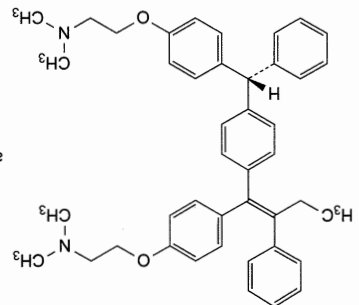
E. 2-[2-[(*EZ*)-1,2-diphenylbut-1-enyl]phenoxy]-*N,N*-dimethylethanamine,



F. 2-[4-[(*Z*)-1,2-diphenylbut-1-enyl]phenoxy]-*N,N*-dimethylethanamine,



G. (2*RS*)-1-[4-[2-(dimethylamino)ethoxy]phenyl]-2-phenylbutan-1-one,



H. 2-[4-[(*RS*)-1-[4-[(2*Z*)-1-[4-2-(dimethylamino)ethoxy]phenyl]phenyl]phenoxy]-*N,N*-dimethylethanamine.

in the mobile phase and dilute to 20.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 20.0 mL with the mobile phase.

Column:

- size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
- stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m);
- temperature: 40 °C.

Mobile phase Dissolve 3.0 g of sodium hydroxide R in a mixture of 8.7 mL of perchloric acid R and 1.9 L of water R; adjust to pH 2.0 with 0.5 M sodium hydroxide and dilute to 2 L with water R; to 1.4 L of this solution, add 600 mL of acetonitrile R.

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 μ L of the test solution and reference solutions (a) and (b).

Run time 1.5 times the retention time of tamsulosin

(retention time = about 6 min).

System suitability: reference solution (b):

- **resolution:** minimum 6 between the peaks due to impurity D and tamsulosin.

Limits:

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.10 per cent); **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

B. Impurities eluting after tamsulosin. Liquid chromatography (2.2.29) as described in test A with the following modifications.

Mobile phase Dissolve 3.0 g of sodium hydroxide R in a mixture of 8.7 mL of perchloric acid R and 1.9 L of water R; adjust to pH 2.0 with 0.5 M sodium hydroxide and dilute to 2 L with water R; add 2 L of acetonitrile R.

Flow rate 1.0 mL/min.

Injection 10 μ L of the test solution and reference solutions (a) and (c).

Run time 5 times the retention time of tamsulosin (retention time = about 2.5 min).

System suitability: reference solution (c):

- **resolution:** minimum 2 between the peaks due to tamsulosin and impurity H.

Limits:

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.10 per cent); **sum of impurities eluting before tamsulosin in test A and after tamsulosin in test B:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Enantiomeric purity

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in methanol R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (b) Dissolve 5.0 mg of tamsulosin racemate CRS in methanol R and dilute to 25.0 mL with the same solvent. Dilute 2.0 mL of this solution to 10.0 mL with methanol R.

Column:

- size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
- stationary phase: silica gel AD for chiral separation R;
- temperature: 40 °C.

Mobile phase diethylamine R, methanol R, anhydrous ethanol R, hexane R (1:150:200:650 V/V/V/V).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 μ L.

Relative retention With reference to tamsulosin (retention time = about 14 min): impurity G = about 0.8.

System suitability: reference solution (b):

- **resolution:** minimum 2 between the peaks due to impurity G and tamsulosin.

Limit:

— **impurity G:** not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

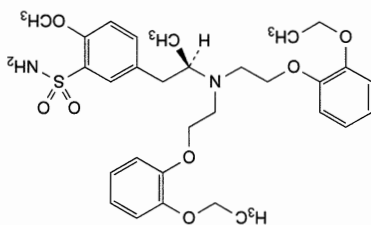
ASSAY

Dissolve 0.350 g in 5.0 mL of anhydrous formic acid R, add 75 mL of a mixture of 2 volumes of acetic anhydride R and 3 volumes of glacial acetic acid R. Titrate immediately with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration. 1 mL of 0.1 M perchloric acid is equivalent to 44.50 mg of $C_{20}H_{29}ClN_2O_5$.

IMPURITIES

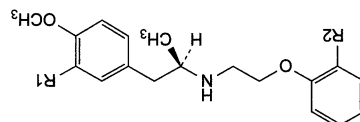
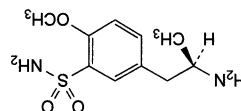
Specified impurities G

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, H, I.

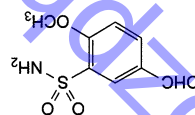


A. 5-[(2R)-2-[[bis[2-(2-ethoxyphenoxy)ethyl]amino]propyl]-2-methoxybenzenesulfonamide,

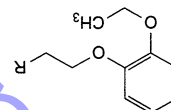
B. 5-[(2*R*)-2-amino]propyl]-2-methoxybenzenesulfonamide,



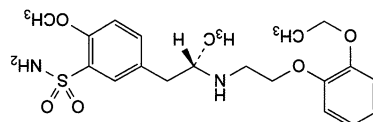
C. $R_1 = SO_2NH_2$, $R_2 = H$: 2-methoxy-5-[(2*R*)-2-[(2-phenoxyethyl)amino]propyl]benzenesulfonamide, $D. R_1 = SO_2NH_2$, $R_2 = OCH_3$: 2-methoxy-5-[(2*R*)-2-[(2-methoxyphenoxy)ethyl]amino]propyl]benzenesulfonamide, $H. R_1 = H$, $R_2 = OC_2H_5$: (2*R*)-*N*-(2-(2-ethoxyphenoxy)ethyl)-1-(4-methoxyphenyl)propan-2-amine,



E. 5-formyl-2-methoxybenzenesulfonamide,



F. $R = NH_2$: 2-(2-ethoxyphenoxy)ethanamine, I. $R = Br$: 1-(2-bromoethoxy)-2-ethoxybenzene,



G. 5-[(2*S*)-2-[(2-ethoxyphenoxy)ethyl]amino]propyl]-2-methoxybenzenesulfonamide.

Ph Eur

Tannic Acid

(Ph. Eur. monograph 1477)

Action and use

Astringent.

Ph Eur

DEFINITION

Mixture of esters of glucose with gallic acid and 3-galloylgallic acid.

CHARACTERS

Appearance

Yellowish-white or slightly brown amorphous light powder or shiny plates.

Solubility

Very soluble in water, freely soluble in acetone, in ethanol (96 per cent) and in glycerol (85 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Dilute 0.1 mL of solution S (see Tests) to 5 mL with water R. Add 0.1 mL of ferric chloride solution R1. A blackish-blue colour is produced which becomes green on the addition of 1 mL of dilute sulfuric acid R.
B. To 1 mL of solution S, add 3 mL of a 1 g/L solution of gelatin R. The mixture becomes turbid and a flocculent precipitate is formed.
C. Dilute 0.1 mL of solution S to 5 mL with water R. Add 0.3 mL of barium hydroxide solution R. A greenish-blue precipitate is formed.

TESTS

Solution S
Dissolve 4.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1).

Dextrins, gum, salts, sugars

To 2 mL of solution S, add 2 mL of ethanol (96 per cent) R. The solution is clear. Add 1 mL of ether R. The solution remains clear for at least 10 min.

Resins

To 5 mL of solution S, add 5 mL of water R. The mixture remains clear (2.2.1) for at least 15 min.

Loss on drying (2.2.32)

Maximum 12.0 per cent, determined on 0.200 g by drying at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

STORAGE

Protected from light.

Ph Eur

Tar

Action and use

Used in treatment of psoriasis.

DEFINITION

Tar is bituminous liquid obtained from the wood of various trees of the family Pinaceae by destructive distillation and is known in commerce as Stockholm Tar.

CHARACTERISTICS

Dark brown or nearly black, semi-liquid; denser than water. Soluble in ethanol (90%), in ether and in fixed and volatile oils.

IDENTIFICATION

A. The aqueous liquid obtained by shaking 1 g with 20 mL of water for 5 minutes is acidic to litmus paper.
B. Carefully add 0.5 g to 10 mL of petroleum spirit (boiling range, 40° to 60°) and allow to stand for 30 minutes. When examined in daylight no fluorescence is produced.

Coal Tar

Action and use

Used in treatment of psoriasis.

Preparations

Coal Tar and Salicylic Acid Ointment

Coal Tar Solution

Strong Coal Tar Solution

Zinc and Coal Tar Paste

DEFINITION

Coal Tar is a product obtained from bituminous coal by

destructive distillation at about 1000°.

CHARACTERISTICS

A nearly black, viscous liquid. On exposure to air, the

viscosity gradually increases. It burns in air with a luminous,

sooty flame. It has a weight per mL of about 1.15 g.

Slightly soluble in water; partly soluble in ethanol, in ether and

in volatile oils.

IDENTIFICATION

A. A saturated solution is alkaline to litmus solution.

B. Carefully add 0.5 g to 10 mL of petroleum spirit (boiling

range, 40° to 60°) and allow to stand for 30 minutes. When

examined in daylight, the supernatant liquid has a blue

fluorescence which becomes more intense when viewed

under ultraviolet light (365 nm).

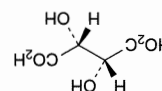
TESTS

Ash

Not more than 2.0%, Appendix XI J.

Tartaric Acid

(Ph. Eur. monograph 0460)



$C_4H_6O_6$

150.1

87-69-4

DEFINITION

(2R,3R)-2,3-Dihydroxybutanedioic acid.

Content

99.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

crystals.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) is strongly acid (2.2.4).

B. It gives the reactions of tartrates (2.3.1).

TESTS

Solution S

Dissolve 5.0 g in distilled water R and dilute to 50 mL with

the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Specific optical rotation (2.2.7)

+ 12.0 to + 12.8 (dried substance).

Dissolve 5.00 g in water R and dilute to 25.0 mL with the

same solvent.

Oxalic acid

Maximum 350 ppm, calculated as anhydrous oxalic acid.

Dissolve 0.80 g in 4 mL of water R. Add 3 mL of hydrochloric

acid R and 1 g of zinc R in granules and boil for 1 min.

Allow to stand for 2 min. Collect the liquid in a test-tube

containing 0.25 mL of a 10 g/L solution of phenylhydrazine

hydrochloride R and heat to boiling. Cool rapidly, transfer to a

graduated cylinder and add an equal volume of hydrochloric

acid R and 0.25 mL of a 50 g/L solution of potassium

ferricyanide R. Shake and allow to stand for 30 min. Any pink

colour in the solution is not more intense than that in a

standard prepared at the same time in the same manner.

using 4 mL of a 0.1 g/L solution of oxalic acid R.

Chlorides (2.4.4)

Maximum 100 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 150 ppm.

Dilute 10 mL of solution S to 15 mL with distilled water R.

Calcium (2.4.3)

Maximum 200 ppm.

To 5 mL of solution S add 10 mL of a 50 g/L solution of

sodium acetate R in distilled water R.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.2 per cent, determined on 1.000 g by drying in

an oven at 105° C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.650 g in 25 mL of water R. Titrate with 1 M

sodium hydroxide using 0.5 mL of phenolphthalein solution R as

indicator, until a pink colour is obtained.

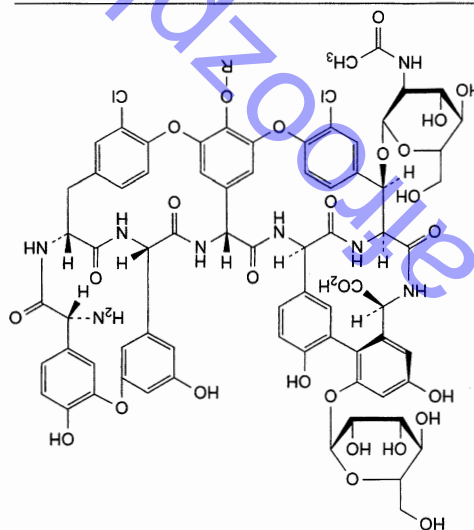
1 mL of 1 M sodium hydroxide is equivalent to 75.05 mg

of $C_4H_6O_6$.

Ph Eur

Teicoplanin

(Ph. Eur. monograph 2358)



Teicoplanin		
R	R'	
A ₂₋₁		
A ₂₋₂		
A ₂₋₃		
A ₂₋₄		
A ₂₋₅		
A ₃₋₁	H	

Action and use

Glycopeptide antibacterial.

DEFINITION

Mixture of glycopeptides produced by certain strains of *Actinoplanes teichomyceticus* sp.; the 6 principal components of the mixture are teicoplanin A₂₋₁ to A₂₋₅ and teicoplanin A₃₋₁. Fermentation product.

Potency

Minimum 900 IU/mg (anhydrous and sodium chloride-free substance).

CHARACTERS

Appearance

Yellowish, amorphous powder.

Solubility

Freely soluble in water, sparingly soluble in dimethylformamide, practically insoluble in ethanol (96 per cent V/V).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Comparison teicoplanin for identification CRS.

B. Examine the chromatograms obtained in the test for composition and related substances.

Results The principal peaks (teicoplanins A₃₋₁, A₂₋₁, A₂₋₂, A₂₋₃, A₂₋₄ and A₂₋₅) in the chromatogram obtained with the test solution are similar in retention time and size to the principal peaks in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₃ or B₄ (2.2.2, Method I). Dissolve 0.8 g in 10 mL of water R.

pH (2.2.3)

6.5 to 7.5.

Dissolve 0.50 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Composition and related substances

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 0.100 g of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of teicoplanin for identification CRS in water R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 10.0 mL with water R. Dilute 1.0 mL of this solution to 20.0 mL with water R.

Reference solution (c) Dissolve 50.0 mg of mestyl oxide CRS in water R and dilute to 25.0 mL with the same solvent. Dilute 1.0 mL of the solution to 10.0 mL with water R. Dilute 1.0 mL of this solution to 100.0 mL with water R.

Column:

size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:

mobile phase A: mix 900 mL of a 3.0 g/L solution of anhydrous sodium dihydrogen phosphate R, adjusted to pH 6.0 with 1 M sodium hydroxide, and 100 mL of acetonitrile R;

mobile phase B: mix 300 mL of a 3.0 g/L solution of anhydrous sodium dihydrogen phosphate R, adjusted to pH 6.0 with 1 M sodium hydroxide, and 700 mL of acetonitrile R;

Time	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 30	100 \rightarrow 50	0 \rightarrow 50
30 - 31	50 \rightarrow 10	50 \rightarrow 90
31 - 35	10	90

Flow rate 2.3 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L.

Identification Use the chromatogram supplied with teicoplanin for identification CRS and the chromatogram obtained with reference solution (a) to identify the groups and impurities.

Relative retention Of groups and impurities with reference to teicoplanin A₂₋₂:
— teicoplanin A₃ group ≤ 0.70 ;

— ticoplanin A₂ group > 0.70 and ≤ 1.25 and within this

group:

— ticoplanin A₂₋₂ = 1;
— ticoplanin A₂₋₁ group < 1;
— ticoplanin A₂₋₃ group > 1 and < 1.12;
— ticoplanin A₂₋₄ = about 1.12;
— ticoplanin A₂₋₅ group > 1.12 and ≤ 1.25;

— impurities > 1.25.

Relative retention Of principal peaks of the groups with reference to ticoplanin A₂₋₂ (retention time = about 18 min): ticoplanin A₂₋₁ = about 0.43;

— resolution: minimum 1.0 between the peaks due to ticoplanin A₂₋₄ and ticoplanin A₂₋₅.

Calculate the percentage content of the different components using the following equations:

$$\text{ticoplanin A}_2 \text{ group} = \frac{S_a + 0.83 \times S_b + S_c}{S_a} \times 100$$

$$\text{ticoplanin A}_{2-2} = \frac{S_2}{S_a + 0.83 \times S_b + S_c} \times 100$$

$$\text{ticoplanin A}_{2-1} \text{ group} = \frac{S_1}{S_a + 0.83 \times S_b + S_c} \times 100$$

$$\text{ticoplanin A}_{2-3} \text{ group} = \frac{S_3}{S_a + 0.83 \times S_b + S_c} \times 100$$

$$\text{ticoplanin A}_{2-4} = \frac{S_4}{S_a + 0.83 \times S_b + S_c} \times 100$$

$$\text{ticoplanin A}_{2-5} \text{ group} = \frac{S_5}{S_a + 0.83 \times S_b + S_c} \times 100$$

$$\text{ticoplanin A}_3 \text{ group} = \frac{0.83 \times S_b}{S_a + 0.83 \times S_b + S_c} \times 100$$

$$\text{impurities} = \frac{S_a + 0.83 \times S_b + S_c}{S_c} \times 100$$

S_a = sum of the areas of the peaks due to ticoplanin A₂ group in the chromatogram obtained with the test solution;
 S_b = sum of the areas of the peaks due to ticoplanin A₃ solution;
 S_c = sum of the areas of the peaks due to ticoplanin A₂₋₁ group in the chromatogram obtained with the test solution;
 S_1 = sum of the areas of the peaks due to ticoplanin A₂₋₁ group in the chromatogram obtained with the test solution;
 S_2 = area of the peak due to ticoplanin A₂₋₂ in the chromatogram obtained with the test solution;
 S_3 = sum of the areas of the peaks due to ticoplanin A₂₋₃ group in the chromatogram obtained with the test solution;
 S_4 = area of the peak due to ticoplanin A₂₋₄ in the chromatogram obtained with the test solution;

S_5 = sum of the areas of the peaks due to ticoplanin A₂₋₅ group in the chromatogram obtained with the test solution.

Limits:

— ticoplanin A₂ group: minimum 80.0 per cent;
— ticoplanin A₂₋₂: 35.0 per cent to 55.0 per cent;
— ticoplanin A₂₋₁ group: maximum 20.0 per cent;
— ticoplanin A₂₋₃ group: maximum 20.0 per cent;
— ticoplanin A₂₋₄: maximum 20.0 per cent;
— ticoplanin A₂₋₅ group: maximum 20.0 per cent;
— ticoplanin A₃ group: maximum 15.0 per cent;
— total of impurities other than mesityl oxide with a relative retention more than 1.25: maximum 5.0 per cent;
— disregard limit: the area of the peak due to ticoplanin A₂₋₂ in the chromatogram obtained with reference solution (b) (0.25 per cent).

Chlorides

Maximum 5.0 per cent, expressed as sodium chloride

(anhydrous substance).

Dissolve 1.000 g in 300 mL of water R, stir and acidify with 2 mL of nitric acid R. Titrate with 0.1 M silver nitrate,

determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M silver nitrate is equivalent to 5.844 mg of NaCl.

Heavy metals (2.4.8)

Maximum 20 ppm.

0.50 g complies with test G. Prepare the reference solution using 100 µL of lead standard solution (100 ppm Pb) R. Filter

the solutions through a membrane filter (nominal pore size 0.45 µm).

Impurity A

Liquid chromatography (2.2.29) as described in the test for composition and related substances with the following modifications.

Injection 20 µL of the test solution and reference solution (c).

Relative retention With reference to ticoplanin A₂₋₂ (retention time = about 18 min): impurity A = about 0.6.

Limits:

— impurity A: maximum twice the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent).

Water (2.5.12)

Maximum 15.0 per cent, determined on 0.300 g.

Bacterial endotoxins (2.6.14)

Less than 0.31 IU/mg.

ASSAY

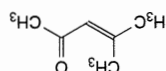
Carry out the microbiological assay of antibiotics (2.7.2), using the diffusion method. Use ticoplanin CRS as the reference substance.

STORAGE

Protected from light, at a temperature of 2 °C to 8 °C.

IMPURITIES

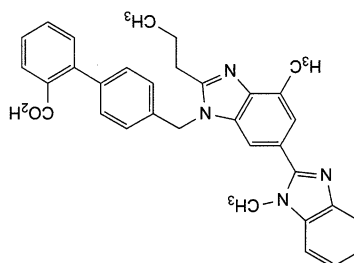
Specified impurities A



A. 4-methylpent-3-en-2-one (mesityl oxide).

Telmisartan

(Ph. Eur. monograph 2154)

C₃₃H₃₀N₄O₂ 514.6 144701-48-4

Action and use
Angiotensin II (AT₁) receptor antagonist.

Ph Eur

DEFINITION

4'-[[4-Methyl-6-(1-methyl-1H-benzimidazol-2-yl)-2-propyl-1H-benzimidazol-1-yl]methyl]biphenyl-2-carboxylic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in methanol, sparingly soluble in methylene chloride. It dissolves in 1 M sodium hydroxide.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.4).

Comparison telmisartan CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in hot anhydrous ethanol R, evaporate to dryness and record new spectra using the residues.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution Y₄ (2.2.2, Method II).

Dissolve 0.5 g in 1 M sodium hydroxide and dilute to 10 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution To 25 mg of the substance to be examined add about 5 mL of methanol R and 100 µL of a 40 g/L solution of sodium hydroxide R. Dissolve with the aid of ultrasound and dilute to 50 mL with methanol R.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with methanol R. Dilute 1.0 mL of this solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve the contents of a vial of telmisartan for system suitability CRS (containing impurities A, B, C, E and F) in 2 mL of methanol R.

Reference solution (c) To 5 mg of telmisartan for peak identification CRS (containing impurity D) add about 5 mL of methanol R and 100 µL of a 40 g/L solution of sodium

hydroxide R. Dissolve with the aid of ultrasound and dilute to 10 mL with methanol R.

Column:

size: l = 0.125 m, Ø = 4.0 mm;

stationary phase: octadecylsilyl silica gel for chromatography R

(5 µm) with a pore size of 10 nm;

temperature: 40 °C.

Mobile phase:

mobile phase A: dissolve 2.0 g of potassium dihydrogen

phosphate R and 3.8 g of sodium pentanesulfonate

monohydrate R1 in water R, adjust to pH 3.0 with dilute

phosphoric acid R and dilute to 1000 mL with water R;

mobile phase B: methanol R2, acetonitrile R1 (20:80 V/V);

Mobile phase B

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 3	70	30
3 - 28	70 → 20	30 → 80

Flow rate 1 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied

with telmisartan for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B, C, E and F;

use the chromatogram supplied with telmisartan for peak

identification CRS and the chromatogram obtained with

reference solution (c) to identify the peak due to impurity D.

Relative retention With reference to telmisartan (retention

time = about 15 min): impurity A = about 0.2;

impurity E = about 0.6; impurity F = about 0.7;

impurity B = about 0.9; impurity C = about 1.5;

impurity D = about 1.6.

System suitability: reference solution (b):

the chromatogram obtained with reference solution (b) is

similar to the chromatogram supplied with telmisartan for

system suitability CRS;

resolution: minimum 3.0 between the peaks due to

impurity B and telmisartan.

Limits:

impurities C, D: for each impurity, not more than twice

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.2 per cent);

impurities A, B: for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

total: not more than 10 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (1.0 per cent);

disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

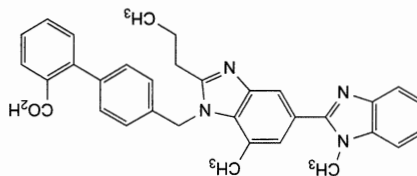
Dissolve 0.190 g in 5 mL of anhydrous formic acid R. Add 75 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 25.73 mg of $C_{15}H_{13}N_2O_2$.

IMPURITIES

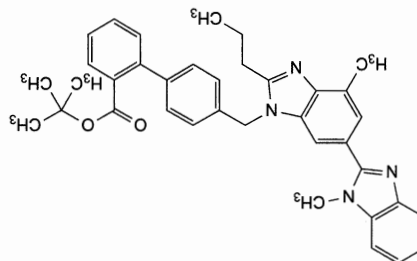
Specified impurities: A, B, C, D.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, F, G, H.

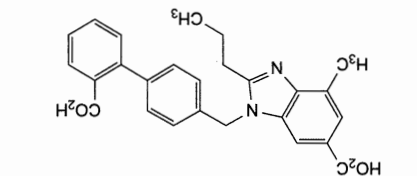
A. 4-methyl-6-(1-methyl-1H-benzimidazol-2-yl)-2-propyl-1H-benzimidazole,



B. 4'-[[7-methyl-5-(1-methyl-1H-benzimidazol-2-yl)-2-propyl-1H-benzimidazol-1-yl]methyl]biphenyl-2-carboxylic acid,

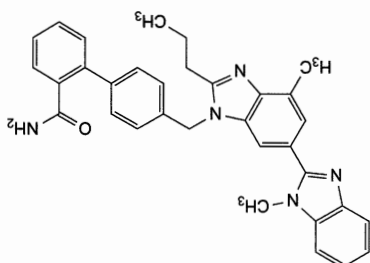
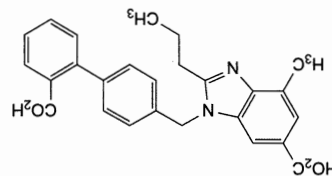


C. 1,1-dimethylethyl 4'-[[4-methyl-6-(1-methyl-1H-benzimidazol-2-yl)-2-propyl-1H-benzimidazol-1-yl]methyl]biphenyl-2-carboxylate,

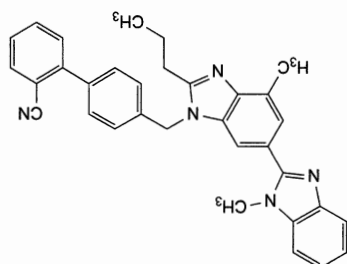


D. unknown structure,

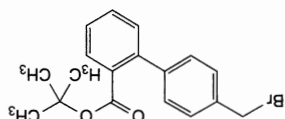
E. 1-[(2'-carboxybiphenyl-4-yl)methyl]-4-methyl-2-propyl-1H-benzimidazol-6-carboxylic acid,



F. 4'-[[4-methyl-6-(1-methyl-1H-benzimidazol-2-yl)-2-propyl-1H-benzimidazol-1-yl]methyl]biphenyl-2-carboxamide,



G. 4'-[[4-methyl-6-(1-methyl-1H-benzimidazol-2-yl)-2-propyl-1H-benzimidazol-1-yl]methyl]biphenyl-2-carbonitrile,

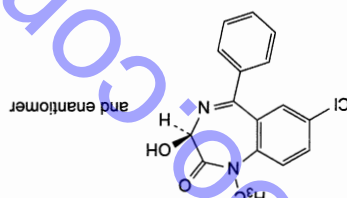


H. 1,1-dimethylethyl 4'-(bromomethyl)biphenyl-2-carboxylate.

Ph Eur

Temazepam

(Ph. Eur. monograph 0954)



$C_{16}H_{13}N_2O_2$ 300.7

846-50-4

Action and use

Benzodiazepine.

Temazepam Oral Solution

Temazepam Tablets

DEFINITION

(3R,5)-7-Chloro-3-hydroxy-1-methyl-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).



Appearance

Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in ethanol (96 per cent).

Infrared absorption spectrophotometry (2.2.24).

Comparison temazepam CRS.

Impurity A

Maximum 0.05 per cent.

Dissolve 0.400 g in methylene chloride R and dilute to 20.0 mL with the same solvent. The absorbance (2.2.25) is

Liquid chromatography (2.2.29).

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 1 volume of *water R* and 9 volumes of *methanol R* and dilute to 50.0 mL with the same mixture of solvents.

100.0 mL with a mixture of 1 volume of *water* R and 9 volumes of *methanol* R. Dilute 2.0 mL of this solution to 10.0 mL with a mixture of 1 volume of *water* R and 9 volumes of *methanol* R.

Reference solution (b) Dissolve 1 mg of oxaepam R, 1 mg of temazepam impurity F CRS and 1 mg of temazepam impurity G CRS in a mixture of 1 volume of water R and 9 volumes of methanol R and dilute to 25 mL with the same mixture of solvents.

Column: size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3.5 μ m).

- mobile phase A: solution containing 4.9 g/L of sodium dihydrogen phosphate R and 0.63 g/L of disodium hydrogen phosphate R (pH 5.6);
- mobile phase B: methanol R;
- mobile phase C: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Mobile phase C (per cent V/V)
0-18	54	39	7
18-25	54 \rightarrow 22	39 \rightarrow 63	7 \rightarrow 15
25-31	22	63	15
31-37	22 \rightarrow 54	63 \rightarrow 39	15 \rightarrow 7

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Relative retention With reference to temazepam (retention time = about 16 min): impurity E = about 0.55; impurity F = about 0.67; impurity G = about 0.73;

Protected from light.

of $C_{16}H_{13}ClN_2O_2$.

Dissolve 0.250 g in 50 mL of nitroethane K. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.20).
1 mL of 0.1 M perchloric acid is equivalent to 30.07 mg of $C_{16}H_{13}ClN_2O_2$.

Maximum 0.1 per cent, determined on 1.0 g.

Loss on drying (2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

(0.05 per cent).

— solution (a) (0.5 per cent);
disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a)

— *total*: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference

(a) chromatogram obtained with reference solution (0.10 per cent);

— *unspecified impurities*: for each impurity, not more than 0.5 times the area of the principal peak in the

than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

corresponding correction factor: impurity $F = 3.2$; impurity $G = 3.1$;

— *correction factors*: for the calculation of contents, multiply the peak areas of the following impurities by the

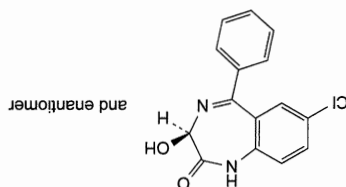
impurity B.

above the baseline of the peak due to impurity G and H^a = height above the baseline of the lowest point of the curve separating this peak from the peak due to

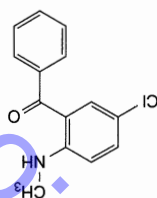
impurity F and impurity G; — *peak-to-valley ratio*: minimum 1.7, where H_p = height

System suitability: reference solution (b): — resolution: minimum 1.5 between the peaks due to

impurity B = about 0.8; impurity D = about 1.2;
impurity C = about 1.3; impurity A = about 1.5.



A. [5-chloro-2-(methylamino)phenyl]phenylmethanone,



Specified impurities: A, B, C, D, E, F, G.

benzodiazepin-2-one (oxazepam),

B. (3*RS*)-7-chloro-3-hydroxy-5-phenyl-1,3-dihydro-2*H*-1,4-

CHARACTERS**Appearance**

Yellow, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in methylene chloride, very slightly soluble in anhydrous ethanol. It dissolves in solutions of acids and alkalis.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison tenoxicam CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of methylene chloride R, evaporate to dryness and record new spectra using the residues.

TESTS**Appearance of solution**

The solution is clear (2.2.1).

Dissolve 0.10 g in methylene chloride R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture Mix equal volumes of acetonitrile R and water R. Adjust to apparent pH 3.2 with dilute phosphoric acid R1.

Test solution Dissolve 35 mg of the substance to be examined in the solvent mixture, sonicate and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

Reference solution (b) Dissolve 7 mg of pyridin-2-amine R (impurity A) in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of tenoxicam impurity mixture CRS (impurities B, G and H) in 1.0 mL of the test solution.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: cyanosilyl silica gel for chromatography R (3.5 μ m);

— temperature: 35 °C.

Mobile phase:

— mobile phase A: mix 25 volumes of methanol R2 and

75 volumes of water R and adjust to apparent pH 3.2 with dilute phosphoric acid R1;

— mobile phase B: mix 25 volumes of water R and

75 volumes of methanol R2 and adjust to apparent pH 3.2 with dilute phosphoric acid R1;

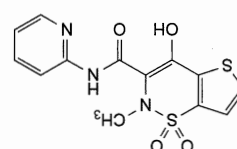
Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	96	4
5 - 16	96 \rightarrow 76	4 \rightarrow 24
16 - 25	76	24

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 230 nm.

Tenoxicam

(Ph Eur monograph 1156)



$C_{13}H_{11}N_3O_4S_2$

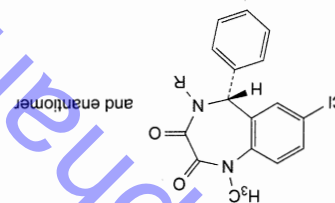
337.4

59804-37-4

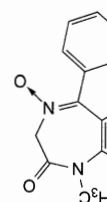


Ph Eur

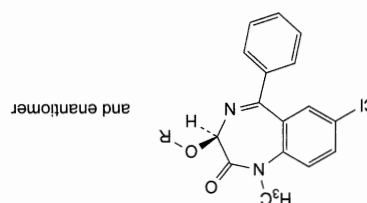
F, R = H: (5R)-7-chloro-1-methyl-5-phenyl-4,5-dihydro-1H-1,4-benzodiazepine-2,3-dione, 1H-1,4-benzodiazepine-2,3-dione, G, R = CH₃: (5R)-7-chloro-1,4-dimethyl-5-phenyl-4,5-dihydro-1H-1,4-benzodiazepine-2,3-dione.



E, 7-chloro-1-methyl-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one 4-oxide.



D, R = CH₃: (3R)-7-chloro-3-methoxy-1-methyl-5-phenyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one, 2,3-dihydro-1H-1,4-benzodiazepin-3-yl acetate,



99.0 per cent to 101.0 per cent (anhydrous substance).

Content

4-Hydroxy-2-methyl-N-(pyridin-2-yl)-2H-thieno[2,3-e]1,2-thiazine-3-carboxamide 1,1-dioxide.

DEFINITION

Ph Eur

Tenoxicam Tablets

Tenoxicam Injection

Preparations

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Action and use

Injection 20 µL

Identification of impurities:

- use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A;
- use the chromatogram supplied with tenoxicam impurity mixture CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities B, G and H; for identification of impurities G and H, which may be inverted in the elution order, take into account the heights of the corresponding peaks in the chromatogram supplied with tenoxicam

impurity mixture CRS.

Relative retention With reference to tenoxicam (retention

time = about 12 min): impurity A = about 0.1;

impurity G = about 0.85; impurity H = about 0.9;

impurity B = about 1.3.

System suitability: reference solution (c):

— resolution: minimum 1.3 between the peaks due to

impurity H (or G if peaks are inverted) and tenoxicam,

and between the peaks due to impurities G and H;

if necessary, optimise the apparent pH of the mobile

phases within the range 3.0–3.4.

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the

corresponding correction factor: impurity A = 0.2;

impurity B = 2.0;

— impurities A, B: for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

0.5 g complies with test C. Prepare the reference solution

using 5 mL of lead standard solution (2 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 5 mL of anhydrous formic acid R.

Add 70 mL of anhydrous acetic acid R. Titrate with 0.1 M

perchloric acid, determining the end-point potentiometrically

(2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 33.74 mg of

C₁₃H₁₁N₃O₄S₂.

STORAGE

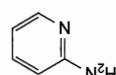
Protected from light.

IMPURITIES

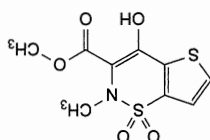
Specified impurities A, B

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or

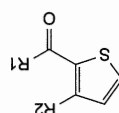
by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C, D, E, F, G, H.



A. pyridin-2-amine,



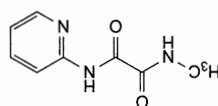
B. methyl 4-hydroxy-2-methyl-2H-thieno[2,3-e]1,2-thiazine-3-carboxylate 1,1-dioxide,



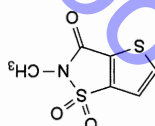
C. R1 = NH-CH₃, R2 = H; N-methylthiophene-2-carboxamide,

H. R1 = OH, R2 = SO₂-NH-CH₃;

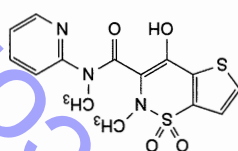
3-[(methylamino)sulfonyl]thiophene-2-carboxylic acid,



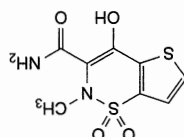
D. N-methyl-N'-(pyridin-2-yl)-ethanediamide,



E. 2-methylthieno[2,3-d]isothiazol-3(2H)-one 1,1-dioxide,



F. 4-hydroxy-N,N-dimethyl-N-(pyridin-2-yl)-2H-thieno[2,3-e]1,2-thiazine-3-carboxamide 1,1-dioxide,



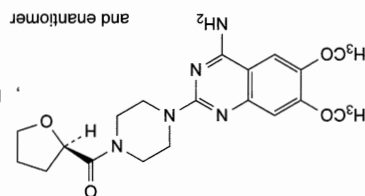
G. 4-hydroxy-2-methyl-2H-thieno[2,3-e]1,2-thiazine-3-carboxamide 1,1-dioxide.

Terazosin Hydrochloride Dihydrate

(Ph. Eur. monograph 2021)



www.webofpharma.com

C₁₉H₂₆ClN₅O₄·2H₂O 459.9 70024-40-7

Action and use

Alpha1-adrenoceptor antagonist.

Ph Eur

DEFINITION

1-(4-Amino-6,7-dimethoxyquinazolin-2-yl)-4-[[[(2RS)-tetrahydrofuran-2-yl]carbonyl]piperazine hydrochloride dihydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly yellow, crystalline powder.

Solubility

Sparingly soluble in water, slightly soluble in methanol, very slightly soluble in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison terazosin hydrochloride dihydrate CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 1.00 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Dilute 10 mL of solution S to 20 mL with water R.

pH (2.2.3)

3.0 to 5.0 for solution S.

Impurities N and O

Liquid chromatography (2.2.29).

Solvent mixture acetonitrile R₁, water R (20:80 V/V).

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of terazosin impurity A CRS and 5.0 mg of terazosin impurity N CRS in acetonitrile R₁ using sonication, add 5.0 mL of the test solution and dilute to 100.0 mL with the solvent mixture.

Reference solution (b) Dilute 10.0 mL of reference solution (a) to 100.0 mL with the solvent mixture.

Column:

— size: l = 0.25 m, Ø = 4.0 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm);
— temperature: 25 °C.
Mobile phase Dissolve 2.80 g of sodium laurylsulfate R in 1000.0 mL of water R and add 11.0 mL of a solution containing 202.4 g/L of triethylamine R and 230.0 g/L of phosphoric acid R; adjust to pH 2.5 with phosphoric acid R; mix 600 volumes of this solution with 400 volumes of acetonitrile R₁.
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 210 nm.
Injection 20 µL.
Run time 4 times the retention time of terazosin.
Relative retention With reference to terazosin (retention time = about 10 min): impurity O = about 0.2; impurity N = about 0.3; impurity A = about 0.4.
System suitability: reference solution (a):
— resolution: minimum 1.5 between the peaks due to impurities A and N.
Limits:
— impurity N: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— impurity O: not more than the area of the peak due to terazosin in the chromatogram obtained with reference solution (b) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.
Reference solution (a) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 5.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (b) Dissolve the contents of a vial of terazosin for system suitability CRS (containing impurities A, B, C, J, K and M) in the mobile phase and dilute to 10 mL with the mobile phase.
Reference solution (c) Dissolve 5.0 mg of terazosin impurity L CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.
Reference solution (d) To 5 mg of terazosin impurity E CRS, add 70 mL of methanol R and 30 mL of water R. Allow to stand for at least 1 h to dissolve the substance.
Use sonication if necessary.

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 µm);

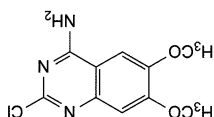
— temperature: 30 °C.

Mobile phase Mix 2 volumes of triethylamine R, 350 volumes of acetonitrile R, and 1650 volumes of a solution containing 6 g/L of sodium citrate R and 14.25 g/L of anhydrous citric acid R.
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 245 nm.
Injection 20 µL.
Run time 4 times the retention time of terazosin.

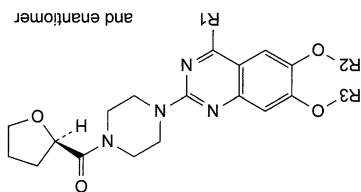
Identification of impurities Use the chromatogram supplied with terazosin for system suitability CRS and the chromatogram

obtained with reference solution (b) to identify the peaks due

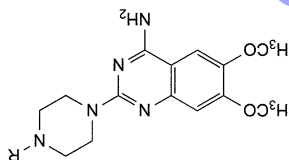
by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, F, G, H, I.



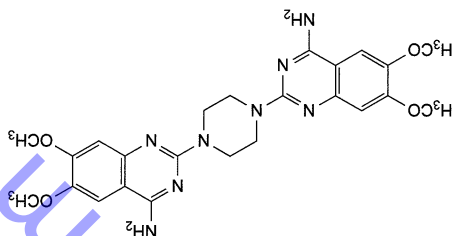
A. 2-chloro-6,7-dimethoxyquinazolin-4-amine,



B. R1 = OH, R2 = R3 = CH₃: 1-(4-hydroxy-6,7-dimethoxyquinazolin-2-yl)-4-[[[(2R,5S)-tetrahydrofuran-2-yl]carbonyl]piperazine,
G. R1 = NH₂, R2 = H, R3 = CH₃: 1-(4-amino-6-hydroxy-7-methoxyquinazolin-2-yl)-4-[[[(2R,5S)-tetrahydrofuran-2-yl]carbonyl]piperazine,
H. R1 = NH₂, R2 = CH₃, R3 = H: 1-(4-amino-7-hydroxy-6-methoxyquinazolin-2-yl)-4-[[[(2R,5S)-tetrahydrofuran-2-yl]carbonyl]piperazine,



C. R = H: 6,7-dimethoxy-2-(piperazin-1-yl)quinazolin-4-amine,
D. R = CHO: 1-(4-amino-6,7-dimethoxyquinazolin-2-yl)-4-formylpiperazine,
F. R = CO-[CH₂]⁴-OH: 1-(4-amino-6,7-dimethoxyquinazolin-2-yl)-4-(5-hydroxypentano-1-yl)piperazine,
J. R = CO-CH(OH)-CH₂-CH₂-CH₃: 1-(4-amino-6,7-dimethoxyquinazolin-2-yl)-4-[(2R,5S)-2-hydroxypentano-1-yl]piperazine,



E. 2,2'-(piperazine-1,4-diyl)bis(6,7-dimethoxyquinazolin-4-amine),

to impurities A, B, C, J, K and M; use the chromatograms obtained with reference solutions (c) and (d) to identify the peaks due to impurities L and B respectively.
Retention time Terazosin = about 11 min.
System suitability: reference solution (b):
— resolution: minimum 1.5 between the peaks due to impurities B and J; if necessary, adjust the proportion of the aqueous component in the mobile phase (an increase in the proportion of the aqueous component increases the retention times);
— the chromatogram obtained is similar to the chromatogram supplied with Terazosin for system suitability CRS; in case of insufficient separation of the impurities, reduce the amount of triethylamine in the mobile phase.
Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity C = 0.7;

— impurities A, C, E, K: for each impurity, not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

— impurity L: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.1 per cent);

— impurities B, J, M: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

7.0 per cent to 8.6 per cent, determined on 0.200 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of methanol R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20). Read the volume added between the 2 points of inflexion.
1 mL of 0.1 M sodium hydroxide is equivalent to 42.39 mg of C₁₉H₁₆ClN₅O₄.

STORAGE

Protected from light.

IMPURITIES

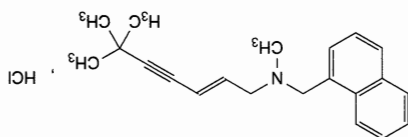
Specified impurities A, B, C, E, J, K, L, M, N, O

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or



Terbinafine Hydrochloride

(Ph. Eur. monograph 1734)



C₂₁H₂₆ClN

327.9

78628-80-5

Action and use

Antifungal.

Preparation

Terbinafine Tablets

Ph Eur

DEFINITION

(2E)-N₆,6-(Trimethyl-N-(naphthalen-1-ylmethyl)hept-2-en-4-yn-1-amine hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Very slightly or slightly soluble in water, freely soluble in anhydrous ethanol and in methanol, slightly soluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison terbinafine hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1) using anhydrous ethanol R as solvent.

TESTS

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Solvent mixture A acetone R, water R (50:50 V/V).

Solvent mixture B acetone R, methanol R (40:60 V/V).

Buffer solution Dilute 2.0 mL of triethylamine R1 to 950 mL with water R. Adjust to pH 7.5 with a mixture of 5 volumes of glacial acetic acid R and 95 volumes of water R and dilute to 1000.0 mL with water R.

Test solution Dissolve 25 mg of the substance to be examined in solvent mixture A and dilute to 50.0 mL with solvent mixture A.

Reference solution (a) Dissolve 5 mg of terbinafine for system suitability CRS (containing impurities B and E) in 10.0 mL of solvent mixture A.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with solvent mixture A. Dilute 1.0 mL of this

solution to 10.0 mL with solvent mixture A.

Column:

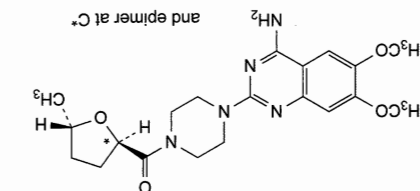
size: $l = 0.15$ m, $\varnothing = 3.0$ mm;

stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

temperature: 40 °C.

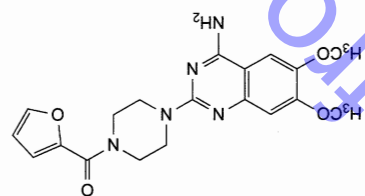
Mobile phase:

mobile phase A: buffer solution, solvent mixture B (30:70 V/V);

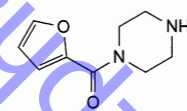


and epimer at C*

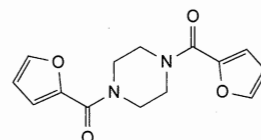
I. 1-(4-amino-6,7-dimethoxyquinazolin-2-yl)-4-[[[(2R,5S)-5-methyltetrahydrofuran-2-yl]carbonyl]piperazine,



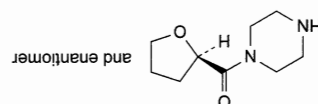
K. 1-(4-amino-6,7-dimethoxyquinazolin-2-yl)-4-(furan-2-ylcarbonyl)piperazine (prazosin),



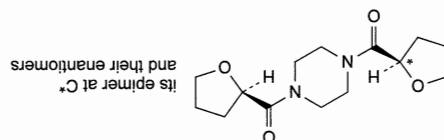
L. 1-(furan-2-ylcarbonyl)piperazine,



M. 1,4-bis(furan-2-ylcarbonyl)piperazine,



N. 1-[[[(2R)-tetrahydrofuran-2-yl]carbonyl]piperazine,



O. 1,4-bis[[tetrahydrofuran-2-yl]carbonyl]piperazine.

Ph Eur

— mobile phase B: buffer solution, solvent mixture B

(5:95 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 4	100	0
4 - 25	100 → 0	0 → 100
25 - 30	0	100

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

Identification of impurities Use the chromatogram supplied

with terbinafine for system suitability CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities B and E.

Relative retention With reference to terbinafine (retention

time = about 15 min); impurity B = about 0.9;

impurity E = about 1.7.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to

impurity B and terbinafine.

Limits:

— correction factor: for the calculation of content, multiply the

peak area of impurity E by 0.5;

— impurity B: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— impurity E: not more than 0.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.05 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R₃ add

5 mL of 0.01 M hydrochloric acid. Titrate with 0.1 M sodium

hydroxide determining the end-point potentiometrically

(2.2.20). Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 32.79 mg of

C₂₁H₂₆ClN.

STORAGE

Protected from light.

IMPURITIES

Specified impurities B, E

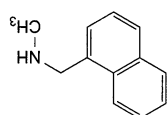
Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

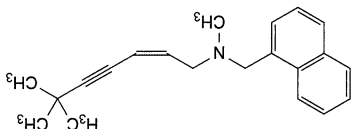
the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

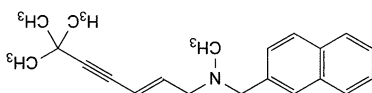
by the general monograph Substances for pharmaceutical use



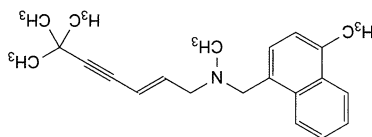
A. N-methyl-C-(naphthalen-1-yl)methanamine,



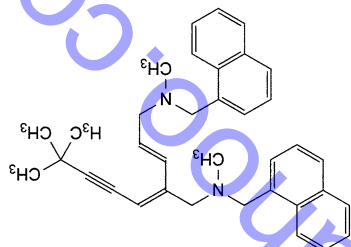
B. (2Z)-N,6,6-trimethyl-N-(naphthalen-1-ylmethyl)hept-2-en-4-yn-1-amine (cis-terbinafine),



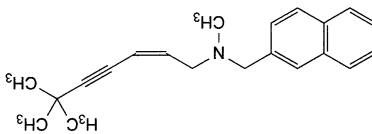
C. (2E)-N,6,6-trimethyl-N-(naphthalen-2-ylmethyl)hept-2-en-4-yn-1-amine (trans-terbinafine),



D. (2Z)-N,6,6-trimethyl-N-[(4-methylnaphthalen-1-yl)methyl]hept-2-en-4-yn-1-amine (4-methylterbinafine),



E. (2E,4E)-4-(4,4-dimethylpent-2-yn-1-ylidene)-N,N'-dimethyl-N,N'-bis(naphthalen-1-ylmethyl)pent-2-ene-1,5-diamine,



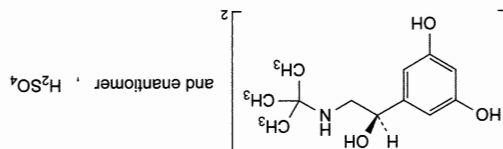
F. (2Z)-N,6,6-trimethyl-N-(naphthalen-2-ylmethyl)hept-2-en-4-yn-1-amine (cis-isoterbinafine).

(2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, D, F.

Terbutaline Sulfate

Terbutaline Sulphate

(Ph. Eur. monograph 0690)



$C_{24}H_{40}N_2O_{10}S$ 548.7 23031-32-5

Action and use
Beta₂-adrenoceptor agonist; bronchodilator.

Preparation
Terbutaline Tablets

Ph. Eur.

DEFINITION

Bis[(1*R*,5*S*)-1-(3,5-dihydroxyphenyl)-2-[(1*S*,1-dimethylethyl)amino]ethanol] sulfate.

CHARACTERS

98.0 per cent to 101.0 per cent (dried substance).

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison *terbutaline sulfate* CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in *aldehyde-free methanol* R, evaporate to dryness and record new spectra using the residues.

B. 5 mL of solution S (see Tests) gives reaction (a) of sulfates (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in *carbon dioxide-free water* R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and its absorbance (2.2.25) at 400 nm in a 2 cm cell is not greater than 0.11.

Acidity

To 10 mL of solution S add 0.05 mL of *methyl red solution* R. Not more than 1.2 mL of 0.01 M *sodium hydroxide* is required to change the colour of the indicator to yellow.

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 75.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 7.5 mg of *terbutaline sulfate* CRS and 22.5 mg of *terbutaline sulfate* CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.



Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 20.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve 4.23 g of *sodium hexanesulfonate* R in 770 mL of 0.050 M *ammonium formate* solution prepared as follows: dissolve 3.15 g of *ammonium formate* R in about 980 mL of *water* R; adjust to pH 3.0 by adding about 8 mL of *anhydrous formic acid* R and dilute to 1000 mL with *water* R; then add 230 mL of *methanol* R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 276 nm.

Injection 20 μ L.

Run time 6 times the retention time of *terbutaline*.

Retention time Impurity C = about 9 min;

terbutaline = about 11 min.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to impurity C and *terbutaline*; if necessary adjust the composition of the mobile phase, decrease the content of *methanol* to increase the retention time.

Limits:

— impurity C: not more than twice the area of the corresponding peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— impurities A, B, D: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— sum of impurities other than C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.02 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105°C for 3 h.

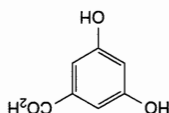
ASSAY

Dissolve 0.400 g in 70 mL of *anhydrous acetic acid* R with heating. Titrate with 0.1 M *perchloric acid*, determining the end-point potentiometrically (2.2.20).

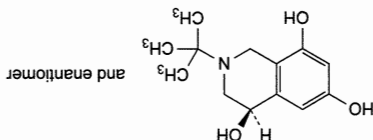
1 mL of 0.1 M *perchloric acid* is equivalent to 54.87 mg of $C_{24}H_{40}N_2O_{10}S$.

IMPURITIES

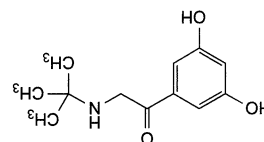
Specified impurities A, B, C, D



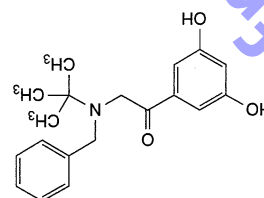
A. 3,5-dihydroxybenzoic acid (α -resorcylic acid),



B. (4*R,S*)-2-(1,1-dimethylethyl)-1,2,3,4-tetrahydroisquinoline-4,6,8-triol,



C. 1-(3,5-dihydroxyphenyl)-2-[(1,1-dimethylethyl)amino]-2-[(1,1-dimethylethyl)amino]ethanone,

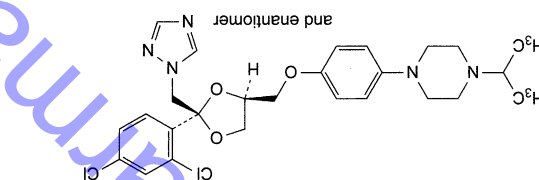


D. 2-[benzyl-(1,1-dimethylethyl)amino]-1-(3,5-dihydroxyphenyl)ethanone.

Ph Eur

Terconazole

(Ph. Eur. monograph 1270)



C₂₆H₃₁Cl₂N₅O₃ 532.5 67915-31-5

Action and use

Antifungal.

Ph Eur

DEFINITION

1-[4-[(2*R*,4*SR*)-2-(2,4-dichlorophenyl)-2-[(1*H*-1,2,4-triazol-1-yl)methyl]-1,3-dioxolan-4-yl]methoxy]phenyl]-4-(1-methylethyl)piperazine.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in acetone, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

First identification A

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison terconazole CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of acetone R, evaporate to dryness in a current of air and record new spectra using the residues.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 30 mg of the substance to be examined

in methanol R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 30 mg of terconazole CRS in

methanol R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 30 mg of terconazole CRS and

30 mg of ketoconazole CRS in methanol R and dilute to 5 mL

with the same solvent.

Plate TLC octadecylsilyl silica gel plate R.

Mobile phase ammonium acetate solution R, dioxan R,

methanol R (20:40:40 V/V/V).

Application 5 µL.

Development In an unsaturated tank over half of the plate.

Drying In a current of warm air for 15 min.

Detection Expose to iodine vapour until the spots appear and

examine in daylight.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with reference

solution (a).

C. To 30 mg in a porcelain crucible add 0.3 g of anhydrous

sodium carbonate R. Heat over an open flame for 10 min.

Allow to cool. Take up the residue with 5 mL of dilute nitric

acid R and filter. To 1 mL of the filtrate add 1 mL of

water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Optical rotation (2.2.7)

–0.10° to +0.10°.

Dissolve 1.0 g in methylene chloride R and dilute to 10 mL

with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be

examined in methanol R and dilute to 10.0 mL with the same

solvent.

Reference solution (a) Dissolve 2.0 mg of ketoconazole CRS

and 2.5 mg of terconazole CRS in methanol R and dilute to

100.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with methanol R. Dilute 5.0 mL of this solution to

20.0 mL with methanol R.

Column:

— size: $l = 0.1$ m, $\varnothing = 4.6$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for

chromatography R (3 µm).

Mobile phase:

— mobile phase A: 3.4 g/L solution of tetrabutylammonium

hydrogen sulfate R;

— mobile phase B: acetonitrile R;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	95 → 50	5 → 50
10 - 15	50	50

Flow rate 2 mL/min.

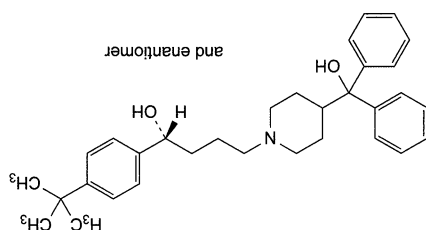
Detection Spectrophotometer at 220 nm.

Injection 10 µL.



Terfenadine

(Ph. Eur. monograph 0955)



$C_{32}H_{41}NO_2$ 471.7 50679-08-8

Action and use

Histamine H₁ receptor antagonist; antihistamine.

Preparations

Terfenadine Oral Suspension

Terfenadine Tablets

Ph. Eur.

DEFINITION

(1*R,S*)-1-[4-(1,1-dimethylethyl)phenyl]-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butan-1-ol.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, freely soluble in methylene chloride, soluble in methanol. It is very slightly soluble in dilute hydrochloric acid.

It shows polymorphism (5.9).

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Melting point (2.2.14): 146 °C to 152 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 50.0 mg in methanol R and dilute to 100.0 mL with the same solvent.

Spectral range 230–350 nm.

Absorption maximum At 259 nm.

Shoulders At 253 nm and 270 nm.

Specific absorbance at the absorption maximum 13.5 to 14.9.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison terfenadine CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 50 mg of terfenadine CRS in methylene chloride R and dilute to 10 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase methanol R, methylene chloride R (10:90 V/V).

Application 10 µL.

Development Over a path of 15 cm.

Limits:

— impurities A, B: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.25 per cent);

— unspecified impurities: for each impurity, not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

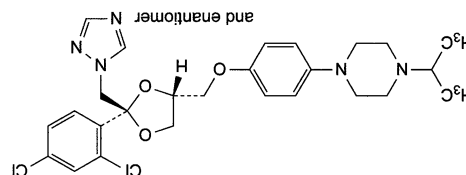
Dissolve 0.150 g in 70 mL of a mixture of anhydrous acetic acid R and 7 volumes of methyl ethyl ketone R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically at the 2nd point of inflexion (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 17.75 mg of C₃₂H₄₁ClN₃O₃.

STORAGE

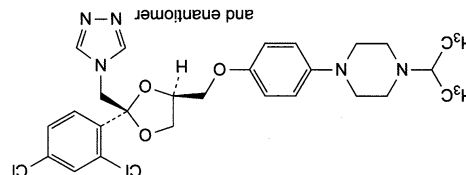
Protected from light.

IMPURITIES

Specified impurities A, B



A. 1-[4-[[[(2*R,S*,4*R,S*)-2-(2,4-dichlorophenyl)-2-[(1*H*,1,2,4-triazol-1-yl)methyl]-1,3-dioxolan-4-yl]methoxy]phenyl]-4-(1-methylethyl)piperazine,



B. 1-[4-[[[(2*R,S*,4*S,R*)-2-(2,4-dichlorophenyl)-2-[(4*H*,1,2,4-triazol-4-yl)methyl]-1,3-dioxolan-4-yl]methoxy]phenyl]-4-(1-methylethyl)piperazine.

Ph. Eur.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS**Related substances**

Liquid chromatography (2.2.29).

Test solution Dissolve 15 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 15 mg of terfenadine impurity A CRS in the mobile phase and dilute to 10.0 mL with the mobile phase. To 5.0 mL of this solution, add 5.0 mL of the test solution and dilute to 50.0 mL with the mobile phase.

Reference solution (c) Dilute 10.0 mL of reference solution (a) to 25.0 mL with the mobile phase.

Reference solution (d) Dissolve 0.1 g of potassium iodide R in the mobile phase and dilute to 100 mL with the mobile phase. Dilute 1 mL of this solution to 100 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dilute 600 mL of acetonitrile R1 to 1 L with diethylammonium phosphate buffer solution pH 6.0 R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 217 nm.

Injection 20 μ L.

Run time 5 times the retention time of terfenadine.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to terfenadine and impurity A;

— mass distribution ratio: minimum 2.0 for the peak due to terfenadine; use potassium iodide R as the unretained compound (reference solution (d)).

Limits:

— impurities A, B, C, D, E, F, G, H, I, J: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c)

(0.2 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.025 times the area of the principal peak in the chromatogram obtained with reference solution (c)

(0.005 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying at 60 °C at a pressure not exceeding 0.5 kPa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.400 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

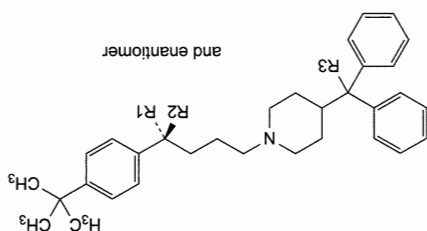
1 mL of 0.1 M perchloric acid is equivalent to 47.17 mg

STORAGE

Protected from light.

IMPURITIES

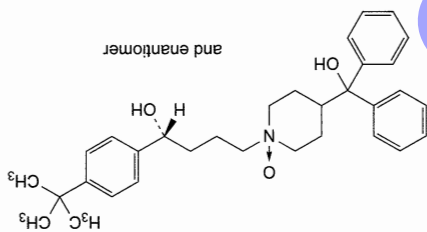
Specified impurities A, B, C, D, E, F, G, H, I, J



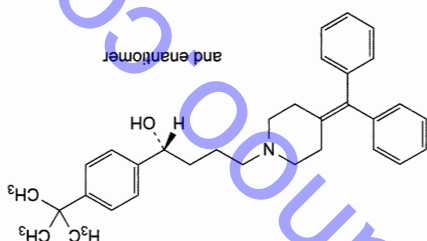
A. R1 + R2 = O, R3 = OH: 1-[4-(1,1-dimethylethyl)phenyl]-4-[4-(hydroxydiphenylmethyl)piperidin-1-yl]butan-1-one,

B. R1 = OH, R2 = R3 = H: (1RS)-1-[4-(1,1-dimethylethyl)phenyl]-4-[4-(diphenylmethyl)piperidin-1-yl]butan-1-ol,

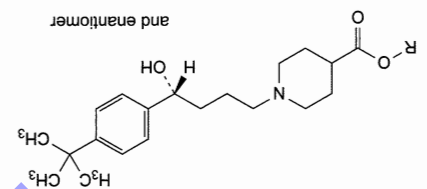
H. R1 = R2 = H, R3 = OH: [1-[4-(1,1-dimethylethyl)phenyl]piperidin-4-yl]diphenylmethanol,



C. 1-[(4RS)-4-[4-(1,1-dimethylethyl)phenyl]-4-(hydroxydiphenylmethyl)piperidin-1-yl]butan-1-oxide,



D. (1RS)-1-[4-(1,1-dimethylethyl)phenyl]-4-[4-(diphenylmethylene)piperidin-1-yl]butan-1-ol,



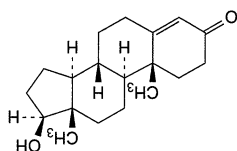
E. R = H: 1-[(4RS)-4-[4-(1,1-dimethylethyl)phenyl]-4-hydroxybutyl]piperidine-4-carboxylic acid,

J. R = C₂H₅: ethyl 1-[(4RS)-4-[4-(1,1-dimethylethyl)phenyl]-4-hydroxybutyl]piperidine-4-carboxylate,



Testosterone

(Ph. Eur. monograph 1373)



$C_{19}H_{28}O_2$ 288.4 58-22-0

Action and use

Androgen.

Preparation

Testosterone Implants

DEFINITION

17 β -Hydroxyandrost-4-en-3-one.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white crystalline powder, or colourless or

yellowish-white crystals.

Solubility

Practically insoluble in water, freely soluble in alcohol and in

methylene chloride, practically insoluble in fatty oils.

mp

About 155 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison testosterone CRS.

TESTS

Specific optical rotation (2.2.7)

+ 106 to + 114 (dried substance).

Dissolve 0.250 g in ethanol R and dilute to 25.0 mL with the

same solvent.

Impurities D and F

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.100 g of the substance to be

examined in methanol R and dilute to 10 mL with the same

solvent.

Reference solution (a) Dissolve 1 mg of stanolone R in

methanol R and dilute to 10 mL with the same solvent.

In 1 mL of this solution, dissolve 10 mg of testosterone for

impurity D identification CRS (testosterone spiked with about

1 per cent of impurity D).

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with methanol R.

Reference solution (c) Dilute 2.0 mL of reference solution (b)

to 10.0 mL with methanol R.

Reference solution (d) Dilute 1.0 mL of reference solution (b)

to 10.0 mL with methanol R.

Plate TLC silica gel F₂₅₄ plate R (6-8 μ m).

Preconditioning (in the dark): add about 5 g of powdered silver

nitrate R to 100 mL of methanol R. Stir the suspension for

30 min. Filter or decant the suspension and immerse the

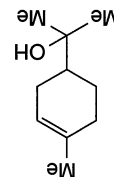
plate in the silver nitrate solution for at least 30 min. Dry at

Terpineol

$C_{10}H_{18}O$

154.3

98-55-5



Terpineol is a mixture of structural isomers in which α -terpineol predominates.

CHARACTERISTICS

A colourless, slightly viscous, liquid which may deposit

crystals; odour, pleasant and characteristic.

Very slightly soluble in water, freely soluble in ethanol (70%);

soluble in ether.

TESTS

Refractive index

1.4800 to 1.4855, Appendix V E.

Weight per mL

0.931 to 0.935 g, Appendix V G.

Low-boiling substances

Not more than 4.0% v/v distils below 214°, Appendix V C.

A pre-conditioned plate can be stored in the dark for

5-7 days.

Mobile phase acetic acid R, ethanol R, dioxan R, methylene

chloride R (1:2:10:90 V/V/V/V).

Application 2 µL.

Development In a saturated tank over 3/4 of the plate.

Drying Allow to stand at room temperature and protected

from light for 30 min.

Detection Spray with a 200 g/L solution of toluidenesulfonic

acid R in ethanol R and heat at 105 °C for 10 min. Examine

in ultraviolet light at 365 nm.

System suitability The chromatogram obtained with reference

solution (a) shows 3 clearly separated spots; impurity D

R_F = about 0.5; testosterone R_F = about 0.65; impurity F

R_F = about 0.7.

Limits:

impurity D: any spot due to impurity D is not more

intense than the spot in the chromatogram obtained with

reference solution (c) (0.2 per cent),

impurity F: any spot due to impurity F is not more intense

than the spot in the chromatogram obtained with

reference solution (d) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be

examined in methanol R and dilute to 10.0 mL with the same

solvent.

Reference solution (a) Dissolve 10 mg of testosterone for system

suitability CRS (containing impurities C and I) in 1 mL of

methanol R.

Reference solution (b) Dilute 1.0 mL of the test solution to

20.0 mL with methanol R. Dilute 1.0 mL of this solution to

10.0 mL with methanol R.

Reference solution (c) Dilute 2.0 mL of reference solution (b)

to 10.0 mL with methanol R.

Column:

size: l = 0.25 m, Ø = 4.6 mm,

stationary phase: spherical end-capped octadecylsilyl silica gel

for chromatography R (5 µm) with a pore size of 15 nm,

temperature: 40 °C.

Mobile phase:

mobile phase A: water for chromatography R, methanol R

(45:55 V/V),

mobile phase B: methanol R,

Time (min)

Mobile phase A (per cent V/V)

Mobile phase B (per cent V/V)

0 100

4 24 100 → 60

24 53 60 → 0

53 55 0

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL.

Relative retention With reference to testosterone (retention

time = about 18 min): impurity G = about 0.6;

impurity H = about 0.8; impurity A = about 0.9;

impurity I = about 0.95; impurity C = about 1.2;

impurity E = about 1.7; impurity J = about 2.1;

impurity B = about 2.5.

System suitability: reference solution (a):

— resolution: minimum baseline separation between the peaks

due to impurity I and testosterone.

Limits: Use the chromatogram obtained with reference

solution (a) to identify the peaks due to impurities C and I:

— correction factor: for the calculation of content, multiply the

peak area of impurity I by 2.9,

impurity C: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.5 per cent),

impurity I: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (c) (0.2 per cent),

impurities A, B, E, G, H, J: for each impurity, not more

than the area of the principal peak in the chromatogram

obtained with reference solution (c) (0.1 per cent),

any other impurity: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (c) (0.1 per cent),

total: not more than 1.2 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.6 per cent),

disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (c)

(0.05 per cent).

Loss on drying (2.3.32)

Maximum 1.0 per cent, determined on 0.500 g by drying in

an oven at 105 °C for 2 h.

ASSAY

Dissolve 50.0 mg in alcohol R and dilute to 100.0 mL with

the same solvent. Dilute 2.0 mL to 100.0 mL with alcohol R.

Measure the absorbance (2.2.25) at the absorption maximum

at 241 nm.

Calculate the content of C₁₉H₂₈O₂ taking the specific

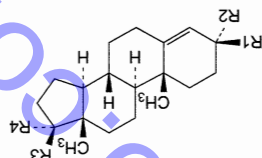
absorbance to be 569.

STORAGE

Protected from light.

IMPURITIES

Specified impurities: A, B, C, D, E, F, G, H, I, J.



A. R1 + R2 = R3 + R4 = O: androst-4-ene-3,17-dione
(androstenedione),
C. R1 + R2 = O, R3 = H, R4 = OH: 17α-hydroxyandrost-
4-en-3-one (epitestosterone),
D. R1 = R3 = OH, R2 = R4 = H: androst-4-ene-3β,17β-
diol (Δ4-androstenediol),
E. R1 + R2 = O, R3 = O, R4 = H: 3-oxoandrost-
4-en-17β-yl acetate (testosterone acetate),

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Practically insoluble in water, very soluble in acetone, in methylene chloride and in anhydrous ethanol, freely soluble in fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison testosterone decanoate CRS.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Dissolve 0.20 g in 20 mL of methanol R.

Specific optical rotation (2.2.7)

+ 75.0 to + 80.0 (dried substance).
Dissolve 0.200 g in anhydrous ethanol R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve the contents of a vial of testosterone decanoate for system suitability CRS (containing impurities A, B, C, D, E and F) in 1 mL of the mobile phase.

Reference solution (b) Dilute 10.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 20.0 mg of testosterone decanoate CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase water R, acetonitrile R (5:95 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (b).

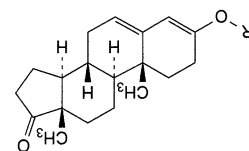
Run time Twice the retention time of testosterone decanoate.

Identification of impurities Use the chromatogram supplied with testosterone decanoate for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E and F.

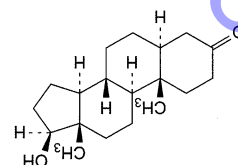
Relative retention With reference to testosterone decanoate (retention time = about 20 min): impurity A = about 0.2;

impurity B = about 0.6; impurities C and G = about 0.79; impurity D = about 0.83; impurity E = about 1.3;

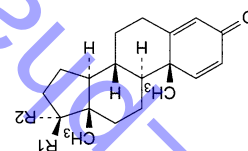
impurity F = about 1.7.



B. R = C₂H₅; 3-ethoxyandrost-3,5-dien-17-one (androstenedione ethylether),
J. R = CH₃; 3-methoxyandrost-3,5-dien-17-one (androstenedione methylether),



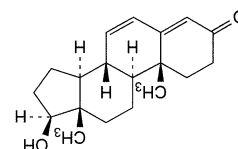
F. 17 β -hydroxy-5 α -androst-3-one (androstanolone, stanolone),



G. R₁ + R₂ = O: androst-1,4-diene-3,17-dione

(androstadienedione).

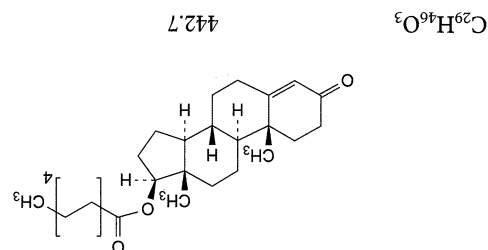
H. R₁ = OH, R₂ = H: 17 β -hydroxyandrost-1,4-dien-3-one (boldenone),



I. 17 β -hydroxyandrost-4,6-dien-3-one (Δ^6 -testosterone).

**Testosterone Decanoate**

(Ph. Eur. monograph 1736)

**Action and use**

Androgen.

DEFINITION

3-Oxoandrost-4-en-17 β -yl decanoate.

Content

97.0 per cent to 102.0 per cent (dried substance).

System suitability: reference solution (a):
— resolution: minimum 1.5 between the peaks due to impurities C and D.
Limits:

— correction factors: for the calculation of content, multiply the peak area of impurity A by 0.7;
— impurities A, B, D, E, F: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);
— sum of impurities C and G: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);
— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Free acid

Dissolve 0.65 g in 10 mL of ethanol (96 per cent) R₃, previously neutralised to bromothymol blue solution R₃, and titrate immediately with 0.01 M sodium hydroxide, using 0.1 mL of bromothymol blue solution R₃ as indicator.

Not more than 0.6 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to blue.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g over diphosphorus pentoxide R at a pressure not exceeding 0.7 kPa.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

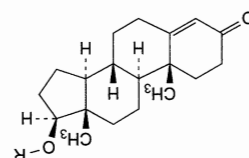
Injection 20 µL of the test solution and reference solution (c). Calculate the percentage content of C₂₉H₄₆O₃ from the declared content of testosterone decanoate CRS.

STORAGE

At a temperature of 2 °C to 8 °C.

IMPURITIES

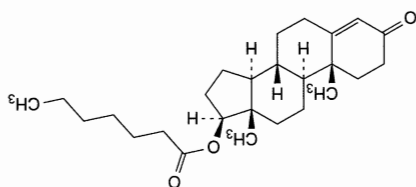
Specified impurities A, B, C, D, E, F, G



A. R = H: testosterone,
B. R = CO-[CH₂]₆-CH₃: 3-oxoandroster-4-en-17-yl
octanoate (testosterone octanoate),
C. R = CO-[CH₂]₇-CH₃: 3-oxoandroster-4-en-17-yl
nonanoate (testosterone nonanoate),
D. R = CO-[CH₂]₈-CH=CH₂: 3-oxoandroster-4-en-17-yl
undec-10-enoate (testosterone undecylenate),
E. R = CO-[CH₂]₉-CH₃: 3-oxoandroster-4-en-17-yl
undecanoate (testosterone undecanoate),
F. R = CO-[CH₂]₁₀-CH₃: 3-oxoandroster-4-en-17-yl
dodecanoate (testosterone laurate),

Testosterone Enantate

(Ph Eur monograph 1048)



C₂₆H₄₀O₃

400.6

315-37-7

Action and use

Androgen.

Preparation

Testosterone Enantate Injection

DEFINITION

3-Oxoandroster-4-en-17-yl heptanoate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Practically insoluble in water, very soluble in anhydrous ethanol, freely soluble in fatty oils.

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Melting point (2.2.14): 34 °C to 39 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison testosterone enantate CRS.

C. Thin-layer chromatography (2.2.27).

Solvent mixture methanol R, methylene chloride R (10:90 V/V).

Test solution Dissolve 5 mg of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 5 mg of testosterone enantate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of testosterone enantate CRS, 5 mg of testosterone decanoate CRS and 5 mg of testosterone isocaproate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase water R, acetonitrile R, 2-propanol R

(20:40:60 V/V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air, then at 100 °C for 10 min; allow to cool.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution (a).

Detection B Spray with alcoholic solution of sulfuric acid R; heat

at 120 °C for 10 min; allow to cool and examine in daylight.

Results B The principal spot in the chromatogram obtained with the test solution is green and is similar in position and size to the principal spot in the chromatogram obtained with the reference solution (a).

System suitability: reference solution (b):
— the chromatogram shows 3 clearly separated principal spots by each method of visualisation.

D. To about 25 mg add 2 mL of a 10 g/L solution of

potassium hydroxide R in methanol R and boil under a reflux condenser for 1 h. Cool. Add 10 mL of water R. Acidify with

dilute hydrochloric acid R until blue litmus paper R turns red.

Filter and wash the precipitate with a small quantity of water R. The residue, after drying at 60 °C at a pressure not

exceeding 0.7 kPa for 3 h, melts (2.2.14) at 150 °C to

TESTS

Specific optical rotation (2.2.7)

+ 81 to + 86 (dried substance).

Dissolve 0.100 g in anhydrous ethanol R and dilute to

Impurity A

Maximum 0.16 per cent.

Dissolve 0.50 g in 10 mL of ethanol (96 per cent) R previously neutralised to bromothymol blue solution R3. Titrate

immediately with 0.01 M sodium hydroxide using 0.1 mL of

bromothymol blue solution R3 as indicator. Not more than

0.6 mL of 0.01 M sodium hydroxide is required to change the

Impurity H

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.100 g of the substance to be

examined in 1.0 mL of ethanol (96 per cent) R.

Reference solution Dissolve 3.0 mg of testosterone enantate

impurity H CRS in 20.0 mL of ethanol (96 per cent) R.

Plate TLC silica gel plate R.

Mobile phase ethyl acetate R, cyclohexane R1 (40:60 V/V).

Application 1 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with a 200 g/L solution of toluenesulfonic

acid R in ethanol (96 per cent) R and heat at 120 °C for

10 min; examine in ultraviolet light at 366 nm.

System suitability: reference solution:

— the chromatogram shows a clearly visible spot due to

impurity H.

Limit: impurity H: any spot due to impurity H is not more

intense than the principal spot in the chromatogram

obtained with the reference solution (0.15 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile

phase.

Reference solution (a) Dissolve with the aid of ultrasound the

contents of a vial of testosterone enantate for system

suitability CRS (containing impurities F and G) in the mobile

phase and dilute to 1.0 mL with the mobile phase.

Reference solution (b) Dilute 2.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 2 mg of testosterone enantate for

peak identification CRS (containing impurity E) in 1.0 mL of

the mobile phase.

Reference solution (d) Dissolve 2 mg of testosterone

caproate CRS (impurity B) and 2 mg of testosterone CRS

(impurity D) in the mobile phase and dilute to 5.0 mL with

the mobile phase. Dilute 1.0 mL of the solution to 100.0 mL

with the mobile phase.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: spherical end-capped dodecylsilyl silica gel

for chromatography R (4 µm).

Mobile phase water R, acetonitrile R (30:70 V/V).

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 242 nm.

Injection 10 µL.

Run time 1.5 times the retention time of testosterone

enantate.

Identification of impurities Use the chromatogram supplied

with testosterone enantate for system suitability CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities F and G; use the

chromatogram supplied with testosterone enantate for peak

identification CRS and the chromatogram obtained with

reference solution (c) to identify the peak due to impurity E;

use the chromatogram obtained with reference solution (d) to

identify the peaks due to impurities B and D.

Relative retention With reference to testosterone enantate

(retention time = about 22 min): impurity D = about 0.1;

impurity B = about 0.7; impurity E = about 0.8;

impurity F = about 0.85; impurity G = about 0.9.

System suitability: reference solution (a):

— resolution: minimum 1.3 between the peaks due to

impurities F and G.

Limits:

— correction factor: for the calculation of content, multiply the

peak area of impurity F by 6.3;

— impurity D: not more than 4 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.4 per cent);

— impurities E, F: for each impurity, not more than 3 times

the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.3 per cent);

— impurity B: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— impurity G: not more than 1.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— *total*: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);
 — *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in a desiccator over *diphosphorus pentoxide* R at a pressure not exceeding 0.7 kPa.

ASSAY

Dissolve 50.0 mg in *anhydrous ethanol* R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with *anhydrous ethanol* R. Measure the absorbance (2.2.25) at the absorption maximum at 241 nm. Calculate the content of $C_{25}H_{38}O_3$ taking the specific absorbance to be 422.

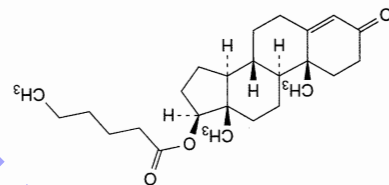
STORAGE

Protected from light, at a temperature of 2 °C to 8 °C.

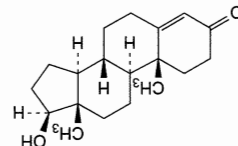
IMPURITIES

Specified impurities A, B, D, E, F, G, H

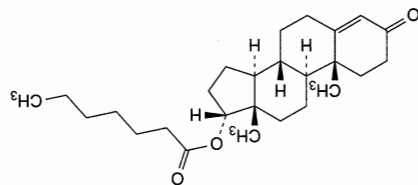
A. heptanoic acid,



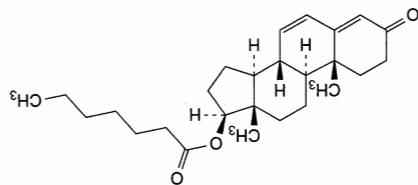
B. 3-oxoandrosta-4-en-17β-yl hexanoate (testosterone caproate),



D. 17β-hydroxyandrosta-4-en-3-one (testosterone),



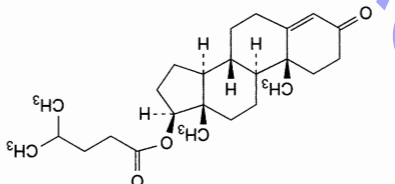
E. 3-oxoandrosta-4-en-17α-yl heptanoate (17α-testosterone enantate),



F. 3-oxoandrosta-4,6-dien-17β-yl heptanoate (Δ6-testosterone enantate),

Testosterone Isocaproate

(Ph. Eur. monograph 1737)



$C_{25}H_{38}O_3$ 386.6

Action and use

Androgen.

Ph Eur

DEFINITION

3-Oxoandrosta-4-en-17β-yl 4-methylpentanoate.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, very soluble in acetone and in methylene chloride, freely soluble in fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison testosterone isocaproate CRS.

B. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained

with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with

reference solution (c).

TESTS

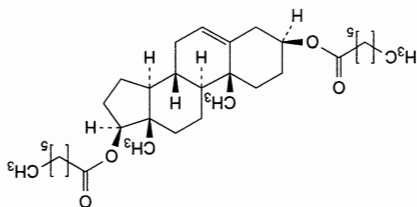
Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

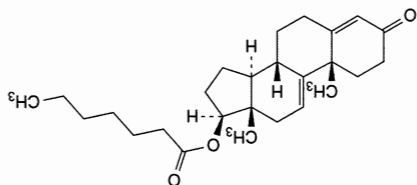


Ph Eur

H. androst-5-ene-3β,17β-diyl diheptanoate.



G. 3-oxoandrosta-4,9(11)-dien-17β-yl heptanoate (Δ9(11)-testosterone enantate),



Dissolve 0.20 g in 20 mL of methanol R.

Specific optical rotation (2.2.7)

+ 82.0 to + 88.0 (dried substance).

Dissolve 0.200 g in anhydrous ethanol R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of testosterone isocaproate for system suitability CRS (containing impurities A, B, C, D, E, F and G) in 10 mL of the mobile phase.

Reference solution (b) Dilute 10.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 20.0 mL with the mobile phase.

Reference solution (c) Dissolve 20.0 mg of testosterone isocaproate CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase water R, acetonitrile R (15:85 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (b).

Run time Twice the retention time of testosterone isocaproate.

Identification of impurities Use the chromatogram supplied with testosterone isocaproate for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, E, F and G.

Relative retention With reference to testosterone isocaproate (retention time = about 14 min): impurity A = about 0.2; impurity B = about 0.4; impurity C = about 0.5; impurity D = about 0.7; impurity G = about 0.8; impurity E = about 1.1; impurity F = about 1.4.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 2.5, where H_p = height above the baseline of the peak due to impurity E and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to testosterone isocaproate.

Limits:

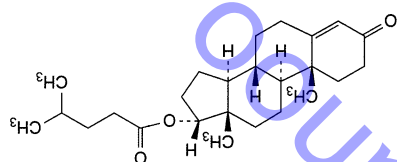
— impurities A, B, C, D, E, F, G: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b)

— unspecified impurities: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b)

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b)

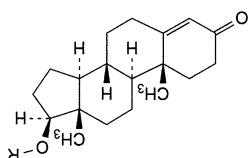
— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent).



G. 3-oxoandroster-4-en-17 α -yl 4-methylpentanoate (epitestosterone isocaproate).

- A. R = H: testosterone,
B. R = CO-CH₃: 3-oxoandroster-4-en-17 β -yl acetate (testosterone acetate),
C. R = CO-C₂H₅: testosterone propionate,
D. R = CO-CH(CH₃)₂: 3-oxoandroster-4-en-17 β -yl 2-methylpropanoate (testosterone isobutyrate),
E. R = CO-[CH₂]₄-CH₃: 3-oxoandroster-4-en-17 β -yl hexanoate (testosterone caproate),
F. R = CO-[CH₂]₅-CH₃: testosterone enantate,



Specified impurities A, B, C, D, E, F, G

IMPURITIES

declared content of testosterone isocaproate CRS. Calculate the percentage content of C₂₅H₃₈O₃ from the Injection 20 μ L of the test solution and reference solution (c) related substances with the following modification.

Liquid chromatography (2.2.29) as described in the test for

ASSAY

diphosphorus pentoxide R at a pressure not exceeding 0.7 kPa.

Maximum 0.5 per cent, determined on 1.000 g over Loss on drying (2.2.32)

required to change the colour of the indicator to blue. Not more than 0.6 mL of 0.01 M sodium hydroxide is

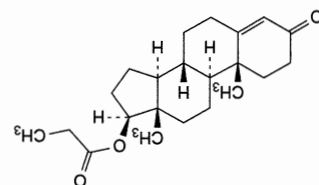
0.1 mL of bromothymol blue solution R3 as indicator.

titrate immediately with 0.01 M sodium hydroxide, using previously neutralised to bromothymol blue solution R3, and Dissolve 0.44 g in 10 mL of ethanol (96 per cent) R,

Free acid

Testosterone Propionate

(Ph. Eur. monograph 0297)

 $C_{22}H_{32}O_3$

344.5

57-85-2

Action and use

Androgen.

Preparations

Testosterone Propionate Injection

Ph Eur

DEFINITION

3-Oxoandrost-4-en-17 β -yl propanoate.

Content

97.5 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder or colourless crystals.

Solubility

Practically insoluble in water, freely soluble in acetone and in ethanol (96 per cent), soluble in fatty oils.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison testosterone propionate CRS.

TESTS

Specific optical rotation (2.2.7)

+ 84 to + 90 (dried substance).

Dissolve 0.250 g in ethanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in methanol R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of testosterone propionate for system suitability CRS (containing impurities A, B and C) in 5.0 mL of methanol R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (c) Dissolve 20.0 mg of testosterone propionate CRS in 50.0 mL of methanol R.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase water R, methanol R (20:80 V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (b).

Run time Twice the retention time of testosterone propionate.

Identification of impurities Use the chromatogram supplied with testosterone propionate for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B and C.

Relative retention With reference to testosterone propionate (retention time = about 8 min): impurity C = about 0.4; impurity A = about 0.7; impurity B = about 1.4.

System suitability: reference solution (a): — peak-to-valley ratio: minimum 3.0, where H_p = height above the baseline of the peak due to impurity B and H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to testosterone propionate.

Limits:

— impurity A: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than 7 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.7 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

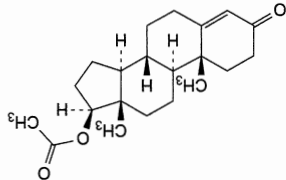
Calculate the percentage content of $C_{22}H_{32}O_3$ taking into account the assigned content of testosterone propionate CRS.

IMPURITIES

Specified impurities A, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: B, D, E.

A. 3-oxoandrost-4-en-17 β -yl acetate (testosterone acetate),

CHARACTERS**Appearance**

White or almost white, slightly hygroscopic, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

It melts at about 148 °C or it may occur in either of 2 other crystalline forms which melt respectively at about 134 °C and 139 °C. Mixtures of these forms melt within the range 134 °C to 147 °C.

IDENTIFICATION

First identification A, B, D

Second identification B, C, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tetracaine hydrochloride CRS.

B. To 10 mL of solution S (see Tests) add 1 mL of ammonium thiocyanate solution R. A white, crystalline precipitate is formed which, after recrystallisation from water R and drying at 80 °C for 2 h, melts (2.2.14) at about 131 °C.

C. To about 5 mg add 0.5 mL of fuming nitric acid R. Evaporate to dryness on a water-bath, allow to cool and dissolve the residue in 5 mL of acetone R. Add 1 mL of 0.1 M alcoholic potassium hydroxide. A violet colour develops.

D. Solution S gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 5.0 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dilute 2 mL of solution S to 10 mL with water R.

pH (2.2.3)

4.5 to 6.5.

Dilute 1 mL of solution S to 10 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use or store them at 2–8 °C.

Solvent mixture acetone/nitile R, water R (20:80 V/V).

Test solution Dissolve 50 mg of the substance to be examined in the solvent mixture and dilute to 50 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of tetracaine for system suitability CRS (containing impurities A, B and C) in 2 mL of the solvent mixture.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

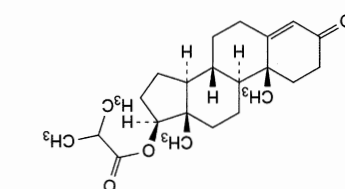
— temperature: 30 °C.

Mobile phase:

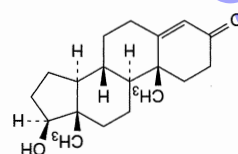
— mobile phase A: dissolve 1.36 g of potassium dihydrogen phosphate R in water R, add 0.5 mL of phosphoric acid R

and dilute to 1000 mL with water R;

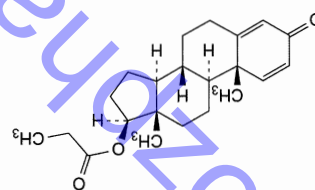
— mobile phase B: acetonitrile R;



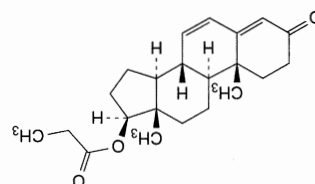
B. 3-oxoandrosta-4-en-17-yl 2-methylpropanoate (testosterone isobutyrate);



C. 17β-hydroxyandrosta-4-en-3-one (testosterone);



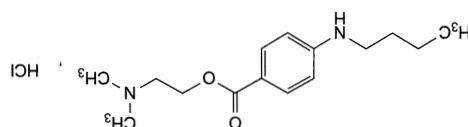
D. 3-oxoandrosta-1,4-dien-17-yl propanoate;



E. 3-oxoandrosta-4,6-dien-17-yl propanoate.

Tetracaine Hydrochloride

(Ph. Eur. monograph 0057)



$C_{15}H_{25}ClN_2O_2$

300.8

136-47-0

Action and use
Local anaesthetic.

Preparation

Tetracaine Eye Drops

DEFINITION

2-(Dimethylamino)ethyl 4-(butylamino)benzoate hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	80	20
3 - 18	80 → 40	20 → 60
18 - 23	40	60

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 300 nm.

Injection 10 µL.

Identification of impurities Use the chromatogram supplied with tetracosactide for system suitability (RS) and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B and C.

Relative retention With reference to tetracosactide (retention time = about 8 min): impurity A = about 0.3;

impurity B = about 1.7; impurity C = about 2.1.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to

tetracosactide and impurity B.

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity B = 0.6;

impurity C = 0.7;

— impurity A: not more than 0.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.05 per cent);

— impurities B, C: for each impurity, not more than the area

of the principal peak in the chromatogram obtained with

reference solution (a) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with test A. Prepare the

reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R and add

5.0 mL of 0.01 M hydrochloric acid. Carry out a

potentiometric titration (2.2.20), using 0.1 M sodium

hydroxide. Read the volume added between the 2 points of

inflection.

1 mL of 0.1 M sodium hydroxide is equivalent to 30.08 mg of

C₁₅H₂₅ClN₂O₂.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A, B, C

DEFINITION

Synthetic tetracosapeptide, in which the sequence of amino acids is the same as that of the first 24 residues of human corticotropin. It increases the rate at which corticoid hormones are secreted by the adrenal glands. It is available as an acetate.

Content

90 per cent to 102 per cent (anhydrous and acetic acid-free substance). By convention, 1 µg of tetracosactide is

equivalent to 1 IU of tetracosactide.

CHARACTERS

Appearance

White or yellow, amorphous powder.

Solubility

Sparingly soluble in water.

IDENTIFICATION

A. Examine the chromatograms obtained in the test for related peptides.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with the reference solution.

B. Amino acid analysis (2.2.56). For hydrolysis use Method 1 and for analysis use Method 1.

Tetracosactide

(Ph. Eur. monograph 0644)



H-Ser - Tyr - Ser - Met - Glu - His - Phe - Arg - Trp - Gly - Lys - Pro - Val - Gly - Lys - Arg - Arg - Pro - Val - Lys - Val - Tyr - Pro - OH

C₁₃₆H₂₁₀N₄₀O₃₁S 2933

16960-16-0

Action and use

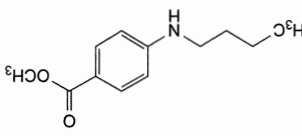
Corticotropic peptide.

Preparations

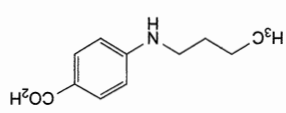
Tetracosactide Injection

Tetracosactide Zinc Injection

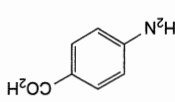
Ph. Eur.



B. 4-(butylamino)benzoic acid,



A. 4-aminobenzoic acid,



Express the content of each amino acid in moles. Calculate the relative proportions of the amino acids, taking that of valine to be equivalent to 3. The values fall within the following limits: lysine 3.5 to 4.7; histidine 0.9 to 1.1; arginine 2.7 to 3.3; serine 1.1 to 2.2; glutamic acid 0.9 to 1.1; proline 2.5 to 3.5; glycine 1.8 to 2.2; methionine 0.9 to 1.1; tyrosine 1.7 to 2.2; phenylalanine 0.9 to 1.1. Not more than traces of other amino acids are present.

TESTS

Specific optical rotation (2.2.7)

— 99 to –109 (anhydrous and acetic acid-free substance).
Dissolve 10.0 mg in 1.0 mL of a mixture of 1 volume of *glacial acetic acid* R and 99 volumes of *water* R.

Absorbance (2.2.25)

determined at the absorption maximum between 240 nm and 280 nm, at 276 nm. The ratio of the absorbance at the maximum at 276 nm to the absorbance at 248 nm is 2.4 to 2.9.
Dissolve 1.0 mg in 0.1 M *hydrochloric acid* and dilute to 5.0 mL with the same acid.

Related peptides

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve an accurately weighed quantity of the substance to be examined in *water* R to obtain the same concentration as in reference solution (a).

Reference solution (a) Dissolve the contents of a vial of *tetracosactide CRS* in *water* R to obtain a concentration of about 1 mg/mL, as indicated in the leaflet provided with the reference standard.

Reference solution (b) In order to prepare impurity A *in situ*, dissolve 1.0 mg of the substance to be examined in 1 mL of a 1 per cent V/V solution of *glacial acetic acid* R, add 50 µL of a mixture of 1 volume of *strong hydrogen peroxide solution* R and 999 volumes of *water* R, and allow to stand for 2 h.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;
— stationary phase: *octadecylsilyl silica gel for chromatography* R (3 µm);
— temperature: 25 °C.

Mobile phase:

— **mobile phase A:** mix 5.0 mL of *glacial acetic acid* R, 60 mL of *acetonitrile* R and 5.0 g of *ammonium sulfate* R and dilute to 1000 mL with *water* R;
— **mobile phase B:** mix 5.0 mL of *glacial acetic acid* R, 310 mL of *acetonitrile* R and 5.0 g of *ammonium sulfate* R and dilute to 1000 mL with *water* R;
— **mobile phase C:** *acetonitrile* R.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)	Mobile phase C (per cent V/V)
0 - 50	55 → 40	45 → 60	0
50 - 50.1	40 → 0	60 → 15	0 → 85
50.1 - 55	0	15	85
55 - 55.1	0 → 55	15 → 45	85 → 0
55.1 - 60	55	45	0

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 275 nm.
Injection 20 µL.

Identification of impurities Use the chromatogram supplied with *tetracosactide CRS* and the chromatogram obtained with reference solution (a) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.
Relative retention With reference to *tetracosactide* (retention time = about 26 min): impurity A = about 0.3; impurity B = about 0.95.
System suitability: reference solution (a):
— *peak-to-valley ratio*: minimum 3, where H_p = height above the baseline of the peak due to impurity B and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to *tetracosactide*.
Limits:
— **impurity A:** maximum 3 per cent;
— **impurity B:** maximum 4 per cent;
— **unspecified impurities:** for each impurity, maximum 2.5 per cent;
— **sum of impurities other than A:** maximum 9 per cent.

Acetic acid (2.5.34)

8.0 per cent to 13.0 per cent.

Test solution Dissolve 10.0 mg of the substance to be examined in a mixture of 5 volumes of mobile phase B and 95 volumes of mobile phase A and dilute to 10.0 mL with the same mixture of mobile phases.

Water (2.5.32)

Maximum 14.0 per cent, determined on 20.0–50.0 mg.

Bacterial endotoxins (2.6.14)

Less than 10 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related peptides.
Calculate the content of $C_{136}H_{210}N_{40}O_{31}S$ using the declared content of *tetracosactide CRS*.

STORAGE

Protected from light, at a temperature of 2 °C to 8 °C.

LABELLING

The label states:
— the mass of peptide in the container;
— where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

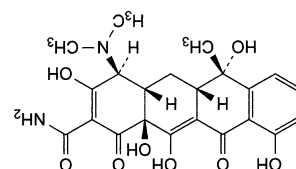
IMPURITIES

Specified impurities A, B

A. *tetracosactide sulfoxide*,
B. unknown structure.

Tetracycline

(Ph. Eur. monograph 0211)

C₂₂H₂₄N₂O₈ 444.4 60-54-8

Action and use

Tetracycline antibacterial.

Ph Eur

DEFINITION

(4S,4aS,5aS,6S,12aS)-4-[(dimethylamino)-3,6,10,12,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydrotetracene-2-carboxamide];

Substance produced by certain strains of *Streptomyces aerofaciens* or obtained by any other means.

Content

88.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Very slightly soluble in water, soluble in ethanol (96 per cent) and in methanol, sparingly soluble in acetone. It dissolves in dilute acid and alkaline solutions.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of tetracycline hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of tetracycline hydrochloride CRS, 5 mg of demeclocycline hydrochloride R and 5 mg of oxytetracycline hydrochloride R in methanol R and dilute to 10 mL with the same solvent.

Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.

Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a 63 g/L solution of oxalic acid R previously adjusted to pH 2 with concentrated ammonia R.

Application 1 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of sulfuric acid R. A violet-red colour develops. Add the solution to 2.5 mL of water R. The colour becomes yellow.

C. Dissolve about 10 mg in a mixture of 1 mL of dilute nitric acid R and 5 mL of water R. Shake and add 1 mL of silver nitrate solution R₂. Any opalescence in the solution is not more intense than that in a mixture of 1 mL of dilute nitric acid R, 5 mL of water R and 1 mL of silver nitrate solution R₂.

TESTS

pH (2.2.3)

3.5 to 6.0.

Suspend 0.1 g in 10 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7)

-260 to -280 (dried substance).

Dissolve 0.250 g in 0.1 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 25.0 mg of the substance to be examined in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (a) Dissolve 25.0 mg of tetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (b) Dissolve 12.5 mg of 4-epitetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Reference solution (c) Dissolve 10.0 mg of anhydrotetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same acid.

Reference solution (d) Dissolve 10.0 mg of 4-epianhydrotetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Reference solution (e) Mix 1.0 mL of reference solution (a), 2.0 mL of reference solution (b) and 5.0 mL of reference solution (d) and dilute to 25.0 mL with 0.01 M hydrochloric acid.

Reference solution (f) Mix 40.0 mL of reference solution (b), 20.0 mL of reference solution (c) and 5.0 mL of reference solution (d) and dilute to 200.0 mL with 0.01 M hydrochloric acid.

Reference solution (g) Dilute 1.0 mL of reference solution (c) to 50.0 mL with 0.01 M hydrochloric acid.

Column:

— size: l = 0.25 m, Ø = 4.6 mm;

— stationary phase: styrene-divinylbenzene copolymer R (8 µm);

— temperature: 60 °C.

Mobile phase Weigh 80.0 g of 2-methyl-2-propanol R and transfer to a 1000 mL volumetric flask with the aid of

200 mL of water R, add 100 mL of a 35 g/L solution of dipotassium hydrogen phosphate R adjusted to pH 9.0 with dilute phosphoric acid R, 200 mL of a 10 g/L solution of tetrabutylammonium hydrogen sulfate R adjusted to pH 9.0 with dilute sodium hydroxide solution R and 10 mL of a 40 g/L solution of sodium edetate R adjusted to pH 9.0 with dilute sodium hydroxide solution R, dilute to 1000.0 mL with water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL; inject the test solution and reference solutions (e), (f) and (g).

System suitability:

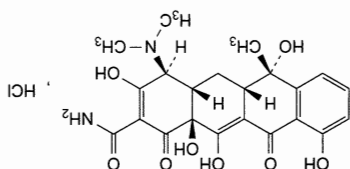
— resolution: minimum 2.5 between the peaks due to impurity A (1st peak) and tetracycline (2nd peak) and

Ph Eur



Tetracycline Hydrochloride

(Ph. Eur. monograph 0210)



64-75-5

480.9

 $C_{22}H_{25}ClN_2O_8$

Action and use
Tetracycline antibacterial.

Preparations
Tetracycline Capsules
Tetracycline Tablets

Ph Eur

DEFINITION

(4S,4aS,5aS,6S,12aS)-4-(Dimethylamino)-3,6,10,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydroretetracene-2-carboxamide hydrochloride. Substances produced by certain strains of *Streptomyces aureofaciens* or obtained by any other means.

Content

95.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Soluble in water, slightly soluble in ethanol (96 per cent), practically insoluble in acetone. It dissolves in solutions of alkali hydroxides and carbonates. Solutions in water become turbid on standing, owing to the precipitation of tetracycline.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of tetracycline hydrochloride CRS in methanol R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of tetracycline hydrochloride CRS, 5 mg of demeclocycline hydrochloride R and

minimum 8.0 between the peaks due to tetracycline and impurity D (3^{rd} peak) in the chromatogram obtained with reference solution (e); if necessary, adjust the concentration of 2-methyl-2-propanol in the mobile phase;
— *signal-to-noise ratio*: minimum 3 for the principal peak in the chromatogram obtained with reference solution (g);
— *symmetry factor*: maximum 1.25 for the peak due to tetracycline in the chromatogram obtained with reference solution (e).

Limits:

— *impurity A*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (5.0 per cent);
— *impurity B* (eluting on the tail of the principal peak): not more than 0.4 times the area of the peak due to impurity A in the chromatogram obtained with reference solution (f) (2.0 per cent);
— *impurity C*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (1.0 per cent);
— *impurity D*: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (0.5 per cent).

Heavy metals (2.4.8)

Maximum 50 ppm.

0.5 g complies with test C. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 13.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

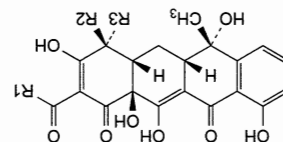
Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{22}H_{24}N_2O_8$.

STORAGE

Protected from light.

IMPURITIES



A. $R_1 = NH_2$, $R_2 = H$, $R_3 = N(CH_3)_2$

(4R,4aS,5aS,6S,12aS)-4-(dimethylamino)-3,6,10,12a-pentahydroxy-6-methyl-1,11-dioxo-1,4,4a,5,5a,6,11,12a-octahydroretetracene-2-carboxamide (4-epitetracycline),

B. $R_1 = CH_3$, $R_2 = N(CH_3)_2$, $R_3 = H$;

(4S,4aS,5aS,6S,12aS)-2-acetyl-4-(dimethylamino)-

3,6,10,12a-pentahydroxy-6-methyl-4a,5a,6,12a-

tetracycline-1,11(4H,5H)-dione (2-acetyl-2-

decarbarmoyltetracycline).

5 mg of oxytetracycline hydrochloride R in methanol R and dilute to 10 mL with the same solvent.
 Plate TLC octadecylsilyl silica gel F₂₅₄ plate R.
 Mobile phase Mix 20 volumes of acetonitrile R, 20 volumes of methanol R and 60 volumes of a 63 g/L solution of oxalic acid R previously adjusted to pH 2 with concentrated ammonium R.
 Application 1 µL.
 Development Over 3/4 of the plate.
 Drying In air.

Detection Examine in ultraviolet light at 254 nm.
 System suitability The chromatogram obtained with reference solution (b) shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

B. To about 2 mg add 5 mL of sulfuric acid R. A violet-red colour develops. Add the solution to 2.5 mL of water R. The colour becomes yellow.
 C. It gives reaction (a) of chlorides (2.3.1).

TESTS

pH (2.2.3)

1.8 to 2.8.

Dissolve 0.1 g in 10 mL of carbon dioxide-free water R.

Specific optical rotation (2.2.7)

–240 to –255 (dried substance).
 Dissolve 0.250 g in 0.1 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 25.0 mg of the substance to be examined in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (a) Dissolve 25.0 mg of tetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solution (b) Dissolve 15.0 mg of 4-epitetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Reference solution (c) Dissolve 10.0 mg of anhydrotetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same acid.

Reference solution (d) Dissolve 10.0 mg of 4-epianhydrotetracycline hydrochloride CRS in 0.01 M hydrochloric acid and dilute to 50.0 mL with the same acid.

Reference solution (e) Mix 1.0 mL of reference solution (a), 2.0 mL of reference solution (b) and 5.0 mL of reference solution (d) and dilute to 25.0 mL with 0.01 M hydrochloric acid.

Reference solution (f) Mix 20.0 mL of reference solution (b), 10.0 mL of reference solution (c) and 5.0 mL of reference solution (d) and dilute to 200.0 mL using 0.01 M hydrochloric acid.

Reference solution (g) Dilute 1.0 mL of reference solution (c) to 50.0 mL with 0.01 M hydrochloric acid.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

stationary phase: styrene-divinylbenzene copolymer R (8 µm);

temperature: 60 °C.

STORAGE

Protected from light. If the substance is sterile, store in a sterile, tamper-proof container.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
 Injection Test solution and reference solution (a).
 Calculate the percentage content of C₂₂H₂₅ClN₂O₈.

Bacterial endotoxins (2.6.14)

Maximum 0.5 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.0 g.
 exceeding 670 Pa for 3 h.
 60 °C over diphosphorus pentoxide R at a pressure not exceeding 670 Pa for 3 h.

Loss on drying (2.2.32)

Maximum 2.0 per cent, determined on 1.000 g by drying at 60 °C over diphosphorus pentoxide R at a pressure not exceeding 670 Pa for 3 h.

Heavy metals (2.4.8)

Maximum 50 ppm.
 0.5 g complies with test C. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

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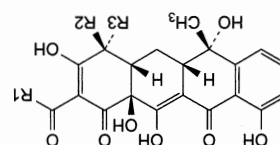
Maximum 50 ppm.
 0.5 g complies with test C. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

Maximum 50 ppm.
 0.5 g complies with test C. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

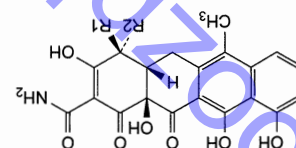
Maximum 50 ppm.
 0.5 g complies with test C. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

Maximum 50 ppm.
 0.5 g complies with test C. Prepare the reference solution using 2.5 mL of lead standard solution (10 ppm Pb) R.

IMPURITIES



A. R1 = NH₂, R2 = H, R3 = N(CH₃)₂; (4*R*,4*S*,5*S*,6*S*,12*S*)-4-(dimethylamino)-3,6,10,12-tetrahydro-1,1-dioxo-1,4,4a,5,5a,6,11,12a-pentahydroxy-6-methyl-1,12-dioxo-1,4,4a,5,12,12a-hexahydroretroterracene-2-carboxamide (4-epitetrazepam), (4-epianhydrotetrazepam), (4*S*,4*S*,5*S*,6*S*,12*S*)-2-acetyl-4-(dimethylamino)-3,6,10,12-pentahydroxy-6-methyl-4a,5a,6,12a-tetrahydroretroterracene-1,11(4*H*,5*H*)-dione (2-acetyl-2-decarbamoyletetrazepam),



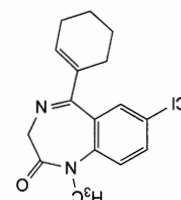
C. R1 = N(CH₃)₂, R2 = H; (4*S*,4*S*,12*S*)-4-(dimethylamino)-3,10,11,12a-tetrahydroxy-6-methyl-1,12-dioxo-1,4,4a,5,12,12a-hexahydroretroterracene-2-carboxamide (anhydrotetrazepam), D. R1 = H, R2 = N(CH₃)₂; (4*R*,4*S*,12*S*)-4-(dimethylamino)-3,10,11,12a-tetrahydroxy-6-methyl-1,12-dioxo-1,4,4a,5,12,12a-hexahydroretroterracene-2-carboxamide (4-epianhydrotetrazepam).

Ph Eur



Tetrazepam

(Ph. Eur. monograph 1738)

C₁₆H₁₇ClN₂O

288.8

10379-14-3

Action and use
Benzodiazepine; hypnotic.

Ph Eur

DEFINITION

7-Chloro-5-(cyclohex-1-enyl)-1-methyl-1,3-dihydro-2*H*-1,4-benzodiazepin-2-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Light yellow or yellow crystalline powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, soluble in acetonitrile.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of tetrazepam.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in acetonitrile *R* and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of the substance to be examined and 5.0 mg of tetrazepam impurity C CRS in acetonitrile *R* and dilute to 10.0 mL with the same solvent.

Dilute 1.0 mL of the solution to 10.0 mL with acetonitrile *R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with acetonitrile *R*. Dilute 1.0 mL of this solution to 10.0 mL with acetonitrile *R*.

Column: 10.0 mL with acetonitrile *R*.

Flow rate: 1.5 mL/min.

Size: 1 = 0.25 m, Ø = 4.6 mm;

Stationary phase: octadecylsilyl silica gel for chromatography *R*

Mobile phase:

mobile phase A: mix 40 volumes of acetonitrile *R* and 60 volumes of a 3.4 g/L solution of potassium dihydrogen phosphate *R*;

mobile phase B: acetonitrile *R*;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 35	100	0
35 - 40	100 → 55	0 → 45
40 - 50	55	45

Flow rate 1.5 mL/min.

Detection A spectrophotometer at 229 nm.

Injection 20 µL.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to tetrazepam and to impurity C.

Limits:

— any impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Chlorides (2.4.4)

Maximum 100 ppm.

Dissolve 0.750 g in 10 mL of methylene chloride *R* and add 15 mL of water *R*. Shake and separate the 2 layers. Dilute 10 mL of the aqueous layer to 15 mL with water *R*.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

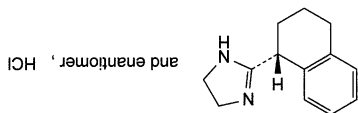
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.



Tetryzoline Hydrochloride

(Ph. Eur. monograph 2101)



C₁₃H₁₇ClN₂ 236.7 522-48-5

Action and use

Adrenoceptor agonist; decongestant.

Ph Eur

DEFINITION

2-[(1*R,S*)-1,2,3,4-Tetrahydronaphthalen-1-yl]-4,5-dihydro-1*H*-imidazole hydrochloride.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, in anhydrous ethanol and in ethanol (96 per cent), practically insoluble in acetone.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tetryzoline hydrochloride CRS.

B. Dissolve 50 mg in 10 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2).

Method II.

Dissolve 1.0 g in water R and dilute to 10 mL with the same solvent.

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 1.0 g of the substance to be examined in a mixture of 25 volumes of 1 M sodium hydroxide and 75 volumes of methanol R and dilute to 10 mL with the same mixture of solvents.

Reference solution Dilute 1.0 mL of the test solution to 100.0 mL with a mixture of 25 volumes of 1 M sodium hydroxide and 75 volumes of methanol R. Dilute 1.0 mL of this solution to 10.0 mL with a mixture of 25 volumes of 1 M sodium hydroxide and 75 volumes of methanol R.

Column:

material: fused silica,

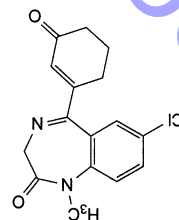
size: $l = 25$ m, $\varnothing = 0.32$ mm,

stationary phase: poly(dimethyl)siloxane R (1 μ m).

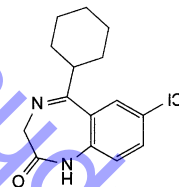
Carrier gas helium for chromatography R.

Split ratio 1:40.

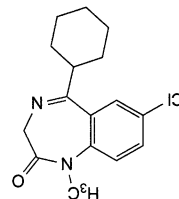
Flow rate 2.5 mL/min.



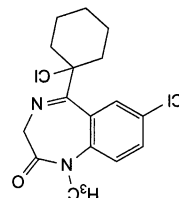
A. 7-chloro-1-methyl-5-(3-oxocyclohexyl)-1,3-dihydro-2H-1,4-benzodiazepin-2-one,



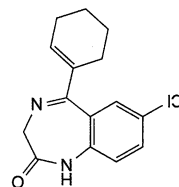
B. 7-chloro-5-cyclohexyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one,



C. 7-chloro-5-cyclohexyl-1-methyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one,



D. 7-chloro-5-(1-chlorocyclohexyl)-1-methyl-1,3-dihydro-2H-1,4-benzodiazepin-2-one,



E. 7-chloro-5-(cyclohex-1-enyl)-1,3-dihydro-2H-1,4-benzodiazepin-2-one.

Ph Eur

IMPURITIES

Protected from light.

STORAGE

C₁₆H₁₇ClN₂O.

1 mL of 0.1 M perchloric acid is equivalent to 28.88 mg of

potentiometrically (2.2.20).

Titrate with 0.1 M perchloric acid, determining the end-point

Dissolve 0.230 g in 50.0 mL of anhydrous acetic acid R.

ASSAY

Temperature:

Time (min)	Temperature (°C)
0 - 8	160
8 - 11	160 → 220
11 - 15	220
Injection port	220
Detector	220

Detection: Flame ionisation.

Injection: 1 µL.

Relative retention: With reference to tetrazoline (retention

time = about 12 min); impurity A = about 0.5.

System suitability: reference solution:

— signal-to-noise ratio: minimum 50 for the principal peak.

Limits:

— impurity A: not more than the area of the principal peak

in the chromatogram obtained with the reference solution

(0.1 per cent),

— any other impurity: not more than the

area of the principal peak in the chromatogram obtained

with the reference solution (0.1 per cent),

— total: not more than twice the area of the principal peak in

the chromatogram obtained with the reference solution

(0.2 per cent),

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with the reference solution

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 100 mL of a mixture of 3 volumes of

anhydrous acetic acid R and 7 volumes of acetic anhydride R.

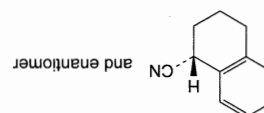
Titrate with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 23.67 mg

of C₁₃H₁₇CIN₂.**IMPURITIES**

Specified impurities: A.



A. (1RS)-1,2,3,4-tetrahydronaphthalene-1-carbonitrile (α-

cyanotetraline).

Ph Eur

Theobroma Oil

Cocoa Butter

DEFINITION

Theobroma Oil is the solid fat obtained from the roasted

seeds of *Theobroma cacao* L.**CHARACTERISTICS**

A yellowish white, solid fat; odour, slight, agreeable and

resembling that of cocoa. Somewhat brittle.

Freely soluble in ether and in petroleum spirit (boiling range,

40° to 60°); slightly soluble in ethanol (96%).

TESTS

Acid value

Not more than 4.0, Appendix X B.

Iodine value

35 to 40 (iodine monochloride method), Appendix X E.

Melting point

31° to 34°, Appendix V A, Method IV. Prepare the

substance being examined in the following manner. Melt

about 30 g in an oven at a temperature of 55° to 60° and

filter through a suitable dry filter paper, maintaining the

temperature between 53° and 60°. Cool with occasional

stirring until the temperature falls to between 32° and 34°.

Stir continuously with a mechanical stirrer until the first signs

of cloudiness appear and continue to stir by hand until the

substance has the consistency of a paste. Immediately

transfer to a vessel previously kept at a temperature of 15° to

22° and allow to stand at this temperature for 24 hours

before carrying out the test.

Refractive index

At 40°, 1.456 to 1.458, Appendix V E.

Saponification value

188 to 196, Appendix X G.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 100 mL of a mixture of 3 volumes of

anhydrous acetic acid R and 7 volumes of acetic anhydride R.

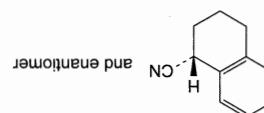
Titrate with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20). Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 23.67 mg

of C₁₃H₁₇CIN₂.**IMPURITIES**

Specified impurities: A.



A. (1RS)-1,2,3,4-tetrahydronaphthalene-1-carbonitrile (α-

cyanotetraline).

Ph Eur

DEFINITION

Theobromine contains not less than 99.0 per cent and not

more than the equivalent of 101.0 per cent of 3,7-dimethyl-

3,7-dihydro-1H-purine-2,6-dione, calculated with reference

to the dried substance.

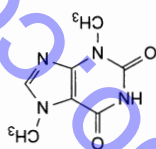
CHARACTERS

A white or almost white powder, very slightly soluble in

water and in anhydrous ethanol, slightly soluble in ammonia.

It dissolves in dilute solutions of alkali hydroxides and in

mineral acids.

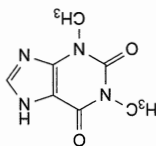


(Ph. Eur. monograph 0298)

Theobromine

Theophylline

(Ph. Eur. monograph 0299)



$C_7H_8N_4O_2$ 180.2 58-55-9

Action and use

Non-selective phosphodiesterase inhibitor (xanthine); treatment of reversible airways obstruction.

Preparations

Aminophylline Injection
Prolonged-release Theophylline Tablets

DEFINITION

1,3-Dimethyl-3,7-dihydro-1H-purine-2,6-dione.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in ethanol (96 per cent). It dissolves in solutions of alkali hydroxides, in ammonia and in mineral acids.

IDENTIFICATION

First identification B, D.

Second identification A, C, D, E.

A. Melting point (2.2.14): 270 °C to 274 °C, determined after drying at 100-105 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of theophylline.

C. Heat 10 mg with 1.0 mL of a 360 g/L solution of

potassium hydroxide R in a water-bath at 90 °C for 3 min,

then add 1.0 mL of diazotised sulfanilic acid solution R. A red

colour slowly develops. Carry out a blank test.

D. Loss on drying (see Tests).

E. It gives the reaction of xanthines (2.3.1).

TESTS

Solution S

Dissolve 0.5 g with heating in carbon dioxide-free water R, cool and dilute to 75 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity

To 50 mL of solution S add 0.1 mL of methyl red solution R. The solution is red. Not more than 1.0 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to yellow.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

IDENTIFICATION

First identification A, C.

Second identification A, C.

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with theophylline CRS.

B. Dissolve about 20 mg in 2 mL of dilute ammonia R1,

warming slightly, and cool. Add 2 mL of silver nitrate solution R2. The solution remains clear. Boil the solution for a few minutes. A white, crystalline precipitate is formed.

C. It gives the reaction of xanthines (2.3.1).

TESTS

Acidity

To 0.4 g add 20 mL of boiling water R and boil for 1 min. Allow to cool and filter. Add 0.05 mL of bromothymol blue solution R1. The solution is yellow or yellowish-green.

Not more than 0.2 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to blue.

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel GF₂₅₄ R as the coating substance.

Test solution To 0.2 g of the finely powdered substance to be

examined add 10 mL of a mixture of 4 volumes of

methanol R and 6 volumes of chloroform R. Heat under a

reflux condenser on a water-bath for 15 min, shaking

occasionally. Cool and filter.

Reference solution Dissolve 5 mg of theobromine CRS in a

mixture of 4 volumes of methanol R and 6 volumes of

chloroform R and dilute to 50 mL with the same mixture of

solvents.

Apply separately to the plate 10 µL of each solution. Develop

over a path of 15 cm using a mixture of 10 volumes of

concentrated ammonia R, 30 volumes of acetone R, 30 volumes

of chloroform R and 40 volumes of butanol R. Allow the plate

to dry in air and examine in ultraviolet light at 254 nm.

Any spot in the chromatogram obtained with the test

solution, apart from the principal spot, is not more intense

than the spot in the chromatogram obtained with the

reference solution (0.5 per cent).

Heavy metals (2.4.8)

1.0 g complies with test C for heavy metals (20 ppm).

Prepare the reference solution using 2 mL of lead standard

solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Not more than 0.5 per cent, determined on 1.000 g by

drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Not more than 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve, with stirring, 0.150 g in 125 mL of boiling water R,

cool to 50-60 °C and add 25 mL of 0.1 M silver nitrate.

Using 1 mL of phenolphthalein solution R as indicator, titrate

dropwise with 0.1 M sodium hydroxide until a pink colour is

obtained.

1 mL of 0.1 M sodium hydroxide is equivalent to 18.02 mg of

$C_7H_8N_4O_2$.

Ph Eur

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of theobromine R in the mobile phase, add 5 mL of the test solution and dilute to 100 mL with the mobile phase. Dilute 5 mL of this solution to 50 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (7 μ m).

Mobile phase Mix 7 volumes of acetonitrile for

chromatography R and 93 volumes of a 1.36 g/L solution of sodium acetate R containing 5.0 mL/L of glacial acetic acid R.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 20 μ L.

Run time 3.5 times the retention time of theophylline.

Relative retention With reference to theophylline (retention

time = about 6 min): impurity C = about 0.3;

impurity B = about 0.4; impurity D = about 0.5;

impurity A = about 2.5.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

Limits:

— impurities A, B, C, D: for each impurity, not more than

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— any other impurity: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.1 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 100 mL of water R, add 20 mL of 0.1 M

silver nitrate and shake. Add 1 mL of bromothymol blue

solution R1. Titrate with 0.1 M sodium hydroxide.

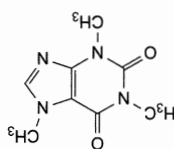
1 mL of 0.1 M sodium hydroxide is equivalent to 18.02 mg of

$C_7H_8N_4O_2$.

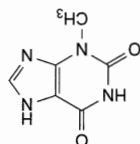
IMPURITIES

Specified impurities A, B, C, D

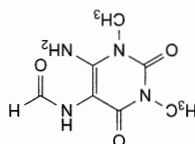
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these



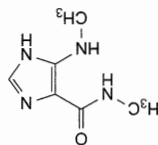
A. 1,3,7-trimethyl-3,7-dihydro-1H-purine-2,6-dione (caffeine),



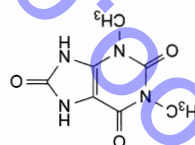
B. 3-methyl-3,7-dihydro-1H-purine-2,6-dione,



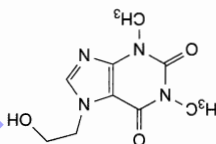
C. N-(6-amino-1,3-dimethyl-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-yl)formamide,



D. N-methyl-5-(methylamino)-1H-imidazole-4-carboxamide (theophyllidine),



E. 1,3-dimethyl-7,9-dihydro-1H-purine-2,6,8(3H)-trione,

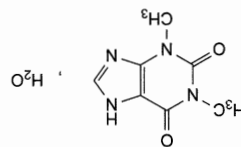


F. 7-(2-hydroxyethyl)-1,3-dimethyl-3,7-dihydro-1H-purine-2,6-dione (etofylline).

Theophylline Hydrate



(Theophylline Monohydrate, Ph Eur monograph 0302)


 $C_7H_8N_4O_2 \cdot H_2O$ 198.2 5967-84-0

Action and use

Non-selective phosphodiesterase inhibitor (xanthine);

treatment of reversible airways obstruction.

Preparations

Aminophylline Injection

Prolonged-release Theophylline Tablets

DEFINITION

1,3-Dimethyl-3,7-dihydro-1H-purine-2,6-dione monohydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in ethanol

(96 per cent). It dissolves in solutions of alkali hydroxides, in

IDENTIFICATION

First identification B, D.

Second identification A, C, D, E.

A. Melting point (2.2.14): 270 °C to 274 °C, determined

after drying at 100–105 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Dry the substance to be examined at 100–105 °C

before use.

Comparison Ph. Eur. reference spectrum of theophylline.

C. Heat 10 mg with 1.0 mL of a 360 g/L solution of

potassium hydroxide R in a water-bath at 90 °C for 3 min,

then add 1.0 mL of diazotised sulfanilic acid solution R. A red

colour slowly develops. Carry out a blank test.

D. Water (see Tests).

E. It gives the reaction of xanthenes (2.3.1).

TESTS

Solution S

Dissolve 0.5 g with heating in carbon dioxide-free water R, cool

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity

To 50 mL of solution S add 0.1 mL of methyl red solution R.

The solution is red. Not more than 1.0 mL of 0.01 M sodium

hydroxide is required to change the colour of the indicator to

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 40.0 mg of the substance to be

examined in the mobile phase and dilute to 20.0 mL with

the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of theobromine R in the

mobile phase, add 5 mL of the test solution and dilute to

100 mL with the mobile phase. Dilute 5 mL of this solution

to 50 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(7 μ m).

Mobile phase Mix 7 volumes of acetonitrile for

chromatography R and 93 volumes of a 1.36 g/L solution of

sodium acetate R containing 5.0 mL/L of glacial acetic acid R.

Flow rate 2.0 mL/min.

Detection Spectrophotometer at 272 nm.

Injection 20 μ L.

Run time 3.5 times the retention time of theophylline.

Relative retention With reference to theophylline (retention

time = about 6 min): impurity C = about 0.3;

impurity B = about 0.4; impurity D = about 0.5;

impurity A = about 2.5.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

theobromine and theophylline.

Limits:

— impurities A, B, C, D: for each impurity, not more than

the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— any other impurity: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.1 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

8.0 per cent to 9.5 per cent, determined on 0.20 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.160 g in 100 mL of water R, add 20 mL of 0.1 M

silver nitrate and shake. Add 1 mL of bromothymol blue

solution R1. Titrate with 0.1 M sodium hydroxide.

IMPURITIES

 $C_7H_8N_4O_2$.

Specified impurities A, B, C, D

Other detectable impurities (the following substances would, if

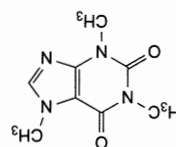
present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

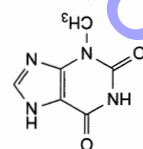
acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

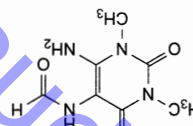
(2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): E, F.



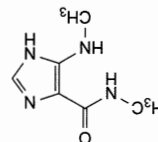
A. 1,3,7-trimethyl-3,7-dihydro-1H-purine-2,6-dione (caffeine).



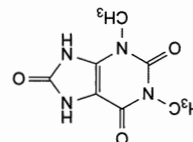
B. 3-methyl-3,7-dihydro-1H-purine-2,6-dione.



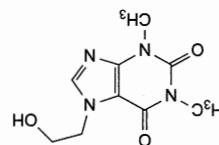
C. N-(6-amino-1,3-dimethyl-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-yl)formamide.



D. N-methyl-5-(methylamino)-1H-imidazole-4-carboxamide (theophylline).



E. 1,3-dimethyl-7,9-dihydro-1H-purine-2,6,8(3H)-trione.

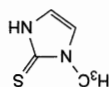


F. 7-(2-hydroxyethyl)-1,3-dimethyl-3,7-dihydro-1H-purine-2,6-dione (etofylline).

Ph Eur

Thiamazole

(Ph. Eur. monograph 1706)



C₄H₆N₂S

114.2

60-56-0

Action and use

Thionamide antithyroid.

Ph Eur

DEFINITION

1-Methyl-1,3-dihydro-2H-imidazole-2-thione.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or pale brown, crystalline powder.

Solubility

Freely soluble in water, freely soluble in methylene chloride, freely soluble or soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, C

Second identification A, B, D

A. Melting point (2.2.14): 143 °C to 146 °C.

B. Dissolve 25 mg in 10 mL of a 0.28 per cent V/V solution of sulfuric acid R and dilute to 50.0 mL with the same solution. Dilute 1.0 mL of this solution to 100.0 mL with a 0.28 per cent V/V solution of sulfuric acid R. Examined between 200 nm and 300 nm (2.2.25), the solution shows 2 absorption maxima, at 211 nm and 251 nm. The ratio of the absorbance measured at the absorption maximum at 251 nm to that measured at the absorption maximum at 211 nm is 2.5 to 2.7.

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison thiamazole CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5.0 mg of the substance to be examined in methanol R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of thiamazole CRS in methanol R and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dissolve 5.0 mg of 2-methylimidazole R in methanol R and dilute to 5.0 mL with the same solvent.

Dilute 1.0 mL of this solution to 2.0 mL with the test solution.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R1, 2-propanol R, toluene R (1:24:75 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— expose the plate to iodine vapour for 30 min;

the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal



spot in the chromatogram obtained with reference solution (a).

TESTS

Solution S

Dissolve 2.0 g in water R and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 0.100 g of the substance to be examined in chloroform R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with chloroform R. Dilute 1.0 mL of this solution to 10.0 mL with chloroform R.

Reference solution (b) Dissolve 5.0 mg of thiamazole impurity A CRS, 5.0 mg of 1-methylimidazole RI and 5.0 mg of thiamazole impurity C CRS in chloroform R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with chloroform R.

Column:

material: fused silica,
size: $l = 30.0$ m, $\varnothing = 0.25$ mm,
stationary phase: poly(dimethyl) (diphenyl)siloxane R with special deactivation for basic compounds (film thickness 0.5 μ m).

Carrier gas helium for chromatography R.

Flow rate 1.5 mL/min.

Split ratio 3:20.

Temperature:

Time (min)	Temperature (°C)	Column
0 - 2	100	100
2 - 7	100 → 250	250
7 - 22	250	150
Detector	250	

Detection Flame ionisation.

Injection 1 μ L.

Relative retention With reference to thiamazole (retention time = about 6.5 min): impurity A = about 0.3;

impurity B = about 0.4; impurity C = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to impurity A and impurity B.

Limits:

— impurities A, B, C: for each impurity, not more than the area of the corresponding peak in the chromatogram

obtained with reference solution (b) (0.1 per cent),

any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.1 per cent),

total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.5 per cent),

disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(0.02 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

12 mL of solution S complies with limit test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

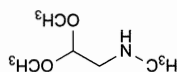
ASSAY

Dissolve 0.250 g in 75 mL of water R. Add 15.0 mL of 0.1 M sodium hydroxide, mix and add with stirring, about 30 mL of 0.1 M silver nitrate. Continue the titration with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

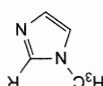
1 mL of 0.1 M sodium hydroxide is equivalent to 11.42 mg of C₁₂H₁₈N₄OS.

IMPURITIES

Specified impurities: A, B, C.



A. 2,2-dimethoxy-N-methylethanamine,

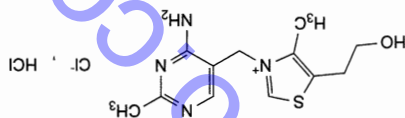


B. R = H: 1-methyl-1H-imidazole,

C. R = SCH₃: 1-methyl-2-(methylsulfanyl)-1H-imidazole.

Thiamine Hydrochloride

(Ph. Eur. monograph 0303)



C₁₂H₁₈Cl₂N₄OS

337.3

67-03-8

Action and use

Vitamin B₁.

Preparations

Thiamine Injection

Thiamine Tablets

Vitamins B and C Injection

DEFINITION

3-[(4-Amino-2-methylpyrimidin-5-yl)methyl]-5-

(2-hydroxyethyl)-4-methylthiazolium chloride hydrochloride.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless crystals.

Solubility

Freely soluble in water, soluble in glycerol, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, C

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison thiamine hydrochloride CRS.

If the spectra obtained show differences, dissolve the substance to be examined and the reference substance separately in water R, evaporate to dryness and record new spectra using the residues.

B. Dissolve about 20 mg in 10 mL of water R, add 1 mL of dilute acetic acid R and 1.6 mL of 1 M sodium hydroxide, heat on a water-bath for 30 min and allow to cool. Add 5 mL of dilute sodium hydroxide solution R, 10 mL of potassium ferricyanide solution R and 10 mL of butanol R and shake vigorously for 2 min. The upper alcoholic layer shows an intense light-blue fluorescence, especially in ultraviolet light at 365 nm. Repeat the test using 0.9 mL of 1 M sodium hydroxide and 0.1 g of anhydrous sodium sulfate R instead of 1.6 mL of 1 M sodium hydroxide. Practically no fluorescence is seen.

C. It gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 2.5 g in distilled water R and dilute to 25 mL with the same solvent.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₇ or GY₇ (2.2.2, Method II).

Dilute 2.5 mL of solution S to 5 mL with water R.

pH (2.2.3)

2.7 to 3.3.

Dilute 2.5 mL of solution S to 10 mL with water R.

Related substances

Liquid chromatography (2.2.29).

Solution A glacial acetic acid R, water R (5:95 V/V).

Test solution Dissolve 0.35 g of the substance to be examined in 15.0 mL of solution A and dilute to 100.0 mL with water R.

Reference solution (a) Dissolve 5 mg of the substance to be examined and 5 mg of thiamine impurity E CRS in 4 mL of solution A and dilute to 25.0 mL with water R. Dilute

5.0 mL of the solution to 25.0 mL with water R.

Reference solution (b) Dilute 1.0 mL of the test solution to 50.0 mL with water R. Dilute 5.0 mL of this solution to 25.0 mL with water R.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m) with a specific surface area of 350 m²/g and a pore size of 10 nm;

— temperature: 45 °C.

Mobile phase:

— mobile phase A: 3.764 g/L solution of sodium hexanesulfonate R adjusted to pH 3.1 with phosphoric acid R;

— mobile phase B: methanol R2;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 25	90 → 70	10 → 30
25 - 33	70 → 50	30 → 50
33 - 40	50	50
40 - 45	50 → 90	50 → 10

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 248 nm.

Injection 25 μ L.

Relative retention With reference to thiamine (retention time = about 30 min): impurity A = about 0.3;

impurity B = about 0.9; impurity C = about 1.2.

System suitability: reference solution (a):

— resolution: minimum 1.6 between the peaks due to impurity E and to thiamine.

Limits:

— any impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— total: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— disregard limit: 0.125 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Sulfates (2.4.13)

Maximum 300 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.400 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.110 g in 5 mL of anhydrous formic acid R and add 50 mL of acetic anhydride R. Titrate immediately with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20) and carrying out the titration within 2 min. Carry out a blank titration.

1 mL of 0.1 M perchloric acid is equivalent to 16.86 mg of C₁₂H₁₈Cl₂N₄O₅.

STORAGE

In a non-metallic container, protected from light.

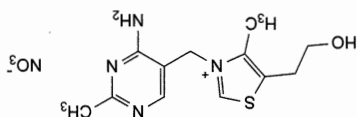
IMPURITIES

Specified impurities A, B, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, E, F, G, H.

Thiamine Nitrate

(Ph. Eur. monograph 0531)

 $C_{12}H_{17}N_5O_4S$

327.4

532-43-4

Action and use

Vitamin B₁.

Ph Eur

DEFINITION

3-[(4-Amino-2-methylpyrimidin-5-yl)methyl]-5-(2-hydroxyethyl)-4-methylthiazolium nitrate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or small, colourless crystals.

Solubility

Sparingly soluble in water, freely soluble in boiling water, slightly soluble in ethanol (96 per cent) and in methanol.

IDENTIFICATION

First identification: A, C.

Second identification: B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of thiamine nitrate.

B. Dissolve about 20 mg in 10 mL of water R, add 1 mL of dilute acetic acid R and 1.6 mL of 1 M sodium hydroxide, heat on a water-bath for 30 min and allow to cool. Add 5 mL of dilute sodium hydroxide solution R, 10 mL of potassium ferricyanide solution R and 10 mL of butanol R and shake vigorously for 2 min. The upper alcoholic layer shows an intense light-blue fluorescence, especially in ultraviolet light at 365 nm. Repeat the test using 0.9 mL of 1 M sodium hydroxide and 0.2 g of sodium sulfite R instead of 1.6 mL of 1 M sodium hydroxide. Practically no fluorescence is produced.

C. About 5 mg gives the reaction of nitrates (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

pH (2.2.3)

6.8 to 7.6 for solution S.

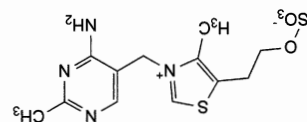
Related substances

Liquid chromatography (2.2.29).

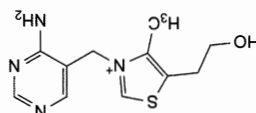
Solution A Add 5 volumes of glacial acetic acid R to 95 volumes of water R and mix.

Test solution Dissolve 0.35 g of the substance to be examined in 15.0 mL of solution A and dilute to 100.0 mL with water R.

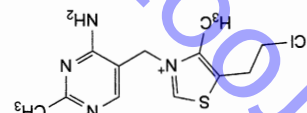
Reference solution (a) Dissolve 5 mg of the substance to be examined and 5 mg of thiamine impurity B CRS in 4 mL of



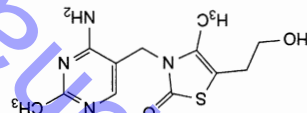
A. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-5-(2-sulfonatooxyethyl)thiazolium (thiamine sulfate ester),



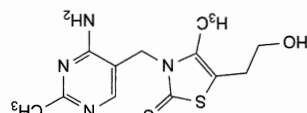
B. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-5-(2-hydroxyethyl)thiazolium (desmethythiamine),



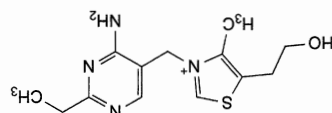
C. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-5-(2-chloroethyl)thiazolium (chlorothiamine),



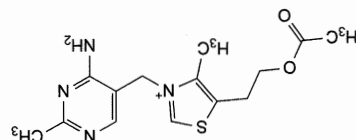
D. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-5-(2-hydroxyethyl)thiazol-2(3H)-one (oxothiamine),



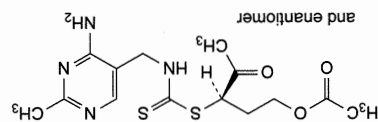
E. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-5-(2-hydroxyethyl)thiazol-2(3H)-thione (thioxothiamine),



F. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-5-(2-hydroxyethyl)thiazolium (ethythiamine),



G. 5-[2-(acetyloxyethyl)-3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methylthiazolium (acetythiamine),



H. (3R)-3-[[[(4-amino-2-methylpyrimidin-5-yl)methyl]thiocarbamoyl]sulfanyl]-4-oxopentyl acetate (ketodithiocarbamate).

Ph Eur



STORAGE

In a non-metallic container, protected from light.

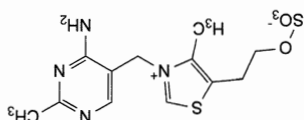
IMPURITIES

Specified impurities A, B, C

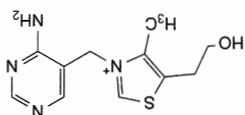
Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

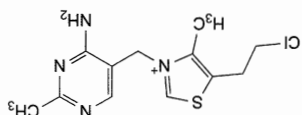
Control of impurities in substances for pharmaceutical use: D, E, F, G, H.



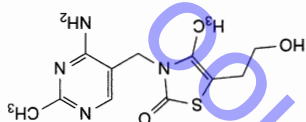
A. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methyl-[2-(sulfonatooxy)ethyl]thiazolium (thiamine sulfate ester),



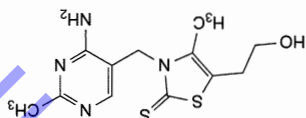
B. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-5-(2-hydroxyethyl)-4-methylthiazolium (desmethylthiamine),



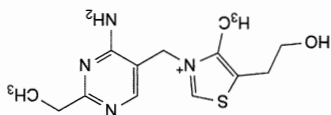
C. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-5-(2-chloroethyl)-4-methylthiazolium (chlorothiamine),



D. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-5-(2-hydroxyethyl)-4-methylthiazol-2(3H)-one (oxothiamine),



E. 3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-5-(2-hydroxyethyl)-4-methylthiazol-2(3H)-thione (thioxothiamine),



F. 3-[(4-amino-2-ethylpyrimidin-5-yl)methyl]-5-(2-hydroxyethyl)-4-methylthiazolium (ethylthiamine),

solution A and dilute to 25.0 mL with water R. Dilute

5.0 mL of this solution to 25.0 mL with water R.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with water R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: spherical end-capped octadecylsilyl silica gel

for chromatography R ($4\ \mu\text{m}$) with a specific surface area of $350\ \text{m}^2/\text{g}$ and a pore size of $10\ \text{nm}$;

— temperature: 45°C .

Mobile phase:

— mobile phase A: 3.764 g/L solution of sodium

hexanesulfonate R adjusted to pH 3.1 with phosphoric

acid R;

— mobile phase B: methanol R2;

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 25	90 \rightarrow 70	10 \rightarrow 30
25 - 33	70 \rightarrow 50	30 \rightarrow 50
33 - 40	50	50
40 - 45	50 \rightarrow 90	50 \rightarrow 10

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 248 nm.

Injection 25 μL .

Relative retention With reference to thiamine (retention

time = about 30 min): impurity A = about 0.3;

impurity B = about 0.9; impurity C = about 1.2.

System suitability: reference solution (a):

— resolution: minimum 1.6 between the peaks due to

impurity B and thiamine.

Limits:

— any impurity: for each impurity, not more than the area of

the principal peak in the chromatogram obtained with

reference solution (b) (1.0 per cent);

— total: not more than 1.5 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (1.5 per cent);

— disregard limit: 0.05 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent); disregard the peak due to the nitrate ion

at the beginning of the chromatogram.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven at 105°C .

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.140 g in 5 mL of anhydrous formic acid R and add

50 mL of acetic anhydride R. Titrate immediately with 0.1 M

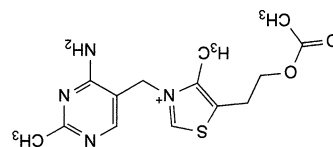
perchloric acid, determining the end-point potentiometrically

(2.2.20) and carrying out the titration within 2 min. Carry

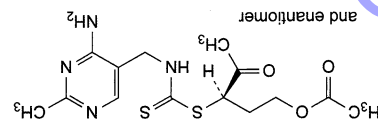
out a blank titration.

1.0 mL of 0.1 M perchloric acid is equivalent to 16.37 mg of

$\text{C}_{12}\text{H}_{17}\text{N}_5\text{O}_4\text{S}$.



C. 5-[2-(acetyloxy)ethyl]-3-[(4-amino-2-methylpyrimidin-5-yl)methyl]-4-methylthiazolium (acetylthiamine),

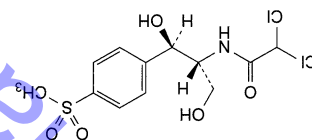


H. (3R,5)-3-[[[(4-amino-2-methylpyrimidin-5-yl)methyl]thiocarbamoyl]-4-oxopentyl]acetate (ketodithiocarbamate).

Ph Eur

Thiamphenicol

(Ph. Eur. monograph 0109)



C₁₂H₁₅Cl₂NO₅

356.2

15318-45-3

Action and use

Antibacterial.

Ph Eur

DEFINITION

2,2-Dichloro-N-[(1R,2R)-2-hydroxy-1-(hydroxymethyl)-2-[4-(methylsulfonyl)phenyl]ethyl]acetamide.

Content

98.0 per cent to 100.5 per cent (dried substance).

CHARACTERS

Appearance

Fine, white or yellowish-white, crystalline powder or crystals.

Solubility

Slightly soluble in water, very soluble in dimethylacetamide,

freely soluble in acetonitrile and in dimethylformamide,

soluble in methanol, sparingly soluble in acetone and in

anhydrous ethanol, slightly soluble in ethyl acetate.

A solution in anhydrous ethanol is dextrorotatory and a

solution in dimethylformamide is laevorotatory.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dry the substance to be examined and the

reference substance at 100-105 °C for 2 h; examine as discs

of potassium bromide R.

Comparison thiamphenicol CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.1 g of the substance to be examined

in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 0.1 g of thiamphenicol CRS in

methanol R and dilute to 10 mL with the same solvent.

Plate silica gel GF₂₅₄ R as the coating substance.

ASSAY

Dissolve 0.300 g in 30 mL of ethanol (96 per cent) R, add

20 mL of a 500 g/L solution of potassium hydroxide R, mix

and heat under a reflux condenser for 4 h. Cool, add

100 mL of water R, neutralise with dilute nitric acid R and

add 5 mL of the same acid in excess. Titrate with 0.1 M

silver nitrate, determining the end-point potentiometrically

(2.2.20), using a silver indicator electrode and a mercurous

sulfate reference electrode or any other appropriate electrode.

Carry out a blank test.

Maximum 0.1 per cent, determined on 2.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 1.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

using 1 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test C. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

Shake 0.5 g with 30 mL of water R for 5 min and filter.

Maximum 200 ppm.

Chlorides (2.4.4)

Maximum 200 ppm.

Shake 0.5 g with 30 mL of water R for 5 min and filter.

Maximum 10 ppm.

1.0 g complies with test C. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 1.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

using 1 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test C. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Melting point (2.2.14)

163 °C to 167 °C.

Absorbance (2.2.25)

Test solution (a) Dissolve 20 mg in water R, heating to about

40 °C, and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 2.5 mL of test solution (a) to

50.0 mL with water R.

Spectral range 240-300 nm for test solution (a); 200-240 nm

for test solution (b).

Absorption maxima At 266 nm and 273 nm for test

solution (a); at 224 nm for test solution (b).

Specific absorbances at the absorption maxima:

— at 266 nm: 25 to 28 for test solution (a),

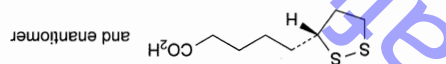
— at 273 nm: 21.5 to 23.5 for test solution (a),

— at 224 nm: 370 to 400 for test solution (b).



Thioctic Acid

(Ph. Eur. monograph 1648)



$C_8H_{14}O_2S_2$ 206.3 1077-28-7

Action and use
Antioxidant.

DEFINITION

5-[(3R_S)-1,2-Dithiolan-3-yl]pentanoic acid.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Very slightly soluble in water, very soluble in dimethylformamide, freely soluble in methanol.

mp

About 61 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison thioctic acid CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1).

Dissolve 0.50 g in a 20 g/L solution of sodium hydroxide R

and dilute to 10 mL with the same solution.

Impurity B

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.20 g of the substance to be examined in dimethylformamide R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of thioctic acid containing

impurity B CRS in dimethylformamide R and dilute to 1.0 mL with the same solvent (1.0 per cent impurity B solution).

Plate TLC silica gel plate R.

Mobile phase 25 per cent V/V solution of ammonia R, water R, ethyl acetate R, propanol R (5:10:40:40 V/V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 50 °C for 20 min.

Detection Expose to iodine vapour for 30 min or until the spots appear.

System suitability: reference solution:

— the chromatogram shows 2 clearly separated principal spots due to impurity B ($R_F = 0.0$) and thioctic acid ($R_F = \text{about } 0.3$).

1 mL of 0.1 M silver nitrate is equivalent to 17.81 mg of $C_{12}H_{15}Cl_2NO_5$.

STORAGE

In an airtight container, protected from light.

Ph Eur



Related substances
Liquid chromatography (2.2.29). Protect the solutions from light.

Solvent mixture A mixture of equal volumes of acetonitrile R1

and a 0.7 g/L solution of potassium dihydrogen phosphate R

previously adjusted to pH 3.6 with phosphoric acid R.

Test solution Dissolve 50.0 mg of the substance to be

examined in the solvent mixture and dilute to 50.0 mL with

the solvent mixture.

Reference solution (a) Dissolve 5 mg of thioctic acid for system

stability CRS (containing impurity A) in the solvent mixture

and dilute to 5 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 50.0 mg of thioctic acid CRS in

the solvent mixture and dilute to 50.0 mL with the solvent

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 µm);

— temperature: 35 °C.

Mobile phase Mix 8 volumes of acetonitrile R1, 41 volumes of

a 0.7 g/L solution of potassium dihydrogen phosphate R

previously adjusted to pH 3.0 with phosphoric acid R, and

51 volumes of methanol R.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 µL.

Run time 2.5 times the retention time of thioctic acid.

Relative retention With reference to thioctic acid (retention

time = about 6 min): impurity A = about 2.2.

System suitability:

— resolution: minimum 6.0 between the peaks due to thioctic

acid and impurity A in the chromatogram obtained with

reference solution (a);

— symmetry factor: maximum 2.0 for the peak due to thioctic

acid in the chromatogram obtained with reference

solution (c).

Limits:

— correction factor: for the calculation of content, multiply the

peak area of impurity A by 0.6;

— impurity A: not more than the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

Loss on drying (2.2.32)

Maximum 0.2 per cent, determined on 1.000 g by drying

in vacuo at 40 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (c).

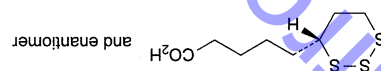
Calculate the percentage content of $C_8H_{14}O_2S_2$ from the peak areas and the declared content of thioctic acid CRS.

STORAGE

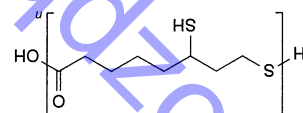
Protected from light.

IMPURITIES

Specified impurities A, B



A. 5-[(4R)-1,2,3-trithian-4-yl]pentanoic acid,

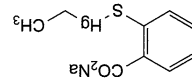


B. α -hydro- ω -hydroxy poly[sulfanediyl(3-sulfanyl-8-oxooctane-1,8-diyl)] (mixture of thioctic acid polymers).

Thiomersal

Thimerosal

(Ph. Eur. monograph 1625)



$C_8H_7HgNaO_2S$

404.8

54-64-8

Action and use

Antiseptic; adjuvant in vaccine formulations.

Ph. Eur.

DEFINITION

Sodium ethyl[2-sulfanylbzenzoato(2-)-O,5]mercurate(1-).

Content

97.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, sparingly soluble or soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Melting point (2.2.14): 103 °C to 115 °C.

Dissolve 0.5 g in water R and dilute to 10 mL with the same solvent. Add 2 mL of dilute hydrochloric acid R. A white precipitate is formed. Wash the precipitate with water R and dry over diphosphorus pentoxide R at a pressure not exceeding 0.7 kPa.

ASSAY

Place 0.5 g in a 100 mL long-necked combustion flask, add 5 mL of sulfuric acid R and heat gently until charring occurs.

exceeding 0.7 kPa for 24 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in a desiccator over diphosphorus pentoxide R at a pressure not exceeding 0.7 kPa for 24 h.

Loss on drying (2.2.32)

milligrams.

m_T = mass of the substance to be examined in solution in milligrams,

m_R = mass of mercuric chloride in the reference

$$\frac{(A_a - A_b - A_c - A_d - A_e) \times m_R \times 0.1847}{(A_a - A_b - A_c - A_d - A_e) \times m_T}$$

compounds, expressed as Hg from the expression:

liquid. Calculate the content of inorganic mercury

Measure the absorbance (2.2.25) of each solution (A_a , A_b , (blank preparation E).

potassium iodide R in flask E. Dilute to 10 mL with water R

preparation D). Place 5 mL of a 332 g/L solution of

potassium iodide R (test preparation B and reference

to 10 mL with a freshly prepared 332 g/L solution of

preparations A and C). Dilute the contents of flasks B and D

contents of flasks A and C to 10 mL with water R (blank

and D add 0.5 mL of the reference solution. Dilute the

solution in flasks A, B, C and D. To each of the flasks C

volumetric flasks A, B, C, D and E. Place 5 mL of the test

Test, reference and blank preparations Label five 10 mL

1.0 mL of the solution to 20.0 mL with water R.

Reference solution Dissolve 95.0 mg of mercuric chloride R in

in water R and dilute to 25.0 mL with the same solvent.

Test solution Dissolve 25 mg of the substance to be examined

Protect the solutions from light throughout the procedure.

Maximum 0.70 per cent.

Inorganic mercury compounds

Dilute 5 mL of solution S to 50 mL with carbon dioxide-free

6.0 to 8.0.

pH (2.2.3)

reference solution B₆ (2.2.2, Method II).

suspension II (2.2.1) and not more intensely coloured than

Solution S is not more opalescent than reference

Appearance of solution

25 mL with the same solvent.

Dissolve 2.0 g in carbon dioxide-free water R and dilute to

Solution S

TESTS

(2.3.1).

D. Solution S (see Tests) gives reaction (a) of sodium

further addition of acid, gives reaction (a) of sulfates (2.3.1).

hydrochloric acid R and filter. 5 mL of the filtrate, without

remaining part of the solution add 10 mL of dilute

solution gives reaction (a) of mercury (2.3.1). To the

solution add 5 mL of dilute nitric acid R. 0.1 mL of this

50 mL of water R to absorb the combustion products. To the

mixture of 1 mL of strong hydrogen peroxide solution R and

C. Treat 50 mg by the oxygen-flask method (2.5.10). Use a

Comparison thiomersal CRS.

B. Infrared absorption spectrophotometry (2.2.24).

— **impurity C**: not more than 6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (3.0 per cent);

— **impurity B**: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (1.0 per cent);

— **impurity D**: not more than 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— **unspecified impurities**: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **total**: not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (a) (5.0 per cent);

— **disregard limit**: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Chlorides (2.4.4)
Maximum 330 ppm.

To 5 mL of solution S add 35 mL of water R and 10 mL of dilute nitric acid R. Shake with 3 quantities, each of 25 mL, of 1,1-dimethylethyl methyl ether R and discard the upper layer. Eliminate the organic solvent from the lower layer by heating on a water-bath. 15 mL of the solution complies with the test for chlorides.

Loss on drying (2.2.32)
Maximum 2.5 per cent, determined on 0.500 g by drying *in vacuo* at 100 °C for 4 h.

Sodium ASSAY
Dissolve 0.400 g in 30 mL of water R. Add 0.1 mL of methyl red solution R and titrate with 0.1 M hydrochloric acid until a red colour is obtained. Boil gently for 2 min. Allow to cool and, if necessary, continue the titration with 0.1 M hydrochloric acid until the red colour is again obtained. 1 mL of 0.1 M hydrochloric acid is equivalent to 2.299 mg of Na.

Thiophental
Dissolve 0.150 g in 5 mL of water R. Add 2 mL of dilute sulfuric acid R and shake with 4 quantities, each of 10 mL, of chloroform R. Combine the chloroform layers, filter and evaporate the filtrate to dryness on a water-bath. Dissolve the residue in 30 mL of previously neutralised dimethylglyoxime R and add 0.1 mL of a 2 g/L solution of thymol blue R in methanol R. Titrate immediately with 0.1 M lithium methoxide until a blue colour is obtained. Protect the solution from atmospheric carbon dioxide during the titration. 1 mL of 0.1 M lithium methoxide is equivalent to 24.23 mg of $C_{11}H_{18}N_2O_2S$.

STORAGE

In an airtight container, protected from light.

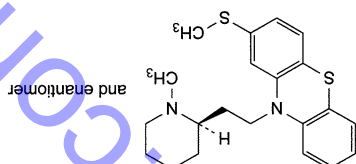
IMPURITIES

Specified impurities B, C, D

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

Thioridazine

(Ph. Eur. monograph 2005)



$C_{21}H_{26}N_2S_2$

370.6

50-52-2

Action and use

Dopamine receptor antagonist; neuroleptic.

CHARACTERS

Appearance

White or almost white powder.

Content

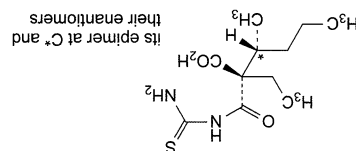
99.0 per cent to 101.0 per cent (dried substance).

10-[2-[(2*RS*)-1-Methylpiperidin-2-yl]ethyl]-2-(methylsulfanyl)-10*H*-phenothiazine



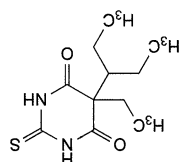
Ph. Eur.

D. mixture of (2*RS*,3*RS*)-2-(carbamothioylcarbamoyl)-2-ethyl-3-methylhexanoic acid and (2*RS*,3*SR*)-2-(carbamothioylcarbamoyl)-2-ethyl-3-methylhexanoic acid.

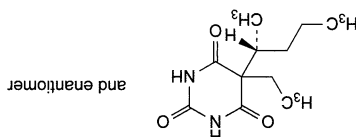


its epimer at C² and their enantiomers

C. 5-ethyl-5-(1-ethylpropyl)-2-thioxo-2,3-dihydropyrimidine-4,6(1*H*,5*H*)-dione,

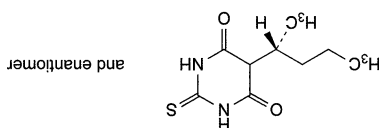


B. 5-ethyl-5-[(1*RS*)-1-methylbutyl]pyrimidine-2,4,6(1*H*,3*H*,5*H*)-trione,



and enantiomer

A. 5-[(1*RS*)-1-methylbutyl]-2-thioxo-2,3-dihydropyrimidine-4,6(1*H*,5*H*)-dione,



and enantiomer

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A.

Solubility

Practically insoluble in water, very soluble in methylene chloride, freely soluble in methanol, soluble in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison thioridazine CRS.

TESTS

Solution S
Dissolve 1.25 g in methanol R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Related substances

Liquid chromatography (2.2.29). Carry out the test as quickly as possible and protected from light.

Test solution Dissolve 20 mg of the substance to be examined in methanol R and dilute to 100 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with methanol R. Dilute 2.0 mL of this solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve the contents of a vial of thioridazine for system suitability CRS (containing impurities A, B, C, D and E) in 1.0 mL of methanol R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R resistant to bases up to pH 11.

Mobile phase:

— mobile phase A: triethylamine R1, acetonitrile R, water R (2:400:600 V/V/V);

— mobile phase B: triethylamine R1, acetonitrile R (2:1000 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	100	0
5 - 35	100 \rightarrow 5	0 \rightarrow 95
35 - 40	5	95

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 25 μ L.

Identification of impurities Use the chromatogram supplied with thioridazine for system suitability CRS and the chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, B, C, D and E. **Relative retention** With reference to thioridazine (retention time = about 30 min): impurity D = about 0.1;

impurity A = about 0.3; impurity C = about 0.4;

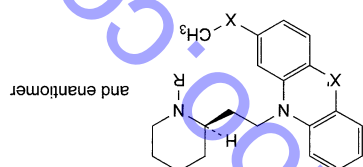
impurity B = about 0.5; impurity E = about 0.6.

System suitability: reference solution (b):

— resolution: minimum 3.5 between the peaks due to impurities C and B.

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 1.9; impurity B = 2.4; impurity C = 0.5; impurity D = 1.5;



Specified impurities A, B, C, D, E.
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): F.

IMPURITIES

Protected from light.

STORAGE

of C₂₁H₂₆N₂S₂.

1 mL of 0.1 M perchloric acid is equivalent to 37.06 mg

potentiometrically (2.2.20).

with 0.1 M perchloric acid, determining the end-point

Dissolve 0.300 g in 60 mL of anhydrous acetic acid R. Titrate

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

50 °C for 4 h.

Maximum 0.5 per cent, determined on 1.000 g in vacuo at

Loss on drying (2.2.32)

using 2 mL of lead standard solution (10 ppm Pb) R.

1.0 g complies with test C. Prepare the reference solution

Maximum 20 ppm.

Heavy metals (2.4.8)

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

(0.5 per cent);

in the chromatogram obtained with reference solution (a)

— total: not more than 5 times the area of the principal peak

with reference solution (a) (0.10 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than

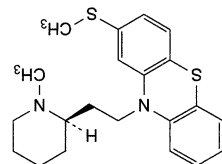
the area of the principal peak in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— impurities A, B, C, D, E: for each impurity, not more than

Thioridazine Hydrochloride

(Ph. Eur. monograph 0586)



and enantiomer, HCl

$C_{21}H_{27}ClN_2S_2$ 407.0 130-61-0

Action and use

Dopamine receptor antagonist; neuroleptic.

Ph Eur

DEFINITION

10-[2-[(2R)-1-Methylpiperidin-2-yl]ethyl]-2-(methylsulfonyl)-10H-phenothiazine hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in methanol, soluble in ethanol

96 per cent.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison thioridazine hydrochloride CRS.

B. 0.2 g gives reaction (b) of chlorides (2.3.1).

TESTS

Carry out all operations protected from light.

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than intensity 6 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Dissolve 1.0 g in methanol R and dilute to 20 mL with the same solvent.

Optical rotation (2.2.7)

−0.10° to +0.10°.

Dissolve 1.0 g in methanol R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test as quickly as possible and protected from light.

Test solution Dissolve 20.0 mg of the substance to be examined in methanol R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with methanol R. Dilute 2.0 mL of this solution to 100.0 mL with methanol R.

Reference solution (b) Dissolve the contents of a vial of thioridazine for system suitability CRS (containing impurities A, B, C, D and E) in 1.0 mL of methanol R.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m) resistant to bases up to pH 11.

STORAGE

Protected from light.

of $C_{21}H_{27}ClN_2S_2$.

1 mL of 0.1 M perchloric acid is equivalent to 40.70 mg

ASSAY

Dissolve 0.300 g in a mixture of 10 mL of anhydrous acetic acid R and 60 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C for 4 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

Maximum 20 ppm.

Dissolve 1.0 g in 20 mL of water R. 12 mL of the solution

complies with test A. Prepare the reference solution using

lead standard solution (1 ppm Pb) R.

Heavy metals (2.4.8)

Maximum 20 ppm.

(0.05 per cent).

the chromatogram obtained with reference solution (a)

— disregard limit: 0.5 times the area of the principal peak in

(0.5 per cent);

in the chromatogram obtained with reference solution (a)

— total: not more than 5 times the area of the principal peak

with reference solution (a) (0.1 per cent);

area of the principal peak in the chromatogram obtained

— unspecified impurities: for each impurity, not more than the

obtained with reference solution (a) (0.1 per cent);

the area of the principal peak in the chromatogram

— impurities A, B, C, D, E: for each impurity, not more than

impurity B = 2.4; impurity C = 0.5; impurity D = 1.5;

corresponding correction factor: impurity A = 1.9;

the peak areas of the following impurities by the

— correction factors: for the calculation of content, multiply

Limits:

impurities C and B.

— resolution: minimum 3.5 between the peaks due to

System suitability: reference solution (b):

impurity F = about 0.9.

impurity B = about 0.5; impurity E = about 0.6;

impurity A = about 0.3; impurity C = about 0.4;

time = about 30 min; impurity D = about 0.1;

Relative retention With reference to thioridazine (retention

due to impurities A, B, C, D and E.

Identification of impurities Use the chromatogram supplied

with thioridazine for system suitability CRS to identify the peaks

Injection 25 μ L.

Detection Spectrophotometer at 275 nm.

Flow rate 1.0 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	100	0
5 - 35	100 \rightarrow 5	0 \rightarrow 95
35 - 40	5	95

(2:1000 V/V);

— mobile phase B: triethylamine R1, acetonitrile R

(2:400:600 V/V/V);

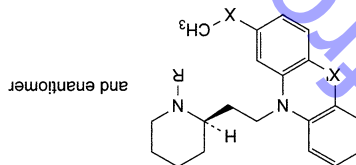
— mobile phase A: triethylamine R1, acetonitrile R, water R

Mobile phase:

IMPURITIES

Specified impurities A, B, C, D, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): F.



A. R = CH₃, X = X' = SO₂; 10-[2-[(2RS)-1-methylpiperidin-2-yl]ethyl]-2-(methylsulfonyl)-10H-phenothiazine 5,5-dioxide,

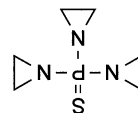
B. R = CH₃, X = SO, X' = S; 10-[2-[(2RS)-1-methylpiperidin-2-yl]ethyl]-2-(methylsulfonyl)-10H-phenothiazine,

C. R = CH₃, X = S, X' = SO; 10-[2-[(2RS)-1-methylpiperidin-2-yl]ethyl]-2-(methylsulfonyl)-10H-phenothiazine 5-oxide,

D. R = CH₃, X = X' = SO; 10-[2-[(2RS)-1-methylpiperidin-2-yl]ethyl]-2-(methylsulfonyl)-10H-phenothiazine 5-oxide,

E. R = CH₃, X = SO₂, X' = S; 10-[2-[(2RS)-1-methylpiperidin-2-yl]ethyl]-2-(methylsulfonyl)-10H-phenothiazine 5-oxide,

F. R = H, X = X' = S; 2-(methylsulfonyl)-10-[2-[(2RS)-piperidin-2-yl]ethyl]-10H-phenothiazine.

Thiotepa

C₆H₁₂N₃P₃S

189.2

52-24-4

Action and use

Cytotoxic alkylating agent.

Preparation

Thiotepa Injection

DEFINITION

Thiotepa is phosphorothioic tri(ethyleneamide). It contains not less than 97.0% and not more than 102.0% of C₆H₁₂N₃P₃S, calculated with reference to the anhydrous substance.

CHARACTERISTICS

Fine, white crystalline flakes.

Freely soluble in water, in chloroform and in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of thiotepa (RS 337).

TESTS**Melting point**

52° to 57°, Appendix V A.

Clarity of solution

A 2.0% w/v solution is clear, Appendix IV A.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following freshly prepared solutions. Solutions (1) and (2) are solutions of the substance being examined in water containing 0.350% w/v and 0.00035% w/v, respectively. For solution (3) dissolve 10 mg of the substance being examined in 2 mL of methanol in a ground-glass-stoppered tube, add 50 µL of a 0.1% v/v solution of orthophosphoric acid, stopper the tube and heat in a water bath at 65° for 50 seconds (generation of methoxythiotepa). Allow the solution to cool and add 1 mL of methanol. For solution (4) dissolve 15 mg of the substance being examined in 10 mL of water, add 1 g of sodium chloride, boil in a water bath for 10 minutes and cool (generation of chloro-adduct).

The chromatographic procedure may be carried out using (a) a stainless steel column (15 cm × 4.6 mm) packed with end-capped octadecylsilyl silica gel for chromatography (5 µm) (Nucleosil C18 is suitable), (b) 15 volumes of acetonitrile and 85 volumes of 0.1M mixed phosphate buffer pH 7.0 as the mobile phase with a flow rate of 1 mL per minute and (c) a detection wavelength of 215 nm.

The chromatogram obtained with solution (3) shows a peak corresponding to methoxythiotepa with a retention time relative to thiotepa of about 1.3 and the chromatogram obtained with solution (4) shows a peak due to the chloro-adduct with a retention time relative to thiotepa of about 3.75. The test is not valid unless the resolution factor between the two principal peaks in the chromatogram obtained with solution (3) is at least 3.

For solution (1) allow the chromatography to proceed for 4 times the retention time of the principal peak. In the chromatogram obtained with solution (1) the area of any peak corresponding to the 'chloro-adduct' (identified from the peak in the chromatogram obtained with solution (4)) is not greater than 1.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.15%), the area of any other secondary peak is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.1%) and the sum of the areas of all the secondary peaks is not greater than twice the area of the principal peak in the chromatogram obtained with solution (2) (0.2%).

Water

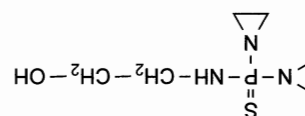
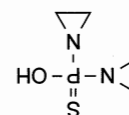
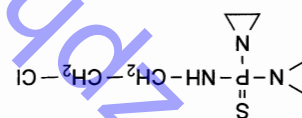
Not more than 0.5% w/w, Appendix IX C. Cool the reagents and titration vessel in ice throughout the procedure and use 1.2 g. Complete the procedure as quickly as possible.

ASSAY

Transfer 0.2 g to an iodine flask with the aid of 50 mL of a 20% w/v solution of sodium thiosulfate and titrate immediately with 0.1M hydrochloric acid VS, using 0.05 mL of methyl orange solution as indicator, until a faint red colour persists for 10 seconds. Stopper the flask, allow to stand for 30 minutes and titrate with 0.1M sodium hydroxide VS using phenolphthalein solution R1 as indicator. Subtract the volume of 0.1M sodium hydroxide VS used from the volume of 0.1M hydrochloric acid VS used. Repeat the operation without the substance being examined. The difference between the titrations represents the amount of hydrochloric acid required. Each mL of 0.1M hydrochloric acid VS is equivalent to 6.307 mg of $C_4H_9NO_3$.

STORAGE

Thiotepa should be stored at a temperature of 2° to 8°. At higher temperatures it polymerises and becomes inactive.

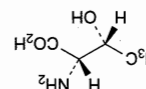
IMPURITIES

C, 'hydroxythiotepa B';

B, 'hydroxythiotepa A';

Threonine

(Ph. Eur. monograph 1049)



$C_4H_9NO_3$

119.1

72-19-5

Action and use
Amino acid.

DEFINITION

(2S,3R)-2-Amino-3-hydroxybutanoic acid.
Fermentation product, extract or hydrolysate of protein.
99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder or colourless crystals.

Solubility

Soluble in water, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

C. Comparison threonine CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a 1 per cent V/V solution of hydrochloric acid R and dilute to 50 mL with the same solution.

Reference solution Dissolve 10 mg of threonine CRS in a 1 per cent V/V solution of hydrochloric acid R and dilute to 50 mL with the same solution.

50 mL with the same solution.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).

Application 5 µL.

Development: Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C for 15 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Mix 1 mL of a 2 g/L solution of the substance to be examined and 1 mL of a 20 g/L solution of sodium periodate R. Add 0.2 mL of piperidine R and 0.1 mL of a 25 g/L solution of sodium nitroprusside R. A blue colour develops that changes to yellow after a few minutes.

TESTS**Solution S**

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

5.0 to 6.5 for solution S.

Specific optical rotation (2.2.7)

−29.0 to −27.6 (dried substance).

Dissolve 1.50 g in water R and dilute to 25.0 mL with the same solvent.

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method I.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of proline R in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of ammonium standard solution (100 ppm NH_4) R to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of isoleucine R (impurity D) and 30 mg of leucine R in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability Reference solution (d):
— resolution: minimum 1.5 between the peaks due to impurity D and leucine.
Calculation of percentage contents:

— for any ninhydrin-positive substance detected at 570 nm, use the concentration of threonine in reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b);

— if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:

— any ninhydrin-positive substance: for each impurity, maximum 0.2 per cent;
— total: maximum 0.5 per cent;
— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

Chlorides (2.4.4)

Maximum 200 ppm.
Dilute 10 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.
Dissolve 0.5 g in distilled water R and dilute to 15 mL with the same solvent.

Ammonium

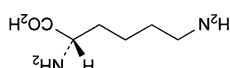
Amino acid analysis (2.2.56) as described in the test for ninhydrin-positive substances with the following modifications.
Injection Test solution, reference solution (c) and blank solution.

Limit:

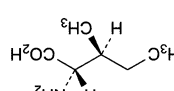
— ammonium at 570 nm: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (2.4.9)

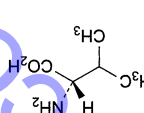
Maximum 10 ppm.



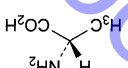
E. (2S)-2,6-diaminohexanoic acid (lysine).



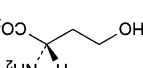
D. (2S,3S)-2-amino-3-methylpentanoic acid (isoleucine).



C. (2S)-2-amino-3-methylbutanoic acid (valine).



A. (2S)-2-amino-4-hydroxybutanoic acid (homoserine).



B. (2S)-2-aminopropanoic acid (alanine).

IMPURITIES
Protected from light.
STORAGE
 $\text{C}_4\text{H}_9\text{NO}_3$.
1 mL of 0.1 M perchloric acid is equivalent to 11.91 mg of (2.2.20).
Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically.

ASSAY

Dissolve 0.100 g in 5 mL of anhydrous formic acid R. Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.
an oven at 105 °C.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Heavy metals (2.4.8)

Maximum 10 ppm.
0.5 g complies with test G. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

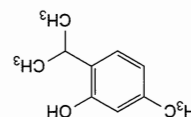
Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Heavy metals (2.4.8)
Maximum 10 ppm.
0.5 g complies with test G. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

In a separating funnel, dissolve 1.0 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min in each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Thymol

(Ph. Eur. monograph 0791)



$C_{10}H_{14}O$ 150.2 89-83-8

DEFINITION

5-Methyl-2-(methylcetyl)phenol.

CHARACTERS

Appearance
Colourless crystals.

Solubility

Very slightly soluble in water, very soluble in ethanol (96 per cent), freely soluble in essential oils and in fatty oils, sparingly soluble in glycerol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification B.
Second identification A, C, D.

A. Melting point (2.2.14): 48 °C to 52 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Dissolve 0.2 g with heating in 2 mL of dilute sodium hydroxide solution R and add 0.2 mL of chloroform R. Heat on a water-bath. A violet colour develops.

D. Dissolve about 2 mg in 1 mL of anhydrous acetic acid R. Add 0.15 mL of sulfuric acid R and 0.05 mL of nitric acid R. A bluish-green colour develops.

TESTS

The solution is not more opalescent than reference suspension IV (2.2.1) and not more intensely coloured than reference solution R₆ (2.2.2, Method II).

Acidity

To 1.0 g in a 100 mL glass-stoppered conical flask add 20 mL of water R. Boil until dissolution is complete, cool and stopper the flask. Shake vigorously for 1 min. Add a few crystals of the substance to be examined to initiate crystallisation. Shake vigorously for 1 min and filter. To 5 mL of the filtrate add 0.05 mL of methyl red solution R and 0.05 mL of 0.01 M sodium hydroxide. The solution is yellow.

Related substances

Gas chromatography (2.2.28).
Test solution Dissolve 0.100 g of the substance to be examined in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1 mL of the test solution to 100 mL with ethanol (96 per cent) R.

Reference solution (b) Dilute 1 mL of reference solution (a) to 10 mL with ethanol (96 per cent) R.

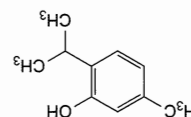
Reference solution (c) Dilute 5 mL of reference solution (b) to 10 mL with ethanol (96 per cent) R.

Column:

— material: glass or steel;

Thymol

(Ph. Eur. monograph 0791)



$C_{10}H_{14}O$ 150.2 89-83-8

DEFINITION

5-Methyl-2-(methylcetyl)phenol.

CHARACTERS

Appearance
Colourless crystals.

Solubility

Very slightly soluble in water, very soluble in ethanol (96 per cent), freely soluble in essential oils and in fatty oils, sparingly soluble in glycerol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification B.
Second identification A, C, D.

A. Melting point (2.2.14): 48 °C to 52 °C.

B. Infrared absorption spectrophotometry (2.2.24).

C. Dissolve 0.2 g with heating in 2 mL of dilute sodium hydroxide solution R and add 0.2 mL of chloroform R. Heat on a water-bath. A violet colour develops.

D. Dissolve about 2 mg in 1 mL of anhydrous acetic acid R. Add 0.15 mL of sulfuric acid R and 0.05 mL of nitric acid R. A bluish-green colour develops.

TESTS

The solution is not more opalescent than reference suspension IV (2.2.1) and not more intensely coloured than reference solution R₆ (2.2.2, Method II).

Acidity

To 1.0 g in a 100 mL glass-stoppered conical flask add 20 mL of water R. Boil until dissolution is complete, cool and stopper the flask. Shake vigorously for 1 min. Add a few crystals of the substance to be examined to initiate crystallisation. Shake vigorously for 1 min and filter. To 5 mL of the filtrate add 0.05 mL of methyl red solution R and 0.05 mL of 0.01 M sodium hydroxide. The solution is yellow.

Related substances

Gas chromatography (2.2.28).
Test solution Dissolve 0.100 g of the substance to be examined in ethanol (96 per cent) R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dilute 1 mL of the test solution to 100 mL with ethanol (96 per cent) R.

Reference solution (b) Dilute 1 mL of reference solution (a) to 10 mL with ethanol (96 per cent) R.

Reference solution (c) Dilute 5 mL of reference solution (b) to 10 mL with ethanol (96 per cent) R.

Column:

— material: glass or steel;

Temperature:

Flow rate 30 mL/min.

Carrier gas nitrogen for chromatography R.

the separation of free fatty acids.

— chromatography R₅ impregnated with a mixture suitable for

— stationary phase: diatomaceous earth for gas

— size: l = 4 m, Ø = 2 mm;

Temperature:

Flow rate 30 mL/min.

Carrier gas nitrogen for chromatography R.

the separation of free fatty acids.

— chromatography R₅ impregnated with a mixture suitable for

— stationary phase: diatomaceous earth for gas

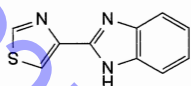
— size: l = 4 m, Ø = 2 mm;

STORAGE

Protected from light.

Tiabendazole

(Ph. Eur. monograph 0866)

 $C_{10}H_7N_3S$

201.2

148-79-8

Action and use

Benzimidazole antihelminthic.

Preparation

Tiabendazole Chewable Tablets

DEFINITION

Tiabendazole contains not less than 98.0 per cent and not more than the equivalent of 101.0 per cent of 2-(thiazol-4-yl)-1H-benzimidazole, calculated with reference to the anhydrous substance.

CHARACTERS

A white or almost white, crystalline powder, practically insoluble in water, slightly soluble in alcohol and in

methylene chloride. It dissolves in dilute mineral acids. It melts at about 300 °C.

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Dissolve 25 mg in 0.1 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 2.0 mL of the solution to 100.0 mL with 0.1 M hydrochloric acid. Examined between 230 nm and 350 nm (2.2.25), the solution shows two absorption maxima, at 243 nm and 302 nm. The ratio of the absorbance measured at the maximum at 302 nm to that measured at the maximum at 243 nm is 1.8 to 2.1.

B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with tabendazole CRS. Examine the substances prepared as discs. C. Examine the chromatograms obtained in the test for related substances in ultraviolet light at 254 nm. The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. Dissolve about 5 mg in 0.1 M hydrochloric acid and dilute to 5 mL with the same acid. Add 3 mg of *p*-phenylenediamine dihydrochloride R and shake until dissolved. Add 0.1 g of zinc powder R, mix, allow to stand for 2 min and add 5 mL of ferric ammonium sulfate solution R2. A bluish-violet colour develops.

TESTS

Related substances

Examine by thin-layer chromatography (2.2.27), using silica gel HF₂₅₄ R as the coating substance.

Test solution (a) Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Test solution (b) Dilute 2 mL of test solution (a) to 20 mL with methanol R.

Reference solution (a) Dissolve 20 mg of tabendazole CRS in methanol R and dilute to 20 mL with the same solvent.

Reference solution (b) Dilute 1 mL of test solution (b) to 10 mL with methanol R.

Reference solution (c) Dilute 1 mL of test solution (b) to 25 mL with methanol R.

Apply separately to the plate 20 µL of each solution. Develop over a path of 15 cm using a mixture of 2.5 volumes of water R, 10 volumes of acetone R, 25 volumes of glacial acetic acid R and 62.5 volumes of toluene R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b)

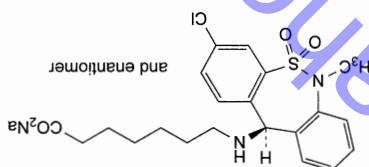
(1.0 per cent) and at most one such spot is more intense than the spot in the chromatogram obtained with reference solution (c) (0.4 per cent).

o-Phenylenediamine

To 5.0 g in a flask fitted with a ground-glass stopper, add 25 mL of a mixture of 1 volume of methanol R and 2 volumes of water R. Shake for 3 min. Filter through a sintered-glass filter (16) (2.1.2) under reduced pressure. To 10 mL of the filtrate add 0.5 mL of hydrochloric acid R and 0.5 mL of acetylacetone R and shake until the solution is clear. The solution is not more intensely coloured than reference solution R₇ (2.2.2, Method I) (10 ppm).

Tianeptine Sodium

(Ph. Eur. monograph 2022)



C₂₁H₂₄ClN₂NaO₄S 458.9

30123-17-2

Action and use

Antidepressant.

Ph. Eur.

DEFINITION

Sodium 7-[[[(1*R*,5*S*)-3-chloro-6-methyl-6,1,1-dihydroindeno[1,2-b]thiazepin-1-yl]amino]heptanoate S,S-dioxide.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish powder, very hygroscopic.

Solubility

Freely soluble in water, in methanol and in methylene chloride.

IDENTIFICATION

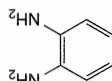
A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of tianeptine sodium.

B. It gives reaction (a) of sodium (2.3.1).

IMPURITIES

Store protected from light.



A. benzene-1,2-diamine.

Ph. Eur.



TESTS

Impurity A

Gas chromatography (2.2.28).

Internal standard solution Dilute 1 mL of ethyl

5-bromovalerate R in ethanol R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of the solution to 250.0 mL with ethanol R.

Test solution Dissolve 0.1000 g of the substance to be examined in the internal standard solution and dilute to 2.0 mL with the same solution.

Reference solution Dissolve 10.0 mg of tianeptine impurity A CRS in the internal standard solution and dilute to 200.0 mL with the same solution.

Column:

material: fused silica,

size: $l = 25$ m, $\varnothing = 0.25$ mm,

stationary phase: poly(cyanopropyl)siloxane R (film thickness 0.2 μ m).

Carrier gas helium for chromatography R.

Linear velocity 26 cm/s.

Split ratio 1:100.

Temperature:

column: 150 °C,

injection port and detector: 210 °C.

Detection Flame ionisation.

Injection 1 μ L.

Run time Twice the retention time of ethyl 5-bromovalerate.

System suitability: reference solution:

elution order: ethanol, ethyl 5-bromovalerate, impurity A,

resolution: minimum 10 between the peaks due to ethyl

5-bromovalerate and impurity A,

signal-to-noise ratio: minimum 20 for the peak due to

impurity A.

Limit:

impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with the reference

solution (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Mix 50 volumes of methanol R and 50 volumes of water for chromatography R.

Test solution Dissolve 50.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 20.0 mL with the solvent mixture.

Reference solution (b) Dissolve 20.0 mg of sodium tianeptine for system suitability CRS in the solvent mixture and dilute to 200.0 mL with the solvent mixture.

Column:

size: $l = 0.15$ m, $\varnothing = 4.6$ mm,

stationary phase: octadecylsilyl silica gel for chromatography R

(3 μ m) with a pore size of 0.01 μ m,

temperature: 30 °C.

Mobile phase:

mobile phase A: mix 21 volumes of methanol R1,

31.5 volumes of acetonitrile R1 and 47.5 volumes of a

2 g/L solution of sodium laurilsulfate R, adjusted to pH 2.5,

with phosphoric acid R,

mobile phase B: mix 20 volumes of methanol R1,

20 volumes of a 2 g/L solution of sodium laurilsulfate R,

adjusted to pH 2.5 with phosphoric acid R and 60 volumes of acetonitrile R1,

adjusted to pH 2.5 with phosphoric acid R and 60 volumes

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 35	100	0
35 - 45	100 → 40	0 → 60
45 - 60	40	60
60 - 70	40 → 100	60 → 0

Flow rate 1 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L.

Relative retention with reference to tianeptine (retention time = about 30 min): impurity C = about 0.4;

impurity D1 = about 0.6; impurity D2 = about 0.8;

impurity E = about 1.1; impurity B = about 1.7.

System suitability: reference solution (b):

resolution: minimum 2.5 between the peaks due to

tianeptine and impurity B.

Limits:

any impurity: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (a) (0.1 per cent),

total: not more than 8 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.4 per cent),

disregard limit: area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.05 per cent).

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.100 g.

ASSAY

Dissolve 0.165 g in 50 mL of anhydrous acetic acid R. Titrate

with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

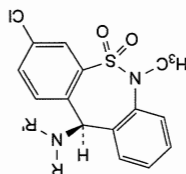
1 mL of 0.1 M perchloric acid is equivalent to 22.95 mg of

STORAGE

In an airtight container.

IMPURITIES

A. Br-[CH₂]₆-CO-O-C₂H₅: ethyl 7-bromohexanoate,



B. R = H, R' = [CH₂]₆-CO-O-C₂H₅: ethyl 7-[[[(11R,5S)-3-chloro-6-methyl-6,11-dihydrodibenzothiazepin-1-yl]amino]heptanoate 5,5-dioxide,
E. R = R' = [CH₂]₆-CO₂H: 7,7'-[[[(11R,5S)-3-chloro-6-methyl-6,11-dihydrodibenzothiazepin-1-yl]imino]dihexanoic acid 5,5-dioxide,

Reference solution Dissolve 20.0 mg of metoclopramide impurity E CRS (impurity C) in methanol R and dilute to 20 mL with the same solvent. Dilute 2.0 mL of the solution to 20 mL with methanol R.

Plate: TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, dioxan R, methanol R, methylene chloride R (2:10:14:90 V/V/V/V).

Application 10 µL.

Development Over 4/5 of the plate.

Drying In air.

Detection Spray with a 2 g/L solution of ninhydrin R in butanol R and heat at 100 °C for 15 min.

Retention factors Impurity C = about 0.1; tiapride = about 0.6.

Limit:

— **impurity C:** any spot due to impurity C is not more intense than the corresponding spot in the chromatogram obtained with the reference solution (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of tiapride hydrochloride CRS and 5.0 mg of napide N-oxide CRS

(impurity D) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:

— **size:** l = 0.25 m, Ø = 4.6 mm;

— **stationary phase:** base-deactivated end-capped octylsilyl silica gel for chromatography R (5 µm);

— **temperature:** 40 °C.

Mobile phase Dissolve 5.44 g of potassium dihydrogen phosphate R and 0.08 g of sodium octanesulfonate R in 780 mL of water R, adjust to pH 2.7 with phosphoric acid R and dilute to 800 mL with water R; add 150 mL of methanol R and 50 mL of acetonitrile R and mix.

Detection Spectrophotometer at 240 nm.

Injection 10 µL.

Run time 3 times the retention time of tiapride.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to

impurity D.

Relative retention With reference to tiapride (retention time = about 7 min): impurity D = about 1.2.

System suitability: reference solution (b):

— **resolution:** minimum 4.0 between the peaks due to tiapride and impurity D.

Calculation of percentage contents:

— for each impurity, use the concentration of tiapride in reference solution (a).

Limits:

— **unspecified impurities:** for each impurity, maximum 0.10 per cent;

— **total:** maximum 0.2 per cent;

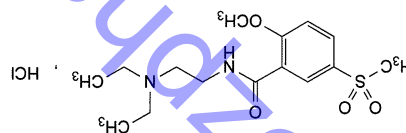
— **reporting threshold:** 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Tiapride Hydrochloride

(Ph. Eur. monograph 1575)



C₁₅H₂₅ClN₂O₄S 364.9 51012-33-0

Action and use

Dopamine receptor antagonist; neuroleptic.

Ph. Eur.

DEFINITION

N-[2-(Diethylamino)ethyl]-2-methoxy-5-(methylsulfonyl)benzamide hydrochloride.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, soluble in methanol, slightly soluble in anhydrous ethanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tiapride hydrochloride CRS.

B. Solution S (see Tests) gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and its absorbance (2.2.25) at 450 nm is not greater than 0.030.

pH (2.2.3)

4.0 to 6.0 for solution S.

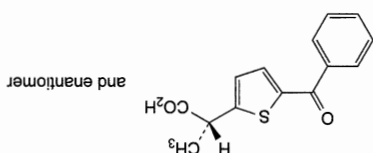
Impurity C

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.400 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Tiaprofenic Acid

(Ph. Eur. monograph 1157)



$C_{14}H_{12}O_3$ 260.3 33005-95-7

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

DEFINITION

(2*R,S*)-2-(5-Benzoylthiophen-2-yl)propanoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, freely soluble in acetone, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification C

Second identification A, B, D

A. Melting point (2.2.14): 95 °C to 99 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 25.0 mg in ethanolic hydrochloric acid R and dilute to 50.0 mL with the same solvent. Dilute 1.0 mL of this solution to 50.0 mL with ethanolic hydrochloric acid R.

Spectral range 220–350 nm.

Absorption maximum At 305 nm.

Shoulder At 262 nm.

Specific absorbance at the absorption maximum 550 to 590.

C. Infrared absorption spectrophotometry (2.2.24).

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 10 mg of naprofenic acid CRS in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of ketoprofen CRS in methylene chloride R and dilute to 10 mL with the same solvent. Dilute 1 mL of this solution to 2 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase acetic acid R, methylene chloride R, acetone R (1:20:80 V/V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 20 mL of anhydrous acetic acid R.

Add 20 mL of acetic anhydride R. Titrate with 0.1 M

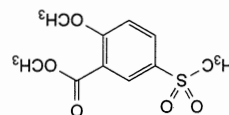
perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 36.49 mg

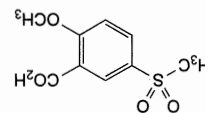
of $C_{15}H_{25}ClN_2O_4S$.

IMPURITIES

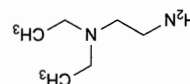
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D.



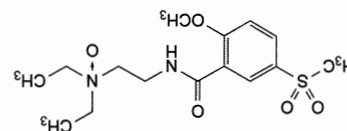
A. methyl 2-methoxy-5-(methylsulfonyl)benzoate,



B. 2-methoxy-5-(methylsulfonyl)benzoic acid,



C. N,N-diethylethane-1,2-diamine,



D. N-[2-(diethylamino)ethyl]-2-methoxy-5-(methylsulfonyl)benzamide (napride N-oxide).

Ph Eur

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Dissolve 2.0 g in ethanol (96 per cent) R and dilute to 20 mL with the same solvent.

Optical rotation (2.2.7)

Dissolve 0.50 g in ethyl acetate R and dilute to 10.0 mL with the same solvent.

Related substances

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 5.0 mL of reference solution (a) to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 10.0 mg of tiaprofenic acid impurity C CRS in the mobile phase and dilute to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 50.0 mL with the mobile phase.

Reference solution (d) Dilute 1.0 mL of reference solution (a) to 2.0 mL with reference solution (c).

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: silica gel for chromatography R (5 μ m).

Mobile phase water R, glacial acetic acid R, hexane R, methylene chloride R (0.25:20:500:500 V/V/V/V); add the water to the acetic acid, then hexane and methylene chloride; sonicate the mixture for 2 min. Do not degas with helium during analysis.

Detection Spectrophotometer at 250 nm.

Injection 20 μ L.

Run time Twice the retention time of tiaprofenic acid.

Relative retention With reference to tiaprofenic acid:

impurity A = about 0.19; impurity B = about 0.43;
impurity C = about 0.86.

System suitability Reference solution (d):

— resolution: minimum 3.0 between the peaks due to impurity C and tiaprofenic acid.

Limits:

— impurity C: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— sum of impurities other than C: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.
2.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 60 °C at a pressure not exceeding 0.7 kPa for 3 h. Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

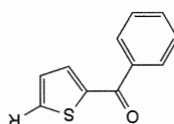
Dissolve 0.250 g in 25 mL of ethanol (96 per cent) R. Add 25 mL of water R and 0.5 mL of phenolphthalein solution R. Titrate with 0.1 M sodium hydroxide. 1 mL of 0.1 M sodium hydroxide is equivalent to 26.03 mg of $C_{14}H_{12}O_3S$.

STORAGE

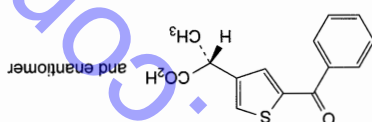
Protected from light.

IMPURITIES

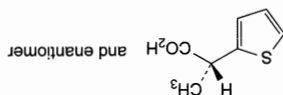
Specified impurities C
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, D, E, F.



A. R = C_2H_5 ; (5-ethylthiophen-2-yl)phenylmethanone,
B. R = $CO-CH_3$; 1-(5-benzoylthiophen-2-yl)ethanone,
F. R = Br; (5-bromothiophen-2-yl)phenylmethanone,



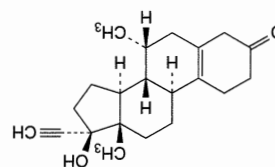
C. (2R,5R)-2-(5-benzoylthiophen-3-yl)propanoic acid,
D. benzoic acid,



E. (2R,5R)-2-(thiophen-2-yl)propanoic acid.

Tibolone

(Ph. Eur. monograph 1739)

 $C_{21}H_{28}O_2$ 312.5

Action and use
Steroid with estrogenic and progestogenic properties.

Preparation

Tibolone Tablets

Ph. Eur.

DEFINITION

17-Hydroxy-7 α -methyl-19-nor-17 α -pregn-5(10)-en-20-yn-3-one.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or crystals.

Solubility

Practically insoluble in water, soluble in acetone and in methanol.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of tibolone.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined in the minimum volume of anhydrous ethanol R, evaporate to dryness on a water-bath and record a new spectrum using the residue.

TESTS

Specific optical rotation (2.2.7)

+ 100 to + 106 (dried substance).

Dissolve 0.250 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Select a brand of acetonitrile such that the formation of possible artefact peaks, eluting after impurity C at relative retentions 0.6 to 0.8, is avoided.

Merck acetonitrile is suitable.

Solvent mixture water R, acetonitrile R1 (25:75 V/V).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 4 mg of tibolone for system suitability CRS (containing impurities A, B, C, D and E) in 1 mL of the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;— stationary phase: spherical end-capped octylsilyl silica gel for chromatography R (5 μ m).

IMPURITIES

At a temperature of 2 °C to 8 °C.

STORAGE

 $C_{21}H_{28}O_2$.

1 mL of 0.1 M sodium hydroxide is equivalent to 31.25 mg of sodium hydroxide, determining the end-point potentiometrically (2.2.20).

Dissolve 0.250 g in 60 mL of tetrahydrofuran R. Add 25 mL of a 100 g/L solution of silver nitrate R. Titrate with 0.1 M

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C for 3 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

(0.05 per cent).

— the chromatogram obtained with reference solution (b) disregard limit: 0.05 times the area of the principal peak in (1.0 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— chromatogram obtained with reference solution (b) 0.1 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than reference solution (b) (0.2 per cent);

— principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurity D: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— impurity C: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity B: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity A: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity B = 1.5; impurity C = 2.1;

— impurity A = 1.7;

— corresponding correction factor: impurity A = 1.7;

— the peak areas of the following impurities by the correction factors: for the calculation of content, multiply

— correction factors: for the calculation of content, multiply

— impurity A: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity B: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity C: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— impurity D: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— the chromatogram obtained with reference solution (b) disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— chromatogram obtained with reference solution (b) 0.1 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than reference solution (b) (0.2 per cent);

— principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurity D: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— impurity C: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity B: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity A: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— the chromatogram obtained with reference solution (b) disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— chromatogram obtained with reference solution (b) 0.1 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than reference solution (b) (0.2 per cent);

— principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurity D: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— impurity C: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity B: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity A: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— the chromatogram obtained with reference solution (b) disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— chromatogram obtained with reference solution (b) 0.1 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than reference solution (b) (0.2 per cent);

— principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— impurity D: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.4 per cent);

— impurity C: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity B: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— impurity A: for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

Solubility

Freely soluble in water, soluble in methanol.

IDENTIFICATION

First identification A, D, E

Second identification B, C, D.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 50 mg of the substance to be examined in 1 mL of water R, add 0.1 mL of hydrochloric acid R1, swirl and allow to stand in iced water for 10 min. Filter the precipitate and rinse with 2 mL of water R. Dissolve in a mixture of 1 volume of water R and 9 volumes of acetone R. Evaporate the solvent almost to dryness, then dry in an oven at 60 °C for 30 min.

Comparison Repeat the operations using ticarcillin monosodium CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 25 mg of ticarcillin monosodium CRS in methanol R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of carbenticillin sodium CRS and 25 mg of ticarcillin monosodium CRS in methanol R and dilute to 5 mL with the same solvent.

Plate TLC silanised silica gel plate R.

Mobile phase Mix 10 volumes of acetone R and 90 volumes of a 154 g/L solution of ammonium acetate R, adjusted to pH 5.0 with glacial acetic acid R.

Application 1 µL.

Development Over a path of 12 cm.

Drying In a current of hot air.

Detection Expose to iodine vapour.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Place about 2 mg in a test-tube about 15 cm long and add 2 mL of sulfuric acid-formaldehyde reagent R. Mix the contents of the tube by swirling; the solution is brown. Place the test-tube in a water-bath for 1 min; a dark reddish-brown colour develops.

D. It gives reaction (a) of sodium (2.3.1).

E. Specific optical rotation (see Tests).

TESTS**Solution S**

Dissolve 2.50 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

pH (2.2.3)

5.5 to 7.5 for solution S.

Specific optical rotation (2.2.7)

+ 172 to + 187 (anhydrous substance).
Dissolve 0.250 g in water R and dilute to 25.0 mL with the same solvent.

CHARACTERS**Appearance**

White or slightly yellow, hygroscopic powder.

Content

89.0 per cent to 102.0 per cent (anhydrous substance).

Semi-synthetic product derived from a fermentation product.

azabicyclo[3.2.0]heptane-2-carboxylate.

3-yl)acetyl]amino]-3,3-dimethyl-7-oxo-4-thia-1-

Disodium (2S,5R,6R)-6-[[[(2R)-2-carboxylato-2-(thiophen-

DEFINITION

Ph Eur

Ticarcillin and Clavulanic Acid Infusion

Preparation

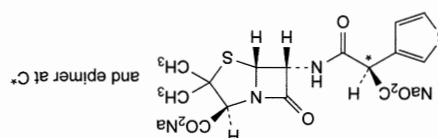
Penicillin antibacterial.

Action and use

C₁₅H₁₄N₂Na₂O₆S₂

428.4

4697-14-7



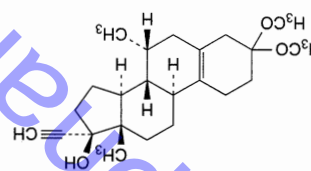
(Ph. Eur. monograph 0956)

Ticarcillin Sodium

Ph Eur

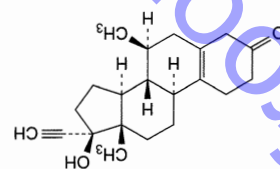
yn-17-ol.

E. 3,3-dimethoxy-7α-methyl-19-nor-17α-pregn-5(10)-en-20-



3-one,

D. 17-hydroxy-7β-methyl-19-nor-17α-pregn-5(10)-en-20-yn-



en-20-yn-3-one,

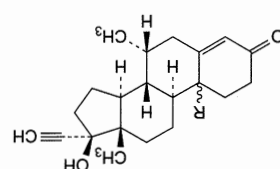
C. R = H: 17-hydroxy-7α-methyl-19-nor-10β,17α-pregn-4-

nor-10β,17α-pregn-4-en-20-yn-3-one,

B. R = O-OH: 10-hydroxy-7α-methyl-19-

pregn-4-en-20-yn-3-one,

A. R = OH: 10,17-dihydroxy-7α-methyl-19-nor-10β,17α-



Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 25.0 mL with mobile phase A.
Reference solution (a) Dissolve 20.0 mg of ticarcillin impurity A CRS in mobile phase A and dilute to 100.0 mL with mobile phase A. Dilute 5.0 mL of this solution to 50.0 mL with mobile phase A.
Reference solution (b) Dilute 1 mL of the test solution to 50.0 mL with mobile phase A.

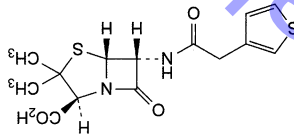
Column:
 — size: $l = 0.25$ m, $\varnothing = 4$ mm;
 — stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).
Mobile phase:
 — mobile phase A: 1.3 g/L solution of ammonium phosphate R adjusted to pH 7.0 with phosphoric acid R;
 — mobile phase B: methanol R, mobile phase A (50:50 V/V);
Detection Spectrophotometer at 220 nm.
Injection 20 μ L.
System suitability: reference solution:
 — resolution: minimum 2.5 between the 2 principal peaks;
 — repeatability: maximum relative standard deviation of 1.0 per cent for the 2 peaks due to ticarcillin after 6 injections.
 Calculate the percentage content of ticarcillin sodium as the sum of the areas of the 2 peaks, multiplying the content of ticarcillin monosodium by 1.054.

STORAGE

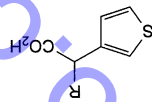
In an airtight container, at a temperature of 2 °C to 8 °C. If the substance is sterile, store in a sterile, tamper-proof container.

IMPURITIES

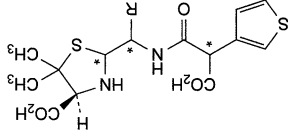
Specified impurities A
 Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E.



A. (2S,5R,6R)-3,3-dimethyl-7-oxo-6-[[[(thiophen-3-yl)acetyl]amino]-4-thia-1-azabicyclo[3.2.0]heptane-2-carboxylic acid (decarboxyticarcillin),



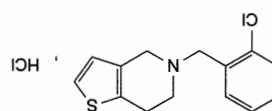
B. R = H: (thiophen-3-yl)acetic acid,
 C. R = CO₂H: 2-(thiophen-3-yl)propanedioic acid (3-thienylmalonic acid),



D. R = CO₂H: (4S)-2-[carboxy[[[2-carboxy-2-(thiophen-3-yl)acetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of ticarcillin),
 E. R = H: (4S)-2-[[[2-carboxy-2-(thiophen-3-yl)acetyl]amino]methyl]-5,5-dimethylthiazolidine-4-carboxylic acid (penilloic acids of ticarcillin).

Ticlopidine Hydrochloride

(Ph. Eur. monograph 1050)

C₁₄H₁₅ClN₂S 300.2 53885-35-1

Action and use

Inhibition of adenosine diphosphate (ADP)-mediated platelet aggregation; antiplatelet drug.

Ph. Eur.

DEFINITION

5-(2-Chlorobenzyl)-4,5,6,7-tetrahydrothieno[3,2-c]pyridine hydrochloride.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparingly soluble in water and in anhydrous ethanol, very slightly soluble in ethyl acetate.

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution (a) Dissolve 40 mg in water R and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 5.0 mL of test solution (a) to 100.0 mL with water R.

Spectral range 250-350 nm for test solution (a); 200-350 nm for test solution (b).

Absorption maxima At 268 nm and 275 nm for test solution (a); at 214 nm and 232 nm for test solution (b).

Absorption ratio $A_{268}/A_{275} = 1.1$ to 1.2 .

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

C. Mix about 6 mg of citric acid R and 0.3 mL of acetic anhydride R. Add about 5 mg of the substance to be examined and heat in a water-bath at 80 °C. A red colour develops.

D. About 20 mg gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.5 g in a 1 per cent V/V solution of hydrochloric acid R and dilute to 20 mL with the same solution.

pH (2.2.3)

3.5 to 4.0.

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).



Heavy metals (2.4.8)
Maximum 10 ppm.

Dissolve 2.0 g in a 85 per cent V/V solution of methanol R and dilute to 20.0 mL with the same solvent. 12 mL of the solution complies with test B. Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

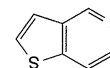
ASSAY

Dissolve 0.150 g in 15 mL of anhydrous acetic acid R. Add 35 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 30.02 mg of $C_{14}H_{15}Cl_2NS$.

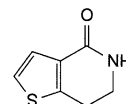
IMPURITIES

Specified impurities F.

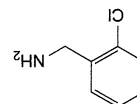
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, G, H, I, J, K, L.



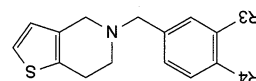
A. thieno[3,2-c]pyridine,



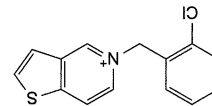
B. 6,7-dihydrothieno[3,2-c]pyridin-4(5H)-one,



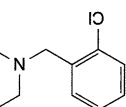
C. (2-chlorophenyl)methanamine,



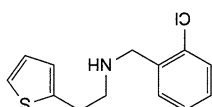
D. R3 = R4 = H: 5-benzyl-4,5,6,7-tetrahydrothieno[3,2-c]pyridine,
G. R3 = Cl, R4 = H: 5-(3-chlorobenzyl)-4,5,6,7-tetrahydrothieno[3,2-c]pyridine,
H. R3 = H, R4 = Cl: 5-(4-chlorobenzyl)-4,5,6,7-tetrahydrothieno[3,2-c]pyridine,



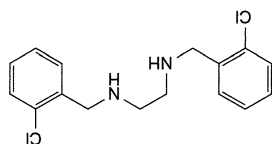
E. 5-(2-chlorobenzyl)thieno[3,2-c]pyridinium,



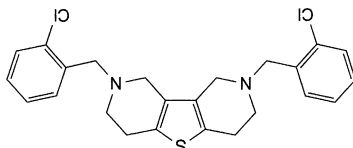
F. 6-(2-chlorobenzyl)-4,5,6,7-tetrahydrothieno[2,3-c]pyridine,



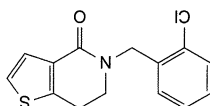
I. N-(2-chlorobenzyl)-2-(thiophen-2-yl)ethanamine,



J. N,N'-bis(2-chlorobenzyl)ethane-1,2-diamine,



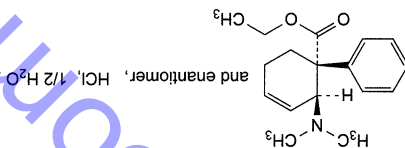
K. 2,8-bis(2-chlorobenzyl)-1,2,3,4,6,7,8,9-octahydrothieno[3,2-c:4,5-c']dipyridine (bis-ticlopidine),



L. 5-(2-chlorobenzyl)-6,7-dihydrothieno[3,2-c]pyridin-4(5H)-one.

Tilidine Hydrochloride Hemihydrate

(Ph. Eur. monograph 1767)



$C_{17}H_{24}ClNO_2 \cdot \frac{1}{2}H_2O$ 318.9

Action and use
Opioid Analgesic.

Ph Eur

DEFINITION

EtHyl (1R,5S,2SR)-2-(dimethylamino)-1-phenylcyclohex-3-enecarboxylate hydrochloride hemihydrate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).
A suitable antioxidant may be added.

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Freely soluble in water, very soluble in methylene chloride, freely soluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of tilidine hydrochloride hemihydrate.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method II).

Acidity or alkalinity

To 20 mL of solution S add 0.2 mL of 0.01 M sodium hydroxide. The pH is not less than 4.1. Add 0.4 mL of 0.01 M hydrochloric acid. The pH is not more than 4.3.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dilute 0.5 mL of the test solution to 100.0 mL with water R.

Reference solution (b) Dilute 2.0 mL of reference solution (a) to 10.0 mL with water R.

Precolumn:

— size: $l = 4$ mm, $\varnothing = 4.0$ mm;

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m).

Column:

— size: $l = 0.125$ m, $\varnothing = 4.0$ mm;

— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix equal volumes of acetonitrile R and a 0.98 g/L solution of ammonium carbonate R.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L.

Run time Twice the retention time of tilidine.

Relative retention With reference to tilidine (retention time = about 11 min): impurity C = about 0.5;

impurity B = about 0.7; impurity A = about 1.5.

Limits:

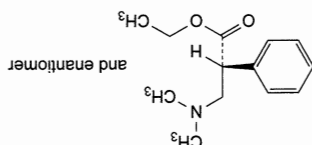
— impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— total: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent);

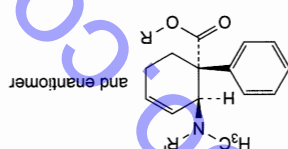
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

D. ethyl (2*RS*)-3-dimethylamino-2-phenylpropionate.

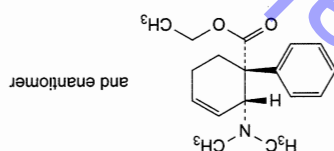


phenylcyclohex-3-enecarboxylate, C. R = C₂H₅, R' = H: ethyl (1*RS*,2*SR*)-2-(methylamino)-1-

phenylcyclohex-3-enecarboxylate, B. R = R' = CH₃: methyl (1*RS*,2*SR*)-2-(dimethylamino)-1-



A. ethyl (1*RS*,2*RS*)-2-(dimethylamino)-1-phenylcyclohex-3-enecarboxylate,



Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D.

IMPURITIES

Specified impurities A, B, C

STORAGE

Protected from light.

1 mL of 0.1 M perchloric acid is equivalent to 30.99 mg of C₁₇H₂₄ClNO₂.

Dissolve 0.250 g in a mixture of 10 mL of anhydrous acetic acid R and 50 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

ASSAY

procedure for the removal of bacterial endotoxins. Less than 0.25 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate Bacterial endotoxins (2.6.14)

2.5 per cent to 3.1 per cent, determined on 0.300 g.

Water (2.5.12)

lead standard solution (2 ppm Pb) R.

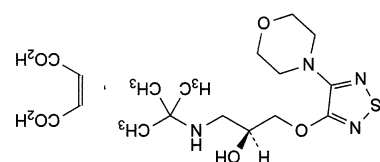
Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using

Maximum 20 ppm.

Heavy metals (2.4.8)

Timolol Maleate

(Ph. Eur. monograph 0572)

 $C_{17}H_{28}N_4O_5$

432.5

26921-17-5

Action and use

Beta-adrenoceptor antagonist.

For Timolol Eye Drops only: treatment of glaucoma.

Preparations

Dorzolamide and Timolol Eye Drops

Timolol Eye Drops

Timolol Tablets

Ph Eur

DEFINITION

(2*S*)-1-[(1,1-Dimethylethyl)amino]-3-[[4-(morpholin-4-yl)-1,2,5-thiadiazol-3-yl]oxy]propan-2-ol (*Z*)-butenedioate.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless crystals.

Solubility

Soluble in water and in ethanol (96 per cent).

mp: about 199 °C, with decomposition.

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Specific optical rotation (2.2.7): -6.2 to -5.7.
Dissolve 1.000 g in 1 M hydrochloric acid and dilute to 10.0 mL with the same acid.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison timolol maleate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.
Reference solution Dissolve 5 mg of timolol maleate CRS in methanol R and dilute to 5 mL with the same solvent.Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase concentrated ammonia R, methanol R, methylene chloride R (1:20:80 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour for 2 h.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

D. Titrate 0.1 g with a mixture of 1 mL of dilute sodium hydroxide solution R and 3 mL of water R. Shake with 3 quantities, each of 5 mL, of ether R. To 0.1 mL of the



Enantiomeric purity

Liquid chromatography (2.2.29). Carry out the test protected from actinic light.

Solvent mixture methylene chloride R, 2-propanol R (10:30 V/V).

Test solution Dissolve 30.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Reference solution (a) Dissolve 30 mg of timolol maleate CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 3 mg of (*R*)-timolol CRS (impurity A) in the solvent mixture and dilute to 10.0 mL with the solvent mixture. Dilute 1.0 mL of the solution to 10.0 mL with the solvent mixture.Reference solution (c) Dilute 1 mL of reference solution (a) to 100 mL with the solvent mixture. Mix 1 mL of this solution with 1 mL of reference solution (b).
Reference solution (d) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture.Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: cellulose derivative of silica gel for chiral separation R (5 µm).

Mobile phase diethylamine R, 2-propanol R, hexane R (2:40:960 V/V/V).

Flow rate 1 mL/min.

Detection Spectrophotometer at 297 nm.

Injection 5 µL.

Elution order Impurity A is eluted first.

System suitability:

— resolution: minimum 4.0 between the peaks due to impurity A and the (*S*)-enantiomer in the chromatogram obtained with reference solution (c);
— the retention times of the principal peaks due to the (*S*)-enantiomer in the chromatograms obtained with the test solution and reference solution (a) are identical.

Limit:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (d) (1.0 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 20 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve the contents of a vial of timolol for system suitability CRS (containing impurities B, C, D and F) in 1.0 mL of mobile phase A.

Reference solution (c) Dissolve 2 mg of the substance to be examined and 20 mg of maleic acid R in 10 mL of acetonitrile R. Evaporate 1 mL of the solution to dryness under a stream of nitrogen R in an amber glass vial. Heat the open vial at 105 °C for 1 h. Reconstitute the residue with 1.0 mL of mobile phase A.

Column:
— size: $l = 0.150$ m, $\phi = 3.9$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase:
— mobile phase A: mixture of equal volumes of methanol R and a 4.32 g/L solution of sodium octanesulfonate R previously adjusted to pH 3.0 with glacial acetic acid R;
— mobile phase B: methanol R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	97.5	2.5
10 - 11	97.5 \rightarrow 70	2.5 \rightarrow 30
11 - 20	70	30

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 295 nm.

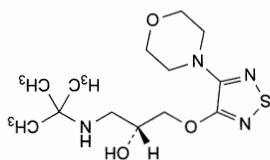
Injection 20 μ L.

Identification of impurities Use the chromatogram supplied obtained with reference solution (b) to identify the peaks due to impurities B, C, D and F; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity E.

Relative retention With reference to timolol (retention time = about 7.5 min): maleic acid = about 0.1; impurity D = about 0.3; impurity E = about 0.4; impurity B = about 0.7; impurity F = about 0.8; impurity C = about 2.1.

System suitability: reference solution (b):
— resolution: minimum 1.5 between the peaks due to impurities B and F.

Limits:
— correction factor: for the calculation of content, multiply the peak area of impurity D by 0.6;
— impurities B, C, D, E, F: for each impurity, not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— total: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);
— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to maleic acid.



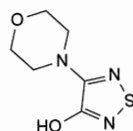
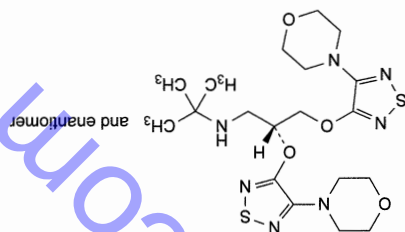
Specified impurities A, B, C, D, E, F
IMPURITIES
Protected from light.
STORAGE
of C₁₇H₂₈N₄O₇S.
1 mL of 0.1 M perchloric acid is equivalent to 43.25 mg
potentiometrically (2.2.20).
with 0.1 M perchloric acid, determining the end-point
Dissolve 0.350 g in 60 mL of anhydrous acetic acid R. Titrate
by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.
impurities for demonstration of compliance. See also 5.10.
Control of impurities in substances for pharmaceutical use: G, H, I, J.

Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.
ASSAY
Dissolve 0.350 g in 60 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 M perchloric acid is equivalent to 43.25 mg of C₁₇H₂₈N₄O₇S.

B. (2RS)-3-[(1,1-dimethylethyl)amino]-2-[[4-(morpholin-4-yl)-1,2,5-thiadiazol-3-yl]oxy]propan-1-ol, and enantiomer

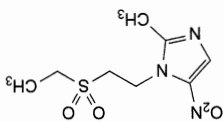
C. (2RS)-N-(1,1-dimethylethyl)-2,3-bis[[4-(morpholin-4-yl)-1,2,5-thiadiazol-3-yl]oxy]propan-1-amine,

D. 4-(morpholin-4-yl)-1,2,5-thiadiazol-3-ol,



Tinidazole

(Ph. Eur. monograph 1051)

 $C_8H_{13}N_3O_4S$

247.3

19387-91-8

Action and use

Antiprotzoal; antibacterial.

DEFINITION

1-[2-(Ethylsulfonyl)ethyl]-2-methyl-5-nitro-1H-imidazole.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Almost white or pale yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone and in

methylene chloride, sparingly soluble in methanol.

IDENTIFICATION

First identification A, C.

Second identification A, B, D, E.

A. Melting point (2.2.14): 125 °C to 128 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 10.0 mg in methanol R and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this

solution to 10.0 mL with methanol R.

Spectral range 220-350 nm.

Absorption maximum At 310 nm.

Specific absorbance at the absorption maximum 340 to 360.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison tinidazole CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 20 mg of tinidazole CRS in methanol R and dilute to 10 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Pre-treatment Heat at 110 °C for 1 h and allow to cool.

Mobile phase butanol R, ethyl acetate R (25:75 V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

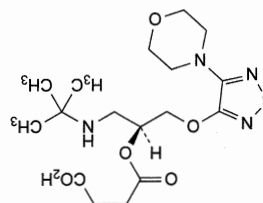
E. To about 10 mg add about 10 mg of zinc powder R,

0.3 mL of hydrochloric acid R and 1 mL of water R. Heat in a

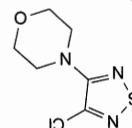
water-bath for 5 min and cool. The solution gives the

reaction of primary aromatic amines (2.3.1).

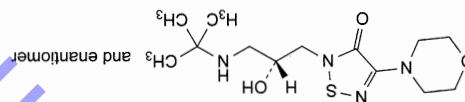
E. (2Z)-4-[(1S)-1-[[[(1S)-1-dimethylethyl]amino]methyl]-2-[(4-morpholin-4-yl)-1,2,5-thiadiazol-3-yl]oxy]ethoxy]-4-oxobut-2-enoic acid,



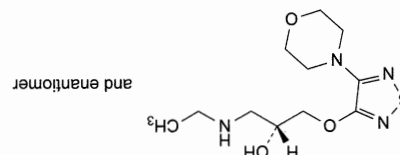
F. 4-(4-chloro-1,2,5-thiadiazol-3-yl)morpholine,



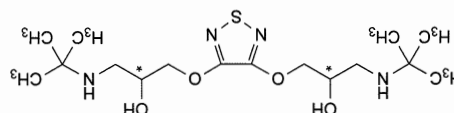
G. 4-(morpholin-4-yl)-1,2,5-thiadiazol-3(2H)-one 1-oxide,



H. 2-[(2RS)-3-[(1,1-dimethylethyl)amino]-2-hydroxypropyl]-4-(morpholin-4-yl)-1,2,5-thiadiazol-3(2H)-one,



I. (2RS)-1-(ethylamino)-3-[[4-(morpholin-4-yl)-1,2,5-thiadiazol-3-yl]oxy]propan-2-ol,



J. 1,1'-[1,2,5-thiadiazol-3,4-diylbis(oxy)]bis[3-[(1,1-dimethylethyl)amino]propan-2-ol].

Ph Eur



TESTS**Appearance of solution**

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Dissolve 1.0 g in acetone R and dilute to 20 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Protect solutions from light.

Test solution Dissolve 10.0 mg of the substance to be examined in 10.0 mL of methanol R and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of tinidazole impurity A CRS and 5.0 mg of tinidazole impurity B CRS in 10.0 mL of methanol R and dilute to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 3.0$ mm;

— stationary phase: octylsilyl silica gel for chromatography R (5 μ m).

Regular column conditioning by subsequent flushing with 50 mL of water R, 100 mL of methanol R, 25 mL of water R and 100 mL of the mobile phase is recommended.

Mobile phase acetonitrile R, methanol R, water R (10:20:70 V/V/V).

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 320 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of tinidazole.

Relative retention With reference to tinidazole (retention

time = about 6 min): impurity A = about 0.6;

impurity B = about 0.7.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to

impurities A and B.

Limits:

— impurities A, B: for each impurity, not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 25 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

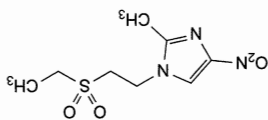
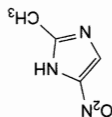
1 mL of 0.1 M perchloric acid is equivalent to 24.73 mg of C₈H₁₃N₃O₄S.

STORAGE

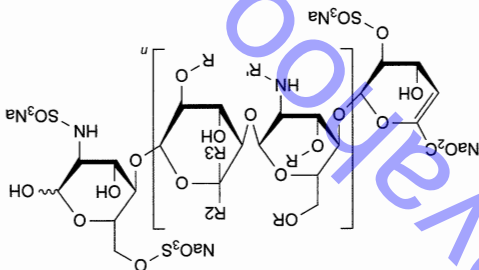
Protected from light.

IMPURITIES

Specified impurities A, B

**Tinzaparin Sodium**

(Ph. Eur. monograph 1271)

**Action and use**

Low molecular weight heparin.

Preparation

Tinzaparin Sodium Injection

DEFINITION

Tinzaparin sodium is the sodium salt of a low-molecular-mass heparin that is obtained by controlled enzymatic depolymerisation of heparin from porcine intestinal mucosa using heparinase from *Flavobacterium heparinum*. The majority of the components have a 2-O-sulfo-4-eneperanosuronic acid structure at the non-reducing end and a 2-N,6-O-disulfo-D-glucosamine structure at the reducing end of their chain.

Tinzaparin sodium complies with the monograph on Low-molecular-mass heparins (0828) with the modifications and additional requirements below.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very slightly soluble in water, very soluble in methylene chloride, freely soluble in alcohol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of tioconazole.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 2.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of tioconazole for system suitability CRS in the mobile phase and dilute to 2.5 mL with the mobile phase.

Column:

size: $l = 0.25$ m, $\phi = 4.6$ mm,

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m) with a specific surface area of

170 m²/g, a pore size of 12 nm and a carbon loading of 10 per cent.

Mobile phase Mix 1 volume of a 1.7 g/L solution of tetrabutylammonium dihydrogen phosphate R previously adjusted to pH 7.4 with dilute ammonia R2 and 3 volumes of methanol R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 218 nm.

Injection 20 μ L.

Run time 2.5 times the retention time of tioconazole.

System suitability: reference solution (b):

— resolution: minimum 1.0 between the peaks due to impurity B and impurity C (locate impurities A, B and C by comparison with the chromatogram provided with tioconazole for system suitability CRS).

Limits:

— correction factors: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.7;

impurity C = 1.7.

— impurities A, B, C: for each impurity, not more than 1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.3 per cent),

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent),

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a)

(1.0 per cent),

— disregard limit: 0.25 times the area of the principal peak in the chromatogram obtained with reference solution (a)

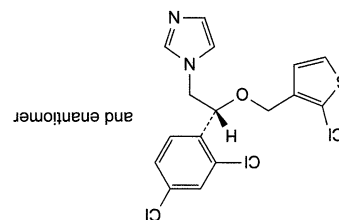
(0.05 per cent).

Water (2.5.12)

Maximum 0.5 per cent, determined on 1.00 g.

Tioconazole

(Ph. Eur. monograph 2074)



C₁₆H₁₃Cl₃N₂O₃ 387.7

65899-73-2

Preparations

Tioconazole Cream

Tioconazole Nail Solution

DEFINITION

1-[(2*RS*)-2-[(2-Chlorothiophen-3-yl)methoxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

TESTS

Appearance of solution

Dissolve 1.0 g in 10 mL of water R. The solution is clear (2.2.1) and not more intensely coloured than intensity 5 of the range of reference solutions of the most appropriate colour (2.2.2, Method II).

Absorbance (2.2.25)

Dissolve 50.0 mg in 100 mL of 0.01 M hydrochloric acid. The specific absorbance, measured at 231 nm and calculated with reference to the dried substance, is 8.0 to 12.5.

TESTS

The mass-average relative molecular mass ranges between 5500 and 7500. The mass percentage of chains lower than 2000 is not more than 10.0 per cent. The mass percentage of chains between 2000 and 8000 ranges between 60.0 and 72.0 per cent. The mass percentage of chains above 8000 ranges between 22.0 and 36.0 per cent.

Carry out identification test C as described in the monograph Low-molecular-mass heparins (0828). The following requirements apply.

Carry out identification test A as described in the monograph Low-molecular-mass heparins (0828) using tinzaparin sodium CRS.

IDENTIFICATION

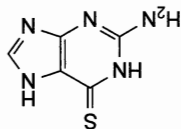
2.5. factor Xa activity to anti-factor IIa activity is between 1.5 and with reference to the dried substance. The ratio of the anti-

120 IU of anti-factor Xa activity per milligram calculated The potency is not less than 70 IU and not more than

The degree of sulfatation is 1.8 to 2.5 per disaccharide unit. 5500 and 7500 with a characteristic value of about 6500.

The mass-average relative molecular mass ranges between 5500 and 7500 with a characteristic value of about 6500.

Tioguanine



$C_5H_5N_5S$ 167.2 154-42-7

Action and use

Purine analogue; cytostatic.

Preparation

Tioguanine Tablets

DEFINITION

Tioguanine is 2-aminopurine-6(1*H*)-thione. It contains not less than 96.0% and not more than 102.0% of $C_5H_5N_5S$, calculated with reference to the dried substance.

CHARACTERISTICS

A pale yellow, crystalline powder. Practically insoluble in water and in ethanol (96%). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

Heat a suitable quantity at 105° at a pressure not exceeding 0.7 kPa for 5 hours. The infrared absorption spectrum of the residue, Appendix II A, is concordant with the reference spectrum of tioguanine (RS 340).

TESTS

Phosphate
To 0.5 g add 2 mL of 5*M* sulfuric acid and heat on a water bath for 5 minutes. Add, dropwise, 5 mL of nitric acid and add 10 mL of water and 0.75 mL of a solution prepared by dissolving 8.3 g of ammonium molybdate in 40 mL of water and adding 35 mL of 5*M* sulfuric acid and sufficient water to produce 100 mL. Add 1.0 mL of strong aminohydroxyapatite-sulfonic acid solution, mix and dilute to 25 mL with water. Measure the absorbance of the resulting solution at 820 nm, Appendix II B, using in the reference cell a solution prepared in the same manner but omitting the substance being examined. The absorbance is not more than that obtained by treating 1.5 mL of phosphate standard solution (100 ppm PO_4) in the same manner, beginning at the words 'add 10 mL of water ...'.

Free sulfur

Dissolve 50 mg in 5 mL of 1*M* sodium hydroxide. The solution is clear, Appendix IV A.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions.

(1) Dissolve 40 mg of the substance being examined in 100 mL of 0.01*M* sodium hydroxide and dilute 1 volume of the resulting solution to 2 volumes with the mobile phase.

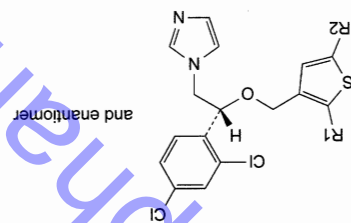
(2) Dilute 1 volume of solution (1) to 100 volumes with a mixture of 1 volume of 0.01*M* sodium hydroxide and 9 volumes of the mobile phase. Further dilute 1 volume of the resulting solution to 5 volumes with the same solvent mixture.

(3) Dissolve 12 mg of guanine BPCRS in 100 mL of 0.01*M* sodium hydroxide and dilute 1 volume of the resulting solution to 20 volumes with the mobile phase.

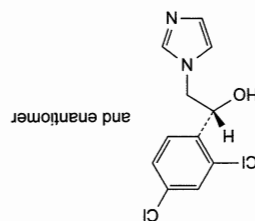
IMPURITIES

Specified impurities A, B, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D.



A. R1 = R2 = H: 1-[(2*RS*)-2-(2,4-dichlorophenyl)-2-[(thiophen-3-yl)methoxy]ethyl]-1*H*-imidazole,
B. R1 = R2 = Cl: 1-[(2*RS*)-2-(2,4-dichlorophenyl)-2-[(2,5-dichlorothiophen-3-yl)methoxy]ethyl]-1*H*-imidazole,
C. R1 = Cl, R2 = Br: 1-[(2*RS*)-2-[(5-bromo-2-chlorothiophen-3-yl)methoxy]-2-(2,4-dichlorophenyl)ethyl]-1*H*-imidazole,



D. (1*RS*)-1-(2,4-dichlorophenyl)-2-(1*H*-imidazol-1-yl)ethanol.

Ph Eur

ASSAY

Dissolve 0.300 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 *M* perchloric acid, determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 *M* perchloric acid is equivalent to 38.77 mg of $C_5H_5N_5S$.

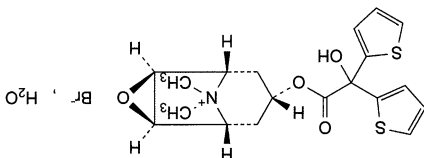
STORAGE

Protected from light.



Tiotropium Bromide Monohydrate

(Ph. Eur. monograph 2420)



$C_{19}H_{22}BrNO_4S_2H_2O$ 490.4

Action and use
Anticholinergic (antimuscarinic) bronchodilator.

Ph Eur

DEFINITION

(1R,2R,4S,5S,7S)-7-[(2-Hydroxy-2,2-dithiophen-2-ylacetyl)oxy]-9,9-dimethyl-3-oxa-9-azoniatricyclo[3.3.1.0^{2,4}]nonane bromide monohydrate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or yellowish-white powder or crystals.

Solubility

Sparsely soluble in water, soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison tiotropium bromide monohydrate CRS.

B. It gives reaction (a) of bromides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).
Dissolve 0.2 g in water R and dilute to 20 mL with the same solvent.

Impurities G and H

Thin-layer chromatography (2.2.27). Prepare the solutions immediately before use.
Solvent mixture Dilute 1 volume of 1 M hydrochloric acid to 100 volumes with methanol R.
Test solution Dissolve 0.40 g of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of tiotropium impurity mixture CRS (40 µg each of impurities G and H) in 1.0 mL of the solvent mixture.

Reference solution (b) Mix 0.1 mL of the test solution with 0.1 mL of reference solution (a).

Plate TLC silica gel F₂₅₄ plate R (2-10 µm).

Mobile phase water R, anhydrous formic acid R, acetonitrile R, methylene chloride R (10:15:35:50 V/V/V/V).

(4) Dissolve 40 mg each of tioguanine BPCRS and guanine BPCRS in 100 mL of 0.01M sodium hydroxide and dilute 1 volume of the resulting solution to 10 volumes with the mobile phase.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (5 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Waters Atlantis dC18 is suitable).

(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2 mL per minute.

(d) Use an ambient column temperature.

(e) Use a detection wavelength of 248 nm.

(f) Inject 10 µL of each solution.

MOBILE PHASE

0.05M anhydrous sodium dihydrogen orthophosphate adjusted to pH 3.0 with orthophosphoric acid.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (4), the resolution factor between the peaks due to tioguanine and guanine is at least 3.0.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to guanine is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (3%);

the area of any other secondary peak is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.2%);

the sum of the areas of any other secondary peaks is not greater than 2.5 times the area of the principal peak in the chromatogram obtained with solution (2) (0.5%);

Disregard any peak with an area less than 0.25 times the area of the principal peak in the chromatogram obtained with solution (2) (0.05%).

Loss on drying

When dried at 105° at a pressure not exceeding 0.7 kPa for 5 hours, loses not more than 1.0% of its weight. Use 1 g.

ASSAY

Carry out the method for liquid chromatography, Appendix III D, using the following solutions.

(1) Dissolve 40 mg of the substance being examined in 100 mL of 0.01M sodium hydroxide and dilute 1 volume of the resulting solution to 10 volumes with the mobile phase.

(2) Dissolve 40 mg of tioguanine BPCRS in 100 mL of 0.01M sodium hydroxide and dilute 1 volume of this solution to 10 volumes with the mobile phase.

(3) Dissolve 40 mg each of tioguanine BPCRS and guanine BPCRS in 100 mL of 0.01M sodium hydroxide and dilute 1 volume of the resulting solution to 10 volumes with the mobile phase.

CHROMATOGRAPHIC CONDITIONS

The chromatographic conditions described under Related substances may be used.

SYSTEM SUITABILITY

The test is not valid unless, in the chromatogram obtained with solution (3), the resolution factor between the peaks due to tioguanine and guanine is at least 3.0.

Application 10 µL of the test solution and reference solution (a) and 20 µL of reference solution (b).

Development Over 2/3 of the plate.

Drying In air.

Detection Expose to iodine vapour until the spots are clearly visible (about 15 min). Remove the plate and examine immediately.

Retention factors Impurity G = about 0.33;

impurity H = about 0.38; tiotropium = about 0.64.

System suitability: reference solution (b):

— the chromatogram shows 3 clearly separated spots.

Limits:

— **impurity G:** any spot due to impurity G is not more intense than the corresponding spot in the chromatogram

obtained with reference solution (a) (0.1 per cent);

— **impurity H:** any spot due to impurity H is not more intense than the corresponding spot in the chromatogram

obtained with reference solution (a) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29). Prepare all solutions protected from light.

Test solution Dissolve 50.0 mg of the substance to be examined in mobile phase B and dilute to 25.0 mL with

mobile phase B.

Reference solution (a) Dissolve 5.0 mg of tiotropium

impurity F CRS in mobile phase B and dilute to 100.0 mL

with mobile phase B. Dilute 1.0 mL of this solution to

25.0 mL with mobile phase B.

Reference solution (b) Dissolve 4 mg of tiotropium for system

suitability CRS (containing impurities A, C and E) in 2.0 mL

of mobile phase B.

Reference solution (c) Dilute 1.0 mL of the test solution to

100.0 mL with mobile phase B. Dilute 1.0 mL of this

solution to 10.0 mL with mobile phase B.

Column:

— size: $l = 0.15$ m, $\varnothing = 3.0$ mm;

— stationary phase: propylsilyl silica gel for chromatography R

(3.5 µm);

— temperature: 50 °C.

Mobile phase:

— **mobile phase A:** dissolve 1.0 g of sodium methanesulfonate

and 5.0 g of potassium dihydrogen phosphate R in about

980 mL of water R, adjust to pH 3.0 with dilute phosphoric

acid R and dilute to 1000 mL with water R;

— **mobile phase B:** methanol R, acetonitrile R, mobile phase A

(10:40:50 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
------------	-------------------------------	-------------------------------

0 - 3 90 10

3 - 17 90 → 80 10 → 20

17 - 28 80 → 25 20 → 75

28 - 30 25 75

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 240 nm.

Injection 5 µL.

Identification of impurities Use the chromatogram supplied

with tiotropium for system suitability CRS and the

chromatogram obtained with reference solution (b) to

identify the peaks due to impurities A, C and E.

IMPURITIES

Specified impurities A, C, E, F, G, H

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: B, D, I, J, K

1 mL of 0.1 M silver nitrate is equivalent to 47.24 mg of $C_{19}H_{22}BrNO_4S_2$.

end-point potentiometrically (2.2.20):

nitric acid R2. Titrate with 0.1 M silver nitrate determining the

Dissolve 0.35 g in 100 mL of water R. Add 10 mL of dilute

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

2.5 per cent to 4.0 per cent, determined on 0.300 g.

Water (2.5.12)

Maximum 10 ppm.

Solvent mixture water R, methanol R (10:90 V/V).

Dissolve 0.50 g of the substance to be examined in 20 mL of the solvent mixture using sonication for about 10 min.

The solution complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution

Maximum 10 ppm.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture water R, methanol R (10:90 V/V).

Dissolve 0.50 g of the substance to be examined in 20 mL of the solvent mixture using sonication for about 10 min.

The solution complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution

Maximum 10 ppm.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent mixture water R, methanol R (10:90 V/V).

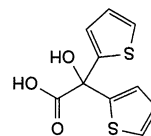
Dissolve 0.50 g of the substance to be examined in 20 mL of the solvent mixture using sonication for about 10 min.

The solution complies with test H. Prepare the reference solution using 0.5 mL of lead standard solution

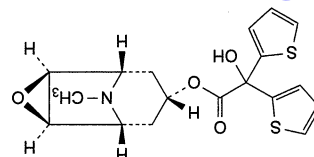
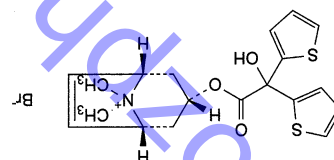
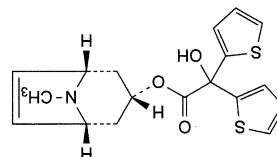
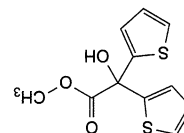
Maximum 10 ppm.

Heavy metals (2.4.8)

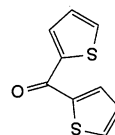
Maximum 10 ppm.



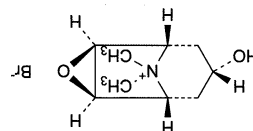
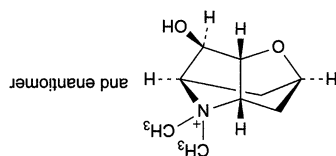
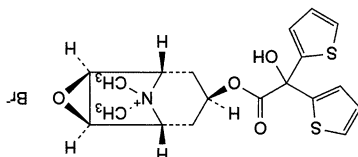
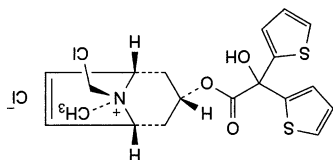
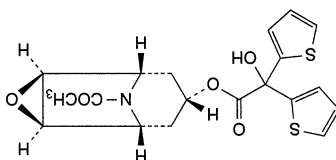
A. 2-hydroxy-2-(1,3-benzodithiol-2-yl)acetic acid,

B. (1*R*,2*R*,4*S*,5*S*,7*S*)-9-methyl-3-oxa-9-azatricyclo[3.3.1.0^{2,4}]nonan-7-yl 2-hydroxy-2-(1,3-benzodithiol-2-yl)acetic acid,C. (1*R*,3*S*,5*S*)-3-[(2-hydroxy-2-(1,3-benzodithiol-2-yl)acetyl)oxy]-8,8-dimethyl-8-azoniabicyclo[3.2.1]oct-6-ene bromide,D. (1*R*,3*S*,5*S*)-8-methyl-8-azabicyclo[3.2.1]oct-6-en-3-yl 2-hydroxy-2-(1,3-benzodithiol-2-yl)acetic acid,

E. methyl 2-hydroxy-2-(1,3-benzodithiol-2-yl)acetic acid,



F. dithiophen-2-ylmethanone,

G. (1*R*,2*R*,4*S*,5*S*,7*S*)-7-hydroxy-9,9-dimethyl-3-oxa-9-azoniatricyclo[3.3.1.0^{2,4}]nonane bromide,H. (1*S*,3*R*,4*R*,5*R*,7*SR*)-4-hydroxy-6,6-dimethyl-2-oxa-6-azoniatricyclo[3.3.1.0^{3,7}]nonane bromide,I. (1*R*,2*R*,4*S*,5*S*,7*T*)-7-[(2-hydroxy-2-(1,3-benzodithiol-2-yl)acetyl)oxy]-9,9-dimethyl-3-oxa-9-azoniatricyclo[3.3.1.0^{2,4}]nonane bromide,J. (1*R*,3*S*,5*S*,8*S*)-8-(chloromethyl)-3-[(2-hydroxy-2-(1,3-benzodithiol-2-yl)acetyl)oxy]-8-methyl-8-azoniabicyclo[3.2.1]oct-6-ene chloride,K. (1*R*,2*R*,4*S*,5*S*,7*S*)-9-acetyl-3-oxa-9-azatricyclo[3.3.1.0^{2,4}]nonan-7-yl 2-hydroxy-2-(1,3-benzodithiol-2-yl)acetic acid.

Titanium Dioxide
(Ph Eur monograph 0150)
TiO₂
79.9
13463-67-7

Action and use
Protective; excipient.
Preparation
Titanium Oxide
Titanium Oxide

DEFINITION
Content
98.0 per cent to 100.5 per cent.
CHARACTERS
Appearance
White or almost white powder.
Solubility
Practically insoluble in water. It does not dissolve in dilute mineral acids but dissolves slowly in hot concentrated sulfuric acid.

IDENTIFICATION

A. When strongly heated, it becomes pale yellow; the colour disappears on cooling.
 B. To 5 mL of solution S2 (see Tests) add 0.1 mL of strong hydrogen peroxide solution R. An orange-red colour appears.
 C. To 5 mL of solution S2 add 0.5 g of zinc R in granules. After 45 min, the mixture has a violet-blue colour.

TESTS

Solution S1
 Shake 20.0 g with 30 mL of hydrochloric acid R for 1 min. Add 100 mL of distilled water R and heat the mixture to boiling. Filter the hot mixture through a hardened filter paper until a clear filtrate is obtained. Wash the filter with 60 mL of distilled water R and dilute the combined filtrate and washings to 200 mL with distilled water R.

Solution S2
 Mix 0.500 g (m) with 5 g of anhydrous sodium sulfate R in a 300 mL long-necked combustion flask. Add 10 mL of water R and mix. Add 10 mL of sulfuric acid R and boil vigorously, with the usual precautions, until a clear solution is obtained. Cool, add slowly a cooled mixture of 30 mL of water R and 10 mL of sulfuric acid R, cool again and dilute to 100.0 mL with water R.

Appearance of solution

Solution S2 is not more opalescent than reference suspension II (2.2.1) and is colourless (2.2.2, Method II).

Acidity or alkalinity

Shake 5.0 g with 50 mL of carbon dioxide-free water R for 5 min. Centrifuge or filter until a clear solution is obtained. To 10 mL of the solution add 0.1 mL of bromothymol blue solution R1. Not more than 1.0 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Water-soluble substances

Maximum 0.5 per cent.
 To 10.0 g add a solution of 0.5 g of ammonium sulfate R in 150 mL of water R and boil for 5 min. Cool, dilute to 200 mL with water R and filter until a clear solution is obtained. Evaporate 100 mL of the solution to dryness in a tared evaporating dish and ignite. The residue weighs a maximum of 25 mg.

Ammony

Maximum 100 ppm.
 To 10 mL of solution S2 add 10 mL of hydrochloric acid R and 10 mL of water R. Cool to 20 °C, if necessary, and add 0.15 mL of sodium nitrite solution R. After 5 min, add 5 mL of a 10 g/L solution of hydroxylamine hydrochloride R and 10 mL of a freshly prepared 0.1 g/L solution of rhodamine B R. Mix thoroughly after each addition. Shake vigorously with 10.0 mL of toluene R for 1 min. Allow to separate and centrifuge for 2 min if necessary. Any pink colour in the toluene phase is not more intense than that in the toluene phase of a standard prepared at the same time in the same manner using a mixture of 5.0 mL of antimony standard solution (1 ppm Sb) R, 10 mL of hydrochloric acid R and 15 mL of a solution containing 0.5 g of anhydrous sodium sulfate R and 2 mL of sulfuric acid R instead of the mixture of 10 mL of solution S2, 10 mL of hydrochloric acid R and 10 mL of water R.

Arsenic (2.4.2, Method A)

Maximum 5 ppm.
 Place 0.50 g in a 250 mL round-bottomed flask, fitted with a thermometer, a funnel with stopcock and a vapour-outlet

tube connected to a flask containing 30 mL of water R. Add 50 mL of water R, 0.5 g of hydrazine sulfate R, 0.5 g of potassium bromide R and 20 g of sodium chloride R. Through the funnel, add dropwise 25 mL of sulfuric acid R, heat and maintain the temperature of the liquid at 110-115 °C for 20 min. Collect the vapour in the flask containing 30 mL of water R. Dilute to 50 mL with water R. 20 mL of the solution complies with the test.

Barium

To 10 mL of solution S1 add 1 mL of dilute sulfuric acid R. After 30 min, any opalescence in the solution is not more intense than that in a mixture of 10 mL of solution S1 and 1 mL of distilled water R.

Iron

Maximum 200 ppm.
 To 8 mL of solution S2 add 4 mL of water R. Mix and add 0.05 mL of bromine water R. Allow to stand for 5 min and remove the excess of bromine with a current of air. Add 3 mL of potassium thiocyanate solution R. Any colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using a mixture of 4 mL of iron standard solution (2 ppm Fe) R and 8 mL of a 200 g/L solution of sulfuric acid R.

Heavy metals (2.4.8)

Maximum 20 ppm.
 To 10 mL of solution S1, add dropwise concentrated ammonia R to adjust to pH 4 and dilute to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

ASSAY

To 300 g of zinc R in granules (710) add 300 mL of a 20 g/L solution of mercuric nitrate R and 2 mL of nitric acid R, shake for 10 min and wash with water R. Pack the amalgamated zinc into a glass tube about 400 mm long and about 20 mm in diameter fitted with a tap and a filter plate. Pass through the column 100 mL of dilute sulfuric acid R followed by 100 mL of water R, making sure that the amalgam is always covered with liquid. Pass slowly at a rate of about 3 mL/min through the column a mixture of 100 mL of dilute sulfuric acid R and 100 mL of water R followed by 100 mL of water R. Collect the eluate in a 500 mL conical flask containing 50.0 mL of a 150 g/L solution of ferric ammonium sulfate R in a mixture of 1 volume of sulfuric acid R and 3 volumes of water R. Add 0.1 mL of ferroin R and titrate immediately with 0.1 M ammonium and cerium nitrate until a greenish colour is obtained (n_1 mL). Pass slowly at a rate of about 3 mL/min through the column a mixture of 50 mL of dilute sulfuric acid R and 50 mL of water R, followed by 20.0 mL of solution S2, a mixture of 50 mL of dilute sulfuric acid R and 50 mL of water R, finally 100 mL of water R. Collect the eluate in a 500 mL conical flask containing 50.0 mL of a 150 g/L solution of ferric ammonium sulfate R in a mixture of 1 volume of sulfuric acid R and 3 volumes of water R. Rinse the lower end of the column with water R, add 0.1 mL of ferroin R and titrate immediately with 0.1 M ammonium and cerium nitrate until a greenish colour is obtained (n_2 mL). Calculate the percentage content of TiO₂ using the following expression:

$$m = \frac{3.99 \times (n_2 - n_1)}{m}$$

m = mass of the substance to be examined used for the preparation of solution S2, in grams.

FUNCTIONALITY-RELATED CHARACTERISTICS

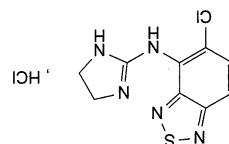
This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristic may be relevant for titanium dioxide used as opacifier in solid oral dosage forms and in preparations for cutaneous application.

Particle-size distribution (2.9.31)

Tizanidine Hydrochloride

(Ph. Eur. monograph 2578)



$C_9H_9ClN_5S$

290.2

64461-82-1

Action and use
Alpha₂-adrenoceptor agonist; skeletal muscle relaxant.

DEFINITION

5-Chloro-N-(4,5-dihydro-1H-imidazol-2-yl)-2,1,3-benzothiadiazol-4-amine hydrochloride.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white, crystalline powder.

Solubility

Soluble in water, very slightly soluble in ethanol (96 per cent), practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tizanidine hydrochloride CRS.

B. Solution S (see Tests) gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

pH (2.2.3)

3.7 to 5.0 for solution S.

Impurity H

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.200 g of the substance to be examined in methanol R and dilute to 5.0 mL with the same solvent.

Reference solution Dissolve 8.0 mg of ethylenediamine R (impurity H) in methanol R and dilute to 100.0 mL with the same solvent.

Plate TLC cellulose plate R.

Mobile phase glacial acetic acid R, water R, methanol R (1:25:75 V/V/V).

Application 5 µL.

Development Over 1/2 of the plate.

Drying In a current of cold air.

Detection Spray with a mixture of 15 volumes of a 20 g/L solution of ninhydrin R in glacial acetic acid R and 85 volumes of butanol R, then heat at 100 °C for 5 min.

Relative retention with reference to tizanidine (R_F = about 0.8): impurity H = about 0.3.

Limit:

— impurity H: any spot due to impurity H is not more intense than the corresponding spot in the chromatogram obtained with the reference solution (0.2 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 30.0 mg of the substance to be examined in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A. Dilute 5.0 mL of the solution to 50.0 mL with mobile phase A.

Reference solution (a) Dissolve 3 mg of tizanidine impurity B CRS in mobile phase A and dilute to 25.0 mL with mobile phase A. Dilute 1.0 mL of the solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with the test solution.

Reference solution (c) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (d) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (e) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (f) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (g) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (h) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (i) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (j) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (k) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (l) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (m) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (n) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (o) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (p) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (q) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (r) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (s) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (t) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (u) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (v) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (w) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (x) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (y) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (z) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (aa) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ab) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ac) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ad) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ae) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (af) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ag) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ah) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ai) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (aj) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Reference solution (ak) Dissolve 30.0 mg of tizanidine CRS in mobile phase A, using sonication if necessary, and dilute to 25.0 mL with mobile phase A.

Detection Spectrophotometer at 230 nm.

Injection 10 µL of the test solution and reference solutions (a)

and (b).

Identification of impurities Use the chromatogram obtained

with reference solution (a) to identify the peak due to

impurity B.

Relative retention With reference to tizanidine (retention

time = about 6 min): impurity B = about 1.3.

System suitability: reference solution (a):

— **resolution:** minimum 4.0 between the peaks due to

tizanidine and impurity B.

Calculation of percentage contents:

— for each impurity, use the concentration of tizanidine in

reference solution (b).

Limits:

— **unspecified impurities:** for each impurity, maximum

0.10 per cent;

— **total:** maximum 0.2 per cent;

— **reporting threshold:** 0.05 per cent.

Heavy metals (2.4.8)

Maximum 20 ppm.

Solvent water R.

0.50 g complies with test H. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modification.

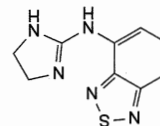
Injection Test solution and reference solution (c).

Calculate the percentage content of C₉H₆Cl₂N₅S taking into account the assigned content of tizanidine hydrochloride CRS.

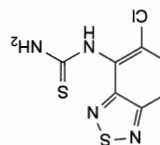
IMPURITIES

Specified impurities H

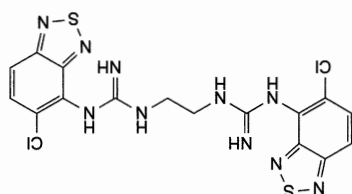
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, G, I.



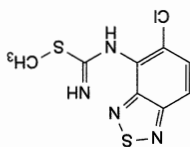
A. N-(4,5-dihydro-1H-imidazo[2-y]-2,1,3-benzothiadiazol-4-amine,



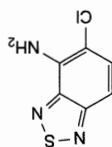
B. N-(5-chloro-2,1,3-benzothiadiazol-4-yl)thiourea,



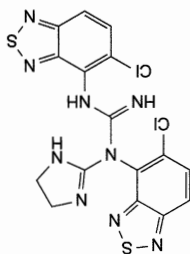
C. 1,1'-ethane-1,2-diylbis[3-(5-chloro-2,1,3-benzothiadiazol-4-yl)guanidine],



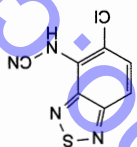
D. methyl N-(5-chloro-2,1,3-benzothiadiazol-4-yl)carbamimidodithioate,



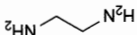
E. 5-chloro-2,1,3-benzothiadiazol-4-amine,



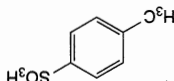
F. 1,3-bis(5-chloro-2,1,3-benzothiadiazol-4-yl)-1-(4,5-dihydro-1H-imidazo[2-y]-2-yl)guanidine,



G. (5-chloro-2,1,3-benzothiadiazol-4-yl)cyanamide,



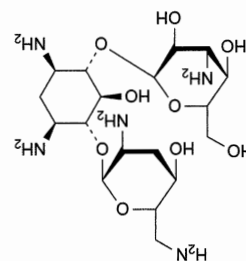
H. ethane-1,2-diamine (ethylenediamine),



I. 4-methylbenzenesulfonic acid (p-toluenesulfonic acid).

Tobramycin

(Ph Eur monograph 0645)



$C_{18}H_{37}N_5O_9$ 467.5 32986-56-4

Action and use
Aminoglycoside antibacterial.

Preparation
Tobramycin Injection

Ph Eur

DEFINITION

4-O-(3-Amino-3-deoxy-α-D-glucopyranosyl)-2-deoxy-6-O-(2,6-diamino-2,3,6-trideoxy-α-D-ribo-hexopyranosyl)-L-streptamine.

Substance produced by *Streptomyces tenebrarius* or obtained by any other means.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

It is produced by methods of manufacture designed to eliminate or minimise substances lowering blood pressure.

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, very slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A.

Second identification B, C.

A. Nuclear magnetic resonance spectrometry (2.2.33).

Preparation 100 g/L solution in deuterium oxide R.

Comparison 100 g/L solution of tobramycin CRS in deuterium oxide R.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in water R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of tobramycin CRS in water R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 4 mg of neomycin sulfate CRS and 4 mg of kanamycin monosulfate CRS in 1 mL of reference solution (a).

Plate TLC silica gel plate R.

Mobile phase methylene chloride R, concentrated ammonia R, methanol R (17:33:50 V/V/V).

Application 5 μL.

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Spray with a mixture of equal volumes of a 2 g/L solution of 1,3-dihydroxybenzophenone R in ethanol

of the detector is set at 35 °C.

durations according to the instrument used. The temperature oxidation and -0.15 V reduction potentials, with pulse cell body, held at respectively + 0.05 V detection, + 0.75 V

electrode and a stainless steel auxiliary electrode which is the gold working electrode, a silver-silver chloride reference

Detection Pulsed amperometric detector or equivalent with a

Flow rate 0.3 mL/min.

polymeric mixing coil.

added pulselessly to the column effluent using a 375 μL

Post-column solution carbonate-free sodium hydroxide solution R diluted 25-fold with carbon dioxide-free water R, which is

Flow rate 1.0 mL/min.

pH 3.0 with dilute phosphoric acid R. Degass.

potassium dihydrogen phosphate R previously adjusted to

stabilised with butylhydroxytoluene R, and 50 mL of 0.2 M

sodium octanesulfonate R, 3 mL/L of tetrahydrofuran R

containing 52 g/L of anhydrous sodium sulfate R, 1.5 g/L of

Mobile phase Mixture prepared with carbon dioxide-free water R

— temperature: 55 °C.

with a pore size of 100 nm.

— stationary phase: styrene-divinylbenzene copolymer R (8 μm)

— size: 1 = 0.25 m, Ø = 4.6 mm.

Column:

to 25.0 mL with the mobile phase.

Reference solution (e) Dilute 10.0 mL of reference solution (a)

to 10.0 mL with the mobile phase.

this solution, add 2.0 mL of reference solution (a) and dilute

sulfate CRS in 20.0 mL of the mobile phase. To 1.0 mL of

Reference solution (d) Dissolve 10.0 mg of kanamycin B

to 50.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (a)

to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of reference solution (a)

the mobile phase and dilute to 100.0 mL with the mobile

Reference solution (a) Dissolve 25.0 mg of tobramycin CRS in

100.0 mL with the mobile phase.

Test solution (b) Dilute 10.0 mL of test solution (a) to

the mobile phase.

Test solution (a) Dissolve 25.0 mg of the substance to be

examined in the mobile phase and dilute to 25.0 mL with

Liquid chromatography (2.2.29).

Related substances

same solvent.

Dissolve 1.00 g in water R and dilute to 25.0 mL with the

+ 138 to + 148 (anhydrous substance).

Specific optical rotation (2.2.7)

Dissolve 1.0 g in 10 mL of carbon dioxide-free water R.

9.0 to 11.0.

pH (2.2.3)

TESTS

heat in a water-bath for 3 min. A violet-blue colour develops.

1 g/L solution of ninhydrin R in ethanol (96 per cent) R and

C. Dissolve about 5 mg in 5 mL of water R. Add 5 mL of a

solution (a).

principal spot in the chromatogram obtained with reference

the test solution is similar in position, colour and size to the

Results The principal spot in the chromatogram obtained with

solution (b) shows 3 major spots which are clearly separated.

System suitability The chromatogram obtained with reference

at 105 °C for 5-10 min.

(96 per cent) R and a 460 g/L solution of sulfuric acid R; heat

NOTE: to prevent problems due to salt precipitation, the

electrochemical cell can be flushed with *water R* overnight.

Injection 20 µL using a refrigerated injector (4–8 °C); inject

test solution (a) and reference solutions (b), (c) and (d).

Run time 1.5 times the retention time of tobramycin.

Relative retention With reference to tobramycin (retention

time = about 18 min): impurity C = about 0.35;

impurity B = about 0.40; impurity A = about 0.70.

System suitability:

— **resolution:** minimum 3.0 between the peaks due to

impurity A and to tobramycin in the chromatogram

obtained with reference solution (d); if necessary, adjust

the concentration of sodium octanesulfonate in the

mobile phase;

— **signal-to-noise ratio:** minimum 10 for the principal peak in

the chromatogram obtained with reference solution (b).

Limits:

— **any impurity:** not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (c) (1.0 per cent) and not more than 1 such peak

has an area greater than the area of the principal peak in

the chromatogram obtained with reference solution (c)

(0.5 per cent);

— **total:** not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (c)

(1.5 per cent);

— **disregard limit:** the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.25 per cent).

2-Methyl-1-propanol (2.4.24, System B)

Maximum 1.0 per cent *m/m*.

Water (2.5.12)

Maximum 8.0 per cent, determined on 0.30 g.

Sulfated ash (2.4.14)

Maximum 0.3 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 2.0 IU/mg, if intended for use in the manufacture

of parenteral preparations without a further appropriate

procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in the test for

related substances with the following modifications.

Injection Test solution (b) and reference solution (e).

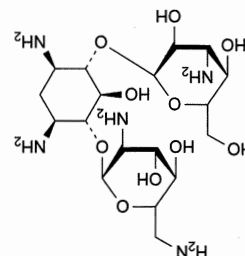
Calculate the percentage content of tobramycin.

STORAGE

If the substance is sterile, store in a sterile, airtight, tamper-

proof container.

IMPURITIES



A. 4-O-(3-amino-3-deoxy-α-D-glucopyranosyl)-2-deoxy-6-O-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-L-streptamine (kanamycin B),

IDENTIFICATION

Practically insoluble in water, freely soluble in acetone, in

anhydrous ethanol, in methylene chloride and in fatty oils.

Solubility

Clear, colourless or yellowish-brown, viscous, oily liquid.

Appearance

CHARACTERS

96.0 per cent to 102.0 per cent.

Content

all-*rac*-2,5,7,8-Tetramethyl-2-(4,8,12-trimethyltridecyl)-3,4-dihydro-2H-1-benzopyran-6-ol.

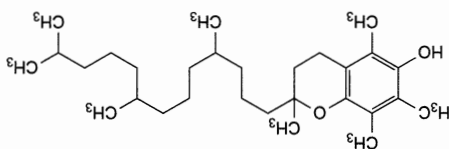
DEFINITION

Ph Eur

Used in prevention and treatment of vitamin E deficiencies.

Action and use

C₂₉H₅₀O₂ 430.7 10191-41-0



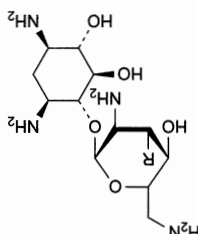
(all-*rac*-α-Tocopherol, Ph Eur monograph 0692)



all-*rac*-Alpha-Tocopherol

Ph Eur

B. R = H: 2-deoxy-4-O-(2,6-diamino-2,3,6-trideoxy-α-D-ribo-hexopyranosyl)-D-streptamine (nebramycin),
C. R = OH: 2-deoxy-4-O-(2,6-diamino-2,6-dideoxy-α-D-glucopyranosyl)-D-streptamine (neamine).



Drying In a current of air.

Development Over 2/3 of the plate.

Application 10 µL.

Mobile phase ether R, cyclohexane R (20:80 *V/V*).

Plate TLC silica gel F₂₅₄ plate R.

of cyclohexane R.

Reference solution Dissolve 10 mg of α-tocopherol CRS in 2 mL

in 2 mL of cyclohexane R.

Test solution Dissolve 10 mg of the substance to be examined

C. Thin-layer chromatography (2.2.27).

Comparison α-tocopherol CRS.

B. Infrared absorption spectrophotometry (2.2.24).

with the same solvent.

Dissolve 2.50 g in anhydrous ethanol R and dilute to 25.0 mL

A. Optical rotation (2.2.7): −0.01° to +0.01°.

Second identification A, C.

First identification A, B.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Related substances

Gas chromatography (2.2.28): use the normalisation

procedure.

Internal standard solution Dissolve 1.0 g of squalane R in cyclohexane R and dilute to 100.0 mL with the same solvent.

Test solution (a) Dissolve 0.100 g of the substance to be examined in 10.0 mL of the internal standard solution.

Test solution (b) Dissolve 0.100 g of the substance to be examined in 10.0 mL of the internal standard solution.

Test solution (c) Dissolve 0.100 g of the substance to be examined in 10 mL of cyclohexane R.

Reference solution (a) Dissolve 0.100 g of α -tocopherol CRS in 10.0 mL of the internal standard solution.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 10 mg of α -tocopherol acetate R in cyclohexane R and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dissolve 10 mg of α -tocopherol for peak identification CRS (containing impurities A and B) in cyclohexane R and dilute to 1 mL with the same solvent.

Reference solution (d) Dilute 1.0 mL of test solution (b) to 100.0 mL with cyclohexane R. Dilute 1.0 mL of this solution to 10.0 mL with cyclohexane R.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.25$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

— column: 280 °C;

— injection port and detector: 290 °C.

Detection Flame ionisation.

Injection 1 μ L of test solution (b) and reference solutions (b), (c) and (d).

Run time Twice the retention time of α -tocopherol.

Identification of impurities Use the chromatogram supplied with α -tocopherol for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to α -tocopherol

(retention time = about 9 min): squalane = about 0.5;

impurity A = about 0.7; impurity B = about 0.8;

impurities C and D = about 1.05 (eluting immediately after the α -tocopherol peak).

System suitability: reference solution (b):

— resolution: minimum 3.5 between the peaks due to α -tocopherol and α -tocopherol acetate.

Limits:

— impurity A: maximum 0.5 per cent;

— impurity B: maximum 1.5 per cent;

— sum of impurities C and D: maximum 1.0 per cent;

— any other impurity: for each impurity, maximum 0.25 per cent;

— total: maximum 2.5 per cent.

— **disregard limit**: the area of the principal peak in the chromatogram obtained with reference solution (d)

(0.1 per cent).

The thresholds indicated under Related substances for

(Table 2034.-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

ASSAY

Gas chromatography (2.2.28) as described in the test for related substances with the following modifications.

Injection Test solution (a) and reference solution (a).

System suitability: reference solution (a):

— symmetry factor: minimum 0.6 for the principal peak.

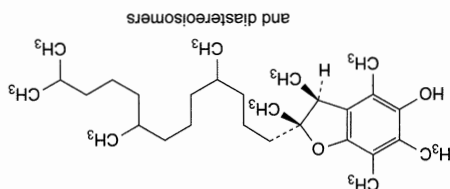
Calculate the percentage content of $C_{29}H_{50}O_2$ from the declared content of α -tocopherol CRS.

STORAGE

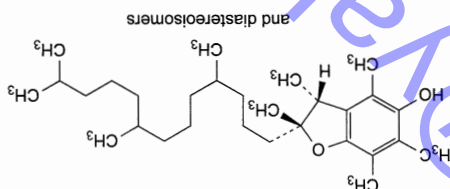
Under an inert gas, protected from light.

IMPURITIES

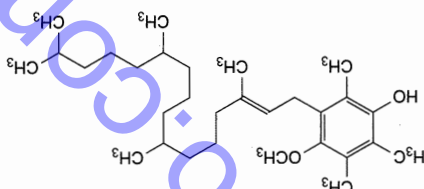
Specified impurities A, B, C, D.



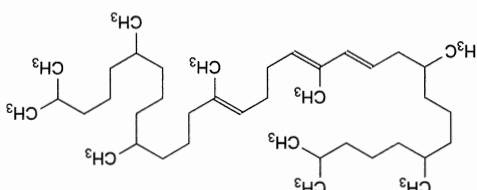
A. all-*rac*-*trans*-2,3,4,6,7-pentamethyl-2-(4,8,12-trimethyltridecyl)-2,3-dihydrobenzofuran-5-ol,



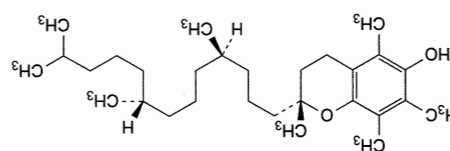
B. all-*rac*-*cis*-2,3,4,6,7-pentamethyl-2-(4,8,12-trimethyltridecyl)-2,3-dihydrobenzofuran-5-ol,



C. 4-methoxy-2,3,6-trimethyl-5-[(all-*RS,E*)-3,7,11,15-tetramethylhexadec-2-enyl]phenol,



D. (all-*RS,all-E*)-2,6,10,14,19,23,27,31-octamethyldodecatriene.

RRR- α -Tocopherol(RRR- α -Tocopherol, Ph Eur monograph 1256) $C_{50}H_{90}O_2$

430.7

59-02-9

Action and use

Used in prevention and treatment of vitamin E deficiencies.

Ph Eur

DEFINITION

(2R)-2,5,7,8-Tetramethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-ol.

Content

94.5 per cent to 102.0 per cent.

CHARACTERS

Appearance

Clear, colourless or yellowish-brown, viscous, oily liquid.

Solubility

Practically insoluble in water, freely soluble in acetone, in anhydrous ethanol, in methylene chloride and in fatty oils.

IDENTIFICATION

First identification A, B

Second identification A, C

A. Optical rotation (2.2.7): $+0.05^\circ$ to $+0.10^\circ$.

Dissolve 2.50 g in anhydrous ethanol R and dilute to 25.0 mL

with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison α -tocopherol CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in 2 mL of cyclohexane R.

Reference solution Dissolve 10 mg of α -tocopherol CRS in 2 mL

of cyclohexane R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase either R, cyclohexane R (20:80 V/V).

Application 10 μ L.

Development Over 2/3 of the plate.

Drying In a current of air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

TESTS

Related substances

Gas chromatography (2.2.28): use the normalisation

procedure.

Internal standard solution Dissolve 1.0 g of squalane R in

cyclohexane R and dilute to 100.0 mL with the same solvent.

Test solution (a) Dissolve 0.100 g of the substance to be

examined in 10.0 mL of the internal standard solution.

Test solution (b) Dissolve 0.100 g of the substance to be

examined in 10.0 mL of cyclohexane R.

Reference solution (a) Dissolve 0.100 g of α -tocopherol CRS in

10.0 mL of the internal standard solution.

Reference solution (b) Dissolve 10 mg of α -tocopherol R and10 mg of α -tocopheryl acetate R in cyclohexane R and dilute to

100.0 mL with the same solvent.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.25$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness

0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

Temperature	Time	Column	Injection port	Detector
(°C)	(min)			
280	0 - 15		290	290

Detection Flame ionisation.

Injection 1 μ L of test solution (b) and reference solution (b).

System suitability: reference solution (b):

— resolution: minimum 3.5 between the peaks due to

 α -tocopherol and α -tocopheryl acetate.

Limits:

— total: maximum 4.0 per cent;

— disregard limit: 0.1 per cent.

The thresholds indicated under Related substances

(Table 2034-1) in the general monograph Substances for

pharmaceutical use (2034) do not apply.

ASSAY

Gas chromatography (2.2.28) as described in the test for

related substances with the following modifications.

Injection Test solution (a) and reference solution (a).

System suitability: reference solution (a):

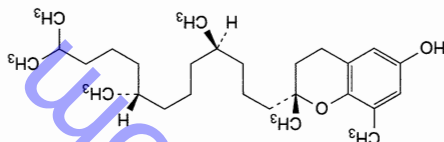
— symmetry factor: minimum 0.6 for the principal peak.

Calculate the percentage content of $C_{50}H_{90}O_2$ taking intoaccount the assigned content of α -tocopherol CRS.

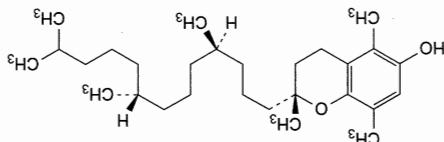
STORAGE

Under an inert gas, protected from light.

IMPURITIES



A. (2R)-2,8-dimethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-

3,4-dihydro-2H-1-benzopyran-6-ol (RRR- δ -tocopherol).

B. (2R)-2,5,8-trimethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-

3,4-dihydro-2H-1-benzopyran-6-ol (RRR- β -tocopherol).

spot in the chromatogram obtained with the reference solution.

TESTS

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

Internal standard solution Dissolve 1.0 g of squalane R in cyclohexane R and dilute to 100.0 mL with the same solvent.

Test solution (a) Dissolve 0.100 g of the substance to be examined in 10.0 mL of the internal standard solution.

Test solution (b) Dissolve 0.100 g of the substance to be examined in 10 mL of cyclohexane R.

Reference solution (a) Dissolve 0.100 g of α -tocopherol acetate CRS in 10.0 mL of the internal standard solution.

Reference solution (b) Dissolve 10 mg of the substance to be examined and 10 mg of α -tocopherol R in cyclohexane R and dilute to 100.0 mL with the same solvent.

Reference solution (c) Dissolve 10 mg of all- α -tocopherol acetate for peak identification CRS (containing impurities A and B) in cyclohexane R and dilute to 1 mL with the same solvent.

Reference solution (d) Dilute 1.0 mL of test solution (b) to 100.0 mL with cyclohexane R. Dilute 1.0 mL of this solution to 10.0 mL with cyclohexane R.

Column: fused silica; size: $l = 30$ m, $\varnothing = 0.25$ mm; stationary phase: poly(dimethyl)siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R. Flow rate 1 mL/min.

Split ratio 1:100. Temperature: column: 280 °C; injection port and detector: 290 °C.

Detection Flame ionisation. Injection 1 μ L of test solution (b) and reference solutions (a), (b), (c) and (d); inject directly onto the column or via a sufficiently inert, glass-lined injection port using an automatic injection device or other reproducible injection method.

Run time Twice the retention time of all- α -tocopherol acetate.

Identification of impurities Use the chromatogram supplied with all- α -tocopherol acetate for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A and B.

Relative retention With reference to all- α -tocopherol acetate (retention time = about 15 min): squalane = about 0.4; impurity A = about 0.7; impurity B = about 0.8; impurity C = about 0.9; impurities D and E = about 1.05 (eluting immediately after the all- α -tocopherol acetate peak).

System suitability: resolution: minimum 3.5 between the peaks due to impurity C and all- α -tocopherol acetate in the chromatogram obtained with reference solution (b); in the chromatogram obtained with reference solution (a), the area of the peak due to impurity C is not greater than 0.2 per cent of the area of the peak due to all- α -tocopherol acetate.

Limits: — impurities A, C: for each impurity, maximum 0.5 per cent;

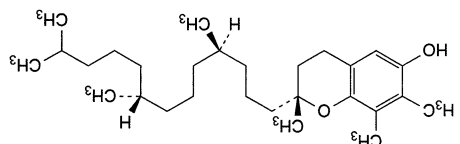
all-rac-Alpha-Tocopherol Acetate

(all-rac-Tocopherol Acetate, Ph Eur monograph 0439)

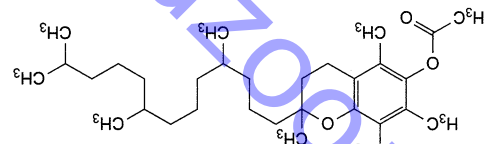


Ph Eur

C. (2R)-2,7,8-trimethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-ol (RRR- γ -tocopherol).



C₃₁H₅₂O₃ 472.7 7695-91-2



Action and use Used in prevention and treatment of vitamin E deficiencies.

Ph Eur

DEFINITION

all-rac-2,5,7,8-Tetramethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-yl acetate.

Content 96.5 per cent to 102.0 per cent.

CHARACTERS

Appearance Clear, colourless or slightly greenish-yellow, viscous, oily liquid.

Solubility Practically insoluble in water, freely soluble in acetone, in anhydrous ethanol and in fatty oils.

IDENTIFICATION

First identification A, B Second identification A, C

A. Optical rotation (2.2.7): -0.01° to $+0.01^{\circ}$. Dissolve 2.50 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24). Comparison α -tocopherol acetate CRS.

C. Thin-layer chromatography (2.2.27). Test solution Dissolve about 10 mg of the substance to be examined in 2 mL of cyclohexane R.

Reference solution Dissolve about 10 mg of α -tocopherol acetate CRS in 2 mL of cyclohexane R.

Plate TLC silica gel F₂₅₄ plate R. Mobile phase ether R, cyclohexane R (20:80 V/V).

Application 10 μ L. Development Over 2/3 of the plate.

Drying In a current of air. Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal

- **impurity B**: maximum 1.5 per cent;
 - **sum of impurities D and E**: maximum 1.0 per cent;
 - **any other impurity**: for each impurity, maximum 0.25 per cent;
 - **total**: maximum 2.5 per cent;
 - **disregard limit**: the area of the principal peak in the chromatogram obtained with reference solution (d) (0.1 per cent).
- The thresholds indicated under Related substances (Table 2034-1) in the general monograph *Substances for pharmaceutical use* (2034) do not apply.

ASSAY

Gas chromatography (2.2.28) as described in the test for related substances with the following modifications.

Injection Test solution (a) and reference solution (a).

System suitability: reference solution (a):

— **symmetry factor**: minimum 0.6 for the principal peak.

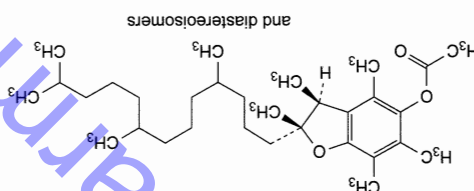
Calculate the percentage content of $C_{31}H_{52}O_3$ from the declared content of α -tocopheryl acetate CRS.

STORAGE

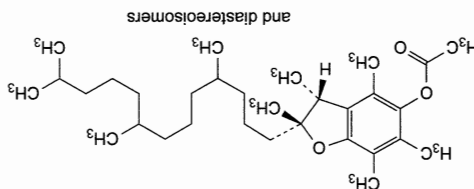
Protected from light.

IMPURITIES

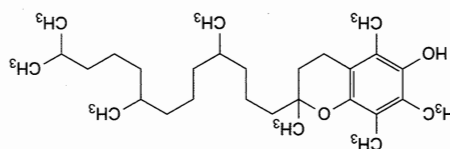
Specified impurities A, B, C, D, E



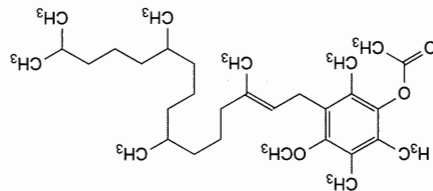
A. all-*rac-trans*-2,3,4,6,7-pentamethyl-2-[(4,8,12-trimethyltridecyl)-2,3-dihydrobenzofuran-5-yl] acetate,



B. all-*rac-cis*-2,3,4,6,7-pentamethyl-2-[(4,8,12-trimethyltridecyl)-2,3-dihydrobenzofuran-5-yl] acetate,



C. all-*rac*-2,5,7,8-tetramethyl-2-[(4,8,12-trimethyltridecyl)-3,4-dihydro-2H-1-benzopyran-6-ol] (all-*rac*- α -tocopherol),



D. 4-methoxy-2,3,6-trimethyl-5-[(all-*RS,E*)-3,7,11,15-tetramethylhexadec-2-enyl]phenyl acetate,

DEFINITION

(2R)-2,5,7,8-Tetramethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-yl acetate.

95.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

Clear, colourless or slightly greenish-yellow, viscous, oily liquid.

Solubility

Practically insoluble in water, freely soluble in acetone, in anhydrous ethanol and in fatty oils, soluble in ethanol (96 per cent).

IDENTIFICATION

First identification A, B

Second identification A, C

A. Optical rotation (2.2.7): $+0.25^\circ$ to $+0.35^\circ$.

Dissolve 2.50 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison α -tocopheryl acetate CRS.

C. Thin-layer chromatography (2.2.27).

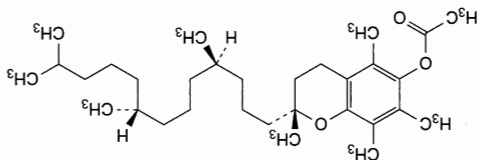
Test solution (a) Dissolve 10 mg of the substance to be examined in 2 mL of cyclohexane R.

Test solution (b) In a ground-glass stoppered tube, dissolve about 10 mg of the substance to be examined in 2 mL of 2.5 M alcoholic sulfuric acid R. Heat on a water-bath for 5 min. Cool and add 2 mL of water R and 2 mL of cyclohexane R. Shake for 1 min. Use the upper layer.

Reference solution (a) Dissolve 10 mg of α -tocopheryl acetate CRS in 2 mL of cyclohexane R.

RRR- α -Tocopheryl Acetate

(RRR- α -Tocopheryl Acetate, Ph Eur monograph 1257)



$C_{31}H_{52}O_3$

472.7

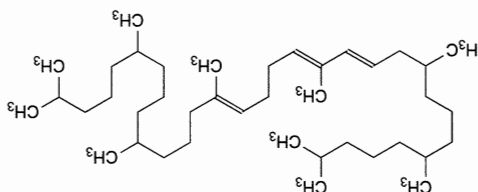
Action and use

Used in prevention and treatment of vitamin E deficiencies.

Ph Eur

Ph Eur

E. (all-*RS,all-E*)-2,6,10,14,19,23,27,31-octamethylidodotriacont-12,14,18-triene.



Reference solution (b) Prepare as described for test solution (b), using α -tocopheryl acetate CRS instead of the substance to be examined.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ether R, cyclohexane R (20:80 V/V).

Application 10 μ L.

Development Over 2/3 of the plate.

Drying In a current of air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with test solution (a) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a). In the chromatograms obtained with test solution (b) and reference solution (b), there may be 2 spots depending on the degree of hydrolysis: the spot with the higher R_f value is due to α -tocopheryl acetate and corresponds to the spot in the chromatogram obtained with reference solution (a); the spot with the lower R_f value is due to α -tocopherol.

TESTS

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

Internal standard solution Dissolve 1.0 g of *squalene* R in cyclohexane R and dilute to 100.0 mL with the same solvent.

Test solution (a) Dissolve 0.100 g of the substance to be examined in 10.0 mL of the internal standard solution.

Test solution (b) Dissolve 0.100 g of the substance to be examined in 10.0 mL of cyclohexane R.

Reference solution (a) Dissolve 0.100 g of α -tocopheryl acetate CRS in 10.0 mL of the internal standard solution.

Reference solution (b) Dissolve 10 mg of α -tocopherol R and 10 mg of α -tocopheryl acetate R in cyclohexane R and dilute to 100.0 mL with the same solvent.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.25$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

Time (min)	Temperature (°C)	Column	Injection port	Detector
0 - 15	280		290	290

Detection Flame ionisation.

Injection 1 μ L of test solution (b) and reference solutions (a) and (b); inject directly onto the column or via a sufficiently inert, glass-lined injection port using an automatic injection device or other reproducible injection method.

System suitability:

— resolution: minimum 3.5 between the peaks due to α -tocopherol and α -tocopheryl acetate in the chromatogram obtained with reference solution (b);

— in the chromatogram obtained with reference solution (a), the area of the peak due to α -tocopherol is not greater

IMPURITIES

Protected from light.

STORAGE

Calculate the percentage content of C₃₁H₅₂O₃ taking into account the assigned content of α -tocopheryl acetate CRS.

— symmetry factor: minimum 0.6 for the principal peak.

System suitability: reference solution (a):

Injection Test solution (a) and reference solution (a).

related substances with the following modifications.

Gas chromatography (2.2.28) as described in the test for

ASSAY

pharmaceutical use (2034) do not apply.

(Table 2034-1) in the general monograph *Substances for*

The thresholds indicated under Related substances

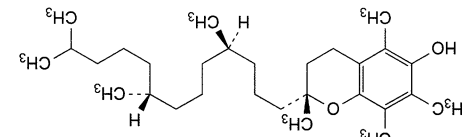
— disregard limit: 0.1 per cent.

— total: maximum 4.0 per cent;

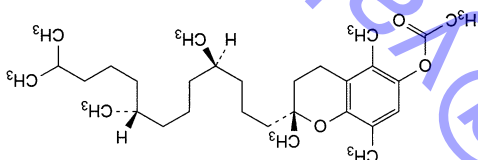
Limits:

α -tocopheryl acetate.

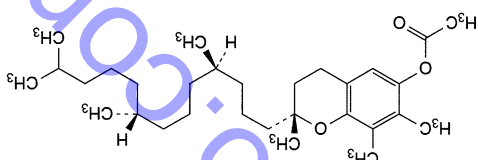
than 0.2 per cent of the area of the peak due to



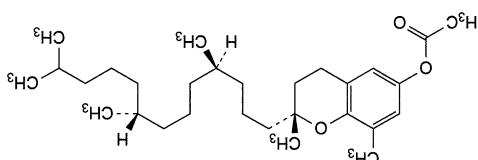
A. (2R)-2,5,7,8-tetramethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-ol (RRR- α -tocopherol).



B. (2R)-2,5,8-trimethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-yl acetate (RRR- β -tocopheryl acetate).



C. (2R)-2,7,8-trimethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-yl acetate (RRR- γ -tocopheryl acetate).



D. (2R)-2,8-dimethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-yl acetate (RRR- δ -tocopheryl acetate).



Alpha Tocopheryl Acetate Concentrate (Powder Form)

(α -Tocopheryl Acetate Concentrate (Powder Form))
Ph Eur monograph 0691)

Action and use

Used in prevention and treatment of vitamin E deficiencies.

Ph Eur

DEFINITION

Preparation obtained either by finely dispersing *all-rac- α -Tocopheryl acetate* (0439) in a suitable carrier of suitable quality (for example gelatin, acacia, carbohydrates, lactoproteins or a mixture thereof) or by adsorbing *all-rac- α -Tocopheryl acetate* (0439) on silicic acid of suitable quality.

Content

90.0 per cent to 115.0 per cent of the α -tocopheryl acetate content stated on the label, which is not less than 25 g per 100 g of concentrate.

CHARACTERS

Appearance

Almost white, yellowish or light brown, small particles.

Solubility

Practically insoluble or swells or forms a dispersion in water, depending on the formulation.

IDENTIFICATION

First identification B.

Second identification A.

A. Thin-layer chromatography (2.2.27).

Test solution To a quantity of the preparation to be examined corresponding to 50 mg of α -tocopheryl acetate add 5 mL of 0.01 M hydrochloric acid and treat with ultrasound at 60 °C. Add 5 mL of anhydrous ethanol R and 10 mL of cyclohexane R, shake for 1 min and centrifuge for 5 min. Use the upper layer.

Reference solution Dissolve 50 mg of α -tocopheryl acetate CRS in cyclohexane R and dilute to 10 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ether R, cyclohexane R (20:80 V/V).

Application 10 μ L.

Development 3/4 of the plate.

Drying In a current of air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

B. Examine the chromatograms obtained in the assay.

Results:

— the principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a);
— in the chromatogram obtained with reference solution (c) no additional principal peak is observed when compared with the chromatogram obtained with the test solution.

ASSAY

Gas chromatography (2.2.28).

Internal standard solution Dissolve 1.0 g of squalane R in cyclohexane R and dilute to 500.0 mL with the same solvent. **Test solution** Weigh accurately a quantity of the preparation to be examined corresponding to about 0.100 g of α -tocopheryl acetate into a 250 mL conical flask. Add 20 mL of 1 M hydrochloric acid and sonicate at 70 °C for 20 min. Add 50 mL of anhydrous ethanol R and 50.0 mL of the internal standard solution. Mix thoroughly for 30 min using a magnetic stirrer. Allow the 2 layers to separate and use the upper layer.

Reference solution (a) Dissolve 0.100 g of α -tocopheryl acetate CRS in 50.0 mL of the internal standard solution. **Reference solution (b)** Dissolve 10 mg of α -tocopheryl R and 10 mg of α -tocopheryl acetate CRS in 5.0 mL of cyclohexane R. **Reference solution (c)** Mix 1.0 mL of the test solution and 1.0 mL of reference solution (a).

Column:

— **material:** fused silica;

— **size:** $l = 30$ m, $\varnothing = 0.25$ mm;

— **stationary phase:** poly(dimethyl)siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 1 mL/min.

Split ratio 1:100.

Temperature:

— **column:** 280 °C;

— **injection port and detector:** 290 °C.

Detection Flame ionisation.

Injection 1 μ L; inject directly onto the column or via a sufficiently inert, glass-lined injection port.

Run time 1.1 times the retention time of α -tocopheryl acetate.

Relative retention With reference to α -tocopheryl acetate (retention time = about 12 min): squalane = about 0.5;

α -tocopherol = about 0.9.

System suitability:

— **resolution:** minimum 3.5 between the peaks due to

chromatogram obtained with reference solution (b);

— in the chromatogram obtained with reference solution (a), the area of the peak due to α -tocopherol is not greater than 0.002 times the area of the peak due to α -tocopheryl acetate (0.2 per cent).

Calculate the percentage content of C₃₁H₅₂O₃ from the declared content of α -tocopheryl acetate CRS.

STORAGE

In an airtight, well-filled container, protected from light.

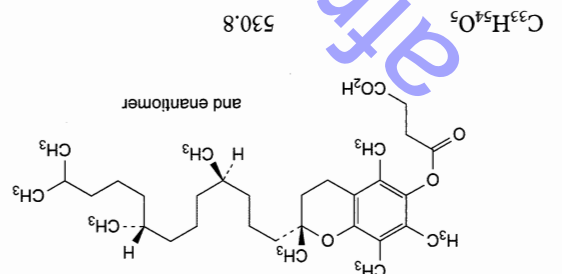
LABELLING

The label states the content of α -tocopheryl acetate, expressed in grams per 100 g of concentrate.

Ph Eur

Alpha Tocopheryl Hydrogen Succinate

(dl- α -Tocopheryl Hydrogen Succinate,
Ph Eur monograph 1258)



Action and use

Used in prevention and treatment of vitamin E deficiencies.

DEFINITION

(2R,5R)-2,5,7,8-Tetramethyl-2-[(4R,8R)-4,8,12-trimethyltridecyl]-3,4-dihydro-2H-1-benzopyran-6-yl hydrogen succinate.

Content

96.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, very soluble in methylene chloride, soluble in acetone and in anhydrous ethanol.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Absorbance (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison RRR- α -tocopheryl hydrogen succinate CRS.

C. Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 10 mg of the substance to be examined in 2 mL of cyclohexane R.

Test solution (b) In a ground-glass-stoppered tube, dissolve 10 mg of the substance to be examined in 2 mL of 2.5 M alcoholic sulfuric acid R. Heat on a water-bath for 5 min. Cool and add 2 mL of water R and 2 mL of cyclohexane R. Shake for 1 min. Use the upper layer.

Reference solution (a) Dissolve 10 mg of RRR- α -tocopheryl hydrogen succinate CRS in 2 mL of cyclohexane R.

Reference solution (b) Prepare as described for test solution (b), using RRR- α -tocopheryl hydrogen succinate CRS instead of the substance to be examined.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase glacial acetic acid R, ether R, cyclohexane R (0.2:20:80 V/V/V).

Application 10 μ L.

Development Over a path of 15 cm.

Drying In a current of air.

Detection A Examine in ultraviolet light at 254 nm.

Results A The principal spot in the chromatogram obtained with test solution (a) is similar in position and size to the principal spot in the chromatogram obtained with reference

TESTS

Optical rotation (2.2.7)

-0.01° to +0.01°.

Dissolve 2.50 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

Absorbance (2.2.25)

Solution A Dissolve 0.150 g in anhydrous ethanol R and dilute to 100 mL with the same solvent.

Test solution (a) Dilute 10.0 mL of solution A to 100.0 mL with anhydrous ethanol R.

Test solution (b) Dilute 20.0 mL of solution A to 50.0 mL with anhydrous ethanol R.

Absorption maximum At 284 nm for test solution (a).

Absorption minimum At 254 nm for test solution (b).

Specific absorbance at the absorption maximum 35 to 38 for test solution (a).

Specific absorbance at the absorption minimum 6.0 to 8.0 for test solution (b).

Acid value (2.5.1)

101 to 108, determined on 1.00 g.

Free tocopherol

Maximum 1.0 per cent.

Dissolve 0.500 g in 100 mL of 0.25 M alcoholic sulfuric acid R. Add 20 mL of water R and 0.1 mL of a 2.5 g/L solution of diphenylamine R in sulfuric acid R. Titrate with 0.01 M ammonium and cerium sulfate until a blue colour is obtained that persists for at least 5 s. Carry out a blank titration.

1 mL of 0.01 M ammonium and cerium sulfate is equivalent to 2.154 mg of free tocopherol.

Related substances

The thresholds indicated under Related substances (Table 2034.-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Heavy metals (2.4.8)

Maximum 20 ppm.

0.50 g complies with test D. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (2.2.28).

Internal standard solution Dissolve 0.300 g of doctriacontane R in hexane R and dilute to 100.0 mL with the same solvent.

S'_{Ti} = area of the peak due to DL- α -tocopheryl hydrogen succinate in the chromatogram obtained in the

interference test.

Measure the areas of the peaks due to RRR- α -tocopheryl hydrogen succinate CRS (S'_T) and dotriacontane (S'_D) in the chromatogram obtained with the test solution.

Determine the response factor (RF) for DL- α -tocopheryl hydrogen succinate from the areas of the peaks due to RRR- α -tocopheryl hydrogen succinate CRS and dotriacontane in the chromatogram obtained with the reference solution, using the following expression:

$$\frac{S'_D \times m_T}{S'_T \times m_D}$$

Calculate the percentage content of DL- α -tocopheryl hydrogen succinate using the following expression:

$$100 \times \frac{S'_D \times m_T}{S'_T \times m_D \times RF}$$

S'_D = area of the peak due to dotriacontane in the

chromatogram obtained with the reference

solution;

$S'_{D(\text{corr})}$ = corrected area of the peak due to

dotriacontane in the chromatogram obtained

with the test solution;

S'_T = area of the peak due to RRR- α -tocopheryl

hydrogen succinate CRS in the chromatogram

obtained with the reference solution;

S'_{Ti} = area of the peak due to DL- α -tocopheryl

hydrogen succinate in the chromatogram

obtained with the test solution;

m_D = mass of dotriacontane in the test solution and

in the reference solution, in milligrams;

m_T = mass of RRR- α -tocopheryl hydrogen succinate

CRS in the reference solution, in milligrams;

m = mass of the substance to be examined in the

test solution, in milligrams.

STORAGE

Protected from light.

Ph Eur

Test solution Weigh 30.0 mg of the substance to be examined

into a 20 mL vial. Add 2.0 mL of methanol R, 1.0 mL of

dimethoxypropane R and 0.1 mL of hydrochloric acid R.

Cap tightly and sonicate. Allow to stand in the dark for

1 h \pm 5 min. Remove from the dark, uncapped and evaporate

just to dryness on a steam bath with the aid of a stream of

nitrogen. Add 10.0 mL of the internal standard solution.

Vortex into solution.

Reference solution Weigh 30.0 mg of RRR- α -tocopheryl hydrogen

succinate CRS into a 20 mL vial. Add 2.0 mL of methanol R,

1.0 mL of dimethoxypropane R and 0.1 mL of hydrochloric

acid R. Cap tightly and sonicate. Allow to stand in the dark

for 1 h \pm 5 min. Remove from the dark, uncapped and

evaporate just to dryness on a steam bath with the aid of a

stream of nitrogen. Add 10.0 mL of the internal standard

solution. Vortex into solution.

Column:

— material: fused silica;

— size: 1 = 15 m, ϕ = 0.32 mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness

0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 3–6 mL/min.

Split ratio 1:10 to 1:20.

Temperature:

Time (min)	Temperature (°C)
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Column	0 - 10	250
	10 - 20	200 \rightarrow 250
Injection port	300	
Detector	330	

Detection Flame ionisation.

Inject 1 μ L; inject directly onto the column or via a glass-

lined injection port using an automatic injection device or

some other reproducible injection method.

System suitability: reference solution.

— resolution: minimum 12.0 between the peaks due to

dotriacontane and DL- α -tocopheryl hydrogen succinate.

Interference test Dissolve 0.100 g of the substance to be

examined in hexane R and dilute to 50.0 mL with the same

solvent. Inject 1 μ L of the solution and record the

chromatogram. If a peak is detected with the same retention

time as that of the peak due to dotriacontane, calculate the

area of this peak relative to the peak area of the substance to

be examined. If the relative peak area is greater than

0.5 per cent, use the corrected peak area $S'_{D(\text{corr})}$ for the

final calculation.

$$S'_{D(\text{corr})} = S'_D - \frac{S'_{Ti}}{S'_T \times S'_D}$$

S'_D = area of the peak due to dotriacontane in the

chromatogram obtained with the test solution;

S'_T = area of the peak with the same retention time as

that of the peak due to dotriacontane in the

chromatogram obtained in the interference test;

S'_{Ti} = area of the peak due to DL- α -tocopheryl hydrogen

succinate in the chromatogram obtained with the

test solution.



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Acid value (2.5.1)

101 to 108, determined on 1.00 g.

Free tocopherol

Maximum 1.0 per cent.

Dissolve 0.500 g in 100 mL of 0.25 M alcoholic sulfuric

acid R. Add 20 mL of water R and 0.1 mL of a 2.5 g/L

solution of diphenylamine R in sulfuric acid R. Titrate with

0.01 M ammonium and cerium sulfate until a blue colour is

obtained that persists for at least 5 s. Carry out a blank

titration.

1 mL of 0.01 M ammonium and cerium sulfate is equivalent to

Related substances

The thresholds indicated under Related substances

(Table 2034-1) in the general monograph Substances for

pharmaceutical use (2034) do not apply.

Heavy metals (2.4.8)

Maximum 20 ppm.

0.50 g complies with test D. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (2.2.28).

Internal standard solution Dissolve 0.300 g of dotriacontane R

in hexane R and dilute to 100.0 mL with the same solvent.

Test solution Weigh 30.0 mg of the substance to be examined

into a 20 mL vial. Add 2.0 mL of methanol R, 1.0 mL of

dimethoxypropane R and 0.1 mL of hydrochloric acid R.

Cap tightly and sonicate. Allow to stand in the dark for

1 h ± 5 min. Remove from the dark, uncapped and evaporate

just to dryness on a steam bath with the aid of a stream of

nitrogen. Add 10.0 mL of the internal standard solution.

Vortex into solution.

Reference solution Weigh 30.0 mg of RRR- α -tocopheryl hydrogen

succinate CRS into a 20 mL vial. Add 2.0 mL of methanol R,

1.0 mL of dimethoxypropane R and 0.1 mL of hydrochloric

acid R. Cap tightly and sonicate. Allow to stand in the dark

for 1 h ± 5 min. Remove from the dark, uncapped and

evaporate just to dryness on a steam bath with the aid of a

stream of nitrogen. Add 10.0 mL of the internal standard

solution. Vortex into solution.

Column:

— material: fused silica;

— size: $l = 15$ m, $\phi = 0.32$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness

0.25 μ m).

Carrier gas helium for chromatography R.

Flow rate 3–6 mL/min.

Split ratio 1:10 to 1:20.

Temperature:

Time (min)	Temperature (°C)
0 - 10	200 → 250
10 - 20	250
Injection port	300
Detector	330

Detection Flame ionisation.

Injection 1 μ L; inject directly onto the column or via a glass-lined injection port using an automatic injection device or some other reproducible injection method.

System suitability: reference solution:

— resolution: minimum 12.0 between the peaks due to

dotriacontane and RRR- α -tocopheryl hydrogen succinate.

Interference test Dissolve 0.100 g of the substance to be

examined in hexane R and dilute to 50.0 mL with the same

solvent. Inject 1 μ L of the solution and record the

chromatogram. If a peak is detected with the same retention

time as that of the peak due to dotriacontane, calculate the

area of this peak relative to the peak area of the substance to

be examined. If the relative peak area is greater than

0.5 per cent, use the corrected peak area $S'_{D(corr)}$ for the

final calculation.

$$S'_{D(corr)} = S'_D - \frac{S_I \times S'_{TI}}{S'_I \times S'_{TI}}$$

Calculate the percentage content of DL- α -tocopheryl hydrogen succinate using the following expression:

$$\frac{100 \times S'_{TI} \times m_D \times RF}{S'_D(corr) \times m}$$

S'_D = area of the peak due to dotriacontane in the

chromatogram obtained with the test solution;

S'_I = area of the peak due to dotriacontane in the

chromatogram obtained in the interference test;

S'_{TI} = area of the peak due to RRR- α -tocopheryl

hydrogen succinate in the chromatogram obtained

with the test solution;

S'_{TI} = area of the peak due to RRR- α -tocopheryl

hydrogen succinate in the chromatogram obtained

in the interference test.

Measure the areas of the peaks due to RRR- α -tocopheryl

hydrogen succinate (S_{TI}) and dotriacontane (S_D) in the

chromatogram obtained with the reference solution and the

areas of the peaks due to RRR- α -tocopheryl hydrogen

succinate (S_{TI}) and dotriacontane (S_D) in the

chromatogram obtained with the test solution.

Determine the response factor (RF) for RRR- α -tocopheryl

hydrogen succinate from the areas of the peaks due to

RRR- α -tocopheryl hydrogen succinate CRS and dotriacontane in

the chromatogram obtained with the reference solution, using

the following expression:

$$\frac{S_D \times m_{TI}}{S_{TI} \times m_D}$$

S_D = area of the peak due to dotriacontane in the

chromatogram obtained with the reference

solution;

$S'_{D(corr)}$ = corrected area of the peak due to

dotriacontane in the chromatogram obtained

with the test solution;

S'_{TI} = area of the peak due to RRR- α -tocopheryl

hydrogen succinate CRS in the chromatogram

obtained with the reference solution;

S'_{TI} = area of the peak due to RRR- α -tocopheryl

hydrogen succinate in the chromatogram

obtained with the test solution;

m_D = mass of dotriacontane in the test solution and

in the reference solution, in milligrams;

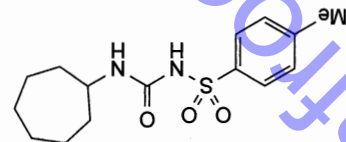
m_T = mass of R,R- α -tocopheryl hydrogen succinate
 CRS in the reference solution, in milligrams;
 = mass of the substance to be examined in the test solution, in milligrams.

STORAGE

Protected from light.

Ph Eur

Tolazamide



$C_{14}H_{21}N_3O_3S$

311.4

1156-19-0

Preparation

Tolazamide Tablets

DEFINITION

Tolazamide is 1-perhydropyrazepin-1-yl-3-tolyl-*p*-sulfonamide. It contains not less than 98.0% and not more than 101.0% of $C_{14}H_{21}N_3O_3S$, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder; odourless or almost odourless.

Very slightly soluble in water; freely soluble in chloroform; soluble in acetone; slightly soluble in ethanol (96%).

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of tolazamide (RS 342).

B. The light absorption, Appendix II B, in the range 230 to 350 nm of a 0.04% w/v solution in ethanol (96%) exhibits maxima at 256, 263 and 275 nm and a shoulder at 268 nm. The absorbances at the maxima are about 0.78, about 0.83 and about 0.62, respectively.

C. Melting point, about 165°, Appendix V A.

TESTS

Heavy metals

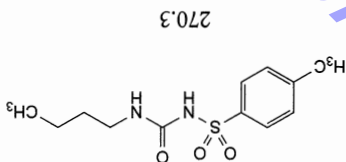
Moisten the residue obtained in the test for Sulfated ash with 1 mL of hydrochloric acid, evaporate to dryness and dissolve the residue in 20 mL of water. 12 mL of the resulting solution complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (1 ppm Pb) to prepare the standard (20 ppm).

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using silica gel G as the coating substance and a mixture of 200 volumes of chloroform, 100 volumes of methanol, 60 volumes of cyclohexane and 23 volumes of 13.5M ammonia as the mobile phase. Apply separately to the plate 10 μ L of each of two solutions in acetone containing (1) 2.0% w/v of the substance being examined and (2) 0.010% w/v of toluene-*p*-sulfonamide. After removal of the

Tolbutamide

(Ph. Eur. monograph 0304)



270.3

64-77-7

Action and use

Inhibition of ATP-dependent potassium channels (sulfonamides); treatment of diabetes mellitus.

Preparation

Tolbutamide Tablets

DEFINITION

1-Butyl-3-[(4-methylphenyl)sulfonyl]urea.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone and in ethanol (96 per cent). It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification A, C.

Second identification A, B, D.

A. Melting point (2.2.14): 126 °C to 130 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution (a) Dissolve 25.0 mg in methanol R and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 10.0 mL of test solution (a) to 250.0 mL with methanol R.

Spectral range 245–300 nm for test solution (a); 220–235 nm for test solution (b).

Absorption maxima At 258 nm, 263 nm and 275 nm for test solution (a); at 228 nm for test solution (b).

Shoulder At 268 nm for test solution (a).

Specific absorbance at the absorption maximum 480 to 520 for test solution (b).

C. Infrared absorption spectrophotometry (2.2.24).

Comparison tolbutamide CRS.

D. To 0.2 g add 8 mL of a 500 g/L solution of sulfuric acid R and heat under a reflux condenser for 30 min. Allow to cool. Crystals are formed which, after recrystallisation from hot water R and drying at 105 °C, melt (2.2.14) at 135 °C to 140 °C.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.2 g in 5 mL of dilute sodium hydroxide solution R and add 5 mL of water R.

pH (2.2.3)

4.5 to 5.5.

To 2.0 g add 50 mL of carbon dioxide-free water R and heat at 70 °C for 5 min. Cool rapidly and filter.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve 10 mg of tolbutamidesulfonamide R (impurity A) and 10 mg of tolbutamidesulfonamide R (impurity B) in the mobile phase and dilute to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 35 volumes of acetonitrile R1 and 65 volumes of a 1.36 g/L solution of potassium dihydrogen phosphate R adjusted to pH 3.5 with phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 230 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of tolbutamide.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurities A and B.

Relative retention With reference to tolbutamide (retention time = about 18 min): impurity B = about 0.2; impurity A = about 0.3.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurities A and B.

Limits:

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 1.0 g in a mixture of 15 volumes of water R and 85 volumes of acetone R and dilute to 20 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (0.5 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with a mixture of 15 volumes of water R and 85 volumes of acetone R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 20 mL of water R and 40 mL of ethanol (96 per cent) R. Titrate with 0.1 M sodium hydroxide, using 1 mL of phenolphthalein solution R as indicator.

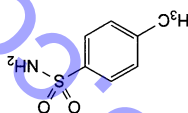
1 mL of 0.1 M sodium hydroxide is equivalent to 27.03 mg of $C_{12}H_{18}N_2O_5S$.

STORAGE

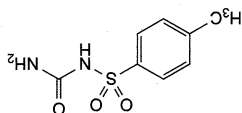
In an airtight container.

IMPURITIES

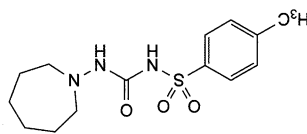
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B, C.



A. (4-methylphenyl)sulfonamide (toluenesulfonamide),



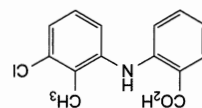
B. 1-[(4-methylphenyl)sulfonyl]urea (toluenesulfonylurea),



C. 1-azepan-1-yl-3-[(4-methylphenyl)sulfonyl]urea (tolazamide).

Tolfenamic Acid

(Ph. Eur. monograph 2039)

C₁₄H₁₂ClNO₂ 261.7 13710-19-5

Action and use

Cyclo-oxygenase inhibitor; analgesic; anti-inflammatory.

Ph Eur

DEFINITION

2-[(3-Chloro-2-methylphenyl)amino]benzoic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in dimethylformamide, sparingly soluble in ethanol and in methylene chloride.

It dissolves in dilute solutions of alkali hydroxides.

mp

About 213 °C.

IDENTIFICATION

First identification B

Second identification A, C

A. Dissolve 20 mg in a mixture of 1 volume of 1 M

hydrochloric acid and 99 volumes of methanol R and dilute to

100 mL with the same mixture of solvents. Dilute 5.0 mL of

the solution to 50 mL with a mixture of 1 volume of 1 M

hydrochloric acid and 99 volumes of methanol R. Examined

absorption maxima, at 286 nm and 345 nm. The ratio of the

absorbance measured at the maximum at 286 nm to that

measured at the maximum at 345 nm is 1.2 to 1.4.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison tolfenamic acid CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 25 mg of the substance to be examined

in a mixture of 1 volume of methanol R and 3 volumes of

methylene chloride R and dilute to 10 mL with the same

mixture of solvents.

Reference solution Dissolve 25 mg of tolfenamic acid CRS in a

mixture of 1 volume of methanol R and 3 volumes of

methylene chloride R and dilute to 10 mL with the same

mixture of solvents.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase glacial acetic acid R, dioxan R, toluene R

(1:25:90 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of warm air.

Detection Ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 5 mL of ethanol R and dilute to 50.0 mL with

the mobile phase.

Reference solution (a) Dissolve 25 mg of 2-chlorobenzoic acid R

and 25 mg of 3-chloro-2-methylaniline R in 5 mL of ethanol R

and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL

of the solution to 50.0 mL with the mobile phase. Dilute

1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to

10.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 100.0 mL with the mobile phase.

Column:

— size: l = 0.25 m, Ø = 4.6 mm,

— stationary phase: spherical end-capped octadecylsilyl silica gel

for chromatography R (5 µm) with a specific surface area of

450 m²/g and a pore size of 8 nm.

Mobile phase glacial acetic acid R, water R, ethanol R

(2:350:650 V/V/V).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 232 nm.

Injection 20 µL.

Run time 3 times the retention time of tolfenamic acid.

Relative retention With reference to tolfenamic acid (retention

time = about 15 min): impurity A = about 0.25;

impurity B = about 0.34.

System suitability: reference solution (a):

— resolution: minimum 2.5 between the peaks due to

impurity A and to impurity B.

Limits:

— impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (a) (0.1 per cent),

— impurity B: not more than half the area of the

corresponding peak in the chromatogram obtained with

reference solution (a) (0.05 per cent),

— any other impurity: not more than the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.1 per cent),

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.5 per cent),

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.01 per cent).

Copper

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method D).

Test solution Place 1.00 g of the substance to be examined in

a silica crucible, moisten with sulfuric acid R, heat cautiously

on a flame for 30 min and then progressively to about

650 ± 50 °C. Continue ignition until all black particles have

disappeared. Allow to cool, dissolve the residue in 0.1 M

hydrochloric acid and dilute to 25.0 mL with the same acid.

Reference solutions Prepare the reference solutions using copper

standard solution (0.1 per cent Cu) R, diluted as necessary

using 0.1 M nitric acid.

Source Copper hollow-cathode lamp.

Wavelength 324.8 nm.

Flame Air-acetylene.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g with the aid of ultrasound in 100 mL of ethanol R. Add 0.1 mL of phenol red solution R and titrate

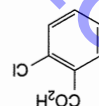
1 mL of 0.1 M sodium hydroxide is equivalent to 26.17 mg of $C_{14}H_{12}ClNO_2$.

STORAGE

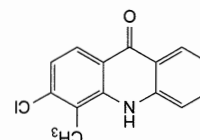
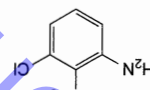
Protected from light.

IMPURITIES

A. 2-chlorobenzoic acid,



B. 3-chloro-2-methylaniline,

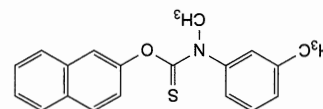


C. 3-chloro-4-methyl-9-oxo-9,10-dihydroacridine.

Ph Eur

Tolnafate

(Ph. Eur. monograph 1158)



$C_{19}H_{17}NO_2$

307.4

2398-96-1

Action and use

Antifungal.

Ph Eur

DEFINITION

O-Naphthalen-2-yl methyl(3-methylphenyl)carbamothioate.

Content

97.0 per cent to 103.0 per cent (dried substance).

CHARACTERS**Appearance**

White or yellowish-white powder.

Related substances

— *impurity D*: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (20 ppm).

Limit:

— *impurity D* in the chromatogram obtained with reference solution (b).
— *symmetry factor*: maximum 1.9 for the peak due to *impurity D* and tolnafate in the chromatogram obtained with reference solution (c);
— *resolution*: minimum 5.0 between the peaks due to

System suitability:

time = about 15 min; *impurity D* = about 0.25.

Relative retention With reference to tolnafate (retention

solution (b); 10 µL of reference solution (c).

Injection 100 µL of the test solution and reference

Detection Spectrophotometer at 254 nm.

Flow rate 1.0 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	70	30
3 - 8	70 → 0	30 → 100
8 - 20	0	100

Mobile phase:
— *mobile phase A*: trifluoroacetic acid R, methanol R, water R (0.1:10:90 V/V/V);
— *mobile phase B*: trifluoroacetic acid R, water R, methanol R (0.1:10:90 V/V/V);
— *stationary phase*: octadecylsilyl silica gel for chromatography R (5 µm).

Column:

— *size*: $l = 0.15$ m, $\varnothing = 4.6$ mm;

methanol R.

examine in 25 mL of *methanol R*. Add 2 mL of this solution to 2 mL of reference solution (a) and dilute to 25 mL with *methanol R*.

Reference solution (c) Dissolve 10 mg of the substance to be examined in 25 mL of *methanol R*. Add 2 mL of this solution to 2 mL of reference solution (a) and dilute to 25 mL with *methanol R*.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with *methylen chloride R*. Take 2.0 mL of this

Reference solution (a) Dissolve 20.0 mg of *N-methyl-m-toluidine R* (impurity D) in 50.0 mL of *methylen chloride R*.

0.01 M hydrochloric acid.

Combine the aqueous phases and dilute to 10.0 mL with 3 quantites, each of 3 mL, of 0.01 M hydrochloric acid.

Test solution Dissolve 0.400 g of the substance to be examined in 2 mL of *methylen chloride R*. Extract with

Liquid chromatography (2.2.29).

Impurity D**TESTS**

Comparison tolnafate CRS.

Infrared absorption spectrophotometry (2.2.24).

IDENTIFICATION

(96 per cent).

Practically insoluble in water, freely soluble in acetone and in methylen chloride, very slightly soluble in ethanol

Solubility

Test solution Dissolve 25 mg of the substance to be examined in 5 mL of methanol R and dilute to 25.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with methanol R. Dilute 1.0 mL of this solution to 10.0 mL with methanol R.

Reference solution (b) Dissolve 5 mg of tolnafate for system suitability CRS (containing resolution component A) in 5.0 mL of methanol R.

Column:
— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;
— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).
Mobile phase:
— mobile phase A: trifluoroacetic acid R, water R, methanol R (0.1:30:70 V/V/V);
— mobile phase B: trifluoroacetic acid R, water R, methanol R (0.1:10:90 V/V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 12	100	0
12 - 30	100 \rightarrow 0	0 \rightarrow 100
30 - 33	0	100

Flow rate 1.0 mL/min.
Detection Spectrophotometer at 254 nm.
Injection 10 μ L.

Relative retention With reference to tolnafate (retention time = about 18 min): resolution component A = about 0.7.
System suitability: reference solution (b):
— resolution: minimum 5.0 between the peaks due to resolution component A and tolnafate.

Limits:
— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);
— **total:** not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);
— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

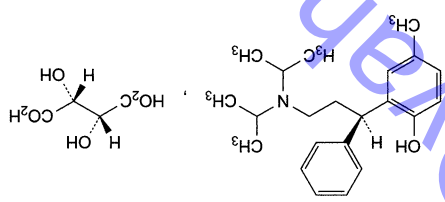
Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.00 g by drying at 60 °C at a pressure not exceeding 0.7 kPa for 3 h.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 50.0 mg in methanol R and dilute to 250.0 mL with the same solvent. Dilute 2.0 mL of this solution to 50.0 mL with methanol R. Measure the absorbance (2.2.25) at the absorption maximum at 257 nm.
Calculate the content of $C_{19}H_{17}NO_5$ taking the specific absorbance to be 720.

STORAGE
Protected from light.
IMPURITIES
Specified impurities D.
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of

Tolerodine Tartrate

(Ph. Eur. monograph 2781)



$C_{26}H_{37}NO_7$ 475.6

124937-52-6

Action and use
Anticholinergic.

DEFINITION

2-[(1*R*)-3-[Bis(1-methylethylamino)-1-phenylpropyl]-4-methylphenol (2*R*,3*R*)-2,3-dihydroxybutanedioate].

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Sparsely soluble in water, slightly soluble in anhydrous ethanol, practically insoluble in heptane.

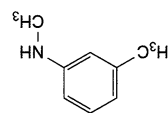
IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

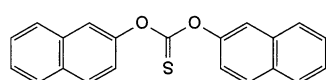
B. Enantiomeric purity (see Tests).

C. It gives reaction (b) of tartrates (2.3.1).

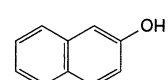
D. *N*,3-dimethylaniline (*N*-methyl-*m*-toluidine).



B. *O*,*O*-dinaphthalen-2-yl carbodithioate.



A. naphthalen-2-ol (β -naphthol).



the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, B.

TESTS

Enantiomeric purity

Liquid chromatography (2.2.29).

Solution A Dissolve 0.29 g of sodium dihydrogen phosphate monohydrate R and 0.47 g of disodium hydrogen phosphate dihydrate R in 1 L of water R. Dissolve 1.04 g of tetrabutylammonium bromide R in this solution.

Test solution Dissolve 4.0 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 2 mg of tolterodine for system

stability CRS (containing impurity F) in the mobile phase

and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dilute 0.5 mL of the test solution to

100.0 mL with the mobile phase.

Column:

— size: $l = 0.10$ m, $\phi = 2$ mm;

— stationary phase: α_1 -acid-glycoprotein silica gel for chiral

separation R (5 μ m).

Mobile phase 2-methylpropanol R, solution A (7:93 V/V).

Flow rate 0.2 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Run time 1.5 times the retention time of tolterodine.

Relative retention With reference to tolterodine (retention

time = about 23 min): impurity F = about 0.9.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 2.5, where H_p = height

above the baseline of the peak due to impurity F and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

tolterodine.

Calculation of percentage content:

— for impurity F, use the concentration of tolterodine in

reference solution (b).

Limit:

— impurity F: maximum 0.6 per cent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in 10 mL of methanol R1, sonicate until dissolution is

complete and dilute to 25.0 mL with methanol R1.

Reference solution (a) Dilute 1.0 mL of the test solution to

10.0 mL with methanol R1.

Reference solution (b) Dissolve 2 mg of tolterodine

impurity E CRS in methanol R1 and dilute to 10.0 mL with

methanol R1. Dilute 1.0 mL of the solution to 20.0 mL with

methanol R1.

Reference solution (c) Dissolve 25 mg of the substance to be

examined in 10 mL of reference solution (b) and dilute to

25.0 mL with methanol R1.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: base-deactivated end-capped octadecylsilyl

silica gel for chromatography R (5 μ m).

Mobile phase:

— mobile phase A: mix 5 volumes of triethylamine R2 and

450 volumes of a 2.88 g/L solution of ammonium

dihydrogen phosphate R3; adjust to pH 5.9 with a

D, E, G.

Control of impurities in substances for pharmaceutical use: A, C,

impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph *Substances for pharmaceutical use*

acceptance criterion for other/unspecified impurities and/or

the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities F

IMPURITIES

$C_{26}H_{37}NO_7$.

1 mL of 0.1 M perchloric acid is equivalent to 47.56 mg of

(2.2.20).

perchloric acid, determining the end-point potentiometrically

sonicate until dissolution is complete. Titrate with 0.1 M

Dissolve 0.400 g in 30.0 mL of anhydrous acetic acid R and

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C for 3 h.

Maximum 0.5 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

to tartaric acid.

— reporting threshold: 0.05 per cent; disregard the peak due

— total: maximum 0.3 per cent;

0.10 per cent;

— unspecified impurities: for each impurity, maximum

Limits:

reference solution (a).

— for each impurity, use the concentration of tolterodine in

Calculation of percentage contents:

impurity E and tolterodine.

— resolution: minimum 1.5 between the peaks due to

System suitability: reference solution (c):

impurity B = about 0.9.

time = about 7 min): tartaric acid = about 0.2;

Relative retention With reference to tolterodine (retention

impurity B.

with reference solution (c) to identify the peak due to

Identification of impurities Use the chromatogram obtained

and (c).

Injection 20 μ L of the test solution and reference solutions (a)

Detection Spectrophotometer at 220 nm.

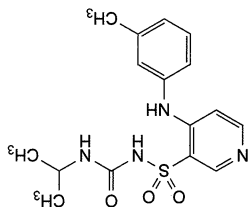
Flow rate 1.5 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 25	100	0
25 - 45	100 → 80	0 → 20

50 per cent V/V solution of phosphoric acid R3;
add 550 volumes of methanol R1;
— mobile phase B: methanol R1;

Anhydrous Torasemide

(Ph Eur monograph 2132)



$C_{16}H_{20}N_4O_3S$ 348.4 56211-40-6

Action and use

Thiazide-like diuretic.

DEFINITION

1-(1-Methylethyl)-3-[[4-[(3-methylphenyl)amino]pyridin-3-yl]sulfonyl]urea.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, slightly soluble in ethanol (96 per cent). It is sparingly soluble in dilute solutions of alkali hydroxides and slightly soluble in dilute acids.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison anhydrous torasemide CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

Liquid chromatography (2.2.29)
Solution A Dissolve 2.7 g of potassium dihydrogen phosphate R in 950 mL of water R, adjust to pH 3.5 with phosphoric acid R and dilute to 1000 mL with water R.
Test solution Dissolve 20.0 mg of the substance to be examined in 15 mL of solution A, cool to room temperature and Add 22.5 mL of solution A, cool to room temperature and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 2.0 mg of torasemide for system suitability CRS (containing impurities A, B, C and D) in 2.5 mL of methanol R and dilute to 5.0 mL with solution A.

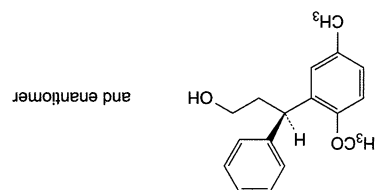
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve the contents of a vial of torasemide impurity E CRS in 0.5 mL of methanol R. Add 0.5 mL of solution A.

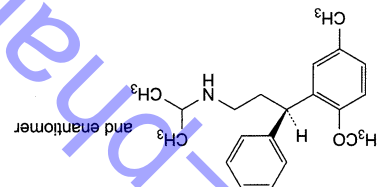
Column:

— size: $l = 0.125$ m, $\phi = 4.0$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
— temperature: 30 °C.

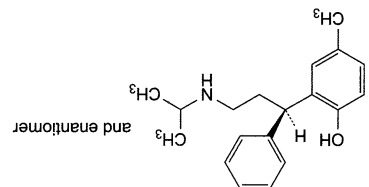
A. (3R,5S)-3-(2-methoxy-5-methylphenyl)-3-phenylpropan-1-ol,



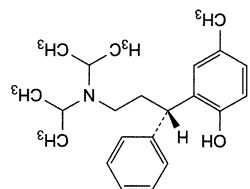
C. (3R,5S)-3-(2-methoxy-5-methylphenyl)-N,N-bis(1-methylethyl)-3-phenylpropan-1-amine,



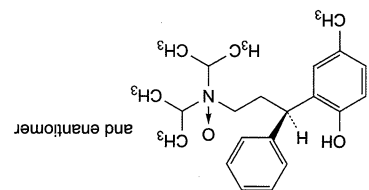
D. (3R,5S)-3-(2-methoxy-5-methylphenyl)-N-(1-methylethyl)-3-phenylpropan-1-amine,



E. 4-methyl-2-[(1R,5S)-3-[(1-methylethyl)amino]-1-phenylpropyl]phenol,



F. 2-[(1S)-3-[bis(1-methylethyl)amino]-1-phenylpropyl]-4-methylphenol,



G. 2-[(1R)-3-[bis(1-methylethyl)amino]-1-phenylpropyl]-4-methylphenol N-oxide.



Mobile phase methanol R, solution A (40:60 V/V).

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 288 nm.

Injection 20 µL.

Run time 2.5 times the retention time of torasemide.

Identification of impurities Use the chromatogram supplied

with torasemide for system suitability CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities A, B, C and D; use the

chromatogram obtained with reference solution (c) to identify

the peak due to impurity E.

Relative retention With reference to torasemide (retention

time = about 10 min): impurity A = about 0.3;

impurity B = about 0.4; impurity C = about 0.5;

impurity E = about 0.7; impurity D = about 2.3.

System suitability:

— resolution: minimum 3.0 between the peaks due to

impurities B and C in the chromatogram obtained with

reference solution (a);

— signal-to-noise ratio: minimum 100 for the principal peak

in the chromatogram obtained with reference solution (b).

Limits:

— correction factors: for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity A = 5.1;

impurity B = 0.76;

— impurity B: not more than 5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.5 per cent);

— impurities A, C, D, E: for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.15 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— total: not more than 6 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.6 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test F. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of anhydrous acetic acid R. Titrate

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 34.84 mg

of C₁₆H₂₀N₄O₃S.

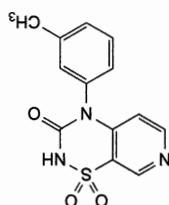
STORAGE

Protected from light.

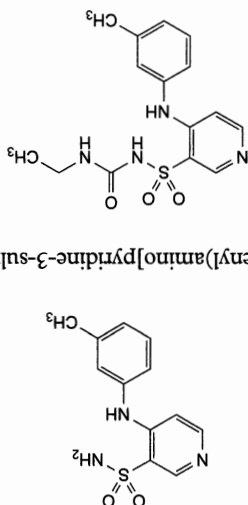
IMPURITIES

Specified impurities A, B, C, D, E

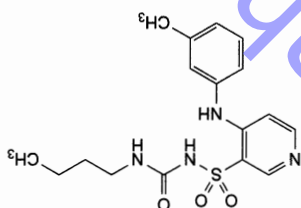
A. 4-(3-methylphenyl)-2H-pyrido[4,3-e]-1,2,4-thiadiazin-3(4H)-one 1,1-dioxide,



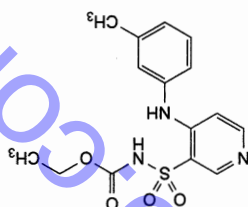
B. 4-[(3-methylphenyl)amino]pyridine-3-sulfonamide,



C. 1-ethyl-3-[[4-[(3-methylphenyl)amino]pyridin-3-yl]sulfonyl]urea,



D. 1-butyl-3-[[4-[(3-methylphenyl)amino]pyridin-3-yl]sulfonyl]urea,

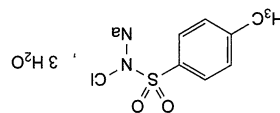


E. ethyl [[4-[(3-methylphenyl)amino]pyridin-3-yl]sulfonyl]carbamate.

Ph Eur

Tosylchloramide Sodium

(Ph. Eur. monograph 0381)



$C_7H_7ClNNaO_2S_3H_2O$ 281.7 127-65-1

Action and use

Antiseptic.

DEFINITION

Sodium N-chloro-4-methylbenzenesulfonimide trihydrate.

Content

98.0 per cent to 103.0 per cent of $C_7H_7ClNNaO_2S_3H_2O$.

CHARACTERS

Appearance

White or slightly yellow, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. Solution S (see Tests) turns red litmus paper R blue and then bleaches it.

B. To 10 mL of solution S add 10 mL of dilute hydrogen peroxide solution R. A white precipitate is formed which dissolves on heating. Filter the hot solution and allow to cool. White crystals are formed which, when washed and dried at 100-105 °C, melt (2.2.14) at 137 °C to 140 °C.

C. Ignite cautiously 1 g, because of the risk of deflagration. Dissolve the residue in 10 mL of water R. The solution gives reaction (a) of chlorides (2.3.1).

D. The solution prepared for identification test C gives reaction (a) of sulfates (2.3.1).

E. The solution prepared for identification test C gives reaction (b) of sodium (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and is colourless (2.2.2, Method II).

pH (2.2.3)

8.0 to 10.0 for solution S.

Ortho compound

To 2 g add 10 mL of water R, mix, add 1 g of sodium metabisulfite R and heat to boiling. Cool to 0 °C, filter rapidly and wash with 3 quantities, each of 5 mL, of iced water R. The precipitate, dried over diphosphorus pentoxide R at a pressure not exceeding 600 Pa, melts (2.2.14) at a minimum of 134 °C.

Residue insoluble in anhydrous ethanol

Maximum 2 per cent.

Shake 1.00 g with 20 mL of anhydrous ethanol R for 30 min, filter on a tared filter, wash any residue with 5 mL of anhydrous ethanol R and dry at 100-105 °C. The residue weighs a maximum of 20 mg.

Tragacanth

(Ph. Eur. monograph 0532)



DEFINITION

Air-hardened, gummy exudate, flowing naturally or obtained by incision from the trunk and branches of *Asragalus* *gummifer* Labill. and certain other species of *Asragalus* from western Asia.

IDENTIFICATION

A. Tragacanth occurs in thin, flattened, ribbon-like, white or pale yellow, translucent strips, about 30 mm long and 10 mm wide and up to 1 mm thick, more or less curved, horny, with a short fracture; the surface is marked by fine longitudinal striae and concentric transverse ridges. It may also contain pieces similar in shape but somewhat thicker, more opaque and more difficult to fracture.

B. Reduce to a powder (355) (2.9.12). The powder is white or almost white and forms a mucilaginous gel with about 10 times its mass of water R. Examine under a microscope using a 50 per cent V/V solution of glycerol R. The powder shows in the gummy mass numerous stratified cellular membranes that turn slowly violet when treated with iodinated zinc chloride solution R. The gummy mass includes starch grains, isolated or in small groups, usually rounded in shape and sometimes deformed, with diameters varying between 4 µm and 10 µm, occasionally up to 20 µm, and a central hilum visible between crossed nicol prisms.

C. Examine the chromatograms obtained in the test for acacia.

Results The chromatogram obtained with the test solution shows 3 zones due to galactose, arabinose and xylose. A faint yellowish zone at the solvent front and a greyish-green zone between the zones due to galactose and arabinose may be present.

D. Moisten 0.5 g of the powdered herbal drug (355) (2.9.12) with 1 mL of ethanol (96 per cent) R and add gradually, while shaking, 50 mL of the mucilage add 5 mL of water R and obtained. To 5 mL of the mucilage add 5 mL of water R and 2 mL of barium hydroxide solution R. A slight flocculent precipitate is formed. Heat on a water-bath for 10 min. An intense yellow colour develops.

ASSAY

Dissolve 0.125 g in 100 mL of water R in a ground-glass-stoppered flask. Add 1 g of potassium iodide R and 5 mL of dilute sulfuric acid R. Allow to stand for 3 min. Titrate with 0.1 M sodium thiosulfate, using 1 mL of starch solution R as indicator.

1 mL of 0.1 M sodium thiosulfate is equivalent to 14.08 mg of $C_7H_7ClNNaO_2S_3H_2O$.

STORAGE

In an airtight container, protected from light.

Ph Eur

Ph Eur

TESTS

Acacia

Thin-layer chromatography (2.2.27).

Test solution To 100 mg of the powdered herbal drug (355) in a thick-walled centrifuge test-tube, add 2 mL of a 100 g/L solution of *trifluoroacetic acid* R, shake vigorously to dissolve the forming gel, stopper the test-tube and heat the mixture at 120 °C for 1 h. Centrifuge the resulting hydrolysate, transfer the clear supernatant carefully into a 50 mL flask, add 10 mL of *water* R and evaporate the solution to dryness under reduced pressure. To the resulting clear film add 0.1 mL of *water* R and 0.9 mL of *methanol* R. Centrifuge to separate the amorphous precipitate, collect the supernatant and, if necessary, dilute to 1 mL with *methanol* R.

Reference solution Dissolve 10 mg of *arabinose* R, 10 mg of *galactose* R, 10 mg of *rhamnose* R and 10 mg of *xylose* R in 1 mL of *water* R and dilute to 10 mL with *methanol* R.

Plate TLC silica gel plate R.

Mobile phase 16 g/L solution of sodium dihydrogen phosphate R, *butanol* R, *acetic acid* R (10:40:50 V/V/V).

Application 10 µL as bands.

Development A Over a path of 10 cm.

Drying A In a current of warm air for a few minutes.

Development B Over a path of 15 cm using the same mobile phase.

Drying B At 110 °C for 10 min.

Detection Spray with *anisaldehyde solution* R and dry at 110 °C for 10 min.

Results The chromatogram obtained with the reference solution shows 4 clearly separated coloured zones due to galactose (greyish-green or green), *arabinose* (yellowish-green), *xylose* (greenish-grey or yellowish-grey) and *rhamnose* (yellowish-green), in order of increasing *R_F* value;

the chromatogram obtained with the test solution does not show a yellowish-green zone corresponding to the zone of *rhamnose* in the chromatogram obtained with the reference solution.

Methylcellulose

Examine the chromatograms obtained in the test for *acacia*. **Results** The chromatogram obtained with the test solution does not show a red zone near the solvent front.

Sterculia gum

A. Place 0.2 g of the powdered herbal drug (355) (2.9.12) in a 10 mL ground-glass-stoppered cylinder graduated in 0.1 mL. Add 10 mL of *ethanol* (60 per cent V/V) R and shake. Any gel formed occupies not more than 1.5 mL.

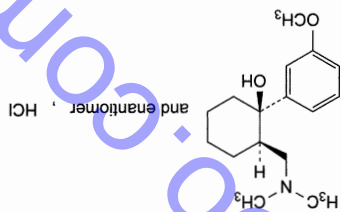
B. To 1.0 g of the powdered herbal drug (355) (2.9.12) add 100 mL of *water* R and shake. Add 0.1 mL of *methyl red* solution R. Not more than 5.0 mL of 0.01 M *sodium hydroxide* is required to change the colour of the indicator.

Foreign matter

Place 2.0 g of the powdered herbal drug (355) (2.9.12) in a 250 mL round-bottomed flask and add 95 mL of *methanol* R. Swirl to moisten the powder and add 60 mL of *hydrochloric acid* R1. Add a few glass beads about 4 mm in diameter and heat on a water-bath under a reflux condenser for 3 h, shaking occasionally. Remove the glass beads and filter the hot suspension *in vacuo* through a sintered-glass filter (160) (2.1.2). Rinse the flask with a small quantity of *water* R and pass the rinsings through the filter. Wash the residue on the filter with about 40 mL of *methanol* R and dry to constant

Tramadol Hydrochloride

(Ph. Eur. monograph 1681)



C₁₆H₂₆ClNO₂ 299.8

36282-47-0

Action and use

Opioid receptor agonist; noradrenaline reuptake inhibitor; analgesic.

µ-Opioid receptor (OP₃, MOR) agonist and noradrenaline reuptake inhibitor; analgesic

Preparations

Tramadol Capsules
Prolonged-release Tramadol Capsules
Prolonged-release Tramadol Tablets



Ph Eur

The label states whether or not the contents are suitable for preparing emulsions.

LABELLING

Absence of *Escherichia coli* (2.6.13).

TAMC: acceptance criterion 10⁴ CFU/g (2.6.12).
TYMC: acceptance criterion 10² CFU/g (2.6.12).

Microbial contamination

Maximum 4.0 per cent.

Total ash (2.4.16)

The average value of the last 3 determinations. **Total ash** (2.4.16) Maximum 4.0 per cent. **Microbial contamination** TAMC: acceptance criterion 10⁴ CFU/g (2.6.12). TYMC: acceptance criterion 10² CFU/g (2.6.12). Absence of *Escherichia coli* (2.6.13). **LABELLING** The label states whether or not the contents are suitable for preparing emulsions. **Action and use** Opioid receptor agonist; noradrenaline reuptake inhibitor; analgesic. **Preparations** Tramadol Capsules Prolonged-release Tramadol Capsules Prolonged-release Tramadol Tablets

Flow time

mass at 110 °C (about 1 h). Allow to cool in a desiccator and weigh. The residue weighs a maximum of 20 mg.

DEFINITION

(1*R*,2*R*)-2-[(1*R*,2*R*)-6-(3-methoxyphenyl)cyclohexanol hydrochloride].

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in methanol, very slightly soluble in acetone.

IDENTIFICATION

First identification B, D

Second identification A, C, D

A. Melting point (2.2.14): 180 °C to 184 °C.

B. Infrared absorption spectrophotometry (2.2.24).

Comparison tramadol hydrochloride CRS.

C. Chromatograms obtained in the test for impurity E.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in water R and dilute to 20 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity

To 10 mL of solution S, add 0.2 mL of methyl red solution R and 0.2 mL of 0.01 M hydrochloric acid. The solution is red.

Not more than 0.4 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to yellow.

Optical rotation (2.2.7)

−0.10° to +0.10°, determined on solution S.

Impurity E

Thin-layer chromatography (2.2.27).

Test solution (a) Dissolve 0.10 g in methanol R and dilute to 2 mL with the same solvent.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with methanol R.

Reference solution (a) Dissolve 25 mg of tramadol hydrochloride CRS in methanol R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of tramadol impurity E CRS in 5 mL of methanol R. Dilute 1 mL of the solution to 10 mL with methanol R.

Reference solution (c) Dissolve 5 mg of tramadol impurity A CRS in 1 mL of reference solution (a).

Plate TLC silica gel F₂₅₄ plate R, prewashed with methanol R. Mobile phase concentrated ammonia R, 2-propanol R, toluene R (1:19:80 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate. Saturate the plate for 20 min with concentrated ammonia R. For this, add concentrated ammonia R to one trough of a twin trough tank. Just before developing, add the mobile phase to the other trough. Place the plate in the chromatographic tank, ensuring

that the layer of silica gel is orientated towards the middle of the tank.

Drying In air.

Detection Expose the plate to iodine vapour for 1 h, examine in ultraviolet light at 254 nm.

System suitability The chromatogram obtained with reference solution (c) shows 2 clearly separated spots.

Limit: test solution (a):

— impurity E: any spot corresponding to impurity E is not more intense and not greater than the spot in the chromatogram obtained with reference solution (b)

(0.2 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.15 g of the substance to be examined in the mobile phase and dilute to 100 mL with the mobile phase.

Reference solution (a) Dilute 2.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of tramadol impurity A CRS in 4.0 mL of the test solution and dilute to 100 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: end-capped base-deactivated octylsilyl silica gel for chromatography R (5 µm).

Mobile phase 295 volumes of acetonitrile R and 705 volumes of a mixture of 0.2 mL of trifluoroacetic acid R and 100 mL of water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 20 µL.

Run time 4 times the retention time of tramadol.

Relative retention With reference to tramadol (retention time = about 5 min): impurity A = about 0.85.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurity A and tramadol.

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.4 per cent);

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution

Water (2.5.12)

(2 ppm Pb) R.

Maximum 0.5 per cent, determined on 1.000 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.180 g in 25 mL of anhydrous acetic acid R and add 10 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 29.98 mg of $C_{16}H_{26}ClNO_2$.

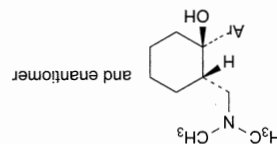
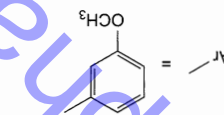
STORAGE

Protected from light.

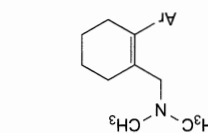
IMPURITIES

Specified impurities A, B

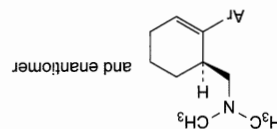
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D.



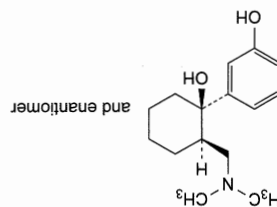
A. (1RS,2SR)-2-[(dimethylamino)methyl]-1-(3-methoxyphenyl)cyclohexanol,



B. [2-(3-methoxyphenyl)cyclohex-1-enyl]-N,N-dimethylmethanamine,



C. (1RS)-[2-(3-methoxyphenyl)cyclohex-2-enyl]-N,N-dimethylmethanamine,



D. (1RS,2RS)-2-[(dimethylamino)methyl]-1-(3-hydroxyphenyl)cyclohexanol,

DEFINITION

N-(5,6,7,8-Tetrahydronaphthalen-1-yl)-4,5-dihydro-1H-imidazol-2-amine hydrochloride monohydrate.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Soluble in water and in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tramazoline hydrochloride monohydrate CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS**Solution S**

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

pH (2.2.3)

4.9 to 6.3 for solution S.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in a mixture of 50 volumes of acetonitrile R and 50 volumes of water R and dilute to 50.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 5.0 mg of tramazoline impurity A CRS and 5.0 mg of tramazoline impurity B CRS in 5 mL of a mixture of 50 volumes of acetonitrile R and 50 volumes of water R and add 5 mL of the test solution.

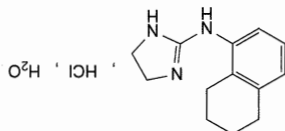
Reference solution (b) Dilute 0.2 mL of reference solution (a) to 100 mL with a mixture of 50 volumes of acetonitrile R and 50 volumes of water R.

Column:

— size: $l = 0.125$ m, $\varnothing = 4$ mm,

Tramazoline Hydrochloride Monohydrate

(Ph. Eur. monograph 1597)



$C_{13}H_{18}ClN_3 \cdot H_2O$ 269.8

74195-73-6

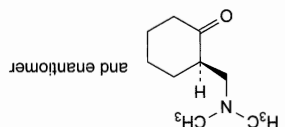
Action and use

Alpha-adrenoceptor agonist.

Ph. Eur.

Ph. Eur.

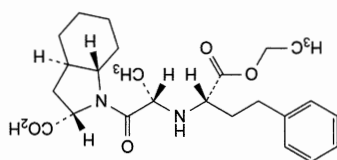
E. (2RS)-2-[(dimethylamino)methyl]cyclohexanone.





Tramadolipril

(Ph. Eur. monograph 2245)



$C_{24}H_{34}N_2O_5$ 430.5 87679-37-6

Action and use

Angiotensin converting enzyme inhibitor.

Preparation

Tramadolipril Capsules

Ph Eur

DEFINITION

(2S,3aR,7aS)-1-[(2S)-2-[[[(1S)-1-(Ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid.

CHARACTERS

99.0 per cent to 101.0 per cent (anhydrous substance).

Appearance

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in anhydrous ethanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison tramadolipril CRS.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution Y₇ (2.2.2 Method II).

Dissolve 1.0 g in methanol R and dilute to 10 mL with the same solvent.

Specific optical rotation (2.2.7)

–16.5 to –18.5 (anhydrous substance).

Dissolve 1.0 g in anhydrous ethanol R and dilute to 50.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase A and dilute to 10.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 100.0 mL with mobile phase A.

Reference solution (b) Dissolve 5 mg of tramadolipril impurity C CRS and 5 mg of tramadolipril impurity D CRS in mobile phase A and dilute to 5 mL with mobile phase A.

Dilute 1 mL of this solution to 20 mL with mobile phase A.

Column:

size: $l = 0.15$ m, $\varnothing = 4.6$ mm,

stationary phase: end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R (3.5 μ m),

temperature: 40 °C.

stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase 2.0 g/L solution of sodium dodecyl sulfate R in a

mixture of 6 volumes of 2-propanol R, 42 volumes of

acetonitrile R and 52 volumes of water R.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 5 μ L.

Run time 3 times the retention time of tramadolipril.

Relative retention With reference to tramadolipril (retention

time = about 6.5 min): impurity A = about 0.71;

impurity B = about 0.86.

System suitability: reference solution (a):

the chromatogram obtained shows 3 clearly separated

peaks,

resolution: minimum 1.5 between tramadolipril and

impurity B.

Limits:

impurity A: not more than 3 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (0.3 per cent);

impurity B: not more than 3 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (0.3 per cent);

any other impurity: not more than the area of the peak due

to impurity B in the chromatogram obtained with

reference solution (b) (0.1 per cent);

sum of other impurities: not more than twice the area of the

peak due to impurity B in the chromatogram obtained

with reference solution (b) (0.2 per cent);

disregard limit: 0.2 times the area of the peak due to

impurity B in the chromatogram obtained with reference

solution (b) (0.02 per cent).

Water (2.5.12)

6.2 per cent to 7.2 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 2.000 g in a mixture of 5 mL of 0.1 M hydrochloric

acid and 75 mL of ethanol (96 per cent) R. Carry out a

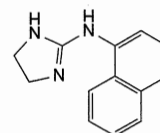
potentiometric titration (2.2.20) using 1 M sodium hydroxide.

Read the volume added between the 2 points of inflexion.

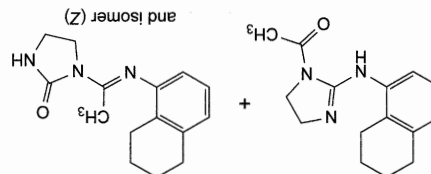
1 mL of 1 M sodium hydroxide is equivalent to 251.8 mg

of $C_{24}H_{34}N_2O_5$.

IMPURITIES



A. N-(naphthalen-1-yl)-4,5-dihydro-1H-imidazol-2-amine,



B. mixture of 1-acetyl-N-(5,6,7,8-tetrahydronaphthalen-1-yl)-4,5-dihydro-1H-imidazol-2-amine and 1-[(E)-1-[(5,6,7,8-tetrahydronaphthalen-1-yl)imino]ethyl]imidazolidin-2-one.

Ph Eur

Mobile phase:
 — mobile phase A: mix 25 volumes of acetonitrile R₁ and 75 volumes of a 6.8 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 2.5 ± 0.1 with phosphoric acid R₃
 — mobile phase B: mix equal volumes of acetonitrile R₁ and a 6.8 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 2.2 ± 0.1 with phosphoric acid R₃

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 20	95	5
20 - 35	95 → 5	5 → 95
35 - 45	5	95

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL.

Relative retention With reference to trandolapril (retention time = about 14.5 min): impurity C = about 2.1; impurity D = about 2.5.

System suitability: reference solution (b):

— resolution: minimum 4 between the peaks due to impurity C and impurity D.

Limits:

— correction factor: for the calculation of content, multiply the peak area of impurity C by 2.2,

— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference

solution (a) (0.2 per cent),

— impurity D: not more than 5 times the area of the principal peak in the chromatogram obtained with

reference solution (a) (0.5 per cent),

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.1 per cent),

— sum of impurities other than D: not more than 5 times the area of the principal peak in the chromatogram obtained

with reference solution (a) (0.5 per cent),

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Palladium

Maximum 5 ppm.

Atomic absorption spectrometry (2.2.23, Method J).

Solvent mixture nitric acid R₁, water R (1:99 V/V).

Test solution T: to the residue of the test for sulfated ash add 3 mL of hydrochloric acid R and 1 mL of fuming nitric acid R.

Cover the crucible with a watch glass and heat at 160-170 °C for 1 h to dissolve the residue. Afterwards continue heating in the open crucible and evaporate the solution. Stop heating before the residue is completely dried, add 1 mL of nitric acid R, heat at 160-170 °C for further 10 min, and after

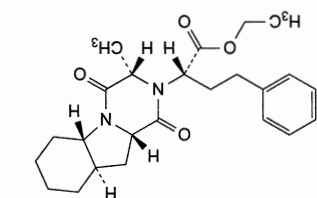
cooling dilute to 10.0 mL with water R.

Reference solutions Prepare reference solutions containing 0.5 µg, 1.0 µg and 1.5 µg of Pd per millilitre by diluting palladium standard solution (500 ppm Pd) R with the solvent mixture.

Source Palladium hollow-cathode lamp.

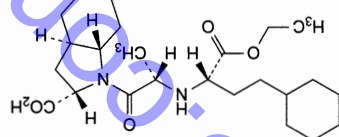
Wavelength 244.8 nm.

Atomisation device Air-acetylene flame.



D, ethyl (2S,2'-(3S,5aS,9aR,10aS)-3-methyl-1,4-dioxodecacyldropropylazino[1,2-a]indol-2(1H)-yl]-4-phenylbutanoate (trandolapril diketopiperazine),

C. (2S,3aR,7aS)-1-[(2S)-2-[(1S)-3-cyclohexyl-1-(ethoxycarbonyl)propyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid (hexahydrotrandolapril),

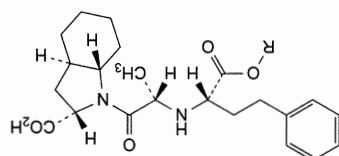


carboxylic acid (trandolaprilate),
 E. R = H: (2S,3aR,7aS)-1-[(2S)-2-[(1S)-1-carboxy-3-phenylpropyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid (isopropyl ester derivative),

[(1-methylethoxy)carbonyl]-3-phenylpropyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid (methyl ester derivative),

B. R = CH(CH₃)₂: (2S,3aR,7aS)-1-[(2S)-2-[(1S)-1-(methoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydro-1H-indole-2-

A. R = CH₃: (2S,3aR,7aS)-1-[(2S)-2-[(1S)-1-



E, F.
 Control of impurities in substances for pharmaceutical use: A, B, impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these by the general monograph Substances for pharmaceutical use acceptance criterion for other/unspecified impurities and/or the tests in the monograph. They are limited by the general present at a sufficient level, be detected by one or other of Other detectable impurities (the following substances would, if Specified impurities C, D

IMPURITIES

Protected from light.

STORAGE

of C₂₄H₃₄N₂O₅.
 1 mL of 0.1 M perchloric acid is equivalent to 43.05 mg

potentiometrically (2.2.20).

Dissolve 0.300 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point

ASSAY

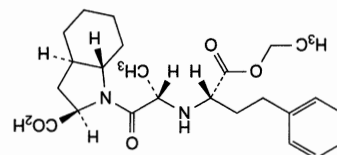
or quartz crucible.

Maximum 0.1 per cent, determined on 2.0 g in a porcelain

Sulfated ash (2.4.14)

Maximum 0.2 per cent, determined on 1.000 g.

Water (2.5.32)

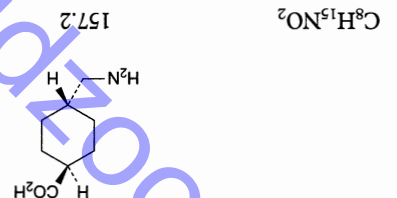


F. (2R,3aR,7aS)-1-[(2S)-2-[[[(1S)-1-(ethoxycarbonyl)-3-phenylpropyl]amino]propanoyl]octahydro-1H-indole-2-carboxylic acid.

Ph Eur

Tranexamic Acid

(Ph. Eur. monograph 0875)



1197-18-8

Action and use

Antifibrinolytic.

Preparations

Tranexamic Acid Injection
Tranexamic Acid Mouthwash
Tranexamic Acid Tablets

Ph Eur

DEFINITION

trans-4-(Aminomethyl)cyclohexanecarboxylic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water and in glacial acetic acid, practically insoluble in acetone and in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison tranexamic acid CRS.

TESTS

pH (2.2.3)

7.0 to 8.0.

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.20 g of the substance to be examined in water R and dilute to 20.0 mL with the same solvent.

Reference solution (a) Dilute 5.0 mL of the test solution to 100.0 mL with water R. Dilute 1.0 mL of this solution to 10.0 mL with water R.

Reference solution (b) Dissolve 20 mg of tranexamic acid CRS (containing impurity C) in water R and dilute to 2 mL with the same solvent.

Reference solution (c) Dissolve 12 mg of 4-aminomethylbenzoic acid R (impurity D) in water R and dilute to 100 mL with the same solvent. Dilute 1 mL of the solution to 50 mL with water R. Dilute 5 mL of this solution to 200 mL with water R.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm or $l = 0.25$ m,

$\phi = 6.0$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Dissolve 11.0 g of anhydrous sodium dihydrogen phosphate R in 500 mL of water R and add 5 mL of triethylamine R and 1.4 g of sodium laurylsulfate R. Adjust to pH 2.5 with dilute phosphoric acid R and dilute to 600 mL with water R. Add 400 mL of methanol R and mix.

Flow rate 0.9 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μ L.

Run time 3 times the retention time of tranexamic acid.

Identification of impurities Use the chromatogram supplied with tranexamic acid CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C; use the chromatogram obtained with reference solution (c) to identify the peak due to impurity D.

Relative retention With reference to tranexamic acid (retention time = about 13 min): impurity C = about 1.1;

impurity D = about 1.3; impurity B = about 1.5;

impurity A = about 2.1.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to tranexamic acid and impurity C.

Limits:

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.2;

impurity C = 0.005; impurity D = 0.006;

impurity A: not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— unspecified impurities: for each impurity, not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— sum of unspecified impurities: not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.025 per cent).

Halides expressed as chlorides (2.4.4)

Maximum 140 ppm.

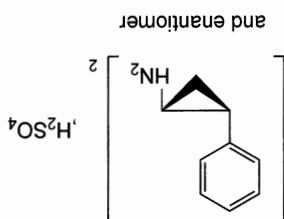
Dissolve 1.2 g in water R and dilute to 50 mL with the same solvent.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of this solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Tranylcypromine Sulfate



(C₉H₁₁N)₂·H₂SO₄ 364.5 13492-01-8

Action and use

Monamine oxidase inhibitor; antidepressant.

Preparation

Tranylcypromine Tablets

DEFINITION

Tranylcypromine Sulfate is (1*RS*,2*SR*)-2-

phenylcyclopropylamine sulfate. It contains not less than 98.0% and not more than 101.0% of (C₉H₁₁N)₂·H₂SO₄, calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white, crystalline powder.

Soluble in water; very slightly soluble in ethanol (96%) and in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of tranylcypromine sulfate (RS 345).

B. Yields the reactions characteristic of sulfates, Appendix VI.

TESTS

Related substances

Carry out the method for gas chromatography, Appendix III B, using the following solutions in solution A. Dissolve 10 mg of 4-chloroaniline (internal standard) in sufficient 0.1M hydrochloric acid to produce 20 mL (solution A).

(1) Add 5 mL of 1M sodium hydroxide to 1 mL of solution A, extract with 10 mL of dichloromethane, add 1 mL of trifuoroacetic anhydride to the dichloromethane extract and allow to stand for 10 minutes. Evaporate the solution at a pressure of 2 kPa using a rotary evaporator and a water bath at 20° and dissolve the residue in 2 mL of dichloromethane. (2) Dissolve 0.1 g of the substance being examined in 5 mL of water, add 1 mL of 5M sodium hydroxide, extract with 10 mL of dichloromethane, add 1 mL of trifuoroacetic anhydride to the dichloromethane extract and allow to stand for 10 minutes. Evaporate the solution at a pressure of 2 kPa using a rotary evaporator and a water bath at 20° and dissolve the residue in 2 mL of dichloromethane. (3) Dissolve 0.1 g of the substance being examined in 5 mL of water, add 1 mL of 5M sodium hydroxide, extract with 10 mL of dichloromethane, add 1 mL of trifuoroacetic anhydride to the dichloromethane extract and allow to stand for 10 minutes. Evaporate the solution at a pressure of 2 kPa using a rotary evaporator and a water bath at 20° and dissolve the residue in 2 mL of dichloromethane.

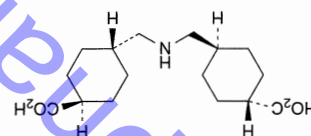
CHROMATOGRAPHIC CONDITIONS

(a) Use a glass column (1.5 m × 4 mm) packed with acid-washed, silanised diatomaceous support (100 to 120 mesh)

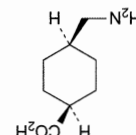
IMPURITIES

Specified impurities A, B

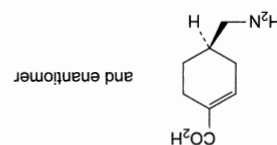
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): C, D.



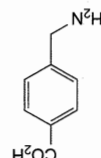
A, trans,trans-4,4'-(iminodimethylene)di(cyclohexanecarboxylic acid),



B, cis-4-(aminomethyl)cyclohexanecarboxylic acid,



C, (RS)-4-(aminomethyl)cyclohex-1-enecarboxylic acid,



D, 4-aminomethylbenzoic acid.

Ph Eur

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.140 g in 20 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 15.72 mg of C₈H₁₅NO₂.

coated with 3% w/w of cyanopropylmethylphenyl methylsilicone fluid (OV-225 is suitable).

(b) Use helium as the carrier gas at 1.7 mL per minute.

(c) Use isothermal conditions maintained at 170°.

LIMITS

In the chromatogram obtained with solution (3), the area of any secondary peak is not greater than that of the peak due to the trifluoroacetyl derivative of 4-chloroaniline (0.5%).

Loss on drying

When dried to constant weight at 105°, loses not more than 0.5% of its weight. Use 1 g.

Sulfated ash

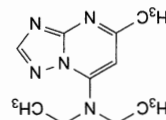
Not more than 0.1%, Appendix IX A.

ASSAY

Carry out Method I for non-aqueous titration, Appendix VIII A, using 0.3 g and determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 36.45 mg of (C₁₅H₁₁N)₂·H₂SO₄.

Trapidil

(Ph. Eur. monograph 1576)



C₁₀H₁₅N₅ 205.3

15421-84-8



Action and use

Antiplatelet agent; vasodilator.

DEFINITION

N₇,N₇-Diethyl-5-methyl-1,2,4-triazolo[1,5-a]pyrimidin-7-amine.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, soluble in ethanol and in methylene chloride.

mp

About 102° C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison trapidil CRS.

TESTS

Solution S

Dissolve 2.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.2 mL of methyl red solution R and 0.2 mL of 0.01 M hydrochloric acid. The solution is red.

Add 0.4 mL of 0.01 M sodium hydroxide. The solution is yellow.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of trapidil impurity A CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of trapidil impurity B CRS in the mobile phase and dilute to 50.0 mL with the mobile phase. Dilute 1.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (c) Mix equal volumes of reference solution (a) and reference solution (b).

Column:

— size: l = 0.125 m, Ø = 4.0 mm,

— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase 50 mL of methanol R, 75 mL of acetonitrile R and 800 mL of a 1.7 g/L solution of potassium dihydrogen phosphate R adjusted to pH 2.45 with phosphoric acid R; dilute to 1000 mL with water R.

Detection Spectrophotometer at 205 nm.

Injection 10 µL.

Run time 3 times the retention time of trapidil.

System suitability:

— resolution: minimum of 4.0 between the peaks due to impurity A and impurity B in the chromatogram obtained with reference solution (c).

Limits:

— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— impurity B: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);

— any other impurity: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— total: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.5 per cent).

— disregard limit: 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.01 per cent).

Chlorides (2.4.4)

Maximum 100 ppm.

Dissolve 0.25 g in 10 mL of water R and dilute to 15 mL with water R. Prepare the standard using 5 mL of chloride standard solution (5 ppm Cl) R.

Ammonium (2.4.1)

Maximum 20 ppm.

0.50 g complies with limit test A. Prepare the standard using 0.1 mL of ammonium standard solution (100 ppm NH₄) R.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in 20 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

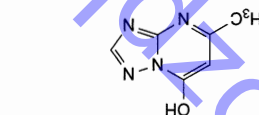
Dissolve 0.180 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 20.53 mg of $C_{10}H_{15}N_5$

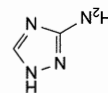
STORAGE

Protected from light.

IMPURITIES

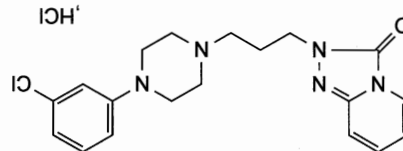


A. 5-methyl-[1,2,4]triazolo[1,5-a]pyrimidin-7-ol,



B. 1,2,4-triazol-3-amine.

Trazodone Hydrochloride



$C_{19}H_{22}ClN_5O \cdot HCl$ 408.3

25332-39-2

Preparations

Trazodone Capsules

Trazodone Tablets

DEFINITION

Trazodone Hydrochloride is 2,3-[4-(3-chlorophenyl)pyridazin-1-yl]propyl-1,2,4-triazolo[4,3-a]pyridin-3(2H)-one hydrochloride. It contains not less than 99.0% and not more than 101.0% of $C_{19}H_{22}ClN_5O \cdot HCl$, calculated with reference to the dried substance.

PRODUCTION

The method of manufacture is such that the level of impurity F, 1-(3-chlorophenyl)pyridazine, is not more than 2.5 ppm when determined by a suitable method.

CHARACTERISTICS

A white or almost white, crystalline powder.

Soluble in water; sparingly soluble in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of trazodone hydrochloride (RS 346). In the preparation of the disc, avoid excessive grinding when triturating the substance being examined with potassium chloride.

B. Yields the reactions characteristic of chlorides, Appendix VI.

TESTS

Acidity
pH of a 1% w/v solution, 3.9 to 4.5, Appendix V L.

Heavy metals

1.0 g complies with limit test C for heavy metals, Appendix VII. Use 2 mL of lead standard solution (10 ppm Pb) to prepare the standard (20 ppm).

3-Chloroaniline

To 10 mL of a 1% w/v solution of the substance being examined in a mixture of equal volumes of water and ethanol add 2 mL of a freshly prepared 5% w/v solution of 4-dimethylaminobenzaldehyde in ethanol and 0.1 mL of hydrochloric acid. Shake well and allow to stand for 5 minutes. Any yellow colour produced at 5 minutes from the preparation of the solution is not more intense than that produced by treating at the same time and in the same manner 10 mL of a 1 µg per mL solution of 3-chloroaniline in a mixture of equal volumes of water and ethanol beginning at the words 'add 2 mL...' (100 ppm).

Related substances

Carry out the procedures protected from light. The combined nominal total content of impurities determined in tests A and B below is not more than 1.0% and no single unknown impurity is more than 0.1%.

A. Carry out the method for liquid chromatography, Appendix III D, using the following solutions in the mobile phase:

(1) 0.1% w/v of the substance being examined.

(2) 0.0001% w/v of the substance being examined.

(3) 0.00005% w/v of the substance being examined.

(4) 0.1% w/v of trazodone hydrochloride impurity standard BPCRS.

CHROMATOGRAPHIC CONDITIONS

(a) Use a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Waters XTerra RP18 or Phenomenex Prodigy are suitable).
(b) Use isocratic elution and the mobile phase described below.

(c) Use a flow rate of 2.0 mL per minute.

(d) Use a column temperature of 40°.

(e) Use a detection wavelength of 254 nm.

(f) Inject 20 µL of each solution.

(g) For solution (1), allow the chromatography to proceed for 3 times the retention time of the principal peak.

MOBILE PHASE

0.4 volumes of diethylamine, 350 volumes of acetonitrile and 650 volumes of water. If necessary, adjust the proportions of acetonitrile and water in the mobile phase to obtain a retention time of about 10 minutes for the principal peak.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with solution (4) closely resembles reference chromatogram A supplied with trazodone hydrochloride impurity standard BPCRS.

LIMITS

In the chromatogram obtained with solution (1):

the area of any peak corresponding to impurity D, identified from reference chromatogram A supplied with trazodone hydrochloride impurity standard BPCRS, is not more than 3 times the area of the principal peak in the chromatogram obtained with solution (2) (0.3%);

the area of any other secondary peak with a retention time of less than or equal to impurity E, identified from reference chromatogram A supplied with trazodone hydrochloride impurity standard BPCRS, is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (0.1%).

Disregard any peak with an area less than that of the principal peak in the chromatogram obtained with solution (3) (0.05%).

B. Carry out the method for liquid chromatography, Appendix III D, using the following solutions in the mobile phase.

- (1) 0.1% w/v of the substance being examined.
- (2) 0.0001% w/v of the substance being examined.
- (3) 0.00005% w/v of the substance being examined.
- (4) 0.1% w/v of trazodone hydrochloride impurity standard BPCRS.

CHROMATOGRAPHIC CONDITIONS

- (a) Use a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Waters XTerra RP18 or Phenomenex Prodigy are suitable).
- (b) Use isocratic elution and the mobile phase described below.
- (c) Use a flow rate of 1.7 mL per minute.
- (d) Use a column temperature of 40°.
- (e) Use a detection wavelength of 254 nm.
- (f) Inject 20 µL of each solution.

For at least 5 times the retention time of the principal peak. (g) For solution (1), allow the chromatography to proceed for at least 5 times the retention time of the principal peak. Obtain a retention time of about 2.5 minutes for the principal peak. 0.4 volumes of diethylamine, 320 volumes of water and 680 volumes of acetonitrile. If necessary, adjust the proportions of acetonitrile and water in the mobile phase to obtain a retention time of about 2.5 minutes for the principal peak.

SYSTEM SUITABILITY

The test is not valid unless the chromatogram obtained with solution (4) closely resembles reference chromatogram B supplied with trazodone hydrochloride impurity standard BPCRS.

LIMITS

In the chromatogram obtained with solution (1):

the area of any secondary peak with a retention time longer than that of impurity E identified from reference chromatogram B, supplied with trazodone hydrochloride impurity standard BPCRS, is not greater than the area of the principal peak obtained with solution (2) (0.1%).

Disregard any peak with an area less than that of the principal peak in the chromatogram obtained with solution (3) (0.05%).

Loss on drying

When dried to constant weight at 105° at a pressure of 3.5 to 6.5 kPa, loses not more than 0.5% of its weight. Use 1 g.

Not more than 0.2%, Appendix IX A.

ASSAY

Carry out the procedure protected from light.

Appendix III D, using the following solutions in the mobile phase.

- (1) 0.01% w/v of the substance being examined.
- (2) 0.01% w/v of trazodone hydrochloride BPCRS.

CHROMATOGRAPHIC CONDITIONS

- (a) Use a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Spherisorb C8 is suitable).
- (b) Use isocratic elution and the mobile phase described below.
- (c) Use a flow rate of 2 mL per minute.
- (d) Use a column temperature of 40°.
- (e) Use a detection wavelength of 254 nm.
- (f) Inject 20 µL of each solution.

MOBILE PHASE

40 volumes of a 0.115% w/v solution of diammonium hydrogen orthophosphate, previously adjusted to pH 6.0 with 10% v/v of orthophosphoric acid or 1M sodium hydroxide, and 60 volumes of methanol.

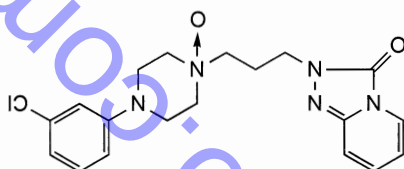
DETERMINATION OF CONTENT

Calculate the content of C₁₉H₂₂ClN₅O₂HCl in the substance being examined using the declared content of C₁₉H₂₂ClN₅O₂HCl in trazodone hydrochloride BPCRS.

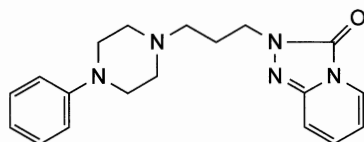
STORAGE

Trazodone Hydrochloride should be kept in an airtight container and protected from light.

IMPURITIES



A. 4-(3-chlorophenyl)-1-[3-(3-oxo-2,3-dihydro-1,2,4-triazolo[4,3-a]pyridin-2-yl)propyl]piperazine N'-oxide

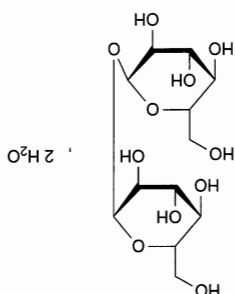


B. 2-[3-(4-phenylpiperazin-1-yl)propyl]-1,2,4-triazolo[4,3-a]pyridin-3(2H)-one



Trehalose Dihydrate

(Ph. Eur. monograph 2297)



$C_{12}H_{22}O_{11} \cdot 2H_2O$ 378.3 6138-23-4 Ph. Eur.

DEFINITION

α -D-Glucopyranosyl α -D-glucopyranoside dihydrate (α , α -trehalose dihydrate). It is obtained by enzymatic modification of starch.

Content

97.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, slightly soluble in methanol, practically insoluble in ethanol (96 per cent).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison trehalose dihydrate CRS.

B. Dissolve 2 g in 5 mL of water R. To 1 mL of this solution add 0.4 mL of a 50 g/L solution of α -naphthol R in ethanol (96 per cent) R and mix thoroughly. Carefully add 2 mL of sulfonic acid R. A violet colour develops at the interface.

C. Dissolve 1 g in 25 mL of water R. To 2 mL of this solution add 1 mL of dilute hydrochloric acid R and mix. Keep the solution for 20 min at room temperature. Add 4 mL of a 40 g/L solution of sodium hydroxide R and 2 mL of a 40 g/L solution of glycine R and mix. Heat the solution in a water-bath for 10 min. No brown colour develops.

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.5 to 6.5 for solution S.

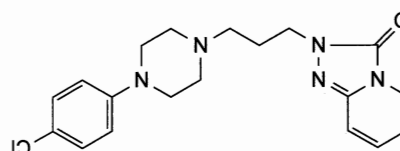
Specific optical rotation (2.2.7)

+ 197 to + 201 (anhydrous substance), determined on solution S.

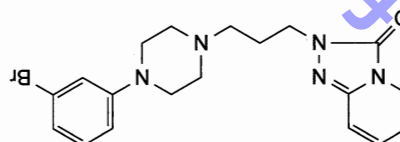
Related substances

Liquid chromatography (2.2.29).

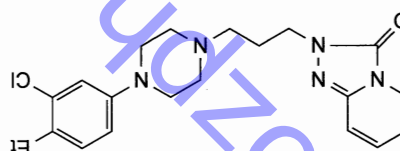
Test solution Dissolve 0.100 g of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.



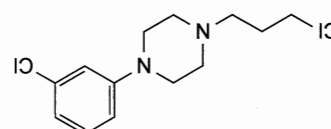
C. 2-3-[4-(4-chlorophenyl)piperazin-1-yl]propyl-1,2,4-triazolo[4,3-a]pyridin-3(2H)-one



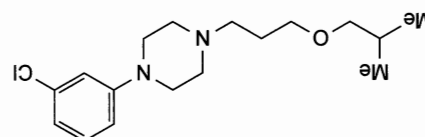
D. 2-3-[4-(3-bromophenyl)piperazin-1-yl]propyl-1,2,4-triazolo[4,3-a]pyridin-3(2H)-one



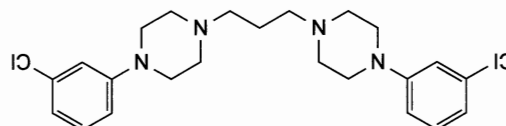
E. 2-3-[4-(3-chloro-4-ethylphenyl)piperazin-1-yl]propyl-1,2,4-triazolo[4,3-a]pyridin-3(2H)-one



F. 1-(3-chloropropyl)-3-chlorophenylpiperazine



G. 3-[4-(3-chlorophenyl)piperazin-1-yl]propyl isobutyl ether



H. 1,3-bis-[4-(3-chlorophenyl)piperazin-1-yl]propane

Reference solution (a) Dissolve 0.100 g of trehalose dihydrate CRS in water R and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with water R.

Reference solution (c) Dilute 5.0 mL of reference solution (b) to 25.0 mL with water R.

Reference solution (d) Dissolve 25 mg of glucose R (impurity A) and 25 mg of maltotriose R in water R, add 2.5 mL of reference solution (a) and dilute to 10.0 mL with water R.

Column:

- size: $l = 0.3$ m, $\varnothing = 8$ mm;
- stationary phase: strong cation-exchange resin (sodium form) R (6 μ m);
- temperature: 80 °C.

Mobile phase water R.

Flow rate 0.4 mL/min.

Detection Refractometer maintained at 40 °C.

Injection 20 μ L of the test solution and reference solutions (b), (c) and (d).

Run time Twice the retention time of trehalose.

Identification of impurities Use the chromatogram obtained with reference solution (d) to identify the peaks due to impurities A and B; impurity B has the same retention time as maltotriose.

Relative retention With reference to trehalose (retention time = about 15 min): impurity B = about 0.9; impurity A = about 1.2.

System suitability Reference solution (d):

- resolution: minimum 1.5 between the peaks due to maltotriose and trehalose.

Limits:

- impurities A, B: for each impurity, not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)
- unspecified impurities: for each impurity, not more than area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent);
- total: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);
- disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.1 per cent).

Chlorides (2.4.4)
Maximum 125 ppm.
Dilute 4 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)
Maximum 200 ppm.
Dilute 7.5 mL of solution S to 15 mL with distilled water R.

Heavy metals (2.4.8)
Maximum 5 ppm.
Dissolve 4.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Soluble starch
Dissolve 1 g in 10 mL of water R. Add 0.1 mL of iodine solution R1. No blue colour develops.

Water (2.5.12)
9.0 per cent to 11.0 per cent, determined on 0.10 g.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

Microbial contamination

If intended for use in the manufacture of parenteral preparations:

- TAMC: acceptance criterion 10^2 CFU/g (2.6.12).

If not intended for use in the manufacture of parenteral preparations:

- TAMC: acceptance criterion 10^3 CFU/g (2.6.12);
- TYMC: acceptance criterion 10^2 CFU/g (2.6.12);
- absence of *Escherichia coli* (2.6.13);
- absence of *Salmoneilla* (2.6.13).

Bacterial endotoxins (2.6.14)

If intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins:

- less than 4 IU/g for parenteral preparations having a concentration of 100 g/L or less of trehalose dihydrate;
- less than 2.5 IU/g for parenteral preparations having a concentration of more than 100 g/L of trehalose dihydrate.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of trehalose from the declared content of trehalose dihydrate CRS.

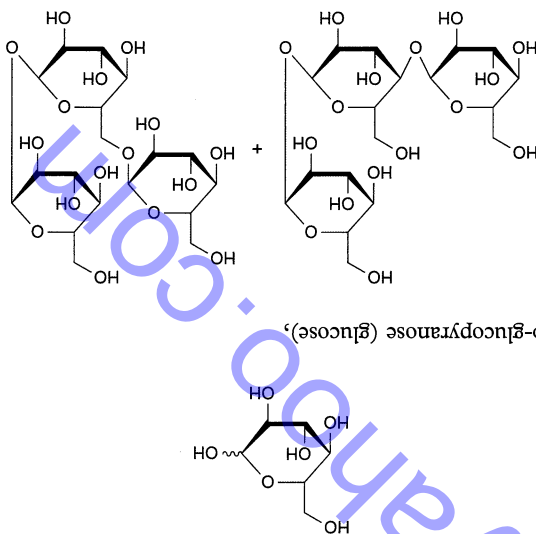
LABELLING

The label states:

- where applicable, the maximum concentration of bacterial endotoxins;
- where applicable, that the substance is suitable for use in the manufacture of parenteral preparations.

IMPURITIES

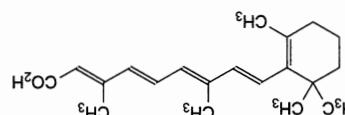
Specified impurities A, B



B, oligosaccharides, mainly glucosyltrehalose: mixture of α -D-glucopyranosyl α -D-glucopyranosyl-(1 \rightarrow 4)- α -D-glucopyranoside (4-O-glucosyltrehalose or α -D-maltosyl α -D-glucoside) and α -D-glucopyranosyl-(1 \rightarrow 6)- α -D-glucopyranosyl α -D-glucopyranoside (6-O-glucosyltrehalose or α -D-isomaltosyl α -D-glucoside).

Tretinoin

(Ph. Eur. monograph 0693)



$C_{20}H_{28}O_2$ 300.4 302-79-4

Action and use

Vitamin A analogue (retinoid); treatment of acne.

Preparation

Tretinoin Gel

DEFINITION

(2E,4E,6E,8E)-3,7-Dimethyl-9-(2,6,6-trimethylcyclohex-1-enyl)nona-2,4,6,8-tetraenoic acid.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow or light orange, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in methylene

chloride, slightly soluble in ethanol (96 per cent).

mp

About 182 °C, with decomposition.

It is sensitive to air, heat and light, especially in solution.

Carry out all operations as rapidly as possible and avoid exposure

to actinic light; use freshly prepared solutions.

IDENTIFICATION

First identification A

Second identification B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison tretinoin CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined

in methylene chloride R and dilute to 10 mL with the same

solvent.

Reference solution Dissolve 10 mg of tretinoin CRS in methylene

chloride R and dilute to 10 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase glacial acetic acid R, acetone R, peroxide-free

ether R, cyclohexane R (2:4:40:54 V/V/V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position and size to the principal

spot in the chromatogram obtained with the reference

solution.

C. Dissolve about 5 mg in 2 mL of antimony trichloride

solution.

Antimony trichloride R. An intense red colour develops and later becomes

violet.

TESTS

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be

examined in methanol R and dilute to 50.0 mL with the same

solvent.

Reference solution (a) Dissolve 10.0 mg of isotretinoin CRS

(impurity A) in methanol R and dilute to 10.0 mL with the

same solvent.

Reference solution (b) Mix 1.0 mL of reference solution (a)

and 0.5 mL of the test solution and dilute to 25.0 mL with

methanol R.

Reference solution (c) Dilute 0.5 mL of the test solution to

100.0 mL with methanol R.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R

(3 µm).

Mobile phase glacial acetic acid R, water R, methanol R

(5:225:770 V/V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 355 nm.

Injection 10 µL.

Run time 1.2 times the retention time of tretinoin.

Identification of impurities Use the chromatogram obtained

with reference solution (a) to identify the peak due to

impurity A.

Relative retention With reference to tretinoin (retention

time = about 29 min): impurity A = about 0.75.

System suitability: reference solution (b):

— resolution: minimum 5.0 between the peaks due to

impurity A and tretinoin.

Limits:

— impurity A: not more than the area of the principal peak

in the chromatogram obtained with reference solution (c)

(0.5 per cent);

— unspecified impurities: for each impurity, not more than

0.4 times the area of the principal peak in the

chromatogram obtained with reference solution (c)

(0.2 per cent);

— total: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (c)

(1.0 per cent);

— disregard limit: 0.1 times the area of the principal peak in

the chromatogram obtained with reference solution (c)

The thresholds indicated under Related substances

(Table 2034-1) in the general monograph Substances for

Pharmaceutical use (2034) do not apply.

Heavy metals (2.4.8)

Maximum 20 ppm.

0.5 g complies with test D. Prepare the reference solution

using 1 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying

in vacuo for 16 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 70 mL of acetone R. Titrate with 0.1 M

tetrabutylammonium hydroxide in 2-propanol, determining the

end-point potentiometrically (2.2.20).

1 mL of 0.1 M tetrabutylammonium hydroxide in 2-propanol is equivalent to 30.04 mg of C₂₀H₃₈O₂.

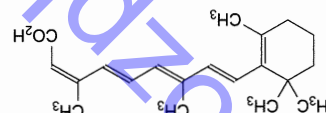
STORAGE

Under an inert gas, in an airtight container, protected from light.

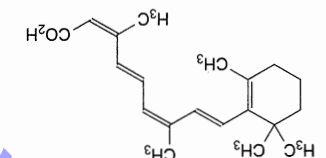
It is recommended that the contents of an opened container be used as soon as possible and any unused part be protected by an atmosphere of inert gas.

IMPURITIES

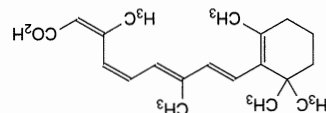
Specified impurities A
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, F, G.



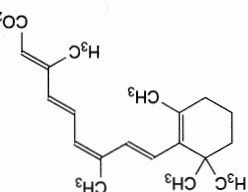
A. (2Z,4E,6E,8E)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-enyl)nona-2,4,6,8-tetraenoic acid (isoretrenoin).



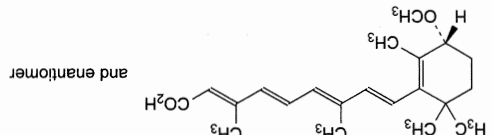
B. (2Z,4E,6Z,8E)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-enyl)nona-2,4,6,8-tetraenoic acid (9,13-di-cis-retinoic acid).



C. (2Z,4Z,6E,8E)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-enyl)nona-2,4,6,8-tetraenoic acid (11,13-di-cis-retinoic acid).



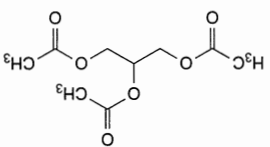
D. (2E,4E,6Z,8E)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-enyl)nona-2,4,6,8-tetraenoic acid (9-cis-retinoic acid).



F. (2E,4E,6E,8E)-9-[(3R,5)-3-methoxy-2,6,6-trimethylcyclohex-1-enyl]-3,7-dimethylnona-2,4,6,8-tetraenoic acid (rac-4-methoxyretrenoin).

Triacetin

(Ph. Eur. monograph 1106)



C₉H₁₄O₆

218.2

102-76-1

Action and use
Antifungal.

Ph Eur

DEFINITION

Propane-1,2,3-triyl triacetate.

Content

97.0 per cent to 100.5 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, colourless, slightly viscous oily liquid.

Solubility

Soluble in water, miscible with ethanol (96 per cent) and toluene.

bp

about 260 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of triacetin.

TESTS

Appearance

It is clear (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Acidity

Dissolve 5.00 g in 25 mL of anhydrous ethanol R, previously neutralised to 0.2 mL of phenolphthalein solution R and add 0.20 mL of 0.1 M sodium hydroxide. The pink colour of the mixture persists for 15 s.

Relative density (2.2.5)

1.159 to 1.164.

Refractive index (2.2.6)

1.429 to 1.432.

Water (2.5.12)

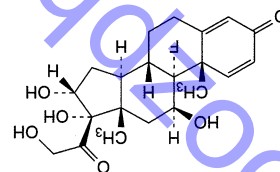
Maximum 0.2 per cent, determined on 5.00 g.

ASSAY

Introduce 0.300 g into a 250 mL borosilicate glass flask fitted with a reflux condenser. Add 25.0 mL of 0.5 M alcoholic potassium hydroxide and a few glass beads. Attach the

Triamcinolone

(Ph. Eur. monograph 1376)



$C_{21}H_{27}FO_6$ 394.4 124-94-7

Action and use

Glucocorticoid.

Preparation

Triamcinolone Tablets

Ph Eur

DEFINITION

9-Fluoro-11 β ,17,21-tetrahydroxyprogna-1,4-diene-3,20-

dione.

Content

97.0 per cent to 103.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, slightly soluble in methanol,

practically insoluble in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison triamcinolone CRS.

If the spectra obtained show differences, dissolve the

substance to be examined and the reference substance

separately in methanol R, evaporate to dryness, dry the

residues at 60 °C at a pressure not exceeding 0.7 kPa and

record new spectra using the residues.

B. Thin-layer chromatography (2.2.27). Prepare the solutions

immediately before use and protect from light. Examine the plate

under ultraviolet light immediately after development.

Test solution Dissolve 10 mg of the substance to be examined

in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of triamcinolone CRS in

methanol R and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of dexamethasone CRS in

solution.

Specific optical rotation (2.2.7)
+ 65 to + 72 (anhydrous substance).
Dissolve 0.100 g in dimethylformamide R and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions

immediately before use and protect from light.

Test solution Dissolve 25.0 mg of the substance to be

examined in a mixture of equal volumes of methanol R and

water R and dilute to 10.0 mL with the same mixture of

solvents.

Reference solution (a) Dissolve 2 mg of triamcinolone CRS and

2 mg of triamcinolone impurity C CRS in a mixture of equal

volumes of methanol R and water R and dilute to 100.0 mL

with the same mixture of solvents.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with a mixture of equal volumes of methanol R and

water R.

Blank methanol R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: base-deactivated end-capped octadecylsilyl

silica gel for chromatography R (5 μ m).

Mobile phase A mixture prepared as follows: in a 1000 mL

volumetric flask mix 525 mL of methanol R with 400 mL of

water R and allow to equilibrate; adjust the volume to

1000 mL with water R and mix again.

Detection Spectrophotometer set at 238 nm.

Injection 20 μ L.

Run time 4.5 times the retention time of triamcinolone.

Retention time Triamcinolone = about 11 min.

System suitability: reference solution (a):

— resolution: minimum of 1.8 between the peaks due to

triamcinolone and to impurity C.

any impurity: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(1 per cent) and not more than 2 such peaks have an area

greater than half the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.5 per cent),

total: not more than twice the area of the principal peak in

the chromatogram obtained with reference solution (b)

(2 per cent).

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent).

Water (2.5.12)

Maximum 1.0 per cent, determined on 0.500 g.

ASSAY

Prepare the solutions immediately before use and protect from light.

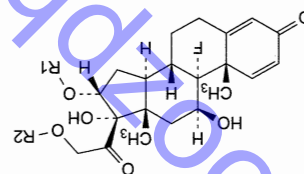
Dissolve 50.0 mg in alcohol R and dilute to 50.0 mL with the same solvent. Dilute 2.0 mL of the solution to 100.0 mL with alcohol R. Measure the absorbance (2.2.25) at the maximum at 238 nm.

Calculate the content of $C_{21}H_{27}FO_6$ taking the specific absorbance to be 389.

STORAGE

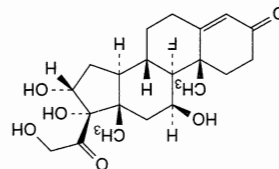
Protected from light.

IMPURITIES



A, R1 = R2 = CO-CH₃: 9-fluoro-11β,17-dihydroxy-3,20-dioxopregna-1,4-diene-16α,21-diyl diacetate (triamcinolone 16,21-diacetate),

B, R1 = H, R2 = CO-CH₃: 9-fluoro-11β,16α,17-trihydroxy-3,20-dioxopregna-1,4-dien-21-yl acetate (triamcinolone 21-acetate),

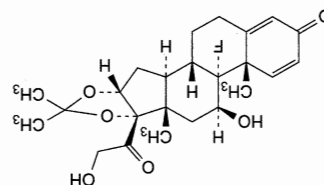


C, 9-fluoro-11β,16α,17,21-tetrahydroxypregna-4-ene-3,20-dione (pregnatriamcinolone).

Ph Eur

Triamcinolone Acetonide

(Ph. Eur. monograph 0533)



$C_{24}H_{31}FO_6$

434.5

76-25-5

Action and use

Glucocorticoid.

Preparations

Triamcinolone Cream
Triamcinolone Acetonide Injection
Triamcinolone Acetonide Nasal Spray
Triamcinolone Ointment
Triamcinolone Dental Paste

DEFINITION
9-Fluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione.

Content
97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in ethanol (96 per cent).

It shows polymorphism (5.9).

IDENTIFICATION

First identification A, C

Second identification B, D

A. Infrared absorption spectrophotometry (2.2.24).

Comparison triamcinolone acetonide CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of methanol R

and evaporate to dryness. Using the residues, prepare halogen salt discs or mulls in liquid paraffin R and record new spectra.

B. Thin-layer chromatography (2.2.27). Prepare the solutions immediately before use and protect from light.

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of triamcinolone acetonide CRS in methanol R and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of triamcinolone hexacetate CRS in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and 8 volumes of methanol R to a mixture of 15 volumes of ether R and 77 volumes of methylene chloride R.

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm, immediately after development.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Examine the chromatograms obtained in the assay.

Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (c).

D. Mix about 5 mg with 45 mg of heavy magnesium oxide R and ignite in a crucible until an almost white residue is obtained (usually less than 5 min). Allow to cool, add 1 mL of water R, 0.05 mL of phenolphthalein solution R1 and about 1 mL of dilute hydrochloric acid R to render the solution colourless. Filter. To a freshly prepared mixture of 0.1 mL of

— *unspecified impurities*: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— *total*: not more than 5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.5 per cent);

— *disregard limit*: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Water (2.5.12)
Maximum 2.0 per cent, determined on 0.500 g.

ASSAY

Carry out the assay protected from light.

Liquid chromatography (2.2.29) as described in the test for related substances with the following modifications.

Mobile phase Mobile phase A.

Injection Test solution and reference solution (c).

Run time 1.5 times the retention time of triamcinolone acetonide.

Retention time Triamcinolone acetonide = about 16 min.

Calculate the percentage content of $C_{24}H_{31}FO_6$ taking into account the assigned content of triamcinolone acetonide CRS.

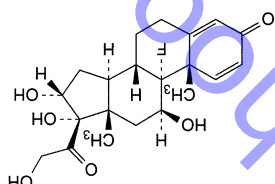
STORAGE

Protected from light.

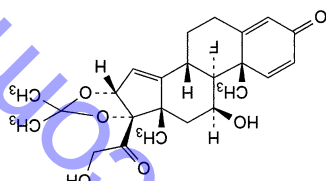
IMPURITIES

Specified impurities B, C

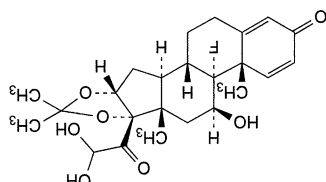
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, D, E, F.



A. 9-fluoro-11β,17,21-tetrahydroxypregna-1,4-diene-3,20-dione (triamcinolone).



B. 9-fluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4,14-triene-3,20-dione (Δ14-triamcinolone acetonide).



C. 9-fluoro-11β,21,21-trihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione (triamcinolone acetonide 21-aldehyde hydrate).

add 1.0 mL of the filtrate. Mix, allow to stand for 5 min and compare the colour of the solution to that of a blank prepared in the same manner. The test solution is yellow and the blank is red.

TESTS

Specific optical rotation (2.2.7)

+ 110 to + 117 (anhydrous substance).
Dissolve 0.100 g in ethanol (96 per cent) R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Test solution Dissolve 25.0 mg of the substance to be examined in mobile phase B and dilute to 25.0 mL with mobile phase B.

Reference solution (a) Dissolve 5 mg of triamcinolone acetonide CRS (containing impurities B and C) in mobile phase B and dilute to 5.0 mL with mobile phase B.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase B. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase B.

Reference solution (c) Dissolve 25.0 mg of triamcinolone acetonide CRS in mobile phase B and dilute to 25.0 mL with mobile phase B.

Column: size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

temperature: 40 °C.

Mobile phase:

— mobile phase A: acetonitrile R, water for chromatography R (32:68 V/V);

— mobile phase B: water for chromatography R, acetonitrile R (35:65 V/V);

Detection Spectrophotometer at 254 nm.

Injection 20 μ L of the test solution and reference solutions (a) and (b).

Identification of impurities Use the chromatogram supplied with triamcinolone acetonide for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and C.

Relative retention With reference to triamcinolone acetonide (retention time = about 16 min): impurity C = about 0.7;

impurity B = about 0.8.

System suitability: reference solution (a):

— *resolution*: minimum 2.5 between the peaks due to impurities C and B.

Limits:

— *impurity* B: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);

— *impurity* C: not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.15 per cent);

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, sparingly soluble in anhydrous ethanol and in methanol.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison triamcinolone hexacetonide CRS.

B. Thin-layer chromatography (2.2.27). Prepare the solutions immediately before use and protect from light.

Test solution Dissolve 10 mg of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of triamcinolone hexacetonide CRS in methanol R and dilute to 20 mL with the same solvent.

Reference solution (b) Dissolve 10 mg of triamcinolone hexacetonide CRS in reference solution (a) and dilute to 10 mL with reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase Add a mixture of 1.2 volumes of water R and 8 volumes of methanol R to a mixture of 15 volumes of either R and 77 volumes of methylene chloride R.

Application 5 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm, immediately after development.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Specific optical rotation (2.2.7)

+ 92 to + 98 (anhydrous substance).
Dissolve 0.100 g in methylene chloride R and dilute to 10.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light.

Test solution Dissolve 25.0 mg of the substance to be examined in methanol R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve 2 mg of triamcinolone hexacetonide CRS and 2 mg of triamcinolone acetate CRS (impurity A) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column: 100.0 mL with the mobile phase.

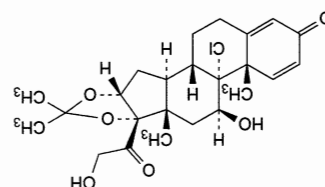
Size: 1 = 0.25 m, Ø = 4.6 mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

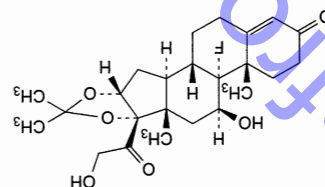
Mobile phase In a 1000 mL volumetric flask mix 750 mL of methanol R with 200 mL of water R and allow to equilibrate; dilute to 1000 mL with water R and mix again.

Flow rate 2 mL/min.

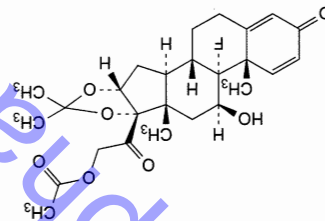
Detection Spectrophotometer at 254 nm.



D. 9-chloro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione (9α-chloro triamcinolone acetate).



E. 9-fluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-4-ene-3,20-dione (1,2-dihydrotriamcinolone acetate).

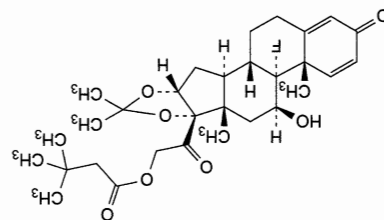


F. 9-fluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3,20-dioxopregna-1,4-dien-2-yl acetate (21-acetate triamcinolone acetate).



Triamcinolone Hexacetonide

(Ph. Eur. monograph 0867)



C₃₀H₄₁FO₇

532.6

5611-51-8

Action and use

Glucocorticoid.

Preparation

Triamcinolone Hexacetonide Injection

Ph. Eur.

DEFINITION

9-fluoro-11β-hydroxy-16α,17-(1-methylethylidenedioxy)-3,20-dioxopregna-1,4-diene-2-yl 3,3-dimethylbutanoate.

Content

97.0 per cent to 103.0 per cent (anhydrous substance).

Equilibration With the mobile phase for about 10 min.

Injection 20 µL.

Run time 3 times the retention time of triamcinolone

hexacetonide.

Retention time Impurity A = about 3 min; triamcinolone

hexacetonide = about 12 min.

System suitability: reference solution (a):

— *resolution*: minimum 20.0 between the peaks due to

impurity A and triamcinolone hexacetonide; if necessary,

adjust the concentration of methanol in the mobile phase.

Limits:

— *impurity A*: not more than 0.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.5 per cent);

— *total*: not more than the area of the principal peak in the

chromatogram obtained with reference solution (b)

(1 per cent);

— the chromatogram obtained with reference solution (b)

(0.05 per cent).

Water (2.5.12)

Maximum 2.0 per cent, determined on 0.50 g.

ASSAY

Dissolve 50.0 mg in ethanol (96 per cent) R and dilute to

50.0 mL with the same solvent. Dilute 2.0 mL of this

solution to 100.0 mL with ethanol (96 per cent) R. Measure

the absorbance (2.2.25) at the absorption maximum at

238 nm.

Calculate the content of $C_{30}H_{41}FO_7$ taking the specific

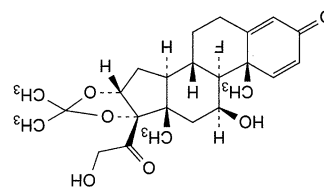
absorbance to be 291.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A

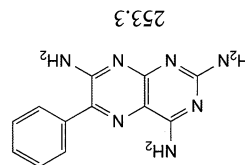


A. 9-fluoro-11β,21-dihydroxy-16α,17-(1-methylethylidenedioxy)pregna-1,4-diene-3,20-dione (triamcinolone acetonide).

Ph Eur

Triamterene

(Ph. Eur. monograph 0058)



396-01-0

$C_{12}H_{11}N_7$

Action and use

Sodium channel blocker; potassium-sparing diuretic.

Preparations

Co-triamterezide Tablets

Triamterene Capsules

DEFINITION

6-Phenylpyridine-2,4,7-triamine.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

Yellow, crystalline powder.

Solubility

Very slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison triamterene CRS.

TESTS

Acidity

Boil 1.0 g with 20 mL of water R for 5 min, cool, filter and

wash the filter with 3 quantities, each of 10 mL, of water R.

Combine the filtrate and washings and add 0.3 mL of

phenolphthalein solution R. Not more than 1.5 mL of 0.01 M

sodium hydroxide is required to change the colour of the

indicator.

Impurity D

Gas chromatography (2.2.28).

Internal standard solution. Dilute 0.1 mL of nitrobenzene R to

100 mL with methanol R. Dilute 1 mL of this solution to

50 mL with methanol R.

Test solution Introduce 0.800 g of the substance to be

examined into a suitable vial, add 5 mL of dimethyl

sulfoxide R and heat until the sample is dissolved (do not heat

to boiling). Allow to cool. Add 5 mL of cold methanol R to

enhance the precipitation of triamterene. Filter and wash the

filter with 5 mL of methanol R. Combine the filtrate and

washings, add 2.0 mL of the internal standard solution and

dilute to 20.0 mL with methanol R.

Reference solution Dissolve 20.0 mg of benzyl cyanide R

(impurity D) in methanol R and dilute to 100.0 mL with the

same solvent. Dilute 5.0 mL of the solution to 50.0 mL with

methanol R. To 2.0 mL of this solution add 2.0 mL of the

internal standard solution and 5 mL of dimethyl sulfoxide R

and dilute to 20.0 mL with methanol R.

Blank solution Dilute 5 mL of dimethyl sulfoxide R to 20 mL

with methanol R.

Column:

— *material*: fused silica;

— *size*: $l = 30$ m, $\varnothing = 0.25$ mm;

— *stationary phase*: macrogol 20 000 R (0.5 µm).

Carrier gas helium for chromatography R.

Flow rate 1.5 mL/min.

Split ratio 1:15.

Temperature:

— *column*: 170 °C;

— *injection port*: 210 °C;

— *detector*: 230 °C.

Detection Flame ionisation.

Injection 1 µL.

Run time Twice the retention time of the internal standard.

Relative retention With reference to the internal standard

(retention time = about 6 min): impurity D = about 1.6.

System suitability: reference solution:

— *resolution*: minimum 2.0 between the peak due to

impurity D and the nearest peak due to the solvent (blank

— *signal-to-noise ratio*: minimum 10 for the peak due to impurity D.

Limit:

— *impurity D*: calculate the ratio (*R*_i) of the area of the peak due to impurity D to the area of the peak due to the internal standard from the chromatogram obtained with the reference solution; from the chromatogram obtained with the test solution, calculate the ratio of the area of the peak due to impurity D to the area of the peak due to the internal standard; this ratio is not greater than *R* (50 ppm).

Related substances

Liquid chromatography (2.2.29).

Test solution: Dissolve 10.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a): Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b): Dissolve 5.0 mg of nitrosotriamteropyrimidine CRS (impurity A) in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (c): Dissolve the contents of a vial of triamterene impurity B CRS in 200 µL of dimethyl sulfoxide *R*. Add 5.0 mL of the test solution and dilute to 50.0 mL with the mobile phase. Filter the solution through a membrane filter (nominal pore size 0.45 µm) before injection.

Column:
— *stationary phase*: spherical end-capped octylsilyl silica gel for chromatography *R* (5 µm).
— *size*: $l = 0.25$ m, $\phi = 4.0$ mm;
— *mobile phase*: butylamine *R*, acetonitrile *R*, methanol *R*, water *R* (2:200:200:600 V/V/V/V), adjusted to pH 5.3 with acetic acid *R*.

Flow rate 1 mL/min.

Detection Spectrophotometer at 320 nm and at 355 nm.

Injection 50 µL.

Relative retention With reference to triamterene (retention time = about 5 min): impurity A = about 0.6;

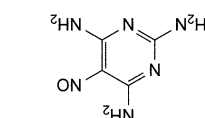
impurity B = about 0.8; impurity C = about 1.7.

System suitability:

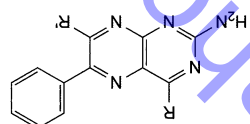
— *resolution*: minimum 1.5 between the peaks due to impurity B and triamterene in the chromatogram obtained with reference solution (c) at 355 nm; if necessary, increase the quantity of water *R* in the mobile phase;
— *signal-to-noise ratio*: minimum 10 for the principal peak in the chromatogram obtained with reference solution (b) at 320 nm.

Limits:

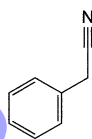
— *correction factors*: for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 1.8; impurity C = 1.5;
— *impurity A* at 320 nm: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (b) (50 ppm);
— *impurities B, C* at 355 nm: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent).



A. 5-nitrosotriamteropyrimidine-2,4,6-triamine (nitrosotriamteropyrimidine),



B. *R* = OH, *R'* = NH₂; 2,7-diamino-6-phenylpyrimidin-4-ol,
C. *R* = NH₂, *R'* = OH; 2,4-diamino-6-phenylpyrimidin-7-ol,



D. phenylacetone nitrile (benzyl cyanide).

IMPURITIES

Protected from light.

STORAGE

1 mL of 0.1 M perchloric acid is equivalent to 25.33 mg of C₁₂H₁₁N₇.

(2.2.20).
perchloric acid, determining the end-point potentiometrically

100 mL of anhydrous acetic acid *R*. Titrate with 0.1 M Dissolve 0.150 g in 5 mL of anhydrous formic acid *R* and add

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in

reference solution (a) (0.05 per cent).

principal peak in the chromatogram obtained with

— *disregard limit* at 355 nm: 0.5 times the area of the

reference solution (a) (0.2 per cent);

principal peak in the chromatogram obtained with

— *total* at 355 nm: not more than twice the area of the

(0.10 per cent);

chromatogram obtained with reference solution (a)

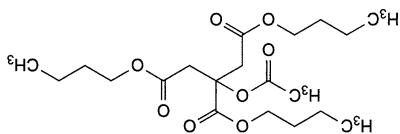
more than the area of the principal peak in the

— *unspecified impurities* at 355 nm: for each impurity, not



Tributyl Acetylacrylate

(Ph. Eur. monograph 1770)



$C_{20}H_{34}O_8$

402.5

77-90-7

Action and use

Excipient.

DEFINITION

Tributyl 2-(acetyloxy)propane-1,2,3-tricarboxylate.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, oily liquid.

Solubility

Not miscible with water, miscible with ethanol (96 per cent) and with methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Preparation Thin films between 2 sodium chloride plates.

Comparison tributyl acetylacrylate CRS.

TESTS

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2).

(Method II).

Acidity

Dilute 10 g with 10 mL of previously neutralised ethanol (96 per cent) R and add 0.5 mL of bromothymol blue solution R₂. Not more than 0.3 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to blue.

Refractive index (2.2.6)

1.442 to 1.445.

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 0.5 g of the substance to be examined in methylene chloride R and dilute to 20 mL with the same solvent.

Reference solution (a) Dissolve 50 mg of tributyl citrate R (impurity A) in methylene chloride R and dilute to 20 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with methylene chloride R. Dilute 1.0 mL of this

solution to 25.0 mL with methylene chloride R.

Reference solution (c) Dissolve the contents of a vial of tributyl acetylacrylate for peak identification CRS (containing impurities B and C) in 1 mL of methylene chloride R.

Column:

material: fused silica;

size: $l = 30$ m, $\varnothing = 0.25$ mm;

stationary phase:

poly[(cyanopropyl)(methyl)[(phenyl)(methyl)]siloxane R (film thickness 0.25 μ m).

STORAGE

Under nitrogen, in an airtight container.

IMPURITIES

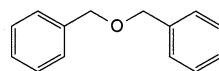
Calculate the sum of the percentage contents of the α -anomer and the β -anomer of ribenoside taking into account the assigned content of ribenoside CRS.

Injection Test solution (b) and reference solution (b). related substances with the following modification.

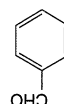
ASSAY

Dilute 5.0 mL of solution S to 20.0 mL with methanol R. 12 mL of the solution complies with test B. Prepare the reference solution using lead standard solution (1 ppm Pb) obtained by diluting lead standard solution (100 ppm Pb) R with methanol R.

D. dibenzyl ether.

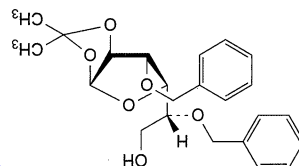


C. benzaldehyde,



glucofuranose,

B. 3,5-di-O-benzyl-1,2-O-(1-methylethylidene)- α -D-



Carrier gas helium for chromatography R.

Linear velocity 36 cm/s.

Split ratio 1:20.

Injection temperature:

Temperature	Time (min)	Column	Injection port	Detector
(°C)	(min)			
70 → 210	0 - 7	7 - 50	250	250

Detection Flame ionisation.

Injection 1 µL; inject via an inert, glass-lined injection port

using an automatic injection device.

Identification of impurities Use the chromatogram supplied

with tributyl acetylacrylate for peak identification CRS and the chromatogram obtained with reference solution (c) to identify

the peaks due to impurities B and C; use the chromatogram

obtained with reference solution (a) to identify the peak due

to impurity A.

Relative retention With reference to tributyl acetylacrylate

(retention time = about 24 min): impurity B = about 0.70;

impurity C = about 0.83; impurity A = about 0.87.

System suitability:

— resolution: minimum 2.0 between the peaks due to

impurity A and tributyl acetylacrylate in the chromatogram

obtained with reference solution (a);

— repeatability: maximum relative standard deviation of

5.0 per cent after 6 injections of reference solution (b).

Limits:

— impurity A: not more than 3 times the area of the

principal peak in the chromatogram obtained with

reference solution (b) (0.6 per cent);

— impurity C: not more than twice the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.4 per cent);

— impurity B: not more than the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.2 per cent);

— unspecified impurities: for each impurity, not more than

0.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(1.0 per cent);

— disregard limit: 0.25 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test F. Prepare the reference solution

using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 0.25 per cent, determined on 2.00 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Introduce 1.500 g into a 250 mL borosilicate glass flask.

Add 25 mL of 2-propanol R, 50 mL of water R, 25.0 mL of

1 M sodium hydroxide and a few glass beads. Heat under a

reflux condenser for 3 h. Allow to cool. Add 1 mL of

IMPURITIES

of $C_{20}H_{34}O_8$.

1 mL of 1 M sodium hydroxide is equivalent to 100.6 mg

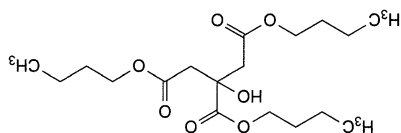
acid. Carry out a blank titration.

phenolphthalein solution R1 and titrate with 1 M hydrochloric

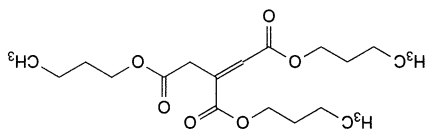
Specified impurities A, B, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

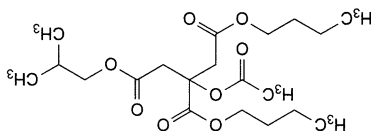
Control of impurities in substances for pharmaceutical use: D, E.



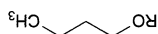
A. tributyl 2-hydroxypropene-1,2,3-tricarboxylate (tributyl citrate),



B. tributyl propene-1,2,3-tricarboxylate (tributyl acrylate),



C. 1,2-dibutyl 3-(2-methylpropyl) 2-(acetyloxy)propane-1,2,3-tricarboxylate,



D. R = H; butan-1-ol,

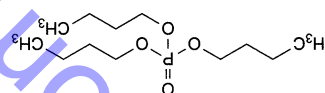
E. R = CO-CH₃; butyl acetate.

Ph Eur



Tributyl Phosphate

(Tri-*n*-butyl Phosphate, Ph Eur monograph 1682)



$C_{12}H_{27}O_4P$

266.3

126-73-8

Action and use

Excipient.

Ph Eur

CHARACTERS

Appearance

Clear, colourless or pale yellow liquid.

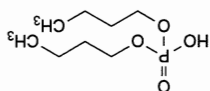
Solubility

Slightly soluble in water, miscible with ethanol (96 per cent).

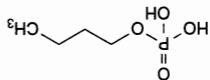
bp

About 289 °C, with decomposition.

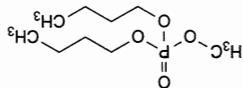
IMPURITIES



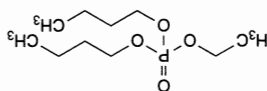
A. dibutyl hydrogen phosphate,



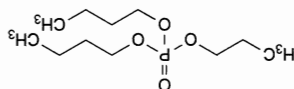
B. butyl dihydrogen phosphate,

C. $\text{H}_3\text{C}-[\text{CH}_2]_3-\text{OH}$: butan-1-ol,

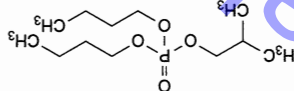
D. dibutyl methyl phosphate,



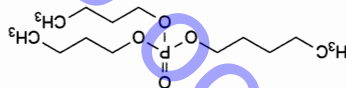
E. dibutyl ethyl phosphate,



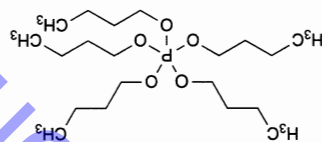
F. dibutyl propyl phosphate,



G. dibutyl 2-methylpropyl phosphate,



H. dibutyl pentyl phosphate,



I. pentabutyl phosphate.

Ph Eur

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison *tri-n*-butyl phosphate CRS.

TESTS

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution Y_6 (2.2.2, Method II).

Acidity

Dissolve 50 mL in 50 mL of ethanol (96 per cent) R previously adjusted with 0.02 M potassium hydroxide or 0.02 M hydrochloric acid to a bluish-green colour, using 0.5 mL of bromothymol blue solution R1 as indicator. Titrate with 0.02 M potassium hydroxide to the initial bluish-green coloration. Not more than 0.8 mL of 0.02 M potassium hydroxide is required.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.

Test solution The substance to be examined.

Reference solution Dissolve 10 mg of the substance to be examined and 10 mg of methyl myristate R in methylene chloride R and dilute to 10 mL with the same solvent.

Column:

material: fused silica;

size: $l = 30$ m, $\varnothing = 0.32$ mm;

stationary phase: poly(dimethyl)siloxane R (5 μm).

Carrier gas helium for chromatography R.

Linear velocity 32 cm/s.

Split ratio 65:1.

Temperature:

column: 250 °C;

injection port and detector: 250 °C.

Detection Flame ionisation.

Injection 1 μL .

Run time Twice the retention time of *tri-n*-butyl phosphate.

System suitability: reference solution:

— resolution: minimum 10 between the peaks due to *tri-n*-butyl phosphate and methyl myristate.

Limits:

any impurity: for each impurity, maximum 0.3 per cent;

total: maximum 0.5 per cent;

disregard limit: 0.01 per cent.

Chlorides (2.4.4)

Maximum 200 ppm.

Dissolve 0.25 g in 15 mL of ethanol (70 per cent V/V) R.

The solution complies with the test. Prepare the reference solution using 10 mL of chloride standard solution

(5 ppm Cl) R and 5 mL of anhydrous ethanol R.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in 13 mL of ethanol (96 per cent) R and dilute to 20.0 mL with water R. 12 mL of the solution complies

with test B. Prepare the reference solution using lead standard solution (2 ppm Pb) obtained by diluting lead

standard solution (100 ppm Pb) R with a mixture of 5 volumes of water R and 13 volumes of ethanol (96 per cent) R.

Water (2.5.32)

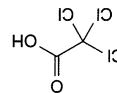
Maximum 0.1 per cent, determined on 1.0 g.

STORAGE

Protected from light.

Trichloroacetic Acid

(Ph. Eur. monograph 1967)

 $C_2HCl_3O_2$

163.4

76-03-9

Preparation

Trichloroacetic Acid Solution

Ph Eur

DEFINITION

2,2,2-Trichloroacetic acid.

Content

98.0 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or almost white, crystalline mass or colourless crystals, very deliquescent.

Solubility

Very soluble in water, in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification: A.

Second identification: B, C

A. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of trichloroacetic acid.

B. To 0.5 mL of solution S (see Tests) add 2 mL of

pyridine R and 5 mL of strong sodium hydroxide solution R.

Shake vigorously and heat in a water-bath at 60-70 °C for

5 min. The upper layer shows an intense red colour.

C. Solution S is strongly acidic (2.2.4).

TESTS

Solution S

Dissolve 2.5 g in water R and dilute to 25 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Chlorides (2.4.4)

Maximum 100 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 20 mL of water R. Titrate with 0.1 M sodium hydroxide, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 16.34 mg of $C_2HCl_3O_2$.

STORAGE

In an airtight container.

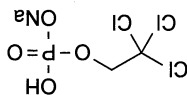
Ph Eur

Triclofos Sodium

 $C_2H_3Cl_3NaO_4P$

251.4

7246-20-0



Action and use
Hypnotic.

Preparation

Triclofos Oral Solution

DEFINITION

Triclofos Sodium is sodium 2,2,2-trichloroethyl hydrogen orthophosphate. It contains not less than 41.3% and not more than 43.2% of Cl and not less than 97.0% and not more than 102.0% of $C_2H_3Cl_3NaO_4P$, both calculated with reference to the dried substance.

CHARACTERISTICS

A white or almost white powder; hygroscopic.

Freely soluble in water; slightly soluble in ethanol (96%); practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of triclofos sodium (RS 350).

B. Yields the reactions characteristic of sodium salts, Appendix VI.

Appendix VI.

C. Heat 0.1 g with 1 g of anhydrous sodium carbonate to a dull residue with water and filter. The filtrate yields the reactions characteristic of chlorides and of phosphates, Appendix VI.

TESTS

Acidity

pH of a 2% w/v solution, 3.0 to 4.5, Appendix V L.

Clarity of solution

A 2.0% w/v solution is clear, Appendix IV A.

Heavy metals

A 10.0% w/v solution complies with limit test A for heavy metals, Appendix VII. Use lead standard solution (2 ppm Pb) to prepare the standard (20 ppm).

Chloride

Dissolve 0.1 g in 50 mL of water. 15 mL of the solution complies with the limit test for chlorides, Appendix VII (0.17%).

Phosphate

Not more than 1.0%, calculated as PO_4 , when determined by the following method. Dissolve 25 mg in 10 mL of water, add 4 mL of 1M sulfuric acid, 1 mL of ammonium molybdate solution and 2 mL of methylaminophenol-sulfite reagent and allow to stand for 15 minutes. Add sufficient water to produce 25 mL, allow to stand for a further 15 minutes and measure the absorbance of a 4 cm layer of the resulting solution at 730 nm, Appendix II B. Calculate the content of phosphate from a calibration curve prepared by treating suitable volumes of a 0.00143% w/v solution of potassium dihydrogen orthophosphate in the same manner.

Loss on drying

When dried at 100° at a pressure not exceeding 0.7 kPa for 3 hours, loses not more than 5.0% of its weight. Use 1 g.

ASSAY

For Cl

Mix 0.25 g with 1 g of anhydrous sodium carbonate in a nickel crucible about 3 cm in diameter, fill the crucible completely

with *anhydrous sodium carbonate* and invert into a nickel crucible about 4 cm in diameter; cover the smaller crucible with *anhydrous sodium carbonate*, well pressed down, using about 25 g of *anhydrous sodium carbonate* in all. Heat for 30 minutes at a dull red heat, cool, transfer to a 400 mL beaker, add 150 mL of *water* and boil gently for 10 minutes. Filter through absorbent cotton into a 600 mL beaker, washing the residue thoroughly with hot *water*, until about 400 mL of filtrate has been collected. Cool, cautiously add *nitric acid* until the solution is neutral to *litmus paper* and add 3 mL of *nitric acid* in excess. Add 50 mL of 0.1M *silver nitrate* VS, allow to stand until precipitation is complete, filter, wash the precipitate with *water* and titrate the combined filtrate and washings with 0.1M *ammonium thiocyanate* VS using *ammonium iron(III) sulfate solution* R2 as indicator. Each mL of 0.1M *silver nitrate* VS is equivalent to 3.545 mg of Cl.

For $C_6H_{15}NO_3$
 Heat 0.2 g in a Kjeldahl flask with 2 mL of *sulfuric acid* and 2.5 mL of *nitric acid* until brown fumes cease to be evolved, cool, add 1 mL of *nitric acid* and heat again. Continue adding *nitric acid* and heating until brown fumes are no longer evolved and the solution is colourless when cold. Heat until dense, white fumes are evolved, cool, transfer the solution to a flask with the aid of 150 mL of *water*, add 50 mL of *citric-molybdic acid solution* and heat slowly to boiling. Swirling the flask continuously, add 25 mL of *guanine solution* at first drop wise and then in a steady stream, heat on a water bath for 5 minutes and cool. Filter, wash the precipitate with *water* until free from acid, transfer the precipitate to a flask with the aid of 100 mL of *water*, add 50 mL of 0.5M *sodium hydroxide* VS and shake until dissolved. Titrate the excess of alkali with 0.5M *hydrochloric acid* VS using *phenolphthalein-thymol blue solution* as indicator. Each mL of 0.5M *sodium hydroxide* VS is equivalent to 4.835 mg of $C_6H_{15}NO_3$. Correct the result for the content of phosphate, as determined by the test described above; each mg of PO_4 is equivalent to 2.65 mg of $C_6H_{15}NO_3$.

Triethanolamine
 (Triolamine, Ph Eur monograph 1577)

OCCN(CCO)CO

149.2
 102-71-6

Action and use
 Pharmaceutical aid.

Ph Eur

DEFINITION
 2,2',2''-Nitrilotriethanol.

Content
 99.0 per cent *m/m* to 103.0 per cent *m/m* of total bases (anhydrous substance).

CHARACTERS
Appearance
 Clear, viscous, colourless or slightly yellow liquid, very hygroscopic.

Solubility
 Miscible with *water* and with *ethanol* (96 per cent), soluble in *methylene chloride*.

IDENTIFICATION
First identification B, C
Second identification A, B, D
 A. Relative density (2.2.5): 1.120 to 1.130.
 B. Refractive index (2.2.6): 1.482 to 1.485.
 C. Examine the chromatograms obtained in the test for related substances.
Results The principal peak in the chromatogram obtained with the test solution is similar in retention time and size to the principal peak in the chromatogram obtained with reference solution (a).
 D. To 1 mL add 0.3 mL of *copper sulfate solution* R. A blue colour develops. Add 2.5 mL of *dilute sodium hydroxide solution* R and heat to boiling. The blue colour remains unchanged.

TESTS
Solution S
 Dissolve 12 g in *water* R and dilute to 20 mL with the same solvent.

Appearance of solution
 Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Related substances
 Gas chromatography (2.2.28).
Internal standard solution Dissolve 5.0 g of 3-aminopropanol R in *water* R and dilute to 100.0 mL with the same solvent.
Test solution Dissolve 10.0 g of the substance to be examined in *water* R. Add 1.0 mL of the internal standard solution and dilute to 100.0 mL with *water* R.
Reference solution (a) Dissolve 1.0 g of triolamine CRS in *water* R and dilute to 10.0 mL with the same solvent.
Reference solution (b) Dissolve 0.1 g of triolamine impurity A CRS, 0.5 g of triolamine impurity B CRS and 0.1 g of triolamine impurity C CRS in *water* R and dilute to 10.0 mL with the same solvent.
 same solvent. To 1.0 mL of this solution add 1.0 mL of the internal standard solution and dilute to 100.0 mL with *water* R.
Column:
 — material: fused silica;
 — size: $l = 25$ m, $\phi = 0.25$ mm;
 — thickness 0.50 μ m).
Carrier gas *helium* for chromatography R.
Flow rate 1 mL/min.
Split ratio 1:35.
Temperature:

Time (min)	Temperature (°C)	Column
0	60	0
0 - 8.5	60 → 230	0 - 8.5
8.5 - 14	230	8.5 - 14
Injection port	260	
Detector	280	

Detection Flame ionisation.

Injection 2 μ L; if necessary inject a blank solution.

Blution order Impurity A, 3-aminopropanol, impurity B, triolamine.

— **resolution:** minimum 2.0 between the peaks due to 3-aminopropanol and impurity A.

Limits:

— **impurity A:** calculate the ratio (R₁) of the area of the peak due to impurity A to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (b); from the chromatogram obtained with the test solution, calculate the ratio of the area of any peak due to impurity A to the area of the peak due to the internal standard; this ratio is not greater than R₁

— **impurity B:** calculate the ratio (R₂) of the area of the peak due to impurity B to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (b); from the chromatogram obtained with the test solution, calculate the ratio of the area of any peak due to impurity B to the area of the peak due to the internal standard; this ratio is not greater than R₂

— **total:** calculate the ratio (R₄) of the area of the peak due to triolamine to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (b); from the chromatogram obtained with the test solution, calculate the ratio of the sum of the areas of any peaks, apart from the principal peak and the peak due to the internal standard, to the area of the peak due to the internal standard; this ratio is not greater than 10 times R₄ (1.0 per cent);

— **disregard limit:** 0.5 times the ratio of the area of the peak due to triolamine to the area of the peak due to the internal standard from the chromatogram obtained with reference solution (b) (0.05 per cent).

Impurity C

Gas chromatography (2.2.28).

Solvent mixture acetone R, chloroform R (10:50 V/V).

Preparation of solid phase extraction columns

Column A Fill a glass chromatography column (l = 400 mm; Ø = 20 mm) fitted with a teflon stopcock and a sintered-glass filter (160) (2.1.2) with 3 g of anhydrous sodium sulfate R and cover with a mixture of 17 g of kieselguhr for chromatography R and 3 g of potassium carbonate R. Settle the column bed by gently tapping the column.

Column B Fill a glass chromatography column (l = 400 mm; Ø = 20 mm) fitted with a teflon stopcock and a sintered-glass filter (160) (2.1.2) with a slurry of 25 g of silica gel for chromatography R (0.063 to 0.200 mm) in the solvent mixture. Apply slight pressure to settle the column and cover the column bed with 5 g of anhydrous sodium sulfate R.

Standard solution (a) Dissolve 50 µL of N-nitrosodihydroethanolamine R (impurity C) in methanol R and dilute to 50.0 mL with the same solvent. Dilute 100 µL of this solution to 100.0 mL with methanol R.

Standard solution (b) Dilute 10.0 mL of standard solution (a) to 50.0 mL with methanol R.

Standard solution (c) Dissolve 50 mg of N-nitrosodihydroethanolamine R in methanol R and dilute to 50.0 mL with the same solvent. Dilute 100 µL of this solution to 100.0 mL with methanol R.

Test solution To 2.000 g of the substance to be examined add 200 µL of methanol R and 0.5 g of sulfamic acid R. Dissolve in 8 mL of water for chromatography R and apply the solution to column A. Rinse the vessel twice with 1.5 mL of water for chromatography R, applying the rinsings to the column as

well. After 15 min of equilibration time elute the column with 100 mL of ethyl acetate R, collecting the eluate in a 250 mL distillation flask. Evaporate the eluate to dryness.

Take up the residue in 1 mL of the solvent mixture, apply to column B and let it settle. Rinse the flask twice with 2 mL of the solvent mixture, apply the rinsings to the column and let it settle. Wash the column with 100 mL of the solvent mixture and discard. Elute the column with 120 mL of acetone R, collecting the eluate in a 250 mL distillation flask. Evaporate the eluate to dryness. Transfer the residue with the aid of a small volume of acetone R into a vial and evaporate again to dryness under a stream of nitrogen R. Dissolve the residue in 100 µL of N-methyltrimethylsilyl-triethanolamine R, add 100 µL of N-methyltrimethylsilyl-triethanolamine R and heat at 70 °C for 1 h.

Reference solution (a) To 1.0 mL of standard solution (a) add 4.0 mL of standard solution (c) and mix. Transfer 500 µL of the solution to a vial and evaporate to dryness under a stream of nitrogen R. Dissolve the residue in 200 µL of N-trimethylpentane for chromatography R, add 200 µL of N-methyltrimethylsilyl-triethanolamine R and heat at 70 °C for 1 h.

Reference solution (b) To 1.0 mL of standard solution (a) add 4.0 mL of standard solution (c) and mix. Transfer 500 µL of the solution to a vial and evaporate to dryness under a stream of nitrogen R. Dissolve the residue in 100 µL of N-methyltrimethylsilyl-triethanolamine R, add 100 µL of N-methyltrimethylsilyl-triethanolamine R and heat at 70 °C for 1 h.

Blank solution In a gas chromatography vial, evaporate 200 µL of methanol R to dryness under a stream of nitrogen R. Dissolve the residue in 100 µL of N-methyltrimethylsilyl-triethanolamine R and heat at 70 °C for 1 h.

Column:
— **material:** fused silica;
— **size:** l = 30 m; Ø = 0.25 mm;
— **stationary phase:** base-deactivated poly(dimethyl) (diphenyl) siloxane R (film thickness 1 µm).
Carrier gas helium for chromatography R.
Flow rate 2 mL/min.
Split ratio 1:10.
Temperature:

Temperature (°C)	Time (min)	Column	Injection port
180 → 280	0 - 5		
280	5 - 10		
220			

Detection: Chemoluminescence;
— dual plasma burner in nitrosamine mode;
— burner temperature: 450 °C;
— oxygen flow rate: 4-5.0 mL/min.

Injection 4 µL.

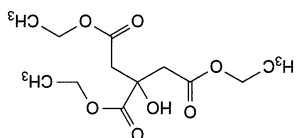
System suitability:

— **resolution:** minimum 1.3 between the peaks due to impurity C and N-nitrosodihydroethanolamine in the chromatogram obtained with reference solution (b);



(Ph. Eur. monograph 1479)

Triethyl Citrate

 $C_{12}H_{20}O_7$

276.3

77-93-0

Action and use

Excipient.

Ph Eur

DEFINITION

Triethyl 2-hydroxypropane-1,2,3-tricarboxylate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

Clear, viscous, colourless or almost colourless, hygroscopic liquid.

Solubility

Soluble in water, miscible with ethanol (96 per cent), slightly soluble in fatty oils.

IDENTIFICATION

First identification A, B

Second identification A, C, D

A. Refractive index (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of triethyl citrate.

C. It gives the reaction of esters (2.3.1).

D. To 0.5 mL add 5 mL of ethanol (96 per cent) R and 4 mL of dilute sodium hydroxide solution R. Boil under reflux for about 10 min. 2 mL of the solution gives the reaction of citrates (2.3.1).

TESTS

Appearance

The substance to be examined is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).

Acidity

Dilute 10 g with 10 mL of previously neutralised ethanol (96 per cent) R, add 0.5 mL of bromothymol blue solution R2. Not more than 0.3 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to blue.

Refractive index (2.2.6)

1.440 to 1.446.

Related substances

Gas chromatography (2.2.28): use the normalisation procedure.
Test solution Dissolve 1.0 mL of the substance to be examined in methylene chloride R and dilute to 50.0 mL with the same solvent.
Reference solution Dissolve 1.0 mL of the substance to be examined and 0.5 mL of methyl tridecanoate R in methylene chloride R, then dilute to 50.0 mL with the same solvent.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— recovery: minimum 50 per cent. The difference between the area of the peak due to impurity C in the chromatogram obtained with reference solution (a) and the area of the corresponding peak in the chromatogram obtained with the test solution is not less than 0.5 times the area of the corresponding peak in the chromatogram.

— impurity C: not more than the difference between the area of the peak due to impurity C in the chromatogram obtained with reference solution (a) and the area of the test solution (24 ppb).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dilute 5 mL of solution S to 30 mL with water R.

The solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.000 g.

Open the titration vessel, introduce the substance to be examined directly into the previously titrated solvent. Stopper the flask immediately.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g. Do not carry out the initial heating on a water-bath.

ASSAY

Dissolve 1.200 g in 75 mL of carbon dioxide-free water R. Add 0.3 mL of methyl red solution R. Titrate with 1 M hydrochloric acid.

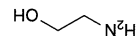
1 mL of 1 M hydrochloric acid is equivalent to 0.149 g of $C_6H_{15}NO_3$.

STORAGE

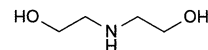
In an airtight container, protected from light.

IMPURITIES

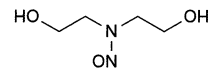
Specified impurities A, B, C



A. 2-aminoethanol (ethanolamine),



B. 2,2'-iminodietanol (dielethanolamine),



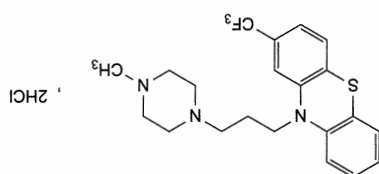
C. 2,2'-(nitrososimino)dietanol (N-nitrosodietethanolamine).

Ph Eur



Trifluoperazine Hydrochloride

(Ph. Eur. monograph 0059)



$C_{21}H_{26}Cl_2F_3N_3S$ 480.4 440-17-5

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Trifluoperazine Tablets

DEFINITION

Trifluoperazine hydrochloride contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 10-[3-(4-methylpiperazin-1-yl)propyl]-2-(trifluoromethyl)-10H-phenothiazine dihydrochloride, calculated with reference to the dried substance.

CHARACTERS

A white to pale yellow, crystalline powder, hygroscopic, freely soluble in water, soluble in alcohol.

It melts at about 242 °C, with decomposition.

IDENTIFICATION

A. Protect the solutions from bright light and measure the absorbances immediately. Dissolve 50 mg in 0.1 M hydrochloric acid and dilute to 500 mL with the same acid. Examined between 280 nm and 350 nm, the solution shows an absorption maximum (2.2.25) at 305 nm. Dilute 5 mL of the solution to 100 mL with 0.1 M hydrochloric acid. Examined between 230 nm and 280 nm, this solution shows an absorption maximum at 255 nm. The specific absorbance at this maximum is about 650.

B. It complies with the identification test for phenothiazines by thin-layer chromatography (2.3.3): use trifluoperazine hydrochloride CRS to prepare the reference solution.

C. Place 0.25 g in a 100 mL separating funnel, add 5 mL of water R and 2 mL of dilute sodium hydroxide solution R. Shake vigorously with 20 mL of ether R. Wash the ether layer with 5 mL of water R, add 0.15 g of maleic acid R and evaporate the ether. The residue, recrystallised from 30 mL of alcohol R and dried, melts (2.2.14) at about 192 °C.

D. Dissolve about 0.5 mg in 1 mL of water R, add 0.1 mL of bromine water R and shake for about 1 min. Add dropwise 1 mL of sulfuric acid R with constant, vigorous agitation. A red colour develops.

E. Dissolve about 50 mg in 5 mL of water R and add 2 mL of nitric acid R. A dark-red colour develops which turns to pale yellow. The solution gives reaction (a) of chlorides (2.3.1).

TESTS

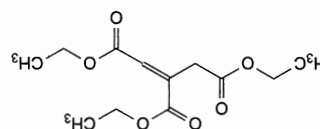
pH (2.2.3)

Dissolve 2.0 g in carbon dioxide-free water R and dilute to 20 mL with the same solvent. The pH of the solution is 1.6 to 2.5.

Related substances

Carry out the test protected from bright light.

A. triethyl propene-1,2,3-tricarboxylate (triethyl aconitate).



IMPURITIES

In an airtight container.

STORAGE

of $C_{12}H_{20}O_7$.

1 mL of 1 M sodium hydroxide is equivalent to 92.1 mg titrate with 1 M hydrochloric acid. Carry out a blank titration. Allow to cool. Add 1 mL of phenolphthalein solution R1 and few glass beads. Heat under a reflux condenser for 1 h. 50 mL of water R, 25.0 mL of 1 M sodium hydroxide and a fitted with a reflux condenser. Add 25 mL of 2-propanol R, Introduce 1.500 g into a 250 mL borosilicate-glass flask

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.25 per cent, determined on 1.000 g.

Water (2.5.12)

(96 per cent) R and water R.

(100 ppm Pb) R with a mixture of equal volumes of ethanol solution (1 ppm) obtained by diluting lead standard test B. Prepare the reference solution using lead standard to 20 mL with water R. 12 mL of the solution complies with Dissolve 4.0 g in 8 mL of ethanol (96 per cent) R and dilute Maximum 5 ppm.

Heavy metals (2.4.8)

— disregard limit: 0.04 per cent.

— total: maximum 0.5 per cent;

— any impurity: for each impurity, maximum 0.2 per cent;

Limits:

citrate and methyl tridecanoate.

— resolution: minimum 1.5 between the peaks due to triethyl

System suitability: reference solution:

Retention time Triethyl citrate = about 13.6 min.

Run time Twice the retention time of triethyl citrate.

Injection 1.0 µL.

Detection Flame ionisation.

— injection port and detector: 220 °C.

— column: 200 °C;

Temperature:

Split ratio About 1:50.

Linear velocity About 26 cm/s.

Carrier gas helium for chromatography R.

— stationary phase: poly(dimethyl)siloxane R (5 µm).

Examine by thin-layer chromatography (2.2.27), using a TLC silica gel GF₂₅₄ plate R.

Test solution Dissolve 0.2 g of the substance to be examined in a mixture of 5 volumes of diethylamine R and 95 volumes of methanol R and dilute to 10 mL with the same mixture of solvents. Prepare immediately before use.

Reference solution Dilute 1 mL of the test solution to 200 mL with a mixture of 5 volumes of diethylamine R and 95 volumes of methanol R.

Apply to the plate 10 µL of each solution. Develop over a path of 12 cm using a mixture of 10 volumes of acetone R, 10 volumes of diethylamine R and 80 volumes of cyclohexane R. Allow the plate to dry in air and examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with the test solution, apart from the principal spot, is not more intense than the spot in the chromatogram obtained with the reference solution (0.5 per cent).

Loss on drying (2.2.32)
Not more than 1.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)
Not more than 0.1 per cent, determined on 1.0 g.

ASSAY
Dissolve 0.200 g in 50 mL of alcohol R and add 5.0 mL of 0.01 M hydrochloric acid. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion.

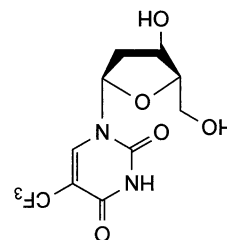
1 mL of 0.1 M sodium hydroxide is equivalent to 48.04 mg of C₂₁H₂₆Cl₂F₃N₃S.

STORAGE
Store in an airtight container, protected from light.

Ph Eur

Trifluridine

Trifluorothymidine



C₁₀H₁₁F₃N₂O₅

296.2

70-00-8

Action and use

Pyrimidine nucleoside analogue; antiviral (herpesviruses).

Preparation

Trifluridine Eye Drops

DEFINITION

Trifluridine is 1-[(2R,4S,5R)-4-hydroxy-5-(hydroxymethyl)oxolan-2-yl]-5-(trifluoromethyl)pyrimidine-2,4(1H,3H)-dione. It contains not less than 98.0% and not more than 102.0% of C₁₀H₁₁F₃N₂O₅, calculated with reference to the dried substance.

CHARACTERISTICS

A white, crystalline powder.

Soluble in water.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of trifluridine (RS 469).
B. In the Assay, the retention time of the principal peak in the chromatogram obtained with solution (1) is similar to that of the principal peak in the chromatogram obtained with solution (2).

TESTS

Specific optical rotation

In a 3% w/v solution in water, + 47 to + 51, Appendix V F, calculated with reference to the dried substance.

Related substances

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in a mixture of 5 volumes of acetonitrile and 95 volumes of a 0.05M solution of sodium dihydrogen orthophosphate which has been adjusted to pH 6.0 with ammonia.

- (1) 0.02% w/v of the substance being examined.
- (2) Dilute 1 volume of solution (1) to 100 volumes.
- (3) Dilute 1 volume of solution (2) to 10 volumes.
- (4) 0.02% w/v of trifluridine impurity standard BPCRS.

CHROMATOGRAPHIC CONDITIONS

- (a) Use a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm) (Inertsil ODS-2 is suitable).
- (b) Use gradient elution and the mobile phase described below.
- (c) Use a flow rate of 1 mL per minute.
- (d) Use an ambient column temperature.
- (e) Use a detection wavelength of 263 nm.
- (f) Inject 20 µL of each solution.

MOBILE PHASE

Mobile phase A 0.05M sodium dihydrogen orthophosphate, adjusted to pH 6.0 with ammonia.

Mobile phase B acetonitrile.

Time (Minutes)	Mobile phase A (% v/v)	Mobile phase B (% v/v)	Comment
0-3	95	5	isocratic
3-10	95→85	5→15	linear gradient
10-15	85	15	isocratic
15-16	85→95	15→5	linear gradient
16-25	95	5	re-equilibration

SYSTEM SUITABILITY

The test is not valid unless:
the chromatogram obtained with solution (4) closely resembles the chromatogram supplied with trifluridine impurity standard BPCRS;
in the chromatogram obtained with solution (4), the resolution factor between the peaks due to 5-carboxy-2'-deoxyuridine (impurity A) and 5-carboxyuracil (impurity C) is at least 1.5.

LIMITS

Identify any peaks in the chromatogram obtained with solution (1) corresponding to impurities C, D and E using solution (4) and multiply the areas of these peaks by the corresponding correction factors: impurity C, 0.6; impurity D, 0.7; impurity E, 0.4.

In the chromatogram obtained with solution (1):

the areas of any peaks corresponding to 5-carboxy-2'-deoxyuridine (impurity A) or 5-(trifluoromethyl)uracil (impurity B) are not greater than half the area of the principal peak in the chromatogram obtained with solution (2) (0.5% of each);

the area of any other secondary peak is not greater than the area of the principal peak in the chromatogram obtained with solution (3) (0.1%);

the sum of the areas of all the secondary peaks is not greater than the area of the principal peak in the chromatogram obtained with solution (2) (1%).

Disregard any peak with an area less than half the area of the peak in the chromatogram obtained with solution (3) (0.05%).

Loss on drying

When dried *in vacuo* at 105° for 4 hours, loses not more than 1.0% of its weight, Appendix IX D, Use 1 g.

ASSAY

Carry out the method for liquid chromatography, Appendix III D, using the following solutions in a mixture of 5 volumes of *acetonitrile* and 95 volumes of a 0.05M solution of *sodium dihydrogen orthophosphate* which has been adjusted to pH 6.0 with *ammonia*.

(1) 0.02% w/v of the substance being examined.

(2) 0.02% w/v of *trifluridine BP*RS.

CHROMATOGRAPHIC CONDITIONS

The chromatographic conditions described under Related substances may be used.

DETERMINATION OF CONTENT

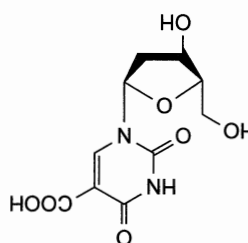
Calculate the content of $C_{10}H_{11}F_3N_2O_5$ in the substance being examined using the declared content of $C_{10}H_{11}F_3N_2O_5$ in *trifluridine BP*RS.

STORAGE

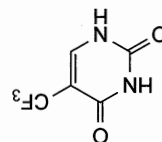
Trifluridine should be protected from light.

IMPURITIES

The impurities limited by the requirements of this monograph include:



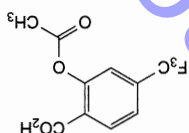
A. 1-[(2R,4S,5R)-4-hydroxy-5-(hydroxymethyl)oxolan-2-yl]-2,4-dioxo-1,2,3,4-tetrahydropyrimidin-5-carboxylic acid (5-carboxy-2'-deoxyuridine; trifluridine related compound A),



B. 5-(trifluoromethyl)pyrimidine-2,4(1H,3H)-dione (5-(trifluoromethyl)uracil; trifluorothymine),

Triflusal

(Ph. Eur. monograph 1377)



$C_{10}H_7F_3O_4$

248.2

322-79-2

Action and use

Thromboxane synthesis inhibitor; antiplatelet drug.

Ph. Eur.

DEFINITION

2-(Acetyloxy)-4-(trifluoromethyl)benzoic acid.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Practically insoluble in water, very soluble in anhydrous ethanol, freely soluble in methylene chloride.

mp

About 118 °C, with decomposition.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison *triflusal* CRS.



TESTS**Related substances**

Liquid chromatography (2.2.29).

Test solution Dissolve 0.200 g of the substance to be

examined in acetonitrile R and dilute to 20.0 mL with the

same solvent. Prepare the solution immediately before use.

Reference solution (a) Dissolve 5.0 mg of trifusal

impurity B CRS in acetonitrile R and dilute to 10.0 mL with

the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a)

to 25.0 mL with acetonitrile R.

Reference solution (c) Dissolve 2.5 mg of the substance to be

examined in acetonitrile R, add 5 mL of reference solution (a)

and dilute to 10.0 mL with acetonitrile R. Prepare the solution

immediately before use.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.0$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (4–5 μ m).

Mobile phase:

— mobile phase A: 0.5 per cent V/V solution of phosphoric

acid R₃

— mobile phase B: acetonitrile R₃

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 20	80 → 30	20 → 70
20 - 25	30	70

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 237 nm.

Injection 10 μ L of the test solution and reference solutions (b)

and (c).

Identification of impurities Use the chromatogram obtained

with reference solution (b) to identify the peak due to

impurity B.

Relative retention With reference to trifusal (retention

time = about 11 min): impurity B = about 1.2.

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to trifusal

and impurity B.

Limits:

— impurity B: not more than 1.5 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (b) (0.3 per cent);

— unspecified impurities: for each impurity, not more than

0.5 times the area of the peak due to impurity B in the

chromatogram obtained with reference solution (b)

(0.10 per cent);

— sum of impurities other than B: not more than 0.5 times the

area of the peak due to impurity B in the chromatogram

obtained with reference solution (b) (0.1 per cent);

— disregard limit: 0.25 times the area of the peak due to

impurity B in the chromatogram obtained with reference

solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in 12 mL of ethanol (96 per cent) R and dilute

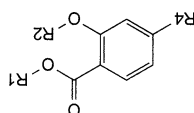
to 20 mL with water R. 12 mL of the solution complies with

test B. Prepare the reference solution using lead standard

solution (1 ppm Pb) obtained by diluting lead standard

solution (100 ppm Pb) R with a mixture of 2 volumes of

water R and 3 volumes of ethanol (96 per cent) R.



A. R1 = H, R2 = CO-CH₃, R4 = CO₂H;

2-(acetyloxy)benzene-1,4-dicarboxylic acid

(2-acetoxyterephthalic acid),

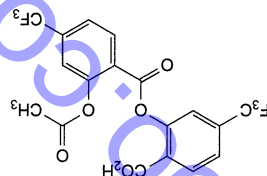
B. R1 = R2 = H, R4 = CF₃; 2-hydroxy-4-

(trifluoromethyl)benzoic acid (4-(trifluoromethyl)salicylic

acid),

C. R1 = R2 = CO-CH₃, R4 = CF₃; acetic 2-(acetyloxy)-4-

(trifluoromethyl)benzoic anhydride,



D. 2-[[2-(acetyloxy)-4-(trifluoromethyl)benzoyl]oxy]-4-

(trifluoromethyl)benzoic acid.

Ph Eur

IMPURITIES

Specified impurities B

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

by the general monograph *Substances for pharmaceutical use*

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, C,

D.

STORAGE

C₁₀H₇F₃O₄.

1 mL of 0.1 M sodium hydroxide is equivalent to 24.82 mg of

potentiometrically (2.2.20).

with 0.1 M sodium hydroxide, determining the end-point

Dissolve 0.200 g in 50 mL of anhydrous ethanol R. Titrate

ASSAY

crucible.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum

crucible.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.00 g by drying

in vacuo.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.00 g by drying

Medium-chain Triglycerides



Fractionated Coconut Oil
(Ph. Eur. monograph 0868)

Action and use

Excipient.
When Medium-chain Triglycerides are prepared from the endosperm of *Cocos nucifera* L. the title fractionated coconut oil may be used.

DEFINITION

Mixture of triglycerides of saturated fatty acids, mainly of caprylic (octanoic) acid and of capric (decanoic) acid. The fatty acids are obtained from the oil extracted from the hard, dried fraction of the endosperm of *Cocos nucifera* L. or from the dried endosperm of *Elaeis guineensis* Jacq.

Content

Minimum 95.0 per cent of saturated fatty acids with 8 and 10 carbon atoms.

CHARACTERS

Appearance

Colourless or slightly yellowish, oily liquid.

Solubility

Practically insoluble in water, miscible with ethanol (96 per cent), with methylene chloride, with light petroleum and with fatty oils.

IDENTIFICATION

First identification B, C

Second identification A, D

A. Heat 3.0 g under a reflux condenser for 30 min with

50 mL of a mixture of equal volumes of ethanol

(96 per cent) R and 2 M alcoholic potassium hydroxide R.

Reserve 10 mL of the mixture for identification test D.

To 40 mL of the mixture add 30 mL of water R, evaporate

the ethanol and acidify the hot solution with 25 mL of dilute

hydrochloric acid R. After cooling, shake with 50 mL of

peroxide-free ether R. Wash the ether layer with 3 quantities,

each of 10 mL, of sodium chloride solution R, dry over

anhydrous sodium sulfate R and filter. Evaporate the ether and

determine the acid value (2.5.1) of the residue, using

0.300 g. The acid value is 350 to 390.

B. Saponification value (see Tests).

C. Composition of fatty acids (see Tests).

D. Evaporate 10 mL of the alcoholic mixture obtained in

identification test A to dryness on a water-bath. Transfer the

residue into a test-tube, add 0.3 mL of sulfuric acid R and

close the test-tube with a stopper through which a U-shaped

glass tube is inserted. One end of the U-tube is dipped into

equal volumes of sulfuric acid R and water R. Heat the test-

tube in a silicone-oil bath at 180 °C for 10 min and collect

the liberated fumes in the tryptophan reagent. Heat the

tryptophan reagent on a water-bath for 1 min. A violet colour

develops.

The substance to be examined is clear (2.2.1) and not more

intensely coloured than reference solution Y₃ (2.2.2).

(Method I).

Method I).

Method I).

Method I).

Method I).

Method I).

Method I).

Method I).

Method I).

Alkaline impurities

Dissolve 2.00 g in a mixture of 1.5 mL of ethanol (96 per cent) R and 3.0 mL of ether R. Add 0.05 mL of bromophenol blue solution R. Not more than 0.15 mL of 0.01 M hydrochloric acid is required to change the colour of the indicator to yellow.

Relative density (2.2.5)

0.93 to 0.96.

Refractive index (2.2.6)

1.440 to 1.452.

Viscosity (2.2.9)

25 mPa.s to 33 mPa.s.

Acid value (2.5.1)

Maximum 0.2.

Hydroxyl value (2.5.3, Method A)

Maximum 10.

Iodine value (2.5.4)

Maximum 1.0.

Peroxide value (2.5.5, Method A)

Maximum 1.0.

Saponification value (2.5.6)

310 to 360.

Unsaponifiable matter (2.5.7)

Maximum 0.5 per cent, determined on 5.0 g.

Composition of fatty acids

Gas chromatography (2.4.22, Method C).

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.32$ mm;

— stationary phase: macrogol 20 000 R (film thickness

0.5 μ m).

Carrier gas: helium for chromatography R.

Flow rate 1.3 mL/min.

Temperature:

Temperature (°C)	Time (min)	Column	Injection port	Detector
70	0 - 1	35 - 50	250	250
70 → 240	1 - 35	240		

Detection: Flame ionisation.

Split ratio 1:100.

Composition of the fatty-acid fraction of the substance:

— caproic acid: maximum 2.0 per cent;

— caprylic acid: 50.0 per cent to 80.0 per cent;

— capric acid: 20.0 per cent to 50.0 per cent;

— lauric acid: maximum 3.0 per cent;

— myristic acid: maximum 1.0 per cent.

Chromium

Maximum 0.05 ppm, if intended for use in parenteral

nutrition.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution: Dissolve 2.0 g of the substance to be examined

in methyl isobutyl ketone R₃ and dilute to 10.0 mL with the

same solvent.

Solution A Dilute 0.100 mL of chromium liposoluble standard solution (1000 ppm Cr) R to 10.0 mL with methyl isobutyl ketone R3.

Stock solution Dilute 0.100 mL of solution A to 10.0 mL with methyl isobutyl ketone R3.

Reference solutions Prepare 3 reference solutions by dissolving for each 2.0 g of the substance to be examined in the minimum volume of methyl isobutyl ketone R3, adding 0.5 mL, 1.0 mL and 2.0 mL, respectively, of stock solution and diluting to 10.0 mL with methyl isobutyl ketone R3.

Source Chromium hollow-cathode lamp.

Wave length 357.8 nm.

Atomic generator Graphite furnace.

Carrier gas argon R.

Copper Maximum 0.1 ppm, if intended for use in parenteral nutrition.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 2.0 g of the substance to be examined in methyl isobutyl ketone R3 and dilute to 10.0 mL with the same solvent.

Solution A Dilute 0.100 mL of copper liposoluble standard solution (1000 ppm Cu) R to 10.0 mL with methyl isobutyl ketone R3.

Stock solution Dilute 0.100 mL of solution A to 10.0 mL with methyl isobutyl ketone R3.

Reference solutions Prepare 3 reference solutions by dissolving for each 2.0 g of the substance to be examined in the minimum volume of methyl isobutyl ketone R3, adding 1.0 mL, 2.0 mL and 4.0 mL, respectively, of stock solution and diluting to 10.0 mL with methyl isobutyl ketone R3.

Source Copper hollow-cathode lamp.

Wave length 324.7 nm.

Atomic generator Graphite furnace.

Carrier gas argon R.

Lead Maximum 0.1 ppm, if intended for use in parenteral nutrition.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 2.0 g of the substance to be examined in methyl isobutyl ketone R3 and dilute to 10.0 mL with the same solvent.

Solution A Dilute 0.100 mL of lead liposoluble standard solution (1000 ppm Pb) R to 10.0 mL with methyl isobutyl ketone R3.

Stock solution Dilute 0.100 mL of solution A to 10.0 mL with methyl isobutyl ketone R3.

Reference solutions Prepare 3 reference solutions by dissolving for each 2.0 g of the substance to be examined in the minimum volume of methyl isobutyl ketone R3, adding 1.0 mL, 2.0 mL and 4.0 mL, respectively, of stock solution and diluting to 10.0 mL with methyl isobutyl ketone R3.

Source Lead hollow-cathode lamp.

Wave length 283.3 nm.

Atomic generator Graphite furnace coated inside with palladium carbide; calcination is carried out in the presence of oxygen at a temperature below 800 °C.

Carrier gas argon R.

Nickel Maximum 0.2 ppm, if intended for use in parenteral nutrition.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 2.0 g of the substance to be examined in methyl isobutyl ketone R3 and dilute to 10.0 mL with the same solvent.

Solution A Dilute 0.100 mL of nickel liposoluble standard solution (1000 ppm Ni) R to 10.0 mL with methyl isobutyl ketone R3.

Stock solution Dilute 0.100 mL of solution A to 10.0 mL with methyl isobutyl ketone R3.

Reference solutions Prepare 3 reference solutions by dissolving for each 2.0 g of the substance to be examined in the minimum volume of methyl isobutyl ketone R3, adding 1.0 mL, 2.0 mL and 4.0 mL, respectively, of stock solution and diluting to 10.0 mL with methyl isobutyl ketone R3.

Source Nickel hollow-cathode lamp.

Wave length 232 nm.

Atomic generator Graphite furnace.

Carrier gas argon R.

Tin Maximum 0.1 ppm, if intended for use in parenteral nutrition.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 2.0 g of the substance to be examined in methyl isobutyl ketone R3 and dilute to 10.0 mL with the same solvent.

Solution A Dilute 0.100 mL of tin liposoluble standard solution (1000 ppm Sn) R to 10.0 mL with methyl isobutyl ketone R3.

Stock solution Dilute 0.100 mL of solution A to 10.0 mL with methyl isobutyl ketone R3.

Reference solutions Prepare 3 reference solutions by dissolving for each 2.0 g of the substance to be examined in the minimum volume of methyl isobutyl ketone R3, adding 1.0 mL, 2.0 mL and 4.0 mL, respectively, of stock solution and diluting to 10.0 mL with methyl isobutyl ketone R3.

Source Tin hollow-cathode lamp.

Wave length 286.3 nm.

Atomic generator Graphite furnace coated inside with palladium carbide.

Carrier gas argon R.

Heavy metals (2.4.8) Maximum 10 ppm, if intended for use other than parenteral nutrition.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12) Maximum 0.2 per cent, determined on 10.00 g.

Total ash (2.4.16) Maximum 0.1 per cent, determined on 2.0 g.

STORAGE In a well-filled container, protected from light.

LABELLING The label states, where applicable, that the substance is intended for use in parenteral nutrition.

Ph Eur

Triglycerol Diisostearate

(Ph. Eur. monograph 2032)

Ph Eur

DEFINITION

Mixture of polyglycerol diesters of mainly isostearic acid, obtained by esterification of polyglycerol and isostearic acid. The polyglycerol consists mainly of triglycerol.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Practically insoluble in water, miscible with ethanol

(96 per cent) and with fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Film between 2 plates of sodium chloride R.

Comparison triglycerol diisostearate CRS.

B. Composition of fatty acids (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₃ ().

Mix 10 mL with 10 mL of ethanol (96 per cent) R.

Acid value (2.5.1)

Maximum 3.0, determined on 1.0 g.

Hydroxyl value (2.5.3, Method A)

180 to 230, determined on 0.25 g.

Iodine value (2.5.4, Method B)

Maximum 5.0.

Peroxide value (2.5.5, Method B)

Maximum 6.0.

Saponification value (2.5.6)

128 to 160.

Composition of fatty acids (2.4.22, Method B)

Use the mixture of calibrating substances in Table 2.4.22-1.

Composition of the fatty-acid fraction of the substance:

— sum of the contents of the fatty acids eluting between palmitic acid and stearic acid: minimum 60.0 per cent;

— sum of the contents of myristic acid, palmitic acid and stearic acid: maximum 11.0 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash

Maximum 0.5 per cent, determined on 1.0 g.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 °C ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting with "Moisten the substance to be examined...".

STORAGE

In an airtight container, protected from light.

Ph Eur

Trihexyphenidyl Hydrochloride

(Ph. Eur. monograph 1626)

Ph Eur

DEFINITION

Mixture of polyglycerol diesters of mainly isostearic acid, obtained by esterification of polyglycerol and isostearic acid. The polyglycerol consists mainly of triglycerol.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Practically insoluble in water, miscible with ethanol

(96 per cent) and with fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Film between 2 plates of sodium chloride R.

Comparison triglycerol diisostearate CRS.

B. Composition of fatty acids (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₃ ().

Mix 10 mL with 10 mL of ethanol (96 per cent) R.

Acid value (2.5.1)

Maximum 3.0, determined on 1.0 g.

Hydroxyl value (2.5.3, Method A)

180 to 230, determined on 0.25 g.

Iodine value (2.5.4, Method B)

Maximum 5.0.

Peroxide value (2.5.5, Method B)

Maximum 6.0.

Saponification value (2.5.6)

128 to 160.

Composition of fatty acids (2.4.22, Method B)

Use the mixture of calibrating substances in Table 2.4.22-1.

Composition of the fatty-acid fraction of the substance:

— sum of the contents of the fatty acids eluting between palmitic acid and stearic acid: minimum 60.0 per cent;

— sum of the contents of myristic acid, palmitic acid and stearic acid: maximum 11.0 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash

Maximum 0.5 per cent, determined on 1.0 g.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 °C ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting with "Moisten the substance to be examined...".

STORAGE

In an airtight container, protected from light.

Ph Eur

Trihexyphenidyl Hydrochloride

(Ph. Eur. monograph 1626)

Ph Eur

DEFINITION

Mixture of polyglycerol diesters of mainly isostearic acid, obtained by esterification of polyglycerol and isostearic acid. The polyglycerol consists mainly of triglycerol.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Practically insoluble in water, miscible with ethanol

(96 per cent) and with fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Film between 2 plates of sodium chloride R.

Comparison triglycerol diisostearate CRS.

B. Composition of fatty acids (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₃ ().

Mix 10 mL with 10 mL of ethanol (96 per cent) R.

Acid value (2.5.1)

Maximum 3.0, determined on 1.0 g.

Hydroxyl value (2.5.3, Method A)

180 to 230, determined on 0.25 g.

Iodine value (2.5.4, Method B)

Maximum 5.0.

Peroxide value (2.5.5, Method B)

Maximum 6.0.

Saponification value (2.5.6)

128 to 160.

Composition of fatty acids (2.4.22, Method B)

Use the mixture of calibrating substances in Table 2.4.22-1.

Composition of the fatty-acid fraction of the substance:

— sum of the contents of the fatty acids eluting between palmitic acid and stearic acid: minimum 60.0 per cent;

— sum of the contents of myristic acid, palmitic acid and stearic acid: maximum 11.0 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash

Maximum 0.5 per cent, determined on 1.0 g.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 °C ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting with "Moisten the substance to be examined...".

STORAGE

In an airtight container, protected from light.

Ph Eur

Trihexyphenidyl Hydrochloride

(Ph. Eur. monograph 1626)

Ph Eur

DEFINITION

Mixture of polyglycerol diesters of mainly isostearic acid, obtained by esterification of polyglycerol and isostearic acid. The polyglycerol consists mainly of triglycerol.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Practically insoluble in water, miscible with ethanol

(96 per cent) and with fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Film between 2 plates of sodium chloride R.

Comparison triglycerol diisostearate CRS.

B. Composition of fatty acids (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₃ ().

Mix 10 mL with 10 mL of ethanol (96 per cent) R.

Acid value (2.5.1)

Maximum 3.0, determined on 1.0 g.

Hydroxyl value (2.5.3, Method A)

180 to 230, determined on 0.25 g.

Iodine value (2.5.4, Method B)

Maximum 5.0.

Peroxide value (2.5.5, Method B)

Maximum 6.0.

Saponification value (2.5.6)

128 to 160.

Composition of fatty acids (2.4.22, Method B)

Use the mixture of calibrating substances in Table 2.4.22-1.

Composition of the fatty-acid fraction of the substance:

— sum of the contents of the fatty acids eluting between palmitic acid and stearic acid: minimum 60.0 per cent;

— sum of the contents of myristic acid, palmitic acid and stearic acid: maximum 11.0 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash

Maximum 0.5 per cent, determined on 1.0 g.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 °C ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting with "Moisten the substance to be examined...".

STORAGE

In an airtight container, protected from light.

Ph Eur

Trihexyphenidyl Hydrochloride

(Ph. Eur. monograph 1626)

Ph Eur

DEFINITION

Mixture of polyglycerol diesters of mainly isostearic acid, obtained by esterification of polyglycerol and isostearic acid. The polyglycerol consists mainly of triglycerol.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Practically insoluble in water, miscible with ethanol

(96 per cent) and with fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Film between 2 plates of sodium chloride R.

Comparison triglycerol diisostearate CRS.

B. Composition of fatty acids (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₃ ().

Mix 10 mL with 10 mL of ethanol (96 per cent) R.

Acid value (2.5.1)

Maximum 3.0, determined on 1.0 g.

Hydroxyl value (2.5.3, Method A)

180 to 230, determined on 0.25 g.

Iodine value (2.5.4, Method B)

Maximum 5.0.

Peroxide value (2.5.5, Method B)

Maximum 6.0.

Saponification value (2.5.6)

128 to 160.

Composition of fatty acids (2.4.22, Method B)

Use the mixture of calibrating substances in Table 2.4.22-1.

Composition of the fatty-acid fraction of the substance:

— sum of the contents of the fatty acids eluting between palmitic acid and stearic acid: minimum 60.0 per cent;

— sum of the contents of myristic acid, palmitic acid and stearic acid: maximum 11.0 per cent.

Water (2.5.12)

Maximum 0.5 per cent, determined on 2.00 g.

Sulfated ash

Maximum 0.5 per cent, determined on 1.0 g.

Heat a silica crucible to redness for 30 min, allow to cool in a desiccator and weigh. Evenly distribute 1.00 g of the substance to be examined in the crucible and weigh. Dry at 100-105 °C for 1 h and ignite in a muffle furnace at 600 °C ± 25 °C until the substance is thoroughly charred. Carry out the test for sulfated ash (2.4.14) on the residue obtained, starting with "Moisten the substance to be examined...".

STORAGE

In an airtight container, protected from light.

Ph Eur

Trihexyphenidyl Hydrochloride

(Ph. Eur. monograph 1626)

Ph Eur

DEFINITION

Mixture of polyglycerol diesters of mainly isostearic acid, obtained by esterification of polyglycerol and isostearic acid. The polyglycerol consists mainly of triglycerol.

CHARACTERS

Appearance

Clear, yellowish, viscous liquid.

Solubility

Practically insoluble in water, miscible with ethanol

(96 per cent) and with fatty oils.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Film between 2 plates of sodium chloride R.

Comparison triglycerol diisostearate CRS.

B. Composition of fatty acids (see Tests).

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₃ ().

Mix 10 mL with 10 mL of ethanol (96 per cent) R.

Acid value (2.5.1)

Maximum 3.0, determined on 1.0 g.

Hydroxyl value (2.5.3, Method A)

TESTS**pH (2.2.3)**

5.2 to 6.2.

Dissolve 0.5 g with heating in 25 mL of carbon dioxide-free water R. Cool to room temperature and dilute to 50 mL with carbon dioxide-free water R.

Optical rotation (2.2.7)

-0.10° to +0.10°.

Dissolve 1.25 g in a mixture of 20 volumes of methanol R and 80 volumes of methylamine R and dilute to 25.0 mL with the same mixture of solvents.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase. Dilute 10.0 mL to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 10.0 mg of trihexyphenidyl impurity A CRS in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 100.0 mL with the mobile phase.

Reference solution (d) To 1 mL of reference solution (b), add 1 mL of the test solution and dilute to 100 mL with the mobile phase.

Column:

size: $l = 0.15$ m, $\phi = 4.6$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 200 mL of water R with 0.2 mL of triethylamine R. Adjust to pH 4.0 with phosphoric acid R and add 800 mL of acetonitrile R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L.

Run time 3 times the retention time of trihexyphenidyl.

System suitability Reference solution (d):

resolution: minimum 4.0 between the peaks due to trihexyphenidyl and to impurity A.

Limits:

impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.5 per cent);

unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

total: not more than 0.5 per cent;
disregard limit: 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

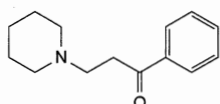
Dissolve 0.250 g in 50 mL of ethanol (96 per cent) R and add 5.0 mL of 0.01 M hydrochloric acid. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium

IMPURITIES $C_{20}H_{32}ClNO$.

1 mL of 0.1 M sodium hydroxide is equivalent to 33.79 mg of

inflixion.

hydroxide. Read the volume added between the 2 points of

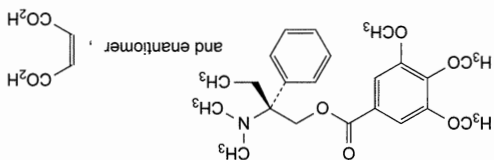


A. 1-phenyl-3-(piperidin-1-yl)propan-1-one.

Ph Eur

Trimebutine Maleate

(Ph Eur monograph 2182)



$C_{26}H_{33}NO_9$ 503.5 34140-59-5

Action and use

Treatment of irritable bowel syndrome.

DEFINITION

(2R,5)-2-(Dimethylamino)-2-phenylbutyl 3,4,5-trimethoxybenzoate (Z)-butenedioate.

Content

99.0 per cent to 101.5 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, soluble in acetonitrile, sparingly soluble in acetone, slightly soluble in ethanol (96 per cent).

mp

About 133 °C.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).
Comparison trimebutine maleate CRS.

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.5 g in acetone R, sonicate and dilute to 100 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture Dissolve 0.24 g of anhydrous sodium dihydrogen phosphate R in 180 mL of water R and adjust to pH 2.5 with dilute phosphoric acid R; dilute to 200 mL with water R. Add 50 mL of acetonitrile R and mix.

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent); disregard any peak due to maleic acid.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in 20 mL of a mixture of 15 volumes of

water R and 85 volumes of doxan R. 12 mL of the solution

complies with test B. Prepare the reference solution using

lead standard solution (1 ppm Pb) obtained by diluting lead

standard solution (100 ppm Pb) R with a mixture of

15 volumes of water R and 85 volumes of doxan R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of anhydrous acetic acid R. Titrate

with 0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 50.35 mg of

$C_{26}H_{33}NO_9$.

IMPURITIES

Specified impurities E.

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

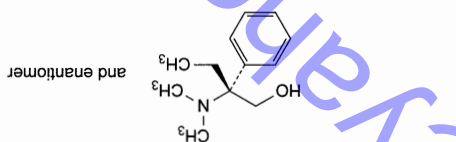
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

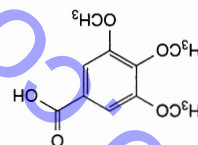
impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use): A, B,

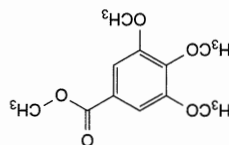
C, D.



A. (2RS)-2-(dimethylamino)-2-phenylbutanol,



B. 3,4,5-trimethoxybenzoic acid,



C. methyl 3,4,5-trimethoxybenzoate,

Test solution Dissolve 10 mg of the substance to be examined

in the solvent mixture and dilute to 10.0 mL with the solvent

mixture.

Reference solution (a) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of methyl

3,4,5-trimethoxybenzoate R (impurity C) in 10.0 mL of the

solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL

with the solvent mixture.

Reference solution (c) Dissolve the contents of a vial of

trimebutine for system suitability CRS (containing impurities D

and E) in 1.0 mL of reference solution (b).

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 μ m);

— temperature: 25 °C.

Mobile phase:

— mobile phase A: dissolve 3.6 g of anhydrous sodium

dihydrogen phosphate R in 990 mL of water R and adjust

to pH 3.0 with phosphoric acid R; dilute to 1000 mL with

water R;

— mobile phase B: acetonitrile R₁;

Time

(min)

Mobile phase A

(per cent V/V)

Mobile phase B

(per cent V/V)

22

78

22 → 35

78 → 65

65 → 60

35 → 40

40

15 - 35

6.5 - 15

3 - 6.5

0 - 3

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 μ L of the test solution and reference solutions (a)

and (c).

Identification of impurities Use the chromatogram supplied

with trimebutine for system suitability CRS and the

chromatogram obtained with reference solution (c) to identify

the peaks due to impurities C, D and E.

Relative retention With reference to trimebutine (retention

time = about 12 min); maleic acid = about 0.17;

impurity E = about 0.9; impurity D = about 1.3;

impurity C = about 1.4.

System suitability: reference solution (c):

— resolution: minimum 1.5 between the peaks due to

impurities D and C;

— peak-to-valley ratio: minimum 10, where H_p = height

above the baseline of the peak due to impurity E and

H_v = height above the baseline of the lowest point of the

curve separating this peak from the peak due to

trimebutine.

Limits:

— impurity E: not more than 6 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.6 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 10 times the area of the principal

peak in the chromatogram obtained with reference

solution (a) (1.0 per cent);

Test solution Dissolve 0.200 g of the substance to be examined in water R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 20.0 mg of trimetazidine for system suitability CRS in water R and dilute to 5.0 mL with the same solvent.

Reference solution (b) Dilute 2.0 mL of the test solution to 100.0 mL with water R. Dilute 5.0 mL of this solution to 100.0 mL with water R.
Reference solution (c) Dilute 25.0 mL of reference solution (b) to 50.0 mL with water R.

Column:
— size: $l = 0.15$ m, $\varnothing = 4.6$ mm,
— stationary phase: spherical octadecylsilyl silica gel for chromatography R (5 μ m) with a pore size of 10 nm,
— temperature: 30 °C.
Mobile phase:
— mobile phase A: mix 357 volumes of methanol R and 643 volumes of a 2.87 g/L solution of sodium heptanesulfonate R adjusted to pH 3.0 with dilute phosphoric acid R,
— mobile phase B: methanol R,

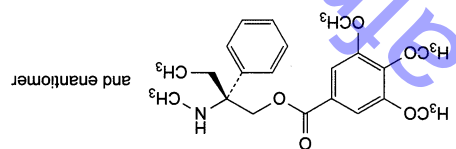
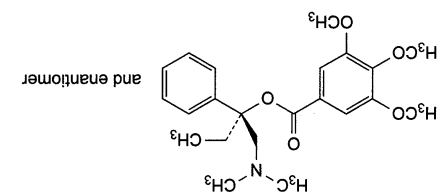
Time
(min)
Mobile phase A (per cent V/V) 95 → 75
Mobile phase B (per cent V/V) 5 → 25
50 - 52
75 → 95
25 → 5

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 240 nm.
Equilibration For at least 1 h with the mobile phase at the initial composition.
Injection 10 μ L.

Relative retention With reference to trimetazidine (retention time = about 25 min): impurity D = about 0.2; impurity C = about 0.4; impurity H = about 0.6; impurities A and I = about 0.9; impurity E = about 0.95; impurity F = about 1.4; impurity B = about 1.8.
System suitability:
— peak-to-valley ratio: minimum 3, where H_p = height above the baseline of the peak due to impurity E and H_b = height above the baseline of the lowest point of the curve separating this peak from the principal peak in the chromatogram obtained with reference solution (a);
— chromatogram obtained with reference solution (a);
— signal-to-noise ratio: minimum 10 for the principal peak in the chromatogram obtained with reference solution (c).

Limits:
— correction factors: for the calculation of contents, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.55; impurity C = 0.37; impurity F = 0.71;
— impurities A, B, C, D, E, F, H, I: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— any other impurity: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);
— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent);
— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (0.2 per cent);



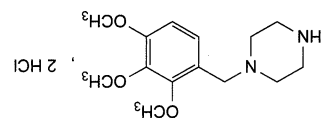
D. (1R,5S)-1-[(dimethylamino)methyl]-1-phenylpropyl 3,4,5-trimethoxybenzoate,
E. (2R,5S)-2-(methylamino)-2-phenylbutyl 3,4,5-trimethoxybenzoate.

Ph Eur



Trimetazidine Hydrochloride

(Trimetazidine Dihydrochloride, Ph Eur monograph 1741)



13171-25-0

Content
98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, crystalline powder, slightly hygroscopic.

Solubility
Freely soluble in water, sparingly soluble in alcohol.

IDENTIFICATION
A. Infrared absorption spectrophotometry (2.2.24).
Comparison Ph. Eur. reference spectrum of trimetazidine dihydrochloride.

B. Dissolve 25 mg in 5 mL of water R. 2 mL of the solution gives reaction (a) of chlorides (2.3.1).

TESTS
Appearance of solution
The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).
Dissolve 1.0 g in water R and dilute to 10 mL with the same solvent.
Related substances
Liquid chromatography (2.2.29).

— *disregard limit*: area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Impurity G

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 22.6 mg of piperazine hydrate R in methanol R and dilute to 100 mL with the same solvent.

Dilute 10 mL of the solution to 100 mL with methanol R.

Plate TLC silica gel plate R.

Mobile phase concentrated ammonia R, alcohol R (20:80 V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying At 100–105 °C for 30 min.

Detection Spray with iodoplatinate reagent R.

Limit:

— *impurity G*: any spot due to impurity G is not more intense than the spot in the chromatogram obtained with

anhydrous piperazine).

Loss on drying (2.2.32)

Maximum 2.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C over diposphorus pentoxide R at a pressure

not exceeding 15 kPa.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.120 g in 50.0 mL of water R. Add 1 mL of nitric acid R and titrate with 0.1 M silver nitrate, determining the

end-point potentiometrically (2.2.20).

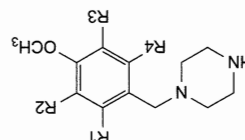
1 mL of 0.1 M silver nitrate is equivalent to 16.96 mg

of $C_{14}H_{24}Cl_2N_2O_3$.

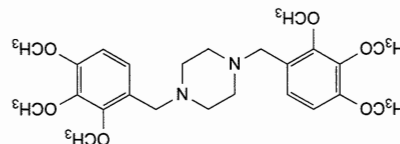
In an airtight container.

IMPURITIES

Specified impurities: A, B, C, D, E, F, G, H, I.



A. R1 = R4 = H, R2 = R3 = OCH₃; 1-(3,4,5-trimethoxybenzyl)piperazine,
E. R1 = R3 = OCH₃, R2 = H; 1-(2,4,5-trimethoxybenzyl)piperazine,
F. R1 = R4 = OCH₃, R2 = H; 1-(2,4,6-trimethoxybenzyl)piperazine,



B. 1,4-bis(2,3,4-trimethoxybenzyl)piperazine,

DEFINITION

3,5,5-Trimethylloxazolidine-2,4-dione.

Content

98.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

Colourless or almost colourless crystals.

Solubility

Soluble in water, very soluble in ethanol (96 per cent).

IDENTIFICATION

First identification B.

Second identification A, C, D.

A. Melting point (2.2.14): 45 °C to 47 °C, determined without previous drying.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs prepared using 3 mg of substance per 0.4 g

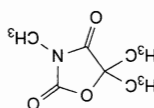
of potassium bromide R.

Comparison trimethadione CRS.

C. To 2 mL of solution S (see Tests) add 1 mL of barium hydroxide solution R. A white precipitate is formed, which dissolves on addition of 1 mL of dilute hydrochloric acid R.

Trimethadione

(Ph. Eur. monograph 0440)



143.1

$C_6H_9NO_3$

127-48-0

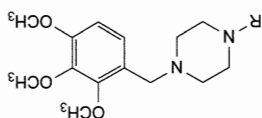
Action and use

Antiepileptic.

(N-methyltrimetazidine).

I. R = CH₃; 1-methyl-4-(2,3,4-trimethoxybenzyl)piperazine

H. R = COOC₂H₅; ethyl 4-(2,3,4-trimethoxybenzyl)piperazine-1-carboxylate,

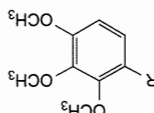


G. piperazine,



D. R = CH₂OH; (2,3,4-trimethoxyphenyl)methanol,

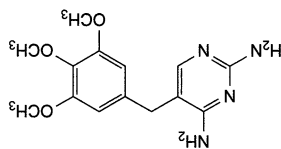
C. R = CHO; 2,3,4-trimethoxybenzaldehyde,





Trimethoprim

(Ph. Eur. monograph 0060)



C₁₄H₁₈N₄O₃ 290.3 738-70-5

Action and use
Dihydrofolate reductase inhibitor; antibacterial.

Preparations
Co-trimoxazole Infusion

Co-trimoxazole Oral Suspension

Paediatric Co-trimoxazole Oral Suspension

Co-trimoxazole Tablets

Dispersible Co-trimoxazole Tablets

Paediatric Co-trimoxazole Tablets

Trimethoprim Oral Suspension

Trimethoprim Tablets

DEFINITION

5-(3,4,5-Trimethoxybenzyl)pyrimidine-2,4-diamine.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or yellowish-white powder.

Solubility

Very slightly soluble in water, slightly soluble in ethanol (96 per cent).

IDENTIFICATION

First identification C.

Second identification A, B, D.

A. Melting point (2.2.14): 199 °C to 203 °C.

B. Dissolve about 20 mg in 0.1 M sodium hydroxide and dilute to 100.0 mL with the same solvent. Dilute 1.0 mL of this solution to 10.0 mL with 0.1 M sodium hydroxide. Examined between 230 nm and 350 nm (2.2.25), the solution shows an absorption maximum at 287 nm. The specific absorbance at the absorption maximum is 240 to 250.

C. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison trimethoprim CRS.

D. Dissolve about 25 mg, heating if necessary, in 5 mL of 0.005 M sulfuric acid and add 2 mL of a 16 g/L solution of potassium permanganate R in 0.1 M sodium hydroxide. Heat to boiling and add to the hot solution 0.4 mL of formaldehyde R. Mix, add 1 mL of 0.5 M sulfuric acid, mix and heat again to boiling. Cool and filter. To the filtrate, add 2 mL of methylene chloride R and shake vigorously. The organic layer, examined in ultraviolet light at 365 nm, shows green fluorescence.

TESTS

Solution S

Dissolve 2.0 g in carbon dioxide-free water R and dilute to 40 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 10 mL of solution S add 0.1 mL of methyl red solution R. Not more than 0.1 mL of 0.01 M hydrochloric acid or 0.01 M sodium hydroxide is required to change the colour of the indicator.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.00 g by drying in a desiccator over anhydrous silica gel R for 6 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Gas chromatography (2.2.28).

Internal standard solution Dissolve 0.125 g of decanol R in anhydrous ethanol R and dilute to 25 mL with the same solvent.

Test solution Dissolve 0.100 g of the substance to be examined in the internal standard solution and dilute to 10.0 mL with the same solution.

Reference solution Dissolve 0.100 g of trimethoprim CRS in the internal standard solution and dilute to 10.0 mL with the same solution.

Column:

material: stainless steel,

size: l = 0.75 m, Ø = 3 mm,

stationary phase: styrene-divinylbenzene copolymer R (125-150 µm).

Carrier gas nitrogen for chromatography R.

Flow rate 20 mL/min.

Temperature:

column: 210 °C,

injection port: 240 °C,

detection: 270 °C.

Detection Flame ionisation.

Injection 1 µL.

Calculate the content of C₁₄H₁₈N₄O₃ from the declared content of trimethoprim CRS.

STORAGE

Protected from light.

TESTS

Appearance of solution

The solution is not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Dissolve 0.5 g in 10 mL of a mixture of 1 volume of water R₃, 4.5 volumes of methanol R and 5 volumes of methylene chloride R.

Related substances

A. Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of trimethoprim for system suitability CRS (containing impurity E) in 1 mL of the mobile phase.

Column:

— size: $l = 0.250$ m, $\varnothing = 4.0$ mm;

— stationary phase: base-deactivated octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 30 volumes of methanol R and 70 volumes of a 1.4 g/L solution of sodium perchlorate R adjusted to pH 3.6 with phosphoric acid R.

Flow rate 1.3 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L loop injector.

Run time 11 times the retention time of trimethoprim.

Relative retention With reference to trimethoprim (retention time = about 5 min): impurity C = about 0.8;

impurity B = about 0.9; impurity A = about 1.5;

impurity D = about 2.0; impurity G = about 2.1;

impurity B = about 2.3; impurity J = about 2.7;

impurity F = about 4.0.

System suitability: reference solution (b):

— resolution: minimum 2.5 between the peaks due to impurity E and trimethoprim.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor:

impurity B = 0.43; impurity E = 0.53;

impurity J = 0.66;

— **any impurity:** not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **total:** not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **disregard limit:** 0.04 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent);

— **disregard limit:** 0.04 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent);

corresponding to impurity H (relative retention = about 10.3).

B. Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 200.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of trimethoprim CRS and 5.0 mg of trimethoprim impurity B CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: nitrite silica gel for chromatography R (5 μ m) with a specific surface area of 350 m²/g and a pore diameter of 10 nm.

Mobile phase Dissolve 1.14 g of sodium hexanesulfonate R in 600 mL of a 13.6 g/L solution of potassium dihydrogen phosphate R; adjust to pH 3.1 with phosphoric acid R and mix with 400 mL of methanol R.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 μ L loop injector.

Run time 6 times the retention time of trimethoprim.

Relative retention With reference to trimethoprim (retention time = about 4 min): impurity H = about 1.8;

impurity I = about 4.9.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to trimethoprim and impurity B.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor:

impurity H = 0.50; impurity I = 0.28;

— **any impurity:** not more than 0.2 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— **total:** not more than 0.4 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— **disregard limit:** 0.04 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent);

— **disregard limit:** 0.04 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.02 per cent); disregard any peak due to impurity B (relative retention = about 1.3).

Impurity K

Gas chromatography (2.2.28).

Test solution Dissolve 0.500 g of the substance to be examined in 35.0 mL of citrate buffer solution pH 5.0 R, add 10.0 mL of 1,1-dimethylethyl methyl ether R, shake thoroughly and centrifuge for 10 min. Use the upper layer.

Reference solution Dilute 5.0 mL of hydrotrolic acid R to 50.0 mL with water R, add 12.5 mg of aniline R and shake thoroughly. Add 10.0 μ L of this solution and 10.0 mL of 1,1-dimethylethyl methyl ether R to 35.0 mL of citrate buffer solution pH 5.0 R, shake thoroughly and centrifuge for 10 min. Use the upper layer.

Column:

— material: fused silica;

— size: $l = 30$ m, $\varnothing = 0.53$ mm;

— stationary phase: poly(dimethyl)siloxane R (film thickness 3 μ m).

Carrier gas helium for chromatography R.

Flow rate 12 mL/min.

Temperature:

— column: 80 °C;

— injection port: 230 °C;

— detector: 270 °C.

Detection Nitrogen-phosphorus detector.

Injection 3 μ L.

Run time 15 min.

System suitability: reference solution:

— **repeatability:** maximum relative standard deviation of 5.0 per cent after 6 injections.

Limit:

— impurity K: not more than the area of the corresponding peak in the chromatogram obtained with the reference solution (5 ppm).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 29.03 mg of $C_{14}H_{18}N_4O_3$.

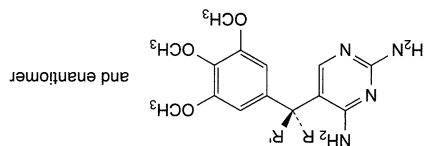
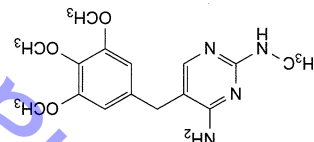
IMPURITIES

By liquid chromatography A: A, B, C, D, E, F, G, H, J.

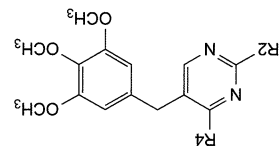
By liquid chromatography B: B, H, I.

By gas chromatography: K.

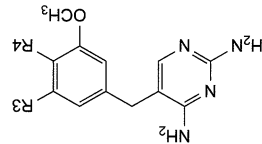
A. N²-methyl-5-(3,4,5-trimethoxybenzyl)pyrimidine-2,4-diamine,



B. R + R' = O: (2,4-diaminopyrimidin-5-yl) (3,4,5-trimethoxyphenyl)methanone,
C. R = OH, R' = H: (R,S)-(2,4-diaminopyrimidin-5-yl) (3,4,5-trimethoxyphenyl)methanol,



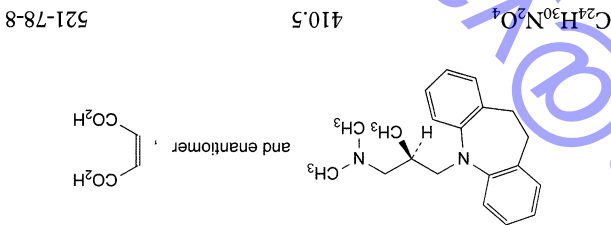
D. R2 = NH2, R4 = OH: 2-amino-5-(3,4,5-trimethoxybenzyl)pyrimidin-4-ol,
E. R2 = OH, R4 = NH2: 4-amino-5-(3,4,5-trimethoxybenzyl)pyrimidin-2-ol,



F. R3 = Br, R4 = OCH3: 5-(3-bromo-4,5-dimethoxybenzyl)pyrimidine-2,4-diamine,
G. R3 = OCH3, R4 = OC2H5: 5-(4-ethoxy-3,5-dimethoxybenzyl)pyrimidine-2,4-diamine,

Trimipramine Maleate

(Ph. Eur. monograph 0534)



521-78-8

410.5

$C_{24}H_{30}N_4O_4$

Action and use

Monamine reuptake inhibitor; tricyclic antidepressant.

Preparation

Trimipramine Tablets

DEFINITION

(2R,S)-3-(10,11-Dihydro-5H-dibenzo[b,j]azepin-5-yl)-N,N,2-trimethylpropan-1-amine (Z)-butenedioate.

Content

98.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water and in ethanol (96 per cent).

IDENTIFICATION

First identification A, C.

Second identification A, B, D, E.

A. Melting point (2.2.14): 140 °C to 144 °C.

B. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 40.0 mg in 100.0 mL with the same acid. Dilute 5.0 mL of this solution to 100.0 mL with 0.01 M hydrochloric acid.

- Spectral range** 230-350 nm.
Absorption maximum At 250 nm.
Shoulder At 270 nm.
- Specific absorbance at the absorption maximum** 205 to 235.
C. Infrared absorption spectrophotometry (2.2.24).
Comparison trimpipramine maleate CRS.
D. Thin-layer chromatography (2.2.27). Prepare the solutions immediately before use.
Test solution Dissolve 0.50 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.
Dilute 1 mL of this solution to 20 mL with methanol R.
Reference solution Dissolve 25 mg of trimpipramine maleate CRS in methanol R and dilute to 10 mL with the same solvent.
Plate TLC silica gel G plate R.
Mobile phase concentrated ammonia R, anhydrous ethanol R, toluene R (0.7:10:90 V/V/V).
Application 5 µL.
Development Over 2/3 of the plate.
Drying In air for 15 min.
Detection Spray with a 5 g/L solution of potassium dichromate R in a mixture of 1 volume of sulfuric acid R and 4 volumes of water R and examine immediately.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.
E. Thin-layer chromatography (2.2.27).
Test solution Dissolve 0.20 g of the substance to be examined in methanol R and dilute to 10 mL with the same solvent.
Reference solution Dissolve 56 mg of maleic acid R in methanol R and dilute to 10 mL with the same solvent.
Plate TLC silica gel GF₂₅₄ plate R.
Mobile phase water R, anhydrous formic acid R, di-isopropyl ether R (3:7:90 V/V/V).
Application 5 µL as bands of 10 mm.
Development Over 2/3 of the plate.
Drying In a current of air for a few minutes and then at 120 °C for 10 min.
Detection Examine in ultraviolet light at 254 nm.
Results The chromatogram obtained with the test solution shows 2 zones: one is on the line of application and the other is similar in position and size to the principal zone in the chromatogram obtained with the reference solution.
- TESTS**
Related substances
Liquid chromatography (2.2.29). Carry out the test protected from light.
Buffer solution pH 7.7 2.64 g/L solution of ammonium phosphate R in water for chromatography R; adjust to pH 7.7 with phosphoric acid R.
Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 100 mL with the mobile phase.
Reference solution (a) Dissolve 5 mg of the substance to be examined and 5 mg of *immodibenzyl* R (impurity F) in the mobile phase and dilute to 10 mL with the mobile phase.
Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.
- Heavy metals** (2.4.8).
Maximum 20 ppm.
Loss on drying (2.2.32)
Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.
- Limits:**
Resolution: minimum 3.5 between the peaks due to trimpipramine and impurity F in the chromatogram obtained with reference solution (a);
Identification of impurities Use the chromatogram supplied with trimpipramine for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D and E; doubling of the peak due to impurity E may be observed.
Relative retention With reference to trimpipramine (retention time = about 27 min): impurity A = about 0.1; impurity B = about 0.3; impurity C = about 0.4; impurity D = about 0.5; impurity F = about 1.4; impurity E = about 1.5.
System suitability:
Resolution: minimum 3.5 between the peaks due to trimpipramine and impurity F in the chromatogram obtained with reference solution (a);
Identification of impurities Use the chromatogram supplied with trimpipramine for peak identification CRS and the chromatogram obtained with reference solution (c) to identify the peaks due to impurities A, B, C, D and E; doubling of the peak due to impurity E may be observed.
Run time 3 times the retention time of trimpipramine.
Injection 20 µL.
Detection Spectrophotometer at 210 nm.
Flow rate 1.5 mL/min.
Mobile phase acetonitrile R₁, buffer solution pH 7.7 (38:62 V/V).
Stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm).
Column:
size: l = 0.125 m, Ø = 4.0 mm;
Injection phase: end-capped octadecylsilyl silica gel for chromatography R₁ in 1 mL of acetonitrile R₁.
Reference solution (c) Dissolve the contents of a vial of trimpipramine for peak identification CRS (containing impurities A, B, C, D and E) in 1 mL of acetonitrile R₁.

ASSAY

Dissolve 0.350 g in 50 mL of anhydrous acetic acid *R*. Titrate with 0.1 *M* perchloric acid determining the end-point potentiometrically (2.2.20).
1 mL of 0.1 *M* perchloric acid is equivalent to 41.05 mg of $C_{24}H_{30}N_2O_4$.

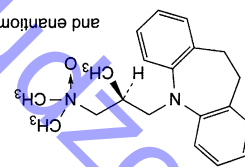
STORAGE

Protected from light.

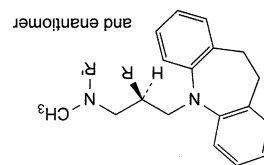
IMPURITIES

Specified impurities A, B, C, D, E, F

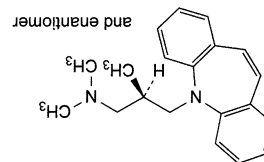
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): G.



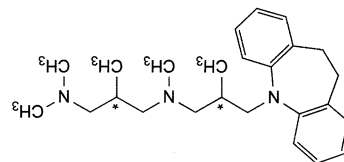
A. (2*RS*)-3-(10,11-dihydro-5*H*-dibenzo[*b,j*]azepin-5-yl)-*N,N*,2-trimethylpropan-1-amine *N*-oxide, and enantiomer



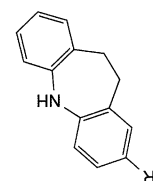
B. *R* = CH_3 , *R'* = H: (2*RS*)-3-(10,11-dihydro-5*H*-dibenzo[*b,j*]azepin-5-yl)-*N,N*,2-trimethylpropan-1-amine, *D*. *R* = H, *R'* = CH_3 : imipramine, and enantiomer



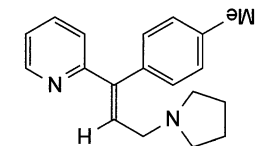
C. (2*RS*)-3-(5*H*-dibenzo[*b,j*]azepin-5-yl)-*N,N*,2-trimethylpropan-1-amine, and enantiomer



E. mixture of the stereoisomers of *N*-[3-(10,11-dihydro-5*H*-dibenzo[*b,j*]azepin-5-yl)-2-methylpropyl]-*N,N'*,*N''*,2-tetramethylpropane-1,3-diamine,



F. *R* = H: 10,11-dihydro-5*H*-dibenzo[*b,j*]azepine, G. *R* = CH_3 : 2-methyl-10,11-dihydro-5*H*-dibenzo[*b,j*]azepine.



$C_{19}H_{22}N_2 \cdot HCl \cdot H_2O$ 332.9 6138-79-0

Action and use

Histamine H_1 receptor antagonist; antihistamine.

Preparation

Triprolidine Tablets

DEFINITION

Triprolidine Hydrochloride is (*E*)-2-(3-pyrrolidin-1-yl-1-*p*-tolylprop-1-en-1-yl)pyridine hydrochloride monohydrate. It contains not less than 98.5% and not more than 101.0% of $C_{19}H_{22}N_2 \cdot HCl$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

A white, crystalline powder. Freely soluble in water; freely soluble in ethanol (96%) and practically insoluble in ether.

IDENTIFICATION

A. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of triprolidine hydrochloride (*RS* 356).

B. Yields reaction A characteristic of chlorides, Appendix VI.

TESTS

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, using the following solutions in methanol.
(1) 1.0% w/v of the substance being examined.
(2) 0.020% w/v of *Z*-triproline hydrochloride *BPCRS*.
(3) 0.010% w/v of the substance being examined.

CHROMATOGRAPHIC CONDITIONS

(a) Use as the coating silica gel F_{254} (Merck silica gel 60 F_{254} plates are suitable).

(b) Use the mobile phase as described below.

(c) Apply 5 μ L of each solution.

(d) Develop the plate to 15 cm.

(e) After removal of the plate, dry in air, examine under ultraviolet light (254 nm).

MOBILE PHASE

Equal volumes of butan-2-one and dimethylformamide.

LIMITS

In the chromatogram obtained with solution (1):

any spot corresponding to *Z*-triprolidine is not more intense than the spot in the chromatogram obtained with solution (2);

and any other *secondary spot* is not more intense than the spot in the chromatogram obtained with solution (3).

Sulfated ash

Not more than 0.1%, Appendix IX A.

Water

4.5 to 6.0% w/w, Appendix IX C. Use 0.4 g.

ASSAY

Carry out Method I for non-aqueous titration,

Appendix VIII A, using 0.25 g dissolved in a mixture of

50 mL of *anhydrous acetic acid* and 0.5 mL of *acetic anhydride*

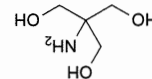
and *crystal violet solution* as indicator. Each mL of

0.1M *perchloric acid VS* is equivalent to 15.74 mg of

$C_{19}H_{22}N_2 \cdot HCl$.

Trometamol

(Ph. Eur. monograph 1053)



$C_4H_{11}NO_3$ 121.1

77-86-1

Action and use
Organic amine proton acceptor; alkalinizing agent.

DEFINITION

Trometamol contains not less than 99.0 per cent and not

more than the equivalent of 100.5 per cent of

aminomethyldinitromethanol, calculated with reference to

the dried substance.

CHARACTERS

A white or almost white, crystalline powder, or colourless

crystals, freely soluble in water, sparingly soluble in alcohol,

very slightly soluble in ethyl acetate.

IDENTIFICATION

First identification B, C.

Second identification A, B, D.

A. Solution S (see Tests) is strongly alkaline (2.2.4).

B. Melting point (2.2.14): 168 °C to 174 °C.

C. Examine by infrared absorption spectrophotometry

(2.2.24), comparing with the spectrum obtained with

trometamol CRS.

D. Examine the chromatograms obtained in the test for

related substances. The principal spot in the chromatogram

obtained with test solution (b) is similar in position, colour

and size to the principal spot in the chromatogram obtained

with reference solution (a).

TESTS

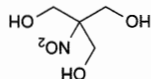
Solution S

Dissolve 2.5 g in *carbon dioxide-free water R* and dilute to

50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).



A. nitromethyldinitromethanol.

IMPURITIES

$C_4H_{11}NO_3$.

1 mL of 0.1 M *hydrochloric acid* is equivalent to 12.11 mg of

colour changes from yellow to red.

Dissolve 0.100 g in 20 mL of *water R*. Add 0.2 mL of *methyl*

red solution R. Titrate with 0.1 M *hydrochloric acid* until the

procedure for the removal of bacterial endotoxins.

Less than 0.03 IU/mg, if intended for use in the manufacture

Bacterial endotoxins (2.6.14)

Not more than 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

drying in an oven at 105 °C.

Not more than 0.5 per cent, determined on 1.000 g by

Loss on drying (2.2.32)

(10 ppm).

Dissolve 1.0 g in *water R* and dilute to 10 mL with the same

Iron (2.4.9)

12 mL of the solution complies with test A for heavy metals

with *hydrochloric acid R1* and dilute to 20 mL with *water R*.

Dissolve 2.0 g in 10 mL of *water R*. Neutralise the solution

Heavy metals (2.4.8)

with the limit test for chlorides (100 ppm).

To 10 mL of solution S add 2.5 mL of *dilute nitric acid R*

and dilute to 15 mL with *water R*. The solution complies

Chlorides (2.4.4)

(1.0 per cent).

chromatogram obtained with reference solution (b)

principal spot, is not more intense than the spot in the

chromatogram obtained with test solution (a), apart from the

After about 10 min examine in daylight. Any spot in the

permananganate R in a 10 g/L solution of *sodium carbonate R*.

at 100 °C to 105 °C. Spray with a 5 g/L solution of *potassium*

ammonia R1 and 90 volumes of 2-propanol R. Dry the plate

path of 10 cm using a mixture of 10 volumes of *dilute*

Apply to the plate 10 µL of each solution. Develop over a

100 mL with *methanol R*.

Reference solution (b) Dilute 1 mL of test solution (a) to

methanol R and dilute to 10 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of *trometamol CRS* in

with *methanol R*.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL

heating, and dilute to 10 mL with *methanol R*.

Test solution (a) Dissolve 0.20 g in 1 mL of *water R*, with

methanol R before applying the solutions.

Examine by thin-layer chromatography (2.2.27), using *silica*

gel G R as the coating substance. Wash the plate with

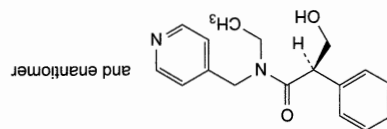
Related substances

The pH of freshly prepared solution S is 10.0 to 11.5.

pH (2.2.3)

Tropicamide

(Ph. Eur. monograph 1159)


 $C_{17}H_{20}N_2O_2$ 284.4 1508-75-4

Action and use

Anticholinergic.

Preparation

Tropicamide Eye Drops

DEFINITION

(2R)-N-Ethyl-3-hydroxy-2-phenyl-N-(pyridin-4-ylmethyl)propanamide.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Slightly soluble in water, freely soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

First identification C.

Second identification A, B, D, E.

A. Melting point (2.2.14): 95 °C to 98 °C.

B. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 20.0 mg in 0.1 M hydrochloric acid and dilute to 50.0 mL with the same acid. Dilute 2.0 mL of this solution to 20.0 mL with 0.1 M hydrochloric acid.

Spectral range 230-350 nm.

Absorption maximum At 254 nm.

Specific absorbance at the absorption maximum 170 to 190.

C. Infrared absorption spectrophotometry (2.2.24).

Comparison tropicamide CRS.

D. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of tropicamide CRS in methylene chloride R and dilute to 10 mL with the same solvent.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase concentrated ammonia R, methanol R, methylene chloride R (0.5:5:95 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the spot in the chromatogram obtained with the reference solution.

E. Dissolve about 5 mg in 3 mL of a mixture of 9 mL of acetic anhydride R, 1 mL of acetic acid R and 0.1 g of citric acid R. Heat on a water-bath for 5-10 min. A reddish-yellow colour is produced.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.1 g in ethanol (96 per cent) R and dilute to 10 mL with the same solvent.

Optical rotation (2.2.7)

-0.1° to +0.1°.

Dissolve 2.5 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 3 mL of acetonitrile R1 and dilute to 50.0 mL with water for chromatography R.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with water for chromatography R. Dilute 1.0 mL of this solution to 10.0 mL with water for chromatography R.

Reference solution (b) Dissolve 5 mg of 4-[(ethylamino)methyl]pyridine R (impurity A), 5.0 mg of tropicamide impurity C CRS and 5.0 mg of tropicamide impurity D CRS in 2 mL of acetonitrile R1 and dilute to 50.0 mL with water for chromatography R. Dilute 1.0 mL of this solution to 10.0 mL with water for chromatography R.

Reference solution (c) Dissolve 5 mg of tropicamide for peak identification CRS (containing impurity B) in 1.0 mL of acetonitrile R1 and dilute to 10.0 mL with water for chromatography R.

Reference solution (d) To 1 mL of reference solution (b) add 1 mL of reference solution (c).

Reference solution (e) Dilute 1.5 mL of reference solution (b) to 10.0 mL with water for chromatography R.

Column:

— size: l = 0.15 m, Ø = 4.6 mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (3 µm);

— temperature: 40 °C.

Mobile phase Dissolve 0.135 g of sodium dodecyl sulfate R and 3.4 mL of phosphoric acid R in 950 mL of water for chromatography R. Adjust to pH 3.0 with strong sodium hydroxide solution R and dilute to 1000 mL with water for chromatography R. Mix 73 volumes of this solution with 27 volumes of acetonitrile R1.

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 210 nm and at 254 nm.

Injection 15 µL.

Run time 3 times the retention time of tropicamide.

Identification of impurities Use the chromatogram obtained with reference solution (c) to identify the peak due to impurity B; use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, C and D.

Relative retention With reference to tropicamide (retention time = about 11 min): impurity C = about 0.4;

impurity A = about 0.5; impurity D = about 0.8;

impurity B = about 2.3.

System suitability Reference solution (d):

— resolution at 210 nm: minimum 2.0 between the peaks due

to impurities C and A;

— resolution at 210 nm: minimum 2.0 between the peaks due

Limits:

— the peak areas of the following impurities by the

corresponding correction factor: impurity A = 0.8;

impurity B = 0.6;

— impurity B at 254 nm: not more than 3 times the area of

the principal peak in the chromatogram obtained with

reference solution (a) (0.3 per cent);

— impurity A at 254 nm: not more than 1.5 times the area of

the principal peak in the chromatogram obtained with

reference solution (a) (0.15 per cent);

— impurity C at 210 nm: not more than the area of the

corresponding peak in the chromatogram obtained with

reference solution (e) (0.15 per cent);

— impurity D at 210 nm: not more than the area of the

corresponding peak in the chromatogram obtained with

reference solution (e) (0.15 per cent);

— unspecified impurities at 254 nm: for each impurity, not

more than the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.10 per cent);

— sum of impurities other than C and D at 254 nm: not more

than 5 times the area of the principal peak in the

chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit at 254 nm: 0.5 times the area of the

principal peak in the chromatogram obtained with

reference solution (a) (0.05 per cent).

Chlorides (2.4.4)

Maximum 100 ppm.

Dissolve 1.0 g with heating in 8 mL of acetic acid R, cool and

dilute to 10 mL with the same acid. Dilute 5 mL of this

solution to 15 mL with water R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 80 °C at a pressure not exceeding 0.7 kPa for 4 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 50 mL of anhydrous acetic acid R.

Add 0.2 mL of naphtholbenzenesulfonic acid solution R and titrate with

0.1 M perchloric acid until the colour changes from orange to

green.

1 mL of 0.1 M perchloric acid is equivalent to 28.44 mg of

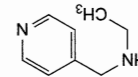
$C_{17}H_{20}N_2O_2$.

STORAGE

Protected from light.

IMPURITIES

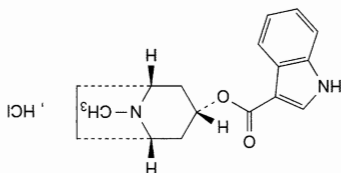
Specified impurities A, B, C, D



A. N-(pyridin-4-ylmethyl)ethanamine,

Tropisetron Hydrochloride

(Ph. Eur. monograph 2102)



$C_{17}H_{21}ClN_2O_2$ 320.8

105826-92-4

Action and use

Serotonin 5HT₃ receptor antagonist; antiemetic.

Ph Eur

DEFINITION

Hydrochloride of (1R,3r,5S)-8-methyl-8-azabicyclo[3.2.1]oct-

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble or soluble in water, sparingly soluble in

ethanol (96 per cent), very slightly soluble in methylene

chloride.

IDENTIFICATION

First identification B, D.

Second identification A, C, D.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50 mg in methanol R and dilute to

25.0 mL with the same solvent. Dilute 1.0 mL of this

solution to 100.0 mL with methanol R.

Spectral range 220-360 nm.

Absorption maxima At 228 nm and 282 nm.

Absorbance ratio A_{228}/A_{282} = 1.3 to 1.4.

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 5 mg of the substance to be examined in a mixture of equal volumes of methanol R and methylene chloride R and dilute to 5 mL with the same mixture of solvents.

Reference solution Dissolve 5 mg of tropisetron hydrochloride CRS in a mixture of equal volumes of methanol R and dilute to 5 mL with the same mixture of solvents.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase anhydrous formic acid R, water R, methanol R, methylene chloride R (2:2:30:70 V/V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In cold air.

Detection A Examine in ultraviolet light at 254 nm.

Detection B Spray first with a solution prepared as follows: dissolve 0.85 g of bisulfite sodium R in a mixture of 10 mL of acetic acid R and 40 mL of water R; to 5 mL of this solution add 5 mL of a 400 g/L solution of potassium iodide R and dilute to 100 mL with water R. Then spray with strong hydrogen peroxide solution R.

Results For both detection methods, the principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

Dissolve 1.00 g in water R and dilute to 20 mL with the same solvent.

Impurity A

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.200 g of the substance to be examined in a mixture of equal volumes of methanol R and methylene chloride R and dilute to 5.0 mL with the same mixture of solvents.

Reference solution (a) Dissolve 5.0 mg of tropine CRS (impurity A) in a mixture of equal volumes of methanol R and methylene chloride R and dilute to 25.0 mL with the same mixture of solvents.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with a mixture of equal volumes of methanol R and methylene chloride R. To 0.1 mL of this solution add 1.0 mL of reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ammonia R, methanol R, methylene chloride R (5:40:60 V/V/V).

Application 10 µL.

Development Over 2/3 of the plate.

Drying In a current of cold air.

Detection Dip the plate in potassium iodobismuthate solution R₁.

System suitability: reference solution (b):

the chromatogram shows 2 clearly separated spots.

N,N'-Dimethylaniline

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Limit:

— impurity A: any spot due to impurity A is not more intense than the spot in the chromatogram obtained with reference solution (a) (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 0.100 g of the substance to be examined in mobile phase A and dilute to 20.0 mL with mobile phase A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with mobile phase A. Dilute 1.0 mL of this solution to 10.0 mL with mobile phase A.

Reference solution (b) Dissolve 5.0 mg of tropisetron impurity B CRS and 5 mg of ethyl indole-3-carboxylate CRS in mobile phase A and dilute to 20.0 mL with mobile phase A.

Dilute 1.0 mL of this solution to 50.0 mL with mobile phase A.

Column:

size: l = 0.25 m, Ø = 4.6 mm;

stationary phase: octylsilyl silica gel for chromatography R

Mobile phase:

mobile phase A: methylaniline R, acetonitrile R, water R, methanol R (0.3:35:400:565 V/V/V/V);

mobile phase B: methylaniline R, acetonitrile R, water R, methanol R (0.3:100:100:800 V/V/V/V);

Time (min)

Mobile phase A (per cent V/V)

Mobile phase B (per cent V/V)

0 - 14

14 - 32

32 - 36

36 - 37

37 - 52

0 - 100

100 - 100

0 - 100

100 - 100

0 - 100

100 - 100

0 - 100

100 - 100

0 - 100

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100 - 100

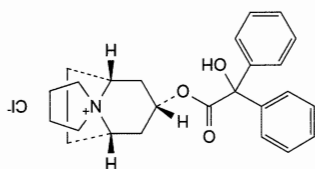
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100 - 100

0 - 100

Trospium Chloride

(Ph. Eur. monograph 1798)

 $C_{25}H_{30}ClNO_3$ 428.0

10405-02-4

Action and use

Anticholinergic.

Preparations

Prolonged-release Trospium Chloride Capsules

Trospium Chloride Tablets

Ph Eur

DEFINITION

(1*R*,3*r*,5*S*)-3-[(Hydroxydiphenylacetyl)oxy]spiro[8-azoniabicyclo[3.2.1]octane-8,1'-pyrrolidinium] chloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Very soluble in water, freely soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison trospium chloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 3.0 g in carbon dioxide-free water R and dilute to 30 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution B₇ (2.2.2, Method II).

pH (2.2.3)

5.0 to 7.0.

Dilute 5 mL of solution S to 50 mL with carbon dioxide-free water R.

Impurity C

Thin-layer chromatography (2.2.27).
Test solution Dissolve 0.20 g of the substance to be examined in 2.0 mL of methanol R.
Reference solution (a) Dissolve 1.0 mg of trospium impurity C CRS in 2.0 mL of methanol R.
Reference solution (b) Dilute 1.0 mL of test solution to 10.0 mL with methanol R. To 50 µL of this solution add 1 mL of reference solution (a).

Plate TLC silica gel plate R.

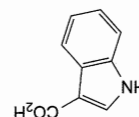
Mobile phase glacial acetic acid R, hydrochloric acid R, acetonitrile R (1:3.5:45 V/V/V).

Application 10 µL as bands.

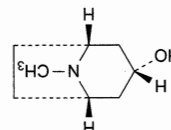
Development Over 2/3 of the plate.

Drying In a current of warm air until the odour of acetic acid is no longer perceptible.

Ph Eur

B. 1*H*-indole-3-carboxylic acid.

(tropine),

A. (1*R*,3*r*,5*S*)-8-methyl-8-azabicyclo[3.2.1]octan-3-ol

Specified impurities A, B.

IMPURITIES

of C₁₇H₂₁ClN₂O₂.

1 mL of 0.1 M perchloric acid is equivalent to 32.08 mg

acid, determining the end-point potentiometrically (2.2.20).
70 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).
Dissolve 0.250 g in 10 mL of anhydrous acetic acid R and add

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 0.3 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Loss on drying (2.2.32)

reference solution (20 ppm).

principal peak in the chromatogram obtained with the — N,N-dimethylamine: not more than the area of the

Limit:

Injection 20 µL.

Detection Spectrophotometer at 248 nm.

Flow rate 1 mL/min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	100	0
10 - 11	100 → 0	0 → 100
11 - 30	0	100
30 - 31	0 → 100	100 → 0
31 - 50	100	0

— mobile phase B: triethylamine R, acetonitrile R, water R; methanol R (0.3:100:100:800 V/V/V/V);

— mobile phase A: triethylamine R, acetonitrile R, water R; methanol R (0.3:35:400:565 V/V/V/V);

Mobile phase:

(5 µm).

— stationary phase: octylsilyl silica gel for chromatography R

— size: l = 0.25 m, Ø = 4.6 mm;

Column:

mobile phase A.

phase A. Dilute 1.0 mL of this solution to 100.0 mL with mobile in mobile phase A and dilute to 100.0 mL with mobile

Reference solution Dissolve 10.0 mg of N,N-dimethylamine R

mobile phase A.

examined in mobile phase A and dilute to 5.0 mL with Test solution Dissolve 250 mg of the substance to be

Detection Spray with potassium iodobismuthate solution R and subsequently with a 5 g/L solution of sodium nitrite R.

System suitability: reference solution (b):

- the chromatogram shows 2 clearly visible and separated zones.

Limit:

- **impurity C:** any zone due to impurity C is not more intense than the zone in the chromatogram obtained with reference solution (a) (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 30.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 6.0 mg of trospium **impurity A** CRS in the mobile phase and dilute to 20.0 mL with the mobile phase.

Reference solution (b) Dissolve 7.5 mg of trospium **impurity B** CRS in the mobile phase and dilute to 5.0 mL with the mobile phase.

Reference solution (c) Dilute a mixture of 0.3 mL of the test solution, 1.0 mL of reference solution (a) and 1.0 mL of reference solution (b) to 100.0 mL with the mobile phase.

Column:

- size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
- stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m);
- temperature: 40 °C.

Mobile phase Mix 1 volume of triethylamine R and 3 volumes of phosphoric acid R with 700 volumes of water R and add 300 volumes of acetonitrile R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 215 nm.

Injection 20 μ L.

Run time 3 times the retention time of trospium.

Relative retention With reference to trospium (retention time = about 10 min): **impurity B** = about 0.7; **impurity A** = about 1.9.

System suitability: reference solution (c):

- resolution: minimum 3 between the peaks due to **impurity B** and trospium.

Limits:

- **impurity B:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.5 per cent);
- **impurity A:** not more than 3 times the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.3 per cent);
- **unspecified impurities:** for each impurity, not more than 0.2 times the area of the peak due to **impurity B** in the chromatogram obtained with reference solution (c) (0.10 per cent);
- **total:** not more than twice the area of the peak due to **impurity B** in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in 50 mL of water R. Titrate with 0.1 M silver nitrate, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M silver nitrate is equivalent to 42.80 mg of $C_{25}H_{30}ClNO_3$.

STORAGE

Protected from light.

IMPURITIES

Specified impurities: A, B, C.

A. hydroxydiphenylacetic acid (benzoic acid),

O=C(O)(c1ccccc1)c2ccccc2

B. (1R,3r,5S)-8-azabicyclo[3.2.1]oct-3-yl hydroxydiphenylacetate,

O=C(O)(c1ccccc1)[C@H]1CC[C@@H]2NCCC[C@H]12c3ccccc3

C. (1R,3r,5S)-3-hydroxyispiro[8-azoniabicyclo[3.2.1]octane,1'-pyrrolidinium],

[O-][N+]1CC[C@H]2C[C@H]1CC[C@H]2C3CCNCC3

Troxerutin

(Ph. Eur. monograph 2133)

OCC1=CC=C(OC2=CC=C(OC3=CC=C(O)C=C3O)C(=O)O2)C(=O)O1

$C_{33}H_{42}O_{19}$ 743

Action and use

Bioflavonoid.

DEFINITION

Mixture of O-hydroxyethylated derivatives of rutin containing minimum 80 per cent of 2-[3,4-bis(2-hydroxyethoxy)phenyl]-3-[[6-O-(6-deoxy- α -L-mannopyranosyl)- β -D-glucopyranosyl]oxy]-5-hydroxy-7-(2-hydroxyethoxy)-4H-1-benzopyran-4-one (tris(hydroxyethyl)rutin).

Content 95.0 per cent to 105.0 per cent (dried substance).

CHARACTERS

Appearance

Yellowish-green, crystalline powder, hygroscopic.

Solubility

Freely soluble in water, slightly soluble in ethanol (96 per cent) and practically insoluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison troxerutin CRS.

B. Examine the chromatograms obtained in the test for composition.

Results The principal peak in the chromatogram obtained with the test solution is similar in position and size to the principal peak in the chromatogram obtained with the reference solution (a).

TESTS

Composition

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 10.0 mg of the substance to be examined in the mobile phase, if necessary using an ultrasonic bath and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 10.0 mg of troxerutin CRS in the mobile phase, if necessary using an ultrasonic bath and dilute to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1 mL of reference solution (a) to 10 mL with the mobile phase. Dilute 1 mL of this solution to 100 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm,

— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase Mix 20 volumes of acetonitrile R and 80 volumes of a 15.6 g/L solution of sodium dihydrogen phosphate R

adjusted to pH 4.4 with dilute phosphoric acid R or dilute sodium hydroxide solution R.

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 350 nm.

Injection 10 μ L.

Run time Twice the retention time of the main compound of troxerutin (tris(hydroxyethyl)rutin).

Relative retention With reference to tris(hydroxyethyl)rutin (retention time = about 25 min): tetrakis(hydroxyethyl)rutin = about 0.5; mono(hydroxyethyl)rutin = about 0.8; bis(hydroxyethyl)rutin = about 1.1.

System suitability: reference solution (a): peak from the peak due to tris(hydroxyethyl)rutin; signal-to-noise ratio: minimum 10 for the principal peak in the chromatogram obtained with reference solution (b).

Limits:

— principal peak: minimum 80 per cent,

— any other peak: for each peak, maximum 5 per cent,

— disregard limit: area of the principal peak in the chromatogram obtained with reference solution (b).

Ethylene oxide

Head-space gas chromatography (2.2.28).

Test solution To 1.00 g of the substance to be examined in a vial, add 1.0 mL of water R. Mix to obtain a homogeneous solution. Heat at 70 °C for 45 min.

Reference solution To 1.00 g of the substance to be examined in a vial, add 50 μ L of ethylene oxide solution R4 and 950 μ L of water R and close tightly. Mix to obtain a homogeneous solution. Heat at 70 °C for 45 min.

Column:

— material: fused silica,

— size: $l = 30$ m, $\varnothing = 0.32$ mm,

— stationary phase:

poly(cyanopropyl) (7) (phenyl) (7) (methyl) (86) siloxane R (film thickness 1 μ m).

Carrier gas helium for chromatography R.

Flow rate 1.1 mL/min.

Static head-space conditions which may be used:

— equilibration temperature: 80 °C,

— equilibration time: 45 min,

— transfer line temperature: 110 °C,

— pressurisation time: 2 min,

— injection time: 12 s.

Temperature:

Time (min)	Temperature (°C)
0 - 5	40
5 - 18	40 \rightarrow 200
Injection port	150
Detector	250

Detection Flame ionisation.

Injection 1.0 mL.

The peak due to ethylene oxide is identified by injecting solutions of ethylene oxide of increasing concentration.

Determine the content of ethylene oxide (ppm) in the substance to be examined using the following expression:

$$\frac{A_1 \times m_1}{A_1 \times m_1 - (A_2 \times m_2) - (A_3 \times m_3)}$$

A_1 = area of the peak due to ethylene oxide in the chromatogram obtained with the test solution,
 A_2 = area of the peak due to ethylene oxide in the chromatogram obtained with the reference solution,
 m_1 = mass of ethylene oxide in the reference solution, in micrograms,
 m_2 = mass of the substance to be examined in the test solution, in grams,
 m_3 = mass of the substance to be examined in the reference solution, in grams.

Limit:

— ethylene oxide: maximum 1 ppm.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test F. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 5.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 4 h.

Sulfated ash (2.4.14) Maximum 0.4 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 100.0 mL of *water R*. Dilute 10.0 mL of this solution to 100.0 mL with *water R*. Dilute 10.0 mL to 100.0 mL with *water R*. Measure the absorbance (2.2.25) at the absorption maximum at 350 nm.

Calculate the percentage content of $C_{33}H_{42}O_{19}$ taking the specific absorbance to be 250.

STORAGE

In an airtight container, protected from light.

Ph Eur

Trypsin

(Ph. Eur. monograph 0694)

9002-07-7

Action and use
Proteolytic enzyme.

Ph Eur

DEFINITION

Trypsin is a proteolytic enzyme obtained by the activation of trypsinogen extracted from the pancreas of mammals. It has an activity of not less than 0.5 microkatal per milligram, calculated with reference to the dried substance. In solution, it has maximum enzymic activity at pH 8; the activity is reversibly inhibited at pH 3, the pH at which it is most stable.

PRODUCTION

The animals from which trypsin is derived must fulfil the requirements for the health of animals suitable for human consumption.

The method of manufacture is validated to demonstrate that the product, if tested, would comply with the following test.

Histamine (2.6.10)

Not more than 1 µg of histamine base per 0.2 microkatal of trypsin activity. Use a 10 g/L solution of the substance to be examined in 0.0015 M borate buffer solution pH 8.0 R inactivated by heating on a water-bath for 30 min. Carry out dilutions with a 9 g/L solution of sodium chloride R.

CHARACTERS

Appearance
White or almost white, crystalline or amorphous powder, hygroscopic if amorphous.

Solubility

Sparsingly soluble in water.

IDENTIFICATION

A. Dilute 1 mL of solution S (see Tests) to 100 mL with *water R*. In a depression in a white spot-plate, mix 0.1 mL of this solution with 0.2 mL of *tosylarginine methyl ester hydrochloride solution R*. A reddish-violet colour develops within 3 min.
B. Dilute 0.5 mL of solution S to 5 mL with *water R*. Add 0.1 mL of a 20 g/L solution of *tosyl-L-tyrosyl-chloromethane hydrochloride R*. Adjust to pH 7.0, shake for 2 h and dilute to 50 mL with *water R*. In one of the depressions of a white spot-plate, mix 0.1 mL of this solution with 0.2 mL of

ASSAY

The activity of trypsin is determined by comparing the rate at which it hydrolyses *benzoylarginine ethyl ester hydrochloride R* with the rate at which *trypsin BRP* hydrolyses the same substrate in the same conditions.
Apparatus Use a reaction vessel of about 30 mL capacity provided with:
— a device that will maintain a temperature of $25.0 \pm 0.1^\circ\text{C}$;
— a stirring device (for example, a magnetic stirrer);
— a lid with holes for the insertion of electrodes, the tip of a burette, a tube for the admission of nitrogen and the introduction of reagents.

An automatic or manual titration device may be used.

For the latter, the burette is graduated in 0.005 mL and the pH meter is provided with a wide-range scale and glass-calomel or glass-silver-silver chloride electrodes.
Test solution Dissolve sufficient of the substance to be examined in 0.001 M *hydrochloric acid* and dilute to 25.0 mL with the same acid in order to obtain a solution containing approximately 700 nanokatals per millilitre.

Reference solution Dissolve 25.0 mg of *trypsin BRP* in 0.001 M *hydrochloric acid* and dilute to 25.0 mL with the same acid. Store the solutions at $0-5^\circ\text{C}$. Warm 1 mL of each solution to about 25°C over 15 min and use 50 µL of each solution

Chymotrypsin

Test solution To 1.8 mL of buffer solution pH 8.0 R add 7.4 mL of *water R* and 0.5 mL of 0.2 M *acetyltirosine ethyl ester R*. While shaking the solution, add 0.3 mL of solution S and start a timer. After exactly 5 min, measure the pH and start a timer. After exactly 5 min, measure the pH (2.2.3).
Reference solution Prepare in the same manner as the test solution, replacing solution S by 0.3 mL of a 0.5 g/L solution of *chymotrypsin BRP*, and measure the pH (2.2.3) exactly 5 min after adding the chymotrypsin.
The pH of the test solution is higher than that of the reference solution.

Loss on drying (2.2.32)

Not more than 5.0 per cent, determined on 0.500 g by drying at 60°C at a pressure not exceeding 0.67 kPa for 2 h.

Microbial contamination

TAMC: acceptance criterion 10^4 CFU/g (2.6.12).
TYMC: acceptance criterion 10^2 CFU/g (2.6.12).

Absence of *Escherichia coli* (2.6.13).

Absence of *Salmonella* (2.6.13).

IDENTIFICATION

First identification A, B.
Second identification A, C, D.
 A. Specific optical rotation (see Tests).
 B. Infrared absorption spectrophotometry (2.2.24).
 C. Thin-layer chromatography (2.2.27).

Comparison tryptophan CRS.
Solvent mixture glacial acetic acid R, water R (50:50 V/V).
Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 50 mL with the solvent mixture.
Reference solution Dissolve 10 mg of tryptophan CRS in the solvent mixture and dilute to 50 mL with the solvent mixture.

Plate TLC silica gel plate R.
Mobile phase glacial acetic acid R, water R, butanol R (20:20:60 V/V/V).

Application 5 µL.
Development: Over 2/3 of the plate.
Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C for 15 min.
Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.
 D. Dissolve about 20 mg in 10 mL of water R. Add 5 mL of dimethylaminobenzaldehyde solution R6 and 2 mL of hydrochloric acid R1. Heat on a water-bath. A purple-blue colour develops.

TESTS

Appearance of solution
 The solution is clear (2.2.1) and not more intensely coloured than reference solution BY₆ (2.2.2, Method II).
 Dissolve 0.1 g in 1 M hydrochloric acid and dilute to 10 mL with the same acid.

Specific optical rotation (2.2.7)

–33.0 to –30.0 (dried substance).
 Dissolve 0.25 g in water R, heating on a water-bath if necessary, and dilute to 25.0 mL with the same solvent.

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method 1. The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.
Solution A dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.
Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.
Reference solution (b) Dissolve 30.0 mg of proline R in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

for each titration. Carry out the titration in an atmosphere of nitrogen. Transfer 10.0 mL of 0.0015 M borate buffer solution pH 8.0 R to the reaction vessel and, while stirring, add 1.0 mL of a freshly prepared 6.86 g/L solution of benzoylarginine ethyl ester hydrochloride R. When the temperature is steady at 25.0 ± 0.1 °C (after about 5 min) adjust to pH 8.0 exactly with 0.1 M sodium hydroxide. Add 50 µL of the test solution and start a timer. Maintain at pH 8.0 by the addition of 0.1 M sodium hydroxide, the tip of the microburette being immersed in the solution; note the volume added every 30 s. Follow the reaction for 8 min. Calculate the volume of 0.1 M sodium hydroxide used per second. Carry out a titration in the same manner using the reference solution and calculate the volume of 0.1 M sodium hydroxide used per second.
 Calculate the activity in microkatal per milligram using the following expression:

$$\frac{m' \times V}{m \times V'} \times A$$

m = mass of the substance to be examined, in milligrams;
m' = mass of tryptophan BRR, in milligrams;
V = volume of 0.1 M sodium hydroxide used per second by the test solution;
V' = volume of 0.1 M sodium hydroxide used per second by the reference solution;
A = activity of tryptophan BRR, in microkatal per milligram.
STORAGE
 In an airtight container, protected from light, at a temperature of 2 °C to 8 °C.

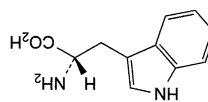
LABELLING

The label states:
 — the activity in microkatal per milligram;
 — for the amorphous substance, that it is hygroscopic.

Ph Eur

Tryptophan

(Ph. Eur. monograph 1272)



73-22-3

204.2

C₁₁H₁₂N₂O₂

Ph Eur

DEFINITION

(2S)-2-Amino-3-(1H-indol-3-yl)propanoic acid.
 Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline or amorphous powder.

Solubility

Sparingly soluble in water, slightly soluble in ethanol (96 per cent). It dissolves in dilute solutions of mineral acids and alkali hydroxides.

Reference solution (c) Dilute 6.0 mL of ammonium standard solution (100 ppm NH_4) R to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A. **Reference solution (d)** Dissolve 30 mg of isoleucine R and 30 mg of leucine R in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Blank solution Solution A. Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids. **System suitability** Reference solution (d): **Resolution:** minimum 1.5 between the peaks due to isoleucine and leucine. **Calculation of percentage contents:**

— for any ninhydrin-positive substance detected at 570 nm, use the concentration of tyryptophan in reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits: — any ninhydrin-positive substance: for each impurity, maximum 0.2 per cent;

— total: maximum 0.5 per cent;

— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Impurity A and other related substances

Liquid chromatography (2.2.29). Prepare the standard, test and reference solutions immediately before use.

Buffer solution pH 2.3 Dissolve 3.90 g of sodium dihydrogen phosphate R in 1000 mL of water R. Add about 700 mL of a 2.9 g/L solution of phosphoric acid R and adjust to pH 2.3 with the same acid solution.

Solvent mixture acetonitrile R, water R (10:90 V/V). **Standard solution** Dissolve 10.0 mg of N-acetyltyryptophan R in the solvent mixture and dilute to 100.0 mL with the solvent mixture. Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

Test solution (a) Dissolve 0.10 g of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.

Test solution (b) Dissolve 0.10 g of the substance to be examined in the standard solution and dilute to 10.0 mL with the standard solution.

Reference solution (a) Dissolve the contents of a vial of 1,1'-ethyldenebis(tryptophan CRS (impurity A) in 1.0 mL of the solvent mixture.

Reference solution (b) Dissolve the contents of a vial of 1,1'-ethyldenebis(tryptophan CRS (impurity A) in 1.0 mL of the standard solution.

Column: — size: $l = 0.25$ m, $\phi = 4.6$ mm; — stationary phase: octadecylsilyl silica gel for chromatography R (5 μm); — temperature: 40 °C.

Mobile phase: — mobile phase A: acetonitrile R, buffer solution pH 2.3 (115:885 V/V); — mobile phase B: acetonitrile R, buffer solution pH 2.3 (350:650 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 10	100	0
10 - 45	100 \rightarrow 0	0 \rightarrow 100
45 - 65	0	100

Flow rate 0.7 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 20 μL of test solutions (a) and (b) and reference solutions (b) and (c).

Retention time Tyryptophan = about 8 min; N-acetyltyryptophan = about 29 min; impurity A = about 34 min.

System suitability: — resolution: minimum 8.0 between the peaks due to N-acetyltyryptophan and impurity A in the chromatogram obtained with reference solution (b); if necessary, adjust the time programme for the elution gradient (an increase in the duration of elution with mobile phase A produces longer retention times and a better resolution);

— signal-to-noise ratio: minimum 15 for the principal peak in the chromatogram obtained with reference solution (c);

— symmetry factor: maximum 3.5 for the peak due to impurity A in the chromatogram obtained with reference solution (b);

— in the chromatogram obtained with test solution (a) there is no peak with the same retention time as N-acetyltyryptophan (in such case correct the area of the N-acetyltyryptophan peak).

Limits: test solution (b): — impurity A: not more than 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (10 ppm);

— sum of impurities with a retention time less than that of tyryptophan: not more than 0.6 times the area of the peak due to N-acetyltyryptophan in the chromatogram obtained with reference solution (b) (100 ppm);

— sum of impurities with a retention time greater than that of tyryptophan and up to 1.8 times the retention time of N-acetyltyryptophan: not more than 1.9 times the area of the peak due to N-acetyltyryptophan in the chromatogram obtained with reference solution (b) (300 ppm);

— disregard limit: 0.02 times the area of the peak due to N-acetyltyryptophan in the chromatogram obtained with reference solution (b); disregard the peak due to N-acetyltyryptophan.

Chlorides (2.4.4)

Dissolve 0.25 g in 3 mL of dilute nitric acid R and dilute to Maximum 200 ppm. 15 mL with water R. The solution, without any further addition of nitric acid, complies with the test.

Sulfates (2.4.13)

Maximum 300 ppm. Dissolve 0.5 g in a mixture of 5 volumes of dilute hydrochloric acid R and 25 volumes of distilled water R, and dilute to 15 mL with the same mixture of solvents.

Ammonium

Amino acid analysis (2.2.56) as described in the test for ninhydrin-positive substances with the following modifications.

Injection: test solution, reference solution (c) and blank solution.

Limit:

— ammonium at 570 nm: not more than the area of the

corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram

obtained with the blank solution.

Iron (2.4.9)

Maximum 20 ppm.

In a separating funnel, dissolve 0.50 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min each time.

To the combined organic layers add 10 mL of water R and shake for 3 min. Examine the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

2.0 g complies with test D. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 3 mL of anhydrous formic acid R.

Add 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically

1 mL of 0.1 M perchloric acid is equivalent to 20.42 mg of $C_{11}H_{12}N_2O_2$.

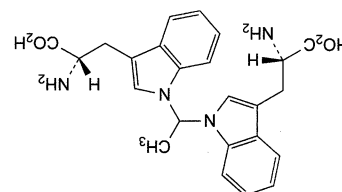
STORAGE

Protected from light.

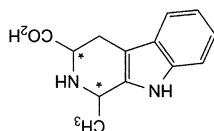
IMPURITIES

Specified impurities A.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E, F, G, H, I, J, K, L.

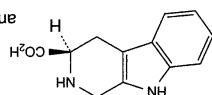


A. 3,3'-[ethane-1,1-diybis(1H-indol-1,3-diyl)]bis[(2S)-2-aminopropanoic acid] (ethane-1,1-diybis(tryptophan))

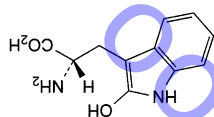


I. 1-methyl-1,2,3,4-tetrahydro-9H-pyrido[3,4-b]indole-3-carboxylic acid,

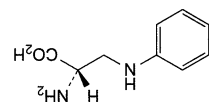
H. (3R)-1,2,3,4-tetrahydro-9H-pyrido[3,4-b]indole-3-carboxylic acid,



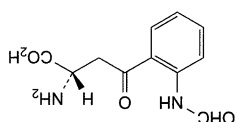
G. (2S)-2-amino-3-(2-hydroxy-1H-indol-3-yl)propanoic acid (2-hydroxytryptophan),



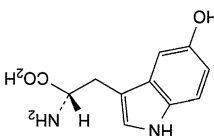
F. (2S)-2-amino-3-(phenylamino)propanoic acid (3-phenylalanine),



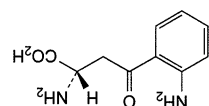
E. (2S)-2-amino-4-[2-(formylamino)phenyl]-4-oxobutanoic acid (N-formylkynurenine),



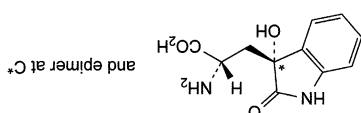
D. (2S)-2-amino-3-(5-hydroxy-1H-indol-3-yl)propanoic acid (5-hydroxytryptophan),



C. (2S)-2-amino-4-(2-aminophenyl)-4-oxobutanoic acid (kynurenine),



B. (2S)-2-amino-3-[(3R)-3-hydroxy-2-oxo-2,3-dihydro-1H-indol-3-yl]propanoic acid (dioxindolylalanine),



IDENTIFICATION

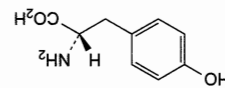
- First identification A, B, C, D, E.**
A. Specific optical rotation (see Tests).
B. Infrared absorption spectrophotometry (2.2.24).
C. Thin-layer chromatography (2.2.27).
Test solution Dissolve 10 mg of the substance to be examined in 1 mL of dilute ammonia R2 and dilute to 50 mL with water R.
- Reference solution.** Dissolve 10 mg of tyrosine CRS in 1 mL of dilute ammonia R2 and dilute to 50 mL with water R.
- Plate** TLC silica gel plate R.
- Mobile phase** concentrated ammonia R1, propanol R (30:70 V/V).
- Application** 5 µL.
- Development** Over 2/3 of the plate.
- Drying** In air.
- Detection** Spray with ninhydrin solution R and heat at 105 °C for 15 min.
- Results** The principal spot in the chromatogram obtained with the Test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.
- D.** To about 50 mg add 1 mL of dilute nitric acid R. A dark red colour is produced within 15 min.
- E.** Dissolve about 30 mg in 2 mL of dilute sodium hydroxide solution R. Add 3 mL of a freshly prepared mixture of equal volumes of a 100 g/L solution of sodium nitrite R and a solution of 0.5 g of sulfanilic acid R in a mixture of 6 mL of hydrochloric acid R1 and 94 mL of water R. An orange-red colour is produced.

TESTS

- Appearance of solution**
 The solution is clear (2.2.1) and not more intensely coloured than reference solution Y7 (2.2.2, Method II).
- Dissolve 0.5 g in dilute hydrochloric acid R and dilute to 20 mL with the same acid.
- Specific optical rotation (2.2.7)**
 –12.3 to –11.0 (dried substance).
- Dissolve 1.25 g in a mixture of equal volumes of dilute hydrochloric acid R and water R and dilute to 25.0 mL with the same mixture of solvents.
- Ninhydrin-positive substances**
 Amino acid analysis (2.2.56). For analysis, use Method 1. The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.
- Solution A** dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.
- Test solution** Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.
- Reference solution (a)** Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.
- Reference solution (b)** Dissolve 30.0 mg of phenylalanine R (impurity A) in solution A and dilute to 100.0 mL with

Tyrosine

(Ph. Eur. monograph 1161)



C₉H₉NO₃ 181.2 60-18-4

Action and use
 Amino acid.

DEFINITION

(2S)-2-Amino-3-(4-hydroxyphenyl)propanoic acid.
 Fermentation product, extract or hydrolysate of protein.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance
 White or almost white, crystalline powder or colourless crystals.

Solubility
 Very slightly soluble in water, practically insoluble in ethanol (96 per cent). It dissolves in dilute mineral acids and in dilute solutions of alkali hydroxides.

of methyl isobutyl ketone R₁, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

0.5 g complies with test G. Prepare the reference solution using 0.5 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.150 g in 5 mL of anhydrous formic acid R.

Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 18.12 mg of C₉H₁₁NO₃.

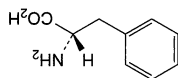
STORAGE

Protected from light.

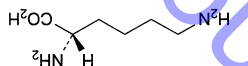
IMPURITIES

Specified impurities A.

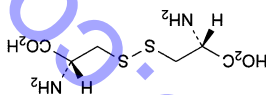
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C.



A. (2S)-2-amino-3-phenylpropanoic acid (phenylalanine).



B. (2S)-2,6-diaminohexanoic acid (lysine).



C. 3,3'-disulfanedibis[(2R)-2-aminopropanoic acid] (cystine).

solution A. Dilute 1.0 mL of the solution to 250.0 mL with

Reference solution (c) Dissolve 30.0 mg of proline R in

1.0 mL of the solution to 250.0 mL with solution A. Dilute

Reference solution (d) Dilute 6.0 mL of ammonium standard

solution (100 ppm NH₄) R to 50.0 mL with solution A. Dilute

1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (e) Dissolve 30 mg of isoleucine R and

30 mg of leucine R in solution A and dilute to 50.0 mL with

solution A. Dilute 1.0 mL of the solution to 200.0 mL with

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference

solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids.

System suitability: reference solution (e):

— resolution: minimum 1.5 between the peaks due to

isoleucine and leucine.

Calculation of percentage contents:

— for impurity A, use the concentration of impurity A in

reference solution (b);

— for any ninhydrin-positive substance detected at 570 nm,

use the concentration of tyrosine in reference solution (a);

— for any ninhydrin-positive substance detected at 440 nm,

use the concentration of proline in reference solution (c);

if a peak is above the reporting threshold at both

wavelengths, use the result obtained at 570 nm for

quantification.

— impurity A at 570 nm: maximum 0.5 per cent;

— any ninhydrin-positive substance: for each impurity,

maximum 0.2 per cent;

— total: maximum 0.6 per cent;

— reporting threshold: 0.05 per cent.

The thresholds indicated under Related substances

(Table 2034.-1) in the general monograph Substances for

pharmaceutical use (2034) do not apply.

Chlorides (2.4.4)

Maximum 200 ppm.

Dissolve 0.25 g in 3 mL of dilute nitric acid R and dilute to

15 mL with water R. The solution, without further addition

of nitric acid, complies with the test.

Sulfates (2.4.13)

Maximum 300 ppm.

Dissolve, with gentle heating, 0.5 g in 5 mL of dilute

hydrochloric acid R and dilute to 15 mL with distilled water R.

Ammonium

Amino acid analysis (2.2.56) as described in the test for

ninhydrin-positive substances with the following

modifications.

Injection Test solution, reference solution (d) and blank

solution.

Limit:

— ammonium at 570 nm: not more than the area of the

corresponding peak in the chromatogram obtained with

reference solution (d) (0.02 per cent), taking into account

the peak due to ammonium in the chromatogram

obtained with the blank solution.

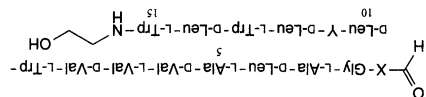
Iron (2.4.9)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of dilute

hydrochloric acid R. Shake with 3 quantities, each of 10 mL,

(Ph. Eur. monograph 1662)



of ethanol (96 per cent) R.
 Plate TLC silica gel F₂₅₄ plate R.
 Mobile phase methanol R, butanol R, water R, acetic acid R,
 butyl acetate R (2.5:7.5:12.20:40 V/V/V/V/V).
 Application 1 µL.
 Development Over 2/3 of the plate.
 Drying In a current of warm air.
 Detection A Examine in ultraviolet light at 254 nm.

Loss on drying (2.2.32)

Maximum 4.0 per cent, determined on 1.000 g by drying under high vacuum at 60 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 1.5 per cent, determined on 1.0 g.

ASSAY

Carry out the microbiological assay of antibiotics (2.7.2)

using the turbidimetric method. Use *gramicidin CRS* as the reference substance.

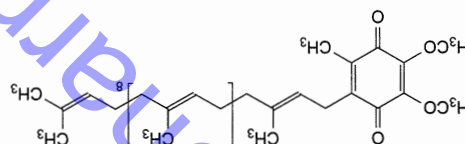
Test solution Prepare a solution of tyrothricin containing about the same amount of gramicidin as the corresponding solution of *gramicidin CRS* i.e. 5 times more concentrated.

STORAGE

In an airtight container, protected from light.

Ubidecarenone

(Ph. Eur. monograph 1578)



$C_{59}H_{90}O_4$ 863 303-98-0

Action and use

Co-enzyme in mitochondrial electron transport.

DEFINITION

2-[(all-*E*)-3,11,15,19,23,27,31,35,39-Decamethyltetraconta-2,6,10,14,18,22,26,30,34,38-decayn-5,6-dimethoxy-3-methyl]benzene-1,4-dione.

Content

97.0 per cent to 103.0 per cent.

CHARACTERS**Appearance**

Yellow or orange, crystalline powder.

Solubility

Practically insoluble in water, soluble in acetone, very slightly soluble in ethanol.

It gradually decomposes and darkens on exposure to light.

mp: about 48 °C.

Carry out all operations avoiding exposure to light.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison *ubidecarenone CRS*.

B. Examine the chromatograms obtained in the test for related substances.

Results The retention time of the principal peak in the chromatogram obtained with the test solution is similar to that of the principal peak in the chromatogram obtained with reference solution (a).

TESTS**Related substances**

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be

examined in 25.0 mL of ethanol R by heating at about 50 °C

for 2 min. Allow to cool.

Reference solution (a) Dissolve 5 mg of *ubidecarenone CRS* in

5 mL of ethanol R by heating at about 50 °C for 2 min.

Allow to cool.

Reference solution (b) Dissolve 2 mg of *ubidecarenone*

impurity D CRS in 2 mL of the test solution by heating at about 50 °C for 2 min. Allow to cool. Dilute 1 mL to 50 mL

with ethanol R.

Reference solution (c) Dilute 1.0 mL of the test solution to

100.0 mL with ethanol R.

Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm,

— stationary phase: octadecylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase ethanol R, methanol R2 (20:80 V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 10 μ L.

Run time 2 times the retention time of *ubidecarenone*.

Relative retention With reference to *ubidecarenone* (retention time = about 12 min): impurity D = about 0.67.

System suitability: reference solution (b):

— resolution: minimum 6.5 between the peaks due to

impurity D and to *ubidecarenone*.

Limits:

— any impurity: not more than half the area of the principal peak in the chromatogram obtained with reference

solution (c) (0.5 per cent),

— total: not more than the area of the principal peak in the

chromatogram obtained with reference solution (c)

(1.0 per cent),

— disregard limit: 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (c)

(0.05 per cent).

Impurity F

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be

examined in 25.0 mL of hexane R.

Reference solution (a) Dissolve the contents of a vial of

ubidecarenone for system suitability CRS in 1 mL of hexane R.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with hexane R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm,

— stationary phase: silica gel for chromatography R (7 μ m).

Mobile phase ethyl acetate R, hexane R (3:97 V/V).

Flow rate 2 mL/min.

Detection Spectrophotometer at 275 nm.

Injection 20 μ L.

Run time 1.2 times the retention time of *ubidecarenone*.

Relative retention With reference to *ubidecarenone* (retention

time = about 10 min): impurity F = about 0.85.

System suitability: reference solution (a):

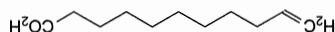
— resolution: minimum 1.5 between the peaks due to

impurity F and to *ubidecarenone*.



Undecenoic Acid

(Undecylenic Acid, Ph Eur monograph 0461)



$C_{11}H_{20}O_2$ 184.3 112-38-9 Ph Eur

DEFINITION

Undec-10-enoic acid.

Content

97.0 per cent to 102.0 per cent.

CHARACTERS

Appearance

White or very pale yellow, crystalline mass or colourless or

Solubility

Practically insoluble in water, freely soluble in ethanol

(96 per cent) and in fatty and essential oils.

IDENTIFICATION

A. Refractive index (2.2.6): 1.447 to 1.450, determined at

25 ± 0.5 °C.

B. Freezing point (2.2.18): 21 °C to 24 °C.

C. To 2.0 g add 2 mL of freshly distilled aniline R and boil

under a reflux condenser for 10 min. Allow to cool and add

30 mL of ether R. Shake with 3 quantities, each of 20 mL, of

dilute hydrochloric acid R and then with 20 mL of water R.

Evaporate the organic layer to dryness on a water-bath. After

recrystallising twice from ethanol (70 per cent V/V) R and

drying *in vacuo* for 3 h, the residue melts (2.2.14) at 66 °C to

68 °C.

D. Dissolve 0.1 g in a mixture of 2 mL of dilute sulfuric

acid R and 5 mL of glacial acetic acid R. Add dropwise

0.25 mL of potassium permanganate solution R. The colour of

the potassium permanganate is discharged.

TESTS

Peroxide value (2.5.5, Method A)

Maximum 10.

Fixed and mineral oils

To 1.0 g add 5 mL of sodium carbonate solution R and 25 mL

of water R and boil for 3 min. The hot solution is not more

opalescent than reference suspension II (2.2.1).

Water-soluble acids

To 1.0 g add 20 mL of water R heated to 35–45 °C and

shake for 2 min. Cool and filter the aqueous layer through a

moistened filter. To 10 mL of the filtrate add 0.1 mL of

phenolphthalein solution R. Not more than 0.1 mL of 0.1 M

sodium hydroxide is required to change the colour of the

indicator.

Degree of unsaturation

Dissolve 85.0 mg in a mixture of 5 mL of dilute hydrochloric

acid R and 30 mL of glacial acetic acid R. Using 0.05 mL of

indigo carmine solution R1 as indicator, added towards the end

of the titration, titrate with 0.0167 M bromide-bromate until

the colour changes from blue to yellow. 8.9 mL to 9.4 mL of

0.0167 M bromide-bromate is required. Carry out a blank

titation.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 0.50 g.

Specified impurities: A, B, C, D, E, F.

IMPURITIES

Store in an airtight container, protected from light.

STORAGE

taking the specific absorbance to be 169.

the maximum at 275 nm. Calculate the content of $C_{59}H_{90}O_4$

50.0 mL with ethanol R. Measure the absorbance (2.2.25) at

50.0 mL with ethanol R. Dilute 2.0 mL of the solution to

Dissolve 50.0 mg in 1.0 mL of hexane R and dilute to

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

reference solution (b) (0.5 per cent).

principal peak in the chromatogram obtained with

— impurity F: not more than 0.5 times the area of the

Limit:

(Z)-isomer).

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

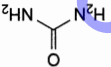
F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

dimethoxy-3-methylbenzene-1,4-dione (ubidecarenone

2,6,10,14,18,22,26,30,34,38-tetracontadecacyl]-5,6-

3,7,11,15,19,23,27,31,35,39-decamethyl-

F. 2-[(2Z,6E,10E,14E,18E,22E,26E,30E,34E,38E)-

<p>ASSAY</p> <p>Dissolve 0.750 g in 10 mL of ethanol (96 per cent) R. Titrate with 0.5 M sodium hydroxide using 0.1 mL of phenolphthalein solution R as indicator, until a pink colour is obtained.</p> <p>1 mL of 0.5 M sodium hydroxide is equivalent to 92.14 mg of $C_{11}H_{20}O_2$.</p> <p>STORAGE</p> <p>In a non-metallic container, protected from light.</p>	<p>Urea</p> <p>(Ph. Eur. monograph 0743)</p>  <p>CH_4N_2O</p> <p>60.1</p> <p>57-13-6</p>	<p>Alkalinity</p> <p>To 2.5 mL of solution S add 7.5 mL of water R, 0.1 mL of methyl red solution R and 0.4 mL of 0.01 M hydrochloric acid. The solution is red to orange.</p> <p>Buret</p> <p>Maximum 0.1 per cent.</p> <p>To 10 mL of solution S add 5 mL of water R, 0.5 mL of a 5 g/L solution of copper sulfate R and 0.5 mL of strong sodium hydroxide solution R. Allow to stand for 5 min. Any reddish-violet colour in the solution is not more intense than that in a standard prepared at the same time in the same manner using 10 mL of a 0.2 g/L solution of buret R.</p> <p>Ammonium (2.4.1)</p> <p>Maximum 500 ppm, determined on 0.1 mL of solution S.</p> <p>Heavy metals (2.4.8)</p> <p>Maximum 10 ppm.</p> <p>Dilute 10 mL of solution S to 20 mL with water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.</p> <p>Loss on drying (2.2.32)</p> <p>Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.</p> <p>Sulfated ash (2.4.14)</p> <p>Maximum 0.1 per cent, determined on 1.0 g.</p> <p>ASSAY</p> <p>Dissolve 0.2000 g in water R and dilute to 50.0 mL with the same solvent. Introduce 1.0 mL of the solution into a combustion flask. Add 4 g of a powdered mixture of 100 g of dipotassium sulfate R, 5 g of copper sulfate R and 2.5 g of selenium R, and 3 glass beads. Wash any adhering particles from the neck into the flask with 5 mL of sulfuric acid R, allowing it to run down the sides of the flask, and mix the contents by rotation. Close the mouth of the flask loosely, for example by means of a glass bulb with a short stem, to avoid excessive loss of sulfuric acid. Heat gradually at first, then increase the temperature until there is vigorous boiling with condensation of sulfuric acid in the neck of the flask; take precautions to prevent the upper part of the flask from becoming overheated. Continue the heating for 30 min. Cool, dissolve the solid material by cautiously adding to the mixture 25 mL of water R, cool again and place in a steam-distillation apparatus. Add 30 mL of strong sodium hydroxide solution R and distil immediately by passing steam through the mixture. Collect the distillate in 15 mL of a 40 g/L solution of boric acid R to which has been added 0.2 mL of methyl red mixed solution R and enough water R to cover the tip of the condenser. Towards the end of the distillation, lower the receiver so that the tip of the condenser is above the surface of the acid. Take precautions to prevent any water on the outer surface of the condenser from reaching the contents of the receiver. Titrate the distillate with 0.01 M sulfuric acid.</p> <p>1 mL of 0.01 M sulfuric acid is equivalent to 0.6006 mg of CH_4N_2O.</p> <p>STORAGE</p> <p>In an airtight container.</p>	<p>Action and use</p> <p>Keratolytic.</p> <p>Preparation</p> <p>Urea Cream</p> <p>Ph Eur</p> <p>DEFINITION</p> <p>Carbamide.</p> <p>Content</p> <p>98.5 per cent to 101.5 per cent (dried substance).</p> <p>CHARACTERS</p> <p>Appearance</p> <p>White or almost white, crystalline powder or transparent crystals, slightly hygroscopic.</p> <p>Solubility</p> <p>Very soluble in water, soluble in ethanol (96 per cent), practically insoluble in methylene chloride.</p> <p>IDENTIFICATION</p> <p>First identification A, B</p> <p>Second identification A, C, D</p> <p>A. Melting point (2.2.14): 132 °C to 135 °C.</p> <p>B. Infrared absorption spectrophotometry (2.2.24).</p> <p>Comparison urea CRS.</p> <p>C. Dissolve 0.1 g in 1 mL of water R. Add 1 mL of nitric acid R. A white, crystalline precipitate is formed.</p> <p>D. Heat 0.5 g in a test tube until it liquefies and the liquid becomes turbid. Cool, dissolve in a mixture of 1 mL of dilute sodium hydroxide solution R and 10 mL of water R and add 0.05 mL of copper sulfate solution R. A reddish-violet colour is produced.</p> <p>TESTS</p> <p>Solution S</p> <p>Dissolve 10.0 g in water R and dilute to 50 mL with the same solvent.</p> <p>Appearance of solution</p> <p>The solution is clear (2.2.1) and colourless (2.2.2, Method II).</p> <p>To 2.5 mL of solution S add 7.5 mL of water R.</p>
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Ph Eur

Urofollitropin

(Ph. Eur. monograph 0958)

97048-13-0



Action and use

Follicle-stimulating hormone.

Preparation

Urofollitropin Injection

Ph Eur

DEFINITION

Urofollitropin is a dry preparation containing menopausal gonadotrophin obtained from the urine of post-menopausal women. It has follicle-stimulating activity and no or virtually no luteinising activity. The potency is not less than 90 International Units of follicle-stimulating hormone (hFSH) per milligram. The ratio of units of luteinising hormone (interstitial-cell-stimulating hormone) [LH(ICSH)] to units of follicle-stimulating hormone is not more than 1/60.

PRODUCTION

It may be prepared by a suitable fractionation procedure followed by immunoadfinity chromatography.

CHARACTERS

Appearance

Almost white or slightly yellowish powder.

Solubility

Soluble in water.

IDENTIFICATION

When administered as prescribed in the assay it causes enlargement of the ovaries of immature female rats.

TESTS

Hepatitis virus antigens

Examined by a suitably sensitive immunochemical method (2.7.1), hepatitis virus antigens are not detected.

HIV antigen

Examined by a suitably sensitive immunochemical method (2.7.1), HIV antigen is not detected.

Residual luteinising activity

The International Units of FSH and LH are the activities contained in stated amounts of the International Standard of human urinary follicle-stimulating hormone and luteinising hormone (interstitial-cell-stimulating hormone) which consists of a mixture of a freeze-dried extract of urine of post-menopausal women with lactose. The equivalence in International Units of the International Standard is stated by the World Health Organization. Use immature female rats approximately 21 days old and having masses such that the difference between the heaviest and the lightest rat is not more than 10 g. Assign the rats at random to 4 equal groups of at least 6 animals. If sets of 4 litter mates are available, assign one litter mate at random from each set to each group and mark according to litter.

Inject subcutaneously into each rat 50 IU of serum gonadotrophin R on the first day and 25 IU of choriotic gonadotrophin R on the fourth day, each in 0.5 mL of phosphate-albumin buffered saline pH 7.2 R. Choose 3 doses of the reference preparation such that the smallest dose produces a depletion of the ovarian ascorbic acid content in all the rats and the largest dose does not produce a maximal depletion in all the rats. Use doses in geometric progression, as an initial approximation, total doses

of 0.5 IU, 1.0 IU and 2.0 IU may be tried although the dose to be used will depend on the sensitivity of the animals. Choose a dose of the preparation to be examined expected to contain 60X IU of follicle-stimulating hormone (hFSH), in which X = the number of IU of hLH in the middle dose of the reference preparation.

Dissolve separately the total quantities of the preparation to be examined and of the reference preparation in 1.0 mL of phosphate-albumin buffered saline pH 7.2 R. Inject into a tail vein to each separate group of rats 6 days after the injection, of choriotic gonadotrophin. Exactly 4 h after the injection, euthanise the rats and remove the ovaries from each animal. Remove any extraneous fluid and tissue from the ovaries and weigh the ovaries immediately.

Treat the combined ovaries from each rat separately, as follows. Crush and homogenise within 2 min in a freshly prepared 25 g/L solution of metaphosphoric acid R at a temperature of 4 °C and dilute to 7 mL with the same solution. Allow the homogenate to stand for 30 min at 4 °C and centrifuge at 4 °C at approximately 2500 g. Filter the supernatant, if necessary, through a 0.22 µm filter.

Prepare a fresh solution consisting of a mixture of 2 mL of a 45.3 g/L solution of sodium acetate R adjusted to pH 7 with acetic acid R, 3 mL of water R and 2 mL of this dichlorophenolindophenol standard solution R. Mix 2 mL of this solution with 2 mL of the clear supernatant. 30 s after mixing, measure the absorbance (2.2.25) of the solution at the maximum at about 520 nm. Use as reference a solution with a known content of ascorbic acid CRS in a 25 g/L solution of metaphosphoric acid R, treated by the same process.

Calculate the amount of ascorbic acid from the ascorbic acid standard curve obtained and express in milligrams per 0.1 g of ovary to obtain the ascorbic acid content of the ovaries. Calculate the mean and its variance of the ascorbic acid content of the ovaries of the rats treated with the preparation to be examined.

For each dose-group of the reference preparation, plot the mean ascorbic acid content of the ovaries as a function of the logarithm of the dose and analyse the regression of the ascorbic acid content on the logarithm of the dose injected, using standard methods of analysis (the method of least squares).

The test is not valid unless: — the slope constant *b* is significant at the 5 per cent level of significance, — for the groups treated with the reference preparation, the sum of squares due to linear regression is equal to at least 95 per cent of the total sum of squares of the ascorbic acid content, — the within-group variance of the ascorbic acid content of the group receiving the preparation to be examined is not significantly different at the 5 per cent level of significance from the within-group variance of the ascorbic acid content of the groups receiving the reference preparation.

The mean ascorbic acid content of the ovaries of the rats treated with the preparation to be examined is not significantly lower than that of the rats treated with the middle dose of the reference preparation (calculated from the regression equation) at the 5 per cent level of significance. **Water** (2.5.32) Maximum 5.0 per cent.

Bacterial endotoxins (2.6.14, Method C)

Less than 0.40 IU per IU of urofollitropin, if intended for use in the manufacture of parenteral preparations without further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

The follicle-stimulating activity of urofollitropin is estimated by comparing under given conditions its effect in enlarging the ovaries of immature rats treated with chorionic gonadotrophin with the same effect of the International Standard preparation of human urinary follicle-stimulating hormone and luteinising hormone or of a reference preparation calibrated in International Units.

The International Units of FSH and LH are the activities contained in stated amounts of the International Standard of human urinary follicle-stimulating hormone and luteinising hormone (interstitial-cell-stimulating hormone) which consists of a mixture of a freeze-dried extract of urine of post-menopausal women with lactose. The equivalence in International Units of the International Standard is stated by the World Health Organization.

Use immature female rats of the same strain, 19 to 28 days old, differing in age by not more than 3 days and having masses such that the difference between the heaviest and the lightest rat is not more than 10 g. Assign the rats at random to 6 equal groups of at least 5 animals. If sets of 6 litter mates are available, assign one litter mate from each set to each group and mark according to litter.

Choose 3 doses of the reference preparation and 3 doses of the preparation to be examined such that the smallest dose produces a positive response in some of the rats and the largest dose does not produce a maximal response in all the rats. Use doses in geometric progression and as an initial approximation total doses of 1.5 IU, 3.0 IU and 6.0 IU may be tried although the dose will depend on the sensitivity of the animals used, which may vary widely.

Dissolve separately the total quantities of the preparation to be examined and of the reference preparation corresponding to the daily doses to be used in sufficient phosphate-albumin buffered saline pH 7.2 R such that the daily dose is administered in a volume of about 0.5 mL. The buffer solution shall contain in the daily dose not less than 14 IU of chorionic gonadotrophin to ensure complete luteinisation. Add a suitable antimicrobial preservative such as 4 g/L of phenol or 0.02 g/L of thiomersal. Store the solutions at $5 \pm 3^\circ\text{C}$.

Inject subcutaneously into each rat the daily dose allocated to its group. Repeat the injection of each dose 24 h and 48 h after the first injection. About 24 h after the last injection, Remove any extraneous fluid and tissue from the ovaries and weigh the 2 combined ovaries of each animal immediately. Calculate the results by the usual statistical methods, using the mass of the 2 combined ovaries as the response. (The precision of the assay may be improved by a suitable correction of the organ mass with reference to the mass of the animal from which it was taken; an analysis of covariance may be used).

The estimated potency is not less than 80 per cent and not more than 125 per cent of the stated potency. The confidence limits ($P = 0.95$) of the estimated potency are not less than 64 per cent and not more than 156 per cent of the stated potency.

STORAGE

In an airtight, tamper-proof container, protected from light, at a temperature of 2°C to 8°C . If the substance is sterile, store in a sterile, airtight, tamper-proof container.

LABELLING

The label states:
— the activity expressed in International Units of follicle-stimulating hormone per container,
— the potency expressed in International Units of follicle-stimulating hormone per milligram.

Ph Eur

Urokinaze

(Ph. Eur. monograph 0695)

9039-53-6

Action and use

Plasminogen activator; fibrinolytic; enzyme.

Ph Eur

DEFINITION

Enzyme, obtained from human urine, that activates plasminogen. It consists of a mixture of low-molecular-mass (LMM) (33 000) and high-molecular-mass (HMM) (54 000) forms, the high-molecular-mass form being predominant.

Potency

Not less than 70 000 IU per milligram of protein.

PRODUCTION

It is produced by validated methods of manufacturing designed to minimise or eliminate vasoactive substances.

CHARACTERS**Appearance**

White or almost white, amorphous powder.

Solubility

Soluble in water.

IDENTIFICATION

A. Place separately in two haemolysis tubes 0.5 mL of citrated human plasma and 0.5 mL of citrated bovine plasma and maintain in a water-bath at 37°C . To each tube add 0.1 mL of a solution containing a quantity of the substance to be examined equivalent to 1000 IU/mL in phosphate buffer solution pH 7.4 R and 0.1 mL of a solution containing a quantity of human thrombin R equivalent to 20 IU/mL in phosphate buffer solution pH 7.4 R. Shake immediately. In both tubes, a clot forms and lyses within 30 min.

B. Carry out identification by a suitable immunodiffusion test.

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 10 mg in 10 mL of water R.

Hepatitis B surface antigen

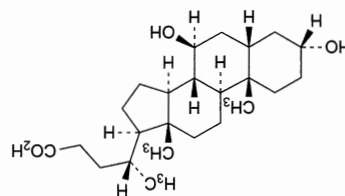
Examine by a suitably sensitive method such as radio-immunoassay. Hepatitis B surface antigen is not detected.

Thromboplastic contaminants

Test solutions Dissolve suitable quantities of the substance to be examined in barbital buffer solution pH 7.4 R to obtain

Ursodeoxycholic Acid

(Ph. Eur. monograph 1275)

C₂₄H₄₀O₄ 392.6 128-13-2

Action and use
Bile acid; treatment of gallstones.

Preparations

Ursodeoxycholic Acid Capsules
Ursodeoxycholic Acid Tablets
Ursodeoxycholic Acid Oral Suspension

DEFINITION

3α,7β-Dihydroxy-5β-cholan-24-oic acid.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Practically insoluble in water, freely soluble in ethanol (96 per cent), slightly soluble in acetone, practically insoluble in methylene chloride.

m.p.

About 202 °C.

IDENTIFICATION**First identification A.**

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison ursodeoxycholic acid CRS.

B. Examine the chromatograms obtained in the test for impurity C.

Results The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve about 10 mg in 1 mL of sulfuric acid R.

Add 0.1 mL of formaldehyde solution R and allow to stand for 5 min. Add 5 mL of water R. The suspension obtained is greenish-blue.

TESTS**Specific optical rotation (2.2.7)**

+ 58.0 to + 62.0 (dried substance).

Dissolve 0.500 g in anhydrous ethanol R and dilute to 25.0 mL with the same solvent.

Impurity C

Thin-layer chromatography (2.2.27).

Solvent mixture water R, acetone R (10:90 V/V).

Test solution (a) Dissolve 0.40 g of the substance to be examined in the solvent mixture and dilute to 10 mL with the solvent mixture.

Test solution (b) Dilute 1 mL of test solution (a) to 10 mL with the solvent mixture.

Reference solution (a) Dissolve 40 mg of ursodeoxycholic acid CRS in the solvent mixture and dilute to 10 mL with the solvent mixture.

Reference solution (b) Dissolve 20 mg of lithocholic acid CRS (impurity C) in the solvent mixture and dilute to 10.0 mL with the solvent mixture (solution A). Dilute 2.0 mL of this solution to 100.0 mL with the solvent mixture.

Reference solution (c) To 5 mL of solution A add 10 mg of chenodeoxycholic acid CRS (impurity A) and dilute to 50 mL with the solvent mixture.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, acetone R, methylene chloride R (1:30:60 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying At 120 °C for 10 min.

Detection Spray immediately with a 47.6 g/L solution of phosphomolybdic acid R in a mixture of 1 volume of sulfuric acid R and 20 volumes of glacial acetic acid R and heat at 120 °C until blue spots appear on a lighter background.

System suitability: reference solution (c):

the chromatogram shows 2 clearly separated principal spots.

Limit: test solution (a):

impurity C: any spot due to impurity C is not more intense than the principal spot in the chromatogram obtained with reference solution (b) (0.1 per cent).

Related substances

Liquid chromatography (2.2.29).

Solvent mixture methanol R, mobile phase (10:90 V/V).

Test solution Dissolve 60 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of ursodeoxycholic acid for system suitability CRS (containing impurities A and H) in 1.0 mL of the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

size: l = 0.25 m, Ø = 4.6 mm;

stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 µm);

temperature: 40 °C ± 1 °C.

Mobile phase Mix 30 volumes of acetonitrile R, 37 volumes of a 0.78 g/L solution of sodium dihydrogen phosphate R adjusted to pH 3 with phosphoric acid R, and 40 volumes of methanol R.

Flow rate 0.8 mL/min.

Detection Refractometer at 35 ± 1 °C.

Injection 150 µL.

Run time 4 times the retention time of ursodeoxycholic acid.

Identification of impurities Use the chromatogram supplied with ursodeoxycholic acid for system suitability CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A and H.

Relative retention With reference to ursodeoxycholic acid (retention time = about 14 min): impurity H = about 0.9; impurity A = about 2.8.

System suitability: reference solution (a):
— **resolution:** minimum 1.5 between the peaks due to impurity H and ursodeoxycholic acid.

Limits:
— **impurity A:** not more than 10 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent);

— **unspecified impurities:** for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **total:** not more than 15 times the area of the principal peak in the chromatogram obtained with reference solution (b) (1.5 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.350 g in 50 mL of ethanol (96 per cent) R, previously neutralised to 0.2 mL of phenolphthalein solution R. Add 50 mL of water R and titrate with 0.1 M sodium hydroxide until a pink colour is obtained.

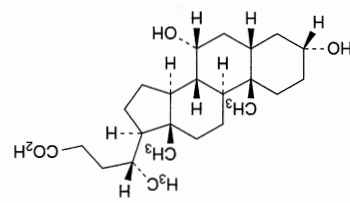
1 mL of 0.1 M sodium hydroxide is equivalent to 39.26 mg of $C_{24}H_{40}O_4$.

IMPURITIES

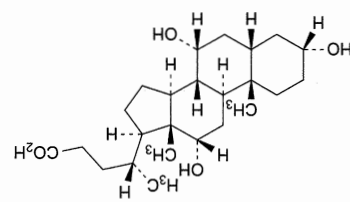
Specified impurities A, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

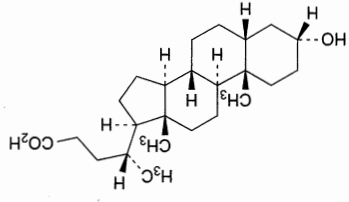
Control of impurities in substances for pharmaceutical use: B, D, E, F, G, H, I.



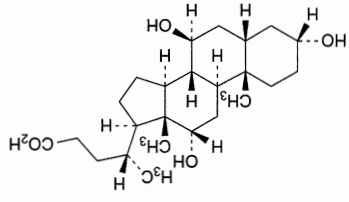
A. 3 α ,7 β -dihydroxy-5 β -cholan-24-oic acid (chenodeoxycholic acid),



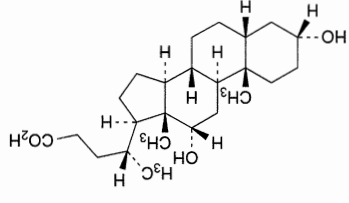
B. 3 α ,7 α ,12 α -trihydroxy-5 β -cholan-24-oic acid (thiocholic acid),



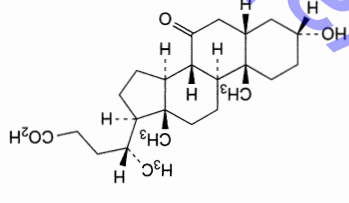
C. 3 α -hydroxy-5 β -cholan-24-oic acid (lithocholic acid),



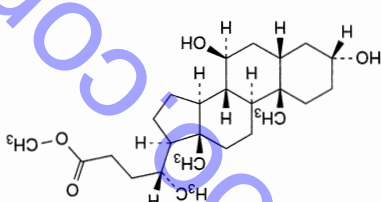
D. 3 α ,7 β ,12 α -trihydroxy-5 β -cholan-24-oic acid (ursocholic acid),



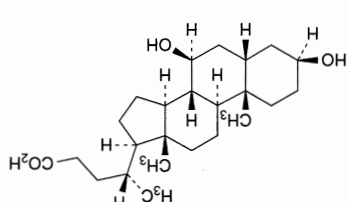
E. 3 α ,12 α -dihydroxy-5 β -cholan-24-oic acid (deoxycholic acid),



F. 3 α -hydroxy-7-oxo-5 β -cholan-24-oic acid,



G. methyl 3 α ,7 β -dihydroxy-5 β -cholan-24-oate,



H. 3 β ,7 β -dihydroxy-5 β -cholan-24-oic acid,

Reference solution (a) Dissolve 5 mg of valaciclovir impurity D CRS, 5.0 mg of valaciclovir impurity E CRS, 5.0 mg of valaciclovir impurity F para-toluenesulfonate CRS in a mixture of 2 mL of water R and 6 mL of ethanol (96 per cent) R, and dilute to 10.0 mL with ethanol (96 per cent) R.

Reference solution (b) Dilute 3.0 mL of reference solution (a) to 10.0 mL with ethanol (96 per cent) R.

Reference solution (c) Dilute 2.0 mL of reference solution (a) to 10.0 mL with ethanol (96 per cent) R.

Reference solution (d) Dilute 0.5 mL of reference solution (a) to 10.0 mL with ethanol (96 per cent) R.

Plate TLC silica gel F₂₅₄ plate R (2–10 µm).

Pre-treatment Wash the plate with methanol R until the solvent front has migrated over at least 4/5 of the plate; allow the plate to dry.

Mobile phase concentrated ammonia R, tetrahydrofuran R, methanol R, methylene chloride R (3:12:34:54 V/V/V/V); use freshly prepared mobile phase.

Application 4 µL of the test solution and reference solutions (b), (c) and (d).

Development Over 4/5 of the plate.

Drying In a current of air.

Detection Examine in ultraviolet light at 254 nm for impurities E and G; spray with a 0.1 g/L solution of fluorescamine R in ethylene chloride R and examine in ultraviolet light at 365 nm for impurity F.

Retention factors Impurity A = about 0; impurity B = about 0.2; valaciclovir = about 0.3; impurity C = about 0.5; impurity D = about 0.6; impurity E = about 0.7; impurity F = about 0.75; impurity G = about 0.79; impurity C is masked by the leading edge of the spot due to valaciclovir; impurities F and G may co-elute, but this does not adversely affect their quantification because they are visualised differently.

System suitability The chromatograms obtained with reference solutions (b), (c) and (d) each show 3 clearly separated spots when examined under ultraviolet light at 254 nm, due to impurities D, E and G.

Limits:

- **impurity E:** any spot due to impurity E is not more intense than the corresponding spot in the chromatogram obtained with reference solution (c) (0.2 per cent);
- **impurity F:** any spot due to impurity F is not more intense than the corresponding spot in the chromatogram obtained with reference solution (b) (0.3 per cent);
- **impurity G:** any spot due to impurity G is not more intense than the corresponding spot in the chromatogram obtained with reference solution (d) (0.05 per cent).

Related substances

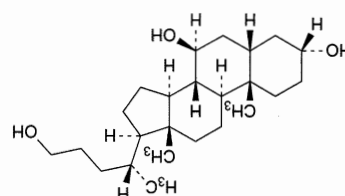
A. Impurities A, B, I and R. Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 50.0 mg of the substance to be examined in a 0.5 per cent V/V solution of hydrochloric acid R and dilute to 100.0 mL with the same solution.

Reference solution (a) Dissolve 2.5 mg of valaciclovir for system suitability CRS (containing impurities A, B, C, D, H, I, J, M and R) in a 0.5 per cent V/V solution of hydrochloric acid R and dilute to 5.0 mL with the same solution.

Reference solution (b) Dissolve 50.0 mg of anhydrous valaciclovir hydrochloride CRS in a 0.5 per cent V/V solution

Ph Eur

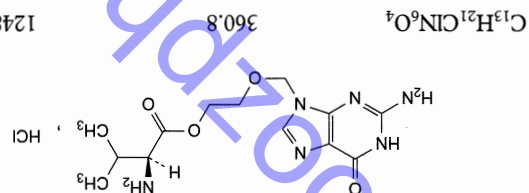


I. 5β-cholane-3α,7β,24-triol.



Anhydrous Valaciclovir Hydrochloride

(Ph. Eur. monograph 1768)



124832-27-5

C₁₃H₂₁ClN₆O₄

360.8

Action and use Purine nucleoside analogue; antiviral (herpesviruses).

Ph Eur

DEFINITION

2-[(2-Amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl 1-valinate hydrochloride.

Content

95.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, slightly soluble in anhydrous ethanol.

It shows polymorphism (5.9).

IDENTIFICATION

Carry out either tests A, B, C or tests A, B, D.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison anhydrous valaciclovir hydrochloride CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of anhydrous ethanol R and evaporate to dryness in a desiccator, under high vacuum, over diphosphorus pentoxide R. Record new spectra using the residues.

B. It gives reaction (a) of chlorides (2.3.1).

C. It complies with the limit for impurity R given in test A for related substances.

D. Optical rotation (2.2.7): laevorotatory.

Dissolve 2.50 g in water R and dilute to 50.0 mL with the same solvent.

TESTS

Impurities E, F and G

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.250 g of the substance to be examined in 2 mL of water R and dilute to 5.0 mL with ethanol (96 per cent) R.

of hydrochloric acid R and dilute to 100.0 mL with the same solution.

Reference solution (c) Dilute 3.0 mL of the test solution to 100.0 mL with a 0.5 per cent V/V solution of hydrochloric acid R. Dilute 1.0 mL of this solution to 100.0 mL with a 0.5 per cent V/V solution of hydrochloric acid R.

Column:

- size: $l = 0.15$ m, $\varnothing = 4.0$ mm;
- stationary phase: crown-ether silica gel for chromatography R (5 μ m);
- temperature: 10 °C.

Mobile phase perchloric acid R, methanol R, water R (0.5:5:95 V/V/V).

Flow rate 0.75 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 μ L of the test solution and reference solution (a).

Run time 1.5 times the retention time of valaciclovir.

Identification of impurities Use the chromatogram supplied with valaciclovir for system suitability CRS and the

chromatogram obtained with reference solution (a) to identify the peaks due to impurities A + B, C + R, D, I

and M.

Relative retention With reference to valaciclovir (retention time = about 21 min): impurities A and B = about 0.2;

impurity I = about 0.4; impurities C and R = about 0.6;

impurity D = about 0.7; impurity M = about 1.3.

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 1.5, where H_p = height above the baseline of the peak due to impurity D and

H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to

impurities C and R.

Limits:

— **correction factor:** for the calculation of content, multiply the peak area of impurities A and B by 0.7;

— **impurity R:** maximum 3.0 per cent; for the calculation, subtract the content of impurity C as determined in

test B for related substances from the content of the coeluting impurities C and R as determined in this

test;

— **sum of impurities A and B:** maximum 2.0 per cent;

— **impurity I:** maximum 0.2 per cent;

— **disregard limit:** the area of the principal peak in the chromatogram obtained with reference solution (c)

(0.03 per cent); disregard any peaks due to impurities other than A + B, C + R or I.

B. Liquid chromatography (2.2.29): use the normalisation procedure. Use the solutions within 24 h of preparation.

Solvent mixture ethanol (96 per cent) R, water R (20:80 V/V).

Test solution Dissolve 40 mg of the substance to be examined in the solvent mixture and dilute to 100 mL with the solvent mixture.

Reference solution (a) Dissolve 2.5 mg of valaciclovir for system suitability CRS (containing impurities A, B, C, D, H, I, J, M and R) in the solvent mixture and dilute to 5.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 20.0 mL with the solvent mixture.

Column:

- size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

- stationary phase: phenylhexylsilica gel for chromatography R (5 μ m);

— temperature: 15 °C.

Mobile phase:

- **mobile phase A:** trifluoroacetic acid R, water R (0.2:100 V/V);
- **mobile phase B:** trifluoroacetic acid R, methanol R2 (0.2:100 V/V);

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 5	90	10
5 - 35	90 → 60	10 → 40

Flow rate 0.8 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 10 μ L.

Identification of impurities Use the chromatogram supplied with valaciclovir for system suitability CRS and the

chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, B, C, D, H, I, J

and M.

Relative retention With reference to valaciclovir (retention time = about 19 min): impurity A = about 0.3;

impurity B = about 0.4; impurity H = about 0.5;

impurity C = about 1.06; impurity I = about 1.09;

impurity D = about 1.2; impurity J = about 1.3;

impurity M = about 1.6.

System suitability: reference solution (a):

— **peak-to-valley ratio:** minimum 2.5, where H_p = height above the baseline of the peak due to impurity C and

H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to

valaciclovir;

— **the chromatogram** obtained is similar to the chromatogram supplied with valaciclovir for system

Limits:

— **impurity M:** maximum 1.5 per cent;

— **impurity D:** maximum 0.5 per cent;

— **impurity C:** maximum 0.3 per cent;

— **impurity H:** maximum 0.1 per cent;

— **impurity J:** maximum 0.1 per cent;

— **unspecified impurities:** for each impurity, maximum 0.05 per cent;

— **disregard limit:** 0.6 times the area of the principal peak in the chromatogram obtained with reference

solution (b) (0.03 per cent); disregard the peaks due to impurities A, B and I.

Limits:

— **total for tests A and B:** maximum 5.0 per cent.

Chloride

9.4 to 9.9 per cent (anhydrous and solvent-free substance).

Dissolve 0.350 g in 100 mL of water R and add 0.2 mL of nitric acid R. Carry out a potentiometric titration (2.2.20), using 0.1 M silver nitrate. Use a silver indicator electrode and a silver-silver chloride reference electrode or a combined

silver electrode. Discard the result from the first titration, which is used to condition the electrodes. Carry out a blank titration.

1 mL of 0.1 M silver nitrate is equivalent to 3.543 mg of Cl.

Palladium

Maximum 10 ppm.

Inductively coupled plasma-atomic emission spectrometry (2.2.57).

Test solution Dissolve 0.1 g in a 2 per cent *V/V* solution of hydrochloric acid *R* in dimethyl sulfoxide *R* and dilute to 10.0 mL with the same solution.

Reference solutions Prepare the reference solutions using a solution containing 1000 µg of Pd per millilitre, diluted as necessary with a 2 per cent *V/V* solution of hydrochloric acid *R* in dimethyl sulfoxide *R*.

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in water *R* and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using 10 mL of lead standard solution (2 ppm Pb) *R*.

Water (2.5.12)

Maximum 2.0 per cent, determined on 0.250 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in test A for related substances with the following modification.

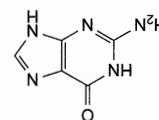
Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{13}H_{21}ClN_5O_4$ from the declared content of anhydrous valaciclovir hydrochloride CRS.

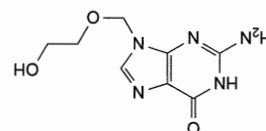
IMPURITIES

Specified impurities A, B, C, D, E, F, G, H, I, J, M, R

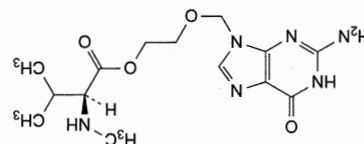
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): K, L, N, O, P, Q.



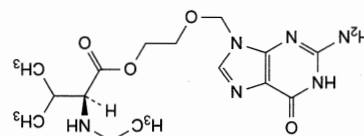
A. 2-amino-1,9-dihydro-6H-purin-6-one (guanine),



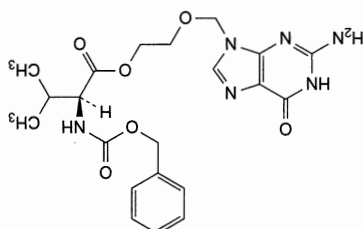
B. 2-amino-9-[(2-hydroxyethoxy)methyl]-1,9-dihydro-6H-purin-6-one (aciclovir),



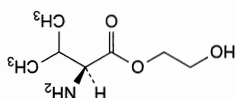
C. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-methyl-L-valinate,



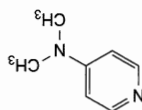
D. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-ethyl-L-valinate,



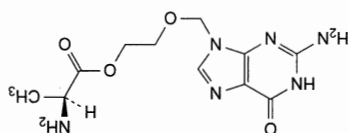
E. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-[(benzyloxy)carbonyl]-L-valinate,



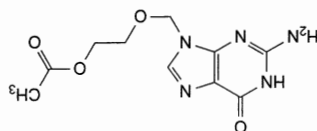
F. 2-hydroxyethyl L-valinate,



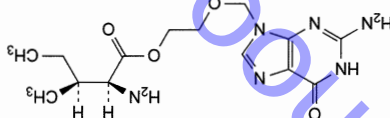
G. N,N-dimethylpyridin-4-amine,



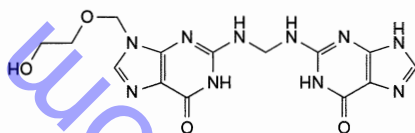
H. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl L-alaninate,



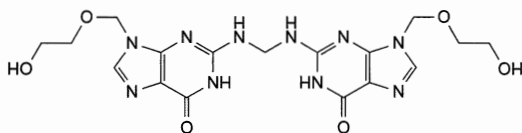
I. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl acetate,



J. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl L-isoleucinate,



K. 9-[(2-hydroxyethoxy)methyl]-2-[[[(6-oxo-6,9-dihydro-1H-purin-2-yl)amino]methyl]amino]-1,9-dihydro-6H-purin-6-one,

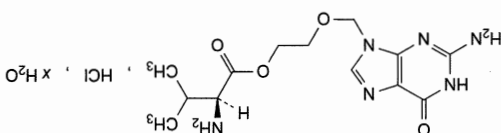


L. 2,2'-(methylenediamino)bis[9-[(2-hydroxyethoxy)methyl]-1,9-dihydro-6H-purin-6-one],



Valaciclovir Hydrochloride, Hydrated

(Ph. Eur. monograph 2751)



$C_{13}H_{21}ClN_6O_4 \cdot xH_2O$ 360.8

(anhydrous substance)

Action and use
Purine nucleoside analogue; antiviral (herpesviruses)

DEFINITION
2-[(2-Amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl L-valinate hydrochloride.

Content
95.0 per cent to 102.0 per cent (anhydrous substance).
It contains a variable quantity of water.

CHARACTERS
Appearance
White or almost white powder, hygroscopic.

Solubility
Freely soluble in water; very slightly soluble in anhydrous ethanol, practically insoluble in acetonitrile.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve the substance to be examined in the minimum volume of water R, evaporate to dryness at room temperature and record the spectrum using the residue.

Comparison Repeat the operations using anhydrous valaciclovir hydrochloride CRS.

B. It complies with the limit for impurity R (see test A for related substances).

C. Water (see Tests).

D. It gives reaction (a) of chlorides (2.3.1).

TESTS

Impurities G and S

Thin-layer chromatography (2.2.27).

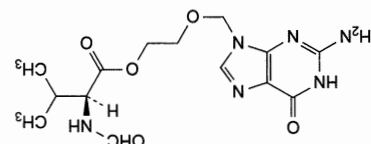
Test solution Dissolve 0.250 g of the substance to be examined in 2 mL of water R and dilute to 5.0 mL with ethanol (96 per cent) R.

Reference solution Dissolve 5.0 mg of valaciclovir impurity G CRS and 5.0 mg of valaciclovir impurity S CRS in a mixture of 2 mL of water R and 6 mL of ethanol (96 per cent) R and dilute to 10.0 mL with ethanol (96 per cent) R. Dilute 0.5 mL of the solution to 10.0 mL with ethanol (96 per cent) R.

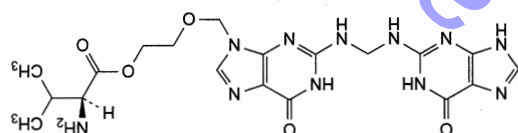
Plate TLC silica gel F₂₅₄ plate R (2-10 µm).

Pretreatment Wash the plate with methanol R until the solvent front has migrated over at least 4/5 of the plate; allow to dry in air.

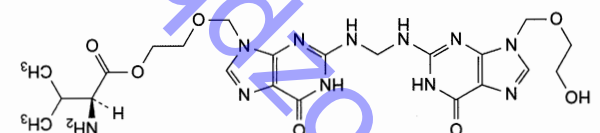
Mobile phase concentrated ammonia R, tetrahydrofuran R, methanol R, methylene chloride R (3:12:34:54 V/V/V/V); use freshly prepared mobile phase.



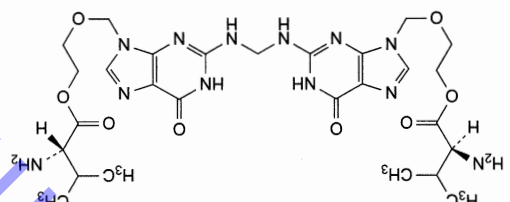
M. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-formyl-L-valinate,



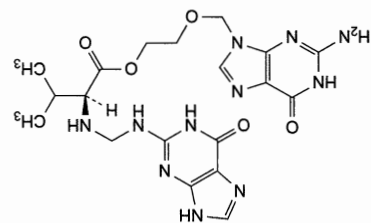
N. 2-[[[6-oxo-2-[[[(6-oxo-6,9-dihydro-1H-purin-2-yl)amino]methyl]amino]-6-oxo-1,6-dihydro-9H-purin-9-yl]methoxy]ethyl] L-valinate,



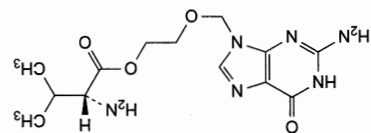
O. 2-[[[2-[[[9-[(2-hydroxyethoxy)methyl]-6-oxo-6,9-dihydro-1H-purin-2-yl]amino]methyl]amino]-6-oxo-1,6-dihydro-9H-purin-9-yl]methoxy]ethyl] L-valinate,



P. 2,2'-[methylenebis[imino(6-oxo-1,6-dihydro-9H-purine-9,2-diyl)methyleneoxy]]diethyl di(L-valinate),



Q. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N'[[[(6-oxo-6,9-dihydro-1H-purin-2-yl)amino]methyl]-L-valinate,



R. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl D-valinate.

Ph Eur

Application 4 µL.**Development** Over 4/5 of the plate.**Drying** In a current of air.**Detection** Examine in ultraviolet light at 254 nm.**Retention factors** Valaciclovir = about 0.3;

impurity S = about 0.7; impurity G = about 0.8.

System suitability The chromatogram obtained with the

reference solution shows 2 clearly separated spots due to

impurities S and G.

Limits:

— impurity G: any spot due to impurity G is not more

intense than the corresponding spot in the chromatogram

obtained with the reference solution (0.05 per cent);

— impurity S: any spot due to impurity S is not more intense

than the corresponding spot in the chromatogram

obtained with the reference solution (0.05 per cent).

Related substances**A. Liquid chromatography (2.2.29):** use the normalisation

procedure.

Test solution Dissolve 50.0 mg of the substance to be

examined in a 0.5 per cent V/V solution of hydrochloric acid R

and dilute to 100.0 mL with the same solution.

Reference solution (a) Dissolve 2.5 mg of valaciclovir for system

suitability CRS (containing impurities A, B, C, D, H, M

and R) in a 0.5 per cent V/V solution of hydrochloric acid R

and dilute to 5.0 mL with the same solution.

Reference solution (b) Dissolve 50.0 mg of anhydrous

valaciclovir hydrochloride CRS in a 0.5 per cent V/V solution

of hydrochloric acid R and dilute to 100.0 mL with the same

Reference solution (c) Dilute 3.0 mL of the test solution to

100.0 mL with a 0.5 per cent V/V solution of hydrochloric

acid R. Dilute 1.0 mL of this solution to 100.0 mL with a

Column:— size: $l = 0.15$ m, $\phi = 4.0$ mm;

— stationary phase: crown-ether silica gel for

chromatography R (5 µm);

— temperature: 10 °C.

Mobile phase perchloric acid R, methanol R, water R

(0.5:5:95 V/V/V).

Flow rate 0.75 mL/min.**Detection** Spectrophotometer at 254 nm.**Injection** 10 µL of the test solution and reference solutions (a)

and (c).

Run time 1.5 times the retention time of valaciclovir.**Identification of impurities** Use the chromatogram supplied

with valaciclovir for system suitability CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities A + B, C + R, D

and M.

Relative retention With reference to valaciclovir (retention

time = about 17 min): impurities A and B = about 0.2;

impurities C and R = about 0.6; impurity D = about 0.7;

impurity M = about 1.3.

System suitability: reference solution (a):— peak-to-valley ratio: minimum 1.5, where H_p = height

above the baseline of the peak due to impurity D and

 H_v = height above the baseline of the lowest point of

the curve separating this peak from the peak due to

impurities C and R.

Limits:

— correction factor: for the calculation of content, multiply

the peak area of impurities A and B by 0.7;

— impurity R: maximum 3.0 per cent; for the calculation,

subtract the content of impurity C as determined in

test B for related substances from the content of the

coeluting impurities C and R as determined in this

test;

— sum of impurities A and B: maximum 2.0 per cent;

— disregard limit: the area of the principal peak in the

chromatogram obtained with reference solution (c)

other than A + B and C + R.

B. Liquid chromatography (2.2.29): use the normalisation

procedure. Use the solutions within 24 h of preparation.

Solvent mixture ethanol (96 per cent) R, water R (20:80 V/V).**Test solution** Dissolve 80 mg of the substance to be examined

in the solvent mixture and dilute to 100.0 mL with the

solvent mixture.

Reference solution (a) Dissolve 1.6 mg of valaciclovir for system

suitability CRS (containing impurities A, B, C, D, H, M

and R) in the solvent mixture and dilute to 2.0 mL with the

solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 20.0 mL with the solvent mixture.

Reference solution (c) Dissolve 2 mg of valaciclovir

impurity P CRS in the solvent mixture and dilute to 25.0 mL

with the solvent mixture. Dilute 1.0 mL of the solution to

100.0 mL with the solvent mixture.

Column:— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped phenylhexylsilica gel for

chromatography R (5 µm);

— temperature: 15 °C.

Mobile phase:

— mobile phase A: trifluoroacetic acid R, water R

(0.2:100 V/V);

— mobile phase B: trifluoroacetic acid R, methanol R2

(0.2:100 V/V);

Time (min)**Mobile phase A** (per cent V/V)**Mobile phase B** (per cent V/V)

0 - 5

5 - 35

35 - 45

60

90 → 60

10 → 40

40

Flow rate 0.8 mL/min.**Detection** Spectrophotometer at 254 nm.**Injection** 10 µL.**Identification of impurities** Use the chromatogram supplied

with valaciclovir for system suitability CRS and the

chromatogram obtained with reference solution (a) to

identify the peaks due to impurities A, B, C, D, H and M;

use the chromatogram obtained with reference solution (c) to

identify the peak due to impurity P.

Relative retention With reference to valaciclovir (retention

time = about 20 min): impurity A = about 0.3;

impurity B = about 0.4; impurity H = about 0.5;

impurity C = about 1.06; impurity D = about 1.2;

impurity M = about 1.6; impurity P = about 2.0.

System suitability: reference solution (a):
— peak-to-valley ratio: minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_v = height separating this peak from the peak due to valaciclovir.

Limits:
— impurity M: maximum 0.6 per cent;
— impurity D: maximum 0.3 per cent;
— impurity C: maximum 0.2 per cent;
— impurities H, P: for each impurity, maximum 0.1 per cent;
— unspecified impurities: for each impurity, maximum 0.05 per cent;
— disregard limit: 0.6 times the area of the principal peak in the chromatogram obtained with reference solution (b) (0.03 per cent); disregard the peaks due to impurities A and B.

Limit:
— total for tests A and B: maximum 4.0 per cent.

Palladium

Maximum 10 ppm.

Inductively coupled plasma-atomic emission spectrometry

(2.2.57).

Test solution Dissolve 0.1 g in a 2 per cent V/V solution of hydrochloric acid R in dimethyl sulfoxide R and dilute to 10.0 mL with the same solution.

Reference solutions Prepare the reference solutions using a solution containing 1000 µg of Pd per millilitre, diluted as necessary with a 2 per cent V/V solution of hydrochloric acid R in dimethyl sulfoxide R.

Heavy metals (2.4.8)

Wavelength 340.5 nm.

Maximum 20 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same solvent. 12 mL of the solution complies with test A. Prepare the reference solution using 10 mL of lead standard solution (2 ppm Pb) R.

Water (2.5.12)

4.5 per cent to 11.0 per cent, determined on 0.100 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in test A for related substances with the following modification.

Injection Test solution and reference solution (b).

Calculate the percentage content of $C_{13}H_{21}ClN_6O_4$ taking into account the assigned content of anhydrous valaciclovir hydrochloride CRS.

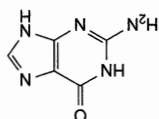
STORAGE

In an airtight container.

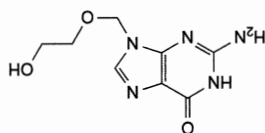
IMPURITIES

Specified impurities A, B, C, D, G, H, M, P, R, S

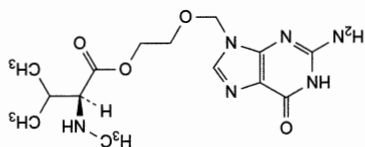
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: 1, J, N.



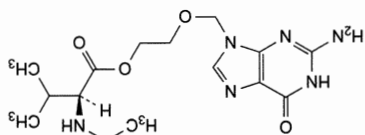
A. 2-amino-1,9-dihydro-6H-purin-6-one (guanine),



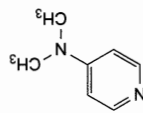
B. 2-amino-9-[(2-hydroxyethoxy)methyl]-1,9-dihydro-6H-purin-6-one (aciclovir),



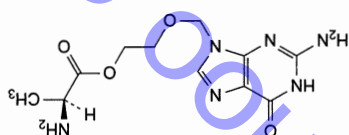
C. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-methyl-L-valinate,



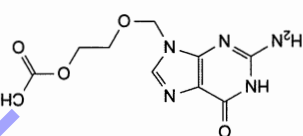
D. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-ethyl-L-valinate,



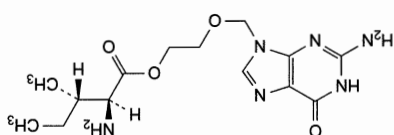
G. N,N-dimethylpyridin-4-amine,



H. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl L-alaninate,



I. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl acetate,

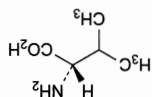


J. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl L-isoleucinate,



(Ph. Eur. monograph 0796)

Valine



117.1

 $C_5H_{11}NO_2$

72-18-4

Action and use

Amino acid.

Ph Eur

DEFINITION

(2S)-2-Amino-3-methylbutanoic acid.

Fermentation product, extract or hydrolysate of protein.

Content

98.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder or colourless

Solubility

Soluble in water, very slightly soluble in ethanol

(96 per cent).

IDENTIFICATION

First identification A, B.

Second identification A, C.

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in a 10.3 g/L solution of hydrochloric acid R and dilute to 50 mL with the same solution.

Reference solution Dissolve 10 mg of valine CRS in a 10.3 g/L

solution of hydrochloric acid R and dilute to 50 mL with the

same solution.

Plate TLC silica gel plate R.

Mobile phase glacial acetic acid R, water R, butanol R

(20:20:60 V/V/V)

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Spray with ninhydrin solution R and heat at 105 °C

for 15 min.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

reference solution.

TESTS

Solution S

Dissolve 2.5 g in water R and dilute to 100 mL with the

same solvent.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured

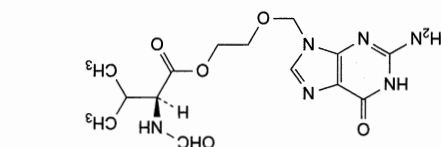
than reference solution BY₆ (2.2.2, Method II).

Specific optical rotation (2.2.7)

+ 26.5 to + 29.0 (dried substance).

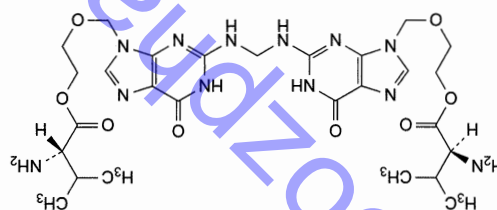
Dissolve 2.00 g in hydrochloric acid R1 and dilute to 25.0 mL

with the same acid.

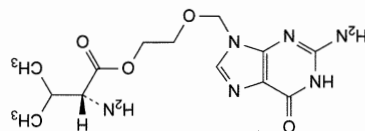


M. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-formyl-L-valinate,

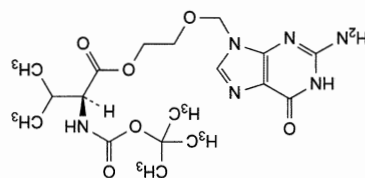
N. 2-[[[6-oxo-2-[[[(6-oxo-6,9-dihydro-1H-purin-2-yl)amino]methyl]amino]-1,6-dihydro-9H-purin-9-yl]methoxy]ethyl] L-valinate,



P. 2,2'-[methylenebis[imino(6-oxo-1,6-dihydro-9H-purine-2,9-diyl)methyleneoxy]]diethyl di(L-valinate),



R. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl D-valinate,



S. 2-[(2-amino-6-oxo-1,6-dihydro-9H-purin-9-yl)methoxy]ethyl N-[(1,1-dimethylethoxy)carbonyl]-L-valinate.

Ph Eur

Ninhydrin-positive substances

Amino acid analysis (2.2.56). For analysis, use Method 1.

The concentrations of the test solution and the reference solutions may be adapted according to the sensitivity of the equipment used. The concentrations of all solutions are adjusted so that the system suitability requirements described in general chapter 2.2.46 are fulfilled, keeping the ratios of concentrations between all solutions as described.

Solution A dilute hydrochloric acid R1 or a sample preparation buffer suitable for the apparatus used.

Test solution Dissolve 30.0 mg of the substance to be examined in solution A and dilute to 50.0 mL with solution A.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with solution A. Dilute 2.0 mL of this solution to 10.0 mL with solution A.

Reference solution (b) Dissolve 30.0 mg of proline R in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Reference solution (c) Dilute 6.0 mL of ammonium standard solution (100 ppm NH₄) R to 50.0 mL with solution A. Dilute 1.0 mL of this solution to 100.0 mL with solution A.

Reference solution (d) Dissolve 30 mg of isoleucine R (impurity B) and 30 mg of leucine R (impurity C) in solution A and dilute to 50.0 mL with solution A. Dilute 1.0 mL of the solution to 200.0 mL with solution A.

Reference solution (e) Dissolve 30.0 mg of isoleucine R (impurity B) in solution A and dilute to 100.0 mL with solution A. Dilute 1.0 mL of the solution to 250.0 mL with solution A.

Blank solution Solution A.

Inject suitable, equal amounts of the test, blank and reference solutions into the amino acid analyser. Run a program suitable for the determination of physiological amino acids. **System suitability** Reference solution (d): — **resolution**: minimum 1.5 between the peaks due to impurities B and C. **Calculation of percentage contents**: — for impurity B, use the concentration of impurity B in reference solution (e); — for any ninhydrin-positive substance detected at 570 nm, use the concentration of valine in reference solution (a); — for any ninhydrin-positive substance detected at 440 nm, use the concentration of proline in reference solution (b); if a peak is above the reporting threshold at both wavelengths, use the result obtained at 570 nm for quantification.

Limits:

— **impurity B** at 570 nm: maximum 0.4 per cent; any ninhydrin-positive substance: for each impurity, maximum 0.2 per cent;

— **total**: maximum 1.0 per cent;

— **reporting threshold**: 0.05 per cent.

The thresholds indicated under Related substances (Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

Chlorides (2.4.4)

Maximum 200 ppm.

Dilute 10 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 300 ppm.

Dissolve 0.5 g in distilled water R and dilute to 15 mL with the same solvent.

Ammonium

Amino acid analysis (2.2.56) as described in the test for ninhydrin-positive substances with the following modifications.

Injection Test solution, reference solution (c) and blank solution.

Limit:

— **ammonium** at 570 nm: not more than the area of the corresponding peak in the chromatogram obtained with reference solution (c) (0.02 per cent), taking into account the peak due to ammonium in the chromatogram obtained with the blank solution.

Iron (2.4.9)

Maximum 10 ppm.

In a separating funnel, dissolve 1.0 g in 10 mL of dilute hydrochloric acid R. Shake with 3 quantities, each of 10 mL, of methyl isobutyl ketone R1, shaking for 3 min each time. To the combined organic layers add 10 mL of water R and shake for 3 min. Use the aqueous layer.

Heavy metals (2.4.8)

Maximum 10 ppm.

Solvent water R.

0.25 g complies with test H. Prepare the reference solution using 0.25 mL of lead standard solution (10 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 3 mL of anhydrous formic acid R. Add 30 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). 1 mL of 0.1 M perchloric acid is equivalent to 11.71 mg of C₅H₁₁NO₂.

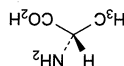
STORAGE

Protected from light.

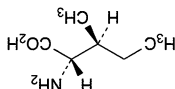
IMPURITIES

Specified impurities B.

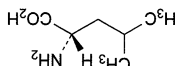
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C



A. (2S)-2-aminopropanoic acid (alanine),



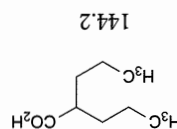
B. (2S,3S)-2-amino-3-methylpentanoic acid (isoleucine),



C. (2S)-2-amino-4-methylpentanoic acid (leucine).

Valproic Acid

(Ph. Eur. monograph 1378)



99-66-1

Action and use
Antiepileptic.

Ph Eur

DEFINITION

2-Propylpentanoic acid.

Content

99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

Colourless or very slightly yellow, clear liquid, slightly

Solubility

Very slightly soluble in water, miscible with ethanol

(96 per cent) and with methylene chloride. It dissolves in

dilute solutions of alkali hydroxides.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison valproic acid CRS.

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Dissolve 2.0 g in dilute sodium hydroxide solution R and dilute

to 10 mL with the same alkaline solution.

Related substances

Gas chromatography (2.2.28).

Test solution Dissolve 0.500 g of the substance to be

examined in heptane R and dilute to 100.0 mL with the same

solvent.

Reference solution (a) Dissolve 5 mg of valproic acid for system

suitability CRS (containing impurity K) in 1.0 mL of

heptane R.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with heptane R.

Column:

— material: wide-bore fused silica;

— size: $l = 30$ m, $\varnothing = 0.53$ mm;

— stationary phase: macrogol 20 000 2-nitroterephthalate R

(film thickness 0.5 μ m).

Carrier gas helium for chromatography R.

Flow rate 8 mL/min.

Temperature:

Time (min)	Temperature (°C)
0 - 5	80
5 - 15	80 \rightarrow 150
15 - 28.3	150 \rightarrow 190
28.3 - 30	190
Injection port	220
Detector	220

Detection Flame ionisation.

Injection 1 μ L.

Relative retention With reference to valproic acid (retention time = about 17 min): impurity K = about 0.97.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to

impurity K and valproic acid.

Limits:

— impurity K: not more than 0.15 times the area of the principal peak in the chromatogram obtained with

reference solution (b) (0.15 per cent);

— unspecified impurities: for each impurity, not more than

0.05 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.05 per cent);

— total: not more than 0.2 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.2 per cent);

— disregard limit: 0.03 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.03 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 2.0 g in ethanol (80 per cent V/V) R and dilute to

20 mL with the same solvent. 12 mL of the solution

complies with test B. Prepare the reference solution using

lead standard solution (2 ppm Pb) obtained by diluting lead

standard solution (100 ppm Pb) R with ethanol

(80 per cent V/V) R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.100 g in 25 mL of ethanol (96 per cent) R.

Add 2 mL of water R. Titrate with 0.1 M sodium hydroxide,

determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M sodium hydroxide is equivalent to 14.42 mg of

C₈H₁₆O₂.

STORAGE

In an airtight container.

IMPURITIES

Specified impurities K

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

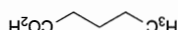
by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

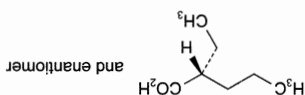
impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: A, B,

C, D, E, F, G, H, I, J, L.



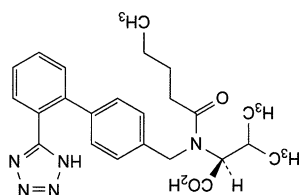
A. pentanoic acid (valeric acid),



B. (2RS)-2-ethylpentanoic acid,

Valsartan

(Ph. Eur. monograph 2423)


 $C_{24}H_{29}N_5O_3$ 435.5 137862-53-4

Action and use

Angiotensin II (AT_1) receptor antagonist.

Preparations

Valsartan Capsules
Valsartan Tablets

Ph Eur

DEFINITION

(2S)-3-Methyl-2-[pentanoyl[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]amino]butanoic acid.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Practically insoluble in water, freely soluble in anhydrous ethanol, sparingly soluble in methylene chloride.

IDENTIFICATION

Carry out either tests A, B or tests A, C.

A. Infrared absorption spectrophotometry (2.2.24).

Comparison valsartan CRS.

B. Enantiomeric purity (see Tests).

C. Specific optical rotation (2.2.7): -69.0 to -64.0 (anhydrous substance).Dissolve 0.200 g in methanol R and dilute to 20.0 mL with the same solvent.

TESTS

Enantiomeric purity

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.Reference solution (a) Dissolve 5 mg of valsartan for peak identification CRS (containing impurity A) in the mobile phase and dilute to 5.0 mL with the mobile phase.Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase.

Column:

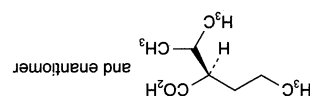
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: silica gel OD for chiral separations R.

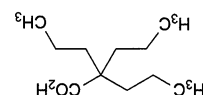
Mobile phase trifluoroacetic acid R, 2-propanol R, hexane R (0.1:15:85 V/V/V).

Flow rate 0.8 mL/min.Detection Spectrophotometer at 230 nm.Injection 10 μ L.

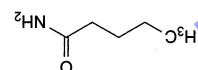
Run time 1.5 times the retention time of valsartan.



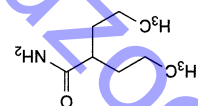
C. (2RS)-2-(1-methylethyl)pentanoic acid,



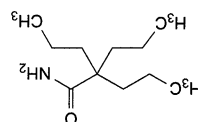
D. 2,2-dipropylpentanoic acid,



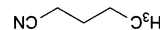
E. pentanamide (valeramide),



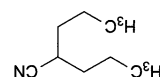
F. 2-propylpentanamide,



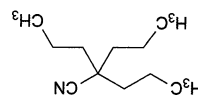
G. 2,2-dipropylpentanamide,



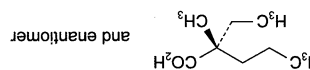
H. pentanenitrile (valeronitrile),



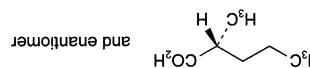
I. 2-propylpentanenitrile,



J. 2,2-dipropylpentanenitrile,



K. (2RS)-2-ethyl-2-methylpentanoic acid,



L. (2RS)-2-methylpentanoic acid.

Ph Eur

Water (2.5.12)
Maximum 2.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.170 g in 70 mL of 2-propanol R. Titrate with 0.1 M tetrabutylammonium hydroxide in 2-propanol, determining the endpoint potentiometrically (2.2.20). Perform all operations under nitrogen.
1 mL of 0.1 M tetrabutylammonium hydroxide in 2-propanol is equivalent to 21.78 mg of $C_{24}H_{29}N_5O_3$.

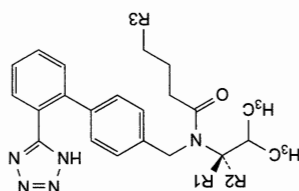
STORAGE

In an airtight container.

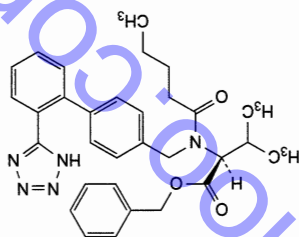
IMPURITIES

Specified impurities A, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B.



A. R1 = H, R2 = CO₂H, R3 = CH₃; (2R)-3-methyl-2-[pentanoyl[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]amino]butanoic acid.
C. R1 = CO₂H, R2 = R3 = H; (2S)-2-[butanoyl[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]amino]butanoic acid.



B. benzyl (2S)-3-methyl-2-[pentanoyl[[2'-(1H-tetrazol-5-yl)biphenyl-4-yl]methyl]amino]butanoate.

Ph Eur

Identification of impurities Use the chromatogram supplied with valsartan for peak identification CRS and the chromatogram obtained with reference solution (a) to identify the peak due to impurity A.

Relative retention With reference to valsartan (retention time = about 13 min): impurity A = about 0.6.

System suitability: reference solution (a):
— resolution: minimum 2.0 between the peaks due to impurity A and valsartan.

Limit:
— impurity A: not more than the area of the principal peak in the chromatogram obtained with reference solution (b) (1.0 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50 mg of the substance to be examined in the mobile phase and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of valsartan for system suitability CRS (containing impurity C) in 1.0 mL of the mobile phase.

Column:
— size: $l = 0.125$ m, $\phi = 3.0$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m).

Mobile phase glacial acetic acid R, acetonitrile R1, water R (1:500:500 V/V/V).

Flow rate 0.4 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 10 μ L.

Run time 6 times the retention time of valsartan.

Identification of impurities Use the chromatogram supplied with valsartan for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peak due to impurity C.

Relative retention With reference to valsartan (retention time = about 5 min): impurity C = about 0.8.

System suitability: reference solution (b):
— resolution: minimum 3.0 between the peaks due to impurity C and valsartan.

Limit:
— impurity C: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— total: not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

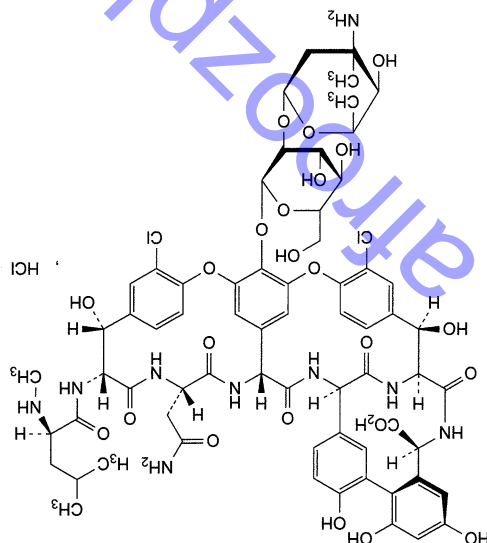
— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)
Maximum 20 ppm.

Dissolve 1.0 g in a mixture of 15 volumes of water R and 85 volumes of acetone R and dilute to 20 mL with the same mixture of solvents. 12 mL of the solution complies with test B. Prepare the reference solution using 10 mL of lead standard solution (1 ppm Pb) R.

Vancomycin Hydrochloride

(Ph. Eur. monograph 1058)



$C_{66}H_{76}ClN_9O_{24}$ 1486 1404-93-9

Action and use
Glycopeptide antibacterial.

Preparations
Vancomycin Capsules
Vancomycin Eye Drops
Vancomycin Infusion
Vancomycin Oral Solution

Ph Eur

DEFINITION

Hydrochloride of a mixture of related glycopeptides, consisting principally of the monohydrochloride of

(3S,6R,7R,22R,23S,26S,30aS,36R,38aR)-3-(2-amino-2-oxoethyl)-44-[[2-O-(3-amino-2,3,6-trideoxy-3-C-methyl-α-L-lyxo-hexopyranosyl)-β-D-glucopyranosyl]oxy]-10,19-dichloro-7,22,28,30,32-pentahydroxy-6-[[[(2R)-4-methyl-2-(methylanino)pentanoyl]amino]-2,5,24,38,39-pentaoxo-2,3,4,5,6,7,23,24,25,26,36,37,38,38a-tetradecahydro-22H-8,11:18,21-dietheno-23,36-(iminomethano)-13,16:31,35-dimetheno-1H,13H-1,6,9]oxadiazacyclohexadecimo[4,5-*m*][10,2,16]benzoxazadiazacyclotetracosine-26-carboxylic acid (vancomycin B).

Substance produced by certain strains of *Amorycolatopsis orientalis* or obtained by any other means.

Potency

Minimum 1050 IU/mg (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic powder.

Solubility

Freely soluble in water, slightly soluble in ethanol

(96 per cent).

IDENTIFICATION

A. Examine the chromatograms obtained in the test for vancomycin B.

Results The principal peak in the chromatogram obtained with test solution (a) is similar in retention time to the principal peak in the chromatogram obtained with reference solution.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and its absorbance (2.2.25) at 450 nm is not greater than 0.10.

Dissolve 2.50 g in water R and dilute to 25.0 mL with the same solvent.

pH (2.2.3)

2.5 to 4.5.

Dissolve 0.50 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Vancomycin B

Liquid chromatography (2.2.29). Use the solutions within 4 h of preparation.

Test solution (a) Dissolve 10.0 mg of the substance to be examined in mobile phase A and dilute to 5.0 mL with mobile phase A.

Test solution (b) Dilute 2.0 mL of test solution (a) to 50.0 mL with mobile phase A.

Test solution (c) Dilute 0.5 mL of test solution (b) to 20.0 mL with mobile phase A.

Reference solution Dissolve the contents of a vial of vancomycin hydrochloride CRS in water R and dilute with the same solvent to obtain a solution containing 0.5 mg/mL. Heat at 65 °C for 24 h. Allow to cool.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase:

mobile phase A: to 4 mL of methylanino R add 1996 mL of water R and adjust to pH 3.2 with phosphoric acid R;

mobile phase B: to 4 mL of methylanino R add 1996 mL of water R and adjust to pH 3.2 with phosphoric acid R;

tetrahydrofuran R and 70 mL of acetonitrile R;

to 920 mL of this solution add 10 mL of

of water R and adjust to pH 3.2 with phosphoric acid R;

to 700 mL of this solution add 10 mL of

tetrahydrofuran R and 290 mL of acetonitrile R;

Mobile phase B

Time (min)

0 - 13

100

100 → 0

0 → 100

Mobile phase A

Mobile phase B

(per cent V/V)

(per cent V/V)

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 20 µL.

System suitability:

— resolution: minimum 5.0 between the 2 principal peaks in the chromatogram obtained with the reference solution;

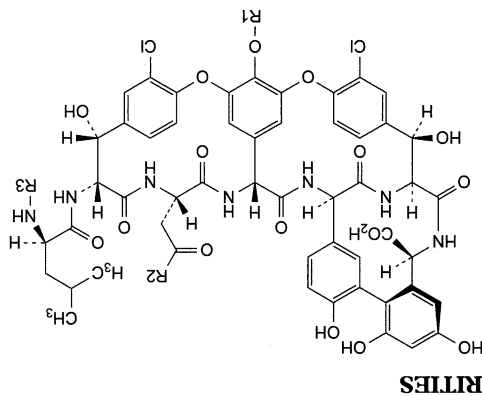
— signal-to-noise ratio: minimum 5 for the principal peak in the chromatogram obtained with test solution (c);

— symmetry factor: maximum 1.6 for the peak due to vancomycin in the chromatogram obtained with test

solution (b).

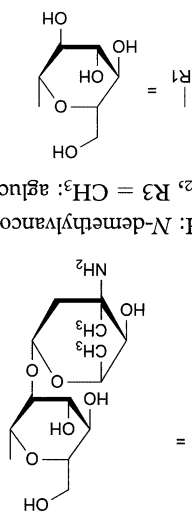
Calculate the percentage content of vancomycin B

hydrochloride using the following expression:

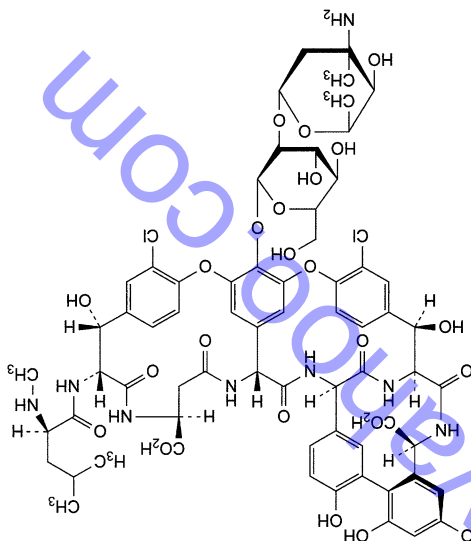


IMPURITIES

A. R₂ = NH₂, R₃ = H: N-demethylvancomycin B,
C. R₁ = H, R₂ = NH₂, R₃ = CH₃: aglucovancomycin B,



D. R₂ = NH₂, R₃ = CH₃: desvancomasaminyivancomycin B,



B. (4S,7R,8R,23R,24S,27S,31aS,37R,39aR)-45-[[2-O-(3-amino-2,3,6-trideoxy-3-C-methyl-α-L-lyxo-hexopyranosyl)-β-D-glucopyranosyl]oxy]-11,20-dichloro-8,23,29,31,33-pentahydroxy-7-[[[(2R)-4-methyl-2-(methylamino)pentanoyl]amino]-2,6,25,39,40-pentaoso-1,2,3,4,5,6,7,8,24,25,26,27,37,38,39,39a-hexadecahydro-12,19,22-dietheno-24,37-(iminomethano)-14,17:32,36-dimetheno-14H-[1,6,10]oxadiazacycloheptadecimo[4,5-m][10,2,16]benzoxadiazacyclotetradecose-4,27-dicarboxylic acid ([β Asp]³vancomycin B).

$$\frac{A_b + \left(\frac{A_i}{25}\right)}{A_b \times 100}$$

A_b = area of the peak due to vancomycin B in the chromatogram obtained with test solution (b);
 A_i = sum of the areas of the peaks due to impurities in the chromatogram obtained with test solution (a).

Limit:

— vancomycin B: minimum 93.0 per cent.

Related substances

Liquid chromatography (2.2.29) as described in the test for vancomycin B with the following modifications.

Injection Test solution (a), (b) and (c).

Calculate the percentage content of each impurity using the following expression:

$$\left(\frac{A_i}{25}\right) \times 100$$

A_i = area of the peak due to an impurity in the

chromatogram obtained with test solution (a);

A_b = area of the peak due to vancomycin B in the

chromatogram obtained with test solution (b);

A_i = sum of the areas of the peaks due to impurities in the chromatogram obtained with test solution (a).

Limits:

— any impurity: for each impurity, maximum 4.0 per cent;

— total: maximum 7.0 per cent;

— disregard limit: the area of the principal peak in the chromatogram obtained with test solution (c)

Heavy metals (2.4.8)

Maximum 30 ppm.

1.0 g complies with test C. Prepare the reference solution using 3.0 mL of lead standard solution (10 ppm Pb) R.

Water (2.5.12)

Maximum 5.0 per cent, determined on 0.500 g.

Sulfated ash (2.4.14)

Maximum 1.0 per cent, determined on 1.00 g.

Bacterial endotoxins (2.6.14)

Less than 0.25 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

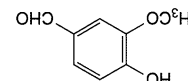
Carry out the microbiological assay of antibiotics (2.7.2). Use vancomycin hydrochloride CRS as the chemical reference substance.

STORAGE

In an airtight container, protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

Vanillin

(Ph. Eur. monograph 0747)



C₈H₈O₃ 152.1 121-33-5

DEFINITION

Vanillin contains not less than 99.0 per cent and not more than the equivalent of 101.0 per cent of 4-hydroxy-3-methoxybenzaldehyde, calculated with reference to the dried substance.

CHARACTERS

White or slightly yellowish, crystalline powder or needles, slightly soluble in water, freely soluble in alcohol and in methanol. It dissolves in dilute solutions of alkali hydroxides.

IDENTIFICATION

First identification B.
A. Melting point (2.2.14): 81 °C to 84 °C.
B. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the spectrum obtained with *vanillin* CRS. Examine the substances prepared as discs. C. Examine the chromatograms obtained in the test for related substances in daylight after spraying. The principal spot in the chromatogram obtained with test solution (b) is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a). D. To 5 mL of a saturated solution of the substance to be examined add 0.2 mL of ferric chloride solution R1. A blue colour is produced. Heat to 80 °C. The solution becomes brown. On cooling, a white precipitate is formed.

TESTS

Appearance of solution
Dissolve 1.0 g in *alcohol* R and dilute to 20 mL with the same solvent. The solution is clear (2.2.1) and not more intensely coloured than reference solution B₆ (2.2.2, Method II).

Related substances

Examine by thin-layer chromatography (2.2.27), using *silica gel* GF₂₅₄ R as the coating substance.

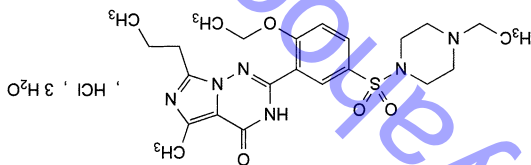
Test solution (a) Dissolve 0.1 g of the substance to be examined in *methanol* R and dilute to 5 mL with the same solvent. **Test solution (b)** Dilute 1 mL of test solution (a) to 10 mL with *methanol* R.

Reference solution (a) Dissolve 10 mg of *vanillin* CRS in *methanol* R and dilute to 5 mL with the same solvent. **Reference solution (b)** Dilute 0.5 mL of test solution (a) to 100 mL with *methanol* R.

Apply to the plate 5 µL of each solution. Develop in an unsaturated tank over a path of 10 cm using a mixture of 0.5 volumes of *anhydrous acetic acid* R, 1 volume of *methanol* R and 98.5 volumes of *methylene chloride* R. Dry the plate in a current of cold air. Examine in ultraviolet light at 254 nm. Any spot in the chromatogram obtained with test solution (a), apart from the principal spot, is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.5 per cent). Spray with

Vardenafil Hydrochloride, Trihydrate

(Ph. Eur. monograph 2782)



C₂₃H₃₃ClN₆O₄S₃H₂O 579.1 330808-88-3

Action and use

Selective inhibitor of cyclic GMP-specific phosphodiesterase type V with vasodilator action; treatment of erectile dysfunction

DEFINITION

2-[2-Ethoxy-5-[(4-ethoxycarbonyl-1-yl)sulfonyl]phenyl]-5-methyl-7-propylimidazo[5,1-f][1,2,4]triazin-4(3H)-one hydrochloride trihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance
White or slightly brown or yellow powder.

Solubility

Slightly soluble in water, freely soluble in anhydrous ethanol, practically insoluble in heptane.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Comparison vardenafil hydrochloride CRS.

B. Water (see Tests).

C. It gives reaction (a) of chlorides (2.3.1).

TESTS

Related substances

Liquid chromatography (2.2.29). Protect the solutions from light.

Solvent mixture acetonitrile R₁, mobile phase A (20:80 V/V). Test solution (a) Dissolve 50.0 mg of the substance to be examined in 20 mL of acetonitrile R₁ and dilute to 100.0 mL with mobile phase A.

Test solution (b) Dilute 15.0 mL of test solution (a) to 50.0 mL with the solvent mixture. Reference solution (a) Dissolve 50.0 mg of vardenafil hydrochloride CRS in 20 mL of acetonitrile R₁ and dilute to 100.0 mL with mobile phase A. Dilute 15.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (b) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (c) Dissolve 5 mg of vardenafil for system suitability CRS (containing impurity A) in 2 mL of acetonitrile R₁ and dilute to 10 mL with mobile phase A.

Column:

— size: $l = 0.25$ m, $\varnothing = 3.0$ mm;

— stationary phase: end-capped polar-embedded octadecylsilyl amorphous organosilica polymer R (5 μ m);

— temperature: 45 °C.

Mobile phase:

— mobile phase A: solution containing 0.7 g/L of disodium hydrogen phosphate dihydrate R and 1.3 g/L of potassium dihydrogen phosphate R₃

— mobile phase B: acetonitrile R₁;

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 2	80	20
2 - 22	80 \rightarrow 25	20 \rightarrow 75
22 - 27	25	75

Flow rate 0.5 mL/min.

Detection Spectrophotometer at 242 nm.

Injection 10 μ L of test solution (a) and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with vardenafil for system suitability CRS and the

chromatogram obtained with reference solution (c) to identify the peak due to impurity A.

Relative retention With reference to vardenafil (retention time = about 16 min): impurity A = about 0.8.

System suitability: reference solution (c):

— resolution: minimum 5.0 between the peaks due to impurity A and vardenafil.

Calculation of percentage contents:

— for each impurity, use the concentration of vardenafil in reference solution (b).

Limits:
— impurity A: maximum 0.15 per cent;
— unspecified impurities: for each impurity, maximum 0.10 per cent;
— total: maximum 0.3 per cent;
— reporting threshold: 0.05 per cent.
Sulfates (2.4.13)
Maximum 400 ppm.
Suspend 0.5 g in 20 mL of a 5.15 g/L solution of hydrochloric acid R and stir for 15 min. Filter if complete dissolution is not obtained.
Water (2.5.12)
8.8 per cent to 10.5 per cent, determined on 60.0 mg.
Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

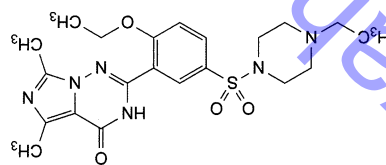
ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.
Injection 10 μ L of test solution (b) and reference solution (a). Calculate the percentage content of C₂₃H₃₃N₆O₄S taking into account the assigned content of vardenafil hydrochloride CRS.

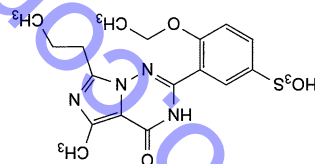
IMPURITIES

Specified impurities A

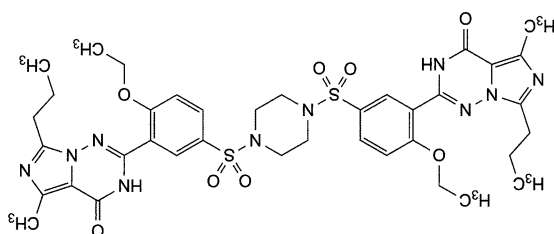
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: B, C.



A. 2-[2-ethoxy-5-[(4-ethylpiperazin-1-yl)sulfonyl]phenyl]-5,7-dimethylimidazo[5,1-b][1,2,4]triazin-4(3H)-one,



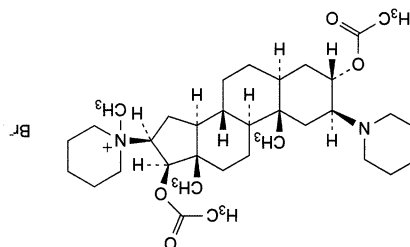
B. 4-ethoxy-3-(5-methyl-4-oxo-7-propyl-3,4-



C. 2,2'-[piperazine-1,4-diylbis[(sulfonyl)(4-ethoxybenzene-1,3-diyl)]]bis[5-methyl-7-propylimidazo[5,1-b][1,2,4]triazin-4(3H)-one].

Vecuronium Bromide

(Ph. Eur. monograph 1769)


 $\text{C}_{34}\text{H}_{57}\text{BrN}_2\text{O}_4$ 638 50700-72-6

Action and use

Non-depolarizing neuromuscular blocker.

Preparation

Vecuronium Bromide for Injection

Ph Eur

DEFINITION

1-[3 α ,17 β -Bis(acetyloxy)-2 β -(piperidin-1-yl)-5 α -androstan-16 β -yl]-1-methylpiperidinium bromide.

Content

99.0 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white crystals or crystalline powder.

Solubility

Slightly soluble in water, freely soluble in methylene chloride, sparingly soluble in acetonitrile and in anhydrous ethanol.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison vecuronium bromide CRS.

C. It gives reaction (a) of bromides (2.3.1).

TESTS

Solution S

Dissolve 0.500 g in a 5.15 g/L solution of hydrochloric acid R and dilute to 50.0 mL with the same solution.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Specific optical rotation (2.2.7)

+ 30.5 to + 35.0 (anhydrous substance), determined on solution S.

Impurity B

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.10 g of the substance to be examined in methylene chloride R and dilute to 5.0 mL with the same solvent.

Reference solution (a) Dissolve 5 mg of the substance to be examined and 5 mg of pancuronium bromide CRS (impurity B) in methylene chloride R and dilute to 5 mL with the same solvent.

Reference solution (b) Dissolve 5.0 mg of pancuronium bromide CRS (impurity B) in methylene chloride R and dilute to 100.0 mL with the same solvent.

Stationary phase TLC silica gel plate R (2-10 μm).
Mobile phase Dissolve 1 g of sodium bromide R in 5 mL of water R. Add 85 mL of 2-propanol R, then 10 mL of acetonitrile R.

Application 1 μL .
Development In an unsaturated tank, over 2/3 of the plate.
Drying In air for 30 min.
Detection Spray with a 2.5 g/L solution of iodine R in a mixture of equal volumes of methanol R and methylene chloride R.
System suitability: reference solution (a):
— the chromatogram obtained shows 2 clearly separated spots.
Limit:
— impurity B: any spot due to impurity B is not more intense than the spot in the chromatogram obtained with reference solution (b) (0.25 per cent).
Related substances
Liquid chromatography (2.2.29). Use freshly prepared solutions.
Test solution Dissolve 40.0 mg of the substance to be examined in a 0.2 g/L solution of hydrochloric acid R in methanol R and dilute to 20.0 mL with the same solution.
Reference solution (a) Dissolve 4 mg of vecuronium for peak identification CRS (containing impurities A, C, D and E) in a 0.2 g/L solution of hydrochloric acid R in methanol R and dilute to 2 mL with the same solution.
Reference solution (b) Dilute 5.0 mL of the test solution to 100.0 mL with a 0.2 g/L solution of hydrochloric acid R in methanol R. Dilute 5.0 mL of this solution to 100.0 mL with a 0.2 g/L solution of hydrochloric acid R in methanol R.
Reference solution (c) Dilute 10.0 mL of reference solution (b) to 50.0 mL with a 0.2 g/L solution of hydrochloric acid R in methanol R.
Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μm);
— temperature: 40 °C.
Mobile phase Mix 50 volumes of an 18.0 g/L solution of tetramethylammonium hydroxide R adjusted to pH 6.5 with phosphoric acid R, 250 volumes of methanol R and 700 volumes of acetonitrile R.
Flow rate 2.0 mL/min.
Detection Spectrophotometer at 210 nm.
Injection 20 μL .
Run time 2.5 times the retention time of vecuronium.
Identification of impurities Use the chromatogram supplied with vecuronium for peak identification CRS and the chromatogram obtained with reference solution (a) to identify the peaks due to impurities A, C, D and E. The elution order may vary, but the quantity of each impurity in the CRS is different so that a clear identification of the impurities is possible.
Relative retention With reference to vecuronium (retention time = about 5 min): impurity C = about 0.8; impurity D = about 0.9; impurity E = about 1.2; impurity A = about 1.3.
System suitability: reference solution (a):
— peak-to-valley ratio: minimum 2.0, where H_p = height above the baseline of the peak due to impurity D and H_o = height above the baseline of the principal peak; if necessary, increase the volume of the buffer solution while simultaneously decreasing the volume of acetonitrile in the mobile phases; do not change the volume of methanol;
— symmetry factor: maximum 3.5 for the principal peak.

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity A = 0.6;

— **impurities A, C, D, E:** for each impurity, not more than the area of the principal peak in the chromatogram

obtained with reference solution (b) (0.25 per cent);

— **unspecified impurities:** for each impurity, not more than twice the area of the principal peak in the chromatogram

obtained with reference solution (c) (0.10 per cent);

— **total:** not more than 2.8 times the area of the principal

peak in the chromatogram obtained with reference

solution (b) (0.7 per cent);

— **disregard limit:** the area of the principal peak in the

chromatogram obtained with reference solution (c) (0.05 per cent).

Water (2.5.12)

Maximum 4.0 per cent, determined on 0.300 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.450 g in 50 mL of *glacial acetic acid R*. Titrate with 0.1 M *perchloric acid*, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M *perchloric acid* is equivalent to 63.8 mg

of $C_{34}H_{57}BrN_2O_4$.

STORAGE

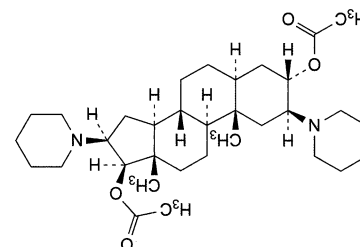
In an airtight container, protected from light and moisture.

IMPURITIES

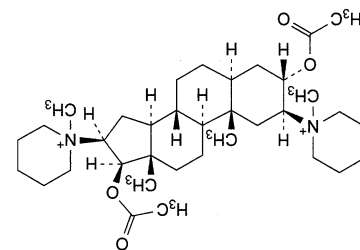
Specified impurities A, B, C, D, E

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): F.



A. 2β,16β-bis(piperidin-1-yl)-5α-androstane-3α,17β-diol diacetate,



B. 1,1'-[3α,17β-bis(acetyloxy)-5α-androstane-2β,16β-diyl]bis(1-methylpiperidinium) (pancuronium),

DEFINITION

Vegetable fatty oils are mainly solid or liquid triglycerides of fatty acids. They may contain small amounts of other lipids such as waxes, free fatty acids, partial glycerides or unsaponifiable matters. Vegetable fatty oils are obtained from the seeds, the fruit or the pit/stone/kernel of various plants by expression and/or solvent extraction, then possibly refined by bleaching and any deodorisation) or physical refining.

Virgin oil An oil obtained from raw materials of special quality by mechanical procedures (e.g. by cold expression or centrifugation).

Refined oil An oil obtained by expression and/or solvent extraction, and subsequently either alkali refining (followed by bleaching and any deodorisation) or physical refining.

Hydrogenated oil An oil obtained by expression and/or solvent extraction, and subsequently either alkali refining or physical

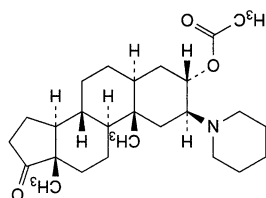
Vegetable Fatty Oils

(Ph. Eur. monograph 1579)

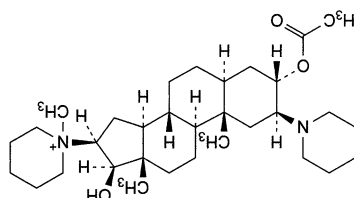
Ph. Eur.



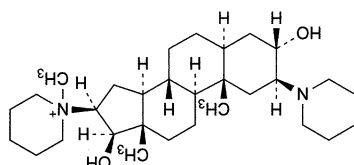
F. 2β-(piperidin-1-yl)-17-oxo-5α-androstane-3α-yl acetate.



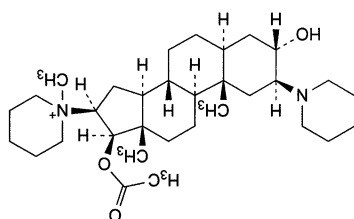
E. 1-[3α-(acetyloxy)-17β-hydroxy-2β-(piperidin-1-yl)-5α-androstane-16β-yl]-1-methylpiperidinium,



D. 1-[3α,17β-dihydroxy-2β-(piperidin-1-yl)-5α-androstane-16β-yl]-1-methylpiperidinium,



C. 1-[17β-(acetyloxy)-3α-hydroxy-2β-(piperidin-1-yl)-5α-androstane-16β-yl]-1-methylpiperidinium,



refining, then possible bleaching, followed by drying,

hydrogenation and subsequent bleaching and deodorisation.

Only alkali-refined oils are used in the manufacture of

parenteral preparations.

PRODUCTION

Measures are taken to ensure that the oil complies with the

limit for benzo[a]pyrene decided by the competent authority.

A limit of 2.0 ppb is set in Commission Regulation (EC)

No. 208/2005.

OBTENTION OF A CRUDE OIL

Where the plant has a high oil content, the oil is generally

obtained by expression under heating followed by an

extraction; where the plant has a low oil content, the oil is

generally obtained by direct extraction.

Mechanical procedures

A. Expression

High-pressure screw-pressing It consists of some or all of the

following steps: cleaning, drying, dehulling or decorticating,

grinding, cooking and flaking.

During *cleaning* the foreign matter is eliminated. *Drying* may

be necessary if the seed moisture content is higher than

desirable for downstream processing. *Decorticating* is useful to

obtain a high-protein meal by reduction of fibre and to

reduce impurities in the oil. *Cooking* serves various purposes:

completion of the breakdown of oil cells, lowering of the

viscosity of the oil, coagulation of the protein in the meal,

adjustment of the moisture level, sterilisation of the seed,

detoxifying undesirable seed constituents (gossypol for

cottonseed) and fixing certain phosphatides in the cake thus

lowering subsequent refining losses. The efficacy of the

expression process is such that only 3 per cent to 6 per cent

of the oil is left in the cake.

Hot screw-pressing The bunches are loaded into cages (for

palm fruit) and moved into a horizontal steriliser with

application of live steam and heating. The purposes of this

steriliser are inactivation of enzymes, loosening of the fruit on

the bunch, coagulation of proteins, etc. After heating in a

digestor, the pulp is fed to a screw-press. The oil is

centrifugally clarified and vacuum-dried.

Pre-pressing followed by solvent extraction The same sequence of

steps is performed as above. The main function of pre-

pressing is to obtain a cake of excellent permeability for the

following solvent extraction stage. The extraction is

performed either in a percolation-type or in an immersion-

type apparatus. The efficacy of the solvent extraction process

is such that residual oil levels in meal are generally below

1 per cent.

B. Centrifugation

Centrifugation separates the oily phase from the aqueous

phases, which contains water-soluble components and residual

solid particles. This operation can be carried out using:

— self-cleaning bowl or disc centrifuges;

— super-decanter, which are horizontal turbines equipped

with a cylindrical bowl that tapers slightly at one end and

which contains a continuously turning screw that scrapes

the sides of the bowl; the screw and the bowl rotate at

different speeds; the solid particles are discarded from the

tapered end of the bowl and the oil flows out from the

other end.

Solvent extraction

Prior to extraction, the following steps are carried out: the

seeds are tempered for about a week at a temperature below

24 °C in order to loosen the hull from the seed and allow the

seed moisture to attain equilibrium, then the seeds are

cleaned, ground, dehulled and flaked. The most widely used

solvent is a mixture of mainly *n*-hexane and methylpentanes

(bp: 65–70 °C) commonly referred to as 'hexane'. Due to the

major fire and explosive risks of this mixture, liquified gases

and supercritical gases may also be used.

REFINING

The objective of refining is to remove impurities and

contaminants of the oil with the least possible damage to the

triglycerides and with minimal loss of oil. The contents of the

following substances are reduced:

— free fatty acids, which may cause deterioration of the oil

by oxidation, a smoked taste when heated and a sharp

flavour (by alkali refining);

— water, which favours the enzymatic hydrolysis reactions

(by alkali refining, drying);

— partial glycerides, which may cause foaming and a bitter

taste (by neutralisation, washing);

— phosphatides and phosphorous compounds, which have

emulsifying properties and may cause deposits, a

darkening of the oil when heated, a cloudy appearance

and bad organoleptic stability (by alkali refining);

— colouring matters such as chlorophyll (by alkali refining)

and carotenoids (by bleaching);

— glycolipids, which may form colloidal solutions with

water;

— free hydrocarbons, paraffin, waxes and resinous materials;

— metals (Fe, Cu, Pb, Sn, P, etc.), which are strong

oxidation catalysts;

— pigments such as gossypol (in cottonseed oil) or

mycotoxins such as aflatoxin (mainly in arachis seeds);

— pesticides;

— oxidation products (aldehydes, peroxides);

— proteins having possible allergic reactions;

— unsaponifiable matters (sterols, tocopherols and other

vitamins);

— polycyclic aromatic hydrocarbons.

Alkali refining

It involves the following steps: degumming if necessary,

neutralisation using alkali, washing and drying.

Degumming During this step of the refining, i.e. treatment

with water and/or phosphoric acid and/or sodium chloride,

the phosphatides, phosphorous compounds and metals are

eliminated. The use of this step depends on the nature of the

oil.

Neutralisation with alkali This step reduces the free-fatty-acid

content below 0.1 per cent; the fatty acids are converted into

oil-insoluble soaps, also called 'soapsacks'. Other substances

may be removed by adsorption on these soaps: mucilaginous

substances, phosphatides, oxidation products, colouring

matters, etc. All substances that become insoluble in the oil

on hydration are removed. Neutralisation with alkali has the

disadvantage of saponifying a portion of neutral oil if the

neutralisation is not well conducted.

Washing This operation consists in removing the excess of

soaps and alkali as well as the remaining traces of metals,

phosphatides and other impurities, using hot water.

Drying The remaining water is eliminated under vacuum

Physical refining

It involves a steam treatment of the oil under high vacuum at

a temperature greater than 235 °C. This technique can only

be applied to oils naturally low in phosphatides and metals

(palm and coconut) or from which phosphatides and metals

Hydrogenated Vegetable Oil

68334-00-9

Action and use

Excipient.

DEFINITION

Hydrogenated Vegetable Oil is a mixture of triglycerides of fatty acids of vegetable origin.

CHARACTERISTICS

An almost white, fine powder at room temperature and a pale yellow, oily liquid above its melting point. Practically insoluble in *water*; soluble in *hexane* and in hot *propan-2-ol*.

IDENTIFICATION

Complies with the tests for Acid value, Iodine value and Saponification value.

TESTS

Melting point
57° to 70°, Appendix V A.

Acid value
Not more than 4.0, Appendix X B.

Iodine value
Not more than 5, Appendix X E, Method B.

Saponification value
175 to 205, Appendix X G, Method II.

Unsaponifiable matter
Not more than 0.8% w/w, Appendix X H.

Heavy metals

2.0 g complies with *limit test C for heavy metals*, Appendix VII. Use 2 mL of *lead standard solution* (10 ppm Pb) to prepare the standard (10 ppm).

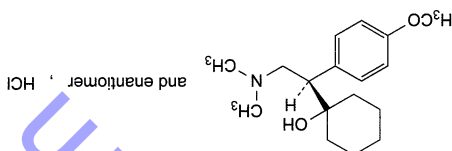
Loss on drying
When dried at 105° for 4 hours, loses not more than 0.1% of its weight. Use 5 g.

STORAGE

Hydrogenated Vegetable Oil should be stored at a temperature of 8° to 25°.

Venlafaxine Hydrochloride

(Ph. Eur. monograph 2119)

C₁₇H₂₈ClNO₂

313.9

99300-78-4

Action and use

Inhibition of 5HT and noradrenaline reuptake; antidepressant.

Preparations

Prolonged-release Venlafaxine Capsules
Prolonged-release Venlafaxine Tablets
Venlafaxine Tablets

phosphoric acid followed by an adsorptive treatment with activated bleaching earth (for sunflower, rapeseed, soyabean). Moreover, it cannot be used for heat-sensitive oils (cottonseed oil), which darken.

Bleaching

The common method of bleaching is by adsorption treatment of the oil, which is generally heated at 90 °C for 30 min under vacuum, with bleaching earth (natural or activated) or carbon (activated or not); synthetic silica adsorbents may also be added. Substances that have not been totally removed during refining are eliminated, for example carotenoids and chlorophyll.

Deodorisation

Deodorisation eliminates odours, volatile substances and any residual extraction solvents; it involves injecting dry vapour into the oil, which is kept under vacuum at a high temperature. Different temperatures are used according to the oil: 200-235 °C for 1.5-3 h or greater than 240 °C for 30 min.

One of the main side reactions is thermic decolourisation due to the destruction of carotenoids when the temperature is greater than 150 °C. This technique provokes a loss of substances that may be distilled (free fatty acids, sterols, tocopherols, part of the refined oil), and may cause *cis-trans* isomerisation of the unsaturated fatty-acid double bonds.

WINTERISATION

Elimination of solids and waxes by filtration at low temperature (also called dewaxing). These solids and waxes could affect the appearance of the oil and cause deposits.

HYDROGENATION

The hydrogenation of the dried and/or bleached oil is performed using a catalyst (e.g. Ni, Pt, Pd), at a temperature of about 100-200 °C under hydrogen pressure. The catalyst is then removed by filtration at 90 °C. The hydrogen must be pure: free of poisons for the catalyst, water-free, and low in carbon dioxide, methane and nitrogen contents. Small amounts of polymers may be obtained. *Trans*-fatty acids are formed during partial hydrogenation.

CHROMATOGRAPHIC PURIFICATION

In high-purity applications, mainly for parenteral uses, the oil may be further purified by passing the oil through a column containing an activated earth. A solvent may sometimes be used to improve the efficiency. High-polarity molecules, such as oxidised materials, acids, alcohols, partial glycerides and free sterols, are preferentially removed. When the oil is used in the manufacture of parenteral preparations, the limits set in the monograph for the acid value, the peroxide value and the water content may be different.

LABELLING

The label states:

— where applicable, that the oil was obtained by expression or extraction;
— where applicable, that the oil is suitable for use in the manufacture of parenteral preparations.

Ph Eur

DEFINITION

1-[(1*R,S*)-2-(*D*-dimethylamino)-1-(4-methoxyphenyl)ethyl]cyclohexanol hydrochloride.

CHARACTERS

99.0 per cent to 101.0 per cent (dried substance).

Appearance

White or almost white powder.

Solubility

Freely soluble in water and in methanol, soluble in anhydrous ethanol, slightly soluble or practically insoluble in acetone.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison venlafaxine hydrochloride CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in 2-propanol R, evaporate to dryness and record new spectra using the residues.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Acidity or alkalinity

Dissolve 0.20 g in carbon dioxide-free water R and dilute to 10 mL with the same solvent. Add 0.05 mL of methyl red solution R and 0.1 mL of 0.01 M hydrochloric acid.

The solution is pink. Not more than 0.2 mL of 0.01 M sodium hydroxide is required to change the colour of the indicator to yellow.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in the mobile phase and dilute to 25.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 10.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve the contents of a vial of venlafaxine for system suitability CRS (containing impurities D and F) in 1.0 mL of the mobile phase.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: end-capped octylsilyl silica gel for chromatography R (5 μ m) with a pore size of 10 nm.

Mobile phase Mix 510 volumes of acetonitrile R and 1490 volumes of a solution prepared as follows: dissolve 17 g of ammonium dihydrogen phosphate R in 1490 mL of water R and adjust to pH 4.4 using phosphoric acid R.

Flow rate 1.2 mL/min.

Detection Spectrophotometer at 225 nm.

Injection 20 μ L.

Run time 10 times the retention time of venlafaxine.

Relative retention With reference to venlafaxine (retention time = about 9 min): impurity D = about 0.9;

impurity F = about 3.4.

System suitability: reference solution (b):

— resolution: minimum 1.5 between the peaks due to impurity D and venlafaxine.

Limits:

— impurity F: not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.1 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (a) (0.2 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

Heavy metals (2.4.8)

Maximum 20 ppm.

Dissolve 1.0 g in 20 mL of water R. 12 mL of the solution complies with test A. Prepare the reference solution using lead standard solution (1 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying *in vacuo* at 80 °C for 3 h.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.250 g in a mixture of 5.0 mL of 0.01 M hydrochloric acid and 50 mL of ethanol (96 per cent) R. Carry out a potentiometric titration (2.2.20), using 0.1 M sodium hydroxide. Read the volume added between the 2 points of inflexion. Carry out a blank titration.

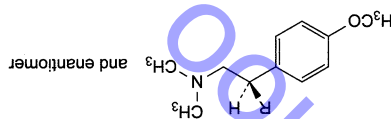
1 mL of 0.1 M sodium hydroxide is equivalent to 31.39 mg of $C_{17}H_{28}ClNO_2$.

IMPURITIES

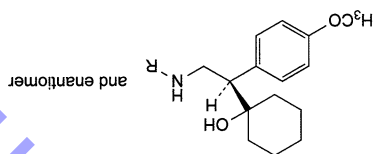
Specified impurities F

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use): A, B, C, D, E, F, G, H.



A. R = H: 2-(4-methoxyphenyl)-N,N-dimethylethanamine, B. R = CO-O-C₂H₅: ethyl (2*R,S*)-3-(dimethylamino)-2-(4-methoxyphenyl)propanoate,



C. R = H: 1-[(1*R,S*)-2-amino-1-(4-methoxyphenyl)ethyl]cyclohexanol, D. R = CH₃: 1-[(1*R,S*)-1-(4-methoxyphenyl)-2-(methylamino)ethyl]cyclohexanol, H. R = CH₂-CH₂-C₆H₄-p-OCH₃: 1-[(1*R,S*)-1-(4-methoxyphenyl)-2-[(2-(4-methoxyphenyl)amino)ethyl]cyclohexanol,

Solubility
Soluble in water, freely soluble in methanol, sparingly soluble in ethanol (96 per cent).
mp: about 144 °C.

IDENTIFICATION

First identification B, D
Second identification A, C, D
A. Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution Dissolve 20.0 mg in 0.01 M hydrochloric acid and dilute to 100.0 mL with the same acid. Dilute 5.0 mL of this solution to 50.0 mL with 0.01 M hydrochloric acid.
Spectral range 210–340 nm.
Absorption maxima At 229 nm and 278 nm.
Shoulder At 282 nm.

Absorbance ratio $A_{278}/A_{229} = 0.35$ to 0.39.

B. Infrared absorption spectrophotometry (2.2.24).
Comparison verapamil hydrochloride CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 5 mL with the same solvent.

Reference solution (a) Dissolve 20 mg of verapamil hydrochloride CRS in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 5 mg of papaverine hydrochloride CRS in reference solution (a) and dilute to 5 mL with the same solution.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase diethylamine R, cyclohexane R (15:85 V/V).

Application 5 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated principal spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference solution (a).

D. It gives reaction (b) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 1.0 g in carbon dioxide-free water R while gently heating and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.5 to 6.0 for solution S.

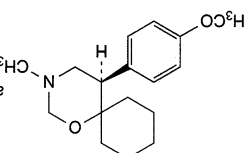
Optical rotation (2.2.7)

–0.10° to +0.10°, determined on solution S.

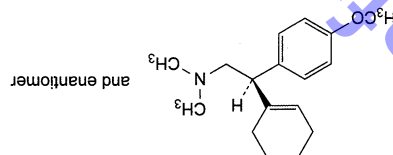
Related substances

Liquid chromatography (2.2.29).

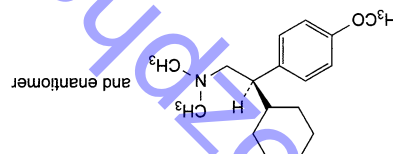
Solvent mixture Mobile phase B, mobile phase A (37:63 V/V).
Test solution Dissolve 25.0 mg of the substance to be examined in the solvent mixture and dilute to 10.0 mL with the solvent mixture.



E. (5RS)-5-(4-methoxyphenyl)-3-methyl-1-oxa-3-azaspiro[5.5]undecane,



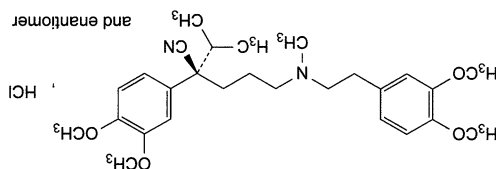
F. (2RS)-2-(cyclohex-1-enyl)-2-(4-methoxyphenyl)-N,N-dimethylethanamine,



G. (2RS)-2-cyclohexyl-2-(4-methoxyphenyl)-N,N-dimethylethanamine.

Verapamil Hydrochloride

(Ph. Eur. monograph 0573)



C₂₇H₃₉ClN₂O₄ 491.1 152-11-4

Action and use
Calcium channel blocker.
Preparations
Verapamil Injection
Verapamil Tablets
Prolonged-release Verapamil Capsules
Prolonged-release Verapamil Tablets

Ph. Eur.

DEFINITION

(2RS)-2-(3,4-Dimethoxyphenyl)-5-[[2-(3,4-dimethoxyphenyl)ethyl](methyl)amino]-2-(1-methylethyl)pentanenitrile hydrochloride.

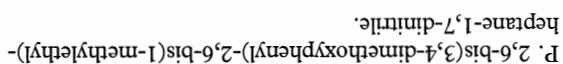
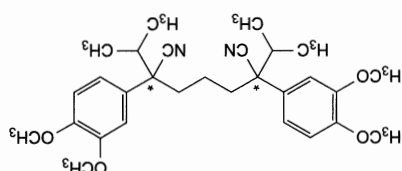
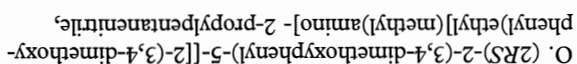
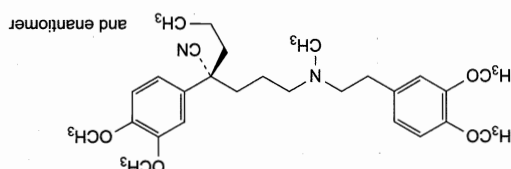
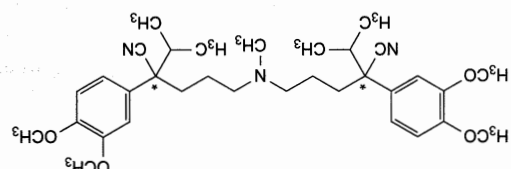
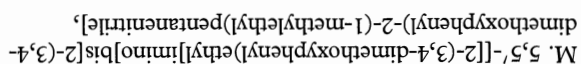
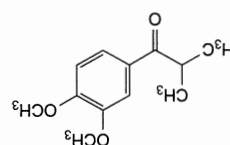
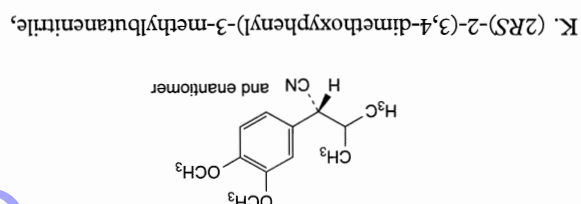
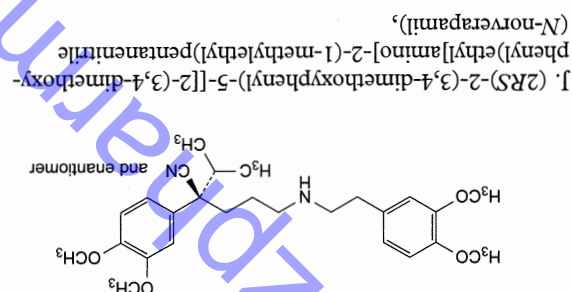
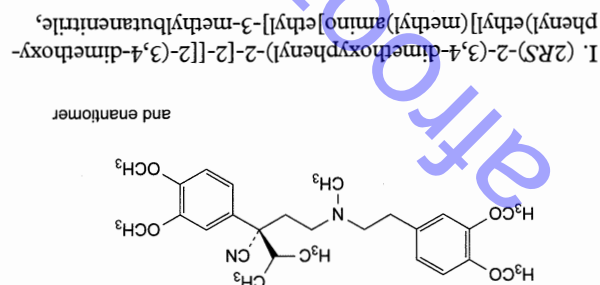
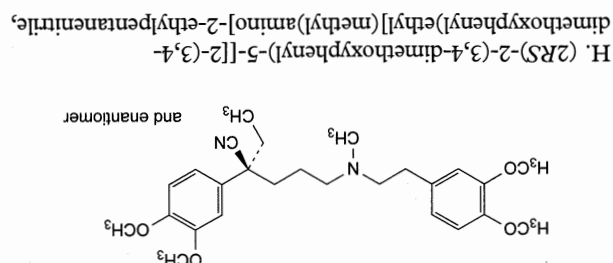
Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.



Ph Eur

Vigabatrin

(Ph Eur monograph 2305)

C₆H₁₁NO₂ 129.2

60643-86-9

Action and use

Antiepileptic.

Preparations

Vigabatrin Oral Powder

Vigabatrin Tablets

Ph Eur

DEFINITION

(4*RS*)-4-Amino-5-enoic acid.

Content

98.5 per cent to 101.5 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white powder.

Solubility

Freely soluble in water, slightly soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison vigabatrin CRS.

TESTS

Impurity D

Liquid chromatography (2.2.29).

Test solution Dissolve 20.0 mg of the substance to be

examined in water R and dilute to 10.0 mL with the same

solvent. To 1.0 mL of the solution add 2.0 mL of a 30.8 g/L

solution of boric acid R adjusted to pH 7.7 with a 500 g/L

solution of sodium hydroxide R, and mix. Add 3.0 mL of a

1.6 g/L solution of (9-fluorenyl)methyl chloroformate R in

acetone R, mix and allow to stand for 5 min. Add 3.0 mL of

ethyl acetate R, shake vigorously for a few seconds and allow

the phases to separate. Use the lower layer within 8 h of

preparation.

Reference solution Dissolve 20.0 mg of vigabatrin

impurity D CRS in water R and dilute to 50.0 mL with the

same solvent. Dilute 1.0 mL of the solution to 10.0 mL with

water R. To 1.0 mL of this solution add 20.0 mg of the

substance to be examined, dissolve in water R and dilute to

10.0 mL with the same solvent. Prepare as for the test

solution, at the same time and in the same manner.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

— stationary phase: phenylsilyl silica gel for chromatography R

(5 μ m).

Mobile phase Mix 25 volumes of acetonitrile R and 75 volumes

of a 4.1 g/L solution of anhydrous sodium acetate R adjusted

to pH 4.2 with glacial acetic acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 263 nm.

Injection 25 μ L.

Run time Twice the retention time of vigabatrin.

Relative retention With reference to vigabatrin (retention

time = about 17 min): (9-fluorenyl)methanol = about 0.4;

impurity D = about 0.6.

System suitability: reference solution:

— resolution: minimum 2.0 between the peaks due to

(9-fluorenyl)methanol and impurity D.

Limit:

— impurity D: not more than 0.5 times the area of the

corresponding peak in the chromatogram obtained with

the reference solution (0.2 per cent).

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 40.0 mg of the substance to be

examined in the mobile phase and dilute to 10.0 mL with

the mobile phase.

Reference solution (a) Dissolve 10 mg of vigabatrin

impurity A CRS and 10.0 mg of vigabatrin impurity B CRS in

the mobile phase and dilute to 10.0 mL with the mobile

phase (solution A). Dilute 3.0 mL of solution A to 50.0 mL

with the mobile phase. Dilute 1.0 mL of this solution to

10.0 mL with the mobile phase.

Reference solution (b) Dissolve 4.0 mg of vigabatrin

impurity B CRS in the mobile phase and dilute to 10.0 mL

with the mobile phase. Dilute 1.0 mL of the solution to

50.0 mL with the mobile phase.

Reference solution (c) To 40 mg of the substance to be

examined add 1.0 mL of solution A and dilute to 10.0 mL

with the mobile phase.

Column 1:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: hexylsilyl silica gel for chromatography R

(5 μ m).

Column 2:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: cation-exchange resin R (10 μ m).

Columns 1 and 2 are coupled in series.

Mobile phase Dissolve 58.5 g of sodium dihydrogen phosphate R

in water R, add 23 mL of phosphoric acid R and dilute to

1000 mL with water R; mix 25 volumes of the solution,

25 volumes of acetonitrile R1 and 950 volumes of water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 μ L.

Run time 2.5 times the retention time of vigabatrin.

Relative retention With reference to vigabatrin (retention

time = about 18 min): impurity E = about 0.5;

impurity A = about 0.8; impurity B = about 1.5.

Identification of impurities Use the chromatogram obtained

with reference solution (a) to identify the peaks due to

impurities A and B; use the chromatogram obtained with

reference solution (b) to identify the peak due to impurity E.

System suitability:

— resolution: minimum 1.5 between the peaks due to

impurity A and vigabatrin in the chromatogram obtained

with reference solution (c);

— signal-to-noise ratio: minimum 20 for the peak due to

impurity E in the chromatogram obtained with reference

solution (b);

— repeatability: maximum relative standard deviation of

5.0 per cent after 5 injections of reference solution (b).

Limits:

— impurities A, B: for each impurity, not more than the area

of the corresponding peak in the chromatogram obtained

with reference solution (a) (0.15 per cent);

— unspecified impurities: for each impurity, not more than

0.5 times the area of the peak due to impurity E in the

chromatogram obtained with reference solution (b)

— total: maximum 0.5 per cent;

— disregard limit: 0.25 times the area of the peak due to

impurity E in the chromatogram obtained with reference

solution (b) (0.05 per cent).

The thresholds indicated under Related substances

(Table 2034-1) in the general monograph Substances for

pharmaceutical use (2034) do not apply.

Heavy metals (2.4.8)

Maximum 10 ppm.

Dissolve 2.0 g in water R and dilute to 20 mL with the same

solvent. 12 mL complies with test A. Prepare the reference

solution using lead standard solution (1 ppm Pb) R.

Water (2.5.12)

Maximum 0.5 per cent, determined on 0.300 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 90 mg in 50 mL of glacial acetic acid R. Titrate with

0.1 M perchloric acid, determining the end-point

potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 12.92 mg

of $C_6H_{11}NO_2$.

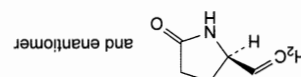
IMPURITIES

Specified impurities: A, B, D.

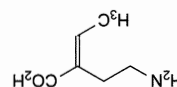
Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of the tests

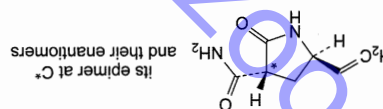
in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities. It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use: C, E, F.



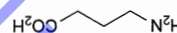
A. (5R)-5-ethenylpyrrolidin-2-one,



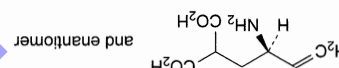
B. (2E)-2-(2-aminoethyl)but-2-enoic acid,



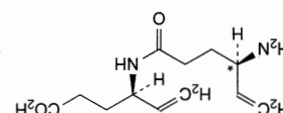
C. 5-ethenyl-2-oxopyrrolidine-3-carboxamide (mixture of the 4 stereoisomers),



D. 4-aminobutanoic acid (GABA),



E. 2-[(2R)-2-aminobut-3-enyl]propanedioic acid,

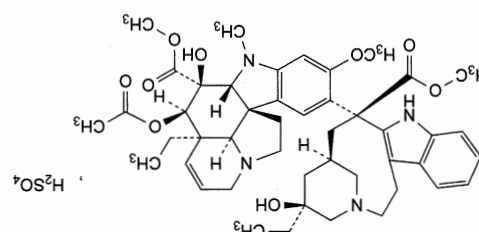


F. 4-[(4-aminohex-5-enyl)amino]hex-5-enoic acid (mixture of the 4 stereoisomers).

Vinblastine Sulfate

Vinblastine Sulfate

(Ph. Eur. monograph 0748)



$C_{46}H_{60}N_4O_{13}S$

909

143-67-9

Vinblastine Injection
Preparation

Vinca alkaloid cytotoxic.

Action and use

Reference solution (a) Dissolve the contents of a vial of vinblastine sulfate CRS in 5.0 mL of water R to obtain a concentration of 1.0 mg/mL.
Reference solution (b) Dissolve 1.0 mg of vinblastine sulfate CRS in 1.0 mL of reference solution (a).
Reference solution (c) Dilute 1.0 mL of reference solution (a) to 50.0 mL with water R.
Test solution Dilute 1.0 mL of solution S (see Tests) to 5.0 mL with water R.
Examine by liquid chromatography (2.2.29).
Keep the solutions in iced water before use.

ASSAY

Not more than 15.0 per cent, determined on 3 mg by thermogravimetry (2.2.34). Heat to 200°C at a rate of 5°C/min, under a stream of nitrogen for chromatography R₂ at a flow rate of 40 mL/min.

Loss on drying

Examine the chromatograms obtained in the assay. In the chromatogram obtained with the test solution, the area of any peak apart from the principal peak is not greater than the area of the principal peak in the chromatogram obtained with reference solution (c) (2.0 per cent) and the sum of the areas of any such peaks is not greater than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (5.0 per cent). Disregard any peak with an area less than that of the peak in the chromatogram obtained with reference solution (d).

Related substances

Dilute 3 mL of solution S to 10 mL with carbon dioxide-free water R. The pH of this solution is 3.5 to 5.0.

pH (2.2.3)

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method 1).

Appearance of solution

Dissolve 50.0 mg in carbon dioxide-free water R and dilute to 10.0 mL with the same solvent.

TESTS

A. Examine by infrared absorption spectrophotometry (2.2.24), comparing with the Ph. Eur. reference spectrum of vinblastine sulfate.
B. Examine the chromatograms obtained in the assay. The principal peak in the chromatogram obtained with the test solution is similar in position and approximate size to the principal peak in the chromatogram obtained with reference solution (a).

IDENTIFICATION

A white or slightly yellowish, crystalline powder, very hygroscopic, freely soluble in water, practically insoluble in alcohol.

CHARACTERS

Vinblastine sulfate contains not less than 95.0 per cent and not more than the equivalent of 104.0 per cent of methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(5S,7R,9S)-5-ethyl-5-hydroxy-9-(methoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-5-hydroxy-8-methoxy-6-methyl-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate sulfate, calculated with reference to the dried substance.

DEFINITION

Ph. Eur.

CHARACTERS

Appearance
White or slightly yellowish, crystalline powder, very hygroscopic.

Solubility

Freely soluble in water, slightly soluble in alcohol.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of vincristine sulfate.

TESTS

Solution S

Dissolve 50.0 mg in carbon dioxide-free water R and dilute to 10.0 mL with the same solvent. Keep the solution in iced water to carry out the test for related substances.

Appearance of solution

Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method I).

pH (2.2.3)

3.5 to 4.5.

Dilute 2 mL of solution S to 10 mL with carbon dioxide-free water R.

Related substances

Liquid chromatography (2.2.29). Keep the solutions in iced water before use.

Test solution Dilute 1.0 mL of solution S to 5.0 mL with water R.

Reference solution (a) Dissolve the contents of a vial of vincristine sulfate CRS in 5.0 mL of water R to obtain a concentration of 1.0 mg/mL.

Reference solution (b) Dissolve 1.0 mg of vincristine sulfate CRS in 1.0 mL of reference solution (a).

Reference solution (c) Dilute 1.0 mL of the test solution to 50.0 mL with water R.

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 20.0 mL with water R.

Precolumn: stationary phase: octylsilyl silica gel for chromatography R.

Column: size: l = 0.25 m, Ø = 4.6 mm,

stationary phase: octylsilyl silica gel for chromatography R (5 µm).

Mobile phase: mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

Mobile phase B: methanol R,

Reference solution (d) Dilute 1.0 mL of reference solution (c) to 20.0 mL with water R.

The chromatographic procedure may be carried out using:

— a stainless steel column 0.25 m long and 4.6 mm in internal diameter packed with octylsilyl silica gel for chromatography R (5 µm). Place between the injector and the column a precolumn packed with suitable silica gel,

— as mobile phase at a flow rate of 1.0 mL/min a mixture of 38 volumes of a 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R,

12 volumes of acetonitrile R and 50 volumes of methanol R,

— as detector a spectrophotometer set at 262 nm,

— a loop injector.

Inject 10 µL of each solution and record the chromatograms for 3 times the retention time of the peak due to vincristine. The assay is not valid unless: in the chromatogram obtained with reference solution (b) the resolution between the peaks due to vincristine and vincblastine is not less than 4; the peak in the chromatogram obtained with reference solution (d) has a signal-to-noise ratio not less than 5. Calculate the percentage content of C₄₆H₆₀N₄O₁₃S from the area of the principal peak in each of the chromatograms obtained with the test solution and reference solution (a) and from the declared content of vincblastine sulfate CRS.

STORAGE

Store in an airtight, glass container, protected from light, at a temperature not exceeding -20 °C. If the substance is sterile, store in a sterile, tamper-proof glass container.

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95.0 per cent to 104.0 per cent (dried substance).

Content

Methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3-ethyl-9-[(5S,7R,9S)-5-ethyl-5-hydroxy-9-(methoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-1,4,5,6,7,8,9,10-octahydro-5,4-b]indol-9-yl]-6-formyl-methanoazacycloundecino[5,4-b]indol-9-yl]-6-formyl-5-hydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate sulfate.

DEFINITION

Ph Eur

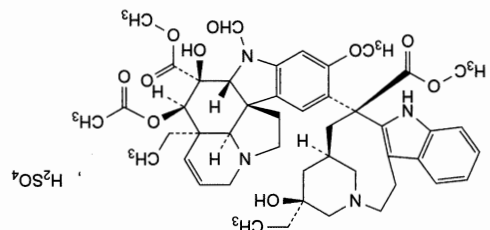
Vincristine Sulfate

Action and use
Vincal alkaloid cytotoxic.

C₄₆H₅₈N₄O₁₃S

923

2068-78-2



Vincristine Sulfate
(Ph. Eur. monograph 0749)

Vincristine Sulfate

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- **total**: not more than 2.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (5.0 per cent),
- **disregard limit**: area of the peak in the chromatogram obtained with reference solution (d) (0.1 per cent).

Loss on drying

Maximum 12.0 per cent, determined on 3 mg by thermogravimetry (2.2.34). Heat the substance to be examined to 200 °C increasing the temperature by 5 °C/min, under a current of nitrogen for chromatography R, at a flow rate of 40 mL/min.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances, with the following modifications.

Mobile phase Mix 30 volumes of a 1.5 per cent *V/V* solution of diethylamine R adjusted to pH 7.5 with phosphoric acid R and 70 volumes of methanol R.

Flow rate 1.0 mL/min.

Calculate the percentage content of $C_{46}H_{58}N_4O_{14}S$ using the chromatogram obtained with reference solution (a) and the declared content of *vincristine sulfate CRS*.

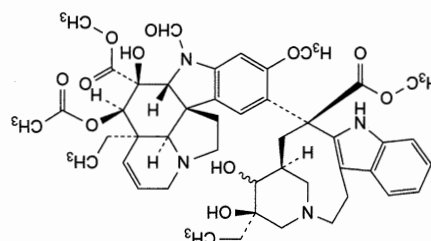
STORAGE

In an airtight, glass container, protected from light, at a temperature not exceeding –20 °C. If the substance is sterile, store in a sterile, airtight, tamper-proof glass container.

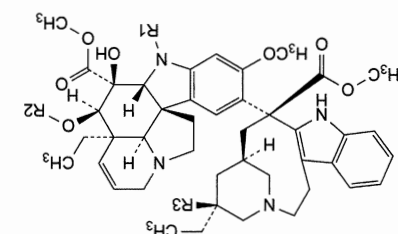
IMPURITIES

Specified impurities A, B, C, D, H

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10, *Control of impurities in substances for pharmaceutical use*: E, F, G.



A. methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(5R,7S,9S)-5-ethyl-5,6-dihydroxy-9-(methoxyoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-6-formyl-5-hydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (3'-hydroxy-VCR),



B. R1 = CHO, R2 = CO-CH₃, R3 = H; methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(5R,7S,9S)-5-ethyl-5,6-dihydroxy-9-(methoxyoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-6-formyl-5-hydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (4'-deoxyvincristine),

C. R1 = H, R2 = CO-CH₃, R3 = OH; methyl (3aR,4R,5S,5aR,10bS,13aR)-4-(acetyloxy)-3a-ethyl-9-[(5S,7R,9S)-5-ethyl-5-hydroxy-9-(methoxyoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-6-formyl-4,5-dihydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (deacetylvincristine),

E. R1 = CH₃, R2 = H, R3 = OH; methyl (3aR,4R,5S,5aR,10bR,13aR)-3a-ethyl-9-[(5S,7R,9S)-5-ethyl-5-hydroxy-9-(methoxyoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-4,5-dihydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (deacetylvinblastine),

H. R1 = CH₃, R2 = CO-CH₃, R3 = OH; vinblastine, (3aR,4R,5S,5aR,10bR,13aR)-3a-ethyl-9-[(5S,7R,9S)-5-ethyl-5-hydroxy-9-(methoxyoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-6-formyl-5-hydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (vincristine),

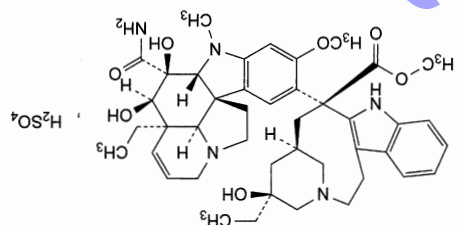
F. R = CH₃; methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(1aS,11S,13S,13aR)-1a-ethyl-11-(methoxyoxycarbonyl)-1a,4,5,10,11,12,13,13a-octahydro-2H-3,13-methano-oxireno[9,10]azacycloundecino[5,4-b]indol-11-yl]-6-formyl-5-hydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (formylvincristine),

G. R = CHO; methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(1aS,11S,13S,13aR)-1a-ethyl-11-(methoxyoxycarbonyl)-1a,4,5,10,11,12,13,13a-octahydro-2H-3,13-methano-oxireno[9,10]azacycloundecino[5,4-b]indol-11-yl]-5-hydroxy-8-methoxy-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (leurosine),

Vindesine Sulfate

Vindesine Sulphate

(Ph. Eur. monograph 1276)



$C_{43}H_{57}N_5O_{15}S$ 852 59917-39-4

Action and use

Vinca alkaloid cytotoxic.

Preparation

Vindesine Injection

Ph. Eur.

DEFINITION

Methyl (5S,7R,9S)-9-[(3aR,4R,5S,5aR,10bR,13aR)-5-carbamoyl-3a-ethyl-4,5-dihydroxy-8-methoxy-6-methyl-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazol-9-yl]-5-ethyl-5-hydroxy-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[4,5-b]indole-9-carboxylate sulfate.

Content

96.0 per cent to 103.0 per cent (dried substance).

CHARACTERS

Appearance
White or almost white, amorphous, hygroscopic substance.

Solubility
Freely soluble in water and in methanol, practically insoluble in cyclohexane.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).
Comparison Ph. Eur. reference spectrum of vindesine sulfate.

TESTS

Solution S
Dissolve 50 mg in carbon dioxide-free water R and dilute to 10 mL with the same solvent.

Appearance of solution
Solution S is clear (2.2.1) and not more intensely coloured than reference solution Y₇ (2.2.2, Method I).

pH (2.2.3)

3.5 to 5.5 for solution S.

Related substances

Liquid chromatography (2.2.29). Keep the solutions in iced water before use.
Test solution Dissolve 10.0 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.
Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with water R.
Reference solution (b) Dissolve 1.0 mg of desacetylvinblastine CRS in water R, add 1.0 mL of the test solution and dilute to 50.0 mL with water R.
Reference solution (c) Dilute 1.0 mL of reference solution (a) to 200.0 mL with water R.

Injection 3 µL.

Detection Flame ionisation.

— injection port and detector: 250 °C.

— column: 170 °C;

Temperature:

Flow rate 60 mL/min.

Carrier gas helium for chromatography R.

copolymer R.

— stationary phase: ethylvinylbenzene-divinylbenzene

— material: glass;

Column:

in 1.0 mL of internal standard solution (b).

Test solution Dissolve 40 mg of the substance to be examined

water R.

internal standard solution (a) and dilute to 50.0 mL with

Reference solution Dissolve 10.0 g of acetaminophen R to 1000 mL

standard solution (a) to 50.0 mL with water R.

Internal standard solution (b) Dilute 10.0 mL of internal

100 mL with water R.

Internal standard solution (a) Dilute 0.500 g of propanol R to

Gas chromatography (2.2.28).

Acetonitrile

(0.01 per cent).

chromatogram obtained with reference solution (c)

— disregard limit: the area of the principal peak in the

(2 per cent);

chromatogram obtained with reference solution (a)

total: not more than the area of the principal peak in the

(1 per cent);

chromatogram obtained with reference solution (a)

0.5 times the area of the principal peak in the

— impurities A, B, C: for each impurity, not more than

Limits:

vindesine.

— symmetry factor: maximum 2.0 for the peak due to

vindesine and desacetylvinblastine;

— resolution: minimum 2.0 between the peaks due to

the retention time of vindesine is less than 40 min;

System suitability: reference solution (b):

Run time Twice the retention time of vindesine.

Injection 200 µL.

Detection Spectrophotometer at 270 nm.

Flow rate 2 mL/min.

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 40	49	51
40 - 49	49 → 30	51 → 70
49 - end	30	70

— mobile phase B: methanol R;

— mobile phase A: 1.5 per cent V/V solution of diethylamine R adjusted to pH 7.4 with phosphoric acid R;

Mobile phase:

(5 µm).

— stationary phase: octadecylsilyl silica gel for chromatography R

Column:

— size: 1 = 0.15 mm, Ø = 4.6 mm;

System suitability: reference solution: — **resolution:** minimum 1.5 between the peaks due to acetonitrile and propanol; — **symmetry factor:** maximum 1.6 for the peak due to acetonitrile.

Limit: — **acetonitrile:** maximum 1.5 per cent *m/m*.

Loss on drying

Maximum 10.0 per cent, determined on 9.00 mg by thermogravimetry (2.2.34). Heat to 200 °C at a rate of 5 °C/min, under a stream of nitrogen for chromatography R at a flow rate of 40 mL/min.

ASSAY

Liquid chromatography (2.2.29). Keep the solutions in iced water before use.

Test solution Dissolve 5.0 mg of the substance to be examined in water R and dilute to 10.0 mL with the same solvent.

Reference solution (a) Dissolve and dilute the entire contents of a vial of vinorelbine sulfate CRS with water R to yield a concentration of approximately 0.50 mg/mL.

Reference solution (b) Add 1.0 mg of desacetylvinblastine CRS to 2.0 mL of reference solution (a).

Column:

— **size:** 1 = 0.15 m, 4.6 mm;

— **stationary phase:** octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 38 volumes of a 1.5 per cent *V/V* solution of diethylamine R, previously adjusted to pH 7.4 with phosphoric acid R, and 62 volumes of methanol R.

Flow rate 1 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 20 µL.

System suitability: reference solution (b): — **resolution:** minimum 1.5 between the peaks due to vinorelbine and desacetylvinblastine;

— **symmetry factor:** maximum 2.0 for the peak due to vinorelbine;

— **repeatability:** maximum relative standard deviation of 1.5 per cent for the peak due to vinorelbine after 5 injections.

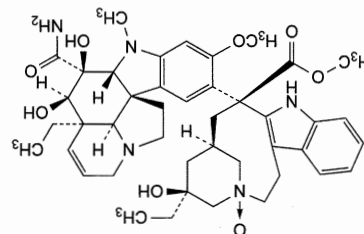
Calculate the percentage content of $C_{43}H_{57}N_5O_{11}$ S from the declared content of vinorelbine sulfate CRS.

STORAGE

In an airtight polypropylene container with a polypropylene cap, at a temperature of –50 °C or below. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

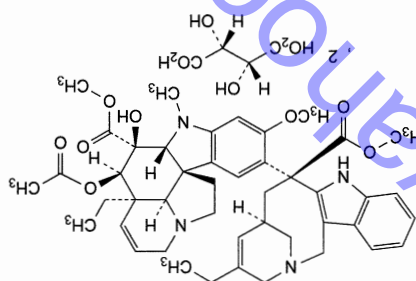
Specified impurities A, B, C



A. vinorelbine 3',N'-oxide,

Vinorelbine Tartrate

(Ph. Eur. monograph 2107)



$C_{53}H_{66}N_4O_{20}$

1079

125317-39-7

Action and use

Vinca alkaloid cytotoxic.

DEFINITION

Methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(6R,8S)-4-ethyl-8-(methoxycarbonyl)-1,3,6,7,8,9-hexahydro-2,6-methano-2H-azacyclodecino[4,3-b]indol-8-yl]-5-hydroxy-8-methoxy-6-methyl-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate dihydrogen bis[(2R,3R)-2,3-dihydroxybutanedioate].

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

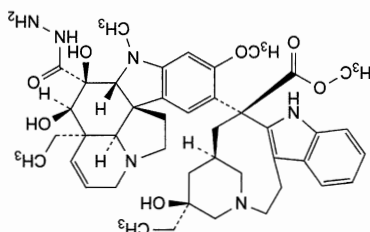
White or almost white powder, hygroscopic.

Solubility

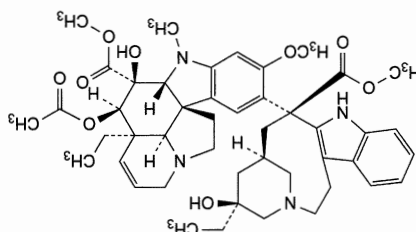
Freely soluble in water and in methanol, practically insoluble in hexane.

Ph. Eur.

C. desacetylvinblastine hydrazide.



B. methyl (3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-[(5S,7R,9S)-5-ethyl-5-hydroxy-9-(methoxycarbonyl)-1,4,5,6,7,8,9,10-octahydro-2H-3,7-methanoazacycloundecino[5,4-b]indol-9-yl]-5-hydroxy-8-methoxy-6-methyl-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate (vinblastine),



IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24). Preparation Dissolve 10 mg in 5 mL of water R. Add 0.5 mL of sodium hydroxide solution R. Extract with 5 mL of methylene chloride R. Dry the organic layer over anhydrous sodium sulfate R, filter and reduce its volume to about 0.5 mL by evaporation and apply to a disc of potassium bromide R. Evaporate and record the spectrum.

Comparison vinorelbine tartrate CRS, treated as described above.

B. It gives reaction (b) of tartrates (2.3.1).

TESTS

Solution S Dissolve a quantity equivalent to 0.140 g of the anhydrous substance in water R and dilute to 10.0 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and its absorbance (2.2.25) at 420 nm is not greater than 0.030.

pH (2.2.3)

3.3 to 3.8 for solution S.

Related substances

Liquid chromatography (2.2.29): use the normalisation procedure.

Test solution Dissolve 35.0 mg of the substance to be examined in the mobile phase and dilute to 25 mL with the mobile phase.

Reference solution (a) Dissolve 7 mg of vinorelbine impurity B CRS in water R and dilute to 50 mL with the same solvent. To 1 mL of this solution add 14 mg of vinorelbine tartrate CRS, dissolve in water R and dilute to 10 mL with the same solvent. Expose this solution for 1 h to a xenon lamp apparatus at a wavelength of 310–880 nm, supplying a dose of 1600 kJ/m² at a fluence rate of 500 W/m² in order to generate impurity A.

Reference solution (b) Dilute 1.0 mL of the test solution to 20.0 mL with water R. Dilute 1.0 mL of this solution to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.15$ m, $\phi = 3.9$ mm,
— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m) with a specific surface area of 125 m²/g, a pore size of 30 nm and a carbon loading of 7 per cent,
— temperature: 35 ± 5 °C.

Mobile phase Dissolve 1.22 g of sodium decanesulfonate R in 620 mL of methanol R and add 380 mL of a 7.80 g/L solution of sodium dihydrogen phosphate R previously adjusted to pH 4.2 with dilute phosphoric acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 267 nm.

Injection 20 μ L.

Run time Twice the retention time of vinorelbine.

Relative retention With reference to vinorelbine (retention time = about 14 min): impurity A = about 0.8;

impurity B = about 1.2.

System suitability:

— **peak-to-valley ratio:** minimum 4, where H_p = height above the baseline of the peak due to impurity B and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to

Wavelength 328.1 nm.

Source Silver hollow-cathode lamp.

Reference solutions Prepare the reference solutions using silver standard solution (5 ppm Ag) R and diluting with a 6.5 per cent V/V solution of lead-free nitric acid R.

examined in 10.0 mL of water R.

Test solution Dissolve 0.500 g of the substance to be

Maximum 5 ppm.

Silver

calculate the content of fluoride.

Introduce the electrodes into the reference solutions and allow to stand for 5 min. Determine the potential difference between the electrodes after 1 min of stabilisation. Using semi-logarithmic paper plot the potential difference obtained for each reference solution as a function of concentration of fluoride. Using exactly the same conditions, determine the potential difference obtained with the test solution and

50 mL with water R.

5.0 mL of total-ionic-strength-adjustment buffer R and dilute to

1.4 mL of fluoride standard solution (10 ppm F) R, add

Reference solutions To 0.6 mL, 0.8 mL, 1.0 mL, 1.2 mL and

adjustment buffer R and dilute to 50 mL with water R.

Test solution Dissolve 0.19 g of the substance to be examined

in 20 mL of water R. Add 5.0 mL of total-ionic-strength-

adjustment buffer R and dilute to 50 mL with water R.

Potentiometry (2.2.36, Method I) using a fluoride-selective

indicator electrode and a silver-silver chloride reference

electrode.

Fluorides

Maximum 50 ppm.

greater than that of the reference solution.

The maximum absorbance value of the test solution is not

650 nm, using the blank solution as compensation liquid.

solution and the reference solution, between 560 nm and

After 45 min, measure the absorbance (2.2.25) of the test

solution of carminic acid R in sulfuric acid R.

warm to room temperature. Add 10.0 mL of a 0.5 g/L

sulfuric acid R while cooling in iced water. Stir and allow to

Blank solution To 2.0 mL of water R slowly add 10.0 mL of

sulfuric acid R.

Add 10.0 mL of a 0.5 g/L solution of carminic acid R in

in iced water. Stir and allow to warm to room temperature.

solution slowly add 10.0 mL of sulfuric acid R while cooling

boric acid R to 100.0 mL with water R. To 2.0 mL of this

Reference solution Dilute 2.5 mL of a 0.572 g/L solution of

acid R in sulfuric acid R.

temperature. Add 10.0 mL of a 0.5 g/L solution of carminic

while cooling in iced water. Stir and allow to warm to room

in 2 mL of water R. Slowly add 10.0 mL of sulfuric acid R

Test solution Dissolve 0.10 g of the substance to be examined

Maximum 50 ppm.

Boron

chromatogram obtained with reference solution (b).

— **disregard limit:** the area of the principal peak in the

— **sum of impurities other than A:** maximum 0.7 per cent,

0.2 per cent,

— **any other impurity:** for each impurity, maximum

— **impurity A:** maximum 0.3 per cent,

Limits:

the chromatogram obtained with reference solution (b).

— **signal-to-noise ratio:** minimum 10 for the principal peak in

solution (a),

vinorelbine in the chromatogram obtained with reference

Atomisation device Air-acetylene flame.

Water (2.5.12)

Maximum 4.0 per cent, determined on 0.250 g.

Bacterial endotoxins (2.6.14)

Less than 2 IU/mg (expressed as vinorelbine base), if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Dissolve 0.350 g in 40 mL of glacial acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 53.96 mg of $\text{C}_{53}\text{H}_{66}\text{N}_4\text{O}_{20}$.

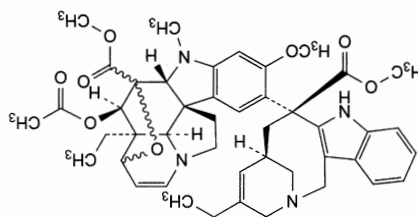
STORAGE

Under an inert gas, protected from light, at a temperature not exceeding -15°C .

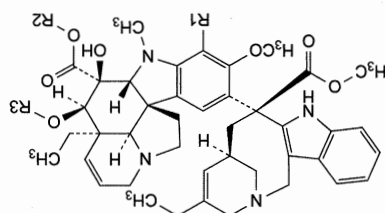
IMPURITIES

Specified impurities A

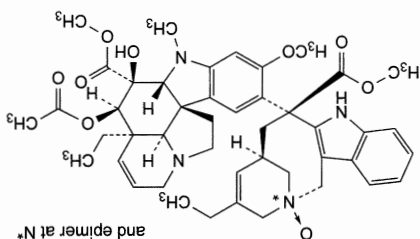
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, C, D, E, F, G, H, I, J.



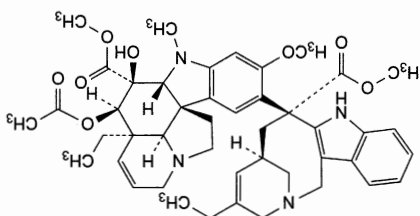
A. methyl (3a), 4R,5R,10bR,13aR,14R,15R,16R,17R,18R,19R,20R,21R,22R,23R,24R,25R,26R,27R,28R,29R,30R,31R,32R,33R,34R,35R,36R,37R,38R,39R,40R,41R,42R,43R,44R,45R,46R,47R,48R,49R,50R,51R,52R,53R,54R,55R,56R,57R,58R,59R,60R,61R,62R,63R,64R,65R,66R,67R,68R,69R,70R,71R,72R,73R,74R,75R,76R,77R,78R,79R,80R,81R,82R,83R,84R,85R,86R,87R,88R,89R,90R,91R,92R,93R,94R,95R,96R,97R,98R,99R,100R,101R,102R,103R,104R,105R,106R,107R,108R,109R,110R,111R,112R,113R,114R,115R,116R,117R,118R,119R,120R,121R,122R,123R,124R,125R,126R,127R,128R,129R,130R,131R,132R,133R,134R,135R,136R,137R,138R,139R,140R,141R,142R,143R,144R,145R,146R,147R,148R,149R,150R,151R,152R,153R,154R,155R,156R,157R,158R,159R,160R,161R,162R,163R,164R,165R,166R,167R,168R,169R,170R,171R,172R,173R,174R,175R,176R,177R,178R,179R,180R,181R,182R,183R,184R,185R,186R,187R,188R,189R,190R,191R,192R,193R,194R,195R,196R,197R,198R,199R,200R,201R,202R,203R,204R,205R,206R,207R,208R,209R,210R,211R,212R,213R,214R,215R,216R,217R,218R,219R,220R,221R,222R,223R,224R,225R,226R,227R,228R,229R,230R,231R,232R,233R,234R,235R,236R,237R,238R,239R,240R,241R,242R,243R,244R,245R,246R,247R,248R,249R,250R,251R,252R,253R,254R,255R,256R,257R,258R,259R,260R,261R,262R,263R,264R,265R,266R,267R,268R,269R,270R,271R,272R,273R,274R,275R,276R,277R,278R,279R,280R,281R,282R,283R,284R,285R,286R,287R,288R,289R,290R,291R,292R,293R,294R,295R,296R,297R,298R,299R,300R,301R,302R,303R,304R,305R,306R,307R,308R,309R,310R,311R,312R,313R,314R,315R,316R,317R,318R,319R,320R,321R,322R,323R,324R,325R,326R,327R,328R,329R,330R,331R,332R,333R,334R,335R,336R,337R,338R,339R,340R,341R,342R,343R,344R,345R,346R,347R,348R,349R,350R,351R,352R,353R,354R,355R,356R,357R,358R,359R,360R,361R,362R,363R,364R,365R,366R,367R,368R,369R,370R,371R,372R,373R,374R,375R,376R,377R,378R,379R,380R,381R,382R,383R,384R,385R,386R,387R,388R,389R,390R,391R,392R,393R,394R,395R,396R,397R,398R,399R,400R,401R,402R,403R,404R,405R,406R,407R,408R,409R,410R,411R,412R,413R,414R,415R,416R,417R,418R,419R,420R,421R,422R,423R,424R,425R,426R,427R,428R,429R,430R,431R,432R,433R,434R,435R,436R,437R,438R,439R,440R,441R,442R,443R,444R,445R,446R,447R,448R,449R,450R,451R,452R,453R,454R,455R,456R,457R,458R,459R,460R,461R,462R,463R,464R,465R,466R,467R,468R,469R,470R,471R,472R,473R,474R,475R,476R,477R,478R,479R,480R,481R,482R,483R,484R,485R,486R,487R,488R,489R,490R,491R,492R,493R,494R,495R,496R,497R,498R,499R,500R,501R,502R,503R,504R,505R,506R,507R,508R,509R,510R,511R,512R,513R,514R,515R,516R,517R,518R,519R,520R,521R,522R,523R,524R,525R,526R,527R,528R,529R,530R,531R,532R,533R,534R,535R,536R,537R,538R,539R,540R,541R,542R,543R,544R,545R,546R,547R,548R,549R,550R,551R,552R,553R,554R,555R,556R,557R,558R,559R,560R,561R,562R,563R,564R,565R,566R,567R,568R,569R,570R,571R,572R,573R,574R,575R,576R,577R,578R,579R,580R,581R,582R,583R,584R,585R,586R,587R,588R,589R,590R,591R,592R,593R,594R,595R,596R,597R,598R,599R,600R,601R,602R,603R,604R,605R,606R,607R,608R,609R,610R,611R,612R,613R,614R,615R,616R,617R,618R,619R,620R,621R,622R,623R,624R,625R,626R,627R,628R,629R,630R,631R,632R,633R,634R,635R,636R,637R,638R,639R,640R,641R,642R,643R,644R,645R,646R,647R,648R,649R,650R,651R,652R,653R,654R,655R,656R,657R,658R,659R,660R,661R,662R,663R,664R,665R,666R,667R,668R,669R,670R,671R,672R,673R,674R,675R,676R,677R,678R,679R,680R,681R,682R,683R,684R,685R,686R,687R,688R,689R,690R,691R,692R,693R,694R,695R,696R,697R,698R,699R,700R,701R,702R,703R,704R,705R,706R,707R,708R,709R,710R,711R,712R,713R,714R,715R,716R,717R,718R,719R,720R,721R,722R,723R,724R,725R,726R,727R,728R,729R,730R,731R,732R,733R,734R,735R,736R,737R,738R,739R,740R,741R,742R,743R,744R,745R,746R,747R,748R,749R,750R,751R,752R,753R,754R,755R,756R,757R,758R,759R,760R,761R,762R,763R,764R,765R,766R,767R,768R,769R,770R,771R,772R,773R,774R,775R,776R,777R,778R,779R,780R,781R,782R,783R,784R,785R,786R,787R,788R,789R,790R,791R,792R,793R,794R,795R,796R,797R,798R,799R,800R,801R,802R,803R,804R,805R,806R,807R,808R,809R,810R,811R,812R,813R,814R,815R,816R,817R,818R,819R,820R,821R,822R,823R,824R,825R,826R,827R,828R,829R,830R,831R,832R,833R,834R,835R,836R,837R,838R,839R,840R,841R,842R,843R,844R,84



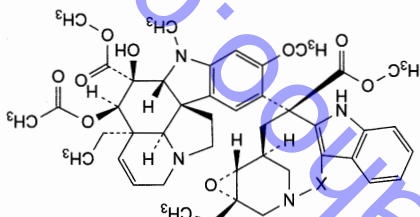
B. RI = R3 = H, R2 = CH3;
(3aR,4R,5S,5aR,10bR,13aR)-3a-ethyl-9-
(methoxycarbonyl)-1,3,6,7,8,9-hexahydro-2H-
zacyclodecino[4,3-b]indol-8-yl]-6-methoxy-
methyl-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-
cd]carbazole-5-carboxylate,
H. RI = R2 = H, R3 = CO-CH3;
(3aR,4R,5S,5aR,10bR,13aR)-4-(acetyloxy)-3a-ethyl-9-
(methoxycarbonyl)-1,3,6,7,8,9-hexahydro-2H-
zacyclodecino[4,3-b]indol-8-yl]-6-methoxy-
methyl-3a,4,5,5a,6,11,12,13a-octahydro-1H-indolizino[8,1-
cd]carbazole-5-carboxylic acid,



ethyl-9-[(6R,8R)-4-ethyl-8-(methoxycarbonyl)-1,3,6,7,8,9-
hexahydro-2,6-methano-2H-benzocyclopenta[4,3-b]indol-8-yl]-
5-hydroxy-8-methyl-3a,4,5,5a,1,1,12,13a-
octahydro-1H-indolizino[8,1-cd]carbazole-5-carboxylate,



$\text{R}_1 = \text{Bz}$, $\text{R}_2 = \text{CH}_3$, $\text{R}_3 = \text{CO-CH}_3$; methyl-
 (3*R*,4*R*,5*S*,5*R*,10*Bz*,13*Ar*)-4-(acetyloxy)-7-bromo-3*a*-ethyl-
 9-(6*R*,8*S*)-4-ethyl-8-(methoxycarbonyl)-1,3,3,6,7,8,9-
 hexahydro-2,6-methano-2-*H*-azacyclodecino[4,3-*b*]indol-8-yl]-
 5-hydroxy-8-methoxy-6-methyl-3*a*,4,5,5*a*,6,11,12,13*a*-
 octahydro-1-*H*-indolizino[8,1-*bc*]carbazole-5-*c*-carboxylate,



E. X = CH₂-CH₂: methyl (1aS,11S,13S,13aR)-11-hydroxy-8-methoxy-5-(methoxycarbonyl)-6-methyl-3a,4,5,5a,6,11,12,13a-octahydro-1*H*-indolizino[8,1-*cd*]carbazol-9-yl]-1a-ethyl-1a,4,5,10,11,12,13a-octahydro-2*H*-3,13-methanooxireno[9,10]azacycloundeceno[5,4-*b*]indole-11-carboxylate (neurosinic),
G. X = CH₂: methyl (1aS,10S,12S,12aR)-10-hydroxy-8-methoxy-5-(methoxycarbonyl)-6-methyl-3a,4,5,5a,6,11,12,13a-octahydro-1*H*-indolizino[8,1-*cd*]carbazol-9-yl]-1a-ethyl-1a,2,4,9,10,11,12,12a-octahydro-3,12-methano-3-*H*-oxireno[8,9]azacyclodeceno[4,3-*b*]indole-10-carboxylate,

IDENTIFICATION

- A. Specific optical rotation (see Tests).
B. Infrared absorption spectrophotometry (2.2.24).
Comparison vincopocetine CRS.

TESTS

Specific optical rotation (2.2.7)

Dissolve 0.25 g in dimethylformamide R and dilute to 25.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).
Test solution Dissolve 50.0 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dilute 1.0 mL of the test solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5.0 mg of vincopocetine

impurity B CRS, 6.0 mg of vincopocetine impurity A CRS,

5.0 mg of vincopocetine impurity C CRS and 5.0 mg of

vincopocetine impurity D CRS in the mobile phase and dilute to

50.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (a)

and 1.0 mL of reference solution (b) to 20.0 mL with the

mobile phase.

Column:
size: $l = 0.25$ m, $\phi = 4.6$ mm;

stationary phase: end-capped octadecylsilyl silica gel for

chromatography R (5 μ m).

Mobile phase 15.4 g/L solution of ammonium acetate R,

acetonitrile R (45:55 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 280 nm.

Injection 15 μ L.

Run time 3 times the retention time of vincopocetine.

Relative retention With reference to vincopocetine (retention

time = about 16 min): impurity A = about 0.4;

impurity D = about 0.68; impurity B = about 0.75;

impurity C = about 0.83.

System suitability: reference solution (c):

— resolution: minimum 2.0 between the peaks due to

impurities D and B.

Limits:

— impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (c) (0.6 per cent);

— impurities B, D: for each impurity, not more than the area

of the corresponding peak in the chromatogram obtained

with reference solution (c) (0.5 per cent);

— impurity C: not more than 0.6 times the area of the

corresponding peak in the chromatogram obtained with

reference solution (c) (0.3 per cent);

— unspecified impurities: for each impurity, not more than the

area of the peak due to vincopocetine in the chromatogram

obtained with reference solution (c) (0.10 per cent);

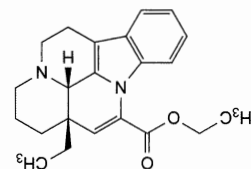
— total: not more than 10 times the area of the peak due to

vincopocetine in the chromatogram obtained with reference

solution (c) (0.05 per cent).

Vincopocetine

(Ph. Eur. monograph 2139)



$C_{22}H_{26}N_2O_2$ 350.5 42971-09-5

Action and use

Vasodilator.

DEFINITION

Ethyl (13aS,13bS)-13a-ethyl-2,3,5,6,13a,13b-hexahydro-1H-indolo[3,2,1-de]pyrido[3,2,1-ij][1,5]naphthyridine-12-carboxylate.

Content

98.5 per cent to 101.5 per cent (dried substance).

CHARACTERS

Appearance

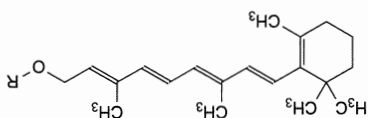
White or slightly yellow, crystalline powder.

Solubility

Practically insoluble in water, soluble in methylene chloride, slightly soluble in anhydrous ethanol.

Vitamin A

(Ph. Eur. monograph 0217)



Substance	R	Molecular formula	M _r
all-(E)-retinol	H	C ₂₀ H ₃₀ O	286.5
all-(E)-retinol acetate	CO-CH ₃	C ₂₂ H ₃₂ O ₂	328.5
all-(E)-retinol propionate	CO-C ₂ H ₅	C ₂₄ H ₃₄ O ₂	342.5
all-(E)-retinol palmitate	CO-C ₁₅ H ₃₁	C ₃₆ H ₆₀ O ₂	524.9

In the British Pharmacopoeia, the term 'Retinol' is used within titles for preparations containing synthetic ester(s) and the term 'Vitamin A' within the title for the preparation containing material of natural origin.

Preparation

Paediatric Vitamins A, C and D Oral Drops

Ph Eur

DEFINITION

Vitamin A refers to a number of substances of very similar structure (including *Z*-isomers) found in animal tissues and possessing similar activity. The principal and biologically most active substance is all-(E)-retinol (all-(E)-3,7-dimethyl-9-(2,6,6-trimethylcyclohex-1-enyl)nona-2,4,6,8-tetraen-1-ol; C₂₀H₃₀O). Vitamin A is generally used in the form of esters such as the acetate, propionate and palmitate.

Synthetic retinol ester refers to an ester (acetate, propionate or palmitate) or a mixture of synthetic retinol esters.

The activity of vitamin A is expressed in retinol equivalents (R.E.). 1 mg R.E. corresponds to the activity of 1 mg of all-(E)-retinol. The activity of the other retinol esters is calculated stoichiometrically, so that 1 mg R.E. of vitamin A corresponds to the activity of:

- 1.47 mg of all-(E)-retinol acetate,
- 1.195 mg of all-(E)-retinol propionate,
- 1.832 mg of all-(E)-retinol palmitate.

International Units (IU) are also used to express the activity of vitamin A. 1 IU of vitamin A is equivalent to the activity of 0.300 µg of all-(E)-retinol. The activity of the other retinol esters is calculated stoichiometrically, so that 1 IU of vitamin A is equivalent to the activity of:

- 0.344 µg of all-(E)-retinol acetate,
- 0.359 µg of all-(E)-retinol propionate,
- 0.550 µg of all-(E)-retinol palmitate,

CHARACTERS

Appearance

Retinol acetate: pale-yellow crystals (mp: about 60 °C). Once melted retinol acetate tends to yield a supercooled melt. Retinol propionate: reddish-brown oily liquid. Retinol palmitate: a fat-like, light yellow solid or a yellow oily liquid, if melted (mp: about 26 °C).

Solubility

All retinol esters are practically insoluble in water, soluble or partly soluble in anhydrous ethanol and miscible with organic solvents.

IMPURITIES

of C₂₂H₂₆N₂O₂.
1 mL of 0.1 M perchloric acid is equivalent to 35.05 mg

potentiometrically (2.2.20).
0.1 M perchloric acid, determining the end-point
acetic anhydride R and anhydrous acetic acid R. Titrate with

ASSAY

Dissolve 0.300 g in 50 mL of a mixture of equal volumes of

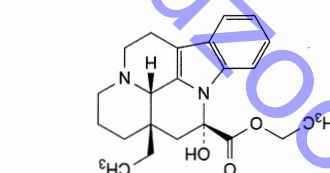
Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

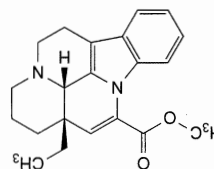
Maximum 0.5 per cent, determined on 1.000 g by drying

Loss on drying (2.2.32)

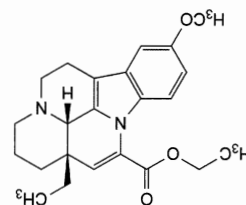
Maximum 0.5 per cent, determined on 1.000 g by drying



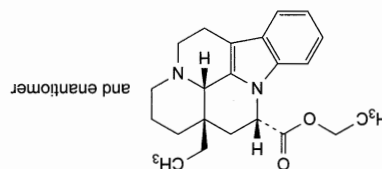
A. ethyl (12S,13aS,13bS)-13a-ethyl-12-hydroxy-2,3,5,6,12,13,13a,13b-octahydro-1H-indolo[3,2,1-de]pyrido[3,2,1-η][1,5]naphthyridine-12-carboxylate (ethyl vincamine),



B. methyl (13aS,13bS)-13a-ethyl-2,3,5,6,13a,13b-hexahydro-1H-indolo[3,2,1-de]pyrido[3,2,1-η][1,5]naphthyridine-12-carboxylate (apovincamine),



C. ethyl (13aS,13bS)-13a-ethyl-10-methoxy-2,3,5,6,13a,13b-hexahydro-1H-indolo[3,2,1-de]pyrido[3,2,1-η][1,5]naphthyridine-12-carboxylate (methoxyvincinapocetine),



D. ethyl (12R,13aR,13bR)-13a-ethyl-2,3,5,6,12,13,13a,13b-octahydro-1H-indolo[3,2,1-de]pyrido[3,2,1-η][1,5]naphthyridine-12-carboxylate (dihydrovincinapocetine).

Ph Eur

Vitamin A and its esters are very sensitive to the action of air, oxidising agents, acids, light and heat. Carry out the assay as rapidly as possible, avoiding exposure to actinic light and air, oxidising agents, oxidation catalysts (e.g. copper, iron), acids and heat; use freshly prepared solutions.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).
Test solution Prepare a solution containing about 3.3 IU of vitamin A per micro litre in cyclohexane R containing 1 g/L of butylhydroxytoluene R.

Reference solution Prepare a 10 mg/mL solution of retinol esters CRS (i.e. 3.3 IU of each ester per micro litre) in cyclohexane R containing 1 g/L of butylhydroxytoluene R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ether R, cyclohexane R (20:80 V/V).

Application 3 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution:

— the chromatogram shows the individual spots of the corresponding esters. The elution order from bottom to top is: retinol acetate, retinol propionate and retinol palmitate.

Results The composition of esters is confirmed by the correspondence of the principal spot or spots of the test solution with those obtained with the reference solution.

B. Related substances (see Tests).

TESTS

Retinol

Thin-layer chromatography (2.2.27).

Test solution Prepare a solution in cyclohexane R, stabilised with a solution containing 1 g/L of butylhydroxytoluene R,

containing about 330 IU of vitamin A per micro litre.

Reference solution Shake 1 mL of the test solution with 20 mL of 0.1 M tetrabutylammonium hydroxide in 2-propanol for 2 min

and dilute to 100 mL with cyclohexane R, stabilised with a solution containing 1 g/L of butylhydroxytoluene R.

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase ether R, cyclohexane R (20:80 V/V).

Application 3 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution:

— in the chromatogram obtained no or only traces of the retinol esters are seen.

Limit Any spot corresponding to retinol in the chromatogram obtained with the test solution is not more intense than the spot in the chromatogram obtained with the reference solution (1.0 per cent).

Related substances

Ultraviolet and visible absorption spectrophotometry (2.2.25).

Test solution The solution described under Activity.

Absorption maximum At 325 nm to 327 nm.

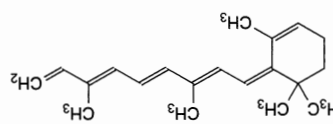
Absorbance ratios:

— A_{370}/A_{326} = maximum 0.14.

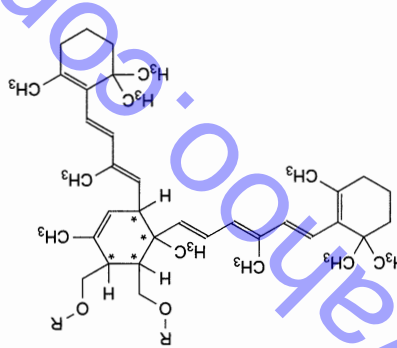
— A_{350}/A_{326} = maximum 0.54.

— A_{300}/A_{326} = maximum 0.60.

B. (3E,5E,7E)-3,7-dimethyl-9-[(1Z)-2,6,6-trimethylcyclohex-2-enylidene]nona-1,3,5,7-tetraene (anhydrous-vitamin A),



A. R = H, CO-CH₃; ketols (Diels-Alder dimers of vitamin A),



IMPURITIES

The label states:
— the number of International Units per gram,
— the name of the ester or esters.

LABELLING

In an airtight container, protected from light.
Once the container has been opened, its contents are to be used as soon as possible; any part of the contents not used at once should be protected by an atmosphere of inert gas.

STORAGE

A_{326} = absorbance at 326 nm,
 m = mass of the substance to be examined, in grams,
 V = total volume to which the substance is to be examined is diluted to give 10 IU/mL to 15 IU/mL,
1900 = factor to convert the specific absorbance of esters of retinol into International Units per gram.

$$A_{326} \times V \times 1900$$

$$100 \times m$$

expression:

The activity of the substance is determined in order to be taken into account for the production of concentrates. Dissolve 25-100 mg, weighed with an accuracy of 0.1 per cent, in 5 mL of pentane R and dilute with 2-propanol R1 to a presumed concentration of 10 IU/mL to 15 IU/mL. Measure the absorbance (2.2.25) at the absorption maximum at 326 nm. Calculate the activity of vitamin A in International Units per gram from the

ACTIVITY

The thresholds indicated under Related substances (Table 2034-1) in the general monograph Substances for pharmaceutical use (2034) do not apply.

CC1=C(C)C(=C/C=C/C(=C/C=C/C(C)COC)/C)C1

Ph Eur

DEFINITION

Solubility

Practically insoluble in *water*; soluble or partly soluble in *ethanol* (96%); miscible with *ether* and with petroleum spirit.

A. Dissolve a quantity containing 10 IU in a mixture of 100 parts of absolute ethanol and 1 part of hydrochloric acid.

chloroform and add 10 mL of *antimony trichloride solution*. A transient bright blue colour is produced immediately.

Acid value
Not more than 2.0, Appendix X B.

Place 1 g in a boiling tube (20 cm \times 2.5 cm) and dissolve in 20 mL of a mixture of 2 volumes of glacial acetic acid and 1 volume of ethanol-free chloroform. Add 1 g of finely

the titrations does not exceed 1.4 mL.

Carry out the method for descending paper chromatography. Appendix III B, using a mixture of 70 volumes of 1,4-dioxan, 15 volumes of methanol and 15 volumes of water containing 1% w/v of butylated hydroxyanisole in the bottom of the tank and as the mobile phase. Saturate the paper with a 10% w/v

not more intense than that of the spot in the corresponding chromatogram obtained with solution (2).

Carry out the test as rapidly as possible, avoiding exposure to actinic light and air, oxidising agents, oxidation catalysts

Appendix II B (Method A). If method A is found not to be valid, examine by liquid chromatography, Appendix III D

Test solution – 1.0 g of a quantity of the substance being examined containing 50,000 IU in a round-bottomed flask, add 3 mL of a freshly prepared 50% w/w solution of potassium hypoxidide

Calculate the content of vitamin A, as all-*trans*-retinol, in IU per gram from the expression:

$$A_{325} \times \frac{1830}{100m} \times V$$

where A_{325} = absorbance at 325 nm,

m = mass of the substance to be examined in grams,
V = total volume of solution containing 10 IU to 15 IU of vitamin A per mL,
1830 = conversion factor for the specific absorbance of all-*trans*-retinol in IU.

The above expression can be used only if A_{325} has a value of not greater than $A_{325, \text{corr}} / 0.970$ where $A_{325, \text{corr}}$ is the corrected absorbance at 325 nm and is given by the equation:

$$A_{325, \text{corr}} = 6.815A_{325} - 2.555A_{310} - 4.260A_{334}$$

where *A* designates the absorbance at the wavelength indicated by the subscript.

If A_{325} has a value greater than $A_{325, \text{corr}} / 0.970$, calculate the content of vitamin A from the expression:

$$A_{325, \text{corr}} \times \frac{100m}{1830 \times V}$$

The assay is not valid unless:
(a) the wavelength of maximum absorption lies between 323 nm and 327 nm and
(b) the absorbance at 300 nm relative to that at 325 nm is at most 0.73.

Method B

Carry out the method for liquid chromatography, Appendix III D, using the following solutions. Prepare solution (1) as follows. To a quantity of the substance being examined containing 50,000 IU in a round-bottomed flask, add 5 mL of a freshly prepared 10% w/v solution of ascorbic acid and 10 mL of a freshly prepared 80% w/v solution of potassium hydroxide and 100 mL of ethanol (96%). Boil under a reflux condenser on a water bath for 15 minutes. Add 100 mL of a 1% w/v solution of sodium chloride and cool. Transfer the solution to a 500 mL separating funnel rinsing the round-bottomed flask with about 75 mL of a 1% w/v solution of sodium chloride and then with 150 mL of a mixture of equal volumes of petroleum spirit (boiling range, 40° to 60°) and ether. Shake for 1 minute. When the layers have separated completely, discard the lower layer and wash the upper layer, first with 50 mL of a 3% w/v solution of potassium hydroxide in a 10% v/v solution of ethanol (96%) and then with three 50 mL quantities of a 1% w/v solution of sodium chloride. Filter the upper layer through 5 g of anhydrous sodium sulfate on a fast filter paper into a 250 mL flask suitable for a rotary evaporator. Wash the funnel with 10 mL of fresh extraction mixture, filter and combine the upper layers. Distill them at a temperature not exceeding 30° under reduced pressure (water ejector) and fill with nitrogen when evaporation is completed. Alternatively evaporate the solvent under a gentle current of nitrogen at a temperature not exceeding 30°. Dissolve the residue in *propan-2-ol*, transfer to a 25 mL volumetric flask and dilute to 25 mL with *propan-2-ol*. Gentle heating in an ultrasonic bath may be required. For solution (2) prepare a solution of retinyl acetate EPCRS in *propan-2-ol* RI so that 1 mL contains about 1000 IU of all-*trans*-retinol.

The exact concentration of solution (2) is assessed by ultraviolet absorption spectrophotometry, Appendix II B. Dilute solution (2) with *propan-2-ol* RI to a presumed concentration of 10 IU per mL to 15 IU per mL and measure the absorbance at 326 nm in matched 1-cm cells using *propan-2-ol* RI as the compensation liquid.

Calculate the content of vitamin A in IU per mL of solution (2) from the following expression, taking into account the assigned content of retinyl acetate EPCRS:

$$A_{326} \times \frac{1900 \times V_2}{100 \times V_1}$$

where A_{326} = absorbance at 326 nm,
 V_2 = volume of the diluted solution, used,
 V_1 = volume of solution (2) used,
1900 = conversion factor for the specific absorbance of retinyl acetate EPCRS in IU.

For solution (3) proceed as described for solution (1) but use 2 mL of solution (2) in place of the substance being examined.

The exact concentration of solution (3) is assessed by ultraviolet absorption spectrophotometry, Appendix II B. Dilute solution (3) with *propan-2-ol* RI to a presumed concentration of 10 IU per mL to 15 IU per mL of all-*trans*-retinol and measure the absorbance at 325 nm in matched 1 cm cells using *propan-2-ol* RI as the compensation liquid. Calculate the content of all-*trans*-retinol in IU per millilitre of solution (3) from the expression:

$$A_{325} \times \frac{100 \times V_3}{1830 \times V_4}$$

where A_{325} = absorbance at 325 nm,
 V_3 = volume of the diluted solution, used,
 V_4 = volume of solution (3) used,
1830 = conversion factor for the specific absorbance of all-*trans*-retinol in IU.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with octadecylsilyl silica gel for chromatography (5 µm to 10 µm), (b) as mobile phase at a flow rate of 1 mL per minute a mixture of 3 volumes of water and 97 volumes of methanol, (c) a detection wavelength of 325 nm, (d) a 10 µL loop injector and (e) an electronic integrator. Inject in triplicate solution (1) and solution (3). The retention time of all-*trans*-retinol is 5 minute ± 1 minute.

The assay is not valid unless (a) the chromatogram obtained with solution (1) shows a peak corresponding to that of all-*trans*-retinol in the chromatogram obtained with solution (3), (b) when using the method of standard additions to solution (1) there is greater than 95% recovery of the added retinyl acetate EPCRS, and (c) the recovery of all-*trans*-retinol in solution (3) as assessed by direct absorption spectrophotometry is greater than 95%. Calculate the content of vitamin A using the following expression:

$$A_1 \times \frac{C \times V}{1} \times \frac{A_2}{m}$$

where A_1 = area of the peak corresponding to all-*trans*-retinol in the chromatogram obtained with solution (1),
 A_2 = area of the peak corresponding to all-*trans*-retinol in the chromatogram obtained with solution (3),
C = concentration of solution (2) in IU per mL,
V = volume of solution (2) used,
1 = conversion factor for the specific absorbance of all-*trans*-retinol in IU.

C	=	concentration of retinyl acetate EPCRS in solution (2) as assessed prior to the saponification in International Units per mL (1000 IU per mL),
V	=	volume of solution (2) treated,
m	=	mass of the substance being examined in solution (1).

STORAGE

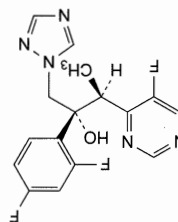
Vitamin A Ester Concentrate (Natural) should be kept in an airtight container, protected from light and stored at a temperature of 8° to 15°. Once the container has been opened its contents should be used as soon as possible; any part of the contents not used at once should be protected by an atmosphere of an inert gas.

LABELLING

The label states (1) the number of IU (Units) of vitamin A per g; (2) the name of the ester or esters; (3) the name and proportion of the principal excipients; (4) the method of restoring the solution if partial crystallisation has occurred.

Voriconazole

(Ph. Eur. monograph 2576)



C₁₆H₁₄F₃N₅O

349.3

137234-62-9

Action and use

Antifungal.

DEFINITION

(2R,3S)-2-(2,4-Difluorophenyl)-3-(5-fluoropyrimidin-4-yl)-1-(1H-1,2,4-triazol-1-yl)butan-2-ol.

Content

97.5 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white powder.

Solubility

Very slightly soluble in water, freely soluble in acetone and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison voriconazole CRS.

B. Enantiomeric purity (see Tests).

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 0.5 g in a 103 g/L solution of hydrochloric acid R and dilute to 20 mL with the same solution.

Enantiomeric purity

Liquid chromatography (2.2.29).

Test solution Dissolve 25.0 mg of the substance to be examined in 2 mL of acetonitrile R and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of voriconazole impurity D CRS in 2 mL of acetonitrile R and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 25 mg of the substance to be examined in 2 mL of acetonitrile R, add 1 mL of reference solution (a) and dilute to 50.0 mL with the mobile phase. Reference solution (c) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: silica gel BC for chiral chromatography R (5 μ m);

— temperature: 30 °C.

Mobile phase Mix 18 volumes of acetonitrile R and 82 volumes of a 0.77 g/L solution of ammonium acetate R previously adjusted to pH 5.0 with glacial acetic acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 256 nm.

Injection 20 μ L of the test solution and reference solutions (b) and (c).

Run time 2.5 times the retention time of voriconazole.

Relative retention With reference to voriconazole (retention time = about 7 min): impurity D = about 1.5.

System suitability: reference solution (b):

— resolution: minimum 4.0 between the peaks due to voriconazole and impurity D.

Limit:

— impurity D: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.2 per cent).

Impurity E

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be examined in 5.0 mL of methanol R and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dissolve 25.0 mg of voriconazole impurity E CRS in 50 mL of methanol R and dilute to 100.0 mL with the mobile phase.

Reference solution (b) Dissolve 17 mg of sodium chloride R in 100.0 mL of water R and dilute to 200.0 mL with the same solvent.

Mix 1 mL of the solution, 1 mL of reference solution (a) and 25 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Reference solution (c)

To 1.0 mL of reference solution (a) add 25 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Reference solution (d)

To 1.0 mL of reference solution (a) add 25 mL of methanol R and dilute to 50.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.0$ mm;

— stationary phase: strongly basic anion-exchange resin for chromatography R (8.5 μ m);

— temperature: 40 °C.

Mobile phase To 1500 mL of water R add 500 mL of methanol R, mix and degas; add about 175 μ L of a 470 g/L solution of sodium hydroxide R and mix.

Flow rate 1.0 mL/min.

Detection Conductivity detector; use a self-regenerating anion suppressor.

Injection 20 µL of the test solution and reference solutions (b) and (c).
Run time Twice the retention time of impurity E.
Relative retention With reference to impurity E (retention time = about 4 min): chloride = about 1.5.

System suitability:
— resolution: minimum 3.5 between the peaks due to impurity E and chloride in the chromatogram obtained with reference solution (b);
— symmetry factor: maximum 1.7 for the peak due to impurity E in the chromatogram obtained with reference solution (c).
Limit:
— impurity E: not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent).

Related substances

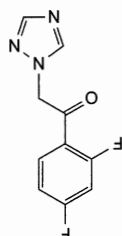
Liquid chromatography (2.2.29).
Test solution (a) Dissolve 50.0 mg of the substance to be examined in the mobile phase, sonicate if necessary, and dilute to 100.0 mL with the mobile phase. Mix well to ensure complete dissolution.
Test solution (b) Dilute 5.0 mL of test solution (a) to 100.0 mL with the mobile phase.
Reference solution (a) Dissolve 50.0 mg of voriconazole CRS in the mobile phase, sonicate if necessary, and dilute to 100.0 mL with the mobile phase. Mix well to ensure complete dissolution. Dilute 5.0 mL of the solution to 100.0 mL with the mobile phase.

Reference solution (b) Suspend 0.100 g of the substance to be examined in 10 mL of a 40 g/L solution of sodium hydroxide R and dilute to 20 mL with the mobile phase; sonicate if necessary. Allow to stand for 30 min. Dilute 1.0 mL of the solution to 100.0 mL with the mobile phase (in situ degradation to obtain impurities A and C).
Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase and mix well. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase and mix well.
Reference solution (d) Dissolve 2 mg of voriconazole impurity B CRS in the mobile phase and dilute to 200 mL with the mobile phase. Dilute 1.0 mL of the solution to 10.0 mL with the mobile phase.

Column:
— size: $l = 0.15$ m, $\varnothing = 3.9$ mm;
— stationary phase: end-capped octadecylsilyl silica gel for chromatography R (4 µm);
— temperature: 35 °C.

Mobile phase Mix 15 volumes of acetonitrile R, 30 volumes of methanol R and 55 volumes of a 1.90 g/L solution of ammonium formate R previously adjusted to pH 4.0 with anhydrous formic acid R while stirring continuously.
Flow rate 1.0 mL/min.
Detection Spectrophotometer at 256 nm.

Injection 20 µL of test solution (a) and reference solutions (b), (c) and (d).
Run time 3 times the retention time of voriconazole.
Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A and C; use the chromatogram obtained with reference solution (d) to identify the peak due to impurity B.
Relative retention With reference to voriconazole (retention time = about 8 min): impurity A = about 0.25; impurity C = about 0.3; impurity B = about 0.6.



A. 1-(2,4-difluorophenyl)-2-((1H-1,2,4-triazol-1-yl)ethanone),

IMPURITIES

Specified impurities A, B, C, D, E.
If the substance is sterile, store in a sterile, tamper-proof container.

STORAGE

If the substance is sterile, store in a sterile, tamper-proof container.
Declared content of voriconazole CRS.
Calculate the percentage content of $C_{16}H_{14}F_2N_5O$ from the Injection Test solution (b) and reference solution (a).

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

ASSAY

Procedure for the removal of bacterial endotoxins.
Less than 0.2 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

Bacterial endotoxins (2.6.14)

Maximum 0.1 per cent, determined on 1.0 g in a platinum crucible.

Sulfated ash (2.4.14)

Maximum 0.4 per cent, determined on 1.00 g.

Water (2.5.12)

using 0.25 mL of lead standard solution (10 ppm Pb) R.
0.250 g complies with test H. Prepare the reference solution Solvent mixture water R, acetone R (30:70 V/V).

Heavy metals (2.4.8)

Maximum 10 ppm.

(0.05 per cent).

the chromatogram obtained with reference solution (c) — disregard limit: 0.5 times the area of the principal peak in maximum 0.5 per cent;

sum of impurities A, B, C, D, E and unspecified impurities: with reference solution (c) (0.10 per cent);

area of the principal peak in the chromatogram obtained with reference solution (c) (0.15 per cent);

chromatogram obtained with reference solution (c) 1.5 times the area of the principal peak in the

impurities A, B, C: for each impurity, not more than impurity B = 2.1; impurity C = 0.7;

corresponding correction factor: impurity A = 0.7; the peak areas of the following impurities by the

— correction factors: for the calculation of content, multiply the peak areas of the following impurities by the

Limit:
impurities A and C.

— resolution: minimum 1.8 between the peaks due to impurities A and C.
System suitability: reference solution (b):

DEFINITION

Sodium 2-oxo-3-[(1*R*,5*S*)-3-oxo-1-phenylbutyl]-2*H*-1-benzopyran-4-olate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, hygroscopic, amorphous powder.

Solubility

Very soluble in water and in ethanol (96 per cent), soluble in acetone, very slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison warfarin sodium CRS.

B. Propan-2-ol (see Tests).

C. It gives reaction (b) of sodium (2.3.1).

TESTS**Appearance of solution**

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 1.0 g in water *R* and dilute to 20 mL with the same solvent.

pH (2.2.3)

7.6 to 8.6.

Dissolve 1.0 g in carbon dioxide-free water *R* and dilute to 100 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Solvent mixture methanol *R*, water *R* (25:75 *V/V*).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of 4-hydroxycoumarin *R* (impurity B) and 2 mg of benzalacetone *R* (impurity C) in 25 mL of methanol *R* and dilute to 100 mL with water *R*.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

stationary phase: cyanosilyl silica gel for chromatography *R* (5 μ m);

temperature: 30 °C.

Mobile phase glacial acetic acid *R*, acetonitrile *R*, water *R* (1:25:75 *V/V/V*).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 μ L.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities B and C.

Relative retention With reference to warfarin (retention time = about 9 min): impurity B = about 0.4;

impurity C = about 0.6.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to impurities B and C.

Warfarin Tablets

Warfarin Oral Suspension

Preparations

(coumarin).

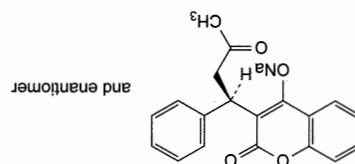
Vitamin K epoxide reductase inhibitor; oral anticoagulant

Action and use

$C_{10}H_{15}NaO_4$

330.3

129-06-6

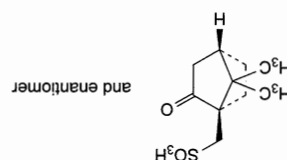


and enantiomer

(Ph. Eur. monograph 0698)

Warfarin Sodium

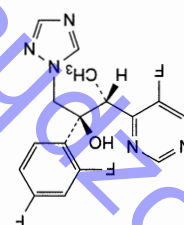
E. [(1*R*,4*S*)-7,7-dimethyl-2-oxobicyclo[2.2.1]hept-1-yl]methanesulfonic acid ((±)-10-camphorsulfonic acid).



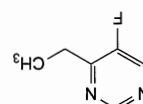
and enantiomer

enantiomer,

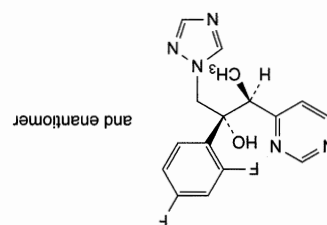
D. (2*S*,3*R*)-2-(2,4-difluorophenyl)-3-(5-fluoropyrimidin-4-yl)-1-(1*H*-1,2,4-triazol-1-yl)butan-2-ol (voriconazole



C. 4-ethyl-5-fluoropyrimidine,



B. (2*R*,3*S*)-2-(2,4-difluorophenyl)-3-pyrimidin-4-yl-1-(1*H*-1,2,4-triazol-1-yl)butan-2-ol,



and enantiomer

Ph Eur

Limits:

— **correction factors:** for the calculation of content, multiply the peak areas of the following impurities by the corresponding correction factor: impurity B = 0.5; impurity C = 0.4;

— **impurities B, C:** for each impurity, not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.15 per cent);

— **unspecified impurities:** for each impurity, not more than area of the principal peak in the chromatogram obtained with reference solution (b) (0.10 per cent);

— **total:** not more than 3 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.3 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (b)

(0.05 per cent);

Phenolic ketones

Dissolve 1.25 g in a 20 g/L solution of sodium hydroxide R

and dilute to 10.0 mL with the same solvent.

The absorbance (2.2.25) is maximum 0.20 measured at 385 nm within 15 min of preparing the solution.

Propan-2-ol (2.4.24, System A)

Maximum 0.5 per cent.

Water (2.5.12)

Maximum 4.0 per cent, determined on 0.750 g.

ASSAY

Dissolve 0.100 g in 0.01 M sodium hydroxide and dilute to 100.0 mL with the same solvent. Dilute 10.0 mL of the solution to 100.0 mL with 0.01 M sodium hydroxide. Dilute 10.0 mL of this solution to 100.0 mL with 0.01 M sodium hydroxide. Measure the absorbance (2.2.25) at the absorbance maximum at 308 nm.

Calculate the percentage content of $C_{19}H_{15}NaO_4$ taking the specific absorbance to be 431.

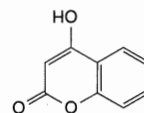
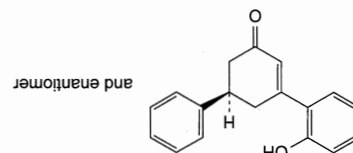
STORAGE

In an airtight container, protected from light.

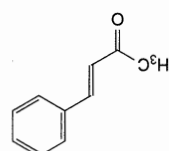
IMPURITIES

Specified impurities B, C

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A.



B. 4-hydroxy-2H-1-benzopyran-2-one (4-hydroxycoumarin),



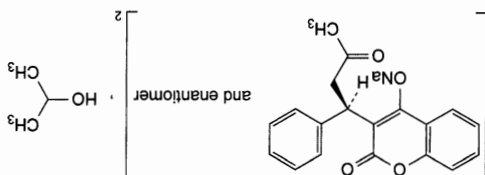
C. (3E)-4-phenylbut-3-en-2-one (benzalacetone).

Ph Eur



Warfarin Sodium Clathrate

(Ph. Eur. monograph 0699)



Action and use

Vitamin K epoxide reductase inhibitor; oral anticoagulant

Preparation

Warfarin Tablets

Ph Eur

DEFINITION

Mixture, in the form of a clathrate, of warfarin sodium (sodium 2-oxo-3-[(1R)-3-oxo-1-phenylbutyl]-2H-1-benzopyran-4-olate) and propan-2-ol in molecular proportions 2:1 (equivalent to about 92 per cent of warfarin sodium).
— **warfarin sodium:** 98.0 per cent to 102.0 per cent (anhydrous and propan-2-ol-free substance);
— **propan-2-ol:** 8.0 per cent to 8.5 per cent.

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent), soluble in acetone, very slightly soluble in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).
Comparison warfarin sodium clathrate CRS.
B. Propan-2-ol (see Tests).
C. It gives reaction (b) of sodium (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and colourless (2.2.2, Method II).

Dissolve 1.0 g in water R and dilute to 20 mL with the same solvent.

pH (2.2.3)

7.6 to 8.6.

Dissolve 1.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Solvent mixture: methanol R, water R (25:75 V/V).

Test solution Dissolve 40.0 mg of the substance to be examined in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of 4-hydroxycoumarin R

(impurity B) and 2 mg of benzalacetone R (impurity C) in

25 mL of methanol R and dilute to 100 mL with water R.

Reference solution (b) Dilute 1.0 mL of the test solution to

100.0 mL with the solvent mixture. Dilute 1.0 mL of this

solution to 10.0 mL with the solvent mixture.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.0$ mm;

— stationary phase: cyanosilyl silica gel for chromatography R

(5 μ m);

— temperature: 30 °C.

Mobile phase glacial acetic acid R, acetonitrile R, water R

(1:25:75 V/V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 260 nm.

Injection 20 μ L.

Run time Twice the retention time of warfarin.

Identification of impurities Use the chromatogram obtained

with reference solution (a) to identify the peaks due to

impurities B and C.

Relative retention With reference to warfarin (retention

time = about 9 min): impurity B = about 0.4;

impurity C = about 0.6.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to

impurities B and C.

Limits:

— **correction factors:** for the calculation of content, multiply

the peak areas of the following impurities by the

corresponding correction factor: impurity B = 0.5;

impurity C = 0.4;

— **impurities B, C:** for each impurity, not more than

1.5 times the area of the principal peak in the

chromatogram obtained with reference solution (b)

(0.15 per cent);

— **unspecified impurities:** for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (b) (0.10 per cent);

— **total:** not more than 3 times the area of the principal peak

in the chromatogram obtained with reference solution (b)

(0.3 per cent);

— **disregard limit:** 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (b)

(0.05 per cent).

Phenolic ketones

Dissolve 1.25 g in a 20 g/L solution of sodium hydroxide R

and dilute to 10.0 mL with the same solvent.

The absorbance (2.2.25) is maximum 0.20 measured at

385 nm within 15 min of preparing the solution.

Propan-2-ol (2.4.24, System A)

8.0 per cent to 8.5 per cent.

Water (2.5.12)

Maximum 0.3 per cent, determined on 2.500 g.

ASSAY

Dissolve 0.100 g in 0.01 M sodium hydroxide and dilute to

100.0 mL with the same solvent. Dilute 10.0 mL of the

PURIFIED WATER IN BULK**PRODUCTION**

Purified water in bulk is prepared by distillation, by ion exchange, by reverse osmosis or by any other suitable method from water that complies with the regulations on water intended for human consumption laid down by the competent authority.

DEFINITION

Water for the preparation of medicines other than those that are required to be both sterile and apyrogenic, unless otherwise justified and authorised.

Purified Water

(Ph. Eur. monograph 0008)

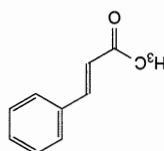
H₂O 18.02

7732-18-5

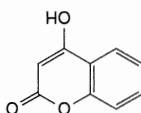


Ph Eur

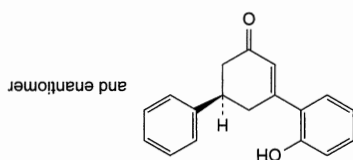
C. (3E)-4-phenylbut-3-en-2-one (benzalacetone).



B. 4-hydroxy-2H-1-benzopyran-2-one (4-hydroxycoumarin),



A. (5R)-3-(2-hydroxyphenyl)-5-phenylcyclohex-2-enone,



Control of impurities in substances for pharmaceutical use): A.

impurities for demonstration of compliance. See also 5.10.

(2034). It is therefore not necessary to identify these

by the general monograph Substances for pharmaceutical use

acceptance criterion for other/unspecified impurities and/or

the tests in the monograph. They are limited by the general

present at a sufficient level, be detected by one or other of

Other detectable impurities (the following substances would, if

Specified impurities B, C

IMPURITIES

In an airtight container, protected from light.

STORAGE

Calculate the percentage content of warfarin sodium

(C₁₉H₁₅NaO₄) taking the specific absorbance to be 431.

maximum at 308 nm.

hydroxide. Measure the absorbance (2.2.25) at the absorption

10.0 mL of this solution to 100.0 mL with 0.01 M sodium

solution to 100.0 mL with 0.01 M sodium hydroxide. Dilute

Purified water in bulk is stored and distributed in conditions designed to prevent growth of micro-organisms and to avoid any other contamination.

Microbiological monitoring

During production and subsequent storage, appropriate measures are taken to ensure that the microbial count is adequately controlled and monitored. Appropriate action levels are set so as to detect adverse trends. Under normal conditions, an appropriate action level is a microbial count of 100 CFU/mL, determined by filtration through a membrane with a nominal pore size not greater than 0.45 µm, using R2A agar and incubating at 30-35 °C for not less than 5 days. The size of the sample is to be chosen in relation to the expected result.

R2A agar
Yeast extract 0.5 g
Proteose peptone 0.5 g
Casein hydrolysate 0.5 g
Glucose 0.5 g
Starch 0.5 g
Dipotassium hydrogen phosphate 0.3 g
Magnesium sulfate, anhydrous 0.024 g
Sodium pyruvate 0.3 g
Agar 15.0 g
Purified water to 1000 mL

Adjust the pH so that after sterilisation it is 7.2 ± 0.2 . Sterilise by heating in an autoclave at 121 °C for 15 min.

Growth promotion of R2A agar

— **Preparation of test strains.** Use standardised stable suspensions of test strains or prepare them as stated in Table 0008-1. Seed lot culture maintenance techniques (seed-lot systems) are used so that the viable micro-organisms used for inoculation are not more than 5 passages removed from the original master seed-lot. Grow each of the bacterial strains separately as described in Table 0008-1. Use buffered sodium chloride-peptone solution pH 7.0 or phosphate buffer solution pH 7.2 to make test suspensions. Use the suspensions within 2 h, or within 24 h if stored at 2-8 °C. As an alternative to preparing and then diluting a fresh suspension of vegetative cells of *Bacillus subtilis*, a stable spore suspension is prepared and then an appropriate volume of the spore suspension is used for test inoculation. The stable spore suspension may be maintained at 2-8 °C for a validated period of time.

— **Growth promotion.** Test each batch of ready-prepared medium and each batch of medium, prepared either from dehydrated medium or from the ingredients described. Inoculate plates of R2A agar separately with a small number (not more than 100 CFU) of the micro-organisms indicated in Table 0008-1. Incubate under the conditions described in the table. Growth obtained must not differ by a factor greater than 2 from the calculated value for a standardised inoculum. For a freshly prepared inoculum, growth of the micro-organisms must be comparable to that obtained with a previously tested and approved batch of medium.

EQUIPMENT

Conductivity cell:

— electrodes of a suitable material such as stainless steel;
— cell constant: the cell constant is generally certified by the supplier and is subsequently verified at suitable intervals using a certified reference solution with a conductivity less than $1500 \mu\text{S}\cdot\text{cm}^{-1}$ or by comparison with a cell having a certified cell constant; the cell constant is confirmed if the value found is within 2 per cent of the certified value, otherwise re-calibration must be performed.
— **Conductometer** Accuracy of $0.1 \mu\text{S}\cdot\text{cm}^{-1}$ or better at the lowest range.

System calibration (conductivity cell and conductometer):
— against one or more suitable certified reference solutions; — accuracy: within 3 per cent of the measured conductivity plus $0.1 \mu\text{S}\cdot\text{cm}^{-1}$.

Conductometer calibration Calibration is carried out for each range of measurement to be used, after disconnection of the conductivity cell, using certified precision resistors or equivalent devices with an uncertainty not greater than 0.1 per cent of the certified value.

If in-line conductivity cells cannot be dismantled, system calibration may be performed against a calibrated conductivity-measuring instrument with a conductivity cell placed close to the cell to be calibrated in the water flow.

PROCEDURE

Measure the conductivity without temperature compensation, recording simultaneously the temperature. Temperature-compensated measurement may be performed after suitable validation.

The water to be examined meets the requirements if the measured conductivity at the recorded temperature is not greater than the value in Table 0008-2.

For temperatures not listed in Table 0008-2, calculate the maximal permitted conductivity by interpolation between the next lower and next higher data points in the table.

Micro-organism	Preparation of the test strain	Growth promotion
<i>Pseudomonas aeruginosa</i>	Casein soyabean digest agar or casein soyabean digest broth	R2A agar
such as: ATCC 9027 NCIMB 8626 CIP 82.118 NBRC 13275	30-35 °C 18-24 h	30-35 °C ≤ 100 CFU ≤ 3 days
<i>Bacillus subtilis</i>	Casein soyabean digest agar or casein soyabean digest broth	R2A agar
such as: ATCC 6633 NCIMB 8054 CIP 52.62 NBRC 3134	30-35 °C 18-24 h	30-35 °C ≤ 100 CFU ≤ 3 days

Table 0008-1. – Growth promotion of R2A agar

Table 0008-2. – Temperature and conductivity requirements

Temperature (°C)	Conductivity (µS-cm ⁻¹)
0	2.4
10	3.6
20	4.3
25	5.1
30	5.4
40	6.5
50	7.1
60	8.1
70	9.1
75	9.7
80	9.7
90	9.7
100	10.2

Heavy metals

If purified water in bulk complies with the requirement for conductivity prescribed for *Water for injections* (0169) in bulk, it is not necessary to carry out the test for heavy metals

prescribed below.

CHARACTERS**Appearance**

Clear and colourless liquid.

TESTS**Nitrates**

Maximum 0.2 ppm.

Place 5 mL in a test-tube immersed in iced water, add

0.4 mL of a 100 g/L solution of potassium chloride R, 0.1 mL

of diphenylamine solution R and, dropwise with shaking, 5 mL

of nitrogen-free sulfuric acid R. Transfer the tube to a water-

bath at 50 °C. After 15 min, any blue colour in the solution

is not more intense than that in a reference solution prepared

at the same time in the same manner using a mixture of

4.5 mL of nitrate-free water R and 0.5 mL of nitrate standard

solution (2 ppm NO₃) R.

Aluminium (2.4.17)

Maximum 10 ppb, if intended for use in the manufacture of

dialysis solutions.

Prescribed solution To 400 mL of the water to be examined

add 10 mL of acetate buffer solution pH 6.0 R and 100 mL of

distilled water R.

Reference solution Mix 2 mL of aluminium standard

solution (2 ppm Al) R, 10 mL of acetate buffer solution

pH 6.0 R and 98 mL of distilled water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R

and 100 mL of distilled water R.

Heavy metals (2.4.8)

Maximum 0.1 ppm.

To 200 mL add 0.15 mL of 0.1 M nitric acid and heat in a

glass evaporating dish on a water-bath until the volume is

reduced to 20 mL. 12 mL of the concentrated solution

complies with test A. Prepare the reference solution using

10 mL of lead standard solution (1 ppm Pb) R and adding

0.075 mL of 0.1 M nitric acid. Prepare the blank solution

adding 0.075 mL of 0.1 M nitric acid.

PURIFIED WATER IN CONTAINERS**DEFINITION**

Purified water in bulk that has been filled and stored in conditions designed to assure the required microbiological quality. It is free from any added substances.

CHARACTERS**Appearance**

Clear and colourless liquid.

TESTS

It complies with the tests prescribed in the section on Purified water in bulk and with the following additional tests.

Acidity or alkalinity

To 10 mL, freshly boiled and cooled in a borosilicate glass flask, add 0.05 mL of methyl red solution R. The solution is not coloured red.

To 10 mL add 0.1 mL of bromothymol blue solution R1.

The solution is not coloured blue.

Oxidisable substances

To 100 mL add 10 mL of dilute sulfuric acid R and 0.1 mL of 0.02 M potassium permanganate and boil for 5 min. The solution remains faintly pink.

Chlorides

To 10 mL add 1 mL of dilute nitric acid R and 0.2 mL of silver nitrate solution R2. The solution shows no change in appearance for at least 15 min.

Sulfates

To 10 mL add 0.1 mL of dilute hydrochloric acid R and 0.1 mL of barium chloride solution R1. The solution shows no change in appearance for at least 1 h.

Ammonium

Maximum 0.2 ppm.

To 20 mL add 1 mL of alkaline potassium tetradodecylmercurate

solution R. After 5 min, examine the solution down the

vertical axis of the tube. The solution is not more intensely

coloured than a standard prepared at the same time by

adding 1 mL of alkaline potassium tetradodecylmercurate solution R

to a mixture of 4 mL of ammonium standard solution

(1 ppm NH₄) R and 16 mL of ammonium-free water R.

Calcium and magnesium

To 100 mL add 2 mL of ammonium chloride buffer solution pH 10.0 R, 50 mg of mordant black 11 trihydrate R and 0.5 mL of 0.01 M sodium edetate. A pure blue colour is produced.

Residue on evaporation

Maximum 0.001 per cent.

Evaporate 100 mL to dryness on a water-bath and dry in an

oven at 100-105 °C. The residue weighs a maximum of

1 mg.

Microbial contamination

TAMC: acceptance criterion 10² CFU/mL (2.6.12).

Use casein soya bean digest agar.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of dialysis solutions.

Ph Eur

Highly Purified Water

(Ph. Eur. monograph 1927)

H₂O 18.02

Ph Eur

DEFINITION

Water intended for use in the preparation of medicinal products where water of high biological quality is needed, except where water for injections (0169) is required.

PRODUCTION

Highly purified water is obtained from water that complies with the regulations on water intended for human consumption laid down by the competent authority.

Current production methods include, for example, double-current reverse osmosis coupled with other suitable techniques such as ultrafiltration and deionisation. Correct operation and maintenance of the system is essential.

In order to ensure the appropriate quality of the water, validated procedures and in-process monitoring of the electrical conductivity and regular microbial monitoring are applied.

Highly purified water is stored in bulk and distributed in conditions designed to prevent growth of micro-organisms and to avoid any other contamination.

Microbiological monitoring

During production and subsequent storage, appropriate measures are taken to ensure that the microbial count is adequately controlled and monitored. Appropriate alert and action levels are set so as to detect adverse trends. Under normal conditions, an appropriate action level is a microbial count of 10 CFU per 100 mL when determined by filtration through a membrane with a nominal pore size not greater than 0.45 µm, using R2A agar, at least 200 mL of highly purified water and incubating at 30–35 °C for not less than 5 days.

R2A agar	0.5 g
Yeast extract	0.5 g
Proteose peptone	0.5 g
Caselin hydrolysate	0.5 g
Glucose	0.5 g
Starch	0.5 g
Dipotassium hydrogen phosphate	0.3 g
Magnesium sulfate, anhydrous	0.024 g
Sodium pyruvate	0.3 g
Agar	15.0 g
Purified water	to 1000 mL

Adjust the pH so that after sterilisation it is 7.2 ± 0.2. Sterilise by heating in an autoclave at 121 °C for 15 min.

lowest range.

— cell constant: the cell constant is generally certified by the supplier and is subsequently verified at suitable intervals using a certified reference solution with a conductivity less than 1500 µS·cm⁻¹ or by comparison with a cell having a certified cell constant; the cell constant is confirmed if the value found is within 2 per cent of the certified value, otherwise re-calibration must be performed.

— electrodes of a suitable material such as stainless steel;

Conductivity cell.

EQUIPMENT

Determine the conductivity off-line or in-line under the following conditions.

Conductivity
Maximum 0.5 mg/L.
Total organic carbon (2.2.44)

Micro-organism	Preparation of the test	Growth promotion
<i>Pseudomonas aeruginosa</i>	Caselin soyabean digest R2A agar agar or caselin soyabean digest broth 30–35 °C ≤ 100 CFU ≤ 3 days	
<i>Bacillus subtilis</i>	Caselin soyabean digest R2A agar agar or caselin soyabean digest broth 30–35 °C ≤ 100 CFU ≤ 3 days	
such as: ATCC 9027 NCIMB 8626 CIP 82.118 NBRC 13275		
such as: ATCC 6633 NCIMB 8054 CIP 52.62 NBRC 3134		

Table 1927.-1. – Growth promotion of R2A agar

— **Growth promotion.** Test each batch of ready-prepared medium and each batch of medium, prepared either from dehydrated medium or from the ingredients described. Inoculate plates of R2A agar separately with a small number (not more than 100 CFU) of the micro-organisms indicated in Table 1927.-1. Incubate under the conditions described in the table. Growth obtained must not differ by a factor greater than 2 from the calculated value for a standardised inoculum. For a freshly prepared inoculum, growth of the micro-organisms must be comparable to that obtained with a previously tested and approved batch of medium.

— **Preparation of test strains.** Use standardised stable suspensions of test strains or prepare them as stated in Table 1927.-1. Seed lot culture maintenance techniques (seed-lot systems) are used so that the viable micro-organisms removed from the original master seed-lot. Grow each of the bacterial strains separately as described in Table 1927.-1. Use buffered sodium chloride-peptone solution pH 7.0 or phosphate buffer solution pH 7.2 to make test suspensions. Use the suspensions within 2 h, or within 24 h if stored at 2–8 °C. As an alternative to preparing and then diluting a fresh suspension of vegetative cells of *Bacillus subtilis*, a stable spore suspension is prepared and then an appropriate volume of the spore suspension is used for test inoculation. The stable spore suspension may be maintained at 2–8 °C for a validated period of time.

System calibration (conductivity cell and conductometer):
— against one or more suitable certified reference solutions;
— accuracy: within 3 per cent of the measured conductivity plus $0.1 \mu\text{S}\cdot\text{cm}^{-1}$.

Conductometer calibration Calibration is carried out for each range of measurement to be used, after disconnection of the conductivity cell, using certified precision resistors or equivalent devices with an uncertainty not greater than 0.1 per cent of the certified value.
If in-line conductivity cells cannot be dismantled, system calibration may be performed against a calibrated conductivity-measuring instrument with a conductivity cell placed close to the cell to be calibrated in the water flow.
Temperature measurement Tolerance $\pm 2^\circ\text{C}$.

PROCEDURE

Stage 1

1. Measure the conductivity without temperature compensation, recording simultaneously the temperature. Temperature-compensated measurement may be performed after suitable validation.
2. Using Table 1927-2, find the closest temperature value that is not greater than the measured temperature. The corresponding conductivity value is the limit at that temperature.
3. If the measured conductivity is not greater than the value in Table 1927-2, the water to be examined meets the requirements of the test for conductivity. If the conductivity is higher than the value in Table 1927-2, proceed with stage 2.

Table 1927-2. – Stage 1
Temperature and conductivity requirements (for non-temperature-compensated conductivity measurements)

Temperature ($^\circ\text{C}$)	Conductivity ($\mu\text{S}\cdot\text{cm}^{-1}$)
0	0.6
5	0.8
10	0.9
15	1.0
20	1.1
25	1.3
30	1.4
35	1.5
40	1.7
45	1.8
50	1.9
55	2.1
60	2.2
65	2.4
70	2.5
75	2.7
80	2.7
85	2.7
90	2.7
95	2.9
100	3.1

Table 1927-3. – Stage 3 – pH and conductivity requirements (for atmosphere and temperature equilibrated samples)

pH	Conductivity ($\mu\text{S}\cdot\text{cm}^{-1}$)
5.0	4.7
5.1	4.1
5.2	3.6
5.3	3.3
5.4	3.0
5.5	2.8
5.6	2.6
5.7	2.5
5.8	2.4
5.9	2.4
6.0	2.4
6.1	2.4
6.2	2.5
6.3	2.4
6.4	2.3
6.5	2.2
6.6	2.1
6.7	2.6
6.8	3.1
6.9	3.8
7.0	4.6

4. Transfer a sufficient amount of the water to be examined (100 mL or more) to a suitable container, and stir the test sample. Adjust the temperature, if necessary, and while maintaining it at $25 \pm 1^\circ\text{C}$, begin vigorously agitating the test sample while periodically observing the conductivity. When the change in conductivity (due to uptake of atmospheric carbon dioxide) is less than $0.1 \mu\text{S}\cdot\text{cm}^{-1}$ per 5 min, note the conductivity.
5. If the conductivity is not greater than $2.1 \mu\text{S}\cdot\text{cm}^{-1}$, the water to be examined meets the requirements of the test for conductivity. If the conductivity is greater than $2.1 \mu\text{S}\cdot\text{cm}^{-1}$, proceed with stage 3.
6. Perform this test within approximately 5 min of the conductivity determination in step 5 under stage 2, while maintaining the sample temperature at $25 \pm 1^\circ\text{C}$. Add a recently prepared saturated solution of *potassium chloride R* to the test sample (0.3 mL per 100 mL of the test sample), and determine the pH (2.2.3) to the nearest 0.1 pH unit.
7. Using Table 1927-3, determine the conductivity limit at the measured pH value in step 6. If the measured conductivity in step 4 under stage 2 is not greater than the conductivity requirements for the pH determined, the water to be examined meets the requirements of the test for conductivity. If either the measured conductivity is greater than this value or the pH is outside the range of 5.0–7.0, the water to be examined does not meet the requirements of the test for conductivity.

Stage 2

CHARACTERS**Appearance**

Clear and colourless liquid.

TESTS**Nitrates**

Maximum 0.2 ppm.

Place 5 mL in a test-tube immersed in iced water, add

0.4 mL of a 100 g/L solution of potassium chloride R, 0.1 mL of diphenylamine solution R and, dropwise with shaking, 5 mL of nitrobenzene-sulfuric acid R. Transfer the tube to a water-

bath at 50 °C. After 15 min, any blue colour in the solution is not more intense than that in a reference solution prepared

at the same time in the same manner using a mixture of 4.5 mL of nitrate-free water R and 0.5 mL of nitrate standard solution (2 ppm NO₃) R.

Aluminium (2.4.17)

Maximum 10 ppb, if intended for use in the manufacture of dialysis solutions.

Prescribed solution To 400 mL of the water to be examined add 10 mL of acetate buffer solution pH 6.0 R and 100 mL of distilled water R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of distilled water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of distilled water R.

Bacterial endotoxins (2.6.14)

Less than 0.25 IU/mL.

LABELLING

The label states, where applicable, that the substance is suitable for use in the manufacture of dialysis solutions.

Ph Eur

Water for injections

(Ph. Eur. monograph 0169)

H₂O

Ph Eur

DEFINITION

Water for the preparation of medicines for parenteral administration when water is used as vehicle (water for injections in bulk) and for dissolving or diluting substances or preparations for parenteral administration (sterilised water for injections).

WATER FOR INJECTIONS IN BULK**PRODUCTION**

Water for injections in bulk is obtained from water that complies with the regulations on water intended for human consumption laid down by the competent authority or from purified water by distillation in an apparatus of which the parts in contact with the water are of neutral glass, quartz or a suitable metal and which is fitted with an effective device to prevent the entrainment of droplets. The correct maintenance of the apparatus is essential. The first portion of the distillate obtained when the apparatus begins to function is discarded and the distillate is collected.

In order to ensure the appropriate quality of the water, validated procedures and in-process-monitoring of the

electrical conductivity and regular microbial monitoring are applied.

Water for injections in bulk is stored and distributed in conditions designed to prevent growth of micro-organisms and to avoid any other contamination.

Microbiological monitoring

During production and subsequent storage, appropriate measures are taken to ensure that the microbial count is adequately controlled and monitored. Appropriate alert and action levels are set so as to detect adverse trends. Under normal conditions, an appropriate action level is a microbial count of 10 CFU per 100 mL when determined by filtration through a membrane with a nominal pore size not greater than 0.45 µm, using R2A agar, using at least 200 mL of water for injections in bulk and incubating at 30–35 °C for not less than 5 days. For aseptic processing, stricter alert levels may need to be applied.

R2A agar

Yeast extract 0.5 g

Proteose peptone 0.5 g

Casein hydrolysate 0.5 g

Glucose 0.5 g

Starch 0.5 g

Dipotassium hydrogen phosphate 0.3 g

Magnesium sulfate, anhydrous 0.024 g

Sodium pyruvate 0.3 g

Agar 15.0 g

Purified water to 1000 mL

Adjust the pH so that after sterilisation it is 7.2 ± 0.2. Sterilise by heating in an autoclave at 121 °C for 15 min.

Growth promotion of R2A agar

suspensions of test strains or prepare them as stated in Table 0169.-1. Seed lot culture maintenance techniques (seed-lot systems) are used so that the viable micro-organisms used for inoculation are not more than 5 passages removed from the original master seed-lot. Grow each of the bacterial strains separately as described in Table 0169.-1. Use buffered sodium chloride-peptone solution pH 7.0 or phosphate buffer solution pH 7.2 to make test suspensions. Use the suspensions within 2 h, or within 24 h if stored at 2–8 °C. As an alternative to preparing and then diluting a fresh suspension of vegetative cells of *Bacillus subtilis*, a stable spore suspension is prepared and then an appropriate volume of the spore suspension is used for test inoculation. The stable spore suspension may be maintained at 2–8 °C for a validated period of time.

Growth promotion. Test each batch of ready-prepared medium and each batch of medium, prepared either from dehydrated medium or from the ingredients described. Inoculate plates of R2A agar separately with a small number (not more than 100 CFU) of the micro-organisms indicated in Table 0169.-1. Incubate under the conditions described in the table. Growth obtained must not differ by a factor greater than 2 from the calculated value for a standardised inoculum. For a freshly prepared inoculum, growth of the micro-organisms must be comparable to that obtained with a previously tested and approved batch of medium.

Table 0169-1. - Growth promotion of R2A agar

Micro-organism	Preparation of the test strain	Growth promotion
<i>Pseudomonas aeruginosa</i>	Casein soyabean digest agar or casein soyabean digest broth 30-35 °C ≤ 100 CFU 18-24 h ≤ 3 days	ATCC 9027 NCIMB 8626 CIP 82.118 NBRC 13275
<i>Bacillus subtilis</i>	Casein soyabean digest R2A agar agar or casein soyabean digest broth 30-35 °C ≤ 100 CFU 18-24 h ≤ 3 days	ATCC 6633 NCIMB 8054 CIP 52.62 NBRC 3134

Total organic carbon (2.2.44)
Maximum 0.5 mg/L.

Conductivity

Determine the conductivity off-line or in-line under the following conditions.

EQUIPMENT

Conductivity cell:

- electrodes of a suitable material such as stainless steel;
- cell constant: the cell constant is generally certified by the supplier and is subsequently verified at suitable intervals using a certified reference solution with a conductivity less than $1500 \mu\text{S}\cdot\text{cm}^{-1}$ or by comparison with a cell having a certified cell constant. The cell constant is confirmed if the value found is within 2 per cent of the certified value, otherwise re-calibration must be performed.
- Conductometer* Accuracy of $0.1 \mu\text{S}\cdot\text{cm}^{-1}$ or better at the lowest range.

System calibration (conductivity cell and conductometer):

- against one or more suitable certified reference solutions;
- accuracy: within 3 per cent of the measured conductivity plus $0.1 \mu\text{S}\cdot\text{cm}^{-1}$.

Conductometer calibration Calibration is carried out for each range of measurement to be used, after disconnection of the conductivity cell, using certified precision resistors or equivalent devices with an uncertainty not greater than 0.1 per cent of the certified value.

If in-line conductivity cells cannot be dismantled, system calibration may be performed against a calibrated conductivity-measuring instrument with a conductivity cell placed close to the cell to be calibrated in the water flow.

Temperature measurement Tolerance $\pm 2^\circ\text{C}$.

PROCEDURE

Stage 1

1. Measure the conductivity without temperature compensation, recording simultaneously the temperature. Temperature-compensated measurement may be performed after suitable validation.
2. Using Table 0169-2, find the closest temperature value that is not greater than the measured temperature. The corresponding conductivity value is the limit at that temperature.
3. If the measured conductivity is not greater than the value in Table 0169-2, the water to be examined meets the requirements of the test for conductivity. If the conductivity is higher than the value in Table 0169-2, proceed with stage 2.

6. Perform this test within approximately 5 min of the conductivity determination in step 5 under stage 2, while maintaining the sample temperature at $25 \pm 1^\circ\text{C}$. Add a recently prepared saturated solution of *potassium chloride K* to the test sample (0.3 mL per 100 mL of the test sample), and determine the pH (2.2.3) to the nearest 0.1 pH unit.
7. Using Table 0169-3, determine the conductivity limit at the measured pH value in step 6. If the measured conductivity in step 4 under stage 2 is not greater than the conductivity requirements for the pH determined, the water to be examined meets the requirements of the test for conductivity. If either the measured conductivity is greater than this value or the pH is outside the range of 5.0-7.0, the water to be examined does not meet the requirements of the test for conductivity.

Stage 3

5. If the conductivity is not greater than $2.1 \mu\text{S}\cdot\text{cm}^{-1}$, the water to be examined meets the requirements of the test for conductivity. If the conductivity is greater than $2.1 \mu\text{S}\cdot\text{cm}^{-1}$, proceed with stage 3.
4. Transfer a sufficient amount of the water to be examined (100 mL or more) to a suitable container, and stir the test sample. Adjust the temperature, if necessary, and while maintaining it at $25 \pm 1^\circ\text{C}$, begin vigorously agitating the test sample while periodically observing the conductivity. When the change in conductivity (due to uptake of atmospheric carbon dioxide) is less than $0.1 \mu\text{S}\cdot\text{cm}^{-1}$ per 5 min, note the conductivity.

Stage 2

Temperature (°C)	Conductivity ($\mu\text{S}\cdot\text{cm}^{-1}$)
0	0.6
5	0.8
10	0.9
15	1.0
20	1.1
25	1.3
30	1.4
35	1.5
40	1.7
45	1.8
50	1.9
55	2.1
60	2.2
65	2.4
70	2.5
75	2.7
80	2.7
85	2.7
90	2.7
95	2.9
100	3.1

Table 0169-2. - Stage 1
Temperature and conductivity requirements (for non-temperature-compensated conductivity measurements)

STERILISED WATER FOR INJECTIONS

DEFINITION

Water for injections in bulk that has been distributed into suitable containers, closed and sterilised by heat in conditions which ensure that the product still complies with the test for bacterial endotoxins. Sterilised water for injections is free from any added substances. Examined in suitable conditions of visibility, it is clear and colourless. Each container contains a sufficient quantity of water for injections to permit the nominal volume to be withdrawn.

TESTS

Acidity or alkalinity

To 20 mL add 0.05 mL of *phenol red solution R*. If the solution is yellow, it becomes red on the addition of 0.1 mL of 0.01 M *sodium hydroxide*; if red, it becomes yellow on the addition of 0.15 mL of 0.01 M *hydrochloric acid*.

Conductivity

Maximum $25 \mu\text{S}\cdot\text{cm}^{-1}$ for containers with a nominal volume of 10 mL or less; maximum $5 \mu\text{S}\cdot\text{cm}^{-1}$ for containers with a nominal volume greater than 10 mL.

Use equipment and the calibration procedure as defined under *Water for injections* in bulk, maintaining the sample temperature at $25 \pm 1^\circ\text{C}$.

Oxidisable substances

For containers with a nominal volume less than 50 mL: heat 100 mL to boiling with 10 mL of *dilute sulfuric acid R*, add 0.4 mL of 0.02 M *potassium permanganate* and boil for 5 min; the solution remains faintly pink.

For containers with a nominal volume equal to or greater than 50 mL: heat 100 mL to boiling with 10 mL of *dilute sulfuric acid R*, add 0.2 mL of 0.02 M *potassium permanganate* and boil for 5 min; the solution remains faintly pink.

Chlorides (2.4.4)

Maximum 0.5 ppm for containers with a nominal volume of 100 mL or less.

15 mL complies with the limit test for chlorides. Prepare the standard using a mixture of 1.5 mL of *chloride standard solution (5 ppm Cl)* R and 13.5 mL of *water R*. Examine the solutions down the vertical axes of the tubes.

For containers with a nominal volume greater than 100 mL, use the following test: to 10 mL add 1 mL of *dilute nitric acid R* and 0.2 mL of *silver nitrate solution R2*. The solution shows no change in appearance for at least 15 min.

Nitrates

Maximum 0.2 ppm.

Place 5 mL in a test-tube immersed in iced water, add 0.4 mL of a 100 g/L solution of *potassium chloride R*, 0.1 mL of *diphenylamine solution R* and, dropwise with shaking, 5 mL of *nitrogen-free sulfuric acid R*. Transfer the tube to a water-bath at 50°C . After 15 min, any blue colour in the solution is not more intense than that in a reference solution prepared at the same time in the same manner using a mixture of 4.5 mL of *nitrate-free water R* and 0.5 mL of *nitrate standard solution (2 ppm NO₃) R*.

Sulfates

To 10 mL add 0.1 mL of *dilute hydrochloric acid R* and 0.1 mL of *barium chloride solution R1*. The solution shows no change in appearance for at least 1 h.

Aluminium (2.4.17)

Maximum 10 ppb, if intended for use in the manufacture of dialysis solutions.

CHARACTERS

Appearance

Clear and colourless liquid.

TESTS

Nitrates

Maximum 0.2 ppm.

Place 5 mL in a test-tube immersed in iced water, add 0.4 mL of a 100 g/L solution of *potassium chloride R*, 0.1 mL of *diphenylamine solution R* and, dropwise with shaking, 5 mL of *nitrogen-free sulfuric acid R*. Transfer the tube to a water-bath at 50°C . After 15 min, any blue colour in the solution is not more intense than that in a reference solution prepared at the same time in the same manner using a mixture of 4.5 mL of *nitrate-free water R* and 0.5 mL of *nitrate standard solution (2 ppm NO₃) R*.

Aluminium (2.4.17)

Maximum 10 ppb, if intended for use in the manufacture of dialysis solutions.

Prescribed solution To 400 mL of the water to be examined add 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of

distilled water R.

Reference solution Mix 2 mL of *aluminium standard solution (2 ppm Al)* R, 10 mL of *acetate buffer solution*

pH 6.0 R and 98 mL of *distilled water R*.

Blank solution Mix 10 mL of *acetate buffer solution pH 6.0 R* and 100 mL of *distilled water R*.

Bacterial endotoxins (2.6.14)

Less than 0.25 IU/mL.

Table 0169-3. – Stage 3
pH and conductivity requirements (for atmosphere- and temperature-equilibrated samples)

pH	Conductivity ($\mu\text{S}\cdot\text{cm}^{-1}$)
5.0	4.7
5.1	4.1
5.2	3.6
5.3	3.3
5.4	3.0
5.5	2.8
5.6	2.6
5.7	2.5
5.8	2.4
5.9	2.4
6.0	2.4
6.1	2.4
6.2	2.5
6.3	2.4
6.4	2.3
6.5	2.2
6.6	2.1
6.7	2.6
6.8	3.1
6.9	3.8
7.0	4.6

liquid. It is stored (where necessary) and distributed under conditions designed to prevent growth of micro-organisms and to avoid other contamination.

For monitoring purposes, the following tests are carried out at regular intervals to demonstrate consistency in the quality of the water used for the preparation of extracts.

Conductivity (2.2.38)

Maximum $2500 \mu\text{S}\cdot\text{cm}^{-1}$, measured at 20°C .

Nitrate

Liquid chromatography (2.2.29).

Test solution The substance to be examined.

Reference solution Dissolve 0.163 g of potassium nitrate R and 0.149 g of potassium bromide R in water R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of the solution to 100.0 mL with water R.

Column:

— size: $l = 0.25\text{ m}$, $\phi = 4\text{ mm}$;

— stationary phase: anion-exchange resin R3.

Mobile phase Dissolve 0.265 g of anhydrous sodium carbonate R and 0.210 g of sodium hydrogen carbonate R in water R and dilute to 1000.0 mL with the same solvent.

Flow rate 1.2 mL/min .

Detection Conductivity detector, using a self-regenerating anion suppressor.

Injection $25\text{ }\mu\text{L}$.

Run time Twice the retention time of nitrate.

Relative retention With reference to nitrate (retention time = about 7 min): bromide = about 0.9 .

System suitability: reference solution:

— resolution: minimum 2.0 between the peaks due to bromide and nitrate.

Limit:

— nitrate: maximum 50 ppm .

Microbiological monitoring

Appropriate measures are taken to ensure that the microbial count is adequately controlled and monitored. Appropriate alert and action levels are set so as to detect adverse trends.

Under normal conditions, an appropriate action level is a microbial count of 100 CFU/mL , determined by filtration through a membrane with a nominal pore size not greater than $0.45\text{ }\mu\text{m}$, using casein soya bean digest agar and incubating at $30\text{--}35^\circ\text{C}$ for not less than 5 days .

The size of the sample is to be chosen in relation to the expected result.

Casein soya bean digest agar

Pancreatic digest of casein

Papain digest of soya bean

Sodium chloride

Agar

Purified water

Adjust the pH so that after sterilisation it is 7.3 ± 0.2 .

Sterilise in an autoclave using a validated cycle.

Growth promotion of casein soya bean digest agar

— Preparation of test strains. Use standardised stable suspensions of test strains or prepare them as stated in Table 2.2.49.-1. Seed lot culture maintenance techniques (seed-lot systems) are used so that the viable micro-organisms used for inoculation are not more than 5 passages removed from the original master seed-lot. Grow each of the bacterial strains separately as described in Table 2.2.49.-1. Use buffered sodium chloride-peptone solution pH 7.0 or phosphate buffer solution pH 7.2 to

Prescribed solution To 400 mL of the water to be examined add 10 mL of acetate buffer solution pH 6.0 R and 100 mL of distilled water R.

Reference solution Mix 2 mL of aluminium standard solution (2 ppm Al) R, 10 mL of acetate buffer solution pH 6.0 R and 98 mL of distilled water R.

Blank solution Mix 10 mL of acetate buffer solution pH 6.0 R and 100 mL of distilled water R.

Ammonium

For containers with a nominal volume less than 50 mL :

maximum 0.6 ppm ; for containers with a nominal volume equal to or greater than 50 mL : maximum 0.2 ppm .

Containers with a nominal volume less than 50 mL : to

20 mL , add 1 mL of alkaline potassium tetraiodomercurate solution R; after 5 min , examine the solution down the vertical axis of the tube; the solution is not more intensely coloured than a standard prepared at the same time by adding 1 mL of alkaline potassium tetraiodomercurate solution R to a mixture of 4 mL of ammonium standard solution (3 ppm NH_4) R and 16 mL of ammonium-free water R.

Containers with a nominal volume equal to or greater than 50 mL : to 20 mL , add 1 mL of alkaline potassium tetraiodomercurate solution R; after 5 min , examine the solution down the vertical axis of the tube; the solution is not more intensely coloured than a standard prepared at the same time by adding 1 mL of alkaline potassium tetraiodomercurate solution R to a mixture of 4 mL of ammonium standard solution (3 ppm NH_4) R and 16 mL of ammonium-free water R.

Calcium and magnesium

To 100 mL , add 2 mL of ammonium chloride buffer solution pH 10.0 R, 50 mg of mordant black 11 nitrate R and 0.5 mL of 0.01 M sodium edetate. A pure blue colour is produced.

Residue on evaporation

Maximum 4 mg (0.004 per cent) for containers with a nominal volume of 10 mL or less; maximum 3 mg (0.003 per cent) for containers with a nominal volume greater than 10 mL .

Evaporate 100 mL to dryness on a water-bath and dry in an oven at $100\text{--}105^\circ\text{C}$.

Particulate contamination: sub-visible particles (2.9.19)

It complies with test A or test B, as appropriate.

Sterility (2.6.1)

It complies with the test for sterility.

Bacterial endotoxins (2.6.14)

Less than 0.25 IU/mL .

Water for Preparation of Extracts

(Ph Eur monograph 2249)

DEFINITION

Water intended for the preparation of Extracts (0765)

complies with the sections Purified water in bulk or Purified water in containers in the monograph Purified water (0008), or is water intended for human consumption of a quality equivalent to that defined in Directive 98/83/EC which is monitored according to the Production section described below.

PRODUCTION

When water intended for human consumption is used as water for preparation of extracts it is a clear, colourless

make test suspensions. Use the suspensions within 2 h, or within 24 h if stored at 2-8 °C. As an alternative to preparing and then diluting a fresh suspension of vegetative cells of *Bacillus subtilis*, a stable spore suspension is prepared and then an appropriate volume of the spore suspension is used for test inoculation. The stable spore suspension may be maintained at 2-8 °C for a validated period of time.

— *Growth promotion*. Test each batch of ready-prepared medium and each batch of medium, prepared either from dehydrated medium or from the ingredients described. Inoculate plates of casein soya bean digest agar separately with a small number (not more than 100 CFU) of the micro-organisms indicated in Table 2249-1. Incubate under the conditions described in this table. Growth obtained must not differ by a factor greater than 2 from the calculated value for a standardised inoculum. For a freshly prepared inoculum, growth of the micro-organisms must be comparable to that obtained with a previously tested and approved batch of medium.

Table 2249-1. – *Growth promotion of casein soya bean digest agar*

Micro-organism	Preparation of the test strain	Growth promotion
<i>Pseudomonas aeruginosa</i>	Casein soya bean digest agar or casein soya bean digest broth	≤ 100 CFU
ATCC 9027	30-35 °C	≤ 3 days
NCIMB 8626	18-24 h	≤ 3 days
CIP 82.118		
NBRC 13275		
<i>Bacillus subtilis</i>	Casein soya bean digest agar or casein soya bean digest broth	≤ 100 CFU
ATCC 6633	30-35 °C	≤ 3 days
NCIMB 8054	18-24 h	≤ 3 days
CIP 52.62		
NBRC 3134		

Ph Eur

Virgin Wheat-germ Oil

(Ph. Eur. monograph 1480)

Ph Eur

DEFINITION

Fatty oil obtained from the germ of the grain of *Triticum aestivum* L. by cold expression or other suitable mechanical means.

CHARACTERS

Appearance

Clear, light yellow or golden-yellow liquid.

Solubility

Practically insoluble in water and in ethanol (96 per cent), miscible with light petroleum (bp: 40-60 °C).

Relative density

About 0.925.

Refractive index About 1.475.

IDENTIFICATION

A. Identification of fatty oils by thin-layer chromatography (2.3.2).

(2.3.2).

Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.

B. Composition of fatty acids (see Tests).

Acid value (2.5.1)
Maximum 20.0.
Peroxide value (2.5.5, Method A)
Maximum 15.0.
Unsaponifiable matter (2.5.7)
Maximum 5.0 per cent, determined on 5.0 g.
Composition of fatty acids (2.4.22, Method C)
Maximum 5.0 per cent, determined on 5.0 g.
Alkaline impurities (2.4.19)
Maximum 5.0 per cent, determined on 5.0 g.
It complies with the test.

TESTS

A. Identification of fatty oils by thin-layer chromatography (2.3.2).
Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.
B. Composition of fatty acids (see Tests).

IDENTIFICATION

Relative density
About 0.925.
Refractive index About 1.475.
A. Identification of fatty oils by thin-layer chromatography (2.3.2).
Results The chromatogram obtained is similar to the corresponding chromatogram shown in Figure 2.3.2.-1.
B. Composition of fatty acids (see Tests).

CHARACTERS

Appearance

Clear, light yellow liquid.

Solubility

Practically insoluble in water and in ethanol (96 per cent), miscible with light petroleum (bp: 40-60 °C).

Relative density

About 0.925.

DEFINITION

Fatty oil obtained from the germ of the grain of *Triticum aestivum* L. by cold expression or by other suitable mechanical means and/or by extraction. It is then refined. A suitable antioxidant may be added.

CHARACTERS

A suitable antioxidant may be added.

Refined Wheat-germ Oil

(Ph. Eur. monograph 1379)

Ph Eur

STORAGE

In an airtight, well-filled container, protected from light.

Water (2.5.32)
Maximum 0.1 per cent, determined on 1.00 g.

Brassicasterol (2.4.23)
Maximum 0.3 per cent in the sterol fraction of the oil.
Composition of fatty acids (2.4.22, Method C)
Maximum 5.0 per cent, determined on 5.0 g.
Unsaponifiable matter (2.5.7)
Maximum 15.0.
Peroxide value (2.5.5, Method A)
Maximum 20.0.
Acid value (2.5.1)
Maximum 20.0.

TESTS

Composition of fatty acids (2.4.22, Method C)
Maximum 5.0 per cent, determined on 5.0 g.
Unsaponifiable matter (2.5.7)
Maximum 15.0.
Peroxide value (2.5.5, Method A)
Maximum 20.0.
Acid value (2.5.1)
Maximum 20.0.

Composition of the fatty-acid fraction of the oil:
— *palmitic acid*: 14.0 per cent to 19.0 per cent,
— *stearic acid*: maximum 2.0 per cent,
— *oleic acid*: 12.0 per cent to 23.0 per cent,
— *linoleic acid*: 52.0 per cent to 59.0 per cent,
— *linolenic acid*: 3.0 per cent to 10.0 per cent,
— *eicosenoic acid*: maximum 2.0 per cent.



Composition of fatty acids (2.4.22, Method C)

Use the mixture of calibrating substances in Table 2.4.22-3.

Composition of the fatty-acid fraction of the oil:

- *palmitic acid*: 14.0 per cent to 19.0 per cent;
- *stearic acid*: maximum 2.0 per cent;
- *oleic acid*: 12.0 per cent to 23.0 per cent;
- *linoleic acid*: 52.0 per cent to 59.0 per cent;
- *linolenic acid*: 3.0 per cent to 10.0 per cent;
- *eicosenoic acid*: maximum 2.0 per cent.

Brassicasterol (2.4.23)

Maximum 0.3 per cent in the sterol fraction of the oil.

Water (2.5.32)

Maximum 0.1 per cent, determined on 1.00 g.

STORAGE

In an airtight, well-filled container, protected from light.

LABELLING*The label states:*

- where applicable, that the substance is suitable for use in the manufacture of parenteral preparations;
- whether the oil is obtained by mechanical means, by extraction or by a combination of the 2.

Ph Eur

Wool Alcohols

Wool Wax Alcohols

(Ph. Eur. monograph 0593)

Preparation

Wool Alcohols Ointment

Ph Eur

DEFINITION

Mixture of sterols and higher aliphatic alcohols from wool

fat. A suitable antioxidant may be added.

Content

Minimum 30.0 per cent of cholesterol.

CHARACTERS**Appearance**

Pale-yellow or brownish-yellow, brittle mass becoming plastic on heating.

Solubility

Practically insoluble in water, soluble in boiling anhydrous ethanol and in methylene chloride, slightly soluble in ethanol (90 per cent V/V).

IDENTIFICATIONDissolve 50 mg in 5 mL of *methylene chloride* R and add1 mL of *acetic anhydride* R and 0.1 mL of *sulfuric acid* R.

Within a few seconds, a green colour develops.

TESTS**Appearance of solution**To 1.0 g add 10 mL of *light petroleum* R1 and shake while warming in a water-bath. The substance dissolves completely. After cooling, the solution is clear (2.2.1).**Alkalinity**Dissolve 2.0 g in 25 mL of hot ethanol (90 per cent V/V) R and add 0.5 mL of *phenolphthalein solution* R1. No red colour develops.**Melting point** (2.2.15)

Minimum 56 °C.

Melt the substance to be examined by heating in a water-bath at a temperature which exceeds the expected melting point by not more than 10 °C; introduce the substance to be examined into the capillary tubes and allow to stand on ice for at least 2 h.

Water-absorption capacity

Place 0.6 g of the substance to be examined and 9.4 g of *white soft paraffin* R in a mortar and melt on a water-bath. Allow to cool and incorporate 20 mL of *water* R, added in portions. Within 24 h no water is released from the almost white, ointment-like emulsion.

Acid value (2.5.1)

Maximum 2.0.

If necessary, heat in a water-bath under a reflux condenser to dissolve the substance to be examined.

Hydroxyl value (2.5.3, Method A)

120 to 180.

Peroxide value (2.5.5, Method A)

Maximum 15.

Take from the substance to be examined wedge-shaped pieces whose base consists of part of the surface. Melt the pieces before carrying out the determination. Before adding 0.5 mL of *saturated potassium iodide solution* R, cool the solution obtained to room temperature.

Saponification value (2.5.6)

Maximum 12.0, determined on 10.00 g. Heat under reflux for 4 h.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 2.000 g by drying in an oven at 105 °C.

Total ash (2.4.16)

Maximum 0.1 per cent.

ASSAY

Gas chromatography (2.2.28). Homogenise the sample before use.

Internal standard solution Dissolve 0.125 g of *pregnenolone isobutyrate* CRS in *heptane* R and dilute to 50.0 mL with the same solvent.

Test solution Dissolve 75.0 mg of the substance to be examined in 10.0 mL of the internal standard solution and dilute to 25.0 mL with *heptane* R.

Reference solution Dissolve 25.0 mg of *cholesterol* CRS in 10.0 mL of the internal standard solution and dilute to 25.0 mL with *heptane* R.

Injection liner:— *packing material*: quartz wool;— *size*: l = 78.5 mm, Ø = 4.0 mm.**Column:**— *material*: fused silica;— *size*: l = 30 m, Ø = 0.25 mm;— *stationary phase*: poly(dimethyl)siloxane R (film thickness 0.25 µm).*Carrier gas helium for chromatography* R.*Flow rate* 1 mL/min.*Split ratio* 1:50.*Temperature:*— *column*: 275 °C;— *injection port*: 285 °C;— *detector*: 300 °C.*Detection* Flame ionisation.

Injection 1 mL.

Relative retention With reference to pregnenolone isobutyrate (retention time = about 8 min): cholesterol = about 1.2.

System suitability: reference solution:

— **resolution:** minimum 5.0 between the peaks due to

pregnenolone isobutyrate and cholesterol.

Calculate the percentage content of cholesterol in the substance to be examined taking into account the assigned content of cholesterol CRS.

STORAGE

In a well-filled container, protected from light.

Ph Eur

Wool Fat

Anhydrous Lanolin

(Ph. Eur. monograph 0134)

Ph Eur

DEFINITION

Purified, anhydrous, waxy substance obtained from the wool of sheep (*Ovis aries*). A suitable antioxidant may be added.

CHARACTERS

Appearance

Yellow, unctuous substance. When melted, it is a clear or almost clear, yellow liquid. A solution in light petroleum is opalescent.

Solubility

Practically insoluble in water, slightly soluble in boiling

anhydrous ethanol.

Characteristic odour.

IDENTIFICATION

A. In a test-tube, dissolve 0.5 g in 5 mL of methylene chloride R and add 1 mL of acetic anhydride R and 0.1 mL of sulfuric acid R. A green colour develops.

B. Dissolve 50 mg in 5 mL of methylamine chloride R, add 5 mL of sulfuric acid R and shake. A red colour develops and an intense green fluorescence appears in the lower layer when examined in daylight, with illumination from behind the

observer.

TESTS

Water-soluble acid or alkaline substances

Melt 5.0 g on a water-bath and shake vigorously for 2 min with 75 mL of water R previously heated to 90–95 °C. Allow to cool and filter through filter paper previously rinsed with water R. To 60 mL of the filtrate (which may not be clear) add 0.25 mL of bromothymol blue solution R1. Not more than 0.2 mL of 0.02 M hydrochloric acid or 0.15 mL of 0.02 M sodium hydroxide is required to change the colour of the indicator.

Water-absorption capacity

Place 10 g of molten wool fat in a mortar and allow to cool to room temperature. Weigh the mortar. Add water R in portions of 0.2–0.5 mL from a burette, stirring vigorously after each addition to incorporate the water R. Instead of a pestle, use a high-density polypropylene cylindrical rod (120 mm long and 10 mm in diameter, for example). The end-point is reached when visible droplets remain which cannot be incorporated. Weigh the mortar again and determine the amount of water absorbed by weight difference. Not less than 20 mL of water R is absorbed.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g dissolved in 25 mL of the prescribed mixture of solvents.

Peroxide value (2.5.5, Method A)

Before adding 0.5 mL of saturated potassium iodide solution R, Maximum 20.

cool the solution obtained to room temperature.

Saponification value (2.5.6)

90 to 105, determined on 2.00 g while heating under reflux for 4 h.

Water-soluble oxidisable substances

To 10 mL of the filtrate obtained in the test for water-soluble acid or alkaline substances add 1 mL of dilute sulfuric acid R and 0.1 mL of 0.02 M potassium permanganate. After 10 min, the solution is not completely decolourised.

Paraffins

Maximum 1.0 per cent.

The tap and cotton plugs used must be free from grease. Prepare a column of anhydrous aluminium oxide 0.23 m long and 20 mm in diameter by adding a slurry of anhydrous

aluminium oxide R and light petroleum R1 to a glass tube fitted

with a tap and containing light petroleum R1 (before use,

dehydrate the anhydrous aluminium oxide by heating it in an

oven at 600 °C for 3 h). Allow to settle and reduce the depth

of the layer of solvent above the column to about 40 mm.

Dissolve 3.0 g of the substance to be examined in 50 mL of

warm light petroleum R1, cool, pass the solution through the

column at a flow rate of 3 mL/min and wash with 250 mL of

light petroleum R1. Concentrate the combined eluate and

washings to low bulk by distillation, evaporate to dryness on

a water-bath and heat the residue at 105 °C for periods of

10 min until 2 successive weighings do not differ by more

than 1 mg. The residue weighs a maximum of 30 mg.

Pesticide residues

Maximum 0.05 ppm for each organochlorine pesticide,

0.5 ppm for each other pesticide and 1 ppm for the sum of

all the pesticides.

All glassware used is thoroughly washed using phosphate-free

detergent as follows. The glassware is immersed in a bath of

detergent solution (5 per cent in deionised water) and allowed to

soak for 24 h. The detergent is washed off with copious amounts

of acetone and hexane for pesticide analysis. It is important to

keep glassware specifically for pesticide analyses, it must not be

mixed up with glassware used for other applications. The glassware

used must be free of chlorinated solvents, plastics and rubber

materials, in particular phthalate plasticisers, oxygenated

compounds and nitrogenated solvents such as acetonitrile.

Use hexane, toluene and acetone for pesticide analysis. Use HPLC

grade reagents for ethyl acetate, cyclohexane and water.

The test consists of the isolation of pesticide residues by size-

exclusion chromatography (2.2.30) followed by solid phase

extraction and identification by gas chromatography coupled

with an electron capture detector or a thermionic detector.

Isolation of the pesticide residues

As detector, use a UV-visible spectrophotometer set at a

wavelength of 254 nm to calibrate the chromatographic

column for gel permeation.

Calibration is extremely important in gel permeation

chromatography (GPC) to check that the pressure, solvent

flow rate, solvent ratio, temperature and column conditions

remain constant. The gel permeation column is to be

calibrated at regular intervals using a standard mixture

prepared as follows: into a 1000 mL volumetric flask,

introduce 50.00 g of maize oil R, 0.20 g of methoxychlor R

magnesium silicate is suitable for use for up to 1 week. It is essential that only deactivated magnesium silicate is used. Take a 6 mL empty solid phase extraction cartridge and weigh into the cartridge 1 g of the deactivated magnesium silicate.

At this stage the GPC fraction still contains about 10 per cent of the substance to be examined, so further clean-up is necessary. A separate isolation procedure is carried out a) for organochlorine and synthetic pyrethroid pesticides and b) for organophosphorus pesticides. Place a preconditioned solid phase extraction cartridge containing 1 g of deactivated *magnesium silicate for pesticide residue analysis R* onto a vacuum manifold.

Condition the cartridge by adding 10 mL of *toluene R* and allowing the solvent to elute through the cartridge. Place the 0.5 mL of the solvent fraction from the evaporating vessel on the preconditioned cartridge. Elute the pesticide fractions from the cartridge using 20 mL of either of the 2 different solvent systems shown below:

a) for determination of the organochlorine and synthetic pyrethroid pesticides: *toluene R*; a very small amount of the substance to be examined is co-eluted;

b) for determination of the organophosphorus pesticides: a mixture of 2 volumes of *acetone R* and 98 volumes of *toluene R*; this solvent system is used to elute all the pesticides including the more polar organophosphorus pesticides;

unfortunately, some of the substance to be examined is co-eluted with this solvent system, which can interfere with the electron capture detector.

Collect the eluate from the extraction cartridges in 25 mL glass vials. Quantitatively transfer the eluate to an evaporating vessel, washing the vial with 3 quantities, each of 10 mL, of *hexane R*.

Place the evaporating vessel on the autoevaporator and evaporate the solid phase extraction fractions down to 0.5 mL. The water-bath temperature is set at 45 °C and the nitrogen pressure is 55 kPa.

Examine the residues by gas chromatography (2.2.28) using electron capture and thermionic detectors as described below.

Recovery Calculate the recovery correction factor (R_{cf}) of the internal standards (*atlimphos R* or *isodm R*) added to the test solution using the following expression:

$$\frac{A_2}{A_1} \times 100$$

A_1 = peak area of an internal standard 1 ppm in solution;
 A_2 = peak area of internal standard extracted from the test solution.

5 mL of the 20 mL test solution containing 1 mL of 2 ppm internal standard concentrated to 0.5 mL is equivalent to 1 ppm of the internal standard in the solution.

If the recovery of the internal standards falls outside the range of 70 per cent to 110 per cent the test is not valid. **Reference solutions** Prepare reference solutions of pesticides using the pesticides standards at a concentration of 0.5 ppm (see composition of reference solutions A to D in Table 0134.-1). Commercially available pesticides may be purchased. The individual standards have a concentration of 10 ppm.

At the same time prepare solutions of pesticides equivalent to the limit of detection of the method (see recommended

and 50.0 mg of *perylene R*. Dilute to 1000.0 mL with a

mixture of equal volumes of *cyclohexane R* and *ethyl acetate R*. To calibrate the column, set the mobile phase at a flow rate of 5 mL/min with a mixture of equal volumes of

cyclohexane R and *ethyl acetate R*. Inject 5 mL of the standard mixture and record the resulting chromatogram. The retention times for the analytes must not vary by more than ± 5 per cent between calibrations. If the retention time shift is greater than ± 5 per cent, take corrective action.

Excessive retention time shifts may be caused by: — poor laboratory temperature control; — the pump containing air; this can be verified by measuring the flow rate: collect 25 mL of column eluate in a volumetric flask and record the time (300 ± 5 s); — a leak in the system.

Changes in pressure, in mobile phase flow rate or in column temperature conditions, as well as column contamination, can affect pesticide retention times and are to be monitored. If the flow rate or column pressure are outside desired bands the precolumn or column is to be replaced.

Test solution In a volumetric flask, dissolve 1 g of the substance to be examined, accurately weighed, in a mixture

of 1 volume of *ethyl acetate R* and 7 volumes of *cyclohexane R*. Add 1 mL of an internal standard (2 ppm, either *isodm R* or *atlimphos R*) and dilute to 20 mL. The internal standard

solutions are used to establish that recoveries of the pesticides from the GPC purification stage, evaporation and solid phase extraction stage are at acceptable levels. Recovery levels of the internal standard solutions from the wool fat are determined by comparing the peak areas of the wool fat extracts with peak areas of solutions of the internal standards.

Precolumn: — size: $l = 0.075$ m, $\varnothing = 21.2$ mm; — stationary phase: styrene-divinylbenzene copolymer *R* (5 μm).

Gel permeation column: — size: $l = 0.3$ m, $\varnothing = 21.2$ mm; — stationary phase: styrene-divinylbenzene copolymer *R* (5 μm).

Mobile phase *ethyl acetate R*, *cyclohexane R* (10:70 V/V). Flow rate 5 mL/min.

Detection Spectrophotometer at 254 nm. Inject 5 mL of the test solution. Discard the first 95 mL (19 min) of eluate containing the substance to be examined. Collect the next 155 mL of eluate (31 min) containing any pesticide residues in an evaporating vessel.

Place the 155 mL of the eluate collected from the gel permeation chromatography column into an evaporating vessel. Place this vessel in an autoevaporator setting the water-bath temperature at 45 °C and the nitrogen pressure at 55 kPa. Evaporate the eluate down to 0.5 mL.

To prepare the solid phase extraction cartridges take some *magnesium silicate for pesticide residue analysis R* and heat it in a muffle furnace at 700 °C for 4 h to remove moisture and polychlorinated biphenyls. Subsequently allow the

magnesium silicate to cool for 2 h and transfer it directly to an oven at 100-105 °C, and allow to stand for 30 min. Transfer the magnesium silicate to a stoppered glass jar and allow to equilibrate for 48 h. This material may be used for up to 2 weeks. After that period the magnesium silicate is to be reactivated, by heating at 600 °C for 2 h in a muffle

furnace. Remove the magnesium silicate from the furnace, cool and store in a stoppered glass jar. The magnesium silicate is deactivated by adding 1 per cent of *water R*. After the water has been added, shake the magnesium silicate

intermittently over 15 min just prior to use. The deactivated

compositions in Table 0134-1). These reference solutions are used to optimise the electron capture detector and thermionic detector to achieve the detection limits of the

package on a computer. The interpretation of pesticides in trace residue analyses is extremely complex. The detectors, particularly the electron capture detector, are prone to interference, both from the substance to be examined itself, and from solvents, reagents and apparatus used in the extraction. These peaks can easily be misinterpreted or quoted as a false positive. Confirmation of pesticides can be achieved by running samples and standards on different capillary columns (see chromatographic systems A or B described below). The peaks can be identified by using the

To identify any pesticide residues compare the chromatograms obtained with reference solutions A to D. The identity of the pesticides can be confirmed by spiking samples or overlaying chromatograms using an integration

content of each pesticide using the following expression:

$$C_p = \frac{P_p \times D \times C_e}{100} \times \frac{R_e}{R_{cf}}$$

C_p = concentration of identified pesticide (ppm);
 P_p = peak area of the individual pesticide in the test sample obtained;

C_e = concentration of the individual pesticide in the external standard (ppm);

P_e = peak area of the individual pesticide in the external standard;

D = dilution factor;

R_{cf} = recovery correction factor.

The dilution factor (D) can be defined as follows:

$$\frac{V_1}{V_2} \times \frac{m \times V_3}{V_2}$$

V_1 = volume of sample obtained after the 2nd evaporation stage;

m = sample weight;

V_2 = GPC injection volume;

V_3 = sample volumetric flask volume.

Chromatographic system A:

Precolumn:

— material: deactivated silica;

— size: $l = 4.5$ m, $\phi = 0.53$ mm.

Column:

— material: fused silica;

— size: $l = 60$ m, $\phi = 0.25$ mm;

— stationary phase: poly(dimethyl) (diphenyl) siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Linear velocity 25 cm/s.

Pressure 180 kPa.

Temperature:

Time (min)	Temperature (°C)
0 - 1	75
1 - 5	75 \rightarrow 175
5 - 30	175 \rightarrow 275
30 - 40	275 \rightarrow 285
40 - 55	285
Injection port	300
Detector	350

Detection Electron capture or thermionic specific detector.

Injection 2 μ L.

Chromatographic system B Which may be used for confirmation analysis:

Precolumn:

— material: deactivated silica;

— size: $l = 4.5$ m, $\phi = 0.53$ mm.

Column:

— material: fused silica;

— size: $l = 60$ m, $\phi = 0.25$ mm;

— stationary phase: poly(cyanopropyl) (7) (phenyl) (86) siloxane R (film thickness 0.25 μ m).

Carrier gas helium for chromatography R.

Linear velocity 25 cm/s.
 Pressure 180 kPa.
 Temperature:

Time (min)	Temperature (°C)
0 - 1	75
1 - 5	75 \rightarrow 175
5 - 30	175 \rightarrow 275
30 - 40	275 \rightarrow 285
40 - 55	285
Injection port	300
Detector	350

Detection Electron capture or thermionic specific detector.
 Injection 2 μ L.

Chlorides

Maximum 150 ppm.

Boil 1.0 g with 20 mL of ethanol (90 per cent V/V) R in a round-bottomed flask fitted with a reflux condenser for 5 min. Cool, add 40 mL of water R and 0.5 mL of nitric acid R and filter. To the filtrate add 0.15 mL of a 10 g/L solution of silver nitrate R in ethanol (90 per cent V/V) R. Allow to stand for 5 min protected from light.

Any opalescence in the solution is not more intense than that in a standard prepared at the same time by adding 0.15 mL of a 10 g/L solution of silver nitrate R in ethanol

(90 per cent V/V) R to a mixture of 0.2 mL of 0.02 M hydrochloric acid, 20 mL of ethanol (90 per cent V/V) R, 40 mL of water R and 0.5 mL of nitric acid R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C for 1 h.

Sulfated ash (2.4.14)

Maximum 0.15 per cent.

Ignite 5.0 g and use the residue to determine the sulfated ash.

STORAGE

At a temperature not exceeding 25 °C.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are

functions of the substance when used as an excipient (see chapter 5.15). Some of the characteristics described in the Functionality-related characteristics section may also be present in the mandatory part of the monograph since they also represent mandatory quality criteria. In such cases, a cross-reference to the tests described in the mandatory part is included in the Functionality-related characteristics section. Control of the characteristics can contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for wool fat used in water-emulsifying ointments and lipophilic creams.

Water-absorption capacity

(see Tests).

Drop point (2.2.17, Method A)
To fill the metal cup, melt the wool fat on a water-bath, cool to about 50 °C, pour into the cup and allow to stand at 15-20 °C for 24 h. The drop point is typically 38 °C to 44 °C.

Ph Eur

Hydrogenated Wool Fat

(Ph. Eur. monograph 0969)

Ph Eur

DEFINITION
Mixture of higher aliphatic alcohols and sterols obtained from the direct, high-pressure, high-temperature hydrogenation of wool fat (0134) during which the esters and acids present are reduced to the corresponding alcohols. A suitable antioxidant may be added.

CHARACTERS

Appearance
White or pale yellow, unctuous substance.

Solubility

Practically insoluble in water, soluble in boiling anhydrous ethanol and in light petroleum.

IDENTIFICATION

First identification B

Second identification A, C

A. Melting point (see Tests).

B. Examine the chromatograms obtained in the test for fatty alcohols and sterols.
Results The principal peaks in the chromatogram obtained with the test solution are similar in retention time and size to the principal peaks in the chromatogram obtained with reference solution (a).
C. Dissolve 50 mg in 5 mL of methylene chloride R and add 1 mL of acetic anhydride R and 0.1 mL of sulfuric acid R. A green colour is produced.

TESTS

Melting point (2.2.15)

45 °C to 55 °C. Allow to stand at 20 °C for 16 h.

Acid value (2.5.1)

Maximum 1.0, determined on 5.0 g.

Hydroxyl value (2.5.3, Method A)

140 to 180.

Saponification value (2.5.6)

Maximum 8.0. Heat under reflux for 4 h.

Fatty alcohols and sterols

Gas chromatography (2.2.28).

Test solution Dissolve 0.25 g of the substance to be examined in 60 mL of anhydrous ethanol R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dissolve 0.25 g of hydrogenated wool fat CRS in 60 mL of anhydrous ethanol R and dilute to 100.0 mL with the same solvent.

Reference solution (b) Dissolve 50 mg of cetyl alcohol CRS and 50 mg of stearyl alcohol CRS in 60 mL of anhydrous ethanol R and dilute to 100.0 mL with the same solvent.

and dilute to 100.0 mL with the same solvent.

50 mg of stearyl alcohol CRS in 60 mL of anhydrous ethanol R and dilute to 100.0 mL with the same solvent.

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50 mg of stearyl alcohol CRS in 60 mL of anhydrous ethanol R and dilute to 100.0 mL with the same solvent.

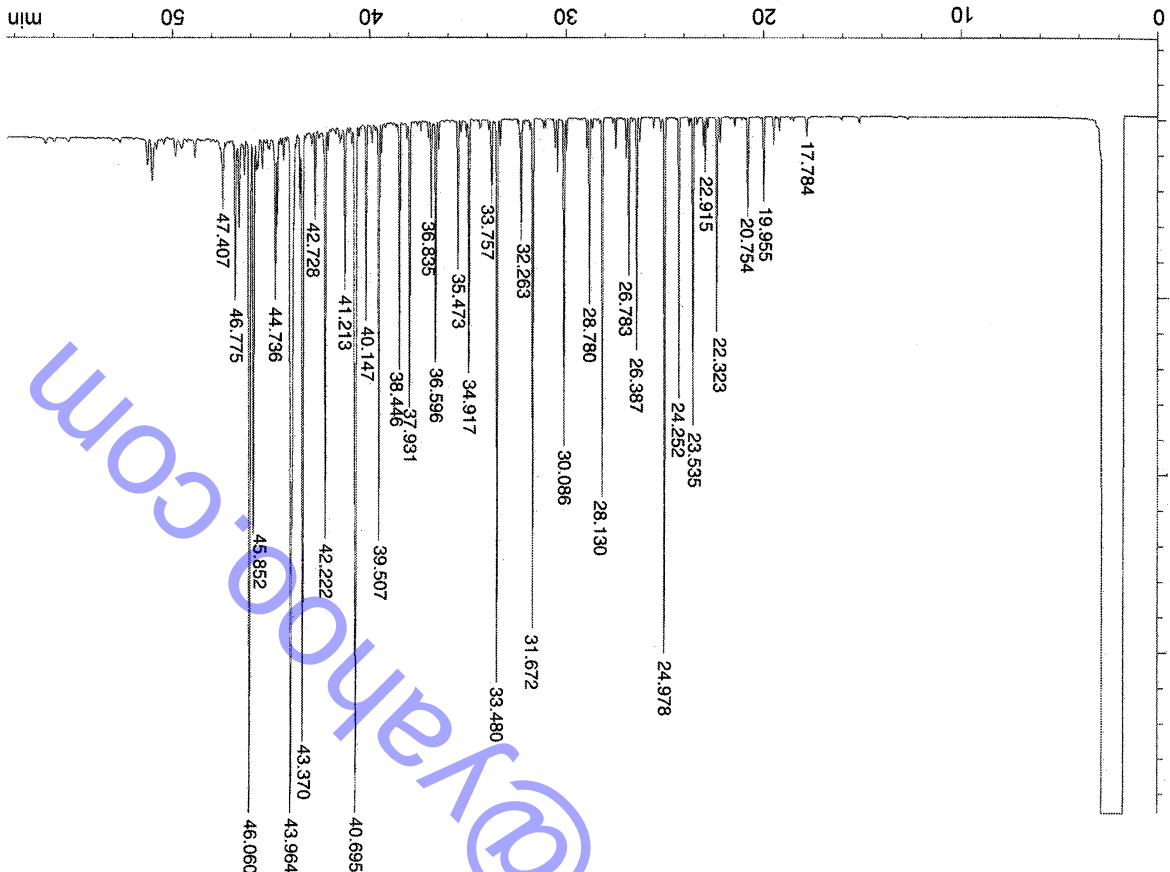


Figure 0969.-1. - Chromatogram for the test for fatty alcohols and sterols in hydrogenated wool fat: reference solution (a)

intense green fluorescence appears in the lower layer when examined in daylight, with illumination from behind the

observer.

TESTS

Water-soluble acid or alkaline substances

Melt 6.7 g on a water-bath and shake vigorously for 2 min

with 75 mL of water R previously heated to 90-95 °C. Allow

to cool and filter through filter paper previously rinsed with

water R. To 60 mL of the filtrate (which may not be clear)

add 0.25 mL of bromothymol blue solution R1. Not more than

0.2 mL of 0.02 M hydrochloric acid or 0.15 mL of 0.02 M

sodium hydroxide is required to change the colour of the

indicator.

Drop point (2.2.17)

38 °C to 44 °C.

To fill the metal cup, melt the residue obtained in the test

for wool-fat content on a water-bath, cool to about 50 °C,

pour into the cup and allow to stand at 15-20 °C for 24 h.

Water-absorption capacity

Place 10 g of the residue obtained in the test for wool-fat

content in a mortar. Add water R in portions of 0.2-0.5 mL

from a burette, stirring vigorously after each addition to

incorporate the water R. The end-point is reached when

visible droplets remain which cannot be incorporated.

Not less than 20 mL of water R is absorbed.

Acid value (2.5.1)

Maximum 0.8, determined on 5.0 g dissolved in 25 mL of

the prescribed mixture of solvents.

Peroxide value (2.5.5, Method A)

Maximum 15.

Saponification value (2.5.6)

67 to 79, determined on 2.00 g while heating under reflux

for 4 h.

Water-soluble oxidisable substances

To 10 mL of the filtrate obtained in the test for water-

soluble acid or alkaline substances add 1 mL of dilute sulfu-

ric acid R and 0.1 mL of 0.02 M potassium permanganate. After

10 min, the solution is not completely decolourised.

Paraffins

Maximum 1.0 per cent.

The tap and cotton plugs used must be free from grease Prepare a

column of anhydrous aluminium oxide 230 mm long and

20 mm in diameter by adding a slurry of anhydrous

aluminium oxide R and light petroleum R1 to a glass tube fitted

with a tap and containing light petroleum R1. Allow to settle

and reduce the depth of the layer of solvent above the

column to about 40 mm. Dissolve 3.0 g of the residue

obtained in the test for wool-fat content in 50 mL of warm

light petroleum R1, cool, pass the solution through the column

at a rate of 3 mL/min and wash with 250 mL of light

petroleum R1. Concentrate the combined eluate and washings

to low bulk by distillation, evaporate to dryness on a water-

bath and heat the residue at 105 °C for periods of 10 min

until 2 successive weighings do not differ by more than 1 mg.

The residue weighs a maximum of 30 mg.

Chlorides

Maximum 115 ppm.

Boil 1.3 g with 20 mL of ethanol (90 per cent V/V) R under a

reflux condenser for 5 min. Cool, add 40 mL of water R and

0.5 mL of nitric acid R and filter. To the filtrate add 0.15 mL

of a 10 g/L solution of silver nitrate R in ethanol

(90 per cent V/V) R. Allow to stand for 5 min, protected from

light. Any opalescence in the solution is not more intense

Hydrous Wool Fat

Laolin

(Ph. Eur. monograph 0135)

Ph Eur

DEFINITION

Mixture of 75 per cent m/m of wool fat and 25 per cent m/m

of water. It is obtained by the gradual addition of water to

melted wool fat with continuous stirring. A suitable

antioxidant may be added.

CHARACTERS

Appearance

Pale yellow, unctuous substance.

IDENTIFICATION

A. In a test-tube, dissolve 0.5 g in 5 mL of methylene

chloride R and add 1 mL of acetic anhydride R and 0.1 mL of

sulfuric acid R. A green colour develops.

B. Dissolve 50 mg in 5 mL of methylene chloride R, add 5 mL

of sulfuric acid R and shake. A red colour develops and an

Ph Eur



In a well-filled container, protected from light.

STORAGE

Maximum 0.1 per cent, determined on 5.0 g.

Total ash (2.4.16)

an oven at 105 °C for 1 h.

Maximum 3.0 per cent, determined on 2.000 g by drying in

Loss on drying (2.2.32)

using 2 mL of lead standard solution (10 ppm Pb) R.

2.0 g complies with test C. Prepare the reference solution

Maximum 10 ppm.

Heavy metals (2.4.8)

obtained with reference solution (b).

cetyl alcohol and stearyl alcohol present in the chromatogram

show enhanced peaks with retention times corresponding to

with reference solution (a) (Figure 0969-1) and it does not

does not differ significantly from the chromatogram obtained

Results The chromatogram obtained with the test solution

Injection 1 µL.

Detection Flame ionisation.

Column	Time (min)	Temperature (°C)
100	0 - 5	100
100 → 300	5 - 45	100 → 300
300	45 - 60	300
325		325
Detector		350

Temperature:

Flow rate 1 mL/min.

Carrier gas helium for chromatography R.

polar phase (film thickness 0.25 µm).

— stationary phase: poly(dimethyl)siloxane R or another non-

— size: 1 = 30 m, Ø = 0.25 mm;

— material: fused silica;

Column:

than that in a standard prepared at the same time by adding 0.15 mL of a 10 g/L solution of silver nitrate R in ethanol (90 per cent V/V) R to a mixture of 0.2 mL of 0.02 M hydrochloric acid, 20 mL of ethanol (90 per cent V/V) R, 40 mL of water R and 0.5 mL of nitric acid R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent.

Ignite 5.0 g and use the residue.

Wool-fat content

72.5 per cent to 77.5 per cent.

In a suitable tared dish containing a glass rod, heat 30.0 g to constant mass on a water-bath, stirring continuously. Weigh the residue.

STORAGE

At a temperature not exceeding 25 °C.

Xanthan Gum

(Ph. Eur. monograph 1277)

11138-66-2

Action and use

Excipient.

Ph Eur

DEFINITION

High-molecular-mass anionic polysaccharide produced by fermentation of carbohydrates with *Xanthomonas campestris*. It consists of a principal chain of $\beta(1 \rightarrow 4)$ -linked D-glucose units with trisaccharide side chains, on alternating anhydroglucose units, consisting of 1 glucuronic acid unit included between 2 mannose units. Most of the terminal units contain a pyruvate moiety and the mannose unit adjacent to the principal chain may be acetylated at C-6. Xanthan gum has a relative molecular mass of approximately 1×10^6 . It exists as the sodium, potassium or calcium salt.

Content

Minimum 1.5 per cent of pyruvoyl groups ($C_3H_3O_2$;

CHARACTERS

Appearance

White or yellowish-white, free-flowing powder.

Solubility

Soluble in water giving a highly viscous solution, practically insoluble in organic solvents.

IDENTIFICATION

A. In a flask, suspend 1 g in 15 mL of 0.1 M hydrochloric acid. Close the flask with a fermentation bulb containing barium hydroxide solution R and heat carefully for 5 min. The barium hydroxide solution shows a white turbidity. B. To 300 mL of water R, previously heated to 80 °C and stirred rapidly with a mechanical stirrer in a 400 mL beaker, add, at the point of maximum agitation, a dry blend of 1.5 g of carob bean gum R and 1.5 g of the substance to be examined. Stir until the mixture forms a solution, and then continue stirring for 30 min or longer. Do not allow the water temperature to drop below 60 °C during stirring. Discontinue stirring and allow the mixture to stand for at least 2 h. A firm rubbery gel forms after the temperature

TESTS

drops below 40 °C but no such gel forms in a 1 per cent control solution of the sample prepared in the same manner but omitting the carob bean gum.

pH (2.2.3)

6.0 to 8.0 for a 10.0 g/L solution.

2-Propanol

Gas chromatography (2.2.28).

Internal standard solution Dilute 0.50 g of 2-methyl-2-propanol R to 500 mL with water R.

Test solution To 200 mL of water R in a 1000 mL round-bottomed flask, add 5.0 g of the substance to be examined and 1 mL of a 10 g/L emulsion of dimethicone R in liquid paraffin R, stopper the flask and shake for 1 h. Distil about 90.0 mL, mix the distillate with 4.0 mL of the internal standard solution and dilute to 100.0 mL with water R.

Reference solution Dilute a suitable quantity of 2-propanol R, accurately weighed, with water R to obtain a solution having a known concentration of 2-propanol of about 1 mg/mL. To 4.0 mL of this solution add 4.0 mL of the internal standard solution and dilute to 100.0 mL with water R.

Column:

— size: $l = 1.8$ m, $\varnothing = 4.0$ mm;

— stationary phase: styrene-divinylbenzene copolymer R (149-177 μ m).

Carrier gas helium for chromatography R.

Flow rate 30 mL/min.

Temperature:

— column: 165 °C;

— injection port and detector: 200 °C.

Detection Flame ionisation.

Injection 5 μ L.

Relative retention With reference to 2-propanol: 2-methyl-2-propanol = about 1.5.

Limit:

— 2-propanol: maximum 750 ppm.

Other polysaccharides

Thin-layer chromatography (2.2.27).

Test solution To 10 mg of the substance to be examined in a thick-walled centrifuge test tube add 2 mL of a 230 g/L solution of trifluoroacetic acid R, shake vigorously to dissolve the forming gel, stopper the test tube, and heat the mixture at 120 °C for 1 h. Centrifuge the hydrolysate, transfer the clear supernatant carefully into a 50 mL flask, add 10 mL of water R and evaporate the solution to dryness under reduced pressure. Take up the residue thus obtained in 10 mL of water R and evaporate to dryness under reduced pressure. Wash 3 times with 20 mL of methanol R and evaporate under reduced pressure. To the resulting clear film which has no odour of acetic acid, add 0.1 mL of water R and 1 mL of methanol R. Centrifuge to separate the amorphous precipitate. Dilute the supernatant, if necessary, to 1 mL with methanol R.

Reference solution Dissolve 10 mg of glucose R and 10 mg of mannose R in 2 mL of water R and dilute to 10 mL with methanol R.

Plate TLC silica gel plate R.

Mobile phase 16 g/L solution of sodium dihydrogen phosphate R, butanol R, acetone R (10:40:50 V/V/V).

Application 5 μ L as bands.

Development Over a path of 15 cm.

Detection Spray with a solution of 0.5 g of diphenylamine R in 25 mL of methanol R to which 0.5 mL of aniline R and 2.5 mL of phosphoric acid R have been added. Heat for 5 min at 120 °C and examine in daylight.

System suitability: reference solution: — the chromatogram shows 2 clearly separated greyish-brown zones due to glucose and mannose in the middle third.

Results The chromatogram obtained with the test solution shows 2 zones corresponding to the zones due to glucose and mannose in the chromatogram obtained with the reference solution. In addition, 1 weak reddish and 2 faint bluish-grey bands may be visible just above the line of application. 1 or 2 bluish-grey bands may also be seen in the upper quarter of the chromatogram. No other bands are visible.

Loss on drying (2.2.32) Maximum 15.0 per cent, determined on 1.000 g by drying in an oven at 105 °C for 2.5 h.

Total ash (2.4.16) 6.5 per cent to 16.0 per cent.

Microbial contamination TAMC: acceptance criterion 10^3 CFU/g (2.6.12). TYMC: acceptance criterion 10^2 CFU/g (2.6.12).

ASSAY **Test solution** Dissolve a quantity of the substance to be examined corresponding to 120.0 mg of the dried substance in water R and dilute to 20.0 mL with the same solvent.

Reference solution Dissolve 45.0 mg of pyruvic acid R in water R and dilute to 500.0 mL with the same solvent. Place 10.0 mL of the test solution in a 50 mL round-bottomed flask, add 20.0 mL of 0.1 M hydrochloric acid and weigh. Boil on a water-bath under a reflux condenser for 3 h. Weigh and adjust to the initial mass with water R. In a separating funnel mix 2.0 mL of the solution with 1.0 mL of dimethylphenylhydrazine-hydrochloric solution R. Allow to stand for 5 min and add 5.0 mL of ethyl acetate R. Shake and allow the solids to settle. Collect the upper layer and shake with 3 quantities, each of 5.0 mL, of sodium carbonate solution R. Combine the aqueous layers and dilute to 50.0 mL with sodium carbonate solution R. Mix. Treat 10.0 mL of the reference solution at the same time and in the same manner as for the test solution.

Immediately measure the absorbance (2.2.25) of the 2 solutions at 375 nm, using sodium carbonate solution R as the compensation liquid. The absorbance of the test solution is not less than that of the reference solution, which corresponds to a content of pyruvoyl groups of not less than 1.5 per cent.

FUNCTIONALITY-RELATED CHARACTERISTICS

This section provides information on characteristics that are recognised as being relevant control parameters for one or more functions of the substance when used as an excipient (see chapter 5.15). This section is a non-mandatory part of the monograph and it is not necessary to verify the characteristics to demonstrate compliance. Control of these characteristics can however contribute to the quality of a medicinal product by improving the consistency of the manufacturing process and the performance of the medicinal product during use. Where control methods are cited, they are recognised as being suitable for the purpose, but other methods can also be used. Wherever results for a particular characteristic are reported, the control method must be indicated.

The following characteristics may be relevant for xanthan gum used as viscosity-increasing agent.

IDENTIFICATION

First identification B.
Second identification A, C.
A. Melting point (2.2.14): 92 °C to 96 °C.
B. Infrared absorption spectrophotometry (2.2.24).
Preparation Mulls in liquid paraffin R.
C. Thin-layer chromatography (2.2.27).
Test solution Dissolve 25 mg of the substance to be examined in water R and dilute to 10 mL with the same solvent.

CHARACTERS

Appearance White or almost white, crystalline powder or crystals.
Solubility Very soluble in water, sparingly soluble in ethanol (96 per cent).

DEFINITION

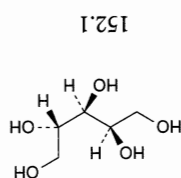
Meso-xylitol.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

Action and use

Sweetening agent.



(Ph. Eur. monograph 1381)

Xylitol



Ph Eur

Powder flow (2.9.36).

Particle-size distribution (2.9.31) or 2.9.38).

See test above.

Apparent viscosity

The following characteristics may be relevant for xanthan gum used as matrix former in prolonged-release tablets

Apparent viscosity The following characteristics may be relevant for xanthan gum used as matrix former in prolonged-release tablets

Stir the preparation at 800 r/min for 2 h whilst maintaining the temperature at 24 ± 1 °C. Determine the viscosity within 15 min at 24 ± 1 °C using a rotating viscometer set at 60 r/min and equipped with a rotating spindle 1.6 mm high and 12.7 mm in diameter which is attached to a shaft cylinder to the lower tip of the shaft should be 25.4 mm and the immersion depth 50.0 mm.

The following characteristics may be relevant for xanthan gum used as matrix former in prolonged-release tablets

low-pitch propeller-type stirrer rotating at 800 r/min. When of potassium chloride R in a 500 mL beaker stirring with a adding the substance take care that agglomerates are destroyed. Add an additional quantity of 44 mL of water R, to rinse any adhering residue from the walls of the beaker. Stir the preparation at 800 r/min for 2 h whilst maintaining the temperature at 24 ± 1 °C. Determine the viscosity within 15 min at 24 ± 1 °C using a rotating viscometer set at 60 r/min and equipped with a rotating spindle 1.6 mm high and 12.7 mm in diameter which is attached to a shaft cylinder to the lower tip of the shaft should be 25.4 mm and the immersion depth 50.0 mm.

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Reference solution (a) Dissolve 25 mg of xylitol CRS in water R and dilute to 10 mL with the same solvent.

Reference solution (b) Dissolve 25 mg of mannitol R and 25 mg of xylitol R in water R and dilute to 10 mL with the same solvent.

Plate TLC silica gel plate R.

Mobile phase water R, ethyl acetate R, propanol R

(10:20:70 V/V/V).

Application 2 µL.

Development Over 3/4 of the plate.

Drying In air.

Detection Spray with 4-aminobenzoic acid solution R, dry in a current of cold air until the acetone is removed, then heat at 100 °C for 15 min; allow to cool, spray with a 2 g/L solution of sodium periodate R, dry in a current of cold air, then heat at 100 °C for 15 min.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension IV (2.2.1) and not more intensely coloured than reference solution BY₇ (2.2.2, Method II).

Dissolve 2.5 g in water R and dilute to 50.0 mL with the same solvent.

Conductivity (2.2.38)

Maximum 20 µS·cm⁻¹.

Dissolve 20.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100.0 mL with the same solvent. Measure the conductivity of the solution while gently stirring with a magnetic stirrer.

Reducing sugars

Maximum 0.2 per cent, calculated as glucose equivalent.

Dissolve 5.0 g in 6 mL of water R with the aid of gentle heat. Cool and add 20 mL of cupri-citric solution R and a few glass beads. Heat so that boiling begins after 4 min and maintain boiling for 3 min. Cool rapidly and add 100 mL of a 2.4 per cent V/V solution of glacial acetic acid R and 20.0 mL of 0.025 M iodine. With continuous shaking, add 25 mL of a mixture of 6 volumes of hydrochloric acid R and 94 volumes of water R and, when the precipitate has dissolved, titrate the excess of iodine with 0.05 M sodium thiosulfate using 1 mL of starch solution R, added towards the end of the titration, as indicator. Not less than 12.8 mL of 0.05 M sodium thiosulfate is required.

Related substances

Gas chromatography (2.2.28).

Internal standard solution Dissolve 5 mg of erythritol R in water R and dilute to 25.0 mL with the same solvent.

Test solution (a) Dissolve 5.000 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.

Test solution (b) Dilute 1.0 mL of test solution (a) to 10.0 mL with water R.

Reference solution (a) Dissolve 5.0 mg each of L-arabinitol CRS (impurity A), galactitol CRS (impurity B), mannitol CRS (impurity C) and sorbitol CRS (impurity D) in water R and dilute to 20.0 mL with the same solvent.

Reference solution (b) Dissolve 50.0 mg of xylitol CRS in water R and dilute to 10.0 mL with the same solvent.

Pipette 1.0 mL of test solutions (a) and (b) and reference solutions (a) and (b) into 4 separate 100 mL round-bottomed flasks. Add 1.0 mL of the internal standard solution to each of the flasks containing test solution (a) or reference solution (a), and 5.0 mL of the internal standard solution to each of the flasks containing test solution (b) or reference solution (b). Evaporate each mixture to dryness in a water-bath at 60 °C with the aid of a rotary evaporator.

Dissolve each dry residue in 1 mL of anhydrous pyridine R, add 1 mL of acetic anhydride R to each flask and boil each solution under reflux for 1 h to complete acetylation.

Column:

— size: l = 30 m, Ø = 0.25 mm;

— stationary phase:

poly(cyanopropylphenyl) (14) (methyl) (86) siloxane R (0.25 µm).

Carrier gas nitrogen R.

Flow rate 1 mL/min.

Split ratio 1:50 to 1:100.

Temperature:

Time (min)

Temperature (°C)

Detection Flame-ionisation.

Injection 1 µL of test solution (a) and reference solution (a) (solutions obtained after derivatisation).

Relative retention With reference to xylitol (retention time = about 1.5 min); internal standard = about 0.6; impurity A = about 0.9; impurity C = about 1.4; impurity B = about 1.45; impurity D = about 1.5.

System suitability: reference solution (a):

— resolution: minimum 2.0 between the peaks due to impurities B and D.

Calculate the percentage content of each related substance in the substance to be examined using the following expression:

$$100 \times \frac{m_s}{m_n} \times \frac{R_n}{R_s}$$

m_s = mass of the particular component in 1 mL of reference solution (a), in milligrams;

m_n = mass of the substance to be examined in 1 mL of test solution (a), in milligrams;

R_s = ratio of the area of the peak due to the particular derivatised component to the area of the peak due to the derivatised internal standard in the chromatogram obtained with reference solution (a);

R_n = ratio of the area of the peak due to the particular derivatised component to the area of the peak due to the derivatised internal standard in the chromatogram obtained with test solution (a).

The sum of the percentage contents of the related substances in the chromatogram obtained with test solution (a) is not greater than 2.0 per cent. Disregard any peak with an area

less.
corresponding to a percentage content of 0.05 per cent or

Lead (2.4.10)

Maximum 0.5 ppm.

Dissolve the substance to be examined in 150.0 mL of the

prescribed mixture of solvents.

Nickel (2.4.15)

Maximum 1 ppm.

Dissolve the substance to be examined in 150.0 mL of the

prescribed mixture of solvents.

Water (2.5.12)

Maximum 1.0 per cent, determined on 1.00 g.

Bacterial endotoxins (2.6.14)

Less than 4 IU/g if the concentration is less than 100 g/L of

xylitol and less than 2.5 IU/g if the concentration is 100 g/L

or more of xylitol, when intended for use in the manufacture of parenteral preparations without a further appropriate

procedure for the removal of bacterial endotoxins.

ASSAY

Gas chromatography (2.2.28) as described in the test for

related substances with the following modifications.

Injection 1 µL of test solution (b) and reference solution (b)

(solutions obtained after derivatisation).

Calculate the percentage content of $C_{16}H_{25}ClN_2$ using the

following expression:

$$T \times \frac{m_t}{m_v} \times \frac{R_v}{R_t}$$

T = declared percentage content of xylitol CRS;

m_t = mass of xylitol CRS in 1 mL of reference solution

(b), in milligrams;

m_v = mass of the substance to be examined in 1 mL of

test solution (b), in milligrams;

R_t = ratio of the area of the peak due to derivatised xylitol

to the area of the peak due to the derivatised internal

standard in the chromatogram obtained with

reference solution (b);

R_v = ratio of the area of the peak due to derivatised xylitol

to the area of the peak due to the derivatised internal

standard in the chromatogram obtained with test

LABELLING

The label states:

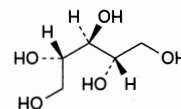
— where applicable, the maximum concentration of bacterial

endotoxins;

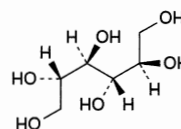
— where applicable, that the substance is suitable for use in

the manufacture of parenteral preparations.

IMPURITIES

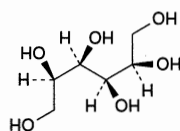


A. L-arabinitol,

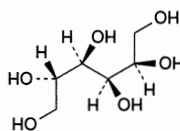


B. meso-galactitol,

C. D-mannitol,

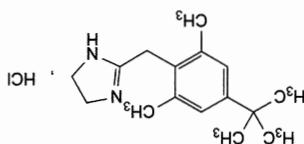


D. D-glucitol (D-sorbitol).



Xylometazoline Hydrochloride

(Ph. Eur. monograph 1162)



$C_{16}H_{25}ClN_2$

280.8

1218-35-5

Action and use

Alpha-adrenoceptor agonist.

Preparation

Xylometazoline Nasal Drops

DEFINITION

2-[4-(1,1-Dimethylethyl)-2,6-dimethylbenzyl]-4,5-dihydro-1H-imidazole hydrochloride.

Content

99.0 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Freely soluble in water, in ethanol (96 per cent) and in

IDENTIFICATION

First identification: A, E.

Second identification: B, C, D, E

A. Infrared absorption spectrophotometry (2.2.24).

Comparison xylometazoline hydrochloride CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in methanol R and dilute to 5 mL with the same solvent.

Reference solution Dissolve 20 mg of xylometazoline

hydrochloride CRS in methanol R and dilute to 5 mL with the

same solvent.

Plate TLC silica gel G plate R.

Mobile phase concentrated ammonia R, methanol R (5:100 V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Ph Eur



Chlorine treatment At the bottom of a chromatographic tank place a beaker containing a mixture of 1 volume of

hydrochloric acid R₁, 1 volume of water R and 2 volumes of a 15 g/L solution of potassium permanganate R. Close the tank

and allow to stand for 15 min. Place the dried plate in the

tank and reclose the tank. Leave the plate in contact with the

chlorine vapour for 5 min. Withdraw the plate and place it in

a current of cold air until the excess of chlorine is removed

and an area of the coating below the points of application

does not give a blue colour with a drop of potassium iodide

and starch solution R.

Detection Spray with potassium iodide and starch solution R.

Results The principal spot in the chromatogram obtained with

the test solution is similar in position, colour and size to the

principal spot in the chromatogram obtained with the

reference solution.

C. Dissolve about 0.5 mg in 1 mL of methanol R.

Add 0.5 mL of a freshly prepared 50 g/L solution of sodium

nitroprusside R and 0.5 mL of a 20 g/L solution of

sodium hydroxide R. Allow to stand for 10 min and add 1 mL

of an 80 g/L solution of sodium hydrogen carbonate R. A violet

colour develops.

D. Dissolve 0.2 g in 1 mL of water R, add 2.5 mL of ethanol

(96 per cent) R and 2 mL of 1 M sodium hydroxide.

Mix thoroughly and examine in ultraviolet light at 365 nm.

The solution shows no fluorescence or at most the same

fluorescence as a blank solution prepared in the same

manner. The identification is not valid unless a solution

prepared in the same manner using naphazoline

hydrochloride CRS instead of the substance to be examined

shows a distinct bluish fluorescence.

E. It gives reaction (a) of chlorides (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured

than reference solution Y₆ (2.2.2, Method II).

Dissolve 2.5 g in water R and dilute to 50.0 mL with the

same solvent.

Acidity or alkalinity

Dissolve 0.25 g in carbon dioxide-free water R and dilute to

25 mL with the same solvent. Add 0.1 mL of methyl red

solution R and 0.1 mL of 0.01 M hydrochloric acid.

The solution is red. Not more than 0.2 mL of 0.01 M sodium

hydroxide is required to change the colour of the indicator to

yellow.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 50.0 mg of the substance to be

examined in water R and dilute to 50.0 mL with the same

solvent. Allow to stand for 1 h before injection.

Reference solution (a) Dilute 5.0 mL of the test solution to

100.0 mL with water R. Dilute 2.0 mL of this solution to

100.0 mL with water R.

Reference solution (b) Dissolve 5.0 mg of xylometazoline

impurity A CRS and 5 mg of the substance to be examined in

10.0 mL of this solution to 50.0 mL with the same solvent. Dilute

Reference solution (c) Dilute 5.0 mL of reference solution (b)

to 50.0 mL with water R.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: end-capped octadecylsilyl silica gel for chromatography with polar incorporated groups R (5 μ m).

Mobile phase:
— mobile phase A: 1.36 g/L solution of potassium dihydrogen phosphate R adjusted to pH 3.0 with phosphoric acid R₃
— mobile phase B: acetonitrile R₁

Time	Mobile phase A	Mobile phase B
(min)	(per cent V/V)	(per cent V/V)
0 - 5	70	30
5 - 20	70 → 15	30 → 85
20 - 35	15	85

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 220 nm.

Injection 10 μ L.

Relative retention With reference to xylometazoline (retention time = about 7.2 min): impurity A = about 0.79.

System suitability: reference solution (b):

— resolution: minimum 2.5 between the peaks due to

impurity A and xylometazoline.

Limits:

— impurity A: not more than the area of the corresponding

peak in the chromatogram obtained with reference

solution (c) (0.2 per cent);

— unspecified impurities: for each impurity, not more than the

area of the principal peak in the chromatogram obtained

with reference solution (a) (0.10 per cent);

— total: not more than 5 times the area of the principal peak

in the chromatogram obtained with reference solution (a)

(0.5 per cent);

— disregard limit: 0.5 times the area of the principal peak in

the chromatogram obtained with reference solution (a)

(0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in

an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.200 g in 25 mL of anhydrous acetic acid R and add

10 mL of acetic anhydride R. Titrate with 0.1 M perchloric

acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 28.08 mg

of C₁₆H₂₅ClN₂.

STORAGE

Protected from light.

IMPURITIES

Specified impurities A

Other detectable impurities (the following substances would, if

present at a sufficient level, be detected by one or other of

the tests in the monograph. They are limited by the general

acceptance criterion for other/unspecified impurities and/or

by the general monograph Substances for pharmaceutical use

(2034). It is therefore not necessary to identify these

impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: B, C,

D, E, F.

Mobile phase water R, methanol R, anhydrous acetic acid R, ethylene chloride R (10:15:25:50 V/V/V/V); measure the volumes accurately since a slight excess of water produces cloudiness.

Application 2 µL; thoroughly dry the points of application.

Development Over a path of 15 cm.

Drying In a current of warm air.

Detection Spray with a 5 g/L solution of thymol R in a mixture of 5 volumes of sulfuric acid R and 95 volumes of ethanol (96 per cent) R. Heat in an oven at 130 °C for 10 min.

System suitability: reference solution (b):

— the chromatogram shows 3 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with reference solution (a).

C. Dissolve 0.1 g in 10 mL of water R. Add 3 mL of cupric tartrate solution R and heat. An orange or red precipitate is formed.

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

Acidity or alkalinity

To 50 mL of solution S add 0.3 mL of phenolphthalein solution R1. The solution is colourless. Not more than 0.2 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator to pink.

Specific optical rotation (2.2.7)

+18.5 to +19.5 (dried substance).

Dissolve 10.0 g in 80 mL of water R, add 1 mL of dilute ammonia R2 and dilute to 100.0 mL with water R. Allow to stand for 30 min.

Chlorides (2.4.4)

Maximum 330 ppm.

Dilute 1.5 mL of solution S to 15 mL with water R.

Heavy metals (2.4.8)

Maximum 20 ppm.

12 mL of solution S complies with test A. Prepare the reference solution using lead standard solution (2 ppm Pb) R.

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C at a pressure not exceeding 0.7 kPa.

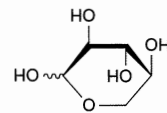
Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Ph Eur

Xylose

(Ph Eur monograph 1278)



58-86-6

C₅H₁₀O₅

Ph Eur

CHARACTERS

D-Xylopyranose.

Appearance

White or almost white, crystalline powder or colourless needles.

Solubility

Freely soluble in water, soluble in hot ethanol (96 per cent).

IDENTIFICATION

First identification A.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison xylose CRS.

B. Thin-layer chromatography (2.2.27).

Solvent mixture water R, methanol R (2:3 V/V).

Test solution Dissolve 10 mg of the substance to be examined in the solvent mixture and dilute to 20 mL with the solvent mixture.

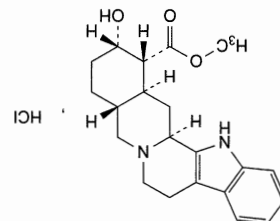
Reference solution (a) Dissolve 10 mg of xylose CRS in the solvent mixture and dilute to 20 mL with the solvent mixture.

Reference solution (b) Dissolve 10 mg of fructose R, 10 mg of glucose R and 10 mg of xylose R in the solvent mixture and dilute to 20 mL with the solvent mixture.

Plate TLC silica gel plate R.

Yohimbine Hydrochloride

(Ph. Eur. monograph 2172)



$C_{21}H_{27}ClN_2O_3$ 390.9 65-19-0

Action and use

Alpha2-adrenoceptor antagonist

DEFINITION

Methyl 17α-hydroxy-yohimbane-16α-carboxylate hydrochloride.

Content

97.0 per cent to 102.0 per cent (dried substance).

CHARACTERS

Appearance

White or slightly yellowish, crystalline powder.

Solubility

Sparsingly soluble in water; practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison yohimbine hydrochloride CRS.

B. It gives reaction (a) of chlorides (2.3.1).

TESTS

Solution S

Dissolve 0.500 g in carbon dioxide-free water R with heating, allow to cool to room temperature and dilute to 50.0 mL with the same solvent.

pH (2.2.3)

3.5 to 5.5 for solution S.

Specific optical rotation (2.2.7)

+101.0 to +105.0 (dried substance), determined on solution S.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions protected from light.

Test solution Dissolve 10.0 mg of the substance to be examined in methanol R and dilute to 50.0 mL with the same solvent.

Reference solution (a) Dissolve 5.0 mg of yohimbine hydrochloride CRS (containing impurities A, F and G) in methanol R and dilute to 25.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with methanol R.

Reference solution (c) Dilute 1.0 mL of reference solution (b) to 10.0 mL with methanol R.

Column:

size: $l = 0.125$ m, $\phi = 4.0$ mm;

(4 μ m);

— temperature: 40 °C.

— stationary phase: octylsilyl silica gel for chromatography R

Mobile phase Mix 50 mL of a 9.08 g/L solution of potassium dihydrogen phosphate R, 100 mL of an 11.88 g/L solution of disodium hydrogen phosphate dihydrate R, 285 mL of acetonitrile R, 4.0 g of sodium laurylsulfate R and 355 mL of water R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 229 nm.

Injection 10 μ L.

Run time 3 times the retention time of yohimbine.

Relative retention With reference to yohimbine (retention time = about 7 min): impurity F = about 0.65;

impurity G = about 0.70; impurity A = about 0.75.

System suitability: reference solution (a):

— peak-to-valley ratio: minimum 1.3, where H_p = height above the baseline of the peak due to impurity G and

H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to

impurity A; and minimum 1.3, where H_p = height above the baseline of the peak due to impurity G and

H_v = height above the baseline of the lowest point of the curve separating this peak from the peak due to

impurity F; not more than 4 times the area of the principal peak in the chromatogram obtained with

reference solution (b) (1.0 per cent);

— impurity F: not more than 4 times the area of the principal peak in the chromatogram obtained with

reference solution (c) (0.4 per cent);

— unspecified impurities: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (c) (0.10 per cent);

— total: not more than twice the area of the principal peak in the chromatogram obtained with reference solution (b) (2.0 per cent);

— disregard limit: 0.5 times the area of the principal peak in the chromatogram obtained with reference solution (c) (0.05 per cent).

Loss on drying (2.2.32)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 105 °C.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution and reference solution (a).

Calculate the percentage content of $C_{21}H_{27}ClN_2O_3$ from the declared content of yohimbine hydrochloride CRS.

STORAGE

In an airtight container, protected from light.

IMPURITIES

Specified impurities A, F, G

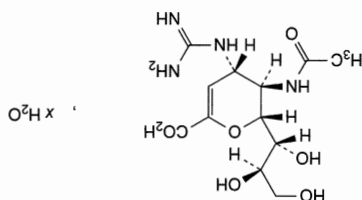
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10.

Control of impurities in substances for pharmaceutical use: B, C, D, E.



Zanamivir Hydrate

(Ph. Eur. monograph 2611)



$C_{12}H_{20}N_4O_7 \cdot xH_2O$

332.3

(anhydrous substance)

SS1942-41-7

Action and use
Neuraminidase inhibitor; treatment of influenza.

Ph Eur

DEFINITION

(2*R*,3*R*,4*S*)-3-(Acetylamino)-4-carbamimidoylamino-2-[(1*R*,2*R*)-1,2,3-trihydroxypropyl]-3,4-dihydro-2*H*-pyran-6-carboxylic acid.

Content

97.0 per cent to 102.0 per cent (dried substance).
It contains a variable quantity of water.

CHARACTERS

Appearance

White or almost white, slightly hygroscopic powder.

Solubility

Slightly soluble in water, practically insoluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24).

Comparison zanamivir hydrate CRS.

TESTS

Specific optical rotation (2.2.7)

+36.0 to +38.5 (dried substance).

Dissolve 0.250 g in 25.0 mL of water *R*; sonicate until dissolution is complete.

Related substances

Liquid chromatography (2.2.29).

Test solution (a) Dissolve 23.0 mg of the substance to be examined in 20 mL of water *R* and dilute to 50.0 mL with 50.0 mL with the mobile phase.

Test solution (b) Dilute 5.0 mL of test solution (a) to 50.0 mL with the mobile phase.

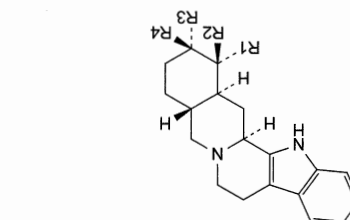
Reference solution (a) Dissolve 23.0 mg of zanamivir for assay CRS in 20 mL of water *R* and dilute to 50.0 mL with acetamintril *R*1. Dilute 5.0 mL of the solution to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of zanamivir for system suitability CRS (containing impurities A, B, C and E) in 6 mL of water *R* and dilute to 10.0 mL with acetamintril *R*1.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the mobile phase. Dilute 1.0 mL of this

solution to 10.0 mL with the mobile phase.

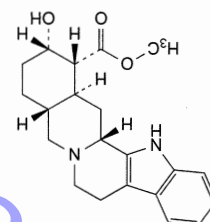
Reference solution (d) Dissolve 3.0 mg of zanamivir impurity F CRS in the mobile phase and dilute to 100.0 mL with the mobile phase.



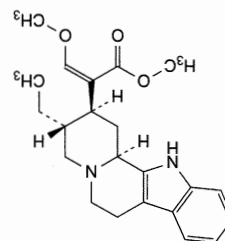
A. R1 = CO-OCH₃, R2 = R3 = H, R4 = OH: methyl 17β-hydroxyyoimbiban-16α-carboxylate (β-yoimbine),

C. R1 = R4 = H, R2 = CO-OCH₃, R3 = OH: methyl 17α-hydroxyyoimbiban-16β-carboxylate (α-yoimbine),

B. methyl 17α-hydroxy-20α-yoimbiban-16β-carboxylate



D. methyl 17α-hydroxy-3β-yoimbiban-16α-carboxylate (pseudo-yoimbine),



E. methyl (2*Z*)-2-[(2*S*,3*R*,12*bS*)-3-ethyl-1,2,3,4,6,7,12,12*b*-octahydroindolo[2,3-*a*]quinolizin-2-yl]-3-methoxyprop-2-enoate,

F. unknown structure,

G. unknown structure.

Ph Eur

Reference solution (e) Dilute 1.0 mL of reference solution (d) to 100.0 mL with the mobile phase. Dilute 3.0 mL of this solution to 20.0 mL with the mobile phase.

Column:
— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;
— stationary phase: amino alkyl vinyl polymer for chromatography R (5 μ m);
— temperature: 30 °C.

Mobile phase 0.7 g/L solution of sulfuric acid R previously adjusted to pH 5.5 with dilute ammonia R3, acetonitrile R1 (40:60 V/V).

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 234 nm.

Preconditioning of the column Prior to first use, rinse with a 0.1 g/L solution of ammonium sulfate R at 1.5 mL/min at 30 °C for about 1 h; prior to each use, rinse with the mobile phase for at least 8 h.

Injection 20 μ L of test solution (a) and reference solutions (b), (c) and (e).

Run time 3 times the retention time of zanamivir.

Identification of impurities Use the chromatogram supplied with zanamivir for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C and E; use the chromatogram obtained with reference solution (e) to identify the peak due to impurity F.

Relative retention With reference to zanamivir (retention time = about 9 min): impurity F = about 0.3; impurity B = about 0.6; impurity C = about 0.75; impurity E = about 0.8; impurity A = about 2.6.

System suitability:
— signal-to-noise ratio: minimum 10 for the principal peak in the chromatogram obtained with reference solution (e);
— peak-to-valley ratio: minimum 2.5, where H_p = height above the baseline of the peak due to impurity E and H_b = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity C in the chromatogram obtained with reference solution (b).

Calculation of percentage contents:
— for impurity F, use the concentration of impurity F in reference solution (e);
— for impurities other than F, use the concentration of zanamivir in reference solution (c).

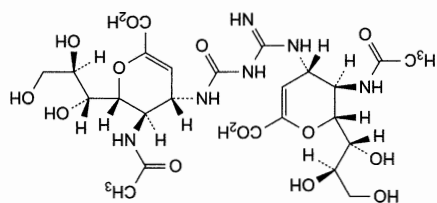
Limits:
— impurity A: maximum 0.5 per cent;
— impurity B: maximum 0.3 per cent;
— impurity C: maximum 0.2 per cent;
— impurity F: maximum 0.01 per cent;
— unspecified impurities: for each impurity, maximum 0.10 per cent;
— total: maximum 1.2 per cent;
— reporting threshold: 0.05 per cent, except for impurity F.

Loss on drying (2.2.32)
4.0 per cent to 9.0 per cent, determined on 1.000 g by drying *in vacuo* at 105 °C.

Sulfated ash (2.4.14)
Maximum 0.1 per cent, determined on 1.0 g.

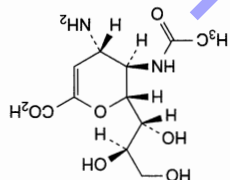
ASSAY
Liquid chromatography (2.2.29) as described in the test for related substances with the following modification.

Injection Test solution (b) and reference solution (a).

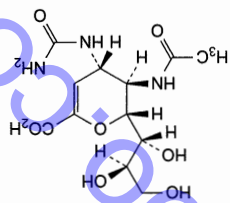


A. (2R,3R,4S)-3-(acetylamino)-4-[[[N-[(2R,3R,4S)-3-

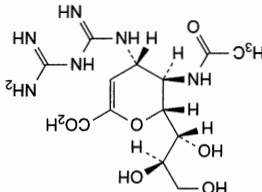
(acetylamino)-6-carboxy-2-[(1R,2R)-1,2,3-trihydroxypropyl]-3,4-dihydro-2H-pyran-6-carboxylic acid, y]carbamimidoyl]carbamoyl]amino]-2-[(1R,2R)-1,2,3-trihydroxypropyl]-3,4-dihydro-2H-pyran-6-carboxylic acid, B. unknown structure,



C. (2R,3R,4S)-3-(acetylamino)-4-amino-2-[(1R,2R)-1,2,3-trihydroxypropyl]-3,4-dihydro-2H-pyran-6-carboxylic acid,



D. (2R,3R,4S)-3-(acetylamino)-4-(carbamoylamino)-2-[(1R,2R)-1,2,3-trihydroxypropyl]-3,4-dihydro-2H-pyran-6-carboxylic acid,



E. (2R,3R,4S)-3-(acetylamino)-4-[[[carbamimidoyl]carbamimidoyl]amino]-2-[(1R,2R)-1,2,3-trihydroxypropyl]-3,4-dihydro-2H-pyran-6-carboxylic acid,

IMPURITIES

In an airtight container, protected from light.

STORAGE

Calculate the percentage content of $C_{12}H_{20}N_4O_7$ taking into account the assigned content of zanamivir for assay CRS.

Specified impurities A, B, C, F
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): D, E, H.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in the minimum volume of water R_1 , evaporate to dryness in a desiccator, under high vacuum over diphosphorus pentoxide R and record new spectra using the residues.

TESTS

Appearance of solution

The solution is not more opalescent than reference suspension I (2.2.1) and not more intensely coloured than reference solution BY₅ (2.2.2, Method II).

Dissolve 0.5 g in 50 mL of water R , heating if necessary.

Specific optical rotation (2.2.7)

+ 60.5 to + 63.0 (dried substance), measured at 25 °C. Dissolve 0.50 g in anhydrous ethanol R and dilute to 50.0 mL with the same solvent.

Related substances

A. Liquid chromatography (2.2.29).

Solvent mixture Mix 4 volumes of acetonitrile R , 20 volumes of methanol R and 76 volumes of a 2 g/L solution of ammonium acetate R previously adjusted to pH 6.8 with dilute acetic acid R .

Test solution (a) Dissolve 20.0 mg of the substance to be examined in the solvent mixture and dilute to 20.0 mL with the solvent mixture.

Test solution (b) Dilute 10.0 mL of test solution (a) to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve 2 mg of thymine R (impurity C) and 2 mg of zidovudine impurity B CRS in the solvent mixture and dilute to 50.0 mL with the solvent mixture.

Reference solution (b) Dissolve 5 mg of zidovudine for system suitability CRS (containing impurities A, G and H) in solvent mixture.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Reference solution (d) Dissolve 20.0 mg of zidovudine CRS in the solvent mixture and dilute to 20.0 mL with the solvent mixture. Dilute 10.0 mL of the solution to 50.0 mL with the solvent mixture.

Reference solution (e) Dissolve 1 mg of zidovudine impurity D CRS in a mixture of 4 volumes of acetonitrile R_1 and 56 volumes of methanol R previously adjusted to pH 6.8 with dilute acetic acid R and dilute to 50.0 mL with the same mixture of solvents. Dilute 5.0 mL of the solution to 10.0 mL with the same mixture of solvents.

Column:

size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R_1 (5 μ m).

Mobile phase:

mobile phase A: 2 g/L solution of ammonium acetate R previously adjusted to pH 6.8 with dilute acetic acid R_1 — mobile phase B: acetonitrile R_1

IDENTIFICATION

A. Specific optical rotation (see Tests).

B. Infrared absorption spectrophotometry (2.2.24). Comparison zidovudine CRS.

Solubility

Sparsely soluble in water, soluble in anhydrous ethanol, practically insoluble in heptane.

It shows polymorphism (5.9).

Appearance

White or slightly brownish powder.

CHARACTERS

97.0 per cent to 102.0 per cent (dried substance).

Content

5-methylpyrimidine-2,4(1H,3H)-dione.

1-(3-Azido-2,3-dideoxy- β -D-erythro-pentofuranosyl)-

DEFINITION

Ph Eur

Abacavir, Zidovudine and Lamivudine Tablets

Zidovudine Infusion

Zidovudine and Lamivudine Tablets

Zidovudine Tablets

Zidovudine Capsules

Preparations

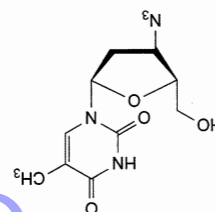
Nucleoside reverse transcriptase inhibitor; antiviral (HIV).

Action and use

C₁₀H₁₃N₅O₄

267.2

30516-87-1



(Ph. Eur. monograph 1059)

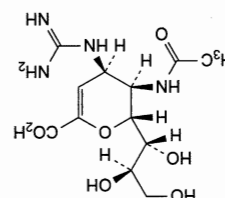
Zidovudine



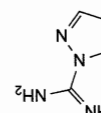
Ph Eur

carboxylic acid.

H. (2R,3R,4R)-3-(acetylamino)-4-carbamimidoylamino-2-[(1R,2R)-1,2,3-trihydroxypropyl]-3,4-dihydro-2H-pyran-6-



F. 1H-pyrazole-1-carboximidamide,



Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 3	95	5
3 - 18	95 → 85	5 → 15
18 - 28	85 → 30	15 → 70
28 - 43	30	70

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 265 nm.

Injection 20 µL of test solution (a) and reference solutions (b), (c) and (e).

Identification of impurities Use the chromatogram supplied with zidovudine for system suitability CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities A, B, C, G and H;

use the chromatogram obtained with reference solution (e) to identify the peak due to impurity D.

Relative retention With reference to zidovudine (retention time = about 16 min): impurity C = about 0.2;

impurity A = about 0.5; impurity H = about 0.95;

impurity B = about 1.05; impurity G = about 1.5;

impurity D = about 2.0.

System suitability: reference solution (b):

— resolution: minimum 2.0 between the peaks due to impurity H and zidovudine; minimum 2.0 between

the peaks due to zidovudine and impurity B.

Calculation of percentage content:

— correction factor: multiply the peak area of impurity C by 0.6;

— for each impurity, use the concentration of zidovudine in reference solution (c).

Limits:

— impurity G: maximum 0.5 per cent;

— impurities A and C: for each impurity, maximum 0.15 per cent;

— unspecified impurities: for each impurity, maximum 0.10 per cent;

— reporting threshold: 0.05 per cent; disregard any peak due to impurity D and any peak eluted after this

impurity.

B. Liquid chromatography (2.2.29).

Test solution Dissolve 0.5 g of the substance to be examined in 10 mL of acetonitrile R1 and dilute to 100.0 mL with the mobile phase.

Reference solution (a) Dissolve 5.0 mg of zidovudine impurity D CRS in acetonitrile R1 and dilute to 10.0 mL with the same solvent.

Reference solution (b) Dilute 1.0 mL of reference solution (a) to 100.0 mL with the mobile phase.

Reference solution (c) Dilute 1.0 mL of reference solution (a) to 50.0 mL with the test solution.

Column:

— size: $l = 0.15$ m, $\phi = 4.6$ mm;

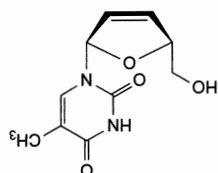
— stationary phase: base-deactivated end-capped octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase water for chromatography R, acetonitrile R1 (30:70 V/V).

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 210 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).



A. 1-[(2R,5S)-5-(hydroxymethyl)-2,4(1H,3H)-dione]-5-methylpyrimidin-2-yl]-

IMPURITIES

Protected from light.

STORAGE

Calculate the percentage content of $C_{10}H_{13}N_5O_4$ taking into account the assigned content of zidovudine CRS.

Injection Test solution (b) and reference solution (d).

Liquid chromatography (2.2.29) as described in test A for related substances with the following modification.

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

an oven at 105 °C.

Maximum 1.0 per cent, determined on 1.000 g by drying in

Loss on drying (2.2.32)

using 1 mL of lead standard solution (10 ppm Pb) R.

1.00 g complies with test H. Prepare the reference solution

Solvent mixture water R, anhydrous methanol R (20:80 V/V).

Maximum 10 ppm.

Heavy metals (2.4.8)

— total for tests A and B: maximum 1.0 per cent.

Limit:

— reporting threshold: 0.05 per cent; disregard any peak eluted before impurity D.

— unspecified impurities: for each impurity, maximum 0.10 per cent;

Limits:

— for each impurity, use the concentration of impurity D in reference solution (b).

Calculation of percentage content:

— resolution: minimum 5.0 between the peaks due to zidovudine and impurity D.

System suitability: reference solution (c):

time = about 1.5 min: impurity D = about 2.5.

Relative retention With reference to zidovudine (retention

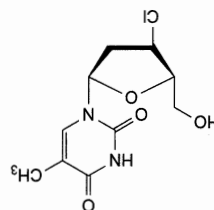
time = about 1.5 min): impurity D = about 2.5.

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to

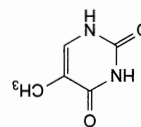
impurity D.

Run time 10 times the retention time of zidovudine.

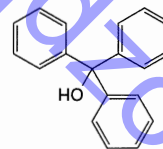
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): B, D, E, F, H, J, K.



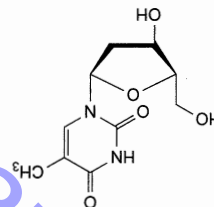
B. 1-(3-chloro-2,3-dideoxy-β-D-erythro-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione,



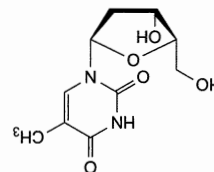
C. 5-methylpyrimidine-2,4(1H,3H)-dione (thymine),



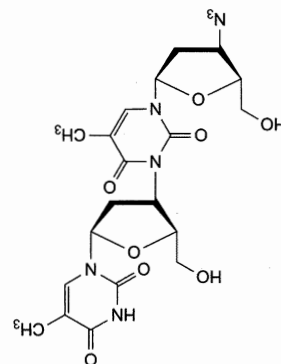
D. triphenylmethanol,



E. 1-(2-deoxy-β-D-erythro-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione (thymidine),



F. 1-(2-deoxy-β-D-threo-pentofuranosyl)-5-methylpyrimidine-2,4(1H,3H)-dione,



G. 1-[(2R,4S,5S)-4-azido-5-(hydroxymethyl)tetrahydrofuran-2-yl]-3-[(2S,3S,5R)-2-(hydroxymethyl)-5-(5-methyl-2,4-dioxo-3,4-dihydropyrimidin-1(2H)-yl)tetrahydrofuran-3-yl]-5-methylpyrimidine-2,4(1H,3H)-dione, H. unknown structure,

Zinc Acetate

(Zinc Acetate Dihydrate, Ph Eur monograph 1482)

$C_4H_6O_4Zn \cdot 2H_2O$ 219.5 5970-45-6

Action and use

Astringent.

Preparation

Erythromycin and Zinc Acetate Lotion

Ph Eur

DEFINITION

Content 99.0 per cent to 101.0 per cent of $C_4H_6O_4Zn \cdot 2H_2O$.

CHARACTERS

Appearance

White or almost white crystalline powder or flakes.

Solubility

Freely soluble in water, soluble in ethanol (96 per cent).

IDENTIFICATION

A. It gives reaction (a) of acetates (2.3.1).

B. It gives the reaction of zinc (2.3.1).

TESTS

Solution S

Dissolve 10.0 g in carbon dioxide-free water R prepared from distilled water R and dilute to 100 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

5.8 to 7.0.

Dilute 10 mL of solution S to 20 mL with carbon dioxide-free water R.

Reducing substances

Boil for 5 min a mixture of 10 mL of solution S, 90 mL of water R, 5 mL of dilute sulfuric acid R and 1.5 mL of a 0.3 g/L solution of potassium permanganate R. The pink colour of the solution remains.

Chlorides (2.4.4)

Maximum 50 ppm.

Dilute 10 mL of solution S with 15 mL of water R.

Sulfates (2.4.13)

Maximum 100 ppm, determined on solution S.

Aluminium

Maximum 5 ppm.

Atomic absorption spectrometry (2.2.23, Method J).

Test solution Dissolve 2.50 g in 20 mL of a 200 g/L solution of cadmium- and lead-free nitric acid R and dilute to 25.0 mL with the same acid solution.

Reference solutions Prepare the reference solutions using

aluminium standard solution (200 ppm Al) R, diluted with a

200 g/L solution of cadmium- and lead-free nitric acid R.

Source Aluminium hollow-cathode lamp.

Wavelength 309.3 nm.

Atomisation device Air-acetylene or acetylene-nitrous oxide

flame.

Arsenic (2.4.2, Method A)

Maximum 2 ppm, determined on 0.5 g.

Maximum 2 ppm.

Atomic absorption spectrometry (2.2.23, Method J).

Test solution Use the solution described in the test for

aluminium.

Reference solutions Prepare the reference solutions using

cadmium standard solution (0.1 per cent Cd) R, diluted with a

200 g/L solution of cadmium- and lead-free nitric acid R.

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Air-acetylene flame.

Copper

Maximum 50 ppm.

Atomic absorption spectrometry (2.2.23, Method J).

Test solution Use the solution described in the test for iron.

Reference solutions Prepare the reference solutions using copper

standard solution (10 ppm Cu) R, diluted with a 200 g/L

solution of cadmium- and lead-free nitric acid R.

Source Copper hollow-cathode lamp.

Wavelength 324.8 nm.

Atomisation device Air-acetylene flame.

Iron

Maximum 50 ppm.

Atomic absorption spectrometry (2.2.23, Method J).

Test solution Dissolve 1.25 g in 20 mL of a 200 g/L solution

of cadmium- and lead-free nitric acid R and dilute to 25.0 mL

with the same acid solution.

Reference solutions Prepare the reference solutions using iron

standard solution (20 ppm Fe) R, diluted with a 200 g/L

solution of cadmium- and lead-free nitric acid R.

Source Iron hollow-cathode lamp.

Wavelength 248.3 nm.

Atomisation device Air-acetylene flame.

Lead

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method J).

Test solution Dissolve 5.00 g in 20 mL of a 200 g/L solution

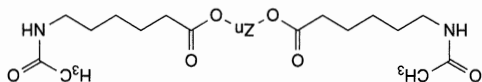
of cadmium- and lead-free nitric acid R and dilute to 25.0 mL

with the same acid solution.

Zinc Acexamate



(Ph. Eur. monograph 1279)



$C_{16}H_{28}N_2O_6Zn$ 409.8

70020-71-2

DEFINITION

Zinc 6-(acetylamino)hexanoate.

Content

97.5 per cent to 101.0 per cent (dried substance).

CHARACTERS

Appearance

White or almost white, crystalline powder.

Solubility

Soluble in water, practically insoluble in acetone and in ethanol (96 per cent). It dissolves in dilute nitric acid.

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison zinc acexamate CRS.

B. 5 mL of solution S (see Tests) gives the reaction of zinc

TESTS

Solution S Dissolve 0.5 g in carbon dioxide-free water R and dilute to

20 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference

suspension IV (2.2.1) and is colourless (2.2.2, Method II).

pH (2.2.3)

5.0 to 7.0 for solution S.

Impurity B

Thin-layer chromatography (2.2.27).

Test solution Dissolve 0.30 g of the substance to be examined

in water R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 15 mg of 6-aminohexanoic acid R

(impurity B) in water R and dilute to 10 mL with the same

solvent. Dilute 1 mL of this solution to 10 mL with water R.

Plate TLC silica gel plate R.

Mobile phase ammonia R, water R, ethanol (96 per cent) R

(2.30:68 V/V/V).

Application 5 µL; allow to dry in air.
Development Over a path of 15 cm.
Drying In a current of warm air.
Detection Spray with ninhydrin solution R and heat at 100-105 °C for 15 min.
Limit:

— **impurity B:** any spot due to impurity B is not more intense than the corresponding spot in the chromatogram obtained with the reference solution (0.5 per cent).

Related substances

Liquid chromatography (2.2.29).
Test solution (a) Dissolve 0.50 g of the substance to be examined in water R and dilute to 100.0 mL with the same solvent.
Test solution (b) To 20.0 mL of test solution (a), add 20 mL of the mobile phase and 0.4 mL of a 100 g/L solution of phosphoric acid R, then dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 40 mg of *N*-acetyl-*ε*-caprolactam R (impurity C) in water R and dilute to 100.0 mL with the same solvent.
Reference solution (b) Dilute 5.0 mL of reference solution (a) to 100.0 mL with water R.
Reference solution (c) Dissolve 20 mg of zinc acexamate impurity A CRS in water R and dilute to 50.0 mL with the same solvent.

Reference solution (d) Dissolve 40 mg of *ε*-caprolactam R (impurity D) in water R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with water R.
Reference solution (e) To 20.0 mL of test solution (a), add 5.0 mL of reference solution (b), 5.0 mL of reference solution (c), 5.0 mL of reference solution (d) and 0.4 mL of a 100 g/L solution of phosphoric acid R, then dilute to 50.0 mL with the mobile phase.

Reference solution (f) To 5.0 mL of reference solution (c), add 5.0 mL of reference solution (b), 5.0 mL of reference solution (d) and 0.4 mL of a 100 g/L solution of phosphoric acid R, then dilute to 50.0 mL with the mobile phase.
Column:
 — size: $l = 0.25$ m, $\varnothing = 4.0$ mm;
 — stationary phase: octadecylsilyl silica gel for chromatography R (5 µm).

Mobile phase Mix 0.2 volumes of phosphoric acid R, 8 volumes of acetonitrile R and 92 volumes of water R, then adjust to pH 4.5 with dilute ammonia R1.
Flow rate 1.2 mL/min.
Detection Spectrophotometer at 210 nm.

Injection 20 µL of test solution (b) and reference solutions (b), (c) and (f).
Run time 8 times the retention time of zinc acexamate.
Elution order Zinc acexamate, impurity D, impurity A, impurity C.

System suitability: reference solution (e):
 — **resolution:** minimum 3.0 between the peaks due to zinc acexamate and impurity D; if necessary, adjust the mobile phase to pH 4.7 with dilute ammonia R1.
Limits:

— **impurity A:** not more than the area of the corresponding peak in the chromatogram obtained with reference solution (f) (2 per cent);

— **impurities C, D:** for each impurity, not more than 1.5 times the area of the corresponding peak in the chromatogram obtained with reference solution (f) (0.15 per cent);
 — **unspecified impurities:** for each impurity, not more than 0.5 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (f) (0.05 per cent);
 — **sum of impurities other than A:** not more than 5 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (f) (0.5 per cent);
 — **disregard limit:** 0.5 times the area of the peak due to impurity C in the chromatogram obtained with reference solution (f) (0.05 per cent).

Arsenic (2.4.2, Method A)
 Maximum 2 ppm, determined on 0.5 g.

Cadmium

Maximum 2 ppm.

Atomic absorption spectrometry (2.2.23, Method D).
Test solution Dissolve 2.50 g in 20 mL of a 200 g/L solution of cadmium- and lead-free nitric acid R and dilute to 25.0 mL with the same acid solution.

Reference solutions Prepare the reference solutions using cadmium standard solution (0.1 per cent Cd) R, diluting with a 200 g/L solution of cadmium- and lead-free nitric acid R.
Source Cadmium hollow-cathode lamp.
Wavelength 228.8 nm.
Atomisation device Air-acetylene flame.

Iron

Maximum 50 ppm.

Atomic absorption spectrometry (2.2.23, Method D).
Test solution Dissolve 1.25 g in 20 mL of a 200 g/L solution of cadmium- and lead-free nitric acid R and dilute to 25.0 mL with the same acid solution.

Reference solutions Prepare the reference solutions using iron standard solution (20 ppm Fe) R, diluting with a 200 g/L solution of cadmium- and lead-free nitric acid R.
Source Iron hollow-cathode lamp.
Wavelength 248.3 nm.
Atomisation device Air-acetylene flame.

Lead

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method D).
Test solution Dissolve 5.00 g in 20 mL of a 200 g/L solution of cadmium- and lead-free nitric acid R and dilute to 25.0 mL with the same acid solution.

Reference solutions Prepare the reference solutions using lead standard solution (0.1 per cent Pb) R, diluting with a 200 g/L solution of cadmium- and lead-free nitric acid R.
Source Lead hollow-cathode lamp.
Wavelength 283.3 nm.
Atomisation device Air-acetylene flame.

Loss on drying (2.2.32)

Maximum 1.0 per cent, determined on 1.000 g by drying in an oven at 105 °C.

ASSAY

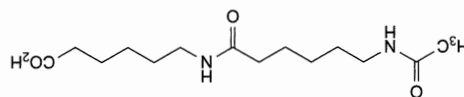
Dissolve 0.400 g in 10 mL of dilute acetic acid R. Carry out the complexometric titration of zinc (2.5.11).
 1 mL of 0.1 M sodium edetate is equivalent to 40.98 mg of $C_{16}H_{28}N_2O_6Zn$.

STORAGE

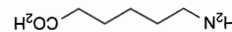
In a non-metallic container.

IMPURITIES

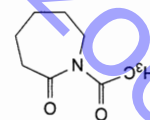
Specified impurities A, B, C, D.



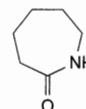
A. 6-[[6-(acetylamino)hexanoyl]amino]hexanoic acid,



B. 6-aminoheptanoic acid (6-aminocaproic acid),



C. 1-acetylhexahydro-2H-azepin-2-one (N-acetyl-ε-caprolactam),



D. hexahydro-2H-azepin-2-one (ε-caprolactam).

Zinc Chloride

(Ph. Eur. monograph 0110)

ZnCl₂

136.3

7646-85-7

**DEFINITION****Content**

95.0 per cent to 100.5 per cent.

CHARACTERS**Appearance**

White or almost white, crystalline powder or cast in white or almost white sticks, deliquescent.

Solubility

Very soluble in water, freely soluble in ethanol (96 per cent)

and in glycerol.

IDENTIFICATION

A. Dissolve 0.5 g in dilute nitric acid R and dilute to 10 mL with the same acid. The solution gives reaction (a) of chlorides (2.3.1).

B. 5 mL of solution S (see Tests) gives the reaction of zinc (2.3.1).

TESTS**Solution S**

To 2.0 g add 38 mL of carbon dioxide-free water R prepared from distilled water R and add dilute hydrochloric acid R dropwise until dissolution is complete. Dilute to 40 mL with carbon dioxide-free water R prepared from distilled water R.

pH (2.2.3)

4.6 to 5.5.

Dissolve 1.0 g in 9 mL of carbon dioxide-free water R, ignoring any slight turbidity.

Oxychlorides

Dissolve 10.0 g in 10 mL of carbon dioxide-free water R.

The solution is not more opalescent than reference suspension II (2.2.1). To 1.5 mL of the solution add 7.5 mL of ethanol (96 per cent) R. The solution may become cloudy within 10 min. Any cloudiness disappears on the addition of 0.2 mL of dilute hydrochloric acid R.

Sulfates (2.4.13)

Maximum 200 ppm.

Dilute 5 mL of solution S to 15 mL with distilled water R.

Prepare the standard using a mixture of 5 mL of sulfate standard solution (10 ppm SO₄) R and 10 mL of distilled water R.**Aluminium, calcium, heavy metals, iron, magnesium**

To 8 mL of solution S add 2 mL of concentrated ammonia R and shake. The solution is clear (2.2.1) and colourless (2.2.2, Method II). Add 1 mL of disodium hydrogen phosphate solution R. The solution remains clear for at least 5 min.

Add 0.2 mL of sodium sulfide solution R. A white precipitate is formed and the supernatant remains colourless.

Ammonium (2.4.1)

Maximum 400 ppm.

Dilute 0.5 mL of solution S to 15 mL with water R.

ASSAY

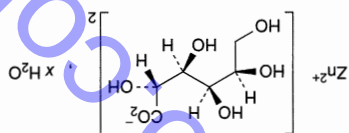
Dissolve 0.250 g in 5 mL of dilute acetic acid R. Carry out the complexometric titration of zinc (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 13.63 mg of ZnCl₂.**STORAGE**

In a non-metallic container.

Zinc Gluconate

(Ph. Eur. monograph 2164)

C₁₂H₂₂ZnO₁₄·xH₂O

455.7

(anhydrous substance)

**DEFINITION**

Anhydrous or hydrated zinc D-gluconate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS**Appearance**

White or almost white, hygroscopic, crystalline powder.

Solubility

Soluble in water, practically insoluble in anhydrous ethanol and in methylene chloride.

IDENTIFICATION

A. Thin-layer chromatography (2.2.27).

Test solution Dissolve 20 mg of the substance to be examined in 1 mL of water R.

Reference solution Dissolve 20 mg of calcium gluconate CRS in 1 mL of water R, heating if necessary in a water-bath at 60 °C.

Plate TLC silica gel plate R (5–40 µm) [or TLC silica gel plate R (2–10 µm)].

Mobile phase concentrated ammonia R, ethyl acetate R, water R, ethanol (96 per cent) R (10:10:30:50 V/V/V/V).

Application 1 µL.

Development Over 3/4 of the plate.

Drying At 100–105 °C for 20 min, then allow to cool to room temperature.

Detection Spray with a solution containing 25 g/L of

ammonium molybdate R and 10 g/L of cerium sulfate R in dilute sulfuric acid R, and heat at 100–105 °C for about 10 min.

Results The principal spot in the chromatogram obtained with the test solution is similar in position, colour and size to the principal spot in the chromatogram obtained with the reference solution.

B. Dissolve 0.1 g in 5 mL of water R. Add 0.5 mL of potassium ferrocyanide solution R. A white precipitate is formed that does not dissolve upon the addition of 5 mL of hydrochloric acid R.

TESTS**Solution S**

Dissolve 1.0 g in water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Sucrose and reducing sugars

Dissolve 0.5 g in a mixture of 2 mL of hydrochloric acid R1 and 10 mL of water R. Boil for 5 min, allow to cool, add 10 mL of sodium carbonate solution R and allow to stand for 10 min. Dilute to 25 mL with water R and filter. To 5 mL of the filtrate add 2 mL of cupri-tartaric solution R and boil for 1 min. Allow to stand for 2 min. No red precipitate is formed.

Chlorides (2.4.4)

Maximum 500 ppm.

Dilute 5 mL of solution S to 15 mL with water R.

Sulfates (2.4.13)

Maximum 500 ppm.

Dissolve 2.0 g in a mixture of 10 mL of acetic acid R and 90 mL of distilled water R.

Cadmium

Maximum 2 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 5.00 g in 20 mL of deionised distilled water R with the aid of ultrasound and dilute to 25.0 mL with the same solvent.

Reference solutions Prepare the reference solutions using cadmium standard solution (0.1 per cent Cd) R, diluting with deionised distilled water R.

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Air-acetylene flame.

Zinc Oxide

(Ph. Eur. monograph 0252)

ZnO

81.4

1314-13-2

**Action and use**

Mild astringent.

Preparations

Zinc Cream
Coal Tar and Zinc Ointment

Zinc Ointment

Zinc and Castor Oil Ointment

Compound Zinc Paste

Zinc and Salicylic Acid Paste

Zinc and Coal Tar Paste

DEFINITION

Content

99.0 per cent to 100.5 per cent (ignited substance).

CHARACTERS**Appearance**

Soft, white or faintly yellowish-white, amorphous powder, free from gritty particles.

Solubility

Practically insoluble in water and in ethanol (96 per cent). It dissolves in dilute mineral acids.

IDENTIFICATION

A. It becomes yellow when strongly heated; the yellow colour disappears on cooling.

B. Dissolve 0.1 g in 1.5 mL of dilute hydrochloric acid R and dilute to 5 mL with water R. The solution gives the reaction of zinc (2.3.1).

TESTS**Alkalinity**

Shake 1.0 g with 10 mL of boiling water R. Add 0.1 mL of phenolphthalein solution R and filter. If the filtrate is red, not

Zinc Stearate

(Ph. Eur. monograph 0306)

557-05-1

Action and use

Excipient.

DEFINITION

Zinc stearate $[(C_{17}H_{35}COO)_2Zn]$, 632 may contain varying proportions of zinc palmitate $[(C_{15}H_{31}COO)_2Zn]$, 576.2 and zinc oleate $[(C_{17}H_{33}COO)_2Zn]$, 628.

Content

10.0 per cent to 12.0 per cent of Zn.

CHARACTERS

Appearance

Light, white or almost white, amorphous powder, free from

Solubility

Practically insoluble in water and in anhydrous ethanol.

IDENTIFICATION

A. Freezing point (2.2.18): minimum 53 °C, determined on the residue obtained in the preparation of solution S (see Tests).

B. Neutralise 5 mL of solution S to red litmus paper R with strong sodium hydroxide solution R. The solution gives the

TESTS

Solution S

To 5.0 g add 50 mL of ether R and 40 mL of a 7.5 per cent V/V solution of cadmium- and lead-free nitric acid R in distilled water R. Heat under a reflux condenser until dissolution is complete. Allow to cool. In a separating funnel, separate the aqueous layer and shake the ether layer with 2 quantities, each of 4 mL, of distilled water R. Combine the aqueous layers, wash with 15 mL of ether R and heat on a water-bath until ether is completely eliminated. Allow to cool and dilute to 50.0 mL with distilled water R (solution S). Evaporate the ether layer to dryness and dry the residue at 105 °C.

Appearance of solution

Solution S is not more intensely coloured than reference solution Y₆ (2.2.2, Method II).

Appearance of solution of fatty acids

Dissolve 0.5 g of the residue obtained in the preparation of solution S in 10 mL of chloroform R. The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₅ (2.2.2, Method II).

Acidity or alkalinity

Shake 1.0 g with 5 mL of ethanol (96 per cent) R and add 20 mL of carbon dioxide-free water R and 0.1 mL of phenol red solution R. Not more than 0.3 mL of 0.1 M hydrochloric acid or 0.1 mL of 0.1 M sodium hydroxide is required to change the colour of the indicator.

Acid value of the fatty acids (2.5.1)

195 to 210.

Dissolve 0.20 g of the residue obtained in the preparation of solution S in 25 mL of the prescribed mixture of solvents.

Chlorides (2.4.4)

Maximum 250 ppm.

Dilute 2 mL of solution S to 15 mL with water R.

more than 0.3 mL of 0.1 M hydrochloric acid is required to change the colour of the indicator.

Carbonates and substances insoluble in acids

Dissolve 1.0 g in 15 mL of dilute hydrochloric acid R. It dissolves without effervescence and the solution is not more opalescent than reference suspension II (2.2.1) and is colourless (2.2.2, Method II).

Arsenic (2.4.2, Method A)

Maximum 5 ppm, determined on 0.2 g.

Cadmium

Maximum 10 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 2.0 g in 14 mL of a mixture of equal volumes of water R and cadmium- and lead-free nitric acid R, boil for 1 min, cool and dilute to 100.0 mL with water R.

Reference solutions Prepare the reference solutions using

cadmium standard solution (0.1 per cent Cd) R and diluting with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.

Source Cadmium hollow-cathode lamp.

Wavelength 228.8 nm.

Atomisation device Air-acetylene or air-propane flame.

Iron (2.4.9)

Maximum 200 ppm.

Dissolve 50 mg in 1 mL of dilute hydrochloric acid R and dilute to 10 mL with water R. Use in this test 0.5 mL of thioglycolic acid R.

Lead

Maximum 50 ppm.

Atomic absorption spectrometry (2.2.23, Method II).

Test solution Dissolve 5.0 g in 24 mL of a mixture of equal volumes of water R and cadmium- and lead-free nitric acid R, boil for 1 min, cool and dilute to 100.0 mL with water R.

Reference solutions Prepare the reference solutions using lead

standard solution (0.1 per cent Pb) R and diluting with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.

Source Lead hollow-cathode lamp.

Wavelength 283.3 nm; 217.0 nm may be used depending on the apparatus.

Atomisation device Air-acetylene flame.

Loss on ignition

Maximum 1.0 per cent, determined on 1.00 g by ignition to constant mass at 500 ± 50 °C.

ASSAY

Dissolve 0.150 g in 10 mL of dilute acetic acid R. Carry out the complexometric titration of zinc (2.5.11).

1 mL of 0.1 M sodium edetate is equivalent to 8.14 mg of ZnO.

Ph Eur



<p>Sulfates (2.4.13) Maximum 0.6 per cent. Dilute 1 mL of solution S to 50 mL with <i>distilled water</i> R. Dilute 12.5 mL of this solution to 15 mL with <i>distilled water</i> R.</p> <p>Cadmium Maximum 5 ppm. Atomic absorption spectrometry (2.2.23, <i>Method II</i>). <i>Test solution</i> Dilute 20.0 mL of solution S to 50.0 mL with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.</p> <p><i>Reference solutions</i> Prepare the reference solutions using cadmium standard solution (0.1 per cent Cd) R and diluting with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.</p> <p><i>Source</i> Cadmium hollow-cathode lamp. <i>Wavelength</i> 228.8 nm. <i>Atomisation device</i> Air-acetylene or an air-propane flame.</p> <p>Lead Maximum 25 ppm. Atomic absorption spectrometry (2.2.23, <i>Method II</i>). <i>Test solution</i> Solution S.</p> <p><i>Reference solutions</i> Prepare the reference solutions using lead standard solution (0.1 per cent Pb) R and diluting with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.</p> <p><i>Source</i> Lead hollow-cathode lamp. <i>Wavelength</i> 283.3 nm. Depending on the apparatus the line at 217.0 nm may be used. <i>Atomisation device</i> Air-acetylene flame.</p> <p>ASSAY To 1.000 g add 50 mL of <i>dilute acetic acid</i> R and boil for at least 10 min or until the layer of fatty acids is clear, adding more <i>water</i> R as necessary to maintain the original volume. Cool and filter. Wash the filter and the flask with <i>water</i> R. Combine the filtrate and washings. Carry out the complexometric titration of zinc (2.5.11). 1 mL of 0.1 M sodium edetate is equivalent to 6.54 mg of Zn.</p> <p>Zinc Sulfate Hexahydrate (Ph. Eur. monograph 1683) ZnSO₄·6H₂O 269.5 13986-24-8</p> <p>Action and use Astringent.</p> <p>Ph Eur</p> <p>DEFINITION Content 99.0 per cent to 104.0 per cent.</p> <p>CHARACTERS Appearance White or almost white, crystalline powder or colourless transparent crystals, efflorescent.</p> <p>Solubility Very soluble in water, practically insoluble in ethanol (96 per cent).</p> <p>IDENTIFICATION A. Solution S (see Tests) gives the reactions of sulfates (2.3.1). B. Solution S gives the reaction of zinc (2.3.1). C. It complies with the limits of the assay.</p> <p>TESTS Solution S Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.</p>	<p>Sulfates (2.4.13) Maximum 0.6 per cent. Dilute 1 mL of solution S to 50 mL with <i>distilled water</i> R. Dilute 12.5 mL of this solution to 15 mL with <i>distilled water</i> R.</p> <p>Cadmium Maximum 5 ppm. Atomic absorption spectrometry (2.2.23, <i>Method II</i>). <i>Test solution</i> Dilute 20.0 mL of solution S to 50.0 mL with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.</p> <p><i>Reference solutions</i> Prepare the reference solutions using cadmium standard solution (0.1 per cent Cd) R and diluting with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.</p> <p><i>Source</i> Cadmium hollow-cathode lamp. <i>Wavelength</i> 228.8 nm. <i>Atomisation device</i> Air-acetylene or an air-propane flame.</p> <p>Lead Maximum 25 ppm. Atomic absorption spectrometry (2.2.23, <i>Method II</i>). <i>Test solution</i> Solution S.</p> <p><i>Reference solutions</i> Prepare the reference solutions using lead standard solution (0.1 per cent Pb) R and diluting with a 3.5 per cent V/V solution of cadmium- and lead-free nitric acid R.</p> <p><i>Source</i> Lead hollow-cathode lamp. <i>Wavelength</i> 283.3 nm. Depending on the apparatus the line at 217.0 nm may be used. <i>Atomisation device</i> Air-acetylene flame.</p> <p>ASSAY To 1.000 g add 50 mL of <i>dilute acetic acid</i> R and boil for at least 10 min or until the layer of fatty acids is clear, adding more <i>water</i> R as necessary to maintain the original volume. Cool and filter. Wash the filter and the flask with <i>water</i> R. Combine the filtrate and washings. Carry out the complexometric titration of zinc (2.5.11). 1 mL of 0.1 M sodium edetate is equivalent to 6.54 mg of Zn.</p> <p>Zinc Sulfate Heptahydrate (Ph. Eur. monograph 0111) ZnSO₄·7H₂O 287.5 7446-20-0</p> <p>Action and use Astringent.</p> <p>Ph Eur</p> <p>Preparations Zinc Sulfate Eye Drops Zinc Sulfate Injection Zinc Sulfate Lotion</p> <p>DEFINITION Content 99.0 per cent to 104.0 per cent.</p> <p>CHARACTERS Appearance White or almost white, crystalline powder or colourless, transparent crystals, efflorescent.</p>
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Appearance of solution
Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)
4.4 to 5.6 for solution S.

Chlorides (2.4.4)
Maximum 300 ppm.
Dilute 3.3 mL of solution S to 15 mL with water R.

Iron (2.4.9)
Maximum 100 ppm.
Dilute 2 mL of solution S to 10 mL with water R. Use in this test 0.5 mL of thioglycolic acid R.

ASSAY
Dissolve 0.200 g in 5 mL of dilute acetic acid R. Carry out the complexometric titration of zinc (2.5.11).

STORAGE
ZnSO₄·6H₂O.
1 mL of 0.1 M sodium edetate is equivalent to 26.95 mg of

In a non-metallic, airtight container.

Zinc Sulfate Monohydrate

(Ph. Eur. monograph 2159)

ZnSO₄·H₂O

Action and use

Astringent.

Preparations

Zinc Sulfate Capsules

Zinc Sulfate Tablets

Ph Eur

DEFINITION

Content

99.0 per cent to 101.0 per cent.

CHARACTERS

Appearance

White or almost white, crystalline powder, or colourless, transparent crystals.

Solubility

Very soluble in water, practically insoluble in ethanol

(96 per cent).

IDENTIFICATION

A. Solution S (see Tests) gives the reactions of sulfates

(2.3.1).
B. Solution S gives the reaction of zinc (2.3.1).

C. It complies with the limits of the assay.

TESTS

Solution S

Dissolve 2.5 g in carbon dioxide-free water R and dilute to 50 mL with the same solvent.

Appearance of solution

Solution S is clear (2.2.1) and colourless (2.2.2, Method II).

pH (2.2.3)

4.0 to 5.6 for solution S.

Chlorides (2.4.4)

Maximum 300 ppm.

Dilute 3.3 mL of solution S to 15 mL with water R.

Alkalinity

Mix 1.0 g with 5 mL of ethanol (96 per cent) R and 0.5 mL of phenol red solution R. Add 50 mL of carbon dioxide-free

TESTS

of zinc (2.3.1).

identification test A and 4 mL of water R gives the reaction

C. A mixture of 1 mL of the aqueous layer obtained in

potassium permanganate solution is discharged.

potassium permanganate solution R. The colour of the

and 5 mL of glacial acetic acid R. Add dropwise 0.25 mL of

B. Dissolve 0.1 g in a mixture of 2 mL of dilute sulfuric acid R

(2.2.14) at 66 °C to 68 °C.

ethanol (70 per cent V/V) R and drying in vacuo for 3 h, melts

a water-bath. The residue, after recrystallisation twice from

20 mL of water R. Evaporate the organic layer to dryness on

each of 20 mL, of dilute hydrochloric acid R and then with

to cool and add 30 mL of ether R. Shake with 3 quantities,

aniline R and boil under a reflux condenser for 10 min. Allow

to dryness. To the residue add 2 mL of freshly distilled

Wash the combined ether layers with water R and evaporate

ether R. Reserve the aqueous layer for identification test C.

sulfuric acid R. Shake with 2 quantities, each of 10 mL, of

A. To 2.5 g add 10 mL of water R and 10 mL of dilute

IDENTIFICATION

116 °C to 121 °C, it may leave a slight solid residue.

mp

Practically insoluble in water and in ethanol (96 per cent).

Solubility

White or almost white, fine powder.

Appearance

CHARACTERS

98.0 per cent to 102.0 per cent (dried substance).

Content

Zinc di(undec-10-enoate).

DEFINITION

Ph Eur

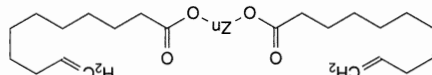
Action and use

Used topically in treatment of fungal infections.

Ph Eur

Zinc Undecenoate

(Zinc Undecylenate, Ph Eur monograph 0539)



C₂₂H₃₈O₄Zn

431.9

557-08-4



Ph Eur

STORAGE

of ZnSO₄·H₂O.

1 mL of 0.1 M sodium edetate is equivalent to 17.95 mg

complexometric titration of zinc (2.5.11).

Dissolve 0.160 g in 5 mL of dilute acetic acid R. Carry out the

ASSAY

Use 0.5 mL of thioglycolic acid R in this test.

Dilute 2 mL of solution S to 10 mL with water R.

Maximum 100 ppm.

Iron (2.4.9)

In a non-metallic container.

5-[2-[4-(1,2-Benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6-chloro-1,3-dihydro-2H-indol-2-one hydrochloride monohydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or slightly pink powder.

Solubility

Practically insoluble in water, slightly soluble in methanol and in methylene chloride.

It shows polymorphism (5.9).

IDENTIFICATION

A. Infrared absorption spectrophotometry (2.2.24).

Comparison ziprasidone hydrochloride monohydrate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

B. Suspend 30 mg in 2 mL of water R, acidify with 0.15 mL of dilute nitric acid R and filter. The clear filtrate gives reaction (a) of chlorides (2.3.1).

TESTS

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light and prepare the solutions immediately before use.

Solvent mixture A water R, methanol R (40:60 V/V).

Solvent mixture B hydrochloric acid R, water R, methanol R (0.04:20:80 V/V/V).

Test solution (a) Dissolve 23 mg of the substance to be examined in solvent mixture A and dilute to 100.0 mL with solvent mixture A.

Test solution (b) Dissolve 23 mg of the substance to be examined in solvent mixture B and dilute to 50.0 mL with solvent mixture B.

Reference solution (a) Dissolve 2.5 mg of ziprasidone for system suitability 1 CRS (containing impurities A, B and C) in solvent mixture A and dilute to 10.0 mL with solvent mixture A.

Reference solution (b) Dilute 1.0 mL of test solution (b) to 100.0 mL with solvent mixture B. Dilute 1.0 mL of this solution to 10.0 mL with solvent mixture B.

Reference solution (c) Dissolve the contents of a vial of ziprasidone for system suitability 2 CRS (containing impurities D and E) in 1.0 mL of solvent mixture B.

A. Column:

— size: $l = 0.15$ m, $\varnothing = 4.6$ mm;

— stationary phase: spherical octylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase:

— mobile phase A: mix 40 volumes of methanol R and 60 volumes of a 6.8 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 2.5 with phosphoric acid R;

— mobile phase B: methanol R;

water R and examine immediately. No reddish colour appears.

Alkali and alkaline-earth metals

Maximum 2.0 per cent.

To 1.0 g add 25 mL of water R and 5 mL of hydrochloric acid R and heat to boiling. Filter whilst hot. Wash the filter and the residue with 25 mL of hot water R. Combine the filtrate and washings and add concentrated ammonia R until alkaline. Add 7.5 mL of thiocyanamide solution R and heat on a water-bath for 30 min. Filter and wash the precipitate with 2 quantities, each of 10 mL, of water R. Combine the filtrate and washings, evaporate to dryness on a water-bath and ignite. The residue weighs a maximum of 20 mg.

Sulfates (2.4.13)

Maximum 500 ppm.

To 0.1 g add a mixture of 2 mL of dilute hydrochloric acid R and 10 mL of distilled water R and heat to boiling. Cool, filter and dilute to 15 mL with distilled water R. Prepare the standard using 5 mL of sulfate standard solution (10 ppm SO_4) R and 10 mL of distilled water R.

Loss on drying (2.2.32)

Maximum 1.5 per cent, determined on 0.500 g by drying in an oven at 105 °C.

Degree of unsaturation

Dissolve 0.100 g in a mixture of 5 mL of dilute hydrochloric acid R and 30 mL of glacial acetic acid R. Using 0.05 mL indigo carmine solution RI, added towards the end of the titration as indicator. Titrate with 0.0167 M bromide-bromate until the colour changes from blue to yellow. 9.1 mL to 9.4 mL of 0.0167 M bromide-bromate is required. Carry out a blank titration.

ASSAY

To 0.350 g add 25 mL of dilute acetic acid R and heat to boiling. Carry out the complexometric titration of zinc (2.5.11).

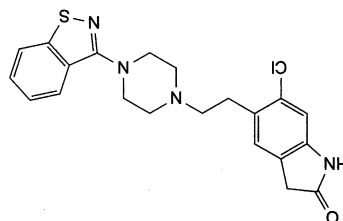
1 mL of 0.1 M sodium edetate is equivalent to 43.19 mg of $\text{C}_{21}\text{H}_{22}\text{Cl}_2\text{N}_4\text{O}_5\text{H}_2\text{O}$.

STORAGE

Protected from light.

Ziprasidone Hydrochloride Monohydrate

(Ph Eur monograph 2421)



$\text{C}_{21}\text{H}_{22}\text{Cl}_2\text{N}_4\text{O}_5\text{H}_2\text{O}$ 467.4 138982-67-9

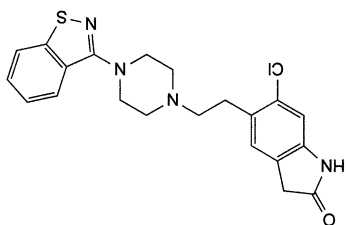
Action and use

Dopamine D_2 receptor antagonist; serotonin 5HT_2 receptor antagonist; neuroleptic.



Ziprasidone Mesilate Trihydrate

(Ph. Eur. monograph 2649)



$C_{22}H_{25}ClN_4O_4S \cdot 3H_2O$ 563.1 199191-69-0

Action and use

Dopamine D_2 receptor antagonist; serotonin 5HT₂ receptor antagonist; neuroleptic.

Ph Eur

DEFINITION

5-[2-[4-(1,2-Benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6-chloro-1,3-dihydro-2H-indol-2-one methanesulfonate trihydrate.

Content

98.0 per cent to 102.0 per cent (anhydrous substance).

PRODUCTION

It is considered that alkylsulfonate esters are genotoxic and are potential impurities in ziprasidone mesilate trihydrate. The manufacturing process should be developed taking into consideration the principles of quality risk management, together with considerations of the quality of starting materials, process capability and validation. The general methods 2.5.37. *Methyl, ethyl and isopropyl methanesulfonate in active substances* and 2.5.39. *Methanesulfonyl chloride in methanesulfonic acid* are available to assist manufacturers.

CHARACTERS

Appearance
White or almost white powder.

Solubility

Very slightly soluble in water, slightly soluble in methanol, practically insoluble in acetonitrile.

It shows polymorphism (5.9).

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison ziprasidone mesilate trihydrate CRS.

If the spectra obtained in the solid state show differences, dissolve the substance to be examined and the reference substance separately in methanol R, evaporate to dryness and record new spectra using the residues.

TESTS

Related substances

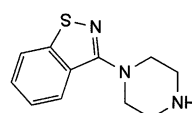
Carry out the tests protected from light and prepare the solutions immediately before use.

A. Liquid chromatography (2.2.29).

Solvent mixture hydrochloric acid R, water for chromatography R, methanol R (0.04:40:60 V/V/V).

Test solution (a) Dissolve 27.0 mg of the substance to be examined in 35 mL of the solvent mixture, sonicate for about 2 min and shake. Dilute to 50.0 mL with the solvent mixture.

A. 3-piperazin-1-yl-1,2-benzisothiazole,



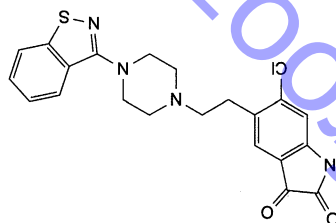
Specified impurities A, B, C, D, E.

IMPURITIES

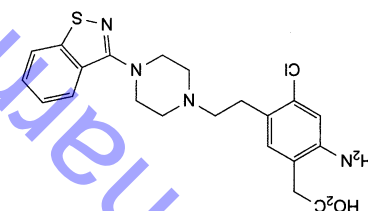
Protected from light.

STORAGE

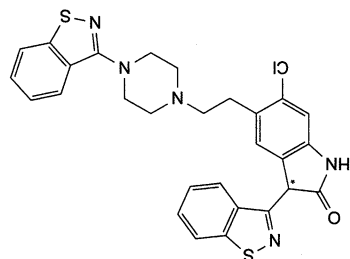
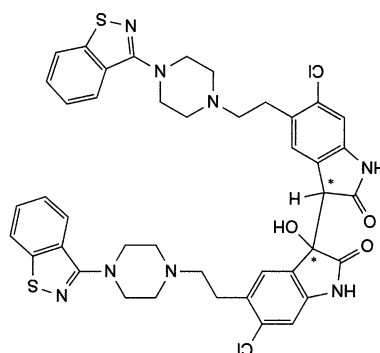
B. 5-[2-[4-(1,2-Benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6-chloro-1H-indole-2,3-dione,



C. 2-[2-amino-5-[2-[4-(1,2-Benzisothiazol-3-yl)piperazin-1-yl]ethyl]-4-chlorophenyl]acetic acid,



D. 5,5'-bis[2-[4-(1,2-Benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6,6'-dichloro-3-hydroxy-1,1',3,3'-tetrahydro-2H,2'H-3,3'-biindole-2,2'-dione,



E. 3-(1,2-Benzisothiazol-3-yl)-5-[2-[4-(1,2-Benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6-chloro-1,3-dihydro-2H-indol-2-one.

Ph Eur

Test solution (b) Dilute 2.0 mL of test solution (a) to 20.0 mL with the solvent mixture.

Reference solution (a) Dissolve 27.0 mg of ziprasidone hydrochloride monohydrate CRS in 35 mL of the solvent mixture, sonicate for about 2 min and shake. Dilute 2.0 mL of the solution to 20.0 mL with the solvent mixture.

Reference solution (b) Dissolve 2.5 mg of ziprasidone for system suitability 1 CRS (containing impurities B and C) in a mixture of 40 volumes of water R and 60 volumes of methanol R and dilute to 10.0 mL with the same mixture of solvents.

Reference solution (c) Dilute 1.0 mL of test solution (a) to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

- size: $l = 0.15$ m, $\phi = 3.9$ mm;
- stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
- temperature: 40 °C.

Mobile phase:

- mobile phase A: mix 40 volumes of methanol R and 60 volumes of a 6.8 g/L solution of potassium dihydrogen phosphate R previously adjusted to pH 3.0 with phosphoric acid R;
- mobile phase B: methanol R;

Detection Spectrophotometer at 229 nm.

Flow rate 1.0 mL/min.

Injection 20 μ L of test solution (a) and reference solutions (b) and (c).

Identification of impurities Use the chromatogram supplied with ziprasidone for system suitability 1 CRS and the chromatogram obtained with reference solution (b) to identify the peaks due to impurities B and C.

Relative retention With reference to ziprasidone (retention time = about 6 min): impurity B = about 0.8; impurity C = about 0.9.

System suitability: reference solution (b):

- peak-to-valley ratio: minimum 1.5, where H_p = height above the baseline for the peak due to impurity C and H_v = height separating this peak from the peak due to the baseline for the peak due to impurity C and H_p = height above the baseline of the lowest point of the curve separating this peak from the peak due to ziprasidone.

Calculation of percentage contents:

- for each impurity, use the concentration of ziprasidone in reference solution (c).

Limits:

- impurity B: maximum 0.15 per cent;
- unspecified impurities: for each impurity, maximum 0.10 per cent;
- reporting threshold: 0.05 per cent; disregard any peak with a retention time greater than 40 min.

Time (min)	Mobile phase A (per cent V/V)	Mobile phase B (per cent V/V)
0 - 20	100	0
20 - 21	100 \rightarrow 0	0 \rightarrow 100
21 - 24	0	100

B. Liquid chromatography (2.2.29).

Solvent mixture hydrochloric acid R, water for chromatography R, methanol R (0.04:20:80 V/V/V).

Test solution Dissolve 27 mg of the substance to be examined in 35 mL of the solvent mixture, sonicate for about 2 min and shake. Dilute to 50.0 mL with the solvent mixture.

Reference solution (a) Dissolve the contents of a vial of ziprasidone for system suitability 2 CRS (containing impurities D and E) in 1.0 mL of the solvent mixture.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the solvent mixture. Dilute 1.0 mL of this solution to 10.0 mL with the solvent mixture.

Column:

- size: $l = 0.15$ m, $\phi = 3.9$ mm;
- stationary phase: end-capped octadecylsilyl silica gel for chromatography R (5 μ m);
- temperature: 35 °C.

Mobile phase Mix 8 volumes of methanol R, 42 volumes of acetonitrile R and 50 volumes of a solution prepared as follows: dissolve 3.4 g of potassium dihydrogen phosphate R and 21.6 g of sodium octanesulfonate R in 900 mL of water R, adjust to pH 3.0 with phosphoric acid R and dilute to 1000 mL with water R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 229 nm.

Injection 20 μ L.

Run time 15 times the retention time of ziprasidone.

Identification of impurities Use the chromatogram obtained with reference solution (a) to identify the peaks due to impurities D and E.

Relative retention With reference to ziprasidone (retention time = about 3 min): impurity E = about 2.6; impurity D = about 7.8.

System suitability: reference solution (a):

- resolution: minimum 5.0 between the peaks due to impurities E and D.

Calculation of percentage contents:

- for each impurity, use the concentration of ziprasidone in reference solution (b).

Limits:

- impurity D: maximum 0.15 per cent;
- unspecified impurities: for each impurity, maximum 0.10 per cent;
- reporting threshold: 0.05 per cent; disregard any peak eluting before the peak due to ziprasidone.

Limit:

- total for tests A and B: maximum 0.2 per cent.

Water (2.5.12)

8.5 per cent to 10.1 per cent, determined on 0.050 g.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

Bacterial endotoxins (2.6.14)

Less than 17.5 IU/mg, if intended for use in the manufacture of parenteral preparations without a further appropriate procedure for the removal of bacterial endotoxins.

ASSAY

Liquid chromatography (2.2.29) as described in test A for related substances with the following modifications.

Injection 20 μ L of test solution (b) and reference solution (a).

System suitability: reference solution (a):

- symmetry factor: maximum 1.7 for the peak due to ziprasidone.

Calculate the percentage content of $C_{22}H_{25}ClN_4O_4S_2$ taking into account the assigned content of ziprasidone hydrochloride monohydrate CRS and a conversion factor of 1.133.

STORAGE

Protected from light. If the substance is sterile, store in a sterile, airtight, tamper-proof container.

IMPURITIES

Specified impurities B, D

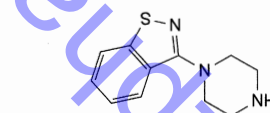
Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph *Substances for pharmaceutical use* (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A, C, E.

By liquid chromatography A, B, C.

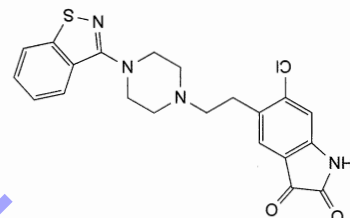
B, D, E.

By liquid chromatography B, D, E.

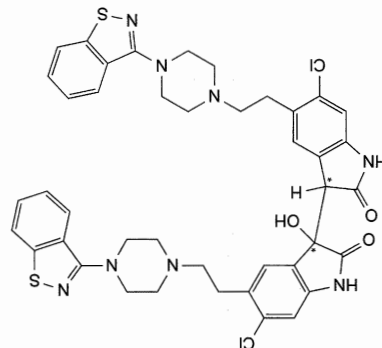
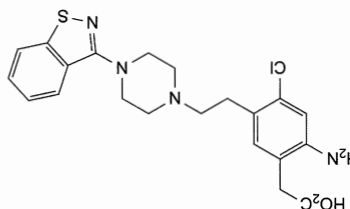
A. 3-piperazin-1-yl-1,2-benzisothiazole,



B. 5-[2-[4-(1,2-benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6-chloro-1H-indole-2,3-dione,



C. 2-[2-amino-5-[2-[4-(1,2-benzisothiazol-3-yl)piperazin-1-yl]ethyl]-4-chlorophenyl]acetic acid,



D. 5,5'-bis[2-[4-(1,2-benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6,6'-dichloro-3-hydroxy-1,1',3,3'-tetrahydro-2H,2'H-3,3'-bimodole-2,2'-dione,

DEFINITION

Bis[N,N-dimethyl-2-[6-methyl-2-(4-methylphenyl)imidazo[1,2-a]pyridin-3-yl]acetamide] (2R,3R)-2,3-dihydroxybutanedioate.

Content

98.5 per cent to 101.0 per cent (anhydrous substance).

CHARACTERS

Appearance

White or almost white, hygroscopic, crystalline powder.

Solubility

Slightly soluble in water, sparingly soluble in methanol, practically insoluble in methylene chloride.

IDENTIFICATION

First identification A, C.

Second identification B, C.

A. Infrared absorption spectrophotometry (2.2.24).

Preparation Dissolve 0.10 g in 10 mL of 0.1 M hydrochloric acid. Add 10 mL of water R. Add dropwise with stirring

1 mL of dilute ammonia R2. Filter and collect the resulting precipitate. Wash the precipitate with water R and then dry at 105 °C for 2 h. Examine the precipitate as a disc.

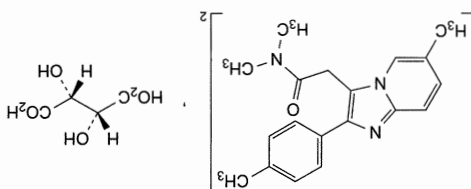
Comparison Repeat the operations using 0.10 g of zolpidem tartrate CRS.

B. Thin-layer chromatography (2.2.27).

Test solution Dissolve 50 mg of the substance to be examined in 5 mL of methanol R, add 0.1 mL of diethylamine R and dilute to 10 mL with methanol R.

Zolpidem Tartrate

(Ph. Eur. monograph 1280)



$C_{22}H_{25}ClN_4O_4S_2$

765

99294-93-6

Action and use

Non-benzodiazepine hypnotic.

Preparation

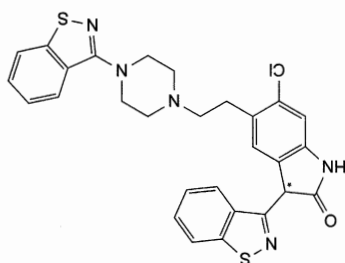
Zolpidem Tablets

Ph Eur



Ph Eur

E. 3-(1,2-benzisothiazol-3-yl)-5-[2-[4-(1,2-benzisothiazol-3-yl)piperazin-1-yl]ethyl]-6-chloro-1,3-dihydro-2H-indol-2-one.



Reference solution (a) Dissolve 50 mg of zolpidem tartrate CRS in 5 mL of methanol R, add 0.1 mL of diethylamine R and dilute to 10 mL with methanol R.

Reference solution (b) Dissolve 50 mg of flunitrazepam CRS in 5 mL of methylene chloride R and dilute to 10 mL with the same solvent. Mix 1 mL of this solution with 1 mL of reference solution (a).

Plate TLC silica gel F₂₅₄ plate R.

Mobile phase dichloromethane R, cyclohexane R, ethyl acetate R (10:45:45 V/V/V).

Application 5 µL.

Development Over 2/3 of the plate.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

System suitability: reference solution (b):

— the chromatogram shows 2 clearly separated spots.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with reference

solution (a).

C. Dissolve about 0.1 g in 1 mL of methanol R heating gently. 0.1 mL of this solution gives reaction (b) of tartrates (2.3.1).

TESTS

Appearance of solution

The solution is clear (2.2.1) and not more intensely coloured than reference solution Y₆ or BY₆ (2.2.2, Method II). Prepare the solutions protected from light and carry out the test as rapidly as possible.

Titrate 0.25 g with 0.125 g of tartaric acid R. Dissolve the mixture in 20 mL of water R and dilute to 25 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29).

Test solution Dissolve 25 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (a) Dissolve 5 mg of zolpidem

impurity A CRS in the mobile phase and dilute to 50.0 mL with the mobile phase.

Reference solution (b) Dissolve 5 mg of the substance to be examined in the mobile phase and dilute to 50.0 mL with the mobile phase. To 10 mL of this solution, add 10 mL of reference solution (a).

Reference solution (c) Dilute 2.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Column:

— size: l = 0.15 m, Ø = 3.9 mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (4 µm).

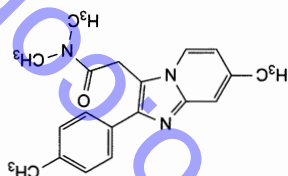
Mobile phase Mix 18 volumes of acetonitrile R, 23 volumes of methanol R and 59 volumes of a 5.6 g/L solution of phosphoric acid R adjusted to pH 5.5 with methylamine R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 254 nm.

Injection 20 µL of the test solution and reference solutions (b) and (c).

Identification of impurities Use the chromatogram obtained with reference solution (b) to identify the peak due to impurity A.



A, N,N-dimethyl-2-[7-methyl-2-(4-methylphenyl)imidazo[1,2-a]pyridin-3-yl]acetamide.

Other detectable impurities (the following substances would, if present at a sufficient level, be detected by one or other of the tests in the monograph. They are limited by the general acceptance criterion for other/unspecified impurities and/or by the general monograph Substances for pharmaceutical use (2034). It is therefore not necessary to identify these impurities for demonstration of compliance. See also 5.10. Control of impurities in substances for pharmaceutical use): A.

IMPURITIES

In an airtight container, protected from light.

STORAGE

1 mL of 0.1 M perchloric acid is equivalent to 38.24 mg of C₂₂H₂₈N₆O₈.

Dissolve 0.300 g in a mixture of 20 mL of anhydrous acetic acid R and 20 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20). Carry out a blank titration.

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 3.0 per cent, determined on 0.50 g.

Water (2.5.12)

(0.05 per cent); disregard any peak due to tartaric acid. the chromatogram obtained with reference solution (c) — disregard limit: 0.25 times the area of the principal peak in (0.2 per cent);

chromatogram obtained with reference solution (c)

— total: not more than the area of the principal peak in the (0.10 per cent);

chromatogram obtained with reference solution (c)

0.5 times the area of the principal peak in the

— unspecified impurities: for each impurity, not more than

Limits:

impurity A and zolpidem.

— resolution: minimum 2.0 between the peaks due to

System suitability: reference solution (b):

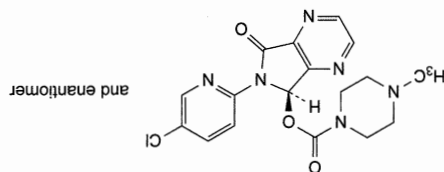
impurity A = about 0.8.

time = about 10 min; tartaric acid = about 0.16;

Relative retention With reference to zolpidem (retention

Zopiclone

(Ph. Eur. monograph 1060)

C₁₇H₁₇ClN₄O₃ 388.8 43200-80-2

Action and use

Non-benzodiazepine hypnotic.

Preparation

Zopiclone Tablets

Ph Eur

DEFINITION

(5R)-6-(5-chloropyridin-2-yl)-7-oxo-6,7-dihydro-5H-pyrido[3,4-b]pyrazin-5-yl 4-methylpiperazine-1-carboxylate.

Content

98.5 per cent to 100.5 per cent.

CHARACTERS

Appearance

White or slightly yellowish powder.

Solubility

Practically insoluble in water, freely soluble in methylene chloride, sparingly soluble in acetone, practically insoluble in ethanol (96 per cent). It dissolves in dilute mineral acids.

mp

About 177 °C, with decomposition.

IDENTIFICATION

First identification B.

Second identification A, C.

A. Ultraviolet and visible absorption spectrophotometry

(2.2.25).

Test solution Dissolve 50.0 mg in a 3.5 g/L solution of hydrochloric acid R and dilute to 100.0 mL with the same solvent. Dilute 2.0 mL of this solution to 100.0 mL with a 3.5 g/L solution of hydrochloric acid R.

Spectral range 220-350 nm.

Absorption maximum At 303 nm.

Specific absorbance at the absorption maximum 340 to 380.

B. Infrared absorption spectrophotometry (2.2.24).

Preparation Discs.

Comparison zopiclone CRS.

C. Thin-layer chromatography (2.2.27).

Test solution Dissolve 10 mg of the substance to be examined in methylene chloride R and dilute to 10 mL with the same solvent.

Reference solution Dissolve 10 mg of zopiclone CRS in methylene chloride R and dilute to 10 mL with the same solvent.

Plate TLC silica gel GF₂₅₄ plate R.

Mobile phase methylamine R, acetone R, ethyl acetate R (2:50:50 V/V/V).

Application 10 µL.

Development Over a path of 15 cm.

Drying In air.

Detection Examine in ultraviolet light at 254 nm.

Results The principal spot in the chromatogram obtained with the test solution is similar in position and size to the principal spot in the chromatogram obtained with the reference solution.

TESTS

Solution S Dissolve 1.0 g in dimethylformamide R and dilute to 20.0 mL with the same solvent.

Appearance of solution

Solution S is not more opalescent than reference suspension II (2.2.1) and not more intensely coloured than appropriate colour (2.2.2, Method II).

Optical rotation (2.2.7)

Dilute 10.0 mL of solution S to 50.0 mL with dimethylformamide R.

Related substances

Liquid chromatography (2.2.29). Prepare the solutions immediately before use.

Test solution Dissolve 40.0 mg of the substance to be examined in the mobile phase and dilute to 10.0 mL with the mobile phase.

Reference solution (a) Dilute 3.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (b) Dilute 1.0 mL of the test solution to 100.0 mL with the mobile phase. Dilute 1.0 mL of this solution to 10.0 mL with the mobile phase.

Reference solution (c) Dissolve 4.0 mg of zopiclone oxide CRS in the mobile phase.

Reference solution (a) in the mobile phase and dilute to 10.0 mL with the mobile phase. To 10.0 mL of this solution, add 1.0 mL of the test solution and dilute to 100.0 mL with the mobile phase.

Column:

— size: $l = 0.25$ m, $\phi = 4.6$ mm;

— stationary phase: octadecylsilyl silica gel for chromatography R (5 µm);

— temperature: 30 °C.

Mobile phase Mix 38 volumes of acetonitrile R and 62 volumes of a solution containing 8.1 g/L of sodium laurylsulfate R and 1.6 g/L of sodium dihydrogen phosphate R adjusted to pH 3.5 with a 10 per cent V/V solution of phosphoric acid R.

Flow rate 1.5 mL/min.

Detection Spectrophotometer at 303 nm.

Injection 20 µL.

Run time 1.5 times the retention time of zopiclone.

Retention time Zopiclone = 27 min to 31 min; if necessary, adjust the concentration of acetonitrile in the mobile phase (increasing the concentration decreases the retention times).

System suitability: reference solution (c):

— resolution: minimum 3.0 between the peaks due to impurity A and zopiclone; if necessary, adjust the mobile phase to pH 4.0 with a 10 per cent V/V solution of phosphoric acid R.

Limits:

— impurities A, B, C: for each impurity, not more than the area of the principal peak in the chromatogram obtained with reference solution (a) (0.3 per cent) and not more than 2 such peaks have an area greater than the area of

the principal peak in the chromatogram obtained with reference solution (b) (0.1 per cent).

2-Propanol

Gas chromatography (2.2.28).

Internal standard solution Dilute 5 mL of ethanol R1 to 100 mL with ethylene chloride R. Dilute 1 mL of this solution to 10 mL with ethylene chloride R.

Test solution Dissolve 0.25 g of the substance to be examined in ethylene chloride R, add 0.5 mL of the internal standard solution and dilute to 5.0 mL with ethylene chloride R.

Reference solution Dilute 4.5 mL of 2-propanol R to 100.0 mL with ethylene chloride R. To 1.0 mL of this solution, add 10.0 mL of the internal standard solution and dilute to 100.0 mL with ethylene chloride R.

Column:

material: fused silica;
size: $l = 10$ m, $\phi =$ about 0.53 mm;
stationary phase: styrene-divinylbenzene copolymer R (film thickness 20 μ m).

Carrier gas helium for chromatography R.

Flow rate 4 mL/min.

Temperature:

Time (min)	Temperature (°C)
0 - 5	50
5 - 10	50 \rightarrow 70
10 - 14	70
14 - 20.5	70 \rightarrow 200
20.5 - 27.5	200
150	
250	

Injection port

Detector

Detection Flame ionisation.

Injection 1 μ L.

Calculate the percentage content m/m of 2-propanol taking its density to be 0.785 g/mL at 20 °C.

Limit:

— 2-propanol: maximum 0.7 per cent m/m.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Sulfated ash (2.4.14)

Maximum 0.1 per cent, determined on 1.0 g.

ASSAY

Dissolve 0.300 g in a mixture of 10 mL of anhydrous acetic acid R and 40 mL of acetic anhydride R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

1 mL of 0.1 M perchloric acid is equivalent to 38.88 mg of $C_{24}H_{27}ClN_2O_5$.

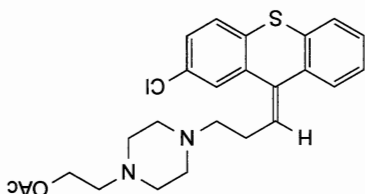
STORAGE

Protected from light.

IMPURITIES

Specified impurities A, B, C.

Zuclopenthixol Acetate



$C_{24}H_{27}ClN_2O_5$

443.0

85721-05-7

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Zuclopenthixol Acetate Injection

DEFINITION

Zuclopenthixol Acetate is (Z)-2-[4-[3-(2-chlorothioxanthen-9-ylidene)propyl]piperazin-1-yl]ethyl acetate. It contains not less than 98.0% and not more than 102.0% of $C_{24}H_{27}ClN_2O_5$, calculated with reference to the dried substance.

CHARACTERISTICS

A yellowish, viscous oil. Very slightly soluble in water; very soluble in dichloromethane, in ethanol (96%) and in ether.

IDENTIFICATION

A. The light absorption of a 0.0015% w/v solution in ethanol (96%), Appendix II B, in the range 210 to 350 nm exhibits two maxima at 230 and 268 nm. The absorbances at the maxima are about 1.18 and 0.51 respectively.
B. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of zuclopenthixol acetate (RS 363).

TESTS

Heavy metals

1.0 g complies with limit test C for heavy metals, Appendix VII (20 ppm). Prepare the standard using 2 mL of lead standard solution (10 ppm Pb).

Related substances
Carry out the method for thin-layer chromatography,

Appendix III A, protected from light, using a silica gel F₂₅₄ precoated plate (Merck silica gel 60 F₂₅₄ plates are suitable), a mixture of 10 volumes of diethylamine, 40 volumes of dichloromethane and 50 volumes of cyclohexane as the mobile phase but using an unlined tank. Apply separately to the plate 4 µL of each of the following solutions. Solutions (1) to (3) contain (1) 0.250% w/v, (2) 0.00025% w/v and (3) 0.000125% w/v respectively of the substance being examined in dichloromethane. Solution (4) contains 0.00050% w/v of 2-chlorothioxanthone BPCRS in dichloromethane. Solution (5) contains 0.000625% w/v of zuclopenthixol hydrochloride BPCRS in dichloromethane containing a few drops of diethylamine. After removal of the plate, allow it to dry in air, spray with a mixture of equal volumes of sulfuric acid and absolute ethanol, heat at 110° for 5 minutes and examine under ultraviolet light (365 nm) immediately. In the chromatogram obtained with solution (1) any spot corresponding to 2-chlorothioxanthone is not more intense than the spot in the chromatogram obtained with solution (4) (0.2%) and any spot corresponding to zuclopenthixol is not more intense than the spot in the chromatogram obtained with solution (5) (0.25%). Any other secondary spot in the chromatogram obtained with solution (1) is not more intense than the spot in the chromatogram obtained with solution (2) (0.1%) and not more than three spots are more intense than the spot in the chromatogram obtained with solution (3) (0.05%).

trans-Isomer

Carry out the method for liquid chromatography, Appendix III D, protected from light, using three solutions in being examined, (2) 0.00046% w/v of trans-clopenthixol acetate dihydrochloride BPCRS (equivalent to 0.00040% w/v of trans-clopenthixol acetate) and (3) 0.020% w/v of the substance being examined and 0.023% w/v of trans-clopenthixol acetate dihydrochloride BPCRS.

The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm × 4.6 mm) packed with silica gel for chromatography (5 µm) (Spherisorb S 5W is suitable), (b) a mixture of 0.03 volume of 13.5M ammonia and 45 volumes of dichloromethane, 45 volumes of heptane and 50 volumes of acetonitrile as the mobile phase with a flow rate of 2 mL per minute and (c) a detection wavelength of 254 nm. Inject 15 µL of each solution. The test is not valid unless the resolution factor between the principal peaks in the chromatogram obtained with solution (3) is at least 2.6. In the chromatogram obtained with solution (1) the area of any peak corresponding to trans-clopenthixol acetate is not greater than the area of the peak in the chromatogram obtained with solution (2) (1%).

Loss on drying

When dried at 60° at a pressure not exceeding 0.7 kPa for 3 hours, loses not more than 0.4% of its weight. Use 1 g.

Sulfated ash

Not more than 0.1%, Appendix IX A.

ASSAY

Dissolve 0.2 g in 50 mL of anhydrous acetic acid and carry out Method I for non-aqueous titration, Appendix VIII A, determining the end point potentiometrically. Each mL of 0.1M perchloric acid VS is equivalent to 22.15 mg of C₂₄H₂₇ClN₂O₂S.

Yellow, viscous, oily liquid.

Appearance

CHARACTERS

98.0 per cent to 102.0 per cent (dried substance).

Content

9-ylidene]propyl]piperazin-1-yl]ethyl decanoate.

2-[4-[3-[(9Z)-2-chloro-9H-thioxanthene-

DEFINITION

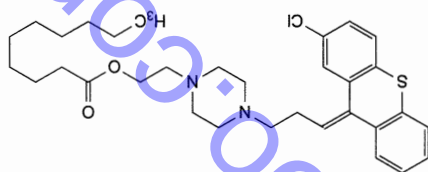
Zuclopenthixol Decanoate Injection

Preparation

Dopamine receptor antagonist; neuroleptic.

Action and use

C₂₄H₂₇ClN₂O₂S 555.2 64053-00-5

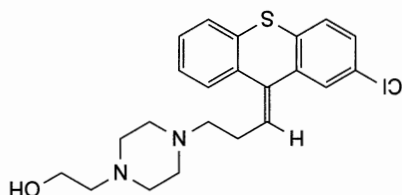


(Ph. Eur. monograph 1707)

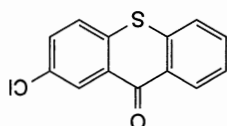
Zuclopenthixol Decanoate



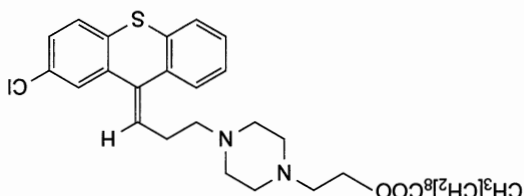
C. zuclopenthixol.



B. 2-chlorothioxanthone,



A. trans-clopenthixol acetate (trans-isomer),



IMPURITIES

Zuclopenthixol Acetate should be protected from light and stored at a temperature not exceeding -20°.

STORAGE

Solubility

Very slightly soluble in water, very soluble in ethanol (96 per cent) and in methylene chloride.

IDENTIFICATION

Infrared absorption spectrophotometry (2.2.24).

Comparison Ph. Eur. reference spectrum of zuclopenthixol decanoate.

TESTS**Appearance of solution**

The solution is clear (2.2.1).

Using an ultrasonic bath, dissolve 1.0 g in ethanol (96 per cent) R and dilute to 20.0 mL with the same solvent.

Related substances

Liquid chromatography (2.2.29). Carry out the test protected from light and prepare the solutions immediately before use.

Solution A Dissolve 8.89 g of docusate sodium R in water R, stirring for about 6–8 h, and dilute to 1000 mL with the same solvent.

Test solution Dissolve 25.0 mg of the substance to be examined in acetonitrile R and dilute to 100.0 mL with the same solvent.

Reference solution (a) Dilute 1.0 mL of the test solution to 100.0 mL with acetonitrile R.

Reference solution (b) Dissolve 5.0 mg of zuclopenthixol impurity B CRS in acetonitrile R and dilute to 100.0 mL with the same solvent. Dilute 5.0 mL of this solution to 100.0 mL with acetonitrile R.

Reference solution (c) Dissolve the contents of a vial of zuclopenthixol for system suitability CRS (zuclopenthixol decanoate containing impurities A, B and C) in 1 mL of methanol R.

Column:

— size: $l = 0.25$ m, $\varnothing = 4.6$ mm;

— stationary phase: spherical end-capped octadecylsilyl silica gel for chromatography R (5 μ m);

— temperature: 40 °C.

Mobile phase Mix 25 volumes of solution A and 75 volumes of anhydrous ethanol R, then add 0.1 volumes of phosphoric acid R.

Flow rate 1.0 mL/min.

Detection Spectrophotometer at 270 nm.

Injection 20 μ L.

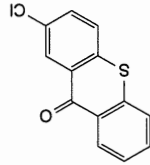
Run time Twice the retention time of zuclopenthixol decanoate.

Identification of impurities Use the chromatogram supplied with zuclopenthixol for system suitability CRS and the chromatograms obtained with reference solutions (b) and (c) to identify the peaks due to impurities A, B and C.

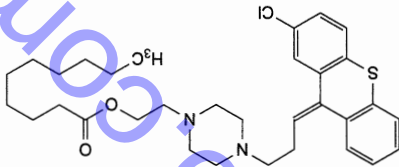
Relative retention With reference to zuclopenthixol decanoate (retention time = about 12 min): impurity C = about 0.4; impurity B = about 0.5; impurity A = about 1.1.

System suitability: reference solution (c):

— **peak-to-valley ratio:** minimum 2.0, where H_p = height above the baseline of the peak due to impurity C and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to impurity B; and minimum 2.5, where H_p = height above the baseline of the peak due to impurity A and H_o = height above the baseline of the lowest point of the curve separating this peak from the peak due to zuclopenthixol decanoate.



B. 2-chloro-9H-thioxanthene-9-one,



A. 2-[4-[3-[(9E)-2-chloro-9H-thioxanthene-9-ylidene]propyl]piperazin-1-yl]ethyl decanoate,

IMPURITIES

Under an inert gas in an airtight container, protected from light, at a temperature not exceeding –20 °C.

STORAGE

1 mL of 0.1 M perchloric acid is equivalent to 27.76 mg of $C_{32}H_{43}ClN_2O_2S$.

Dissolve 0.250 g in 50 mL of anhydrous acetic acid R. Titrate with 0.1 M perchloric acid, determining the end-point potentiometrically (2.2.20).

ASSAY

Maximum 0.1 per cent, determined on 1.0 g.

Sulfated ash (2.4.14)

Maximum 0.5 per cent, determined on 1.000 g by drying in an oven at 60 °C at a pressure not exceeding 0.7 kPa for 3 h.

Loss on drying (2.2.32)

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

Heavy metals (2.4.8)

Maximum 20 ppm.

1.0 g complies with test C. Prepare the reference solution using 2 mL of lead standard solution (10 ppm Pb) R.

— **total:** not more than 1.5 times the area of the principal peak in the chromatogram obtained with reference solution (a) (1.5 per cent);

— **disregard limit:** 0.05 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.05 per cent).

— **unspecified impurities:** for each impurity, not more than 0.1 times the area of the principal peak in the chromatogram obtained with reference solution (a) (0.10 per cent);

— **reference solution (a)** (0.3 per cent);

— **reference solution (b)** (0.2 per cent);

— **reference solution (c)** (0.3 per cent);

— **reference solution (d)** (0.2 per cent);

— **reference solution (e)** (0.2 per cent);

— **reference solution (f)** (0.2 per cent);

— **reference solution (g)** (0.2 per cent);

— **reference solution (h)** (0.2 per cent);

— **reference solution (i)** (0.2 per cent);

— **reference solution (j)** (0.2 per cent);

— **reference solution (k)** (0.2 per cent);

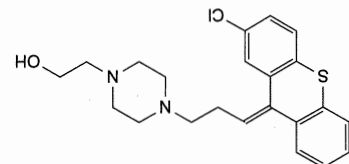
— **reference solution (l)** (0.2 per cent);

— **reference solution (m)** (0.2 per cent);

— **reference solution (n)** (0.2 per cent);

— **reference solution (o)** (0.2 per cent);

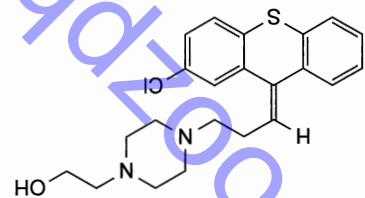
— **reference solution (p)** (0.2 per cent);



C. 2-[4-[3-[(9Z)-2-chloro-9H-thioxanthene-9-ylidene]propyl]piperazin-1-yl]ethanol.

Ph Eur

Zuclopenthixol Hydrochloride



$C_{22}H_{25}ClN_2OS \cdot 2HCl$ 473.9 633-59-0

Action and use

Dopamine receptor antagonist; neuroleptic.

Preparation

Zuclopenthixol Tablets

DEFINITION

Zuclopenthixol Hydrochloride is (Z)-2-4-[3-

(2-chlorothioxanthene-9-ylidene)propyl]piperazin-1-ylethanol dihydrochloride. It contains not less than 98.0% and not more than 101.0% of $C_{22}H_{25}ClN_2OS \cdot 2HCl$, calculated with reference to the anhydrous substance.

CHARACTERISTICS

An off-white, granular powder.

Very soluble in water; sparingly soluble in ethanol (96%); very slightly soluble in ether.

IDENTIFICATION

A. The light absorption of a 0.0015% w/v solution in ethanol (96%), Appendix II B, in the range 210 to 350 nm exhibits two maxima at 230 and 268 nm. The absorbances at the maxima are about 1.05 and 0.42, respectively.

B. The infrared absorption spectrum, Appendix II A, is concordant with the reference spectrum of zuclopenthixol hydrochloride (RS 365).

C. Yields reaction A characteristic of chlorides, Appendix VI.

TESTS

Acidity

pH of a 1% w/v solution, 2.0 to 3.0, Appendix V L.

Heavy metals

1.0 g complies with limit test C for heavy metals, Appendix VII

solution (10 ppm Pb).

Related substances

Carry out the method for thin-layer chromatography, Appendix III A, protected from light, using a TLC silica gel F_{254} plate (Merck plates are suitable), a mixture of 10 volumes of ethyl acetate and 90 volumes of cyclohexane as the mobile phase but using an unlined tank and allowing the mobile phase to ascend 10 cm above the line of application.

Apply separately to the plate 4 μ L of each of the following solutions. Solutions (1) to (3) contain (1) 2.5% w/v, (2) 0.0025% w/v and (3) 0.00125% w/v respectively of the substance being examined in methanol containing a few drops of diethylamine. Solution (4) contains 0.0025% w/v of 2-chlorothioxanthone BPCRS in methanol. Solution (5) contains 0.0025% w/v of 9-allyl-2-chlorothioxanthene-9-ol BPCRS in methanol. After removal of the plate, allow it to dry in air, spray with a mixture of equal volumes of sulfuric acid and absolute ethanol, heat at 110° for 5 minutes and examine under ultraviolet light (365 nm) immediately. In the chromatogram obtained with solution (1) any spots corresponding to 2-chlorothioxanthone and to 9-allyl-2-chlorothioxanthene are not more intense than the spots in the chromatograms obtained with solutions (4) and (5) respectively (0.1% each), any other secondary spot is not more intense than the spot in the chromatogram obtained with solution (2) (0.1%) and not more than four such spots are more intense than the spot in the chromatogram obtained with solution (3) (0.05%).

Free amine

Carry out the method for thin-layer chromatography, Appendix III A, protected from light, using a silica gel F_{254} precoated plate (Merck silica gel 60 F_{254} plates are suitable) and a mixture of 2 volumes of water, 10 volumes of 13.5M ammonia, 20 volumes of butan-1-ol and 65 volumes of acetone as the mobile phase. Apply separately to the plate 4 μ L of each of the following solutions. Solution (1) is a 0.25% w/v solution of the substance being examined in ethanol (96%); add a few drops of diethylamine to aid dissolution. For solution (2), dilute 1 volume of solution (1) to 400 volumes with ethanol (96%). After removal of the plate, allow it to dry in air, spray with a mixture of equal volumes of sulfuric acid and absolute ethanol, heat at 110° for 5 minutes and examine under ultraviolet light (365 nm) immediately. In the chromatogram obtained with solution (1) any secondary spot of the same colour and at an R_f value lower than that of the principal spot is not more intense than the spot in the chromatogram obtained solution (2) (0.25%).

trans-Isomer

Carry out the method for liquid chromatography, Appendix III D, protected from light, using the following solutions. For solution (1) dissolve 10 mg of the substance being examined in 5 mL of water, add 25 mL of the mobile phase, shake thoroughly for 1 minute and dilute to 50 mL with the mobile phase; use the upper layer. For solution (2) prepare a 0.004% w/v solution of trans-clopenthixol hydrochloride BPCRS in water, shake 5 mL of this solution with 25 mL of the mobile phase for 1 minute and dilute to 50 mL with the mobile phase; use the upper layer. For solution (3) dissolve 10 mg of the substance being examined in 5 mL of a 0.004% w/v solution of trans-clopenthixol hydrochloride BPCRS, add 25 mL of the mobile phase, shake thoroughly for 1 minute, dilute to 50 mL with the mobile phase and use the upper layer. The chromatographic procedure may be carried out using (a) a stainless steel column (25 cm \times 4.6 mm) packed with silica gel for chromatography (5 μ m) (Sphersorb S 5W is suitable), (b) a mixture of 0.2 volume of water, 0.3 volume of 13.5M ammonia, 15 volumes of propan-2-ol and 85 volumes of n-heptane as the mobile phase with a flow rate of 1.5 mL per minute and (c) and a detection wavelength of 254 nm. The test is not valid unless the chromatogram obtained with solution (3) shows a similar resolution to that in the reference chromatogram supplied with trans-clopenthixol hydrochloride BPCRS.

In the chromatogram obtained with solution (1) the area of any peak corresponding to *trans*-clopenthixol hydrochloride is not greater than 1.5 times the area of the peak in the chromatogram obtained with solution (2) (3%).

Water

Not more than 2.5%, determined on 0.5 g, Appendix IX C.

Sulfated ash

Not more than 0.1%, Appendix IX A.

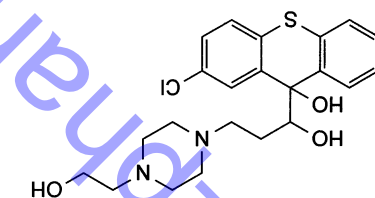
ASSAY

Dissolve 0.5 g in 50 mL of anhydrous acetic acid and carry out Method I for non-aqueous titration, Appendix VIII A, using crystal violet solution as indicator. Each mL of 0.1M perchloric acid VS is equivalent to 23.70 mg of $C_{22}H_{27}Cl_3N_2OS$.

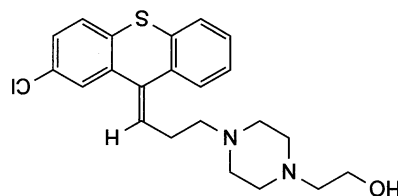
STORAGE

Zuclopenthixol Hydrochloride should be protected from light.

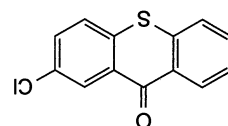
IMPURITIES



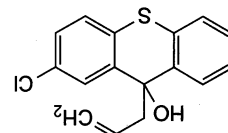
A. 2-chloro-9-(1-hydroxy-3-(4-(2-hydroxyethyl)piperazin-1-yl)propyl)thioxanthene-9-ol,



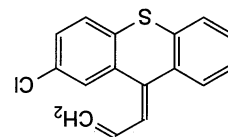
B. *trans*-clopenthixol,



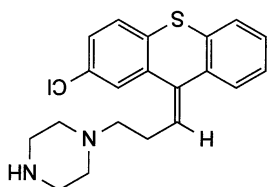
C. 2-chlorothioxanthone,



D. 9-allyl-2-chlorothioxanthene-9-ol,



E. 9-allylidene-2-chlorothioxanthene,



F. 2-chloro-9-(3-piperazin-1-ylpropylidene)thioxanthene.